

STUDY TITLE: Profiling Memory T cells Across Human Tissues

PROTOCOL NAME: CyTOF Detailed Lab Protocol

SUMMARY: Preparation and staining of cryopreserved single cell suspensions from human tissue samples for CyTOF analysis

1. Thaw cryopreserved PBMC samples

- a. Place cryovial in 37°C waterbath until ice is partially melted
- b. Transfer to 15ml conical tube
- c. Slowly dilute with 8mL of pre-warmed (37°C) Thaw Media (RPMI-1640 + 10% FBS + 50U/mL Benzonase)
- d. Centrifuge at 350g for 5mins
- e. Resuspend in 5mL of Thaw Media
- f. Count and assess viability
- g. Normalize cell counts to stain 1 million cells per tissue

2. Viability staining

- a. Resuspend samples in 500uL of Rh103 viability media (RPMI-1640 + 10% FBS + 1uM Rh103)
- b. Incubate for 20mins at 37°C
- c. Wash 1X in cell staining media (CSM: PBS + 0.1% BSA)

3. Barcode and pool cells from paired tissues

- a. Barcode cells from each tissue by staining with CD45 antibodies conjugated to monoisotopic cisplatin CD45_194Pt, CD45_195Pt, CD45_196Pt, CD45_198Pt (diluted 1:100 in CSM)
- b. Incubate for 15mins at 4°C
- c. Wash 2 times with 1mL of CSM
- d. Combine cells from all tissues into a single tube

4. Antibody panel staining

- a. Prepare 2X “Antibody Staining Panel”* in CSM and filter through 0.1micron Amicon centrifugal filter to remove aggregates (12,000g for 1 min).
- b. Resuspend each barcoded sample in 50uL CSM containing 1:50 dilution of TruStain FcX (Biolegend, Cat# 422302), and add 50uL of the filtered surface antibody cocktail
- c. Incubate for 30mins at 4°C
- d. Wash 2 times with 1mL CSM and remove supernatant
- e. Resuspend cell pellets in 500uL of 1X FoxP3 Fixation/Permeabilization buffer (eBioscience, Cat # 00-5523-00)

- f. Incubate for 30mins at 4°C
- g. Wash 2X with 1X Permash buffer (eBioscience, Cat # 00-5523-00)
- h. Resuspend pellet in 100uL of Permash buffer containing a 1:100 dilution of intracellular staining antibodies
- i. Incubate for 30mins at 4°C
- j. Wash 2X with 1X Permash buffer (eBioscience, Cat # 00-5523-00)
- k. Resuspend 400uL of 0.125nM Intercalator_Ir diluted in PBS containing 2% formaldehyde (freshly-diluted from 16% ampule stock)
- l. Incubate for 30mins at 4°C
- m. Wash 1X in 1mL PBS + 0.1% BSA
- n. Remove supernatant, resuspend cells in residual volume and store at 4°C until acquisition

5. Data acquisition

- a. Immediately prior to acquisition, wash cells with deionized water and count
- b. Resuspend in deionized water with a 1/20 dilution of EQ Four Element Calibration Beads (Fluidigm Inc.)
- c. Acquire on a CyTOF2 mass cytometer (Fluidigm Inc.) equipped with a SuperSampler fluidics system (Victorian Airships) at an event rate of <400evt/s

6. Initial data processing

- a. Normalize data using bead-based normalization algorithm in Helios acquisition software (Fluidigm Inc.)
- b. Exclude EQ Four Element Beads on the basis of Ir-193 and Ce-140
- c. Exclude dead cells on the basis of Rh103 positivity
- d. Debarcode cells from each tissue by Boolean gating based on corresponding Pt CD45 channels.

*ImmPort Reagent Set Name: “Miron et al. 2018 CyTOF 35 Panel”