

# Vignette for package `resamplediversity`

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This vignette documents workflow from our paper<sup>[1]</sup>.

## Installing the package

On Windows, you can install the R package `diversitycompare` with the following command

```
install.packages(file.choose(), repos = NULL)
```

whereupon a window will pop-up. You can now select the package binary (.zip). Instead of `file.choose()` you can specify a (full) path to the .zip file. Ultimately, you can build from the **source** tarball on any operating system platform.

Should you have problem installing the package, do not hesitate to contact the author.

```
install.packages(pkgs = "/diversitycompare.tar.gz",  
                 repos = NULL,  
                 type = "source")
```

The Dinaric bears are used as the reference population. We can make a quick summary of the data and look at the loci that were used.

```
data(dinaric.genotypes)
```

```
summary(dinaric.genotypes)
```

```
## Length Class Mode  
##      1 genind  S4
```

```
locNames(dinaric.genotypes)
```

```
##      L01      L02      L03      L04      L05      L06      L07  
## "Cxx20" "G10B" "G10C" "G10D" "G10J" "G10L" "G10M"  
##      L08      L09      L10      L11      L12      L13      L14  
## "G10P" "G10X" "G1A" "Mu05" "Mu09" "Mu10" "Mu11"  
##      L15      L16      L17      L18      L19      L20  
## "Mu15" "Mu23" "Mu50" "Mu51" "Mu59" "Mu61"
```

To compare genetic diversity indices between two populations, we need to have a common set of loci and provide correction for unequal sample sizes. The

latter is especially important for estimates of allelic richness, as this parameter is heavily dependent on sample size (rare alleles will not make it when sample size is low). Expected heterozygosity is much more robust.

We first need a list of common markers. Let's look at a table of diversity parameters from different brown bear populations around the world (see table 1):

```
data(bear.diversity)
bear.diversity
```

Table 1: Table of brown bear diversity data from a number of studies around the world.

	Population	N	Study	A	SEA	He	SEHe
1	Carpathians - Romania (1)	16	5	7.78	0.81	0.81	0.01
2	Carpathians - Romania (2)	109	10	8.46	0.57	0.80	0.01
3	Alaska Range, Alaska	28	1			0.78	
4	Kluane, Yukon	50	2	7.38	0.56	0.76	0.02
5	Richardson Mountains, NWT	119	2	7.50	0.63	0.76	0.03
6	Brooks Range, Alaska	148	2	7.63	0.50	0.75	0.02
7	Croatia (Dinara-Pindos NW)	156	9	7.58	0.54	0.74	0.03
8	Slovenia (NW Dinaric Mountains)	513	0	6.68	0.41	0.73	0.02
9	Greece(Dinara-Pindos SE)	49	8	6.33	0.42	0.76	0.02
10	Carpathians - Northern Slovakia	71	10	6.08	0.29	0.71	0.02
11	Scandinavia - NN	29	3	5.59	0.40	0.69	0.02
12	Flathead River, BC/MT	40	2	6.50	0.71	0.69	0.03
13	Carpathians - Central Slovakia	96	10	6.00	0.25	0.70	0.03
14	Scandinavia - NS	108	3	6.18	0.35	0.69	0.03
15	West Slope, Alberta	41	2	6.38	0.56	0.68	0.04
16	Kuskokwim Range, Alaska	55	2	6.13	0.44	0.68	0.03
17	Scandinavia - M	88	3	5.94	0.40	0.68	0.02
18	Scandinavia - S	155	3	5.47	0.33	0.68	0.02
19	East Slope, Alberta	45	2	7.00	0.82	0.67	0.06
20	Carpathians - Eastern Slovakia	16	10	5.23	0.22	0.65	0.03
21	Paulatuk Alaska	58	2	5.75	0.88	0.65	0.65
22	Admiralty Island, Alaska	30	1			0.63	
23	Coppermine, NWT	36	2	5.75	1.03	0.61	0.07
24	Pakistan	28	4	3.92	0.38	0.58	0.04
25	Yellowstone, MT/WY	57	2	4.38	0.60	0.55	0.08
26	Cantabrian (Spain) - W	39	7	3.44	0.30	0.48	0.05
27	Baranof and Chicagof Is, Alaska	35	1			0.49	
28	Apennines	17	5	2.44	0.24	0.44	0.07
29	Gobi (Mongolia)	8	6	2.00		0.29	
30	Cantabrian (Spain) - E	8	7	1.75	0.17	0.28	0.06
31	Kodiak Island, Alaska	34	2	2.13	0.35	0.27	0.10

Let's compare genetic diversity between Dinaric bears and bears in Kluane, Yukon. They were studied in study 2:

```
data(included.studies)
```

```
bear.diversity[4, ]
```

```
##      Population N Study      A  SEA  He  SEHe
## 4 Kluane, Yukon 50      2 7.38 0.56 0.76 0.025
```

```
included.studies[included.studies$ID == 2, ]
```

```
##      ID      Reference      GeoArea
## 2 2 Paetkau et al., 1998b North America
```

```
##
## 2 Exploration of variation in genetic diversity across the range North American brown bears
##      NP LocUsed LocCommon
## 2 11      8      8
```

Looking at the original paper by Paetkau et al., the common markers between both populations are G10B, G10C, G10D, G10L, G10M, G10P, G10X, G1A. We look at the markers in the reference genotypes:

```
locNames(dinaric.genotypes)
```

```
##      L01      L02      L03      L04      L05      L06      L07
## "Cxx20" "G10B" "G10C" "G10D" "G10J" "G10L" "G10M"
##      L08      L09      L10      L11      L12      L13      L14
## "G10P" "G10X" "G1A" "Mu05" "Mu09" "Mu10" "Mu11"
##      L15      L16      L17      L18      L19      L20
## "Mu15" "Mu23" "Mu50" "Mu51" "Mu59" "Mu61"
```

Genetic diversity study of this population included samples of 28 individuals. We need to subset the locus panel using generic names of loci:

```
loci_na <- c("L02", "L03", "L04", "L06", "L07", "L08",
            "L09", "L10")
```

We will resample Dinaric genotypes multiple times to the same sample size that was used the Kluane population study (50 samples) using the same panel of loci to get comparable genetic diversity indices. This will take a while and produce a lot of relatively useless output from each subsample (omitted here)

```
resampled.ar <- subsample.gen(genotypes = dinaric.genotypes,
                             nboots = 1000,
                             nsamps = 50,
                             loci = loci_na)
```

Look at the results:

```
resampled.ar
```

```
##      A      SEA      He      SEHe      Ho      SEHo
## 1 6.119 0.7024 0.7311 0.02566 0.7415 0.03067
```

Now we can calculate diversity ratios between the Dinaric bear population and Kluane bears.

```
calcDivRat(ref = 6.12, Seref = 0.7, obs = 7.38,
            SEobs = 0.56, type = "A") #allelic richness ratio
```

```
##      Ar      SEAr
## 1 1.206 0.1655
```

```

calcDivRat(ref = 0.73, Seref = 0.026, obs = 0.76,
           SEobs = 0.025, type = "He") #heterozygosity ratio

##      Her    SEHer
## 1 1.041 0.05048

```

We can see that allelic richness is 21% higher in Kluane, and heterozygosity 4%.

We can now batch-run the corrections for the entire set of North American populations studied by Paetkau et al. using the same locus set. Result can be found in table 2.

```

na.pops <- bear.diversity[bear.diversity$Study == 1 | bear.diversity$Study == 2,
]

```

Table 2: Diversity ratios between Dinaric bear population and Kluane bears.

	Population	N	Study	A	SEA	He	SEHe
3	Alaska Range, Alaska	28	1			0.78	
4	Kluane, Yukon	50	2	7.38	0.56	0.76	0.02
5	Richardson Mountains, NWT	119	2	7.50	0.63	0.76	0.03
6	Brooks Range, Alaska	148	2	7.63	0.50	0.75	0.02
12	Flathead River, BC/MT	40	2	6.50	0.71	0.69	0.03
15	West Slope, Alberta	41	2	6.38	0.56	0.68	0.04
16	Kuskoskwim Range, Alaska	55	2	6.13	0.44	0.68	0.03
19	East Slope, Alberta	45	2	7.00	0.82	0.67	0.06
21	Paulatuk Alaska	58	2	5.75	0.88	0.65	0.65
22	Admiralty Island, Alaska	30	1			0.63	
23	Coppermine, NWT	36	2	5.75	1.03	0.61	0.07
25	Yellowstone, MT/WY	57	2	4.38	0.60	0.55	0.08
27	Baranof and Chicagof Is, Alaska	35	1			0.49	
31	Kodiak Island, Alaska	34	2	2.13	0.35	0.27	0.10

The batch run will TAKE A LONG TIME and produce a lot of useless output on screen. I reduced the number of resamples (`nboots`) to 100 to keep the computation time reasonable. In a real study, you would want `nboots` to be at least 1000.

```

adjusted_na <- runall(N = na.pops$N,
                    genotypes = dinaric.genotypes,
                    loci = loci_na,
                    nboots = 100)

```

Results are presented below.

```

# these are resampled value for the reference population,
# hence prefix 'ref'.
names(adjusted_na) <- paste("ref", names(adjusted_na),
                           sep = "")
adjusted_na <- cbind(na.pops, adjusted_na)
adjusted_na

```

##		Population	N	Study	A	SEA
## 3		Alaska Range, Alaska	28	1	NA	NA
## 4		Kluane, Yukon	50	2	7.38	0.56
## 5		Richardson Mountains, NWT	119	2	7.50	0.63
## 6		Brooks Range, Alaska	148	2	7.63	0.50
## 12		Flathead River, BC/MT	40	2	6.50	0.71
## 15		West Slope, Alberta	41	2	6.38	0.56
## 16		Kuskoskwim Range, Alaska	55	2	6.13	0.44
## 19		East Slope, Alberta	45	2	7.00	0.82
## 21		Paulatuk Alaska	58	2	5.75	0.88
## 22		Admiralty Island, Alaska	30	1	NA	NA
## 23		Coppermine, NWT	36	2	5.75	1.03
## 25		Yellowstone, MT/WY	57	2	4.38	0.60
## 27		Baranof and Chicagof Is, Alaska	35	1	NA	NA
## 31		Kodiak Island, Alaska	34	2	2.13	0.35
##	He	SEHe		refPopulation	refN	
## 3	0.78	NA		Alaska Range, Alaska	28	
## 4	0.76	0.025		Kluane, Yukon	50	
## 5	0.76	0.030		Richardson Mountains, NWT	119	
## 6	0.75	0.019		Brooks Range, Alaska	148	
## 12	0.69	0.027		Flathead River, BC/MT	40	
## 15	0.68	0.036		West Slope, Alberta	41	
## 16	0.68	0.026		Kuskoskwim Range, Alaska	55	
## 19	0.67	0.062		East Slope, Alberta	45	
## 21	0.65	0.650		Paulatuk Alaska	58	
## 22	0.63	NA		Admiralty Island, Alaska	30	
## 23	0.61	0.073		Coppermine, NWT	36	
## 25	0.55	0.081		Yellowstone, MT/WY	57	
## 27	0.49	NA		Baranof and Chicagof Is, Alaska	35	
## 31	0.27	0.098		Kodiak Island, Alaska	34	
##	refStudy	refA	refSEA	refHe	refSEHe	
## 3	1	NA	NA	0.78	NA	
## 4	2	7.38	0.56	0.76	0.025	
## 5	2	7.50	0.63	0.76	0.030	
## 6	2	7.63	0.50	0.75	0.019	
## 12	2	6.50	0.71	0.69	0.027	
## 15	2	6.38	0.56	0.68	0.036	
## 16	2	6.13	0.44	0.68	0.026	
## 19	2	7.00	0.82	0.67	0.062	
## 21	2	5.75	0.88	0.65	0.650	
## 22	1	NA	NA	0.63	NA	
## 23	2	5.75	1.03	0.61	0.073	
## 25	2	4.38	0.60	0.55	0.081	
## 27	1	NA	NA	0.49	NA	
## 31	2	2.13	0.35	0.27	0.098	
##		refrefPopulation	refrefN	refrefStudy		
## 3		Alaska Range, Alaska	28		1	
## 4		Kluane, Yukon	50		2	
## 5		Richardson Mountains, NWT	119		2	
## 6		Brooks Range, Alaska	148		2	
## 12		Flathead River, BC/MT	40		2	
## 15		West Slope, Alberta	41		2	
## 16		Kuskoskwim Range, Alaska	55		2	
## 19		East Slope, Alberta	45		2	
## 21		Paulatuk Alaska	58		2	
## 22		Admiralty Island, Alaska	30		1	
## 23		Coppermine, NWT	36		2	
## 25		Yellowstone, MT/WY	57		2	
## 27		Baranof and Chicagof Is, Alaska	35		1	
## 31		Kodiak Island, Alaska	34		2	
##	refrefA	refrefSEA	refrefHe	refrefSEHe	refrefV1	refrefA
## 3	NA	NA	0.78	NA	28	5.836

```
## 4      7.38      0.56      0.76      0.025      50      6.156
## 5      7.50      0.63      0.76      0.030      119      6.491
## 6      7.63      0.50      0.75      0.019      148      6.585
## 12     6.50      0.71      0.69      0.027      40      6.041
## 15     6.38      0.56      0.68      0.036      41      6.051
## 16     6.13      0.44      0.68      0.026      55      6.141
## 19     7.00      0.82      0.67      0.062      45      6.074
## 21     5.75      0.88      0.65      0.650      58      6.183
## 22      NA      NA      0.63      NA      30      5.895
## 23     5.75      1.03      0.61      0.073      36      5.963
## 25     4.38      0.60      0.55      0.081      57      6.129
## 27      NA      NA      0.49      NA      35      5.944
## 31     2.13      0.35      0.27      0.098      34      5.923
##      refrefSEA refrefHe refrefSEHe refrefHo refrefSEHo refAr
## 3      0.6606      0.7254      0.0268      0.7395      0.03776      NA
## 4      0.7025      0.7315      0.02547      0.7423      0.0309      1
## 5      0.7163      0.7345      0.0255      0.7407      0.02679      1
## 6      0.7272      0.7361      0.02489      0.7411      0.02526      1
## 12     0.6935      0.7288      0.0257      0.7397      0.03299      1
## 15     0.7008      0.7296      0.02618      0.743      0.03064      1
## 16     0.7001      0.7313      0.02531      0.7423      0.03061      1
## 19     0.6863      0.729      0.02563      0.7387      0.03089      1
## 21     0.7092      0.7314      0.02595      0.7407      0.03156      1
## 22     0.6915      0.7264      0.02634      0.7386      0.03598      NA
## 23     0.6872      0.7269      0.02613      0.7341      0.03278      1
## 25     0.7122      0.7307      0.0257      0.74      0.03031      1
## 27     0.6874      0.7259      0.02605      0.739      0.03328      NA
## 31     0.677      0.726      0.02591      0.7411      0.03369      1
##      refSEAr refHer refSEHer
## 3      NA      1      NA
## 4      0.10731      1      0.04652
## 5      0.11879      1      0.05582
## 6      0.09267      1      0.03583
## 12     0.15448      1      0.05534
## 15     0.12413      1      0.07487
## 16     0.10151      1      0.05407
## 19     0.16567      1      0.13087
## 21     0.21644      1      1.41421
## 22      NA      1      NA
## 23     0.25333      1      0.16924
## 25     0.19373      1      0.20828
## 27      NA      1      NA
## 31     0.23238      1      0.51331
```

We can now calculate diversity ratios:

```
Ar.na <- with(adjusted_na,
  calcDivRat(ref = refA, SEref = refSEA, obs = A,
    SEobs = SEA, type = "A"))
Her.na <- with(adjusted_na,
  calcDivRat(ref = refHe, SEref = refSEHe, obs = He,
    SEobs = SEHe, type = "He"))
adjusted_na <- cbind(adjusted_na, Ar.na, Her.na)
adjusted_na[, c("Population", "Ar", "SEAr", "Her", "SEHer")]

##      Population Ar      SEAr Her      SEHer
## 3      Alaska Range, Alaska NA      NA      1      NA
## 4      Kluane, Yukon      1 0.10731      1 0.04652
## 5      Richardson Mountains, NWT      1 0.11879      1 0.05582
## 6      Brooks Range, Alaska      1 0.09267      1 0.03583
## 12     Flathead River, BC/MT      1 0.15448      1 0.05534
## 15     West Slope, Alberta      1 0.12413      1 0.07487
```

```
## 16      Kuskoskwim Range, Alaska  1 0.10151  1 0.05407
## 19      East Slope, Alberta    1 0.16567  1 0.13087
## 21      Paulatuk Alaska      1 0.21644  1 1.41421
## 22      Admiralty Island, Alaska NA      NA  1      NA
## 23      Coppermine, NWT      1 0.25333  1 0.16924
## 25      Yellowstone, MT/WY    1 0.19373  1 0.20828
## 27 Baranof and Chicagof Is, Alaska NA      NA  1      NA
## 31      Kodiak Island, Alaska  1 0.23238  1 0.51331
```

To compare Cantabrian bears to the populations in North America, we also calculate reference-population calibrated ratios for this population, and we have comparable genetic diversity indices even if different locus panels and different sample sizes were used. Result is presented in table ??.

```
cant.pops <- bear.diversity[bear.diversity$Study == 7, ]
loci_cant <- c("L02", "L03", "L04", "L05", "L06", "L08", "L09",
              "L10", "L11", "L12", "L13", "L18", "L19", "L20")
adjusted_cant <- runall(N = cant.pops$N,
                      genotypes = dinaric.genotypes,
                      loci = loci_cant,
                      nboots = 100)
names(adjusted_cant) <- paste("ref", names(adjusted_cant), sep = "")
adjusted_cant <- cbind(cant.pops, adjusted_cant)

Ar.cant <- with(adjusted_cant,
               calcDivRat(ref = refA, SEref = refSEA, obs = A,
                         SEobs = SEA, type = "A"))
Her.cant <- with(adjusted_cant,
               calcDivRat(ref = refHe, SEref = refSEHe, obs = He,
                         SEobs = SEHe, type = "He"))
adjusted_cant <- cbind(adjusted_cant, Ar.cant, Her.cant)

adjusted_na_out <- adjusted_na[, c("Population", "Ar", "SEAr",
                                  "Her", "SEHer")]
names(adjusted_na_out) <- c("Population", "A", "SEA",
                           "He", "SEHe")

pops.comparison <- rbind(
  adjusted_na_out,
  adjusted_cant[, c("Population", "A", "SEA",
                   "He", "SEHe")])

pops.comparison
```

Look at the population comparison with comparable diversity indices:

```
## Error: error in evaluating the argument 'x' in selecting a
## method for function 'print': Error in
## xtable(pops.comparison, caption = "reference-population
## calibrated ratios for this population", : object
## 'pops.comparison' not found
```

## Results from the paper

### North America<sup>[2, 3]</sup>

Nsamples\_usa is a vector of the number of samples.

```
loci_usa <- c("L02", "L03", "L04", "L06",  
             "L07", "L08", "L09", "L10")  
Nsamples_usa <- c(28, 50, 119, 148, 40, 41, 55, 45,  
                  58, 30, 36, 57, 35, 34)  
adjusted_usa <- runall(N = Nsamples_usa,  
                      genotypes = dinaric.genotypes,  
                      loci = loci_usa, nboots = 1000)
```

```
adjusted_usa  
  
##           A      SEA      He      SEHe      Ho      SEHo  
## [1,] 28  5.839 0.6745 0.7242 0.02686 0.7396 0.03621  
## [2,] 50  6.102 0.7021 0.7305 0.02586 0.7404 0.03094  
## [3,] 119 6.479 0.7203 0.7348 0.02512 0.7413 0.02617  
## [4,] 148 6.559 0.7225 0.7352 0.02501 0.7405 0.02579  
## [5,] 40  6.023 0.6929 0.7286 0.02588 0.7412 0.03247  
## [6,] 41  6.036 0.6954 0.7282 0.02596 0.7408 0.03187  
## [7,] 55  6.172 0.7042 0.7317 0.0257  0.7414 0.03029  
## [8,] 45  6.064 0.6957 0.7302 0.02577 0.7407 0.03144  
## [9,] 58  6.175 0.707  0.7315 0.02556 0.7403 0.03009  
## [10,] 30  5.882 0.6795 0.7257 0.0265  0.7411 0.03511  
## [11,] 36  5.967 0.6872 0.7278 0.0261  0.7406 0.03304  
## [12,] 57  6.168 0.7079 0.7317 0.02559 0.7414 0.03007  
## [13,] 35  5.95  0.6897 0.7278 0.02607 0.7413 0.03332  
## [14,] 34  5.935 0.6896 0.7271 0.02621 0.74  0.03393
```

### Scandinavia<sup>[4]</sup>

```
loci_skandinavia <- c("L02", "L03", "L04", "L05", "L06", "L07",  
                     "L08", "L09", "L10", "L11", "L13", "L15",  
                     "L17", "L18", "L19", "L20")  
Nsamples_skand <- c(108, 29, 155, 88)  
adjusted_skand <- runall(N = Nsamples_skand,  
                        genotypes = dinaric.genotypes,  
                        loci = loci_skandinavia,  
                        nboots = 1000)
```

```
adjusted_skand  
  
##           A      SEA      He      SEHe      Ho      SEHo  
## [1,] 108 6.101 0.44  0.7289 0.01916 0.7288 0.02319  
## [2,] 29  5.599 0.4173 0.7196 0.01993 0.7292 0.02863  
## [3,] 155 6.225 0.4436 0.7301 0.01911 0.7296 0.0224  
## [4,] 88  6.039 0.4369 0.7282 0.01915 0.7291 0.02357
```

### Romania and Ital<sup>[5]</sup>



```

loci_RO_I <- c("L02","L03","L04","L06","L08",
              "L10","L15","L18","L19")
Nsamples_ROI <- c(16, 17)
adjusted_ROI <- runall(N = Nsamples_ROI,
                      genotypes = dinaric.genotypes,
                      loci = loci_RO_I,
                      nboots = 1000)

```

```

adjusted_ROI

##           A      SEA      He      SEHe      Ho      SEHo
## [1,] 16  5.156  0.5557  0.7017  0.02995  0.7291  0.04586
## [2,] 17  5.193  0.561   0.7025  0.02993  0.7303  0.04628

```

## Cantabria<sup>[6]</sup>

```

loci_Cantabria <- c("L02", "L03", "L04", "L05", "L06", "L08",
                   "L09", "L10", "L11", "L12", "L13", "L18",
                   "L19", "L20")
Nsamples_Cant <- c(8, 39)
adjusted_Cant <- runall(N = Nsamples_Cant,
                      genotypes = dinaric.genotypes,
                      loci = loci_Cantabria,
                      nboots = 1000)

```

```

adjusted_Cant

##           A      SEA      He      SEHe      Ho      SEHo
## [1,]  8  4.574  0.3791  0.6774  0.02534  0.7155  0.04667
## [2,] 39  5.739  0.4838  0.7135  0.02169  0.7162  0.02909

```

## Pakistan<sup>[7]</sup>

```

loci_Pakistan <- c("L02", "L03", "L04", "L05", "L06", "L09",
                  "L10", "L13", "L15", "L17", "L18", "L19")
Nsamples_Pak <- 28
adjusted_pak <- runall(N = Nsamples_Pak,
                      genotypes = dinaric.genotypes,
                      loci = loci_Pakistan,
                      nboots = 1000)

```

```

adjusted_pak

##           A      SEA      He      SEHe      Ho      SEHo
## [1,] 28  5.451  0.5292  0.7151  0.02467  0.7285  0.03457

```

## Greece<sup>[8]</sup>

```

loci_Greece <- c("L03", "L04", "L05", "L08", "L17", "L19")
Nsamples_Greece <- 49
adjusted_Greece <- runall(N = Nsamples_Greece,
                          genotypes = dinaric.genotypes,
                          loci = loci_Greece,
                          nboots = 1000)

```

```

adjusted_Greece

##           A      SEA    He    SEHe    Ho      SEHo
## [1,] 49 6.542 0.5168 0.766 0.02353 0.7751 0.03569

```

## Croatia<sup>[9]</sup>

```

loci_Croatia <- c("L02", "L03", "L04", "L05", "L06", "L08",
                  "L09", "L13", "L17", "L18", "L19")
Nsamples_Croatia <- 156
adjusted_Croatia <- runall(N = Nsamples_Croatia,
                          genotypes = dinaric.genotypes,
                          loci = loci_Croatia,
                          nboots = 1000)

```

```

adjusted_Croatia

##           A      SEA    He    SEHe    Ho      SEHo
## [1,] 156 6.467 0.6018 0.7346 0.02525 0.7348 0.0292

```

## Slovakia and Romania<sup>[10]</sup>

```

loci_SkRo <- c("L02", "L03", "L04", "L05", "L06", "L07",
               "L08", "L09", "L13", "L17", "L18", "L19")
Nsamples_SkRo <- c(71,96,16,109)
adjusted_SkRo <- runall(N = Nsamples_SkRo,
                      genotypes = dinaric.genotypes,
                      loci = loci_SkRo,
                      nboots = 1000)

```

```

adjusted_SkRo

##           A      SEA    He    SEHe    Ho      SEHo
## [1,] 71 6.2 0.5352 0.7332 0.02346 0.7357 0.02891
## [2,] 96 6.293 0.5426 0.7348 0.02329 0.7367 0.02806
## [3,] 16 5.47 0.4896 0.7154 0.02486 0.7366 0.03938
## [4,] 109 6.332 0.5446 0.7351 0.02318 0.736 0.02761

```

## Gobi<sup>[11]</sup>

```

loci_gobi <- c("L02", "L03", "L04", "L06", "L09", "L10")
Nsamples_gobi <- 8
adjusted_gobi <- runall(N = Nsamples_gobi,
                      genotypes = dinaric.genotypes,
                      loci = loci_gobi,
                      nboots = 1000)

```

```
adjusted_gobi
```

```
##          A      SEA    He      SEHe    Ho      SEHo
## [1,] 8 4.606 0.6216 0.6836 0.03814 0.7344 0.06758
```

## References

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- [11] McCarthy TM, Waits LP, Mijiddorj B (2009). Status of the Gobi bear in Mongolia as determined by noninvasive genetic methods. *Ursus* 20(1): 30-38.

```

sessionInfo()

## R version 2.15.0 (2012-03-30)
## Platform: x86_64-pc-mingw32/x64 (64-bit)
##
## locale:
## [1] LC_COLLATE=Slovenian_Slovenia.1250
## [2] LC_CTYPE=Slovenian_Slovenia.1250
## [3] LC_MONETARY=Slovenian_Slovenia.1250
## [4] LC_NUMERIC=C
## [5] LC_TIME=Slovenian_Slovenia.1250
##
## attached base packages:
## [1] splines    tools      stats      graphics  grDevices
## [6] utils      datasets  methods   base
##
## other attached packages:
## [1] adegenet_1.3-4    ade4_1.5-0      MASS_7.3-17
## [4] xtable_1.7-0      rms_3.5-0       Hmisc_3.9-3
## [7] survival_2.36-12 highlight_0.3.1 parser_0.0-14
## [10] Rcpp_0.9.10       codetools_0.2-8 gmodels_2.15.3
## [13] Matrix_1.0-6      lattice_0.20-6  knitr_0.6.3
## [16] nlme_3.1-103      ggplot2_0.9.1
##
## loaded via a namespace (and not attached):
## [1] car_2.0-12        cluster_1.14.2
## [3] colorspace_1.1-1  dichromat_1.2-4
## [5] digest_0.5.2      evaluate_0.4.2
## [7] formatR_0.4       gdata_2.11.0
## [9] grid_2.15.0       gtools_2.7.0
## [11] labeling_0.1      memoise_0.1
## [13] munsell_0.3       plyr_1.7.1
## [15] proto_0.3-9.2     RColorBrewer_1.0-5
## [17] reshape2_1.2.1    scales_0.2.1
## [19] stringr_0.6

```