

# Other chromatin capture/targeted sequencing

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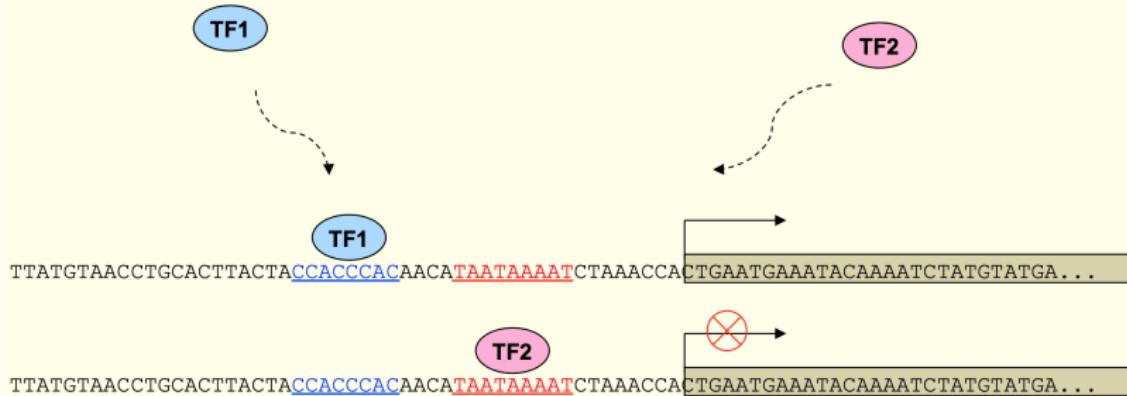
2021-04-19

# Transcription factor binding regulates gene expression

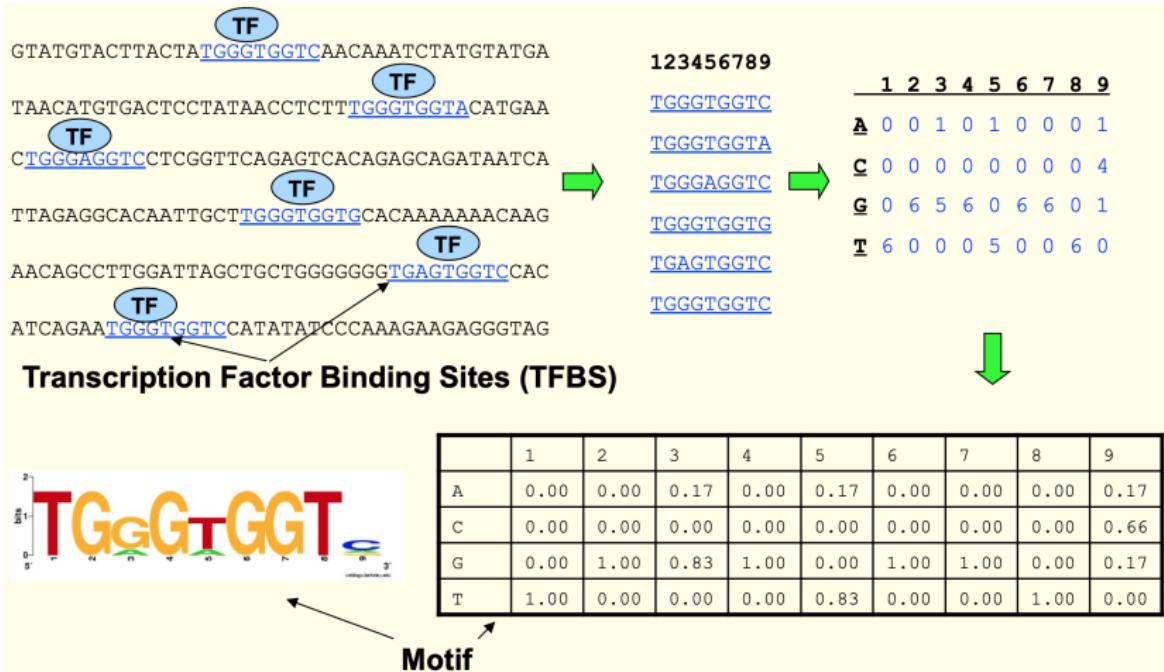
Transcription factors (TF):



Transcription factor binding sites (TFBS): CCACCCAC, TAATAAAAT



# Transcription factors recognize specific motifs

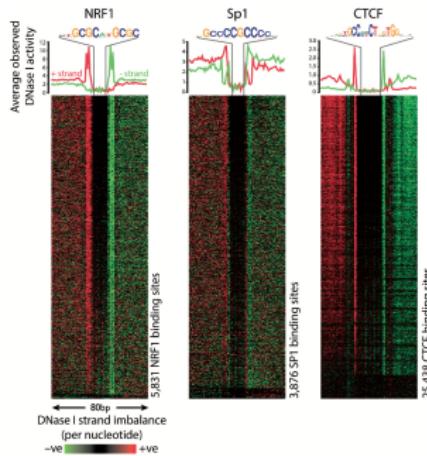


# Other “captured/targeted” sequencing technologies

- Enrich and then sequence selected genomic regions.
  - **DNase-seq, MNase-seq, FAIRE-seq, ATAC-seq:** detect open chromatin sites.
  - **CUT&RUN, CUT&TAG:** improved ChIP-seq profiling.
  - **MeDIP-seq:** measure methylated DNA.
  - **GRO-seq:** map the position, amount and orientation of transcriptionally engaged RNA polymerases.
  - **Ribo-seq:** detect ribosome occupancy on mRNA. This is captured RNA-seq.

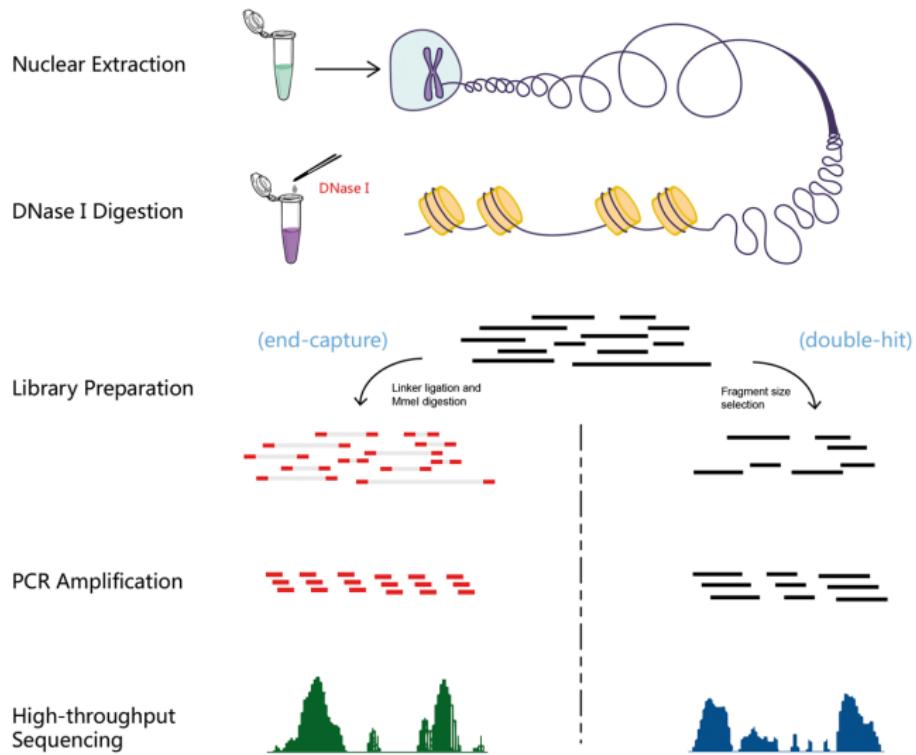
# DNAse-seq

- A widely used approach in gene regulation studies uses DNase I as a tool to identify DNase I Hypersensitive Sites (DHSs) within chromatin
- DHSs represent open chromatin regions that are normally only accessible at sites of active regulatory elements such as transcriptional enhancers

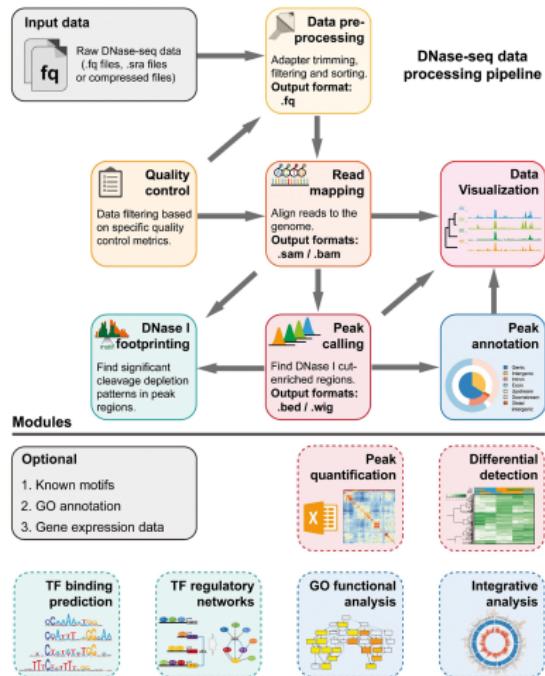


Cockerill,P.N. (2011) Structure and function of active chromatin and DNase I hypersensitive sites. FEBS J., 278, 2182–2210.

# Overview of DNase-seq experimental protocols



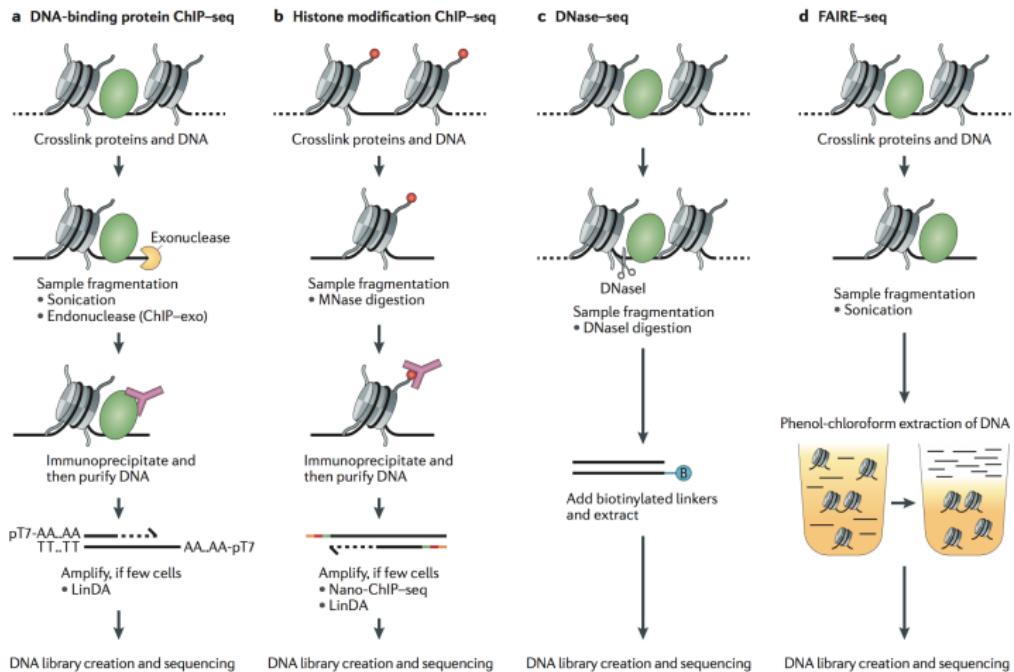
# General analysis pipeline for sequence-tag experiments



## Other technologies to assess open chromatin

- **FAIRE-seq** - Formaldehyde-Assisted Isolation of Regulatory Elements followed by sequencing.
- **MNase-seq** - Micrococcal Nuclease-assisted assessment of open chromatin.
- **ATAC-seq** - Assay for transposase- accessible chromatin using sequencing. Tn5 transposase is used to transpose sequencing adapters into the genomic DNA

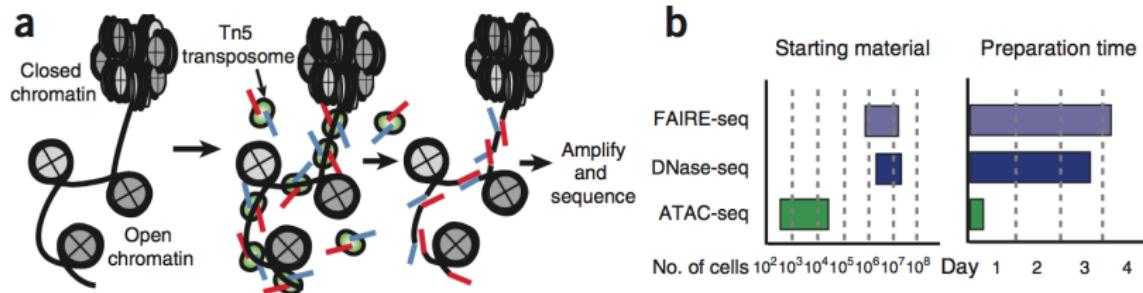
# Comparison of experimental protocols



<https://zhonglab.gitbook.io/3dgenome/chap2-experiment-tools-for-exploring-genome-interaction/2.2-primary-order>

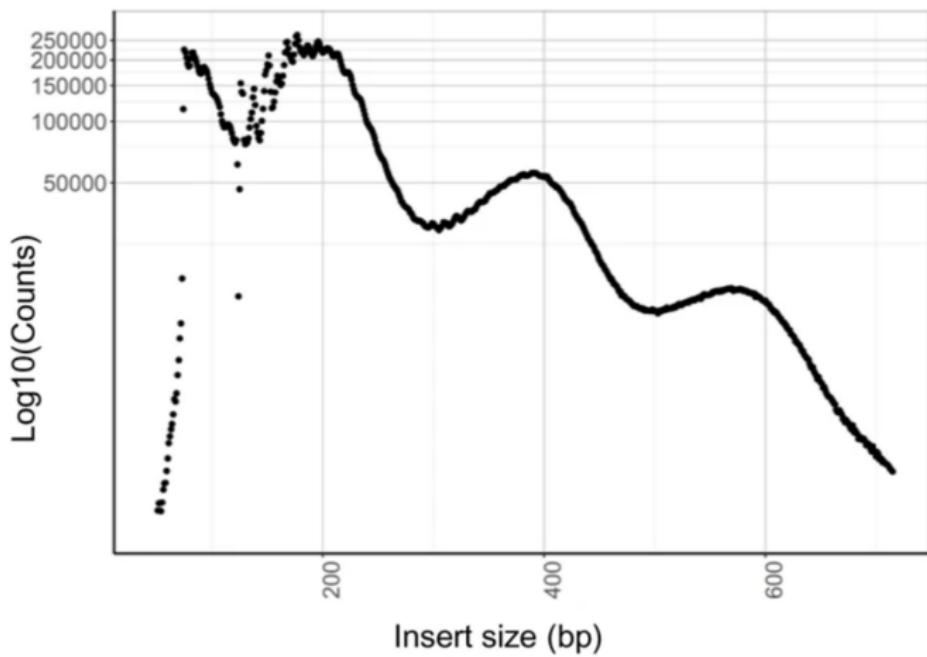
# ATAC-seq: finding open chromatin regions

- ATAC-seq is an ensemble measure of open chromatin that uses the prokaryotic Tn5 transposase to tag regulatory regions by inserting sequencing adapters into accessible regions of the genome



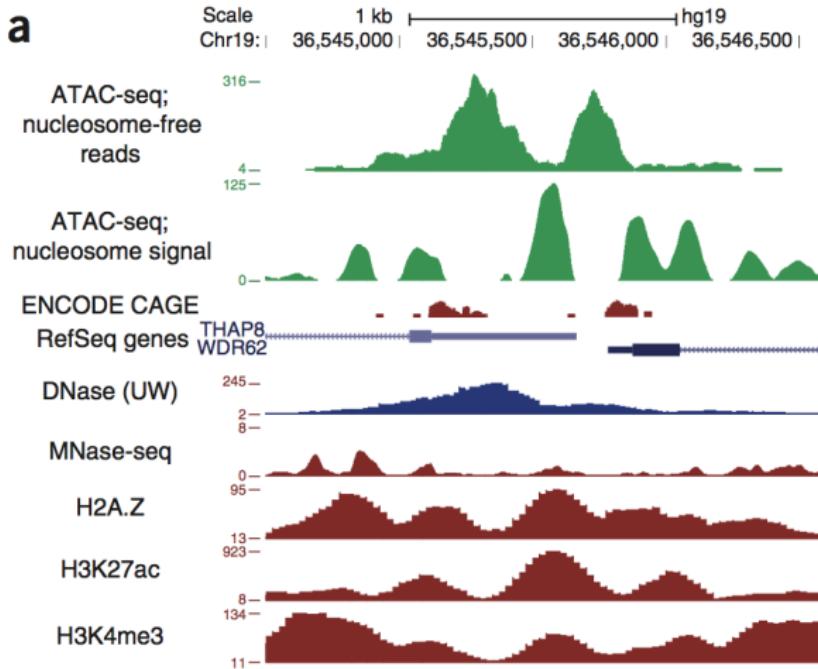
Jason D Buenrostro et al., "Transposition of Native Chromatin for Fast and Sensitive Epigenomic Profiling of Open Chromatin, DNA-Binding Proteins and Nucleosome Position," Nature Methods 10, no. 12 (December 2013): 1213–18, <https://doi.org/10.1038/nmeth.2688>.

# ATAC-seq: revealing nucleosome positioning



<https://www.nature.com/articles/s41598-019-44076-8>

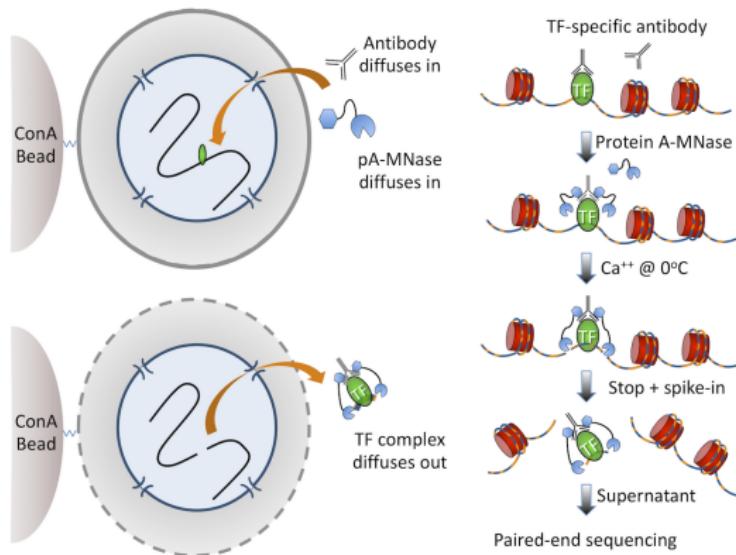
# ATAC-seq: finding open chromatin regions



Jason D Buenrostro et al., "Transposition of Native Chromatin for Fast and Sensitive Epigenomic Profiling of Open Chromatin, DNA-Binding Proteins and Nucleosome Position," Nature Methods 10, no. 12 (December 2013): 1213–18,  
<https://doi.org/10.1038/nmeth.2688>.

# CUT&RUN

- Cleavage Under Targets and Release Using Nuclease
- Antibody-targeted controlled cleavage by micrococcal nuclease



<https://data.4dnucleome.org/experiment-types/cut-n-run/>

# CUT&TAG

- Cleavage Under Targets and Tagmentation
- Tn5 transposase conjugated with adapters inserts them directly into cut sequences

CUT&Tag vs. CUT&RUN vs. ChIP-Seq

|                                    | CUT&Tag  | CUT&RUN  | ChIP-Seq   |
|------------------------------------|--|--|--|
| Performed Under Native Conditions? | Yes  | Yes  | No   |
| Chromatin Fragmentation Method     | Tn5-based tagmentation   | MNase digestion  | Sonication   |
| Cell Number Requirements           | 5,000-500,000 cells  | 500,000 cells  | 1-10 million cells   |
| Sequencing Depth Required *        | 2 million reads  | 8 million reads  | 20-50 million reads  |
| Integrated Library Preparation?    | Yes, uses tagmentation   | No, separate library prep required   | No, separate library prep required   |
| Compatible Targets                 | Primarily histone modifications, some transcription factors and co-factors | Wide range of histone modifications, transcription factors, and co-factors | Wide range of histone modifications, transcription factors, and co-factors |
| Workflow Length                    | 1-2 days   | 1-2 days   | 2-3 days   |

\* Kaya-Okur et al. *Nature Communications* (2019) 10:1930

## More

- ChIP-seq analysis notes from Ming Tang,  
<https://github.com/crazyhottommy/ChIP-seq-analysis>
- Notes on ChIP-seq and other-seq-related tools,  
[https://github.com/mdozmorov/ChIP-seq\\_notes](https://github.com/mdozmorov/ChIP-seq_notes)