

SYSTEMATIC MAP

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What is the available evidence for the range of applications of genome-editing as a new tool for plant trait modification and the potential occurrence of associated off-target effects: a systematic map

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Abstract

Background: Within the last decades, genome-editing techniques such as CRISPR/Cas, TALENs, Zinc-Finger Nucleases, Meganucleases, Oligonucleotide-Directed Mutagenesis and base editing have been developed enabling a precise modification of DNA sequences. Such techniques provide options for simple, time-saving and cost-effective applications compared to other breeding techniques and hence genome editing has already been promoted for a wide range of plant species. Although the application of genome-editing induces less unintended modifications (off-targets) in the genome compared to classical mutagenesis techniques, off-target effects are a prominent point of criticism as they are supposed to cause unintended effects, e.g. genomic instability or cell death. To address these aspects, this map aims to answer the following question: What is the available evidence for the range of applications of genome-editing as a new tool for plant trait modification and the potential occurrence of associated off-target effects? This primary question will be considered by two secondary questions: One aims to overview the market-oriented traits being modified by genome-editing in plants and the other explores the occurrence of off-target effects.

Methods: A literature search in nine bibliographic databases, Google Scholar, and 47 web pages of companies and governmental agencies was conducted using predefined and tested search strings in English language. Articles were screened on title/abstract and full text level for relevance based on pre-defined inclusion criteria. The relevant information of included studies were mapped using a pre-defined data extraction strategy. Besides a descriptive summary of the relevant literature, a spreadsheet containing all extracted data is provided.

Results: Altogether, 555 relevant articles from journals, company web pages and web pages of governmental agencies were identified containing 1328 studies/applications of genome-editing in model plants and agricultural crops in the period January 1996 to May 2018. Most of the studies were conducted in China followed by the USA. Genome-editing was already applied in 68 different plants. Although most of the studies were basic research, 99 different market-oriented applications were identified in 28 different crops leading to plants with improved food and feed quality, agronomic value like growth characteristics or increased yield, tolerance to biotic and abiotic stress, herbicide tolerance or industrial benefits. 252 studies explored off-target effects. Most of the studies were conducted using CRISPR/Cas. Several studies firstly investigated whether sites in the genome show similarity to the target sequence and secondly analyzed these potential off-target sites by sequencing. In around 3% of the analyzed potential off-target

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sites, unintended mutations were detected. Only a few studies conducted off-target analyses using unbiased detection methods (e.g. whole genome sequencing). No off-target effects that could be correlated to the genome-editing process were identified in these studies.

Conclusions: The rapid adoption in plant breeding was demonstrated by a considerable number of market oriented applications (crops and traits) described in publications worldwide. Studies investigating off-target effects are very heterogeneous in their structure and design. Therefore, an in-depth assessment regarding their weight of evidence is mandatory.

Keywords: New plant breeding techniques, Gene editing, Targeted genome modification, Mutagenesis, Unintended effects, Off-target mutation, Evidence map, Evidence synthesis

Background

Technological progress in agriculture and plant breeding has contributed significantly to a stable food supply and formed the basis for high yields as well as the production of high-quality agricultural products [1]. However, in an ever-changing world, new challenges will be encountered within the next decades. Aside the demands of the growing global population and limited fossil resources, climate change is a driver of breeding efforts as it is associated with increased extreme weather events like droughts or floods as well as changing dynamics of pests and diseases. Agriculture needs to ensure and increase the world agricultural production to serve extended demands with limited environmental resources like soil and water [2]. In contrast, intensification of agriculture causes considerable impact on nature, as naturally diverse landscapes are replaced by arable land for the cultivation of few plant species. Biodiversity is threatened through habitat loss and pesticide use, which in turn is considered to increase disease and pest pressure [1]. Additionally, these impacts of agriculture on the environment are becoming increasingly important in societal debates.

To meet all these challenges, improved crop varieties may be developed and integrated into a sustainable farming system considering their economic, environmental and social impacts [2]. Examples for a sustainable farming system are the preservation of natural soil fertility through suitable crop rotations, fertilization and plant protection according to the principles of integrated cultivation or a balanced and species-appropriate animal husbandry [3]. Additionally, plant breeding is of crucial importance to manage environmental impacts on cultivation systems by providing varieties resistant to plant diseases or pests, tolerant to abiotic stress and more broadly support a “greener production”—in time. This may reduce pesticide use and result in less intense management efforts (e.g. irrigation) [4]. Further yield improvement can increase the yield per hectare and may open land management options e.g. balance with areas for nature conservation [1]. Nevertheless, there is evidence that increased yields may lead to a rebound-effect

meaning that yield improvement conserves the rate of land clearance, but the effect is smaller than it could be [5]. One example for the rebound-effect in agriculture is the Green Revolution [6]. As a result of this revolution, yields increased, saving ecosystems from conversion to agricultural land. However, the effect was much smaller than expected. One explanation for this effect may be that increased productivity due to new technologies also increases the profitability of agriculture compared with alternative land use [6].

Plant breeding essentially relies on the utilization of genetic variation within the breeding material that can be used for crossing and selection to develop improved varieties. New genetic variation occurs naturally by spontaneous mutations that enable some individual plants in a population to adapt to changing environmental conditions. However, as the mutation rate is fairly low and random, plant breeders and scientists have artificially induced mutations for several decades [7]. The first generation of mutation breeding used chemical and physical mutagens to generate a plurality of nonspecific mutations. The increased mutation rate results in plants with a few positive, a lot of neutral and several negative characteristics. Thus, laborious backcrossing and selection steps are necessary in order to select for a target trait in a desired genetic background (maternal variety). Nevertheless, to date 3282 mutant varieties from 225 different plant species have been generated through undirected mutagenesis and officially but voluntarily registered [8].

Glossary: [1, 9, 10]

Backcrossing

Backcrossing is a crossing of a hybrid with one of its parents in order to achieve offspring that are genetically closer to the selected parent. This way, desired heterologous traits from the hybrid can be transferred into the genetic background of a parental line. Since crossing recombines all genes, many backcrosses are necessary to achieve considerable dilution of unwanted genes from the hybrid.

Mutagen

A mutagen is an agent that increases the mutation rate within an organism or cell, e.g. X-rays, gamma-rays or chemicals like ethyl methane sulfonate (EMS).

Mutation breeding

A plant breeding approach using mutagens to enhance genetic variation. The resulting random mutations can generate new gene variations with positive traits that can be selected for further breeding. However, several of these mutations are negative and diminish the viability of the plant.

Off-target effect due to genome editing

Unintended cleavage and mutations at untargeted genomic sites with similar but not identical sequences compared to the target site.

Selection

A process in breeding by which the breeder chooses only those individuals that show desired trait(s).

In 1983, the first recombinant DNA was delivered to plant cells using *Agrobacterium tumefaciens* [11, 12]. From this time on, it is possible to work at a single gene level with genetic material from any organism, generating plants that cannot be bred conventionally. Nevertheless, the induced mutations using chemical and physical mutagens as well as the “classical” transgenic approach show limited efficiencies and unintended side effects due to the random targeting [13].

In recent years, genome-editing techniques have been developed enabling a more precise modification of DNA sequences in a site-directed manner [10]. To date, genome-editing comprises three molecular approaches that efficiently induce targeted alterations in genomes: (i) Site-directed nucleases (SDN), including Meganucleases (MN), Zinc-Finger Nucleases (ZFN), Transcription Activator-Like Effector Nucleases (TALENs) and Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein (CRISPR/Cas), (ii) Oligonucleotide-directed mutagenesis (ODM) and (iii) base editing (BE). A detailed description of the single techniques is summarized in Additional file 1. Site-directed nucleases induce double-strand breaks (DSBs) in the DNA which are subsequently repaired by the autochthonous cellular mechanisms. The type of repair can be categorized in three main types [10, 14–16]:

1. No template is added and the DSB is repaired by autochthonous cellular mechanisms [in most cases non-homologous end joining (NHEJ)] resulting in small insertion-deletion (indel) mutations. This approach is defined as SDN1.
2. A repair template is added which, except for a few nucleotides, is identical to the sequences in which the DSB is introduced. Then, the DSB is repaired via homology-directed repair (HDR), causing nucleotide substitution or, depending on the template used, targeted indels of a specific size. This approach is defined as SDN2.
3. The repair template harbors a recombinant DNA sequence additional to the homologous sequences in which the DSB is made and the break is repaired via HDR, resulting in more complex alterations, i.e. the insertion of foreign genes. This approach is defined as SDN3.

The Oligonucleotide-Directed Mutagenesis technique does not induce a DSB. Instead the introduced oligonucleotide binds to the targeted DNA sequences and this sequence is then modified by the cellular mechanism of mismatch repair [10]. Base editing is a recently developed approach enabling a targeted switch of DNA bases in a given frame into another without any DSB [17].

Genome-editing offers substantial advantages compared to previous mutation breeding techniques and conventional genetic engineering in terms of speed and precision. It provides the opportunity to selectively mutate or modify one or a few genes (SDN1, SDN2). In addition, it is now possible to precisely modify or selectively replace (SDN3) entire genes from both closely as well as distantly related organisms [10]. By the use of conventional genetic engineering, traces of recombinant DNA, from the used gene shuttle (e.g. bacteria, virus or plasmid), persist in the modified organism leading to clearly characterized genetically modified organisms. In contrast, by applying genome-editing, it is possible to modify crops without inserting foreign DNA sequences at all [18]. Therefore, some countries like USA, Canada, Brasilia, Argentina and others reduced the regulatory burden for plant breeders [19]. Due to the simplicity, time-saving and cost-effective application of genome-editing, it has already been applied in a wide range of cultivars. Genome-editing has been used for:

- i. Analyzing gene functions (e.g. effect of the RAV2 gene for salt stress in rice [20]).
- ii. Improvement of product quality (e.g. improved oil quality in soybean [21]).
- iii. Development of disease resistant varieties (e.g. virus resistant cucumber [22]).

- iv. Developing of herbicide tolerant varieties (e.g. resistance to the herbicide chlorsulfuron in canola [23]).
- v. Improved adaption to abiotic stress, (e.g. drought tolerance in maize [24]).

Even in plants like hexaploid wheat, that were so far largely inaccessible for targeted genetic alterations, the simultaneous mutation of all six alleles was successfully performed [25]. These successful applications are opening up new dimensions for the scientific plant breeding and agricultural community.

Compared to randomly induced mutations by chemicals or irradiation, the number of unintended mutations (off-target effects) is broadly reduced by genome-editing techniques [10]. Nevertheless, their application does not completely or per se exclude the occurrence of off-target effects. Off-target effects caused by genome editing most likely occur in DNA sequences that are similar (not identical) to the targeted one but are located at another site in the genome. Mainly, they occur due to the lack of exclusiveness and/or length of the recognition site [10]. When analyzing off-target effects, one distinguishes between biased and unbiased detection methods [26]. To date, the predominant approach for identifying off-target effects is the biased approach consisting of two steps: (i) Using sequence alignment programs, sites in the genome with high similarity to the target sequence are identified, which are designated as potential off-target sites. Several different tools like BLAST [27], Cas-OFFinder [28] or CRISPR-P [29, 30] are used to identify these potential off-target sites. (ii) The identified individual DNA sequences (potential off-target sites) are then analyzed for undesired mutations (off-target effects) using various detection methods like mismatch-sensitivity endonuclease assays, Sanger sequencing or targeted deep sequencing [31]. All detection methods have their specific advantages and disadvantages which are addressed in several reviews (e.g. [26, 32]). In contrast to the biased detection methods, unbiased ones are used to identify off-target effects in a completely unrestricted way. Therefore, it requires genome-wide sequencing to identify off-target mutations anywhere in the genome and de novo define off-target sites [26, 32]. Depending on the detection method being used, the results of identifying off-target mutations vary widely. Although genome-editing techniques induce much less off-target effects compared to classical mutagenesis techniques, these are an important point of criticism as they may possibly cause genomic instability, cytotoxicity and cell death [33–35].

This systematic map facilitates an objective debate by informing interested stakeholder communities in a transparent and retraceable manner about the status of research, the progress of genome-editing in plants and

the available evidence for the potential occurrence of associated off-target effects. Furthermore, risk assessors and decision makers are depending on the provision of a reliable body of evidence to support conclusions about potential risks being associated with the application of genome-editing. Thus, an overview of the available evidence on the occurrence of off-target effects could be of crucial importance.

Stakeholder engagement

The systematic map question, the secondary questions and the scope was designed by the review team reflecting discussions with policy makers, authorities, regulators and academia requesting a broad overview on the available evidence about the application of genome editing in plants and the potential occurrence of off-target effects. Throughout the review process there was no stakeholder engagement. As indicated in the systematic map protocol the results of the map were discussed on a conference with different stakeholders from this field, including besides others plant breeders, federal authorities, academia, farmer organizations and processing industry. Stakeholder remarks are taken into account when preparing a systematic review based on the results of this map.

Objectives of the map

As genome-editing techniques are a promising tool to revolutionize plant breeding, they are of particular relevance to scientists, breeders, farmers but also to decision and policy makers with regards to the broader agricultural management and future challenges. Therefore, we wanted to provide a comprehensive and transparent overview of the available evidence base concerning the effects of genome-editing in plants. The main objectives were:

- Overview of the traits modified by genome-editing in model plants as well as in crops produced for agricultural production.
- Overview of the available evidence about the occurrence of off-target effects due to the use of genome-editing techniques in model plants as well as in crops produced for agricultural production.
- Identification of the geographical distribution of genome-editing activities in plants worldwide.
- Identification of the volume of the available literature, evidence clusters and key characteristics of the evidence base to inform interested stakeholder communities.
- Identification of knowledge gaps concerning the occurrence of off-target effects in order to inform

- decision makers which future research might be needed for a risk assessment.
- Assessment whether a specific section of the available evidence base is suitable for an in-depth analyses by a systematic review.

The primary question of the systematic map was: “What is the available evidence for the range of applications of genome-editing as a new tool for plant trait modification and the potential occurrence of associated off-target effects”?

To answer this primary question, it was subdivided into two secondary questions related to (1) the traits modified by genome-editing and (2) the occurrence of off-target effects due to the use of genome-editing.

Secondary question one

“What are the traits modified by genome-editing in model plants as well as in crops produced for agricultural production?”

Population: Any model plant or crop produced for agricultural production.

Intervention: One of the following genome-editing techniques was used to induce an alteration in the plant genome: Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein (CRISPR/Cas), Transcription Activator-Like Effector Nucleases (TALENs), Meganucleases (MN), Zinc-Finger Nucleases (ZFN), Oligonucleotide-Directed Mutagenesis (ODM), base editing (BE).

Outcome: The alteration of the genome (i.e. insertion, deletion or substitution of nucleotides) induced by the use of a genome-editing technique.

Secondary question two

“What is the available evidence for the potential occurrence of associated off-target effects due to the use of genome-editing in model plants as well as in crops produced for agricultural production?”

Population: Any model plant or crop produced for agricultural production.

Intervention: One of the following genome-editing techniques was used to induce an alteration in the plant genome: Clustered

Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein (CRISPR/Cas), Transcription Activator-Like Effector Nucleases (TALENs), Meganucleases (MN), Zinc-Finger Nucleases (ZFN), Oligonucleotide-Directed Mutagenesis (ODM), base editing (BE). The alteration of the genome (i.e. insertion, deletion or substitution of nucleotides) induced by the use of a genome-editing technique. Additionally, the occurrence of off-target effects was assessed.

Outcome:

Methods

The methods used to conduct this systematic map were based on the Collaboration for Environmental Evidence (CEE) systematic review guidelines [36]. Detailed information about the methods used to perform this systematic map are presented in the published protocol [37]. A brief summary of these methods is provided here.

Search for articles

The search string was composed of two parts: The first part defined the population of interest comprising less specific terms like crop, plant or seed as well as specific model plants and crops including their English and Latin names. The second part defined the intervention, i.e. the genome-editing technique applied to induce an alteration in the plant genome (CRISPR, TALENs, ZFN, MN, ODM or BE). To test the comprehensiveness of the search strategy a scoping search was carried out and the identified records were tested against an a priori defined test library with articles of known relevance. Details on search settings and subscriptions can be found in Additional file 2. The following bibliographic databases were searched whereby the search string was adapted to the specific needs of each database to which it was applied to:

- Scopus
- PubMed
- Science direct
- Agris
- Web of Science (WoS)
- Biological Abstracts
- BIOSIS Previews
- CAB Abstracts
- SciELO Citation Index

In addition, Google Scholar (<https://scholar.google.com>) was searched using 30 different combinations

of different (model) plants and genome-editing terms. The first 20 hits organized by relevance, of each search term were examined for relevance. Furthermore, a total amount of 47 web pages of companies working with genome-editing and the USDA database “Am I regulated?” (<https://www.aphis.usda.gov/aphis/ourfocus/biotechnology/am-i-regulated>) were searched to identify grey literature. Finally, the bibliographies of 107 review articles identified by the literature search were screened to identify further relevant papers. In May 2018, the search string was applied in order to identify articles published after 1996, when the first study about a genome-editing technique was published.

Article screening and study eligibility criteria

Screening process

Before applying the selection criteria at title/abstract and then at full text level, a consistency check was conducted by all four participating reviewers aiming to determine the inter-reviewer agreement. The level of agreement was tested formally using a kappa test [38]. 100 references retrieved by the search were randomly explored at title/abstract level leading to a kappa value of 0.48. After discussing all disagreements, a second consistency check was carried out using another randomly allocated 100 references resulting in a kappa value of 0.71. After title/abstract screening, potentially relevant articles were checked at full text level. A list of unobtainable articles (Additional file 3) and articles excluded at full text level with the reason for exclusion (Additional file 4) are provided. For conducting the consistency check as well as for the screening process at title/abstract and full text level the open-access and non-profit database CADIMA was used [39]. Two members of the review team are authors of a few articles retrieved by the review process. However, as none of these papers comprise primary data, their articles were excluded at title/abstract level. Nevertheless, the two coauthors routinely screened literature covering the issues addressed in this map.

Eligibility criteria

An article had to meet all the following inclusion criteria in order to enter into the systematic map:

Eligible populations:

Any model plant or crop produced for agricultural production as well as higher fungi was used.

Eligible interventions:

At least one of the following genome-editing techniques was used to induce an alteration in

the plant genome: Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein (CRISPR/Cas), Transcription Activator-like Effector Nuclease (TALENs), Zinc-Finger Nuclease (ZFN), Meganuclease (MN), Oligonucleotide-Directed Mutagenesis (ODM), base editing (BE).

Eligible outcome for secondary question one:

An alteration in the plant genome was reported (insertion, deletion or substitution) due to the use of a genome-editing technique.

Eligible outcome for secondary question two:

The occurrence of off-target effects was assessed.

Eligible type of data:

Only those references were included which comprise primary data referring to the use of a genome-editing technique to induce a sequence alteration in the plant genome.

Eligible languages:

References in German and English languages were included. Articles in other languages were included when besides title and abstract, further parts of the article, like figures or tables, were in English or German and the provided information allowed for a definite judgment of their relevance.

Study validity assessment

The aim of this systematic map was to provide a broad overview of the progress on genome-editing in plants as well as the examination of associated off-target effects. The validity of the included studies (critical appraisal)

was not assessed. However, in order to facilitate the decision on the potential of a subsequent systematic review, data being indicative for the validity of an included study were extracted.

Data coding strategy

Articles in which several genome-editing techniques were applied, different plants were used or different genes were addressed have been subdivided in distinct studies. While articles were screened for relevance at title/abstract and full text level, the relevant data were finally extracted at study level.

The data of each included study was extracted in one row in the excel file for the following superordinate categories:

1. Bibliographic information (reference type, authors, year of publication, title, abstract, keywords, periodical, issue number, page range, volume, DOI/ISBN, corresponding author and the name of the country the corresponding author is located).
2. Information answering secondary question one about traits modified by genome-editing (genome-editing technique, plant species, sequence identifier, trait, type of alteration, progress in research, key topic).
3. Information answering secondary question two about the occurrence of off-target effects due to the use of genome-editing (search for off-target effects, prediction of potential off-target effects (*in silico*), the prediction method used, identification of potential off-target sites, detection of off-target effects (biased/unbiased), detection method, amount of identified off-target effects (biased/unbiased)).

Data were extracted according to the systematic map protocol [37]. Data from a subset of 20 studies were extracted by two reviewers independently to assess the consistency of the extraction process across the reviewers. Discrepancies were discussed and clarified within the whole review team. Then, the data of the other studies were extracted by one reviewer and cross-checked by a second one to minimize the introduction of human error. In case of missing data, “no information” has been noted under the respective category. If data were in another language than English or German, it is stated as “language”.

Data mapping method

The overview of research activities is provided in a narrative report and visualized in tables and figures. In addition, relevant studies and the extracted data are catalogued in Additional file 5 (to answer secondary question one) and Additional file 6 (to answer secondary question

two) as well as provided in a searchable database that is freely accessible on the web page <https://www.dialog-gea.de/de/service/repositorium>.

Deviations from the systematic map protocol

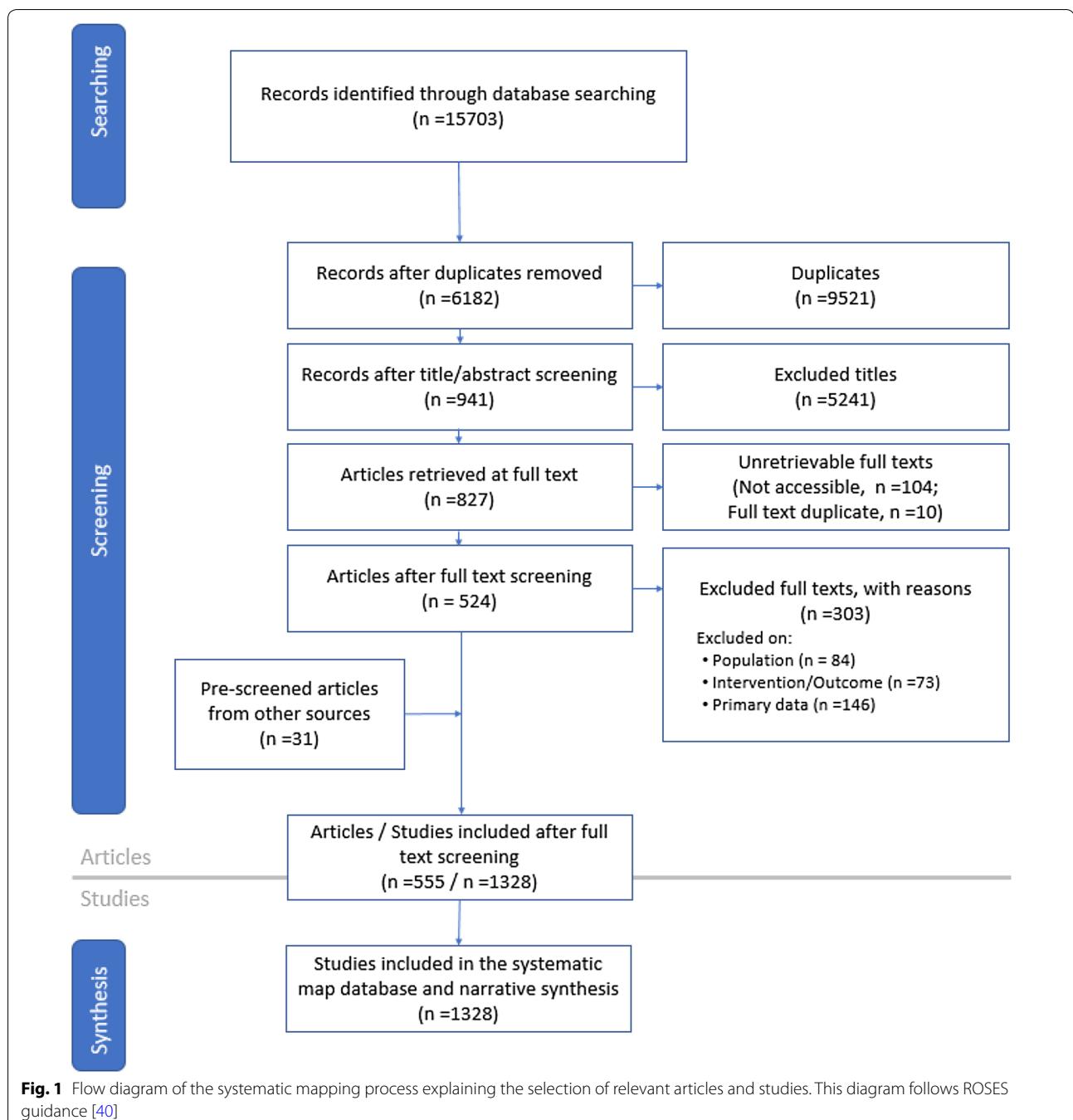
The methods used in this map deviate from the protocol in the following aspects:

- i. Contrary to what was stated in the protocol, the recently developed base editing method was not excluded. To identify all base editing studies the second part of the search string (intervention) was extended by the terms “base-editing” and “base editing”.
- ii. Articles in other languages than English or German were included if further parts than title and abstract of the article like figures or tables were in English or German and the provided information allowed for a definite judgment of their relevance.
- iii. In the course of data extraction we noticed that it was not possible to properly categorize the progress in research to the three classes indicated in the systematic map protocol [37]. Therefore, we decided to classify the studies as either basic research or market-oriented application. To be flagged as market-oriented, a study had to meet three criteria:
 1. Genome-editing was applied in an agricultural crop.
 2. A trait was addressed that may be of interest for commercialization (market-oriented trait).
 3. The targeted trait is expressed in the edited plant grown.
 All studies that did not meet all three criteria were classified as basic research.
- vi. Contrary to what was stated in the protocol, no EndNote database of all studies included in the systematic map was attached to this systematic map. Instead, one excel file for each secondary question was provided as additional file containing all included studies and the extracted data.

Results

Review descriptive statistics

Figure 1 presents the systematic mapping process of articles and studies in a flow diagram. From January 1996 until May 2018, in total 15,703 records were identified from ten bibliographic databases, Google Scholar, the targeted search on 47 company web pages, the USDA-database “Am I regulated?” and the screening of 107 review articles. After removing duplicates (n=9521),



6182 articles remained and were screened on title/abstract level. Main reasons for exclusion at this stage were the application of genome-editing in animals and the absence of primary data. 941 articles passed the inclusion criteria on title/abstract level or were rated as “unclear” and remained included for full text screening. The application of the inclusion criteria at full text level resulted in 524 relevant articles. Searching for grey

literature on company websites and websites from governmental authorities identified another 31 further relevant documents and web pages that were considered during data extraction. Out of these 555 records, a total amount of 1328 studies were extracted and formed the basis for this systematic map. A list of all studies and the extracted data to answer secondary question one is provided in Additional file 5. Additional file 6 provides a list

of all studies and the extracted data to answer secondary question two. A ROSES reporting form is included in Additional file 7.

General overview of the application of genome-editing in model plants and crops

Studies per year

In the mapping period between January 1996 and May 2018, a total amount of 1328 studies were identified. As shown in Fig. 2, the number of studies using TALENs, ZFN, ODM, MN and BE has remained on a relatively low level. In contrast, the number of studies on CRISPR/Cas has risen sharply soon after the system was applied for the first time in plants in 2013. Nearly 85% of the studies were published since 2015 indicating the rapid dissemination and development of these techniques within the last few years.

As shown in Table 1, 26 studies used MN, 27 studies used ODM, 42 studies used BE, 73 studies used ZFN, 128 studies used TALENs and 1032 studies used CRISPR/Cas. When using CRISPR/Cas, the most frequently used nuclease was Cas9 ($n=986$) followed with a large distance by Cas12a (also known as Cpf1) ($n=46$).

Geographical distribution of genome-editing studies

The number of studies per country was calculated based on the country the corresponding author is located at. In

case of grey literature, the study was accounted based on the country the company is located at. Multiple assignments are possible if the corresponding author is affiliated with institutions in different countries or if more than one author was indicated being corresponding author. Therefore, the sum of studies accounted for different countries is higher ($n=1494$) than the total amount of studies identified ($n=1328$). Asia is the leading continent when applying and publicizing genome-editing in plants ($n=784$ studies, 53%) followed by North America ($n=508$ studies, 34%), Europe ($n=189$ studies, 13%), Australia ($n=6$ studies, <1%), South America ($n=4$ studies, <1%) and Africa ($n=3$ studies, <1%) (Fig. 3). In total, publications from 33 countries were identified. As shown in Fig. 3, China has a substantial lead in the number of studies ($n=599$ studies, 40%) followed by the USA ($n=487$ studies, 33%), Japan (94 studies, 6%) and Germany ($n=88$ studies, 6%).

Genome-editing applications in model plants and crops

Around two-third of the studies ($n=907$; 68%) were conducted on agricultural crops and one-third ($n=421$; 32%) on model organisms. However, it is worth noting that rice is an important crop plant but it is also used as model species because of its genome size and because embryogenic rice cultures can be easily prepared, transformed and rapidly regenerated into fertile plants. In total, 51

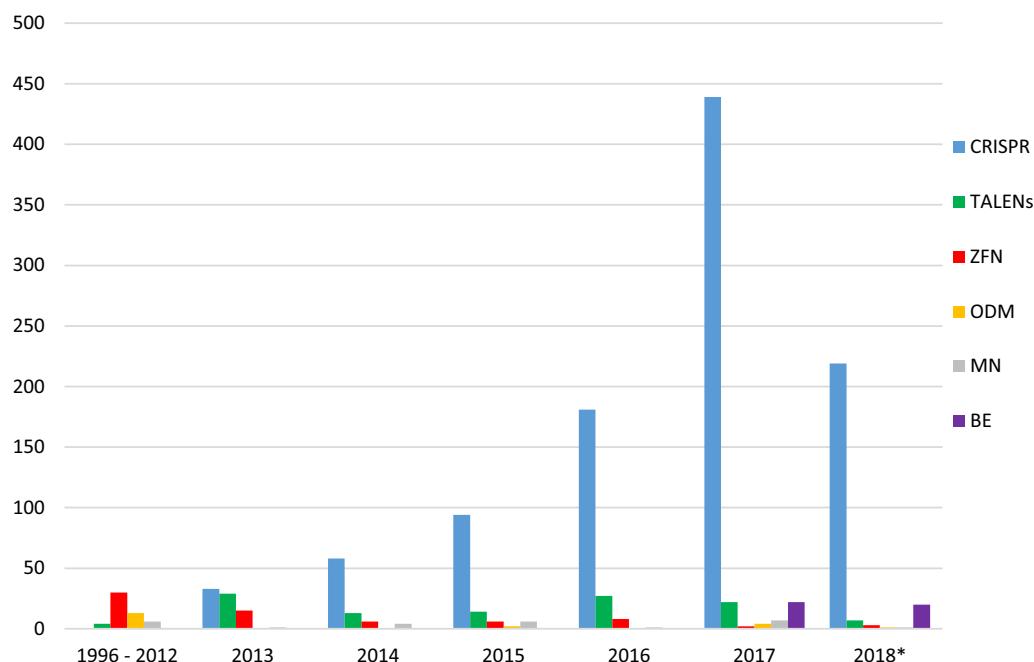


Fig. 2 The number of genome-editing applications published per year. *Only January–May 2018; CRISPR/Cas Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein, TALENs Transcription Activator-Like Effector Nucleases, ZFN Zinc-Finger Nucleases, ODM Oligo-Directed Mutagenesis, MN Meganucleases, BE base editing

Table 1 Heat map showing number of studies performed with the different genome-editing techniques (rows) and the year the studies were published (columns) (1996–May 2018)

	CRISPR/Cas	TALENs	ZFN	ODM	MN	BE	# studies
1996	0	0	0	0	0	0	0
1997	0	0	0	0	0	0	0
1998	0	0	0	0	0	0	0
1999	0	0	0	4	0	0	4
2000	0	0	0	1	0	0	1
2001	0	0	0	0	0	0	0
2002	0	0	0	0	0	0	0
2003	0	0	0	6	1	0	7
2004	0	0	0	1	0	0	1
2005	0	0	2	0	0	0	2
2006	0	0	1	1	0	0	2
2007	0	0	0	0	0	0	0
2008	0	0	4	0	0	0	4
2009	0	0	10	0	2	0	12
2010	0	0	5	0	1	0	6
2011	0	2	10	0	0	0	12
2012	0	2	1	0	2	0	5
2013	33	29	15	0	1	0	78
2014	58	13	6	0	4	0	81
2015	94	14	6	2	6	0	122
2016	181	27	8	0	1	0	217
2017	439	22	2	4	7	22	496
2018*	219	7	3	1	1	20	251
No information	8	12	0	7	0	0	27
# studies	1032	128	73	27	26	42	1328

*Only January–May 2018

CRISPR/Cas Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein, TALENs Transcription Activator-Like Effector Nucleases, ZFN Zinc-Finger Nucleases, ODM Oligo-Directed Mutagenesis, MN Meganucleases, BE base editing

species and subspecies of different agricultural crops and 17 species/subspecies of model plants were under investigation, with the majority of studies focusing on rice ($n=465$) followed by the model organisms *Arabidopsis* ($n=218$) and tobacco ($n=107$). Besides to these, tomato ($n=84$) is most commonly studied followed by maize ($n=77$), wheat ($n=63$) and soybean ($n=53$) (Fig. 4).

Type of alteration in the plant genome

As shown in Fig. 5, the majority of studies ($n=1223$; 92%) describe the induction of point mutations or indels comparable to spontaneous mutations or undirected mutagenesis. This was mainly achieved with SDN1 ($n=1154$) where no repair template was added and the DSB was repaired by NHEJ. Additionally, the induction of point mutations (PM) using the ODM technique ($n=27$) or BE ($n=42$) leads to point mutations comparable to SDN1. Only 36 studies (3%) added a repair template that,

except for a few nucleotides, was identical to the targeted sequence in which the DSB was introduced leading to a DSB repair via homology-directed repair (SDN2). In 68 studies (5%), a repair template was added that harbors a recombinant DNA sequence additional to the homologous sequences and the DSB was repaired via homology-directed repair (SDN3).

Secondary question 1: “What are the traits modified by genome-editing in model plants as well as in crops produced for agricultural production?”

In total, 193 studies were allocated as market-oriented applications. However, different scientists studied the same crop species and trait. Considering this, a total amount of 99 different applications in 28 different plant species remained. These market-oriented applications build the basis to answer secondary question 1. Figure 6 categorizes the market-oriented applications to different

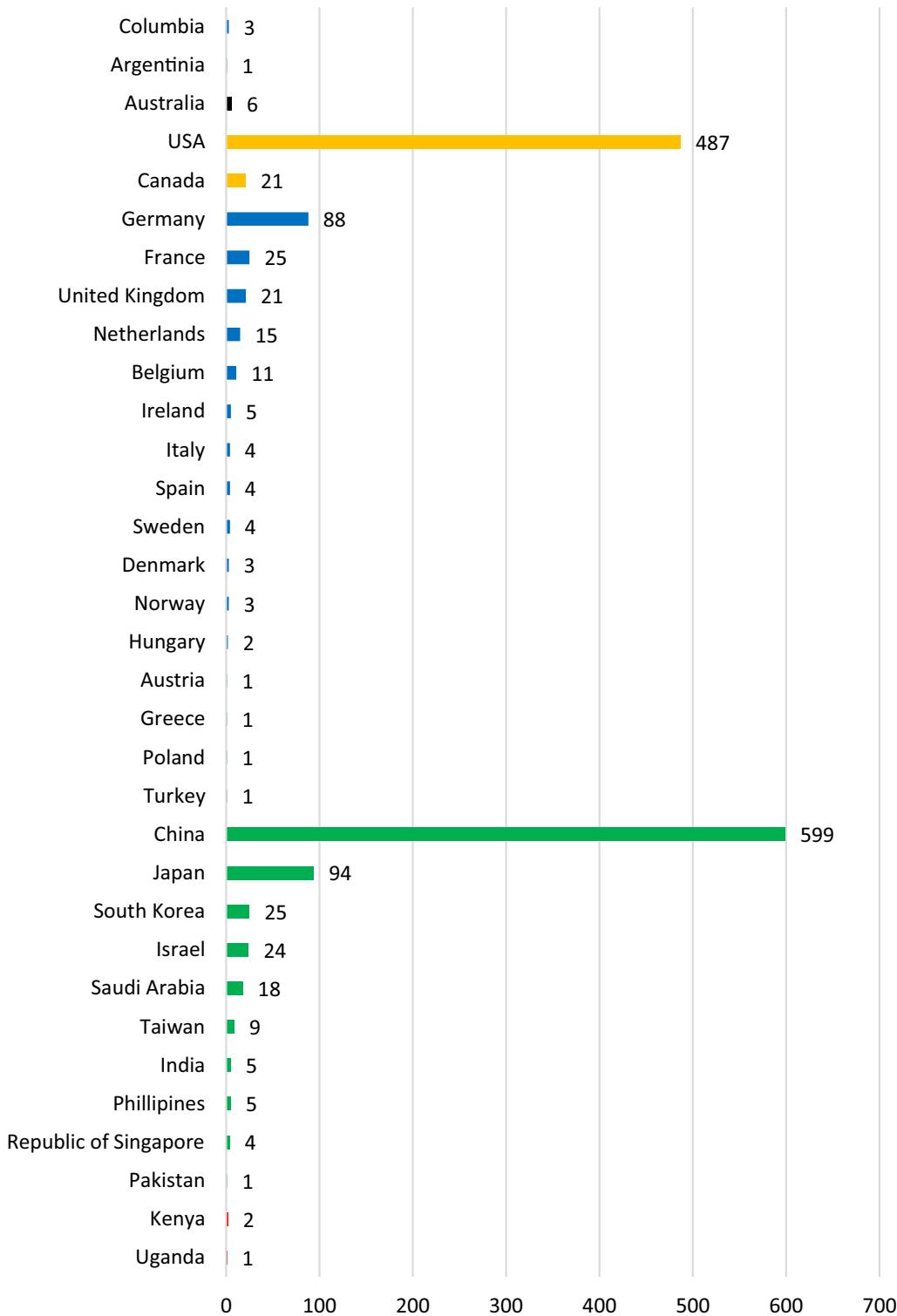
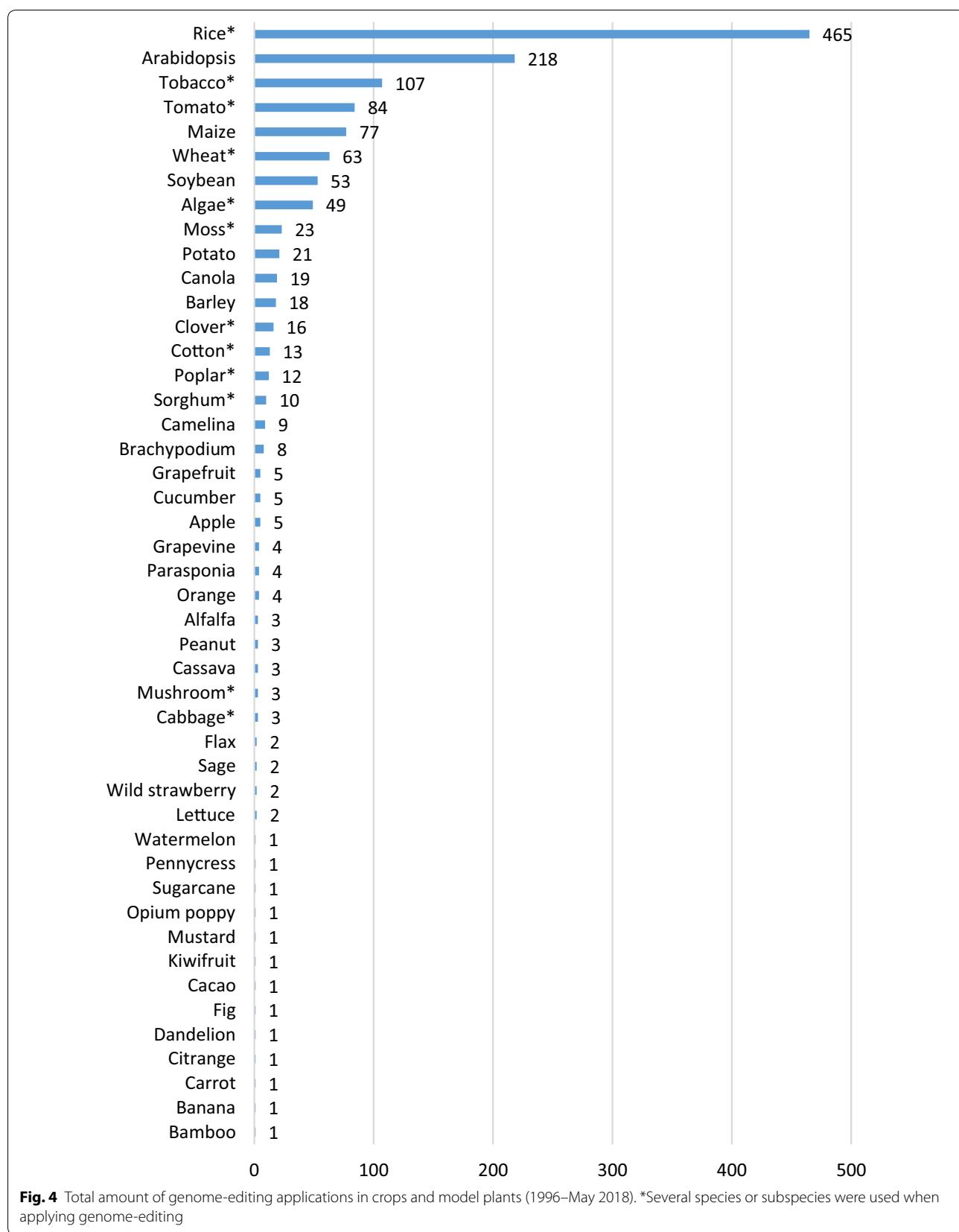
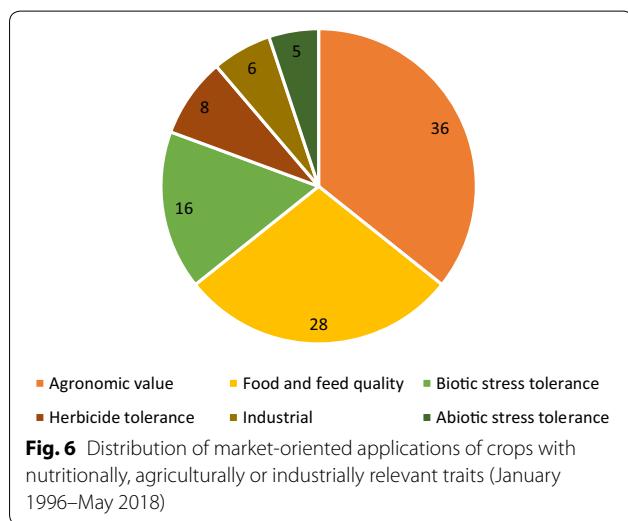
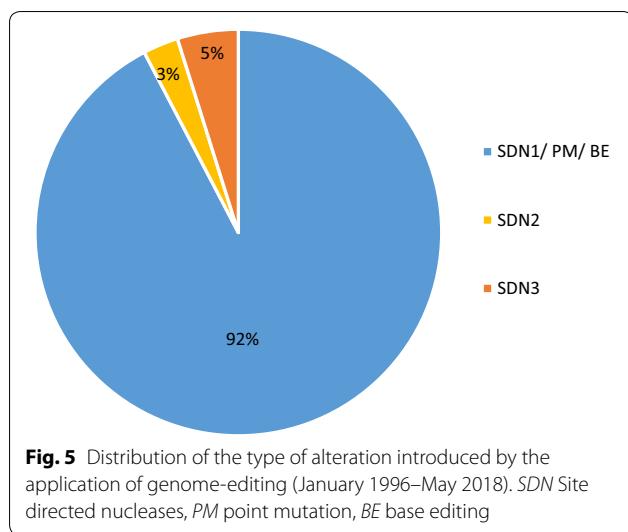


Fig. 3 Number of studies per country* in the systematic map database (grouped by continent; January 1996–May 2018). *Identified by the corresponding author(s); South America: Grey; Australia: Black; North America: Orange; Europe: Blue; Asia: Green; Africa: Red





groups of traits. Most of the market-oriented applications ($n=36$) are related to an improved agronomic value (Table 2), followed by 28 applications with an improved food and feed quality (Table 3). For biotic stress tolerance, 16 different applications were identified (Table 4), for herbicide tolerance eight applications (Table 5), for industrial utilization six applications (Table 6) and for abiotic stress tolerance five applications (Table 7). For more detailed information about the analyzed traits, see the respective tables.

As shown in Fig. 7 most of the market-oriented applications were applied in rice ($n=29$), followed by tomato ($n=16$), maize ($n=10$), potato ($n=6$), wheat ($n=6$), soybean ($n=4$) and canola ($n=4$). In 21 other agricultural relevant crops, one or two market-oriented applications were identified.

Secondary question 2: "What is the available evidence for the potential occurrence of associated off-target effects due to the use of genome-editing in model plants as well as in crops produced for agricultural production?"

In total, 252 studies from 161 articles were identified in which the occurrence of off-target effects was assessed. Table 8 maps the number of analyzed off-target effects for different genome-editing techniques and different plant species. Most of the off-target analyses were conducted in CRISPR/Cas studies ($n=228$) followed by TALENs studies ($n=9$), BE studies ($n=9$) and ZFN studies ($n=4$). Solely in one ODM and in one MN study off-target effects were investigated. Most off-target effects were analyzed in rice ($n=93$), followed by tomato ($n=28$), *Arabidopsis* ($n=23$) and soybean ($n=15$) (Table 8).

Off-target effects considered for CRISPR/Cas-systems

More than 90% of the studies, in which off-target effects were assessed, were conducted with CRISPR/Cas. Figure 8 provides an overview of the applied approaches to identify off-target effects. In total, 228 CRISPR/Cas studies dealt with the analysis of off-target effects. 205 studies predicted potential off-target sites and 195 of these identified potential off-target sites. Solely in 188 studies, these potential off-target sites were further analyzed for the occurrence of off-target effects using biased detection methods. In addition, 23 studies with already known potential off-target sites were assessed using biased detection methods. So, in total, 211 studies analyzed potential off-target sites using biased detection methods. Solely, nine studies searched for off-target mutations in a completely unrestricted way using unbiased detection methods. An overview of all identified CRISPR/Cas studies, in which off-target effects were addressed, is provided in Additional file 6 including all extracted data.

Prediction of potential off-target sites by CRISPR/Cas

Different prediction tools can be used to identify potential off-target sites. All of them have in common that based on sequence alignment programs DNA-sequences are identified in which unintended mutations could occur due to high similarity between the targeted sequence and the potential off-target site. However, the prediction of a potential off-target site is not equated with a real off-target mutation. It has to be shown in a follow up step by verifying the potential off-target site using biased detection methods. 205 CRISPR/Cas studies searched for potential off-target sites. As shown in Fig. 9, many different prediction tools were used to identify these sites. Mainly, three tools were used to predict potential off-target sites. BLAST was used 54 times, CRISPR-P 51 times and CasOFF-Finder 31 times. 12 other prediction tools were used in 31 studies (for detailed information see

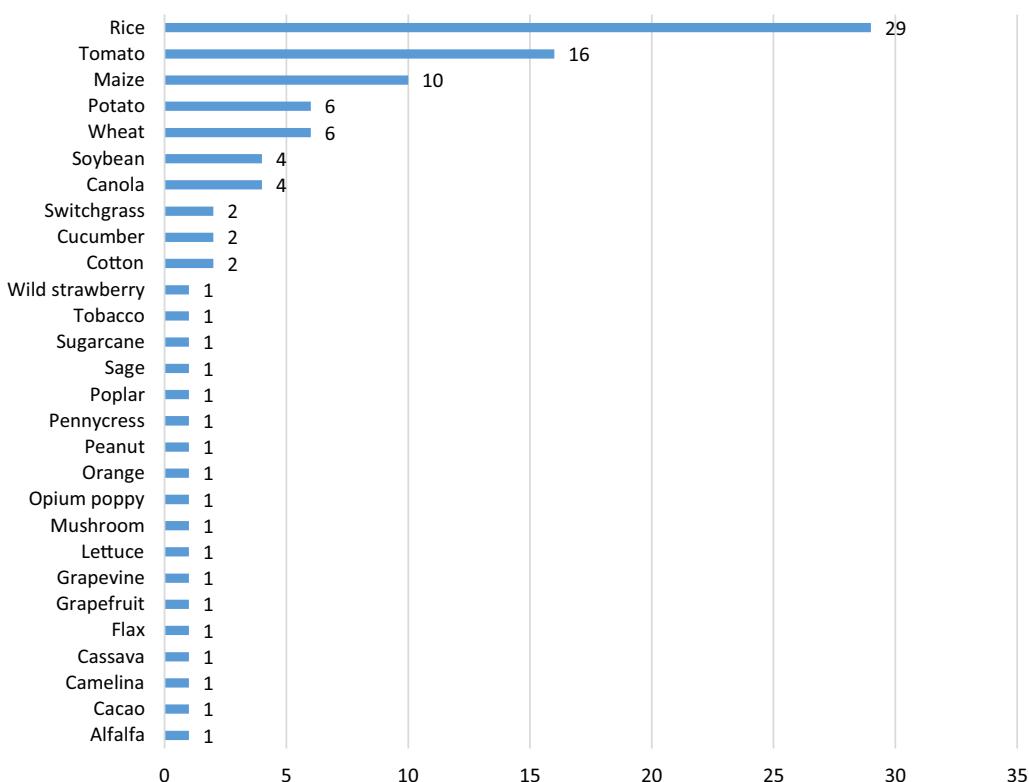


Fig. 7 Total amount of genome-editing applications in crops with market-oriented traits (January 1996–May 2018)

Additional file 6). 41 studies did not provide any details about the used tool(s). Three studies used two different prediction tools to identify potential off-target sites.

The number of predicted off-target sites varies widely between studies from zero to 4265. Several reasons for this broad heterogeneity exist, which were not extracted in detail within this map but will be elucidated in the discussion.

Detection of off-target effects—biased

Targeted sequencing to pre-selected sites was applied in 211 CRISPR/Cas studies to detect off-target effects. As shown in Fig. 8, the predicted off-target sites were frequently analyzed for the occurrence of off-target effects using biased detection methods. In a few studies, potential off-target sites were already known and analyzed without using sequence alignment programs a priori. Figure 10 displays the different detection methods applied to identify off-target mutations. In the large majority of studies, off-target effects were detected using a PCR followed by sequencing ($n=137$). Only a few studies used the detection methods RE-PCR assay ($n=16$), targeted deep sequencing ($n=15$), enzyme mismatch cleavage assay ($n=13$) and CAPS analyses ($n=13$). In 11 studies,

no information about the used detection method was provided.

Taking all CRISPR/Cas studies together, 1738 different potential off-target sites were analyzed using targeted sequencing. Off-target effects were identified in 55 of these sites, indicating that in around 3% of the analyzed sequences off-target mutations were detected. In another six studies, no information was provided about the amount of analyzed off-target sites but off-target mutations were not identified either.

Considering the different plant species, most of the CRISPR/Cas studies using biased detection methods were conducted in rice ($n=77$), followed by tomato ($n=23$), *Arabidopsis* ($n=21$), different moss species ($n=13$) and soybean ($n=12$) (Table 9). In rice, a total amount of 291 potential off-target sites were analyzed and in 25 of these sites, off-target mutations were detected. In contrast, studies conducted in tomatoes solely reported the identification of one off-target mutation when analyzing 222 potential off-target sites.

In one study, a different approach was chosen to assess the occurrence of off-target effects [175]. In this study, a series of mismatches were introduced at the sgRNA followed by analyzing whether the targeted sequence was

Table 2 Genome-editing in plants for modifying agronomically relevant traits (1996–May 2018)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Canola	Christian-Albrechts-University of Kiel, Germany	Increased yield	Increased shatter resistance to avoid seed loss during mechanical harvest	CRISPR/Cas9 SDN1	[41]
Canola	Huazhong Agricultural University, China	Increased yield	Increased seeds number per husk, higher seed weight	CRISPR/Cas9 SDN1	[42]
Cotton	Anhui Agricultural University, China; Chinese Academy of Agricultural Sciences, China	Growth characteristics	Improved root growth under high and low nitrogen conditions	CRISPR/Cas9 SDN1	[43]
Cucumber	Chinese Academy of Agricultural Sciences, China	Growth characteristics	Only female flowers	CRISPR/Cas9 SDN1	[44]
Lettuce	University of California, USA	Increased yield	Germination at high temperature	CRISPR/Cas9 SDN1	[45]
Maize	Benson Hill Biosystems, USA	Increased yield	Increased photosynthesis efficiency	Meganuclease SDN3	[46]
Maize	University of Wisconsin, USA	Growth characteristics	Early flowering under long day conditions	CRISPR/Cas9 SDN1	[47]
Maize	DuPont Pioneer, USA	Growth characteristics	Male sterility	CRISPR/Cas9 SDN1	[48, 49]
	University of Science and Technology Beijing, China; Beijing Solidwill Sci-Tech Co. Ltd, China			CRISPR/Cas9 SDN1	[4]
	Chinese Academy of Sciences, China			CRISPR/Cas9 SDN1	[50]
Maize	Syngenta Seeds, USA	Growth characteristics	Haploid induction	TALENs SDN1	[51]
Potato	Collectis Plant Science, USA	Storage characteristics	Improved cold storage and processing traits (reduced sugars/reduced levels of acrylamide)	TALENs SDN1	[52]
Rice	Chinese Academy of Sciences, China	Increased yield	Altered grain number per panicle	CRISPR/Cas9 SDN1	[53]
	National Rice Research Institute, China			CRISPR/Cas9 SDN1	[54]
Rice	Chinese Academy of Sciences, China	Increased yield	Seed size/increased seed weight	CRISPR/Cas9 SDN1	[53]
	Anhui Academy of Agricultural Sciences, China			CRISPR/Cas9 SDN1	[55]
	Fudan University, China			CRISPR/Cas9 SDN1	[56]
	Yangzhou University, China			CRISPR/Cas9 SDN1	[57]
	Agronomy College of Henan Agricultural University, China			CRISPR/Cas9 SDN1	[58]
	Chinese Academy of Agricultural Sciences, China; Yangzhou University, China			CRISPR/Cas9 SDN1	[59]
Rice	Chinese Academy of Sciences, China	Growth characteristics	Increased plant height, improved tiller-production, erect panicle, increased biomass	CRISPR/Cas9; SDN1	[53, 60]
	Wuhan Institute of Bioengineering; Huazhong Agricultural University, China			CRISPR/Cas9 SDN1	[60]
	Sichuan Agricultural University			CRISPR/Cas9 SDN1	[61]
	Chinese Academy of Agricultural Sciences, China; Yangzhou University, China			CRISPR/Cas9 SDN1	[59]

Table 2 (continued)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Rice	Chinese Academy of Agricultural Sciences, China; Jiangsu Academy of Agricultural Sciences, China	Growth characteristics	Early maturing	CRISPR/Cas9 SDN1	[62]
Rice	Kyung Hee University, South Korea	Growth characteristics	Male sterility	CRISPR/Cas9 SDN1	[63]
	Shanghai Jiao Tong University, China			CRISPR/Cas9 SDN1	[64]
	South China Agricultural University, China			CRISPR/Cas9 SDN1	[65, 66]
	Sichuan Agricultural University, China			CRISPR/Cas9 SDN1	[67, 68]
Rice	Chinese Academy of Sciences, China; University of Chinese Academy of Sciences, China	Increased yield	Regulation of pollen tube growth	CRISPR/Cas9 SDN1	[69]
Rice	China Agricultural University, China	Storage characteristics	Increased seed storage	TALENs SDN1	[70]
Rice	China National Rice Research Institute, China; China Three Gorges University, China	Increased yield	Increased seed setting rate	CRISPR/Cas9 SDN1	[71]
Rice	Anhui Academy of Agricultural Sciences, China	Increased yield	Longer panicle	CRISPR/Cas9 SDN1	[55]
Rice	Nanjing Agricultural University, China	Increased yield	Grain yield, regulation of seed development	CRISPR/Cas9 SDN1	[72]
Rice	Chinese Academy of Sciences, China	Growth characteristics	Decreased plant height	CRISPR/Cas9 SDN1	[73]
	Syngenta Biotechnology, China			CRISPR/Cas9 SDN1	[74]
Rice	Wuhan Institute of Bioengineering, China; Huazhong Agricultural University, China	Increased yield	Increased nitrogen utilization efficiency	CRISPR/Cas9 SDN1	[60]
Rice	Hunan Normal University, China	Growth characteristics	Regulation of seed dormancy, stomatal opening, plant developmental, abiotic stress tolerance and leaf senescence	CRISPR/Cas9 SDN1	[75]
Soybean	Chinese Academy of Agricultural Sciences, China	Growth characteristics	Late flowering	CRISPR/Cas9 SDN1	[76]
Switchgrass	Iowa State University, USA	Growth characteristics	Bushy phenotype	CRISPR/Cas9 SDN1	[77]
Tomato	National Food Research Institute, Japan	Increased yield	Regulating fruit ripening	CRISPR/Cas9 SDN1	[78]
Tomato	University of Minnesota, USA	Growth characteristics	Bigger seedlings	TALENs SDN1	[79]
Tomato	Cold Spring Harbor Laboratory, USA; Max Planck Institute for Plant Breeding Research, Germany; Université Paris-Scalay, France	Growth characteristics	Early flowering	CRISPR/Cas9 SDN1	[80]
Tomato	University of Florida, USA	Growth characteristics	Easy separation of fruit and stem	CRISPR/Cas9 SDN1	[81]
Tomato	Cold Spring Harbor Laboratory, USA	Increased yield	Fruit size	CRISPR/Cas9 SDN1	[82]
Tomato	Cold Spring Harbor Laboratory, USA	Increased yield	Highly branched inflorescence and formation of multiple flowers	CRISPR/Cas9 SDN1	[82]
Tomato	Weizmann Institute of Science, Israel	Growth characteristics	Yellow fruit color	CRISPR/Cas9 SDN1	[83]
	Weizmann Institute of Science, Israel			CRISPR/Cas9 SDN 3	[84]

Table 2 (continued)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Tomato	Weizmann Institute of Science, Israel	Growth characteristics	Orange fruit color	CRISPR/Cas9 SDN3	[84]
Tomato	Academy of Agriculture and Forestry Sciences; Chinese Academy of Sciences, China	Growth characteristics	Pink fruit color	CRISPR/Cas9 SDN1	[85]
Wheat	Kansas State University, USA Chinese Academy of Sciences, China	Increased yield	Bigger grains, increased grain weight	CRISPR/Cas9 SDN1 CRISPR/Cas9 SDN1	[86] [87]
Wild strawberry	University of Maryland, USA	Growth characteristics	Faster seedling growth	CRISPR/Cas9 SDN1	[88]

TALENs Transcription Activator-Like Effector Nucleases, CRISPR/Cas9 Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9, SDN Site directed nucleases

successfully mutated despite the mismatch(es) between the sgRNA and the targeted sequence. Additionally, in this study, the off-target patterns between two PAMs (NGG and NAG) were compared. 22 times the sgRNA was designed in a way that it contained one mismatch to the targeted sequence. 15 of these altered sgRNA induced a DSB at the targeted sequence. 14 times the sgRNA contained two mismatches to the targeted sequence and four of these sgRNA induced a DSB in the targeted sequence. Moreover, eight times the sgRNA contained three mismatches. In these cases, no mutation was identified in the targeted sequence [175]. According to the prediction of off-target effects, the summary provided here does not allow any conclusions to be drawn due to broad heterogeneity.

Heterogeneity in CRISPR/Cas-studies regarding the evidence how to predict and detect potential off-target effects

The number of potential off-target sites called “identified” by the respective authors varies widely between zero and 4265. Several reasons were identified that could explain this broad heterogeneity:

- In total, potential off-target sites were investigated in over 30 different plant species and subspecies. The different genome sizes and the different number of chromosome sets of the individual plants vary widely which influence the number of potential off-target sites.
- To identify potential off-target sites in CRISPR/Cas-studies 15 different prediction tools were applied.
- For the detection of off-target effects, various methods have been used, but all of them show their

specific advantages and disadvantages and could affect the occurrence of off-target effects [26, 32].

- The number of hypothetically tolerated mismatches between the target sequence and the potential off-target sites which is an exercise in combinatorics. Ali et al. [176] identified a total amount of 4265 potential off-target sites in the model organism *Nicotiana benthamiana* for a CRISPR/Cas9 sgRNA. To identify candidate off-target sites, the genome was screened allowing one to seven mismatches to the target sequence. Hence, the more mismatches are tolerated in prediction the higher is the number of potential off-target sites (in the paper: one, two or three mismatches tolerated: No potential off-target sites, four mismatches tolerated: One potential off-target site, five mismatches tolerated: 60 potential off-target sites, six mismatches tolerated: 515 potential off-target sites, seven mismatches tolerated: 3689 potential off-target sites). This indicates that the number of potential off-target sites strongly depends on the number of hypothetically tolerated mismatches. Throughout the available literature, the number of tolerated mismatches predetermined by the researchers was very heterogeneous ranging up to 13 mismatches [177].
- Individual studies also deviate between different structural assumptions when determining potential off-target sites. In most studies, a potential off-target was only counted as such if a PAM followed the potential off-target site. However, in some studies, potential off-target sites were assigned as such without being followed by a PAM (e.g. [44, 178]), although at these sites no DSB can be induced with the specific CRISPR-nucleases used.

Table 3 Genome-editing in plants for improved food and feed quality (1996–May 2018)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Alfalfa	Calyxt, Inc., USA	Product quality	Reduced lignin content	TALENs SDN1	[89]
Camelina sativa	Montana State University, USA	Product quality	Increased levels of oleic acid and α-linolenic acid	CRISPR/Cas9 SDN1	[90]
	University Nebraska, USA		Increased levels of oleic acid, decreased levels of fatty acids	CRISPR/Cas9 SDN1	[91]
	Université Paris-Saclay, France		Increased levels of oleic acid, decreased levels of fatty acids	CRISPR/Cas9 SDN1	[92]
	Kansas State University, USA		Lower oil content	CRISPR/Cas9 SDN1	[93]
Canola	Tamagawa University, Japan	Product quality	Altered fatty acid composition	CRISPR/Cas9 SDN1	[94]
Maize	Du Pont Pioneer, USA;	Product quality	Waxy corn, improved starch production	CRISPR/Cas9 SDN1	[95]
	Chinese Academy of Agricultural Sciences, China		Waxy phenotype, abolition of amylose	CRISPR/Cas9 SDN1	[96]
Maize	Agrivida, USA	Product quality	Higher levels of starch in their leaves and stalks	Meganucleases SDN1	[97]
Maize	Dow AgroScience, USA	Product quality	Reduced phytate production + herbicide tolerance	ZFN SDN3	[98, 99]
Mushroom	Penn State University, USA	Product quality	Non-browning mushroom	CRISPR/Cas9 SDN1	[100]
Opium poppy	Cankiri Karatekin University, Turkey; Dokuz Eylul University, Turkey	Product quality	Reduced morphine and thebaine content	CRISPR/Cas9 SDN1	[101]
Peanut	Guangdong Academy of Agricultural Sciences, China	Product quality	Increased oleic acid content, decreased linoleic acid content	TALENs SDN1	[102]
Potato	Calyxt, USA	Product quality	Non-browning potato	TALENs SDN1	[103]
Potato	Simplot Plant Science, USA	Product quality	Reduced black spottiness	TALENs SDN1	[104]
Potato	RIKEN Center for Sustainable Resource Science, Japan; Chiba University, Japan	Product quality	Reduction of harmful ingredients (glycoalkaloids)	TALENs SDN1	[105]
	Kobe University, Japan		Complete abolition of glycoalkaloids (bitter taste)	CRISPR/Cas9 SDN1	[106]
Rice	Chinese Academy of Sciences, China	Product quality	Fragrant rice	TALENs SDN1	[107]
	Chinese Academy of Agricultural Sciences, China; Yangzhou University, China			CRISPR/Cas9 SDN1	[59]
Rice	Chinese Academy of Agricultural Sciences, China; University of California, USA	Product quality	Increased contents benefitting human health (increased amylose content)	CRISPR/Cas9 SDN1	[108]
Rice	Huazhong Agricultural University, China	Product quality	Reduced contents harming human health (arsenic content)	CRISPR/Cas9 SDN1	[109]
	Sun Yat-sen University, China			CRISPR/Cas9 SDN1	[110]
Rice	National Agriculture and Food Research Organization, Japan	Product quality	Altered fatty acid composition	CRISPR/Cas9 SDN1	[111]
Rice	Université Montpellier, France	Product quality	Reduced contents harming human health (cesium content)	CRISPR/Cas9 SDN1	[112]
Rice	Hunan Agricultural University, Hunan Hybrid Rice Research Center, Normal University, China	Product quality	Reduced contents harming human health (cadmium content in plants)	CRISPR/Cas9 SDN1	[113]
Rice	Chinese Academy of Sciences, Shanghai, China; Purdue University, West Lafayette, USA	Product quality	Waxy rice	CRISPR/Cas9 SDN1	[114]

Table 3 (continued)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Sage	Second Military Medical University, China	Product quality	Reduced phenolic acid content	CRISPR/Cas9 SDN1	[115]
Soybean	Cellectis plant science Inc., USA/Calyxt, USA	Product quality	High oleic content, low linoleic content	TALENs SDN1	[21, 116–118]
Tomato	Agricultural Research Organization, Israel	Product quality	Seedless tomato	CRISPR/Cas9 SDN1	[119]
	Tokushima University, Japan			CRISPR/Cas9 SDN1	[120]
Tomato	University of Tsukuba, Japan	Product quality	Increased contents benefitting human health (increased GABA content)	CRISPR/Cas9 SDN1	[121]
	China Agricultural University, China			CRISPR/Cas9 SDN1	[122]
Tomato	China Agricultural University, China	Product quality	Increased contents benefitting human health (increased lycopene content)	CRISPR/Cas9 SDN1	[123]
Tomato	Xinjiang Academy of Agricultural Science, China	Product quality	Improved shelf life	CRISPR/Cas9 SDN1	[124]
Wheat	Calyxt, Inc., USA	Product quality	Increased nutritional value	TALENs SDN1	[125]
Wheat	Instituto de Agricultura Sostenible (IAS-CSIC), Spain; University of Minnesota, USA	Product quality	Reduced gluten content	CRISPR/Cas9 SDN1	[126]
Wheat (durum)	Instituto de Agricultura Sostenible (IAS-CSIC), Spanien; University of Minnesota, USA	Product quality	Reduced gluten content	CRISPR/Cas9 SDN1	[126]

TALENs Transcription Activator-Like Effector Nucleases, CRISPR/Cas9 Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9, ZFN Zinc-Finger Nucleases, SDN Site directed nucleases

- vi. In some studies, the sgRNA was selected taking into account that no potential off-target effect should occur. Care was taken that, apart from the target sequence, no other sites in the genome possess a similar sequence that could result in an off-target mutation. Thus, the predicted number of potential off-target sites is lower or zero in these studies compared to studies, in which potential off-target sites were not considered a priori in the sequence selection.

Detection of off-target effects—unbiased

Nine CRISPR/Cas studies were identified using whole genome sequencing (WGS) as an unbiased detection method to identify genome-wide off-target effects (see Additional file 6). No off-target mutations were detected in any of these studies. One study compared both biased and unbiased detection methods. While using unbiased detection methods, no off-target effects were detected, but biased methods detected one off-target [179]. An explanation for this could be that WGS is able to detect higher frequency off-target effects only, but lacks the sensibility required to detect off-target mutations in bulk population [26].

Off-target effects considered for TALENs systems

In the period until May 2018, nine TALENs studies were identified addressing off-target effects. Additional file 6 provides an overview of these studies including all extracted data according to the systematic map protocol. Figure 11 maps the different approaches used to analyze off-target effects. Five studies predicted potential off-target sites. Two times, the TAL Effector Nucleotide Targeter 2.0 was used and one time each of the tools PROGNONS, kmasker and Arabidopsis Information Resource PatMatch. The number of identified potential off-target sites varies widely between zero and 18. In one study, no precise information was given and it was just indicated that many potential off-target sites were identified. The four studies that identified potential off-target sites investigated these for the occurrence of off-target effects using biased detection methods. In addition, three studies with already known potential off-target sites assessed these sites. Different tools were used to examine whether off-target effects occurred. Two studies used enzyme mismatch cleavage assay and one study each PCR + Sequencing, PCR + CAPS and RE-PCR assay. In two studies, no detailed information about the applied detection method was provided. In total, 31 potential off-target sites were analyzed for the occurrence of off-target

Table 4 Genome-editing in plants for increased resistance to biotic stress (1996–May 2018)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Cacao	Pennsylvania State University, USA	Fungal resistance	Resistance to <i>Phytophthora tropicalis</i>	CRISPR/Cas9 SDN1	[127]
Cucumber	Volcani Center, Israel	Virus resistance	Immunity to cucumber vein yellowing virus (<i>Ipoavirus</i>) infection and resistance to the potyviruses Zucchini yellow mosaic virus and Papaya ring spot mosaic virus-W	CRISPR/Cas9 SDN1	[22]
Grapefruit	University of Florida, USA	Bacterial resistance	Resistance to citrus canker	CRISPR/Cas9 SDN1	[128, 129]
Grapevine	Northwest A&F University and Ministry of Agriculture, China	Fungal resistance	Resistance to <i>Botrytis cinerea</i>	CRISPR/Cas9 SDN1	[130]
Maize	Du Pont Pioneer, USA	Fungal resistance	Resistance to Northern Leaf Blight (NLB)	CRISPR/Cas9 (Cisgenesis) SDN3	[131]
Orange	Chinese Academy of Agricultural Sciences and National Center for Citrus Variety Improvement; Southwest University, China	Bacterial resistance	Resistance to citrus canker	CRISPR/Cas9 SDN1	[132]
Rice	Chinese Academy of Agriculture, China	Fungal resistance	Resistance to rice blast	CRISPR/Cas9 SDN1	[133]
Rice	Iowa State University, USA	Bacterial resistance	Resistance to bacterial blight	CRISPR/Cas9 SDN1	[134]
	IRD-CIRAD-Université, France			TALENs SDN1	[135]
	Iowa State University, USA			TALENs SDN1	[136]
	National University of Singapore, Singapore			TALENs SDN1	[137]
	Chinese Academy of Sciences, China			TALENs SDN1	[138]
	National Center for Plant Gene Research, China; Sichuan Agricultural University, China			CRISPR/Cas9 SDN1	[139]
Rice	Iowa State University, USA	Fungal resistance	Resistance to powdery mildew	TALENs SDN1	[140]
Rice	Shanghai Jiao Tong University, China; Yunnan Academy of Agricultural Sciences, China	Bacterial resistance	Resistance to the pathogen Xoc RS105	TALENs SDN1	[141]
Rice	Sichuan Agricultural University, China	Bacterial resistance/ fungal resistance	Resistance to bacterial blight and rice blight	CRISPR/Cas9 SDN1	[61]
Rice	International Rice Research Institute (IRRI), Philippines	Virus resistance	Resistance to rice tungro disease (RTD)	CRISPR/Cas9 SDN1	[142]
Tomato	Max Planck Institute for Developmental Biology, Germany; Norwich Research Park, UK	Fungal resistance	Resistance to powdery mildew	CRISPR/Cas9 SDN1	[143]
Tomato	King Abdullah University of Science and Technology, Saudi Arabia	Virus resistance	Resistance to tomato yellow leaf virus	CRISPR/Cas9 SDN1	[144]
Tomato	University of California, USA	Bacterial resistance	Resistance to different pathogens including <i>P. syringae</i> , <i>P. capsici</i> and <i>Xanthomonas</i> spp.	CRISPR/Cas9 SDN1	[145]
Wheat	Chinese Academy of Sciences, China	Fungal resistance	Resistance to powdery mildew	TALENs SDN1	[25]
	Chinese Academy of Sciences, China			CRISPR/Cas9 SDN1	[146]
	Calyxt, Inc., USA			TALENs SDN1	[147]

TALENs Transcription Activator-Like Effector Nucleases, CRISPR/Cas9 Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9, SDN Site directed nucleases

Table 5 Genome-editing for generating herbicide tolerant plants (1996–May 2018)

Plants	Developer, producer, country	Trait ^a	Specification	Technological specification	References
Canola	Cibus, Canada; Cibus, USA Bayer BioScience N.V., Belgium	Herbicide tolerance –	ODM		[23] [148]
Cassava	Donald Danforth Plant Science Center, St. Louis, USA	Herbicide tolerance –	CRISPR/Cas9 SDN3		[149]
Cotton	Bayer CropScience N.V., Belgium	Herbicide tolerance –	Mega-nucleases SDN3		[150]
Flax	Cibus, USA	Herbicide tolerance –	CRISPR/Cas9 SDN1		[151]
Maize	DuPont Pioneer, USA Dow AgroScience, USA Pioneer Hi-Bred International, USA	Herbicide tolerance –	CRISPR/Cas9 SDN1, SDN2, SDN3 ZFN SDN3 ODM		[48, 49] [99, 152] [153, 154]
Potato	Michigan State University, USA	Herbicide tolerance –	CRISPR/Cas9, TALENs SDN2		[155]
Rice	Chinese Academy of Sciences, China Chinese Academy of Sciences, China; Huazhong Agricultural University, China; University of California San Diego, USA Zhejiang University, China Tohoku University, Japan Kobe University, Japan; University of Tsukuba, Japan King Abdullah University of Science and Technology, Saudi Arabia	Herbicide tolerance –	CRISPR/Cas9 SDN2 CRISPR/Cas9 SDN2 TALENs SDN2 ODM BE CRISPR/Cas9 SDN2		[156] [157] [158] [159] [160, 161] [162]
Soybean	DuPont Pioneer, USA	Herbicide tolerance –	CRISPR/Cas9 SDN2 CRISPR/Cas9 SDN3		[163] [164]

^a No detailed breakdown regarding chemical agents

TALENs Transcription Activator-Like Effector Nucleases, CRISPR/Cas9 Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9, ZFN Zinc-Finger Nuclease, ODM Oligo-Directed Mutagenesis, SDN Site directed nucleases, BE base editing

Table 6 Genome-editing in plants for industrial utilization (1996–May 2018)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Pennycress	Illinois State University, USA	Product quality	Altered oil composition	CRISPR/Cas9 SDN1	[165]
Poplar	University of Georgia, USA	Product quality	Stem wood discoloration due to lignin reduction	CRISPR/Cas9 SDN1	[166]
Potato	Swedish University of Agricultural Sciences, Sweden	Product quality	Improved starch quality	CRISPR/Cas9 SDN1	[167]
Sugarcane	University of Florida, USA	Product quality	Reduced lignin content	TALENs SDN1	[168, 169]
Switchgrass	Noble Research Institute, USA	Product quality	Reduced lignin content	CRISPR/Cas9	[170]
Tobacco	North Carolina State University, USA	Product quality	Reduced nicotine content	Meganucleases SDN1	[171]

CRISPR/Cas9 Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9, TALENs Transcription Activator-Like Effector Nucleases, SDN Site directed nucleases

effects and one of these sites contained an off-target mutation. In one study, an unbiased search for off-target effects was conducted [180]. Using Whole Genome Sequencing (WGS), three off-target mutations were identified which were not present in the wild-type sample. However, the off-target sequences showed no similarity to the TALENs binding sites. Therefore, the occurrence

of these mutations cannot be ruled out to be spontaneous ones or sequencing errors [180].

Off-target effects considered for Zinc-Finger Nucleases systems

Four studies dealt with the analyses of off-target effects when applying ZFN in plants. Detailed information is

Table 7 Genome-editing in plants to improve tolerance to abiotic stress (1996–May 2018)

Plant	Developer, producer, country	Trait	Specification	Technological specification	References
Maize	Ghent University, Belgium; Center for Plant Systems Biology, Belgium; Jomo Kenyatta University of Agriculture and Technology, Kenya	Drought tolerance	–	CRISPR/Cas9 SDN1	[172]
	DuPont Pioneer, USA			CRISPR/Cas9 SDN3	[24, 164]
Rice	Anhui Academy of Agricultural Sciences, China	Salt tolerance	–	CRISPR/Cas9 SDN1	[20]
Rice	Huazhong Agricultural University, China	Arsenic tolerance	–	CRISPR/Cas9 SDN1	[109]
	Sun Yat-sen University, China			CRISPR/Cas9 SDN1	[110]
Soybean	USDA-ARS, USA	Drought and salt tolerance	–	CRISPR/Cas9 SDN1	[173]
Wheat	Montana State University, USA	Drought tolerance	–	CRISPR/Cas9 SDN1	[174]

CRISPR/Cas9 Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9, SDN Site directed nucleases

provided in Additional file 6. All studies identified putative off-target sites that are most closely related to the target sequences. Two times, the database PLantGDB was used to identify potential off-target sites and two times no detailed information was provided. In total, 10 potential off-target sites were detected which were then screened for the occurrence of off-target effects. No off-target mutations were detected in any of these analyzed, potential off-target sites.

Off-target effects considered for Meganuclease systems

Solely one MN study was identified that analyzed off-target effects (Additional file 6). In this study, a homologous sequence to the target one was identified that differs solely in two nucleotides. However, when screening this potential off-target site no modification was identified.

Off-target effects considered for Oligonucleotide-Directed Mutagenesis systems

One study addressed the occurrence of off-target effects in rice using the ODM technique (Additional file 6). Beside the targeted Acetolactate synthase (ALS) sequence, the whole coding region of ALS gene was sequenced but no further mutation was detected.

Off-target effects considered for base editing systems

In total, nine BE studies analyzed the occurrence of off-target effects (Additional file 6). However, one study could only rarely be evaluated due to language barriers [181]. The approach used to identify off-target effects was similar in all studies (Fig. 12). In a first step, potential off-target sites were predicted using different tools (2× CRISPR-GE tool, 2× CRISPR-P, 1× CasOff-Finder, 3×

no information). According to the other genome-editing techniques, the number of potential off-target sites varies widely between one and nine. In a second step, all predicted potential off-target sites were sequenced for the occurrence of off-target effects using PCR + sequencing method ($n=6$), targeted deep sequencing ($n=2$) or enzyme mismatch cleavage assay ($n=1$). One off-target mutation was identified.

Discussion

Market oriented applications of genome-editing

This map documents the state of evidence for the application of genome-editing as a new tool for the modification of plant traits and the associated potential occurrence of off-target effects. The publication rate of primary research has risen sharply since the CRISPR/Cas technique was first applied in plants in 2013. It is worth mentioning that in total primary studies from 33 countries were identified but nearly three quarter of these studies originate from either China (40%) or the USA (33%). For comparison, Japan and Germany published around 6% of the studies each and no other country contributed more than 2% of the total. Summarizing the number of published studies by continent, more than 50% of the studies were conducted in an Asian country (53%), around one-third in North American countries (34%) and only 13% in European countries. As the genome-editing techniques, especially CRISPR/Cas, were just recently developed, the large majority of the existing applications represent basic research. Nevertheless, almost 100 different applications aimed to produce beneficial agricultural traits in 28 different agricultural crops. We determined that the majority of such “market-oriented” applications have

Table 8 Overview of off-target studies in relation to different genome-editing techniques and plant species (January 1996–May 2018)

	CRISPR/Cas	TALENs	ZFN	ODM	MN	BE	# studies
Rice	82	2	0	1	0	8	93
Tomato	25	1	1	0	0	1	28
Arabidopsis	23	2	2	0	0	0	27
Soybean	13	2	0	0	0	0	15
Moss	13	0	0	0	0	0	13
Wheat	11	0	0	0	0	0	11
Tobacco	9	0	0	0	1	0	10
Maize	7	0	1	0	0	0	8
Algae	4	0	0	0	0	0	4
Canola	4	0	0	0	0	0	4
Cotton	4	0	0	0	0	0	4
Clover	3	0	0	0	0	0	3
Grapevine	3	0	0	0	0	0	3
Orange	3	0	0	0	0	0	3
Poplar	3	0	0	0	0	0	3
Cucumber	2	0	0	0	0	0	2
Grapefruit	2	0	0	0	0	0	2
Lettuce	2	0	0	0	0	0	2
Sage	2	0	0	0	0	0	2
Barley	1	1	0	0	0	0	2
Potato	1	1	0	0	0	0	2
Alfalfa	1	0	0	0	0	0	1
Cabbage	1	0	0	0	0	0	1
Cacao	1	0	0	0	0	0	1
Camelina	1	0	0	0	0	0	1
Carrot	1	0	0	0	0	0	1
Citrangle	1	0	0	0	0	0	1
Flax	1	0	0	0	0	0	1
Kiwifruit	1	0	0	0	0	0	1
Opium poppy	1	0	0	0	0	0	1
Watermelon	1	0	0	0	0	0	1
Wild strawberry	1	0	0	0	0	0	1
# studies	228	9	4	1	1	9	252

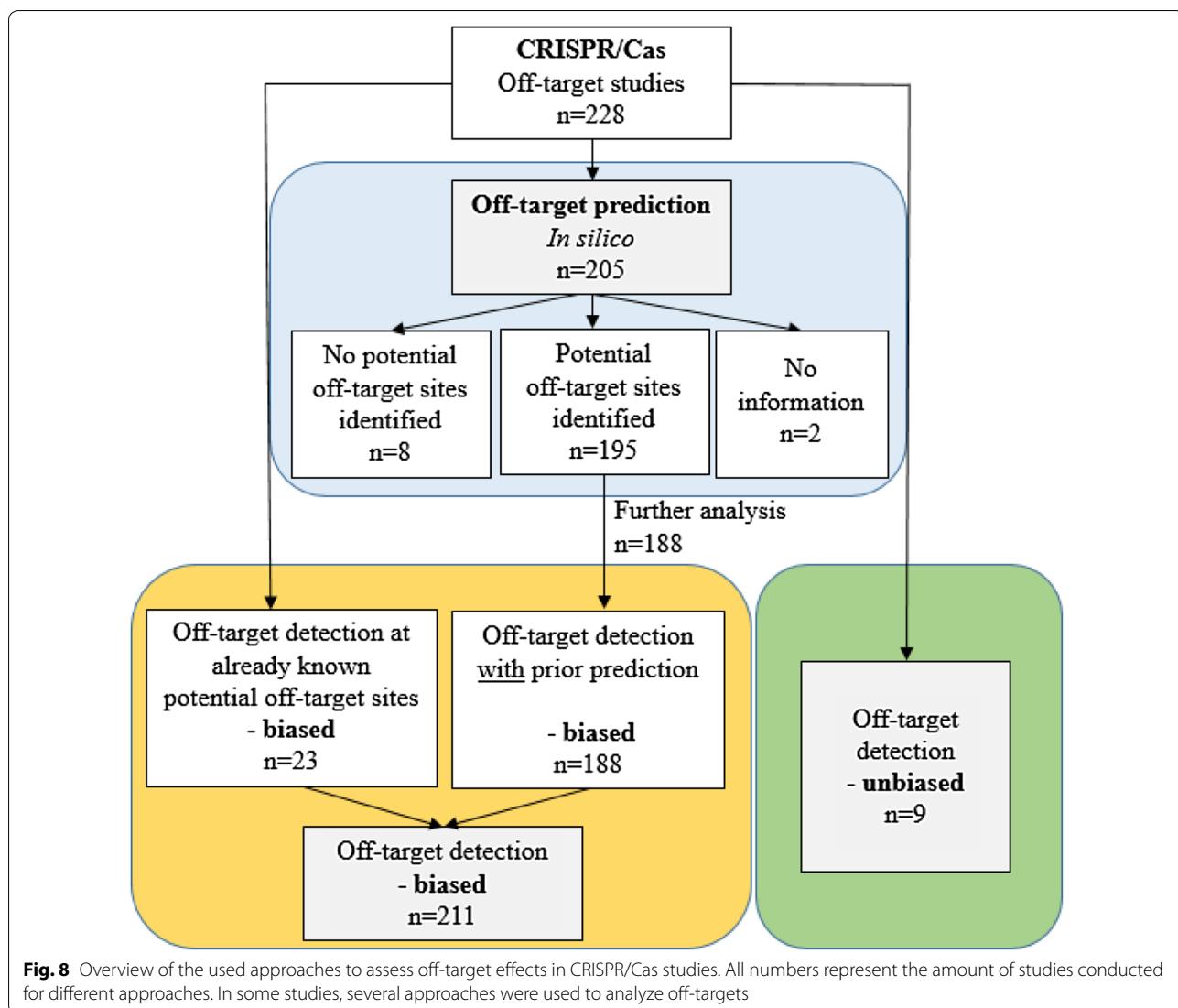
CRISPR/Cas Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein, **TALENs** Transcription Activator-Like Effector Nucleases, **ZFN** Zinc-Finger Nuclease, **ODM** Oligo-Directed Mutagenesis, **MN** Meganucleases, **BE** base editing

been carried out in economically important crops such as maize, rice, wheat and soybeans, but less economically important crops such as cucumber, lettuce, peanut or grapefruit have been worked on as well. The market-oriented applications address several breeding objectives including yield improvement, improved growth characteristics, improved food and feed quality, tolerances to biotic and abiotic stress, herbicide tolerance and industrial utilization. This indicates that genome-editing is able to address beneficial traits for the agricultural value

chain and could contribute to food security and the environmental management. To confirm this hypothesis a systematic review would be required to quantify such effects.

Off-target effects of genome-editing

252 studies investigated the occurrence of off-target effects following a genome-editing application in plants. Most of these studies were performed with the genome-editing technique CRISPR/Cas ($n=228$), whereas only a



few studies used TALENs ($n=9$), BE ($n=9$), ZFN ($n=4$), MN ($n=1$) or ODM ($n=1$). Reasons for these findings are that the total number of studies in which CRISPR/Cas was applied is much larger compared to the other genome-editing techniques. Additionally, CRISPR/Cas is more susceptible to off-target effects compared to other nuclease techniques such as ZFN and TALENs, as it works as a monomer, whereas ZFN and TALENs work as dimers [182]. In addition, the sgRNA used in CRISPR/Cas-applications is able to tolerate several mismatches leading to the induction of a DSB at a site in the DNA that is similar but not identical to the targeted sequence [32]. In most CRISPR/Cas studies biased detection methods were used, meaning that potential off-target sites were first predicted using bioinformatics programs followed by targeted sequencing of these sites for the

occurrence of off-target effects. The biased method suggests a very broad data basis, but as shown in the result section, individual studies are very heterogeneous in their structure and design. Heterogeneity was identified regarding the plant species, the CRISPR-variant, the prediction tools and detection methods used, the amount of tolerated mismatches and the chosen sgRNA. In order to allow any conclusions to be drawn about the occurrence of off-target effects a more in-depth analysis e.g. by a systematic review is mandatory.

Knowledge gaps

Regarding secondary question 2, several topics have been identified representing knowledge gaps where no studies or only a small number of studies exist. A knowledge gap exists for the analysis of off-target effects in TALENs, ZFN, MN, ODM and BE. Only nine studies have been

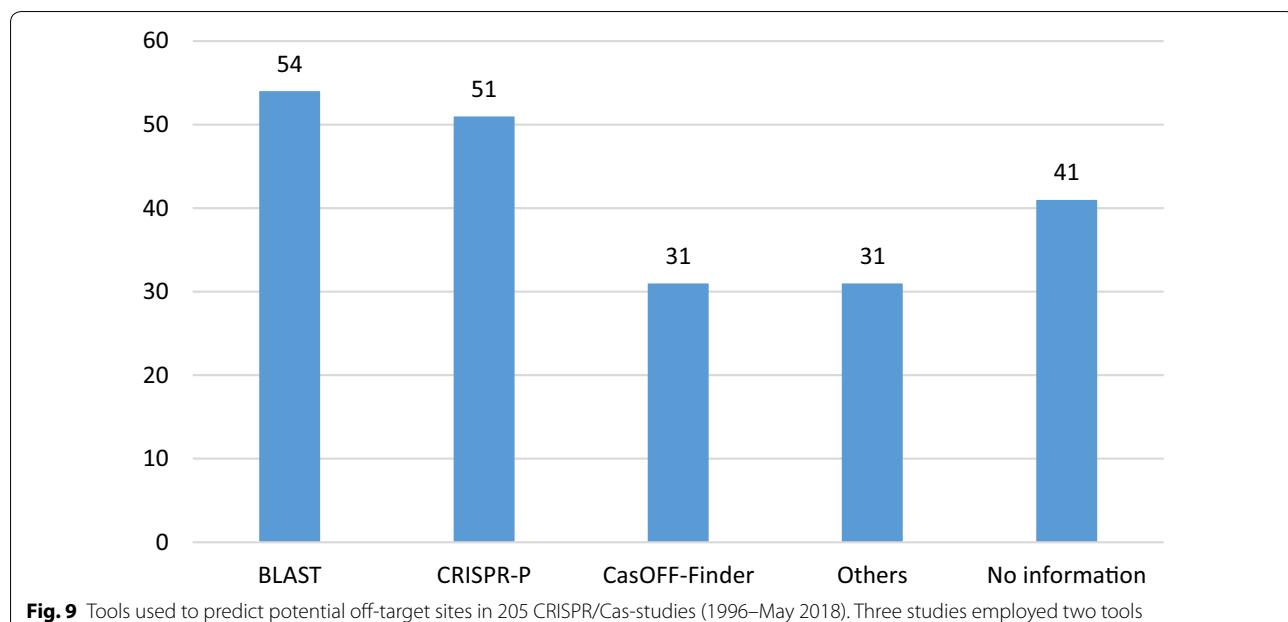


Fig. 9 Tools used to predict potential off-target sites in 205 CRISPR/Cas-studies (1996–May 2018). Three studies employed two tools

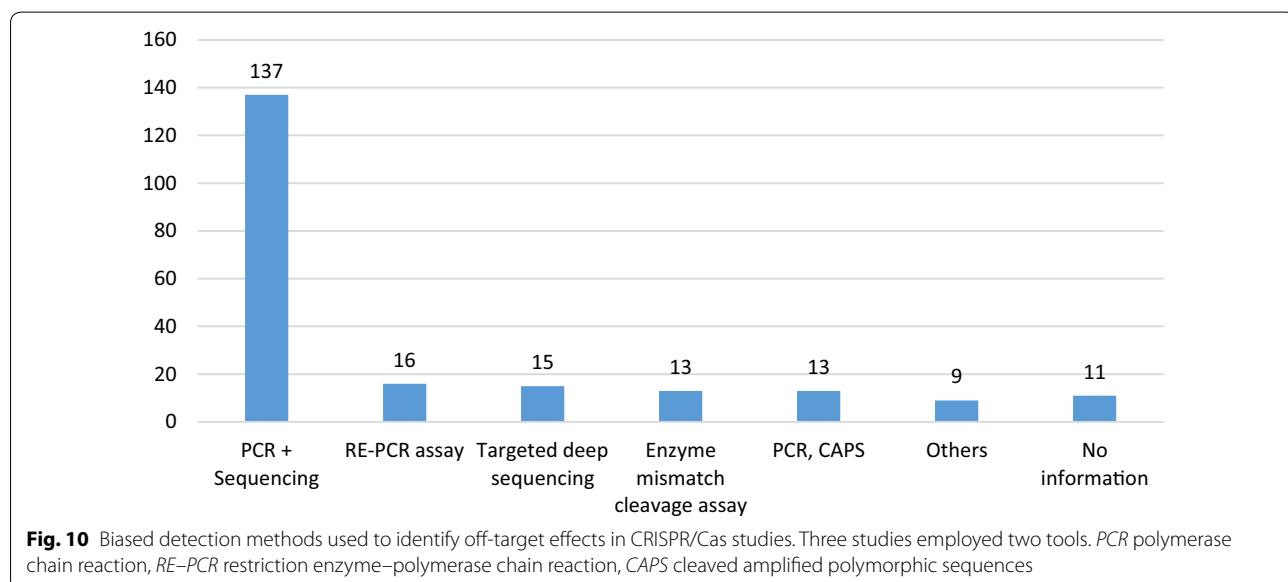


Fig. 10 Biased detection methods used to identify off-target effects in CRISPR/Cas studies. Three studies employed two tools. PCR polymerase chain reaction, RE-PCR restriction enzyme–polymerase chain reaction, CAPS cleaved amplified polymorphic sequences

identified analyzing the occurrence of off-target effects in TALENs and BE studies. The amount of off-target studies for ZFN ($n=4$), ODM ($n=1$) and MN ($n=1$) were even lower. For a more in-depth off-target analysis regarding these techniques, further research and more primary studies are needed. However, the CRISPR/Cas technique is applied much more frequently because of efficiency, time saving and cost-effectiveness compared to TALENs, ZFN, ODM and MN [182]. Therefore, it is questionable what additional research effort is justified. BE differs as it

has great potential to support plant breeding, though it is not broadly established yet. It is based on the CRISPR/Cas9 system but enables exchanging individual base pairs at specific sites without inducing a DSB. Our map shows that only a few BE studies addressed the occurrence of off-target effects so far. The likely reason is that BE is a “young” method in plants, and its first publication was in 2017. Further research about BE including the analysis of off-target effects is highly recommended. Another knowledge gap affecting all genome-editing techniques is

Table 9 Overview of the amount of studies analyzing potential off-target sites, the amount of analyzed potential off-target sites and the amount of identified off-target effects using biased detection methods in different plant species

Plant species	Amount of studies analyzing preselected potential off-target sites	Amount of analyzed potential off-target sites—biased	Amount of identified off-target effects—biased
Rice	77	296	25
Tomato	23	213	1
Arabidopsis	21	229	4
Moss	13	58	0
Soybean	12	106	6
Tobacco	9	50	1
Wheat	8	85	6
Maize	7	23	2
Algae	4	357	0
Canola	4	73	0
Cotton	4	58	0
Orange	3	21	6
Grapevine	3	13	0
Poplar	3	6	0
Grapefruit	2	15	0
Cucumber	2	9	0
Clover	2	3	0
Lettuce	1	91	0
Cacao	1	9	0
Flax	1	8	0
Barley	1	4	1
Kiwifruit	1	4	0
Camelina	1	3	0
Citrangle	1	3	0
Watermelon	1	3	0
Cabbage	1	2	1
Carrot	1	1	1
Potato	1	1	0
Sage	1	1	0
Wild strawberry	1	1	0
Alfalfa	1	No information	0

the genome-wide analysis of off-target effects in a completely unrestricted way. Only nine studies addressed this aspect using the CRISPR/Cas technique and one using the TALENs technique. No study analyzed off-target effects using unbiased detection methods in ZFN, ODM, MN and BE studies.

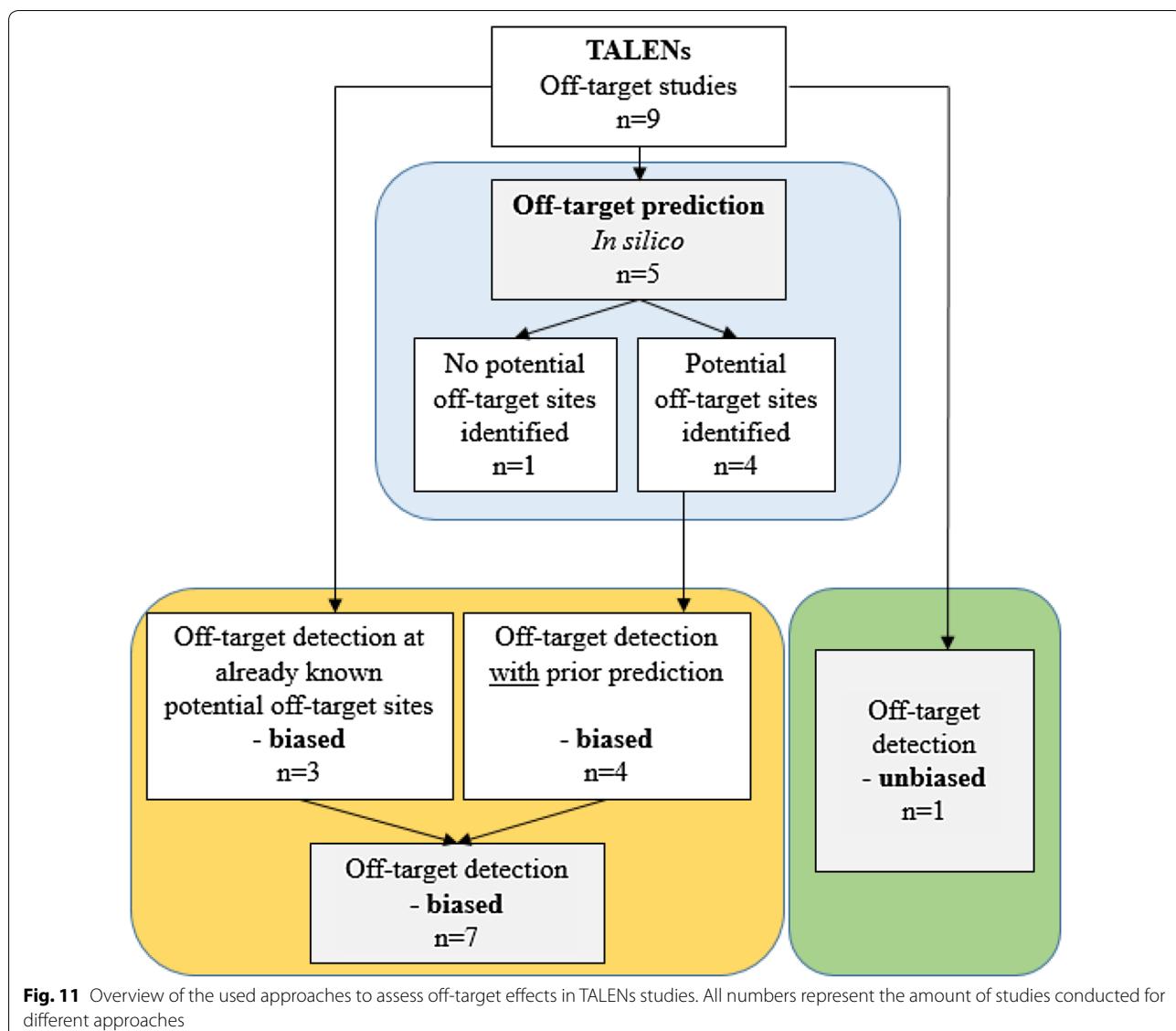
Knowledge clusters

The analysis of off-target effects in CRISPR/Cas studies using biased detection methods represents a knowledge cluster. This cluster is amenable for a critical appraisal serving the interest of researchers and decision-makers. The following parameters could affect the occurrence of off-targets and are worth to be analyzed in-depth:

- Genome composition, size and ploidy level of different plant species.
- Amount of tolerated mismatches at the potential off-target sites.
- Quality of available biased detection methods.
- CRISPR variants (e.g. Nickase).
- Chosen sgRNA.
- Methodology of SDN delivery.

Limitations of the systematic map

References were only searched in German and English language. Publications in an Asian language, in which only the abstract was available in English, were also



identified but could not be included in this map for the extraction of detailed data. Since most of the studies were found in scientific journals, a bias in the pool of articles found is that publications from countries that probably use genome editing but do not publish in either English or German language are not represented. For example, it can be assumed that in South American countries, considerably more research is done than the identified literature suggests. Identifying grey literature as well as manually identifying scientific journals publishing in the local language(s) was also not possible due to language barriers.

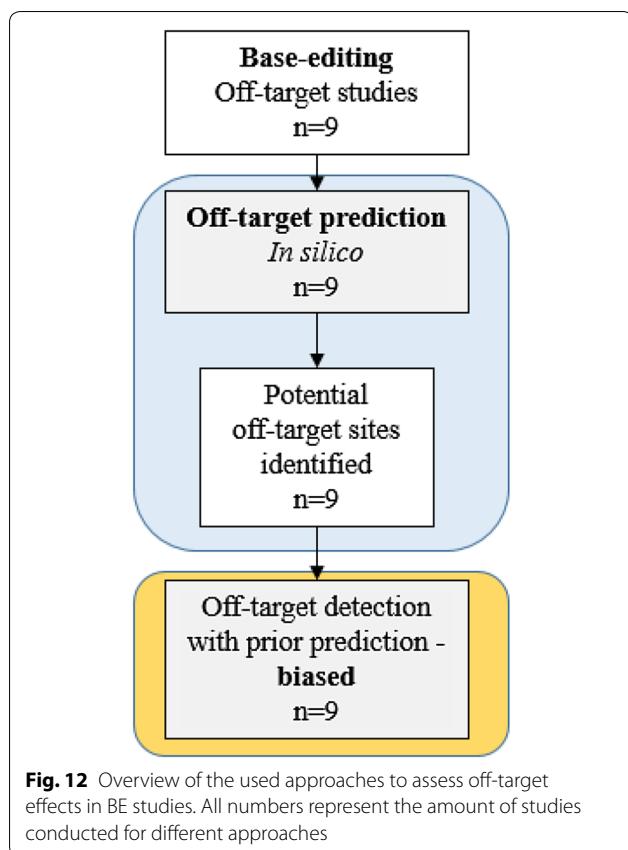
Additionally, the full text of 104 articles, that have been rated as relevant on title/abstract level, were un-retrievable and were therefore not included in the systematic map (see Additional file 3).

It has been demonstrated that the individual studies addressing off-target effects differ widely in design and implementation. Therefore, no reliable conclusions about the occurrence of off-target effects can be drawn based on the results of this map. A critical appraisal of the individual studies in form of a systematic review is recommended.

Conclusions

Implications for policy/management

This systematic map identified substantial bodies of evidence regarding the applications of different genome-editing techniques in plants as well as the occurrence of off-target effects. Until May 2018, almost 100 market-oriented applications were identified including improved food and feed quality, yield improvement, altered growth



characteristics, resistance against biotic and abiotic stress, herbicide tolerance and industrial utilization. The wide range of different applications addressing all parts in the agricultural value chain and the application in many different plant species indicates that genome-editing became a promising tool to breed varieties that are better adapted to the needs of agriculture and to enable a valuable contribution to food security and the environment.

A decisive factor that impacts the use of genome-editing is the acceptance by consumers and retailers for products derived from innovative plant breeding methods. A prominent point of criticism in this context is the occurrence of off-target effects. Since plant breeding is the contextual background, it is worth to compare the occurrence of off-target effects in the context of naturally occurring mutations and routinely used breeding techniques like regular crossing or undirected mutagenesis using chemical mutagens or irradiation [10].

Implication for research

Results of this systematic map determined that different approaches were used to analyze off-target effects depending on the plant species and with regard to the CRISPR-variant, the prediction tools and detection methods used, the amount of tolerated mismatches and

the chosen sgRNA. A critical appraisal in the course of a systematic review could help to identify parameters in order to further reduce the occurrence of off-target effects. The identified knowledge cluster for the detection of off-target effects in CRISPR-studies using biased detection methods would be suitable to address this.

The results of this map further identified a knowledge gap regarding the analysis of off-target effects using unbiased detection tools. To increase this data basis and to evaluate whether an unbiased off-target analysis has an added value compared to biased off-target analysis, more studies should assess this aspect, in future.

Additional files

Additional file 1. Detailed description of the genome-editing techniques.

Additional file 2. List of peer-reviewed and grey literature sources searched.

Additional file 3. List of un-retrievable articles.

Additional file 4. List of articles excluded at full text assessment with the reason for exclusion.

Additional file 5. List of relevant studies and the systematic map database to answer secondary question 1.

Additional file 6. List of relevant studies and the systematic map database to answer secondary question 2.

Additional file 7. ROSES Reporting Form.

Acknowledgements

The authors wish to thank Maren Fischer for carefully reading the manuscript.

Authors' contributions

The review process was coordinated by DM. DM, FH, TS and DK conducted the screening of articles and data extraction. DM drafted the report. FH, TS, CK and RW assisted in editing and revising the draft. All authors read and approved the final manuscript.

Funding

This systematic map is part of the ELSA-GEA project funded by the Federal Ministry of Education and Research (BMBF) (01GP1613B).

Availability of data and materials

The datasets generated and/or analyzed during the current study are available at the ELSA-GEA homepage <https://www.dialog-gea.de/de/service/repository>.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The reviewers Frank Hartung (FH) and Thorben Sprink (TS) are authors of a few research studies included in the review process. However, as none of these studies comprise primary data, their articles were excluded at title/abstract level.

Received: 28 March 2019 Accepted: 27 June 2019

Published online: 12 July 2019

References

- Becker H. Pflanzenzüchtung. 2nd ed. Stuttgart: UTB; 2011.
- Ronald P. Plant genetics, sustainable agriculture and global food security. *Genetics*. 2011;188:11–20. <https://doi.org/10.1534/genetics.111.128553>.
- FAO—Committee on Agriculture (17th session). 06.03.2003. <http://www.fao.org/docrep/meeting/006/y8704e.htm>. Accessed 25 Sept 2018.
- Xie K, Wu S, Li Z, Zhou Y, Zhang D, Dong Z, et al. Map-based cloning and characterization of Zea mays male sterility33 (ZmMs33) gene, encoding a glycerol-3-phosphate acyltransferase. *Theor Appl Genet*. 2018;131:1363–78. <https://doi.org/10.1007/s00122-018-3083-9>.
- Phalan B. What have we learned from the land sparing-sharing model? *Sustainability*. 2018;10:1760. <https://doi.org/10.3390/su10061760>.
- Stevenson JR, Villoria N, Byerlee D, Kelley T, Maredia M. Green Revolution research saved an estimated 18 to 27 million hectares from being brought into agricultural production. *Proc Natl Acad Sci USA*. 2013;110:8363–8. <https://doi.org/10.1073/pnas.1208065110>.
- Songstad DD, Petolino JF, Voytas DF, Reichert NA. Genome editing of plants. *Crit Rev Plant Sci*. 2017;36:1–23. <https://doi.org/10.1080/07352689.2017.1281663>.
- International Atomic Energy Agency (IAEA). Mutant variety database. <https://mvdb.iaea.org/#!Search?page=1&size=15&sortby=Name&sort=ASC>. Accessed 25 Sept 2018.
- Schlegel RHJ. Dictionary of plant breeding. 2nd ed. Boca Raton: CRC Press; 2009.
- SAM High Level Group of Scientific Advisors. New techniques in agricultural biotechnology. 2017.
- Barton KA, Binns AN, Matzke AJM, Chilton M-D. Regeneration of intact tobacco plants containing full length copies of genetically engineered T-DNA, and transmission of T-DNA to R1 progeny. *Cell*. 1983;32:1033–43. [https://doi.org/10.1016/0092-8674\(83\)90288-X](https://doi.org/10.1016/0092-8674(83)90288-X).
- Herrera-Estrella L, Depicker A, van Montagu M, Schell J. Expression of chimaeric genes transferred into plant cells using a Ti-plasmid-derived vector. *Nature*. 1983;303:209–13. <https://doi.org/10.1038/303209a0>.
- Govindan G, Ramalingam S. Programmable site-specific nucleases for targeted genome engineering in higher eukaryotes. *J Cell Physiol*. 2016;231:2380–92. <https://doi.org/10.1002/jcp.25367>.
- European Food Safety Authority (EFSA). Scientific opinion addressing the safety assessment of plants developed using Zinc Finger Nuclease 3 and other Site-Directed Nucleases with similar function. EFSA J. 2012;10:2943.
- van de Wiel CCM, Schaart JG, Lotz LAP, Smulders MJM. New traits in crops produced by genome editing techniques based on deletions. *Plant Biotechnol Rep*. 2017;11:1–8. <https://doi.org/10.1007/s11816-017-0425-z>.
- Podevin N, Davies HV, Hartung F, Nogué F, Casacuberta JM. Site-directed nucleases: a paradigm shift in predictable, knowledge-based plant breeding. *Trends Biotechnol*. 2013;31:375–83. <https://doi.org/10.1016/j.tibtech.2013.03.004>.
- Komor AC, Kim YB, Packer MS, Zuris JA, Liu DR. Programmable editing of a target base in genomic DNA without double-stranded DNA cleavage. *Nature*. 2016;533:420–4. <https://doi.org/10.1038/nature17946>.
- Nationale Akademie der Wissenschaften Leopoldina, Deutsche Forschungsgemeinschaft, acatech – Deutsche Akademie der Technikwissenschaften, Union der deutschen Akademien der Wissenschaften. The opportunities and limits of genome editing. 2015.
- Bömeke O, Kahrmann J, Matthies A. Detaillierte Übersicht zum regulatorischen Status der neuen molekulärbiologischen Techniken (NMT) in ausgewählten Drittstaaten. https://www.bmel.de/DE/Landwirtschaft/Pflanzenbau/Gentechnik/_Texte/Neue_molekulärbiologische_Techniken.html. Accessed 21 Feb 2019.
- Duan Y-B, Li J, Qin R-Y, Xu R-F, Li H, Yang Y-C, et al. Identification of a regulatory element responsible for salt induction of rice OsRAV2 through ex situ and in situ promoter analysis. *Plant Mol Biol*. 2016;90:49–62. <https://doi.org/10.1007/s11103-015-0393-z>.
- Demorest ZL, Coffman A, Baltes NJ, Stoddard TJ, Clasen BM, Luo S, et al. Direct stacking of sequence-specific nuclease-induced mutations to produce high oleic and low linolenic soybean oil. *BMC Plant Biol*. 2016;16:225. <https://doi.org/10.1186/s12870-016-0906-1>.
- Chandrasekaran J, Brumin M, Wolf D, Leibman D, Klap C, Pearlsman M, et al. Development of broad virus resistance in non-transgenic cucumber using CRISPR/Cas9 technology. *Mol Plant Pathol*. 2016;17:1140–53. <https://doi.org/10.1111/mpp.12375>.
- Gocal GFW, Schöpke C, Beetham PR. Oligo-mediated targeted gene editing. In: Zhang F, Puchta H, Thomson JG, editors. Advances in new technology for targeted modification of plant genomes. New York: Springer; 2015. p. 73–89. https://doi.org/10.1007/978-1-4939-2556-8_5.
- Shi J, Gao H, Wang H, Lafitte HR, Archibald RL, Yang M, et al. ARGOS8 variants generated by CRISPR–Cas9 improve maize grain yield under field drought stress conditions. *Plant Biotechnol J*. 2017;15:207–16. <https://doi.org/10.1111/pbi.12603>.
- Wang Y, Cheng X, Shan Q, Zhang Y, Liu J, Gao C, Qiu J-L. Simultaneous editing of three homoeoalleles in hexaploid bread wheat confers heritable resistance to powdery mildew. *Nat Biotechnol*. 2014;32:947–51. <https://doi.org/10.1038/nbt.2969>.
- Martin F, Sánchez-Hernández S, Gutiérrez-Guerrero A, Pinedo-Gomez J, Benabdellah K. Biased and unbiased methods for the detection of off-target cleavage by CRISPR/Cas9: an overview. *Int J Mol Sci*. 2016. <https://doi.org/10.3390/ijms17091507>.
- Altschul S, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol*. 1990;215:403–10. <https://doi.org/10.1006/jmbi.1990.9999>.
- Bae S, Park J, Kim J-S. Cas-OFFinder: a fast and versatile algorithm that searches for potential off-target sites of Cas9 RNA-guided endonucleases. *Bioinformatics*. 2014;30:1473–5. <https://doi.org/10.1093/bioinformatics/btu048>.
- Lei Y, Lu L, Liu H-Y, Li S, Xing F, Chen L-L. CRISPR-P: a web tool for synthetic single-guide RNA design of CRISPR-system in plants. *Mol Plant*. 2014;7:1494–6. <https://doi.org/10.1093/mp/ssu044>.
- Liu H, Ding Y, Zhou Y, Jin W, Xie K, Chen L-L. CRISPR-P 2.0: an improved CRISPR–Cas9 tool for genome editing in plants. *Mol Plant*. 2017;10:530–2. <https://doi.org/10.1016/j.molp.2017.01.003>.
- Tycko J, Myer VE, Hsu PD. Methods for optimizing CRISPR–Cas9 genome editing specificity. *Mol Cell*. 2016;63:355–70. <https://doi.org/10.1016/j.molcel.2016.07.004>.
- Zischewski J, Fischer R, Bortesi L. Detection of on-target and off-target mutations generated by CRISPR/Cas9 and other sequence-specific nucleases. *Biotechnol Adv*. 2017;35:95–104. <https://doi.org/10.1016/j.biotechadv.2016.12.003>.
- Agapito-Tenfen SZ, Wikmark O-G. Current status of emerging technologies for plant breeding. Biosafety and knowledge gaps of site directed nucleases and oligonucleotide-directed mutagenesis. *GenØk Centre for Biosafety*: Tromsø; 2015.
- Zhang X-H, Tee LY, Wang X-G, Huang Q-S, Yang S-H. Off-target effects in CRISPR/Cas9-mediated genome engineering. *Mol Ther Nucleic Acids*. 2015;4:e264. <https://doi.org/10.1038/mtna.2015.37>.
- Kanchiswamy CN, Maffei M, Malnoy M, Velasco R, Kim J-S. Fine-tuning next-generation genome editing tools. *Trends Biotechnol*. 2016;34:562–74. <https://doi.org/10.1016/j.tibtech.2016.03.007>.
- Collaboration for Environmental Evidence. Guidelines for systematic review and evidence synthesis in environmental management, Version 4.2. Environmental Evidence. 2013. <http://www.environmentalevidence.e.org/Documents/Guidelines/Guidelines4.2.pdf>. Accessed 13 Sept 2018.
- Modrzejewski D, Hartung F, Sprink T, Krause D, Kohl C, Schiemann J, Wilhelm R. What is the available evidence for the application of genome editing as a new tool for plant trait modification and the potential occurrence of associated off-target effects: a systematic map protocol. *Environ Evid*. 2018;7:11. <https://doi.org/10.1186/s13750-018-0130-6>.
- Cohen J. A coefficient of agreement for nominal scales. *Educ Psychol Meas*. 1960;20:37–46. <https://doi.org/10.1177/001316446002000104>.
- Kohl C, McIntosh EJ, Unger S, Haddaway NR, Kecke S, Schiemann J, Wilhelm R. Online tools supporting the conduct and reporting of systematic reviews and systematic maps: a case study on CADIMA and review of existing tools. *Environ Evid*. 2018;7:2420. <https://doi.org/10.1186/s13750-018-0115-5>.
- Haddaway NR, Macura B, Whaley P, Pullin AS. ROSES for Systematic Map Reports. Version 1.0. 2017. <https://doi.org/10.6084/m9.figshare.5897299>.
- Braatz J, Harloff H-J, Mascher M, Stein N, Himmelbach A, Jung C. CRISPR–Cas9 targeted mutagenesis leads to simultaneous modification of different homoeologous gene copies in polyploid oilseed rape

- (*Brassica napus*). Plant Physiol. 2017;174:9. <https://doi.org/10.1104/pp.17.00426>.
42. Yang Y, Zhu K, Li H, Han S, Meng Q, Khan SU, et al. Precise editing of CLAVATA genes in *Brassica napus* L. regulates multilocular silique development. Plant Biotechnol J. 2018;16:1322–35. <https://doi.org/10.1111/pbi.12872>.
 43. Wang Y, Meng Z, Liang C, Meng Z, Wang Y, Sun G, et al. Increased lateral root formation by CRISPR/Cas9-mediated editing of arginase genes in cotton. Sci China Life Sci. 2017;60:524–7. <https://doi.org/10.1007/s11427-017-9031-y>.
 44. Hu B, Li D, Liu X, Qi J, Gao D, Zhao S, et al. Engineering non-transgenic gynoecious cucumber using an improved transformation protocol and optimized CRISPR/Cas9 system. Mol Plant. 2017;10:1575–8. <https://doi.org/10.1016/j.molp.2017.09.005>.
 45. Bertier LD, Ron M, Huo H, Bradford KJ, Britt AB, Michelmore RW. High-resolution analysis of the efficiency, heritability, and editing outcomes of CRISPR/Cas9-induced modifications of NCED4 in lettuce (*Lactuca sativa*). G3. 2018;8:1513–21. <https://doi.org/10.1534/g3.117.300396>.
 46. United States Department of Agriculture (USDA). 2015. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/15-013-01air.pdf. Accessed 25 Aug 2018.
 47. Huang C, Sun H, Xu D, Chen Q, Liang Y, Wang X, et al. ZmCCT9 enhances maize adaptation to higher latitudes. Proc Natl Acad Sci USA. 2018;115:E334–41. <https://doi.org/10.1073/pnas.1718058115>.
 48. Svitashov S, Young JK, Schwartz C, Gao H, Falco SC, Cigan AM. Targeted mutagenesis, precise gene editing, and site-specific gene insertion in maize using Cas9 and guide RNA. Plant Physiol. 2015;169:931–45. <https://doi.org/10.1104/pp.15.00793>.
 49. Svitashov S, Schwartz C, Lenderts B, Young JK, Mark Cigan A. Genome editing in maize directed by CRISPR–Cas9 ribonucleoprotein complexes. Nat Commun. 2016;7:1–7. <https://doi.org/10.1038/ncomms13274>.
 50. Li J, Zhang H, Si X, Tian Y, Chen K, Liu J, et al. Generation of thermo-sensitive male-sterile maize by targeted knockout of the ZmTMS5 gene. J Genet Genom. 2017;44:465–8. <https://doi.org/10.1016/j.jgg.2017.02.002>.
 51. Kelliher T, Starr D, Richbourg L, Chintamanani S, Delzer B, Nuccio ML, et al. MATRILINEAL, a sperm-specific phospholipase, triggers maize haploid induction. Nature. 2017;542:105–9. <https://doi.org/10.1038/nature20827>.
 52. Clasen BM, Stoddard TJ, Luo S, Demorest ZL, Li J, Cedrone F, et al. Improving cold storage and processing traits in potato through targeted gene knockout. Plant Biotechnol J. 2016;14:169–76. <https://doi.org/10.1111/pbi.12370>.
 53. Li M, Li X, Zhou Z, Wu P, Fang M, Pan X, et al. Reassessment of the four yield-related genes Gn1a, DEP1, GS3, and IPA1 in rice using a CRISPR/Cas9 system. Front Plant Sci. 2016;7:1–13. <https://doi.org/10.3389/fpls.2016.00377>.
 54. Shen L, Wang C, Fu Y, Wang J, Liu Q, Zhang X, et al. QTL editing confers opposing yield performance in different rice varieties. J Integr Plant Biol. 2016. <https://doi.org/10.1111/jipb.12501>.
 55. Xu R, Yang Y, Qin R, Li H, Qiu C, Li L, et al. Rapid improvement of grain weight via highly efficient CRISPR/Cas9-mediated multiplex genome editing in rice. J Genet Genom. 2016;43:529–32. <https://doi.org/10.1016/j.jgg.2016.07.003>.
 56. Hu Z, Lu S-J, Wang M-J, He H, Sun L, Wang H, et al. A novel QTL q TGW3 encodes the GSK3/SHAGGY-like kinase OsGSK5/OsSK41 that interacts with OsARF4 to negatively regulate grain size and weight in rice. Mol Plant. 2018;11:736–49. <https://doi.org/10.1016/j.molp.2018.03.005>.
 57. Shen L, Li J, Fu Y, Wang J, Hua Y, Jiao X, Yan C, Wang K. Orientation improvement of grain length and grain number in rice by using CRISPR/Cas9 system. Chin J Rice Sci. 2017. <https://doi.org/10.16819/j.1001-7216.2017.7029>.
 58. Ji X, Li F, Yan Y, Sun HZ, Zhang J, Li JZ, et al. CRISPR/Cas9 system-based editing of phytochrome-interacting factor OsPIL15. Sci Agric Sin. 2017. <https://doi.org/10.3864/j.issn.0578-1752.2017.15.002>.
 59. Shen L, Hua Y, Fu Y, Li J, Liu Q, Jiao X, et al. Rapid generation of genetic diversity by multiplex CRISPR/Cas9 genome editing in rice. Sci China Life Sci. 2017;60:506–15. <https://doi.org/10.1007/s11427-017-9008-8>.
 60. Lu K, Wu B, Wang J, Zhu W, Nie H, Qian J, et al. Blocking amino acid transporter OsAAP3 improves grain yield by promoting outgrowth buds and increasing tiller number in rice. Plant Biotechnol J. 2018;50:1416. <https://doi.org/10.1111/pbi.12907>.
 61. Liao Y, Bai Q, Xu P, Wu T, Guo D, Peng Y, et al. Mutation in rice abscisic acid2 results in cell death, enhanced disease-resistance, altered seed dormancy and development. Front Plant Sci. 2018;9:1248. <https://doi.org/10.3389/fpls.2018.00405>.
 62. Li X, Zhou W, Ren Y, Tian X, Lv T, Wang Z, et al. High-efficiency breeding of early-maturing rice cultivars via CRISPR/Cas9-mediated genome editing. J Genet Genom. 2017;44:175–8. <https://doi.org/10.1016/j.jgg.2017.02.001>.
 63. Lee S-K, Eom J-S, Hwang S-K, Shin D, An G, Okita TW, Jeon J-S. Plastidic phosphoglucomutase and ADP-glucose pyrophosphorylase mutants impair starch synthesis in rice pollen grains and cause male sterility. J Exp Bot. 2016;67:5557–69. <https://doi.org/10.1093/jxb/erv324>.
 64. Li Q, Zhang D, Chen M, Liang W, Wei J, Qi Y, Yuan Z. Development of japonica photo-sensitive genic male sterile rice lines by editing carbon starved anther using CRISPR/Cas9. J Genet Genom. 2016;43:415–9. <https://doi.org/10.1016/j.jgg.2016.04.011>.
 65. Xie Y, Niu B, Long Y, Li G, Tang J, Zhang Y, et al. Suppression or knockout of SaF/SaM overcomes the Sa-mediated hybrid male sterility in rice. J Integr Plant Biol. 2017;59:669–79. <https://doi.org/10.1111/jipb.12564>.
 66. Zhou H, He M, Li J, Chen L, Huang Z, Zheng S, et al. Development of commercial thermo-sensitive genic male sterile rice accelerates hybrid rice breeding using the CRISPR/Cas9-mediated TMS5 editing system. Sci Rep. 2016;6:1–12. <https://doi.org/10.1038/srep37395>.
 67. Zou T, He Z, Qu L, Liu M, Zeng J, Liang Y, et al. Knockout of OsACOS12 caused male sterility in rice. Mol Breed. 2017;37:437. <https://doi.org/10.1007/s11032-017-0722-9>.
 68. Zou T, Xiao Q, Li W, Luo T, Yuan G, He Z, et al. OsLAP6/OsPKS1, an orthologue of *Arabidopsis* PKSA/LAP6, is critical for proper pollen exine formation. Rice. 2017;10:615. <https://doi.org/10.1186/s12284-017-0191-0>.
 69. Liu L, Zheng C, Kuang B, Wei L, Yan L, Wang T. Receptor-like kinase RUPO interacts with potassium transporters to regulate pollen tube growth and integrity in rice. PLoS Genet. 2016;12:e1006085. <https://doi.org/10.1371/journal.pgen.1006085>.
 70. Ma L, Zhu F, Li Z, Zhang J, Li X, Dong J, Wang T. TALEN-based mutagenesis of lipoxygenase LOX3 enhances the storage tolerance of rice (*Oryza sativa*) seeds. PLoS ONE. 2015;10:e0143877. <https://doi.org/10.1371/journal.pone.0143877>.
 71. Qian W, Wu C, Fu Y, Hu G, He Z, Liu W. Novel rice mutants overexpressing the brassinosteroid catabolic gene CYP73A4. Plant Mol Biol. 2017;93:197–208. <https://doi.org/10.1007/s11103-016-0558-4>.
 72. Yuan J, Chen S, Jiao W, Wang L, Wang L, Ye W, et al. Both maternally and paternally imprinted genes regulate seed development in rice. New Phytol. 2017;216:373–87. <https://doi.org/10.1111/nph.14510>.
 73. Lu Y, Zhu J-K. Precise editing of a target base in the rice genome using a modified CRISPR/Cas9 system. Mol Plant. 2017;10:523–5. <https://doi.org/10.1016/j.molp.2016.11.013>.
 74. Wang Y, Geng L, Yuan M, Wei J, Jin C, Li M, et al. Deletion of a target gene in Indica rice via CRISPR/Cas9. Plant Cell Rep. 2017;36:1333–43. <https://doi.org/10.1007/s00299-017-2158-4>.
 75. Huang Y, Guo Y, Liu Y, Zhang F, Wang Z, Wang H, et al. 9-cis-Epoxy-carotenoid dioxygenase 3 regulates plant growth and enhances multi-abiotic stress tolerance in rice. Front Plant Sci. 2018;9:1248. <https://doi.org/10.3389/fpls.2018.00162>.
 76. Cai Y, Chen L, Liu X, Guo C, Sun S, Wu C, et al. CRISPR/Cas9-mediated targeted mutagenesis of GmFT2a delays flowering time in soya bean. Plant Biotechnol J. 2017. <https://doi.org/10.1111/pbi.12758>.
 77. Liu Y, Merrick P, Zhang Z, Ji C, Yang B, Fei S-Z. Targeted mutagenesis in tetraploid switchgrass (*Panicum virgatum* L.) using CRISPR/Cas9. Plant Biotechnol J. 2018;16:381–93. <https://doi.org/10.1111/pbi.12778>.
 78. Ito Y, Nishizawa-Yokoi A, Endo M, Mikami M, Toki S. CRISPR/Cas9-mediated mutagenesis of the RIN locus that regulates tomato fruit ripening. Biochem Biophys Res Commun. 2015;467:76–82. <https://doi.org/10.1016/j.bbrc.2015.09.117>.
 79. Lor VS, Starker CG, Voytas DF, Weiss D, Olszewski NE. Targeted mutagenesis of the tomato PROCERA gene using transcription activator-like effector nucleases. Plant Physiol. 2014;166:1288–91. <https://doi.org/10.1104/pp.114.247593>.
 80. Soyk S, Müller NA, Park SJ, Schmalenbach I, Jiang K, Hayama R, et al. Variation in the flowering gene SELF PRUNING 5G promotes day-neutrality

- and early yield in tomato. *Nat Genet.* 2017;49:162–8. <https://doi.org/10.1038/ng.3733>.
81. United States Department of Agriculture (USDA). 2018. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/18-051-01_air_response_signed.pdf. Accessed 25 Aug 2018.
 82. Rodríguez-Leal D, Lemmon ZH, Man J, Bartlett ME, Lippman ZB. Engineering quantitative trait variation for crop improvement by genome editing. *Cell.* 2017;171(470–480):e8. <https://doi.org/10.1016/j.cell.2017.08.030>.
 83. Filler Hayut S, Melamed Bessudo C, Levy AA. Targeted recombination between homologous chromosomes for precise breeding in tomato. *Nat Commun.* 2017;8:15605. <https://doi.org/10.1038/ncomms15605>.
 84. Dahan-Meir T, Filler-Hayut S, Melamed-Bessudo C, Bocobza S, Czosnek H, Aharoni A, Levy AA. Efficient in planta gene targeting in tomato using geminiviral replicons and the CRISPR/Cas9 system. *Plant J.* 2018;95:5–16. <https://doi.org/10.1111/tpj.13932>.
 85. Deng L, Wang H, Sun C, Li Q, Jiang H, Du M, et al. Efficient generation of pink-fruited tomatoes using CRISPR/Cas9 system. *J Genet Genom.* 2018;45:51–4. <https://doi.org/10.1016/j.jgg.2017.10.002>.
 86. Wang W, Pan Q, He F, Akhunova A, Chao S, Trick H, Akhunov E. Transgenerational CRISPR–Cas9 activity facilitates multiplex gene editing in allopolyploid wheat. *CRISPR J.* 2018;1:65–74. <https://doi.org/10.1089/crispr.2017.0010>.
 87. Zhang Y, Li D, Zhang D, Zhao X, Cao X, Dong L, et al. Analysis of the functions of TaGW2 homoeologs in wheat grain weight and protein content traits. *Plant J.* 2018;94:857–66. <https://doi.org/10.1111/tpj.13903>.
 88. Zhou J, Wang G, Liu Z. Efficient genome editing of wild strawberry genes, vector development and validation. *Plant Biotechnol J.* 2018;16:61292. <https://doi.org/10.1111/pbi.12922>.
 89. United States Department of Agriculture (USDA). 2017. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/17-038-02_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
 90. Ozseyan ME, Kang J, Mu X, Lu C. Mutagenesis of the FAE1 genes significantly changes fatty acid composition in seeds of *Camelina sativa*. *Plant Physiol Biochem.* 2018;123:1–7. <https://doi.org/10.1016/j.plaphy.2017.11.021>.
 91. Jiang WZ, Henry IM, Lynagh PG, Comai L, Cahoon EB, Weeks DP. Significant enhancement of fatty acid composition in seeds of the allohexaploid, *Camelina sativa*, using CRISPR/Cas9 gene editing. *Plant Biotechnol J.* 2017;15:648–57. <https://doi.org/10.1111/pbi.12663>.
 92. Morineau C, Belloc Y, Tellier F, Gissot L, Kelemen Z, Nogué F, Faure J-D. Selective gene dosage by CRISPR–Cas9 genome editing in hexaploid *Camelina sativa*. *Plant Biotechnol J.* 2017;15:729–39. <https://doi.org/10.1111/pbi.12671>.
 93. Aznar-Moreno JA, Durrett TP. Simultaneous targeting of multiple gene homologs to alter seed oil production in *Camelina sativa*. *Plant Cell Physiol.* 2017;58:1260–7. <https://doi.org/10.1093/pcp/pcx058>.
 94. Okuzaki A, Ogawa T, Koizuka C, Kaneko K, Inaba M, Imamura J, Koizuka N. CRISPR/Cas9-mediated genome editing of the fatty acid desaturase 2 gene in *Brassica napus*. *Plant Physiol Biochem.* 2018. <https://doi.org/10.1016/j.plaphy.2018.04.025>.
 95. United States Department of Agriculture (USDA). 2015. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/15-352-01_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
 96. Qi X, Dong L, Liu C, Mao L, Liu F, Zhang X, et al. Systematic identification of endogenous RNA polymerase III promoters for efficient RNA guide-based genome editing technologies in maize. *Crop J.* 2018;6:314–20. <https://doi.org/10.1016/j.cj.2018.02.005>.
 97. United States Department of Agriculture (USDA). 2015. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/15-078-02_air_inquiry.pdf. Accessed 25 Aug 2018.
 98. United States Department of Agriculture (USDA). 2010. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/DOW_Email_9620to_Susan_%20Kohler_032010.pdf. Accessed 25 Aug 2018.
 99. Shukla VK, Doyon Y, Miller JC, DeKelver RC, Moehle EA, Worden SE, et al. Precise genome modification in the crop species *Zea mays* using zinc-finger nucleases. *Nature.* 2009;459:437–41. <https://doi.org/10.1038/nature07992>.
 100. United States Department of Agriculture (USDA). 2015. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/15-321-01_air_inquiry.pdf. Accessed 25 Aug 2018.
 101. Alagoz Y, Gurkok T, Zhang B, Unver T. Manipulating the biosynthesis of bioactive compound alkaloids for next-generation metabolic engineering in opium poppy using CRISPR–Cas 9 genome editing technology. *Sci Rep.* 2016;6:30910–8. <https://doi.org/10.1038/srep30910>.
 102. Wen S, Liu H, Li X, Chen X, Hong Y, Li H, et al. TALEN-mediated targeted mutagenesis of fatty acid desaturase 2 (FAD2) in peanut (*Arachis hypogaea* L.) promotes the accumulation of oleic acid. *Plant Mol Biol.* 2018;97:177–85. <https://doi.org/10.1007/s11103-018-0731-z>.
 103. United States Department of Agriculture (USDA). 2016. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/16-090-01_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
 104. United States Department of Agriculture (USDA). 2016. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/16-320-01_air_inquiry.pdf. Accessed 25 Aug 2018.
 105. Sawai S, Ohyama K, Yasumoto S, Seki H, Sakuma T, Yamamoto T, et al. Sterol side chain reductase 2 is a key enzyme in the biosynthesis of cholesterol, the common precursor of toxic steroidal glycoalkaloids in potato. *Plant Cell.* 2014;26:3763–74. <https://doi.org/10.1105/tpc.114.130096>.
 106. Nakayasu M, Akiyama R, Lee HJ, Osakabe K, Osakabe Y, Watanabe B, et al. Generation of α-solanine-free hairy roots of potato by CRISPR/Cas9 mediated genome editing of the St16DOX gene. *Plant Physiol Biochem.* 2018. <https://doi.org/10.1016/j.plaphy.2018.04.026>.
 107. Shan Q, Zhang Y, Chen K, Zhang K, Gao C. Creation of fragrant rice by targeted knockout of the OsBADH2 gene using TALEN technology. *Plant Biotechnol J.* 2015;13:791–800. <https://doi.org/10.1111/pbi.12312>.
 108. Sun Y, Jiao G, Liu Z, Zhang X, Li J, Guo X, et al. Generation of high-amyllose rice through CRISPR/Cas9-mediated targeted mutagenesis of starch branching enzymes. *Front Plant Sci.* 2017;8:1–15. <https://doi.org/10.3389/fpls.2017.00298>.
 109. Ye Y, Li P, Xu T, Zeng L, Cheng D, Yang M, et al. OsPT4 contributes to arsenate uptake and transport in rice. *Front Plant Sci.* 2017;8:311. <https://doi.org/10.3389/fpls.2017.02197>.
 110. Wang F-Z, Chen M-X, Yu L-J, Xie L-J, Yuan L-B, Qi H, et al. OsARM1, an R2R3 MYB transcription factor, is involved in regulation of the response to arsenic stress in rice. *Front Plant Sci.* 2017;8:1868. <https://doi.org/10.3389/fpls.2017.01868>.
 111. Abe K, Araki E, Suzuki Y, Toki S, Saika H. Production of high oleic/low linoleic rice by genome editing. *Plant Physiol Biochem.* 2018. <https://doi.org/10.1016/j.plaphy.2018.04.033>.
 112. Nieves-Cordones M, Mohamed S, Tanoi K, Kobayashi NI, Takagi K, Vernet A, et al. Production of low-Cs+ rice plants by inactivation of the K+-transporter OsHAK1 with the CRISPR–Cas system. *Plant J.* 2017;92:43–56. <https://doi.org/10.1111/tpj.13632>.
 113. Tang L, Mao B, Li Y, Lv Q, Zhang L, Chen C, et al. Knockout of OsNramp5 using the CRISPR/Cas9 system produces low Cd-accumulating indica rice without compromising yield. *Sci Rep.* 2017;7:14438. <https://doi.org/10.1038/s41598-017-14832-9>.
 114. Zhang J, Zhang H, Botella JR, Zhu J-K. Generation of new glutinous rice by CRISPR/Cas9-targeted mutagenesis of the Waxy gene in elite rice varieties. *J Integr Plant Biol.* 2018;60:369–75. <https://doi.org/10.1111/jipb.12620>.
 115. Zhou Z, Tan H, Li Q, Chen J, Gao S, Wang Y, et al. CRISPR/Cas9-mediated efficient targeted mutagenesis of RAS in *Salvia miltiorrhiza*. *Phytochemistry.* 2018;148:63–70. <https://doi.org/10.1016/j.phytochem.2018.01.015>.
 116. United States Department of Agriculture (USDA). 2014. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/cellectis_air_fad2k_0_soy_cbidel.pdf. Accessed 25 Aug 2018.
 117. United States Department of Agriculture (USDA). 2015. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/15-071-01air.pdf. Accessed 25 Aug 2018.
 118. Haun W, Coffman A, Clasen BM, Demorest ZL, Lowy A, Ray E, et al. Improved soybean oil quality by targeted mutagenesis of the fatty acid desaturase 2 gene family. *Plant Biotechnol J.* 2014;12:934–40. <https://doi.org/10.1111/pbi.12201>.
 119. Klap C, Yeshayahou E, Bolger AM, Arazi T, Gupta SK, Shabtai S, et al. Tomato facultative parthenocarpy results from SLAGAMOUS-LIKE 6 loss

- of function. *Plant Biotechnol J*. 2017;15:634–47. <https://doi.org/10.1111/pbi.12662>.
120. Ueta R, Abe C, Watanabe T, Sugano SS, Ishihara R, Ezura H, et al. Rapid breeding of parthenocarpic tomato plants using CRISPR/Cas9. *Sci Rep*. 2017;7:507. <https://doi.org/10.1038/s41598-017-00501-4>.
 121. Lee J, Nonaka S, Takayama M, Ezura H. Utilization of a genome-edited tomato (*Solanum lycopersicum*) with high gamma aminobutyric acid content in hybrid breeding. *J Agric Food Chem*. 2018;66:963–71. <https://doi.org/10.1021/acs.jafc.7b05171>.
 122. Nonaka S, Arai C, Takayama M, Matsukura C, Ezura H. Efficient increase of γ-aminobutyric acid (GABA) content in tomato fruits by targeted mutagenesis. *Sci Rep*. 2017;7:7057. <https://doi.org/10.1038/s41598-017-06400-y>.
 123. Li X, Wang Y, Chen S, Tian H, Fu D, Zhu B, et al. Lycopene is enriched in tomato fruit by CRISPR/Cas9-mediated multiplex genome editing. *Front Plant Sci*. 2018;9:179. <https://doi.org/10.3389/fpls.2018.00559>.
 124. Yu Q-H, Wang B, Li N, Tang Y, Yang S, Yang T, et al. CRISPR/Cas9-induced targeted mutagenesis and gene replacement to generate long-shelf life tomato lines. *Sci Rep*. 2017;7:818. <https://doi.org/10.1038/s41598-017-12262-1>.
 125. United States Department of Agriculture (USDA). 2017. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/17-038-01_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
 126. Sánchez-León S, Gil-Humanes J, Ozuna CV, Giménez MJ, Sousa C, Voytas DF, Barro F. Low-gluten, nontransgenic wheat engineered with CRISPR/Cas9. *Plant Biotechnol J*. 2017. <https://doi.org/10.1111/pbi.12837>.
 127. Fister AS, Landheir L, Maximova SN, Guiltinan MJ. Transient expression of CRISPR/Cas9 machinery targeting TcNPR3 enhances defense response in *Theobroma cacao*. *Front Plant Sci*. 2018;9:47. <https://doi.org/10.3389/fpls.2018.00268>.
 128. Jia H, Orbović V, Jones JB, Wang N. Modification of the PthA4 effector binding elements in Type I CsLOB1 promoter using Cas9/sgRNA to produce transgenic Duncan grapefruit alleviating XccΔPthA4:dCsLOB1.3 infection. *Plant Biotechnol J*. 2016;14:1291–301. <https://doi.org/10.1111/pbi.12495>.
 129. Jia H, Zhang Y, Orbović V, Xu J, White FF, Jones JB, Wang N. Genome editing of the disease susceptibility gene CsLOB1 in citrus confers resistance to citrus canker. *Plant Biotechnol J*. 2017;15:817–23. <https://doi.org/10.1111/pbi.12677>.
 130. Wang X, Tu M, Wang D, Liu J, Li Y, Li Z, et al. CRISPR/Cas9-mediated efficient targeted mutagenesis in grape in the first generation. *Plant Biotechnol J*. 2018;16:844–55. <https://doi.org/10.1111/pbi.12832>.
 131. United States Department of Agriculture (USDA). 2017. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/17-076-01_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
 132. Peng A, Chen S, Lei T, Xu L, He Y, Wu L, et al. Engineering canker-resistant plants through CRISPR/Cas9-targeted editing of the susceptibility gene CsLOB1 promoter in citrus. *Plant Biotechnol J*. 2017;15:1509–19. <https://doi.org/10.1111/pbi.12733>.
 133. Wang F, Wang C, Liu P, Lei C, Hao W, Gao Y, et al. Enhanced rice blast resistance by CRISPR/Cas9-targeted mutagenesis of the ERF transcription factor gene OsERF922. *PLoS ONE*. 2016;11:e0154027. <https://doi.org/10.1371/journal.pone.0154027>.
 134. Zhou J, Peng Z, Long J, Sosso D, Liu B, Eom J-S, et al. Gene targeting by the TAL effector PthXo2 reveals cryptic resistance gene for bacterial blight of rice. *Plant J*. 2015;82:632–43. <https://doi.org/10.1111/tpj.12838>.
 135. Blanvillain-Baufumé S, Reschke M, Solé M, Auguy F, Doucoure H, Szurek B, et al. Targeted promoter editing for rice resistance to *Xanthomonas oryzae* pv. *oryzae* reveals differential activities for SWEET14-inducing TAL effectors. *Plant Biotechnol J*. 2017;15:306–17. <https://doi.org/10.1111/pbi.12613>.
 136. Li T, Liu B, Spalding MH, Weeks DP, Yang B. High-efficiency TALEN-based gene editing produces disease-resistant rice. *Nat Biotechnol*. 2012;30:390–2. <https://doi.org/10.1038/nbt.2199>.
 137. Wang J, Tian D, Gu K, Yang X, Wang L, Zeng X, Yin Z. Induction of Xa10-like genes in rice cultivar nipponbare confers disease resistance to rice bacterial blight. *Mol Plant Microbe Interact*. 2017;30:466–77. <https://doi.org/10.1094/MPMI-11-16-0229-R>.
 138. Xie C, Zhang G, Zhang Y, Song X, Guo H, Chen X, Fang R. SRWD1, a novel target gene of DELLA and WRKY proteins, participates in the development and immune response of rice (*Oryza sativa* L.). *Sci Bull*. 2017;62:1639–48. <https://doi.org/10.1016/j.scib.2017.12.002>.
 139. Zhou X, Liao H, Chern M, Yin J, Chen Y, Wang J, et al. Loss of function of a rice TPR-domain RNA-binding protein confers broad-spectrum disease resistance. *Proc Natl Acad Sci USA*. 2018;115:3174–9. <https://doi.org/10.1073/pnas.1705927115>.
 140. United States Department of Agriculture (USDA). 2014. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/air_isu_ting_rice.pdf. Accessed 25 Aug 2018.
 141. Cai L, Cao Y, Xu Z, Ma W, Zakria M, Zou L, et al. A transcription activator-like effector Tal7 of *Xanthomonas oryzae* pv. *oryzicola* activates rice gene Os09g29100 to suppress rice immunity. *Sci Rep*. 2017;7:5089. <https://doi.org/10.1038/s41598-017-04800-8>.
 142. Macovei A, Sevilla NR, Cantos C, Jonson GB, Slamet-Loedin I, Čermák T, et al. Novel alleles of rice elf4G generated by CRISPR/Cas9-targeted mutagenesis confer resistance to Rice tungro spherical virus. *Plant Biotechnol J*. 2018;47:417. <https://doi.org/10.1111/pbi.12927>.
 143. Nekrasov V, Wang C, Win J, Lanz C, Weigel D, Kamoun S. Rapid generation of a transgene-free powdery mildew resistant tomato by genome deletion. *Sci Rep*. 2017;7:482. <https://doi.org/10.1038/s41598-017-00578-x>.
 144. Mahfouz M, Tashkandi M, Ali Z, Aljedaani F, Shami A. Engineering resistance against tomato yellow leaf curl virus via the CRISPR/Cas9 system in tomato. 2017. <https://doi.org/10.1101/237735>.
 145. Toledo Thomazella DP de, Brail Q, Dahlbeck D, Staskawicz BJ. CRISPR–Cas9 mediated mutagenesis of a DMR6 ortholog in tomato confers broad-spectrum disease resistance. 2016;1–23. <https://doi.org/10.1101/064824>.
 146. Zhang Y, Bai Y, Wu G, Zou S, Chen Y, Gao C, Tang D. Simultaneous modification of three homoeologs of TaEDR1 by genome editing enhances powdery mildew resistance in wheat. *Plant J*. 2017;91:714–24. <https://doi.org/10.1111/tpj.13599>.
 147. United States Department of Agriculture (USDA). 2015. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/15-238-01_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
 148. Ruiter R, van den Brande I, Stals E, Delaure S, Cornelissen M, D'Halluin K. Spontaneous mutation frequency in plants obscures the effect of chimeroplasty. *Plant Mol Biol*. 2003;53:675–89. <https://doi.org/10.1023/b:plan.0000019111.96107.01>.
 149. Hummel AW, Chauhan RD, Cermak T, Mutka AM, Vijayaraghavan A, Boyher A, et al. Allele exchange at the EPSPS locus confers glyphosate tolerance in cassava. *Plant Biotechnol J*. 2018;16:1275–82. <https://doi.org/10.1111/pbi.12868>.
 150. D'Halluin K, Vanderstraeten C, van Hulle J, Rosolowska J, van den Brande I, Pennewaert A, et al. Targeted molecular trait stacking in cotton through targeted double-strand break induction. *Plant Biotechnol J*. 2013;11:933–41. <https://doi.org/10.1111/pbi.12085>.
 151. Sauer NJ, Narváez-Vásquez J, Mozoruk J, Miller RB, Warburg ZJ, Woodward MJ, et al. Oligonucleotide-mediated genome editing provides precision and function to engineered nucleases and antibiotics in plants. *Plant Physiol*. 2016;170:1917–28. <https://doi.org/10.1104/pp.15.01696>.
 152. Ainley WM, Sastry-Dent L, Welter ME, Murray MG, Zeitzer B, Amora R, et al. Trait stacking via targeted genome editing. *Plant Biotechnol J*. 2013;11:1126–34. <https://doi.org/10.1111/pbi.12107>.
 153. Zhu T, Peterson DJ, Tagliani L, Clair G, Baszcynski CL, Bowen B. Targeted manipulation of maize genes *in vivo* using chimeric RNA/DNA oligonucleotides. *Proc Natl Acad Sci*. 1999;96:8768–73. <https://doi.org/10.1073/pnas.96.15.8768>.
 154. Zhu T, Mettenburg K, Peterson DJ, Tagliani L, Baszcynski CL. Engineering herbicide-resistant maize using chimeric RNA/DNA oligonucleotides. *Nat Biotechnol*. 2000;18:555–8. <https://doi.org/10.1038/75435>.
 155. Butler NM, Baltes NJ, Voytas DF, Douches DS. Geminivirus-mediated genome editing in potato (*Solanum tuberosum* L.) using sequence-specific nucleases. *Front Plant Sci*. 2016;7:1–13. <https://doi.org/10.3389/fpls.2016.01045>.
 156. Li J, Meng X, Zong Y, Chen K, Zhang H, Liu J, et al. Gene replacements and insertions in rice by intron targeting using CRISPR–Cas9. *Nat Plants*. 2016;2:1–6. <https://doi.org/10.1038/nplants.2016.139>.
 157. Sun Y, Zhang X, Wu C, He Y, Ma Y, Hou H, et al. Engineering herbicide-resistant rice plants through CRISPR/Cas9-mediated homologous

- recombination of acetolactate synthase. Mol Plant. 2016;9:628–31. <https://doi.org/10.1016/j.molp.2016.01.001>.
158. Wang M, Liu Y, Zhang C, Liu J, Liu X, Wang L, et al. Gene editing by co-transformation of TALEN and chimeric RNA/DNA oligonucleotides on the rice OsEPSPS gene and the inheritance of mutations. PLoS ONE. 2015;10:e0122755. <https://doi.org/10.1371/journal.pone.0122755>.
159. Okuzaki A, Toriyama K. Chimeric RNA/DNA oligonucleotide-directed gene targeting in rice. Plant Cell Rep. 2004;22:509–12. <https://doi.org/10.1007/s00299-003-0698-2>.
160. Shimatani Z, Kashojo S, Takayama M, Terada R, Arazoe T, Ishii H, et al. Targeted base editing in rice and tomato using a CRISPR–Cas9 cytidine deaminase fusion. Nat Biotechnol. 2017;35:441–3. <https://doi.org/10.1038/nbt.3838>.
161. Shimatani Z, Fujikura U, Ishii H, Matsui Y, Suzuki M, Ueke Y, et al. Inheritance of co-edited genes by CRISPR-based targeted nucleotide substitutions in rice. Plant Physiol Biochem. 2018. <https://doi.org/10.1016/j.plaphy.2018.04.028>.
162. Butt H, Eid A, Ali Z, Atia MAM, Mokhtar MM, Hassan N, et al. Efficient CRISPR/Cas9-mediated genome editing using a chimeric single-guide RNA molecule. Front Plant Sci. 2017;8:1441. <https://doi.org/10.3389/fpls.2017.01441>.
163. Li Z, Liu Z-B, Xing A, Moon BP, Koellhoffer JP, Huang L, et al. Cas9-guide RNA directed genome editing in soybean. Plant Physiol. 2015;169:960–70. <https://doi.org/10.1104/pp.15.00783>.
164. Chilcoat D, Liu Z-B, Sander J. Use of CRISPR/Cas9 for crop improvement in maize and soybean. Prog Mol Biol Transl Sci. 2017;149:27–46. <https://doi.org/10.1016/bs.pmbts.2017.04.005>.
165. United States Department of Agriculture (USDA). 2018. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/18-036-01_a1_air_inquiry_cbidel.pdf. Accessed 25 Aug 2018.
166. Zhou X, Jacobs TB, Xue L-J, Harding SA, Tsai C-J. Exploiting SNPs for biallelic CRISPR mutations in the outcrossing woody perennial *Populus* reveals 4-coumarate:CoA ligase specificity and redundancy. New Phytol. 2015;208:298–301. <https://doi.org/10.1111/nph.13470>.
167. Andersson M, Turesson H, Nicolia A, Fält A-S, Samuelsson M, Hofvander P. Efficient targeted multiallelic mutagenesis in tetraploid potato (*Solanum tuberosum*) by transient CRISPR–Cas9 expression in protoplasts. Plant Cell Rep. 2017;36:117–28. <https://doi.org/10.1007/s00299-016-0262-3>.
168. Jung JH, Altpeter F. TALEN mediated targeted mutagenesis of the caffeic acid O-methyltransferase in highly polyploid sugarcane improves cell wall composition for production of bioethanol. Plant Mol Biol. 2016;92:131–42. <https://doi.org/10.1007/s11103-016-0499-y>.
169. Kannan B, Jung JH, Moxley GW, Lee S-M, Altpeter F. TALEN-mediated targeted mutagenesis of more than 100 COMT copies/alleles in highly polyploid sugarcane improves saccharification efficiency without compromising biomass yield. Plant Biotechnol J. 2018;16:856–66. <https://doi.org/10.1111/pbi.12833>.
170. Park J-J, Yoo CG, Flanagan A, Pu Y, Debnath S, Ge Y, et al. Defined tetra-allelic gene disruption of the 4-coumarate: coenzyme A ligase 1 (Pv4CL1) gene by CRISPR/Cas9 in switchgrass results in lignin reduction and improved sugar release. Biotechnol Biofuels. 2017;10:284. <https://doi.org/10.1186/s13068-017-0972-0>.
171. United States Department of Agriculture (USDA). 2011. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/17-126-01_air_inquiry.pdf. Accessed 25 Aug 2018.
172. Njuguna E, Coussens G, Aesaert S, Neyt P, Anami S, van Lijsebettens M. Modulation of energy homeostasis in maize and *Arabidopsis* to develop lines tolerant to drought, genotoxic and oxidative stresses. Afrika Focus. 2018. <https://doi.org/10.21825/af.v30i2.8080>.
173. United States Department of Agriculture (USDA). 2017. https://www.aphis.usda.gov/biotechnology/downloads/reg_loi/17-219-01_air_inquiry.pdf. Accessed 25 Aug 2018.
174. Kim D, Alptekin B, Budak H. CRISPR/Cas9 genome editing in wheat. Funct Integr Genom. 2018;18:31–41. <https://doi.org/10.1007/s10142-017-0572-x>.
175. Meng X, Hu X, Liu Q, Song X, Gao C, Li J, Wang K. Robust genome editing of CRISPR–Cas9 at NAG PAMs in rice. Sci China Life Sci. 2018;61:122–5. <https://doi.org/10.1007/s11427-017-9247-9>.
176. Ali Z, Abul-faraj A, Piatek M, Mahfouz MM. Activity and specificity of TRV-mediated gene editing in plants. Plant Signal Behav. 2015;10:e1044191. <https://doi.org/10.1080/1592324.2015.1044191>.
177. Shen C, Que Z, Xia Y, Tang N, Li D, He R, Cao M. Knock out of the annexin gene OsAnn3 via CRISPR/Cas9-mediated genome editing decreased cold tolerance in rice. J Plant Biol. 2017;60:539–47. <https://doi.org/10.1007/s12374-016-0400-1>.
178. Tian S, Jiang L, Gao Q, Zhang J, Zong M, Zhang H, et al. Efficient CRISPR/Cas9-based gene knockout in watermelon. Plant Cell Rep. 2017;36:399–406. <https://doi.org/10.1007/s00299-016-2089-5>.
179. Zhang H, Zhang J, Wei P, Zhang B, Gou F, Feng Z, et al. The CRISPR/Cas9 system produces specific and homozygous targeted gene editing in rice in one generation. Plant Biotechnol J. 2014;12:797–807. <https://doi.org/10.1111/pbi.12200>.
180. Forner J, Pfeiffer A, Langenecker T, Manavella PA, Manavella P, Lohmann JU. Germline-transmitted genome editing in *Arabidopsis thaliana* using TAL-effector-nucleases. PLoS ONE. 2015;10:e0121056. <https://doi.org/10.1371/journal.pone.0121056>.
181. Ren B, Yan F, Kuang Y, Li N, Zhang D, Lin H, Zhou H. Specificity and inheritance of rBE3 and rBE4 endonuclease-induced gene modifications in rice. Chin J Biotechnol 2007;33(10):1776–85.
182. Khandagale K, Nadaf A. Genome editing for targeted improvement of plants. Plant Biotechnol Rep. 2016;10:327–43. <https://doi.org/10.1007/s11816-016-0417-4>.

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