

Network analysis of metabolic subsystems

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Abstract. Subsystems are parts of a metabolism that perform different important tasks in a cell. In this article, we explore these subsystems from a network science point of view. We attempt to find ways of detecting subsystems in a metabolic network using community detection algorithms. We use the Louvain modularity optimization algorithm and the Clauset-Newman-Moore algorithm as a baseline against which we compare the effectiveness of a motif-based community detection approach. We also attempt to explain the roles different motifs play in a metabolic network. We present the results on a metabolic network of the Chinese hamster ovary cell, a mammalian cell that is commonly used in biomedical research and in biotechnology.

1 Introduction

Since the turn of the century, life sciences have been evolving rapidly. Advances in data acquisition, storage and analysis technology have allowed scientist to gather immense amounts of data and build complex models from it [9]. These complicated models have brought people of various backgrounds, such as physics, mathematics and computer science into the field of biology.

One such field is network science, which is often used to analyze different kinds of networks that appear in the various subfields of modern biology, including ecology [16], systems biology [1] and neuroscience [18].

Metabolic networks [10] are used to model the metabolisms of various organisms. They are usually represented with a bipartite graph composed of two types of vertices: reactions and chemicals produced and consumed by the reactions. The edges in such a network connect chemicals to reactions. Furthermore, the edges are directed indicating whether the chemical was produced or consumed. A third kind of vertex can be added to represent enzymes that catalyze the reaction, but do not directly partake in it. Other commonly used representations are simplified representations, where one of the types of vertices is omitted [15].

In this article, we analyze the subsystems in a metabolic network of the Chinese hamster ovary cell. We explore different methods of detecting the subsystems, focusing on an approach based on network motifs [2]. We expect this algorithm to outperform other commonly used community detection algorithms such as Louvain optimization or the Clauset-Newman-Moore algorithm. We also analyse the frequencies of different motifs in the network and attempt to offer an interpretation of the results.

2 Related works

Jeong et. al. [10] offer a general overview of the organization and structure of metabolic networks. The authors compare the metabolic networks of 43 different organisms and find that metabolic networks belong to the class of scale-free networks. Moreover, they find that the network diameter is consistent across all of the analyzed networks, irrespective of the number of substrates found in the given species.

Holme et. al. [8] present a method to decompose metabolic networks into subnetworks based on the global network structure. They use a modified Girvan-Newman algorithm [5] to construct a hierarchical clustering tree. They argue that instead of looking at a particular partitioning of a network, we should look at the hierarchical clustering tree as whole, since well-defined subnetworks appear at different levels of the hierarchy.

A framework for clustering networks based on higher order structures (e.g. motifs) is introduced in [2]. The authors of the article present an algorithm that performs community detection by using three node motifs. It works by cutting the network into communities in a way that minimizes the ratio of the number of motifs of a particular type cut to the number of nodes in instances of the motif.

3 Methods

In this article, we analyse a metabolic network of the Chinese hamster ovary (CHO) cell. The CHO cell is frequently used in biological and medical research and in the production of biopharmaceuticals [7].

We use a whole-cell metabolic network of the Chinese hamster ovary (CHO) cell that was taken from the BiGG database [11, 7]. The original network contains 4,456

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metabolites that take part in 6,663 reactions. The reactions and metabolites are annotated with additional meta-data, such as name, subsystem, BiGG ID etc.

The network was simplified to a simple directed graph, where reactions are represented with nodes. If one reaction produces a metabolite that is used by another reaction, they are connected by an arc. This network has 6,663 nodes and 656,609 arcs. If we treat as an undirected network, it has 546,208 edges.

We use two commonly used network community detection algorithms, namely the Louvain modularity optimization algorithm [3] and the Clauset-Newman-Moore algorithm [4], as the baseline and compare them to motif-based clustering methods [2].

The Louvain modularity optimization algorithm is a method of community detection which optimizes the modularity of a network. Modularity [5] is a value which measures the density of edges within communities to edges outside communities. Optimizing this value should give us a good partitioning of a given network.

Clauset-Newman-Moore algorithm also optimizes the modularity measure. Starting with each vertex as a single community it repeatedly joins together two communities which will give the largest increase in modularity. It builds a dendrogram which represents the hierarchical decomposition of a network into communities at all levels.

The motif-based clustering method, described in Section 2, motif counting and the Clauset-Newman-Moore algorithm were taken from the SNAP library [13] and the Louvain modularity optimization algorithm was taken from the NetworkX Python library [6].

We present those algorithms because they scale well to large networks. Using Infomap [17] and the Girvan-Newman algorithm [5] proves to be unfeasible, since with this network, those algorithms do not terminate even after several days of running.

4 Results

The network has a very large connected component of 6,036 nodes, while the other components are very small, as they are composed of at most four nodes. The largest connected component contains a strongly connected component of 5,307 nodes, while the other nodes are isolated. These probably represent sources and sinks of the metabolism.

The network appears to have a scale-free structure. Its in-degree, out-degree and degree distributions are plotted in Figure 1. Some commonly used network properties, namely the clustering coefficient, the network’s effective diameter, its density and average degree, are presented in Table 1. The clustering coefficient is computed as

$$\langle C \rangle = \frac{3 \times \# \text{triangles}}{\# \text{connected triplets}} . \quad (1)$$

The effective diameter E_{90} measures the maximum number of hops needed to reach 90% of the network [12].

The graph density is defined as

$\langle C \rangle$	E_{90}	ρ	$\langle k \rangle$
0.012	15	0.015	194.1

Table 1. The clustering coefficient $\langle C \rangle$, effective diameter E_{90} , the density ρ and the average degree $\langle k \rangle$ of the network.

$$\rho = \frac{|E|}{|V|(|V| - 1)} , \quad (2)$$

where $|E|$ is the number of edges and $|V|$ is the number of vertices in the directed network.

First, we count the appearances of all 13 directed three-node motifs in the network. The significances of the motifs were calculated according to the following formula [14]:

$$Z_i = \frac{n_i - \mu_i}{\sigma_i} , \quad (3)$$

where n_i is the count of the i -th motif in the CHO metabolic network and μ_i and σ_i are the mean and standard deviation of the counts for ten instances of Erdős-Rény random networks.

These scores are presented in the second row of Table 2. Another option of evaluating the significances of the motifs would be to present the p -values, but since the numbers of occurrences of motifs in the CHO network are so dissimilar to the random graph, all those values would be effectively zero.

The normalized mutual information scores of the motif based clustering approach are listed in the last row of Table 2. We have calculated these scores by comparing the algorithm’s results and the actual subsystems listed in the network annotation. The results are not the best, but still much better than the Louvain and the Clauset-Newman-Moore method, listed in Table 3. We partly attribute the quality of the results to the fact that most biological networks tend to be noisy.

The results show that the motif M4 is the most over-represented motif in the network, while also being the motif that works best with clustering. The motif is a fully connected triangle, which represents groups of tightly coupled two-way chemical reactions. It is extremely rare in random graphs - in our case, it appeared somewhere from zero to five times, while the metabolic network contains over a million and a half. This motif might be important for the robustness of the metabolism. If there is a shortage of one of the metabolites, the triangular structure assures that it is quickly replaced by the neighboring reactions, effectively preventing such shortages from happening.

Another thing we see from the results is that triangular motifs, motifs in which all three nodes are connected in some way, with the exception of M5, tend to work better with clustering than wedge shaped motifs. This is also probably a consequence of tight coupling between the reactions that form a subsystem. The wedge shaped motifs probably mostly represent bridges between subsystems.














motif													
Z	-379.0	496.4	6,523	1,171,385	1,055	3,566	4,604	1,411	-867.2	2,599	1,293	1,387	40,286
NMI	0.44	0.40	0.48	0.64	0.23	0.43	0.46	0.11	0.09	0.09	0.20	0.23	0.42

Table 2. Motif significance Z compared to a random network and normalized mutual information score for motif clustering using each of the 13 motifs.

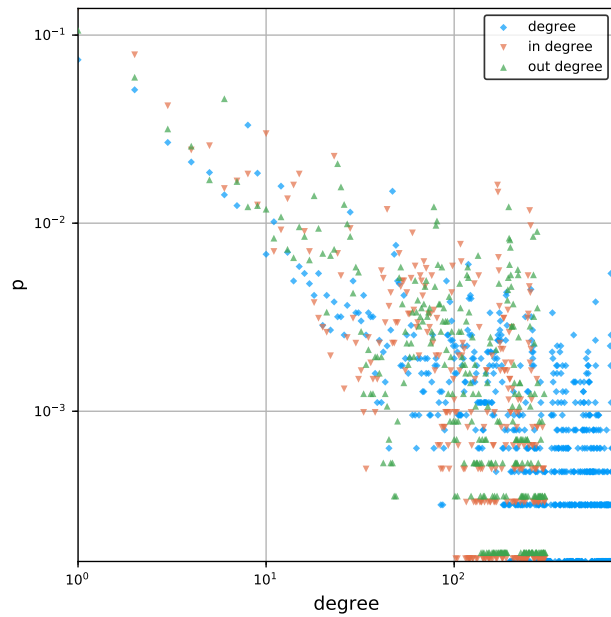


Fig. 1. The in-degree, out-degree and degree distributions of the network, plotted on a log-log scale.

Algorithm	Louvain Modularity	Clauset-Newman-Moore
NMI	0.1	0.27

Table 3. The normalized mutual information scores of the other algorithms we used.

5 Conclusion

Our analysis shows that three-node motifs play an important role in the structure of metabolic networks. The analyzed network contains a very large amount of fully-connected triplets of reactions. The number of such motifs found in this network is thousands and in one case even over a million times larger than what one would find in a random graph.

In terms of subsystem detection, the motif-based community detection algorithm outperforms traditionally used algorithms, while remaining computationally efficient. It works best when clustering with densely connected motifs, which again shows that such motifs play an important role in the structure of a metabolism.

Further work could include checking how this community detection algorithm performs with larger motifs and exploring the roles those larger motifs play in a metabolic network.

6 Authors contributions

All the authors were involved in the preparation of the manuscript. All the authors have read and approved the final manuscript.

References

1. Albert-Laszlo Barabasi and Zoltan N Oltvai. Network biology: understanding the cell's functional organization. *Nature reviews genetics*, 5(2):101–113, 2004.
2. Austin R Benson, David F Gleich, and Jure Leskovec. Higher-order organization of complex networks. *Science*, 353(6295):163–166, 2016.
3. Vincent D Blondel, Jean-Loup Guillaume, Renaud Lambiotte, and Etienne Lefebvre. Fast unfolding of communities in large networks. *Journal of statistical mechanics: theory and experiment*, 2008(10):P10008, 2008.
4. Aaron Clauset, Mark EJ Newman, and Cristopher Moore. Finding community structure in very large networks. *Physical review E*, 70(6):066111, 2004.
5. Michelle Girvan and Mark EJ Newman. Community structure in social and biological networks. *Proceedings of the national academy of sciences*, 99(12):7821–7826, 2002.
6. Aric A. Hagberg, Daniel A. Schult, and Pieter J. Swart. Exploring network structure, dynamics, and function using NetworkX. In *Proceedings of the 7th Python in Science Conference (SciPy2008)*, pages 11–15, Pasadena, CA USA, August 2008.
7. Hooman Hefzi, Kok Siong Ang, Michael Hanscho, Aarash Bordbar, David Ruckerbauer, Meiyappan Lakshmanan, Camila A Orellana, Deniz Baycin-Hizal, Yingxiang Huang, Daniel Ley, et al. A consensus genome-scale reconstruction of chinese hamster ovary cell metabolism. *Cell Systems*, 3(5):434–443, 2016.
8. Petter Holme, Mikael Huss, and Hawoong Jeong. Subnetwork hierarchies of biochemical pathways. *Bioinformatics*, 19(4):532–538, 2003.
9. B. Ingalls. *Mathematical Modelling in Systems Biology: An Introduction*. Applied Mathematics, University of Waterloo, 2012.
10. Hawoong Jeong, Bálint Tombor, Réka Albert, Zoltan N Oltvai, and A-L Barabási. The large-scale organization of metabolic networks. *Nature*, 407(6804):651–654, 2000.
11. Zachary A. King, Justin Lu, Andreas Dräger, Philip Miller, Stephen Federowicz, Joshua A. Lerman, Ali Ebrahim, Bernhard O. Palsson, and Nathan E. Lewis. Bigg models: A platform for integrating, standardizing and sharing genome-scale models. *Nucleic Acids Research*, 44(D1):D515, 2016.

12. Jure Leskovec, Jon Kleinberg, and Christos Faloutsos. Graph evolution: Densification and shrinking diameters. *ACM Transactions on Knowledge Discovery from Data (TKDD)*, 1(1):2, 2007.
13. Jure Leskovec and Rok Sosič. Snap: A general-purpose network analysis and graph-mining library. *ACM Transactions on Intelligent Systems and Technology (TIST)*, 8(1):1, 2016.
14. Ron Milo, Shai Shen-Orr, Shalev Itzkovitz, Nadav Kashtan, Dmitri Chklovskii, and Uri Alon. Network motifs: simple building blocks of complex networks. *Science*, 298(5594):824–827, 2002.
15. M. Newman. *Networks: An Introduction*. OUP Oxford, 2010.
16. Stephen R Proulx, Daniel EL Promislow, and Patrick C Phillips. Network thinking in ecology and evolution. *Trends in Ecology & Evolution*, 20(6):345–353, 2005.
17. Martin Rosvall and Carl T Bergstrom. Maps of random walks on complex networks reveal community structure. *Proceedings of the National Academy of Sciences*, 105(4):1118–1123, 2008.
18. Olaf Sporns. Contributions and challenges for network models in cognitive neuroscience. *Nature neuroscience*, 17(5):652–660, 2014.