

# Dewlap color variation in *Anolis sagrei* is maintained among habitats within islands of the West Indies

## Abstract

Animal signals evolve in an ecological context. Locally adapting animal sexual signals can be especially important for initiating or reinforcing reproductive isolation during the early stages of speciation. Previous studies have demonstrated that dewlap color in *Anolis* lizards can be highly variable between populations in relation to both biotic and abiotic adaptive drivers at relatively large geographical scales. Here, we investigated differentiation of dewlap coloration among habitat types at a small spatial scale, within multiple islands of the West Indies, to test the hypothesis that similar local adaptive processes occur over smaller spatial scales. We explored variation in dewlap coloration in the most widespread species of anole, *Anolis sagrei*, across three characteristic habitats spanning the Bahamas and the Cayman Islands, namely beach scrub, primary coppice forest and mangrove forest. Using reflectance spectrometry paired with supervised machine learning, we found significant differences in spectral properties of the dewlap between habitats within small islands, sometimes over very short distances. Passive divergence in dewlap phenotype associated with isolation-by-distance did not seem to explain our results. On the other hand, these habitat-specific dewlap differences varied in magnitude and direction across islands, and thus our primary test for adaptation – parallel responses across islands – was not supported. We suggest that neutral processes or selection could be involved in several ways, including sexual selection. Our results shed new light on the scale at which signal color polymorphism can be maintained in the presence of gene flow, and the relative role of local adaptation and other processes in driving these patterns.

**Keywords** — reflectance, adaptation, sexual signal, machine learning, polymorphism

## Introduction

The staggering diversity of animal communication signals has long been of interest to evolutionary biologists. Animals use chemical, mechanical, electromagnetic, and visual signals to communicate in a wide variety of contexts, including, competition for mates, species recognition, aposematism, and cooperation (Bradbury and Vehrenamp, 2011). A primary evolutionary factor shaping communication signals is the sensory system and behavior of recipients (the sensory drive hypothesis; Endler and McLellan 1988; Endler 1992, 1998). Over the past decades, scientists have established that signals evolve in an ecological context and are dependent on environmental conditions (Endler, 1992, 1993a,b). Just as different habitats may favor different combinations of ecomorphological traits to maximize performance and fitness (Arnold, 1983), they may also shape different forms of a signal, so as to maximize its transmission and detection (e.g. Seehausen 1997), or reduce its detection by unintended recipients such as predators (Endler, 1984, 1990, 1991; Halfwerk et al., 2014). This selective pressure may drive the local adaptation of communication signals.

One potential barrier to the maintenance of localized signal divergence is the homogenizing effect of gene flow. Population genetics theory suggests that gene flow may counteract local adaptation between localities and prevent divergence altogether, especially at small spatial scales, because of the inflow of maladapted alleles or because of the breaking of linkage between coevolving loci (Felsenstein, 1976; García-Ramos and Kirkpatrick, 1997; Dieckmann and Doebeli, 1999; Lenormand, 2002; Hendry et al., 2007a). This genetic homogenization has been confirmed empirically in systems such as stick insects (Nosil and Crespi, 2004) and stickleback (Hendry et al., 2007b). Yet, examples of microgeographic adaptation, i.e. adaptation at smaller scales than the range of dispersal, exist, highlighting the potential of some organisms to respond to selection in the face of

gene flow (see Richardson et al. 2014 and references therein). Examples include small scale adaptation in fragmented areas in Australian fruit flies (Willi and Hoffmann, 2012), and local adaptation to predation pressure in North American salamanders (Richardson and Urban, 2013). Therefore, despite evidence that local adaptation may be particularly difficult at small spatial scales where gene flow tends to cause adjoining populations to remain genetically homogeneous, the potential adaptive response of species traits, in particular communication signals, to localized differences in habitats remains relatively unknown (Richardson et al., 2014). Lizards of the neotropical genus *Anolis* are an excellent group for studying the eco-evolutionary dynamics of local adaptation and natural selection (Losos, 2009). A particularly conspicuous trait of anoles is their dewlap, an extensible flap of skin that is typically sexually dimorphic and used as a communication signal in courtship (Sigmund, 1983; Driessens et al., 2014, 2015) and territorial displays (Losos, 1985; Macedonia and Stamps, 1994; Macedonia et al., 2013) as well as in predator deterrence (Leal and Rodríguez-Robles, 1995, 1997; Leal and Rodriguez-Robles, 1997). Dewlap characteristics vary widely among the approximately 400 species of the genus (Nicholson et al., 2007). Interspecific variation in dewlap coloration is implicated in species recognition (Rand and Williams, 1970; Williams, 1969; Williams and Rand, 1977; Losos, 1985; Macedonia and Stamps, 1994; Fleishman, 2000; Macedonia et al., 2013), and this function could have had a role in initiating or reinforcing reproductive isolation during speciation (Lambert et al., 2013; Geneva et al., 2015; Ng et al., 2017).

Within species, studies have shown a link between variation in dewlap coloration and differences in habitats or climatic conditions (Macedonia, 2001; Leal and Fleishman, 2002; Thorpe and Stenson, 2002; Thorpe, 2002; Leal and Fleishman, 2004; Vanhooydonck et al., 2009; Ng et al., 2012, 2013, 2016; Vanhooydonck et al., 2009; Driessens et al., 2017). Some studies suggest that those differences may be adaptive and that dewlaps may have evolved to maximize detectability given local light conditions (Fleishman and Persons, 2001; Leal and Fleishman, 2002, 2004). Although this claim is further supported by recent findings that dewlap colors are perceived differently under different levels of shading (Fleishman et al., 2020), other studies found conflicting patterns of between-habitat variation that did not support the sensory drive hypothesis (Fleishman et al., 2009; Ng et al., 2012; Macedonia et al., 2014).

Previous studies investigating variation in anole dewlaps compared populations at relatively large geographical scales, e.g. between islands (Vanhooydonck et al., 2009; Driessens et al., 2017) or within large islands such as Puerto Rico (Leal and Fleishman, 2004) or Hispaniola (Ng et al., 2012, 2016). These large scales and marine barriers should reduce gene flow (Ng and Glor, 2011; Lambert et al., 2013; Richardson et al., 2014; Ng et al., 2017). That said, examples do exist of divergence in dewlap coloration at smaller scales or between populations with high degrees of gene flow (Thorpe and Stenson, 2002; Thorpe, 2002; Stapley et al., 2011; Ng et al., 2016).

*Anolis sagrei* is widespread across islands of the West Indies (Reynolds et al., 2020). It has been the subject of numerous studies concerning local adaptation (Losos et al., 1994, 1997, 2001; Kolbe et al., 2012), biological invasion (Kolbe et al., 2008), and sexual selection (Tokarz, 2002; Tokarz et al., 2005; Tokarz, 2006; Driessens et al., 2014; Steffen and Guyer, 2014; Driessens et al., 2015) among many other topics. Between-island variation in the mainly orange-red color of its dewlap was shown to be better explained by climatic variables such as annual precipitation and solar radiation (proposed to affect the average vegetation type on each island and among other things, its ambient light environment, Driessens et al. 2017), than by proxies for biotic factors such as sexual selection or predation pressure (Vanhooydonck et al., 2009; Baeckens et al., 2018). How intra-island differences in habitat may contribute to the diversity of dewlap coloration, however, remains unexplored, and may reveal new insights into the scale of local differentiation despite gene flow.

Here, we analyzed the color characteristics of *A. sagrei* dewlaps within nine islands in the Bahamas and Cayman Islands. These island systems presently, if not historically, comprise relatively small islands, with no major geographic barriers within islands limiting dispersal for this species (Kamath and Losos, 2018). These islands all share three characteristic native West Indian small-island habitat-types – beach scrub bush, closed-canopy primary coppice forest, and mangrove forest – that are often spatially intermingled. These habitats contrast in environmental parameters including vegetation community, light irradiance, humidity, and temperature (Howard, 1950;

Schoener, 1968). The Cayman Islands and the Bahamas have been colonized independently by *A. sagrei* from Cuba (van de Schoot 2016 unpublished thesis; Reynolds et al. 2020), such that these archipelagos constitute an ideal suite of natural replicates to explore within-island dewlap diversity across multiple islands. 107  
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Our sampling design included sites in close proximity; the median distance between two sites within an island was 11.2km. Combining reflectance spectrometry and supervised machine learning, we tested for divergence in dewlap phenotype between habitats within islands and between islands across part of the range of *A. sagrei*. We predicted that if light conditions in the environment indeed drive color evolution, dewlaps should be most similar between beach scrub and mangrove forest, which both have high levels of light irradiance, compared to the darker, closed-canopy coppice forest. If detectability is maximized given the local conditions, we expected darker and more contrasting dewlaps in high irradiance habitats. Finally, if habitat characteristics are strong determinants of dewlap color variation, similar patterns should be observed across multiple islands (Harvey and Pagel, 1991; Losos, 2011). 112  
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122 **Methods**

123 **Data collection**

124 We sampled 466 male *A. sagrei* from seven islands in the Bahamas Archipelago – Abaco, North  
125 Andros, South Andros, South Bimini, Eleuthera, Long Island, and Ragged Island – and two in the  
126 Cayman Islands – Cayman Brac and Little Cayman (Figure S1A). These islands were chosen to  
127 span the breadth of the West Indian range of *A. sagrei*, because they have highly similar habitat  
128 types, and because the *A. sagrei* on each island group are derived from ancient and distinct  
129 colonization events from Cuba (i.e. relatively evolutionarily independent, Reynolds et al. 2020).  
130 Three habitats were sampled on each island based on characterizations by Howard (1950) and  
131 Schoener (1968). Each habitat is clearly distinguishable by its dominant vegetation type — xeric  
132 beach scrub (open, relatively dry habitat consisting of low vegetation or isolated trees), primary  
133 coppice forest (closed-canopy forest) and mangrove forest (wet coastal habitat with trees growing  
134 in brackish water and high light penetration, although lizards were sampled in dry soil areas).  
135 Sample sizes are given in Table S1. Our sampling design enabled us to test for differences between  
136 habitats at a coarse and fine geographical scale. The median distance between two localities within  
137 an island was ~ 11km (Figure S1B), and 80.3% of all pairwise distances within islands were less  
138 than 50km. Additionally, there are no major barriers to dispersal (such as mountains or grassland)  
139 on any of the islands that we sampled.

140 **Reflectance measurements**

141 We measured reflectance between 300nm and 700nm wavelength, a range from ultraviolet to red  
142 that encompasses the colors visible to most lizards and vertebrates in general (Lazareva et al.,  
143 2012). Measurements were taken with an Ocean Optics USB4000 spectrometer, a pulsed Xenon  
144 light source (PX-2, Ocean Optics, Largo, FL, USA) and a reflectance probe protected by a black  
145 anodized aluminum sheath. Measurements were taken with a 45-degree inclination to prevent  
146 specular reflection (Endler, 1990). The device was regularly standardized with a Spectralon white  
147 standard (Labsphere, North Sutton, NH, USA). Reflectance was measured at the center of the  
148 dewlap. Reflectance curves were smoothed using the R package pavo (Maia et al., 2013) as well  
149 as with custom R functions, down to one reflectance value at each nanometer in wavelength from  
150 300 to 700nm.

151 **Analysis**

152 We tested for detectable differences in dewlap coloration between populations from different habi-  
153 tats across islands by following an analytic pipeline in several steps. First, we used multivariate  
154 analyses of variance to assess the relative contributions of islands, habitats and habitat-by-island  
155 interactions on the partitioning of variation in color space. Second, and provided that habitat-by-  
156 island interactions were found, we investigated habitat-differences in dewlap color for each island  
157 separately using machine learning classification. Third, for each island where multivariate differ-  
158 ences were detected using our machine learning pipeline, we used univariate analyses of variance  
159 to decompose the signal among the different dimensions of color space. Fourth, for each signifi-  
160 cant between-habitat variation found in univariate analyses, we used post-hoc tests to determine  
161 which habitats were responsible for the differences. Last, to get insights into the spatial scale of  
162 phenotypic variation, for each significant contrast between two habitats detected along a given  
163 dimension on a given island, we performed multiple pairwise Wilcoxon tests to assess differences  
164 in dewlap coloration among the sites involved in that significant contrast, and recorded the geo-  
165 graphical distance between sites that were found significant. In parallel, we tested a possible effect  
166 of isolation-by-distance, an alternative cause of phenotypic divergence between localities, based on  
167 diffusion approximation and dispersal distance, irrespective of habitat types. We did so using a  
168 permutation test to assess the significance of the correlation between geographical distances and  
169 phenotypic distances among sites within each island.

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171 All analyses in this study were performed in R 3.6.1 (R Core Team, 2019).

## Dimensionality reduction

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Because neighboring wavelengths are highly collinear and redundant in reflectance, we reduced the dimensionality of the data using principal component analysis (PCA), as per Cuthill et al. (1999) and Leal and Fleishman (2002). We performed PCA on data from all islands combined, as well as on each island separately and systematically retained the first four principal components (PC), which together always explained more than 88.8% of the variance across islands (Table S2). PCs need not represent the same wavelengths across islands because they are fitted on different datasets. Nevertheless, PC1 was highly collinear with brightness for all islands (Figure S11), while the other PCs captured the chromatic variation (i.e. irrespective of brightness) in dewlap color.

## Among-island variance partitioning

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We performed a two-way nonparametric multivariate analysis of variance (PERMANOVA, Anderson 2001, R package vegan, Oksanen et al. 2019) to identify differences in coloration between islands, habitats and habitats within islands, using principal components fitted on data from all islands together. We used a nonparametric test because although no multivariate outliers were detected based on the Mahalanobis distance, the assumption of multivariate normality was violated in several habitats on several islands (Henze-Zirkler's test, Henze and Zirkler 1990, R package MVN, Korkmaz et al. 2014,  $P < 0.05$ , Table S3).

## Within-island machine learning

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We performed a machine learning classification analysis on the first four principal components within each island separately, using random forests (Breiman, 2001). Random forests are a versatile, intuitive, and powerful algorithm commonly used in machine learning, using decision trees to predict the labels of particular observations based on their multivariate coordinates. These coordinates, or variables, are passed through a series of successive decision nodes, each examining a given variable of any given observation (James et al., 2013). The prediction for each observation is an aggregate over a large number of decision trees, each tree being trained on a subset of observations sampled with replacement from the dataset, and each tree being allowed to examine only a subset of the variables. This allows the random forest to overcome the individual errors of all trees in the predictions it makes.

To detect differences in dewlap coloration between habitats, we measured the success of random forests in reassigning individual lizards to their correct habitat of origin, based solely on their principal component scores. In machine learning, this so-called cross-validation procedure is typically done in two steps (James et al., 2013). First, a random forest is trained in recognizing features of dewlap coloration most associated with the different habitats, by being presented with multiple observations, making predictions about them, and updating its own decision rules based on whether the prediction deviates from the truth. Then, once trained, the patterns that the random forest has learned to recognize are tested by presenting new, previously unseen observations to the random forest, and measuring the proportion of correct predictions. This proportion, or success score, can then be statistically assessed against random guessing using a binomial test.

The cross-validation procedure requires that the data be split into a training set and a testing set. To remove any bias due to the set that is being sampled for training, it is common practice to use k-fold cross-validation (James et al., 2013), where the data are split into  $k$  random bins and  $k$  independent machines are trained, each taking one of the bins as a testing set and the rest for training, and where classification success is measured by summing all correct classifications from the  $k$  machines.

Here, we used a k-fold cross-validation procedure with  $k = 5$ , where each training set consisted of 80% of the data and the machine was tested on the remaining 20%. Each training set was conditioned on containing at least five lizards from each of the three habitats. We also down-sampled the training set to the sample size of the least represented habitat, to ensure that the different habitats were equally represented. To further remove any bias due to the specific random split into the different bins, we replicated each k-fold cross-validation five times. We then averaged the five resulting confusion matrices across replicates, where each confusion matrix shows the number

of lizards from each habitat reassigned into each habitat. For each island, we then used the average proportion of correctly reassigned lizards (i.e. the proportion of observations on the diagonal of the average confusion matrix) as an estimate of classification success. This score was tested against random guessing by comparing it to a binomial distribution with number of trials being the number of lizards on that island and success probability 1/3, representing the rate of successful classification by chance when three habitats are involved.

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We used the machine learning fitting functions in the R package rminer (Cortez, 2020), which calls random forest routines from the randomForest package (Liaw and Wiener 2002, implementation from the original random forest algorithm Breiman, 2001). For each random forest, we optimized the number of trees in the forest and the number of variables examined by each tree using the grid hyperparameter search procedure implemented in rminer, to choose between two numbers of trees (500 or 1,000) and four numbers of principal components examined per tree (1 to 4), using rminer's ordered holdout validation method with 2/3 of the data used for training.

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We validated the results of our analysis by using two other widely used machine learning classification methods: linear discriminant analysis and support vector machines (Cristianini and Shawe-Taylor, 2000; James et al., 2013), both accessible in rminer (Cortez, 2020).

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To know which wavelengths were most used to assign data points to each habitat, we trained another set of random forests, this time directly on reflectance data (taken every 5nm from 300 to 700nm) instead of principal components. We recorded the relative importance of each wavelength for each habitat, as measured by the mean decrease in accuracy during wavelength permutation, implemented in the randomForest package (Liaw and Wiener, 2002).

## 250 Univariate analyses

For each island where significant differences in dewlap coloration were detected between habitats, we used multiple univariate analyses of variance (ANOVA) to identify possible principal components underlying the observed differences. We constructed our ANOVA models in two steps, as per Zuur (2009). In a first step, we accounted for heterogeneity of variances across groups by systematically comparing the goodness-of-fit of an ANOVA model estimated with ordinary least squares (OLS) with that of a model estimated with generalized least squares (GLS), which allowed one estimate of residual variance per habitat (using the R package nlme, Pinheiro and Bates 2000; Pinheiro et al. 2020). Both models were fitted with restricted maximum likelihood (REML). Goodness-of-fit was estimated using Akaike's Information Criterion corrected for small sample sizes (AICc, R package MuMIn, Bartoń 2019), and the estimation method yielding the lowest AICc was retained. In a second step, we re-fitted the retained model with maximum likelihood (ML) to test for the effect of habitat type using likelihood ratio tests (LRT) between a model including a habitat-term and a null model lacking the habitat-term.

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We evaluated the normality of the standardized residuals (residuals divided by their standard error, which can differ among habitats in a GLS model) of each fitted ANOVA model using Shapiro-Wilk's test, with P-values adjusted for multiple testing using the Benjamini-Hochberg correction (Benjamini and Hochberg, 1995). In cases where significant deviations from normality were detected ( $P_{adj} < 0.05$ , Table S4) we performed Kruskal-Wallis's nonparametric test to back up the ANOVA results.

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To know which habitat-populations were different from which in dewlap coloration, we performed different post-hoc multiple comparison tests (all implemented in the PMCMRplus package, Pohlert, 2020), depending on which assumptions were met. In cases where normality and homoscedasticity were met (i.e. OLS-ANOVA was the best fit), we used Tukey's honest significant difference test. When normality was met but not homoscedasticity (i.e. GLS-ANOVA was the best fit), we used Dunnett's T3 test. Finally, whenever we used Kruskal-Wallis's test because the ANOVA residuals were not normally distributed, we used Nemenyi's test for post-hoc comparisons.

## Spatial autocorrelation

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We tested for within-island spatial autocorrelation between the geographical distances among sampling sites and their Euclidean distances in multivariate color space (mean PC1 to PC4 per site, Table S5), regardless of habitat type. For this, we performed Mantel's test (Legendre and Legendre 2012, R package vegan; Oksanen et al. 2019) on each island, using 999 permutations and geographical distances computed as geodesic distances from latitude and longitude data (R package geosphere, Hijmans 2019).

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## Site differences

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In this study, we were interested in the minimum spatial scale at which significant differences between habitats could be detected within islands. We performed multiple pairwise nonparametric Wilcoxon-Mann-Whitney tests (Hollander et al., 2013) to compare dewlap coloration between sites with different habitat types, for each pair of habitats and each variable where significant differences were detected with our analyses of variance. The P-values were adjusted using a Benjamini-Hochberg correction for multiple testing (Benjamini and Hochberg, 1995).

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294 **Results**

295 We tested for variation in *A. sagrei* dewlap coloration between populations living in three charac-  
296 teristic habitat types across nine islands that span the West Indian range of the brown anole (beach  
297 scrub, primary coppice and mangroves). We found that most of the variation in coloration was  
298 partitioned between islands (two-way PERMANOVA,  $F(df = 8) = 43.7$ ,  $P = 0.0001$ , explained  
299 variance  $R^2 = 40.9\%$ ). Nonetheless, we did find evidence for differences in dewlap coloration  
300 between habitat types, and those were mostly island-specific (habitat-by-island interaction term,  
301  $F(16) = 3.53$ ,  $P = 0.0001$ ,  $R^2 = 6.6\%$ ), with a significant portion of the variation explained by  
302 an habitat effect across all islands, but this effect was relatively small ( $F(2) = 4.7$ ,  $P = 0.0001$ ,  
303  $R^2 = 1.1\%$ ).

304 We subsequently tested for differences in dewlap coloration between habitat-populations within  
305 each island, using within-island principal component scores (to maximize the variation captured  
306 for each island, see Methods). Our within-island random forest classification analyses revealed  
307 detectable differences in dewlap coloration on eight out of the nine islands in our sample: Abaco,  
308 Bimini, Cayman Brac, Eleuthera, Little Cayman, Long Island, North Andros and South Andros.  
309 The accuracy of random forest classification exceeded random expectation more often than ex-  
310 pected by chance for all these islands (Table 1). Accuracy was as high as 73% for Cayman Brac.  
311 We obtained similar results using other machine learning approaches such as support vector ma-  
312 chines (Table S7) and linear discriminant analysis (Table S8), except that these methods did not  
313 detect significant differences on Eleuthera and North Andros. We describe in details the specific  
314 differences detected on each island in the Appendix, and focus here on the general patterns emerg-  
315 ing from our data.

316 Overall, we found significant differences in dewlap coloration between populations that were  
317 often in close geographical proximity. On Bimini, notably, we found a significant difference be-  
318 tween dewlaps from beach scrub and primary coppice forest, at a distance of a few hundred meters,  
319 making this contrast the smallest geographical scale at which differences in coloration were found  
320 in our study (Fig. S3). We also detected significant differences in dewlap coloration at distances  
321 below one kilometer on Abaco (Fig. S2G), and at distances between one and ten kilometers on  
322 Bimini (Fig. S3G), Cayman Brac (Fig. S4G), Little Cayman (Fig. S6G), Long Island (Fig. S7G)  
323 and South Andros (Fig. S10G).

324 We found evidence of spatial autocorrelation in dewlap coloration between the sites within is-  
325 lands for Abaco (Table 2), suggesting that populations from closer sites tend to have more similar  
326 dewlaps on this island than expected by chance. Abaco was the island we sampled at the largest  
327 scale, with some sites nearly a hundred kilometers away from each other (Fig. 2A). That said, some  
328 sites were also in close proximity, and significant differences in coloration were detected between  
329 habitats sometimes less than a kilometer away (Fig. S2G), suggesting that differences in dewlap  
330 coloration between distant sites may be partly attributable to isolation-by-distance, but this may  
331 not necessarily be the case for sites in close proximity. We did not find evidence for spatial auto-  
332 correlation on other islands than Abaco (although Eleuthera was nearly significant, Table 2).

333 A striking feature of our data was inconsistencies in between-habitat differences among islands,  
334 in terms of which habitats differ from which, which dimensions of coloration were involved, and  
335 in which direction. For example, while on Cayman Brac the random forests could well distinguish  
336 between all three habitats (Fig. S4D), on Abaco dewlaps from beach scrub and primary coppice  
337 were often mistaken, and on Bimini beach scrub dewlaps were more often classified into primary  
338 coppice or mangrove than into beach scrub (Fig. S3D). In terms of variable importance, for multi-  
339 ple islands the random forests used information in the UV range to discriminate between at least  
340 some habitats, particularly on Abaco (Fig. S2F), Bimini (Fig. S3F), Cayman Brac (Fig. S4F),  
341 Little Cayman (Fig. S6F) and Long Island (Fig. S7F), but differences in UV reflectance involved  
342 different habitats and were in different directions among these islands.

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## Discussion

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Two main insights follow from our results. First, we detected significant differences in dewlap coloration between habitats within seven out of the nine islands investigated (excluding North Andros where the follow-up univariate analyses were not significant), suggesting a putatively high potential for local differentiation of dewlap coloration in *Anolis sagrei*. Second, we found differences in coloration along different dimensions of color space, and in different directions, on different islands.

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Detectable differences in dewlap color between populations are surprising, as habitats were often in close geographical proximity to each other (as close as a few hundred meters on Bimini and most of the time within ten kilometers). Indeed, given that (1) the populations were continuously distributed between the habitats, (2) populations from different habitats were not monophyletic with respect to mitochondrial haplotypes (van de Schoot 2016 unpublished thesis), and (3) *A. sagrei* is highly mobile (Kamath and Losos, 2018), we would have expected more homogeneous distributions of color phenotypes within islands due to gene flow, with fewer differences between populations, especially those in close proximity.

Several scenarios could account for these findings. One explanation is an adaptive one: populations living in different habitats could be phenotypically adapted to their local environmental conditions. Given that the brightly colored dewlap of *A. sagrei* is used as a communication signal, its color may be a target for selection if the transmission or perception of the signal differs from one habitat to another, for example because of differences in ambient light, according to the sensory-drive hypothesis (Endler and McLellan, 1988; Endler, 1992, 1998). The sensory-drive hypothesis has been tested multiple times for dewlap coloration in *Anolis* lizards, with mixed results. Some authors found support for it (Leal and Fleishman, 2002, 2004), while others found differences in dewlap coloration between habitats inconsistent with the sensory-drive hypothesis (Fleishman et al., 2009; Ng et al., 2012).

If our results were an example of sensory drive, we would have expected to see consistent differences between populations from different habitats across islands, given the apparent environmental consistency each of the three habitat types across the islands we sampled. In particular, we would have expected divergence in line with increased detectability given local light conditions, such as the high contrasts with background vegetation found in the UV range in Leal and Fleishman (2002) and Leal and Fleishman (2004). We might also have expected mangrove and beach scrub lizards, both inhabiting areas with high light penetration, to have more similar dewlaps, and to differ significantly from lizards from the coppice habitat, where irradiance is low. Instead, we found inconsistencies in the way dewlap color differed between habitats across islands. While short-wavelengths (UV reflectance) were often involved in color differences, they were not involved on all islands where significant differences were detected. On some islands, other or additional variables differed, such as brightness, red reflectance or the reflectance at the ends of the spectrum visible to *Anolis* lizards (UV and red, Lazareva et al. 2012) relative to intermediate wavelengths (blue-to-yellow). Similar portions of the spectrum were sometimes involved in opposite directions on different islands, such as on Abaco and Cayman Brac, where mangrove lizards had a higher UV-reflectance than beach scrub lizards on the former, but a lower UV-reflectance on the latter. Overall, the observed heterogeneity of divergence patterns across islands provides no support to a sensory-drive explanation.

It is presently not known if the reported differences in coloration have a genetic basis. Yet, we find it unlikely that these differences arose through phenotypic plasticity, as although the carotenoids that partly make up the red and orange colors of anole dewlaps must be found in the diet (Goodwin, 1984; Hill et al., 2002; Hill and McGraw, 2006), studies testing the effect of carotenoid deprivation (Steffen et al., 2010; Ng et al., 2013) and heritability (Cox et al., 2017) of dewlap coloration in *A. sagrei* and *A. distichus* (another species with a carotenoid-based dewlap), found little support for phenotypic and developmental plasticity in dewlap coloration. One exception is a study demonstrating that lizards heavily parasitized by skin mites had duller dewlaps (Cook et al., 2013), but we found no sign of that in our study.

We found no evidence for a role of genetic drift in explaining the observed patterns either. First,

405 *A. sagrei* was distributed across the islands continuously, usually at relatively high population  
406 densities, rather than in small and isolated populations where drift might be expected to have a  
407 strong effect. Second, we found no evidence of isolation-by-distance except on Abaco, which was  
408 sampled at the largest geographical scale, with sites nearly a hundred kilometers apart from each  
409 other. Hence, while isolation-by-distance may explain long-range differences on this island, most of  
410 the differences among habitats across the rest of the sampling region are unlikely to be explained  
411 by genetic drift, as habitats were often in close proximity (less than 10km).

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413 Genetic drift ~~may~~ contribute to some of the observed variation. Indeed, while only Abaco  
414 showed significant patterns consistent with isolation-by-distance (which may emerge under limited  
415 dispersal and drift, Wright 1943; Kimura and Weiss 1964; Slatkin 1987), there may have been  
416 too few sites on most islands to conclusively detect it, and even then, the absence of detectable  
417 isolation-by-distance may not necessarily constitute evidence for the absence of drift. Besides, spa-  
418 tial autocorrelation was the strongest on islands sampled at the largest scales (e.g. some sites on  
419 Abaco were nearly 100km apart, and Eleuthera – the second strongest signal, albeit nonsignificant  
420 – had sites more than 30km apart), such that it is possible that neutral processes and/or dispersal  
421 limitations might ~~in fact~~ contribute to shaping variation over long distances. That said, many  
422 significant differences were found between habitats in close proximity, contrary to what would be  
423 expected under isolation-by-distance, including on islands where spatial autocorrelation was de-  
424 tected. Moreover, *A. sagrei* was distributed across the islands continuously, usually at relatively  
425 high population densities, rather than in small and isolated populations particularly prone to drift.  
426 Together with the fact that isolation-by-distance may not necessarily only emerge from drift (e.g.  
427 if there is a spatial environmental gradient), this indicates that genetic drift may have limited  
428 potential to explain the differences observed between habitats, at least at a local scale.

429

430 In this study, we found larger differences among than within islands, a pattern already reported  
431 and linked to climatic conditions (Driessens et al., 2017) and to densities of predators and of anole  
432 congeners (Vanhooijdonck et al., 2009; Baeckens et al., 2018). Differences among habitats within  
433 islands, however, are still difficult to account for. Remaining hypotheses may include, for exam-  
434 ple, runaway sexual selection (i.e. arbitrary preferences of females for some colors over others,  
435 Andersson 1994) arbitrarily operating in different directions across islands, but no evidence so  
436 far suggests that dewlap is a target of mate choice in anoles (Tokarz, 2002; Tokarz et al., 2005;  
437 Lailvaux and Irschick, 2006; Nicholson et al., 2007). Another hypothesis is that the different ge-  
438 netic constitutions of different islands, perhaps resulting from founder effects (the islands have  
439 been colonized independently, van de Schoot 2016 unpubl.; Driessens et al. 2017; Reynolds et al.  
440 2020), may have predisposed populations to adapt differently to similarly selective circumstances.  
441 That said, signalling can be a multifaceted behavior, and while we focused on dewlap color here,  
442 other potentially important elements of behavioral interactions include dewlap size, dewlap pat-  
443 ttering, display activity as well as behaviors associated with dewlap extensions such as headbobs  
444 and pushups (Vanhooijdonck et al., 2005; Driessens et al., 2014, 2015; Lailvaux et al., 2015). These  
445 have been linked in various ways (and not always consistently across studies) not only with envi-  
446 ronmental variables but also with proxies for sexual selection, predation and species recognition  
447 (Vanhooijdonck et al., 2005; Lailvaux and Irschick, 2007; Vanhooydonck et al., 2009; Driessens  
448 et al., 2017; Baeckens et al., 2018), which do vary among islands but may also vary within islands,  
449 and possibly in interaction with dewlap color. Either way, new data would be needed to test these  
450 hypotheses.

451

452 Altogether, our results show that dewlap color of *A. sagrei* commonly varies between habitat  
453 types, even in close geographical proximity, within islands of the West Indies. However, coloration  
454 differs in different ways across similar habitats from one island to another. We discussed several  
455 non-mutually exclusive mechanisms that could explain these observations. Nevertheless, heteroge-  
456 neous patterns of divergence across islands do not support an adaptive sensory-drive scenario, and  
457 we propose that within-island dewlap color variation may be underlain by a more subtle mosaic of  
factors.

458

## 459 Figures

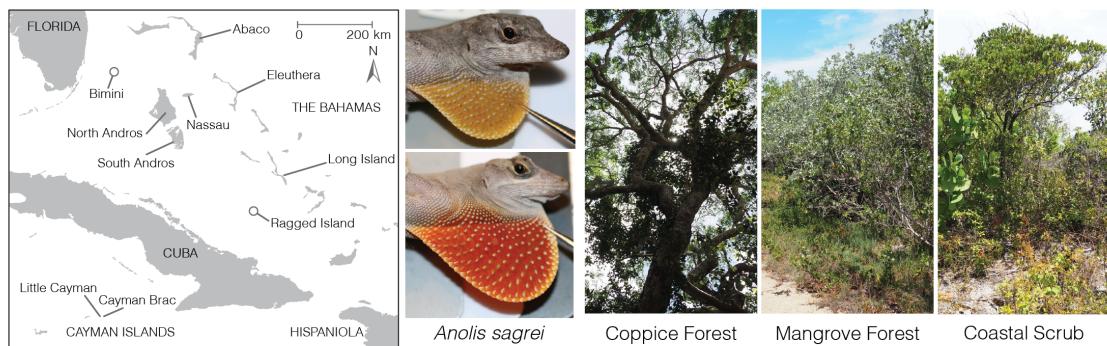


Figure 1: Overview of our study design, including a map of the Bahamas and the Cayman Islands, on which are indicated the nine islands we sampled, two representatives of our study species *Anolis sagrei* with their dewlaps deployed, and the three types of habitats we considered on each island.

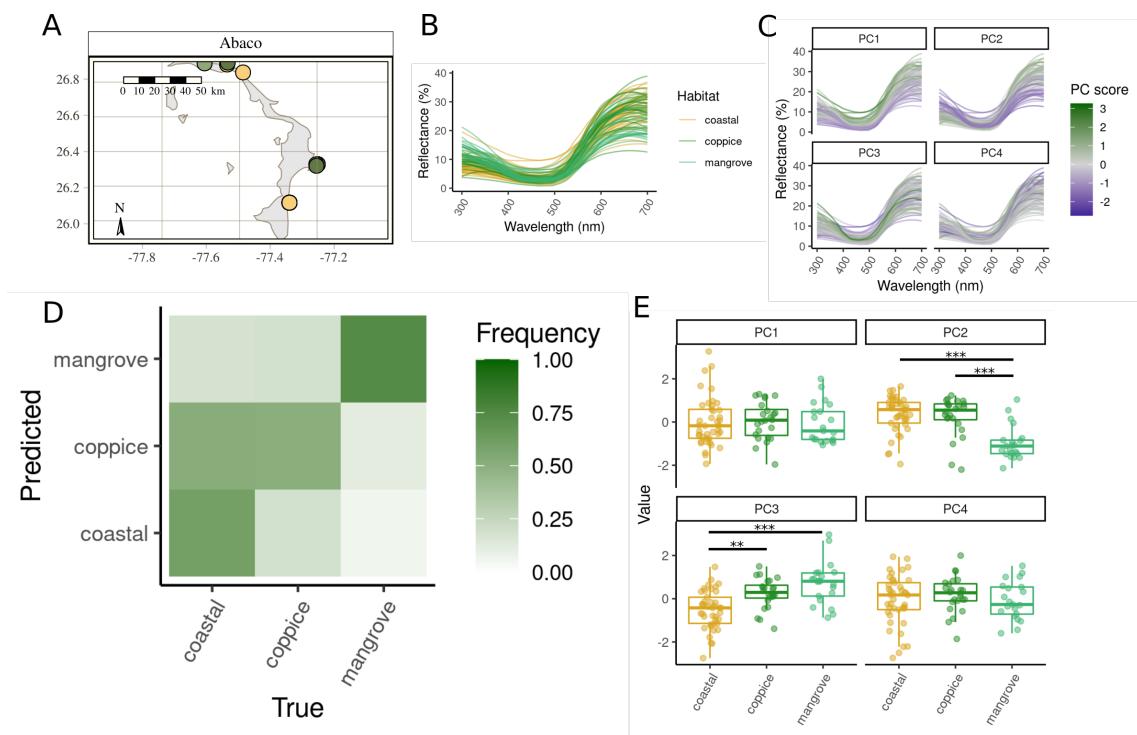


Figure 2: Comparison of dewlap coloration across habitats on Abaco. (A) Map of the island with the sampling sites colored by habitat. (B) Reflectance profiles of all the dewlaps on the island. (C) How reflectance profiles map onto the within-island principal components. (D) Confusion matrix showing the proportion of lizards from each (true) habitat reassigned to each (predicted) habitat by the random forests, based on the first four within-island principal components and averaged across replicates. Each column sums to one. (E) Within-island principal component scores across habitats. Bars indicate significant contrasts. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .

460 **Tables**

Table 1: Random forest classification results. For each island are shown the sample size ( $N$ ) and the proportion of correctly reassigned observations (or success score). P-values were computed using a binomial test and assess the significance of the observed success score relative to the score expected under random guessing. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ , \*\*\*,  $P < 0.001$ .

Island	$N$	Score	$P$
Abaco	86	0.612	< 0.0001 ***
Bimini	67	0.510	0.0011 **
Cayman Brac	50	0.728	< 0.0001 ***
Eleuthera	56	0.493	0.0072 **
Little Cayman	45	0.649	< 0.0001 ***
Long Island	53	0.634	< 0.0001 ***
North Andros	28	0.507	0.0216 *
Ragged Island	50	0.368	0.2874
South Andros	31	0.484	0.0270 *

Table 2: Mantel's test of spatial autocorrelation. For each island are shown the correlation (Pearson's  $\rho$ ) between the matrix of phenotypic distances between populations from each site and the matrix of geographic distances between sites, where phenotypic distances are Euclidean distances between the mean phenotypes of each site in the multivariate space consisting of the first four within-island principal components. P-values assess the significance of the observed correlation against the correlation expected if population means were randomly permuted among sites (999 permutations). \*,  $P < 0.05$ .

Island	$\rho$	$P$
Abaco	0.448	0.033 *
Bimini	0.810	0.250
Cayman Brac	-0.737	0.833
Eleuthera	0.844	0.058
Little Cayman	-0.042	0.667
Long Island	0.367	0.333
North Andros	0.051	0.667
Ragged Island	-0.363	0.708
South Andros	-0.979	1.000

Table 3: Significance of habitat differences in dewlap coloration, using ANOVA for all islands where significant multivariate differences in dewlap coloration were detected by random forests. Model, best-fitting model (either OLS or GLS). AICc, corrected AIC score of the best-fitting model.  $\Delta\text{AICc}$ , difference in AICc between the best-fitting model and the OLS-model. AICcw, AICc weight. Log-lik., log-likelihood.  $\chi^2$ , likelihood ratio. df, degrees of freedom. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .

Island	Variable	AICc	$\Delta\text{AICc}$	AICcw	Model	Log-lik.	$\chi^2$	df	P
Abaco	PC1	255.81	2.16	0.746	OLS	-121.45	0.14	2	0.9308
Abaco	PC2	225.32	4.02	0.882	OLS	-105.66	31.74	2	< 0.0001 ***
Abaco	PC3	229.53	2.01	0.732	OLS	-107.84	27.37	2	< 0.0001 ***
Abaco	PC4	254.64	0.78	0.596	OLS	-120.85	1.36	2	0.5070
Bimini	PC1	194.16	0.77	0.595	OLS	-90.87	7.40	2	0.0248 *
Bimini	PC2	193.49	1.29	0.656	OLS	-90.52	8.09	2	0.0175 *
Bimini	PC3	184.22	-0.23	0.529	GLS	-83.46	10.39	2	0.0056 **
Bimini	PC4	200.91	3.54	0.854	OLS	-94.40	0.33	2	0.8499
Cayman Brac	PC1	136.64	-4.05	0.884	GLS	-59.29	13.81	2	0.0010 **
Cayman Brac	PC2	144.75	3.51	0.853	OLS	-66.24	8.41	2	0.0149 *
Cayman Brac	PC3	127.13	2.77	0.800	OLS	-56.86	27.16	2	< 0.0001 ***
Cayman Brac	PC4	147.37	4.33	0.897	OLS	-67.63	5.63	2	0.0600
Eleuthera	PC1	168.72	2.42	0.770	OLS	-78.46	1.00	2	0.6074 **
Eleuthera	PC2	160.03	-2.20	0.750	GLS	-70.89	11.34	2	0.0034 **
Eleuthera	PC3	163.49	-0.20	0.525	GLS	-72.69	5.57	2	0.0617
Eleuthera	PC4	164.08	3.49	0.852	OLS	-76.01	5.89	2	0.0525
Little Cayman	PC1	130.60	2.50	0.777	OLS	-59.26	8.18	2	0.0167 *
Little Cayman	PC2	112.66	-3.61	0.859	GLS	-46.74	29.76	2	< 0.0001 ***
Little Cayman	PC3	118.32	1.41	0.669	OLS	-52.68	21.34	2	< 0.0001 ***
Little Cayman	PC4	135.58	2.53	0.780	OLS	-61.92	2.85	2	0.2410
Long Island	PC1	154.54	-2.09	0.740	GLS	-68.62	2.91	2	0.2331
Long Island	PC2	155.80	-3.08	0.823	GLS	-68.92	4.52	2	0.1043
Long Island	PC3	150.54	3.67	0.862	OLS	-69.08	11.24	2	0.0036 **
Long Island	PC4	155.05	2.38	0.767	OLS	-71.47	6.46	2	0.0395 *
North Andros	PC1	88.64	0.27	0.534	OLS	-38.84	0.75	2	0.6864
North Andros	PC2	85.36	2.17	0.748	OLS	-37.01	4.42	2	0.1100
North Andros	PC3	85.31	5.82	0.948	OLS	-36.98	4.48	2	0.1065
North Andros	PC4	88.45	4.83	0.918	OLS	-38.74	0.96	2	0.6194
South Andros	PC1	95.12	0.44	0.554	OLS	-41.93	3.10	2	0.2125 *
South Andros	PC2	89.93	-0.05	0.506	GLS	-35.84	7.76	2	0.0206 *
South Andros	PC3	87.21	-6.14	0.956	GLS	-34.05	10.35	2	0.0056 **
South Andros	PC4	83.01	2.94	0.813	OLS	-35.23	16.51	2	0.0003 ***

## 461 References

- 462 Anderson, M. J. (2001). A new method for non-parametric multivariate analysis of variance.  
463 *Austral Ecology*, 26(1):32–46.
- 464 Andersson, M. B. (1994). *Sexual Selection*. Monographs in Behavior and Ecology. Princeton  
465 University Press, Princeton, N.J.
- 466 Arnold, S. J. (1983). Morphology, Performance and Fitness. *American Zoologist*, 23(2):347–361.
- 467 Baeckens, S., Driessens, T., and Van Damme, R. (2018). The brown anole dewlap revisited:  
468 Do predation pressure, sexual selection, and species recognition shape among-population signal  
469 diversity? *PeerJ*, 6:e4722.
- 470 Bartoń, K. (2019). MuMIn: Multi-Model Inference.
- 471 Benjamini, Y. and Hochberg, Y. (1995). Controlling the false discovery rate: A practical and  
472 powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B*, 57:289–  
473 300.
- 474 Bradbury, J. W. and Vehrencamp, S. L. (2011). *Principles of Animal Communication*. Sinauer  
475 Associates, Sunderland, Mass, 2nd ed edition.
- 476 Breiman, L. (2001). Random forests. *Machine Learning*, 45(1):5–32.
- 477 Cook, E. G., Murphy, T. G., and Johnson, M. A. (2013). Colorful displays signal male quality in  
478 a tropical anole lizard. *Naturwissenschaften*, 100(10):993–996.
- 479 Cortez, P. (2020). Rminer: Data Mining Classification and Regression Methods.
- 480 Cox, R. M., Costello, R. A., Camber, B. E., and McGlothlin, J. W. (2017). Multivariate genetic  
481 architecture of the *Anolis* dewlap reveals both shared and sex-specific features of a sexually  
482 dimorphic ornament. *Journal of Evolutionary Biology*, 30(7):1262–1275.
- 483 Cristianini, N. and Shawe-Taylor, J. (2000). *An Introduction to Support Vector Machines and  
484 Other Kernel-based Learning Methods*. Cambridge University Press, first edition.
- 485 Cuthill, I. C., Bennett, A. T. D., Partridge, J. C., and Maier, E. J. (1999). Plumage Reflectance and  
486 the Objective Assessment of Avian Sexual Dichromatism. *The American Naturalist*, 153(2):183–  
487 200.
- 488 Dieckmann, U. and Doebeli, M. (1999). On the origin of species by sympatric speciation. *Nature*,  
489 400(6742):354–357.
- 490 Driessens, T., Baeckens, S., Balzarolo, M., Vanhooydonck, B., Huyghe, K., and Van Damme, R.  
491 (2017). Climate-related environmental variation in a visual signalling device: The male and  
492 female dewlap in *Anolis sagrei* lizards. *Journal of Evolutionary Biology*, 30(10):1846–1861.
- 493 Driessens, T., Huyghe, K., Vanhooydonck, B., and Van Damme, R. (2015). Messages conveyed by  
494 assorted facets of the dewlap, in both sexes of *Anolis sagrei*. *Behavioral Ecology and Sociobiology*,  
495 69(8):1251–1264.
- 496 Driessens, T., Vanhooydonck, B., and Van Damme, R. (2014). Deterring predators, daunting  
497 opponents or drawing partners? Signaling rates across diverse contexts in the lizard *Anolis  
498 sagrei*. *Behavioral Ecology and Sociobiology*, 68(2):173–184.
- 499 Endler, J. (1998). Sensory ecology, receiver biases and sexual selection. *Trends in Ecology &  
500 Evolution*, 13(10):415–420.
- 501 Endler, J. A. (1984). Natural and sexual selection on color patterns in poeciliid fishes. In Balon,  
502 E. K. and Zaret, T. M., editors, *Evolutionary Ecology of Neotropical Freshwater Fishes*, vol-  
503 ume 3, pages 95–111. Springer Netherlands, Dordrecht.
- 504 Endler, J. A. (1990). On the measurement and classification of colour in studies of animal colour  
505 patterns. *Biological Journal of the Linnean Society*, 41(4):315–352.

Endler, J. A. (1991). Variation in the appearance of guppy color patterns to guppies and their predators under different visual conditions. <i>Vision Research</i> , 31(3):587–608.	506 507
Endler, J. A. (1992). Signals, Signal Conditions, and the Direction of Evolution. <i>The American Naturalist</i> , 139:S125–S153.	508 509
Endler, J. A. (1993a). The Color of Light in Forests and Its Implications. <i>Ecological Monographs</i> , 63(1):1–27.	510 511
Endler, J. A. (1993b). Some general comments on the evolution and design of animal communication systems. <i>Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences</i> , 340(1292):215–225.	512 513 514
Endler, J. A. and McLellan, T. (1988). The Processes of Evolution: Toward a Newer Synthesis. <i>Annual Review of Ecology and Systematics</i> , 19(1):395–421.	515 516
Felsenstein, J. (1976). The Theoretical Population Genetics of Variable Selection and Migration. <i>Annual Review of Genetics</i> , 10(1):253–280.	517 518
Fleishman, L. J. (2000). <i>Signal Function, Signal Efficiency and the Evolution of Anoline Lizard Dewlap Color</i> , pages 209–236. Tapir Academic, Trondheim.	519 520
Fleishman, L. J., Leal, M., and Persons, M. H. (2009). Habitat light and dewlap color diversity in four species of Puerto Rican anoline lizards. <i>Journal of Comparative Physiology A</i> , 195(11):1043–1060.	521 522 523
Fleishman, L. J. and Persons, M. (2001). The influence of stimulus and background colour on signal visibility in the lizard <i>Anolis cristatellus</i> . <i>The Journal of Experimental Biology</i> , 204(Pt 9):1559–1575.	524 525 526
Fleishman, L. J., Wadman, C. S., and Maximov, K. J. (2020). The interacting effects of total light intensity and chromatic contrast on visual signal visibility in an <i>Anolis</i> lizard. <i>Animal Behaviour</i> , page S0003347220302037.	527 528 529
García-Ramos, G. and Kirkpatrick, M. (1997). Genetic Models of Adaptation and Gene Flow in Peripheral Populations. <i>Evolution</i> , 51(1):21–28.	530 531
Geneva, A. J., Hilton, J., Noll, S., and Glor, R. E. (2015). Multilocus phylogenetic analyses of Hispaniolan and Bahamian trunk anoles ( <i>distichus</i> species group). <i>Molecular Phylogenetics and Evolution</i> , 87:105–117.	532 533 534
Goodwin, T. W. (1984). <i>The Biochemistry of the Carotenoids</i> . Springer Netherlands, Dordrecht.	535
Halfwerk, W., Jones, P. L., Taylor, R. C., Ryan, M. J., and Page, R. A. (2014). Risky Ripples Allow Bats and Frogs to Eavesdrop on a Multisensory Sexual Display. <i>Science</i> , 343(6169):413–416.	536 537
Harvey, P. H. and Pagel, M. D. (1991). <i>The Comparative Method in Evolutionary Biology</i> . Oxford Series in Ecology and Evolution. Oxford University Press, Oxford ; New York.	538 539
Hendry, A. P., Day, T., and Taylor, E. B. (2007a). Population Mixing and the Adaptive Divergence of Quantitative Traits in Discrete Populations: A Theoretical Framework for Empirical Tests. <i>Evolution</i> , 55(3):459–466.	540 541 542
Hendry, A. P., Taylor, E. B., and McPhail, J. D. (2007b). Adaptive Divergence and the Balance Between Selection and Gene Flow: Lake and Stream Stickleback in the Misty System. <i>Evolution</i> , 56(6):1199–1216.	543 544 545
Henze, N. and Zirkler, B. (1990). A class of invariant consistent tests for multivariate normality. <i>Communications in Statistics - Theory and Methods</i> , 19(10):3595–3617.	546 547
Hijmans, R. J. (2019). Geosphere: Spherical Trigonometry.	548
Hill, G. E., Inouye, C. Y., and Montgomerie, R. (2002). Dietary carotenoids predict plumage coloration in wild house finches. <i>Proceedings of the Royal Society of London. Series B: Biological Sciences</i> , 269(1496):1119–1124.	549 550

- 552 Hill, G. E. and McGraw, K. J., editors (2006). *Bird Coloration*. Harvard University Press, Cambridge, Mass.
- 554 Hollander, M., Wolfe, D. A., and Chicken, E. (2013). *Nonparametric Statistical Methods*. Wiley  
555 Series in Probability and Statistics. John Wiley & Sons, Inc, Hoboken, New Jersey, third edition  
556 edition.
- 557 Howard, R. A. (1950). Vegetation of the Bimini Island Group: Bahamas, B. W. I. *Ecological  
558 Monographs*, 20(4):317–349.
- 559 James, G., Witten, D., Hastie, T., and Tibshirani, R. (2013). *An Introduction to  
560 Statistical Learning*, volume 103 of *Springer Texts in Statistics*. Springer New York, New York,  
561 NY.
- 562 Kamath, A. and Losos, J. B. (2018). Estimating encounter rates as the first step of sexual se-  
563 lection in the lizard *Anolis sagrei*. *Proceedings of the Royal Society B: Biological Sciences*,  
564 285(1873):20172244.
- 565 Kimura, M. and Weiss, G. H. (1964). The Stepping Stone Model of Population Structure and the  
566 Decrease of Genetic Correlation with Distance. *Genetics*, 49(4):561–576.
- 567 Kolbe, J. J., Larson, A., Losos, J. B., and de Queiroz, K. (2008). Admixture determines genetic  
568 diversity and population differentiation in the biological invasion of a lizard species. *Biology  
569 Letters*, 4(4):434–437.
- 570 Kolbe, J. J., Leal, M., Schoener, T. W., Spiller, D. A., and Losos, J. B. (2012). Founder Ef-  
571 fects Persist Despite Adaptive Differentiation: A Field Experiment with Lizards. *Science*,  
572 335(6072):1086–1089.
- 573 Korkmaz, S., Goksuluk, D., and Zararsiz, G. (2014). MVN: An R Package for Assessing Multi-  
574 variate Normality. *The R Journal*, 6(2):151–162.
- 575 Lailvaux, S. P. and Irschick, D. J. (2006). No Evidence for Female Association with High-  
576 Performance males in the Green Anole Lizard, *Anolis carolinensis*. *Ethology*, 112(7):707–715.
- 577 Lailvaux, S. P. and Irschick, D. J. (2007). The Evolution of Performance-Based Male Fighting  
578 Ability in Caribbean Anolis Lizards. *The American Naturalist*, 170(4):573–586.
- 579 Lailvaux, S. P., Leifer, J., Kircher, B. K., and Johnson, M. A. (2015). The incredible shrinking  
580 dewlap: Signal size, skin elasticity, and mechanical design in the green anole lizard (*Anolis  
581 carolinensis*). *Ecology and Evolution*, 5(19):4400–4409.
- 582 Lambert, S. M., Geneva, A. J., Luke Mahler, D., and Glor, R. E. (2013). Using genomic data  
583 to revisit an early example of reproductive character displacement in Haitian *Anolis* lizards.  
584 *Molecular Ecology*, 22(15):3981–3995.
- 585 Lazareva, O. F., Shimizu, T., and Wasserman, E. A. (2012). *How Animals See the  
586 World**Comparative Behavior, Biology, and Evolution of Vision*. Oxford University Press.
- 587 Leal, M. and Fleishman, L. J. (2002). Evidence for habitat partitioning based on adaptation to  
588 environmental light in a pair of sympatric lizard species. *Proceedings of the Royal Society of  
589 London. Series B: Biological Sciences*, 269(1489):351–359.
- 590 Leal, M. and Fleishman, L. J. (2004). Differences in Visual Signal Design and Detectability between  
591 Allopatric Populations of *Anolis* Lizards. *The American Naturalist*, 163(1):26–39.
- 592 Leal, M. and Rodríguez-Robles, J. A. (1995). Antipredator Responses of *Anolis cristatellus* (Sauria:  
593 Polychrotidae). *Copeia*, 1995(1):155–161.
- 594 Leal, M. and Rodriguez-Robles, J. A. (1997). Antipredator Responses of the Puerto Rican Giant  
595 Anole, *Anolis cuvieri* (Squamata: Polychrotidae). *Biotropica*, 29(3):372–375.
- 596 Leal, M. and Rodríguez-Robles, J. A. (1997). Signalling displays during predator-prey interactions  
in a Puerto Rican anole, *Anolis cristatellus*. *Animal Behaviour*, 54(5):1147–1154.

Legendre, P. and Legendre, L. (2012). <i>Numerical Ecology</i> . Number 24 in <i>Developments in Environmental Modelling</i> . Elsevier, Amsterdam, third english edition edition.	598 599
Lenormand, T. (2002). Gene flow and the limits to natural selection. <i>Trends in Ecology &amp; Evolution</i> , 17(4):183–189.	600 601
Liaw, A. and Wiener, M. (2002). Classification and Regression by randomForest. <i>R News</i> , 2(3):18–22.	602 603
Losos, J. B. (1985). An Experimental Demonstration of the Species-Recognition Role of <i>Anolis</i> Dewlap Color. <i>Copeia</i> , 1985(4):905–910.	604 605
Losos, J. B. (2009). <i>Lizards in an Evolutionary Tree: Ecology and Adaptive Radiation of Anoles</i> . University of California Press.	606 607
Losos, J. B. (2011). Convergence, Adaptation, and Constraint. <i>Evolution</i> , 65(7):1827–1840.	608
Losos, J. B., Irschick, D. J., and Schoener, T. W. (1994). Adaptation and Constraint in the Evolution of Specialization of Bahamian <i>Anolis</i> Lizards. <i>Evolution</i> , 48(6):1786–1798.	609 610
Losos, J. B., Schoener, T. W., Warheit, K. I., and Creer, D. (2001). Experimental studies of adaptive differentiation in Bahamian <i>Anolis</i> lizards. <i>Genetica</i> , 112–113:399–415.	611 612
Losos, J. B., Warheit, K. I., and Schoener, T. W. (1997). Adaptive differentiation following experimental island colonization in <i>Anolis</i> lizards. <i>Nature</i> , 387(6628):70–73.	613 614
Macedonia, J. M. (2001). Habitat light, colour variation, and ultraviolet reflectance in the Grand Cayman anole, <i>Anolis conspersus</i> . <i>Biological Journal of the Linnean Society</i> , 73(3):299–320.	615 616
Macedonia, J. M., Clark, D. L., Riley, R. G., and Kemp, D. J. (2013). Species recognition of color and motion signals in <i>Anolis grahami</i> : Evidence from responses to lizard robots. <i>Behavioral Ecology</i> , 24(4):846–852.	617 618 619
Macedonia, J. M., Clark, D. L., and Tamasi, A. L. (2014). Does Selection Favor Dewlap Colors that Maximize Detectability? A Test with Five Species of Jamaican <i>Anolis</i> Lizards. <i>Herpetologica</i> , 70(2):157–170.	620 621 622
Macedonia, J. M. and Stamps, J. A. (1994). Species Recognition in <i>Anolis grahami</i> (Sauria, Iguanidae): Evidence from Responses to Video Playbacks of Conspecific and Heterospecific Displays. <i>Ethology</i> , 98(3-4):246–264.	623 624 625
Maia, R., Eliason, C. M., Bitton, P.-P., Doucet, S. M., and Shawkey, M. D. (2013). Pavo: An R package for the analysis, visualization and organization of spectral data. <i>Methods in Ecology and Evolution</i> , pages n/a–n/a.	626 627 628
Ng, J., Geneva, A. J., Noll, S., and Glor, R. E. (2017). Signals and Speciation: <i>Anolis</i> Dewlap Color as a Reproductive Barrier. <i>Journal of Herpetology</i> , 51(3):437–447.	629 630
Ng, J. and Glor, R. E. (2011). Genetic differentiation among populations of a Hispaniolan trunk anole that exhibit geographical variation in dewlap colour. <i>Molecular Ecology</i> , 20(20):4302–4317.	631 632
Ng, J., Kelly, A. L., MacGuigan, D. J., and Glor, R. E. (2013). The Role of Heritable and Dietary Factors in the Sexual Signal of a Hispaniolan <i>Anolis</i> Lizard, <i>Anolis distichus</i> . <i>Journal of Heredity</i> , 104(6):862–873.	633 634 635
Ng, J., Landeen, E. L., Logsdon, R. M., and Glor, R. E. (2012). Correlation Between <i>Anolis</i> Lizard Dewlap Phenotype and Environmental Variation Indicates Adaptive Divergence of a Signal Important to Sexual Selection and Species Recognition. <i>Evolution</i> , 67(2):573–582.	636 637 638
Ng, J., Ossip-Klein, A. G., and Glor, R. E. (2016). Adaptive signal coloration maintained in the face of gene flow in a Hispaniolan <i>Anolis</i> Lizard. <i>BMC Evolutionary Biology</i> , 16(1):193.	639 640
Nicholson, K. E., Harmon, L. J., and Losos, J. B. (2007). Evolution of <i>Anolis</i> Lizard Dewlap Diversity. <i>PLoS ONE</i> , 2(3):e274.	641

- 643 Nosil, P. and Crespi, B. J. (2004). Does Gene Flow Constrain Adaptive Divergence or Vice Versa? A  
 644 Test Using Ecomorphology and Sexual Isolation in *Timema cristinae* Walking-Sticks. *Evolution*,  
 645 58(1):102–112.
- 646 Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P. R.,  
 647 O'Hara, R. B., Simpson, G. L., Solymos, P., Stevens, M. H. H., Szoecs, E., and Wagner, H.  
 648 (2019). Vegan: Community Ecology Package. R package version 2.5-6.
- 649 Pinheiro, J. and Bates, D. (2000). *Mixed-Effects Models in S and S-PLUS*. Statistics and Com-  
 650 puting. Springer-Verlag, New York.
- 651 Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., and Team, R. C. (2020). Nlme: Linear and  
 652 Nonlinear Mixed Effects Models.
- 653 Pohlert, T. (2020). PMCMRplus: Calculate Pairwise Multiple Comparisons of Mean Rank Sums  
 654 Extended. R package version 1.4.4.
- 655 R Core Team (2019). R: A Language and Environment for Statistical Computing. R Foundation  
 656 for Statistical Computing.
- 657 Rand, A. S. and Williams, E. E. (1970). An Estimation of Redundancy and Information Content  
 658 of Anole Dewlaps. *The American Naturalist*, 104(935):99–103.
- 659 Reynolds, R. G., Kolbe, J. J., Glor, R. E., López-Darias, M., Gómez Pourroy, C. V., Harrison,  
 660 A. S., Queiroz, K., Revell, L. J., and Losos, J. B. (2020). Phylogeographic and phenotypic  
 661 outcomes of brown anole colonization across the Caribbean provide insight into the beginning  
 662 stages of an adaptive radiation. *Journal of Evolutionary Biology*, 33(4):468–494.
- 663 Richardson, J. L. and Urban, M. C. (2013). Strong Selection Barriers Explain Microgeographic  
 664 Adaptation in Wild Salamander Populations. *Evolution*, 67(6):1729–1740.
- 665 Richardson, J. L., Urban, M. C., Bolnick, D. I., and Skelly, D. K. (2014). Microgeographic adap-  
 666 tation and the spatial scale of evolution. *Trends in Ecology & Evolution*, 29(3):165–176.
- 667 Schoener, T. W. (1968). The *Anolis*Lizards of Bimini: Resource Partitioning in a Complex Fauna.  
 668 *Ecology*, 49(4):704–726.
- 669 Seehausen, O. (1997). Cichlid Fish Diversity Threatened by Eutrophication That Curbs Sexual  
 670 Selection. *Science*, 277(5333):1808–1811.
- 671 Sigmund, W. R. (1983). Female Preference for *Anolis carolinensis* Males as a Function of Dewlap  
 672 Color and Background Coloration. *Journal of Herpetology*, 17(2):137–143.
- 673 Slatkin, M. (1987). Gene Flow and the Geographic Structure of Natural Populations. *Science*,  
 674 236(4803):787–792.
- 675 Stapley, J., Wordley, C., and Slate, J. (2011). No Evidence of Genetic Differentiation Between  
 676 Anoles With Different Dewlap Color Patterns. *Journal of Heredity*, 102(1):118–124.
- 677 Steffen, J. E. and Guyer, C. C. (2014). Display behaviour and dewlap colour as predictors of  
 678 contest success in brown anoles: Dewlap Colour and Behaviour in Contests. *Biological Journal  
 679 of the Linnean Society*, 111(3):646–655.
- 680 Steffen, J. E., Hill, G. E., and Guyer, C. (2010). Carotenoid Access, Nutritional Stress, and the  
 681 Dewlap Color of Male Brown Anoles. *Copeia*, 2010(2):239–246.
- 682 Thorpe, R. S. (2002). Analysis of Color Spectra in Comparative Evolutionary Studies: Molecular  
 683 Phylogeny and Habitat Adaptation in the St. Vincent Anole (*Anolis trinitatis*). *Systematic  
 684 Biology*, 51(4):554–569.
- 685 Thorpe, R. S. and Stenson, A. G. (2002). Phylogeny, Paraphyly and Ecological Adaptation of  
 686 the Colour and Pattern in the Anolis Roquet Complex on Martinique: Interaction Between  
 687 Phylogeny and Adaptation. *Molecular Ecology*, 12(1):117–132.

- Tokarz, R. R. (2002). An Experimental Test of the Importance of the Dewlap in Male Mating Success in the Lizard *Anolis sagrei*. *Herpetologica*, 58(1):87–94. 688  
689
- Tokarz, R. R. (2006). Importance of Prior Physical Contact with Familiar Females in the Development of a Male Courtship and Mating Preference for Unfamiliar Females in the Lizard *Anolis Sagrei*. *Herpetologica*, 62(2):115–124. 690  
691  
692
- Tokarz, R. R., Paterson, A. V., and McMann, S. (2005). Importance of Dewlap Display in Male Mating Success in Free-Ranging Brown Anoles (*Anolis sagrei*). *Journal of Herpetology*, 39(1):174–177. 693  
694  
695
- van de Schoot, M. (2016). Within and between Island Radiation and Genetic Variation in Anolis Sagrei. PhD thesis, Wageningen University, Wageningen, The Netherlands. 696  
697
- Vanhooydonck, B., Herrel, A., Meyers, J. J., and Irschick, D. J. (2009). What determines dewlap diversity in *Anolis* lizards? An among-island comparison. *Journal of Evolutionary Biology*, 22(2):293–305. 698  
699  
700
- Vanhooydonck, B., Herrel, A. Y., Van Damme, R., and Irschick, D. J. (2005). Does dewlap size predict male bite performance in Jamaican *Anolis* lizards? *Functional Ecology*, 19(1):38–42. 701  
702
- Willi, Y. and Hoffmann, A. A. (2012). Microgeographic adaptation linked to forest fragmentation and habitat quality in the tropical fruit fly *Drosophila birchii*. *Oikos*, 121(10):1627–1637. 703  
704
- Williams, E. E. (1969). The Ecology of Colonization as Seen in the Zoogeography of Anoline Lizards on Small Islands. *The Quarterly Review of Biology*, 44(4):345–389. 705  
706
- Williams, E. E. and Rand, A. S. (1977). Species Recognition, Dewlap Function and Faunal Size. *American Zoologist*, 17(1):261–270. 707  
708
- Wright, S. (1943). Isolation by Distance. *Genetics*, 28(2):114–138. 709
- Zuur, A. F., editor (2009). Mixed Effects Models and Extensions in Ecology with R. Statistics for Biology and Health. Springer, New York, NY. 710

## 712 Appendix

713 Here we describe more precisely the patterns identified on each island.

714 On Abaco, dewlaps from the mangrove habitat were the best discriminated, while dewlaps from  
715 the beach scrub habitat were often mistaken for dewlaps from the coppice habitat (Fig. 2D). Im-  
716 portance analysis revealed that beach scrub and mangrove lizards mostly differed in reflectance in  
717 the ultraviolet (UV) end of the spectrum (below 400nm, Fig. S2F), where mangrove dewlaps had  
718 higher UV reflectance relative to beach scrub lizards, and coppice lizards had an intermediate UV  
719 reflectance between the two other habitats (Fig. 2B). Consistent with this, our analyses of variance  
720 detected significantly lower PC2 scores in mangrove lizards than in the two other habitats (Fig.  
721 2E, Table 3), representing a higher UV-reflectance relative to red (Fig. 2C). Beach scrub lizards  
722 also scored lower on PC3 (Fig. 2E, Table 3), indicating less curvature of the reflectance profile  
723 and relatively higher reflectance at intermediate wavelengths (blue-to-yellow) than at the ends of  
724 the range (Fig. 2C). Differences were detected between sites both at large ( $\sim 100\text{km}$ ) and short  
725 ( $< 1\text{km}$ ) distances (Fig. S2G). Abaco was the only island where we detected significant spatial  
726 autocorrelation (Table 2), that is, sites that were closer geographically tended to have populations  
727 of lizards with more similar dewlap colors.

729 On Bimini, the random forests mostly correctly classified lizards from the coppice and mangrove  
730 habitats while often misclassifying lizards from the beach scrub habitat (Fig. S3D). Relatively flat  
731 importance profiles for beach scrub lizards suggested that brightness was used instead of a partic-  
732 ular wavelength to identify some of the beach scrub dewlaps (Fig. S3F). Indeed, some beach scrub  
733 dewlaps were substantially brighter than the rest (Fig. S3B, C), a pattern that was captured by  
734 our analysis of variance along PC1 (i.e. brightness, Fig. S3C, E, Table 3). The random forests also  
735 used UV reflectance to discriminate between coppice and mangrove dewlaps (Fig. S3F), which  
736 could reflect the significant difference we detected along PC3 between these two habitats (Fig.  
737 S3C, E, Table 3). Beach scrub lizards were characterized by elevated red reflectance relative to  
738 UV (as represented by PC2, (Fig. S3C, E)), and beach scrub and mangrove lizards were charac-  
739 terized by a more even distribution of the reflectance along the spectrum (as represented by PC3,  
740 (Fig. S3C, E)), in contrast to coppice lizards which harbored a stronger curvature at interme-  
741 diate wavelengths (Fig. S3C, E). On this island, the beach scrub and coppice habitats were separated  
742 by a few hundred meters, making this contrast the smallest geographical scale at which differences  
743 in coloration were found in our study (Fig. S3G).

745 On Cayman Brac, all three habitats could be well discriminated against each other (Fig. S4D),  
746 with UV reflectance appearing to be an important variable differentiating beach scrub and man-  
747 grove dewlaps (Fig. S4F). In contrast, coppice dewlaps had a relatively flat importance profile,  
748 suggesting that brightness made them more distinct rather than any particular wavelength (Fig.  
749 S4F). Consistent with this, coppice dewlaps were significantly different from all other dewlaps  
750 along PC1 (Fig. S4E, Table 3). At a distance between 2 and 3km (Fig. S4G), dewlaps in the  
751 beach scrub habitat reflected more red light (as represented by PC2, Fig. S4C, E) and more UV  
752 (as represented by PC3, Fig. S4C, E) than in the mangrove habitat. Coppice lizards were also  
753 characterized by a higher UV reflectance than mangrove dewlaps (PC3, Fig. S4C, E, Table 3).

755 On Eleuthera, although random forests detected between-habitat differences in dewlap color,  
756 other approaches did not (Tables S8 and S7), suggesting that the differences may be small. Consis-  
757 tent with this, the only significant univariate difference detected was for PC2 between beach scrub  
758 and mangrove lizards, where beach scrub lizards had higher levels of red reflectance and mangrove  
759 lizards higher levels of UV reflectance (Fig. S5C, E, Table 3).

761 Little Cayman was characterized by a better discrimination of mangrove lizards from the rest  
762 than between beach scrub and coppice lizards (Fig. S6D). Mangrove dewlaps were most distinct  
763 with respect to their reflectance in short wavelengths (Fig. S6F), with significantly lower UV re-  
764 flectance (as represented by PC2, Fig. S6C, E, Table 3). Beach scrub lizards were characterized  
765 by brighter dewlaps than coppice lizards (PC1), and also more convex curves, i.e. slightly higher  
766 UV and red reflectance (as represented by higher PC3 scores), than lizards from the other two  
767 habitats (Fig. S6C, E, Table 3).

On Long Island the three habitats were relatively well discriminated (Fig. S7D). Importance profiles indicated that short wavelengths were used to discriminate between beach scrub and mangrove lizards (Fig. S7F). Beach scrub lizards had more curved reflectance profiles than in either of the two other habitats, with higher levels of UV and red reflectance relative to intermediate wavelengths (PC3, Fig. S7C, E, Table 3). Beach scrub lizards also differed from mangrove lizards along PC4 (Fig. S7E), which represented a rather small portion of the variance not already explained by the first three principal components, and is therefore difficult to interpret (Fig. S7C). Coppice lizards were significantly darker than mangrove and beach scrub lizards (PC1, Fig. S7C, E, Table 3).

On North Andros, although the random forest classification was significant ( $P = 0.0216$ , Table 1) and the average confusion matrix indicated that lizards from beach scrub were particularly well predicted (Fig. S8D), no significant univariate differences were detected along any of the four PCs (Fig. S8E, Table 3). Importance analysis of full-spectrum random forests showed higher importance scores near the UV-end of the spectrum in discriminating beach scrub dewlaps from the rest (Fig. S8F). Besides, reflectance curves of beach scrub dewlaps appeared more similar to each other in the UV range than dewlaps from other habitats (Fig. S8B), suggesting that the machines may have used this low within-habitat variance, as opposed to between-habitat differences in means, to correctly classify beach scrub lizards. A small sample size on this island may also have contributed to a lack of power in detecting univariate differences using analyses of variance (Table S1).

On South Andros beach scrub and coppice dewlaps could be discriminated better against each other than with mangrove dewlaps (Fig. S10D), with importance profiles supporting UV-reflectance as a predictor of coppice lizards (Fig. S10F). Coppice lizards had more curved reflectance profiles than beach scrub lizards (PC3), and lizards from both habitats differed along PC4, which is again more difficult to interpret (Fig. S10C, E, Table 3). Beach scrub lizards also differed from mangrove lizards in PC4 (Fig. S10C, E, Table 3).

Classification success was not significantly better than expected by chance on Ragged Island (Table 1) where nearly no habitat could be differentiated from any other based on reflectance.

## Supplementary Figures

800

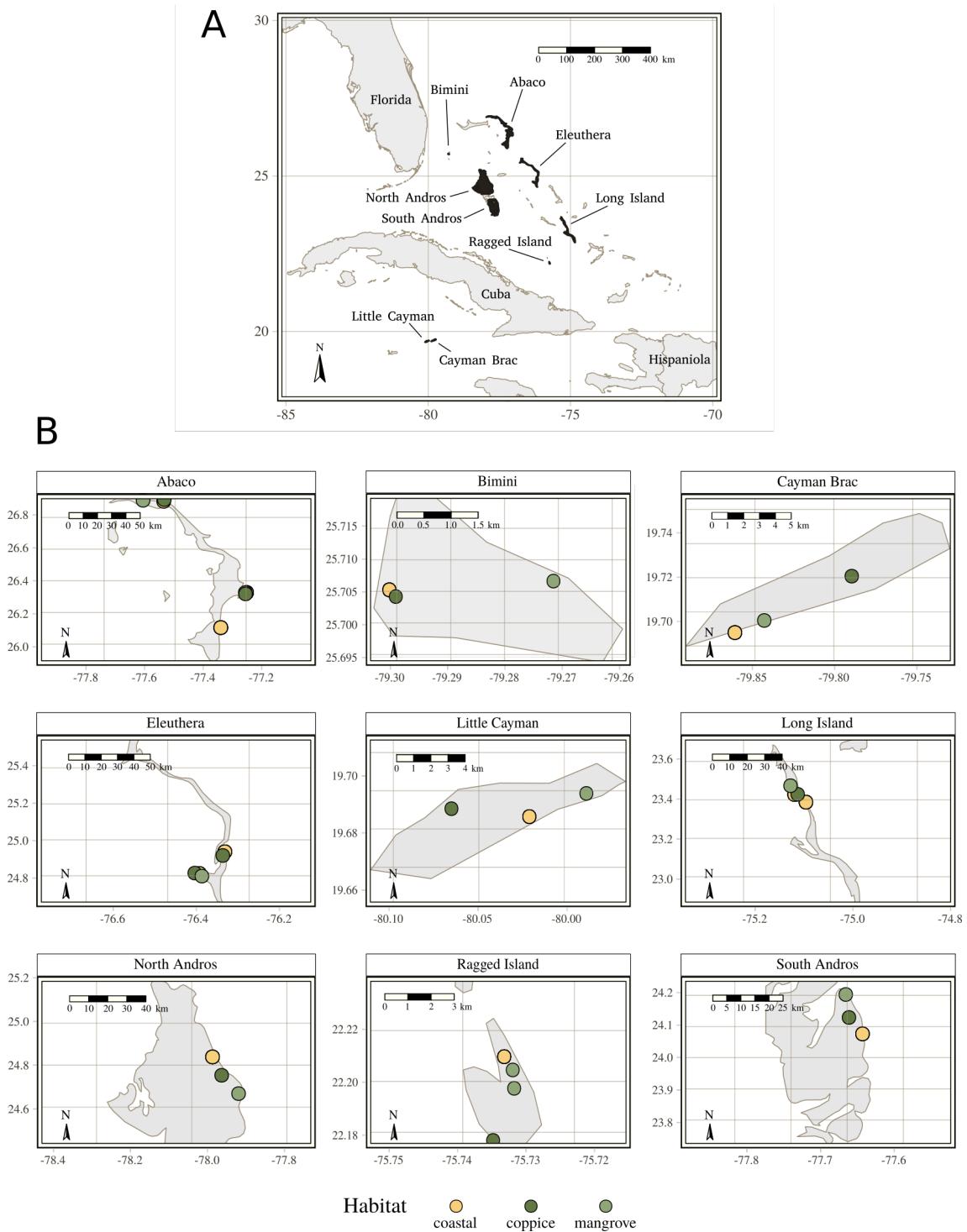


Figure S1: Maps of the islands. (A) Map of the West Indies with sampled islands highlighted in black. (B) Sampling sites within islands colored after their respective habitat types.

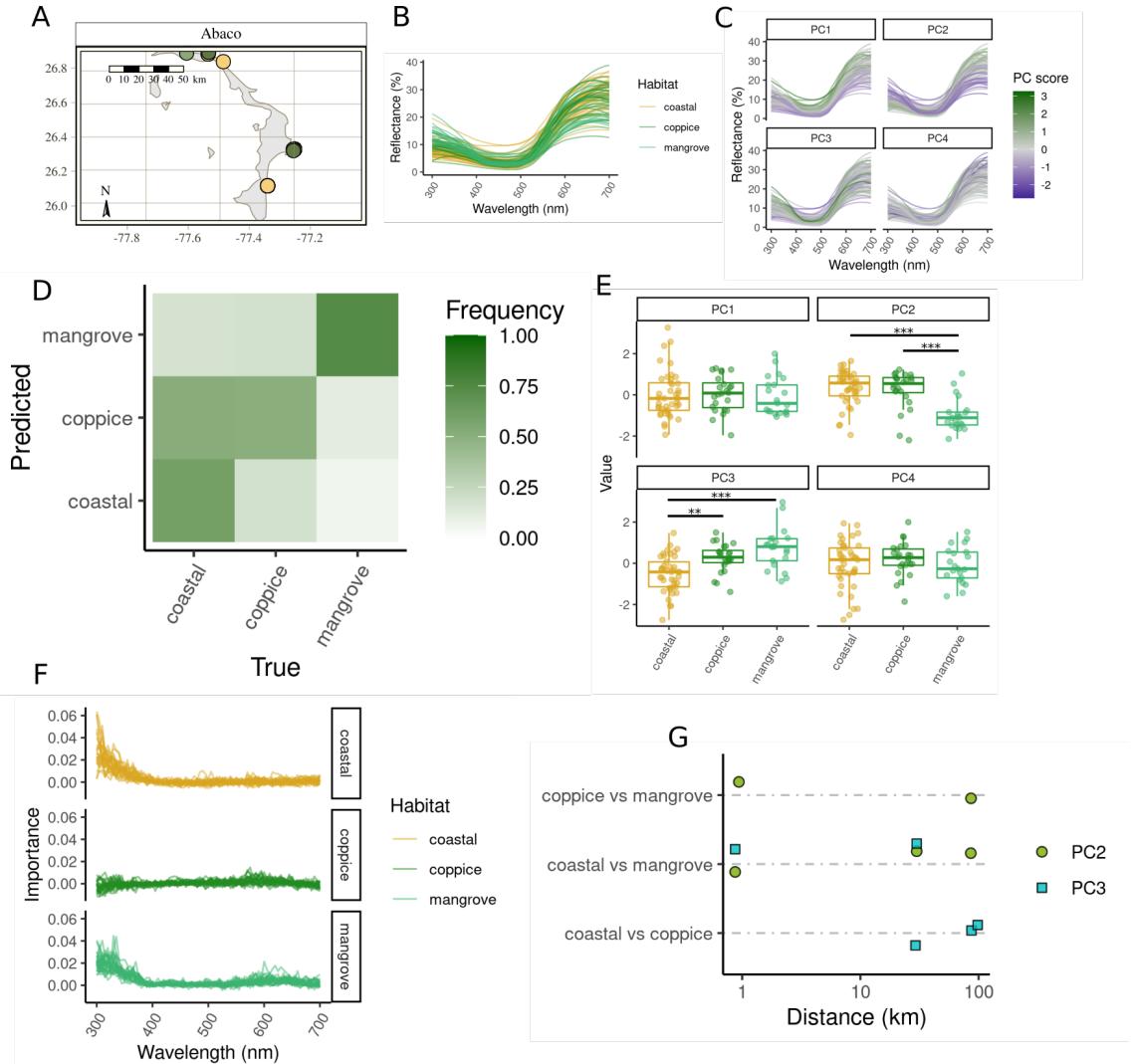


Figure S2: Comparison of dewlap coloration across habitats on Abaco, with extended results. (A–E) Legend as per Figure 2. (F) One-dimensional sensitivity analysis showing the relative importance (mean decrease in accuracy) of the various wavelengths in random forest classification of the whole spectrum. (G) Geographical distance between sites where significant differences were detected in within-island principal component scores (Wilcoxon test, Benjamini-Hochberg correction,  $P < 0.05$ ), including only pairs of sites whose habitats were involved in between-habitat dewlap differences.

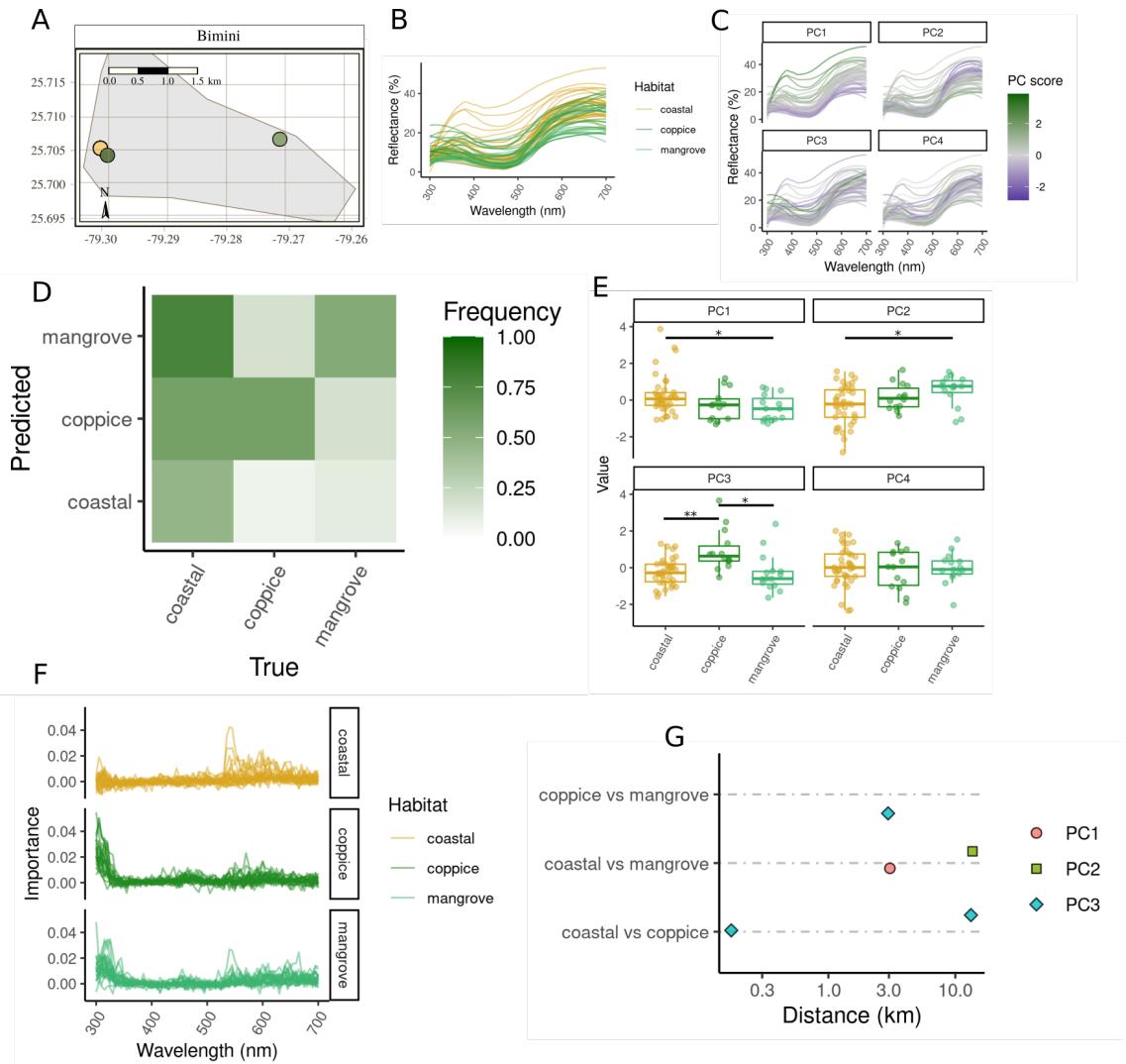


Figure S3: Comparison of dewlap coloration across habitats on Bimini. Legend is as per Figure S2.

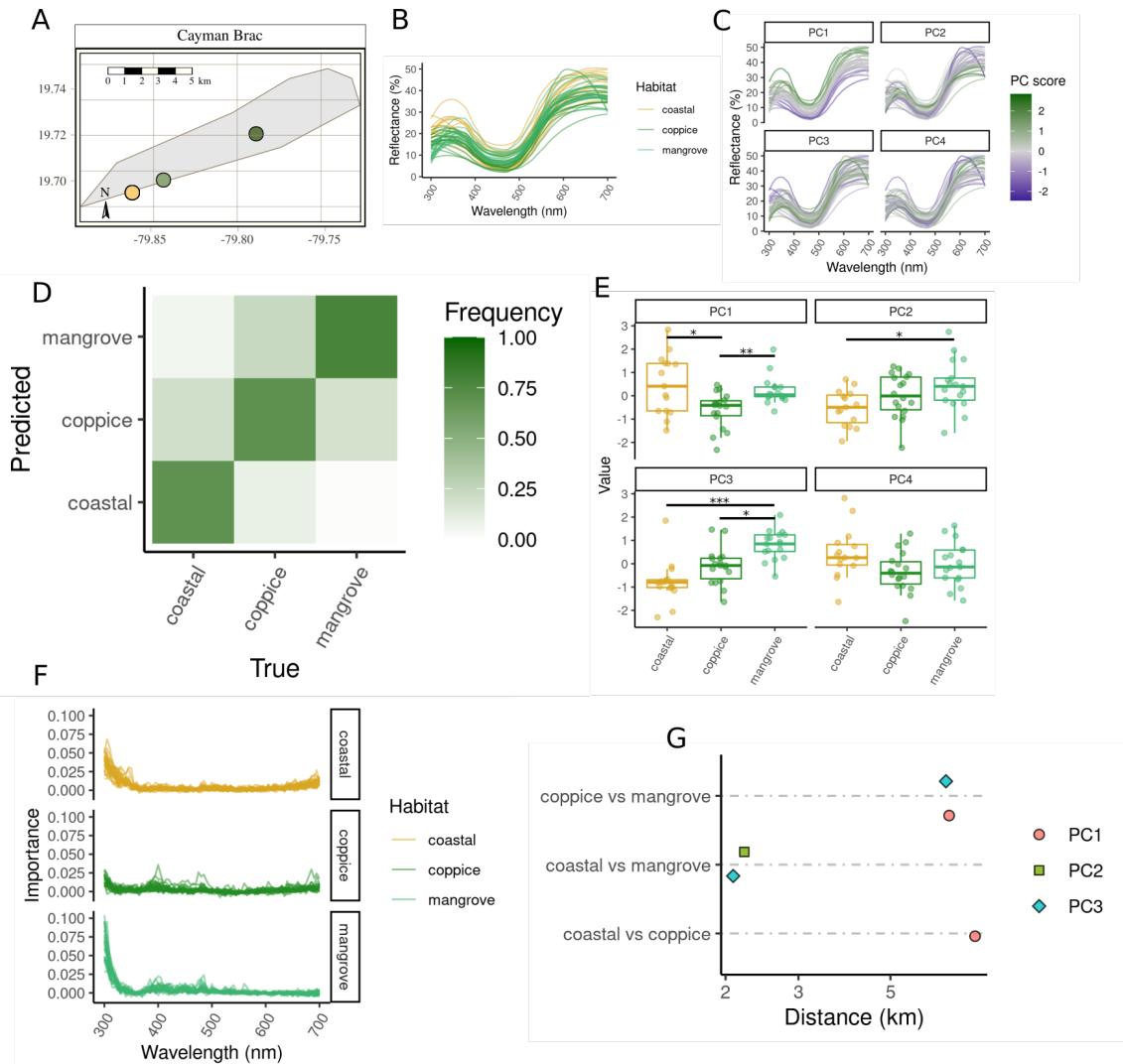


Figure S4: Comparison of dewlap coloration across habitats on Cayman Brac. Legend is as per Figure S2.

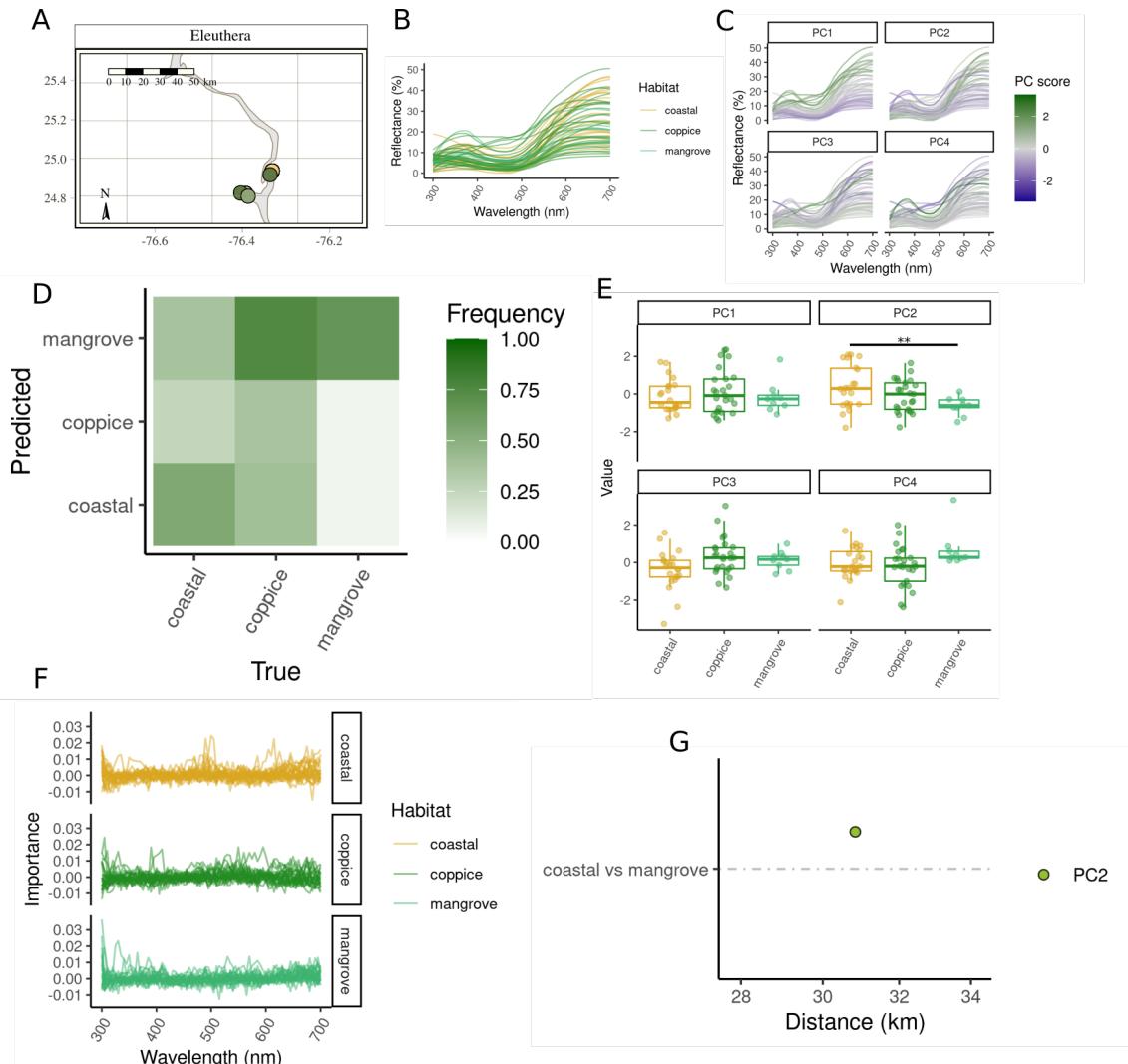


Figure S5: Comparison of dewlap coloration across habitats on Eleuthera. Legend is as per Figure S2.

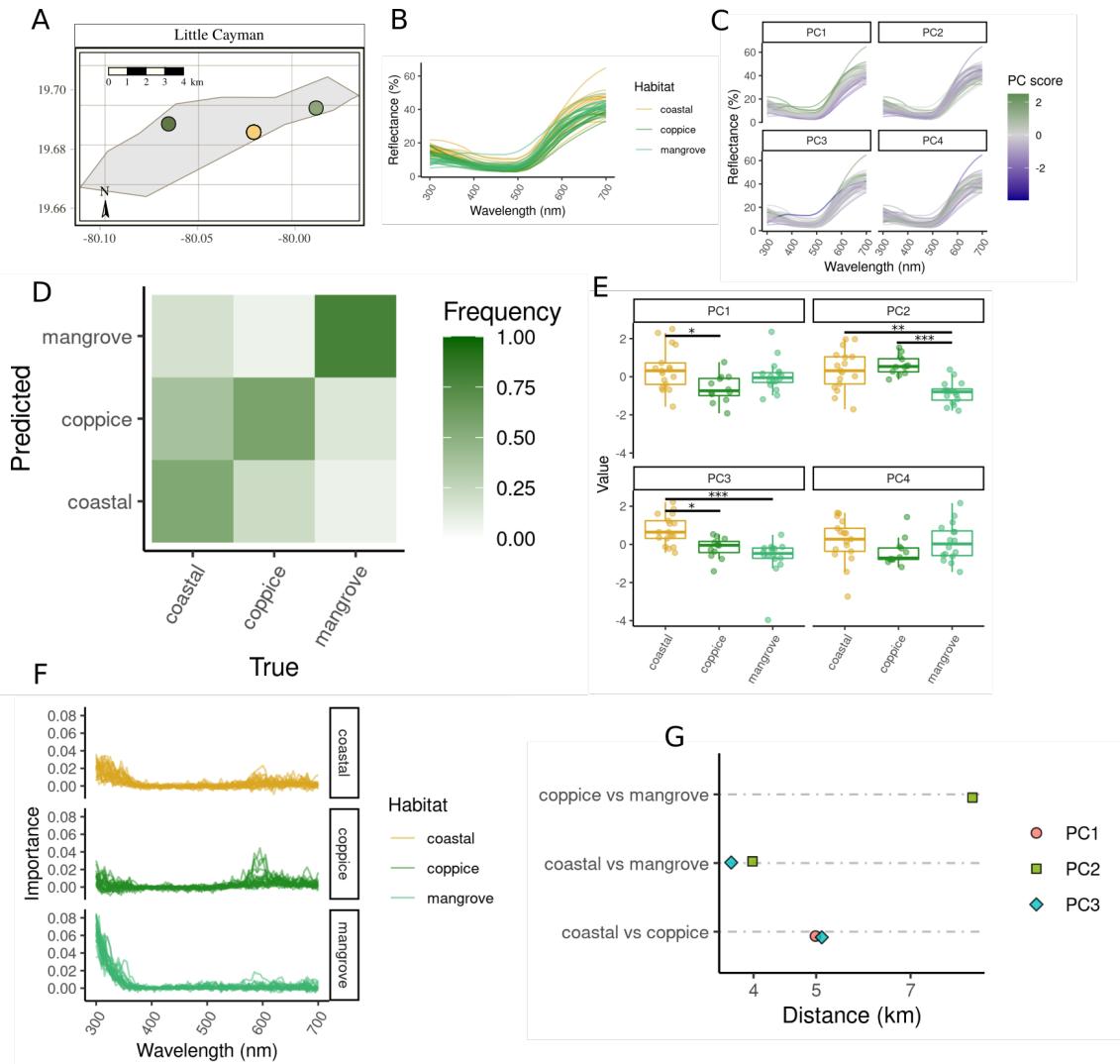


Figure S6: Comparison of dewlap coloration across habitats on Little Cayman. Legend is as per Figure S2.

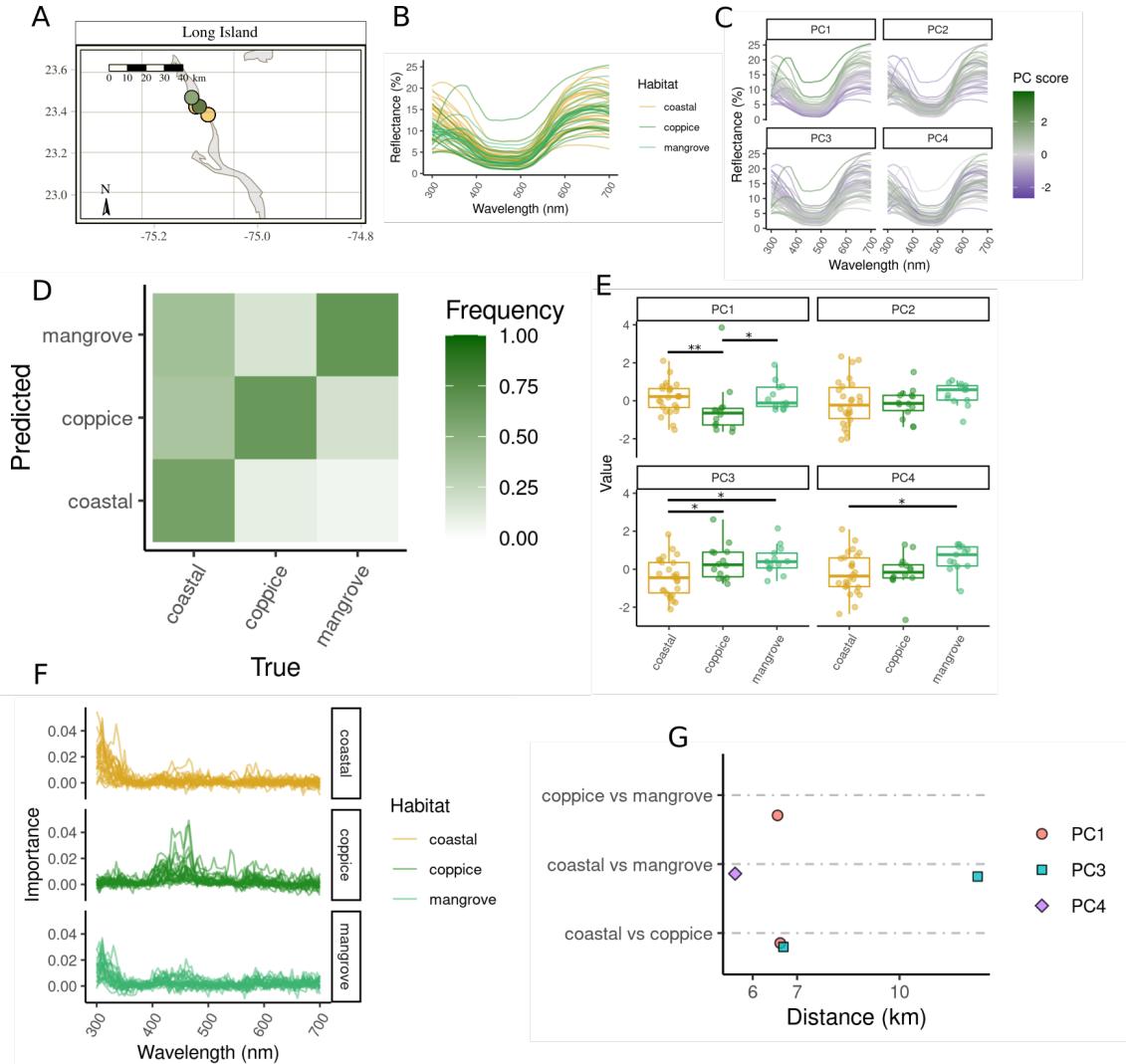


Figure S7: Comparison of dewlap coloration across habitats on Long Island. Legend is as per Figure S2.

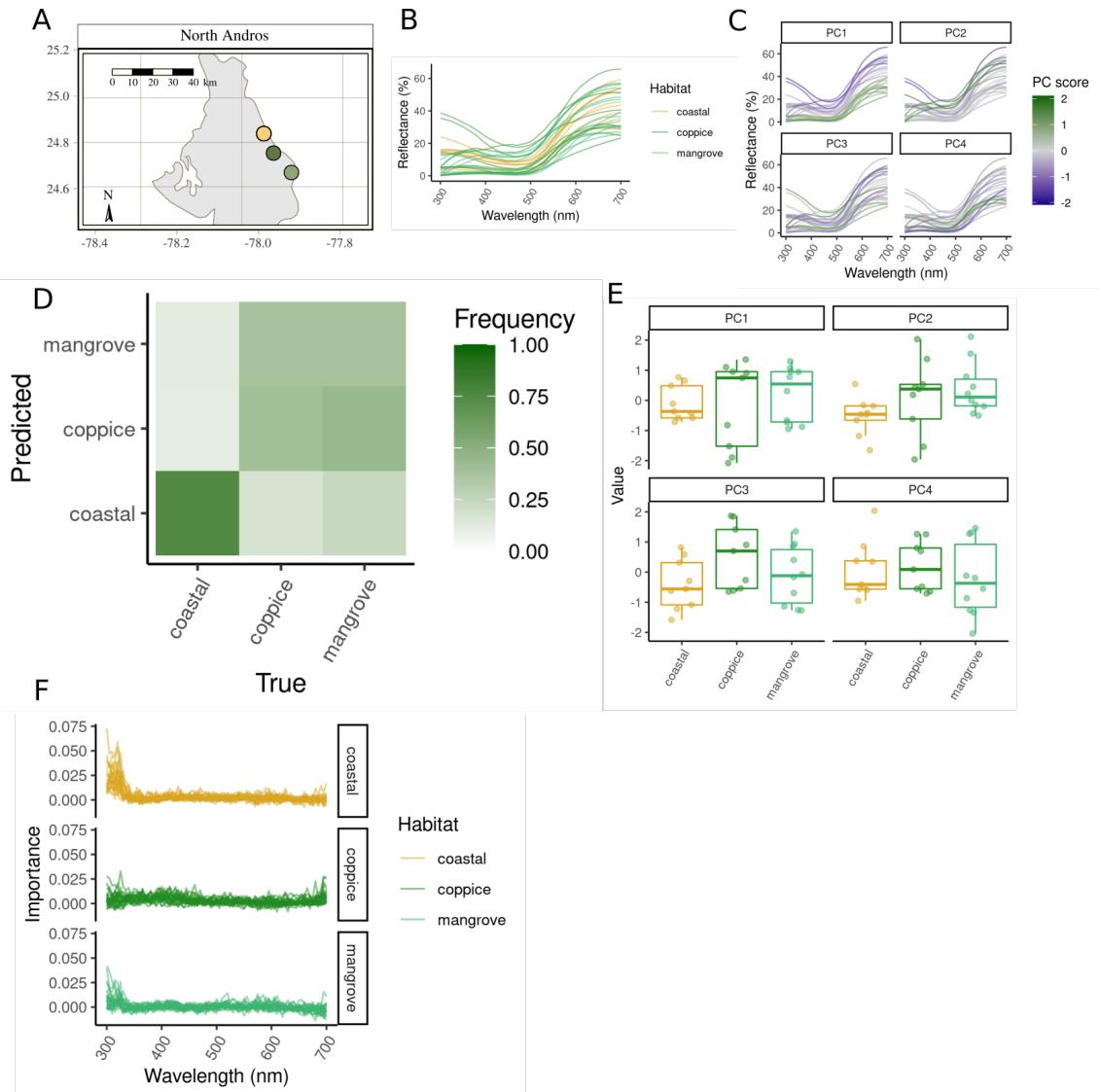


Figure S8: Comparison of dewlap coloration across habitats on North Andros. Legend is as per Figure S2, but without panel G.

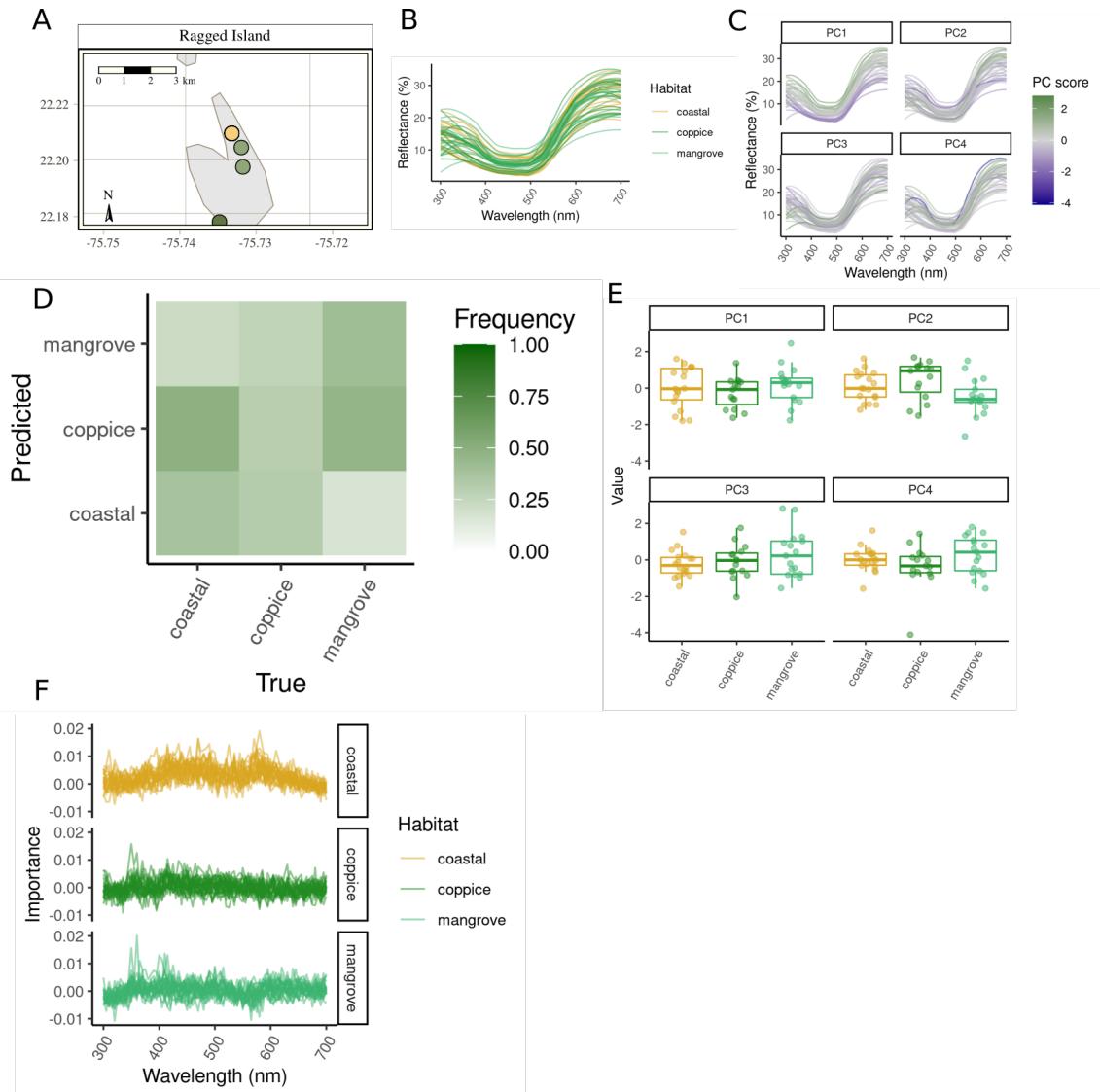


Figure S9: Comparison of dewlap coloration across habitats on Ragged Island. Legend is as per Figure S2, but without panel G.

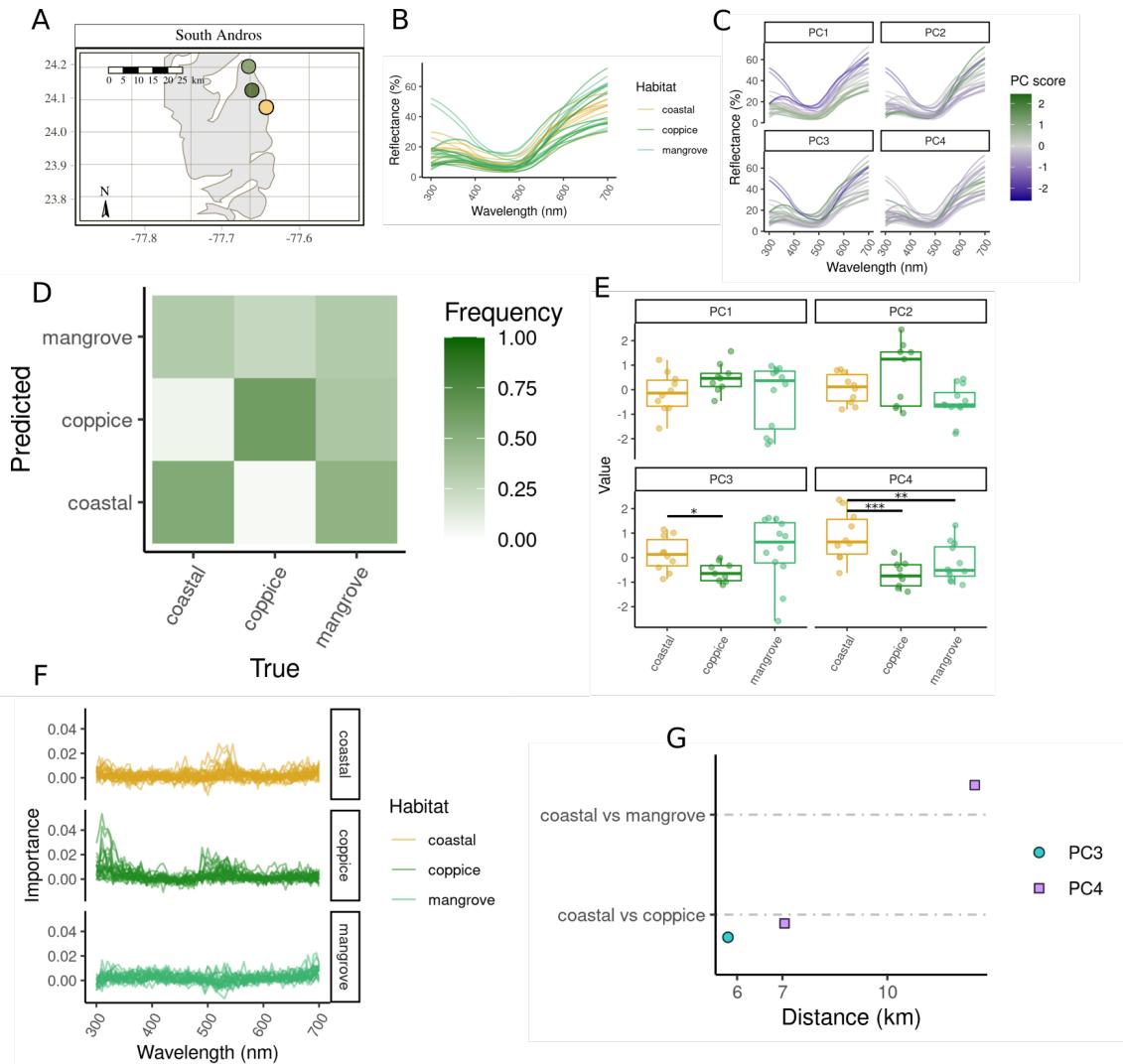


Figure S10: Comparison of dewlap coloration across habitats on South Andros. Legend is as per Figure S2.

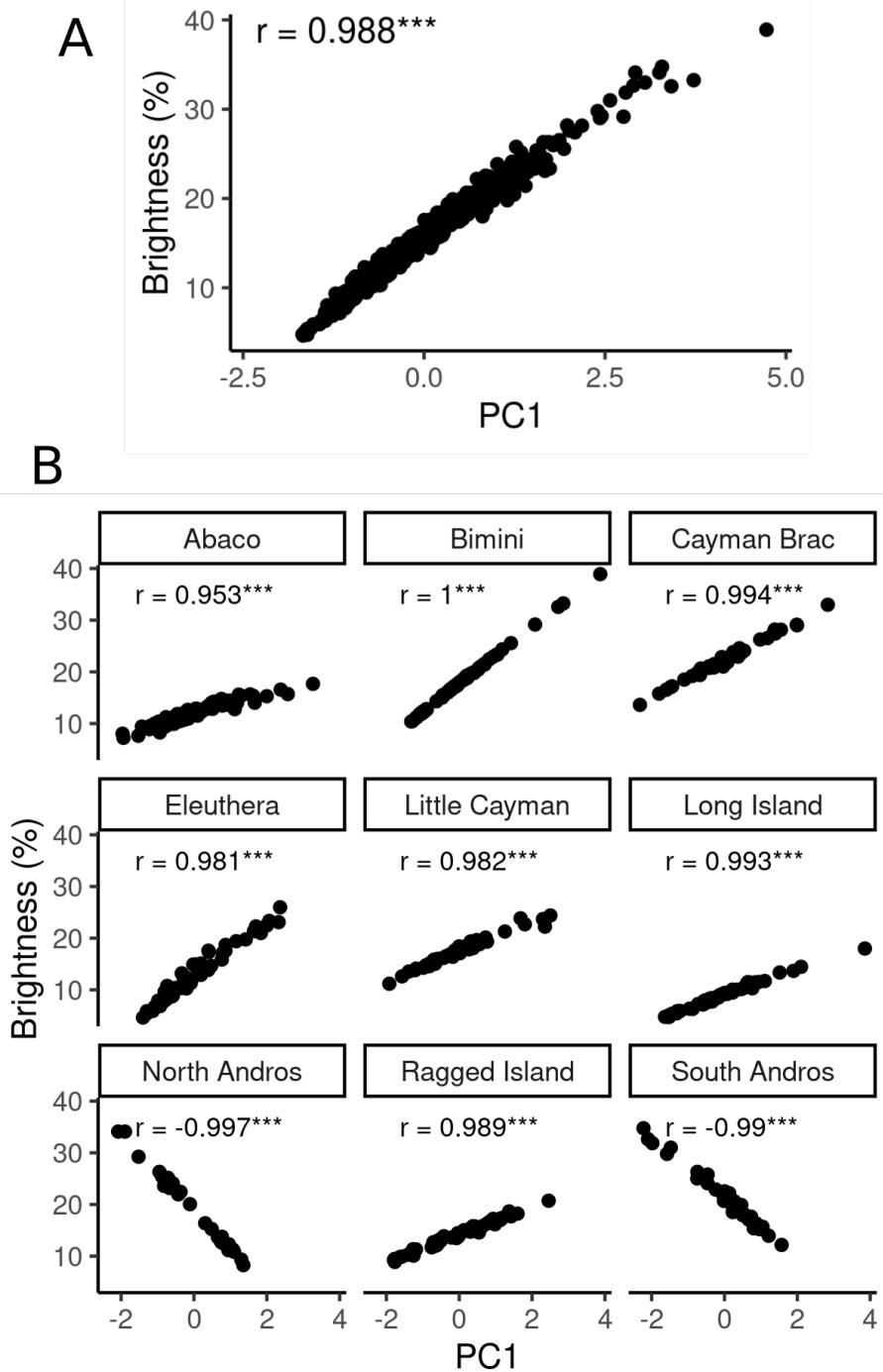


Figure S11: PC1 captures brightness across all islands. (A) Correlation between dewlap brightness (as measured by the mean reflectance from 300 to 700nm in wavelength) and PC1 score across all islands. (B) Correlation between brightness and within-island PC1, for each island. Pearson's correlation coefficients are reported. \*\*\*,  $P < 0.001$ .

## Supplementary Tables

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Table S1: Number of lizards sampled in each habitat on each island.

	coastal	coppice	mangrove
Abaco	41	24	21
Bimini	38	14	15
Cayman Brac	15	18	17
Eleuthera	22	25	9
Little Cayman	17	12	16
Long Island	26	14	13
North Andros	9	9	10
Ragged Island	18	15	17
South Andros	10	9	12

Table S2: Proportion of variance explained by the first four principal components on each island, as well as in the PCA performed on all islands together (last row).

Island	Total	PC1	PC2	PC3	PC4
Abaco	0.906	0.400	0.279	0.147	0.079
Bimini	0.921	0.502	0.208	0.160	0.051
Cayman Brac	0.888	0.438	0.190	0.155	0.105
Eleuthera	0.926	0.490	0.233	0.138	0.066
Little Cayman	0.907	0.441	0.212	0.176	0.078
Long Island	0.925	0.515	0.205	0.161	0.043
North Andros	0.937	0.560	0.170	0.152	0.054
Ragged Island	0.907	0.483	0.226	0.127	0.072
South Andros	0.948	0.488	0.247	0.146	0.067
All islands	0.913	0.473	0.197	0.164	0.079

Table S3: Henze-Zirkler's test of multivariate normality, performed on global principal components (i.e. fitted on data from all islands together) in each habitat and on each island. The number of outlier points detected based on the Mahalanobis distance is reported.  $HZ$ , test statistic. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .

Island	Habitat	Outliers	$HZ$	$P$
Abaco	coastal	0	1.099	0.0027 **
Abaco	coppice	0	1.074	0.0022 **
Abaco	mangrove	0	1.063	0.0023 **
Bimini	coastal	0	1.281	< 0.0001 ***
Bimini	coppice	0	0.850	0.0482 *
Bimini	mangrove	0	1.191	0.0001 ***
Cayman Brac	coastal	0	0.647	0.5311
Cayman Brac	coppice	0	0.701	0.3940
Cayman Brac	mangrove	0	0.657	0.5357
Eleuthera	coastal	0	1.614	< 0.0001 ***
Eleuthera	coppice	0	1.481	< 0.0001 ***
Eleuthera	mangrove	0	0.729	0.1423
Little Cayman	coastal	0	0.624	0.6599
Little Cayman	coppice	0	0.638	0.4867
Little Cayman	mangrove	0	0.873	0.0413 *
Long Island	coastal	0	0.824	0.1468
Long Island	coppice	0	0.923	0.0150 *
Long Island	mangrove	0	0.773	0.1289
North Andros	coastal	0	0.658	0.3174
North Andros	coppice	0	0.763	0.0900
North Andros	mangrove	0	0.668	0.3185
Ragged Island	coastal	0	0.756	0.2268
Ragged Island	coppice	0	0.797	0.1115
Ragged Island	mangrove	0	0.542	0.9022
South Andros	coastal	0	0.660	0.3451
South Andros	coppice	0	0.659	0.3154
South Andros	mangrove	0	0.911	0.0144 *

Table S4: Shapiro-Wilk's test of univariate normality of the standardized residuals from OLS and GLS-ANOVAs performed on each island where significant differences were detected by random forest classification.  $W$ , test statistic. \*,  $P_{adj} < 0.05$ ; \*\*,  $P_{adj} < 0.01$ ; \*\*\*,  $P_{adj} < 0.001$ .

Island	Variable	$W$	$P$	$P_{adj}$
Abaco	PC1	0.961	0.0109	0.0497 *
Abaco	PC2	0.960	0.0089	0.0473 *
Abaco	PC3	0.988	0.5938	0.7308
Abaco	PC4	0.982	0.2709	0.4816
Bimini	PC1	0.890	0.0000	0.0008 ***
Bimini	PC2	0.984	0.5179	0.6906
Bimini	PC3	0.959	0.0281	0.0997
Bimini	PC4	0.980	0.3386	0.5418
Cayman Brac	PC1	0.986	0.8236	0.8785
Cayman Brac	PC2	0.989	0.9299	0.9299
Cayman Brac	PC3	0.934	0.0079	0.0473 *
Cayman Brac	PC4	0.981	0.5926	0.7308
Eleuthera	PC1	0.930	0.0031	0.0330 *
Eleuthera	PC2	0.975	0.3090	0.5204
Eleuthera	PC3	0.981	0.4989	0.6906
Eleuthera	PC4	0.970	0.1788	0.4082
Little Cayman	PC1	0.955	0.0821	0.2539
Little Cayman	PC2	0.982	0.6854	0.7833
Little Cayman	PC3	0.891	0.0005	0.0081 **
Little Cayman	PC4	0.977	0.4858	0.6906
Long Island	PC1	0.937	0.0074	0.0473 *
Long Island	PC2	0.989	0.9039	0.9299
Long Island	PC3	0.971	0.2170	0.4341
Long Island	PC4	0.983	0.6261	0.7420
North Andros	PC1	0.937	0.0952	0.2539
North Andros	PC2	0.978	0.8075	0.8785
North Andros	PC3	0.905	0.0147	0.0587
North Andros	PC4	0.949	0.1913	0.4082
South Andros	PC1	0.941	0.0879	0.2539
South Andros	PC2	0.946	0.1199	0.2952
South Andros	PC3	0.965	0.3950	0.6019
South Andros	PC4	0.957	0.2480	0.4668

Table S5: Locations of the sampling sites across islands, with mean within-island principal component scores per site.

Island	Longitude	Latitude	Habitat	PC1	PC2	PC3	PC4
Abaco	-77.7	26.9	mangrove	1.006	0.117	-0.057	-1.259
Abaco	-77.6	26.9	coastal	0.155	0.532	0.079	-2.046
Abaco	-77.6	26.9	coppice	-0.084	0.033	0.636	-0.652
Abaco	-77.2	26.1	coastal	0.304	-0.093	-0.987	-0.133
Abaco	-77.0	26.3	mangrove	-0.316	-1.224	0.899	0.185
Abaco	-77.0	26.3	coppice	0.092	0.314	0.124	0.483
Abaco	-77.0	26.3	coastal	-0.337	0.750	-0.207	0.540
Bimini	-79.3	25.6	coastal	-0.263	-1.270	-0.185	0.164
Bimini	-79.3	25.7	coastal	0.468	0.083	-0.242	0.011
Bimini	-79.3	25.7	coppice	-0.270	0.171	0.926	-0.124
Bimini	-79.3	25.7	mangrove	-0.447	0.533	-0.288	-0.014
Cayman Brac	-79.9	19.7	coastal	0.483	-0.523	-0.781	0.443
Cayman Brac	-79.8	19.7	mangrove	0.219	0.453	0.810	-0.015
Cayman Brac	-79.8	19.7	coppice	-0.610	0.008	-0.114	-0.355
Eleuthera	-76.3	24.8	coppice	-0.123	-0.857	0.045	0.117
Eleuthera	-76.3	24.8	coastal	-0.520	-0.642	-0.492	-0.538
Eleuthera	-76.3	24.8	mangrove	-0.163	-0.651	0.126	0.685
Eleuthera	-76.2	24.9	coppice	0.391	0.602	0.540	-0.539
Eleuthera	-76.1	24.9	coastal	0.254	1.228	-0.315	0.402
Little Cayman	-80.1	19.7	coppice	-0.621	0.614	-0.179	-0.404
Little Cayman	-80.0	19.7	coastal	0.395	0.355	0.743	0.175
Little Cayman	-80.0	19.7	mangrove	0.047	-0.838	-0.655	0.118
Long Island	-75.2	23.5	mangrove	0.207	0.366	0.484	0.587
Long Island	-75.2	23.4	coastal	-0.097	-0.798	-0.140	-0.347
Long Island	-75.2	23.4	coppice	-0.485	-0.133	0.366	-0.148
Long Island	-75.1	23.4	coastal	0.316	0.317	-0.626	-0.130
North Andros	-77.9	24.8	coastal	-0.098	-0.516	-0.403	0.066
North Andros	-77.8	24.8	coppice	-0.139	0.090	0.519	0.192
North Andros	-77.8	24.7	mangrove	0.213	0.384	-0.104	-0.232
Ragged Island	-75.7	22.2	coppice	-0.269	0.440	-0.088	-0.360
Ragged Island	-75.7	22.2	coastal	0.049	0.076	-0.221	0.029
Ragged Island	-75.7	22.2	mangrove	0.246	-0.273	0.118	0.933
Ragged Island	-75.7	22.2	mangrove	0.099	-0.749	0.588	-0.636
South Andros	-77.6	24.2	mangrove	-0.250	-0.539	0.316	-0.211
South Andros	-77.6	24.1	coppice	0.466	0.657	-0.586	-0.687
South Andros	-77.5	24.1	coastal	-0.120	0.055	0.149	0.872

Table S6: Nonparametric Kruskal-Wallis tests performed on each variable on each island where deviations from normality were detected. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .

Island	Variable	$\chi^2$	df	$P$
Abaco	PC1	0.74	2	0.6924
Abaco	PC2	23.13	2	< 0.0001 ***
Bimini	PC1	7.38	2	0.0250 *
Cayman Brac	PC3	22.46	2	< 0.0001 ***
Eleuthera	PC1	0.29	2	0.8666
Little Cayman	PC3	19.95	2	< 0.0001 ***
Long Island	PC1	10.98	2	0.0041 **

Table S7: Support vector machine classification results. Legend is as per Table 1.

Island	<i>N</i>	Score	<i>P</i>	
Abaco	86	0.581	< 0.0001	***
Bimini	67	0.555	< 0.0001	***
Cayman Brac	50	0.708	< 0.0001	***
Eleuthera	56	0.443	0.0513	
Little Cayman	45	0.711	< 0.0001	***
Long Island	53	0.664	< 0.0001	***
North Andros	28	0.429	0.1039	
Ragged Island	50	0.424	0.0756	
South Andros	31	0.574	0.0040	**

Table S8: Linear discriminant analysis classification results. Legend is as per Table 1.

Island	<i>N</i>	Score	<i>P</i>	
Abaco	86	0.644	< 0.0001	***
Bimini	67	0.585	< 0.0001	***
Cayman Brac	50	0.780	< 0.0001	***
Eleuthera	56	0.425	0.0871	
Little Cayman	45	0.738	< 0.0001	***
Long Island	53	0.574	0.0001	***
North Andros	28	0.414	0.1911	
Ragged Island	50	0.400	0.1259	
South Andros	31	0.587	0.0013	**