
Generalist Equivariant Transformer

Towards 3D Molecular Interaction Learning

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Abstract

Many processes in biology and drug discovery involve various 3D interactions between molecules, such as protein and protein, protein and small molecule, etc. Given that different molecules are usually represented in different granularity, existing methods usually encode each type of molecules independently with different models, leaving it defective to learn the various underlying interaction physics. In this paper, we first propose to universally represent an arbitrary 3D complex as a geometric graph of sets, shedding light on encoding all types of molecules with one model. We then propose a Generalist Equivariant Transformer (GET) to effectively capture both domain-specific hierarchies and domain-agnostic interaction physics. To be specific, GET consists of a bilevel attention module, a feed-forward module and a layer normalization module, where each module is E(3) equivariant and specialized for handling sets of variable sizes. Notably, in contrast to conventional pooling-based hierarchical models, our GET is able to retain fine-grained information of all levels. Extensive experiments on the interactions between proteins, small molecules and RNA/DNAs verify the effectiveness and generalization capability of our proposed method across different domains.

1. Introduction

Molecular interactions (Tomasi & Persico, 1994), which describe attractive or repulsive forces between molecules

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and between non-bonded atoms, are crucial in the research of chemistry, biochemistry and biophysics, and come as foundation processes of various downstream applications, including drug discovery, material design, etc (Sapoval et al., 2022; Tran et al., 2023; Vamathevan et al., 2019). There are different types of molecular interactions, and this paper mainly focuses on the ones that exist in bimolecular complexes, consisting of proteins, small molecules or RNA/DNAs. Specifically, to better capture their physical effects, we study molecular interactions via 3D geometry where atom coordinates are always provided.

Modeling molecular interaction relies heavily on appropriate representation of molecules. Recent studies apply Graph Neural Networks (GNNs) for this purpose (Gilmer et al., 2017; Jin et al., 2018). This is motivated by the fact that graphs naturally represent molecules, by considering atoms as nodes and inter-atom interactions or bonds as edges. When further encapsulating 3D atom coordinates, geometric graphs (Gasteiger et al., 2020b; Schütt et al., 2017; Stärk et al., 2022) are used in place of conventional graph modeling that solely encodes topology. To process geometric graphs, equivariant GNNs, a new kind of GNNs that meet E(3) equivariance regarding translation, rotation and reflection are proposed, which exhibit promising performance in molecule interaction tasks (Kong et al., 2022b; Luo et al., 2022; Townshend et al., 2020; Zhang et al., 2022a).

Despite the encouraging progress, there still lacks a desirable and unified form of cross-domain molecular representation in molecular interaction. The molecules of different domains like small molecules, proteins, and RNA/DNAs are usually represented in different granularity, which consist of atoms, residues, and nucleobases, respectively. Existing approaches typically design domain-specific representations and model each of the interacting instances independently (Somnath et al., 2021; Wang et al., 2022a), which are defective in learning the various underlying interaction physics. Therefore, designing unified cross-domain molecular representation is demanded, which, however, is non-trivial. For one thing, directly applying unshared block-level graphs, whose nodes correspond to domain-specific building blocks, leads to limited transferability of the model from one domain to another. For another, decomposing all molecules

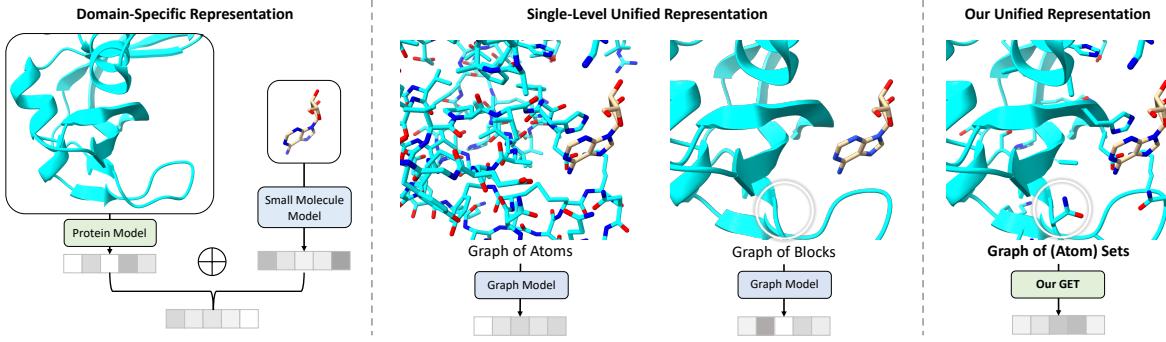


Figure 1. Domain-specific representations and unified representations in molecular interaction.

into atom-level graphs discards the block specificity (*e.g.* which residue each atom belongs to) and overlook valuable heuristics for representation learning. It is still an open problem in designing a universal representation and a generalist model thereon to capture both the block-level specificity and atom-level shareability.

In this paper, we tackle this problem by modeling a complex involved in molecular interaction as *a geometric graph of sets*. This representation follows a bilevel design: in the top level, a complex is represented as a geometric graph of blocks; in the bottom level, each block contains a set of atomic instances. It is nontrivial to process such bilevel geometric graphs, as the model should handle blocks of variable sizes and ensure certain specific geometries. To this end, we propose *Generalist Equivariant Transformer* (*GET*), which consists of the three modules: bilevel attention module, feed-forward module and layer normalization module. To be specific, the bilevel attention module updates the information of each atom by adopting both sparse block-level and dense atom-level attentions. The feed-forward module is to inject the intra-block geometry to each atom, and the layer normalization module is proposed to stabilize and accelerate the training. All the modules are $E(3)$ -equivariant regarding the 3D coordinates, permutation-invariant regarding all atoms within each block, and work regardless of the varying block size. We compare our method with other representation approaches in Figure 1.

Notably, our formulation of graph of sets is relevant to conventional pooling-based hierarchical models based on graph of graphs (Jin et al., 2022). Nevertheless, these hierarchical architecture are usually inefficient and will blot out the fine-grained information after certain pooling-based aggregation, while our *GET* is able to retain both the atom-level and block-level information.

We conduct experiments on various molecular interactions between proteins, small molecules and RNA/DNAs. The results exhibit the superiority of our *GET* on the proposed unified representation over traditional methods including domain-specific independent models, single-level unified

representations and hierarchical models. More excitingly, we identify strong potential of *GET* in capturing and transferring shared knowledge across different domains, and enabling zero-shot performance on RNA/DNA-ligand binding affinity prediction.

2. Related Work

Molecular Interaction and Representation Various types of molecules across different domains (Du et al., 2016; Elfiky, 2020; Jones & Thornton, 1996) can form interactions, the strength of which are usually measured by the energy gap between the unbound and bound states of the molecules (*i.e.* affinity) (Gilson et al., 1997). We primarily investigate interactions between two proteins (Jones & Thornton, 1996) and between a protein and a small molecule (Du et al., 2016), both of which are widely explored in the machine learning community (Kong et al., 2022a; Luo et al., 2023; 2022; Somnath et al., 2021; Stärk et al., 2022; Wang et al., 2022a). Furthermore, we embark on a pioneering effort to involve RNA/DNAs, which is a challenging endeavor due to the limited availability of such data. Small molecules are usually represented by graphs where nodes are atoms (Atz et al., 2021; Hoogeboom et al., 2022; Xu et al., 2022; Zaidi et al., 2022), but there are also explorations on subgraph-level decomposition of molecules by mining motifs (Geng et al., 2023; Jin et al., 2018; Kong et al., 2022b). Proteins are built upon residues, which are predefined sets of atoms (Richardson, 1981), and thus have mainly two categories of representations according to the granularity of graph nodes: atom-level and residue-level. Atom-level representations, as the name suggests, decompose proteins into single atoms (Townshend et al., 2020) and discard the hierarchy of proteins. Residue-level representations either exert pooling on the atoms (Jin et al., 2022), or directly use residue-specific features (Anand & Achim, 2022; Shi et al., 2022; Somnath et al., 2021; Wang et al., 2022a) which is limited to proteins. Similarly, RNA/DNAs also have atom-level and nucleobase-level representations (Avsec et al., 2021; Watson & Crick, 1953). Despite the differences in building blocks, the basic units (*i.e.* atoms) are shared across

different molecular domains, and so do the fundamental interaction physics. Therefore, it is valuable to construct a unified representation for different molecular domains, which is explored in this paper.

Equivariant Network Equivariant networks integrate the symmetry of 3D world, namely E(3)-equivariance, into the models, and thus are widely used in geometric learning (Han et al., 2022). A line of methods rely on preprocessing 3D coordinates into invariant features, *e.g.* pairwise distances (Schütt et al., 2017; Choukroun & Wolf, 2021), angles (Gasteiger et al., 2020b;a; 2021; Liu et al., 2021b), to achieve invariant outputs. More recent works also keep track of equivariant features to achieve stronger expressivity (Joshi et al., 2023), either via scalarization (Schütt et al., 2021; Thölke & De Fabritiis, 2022; Du et al., 2023) or irreducible representations (Thomas et al., 2018; Batzner et al., 2022; Liao & Smidt, 2022; Batatia et al., 2022; Musaelian et al., 2023). Our work is inspired by multi-channel equivariant graph neural networks (Huang et al., 2022; Kong et al., 2022b) which assign each node with a coordinate matrix. However, they require a fixed number of channels (*i.e.* constant number of rows in the coordinate matrix) and lack invariance w.r.t to the permutations of the coordinates, which limits their application here as each building block is an unordered set of atoms with variable size. Moreover, the node features are still limited to single vector form (Thölke & De Fabritiis, 2022; Liao & Smidt, 2022), which is unable to accommodate all-atom representations in single blocks. In contrast, our proposed model is designed to handle geometric graphs of sets where each node contains an unordered set of 3D instances with a different size, which fits perfectly with the concept of building blocks in molecules.

3. Method

We start by illustrating the proposed unified representation for molecules in § 3.1. Then we introduce GET in § 3.2. Each layer of GET consists of the three types of E(3)-equivariant modules: a bilevel attention module, a feed-forward module, and a layer normalization after each previous module. The overall concepts are depicted in Figure 2 and the detailed scheme is presented in Appendix B.

3.1. Unified Representation: Geometric Graphs of Sets

Graphs come as a central tool for molecular representations, and different kind of graphs is applied in different case. For instance, small molecules can be represented as single-level graphs, where each node is an atom, while proteins (RNA/DNAs) correspond to two-level graphs, where each node is a residue (nucleobase) that consists of a variable number of atoms. To better characterize the interaction between different molecules, below we propose a unified molecular representation.

Given a complex consisting of a set of atoms \mathbb{A} , we first identify a set of blocks (*i.e.* subsets) from \mathbb{A} according to some predefined notions (*e.g.* residues for proteins). Then the complex is abstracted as a geometric graph of sets $\mathcal{G} = (\mathcal{V}, \mathcal{E})$, where $\mathcal{V} = \{(\mathbf{H}_i, \vec{\mathbf{X}}_i) | 1 \leq i \leq B\}$ includes all B blocks and $\mathcal{E} = \{(i, j, e_{ij}) | 1 \leq i, j \leq B\}$ includes all edges between blocks¹, where $e_{ij} \in \mathbb{R}^{d_e}$ distinguishes the type of the edge as intra-molecular or inter-molecular connection. In each block composed of n_i atoms, $\mathbf{H}_i \in \mathbb{R}^{n_i \times d_h}$ denotes a set of atom feature vectors and $\vec{\mathbf{X}}_i \in \mathbb{R}^{n_i \times 3}$ denotes a set of 3D atom coordinates. To be specific, the p -th row of \mathbf{H}_i , which is the feature vector of atom p , sums up the trainable embeddings of atom types $a_i[p]$, block types b_i , and atom position codes $p_i[p]$ (see Appendix A), namely, $\mathbf{H}_i[p] = \text{Embed}(a_i[p]) + \text{Embed}(b_i) + \text{Embed}(p_i[p])$, $1 \leq p \leq n_i$. To reduce the computational complexity, we construct \mathcal{E} via k-nearest neighbors ($k = 9$) according to the block distance which is defined as the minimum distance between inter-block atom pairs:

$$d(i, j) = \min\{\|\vec{\mathbf{X}}_i[p] - \vec{\mathbf{X}}_j[q]\|_2 \mid 1 \leq p \leq n_i, 1 \leq q \leq n_j\}. \quad (1)$$

Overall, the block-level geometry derived from atomic interactions defines the connectivity of the graph, while the atom-level instances compose the unordered matrix-form node features with variable sizes. As we will observe in the next section, the above bilevel design allows our model to capture sparse interactions for the top level and dense interactions for the bottom level, achieving a desirable integration of different granularities. We will also demonstrate in the experiments that the representation can be easily extended to arbitrary block definitions (*e.g.* subgraph-decomposition of small molecules (Kong et al., 2022b; Geng et al., 2023)).

Connection to Single-Level Representations By restricting the blocks to one-atom subsets, we can obtain the **atom-level** representation where each node is one atom, and correspondingly both \mathbf{H}_i and $\vec{\mathbf{X}}_i$ are downgraded to row vectors as $n_i \equiv 1$. If we retain the building blocks but replace \mathbf{H}_i and $\vec{\mathbf{X}}_i$ with their centroids, then we obtain the **block-level** representation where the atoms in the same block are pooled into one single instance. Both single-level representations assign a vector and a 3D coordinate to each node, hence can be fed into most structural learning models (Gasteiger et al., 2020b; Satorras et al., 2021; Schütt et al., 2017; Thölke & De Fabritiis, 2022). On the contrary, the proposed bilevel representation requires the capability of processing E(3)-equivariant feature matrices (\mathbf{H}_i and $\vec{\mathbf{X}}_i$) with a variable number of rows, which cannot be directly processed by existing models. Additionally, within each block, the rows of \mathbf{H}_i and $\vec{\mathbf{X}}_i$ are indeed elements in a set and their update

¹We have added self-loops to reflect self-interactions between the atoms in each block.

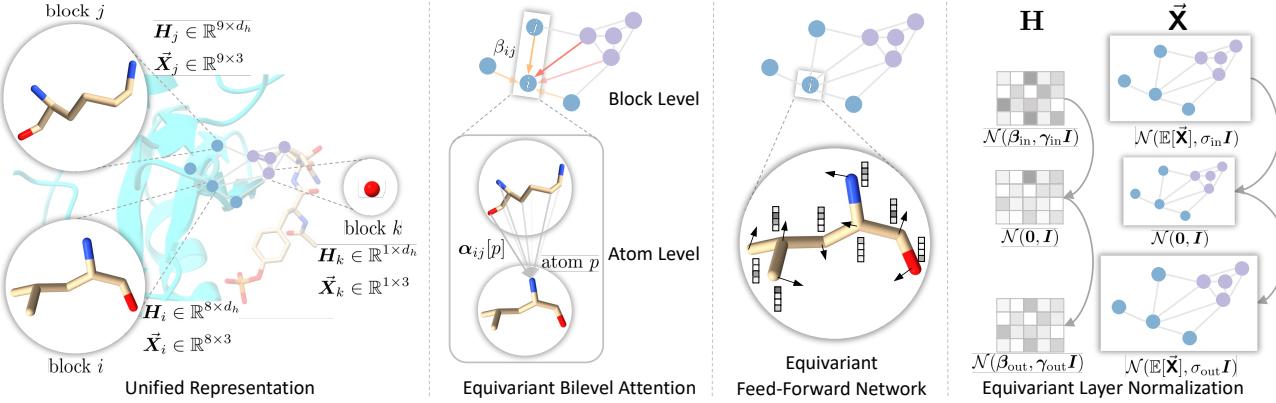


Figure 2. Overview of the unified representation and the equivariant modules in our Generalist Equivariant Transformer (GET). From left to right: The unified representation treats molecules as geometric graphs of sets according to predefined building blocks; The bilevel attention module captures both sparse block-level and dense atom-level interactions via an equivariant attention mechanism; The feed-forward network injects the block-level information into the intra-block atoms; The layer normalization transforms the input distribution with trainable scales and offsets.

should be unaffected by the row order. Luckily, the above challenges are well handled by our Generalist Equivariant Transformer proposed in the next section.

Comparison with Hierarchical Representations Previous studies (Jin et al., 2022) model proteins in a hierarchical manner, where the atom-level features within each residue are first pooled as the residue-level features that will be processed via the message passing over the graph of residues. In contrast to these hierarchical methods, our bilevel representation retains the information of both the atom-level and residue-level features simultaneously for attention-based message passing. Namely, the proposed matrix-form representation inherently preserves more spatial information than the hierarchical counterparts, since the fine-grained atomic distances are explicitly retained throughout the entire message passing process.

3.2. Generalist Equivariant Transformer

Upon the unified representation, we propose GET to model the structure of the input complex. As mentioned above, one beneficial property of GET is that it can tackle blocks of variable sizes emphi.e. matrix-form representations with variable shapes), while conventional explorations focus on vector-form features with fixed sizes. Besides, GET is sophisticatedly designed to ensure E(3)-equivariance and intra-block permutation invariance to handle the symmetry. Specifically, each layer of GET first exploits an equivariant bilevel attention module to capture both sparse interactions in block level and dense interactions in atom level. Then an equivariant feed-forward module updates each atom with the geometry of its block. Finally, a novel equivariant layer normalization is implemented on both the hidden states and coordinates. We present a detailed scheme of GET in Appendix B.

Equivariant Bilevel Attention Module Given two blocks \$i\$ and \$j\$ of \$n_i\$ and \$n_j\$ atoms, respectively, we first obtain the query, the key, and the value matrices as follows:

$$\mathbf{Q}_i = \mathbf{H}_i \mathbf{W}_Q, \quad \mathbf{K}_j = \mathbf{H}_j \mathbf{W}_K, \quad \mathbf{V}_j = \mathbf{H}_j \mathbf{W}_V, \quad (2)$$

where \$\mathbf{W}_Q, \mathbf{W}_K, \mathbf{W}_V \in \mathbb{R}^{d_h \times d_r}\$ are trainable parameters. We denote \$\vec{\mathbf{X}}_{ij} \in \mathbb{R}^{n_i \times n_j \times 3}\$ and \$\mathbf{D}_{ij} \in \mathbb{R}^{n_i \times n_j}\$ as the relative coordinates and distances between any atom pair in block \$i\$ and \$j\$, namely, \$\vec{\mathbf{X}}_{ij}[p, q] = \vec{\mathbf{X}}_i[p] - \vec{\mathbf{X}}_j[q], \mathbf{D}_{ij}[p, q] = \|\vec{\mathbf{X}}_{ij}[p, q]\|_2\$.

The **atom-level cross attention values** from \$j\$ to \$i\$ are calculated by:

$$\mathbf{R}_{ij}[p, q] = \phi_A(\mathbf{Q}_i[p], \mathbf{K}_j[q], \text{RBF}(\mathbf{D}_{ij}[p, q]), e_{ij}), \quad (3)$$

$$\alpha_{ij} = \text{Softmax}(\mathbf{R}_{ij} \mathbf{W}_A). \quad (4)$$

Here, \$e_{ij}\$ is the optional edge feature to distinguish between intra-molecule edges and inter-molecule edges; \$\phi_A\$ is a 2-layer Multi-Layer Perceptron (MLP) with SiLU (Hendrycks & Gimpel, 2016) activation; RBF (Gasteiger et al., 2020b) embeds the distance with radial basis functions (definition in Appendix B); \$\mathbf{R}_{ij} \in \mathbb{R}^{n_i \times n_j \times d_r}\$ represents the relations between each atom pair in block \$i\$ and \$j\$, which are later mapped to scalars by \$\mathbf{W}_A \in \mathbb{R}^{d_r \times 1}\$ to obtain the atom-level cross attentions \$\alpha_{ij} \in \mathbb{R}^{n_i \times n_j}\$ between the two blocks through Softmax along the columns of \$\mathbf{R}_{ij} \mathbf{W}_A \in \mathbb{R}^{n_i \times n_j}\$.

The **block-level cross attention value** from \$j\$ to \$i\$ is given by the following equations:

$$r_{ij} = \frac{1}{n_i n_j} \sum_{p=1}^{n_i} \sum_{q=1}^{n_j} \mathbf{R}_{ij}[p, q], \quad (5)$$

$$\beta_{ij} = \frac{\exp(r_{ij} \mathbf{W}_B)}{\sum_{j \in \mathcal{N}(i)} \exp(r_{ij} \mathbf{W}_B)}, \quad (6)$$

where $\mathbf{W}_B \in \mathbb{R}^{d_r \times 1}$, and $\mathcal{N}(i)$ denotes the neighborhood blocks of i . Basically, $\mathbf{r}_{ij} \in \mathbb{R}^{d_r}$ represents the global relation between i and j after aggregating all values in \mathbf{R}_{ij} , which is then mapped to a scalar to obtain the block-level cross attentions β_{ij} through Softmax in $\mathcal{N}(i)$.

With the atom-level and the block-level attentions at hand, we are ready to update both the hidden states and coordinates for each atom p in block i :

$$\mathbf{m}_{ij,p} = \alpha_{ij}[p] \cdot \phi_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])) \quad (7)$$

$$\vec{\mathbf{m}}_{ij,p} = \alpha_{ij}[p] \cdot (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p]))) \quad (8)$$

$$\mathbf{H}'_i[p] = \mathbf{H}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \phi_m(\mathbf{m}_{ij,p}), \quad (9)$$

$$\vec{\mathbf{X}}'_i[p] = \vec{\mathbf{X}}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} (\sigma_m(\mathbf{m}_{ij,p}) \cdot \vec{\mathbf{m}}_{ij,p}), \quad (10)$$

where, \parallel specifies the concatenation along the second dimension; ϕ_v, ϕ_m, σ_v , and σ_m are all MLPs, ϕ_v and σ_v are applied for each row of the input matrix independently; \odot computes the element-wise multiplication. It is verified that the shape of the updated variables \mathbf{H}'_i and $\vec{\mathbf{X}}'_i$ keeps the same regardless of the value of the block size n_j . In addition, since the attentions α_{ij} and β_{ij} are E(3)-invariant, the update of $\vec{\mathbf{X}}'_i$ is E(3)-equivariant. It can also be observed that the update is independent to the atom permutation of each block. We provide detailed proofs in Appendix C.

Equivariant Feed-Forward Network This module updates \mathbf{H}_i and $\vec{\mathbf{X}}_i$ for each atom individually. We denote each row of \mathbf{H}_i as \mathbf{h} , and $\vec{\mathbf{X}}_i$ as $\vec{\mathbf{x}}$. We first calculate the centroids of the block:

$$\mathbf{h}_c = \text{centroid}(\mathbf{H}_i), \quad \vec{\mathbf{x}}_c = \text{centroid}(\vec{\mathbf{X}}_i). \quad (11)$$

Then we obtain the relative coordinate $\Delta\vec{\mathbf{x}}$ between each atom and the centroid to derive corresponding distance representation \mathbf{r} :

$$\Delta\vec{\mathbf{x}} = \vec{\mathbf{x}} - \vec{\mathbf{x}}_c, \quad \mathbf{r} = \text{RBF}(\|\Delta\vec{\mathbf{x}}\|_2), \quad (12)$$

The centroids and the distance representation are subsequently integrated into the updating process of \mathbf{h} and $\vec{\mathbf{x}}$ to let each atom be aware of the geometric context of its block, where ϕ_h, σ_x are MLPs:

$$\mathbf{h}' = \mathbf{h} + \phi_h(\mathbf{h}, \mathbf{h}_c, \mathbf{r}), \quad (13)$$

$$\vec{\mathbf{x}}' = \vec{\mathbf{x}} + \Delta\vec{\mathbf{x}} \sigma_x(\mathbf{h}, \mathbf{h}_c, \mathbf{r}), \quad (14)$$

Equivariant Layer Normalization Layer normalization is known to stabilize and accelerate the training of deep neural networks (Ba et al., 2016; Vaswani et al., 2017). The challenge here is that we need to additionally consider E(3)-equivariance when normalizing the coordinates. To this

end, we first extract the centroid of the entire graph as $\mathbb{E}[\vec{\mathbf{X}}]$, where $\vec{\mathbf{X}}$ collects the coordinates of all atoms in all blocks. Then we exert layer normalization on the hidden vectors and coordinates of individual atoms as follows:

$$\mathbf{h}' = \frac{\mathbf{h} - \mathbb{E}[\mathbf{h}]}{\sqrt{\text{Var}[\mathbf{h}]}} \cdot \gamma + \beta, \quad (15)$$

$$\vec{\mathbf{x}}' = \frac{\vec{\mathbf{x}} - \mathbb{E}[\vec{\mathbf{X}}]}{\sqrt{\text{Var}[\vec{\mathbf{X}} - \mathbb{E}[\vec{\mathbf{X}}]]}} \cdot \sigma + \mathbb{E}[\vec{\mathbf{X}}], \quad (16)$$

where γ, β , and σ are learnable parameters, and $\text{Var}[\vec{\mathbf{X}}]$ calculates the variation of all atom coordinates with respect to the centroid. Therefore, the coordinates, after subtracting the centroid of all atoms, are first normalized to standard Gaussian distribution and then scaled with σ before recovering the centroid. In addition, to further reflect the rescaling of the coordinates into hidden features, we inject the following update before applying the above layer normalization:

$$\mathbf{h} = \mathbf{h} + \phi_{LN}(\text{RBF}(\sigma / \sqrt{\text{Var}[\vec{\mathbf{X}}]})), \quad (17)$$

where ϕ_{LN} is an MLP. In contrast to existing literature which only implements layer normalization on E(3)-invariant features (Thölke & De Fabritiis, 2022; Liao & Smidt, 2022) or node-wise velocities (Zaidi et al., 2022), ours works on both E(3)-invariant features and E(3)-equivariant coordinates.

Thanks to the E(3)-equivariance of each module, GET, which is the cascading of these modules in each layer, also conforms to the symmetry of the 3D world. We provide the proof in Appendix C and complexity analysis in Appendix D.

4. Experiments

In this section, we aim to answer the following three questions via empirical experiments: (1) Does modeling complexes with unified representation better captures the geometric interactions than treating each interacting entity independently with domain-specific representations (§ 4.1)? (2) Is the proposed unified representation more expressive than vanilla single-level representations or pooling-based hierarchical methods (§ 4.2)? (3) Can the proposed method generalize to different domains by learning the various underlying interaction physics (§ 4.3)?

We conduct experiments on prediction of binding between proteins, small molecules and nucleic acids. Thus, we adopt three widely used metrics for quantitative evaluation (Townshend et al., 2020; Liu et al., 2021a; Luo et al., 2023; Notin et al., 2022): **RMSE** is the Root Mean Square Error of the predicted value; **Pearson Correlation** (Cohen et al., 2009) measures the linear correlation between the predicted values and the target values; **Spearman Correlation** (Hauke

Table 1. The mean and the standard deviation of three runs on the PDBbind benchmark. The best results are marked in bold and the second best are underlined. The results of baselines are borrowed from Wang et al. (2022a). Baselines encoding the complexes with one model are marked with *.

Model	RMSE \downarrow	Pearson \uparrow	Spearman \uparrow
DeepDTA (Öztürk et al., 2018)	1.866 ± 0.080	0.472 ± 0.022	0.471 ± 0.024
Bepler and Berger (Bepler & Berger, 2019)	1.985 ± 0.006	0.165 ± 0.006	0.152 ± 0.024
TAPE (Rao et al., 2019)	1.890 ± 0.035	0.338 ± 0.044	0.286 ± 0.124
ProtTrans (Elnaggar et al., 2022)	1.544 ± 0.015	0.438 ± 0.053	0.434 ± 0.058
MaSIF (Gainza et al., 2020)	1.484 ± 0.018	0.467 ± 0.020	0.455 ± 0.014
IEConv (Hermosilla et al., 2020)	1.554 ± 0.016	0.414 ± 0.053	0.428 ± 0.032
Holoprot-Full Surface (Somnath et al., 2021)	1.464 ± 0.006	0.509 ± 0.002	0.500 ± 0.005
Holoprot-Superpixel (Somnath et al., 2021)	1.491 ± 0.004	0.491 ± 0.014	0.482 ± 0.032
ProNet-Amino Acid (Wang et al., 2022a)	1.455 ± 0.009	0.536 ± 0.012	0.526 ± 0.012
ProNet-Backbone (Wang et al., 2022a)	1.458 ± 0.003	0.546 ± 0.007	0.550 ± 0.008
ProNet-All-Atom (Wang et al., 2022a)	1.463 ± 0.001	0.551 ± 0.005	0.551 ± 0.008
GVP* (Jing et al., 2021)	1.594 ± 0.073	-	-
Atom3D-3DCNN* (Townshend et al., 2020)	<u>1.416 ± 0.021</u>	0.550 ± 0.021	<u>0.553 ± 0.009</u>
Atom3D-ENN* (Townshend et al., 2020)	1.568 ± 0.012	0.389 ± 0.024	0.408 ± 0.021
Atom3D-GNN* (Townshend et al., 2020)	1.601 ± 0.048	0.545 ± 0.027	0.533 ± 0.033
GET* (ours)	1.364 ± 0.009	0.596 ± 0.006	0.573 ± 0.007

& Kossowski, 2011) measures the correlation between the rankings given by the predicted and the target values

4.1. Comparison to Domain-Specific Representations

We evaluate our method on the prediction of binding affinity between proteins and small molecules against state-of-the-art two-branch models with domain-specific representations from existing literature (Somnath et al., 2021; Wang et al., 2022a). We follow Somnath et al. (2021); Wang et al. (2022a) to conduct experiments on the well-established PDBbind (Wang et al., 2004; Liu et al., 2015) and split the dataset (4,709 biomolecular complexes) according to sequence identity of the protein with 30% as the threshold. Details of the experiments are provided in Appendix F.

Results Table 1 shows that our GET surpasses the baselines by a large margin. Compared to the baselines which encode proteins and small molecules independently with delicately designed domain-specific models, our unified representation enables unified geometric learning with only one model, which better captures the interactive geometric information between the protein and the small molecule. Notably, among models with one encoder (Jing et al., 2021; Townshend et al., 2020), our method also achieves significant improvement since our unified representation retains domain-specific hierarchies instead of decomposing all types of molecules into atomic graphs.

4.2. Comparison to Vanilla Unified Representations

Next, we compare the proposed unified representation with three vanilla unified representations: (1) **Block**-level methods assign each building block to one node where the definition of building block is domain-specific (*e.g.* each residue

in the proteins is one node); (2) **Atom**-level methods treats all kinds of molecules as graphs of atoms; (3) **Hierarchical** methods first implement message passing on atom-level graphs, then obtain the block-level representations by pooling for further message passing on the block-level graphs (Jin et al., 2022).

Baselines These vanilla unified representations are compatible with most geometric graph models in existing literature, thus we adopt the following representative models as backbone for comparison. **SchNet** (Schütt et al., 2017), **DimeNet++** (Gasteiger et al., 2020b;a), and **GemNet** (Gasteiger et al., 2021) build invariant models based on invariant geometric features (*i.e.* distances and angles). **EGNN** (Satorras et al., 2021), TorchMD-Net (**ET**) (Thölke & De Fabritiis, 2022), and **LEFTNet** (Du et al., 2023) keep track of equivariant features and are implemented directly on 3D coordinates via scalarization (Han et al., 2022). **MACE** (Batatia et al., 2022) and **Equiformer** (Liao & Smidt, 2022) leverage spherical harmonics and irreducible representations (Thomas et al., 2018) to compose equivariant models.

Dataset To this end, we evaluate the models on prediction of protein-protein affinity and ligand-binding affinity. For **Protein-Protein Affinity** (PPA), we adopt the Protein-Protein Affinity Benchmark Version 2 (Kastritis et al., 2011; Vreven et al., 2015) as the test set, which categorizes 176 diversified protein-protein complexes into three difficulty levels (*i.e.* Rigid, Medium, Flexible) according to the conformation change of the proteins from the unbound to the bound state (Kastritis et al., 2011). The Flexible split is the most challenging as the proteins undergo large conformation change upon binding. As for training, we obtain

Table 2. The mean and the standard deviation of three runs on PPA and LBA prediction. The best results are marked in bold and the second best are underlined. Baselines that fail to process atomic graphs due to high complexity are marked with OOM (out of memory).

Repr.	Model	PPA		LBA		
		Pearson \uparrow	Spearman \uparrow	RMSE \downarrow	Pearson \uparrow	Spearman \uparrow
Block	SchNet	0.439 \pm 0.016	0.427 \pm 0.012	1.406 \pm 0.020	0.565 \pm 0.006	0.549 \pm 0.007
	DimeNet++	0.323 \pm 0.025	0.317 \pm 0.031	1.391 \pm 0.020	0.576 \pm 0.016	0.569 \pm 0.016
	EGNN	0.381 \pm 0.021	0.382 \pm 0.022	1.409 \pm 0.015	0.566 \pm 0.010	0.548 \pm 0.012
	ET	0.424 \pm 0.021	0.415 \pm 0.027	1.367 \pm 0.037	0.599 \pm 0.017	0.584 \pm 0.025
	GemNet	0.387 \pm 0.023	0.393 \pm 0.027	1.393 \pm 0.036	0.569 \pm 0.027	0.553 \pm 0.026
	MACE	0.470 \pm 0.015	0.466 \pm 0.011	1.385 \pm 0.006	0.599 \pm 0.010	0.580 \pm 0.014
	Equiformer	0.484 \pm 0.007	<u>0.496 \pm 0.007</u>	1.350 \pm 0.019	0.604 \pm 0.013	0.591 \pm 0.012
	LEFTNet	0.452 \pm 0.013	0.452 \pm 0.013	1.377 \pm 0.013	0.588 \pm 0.011	0.576 \pm 0.010
Atom	SchNet	0.369 \pm 0.007	0.404 \pm 0.016	1.357 \pm 0.017	0.598 \pm 0.011	0.592 \pm 0.015
	DimeNet++	OOM	OOM	1.439 \pm 0.036	0.547 \pm 0.015	0.536 \pm 0.016
	EGNN	0.302 \pm 0.010	0.349 \pm 0.009	1.358 \pm 0.000	0.599 \pm 0.002	0.587 \pm 0.004
	ET	0.401 \pm 0.005	0.436 \pm 0.004	1.381 \pm 0.013	0.591 \pm 0.007	0.583 \pm 0.009
	GemNet	OOM	OOM	OOM	OOM	OOM
	MACE	0.463 \pm 0.052	0.449 \pm 0.052	1.411 \pm 0.029	0.579 \pm 0.009	0.563 \pm 0.012
	Equiformer	OOM	OOM	OOM	OOM	OOM
	LEFTNet	0.448 \pm 0.046	0.431 \pm 0.046	1.343 \pm 0.004	0.610 \pm 0.004	0.598 \pm 0.003
Hierarchical	SchNet	0.438 \pm 0.017	0.424 \pm 0.016	1.370 \pm 0.028	0.590 \pm 0.017	0.571 \pm 0.028
	DimeNet++	OOM	OOM	1.388 \pm 0.010	0.582 \pm 0.009	0.574 \pm 0.007
	EGNN	0.386 \pm 0.021	0.390 \pm 0.016	1.380 \pm 0.015	0.586 \pm 0.004	0.568 \pm 0.004
	ET	0.401 \pm 0.005	0.438 \pm 0.029	1.383 \pm 0.009	0.580 \pm 0.008	0.564 \pm 0.004
	GemNet	OOM	OOM	OOM	OOM	OOM
	MACE	0.466 \pm 0.020	0.470 \pm 0.016	1.372 \pm 0.021	0.612 \pm 0.010	0.592 \pm 0.010
	Equiformer	OOM	OOM	OOM	OOM	OOM
	LEFTNet	0.445 \pm 0.024	0.446 \pm 0.029	1.366 \pm 0.016	0.592 \pm 0.014	0.580 \pm 0.011
Unified	GET (ours)	0.514 \pm 0.011	0.533 \pm 0.011	<u>1.327 \pm 0.005</u>	<u>0.620 \pm 0.004</u>	<u>0.611 \pm 0.003</u>
	GET-PS (ours)	-	-	<u>1.309 \pm 0.012</u>	<u>0.633 \pm 0.008</u>	<u>0.642 \pm 0.009</u>

2,500 complexes with annotated binding affinity (K_i or K_d) from PDBbind (Wang et al., 2004) and split the dataset according to sequence identity on a threshold of 30%. For **Ligand-Binding Affinity** (LBA), we use the LBA dataset and its splits in Atom3D benchmark (Townshend et al., 2020), where there are 3507, 466, and 490 complexes in the training, the validation, and the test sets. Although this dataset overlaps with the PDBbind benchmark in terms of the included complexes, the different data processing methods lead to deviations in model performance from the PDBbind benchmark. Thus, for fair comparison with previous literature, we follow their settings to use PDBbind in § 4.1 and Atom3D here. Details are provided in Appendix F.

Results We report the mean and the standard deviation of the metrics across 3 runs for PPA and LBA in Table 2. Details on different difficulty levels for PPA are included in Appendix N due to the space limit. Inspiringly, it reads that our GET with the proposed unified representation achieves significantly better performance compared with the baselines with either single-level representations or hierarchical pooling, no matter the interacting partners are macro molecules (*i.e.* proteins) or small molecules. This confirms the superiority of our method, which comes from a desirable integration of different granularities. Further, to show the flexibility of the proposed unified representation, as men-

tioned in § 3.1, we add **GET-PS**, which defines the blocks in small molecules as principal subgraphs (Kong et al., 2022b) instead of atoms. GET-PS receives obvious gains over GET since fragments in small molecules usually contribute to interactions as a whole (Hajduk & Greer, 2007).

4.3. Generalization Across Different Domains

Finally, we explore whether our model is able to find various underlying physics that can generalize across different domains by mix-domain training and zero-shot testing.

Data Augmentation from Different Domains We mix the dataset of protein-protein affinity and protein-ligand affinity for training, and evaluate the models on the test set of the two domains, respectively. We also benchmark ET, MACE, and LEFTNet, which exhibit competitive performance and efficiency in § 4.2, under the same setting for comparison. We present the results in Table 3, and include detailed results in Appendix K. The results demonstrate that our method obtains benefits from the mixed training set on both PPA and LBA, while the baselines receive negative impact in most cases. Additional results on mixing PDBbind and PPA are provided in Table 5, where our GET exhibits significant improvement from the mixed training set. These phenomena well demonstrate the generalization ability of the proposed GET equipped with the unified representation.

Table 3. The mean and the standard deviation of Pearson Correlation from three runs on protein-protein affinity (PPA) and ligand binding affinity (LBA). Methods with the suffix ”-mix” are trained on the mixed dataset of PPA and LBA. The best results are marked in **bold** and the second best are underlined.

Repr.	Model	PPA-All	LBA
Block	ET	0.424 ± 0.021	0.599 ± 0.017
	ET-mix	0.457 ± 0.011	0.586 ± 0.012
	MACE	0.470 ± 0.015	0.599 ± 0.010
	MACE-mix	0.372 ± 0.042	0.590 ± 0.018
	LEFTNet	0.452 ± 0.013	0.588 ± 0.011
	LEFTNet-mix	0.450 ± 0.008	0.543 ± 0.005
Atom	ET	0.401 ± 0.005	0.591 ± 0.007
	ET-mix	0.382 ± 0.029	0.566 ± 0.061
	MACE	0.463 ± 0.052	0.579 ± 0.009
	MACE-mix	0.444 ± 0.024	0.580 ± 0.030
	LEFTNet	0.448 ± 0.046	0.610 ± 0.004
	LEFTNet-mix	0.476 ± 0.023	0.579 ± 0.014
Hier.	ET	0.438 ± 0.026	0.580 ± 0.008
	ET-mix	0.412 ± 0.035	0.569 ± 0.017
	MACE	0.466 ± 0.020	0.612 ± 0.010
	MACE-mix	0.324 ± 0.076	0.588 ± 0.011
	LEFTNet	0.445 ± 0.024	0.592 ± 0.014
	LEFTNet-mix	0.472 ± 0.020	0.556 ± 0.001
Unified	GET (ours)	0.514 ± 0.011	0.620 ± 0.004
	GET-mix (ours)	0.519 ± 0.004	0.622 ± 0.006

Zero-Shot Prediction of DNA/RNA-Ligand Affinity A more practical and meaningful, yet also more challenging scenario is ligand (small molecule) binding on nucleic acids (RNA/DNA), the data of which are scarce and expensive to obtain. We use the 149 data points available in PDBbind (Wang et al., 2004) as the zero-shot test set, and train a model on binding data from other domains in PDBbind (*i.e.* protein-protein, protein-ligand and RNA/DNA-protein). Table 4 show that GET exhibits remarkable zero-shot performance, achieving amazing generalizability across different domains on molecular interaction.

Table 4. Zero-shot performance on DNA/RNA-ligand binding affinity prediction across three runs. The best results are marked in **bold** and the second best are underlined.

Repr.	Model	Pearson \uparrow	Spearman \uparrow
Block	ET	0.217 ± 0.059	0.185 ± 0.051
	MACE	0.004 ± 0.045	0.045 ± 0.034
	LEFTNet	0.279 ± 0.127	0.252 ± 0.082
Atom	ET	0.150 ± 0.034	0.198 ± 0.043
	MACE	-0.005 ± 0.079	0.027 ± 0.083
	LEFTNet	0.271 ± 0.062	0.279 ± 0.062
Hierarchical	ET	0.348 ± 0.047	0.302 ± 0.028
	MACE	0.002 ± 0.055	0.041 ± 0.030
	LEFTNet	0.279 ± 0.122	0.259 ± 0.077
Unified	GET	0.450 ± 0.054	0.362 ± 0.041

Both experiments confirm the potential of our model to discover various underlying principles of molecular interactions capable of generalizing across diverse domains. Additionally, we also validate the superiority of GET on tasks

Table 5. The mean and the standard deviation of three runs on PDBbind benchmark. GET-mix is trained on the mixed dataset of PPA and PDBbind. The best results are marked in **bold** and the second best are underlined.

Model	RMSE \downarrow	Pearson \uparrow	Spearman \uparrow
Holoprot-Full Surface	1.464 ± 0.006	0.509 ± 0.002	0.500 ± 0.005
Holoprot-Superpixel	1.491 ± 0.004	0.491 ± 0.014	0.482 ± 0.032
ProNet-Amino Acid	1.455 ± 0.009	0.536 ± 0.012	0.526 ± 0.012
ProtNet-Backbone	1.458 ± 0.003	0.546 ± 0.007	0.550 ± 0.008
ProtNet-All-Atom	1.463 ± 0.001	<u>0.551 ± 0.005</u>	<u>0.551 ± 0.008</u>
GET (ours)	1.364 ± 0.009	0.596 ± 0.006	0.573 ± 0.007
GET-mix (ours)	1.350 ± 0.025	0.604 ± 0.014	0.600 ± 0.025

concerning bare molecules by experiments on protein property prediction. These results, although beyond the primary scope of this paper, are provided in Appendix J for readers interested in this topic.

5. Analysis

5.1. Generalization to Low-Quality Structures

In many real-world scenarios, the 3D structures of the molecular complexes are not available. While they might be obtained from structure prediction models (*e.g.* AlphaFold (Jumper et al., 2021)), the predicted results are likely to contain errors. To test the generalization of our GET under such scenarios, we add noise of different scales on the crystal structures to mimic the predicted structures of different quality. We conduct experiments on Ligand-Binding Affinity (LBA) prediction, and present the results in Table 6.

Table 6. Results on Ligand-Binding Affinity (LBA) prediction under different scales of structural error. The mean and standard deviations are reported across three parallel runs.

Error Scale (Å)	RMSE \downarrow	Pearson \uparrow	Spearman \uparrow
0.0	1.327 ± 0.005	0.620 ± 0.004	0.611 ± 0.003
0.1	1.331 ± 0.006	0.618 ± 0.003	0.607 ± 0.006
0.5	1.334 ± 0.006	0.616 ± 0.003	0.605 ± 0.004
1.0	1.328 ± 0.005	0.619 ± 0.003	0.608 ± 0.004
2.0	1.338 ± 0.009	0.614 ± 0.005	0.604 ± 0.005
3.0	1.342 ± 0.004	0.610 ± 0.003	0.600 ± 0.004
5.0	1.375 ± 0.006	0.592 ± 0.003	0.587 ± 0.004

It reads that the model achieves relatively stable performance within a 3.0 Å coordinate error range. However, larger errors may lead to a further drop in performance. Given that current commercial softwares claim to be able to predict complex structures within 2.0 Å with above 90% success rate (Friesner et al., 2004; Su et al., 2018), we conclude that the framework has considerable potential to generalize to settings where the 3D structures are inferred computationally. Nevertheless, we have to admit that this is much more challenging for other types of complexes (*e.g.* protein-protein), as their binding structures are more difficult to be predicted precisely.

5.2. Ablation Study

We conduct ablation study by removing the following modules: the layer normalization (w/o LN); the equivariant normalization on coordinates in the LN (w/o equivLN); the reflection of rescaling information in hidden features in Eq. 17 (w/o EmbedScale); the equivariant feed-forward network (w/o FFN); both LN and FFN (w/o LN & FFN). The results are presented in Table 7.

Table 7. Ablation study of each module in our proposed Generalist Equivariant Transformer (GET), where LN and FFN are abbreviations for LayerNorm and Feed-Forward Network, respectively. We mark the best results in bold and underline the second best results.

Model	Pearson↑	Spearman↑
	PPA-All	
GET-mix	0.519 ± 0.004	0.537 ± 0.003
GET	<u>0.514 ± 0.011</u>	0.533 ± 0.011
w/o LN	0.366 ± 0.024	0.426 ± 0.032
w/o equivLN	0.368 ± 0.025	0.426 ± 0.032
w/o EmbedScale	0.490 ± 0.027	0.507 ± 0.030
w/o FFN	0.494 ± 0.010	0.510 ± 0.010
w/o LN & FFN	0.360 ± 0.018	0.423 ± 0.024
LBA		
GET-mix	0.622 ± 0.006	0.615 ± 0.008
GET	<u>0.620 ± 0.004</u>	0.611 ± 0.003
w/o LN	0.589 ± 0.007	0.593 ± 0.008
w/o equivLN	0.591 ± 0.008	0.597 ± 0.009
w/o EmbedScale	0.591 ± 0.002	0.586 ± 0.003
w/o FFN	0.593 ± 0.008	0.601 ± 0.012
w/o LN & FFN	0.589 ± 0.009	0.596 ± 0.008

The ablations of the modules reveal following regularities: (1) Removing either the entire layer normalization or only the equivariant normalization on coordinates introduces instability in training, which not only leads to higher variance across different experiments, but also induces adverse impacts in some tasks like PPA; (2) Not reflecting the rescaling information in the hidden features has an adverse effect on the performance as the scale of the coordinates also carries essential information for learning the interaction physics; (3) The removal of the equivariant feed-forward module incurs detriment to the overall performance, indicating the necessity of the FFN to encourage intra-block geometrical communications between atoms. We additionally include analysis on attention in Appendix L. Further sensitivities on hidden size, number of layers, and K-nearest neighbors are included in Appendix H and I.

6. Limitations

First, current evaluations mainly focus on prediction tasks in molecular interactions. Generative tasks are another major branch in learning molecular interactions (Luo et al., 2021; Liu et al., 2022; Peng et al., 2022). Designing generative algorithms for the proposed unified representation is non-trivial, and we leave this for future work. Further, it is also possible to generalize atom-level knowledge in other scenarios apart from molecular interactions (*e.g.* tasks concerning

bare molecules). For instance, universal pretraining on different molecular domains, which needs careful design of the unsupervised task, hence we also leave this for future work.

7. Conclusion

In this paper, we explore the unified representation of molecules as geometric graphs of sets, which enables all-atom representations while preserving the heuristic building blocks of different molecules. To model the unified representaion, we propose a Generalist Equivariant Transformer (GET) to accommodate matrix-form node features and coordinates with E(3)-equivariance and permutation invariance. Each layer of GET consists of a bilevel attention module, a feed-forward module, and an equivariant layer normalization after each of the previous two modules. Experiments on molecular interactions demonstrate the superiority of learning unified representation with our GET compared to single-level representations and existing baselines. Further explorations on mixing molecular types reveal the ability of our method to learn generalizable molecular interaction mechanisms, which could inspire future research on universal representation learning of molecules.

Software and Data

Codes for our GET as well as the experiments are available at <https://github.com/THUNLP-MT/GET>.

Impact Statement

Our work on unified molecular representation learning has practical implications across various domains, including drug discovery, biotechnology, as well as advancing the design of Transformers (Zhao et al., 2023; Meng et al., 2023; Xu et al., 2024). By introducing a unified model capable of accommodating data from different molecular domains, we address the challenge of data scarcity and enhance predictive performance in specific domains suffering from limited data availability. For instance, in affinity prediction, our model leverages abundant data from diverse domains to enhance predictive performance, as demonstrated in our zero-shot experiments on RNA-ligand affinity prediction. This adaptability to new domains without specific training data showcases the practical utility of our approach in scenarios where data is limited. We include more discussions in Appendix M. As for societal consequences, we feel none must be specifically highlighted here.

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References

- Alford, R. F., Leaver-Fay, A., Jeliazkov, J. R., O'Meara, M. J., DiMaio, F. P., Park, H., Shapovalov, M. V., Renfrew, P. D., Mulligan, V. K., Kappel, K., et al. The rosetta all-atom energy function for macromolecular modeling and design. *Journal of chemical theory and computation*, 13(6):3031–3048, 2017.
- Anand, N. and Achim, T. Protein structure and sequence generation with equivariant denoising diffusion probabilistic models. *arXiv preprint arXiv:2205.15019*, 2022.
- Atz, K., Grisoni, F., and Schneider, G. Geometric deep learning on molecular representations. *Nature Machine Intelligence*, 3(12):1023–1032, 2021.
- Avsec, Ž., Agarwal, V., Visentin, D., Ledsam, J. R., Grabska-Barwinska, A., Taylor, K. R., Assael, Y., Jumper, J., Kohli, P., and Kelley, D. R. Effective gene expression prediction from sequence by integrating long-range interactions. *Nature methods*, 18(10):1196–1203, 2021.
- Ba, J. L., Kiros, J. R., and Hinton, G. E. Layer normalization. *arXiv preprint arXiv:1607.06450*, 2016.
- Baldassarre, F., Menéndez Hurtado, D., Elofsson, A., and Azizpour, H. Graphqa: protein model quality assessment using graph convolutional networks. *Bioinformatics*, 37(3):360–366, 2021.
- Ballester, P. J. and Mitchell, J. B. A machine learning approach to predicting protein–ligand binding affinity with applications to molecular docking. *Bioinformatics*, 26(9):1169–1175, 2010.
- Batatia, I., Kovacs, D. P., Simm, G., Ortner, C., and Csányi, G. Mace: Higher order equivariant message passing neural networks for fast and accurate force fields. *Advances in Neural Information Processing Systems*, 35: 11423–11436, 2022.
- Batzner, S., Musaelian, A., Sun, L., Geiger, M., Mailoa, J. P., Kornbluth, M., Molinari, N., Smidt, T. E., and Kozinsky, B. E (3)-equivariant graph neural networks for data-efficient and accurate interatomic potentials. *Nature communications*, 13(1):2453, 2022.
- Bepler, T. and Berger, B. Learning protein sequence embeddings using information from structure. *arXiv preprint arXiv:1902.08661*, 2019.
- Chanussot*, L., Das*, A., Goyal*, S., Lavril*, T., Shuaibi*, M., Riviere, M., Tran, K., Heras-Domingo, J., Ho, C., Hu, W., Palizhati, A., Sriram, A., Wood, B., Yoon, J., Parikh, D., Zitnick, C. L., and Ulissi, Z. Open catalyst 2020 (oc20) dataset and community challenges. *ACS Catalysis*, 2021. doi: 10.1021/acscatal.0c04525.
- Choukroun, Y. and Wolf, L. Geometric transformer for end-to-end molecule properties prediction. *arXiv preprint arXiv:2110.13721*, 2021.
- Cohen, I., Huang, Y., Chen, J., Benesty, J., Benesty, J., Chen, J., Huang, Y., and Cohen, I. Pearson correlation coefficient. *Noise reduction in speech processing*, pp. 1–4, 2009.
- Du, W., Du, Y., Wang, L., Feng, D., Wang, G., Ji, S., Gomes, C., and Ma, Z.-M. A new perspective on building efficient and expressive 3d equivariant graph neural networks. *arXiv preprint arXiv:2304.04757*, 2023.
- Du, X., Li, Y., Xia, Y.-L., Ai, S.-M., Liang, J., Sang, P., Ji, X.-L., and Liu, S.-Q. Insights into protein–ligand interactions: mechanisms, models, and methods. *International journal of molecular sciences*, 17(2):144, 2016.
- Elfiky, A. A. Anti-hcv, nucleotide inhibitors, repurposing against covid-19. *Life sciences*, 248:117477, 2020.
- Elnaggar, A., Heinzinger, M., Dallago, C., Rehawi, G., Wang, Y., Jones, L., Gibbs, T., Feher, T., Angerer, C., Steinegger, M., Bhowmik, D., and Rost, B. Prottrans: Toward understanding the language of life through self-supervised learning. *IEEE Transactions on Pattern Analysis and Machine Intelligence*, 44(10):7112–7127, 2022. doi: 10.1109/TPAMI.2021.3095381.
- Fey, M. and Lenssen, J. E. Fast graph representation learning with pytorch geometric. *arXiv preprint arXiv:1903.02428*, 2019.
- Friesner, R. A., Banks, J. L., Murphy, R. B., Halgren, T. A., Klicic, J. J., Mainz, D. T., Repasky, M. P., Knoll, E. H., Shelley, M., Perry, J. K., et al. Glide: a new approach for rapid, accurate docking and scoring. 1. method and assessment of docking accuracy. *Journal of medicinal chemistry*, 47(7):1739–1749, 2004.
- Gainza, P., Svärrisson, F., Monti, F., Rodola, E., Boscaini, D., Bronstein, M., and Correia, B. Deciphering interaction fingerprints from protein molecular surfaces using geometric deep learning. *Nature Methods*, 17(2):184–192, 2020.
- Gasteiger, J., Giri, S., Margraf, J. T., and Günnemann, S. Fast and uncertainty-aware directional message passing for non-equilibrium molecules. *arXiv preprint arXiv:2011.14115*, 2020a.
- Gasteiger, J., Groß, J., and Günnemann, S. Directional message passing for molecular graphs. *arXiv preprint arXiv:2003.03123*, 2020b.
- Gasteiger, J., Becker, F., and Günnemann, S. Gemnet: Universal directional graph neural networks for molecules.

- Advances in Neural Information Processing Systems*, 34: 6790–6802, 2021.
- Geng, Z., Xie, S., Xia, Y., Wu, L., Qin, T., Wang, J., Zhang, Y., Wu, F., and Liu, T.-Y. De novo molecular generation via connection-aware motif mining. *arXiv preprint arXiv:2302.01129*, 2023.
- Gilmer, J., Schoenholz, S. S., Riley, P. F., Vinyals, O., and Dahl, G. E. Neural message passing for quantum chemistry. In *International conference on machine learning*, pp. 1263–1272. PMLR, 2017.
- Gilson, M. K., Given, J. A., Bush, B. L., and McCammon, J. A. The statistical-thermodynamic basis for computation of binding affinities: a critical review. *Biophysical journal*, 72(3):1047–1069, 1997.
- Gligorijević, V., Renfrew, P. D., Kosciolek, T., Leman, J. K., Berenberg, D., Vatanen, T., Chandler, C., Taylor, B. C., Fisk, I. M., Vlamakis, H., et al. Structure-based protein function prediction using graph convolutional networks. *Nature communications*, 12(1):3168, 2021.
- Hajduk, P. J. and Greer, J. A decade of fragment-based drug design: strategic advances and lessons learned. *Nature reviews Drug discovery*, 6(3):211–219, 2007.
- Han, J., Rong, Y., Xu, T., and Huang, W. Geometrically equivariant graph neural networks: A survey. *arXiv preprint arXiv:2202.07230*, 2022.
- Hauke, J. and Kossowski, T. Comparison of values of pearson’s and spearman’s correlation coefficients on the same sets of data. *Quaestiones geographicae*, 30(2):87–93, 2011.
- Hendrycks, D. and Gimpel, K. Gaussian error linear units (gelus). *arXiv preprint arXiv:1606.08415*, 2016.
- Henikoff, S. and Henikoff, J. G. Amino acid substitution matrices from protein blocks. *Proceedings of the National Academy of Sciences*, 89(22):10915–10919, 1992.
- Hermosilla, P. and Ropinski, T. Contrastive representation learning for 3d protein structures. *arXiv preprint arXiv:2205.15675*, 2022.
- Hermosilla, P., Schäfer, M., Lang, M., Fackelmann, G., Vázquez, P. P., Kozlíková, B., Krone, M., Ritschel, T., and Ropinski, T. Intrinsic-extrinsic convolution and pooling for learning on 3d protein structures. *arXiv preprint arXiv:2007.06252*, 2020.
- Hoogeboom, E., Satorras, V. G., Vignac, C., and Welling, M. Equivariant diffusion for molecule generation in 3d. In *International Conference on Machine Learning*, pp. 8867–8887. PMLR, 2022.
- Huang, W., Han, J., Rong, Y., Xu, T., Sun, F., and Huang, J. Equivariant graph mechanics networks with constraints. *arXiv preprint arXiv:2203.06442*, 2022.
- Jiménez, J., Skalic, M., Martinez-Rosell, G., and De Fabritiis, G. K deep: protein–ligand absolute binding affinity prediction via 3d-convolutional neural networks. *Journal of chemical information and modeling*, 58(2):287–296, 2018.
- Jin, W., Barzilay, R., and Jaakkola, T. Junction tree variational autoencoder for molecular graph generation. In *International conference on machine learning*, pp. 2323–2332. PMLR, 2018.
- Jin, W., Barzilay, R., and Jaakkola, T. Antibody-antigen docking and design via hierarchical equivariant refinement. *arXiv preprint arXiv:2207.06616*, 2022.
- Jing, B., Eismann, S., Suriana, P., Townshend, R. J., and Dror, R. Learning from protein structure with geometric vector perceptrons. *arXiv preprint arXiv:2009.01411*, 2020.
- Jing, B., Eismann, S., Soni, P. N., and Dror, R. O. Equivariant graph neural networks for 3d macromolecular structure. *arXiv preprint arXiv:2106.03843*, 2021.
- Jones, S. and Thornton, J. M. Principles of protein-protein interactions. *Proceedings of the National Academy of Sciences*, 93(1):13–20, 1996.
- Joshi, C. K., Bodnar, C., Mathis, S. V., Cohen, T., and Lio, P. On the expressive power of geometric graph neural networks. In Krause, A., Brunskill, E., Cho, K., Engelhardt, B., Sabato, S., and Scarlett, J. (eds.), *Proceedings of the 40th International Conference on Machine Learning*, volume 202 of *Proceedings of Machine Learning Research*, pp. 15330–15355. PMLR, 23–29 Jul 2023. URL <https://proceedings.mlr.press/v202/joshi23a.html>.
- Jumper, J., Evans, R., Pritzel, A., Green, T., Figurnov, M., Ronneberger, O., Tunyasuvunakool, K., Bates, R., Žídek, A., Potapenko, A., et al. Highly accurate protein structure prediction with alphafold. *Nature*, 596(7873):583–589, 2021.
- Kastritis, P. L., Moal, I. H., Hwang, H., Weng, Z., Bates, P. A., Bonvin, A. M., and Janin, J. A structure-based benchmark for protein–protein binding affinity. *Protein Science*, 20(3):482–491, 2011.
- Kong, X., Huang, W., and Liu, Y. Conditional antibody design as 3d equivariant graph translation. *arXiv preprint arXiv:2208.06073*, 2022a.

- Kong, X., Huang, W., Tan, Z., and Liu, Y. Molecule generation by principal subgraph mining and assembling. *Advances in Neural Information Processing Systems*, 35: 2550–2563, 2022b.
- Liao, Y.-L. and Smidt, T. Equiformer: Equivariant graph attention transformer for 3d atomistic graphs. *arXiv preprint arXiv:2206.11990*, 2022.
- Liu, M., Luo, Y., Uchino, K., Maruhashi, K., and Ji, S. Generating 3d molecules for target protein binding. In *International Conference on Machine Learning*, pp. 13912–13924. PMLR, 2022.
- Liu, X., Luo, Y., Li, P., Song, S., and Peng, J. Deep geometric representations for modeling effects of mutations on protein-protein binding affinity. *PLoS computational biology*, 17(8):e1009284, 2021a.
- Liu, Y., Wang, L., Liu, M., Zhang, X., Oztekin, B., and Ji, S. Spherical message passing for 3d graph networks. *arXiv preprint arXiv:2102.05013*, 2021b.
- Liu, Z., Li, Y., Han, L., Li, J., Liu, J., Zhao, Z., Nie, W., Liu, Y., and Wang, R. Pdb-wide collection of binding data: current status of the pdbsbind database. *Bioinformatics*, 31(3):405–412, 2015.
- Luo, S., Guan, J., Ma, J., and Peng, J. A 3d generative model for structure-based drug design. *Advances in Neural Information Processing Systems*, 34:6229–6239, 2021.
- Luo, S., Su, Y., Peng, X., Wang, S., Peng, J., and Ma, J. Antigen-specific antibody design and optimization with diffusion-based generative models. *bioRxiv*, pp. 2022–07, 2022.
- Luo, S., Su, Y., Wu, Z., Su, C., Peng, J., and Ma, J. Rotamer density estimator is an unsupervised learner of the effect of mutations on protein-protein interaction. *bioRxiv*, pp. 2023–02, 2023.
- Meng, L., Wen, M., Le, C., Li, X., Xing, D., Zhang, W., Wen, Y., Zhang, H., Wang, J., Yang, Y., et al. Offline pre-trained multi-agent decision transformer. *Machine Intelligence Research*, 20(2):233–248, 2023.
- Musaelian, A., Batzner, S., Johansson, A., Sun, L., Owen, C. J., Kornbluth, M., and Kozinsky, B. Learning local equivariant representations for large-scale atomistic dynamics. *Nature Communications*, 14(1):579, 2023.
- Notin, P., Dias, M., Frazer, J., Hurtado, J. M., Gomez, A. N., Marks, D., and Gal, Y. Tranception: protein fitness prediction with autoregressive transformers and inference-time retrieval. In *International Conference on Machine Learning*, pp. 16990–17017. PMLR, 2022.
- Öztürk, H., Özgür, A., and Ozkirimli, E. Deepdta: deep drug–target binding affinity prediction. *Bioinformatics*, 34(17):i821–i829, 2018.
- Peng, X., Luo, S., Guan, J., Xie, Q., Peng, J., and Ma, J. Pocket2mol: Efficient molecular sampling based on 3d protein pockets. In *International Conference on Machine Learning*, pp. 17644–17655. PMLR, 2022.
- Ragoza, M., Hochuli, J., Idrobo, E., Sunseri, J., and Koes, D. R. Protein–ligand scoring with convolutional neural networks. *Journal of chemical information and modeling*, 57(4):942–957, 2017.
- Rao, R., Bhattacharya, N., Thomas, N., Duan, Y., Chen, P., Canny, J., Abbeel, P., and Song, Y. Evaluating protein transfer learning with tape. *Advances in neural information processing systems*, 32, 2019.
- Richardson, J. S. The anatomy and taxonomy of protein structure. *Advances in protein chemistry*, 34:167–339, 1981.
- Sapoval, N., Aghazadeh, A., Nute, M. G., Antunes, D. A., Balaji, A., Baraniuk, R., Barberan, C., Dannenfelser, R., Dun, C., Edrisi, M., et al. Current progress and open challenges for applying deep learning across the biosciences. *Nature Communications*, 13(1):1728, 2022.
- Satorras, V. G., Hoogeboom, E., and Welling, M. E (n) equivariant graph neural networks. In *International conference on machine learning*, pp. 9323–9332. PMLR, 2021.
- Schütt, K., Kindermans, P.-J., Sauceda Felix, H. E., Chmiela, S., Tkatchenko, A., and Müller, K.-R. Schnet: A continuous-filter convolutional neural network for modeling quantum interactions. *Advances in neural information processing systems*, 30, 2017.
- Schütt, K., Unke, O., and Gastegger, M. Equivariant message passing for the prediction of tensorial properties and molecular spectra. In *International Conference on Machine Learning*, pp. 9377–9388. PMLR, 2021.
- Shanehsazzadeh, A., Belanger, D., and Dohan, D. Is transfer learning necessary for protein landscape prediction? *arXiv preprint arXiv:2011.03443*, 2020.
- Shi, C., Wang, C., Lu, J., Zhong, B., and Tang, J. Protein sequence and structure co-design with equivariant translation. *arXiv preprint arXiv:2210.08761*, 2022.
- Somnath, V. R., Bunne, C., and Krause, A. Multi-scale representation learning on proteins. *Advances in Neural Information Processing Systems*, 34:25244–25255, 2021.

- Stärk, H., Beaini, D., Corso, G., Tossou, P., Dallago, C., Günemann, S., and Liò, P. 3d infomax improves gnns for molecular property prediction. In *International Conference on Machine Learning*, pp. 20479–20502. PMLR, 2022.
- Steinegger, M. and Söding, J. Mmseqs2 enables sensitive protein sequence searching for the analysis of massive data sets. *Nature biotechnology*, 35(11):1026–1028, 2017.
- Su, M., Yang, Q., Du, Y., Feng, G., Liu, Z., Li, Y., and Wang, R. Comparative assessment of scoring functions: the casf-2016 update. *Journal of chemical information and modeling*, 59(2):895–913, 2018.
- Thölke, P. and De Fabritiis, G. Torchmd-net: equivariant transformers for neural network based molecular potentials. *arXiv preprint arXiv:2202.02541*, 2022.
- Thomas, N., Smidt, T., Kearnes, S., Yang, L., Li, L., Kohlhoff, K., and Riley, P. Tensor field networks: Rotation-and translation-equivariant neural networks for 3d point clouds. *arXiv preprint arXiv:1802.08219*, 2018.
- Tomasi, J. and Persico, M. Molecular interactions in solution: an overview of methods based on continuous distributions of the solvent. *Chemical Reviews*, 94(7):2027–2094, 1994.
- Townshend, R. J., Vögele, M., Suriana, P., Derry, A., Powers, A., Laloudakis, Y., Balachandar, S., Jing, B., Anderson, B., Eismann, S., et al. Atom3d: Tasks on molecules in three dimensions. *arXiv preprint arXiv:2012.04035*, 2020.
- Tran, R., Lan, J., Shuaibi, M., Wood, B. M., Goyal, S., Das, A., Heras-Domingo, J., Kolluru, A., Rizvi, A., Shoghi, N., et al. The open catalyst 2022 (oc22) dataset and challenges for oxide electrocatalysts. *ACS Catalysis*, 13(5):3066–3084, 2023.
- Vamathevan, J., Clark, D., Czodrowski, P., Dunham, I., Ferran, E., Lee, G., Li, B., Madabhushi, A., Shah, P., Spitzer, M., et al. Applications of machine learning in drug discovery and development. *Nature reviews Drug discovery*, 18(6):463–477, 2019.
- Vaswani, A., Shazeer, N., Parmar, N., Uszkoreit, J., Jones, L., Gomez, A. N., Kaiser, Ł., and Polosukhin, I. Attention is all you need. *Advances in neural information processing systems*, 30, 2017.
- Vreven, T., Moal, I. H., Vangone, A., Pierce, B. G., Kastritis, P. L., Torchala, M., Chaleil, R., Jiménez-García, B., Bates, P. A., Fernandez-Recio, J., et al. Updates to the integrated protein–protein interaction benchmarks: docking benchmark version 5 and affinity benchmark version 2. *Journal of molecular biology*, 427(19):3031–3041, 2015.
- Wang, L., Liu, H., Liu, Y., Kurtin, J., and Ji, S. Learning protein representations via complete 3d graph networks. *arXiv preprint arXiv:2207.12600*, 2022a.
- Wang, R., Fang, X., Lu, Y., and Wang, S. The pdbsbind database: Collection of binding affinities for protein-ligand complexes with known three-dimensional structures. *Journal of medicinal chemistry*, 47(12):2977–2980, 2004.
- Wang, Z., Combs, S. A., Brand, R., Calvo, M. R., Xu, P., Price, G., Golovach, N., Salawu, E. O., Wise, C. J., Ponnapalli, S. P., et al. Lm-gvp: an extensible sequence and structure informed deep learning framework for protein property prediction. *Scientific reports*, 12(1):6832, 2022b.
- Watson, J. D. and Crick, F. H. The structure of dna. In *Cold Spring Harbor symposia on quantitative biology*, volume 18, pp. 123–131. Cold Spring Harbor Laboratory Press, 1953.
- Xu, G., Hou, Q., and Cheng, M.-M. Dual frequency transformer for efficient sdr-to-hdr translation. *Machine Intelligence Research*, pp. 1–11, 2024.
- Xu, M., Yu, L., Song, Y., Shi, C., Ermon, S., and Tang, J. Geodiff: A geometric diffusion model for molecular conformation generation. *arXiv preprint arXiv:2203.02923*, 2022.
- Zaidi, S., Schaarschmidt, M., Martens, J., Kim, H., Teh, Y. W., Sanchez-Gonzalez, A., Battaglia, P., Pascanu, R., and Godwin, J. Pre-training via denoising for molecular property prediction. *arXiv preprint arXiv:2206.00133*, 2022.
- Zhang, Y., Cai, H., Shi, C., Zhong, B., and Tang, J. E3bind: An end-to-end equivariant network for protein-ligand docking. *arXiv preprint arXiv:2210.06069*, 2022a.
- Zhang, Z., Xu, M., Jamasb, A., Chenthamarakshan, V., Lozano, A., Das, P., and Tang, J. Protein representation learning by geometric structure pretraining. *arXiv preprint arXiv:2203.06125*, 2022b.
- Zhao, Y., Zhang, J., and Zong, C. Transformer: A general framework from machine translation to others. *Machine Intelligence Research*, 20(4):514–538, 2023.

A. Atom Position Code

Certain types of molecules have conventional position codes to distinguish different status of the atoms in the same block. For example, in the protein domain, where building blocks are residues, each atom in a residue is assigned a position code ($\alpha, \beta, \gamma, \delta, \varepsilon, \zeta, \eta, \dots$) according to the number of chemical bonds between it and the alpha carbon (i.e. C_α). As these position codes provide meaningful heuristics of intra-block geometry, we also include them as a component of the embedding. For other types of molecules without such position codes (e.g. small molecules), we assign a [BLANK] type for positional embedding.

B. Scheme of the Generalist Equivariant Transformer

We depict the overall workflow and the details of the equivariant bilevel attention module in Figure 3.

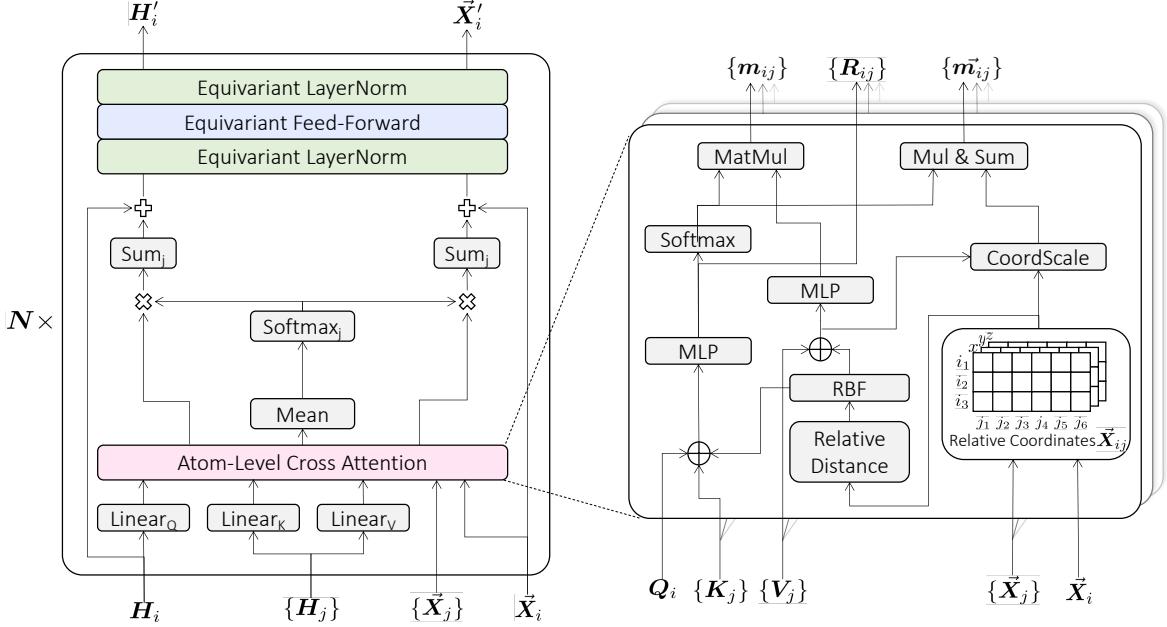


Figure 3. The scheme of a layer of Generalist Equivariant Transformer, where block i ($\mathbf{H}_i, \vec{\mathbf{X}}_i$) is updated by its neighbors ($\{\mathbf{H}_j\}, \{\vec{\mathbf{X}}_j\}$, $j \in \mathcal{N}_b(i)$). \times , $+$, and \oplus denote multiplication, addition and concatenation, respectively. (Left) The overall workflow of a layer and details of the block-level attention. (Right) The details of the atom-level cross attention. GET is composed of N such layers.

RBF embeds distances into d_{rbf} -dimensional vectors through radial basis functions:

$$\text{RBF}(d)_k = u\left(\frac{d}{c}\right) \cdot \exp\left(-\frac{|d - \mu_k|}{2\gamma}\right), 1 \leq k \leq d_{\text{rbf}} \quad (18)$$

where μ_k is uniformly distributed in $[0, c]$ ($c = 7.0$ in our paper), $\gamma = \frac{c}{d_{\text{rbf}}}$, and $u(d)$ is the polynomial envelope function for cutting off large distances (Gasteiger et al., 2020b) ($p = 5$ in our paper):

$$u(x) = 1 - \frac{(p+1)(p+2)}{2}x^p + p(p+2)x^{p+1} - \frac{p(p+1)}{2}x^{p+2} \quad (19)$$

C. Proof of E(3)-Equivariance and Intra-Block Permutation Invariance

Theorem C.1 (E(3)-Equivariance and Intra-Block Permutation Invariance). Denote the proposed Equivariant Transformer as $\{\mathbf{H}'_i, \vec{\mathbf{X}}'_i\} = \text{GET}(\{\mathbf{H}_i, \vec{\mathbf{X}}_i\})$, then it conforms to E(3)-Equivariance and Intra-Block Permutation Invariance. Namely, $\forall g \in E(3), \forall \{\pi_i \in S_{n_i} | 1 \leq i \leq B\}$, where B is the number of blocks in the input and S_{n_i} includes all permutations on n_i elements, we have $\{\pi_i \cdot \mathbf{H}'_i, \pi_i \cdot g \cdot \vec{\mathbf{X}}'_i\} = \text{GET}(\{\pi_i \cdot \mathbf{H}_i, \pi_i \cdot g \cdot \vec{\mathbf{X}}_i\})$.

The Generalist Equivariant Transformer (GET) is the cascading of the three types of modules: bilevel attention, feed-forward network, and layer normalization. Further, the E(3)-equivariance and the intra-block permutation invariance are disentangled. Therefore, the proof of its E(3)-equivariance and its invariance respect to the intra-block permutations can be decomposed into proof of these two properties on each module, which we present below.

C.1. Proof of E(3)-Equivariance

First we give the definition of E(3)-equivariance as follows:

Definition C.2 (E(3)-equivariance). A function $\phi : \mathbb{X} \rightarrow \mathbb{Y}$ conforms E(3)-equivariance if $\forall g \in E(3)$, the equation $\rho_{\mathbb{Y}}(g)\mathbf{y} = \phi(\rho_{\mathbb{X}}(g)\mathbf{x})$ holds true, where $\rho_{\mathbb{X}}$ and $\rho_{\mathbb{Y}}$ instantiate g in \mathbb{X} and \mathbb{Y} , respectively. A special case is E(3)-invariance where $\rho_{\mathbb{Y}}$ constantly outputs identity transformation (i.e. $\rho_{\mathbb{Y}}(g) \equiv I$).

Given $g \in E(3)$ and $\vec{\mathbf{x}} \in \mathbb{R}^3$, we can instantiate g as $g \cdot \vec{\mathbf{x}} := Q\vec{\mathbf{x}} + \vec{t}$, where $Q \in \mathbb{R}^{3 \times 3}$ is an orthogonal matrix and $\vec{t} \in \mathbb{R}^3$ is a translation vector. Implementing g on a coordinate matrix $\vec{\mathbf{X}} \in \mathbb{R}^{n \times 3}$ means transforming each coordinate (i.e. each row) with g .

Then we prove the E(3)-equivariance of each module in GET as follows:

Lemma C.3. Denote the bilevel attention module as $\{\mathbf{H}'_i, \vec{\mathbf{X}}'_i\} = \text{Att}(\{\mathbf{H}_i, \vec{\mathbf{X}}_i\})$, then it is E(3)-equivariant. Namely, $\forall g \in E(3)$, we have $\{\mathbf{H}'_i, g \cdot \vec{\mathbf{X}}'_i\} = \text{Att}(\{\mathbf{H}_i, g \cdot \vec{\mathbf{X}}_i\})$.

Proof. The key to the proof of Lemma C.3 is to prove that the propagation in Eq. 2-10 is E(3)-invariant on \mathbf{H}_i and E(3)-equivariant on $\vec{\mathbf{X}}_i$. Obviously, the correlation \mathbf{R}_{ij} between block i and block j in Eq. 3 is E(3)-invariant because all the inputs, that is, the query, the key, and the distance matrices, are not affected by the geometric transformation g . Therefore, we can immediately derive that the atom-level cross attention α_{ij} in Eq. 4 is E(3)-invariant. Similarly, the block-level attention β_{ij} in Eq. 6 is E(3)-invariant because it only operates on \mathbf{r}_{ij} in Eq. 5 which aggregates α_{ij} and the edge feature. Finally, we can derive the E(3)-invariance on \mathbf{H} and the E(3)-equivariance on $\vec{\mathbf{X}}$:

$$\begin{aligned}
 \mathbf{H}'_i[p] &= \mathbf{H}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \phi_m(\mathbf{m}_{ij,p}), \\
 &= \mathbf{H}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \phi_m(\alpha_{ij}[p] \cdot \phi_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])), \\
 g \cdot \vec{\mathbf{X}}'_i[p] &= g \cdot \left(\vec{\mathbf{X}}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} (\bar{\mathbf{m}}_{ij,p} \odot \sigma_m(\mathbf{m}_{ij,p})) \right) \\
 &= g \cdot \left(\vec{\mathbf{X}}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \left(\alpha_{ij}[p] \cdot (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])) \odot \sigma_m(\mathbf{m}_{ij,p})) \right) \right) \\
 &= Q \left(\vec{\mathbf{X}}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \left(\alpha_{ij}[p] \cdot (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])) \odot \sigma_m(\mathbf{m}_{ij,p})) \right) \right) + \vec{t} \\
 &= (Q\vec{\mathbf{X}}_i[p] + \vec{t}) + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \\
 &\quad \left(\alpha_{ij}[p] \cdot \left(\begin{bmatrix} Q(\vec{\mathbf{X}}_i[p] - \vec{\mathbf{X}}_j[1]) \\ \vdots \\ Q(\vec{\mathbf{X}}_i[p] - \vec{\mathbf{X}}_j[n_j]) \end{bmatrix} \odot \sigma_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])) \right) \odot \sigma_m(\mathbf{m}_{ij,p}) \right) \\
 &= (Q\vec{\mathbf{X}}_i[p] + \vec{t}) + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \\
 &\quad \left(\alpha_{ij}[p] \cdot \left(\begin{bmatrix} Q\vec{\mathbf{X}}_i[p] + \vec{t} - (Q\vec{\mathbf{X}}_j[1] + \vec{t}) \\ \vdots \\ Q\vec{\mathbf{X}}_i[p] + \vec{t} - (Q\vec{\mathbf{X}}_j[n_j] + \vec{t}) \end{bmatrix} \odot \sigma_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])) \right) \odot \sigma_m(\mathbf{m}_{ij,p}) \right) \\
 &= g \cdot \vec{\mathbf{X}}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \\
 &\quad \left(\alpha_{ij}[p] \cdot \left(\begin{bmatrix} g \cdot \vec{\mathbf{X}}_i[p] - g \cdot \vec{\mathbf{X}}_j[1] \\ \vdots \\ g \cdot \vec{\mathbf{X}}_i[p] - g \cdot \vec{\mathbf{X}}_j[n_j] \end{bmatrix} \odot \sigma_v(\mathbf{V}_j \parallel \text{RBF}(\mathbf{D}_{ij}[p])) \right) \odot \sigma_m(\mathbf{m}_{ij,p}) \right),
 \end{aligned}$$

which concludes the proof of Lemma C.3. \square

Lemma C.4. Denote the equivariant feed-forward network as $\{\mathbf{H}'_i, \vec{\mathbf{X}}'_i\} = \text{EFFN}(\{\mathbf{H}_i, \vec{\mathbf{X}}_i\})$, then it is E(3)-equivariant. Namely, $\forall g \in E(3)$, we have $\{\mathbf{H}'_i, g \cdot \vec{\mathbf{X}}'_i\} = \text{EFFN}(\{\mathbf{H}_i, g \cdot \vec{\mathbf{X}}_i\})$.

Proof. The proof of Lemma C.4 focuses on the single-atom updates in Eq. 11-14. First, it is easy to obtain the E(3)-equivariance of the centroid in Eq. 11:

$$g \cdot \vec{x}_c = g \cdot \text{centroid}(\vec{X}_i) = \text{centroid}(g \cdot \vec{X}_i).$$

Then we have the following equation on the relative coordinate $\Delta\vec{x}$ in Eq. 12:

$$\mathbf{Q}\Delta\vec{x} = (\mathbf{Q}\vec{x} + \vec{t}) - (\mathbf{Q}\vec{x}_c + \vec{t}) = g \cdot \vec{x} - g \cdot \vec{x}_c.$$

We can immediately obtain the E(3)-invariance of r in Eq. 12:

$$r = \text{RBF}(\|\mathbf{Q}\Delta\vec{x}\|_2) = \text{RBF}(\sqrt{(\mathbf{Q}\Delta\vec{x})^\top(\mathbf{Q}\Delta\vec{x})}) = \text{RBF}(\sqrt{\Delta\vec{x}^\top \mathbf{Q}^\top \mathbf{Q} \Delta\vec{x}}) = \text{RBF}(\|\Delta\vec{x}\|_2).$$

Finally we can derive the E(3)-invariance on \mathbf{h} and the E(3)-equivariance on \vec{x} :

$$\begin{aligned} \mathbf{h}' &= \mathbf{h} + \phi_h(\mathbf{h}, \mathbf{h}_c, \mathbf{r}), \\ g \cdot \vec{x}' &= g \cdot (\vec{x} + \Delta\vec{x}\phi_x(\mathbf{h}, \mathbf{h}_c, \mathbf{r})) \\ &= \mathbf{Q}(\vec{x} + \Delta\vec{x}\phi_x(\mathbf{h}, \mathbf{h}_c, \mathbf{r})) + \vec{t} \\ &= \mathbf{Q}\vec{x} + \vec{t} + \mathbf{Q}\Delta\vec{x}\phi_x(\mathbf{h}, \mathbf{h}_c, \mathbf{r}) \\ &= g \cdot \vec{x} + (g \cdot \vec{x} - g \cdot \vec{x}_c)\phi_x(\mathbf{h}, \mathbf{h}_c, \mathbf{r}) \\ &= g \cdot \vec{x} + (g \cdot \vec{x} - \text{centroid}(g \cdot \vec{X}_i))\phi_x(\mathbf{h}, \mathbf{h}_c, \mathbf{r}), \end{aligned}$$

which concludes the proof of Lemma C.4 \square

Lemma C.5. Denote the equivariant layer normalization as $\{\mathbf{H}'_i, \vec{X}'_i\} = \text{ELN}(\{\mathbf{H}_i, \vec{X}_i\})$, then it is E(3)-equivariant. Namely, $\forall g \in E(3)$, we have $\{\mathbf{H}'_i, g \cdot \vec{X}'_i\} = \text{ELN}(\{\mathbf{H}_i, g \cdot \vec{X}_i\})$.

Proof. Since the layer normalization is implemented on the atom level, namely each row of the coordinate matrix in a node, we again only need to concentrate on the single-atom normalization in Eq. 15-16. The key points lie in the E(3)-equivariance of $\mathbb{E}[\vec{\mathbf{X}}]$ and the E(3)-invariance of $\text{Var}[\vec{\mathbf{X}} - \mathbb{E}[\vec{\mathbf{X}}]]$. The first one is obvious because $\mathbb{E}[\vec{\mathbf{X}}]$ is the centroid of the coordinates of all atoms:

$$g \cdot \mathbb{E}[\vec{\mathbf{X}}] = g \cdot \text{centroid}(\vec{\mathbf{X}}) = \text{centroid}(g \cdot \vec{\mathbf{X}}) = \mathbb{E}[g \cdot \vec{\mathbf{X}}].$$

Suppose there are N atoms in total, then we can prove the E(3)-invariance of the variance as follows:

$$\begin{aligned} \text{Var}[\vec{\mathbf{X}} - \mathbb{E}[\vec{\mathbf{X}}]] &= \frac{\sum_{i=1}^N (x_i - \bar{x})^2 + \sum_{i=1}^N (y_i - \bar{y})^2 + \sum_{i=1}^N (z_i - \bar{z})^2}{3N} \\ &= \frac{\sum_{i=1}^N [(x_i - \bar{x})^2 + (y_i - \bar{y})^2 + (z_i - \bar{z})^2]}{3N} \\ &= \frac{\sum_{i=1}^N (\vec{x}_i - \mathbb{E}[\vec{\mathbf{X}}])^\top (\vec{x}_i - \mathbb{E}[\vec{\mathbf{X}}])}{3N} \\ &= \frac{\sum_{i=1}^N (\vec{x}_i - \mathbb{E}[\vec{\mathbf{X}}])^\top \mathbf{Q}^\top \mathbf{Q} (\vec{x}_i - \mathbb{E}[\vec{\mathbf{X}}])}{3N} \\ &= \frac{\sum_{i=1}^N (\mathbf{Q}\vec{x}_i - \mathbf{Q}\mathbb{E}[\vec{\mathbf{X}}])^\top (\mathbf{Q}\vec{x}_i - \mathbf{Q}\mathbb{E}[\vec{\mathbf{X}}])}{3N} \\ &= \frac{\sum_{i=1}^N (g \cdot \vec{x}_i - g \cdot \mathbb{E}[\vec{\mathbf{X}}])^\top (g \cdot \vec{x}_i - g \cdot \mathbb{E}[\vec{\mathbf{X}}])}{3N} \\ &= \frac{\sum_{i=1}^N (g \cdot \vec{x}_i - \mathbb{E}[g \cdot \vec{\mathbf{X}}])^\top (g \cdot \vec{x}_i - \mathbb{E}[g \cdot \vec{\mathbf{X}}])}{3N} \\ &= \text{Var}[g \cdot \vec{\mathbf{X}} - \mathbb{E}[g \cdot \vec{\mathbf{X}}]]. \end{aligned}$$

Therefore, we can finally derive the E(3)-invariance on \mathbf{h} and the E(3)-equivariance on \vec{x} in Eq. 15-16:

$$\begin{aligned}
 \mathbf{h} &= \frac{\mathbf{h} - \mathbb{E}[\mathbf{h}]}{\sqrt{\text{Var}[\mathbf{h}]}} \cdot \gamma + \beta, \\
 g \cdot \vec{x} &= g \cdot \left(\frac{\vec{x} - \mathbb{E}[\vec{\mathbf{X}}]}{\sqrt{\text{Var}[\vec{\mathbf{X}} - \mathbb{E}[\vec{\mathbf{X}}]]}} \cdot \sigma + \mathbb{E}[\vec{\mathbf{X}}] \right) = \frac{Q\vec{x} - Q\mathbb{E}[\vec{\mathbf{X}}]}{\sqrt{\text{Var}[\vec{\mathbf{X}} - \mathbb{E}[\vec{\mathbf{X}}]]}} \cdot \sigma + Q\mathbb{E}[\vec{\mathbf{X}}] + \vec{t} \\
 &= \frac{Q\vec{x} + \vec{t} - (Q\mathbb{E}[\vec{\mathbf{X}}] + \vec{t})}{\sqrt{\text{Var}[\vec{\mathbf{X}} - \mathbb{E}[\vec{\mathbf{X}}]]}} \cdot \sigma + (Q\mathbb{E}[\vec{\mathbf{X}}] + \vec{t}) = \frac{g \cdot \vec{x} - g \cdot \mathbb{E}[\vec{\mathbf{X}}]}{\sqrt{\text{Var}[g \cdot \vec{\mathbf{X}} - \mathbb{E}[g \cdot \vec{\mathbf{X}}]]}} \cdot \sigma + g \cdot \mathbb{E}[\vec{\mathbf{X}}] \\
 &= \frac{g \cdot \vec{x} - \mathbb{E}[g \cdot \vec{\mathbf{X}}]}{\sqrt{\text{Var}[g \cdot \vec{\mathbf{X}} - \mathbb{E}[g \cdot \vec{\mathbf{X}}]]}} \cdot \sigma + \mathbb{E}[g \cdot \vec{\mathbf{X}}],
 \end{aligned}$$

which concludes the proof of Lemma C.5. \square

With Lemma C.3-C.5 at hand, it is obvious to deduce the E(3)-equivariance of the GET layer.

C.2. Proof of Intra-Block Permutation Invariance

Obviously, the feed-forward network and the layer normalization are invariant to intra-block permutations because they are implemented on single atoms and the only incorporated multi-atom operation is averaging, which is invariant to the permutations. Therefore, the proof narrows down to the intra-block permutation invariance of the bilevel attention module.

Lemma C.6. Denote the bilevel attention module as $\{\mathbf{H}'_i, \vec{\mathbf{X}}'_i\} = \text{Att}(\{\mathbf{H}_i, \vec{\mathbf{X}}_i\})$, then it conforms to intra-block permutation invariance. Namely, $\forall \{\pi_i \in S_{n_i} | 1 \leq i \leq B\}$, where B is the number of blocks in the input and S_{n_i} includes all permutations on n_i elements, we have $\{\pi_i \cdot \mathbf{H}'_i, \pi_i \cdot \vec{\mathbf{X}}'_i\} = \text{Att}(\{\pi_i \cdot \mathbf{H}_i, \pi_i \cdot \vec{\mathbf{X}}_i\})$.

Proof. Denote the permutation of block i as π_i , then it can be instantiated as the multiplication of a series of elementary row-switching matrices $\mathbf{P}_i = \mathbf{P}_i^{(m_i)} \mathbf{P}_i^{(m_i-1)} \dots \mathbf{P}_i^{(1)}$. For example, we have $\pi_i \cdot \mathbf{H}_i = \mathbf{P}_i \mathbf{H}_i$, $\pi_i \cdot \vec{\mathbf{X}}_i = \mathbf{P}_i \vec{\mathbf{X}}_i$. Here we first prove an elegant property of \mathbf{P}_i , which we will use in the later proof:

$$\begin{aligned}
 \mathbf{P}_i^\top \mathbf{P}_i &= (\mathbf{P}_i^{(m_i)} \mathbf{P}_i^{(m_i-1)} \dots \mathbf{P}_i^{(1)})^\top (\mathbf{P}_i^{(m_i)} \mathbf{P}_i^{(m_i-1)} \dots \mathbf{P}_i^{(1)}) \\
 &= \mathbf{P}_i^{(1)\top} \dots \mathbf{P}_i^{(m_i-1)\top} \mathbf{P}_i^{(m_i)\top} \mathbf{P}_i^{(m_i)} \mathbf{P}_i^{(m_i-1)} \dots \mathbf{P}_i^{(1)} \\
 &= \mathbf{P}_i^{(1)\top} \dots \mathbf{P}_i^{(m_i-1)\top} \mathbf{I} \mathbf{P}_i^{(m_i-1)} \dots \mathbf{P}_i^{(1)} \\
 &= \dots \\
 &= \mathbf{I}
 \end{aligned}$$

Given arbitrary permutations on each block, we have the permuted query, key, and value matrices:

$$\begin{aligned}
 \mathbf{P}_i \mathbf{Q}_i &= \mathbf{P}_i \mathbf{H}_i \mathbf{W}_Q = (\pi_i \cdot \mathbf{H}_i) \mathbf{W}_Q, \\
 \mathbf{P}_i \mathbf{K}_i &= \mathbf{P}_i \mathbf{H}_i \mathbf{W}_K = (\pi_i \cdot \mathbf{H}_i) \mathbf{W}_K, \\
 \mathbf{P}_i \mathbf{V}_i &= \mathbf{P}_i \mathbf{H}_i \mathbf{W}_V = (\pi_i \cdot \mathbf{H}_i) \mathbf{W}_V.
 \end{aligned}$$

The distance matrix \mathbf{D}_{ij} is also permuted as $\mathbf{P}_i \mathbf{D}_{ij} \mathbf{P}_j^\top$. Therefore, the atom-level attention α_{ij} in Eq. 4 is also permuted

as $\mathbf{P}_i \alpha_{ij} \mathbf{P}_j^\top$, and the messages in Eq. 7-8 are permuted as:

$$\begin{aligned}\mathbf{P}_i \mathbf{m}_{ij} &= \mathbf{P}_i \begin{bmatrix} \alpha_{ij}[1] \cdot \phi_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[1])) \\ \vdots \\ \alpha_{ij}[n_i] \cdot \phi_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[n_i])) \end{bmatrix} = \mathbf{P}_i \begin{bmatrix} \alpha_{ij}[1] \mathbf{P}_j^\top \mathbf{P}_j \phi_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[1])) \\ \vdots \\ \alpha_{ij}[n_i] \mathbf{P}_j^\top \mathbf{P}_j \phi_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[n_i])) \end{bmatrix} \\ \mathbf{P}_i \vec{\mathbf{m}}_{ij} &= \mathbf{P}_i \begin{bmatrix} \alpha_{ij}[1] \cdot (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[1]))) \\ \vdots \\ \alpha_{ij}[n_i] \cdot (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[n_i]))) \end{bmatrix} \\ &= \mathbf{P}_i \begin{bmatrix} \alpha_{ij}[1] \mathbf{P}_j^\top \mathbf{P}_j (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[1]))) \\ \vdots \\ \alpha_{ij}[n_i] \mathbf{P}_j^\top \mathbf{P}_j (\vec{\mathbf{X}}_{ij}[p] \odot \sigma_v(\mathbf{V}_j \| \text{RBF}(\mathbf{D}_{ij}[n_i]))) \end{bmatrix},\end{aligned}$$

The block-level attention β_{ij} in Eq. 6 remains unchanged as the average of \mathbf{R}_{ij} in obtaining \mathbf{r}_{ij} eliminates the effect of permutations. Finally, we can derive the intra-block permutation invariance as follows:

$$\begin{aligned}\mathbf{P}_i \mathbf{H}'_i &= \mathbf{P}_i \left(\mathbf{H}_i + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \phi_m(\mathbf{m}_{ij}) \right) = \mathbf{P}_i \mathbf{H}_i + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \phi_m(\mathbf{P}_i \mathbf{m}_{ij}), \\ \mathbf{P}_i \vec{\mathbf{X}}'_i &= \mathbf{P}_i \left(\vec{\mathbf{X}}_i + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \vec{\mathbf{m}}_{ij} \odot \sigma_m(\vec{\mathbf{m}}_{ij}) \right) = \mathbf{P}_i \vec{\mathbf{X}}_i + \sum_{j \in \mathcal{N}(i)} \beta_{ij} (\mathbf{P}_i \vec{\mathbf{m}}_{ij}) \odot \sigma_m(\mathbf{P}_i \vec{\mathbf{m}}_{ij})\end{aligned}$$

which concludes Lemma C.6. \square

D. Complexity Analysis

To discuss the scalability of the model, we additionally provide the complexity analysis as follows. The main complexity lies in the attention-based message passing module. Suppose block i and block j have n_i and n_j atoms, respectively. Since the attention module implements bipartite cross attention between block pairs, there are a total of $n_i n_j$ attention edges between block i and block j . Therefore, the exact complexity should be $O(\sum_{i \in \mathcal{V}} \sum_{j \in \mathcal{N}(i)} n_i n_j)$, where \mathcal{V} includes all nodes and $\mathcal{N}(i)$ includes all neighbors of block i . Since we use K nearest neighbors to construct graphs in block level, we have $\mathcal{N}(i) \leq K$. Denote the maximum number of atoms in a single block is C (in natural proteins we have $C = 14$), we have $n_i \leq C$. Therefore, we have $\sum_{i \in \mathcal{V}} \sum_{j \in \mathcal{N}(i)} n_i n_j \leq \sum_{i \in \mathcal{V}} K C^2 = K C^2 |\mathcal{V}|$, namely, the complexity should be bounded by $O(K C^2 |\mathcal{V}|)$, which is linear to the number of blocks in the graph. A linear complexity means the algorithm should be easy to scale to larger molecular systems.

Practically, the complexity can be further optimized by selecting only k nearest neighbors of each atom in message passing between block i and block j . With this sparse attention, the complexity is $O(\sum_{i \in \mathcal{V}} \sum_{j \in \mathcal{N}(i)} k n_i) \leq O(k K N)$, where N is the total number of atoms.

E. Ligand Efficacy Prediction

We additionally provide the evaluation results on Ligand Efficacy Prediction (LEP). This task requires identifying a given ligand as the "activator" or the "inactivator" of a functional protein. Specifically, given the two complexes where the ligand interacts with the active and the inactive conformation of the protein respectively, the models need to distinguish which one is more favorable. To this end, we first obtain the graph-level representations of the two complexes. Then we concatenate the two representations to do a binary classification. We use two metrics for evaluation: the area under the receiver operating characteristic (**AUROC**) and the area under precision-recall curve (**AUPRC**).

Dataset We follow the LEP dataset and its splits in the Atom3D benchmark (Townshend et al., 2020), which includes 27 functional proteins and 527 ligands known as activator or inactivator to a certain protein. The active and the inactive

complexes are generated by Glide (Friesner et al., 2004). The splits of the training, the validation, and the test sets are based on the functional proteins to ensure generalizability.

Table 8. The mean and the standard deviation of three runs on ligand efficacy prediction. The best results are marked in bold and the second best are underlined.

Repr.	Model	AUROC↑	AUPRC↑
Block	SchNet	0.732 ± 0.022	0.718 ± 0.031
	DimeNet++	0.669 ± 0.014	0.609 ± 0.036
	EGNN	0.746 ± 0.017	0.755 ± 0.031
	ET	0.744 ± 0.034	0.721 ± 0.052
Atom	SchNet	0.712 ± 0.026	0.639 ± 0.033
	DimeNet++	0.589 ± 0.049	0.503 ± 0.020
	EGNN	0.711 ± 0.020	0.643 ± 0.041
	ET	0.677 ± 0.004	0.636 ± 0.054
Hierarchical	SchNet	0.736 ± 0.020	0.731 ± 0.048
	DimeNet++	0.579 ± 0.118	0.517 ± 0.100
	EGNN	0.724 ± 0.027	0.720 ± 0.056
	ET	0.717 ± 0.033	0.724 ± 0.055
Unified	GET (ours)	0.761 ± 0.012	0.751 ± 0.012

Results We present the mean and the standard deviation of the metrics across three runs in Table 8. LEP requires distinguishing the active and inactive conformations of the receptor, thus it is essential to capture the block-level geometry of the protein in addition to the atom-level receptor-ligand interactions. The unified representation excels at learning the bilevel geometry, therefore, naturally, we observe obvious gains on the metrics of our method compared to the baselines.

F. Implementation Details

We conduct experiments on 1 GeForce RTX 2080 Ti GPU with 12G memory except the zero-shot evaluation on RNA/DNA-ligand affinity which needs 2 GPUs. Each model is trained with Adam optimizer and exponential learning rate decay. To avoid unstable checkpoints from early training stages, we select the latest checkpoint from the saved top- k checkpoints on the validation set for testing. Since the number of blocks varies a lot in different samples, we set a upperbound of the number of blocks to form a dynamic batch instead of using a static batch size. We use $k = 9$ for constructing the k-nearest neighbor graph in § 3.1 and set the size of the RBF kernel (d_{rbf}) to 32. We give the description of the hyperparameters in Table 9 and their values for each task in Table 10.

Table 9. Descriptions of the hyperparameters.

hyperparameter	description
d_h	Hidden size
d_r	Radial size for the attention module
lr	Learning rate
final_lr	Final learning rate
max_epoch	Maximum of epochs to train
save_topk	Number of top- k checkpoints to save
n.layers	Number of layers
max_n_vertex	Upperbound of the number of nodes in a batch

Table 10. Hyperparameters for our GET on each task.

hyperparameter	PPA	LBA	LEP	hyperparameter	PPA	LBA	LEP
GET							
d_h	128	64	128	d_r	16	32	64
lr	10^{-4}	10^{-3}	5×10^{-4}	final_lr	10^{-4}	10^{-6}	10^{-4}
max_epoch	20	10	90	save_topk	3	3	7
n.layers	3	3	3	max_n_vertex	1500	2000	1500
GET-mix							
d_h	128	128	-	d_r	16	16	-
lr	5×10^{-5}	5×10^{-5}	-	final_lr	5×10^{-5}	10^{-6}	-
max_epoch	20	20	-	save_topk	3	3	-
n.layers	3	3	-	max_n_vertex	1500	1500	-

F.1. PDBbind Benchmark

We follow Somnath et al. (2021); Wang et al. (2022a) to conduct experiments on the well-established PDBbind (Wang et al., 2004; Liu et al., 2015) and use the split with sequence identity threshold of 30% which should barely have data leakage problem. A total of 4709 complexes are first filtered by resolution and then splitted into 3507, 466, 490 for training, validation, and testing (Somnath et al., 2021). We directly borrow the results of the baselines from Wang et al. (2022a). For our model, we set $d_h = 64$, $d_r = 64$, $\text{lr} = 10^{-3}$, $\text{final_lr} = 10^{-4}$, and $\text{max_n_vertex} = 1500$.

F.2. Protein-Protein Affinity

Here we illustrate the setup for protein-protein affinity prediction with more details.

We adopt the Protein-Protein Affinity Benchmark Version 2 (Kastritis et al., 2011; Vreven et al., 2015) as the test set, which contains 176 diversified protein-protein complexes with annotated affinity collected from existing literature. These complexes are further categorized into three difficulty levels (i.e. Rigid, Medium, Flexible) according to the conformation change of the proteins from the unbound to the bound state (Kastritis et al., 2011), among which the Flexible split is the most challenging as the proteins undergo large conformation change upon binding.

As for training, we first filter out 2,500 complexes with annotated binding affinity (K_i or K_d) from PDBbind (Wang et al., 2004). Then we use MMseqs2 (Steinegger & Söding, 2017) to cluster the sequences of these complexes together with the test set by dividing complexes with sequence identity above 30% into the same cluster, where sequence identity is calculated based on the BLOSUM62 substitution matrix (Henikoff & Henikoff, 1992). The complexes that shares the same clusters with the test set are dropped to prevent data leakage, after which we finally obtained 2,195 valid complexes. We split these complexes into training set and validation set with a ratio of 9:1 with respect to the number of clusters. Following previous literature (Ballester & Mitchell, 2010; Jiménez et al., 2018; Ragoza et al., 2017), we predict the negative log-transformed value (pK) instead of direct regression on the affinity.

G. Baselines

G.1. Implementation

In this section, we describe the implementation details of different baselines. All the baselines are designed for structural learning on graphs whose nodes are represented as one feature vector and one coordinate. Therefore, for **block-level** representation, we average the embeddings and the coordinates of the atoms in each block before feeding the graph to the baselines. For **atom-level** representation, each node is represented as the embedding and the coordinate of each atom. For **Hierarchical** methods, we first implement message passing on atom-level graphs, then average the embeddings and coordinates within each block before conducting block-level message passing (Jin et al., 2022). For fair comparison, the number of layers in each model is set to 3, except MACE and Equiformer, which are quite unstable in training with more than 2 layers. We present other hyperparameters in Table 11.

For **SchNet** (Schütt et al., 2017) and **DimeNet++** (Gasteiger et al., 2020b;a), we use the implementation in PyTorch Geometric (Fey & Lenssen, 2019). For EGNN (Satorras et al., 2021), **ET** (Thölke & De Fabritiis, 2022), **MACE** (Batatia et al., 2022), and **LEFTNet** (Du et al., 2023), we directly use the official open-source codes provided in their papers. For **GemNet** (Gasteiger et al., 2021) and **Equiformer** (Liao & Smidt, 2022), we use the implementation in open-source projects, the Open Catalyst Project (Chanussot* et al., 2021) and equiformer-pytorch², respectively. For fair comparison with SchNet and ET, we project the edge feature into the same shape as the distance feature in these models, and add the edge feature to the distance feature. It is also worth mentioning that due to the high complexity of angular features (DimeNet++, GemNet) and irreducible representations (MACE, Equiformer), these models need 2 GeForce RTX 2080 Ti GPUs for training on atomic graphs. Even so, some of them still fail to run the experiments due to the limitation of the GPU memory (i.e. GemNet and Equiformer).

G.2. Number of Parameters and Training Efficiency

We further provide the number of parameters and training efficiency for the baselines as well as our GET in Table 12.

²<https://github.com/lucidrains/equiformer-pytorch/tree/main>

³Equiformer is quite unstable and extremely memory intensive with large widths and depths.

Table 11. Hyperparameters for each baseline on each task.

hyperparameter	PPA	LBA	LEP	hyperparameter	PPA	LBA	LEP
SchNet							
d_h	128	64	64	max_n_vertex	1500	1500	1500
lr	10^{-3}	5×10^{-4}	10^{-3}	final_lr	10^{-4}	10^{-5}	10^{-4}
max_epoch	20	60	65	save_topk	3	5	5
SchNet-mix							
d_h	128	128	-	max_n_vertex	1500	1500	-
lr	5×10^{-5}	5×10^{-5}	-	final_lr	5×10^{-5}	5×10^{-5}	-
max_epoch	20	20	-	save_topk	3	3	-
DimeNet++							
d_h	128	64	64	max_n_vertex	1500	1500	1500
lr	10^{-3}	5×10^{-4}	10^{-3}	final_lr	10^{-4}	10^{-5}	10^{-4}
max_epoch	20	60	65	save_topk	3	5	5
EGNN							
d_h	128	64	64	max_n_vertex	1500	1500	1500
lr	10^{-3}	5×10^{-4}	10^{-3}	final_lr	10^{-4}	10^{-5}	10^{-4}
max_epoch	20	60	65	save_topk	3	5	5
ET							
d_h	128	64	64	max_n_vertex	1500	1500	1500
lr	10^{-3}	5×10^{-4}	10^{-3}	final_lr	10^{-4}	10^{-5}	10^{-4}
max_epoch	20	20	65	save_topk	3	3	5
ET-mix							
d_h	128	128	-	max_n_vertex	1500	1500	-
lr	5×10^{-5}	5×10^{-5}	-	final_lr	5×10^{-5}	5×10^{-5}	-
max_epoch	20	20	-	save_topk	3	3	-
GemNet							
d_h	128	64	-	max_n_vertex	1000	2000	-
lr	10^{-4}	10^{-3}	-	final_lr	10^{-4}	10^{-6}	-
max_epoch	20	10	-	save_topk	3	3	-
MACE							
d_h	128	64	-	max_n_vertex	1500	1500	-
lr	10^{-4}	10^{-3}	-	final_lr	10^{-4}	10^{-6}	-
max_epoch	20	20	-	save_topk	3	3	-
MACE-mix							
d_h	128	128	-	max_n_vertex	1500	1500	-
lr	5×10^{-5}	5×10^{-5}	-	final_lr	5×10^{-5}	5×10^{-5}	-
max_epoch	20	20	-	save_topk	3	3	-
Equiformer ³							
d_h	32	32	-	max_n_vertex	400	1000	-
lr	10^{-4}	10^{-3}	-	final_lr	10^{-4}	10^{-6}	-
max_epoch	10	10	-	save_topk	3	3	-
LEFTNet							
d_h	128	64	-	max_n_vertex	1000	2000	-
lr	10^{-4}	10^{-3}	-	final_lr	10^{-4}	10^{-6}	-
max_epoch	20	10	-	save_topk	3	3	-
LEFTNet-mix							
d_h	128	128	-	max_n_vertex	1000	1000	-
lr	10^{-4}	10^{-4}	-	final_lr	10^{-4}	10^{-4}	-
max_epoch	20	20	-	save_topk	3	3	-

When comparing GET to simpler yet weaker baselines (e.g., SchNet and EGNN), it is evident that GET may have more parameters and a slower training speed. However, it is crucial to note that GET exhibits competitive parameter and computation efficiency when compared with more complex yet stronger baselines, such as Equiformer, MACE, and LEFTNet. It's worth mentioning that a significant portion of parameters in GET is attributed to the Feedforward Neural Network (FFN) that projects latent features to higher dimensions in intermediate layers, aligning with the structure of vanilla Transformers. Without this part, GET has the least parameters among all the models. Nevertheless, adding FFN should not harm the efficiency much as the time cost mainly comes from message passing over edges, which is proportional to the number of edges, while time complexity of FFN is proportional to the number of nodes whose value is much smaller than the number of edges.

Moreover, the throughput of GET is comparable to both atom-level and hierarchical counterparts. This aligns with the design of the model, which considers both block-level and atom-level geometry. We also present the complexity analysis in Appendix D to further elucidate the efficiency of our approach, showing its linear complexity concerning the number of nodes in the graph. This characteristic indicates favorable scalability to large graphs in practical settings.

Table 12. Number of parameters and training speed for baselines and our GET.

Repr.	Model	PPA		LBA		LEP	
		Parameter	Sec. / Batch	Parameter	Sec. / Batch	Parameter	Sec. / Batch
Block	SchNet	0.25M	0.054	0.15M	0.040	0.14M	0.118
	DimeNet++	1.53M	0.233	0.40M	0.189	0.40M	0.263
	EGNN	0.43M	0.054	0.12M	0.035	0.11M	0.130
	ET	0.71M	0.072	0.20M	0.050	0.20M	0.133
	GemNet	1.35M	0.225	0.69M	0.179	-	-
	MACE	12.9M	0.296	1.97M	0.285	-	-
	Equiformer	0.56M	1.846	0.56M	1.364	-	-
	LEFTNet	1.57M	0.088	0.43M	0.068	-	-
Atom	SchNet	0.25M	0.109	0.15M	0.050	0.14M	0.123
	DimeNet++	1.56M	OOM	0.40M	0.435	0.40M	0.357
	EGNN	0.43M	0.145	0.12M	0.060	0.11M	0.139
	ET	0.71M	0.217	0.20M	0.079	0.20M	0.145
	GemNet	1.35M	OOM	0.69M	OOM	-	-
	MACE	12.9M	1.259	1.97M	0.535	-	-
	Equiformer	0.56M	OOM	0.56M	OOM	-	-
	LEFTNet	1.57M	0.472	0.43M	0.177	-	-
Hierarchical	SchNet	0.37M	0.127	0.21M	0.081	0.20M	0.088
	DimeNet++	3.07M	OOM	0.60M	0.622	0.60M	0.633
	EGNN	0.61M	0.143	0.17M	0.077	0.17M	0.100
	ET	1.00M	0.184	0.30M	0.104	0.29M	0.119
	GemNet	2.64M	OOM	1.37M	OOM	-	-
	MACE	25.7M	0.821	3.91M	0.426	-	-
	Equiformer	1.10M	OOM	1.10M	OOM	-	-
	LEFTNet	3.10M	0.307	0.85M	0.129	-	-
Unified	GET (w/o FFN)	0.23M	0.291	0.09M	0.193	0.20M	0.155
	GET	2.50M	0.339	0.69M	0.237	1.60M	0.192

H. Sensitivity to Width and Depth

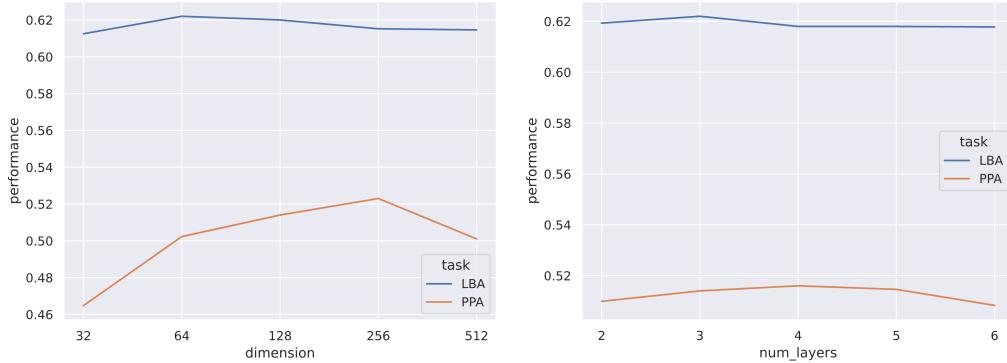


Figure 4. Performance with respect to the dimensions of the hidden layers (left) and the number of layers (right) on protein-protein affinity (PPA) and ligand-binding affinity (LBA).

We show the performance with respect to dimensions of the hidden layers and the number of layers in Figure 4 on protein-protein affinity (PPA) and ligand-binding affinity (LBA). The performance is not so sensitive to both width and depth on LBA, while it is relatively more sensitive to width on PPA. Nevertheless, the common trend is that performance increases first and then decreases with dimension getting larger or the network getting deeper, so it is still necessary to find the suitable size of the model that the dataset can support.

I. Sensitivity to K-Nearest Neighbors

We adjust the number of nearest neighbors (*i.e.* the hyperparameter k) and depict the change of performance in Figure 5. It reads that the performance slightly decreases when k is too small, but reaches saturation quickly as k increases. This suggests that further increases of k do not significantly impact performance. Further analysis for sensitivity on hidden size and number of layers is included in Appendix H.

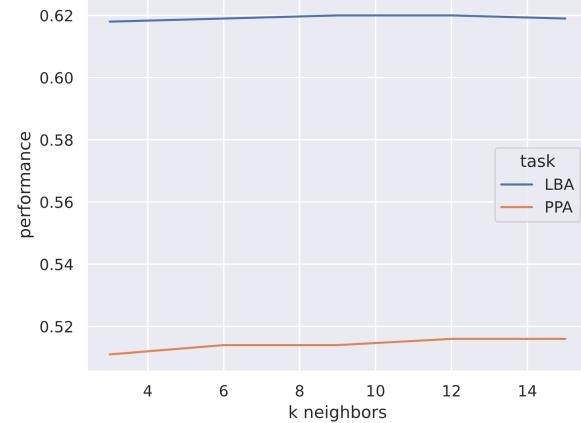


Figure 5. Performance (Pearson Correlation) with respect to the number of nearest neighbors on protein-protein affinity (PPA) and ligand-binding affinity (LBA).

J. Results for Protein Property Prediction

Though tasks concerning bare molecules are out of the main scope of this paper, we still include the results on protein property prediction here, in case the readers are curious of the capability of GET on tasks without molecular interactions. Following Zhang et al. (2022b), we evaluate our GET on **Enzyme Commission** (EC) number prediction, which requires predicting the characteristic of the given protein when acting as a catalyst. The EC numbers form 538 binary classification tasks, and comprises 15550, 1729, and 1919 proteins for training, validation, and testing, respectively. Aligning with Zhang et al. (2022b), we construct the residue-level graph of proteins and utilizes the edge features from IEConv (Hermosilla et al., 2020) and validate the models with F1 Max and AUPRC. Table 13 demonstrates that our GET achieves remarkable performance compared with the baselines, including the GearNet series (Zhang et al., 2022b) which are tailored for representation learning on proteins, indicating the strong modeling capability of our proposed GET.

Table 13. Evaluation on Enzyme Commission number prediction. Baseline results are borrowed from Zhang et al. (2022b).

Model	F1 Max↑	AUPRC↑
CNN (Shanehsazzadeh et al., 2020)	0.545	0.526
ResNet (Rao et al., 2019)	0.605	0.590
Transformer (Rao et al., 2019)	0.238	0.218
GVP (Jing et al., 2020)	0.489	0.482
GraphQA (Baldassarre et al., 2021)	0.509	0.543
New IEConv (Hermosilla & Ropinski, 2022)	0.735	0.775
DeepFRI (Gligorjević et al., 2021)	0.631	0.547
LM-GVP (Wang et al., 2022b)	0.664	0.710
GearNet-IEConv (Zhang et al., 2022b)	0.810	0.835
GearNet-Edge-IEConv (Zhang et al., 2022b)	0.810	0.843
GET-IEConv (ours)	0.835	0.866

K. Detailed Results of Universal Learning of Molecular Interaction Affinity

We provide the mean and the standard deviation of three parallel experiments on the universal learning of molecular interaction affinity (§ 4.3) in Tables 14 and 15.

Table 14. The mean and the standard deviation of three runs on protein-protein affinity prediction. Methods with the suffix ”-mix” are trained on the mixed dataset of protein-protein affinity and ligand binding affinity. The best results are marked in bold and the second best are underlined.

Repr.	Model	Rigid	Medium	Flexible	All
Pearson↑					
Block	SchNet	0.542 ± 0.012	0.504 ± 0.020	0.102 ± 0.019	0.439 ± 0.016
	SchNet-mix	0.553 ± 0.029	0.507 ± 0.011	0.093 ± 0.041	0.434 ± 0.011
	ET	0.575 ± 0.041	0.470 ± 0.024	0.087 ± 0.024	0.424 ± 0.021
	ET-mix	0.579 ± 0.028	0.502 ± 0.019	0.179 ± 0.044	0.457 ± 0.011
	MACE	0.621 ± 0.022	0.450 ± 0.027	0.307 ± 0.041	0.470 ± 0.015
	MACE-mix	0.572 ± 0.135	0.353 ± 0.040	0.170 ± 0.046	0.372 ± 0.042
	LEFTNet	0.563 ± 0.035	0.497 ± 0.018	0.202 ± 0.016	0.452 ± 0.013
Atom	LEFTNet-mix	0.522 ± 0.042	0.544 ± 0.009	0.152 ± 0.058	0.450 ± 0.008
	SchNet	0.592 ± 0.007	0.522 ± 0.010	-0.038 ± 0.016	0.369 ± 0.007
	SchNet-mix	0.625 ± 0.017	0.520 ± 0.021	-0.012 ± 0.049	0.421 ± 0.019
	ET	0.609 ± 0.023	0.486 ± 0.004	0.049 ± 0.009	0.401 ± 0.005
	ET-mix	0.618 ± 0.048	0.444 ± 0.027	0.057 ± 0.125	0.382 ± 0.029
	MACE	0.653 ± 0.066	0.499 ± 0.053	0.241 ± 0.061	0.463 ± 0.052
	MACE-mix	0.579 ± 0.009	0.484 ± 0.056	0.197 ± 0.021	0.444 ± 0.024
Hierarchical	LEFTNet	0.583 ± 0.080	0.510 ± 0.029	0.243 ± 0.091	0.448 ± 0.046
	LEFTNet-mix	0.688 ± 0.021	0.532 ± 0.021	0.244 ± 0.061	0.476 ± 0.023
	SchNet	0.542 ± 0.028	0.507 ± 0.020	0.098 ± 0.011	0.438 ± 0.017
	SchNet-mix	0.524 ± 0.031	0.515 ± 0.011	0.135 ± 0.077	0.429 ± 0.025
	ET	0.572 ± 0.051	0.498 ± 0.025	0.101 ± 0.093	0.438 ± 0.026
	ET-mix	0.494 ± 0.100	0.501 ± 0.007	0.130 ± 0.055	0.412 ± 0.035
	MACE	0.616 ± 0.069	0.461 ± 0.050	0.275 ± 0.032	0.466 ± 0.020
Unified	MACE-mix	0.525 ± 0.122	0.336 ± 0.067	0.060 ± 0.114	0.324 ± 0.076
	LEFTNet	0.533 ± 0.059	0.494 ± 0.026	0.165 ± 0.031	0.445 ± 0.024
	LEFTNet-mix	0.594 ± 0.059	0.543 ± 0.016	0.166 ± 0.109	0.472 ± 0.020
	GET (ours)	0.670 ± 0.017	0.512 ± 0.010	0.381 ± 0.014	0.514 ± 0.011
	GET-mix (ours)	0.697 ± 0.003	<u>0.533 ± 0.004</u>	0.389 ± 0.009	0.519 ± 0.004
Spearman↑					
Block	SchNet	0.476 ± 0.015	0.520 ± 0.013	0.068 ± 0.009	0.427 ± 0.012
	SchNet-mix	0.497 ± 0.044	0.527 ± 0.009	0.042 ± 0.031	0.426 ± 0.007
	ET	0.552 ± 0.039	0.482 ± 0.025	0.090 ± 0.062	0.415 ± 0.027
	ET-mix	0.550 ± 0.039	0.524 ± 0.019	0.188 ± 0.070	0.472 ± 0.019
	MACE	0.596 ± 0.047	0.450 ± 0.014	0.306 ± 0.029	0.466 ± 0.011
	MACE-mix	0.526 ± 0.129	0.366 ± 0.023	0.193 ± 0.030	0.370 ± 0.030
	LEFTNet	0.515 ± 0.039	0.492 ± 0.020	0.193 ± 0.023	0.452 ± 0.013
Atom	LEFTNet-mix	0.505 ± 0.048	0.543 ± 0.028	0.147 ± 0.086	0.439 ± 0.014
	SchNet	0.546 ± 0.005	0.512 ± 0.007	0.028 ± 0.032	0.404 ± 0.016
	SchNet-mix	0.557 ± 0.042	0.516 ± 0.033	0.036 ± 0.010	0.428 ± 0.022
	ET	0.582 ± 0.025	0.487 ± 0.002	0.117 ± 0.008	0.436 ± 0.004
	ET-mix	0.608 ± 0.040	0.453 ± 0.037	0.058 ± 0.135	0.394 ± 0.027
	MACE	0.619 ± 0.037	0.487 ± 0.049	0.221 ± 0.064	0.449 ± 0.052
	MACE-mix	0.504 ± 0.047	0.483 ± 0.064	0.226 ± 0.046	0.449 ± 0.029
Hierarchical	LEFTNet	0.524 ± 0.074	0.508 ± 0.038	0.189 ± 0.066	0.431 ± 0.046
	LEFTNet-mix	0.634 ± 0.060	0.518 ± 0.026	0.216 ± 0.044	0.455 ± 0.020
	SchNet	0.476 ± 0.017	0.523 ± 0.014	0.072 ± 0.021	0.424 ± 0.016
	SchNet-mix	0.487 ± 0.027	0.532 ± 0.007	0.096 ± 0.053	0.412 ± 0.024
	ET	0.547 ± 0.045	0.516 ± 0.019	0.100 ± 0.111	0.438 ± 0.029
	ET-mix	0.446 ± 0.116	0.499 ± 0.009	0.143 ± 0.090	0.408 ± 0.042
	MACE	0.580 ± 0.075	0.476 ± 0.048	0.282 ± 0.036	0.470 ± 0.016
Unified	MACE-mix	0.484 ± 0.098	0.340 ± 0.061	0.086 ± 0.125	0.324 ± 0.076
	LEFTNet	0.476 ± 0.082	0.494 ± 0.037	0.151 ± 0.019	0.446 ± 0.029
	LEFTNet-mix	0.572 ± 0.072	<u>0.553 ± 0.029</u>	0.143 ± 0.124	0.473 ± 0.029
Unified	GET (ours)	0.622 ± 0.030	0.533 ± 0.014	0.363 ± 0.017	0.533 ± 0.011
	GET-mix (ours)	<u>0.632 ± 0.025</u>	0.555 ± 0.008	0.391 ± 0.007	0.537 ± 0.003

Table 15. The mean and the standard deviation of three runs on ligand binding affinity prediction. Methods with the suffix “-mix” are trained on the mixed dataset of protein-protein affinity and ligand binding affinity. The best results are marked in bold and the second best are underlined.

Repr.	Model	LBA		
		RMSE↓	Pearson↑	Spearman↑
Block	SchNet	1.406 ± 0.020	0.565 ± 0.006	0.549 ± 0.007
	SchNet-mix	1.385 ± 0.016	0.573 ± 0.011	0.553 ± 0.012
	ET	1.367 ± 0.037	0.599 ± 0.017	0.584 ± 0.025
	ET-mix	1.423 ± 0.054	0.586 ± 0.012	0.567 ± 0.019
	MACE	1.385 ± 0.006	0.599 ± 0.010	0.580 ± 0.014
	MACE-mix	1.449 ± 0.050	0.590 ± 0.018	0.576 ± 0.010
	LEFTNet	1.377 ± 0.013	0.588 ± 0.011	0.576 ± 0.010
	LEFTNet-mix	1.433 ± 0.016	0.543 ± 0.005	0.532 ± 0.010
Atom	SchNet	1.357 ± 0.017	0.598 ± 0.011	0.592 ± 0.015
	SchNet-mix	1.365 ± 0.010	0.589 ± 0.006	0.575 ± 0.009
	ET	1.381 ± 0.013	0.591 ± 0.007	0.583 ± 0.009
	ET-mix	1.448 ± 0.122	0.566 ± 0.061	0.564 ± 0.059
	MACE	1.411 ± 0.029	0.579 ± 0.009	0.563 ± 0.012
	MACE-mix	1.420 ± 0.037	0.580 ± 0.030	0.568 ± 0.026
	LEFTNet	1.343 ± 0.004	0.610 ± 0.004	0.598 ± 0.003
	LEFTNet-mix	1.436 ± 0.019	0.579 ± 0.014	0.561 ± 0.016
Hierarchical	SchNet	1.370 ± 0.028	0.590 ± 0.017	0.571 ± 0.028
	SchNet-mix	1.403 ± 0.010	0.572 ± 0.004	0.554 ± 0.004
	ET	1.383 ± 0.009	0.580 ± 0.008	0.564 ± 0.004
	ET-mix	1.421 ± 0.032	0.569 ± 0.017	0.558 ± 0.017
	MACE	1.372 ± 0.021	0.612 ± 0.010	0.592 ± 0.010
	MACE-mix	1.432 ± 0.019	0.588 ± 0.011	0.572 ± 0.010
	LEFTNet	1.366 ± 0.016	0.592 ± 0.014	0.580 ± 0.011
	LEFTNet-mix	1.486 ± 0.081	0.556 ± 0.001	0.545 ± 0.005
Unified	GET (ours)	1.327 ± 0.005	<u>0.620 ± 0.004</u>	<u>0.611 ± 0.003</u>
	GET-mix (ours)	<u>1.329 ± 0.008</u>	0.622 ± 0.006	0.615 ± 0.008

L. Attention Visualization

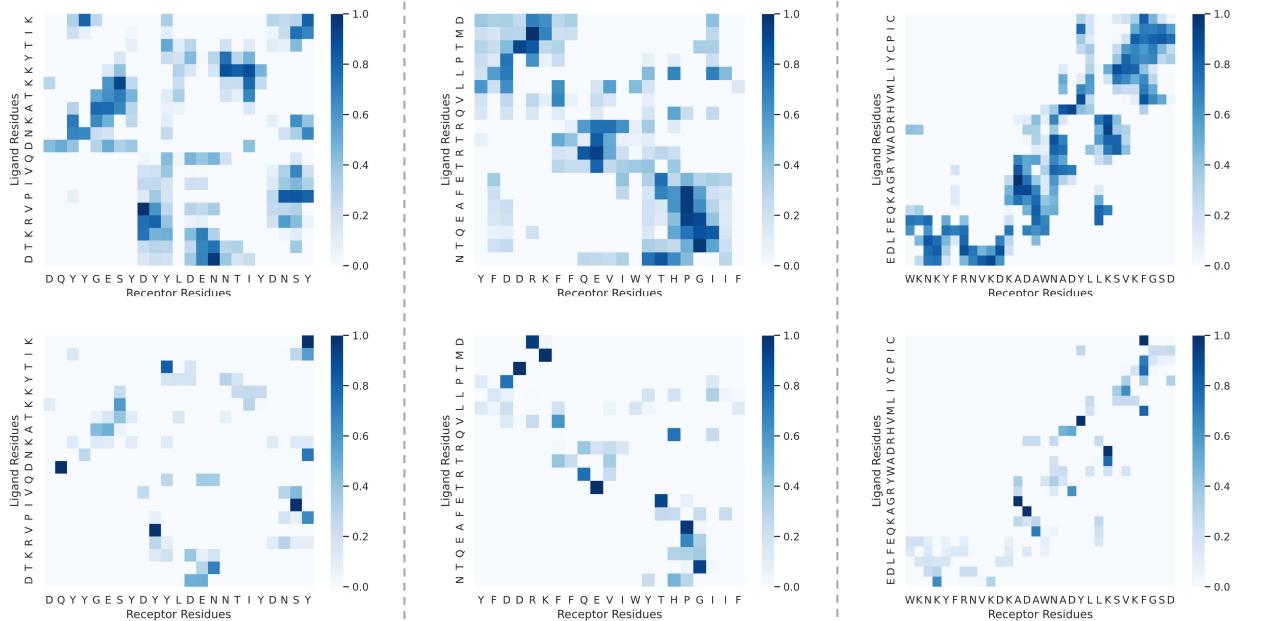


Figure 6. Attention weights of GET (upper row) and energy contributions given by Rosetta (lower row). PDB identities of the complexes are 1ahw, 1b6c, and 1gxd from left to right.

We visualize the attention weights between blocks on the interface of protein-protein complexes and compare them with the energy contributions calculated by Rosetta (Alford et al., 2017), which uses physics-based force fields. It is observed that the hot spots of attention weights largely agree with those predicted by Rosetta. We present three examples in Figure 6. The values are normalized by the maximum value (v_{\max}) and the minimum value (v_{\min}) in each figure (i.e. $v' = \frac{v - v_{\min}}{v_{\max} - v_{\min}}$)

M. Implications in Practical Applications

Firstly, we believe that the introduction of a unified molecular representation marks a significant stride in the field of geometric molecular representation learning. The challenge of data scarcity, primarily stemming from the high costs associated with wetlab experiments, has long hindered progress in this domain. Our approach posits that, despite the limited availability of data in specific molecular domains, the underlying interaction mechanisms are shared across diverse domains. Consequently, we propose a unified model capable of accommodating data from different molecular domains, presenting a promising solution to the challenge of data scarcity. The keypoint of this strategy lies in the invention of unified molecular representations and corresponding models that exhibit robust generalization across different molecular domains. Our work serves as a first step towards this vision, demonstrating that our GET benefits from training on mixed data across different domains and exhibits exceptional zero-shot ability even on entirely unseen domains.

Secondly, in practical applications such as affinity prediction, our model offers a valuable tool for leveraging abundant data from other domains to enhance predictive performance in specific, often cutting-edge, domains that suffer from data scarcity. We illustrate this feasibility through our zero-shot experiments on RNA-ligand affinity prediction in Section 4.3. By demonstrating the adaptability of our model to a new domain without the need for specific training data, we showcase the practical utility of our approach in scenarios where data is limited.

N. Detailed Results of Protein-Protein Affinity

We show the detailed mean and standard deviation of three runs on all test splits of protein-protein affinity in Table 16.

Table 16. The mean and the standard deviation of three runs on protein-protein affinity prediction. The best results are marked in bold and the second best are underlined. Baselines that fail to process atomic graphs due to high complexity are marked with OOM (out of memory)

Repr.	Model	Rigid	Medium	Flexible	All
Pearson↑					
Block	SchNet	0.542 ± 0.012	0.504 ± 0.020	0.102 ± 0.019	0.439 ± 0.016
	DimeNet++	0.487 ± 0.087	0.367 ± 0.043	0.152 ± 0.078	0.323 ± 0.025
	EGNN	0.437 ± 0.023	0.436 ± 0.028	0.094 ± 0.049	0.381 ± 0.021
	ET	0.575 ± 0.041	0.470 ± 0.024	0.087 ± 0.024	0.424 ± 0.021
	GemNet	0.480 ± 0.061	0.425 ± 0.051	0.086 ± 0.048	0.387 ± 0.023
	MACE	0.621 ± 0.022	0.450 ± 0.027	0.307 ± 0.041	0.470 ± 0.015
	Equiformer	0.630 ± 0.024	0.503 ± 0.015	<u>0.298 ± 0.017</u>	<u>0.484 ± 0.007</u>
	LEFTNet	0.563 ± 0.035	0.497 ± 0.018	0.202 ± 0.016	0.452 ± 0.013
Atom	SchNet	0.592 ± 0.007	0.522 ± 0.010	-0.038 ± 0.016	0.369 ± 0.007
	DimeNet++	OOM			
	EGNN	0.497 ± 0.027	0.452 ± 0.012	-0.054 ± 0.013	0.302 ± 0.010
	ET	0.609 ± 0.023	0.486 ± 0.004	0.049 ± 0.009	0.401 ± 0.005
	GemNet	OOM			
	MACE	<u>0.653 ± 0.066</u>	0.499 ± 0.053	0.241 ± 0.061	0.463 ± 0.052
	Equiformer	OOM			
	LEFTNet	0.583 ± 0.080	0.510 ± 0.029	0.243 ± 0.091	0.448 ± 0.046
Hierarchical	SchNet	0.542 ± 0.028	0.507 ± 0.020	0.098 ± 0.011	0.438 ± 0.017
	DimeNet++	OOM			
	EGNN	0.461 ± 0.018	0.440 ± 0.024	0.089 ± 0.051	0.386 ± 0.021
	ET	0.572 ± 0.051	0.498 ± 0.025	0.101 ± 0.093	0.438 ± 0.026
	GemNet	OOM			
	MACE	0.616 ± 0.069	0.461 ± 0.050	0.275 ± 0.032	0.466 ± 0.020
	Equiformer	OOM			
	LEFTNet	0.533 ± 0.059	0.494 ± 0.026	0.165 ± 0.031	0.445 ± 0.024
Unified	GET (ours)	0.670 ± 0.017	0.512 ± 0.010	0.381 ± 0.014	0.514 ± 0.011
Spearman↑					
Block	SchNet	0.476 ± 0.015	0.520 ± 0.013	0.068 ± 0.009	0.427 ± 0.012
	DimeNet++	0.466 ± 0.088	0.368 ± 0.037	0.171 ± 0.054	0.317 ± 0.031
	EGNN	0.364 ± 0.043	0.455 ± 0.026	0.080 ± 0.038	0.382 ± 0.022
	ET	0.552 ± 0.039	0.482 ± 0.025	0.090 ± 0.062	0.415 ± 0.027
	GemNet	0.420 ± 0.072	0.446 ± 0.059	0.066 ± 0.058	0.393 ± 0.027
	MACE	0.596 ± 0.047	0.450 ± 0.014	<u>0.306 ± 0.029</u>	0.466 ± 0.011
	Equiformer	0.560 ± 0.015	<u>0.530 ± 0.017</u>	0.251 ± 0.002	<u>0.496 ± 0.007</u>
	LEFTNet	0.515 ± 0.039	0.492 ± 0.020	0.193 ± 0.023	0.452 ± 0.013
Atom	SchNet	0.546 ± 0.005	0.512 ± 0.007	0.028 ± 0.032	0.404 ± 0.016
	DimeNet++	OOM			
	EGNN	0.450 ± 0.042	0.438 ± 0.021	0.027 ± 0.030	0.349 ± 0.009
	ET	0.582 ± 0.025	0.487 ± 0.002	0.117 ± 0.008	0.436 ± 0.004
	GemNet	OOM			
	MACE	<u>0.619 ± 0.037</u>	0.487 ± 0.049	0.221 ± 0.064	0.449 ± 0.052
	Equiformer	OOM			
	LEFTNet	0.524 ± 0.074	0.508 ± 0.038	0.189 ± 0.066	0.431 ± 0.046
Hierarchical	SchNet	0.476 ± 0.017	0.523 ± 0.014	0.072 ± 0.021	0.424 ± 0.016
	DimeNet++	OOM			
	EGNN	0.387 ± 0.023	0.461 ± 0.020	0.078 ± 0.043	0.390 ± 0.016
	ET	0.547 ± 0.045	0.516 ± 0.019	0.100 ± 0.111	0.438 ± 0.029
	GemNet	OOM			
	MACE	0.580 ± 0.075	0.476 ± 0.048	0.282 ± 0.036	0.470 ± 0.016
	Equiformer	OOM			
	LEFTNet	0.476 ± 0.082	0.494 ± 0.037	0.151 ± 0.019	0.446 ± 0.029
Unified	GET (ours)	0.622 ± 0.030	0.533 ± 0.014	0.363 ± 0.017	0.533 ± 0.011