An R package for analyses of bioassays and probit graphs

08 July 2021

Summary

Dose-response relationships (also known as exposure—response relationships) reflect the effects of a substance (most of the time a xenobiotic or a chemical) on organisms (populations, tissues or cells). Dose-response analyses are widely used in broad research areas, from medicine and physiology to vector control and pest management in agronomy. Further, reporting the response of organisms to stressors is an essential component of many public policies (e.g., health, environment). An ideal example is monitoring of resistance to xenobiotics. Since the 1950s, xenobiotics (e.g., insecti-, pesti-, fungicides) have been widely used to control populations of vectors or pests. As a response, resistance mechanisms have been selected in targeted populations, undermining their efficiency. Establishing and comparing the resistance level of various populations to various xenobiotics is at the core of world health organization (WHO) recommendations in order to define / adjust vector control strategies. It is usually done by exposing batches of individuals (adults or larvae) to varying doses of the xenobiotic to assess their responses (mortality or knock-down effect). Despite the availability of statistical approaches for such analyses, there had been a lack of easily accessible analytical infrastructure for it (the traditionally-used software Probit ran in Basic and several labs kept an old computer for it). In 2013, we developed an R script with a robust statistical background to ease the dose-mortality relationship analysis. It has been used in many studies (e.g., Alout et al. (2016); pocquet2014; Badolo et al. (2019); Assogba et al. (2016); Yaméogo et al. (2021); Epelboin et al. (2021); Perrier et al. (2021), and is now recommended as good practice by the ANSES (the French national agency for health and environment safety). In order to make it even more user friendly, we have now developed it into an R package called 'BioRssay' with more flexibility and improved presentation of results.

Statement of need

'BioRssay' is a comprehensive compilation of scripts in R language (Core and Team 2020) designed to analyze dose-response relationships (or exposure-response: mortality, knock-down effect, etc.) from bioassays of one or more strains, lines, populations (but also, cells etc.). This package provides a complete analytic workflow from data quality assessment to statistical analyses and data visualization. In the first steps, base-mortality in the controls (i.e., mortality linked to the experiment itself, not the exposure) is taken into account by adjusting the data following Abott's correction (Abbott and others 1925). Data are then analyzed using a generalized linear model (probit-link function) to generate mortality-dose regressions (which take overdispersion into account and allow for mortality of 0 or 1). Linearity of the log-dose response for each population is then tested using a chi-square test between model predictions and observed data (significant deviations from linearity may reflect mixed populations for example). By default, doses lethal for 25%, 50% and 95% of the populations (LD25, LD50 and LD95 respectively) are computed with their 95% confidence intervals (CI), following Johnson et al. (2013) approach, which allows taking the heterogeneity of the data into account (Finney 1971). Probit analysis. Cambridge: Cambridge University Press. 350 p.). Otherwise, the user has the option to specify any LD level. Likelihood ratio tests (LRT) are then implemented to test for statistical significance of the differences in response between different populations/strains, and when necessary, Holm-Bonferroni method (Holm 1979). is performed to control the family-wise error inherent to multiple testing. Finally, the resistance ratios for the required LD(s) (RR25, RR50 and RR95, by default). i.e., the LD(s) for a given population divided by the LD(s) of the population with the lowest one (usually the susceptible reference), are calculated according to Robertson and Preisler (n.d.), with 95% confidence intervals. Customizable plots of the probit-transformed regressions are also drawn (e.g., with or without the desired confidence intervals).

Citations

Acknowledgements

We want to thank Jérôme Chopard, Haoues Alout, Mylène Weill and Nicole Pasteur for their valuable comments on earlier versions of the script and its outputs.

References

Abbott, Walter S, and others. 1925. "A Method of Computing the Effectiveness of an Insecticide." *J. Econ. Entomol* 18 (2): 265–67.

Alout, Haoues, Pierrick Labbé, A Berthomieu, Patrick Makoundou, Philippe Fort, N Pasteur, and Mylene Weill. 2016. "High Chlorpyrifos Resistance in Culex Pipiens Mosquitoes: Strong Synergy Between Resistance Genes." *Heredity* 116 (2): 224–31.

Assogba, Benoît S, Pascal Milesi, Luc S Djogbénou, Arnaud Berthomieu, Patrick Makoundou, Lamine S Baba-Moussa, Anna-Sophie Fiston-Lavier, Khalid Belkhir, Pierrick Labbe, and Mylène Weill. 2016. "The Ace-1 Locus Is Amplified in All Resistant Anopheles Gambiae Mosquitoes: Fitness Consequences of Homogeneous and Heterogeneous Duplications." *PLoS Biology* 14 (12): e2000618.

Badolo, Athanase, Aboubacar Sombié, Patricia M Pignatelli, Aboubakar Sanon, Félix Yaméogo, Dimitri W Wangrawa, Antoine Sanon, Hirotaka Kanuka, Philip J McCall, and David Weetman. 2019. "Insecticide Resistance Levels and Mechanisms in Aedes Aegypti Populations in and Around Ouagadougou, Burkina Faso." *PLoS Neglected Tropical Diseases* 13 (5): e0007439.

Core, R, and R Team. 2020. "A Language and Environment for Statistical Computing, R Foundation for Statistical Computing; 2013."

Epelboin, Yanouk, Lanjiao Wang, Quentin Giai Gianetto, Valérie Choumet, Pascal Gaborit, Jean Issaly, Amandine Guidez, et al. 2021. "CYP450 Core Involvement in Multiple Resistance Strains of Aedes Aegypti from French Guiana Highlighted by Proteomics, Molecular and Biochemical Studies." *Plos One* 16 (1): e0243992.

Finney, David John. 1971. "Probit Analysis."

Holm, Sture. 1979. "A Simple Sequentially Rejective Multiple Test Procedure." Scandinavian Journal of Statistics, 65–70.

Johnson, Reed M, Lizette Dahlgren, Blair D Siegfried, and Marion D Ellis. 2013. "Acaricide, Fungicide and Drug Interactions in Honey Bees (Apis Mellifera)." *PloS One* 8 (1): e54092.

Perrier, Stéphane, Eléonore Moreau, Caroline Deshayes, Marine El-Adouzi, Delphine Goven, Fabrice Chandre, and Bruno Lapied. 2021. "Compensatory Mechanisms in Resistant Anopheles Gambiae Acerkis and Kdrkis Neurons Modulate Insecticide-Based Mosquito Control." Communications Biology 4 (1): 1–16.

Robertson, JL, and HK Preisler. n.d. "Pesticide Bioassays with Arthropods. 1992." CRC Boca Raton, FL.

Yaméogo, Félix, Dimitri Wendgida Wangrawa, Aboubacar Sombié, Antoine Sanon, and Athanase Badolo. 2021. "Insecticidal Activity of Essential Oils from Six Aromatic Plants Against Aedes Aegypti, Dengue Vector from Two Localities of Ouagadougou, Burkina Faso." Arthropod-Plant Interactions, 1–8.