

Gene Expression Platforms for Global Co-Expression Analyses A Comparison of spotted cDNA microarrays, Affymetrix microarrays, and SAGE

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cDNA Microarrays

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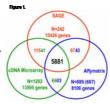
1. Abstract

We have conducted a comparison of three major expression platforms: cDNA microarray, oligonucleotide microarray and serial analysis of gene expression (SAGE) using large sets of available data for Homo angions. Several studies have compared two of the three platforms to evaluate the consistency of expression profiles for a single tissue or sample patterns. To this end we analyzed a recently published data set of 10 290 cDNA microarray (cDNA) experiments (Stuart et al. 2003), 242 SAGE libraries from the Gene Expression Dambus (GEO), and 607 AMPmetry (RIC-1133A) disponselotide microarray to the control of the contr



2. Gene Expression Data

Human gene expression data for three major expression platforms (see sidebar) were collected. We user sidebary were collected. We user (2022 cDNA) microarray experiments (Stuart et al., 2000), 242 SAGE libraries from collegen co





3. Methods

Gene Expression Analysis [sections 4-6]

Pearson correlations between genes were calculated using a modified version of the C clustering library (De Hoon $et\,al.,2004$). Correlations of correlations were calculated using the R statistical package (v. 1.8.1) and plotted with the R hexbin function.

Internal Consistency Markysis (section 4).
To evaluate the consistency of ocexpression observed with each
platform, we divide the experiments
available and determine co-expression
for each subset independently. The
results are then compared by
calculating a correlation of the gene
correlations. Division was performed
correlations. Division was performed
correlations. semi-randomly such that similar or replicate experiments remain together. If the platform consistently the control of the cost of the cost

Figure 2. Set of X express	ion experiments			
	+			
Divide experiments into two groups				
Randomize gene	expression values			
Set of X/2 expression	Set of X/2 expression			
experiments	experiments			
+	+			
Calculate all pairwise Pearson correlations	Calculate all pairwise Pearson correlations			
	n correlations for ene pair			
Graph and determine overall correlation				
,				

A correlation of gene correlations was determined as for the internal consistency analysis using the entire experiment set for each platform, and comparing correlations between platforms. If the two platforms being compared report the same distance between each gene pair, the overall correlation between platforms should be near 1.

Ranked Best Match Analysis (section 6)

Instead of considering the actual Pureson correlation between each gene pair and comparing between patterns, the correlation rank was considered. For example, it may be that for gene A, SAGE experiments identify its most similar gene (in terms of expression patterns to be gene B with a Person correlation of 0.9. The cBNA correlation

Gene Omiology Analysis (section 7)
The Gene Ontology (GO) MySQL database dump (release 200402 of associdi) was downloaded and a CO MySQL database was constructed. The most specific GO with the contraction of the contractio

4. Internal Consistency Analysis

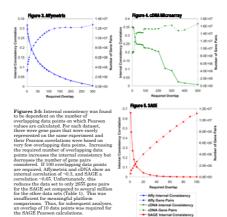
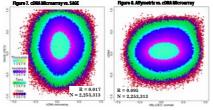


Table 1. Summary of Internal consistency results						
Platform	Division	Overlap	Gene pairs	r _c		
SAGE	Random	10	2168919	0.355		
		100	2655	0.770		
	By tissue	10	1734959	0.150		
	Cancer/Normal	10	1631419	0.204		
		80	1253	0.707		
Affymetrix	Random	100	4172389	0.942		
	By GSE series	100	3259181	0.255		
cDNA Microarray	Random	100	10426666	0.890		
	By author	100	10355435	0.287		

Table 1: Internal consistency was performed using a variety of different methods to determine the effect of (1) similar or replicate experiments; and (2) cancer samples in the dataset. A purely random in the dataset. A purely random division of samples was found to produce an unrealistically high internal consistency because of the presence of replicate experiments in each subset. Therefore, samples were divided semi-randomly keeping samples from the same tissue, experimental series, or publication author together. The presence of Cancer samples did not have a significant impact on oc. nave a significant impact on co-

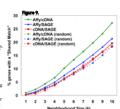
5. Platform Comparison Analysis

Figures 6-8: Poor levels of consistency were observed between platforms. Each point on the plots represents as hand gene pairs, and its coordinates represent the correlation of those plots represents as hand gene pairs, and its coordinates represent the correlation of the plots netrix vs. SAGE more likely explanation is that each platform identifies different co-expression patterns because the available data for each platform represents different tissue sources and experimental conditions. Yet another possibility is that few genes are actually consistently oc-expressed in biological syster N = 2,253,313



6. Ranked Best Match Analysis

Figure 9: The ranked best match analysis shows that different expression platforms do sometimes identify the same co-expressed genes. The Affymetrix versus eDNA 272% of genes having a co-expressed period of Pearson rank 10 or better confirmed by both platforms compared to 19.5% for random data. Affymetrix versus SAGE agreed for 21.4% of genes compared to 18.3% for random, and genes compared to 18.3% for random, and cDNA versus SAGE for 19.6% compared to cDNA versus SAGE for 19.6% compared to 18.6% for random. The high percentages of gene pairs in agreement for random data are the result of our minimum overlag criteria. Affect of the result of our minimum overlag criteria. Affect and cDNA) or 10 (for SAGE) overlapping data points. Some genes will have only a few gene pairs that meet this minimum. Thus, having a shared match within a rank of 10 for the two platforms will occur easily by chance. It is the difference from random, not the



7. Gene Ontology (GO) Analysis

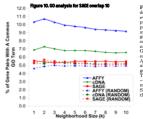


Figure 10: GO biological pr domain knowledge was used to evaluate gene co-expression predictions for each platform. The proportion of gene pairs annotated at a specific common GO term for given gene were enumerated and compared against the maximum compared against the maximum number of gene pairs that share GO terms for a given gene across each neighborhood distance. Affymetrix placed 9-11% (4-7% above random) of its co-expressed gene pairs at common GO terms. The cDNA microarray data placed that placed the common GO terms. The SAGE nerformance was no better maximum GO placements. The SAGE performance was no better than random data.

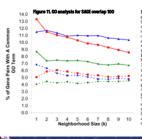


Figure 11: If 100 overlapping data points are required for SAGE (instead of 10) its performance against GO visibly improves. An overlap of 10 for SAGE was originally chosen so that a sufficient number of gene pairs to the Affv and cDNA datasets However, this may have unfairly biased the SAGE results. When 100 overlapping data points are required, the SAGE data actually required, the SAGE data actuall showed the highest internal consistency (Figure 5) and performed as well or better than Affy and cDNA in the GO placement analysis (Figure 11). Therefore, we expect the SAGE evaluation to improve as publicly available SAGE data increases to levels comparable to current and cDNA levels (~500-1000

8. Conclusions

- Co-expressed genes can be identified based on large-scale gene expression data.
 > Measures of internal consistency range from 0.255 to 0.942 for Affymetrix, 0.257 to 0.890 for dDNA microarrays and 0.155 to 0.770 for SAGE depending on bow the data is divided.
 > Direct comparison of correlation values between platforms yields poor correlations (RC0.1)
 Co-expression dendiried by larger set of overlapping data will be most reliable (with more than 0.250 platforms).
 > Comparison of gene rank shows significant overlap in oc expressed pairs identified by different platforms, particularly between Affymetrix and cDNA.
 2 Gene pairs identified as occupressed are more likely to share the same GO biological process.
 Affymetrix microarrays consistently identify the most oc-expressed genes that are confirmed by GO of SAGE experiments few gene pairs have sufficient overlap.

Acknowledgments

funding | Natural Sciences and Engineering Council of Canada (for OG and EP); Michael Smith Foundation for Health Research (for OG, SJ and EP); CIHR/MSFHR Bioinformatics Training Program (for DF); Killam Trusts (for EP); Genome BC

ces | 1. Stuart et al. 2003. Science. 302(5643):249-255; 2. De Hoon et al. 2004.
ormatics. Feb 10 [epub ahead of print]; 3. Shannon et al. (2003). Genome Res 13:2498-