SamSrf X – Users guide

This manual replaces the longer cookbook that was included in previous SamSrf versions. You can still find this cookbook in the *SamSrf/Documents* folder. Most of the information in there should be correct but many individual steps described are probably out of date. There is also a separate document introducing the concepts of pRF analysis.

The present manual describes the basic steps for a standard pRF analysis that you can run using the *SamSrfX* GUI. The GUI itself also gives detailed descriptions of the various model parameters you can specify. That should help you figure out more advanced analyses.

More advanced users may still wish to run analyses directly in Matlab and code up batch analysis looping through all data sets etc. This functionality has not changed.

Since SamSrf 10.3, we also added standalone versions of several tools, including a command line interface for running pRF or CF analyses. This can be run via the *SamSrfAnalysis* function. Please refer to the specific user manual for that.

1. To begin using SamSrfX, either launch the SamSrfX standalone app or run SamSrfX in the Matlab command window.

The SamSrfX GUI looks something like this. The menu bar at the top lets you access several functions.

2. For starters, you may want to use the *Working Folder* menu to navigate to the folder where you want the pRF map files to be created.

While this step is not strictly necessary, it makes things more convenient for you when selecting files.

We are assuming a directory structure used by the *SamPenDu* lab in the past, where functional data are kept in a subfolder of the subject's *FreeSurfer* folder. But this is not mandatory either; you could also have functional and structural data in separate places (as in the BIDS structure).

Here, we navigated to *X001/prf* which is a subfolder in our example dataset.

SamSrf X Analysis GUI v9.96 Working Folder ~ Data Files ~ Surf Fo	older ~ Region of Interest ~ Model S	pecification ~ Apertures ~ Hemodynamic Response ~ Connective Fields ~ Model Fitting	— □ X ng ~ Miscellaneous		
Standard_2D_Gaussian_pRF		prf_gaussian_rf(x0,y0,Sigma)	HRF: de Haas canonical		
Algorithm: samsrf_fit_prf					
Parameter Scaled	Positive				
x0			(F)		
y0 ~			(a)		
Sigma			š lo \		
	·		0 5 10 15 20 25 30 Time (s)		
Field	Value		Region of Interest:		
	pRF_Gaussian	**************************************	< None selected >		
Prf_Function	1x1 function_handle	Welcome to the Seriously Annoying MatLab Surfer Analysis Tool! by D.S. Schwarzkopf from the University of Auckland, New Zealand			
Scaling_Factor	Nat		< No files selected >		
TR					
Hrf		***************************************			
Aperture_File	0x0 char	1			
Polar_Search_Space	✓	1			
Param1	1x36 double	2			
Param2	1x29 double	2			
Param3	1x34 double				
Downsample_Predictions					
Noise_Ceiling_Threshold	(
Seed_Fine_Fit	0x0 char				
Coarse_Fit_Only					
Smoothed_Coarse_Fit					
Fine_Fit_Threshold	0.010				
Coarse_Fit_Block_Size	1000				
Only_Positive_Coarse_Fits					
Replace_Bad_Fits					
Coarse_Fit_Percentile	10	0			
Aperture_Mean_Response					
Compressive_Nonlinearity			Subject surf folder:		
			< None selected >		
			Average Normalise Export		

3. Next, we need to select the data. Under the *Data Files* menu, you have several options:

Your functional data should have have been motion corrected and coregistered to the anatomical T1 image already.

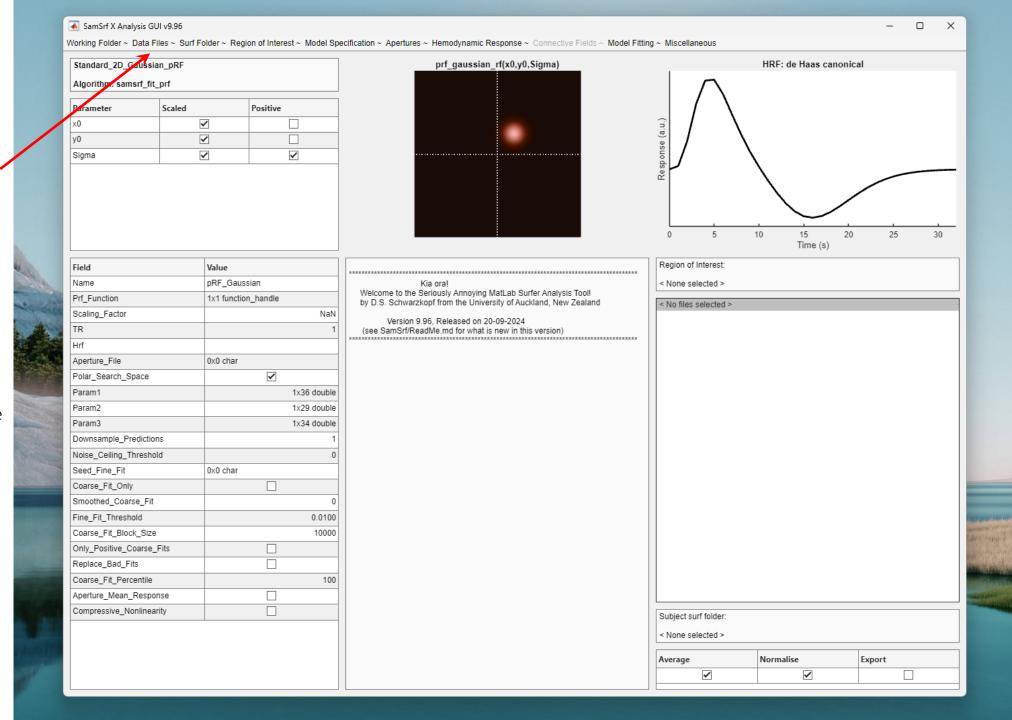
You now have three options:

- Select hemisphere .GII files
- Select bilateral .GII files
- Select volume .NII files

If you choose the bilateral option, a dialog will ask you to specify the *left hemisphere* files, and it will automatically combine these with the corresponding right hemisphere files.

Our functional data files are in *X001/func*.

Note: SamSrfX expects .GII files to be named following the same convention as FreeSurfer files: The prefix Ih and rh indicate left and right hemisphere files, respectively.

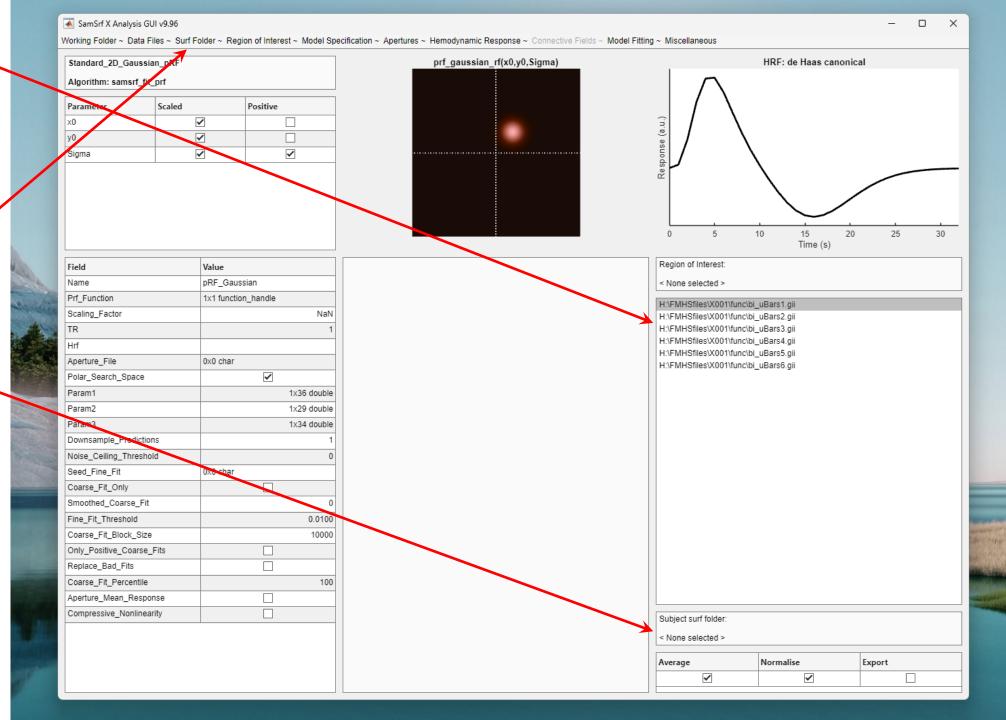


Here, we used the bilateral option. You can now see the *lh_*.GII* files in the data file list.

For surface-based analysis, we also need the *surf* folder of the subject's FreeSurfer recon.

4. Select this via the Surf Folder menu. If your data are structured in the normal SamSrf way, the dialog that opens will already point to the correct place so you just need to confirm.

Once confirmed, the surf folder will appear below the file list.



Many pRF analyses can be rather time-intensive. We therefore often restrict them to a region of interest (ROI), such as a roughly defined occipital region. This is optional and you need to decide which parts of the brain are most important to analyse.

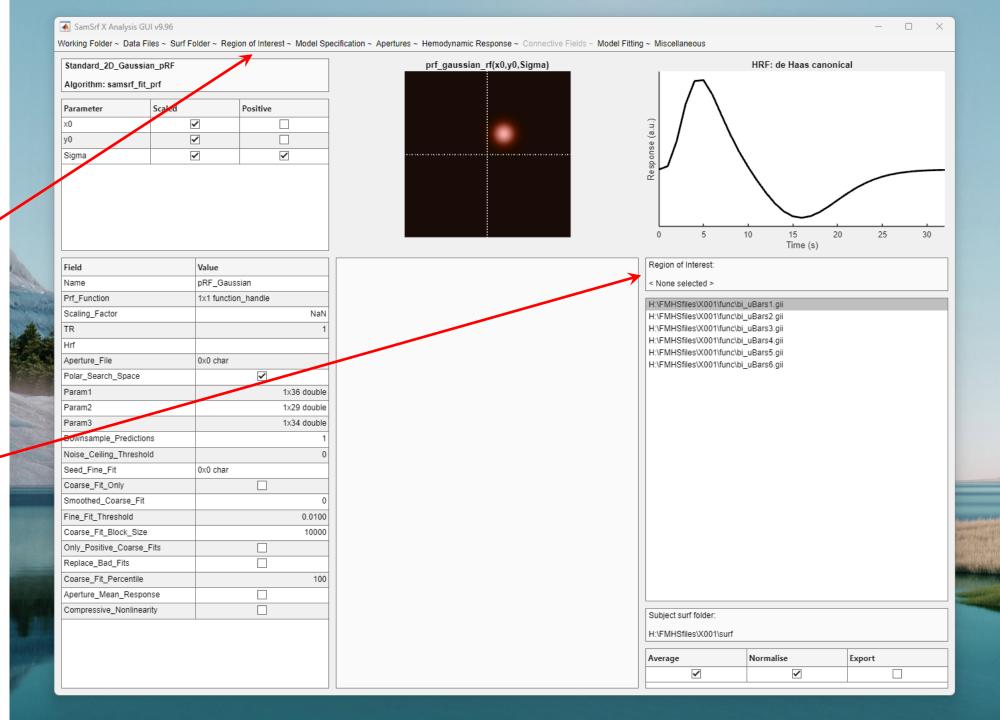
5. You can automatically generate a ROI in the *Region of Interest* menu via *Create Occipital ROIs*.

This will create a occipital ROIs for each hemisphere as well as a combined (bilateral) ROI called occ.label.

Alternatively, you can create your ROIs yourself.

6. Next, use Region of Interest -> Select ROI Label to select the ROI you want to analyse.

Once confirmed, this ROI will then appear above the file list.



Now things get real. You need to specify the model for your analysis. The model contains all the parameters and options of the analysis you want to run.

SamSrfX GUI currently supports three algorithms:

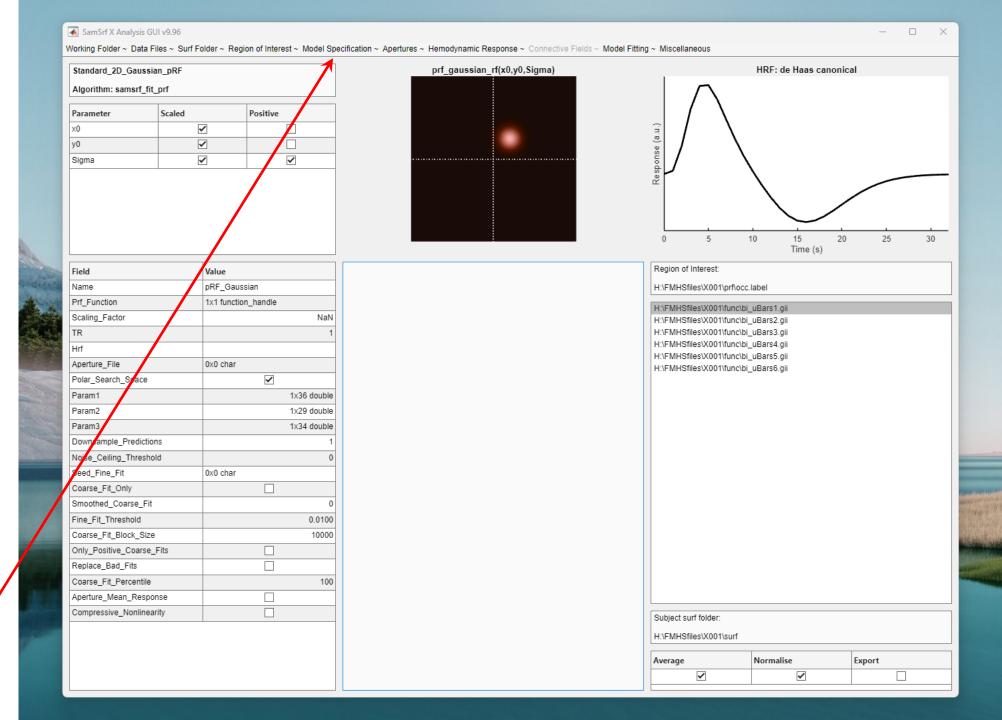
- Forward model pRF
- Reverse correlation pRF
- Reverse correlation CF

Each of these algorithm comes with different model options.

By default, the GUI opens with a standard 2D Gaussian pRF model and for many people there won't be any reason to change this.

However, you may want to use a different algorithm or use another pRF model (e.g. the difference-of-Gaussians or elliptical pRF model).

7. Choose an algorithm or predetermined model using the *Model Specification* menu.



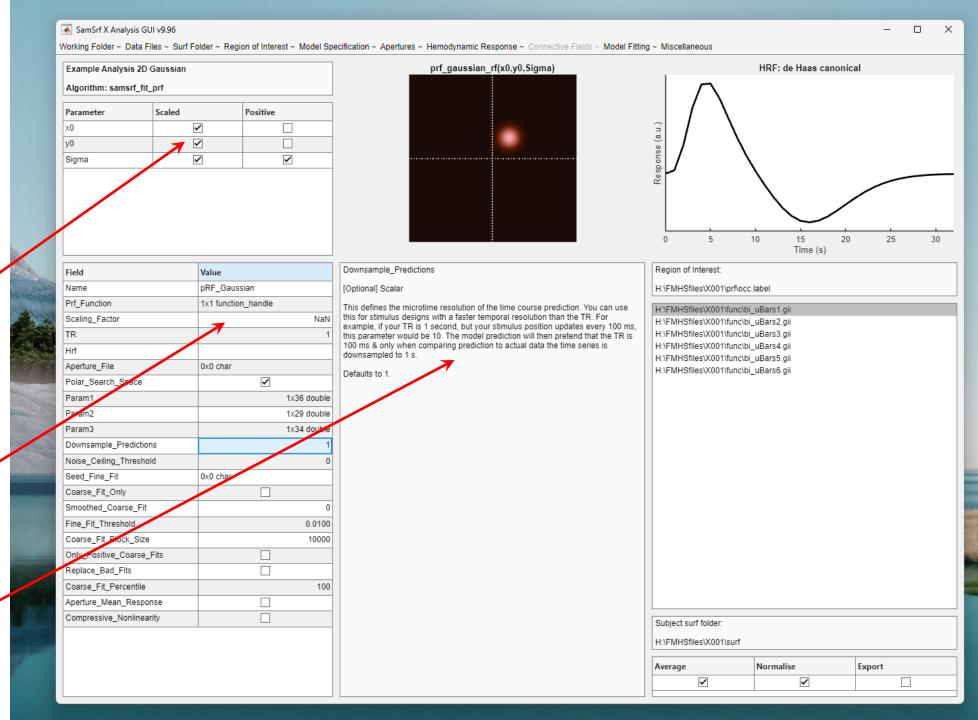
For now, we stick to the standard 2D Gaussian pRF model.

The model parameters/options are listed in the tables on the left side of the GUI.

The top table shows the free parameters of the pRF model and some check boxes for them. You can rename the parameters, but you cannot add or remove parameters here. For this you would need to load another model with a different number of parameters.

Other options are shown in the bottom table. You can edit all of the values, check boxes, and file names in the right column. Some rows cannot be edited, however (function handles and vectors).

8. Clicking on any cells in these tables will show you some detailed descriptions about them in the info field in the centre.



For the conventional Gaussian model, you won't need to tweak many of the parameters.

There are only a few things that are mandatory to define before you can run an analysis:

9. You must define the stimulus apertures corresponding to the sequence in your data. You can select the aperture file either through the *Load Apertures* in the *Apertures* menu or by clicking the cell in the table.

In the menu, View Apertures allows you to visualise the apertures so you can check they are correct.

(Note: In previous versions, the forward modelling pRF algorithm expected vectorised apertures. This step is now done internally so you don't need to worry about it. You can still select vectorised apertures if you want. It won't matter either way.)

Example Analysis 2	PD Gaussian		prf_gaussian_rf(x0,y0,Sigma)	HRF: de Haas canonical		
Algorithm: samsrf_	fit_prf					
Parameter	Scaled	Positive				
x0	✓			(i) \ \		
у0	✓					
Sigma	V	✓		Response (a.u.)		
				0 5 10 15 20 25 Time (s)		
Field	Value			Region of Interest:		
Name	pRF_Gaus	ssian	[Optional] Scalar	H:\FMHSfiles\X001\prf\occ.label		
Prf_Function	1x1 functio					
Scaling_Factor		– N	This defines the microtime resolution of the time course prediction. You can use this for stimulus designs with a faster temporal resolution than the TR. For	H:\FMHSfiles\X001\func\bi_uBars1.gii H:\FMHSfiles\X001\func\bi_uBars2.gii		
TR			example, if your TR is 1 second, but your stimulus position updates every 100 ms, this parameter would be 10. The model prediction will then pretend that the TR is	,		
Hrf			100 ms & only when comparing prediction to actual data the time series is downsampled to 1 s.	H:\FMHSfiles\X001\func\bi_uBars4.gii		
Aperture_File	0xo che			H:\FMHSfiles\X001\func\bi_uBars5.gii H:\FMHSfiles\X001\func\bi_uBars6.gii		
Polar_Search_Space	9	✓	Defaults to 1.	H. IF MINOSINES XXXX TRUITCIDI_ubarsv.gii		
Param1		1x36 doul				
Param2		1x29 doul				
Param3		1x34 doul				
Downsample_Predic	tions					
Noise_Ceiling_Thres	hold					
Seed_Fine_Fit	0x0 char					
Coarse_Fit_Only						
Smoothed_Coarse_F	Fit					
Fine_Fit_Threshold		0.01				
Coarse_Fit_Block_S	ize	100				
Only_Positive_Coars	e_Fits					
Replace_Bad_Fits						
Coarse_Fit_Percenti	le	1				
Aperture_Mean_Res	ponse					
	earity			Subject surf folder:		
				Gubject Suri 1010cl.		
Compressive_Nonlin				H:\FMHSfiles\X001\surf		
				H:\FMHSfiles\X001\surf Average Normalise Export		

The other mandatory parameter you must define is the scaling factor. In 2D retinotopic mapping designs this is the maximum eccentricity of your stimulus.

10. Change the *Scaling_Factor* parameter to the desired value.

This is a critical parameter because the search space for the coarse-fitting stage will depend on this value. It also affects the exclusion criteria for bad pRF fits.

■ SamSrf X Analysis GUI v9.96 Working Folder ~ Data Files ~ Surf Folder ~ Region of Interest ~ Model Specification ~ Apertures ~ Hemodynamic Response ~ Connective Fields ~ Model Fitting ~ Miscellaneous HRF: de Haas canonical Example Analysis 2D Gaussian prf gaussian rf(x0,y0,Sigma) Algorithm: samsrf fit prf Scaled Positive Parameter ~ x0 y0 ~ Sigma ~ **~** 10 20 25 30 Time (s) Downsample_Predictions Region of Interest: Field [Optional] Scalar H:\FMHSfiles\X001\prf\occ.label Prf_Function 1x1 function han This defines the microtime resolution of the time course prediction. You can use H:\FMHSfiles\X001\func\bi uBars1.gii Scaling Factor this for stimulus designs with a faster temporal resolution than the TR. For H:\FMHSfiles\X001\func\bi uBars2.gii example, if your TR is 1 second, but your stimulus position updates every 100 ms, H:\FMHSfiles\X001\func\bi uBars3.gii this parameter would be 10. The model prediction will then pretend that the TR is 100 ms & only when comparing prediction to actual data the time series is H:\FMHSfiles\X001\func\bi uBars4.gii Hrf downsampled to 1 s. H:\FMHSfiles\X001\func\bi_uBars5.gii Aperture_File 0x0 char H:\FMHSfiles\X001\func\bi_uBars6.gii Defaults to 1. Polar Search Space Param1 1x36 double Param2 1x29 double Param3 1x34 double Downsample Predictions Noise_Ceiling_Threshold Seed Fine Fit 0x0 char Coarse_Fit_Only Smoothed Coarse Fit Fine_Fit_Threshold 0.0100 Coarse_Fit_Block_Size 10000 Only_Positive_Coarse_Fits Replace Bad Fits Coarse_Fit_Percentile 100 Aperture_Mean_Response Compressive_Nonlinearity Subject surf folder: H:\FMHSfiles\X001\surf Normalise Export Average

In theory, we are now ready to run the analysis. If you click on the *Model Fitting menu*, it will give you a completeness check of all the mandatory parameters and this should tell you that you are ready.

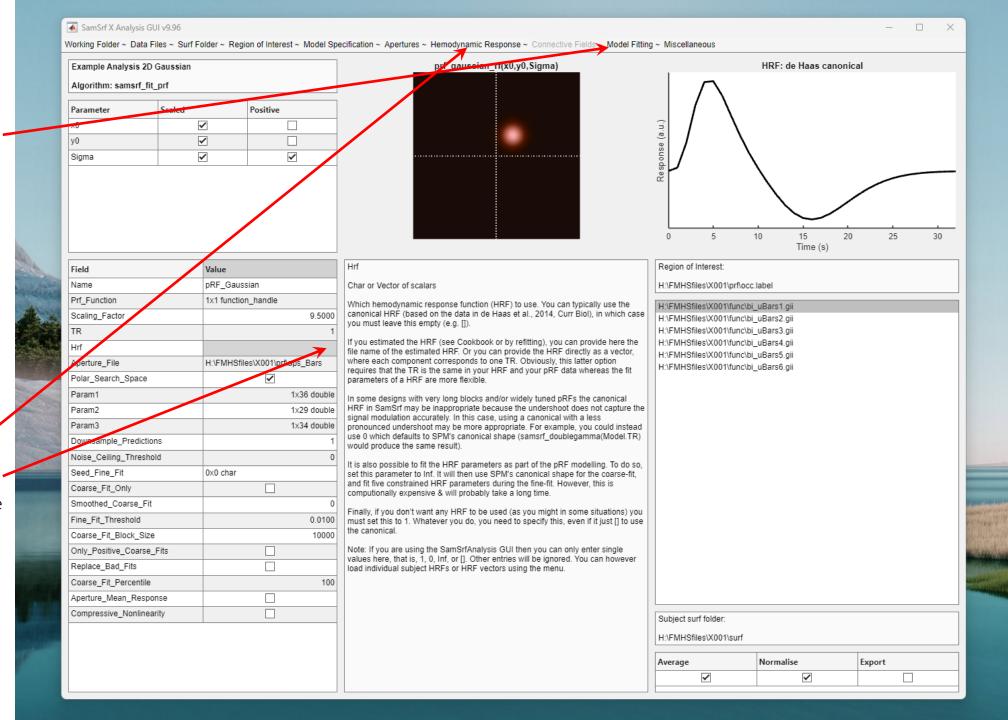
It will also warn you if you didn't define a ROI but that doesn't stop you from running the analysis.

However, there could be many other options you can define. You can also chose a different approach for the hemodynamic response function.

11. Set the HRF, if desired.

You can either change the value in the table (see info field for the available options) or you can load a prefit HRF from a file via the *Hemodynamic Response* menu at the top.

However, we stick with the default, an empty *Hrf* field: *de Haas canonical HRF*



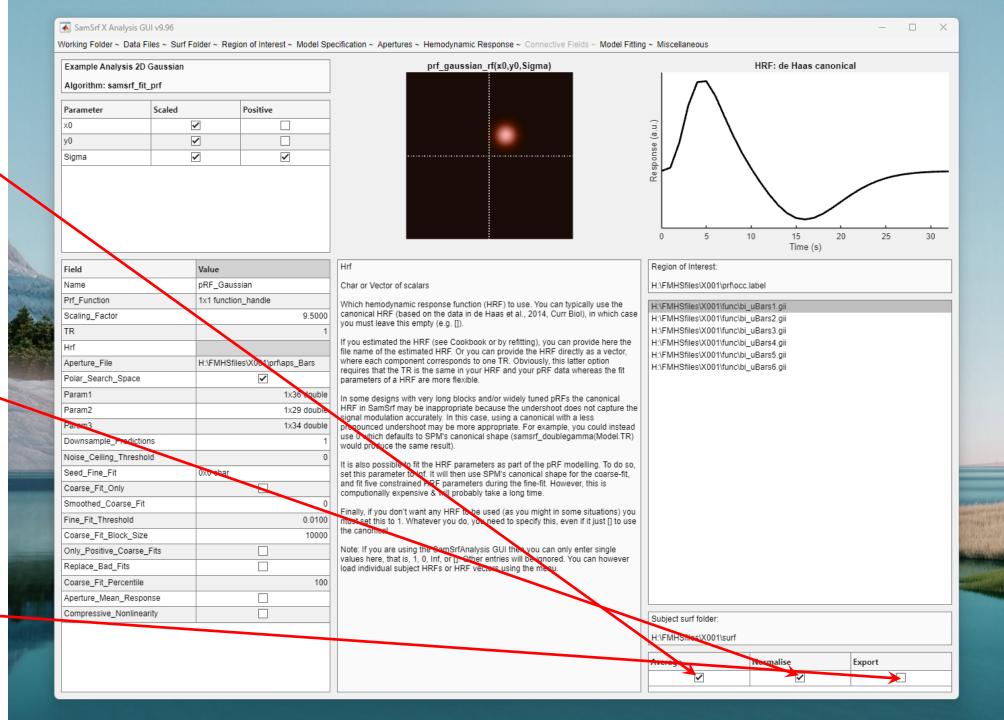
Finally, there are some options for how the data are treated prior to the analysis and what happens after pRF fitting.

- 12. Unchecking the Average box means run will be concatenated instead of being averaged!

 Most conventional pRF studies use the same temporal sequence for stimuli in every run. But if the sequence differs between runs you must concatenate them.
- 13. Usually, the time series in each run is detrended and normalised. Uncheck this box if that is not desired.

pRF maps will be saved in a .MAT file inside the working folder. This format is convenient for many reasons, but you may wish to export data to be used in other tools (e.g. FreeView).

14. Check this box to export data in the same format you use for input (i.e. either .G/l or .N/l).

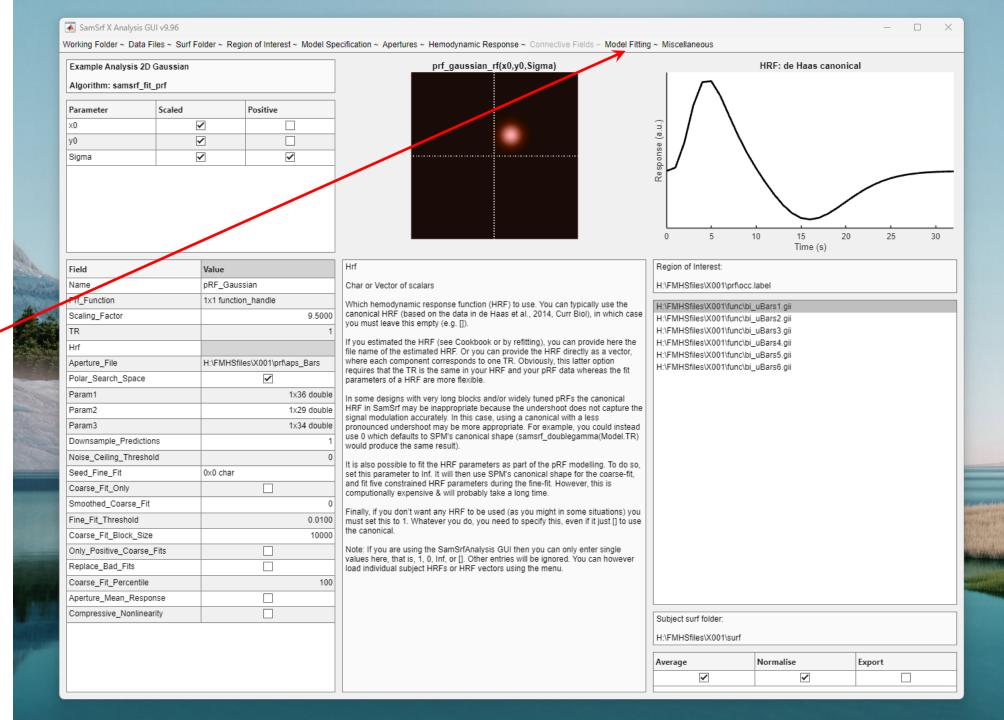


Now you can run the analysis. As already mentioned, when you click on *Model Fitting* it will perform a completeness check that all necessary information has been defined.

Moreover, as the specification is complete now, the *Fit Model* option in that menu should now be available.

15. Click Fit Model to start the analysis. This will then start by converting the .GII files into SamSrf format, in this case as bilateral surface data.

Following that, the actual pRF analysis commences.

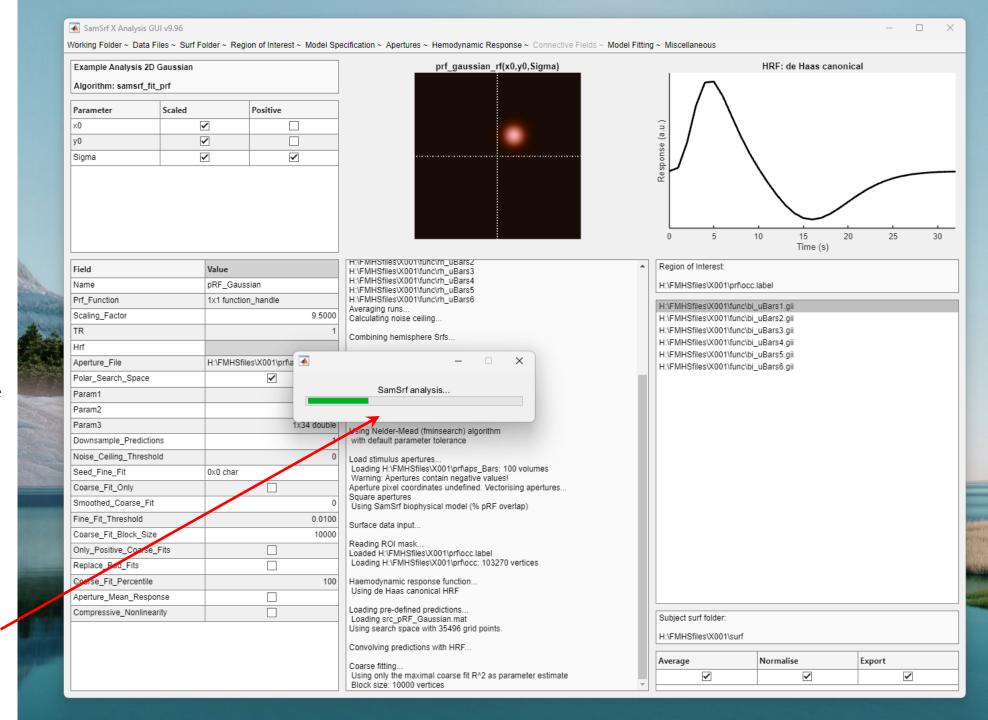


Intermission

Some pRF analysis only takes a few minutes but other can be hours or even days. Just how long it takes depends on:

- How many voxels or vertices you are analysing (so e.g. how large the ROI is)
- How many time points in the time series (therefore concatenated data take longer than averaged)
- Whether you are filtering the data based on SNR ratios
- How many free parameters your model has
- The processing power and amount of system memory

For the protracted phases of the analysis (like fine-fitting pRFs) the progress bar will give you an idea how long it may take...



Either way, if all goes to plan your analysis should eventually be completed.

16. The *Miscellaneous* menu has a few options for you to inspect the data directly in the GUI.

For example, you can use the *Map Display Tool* to look at the surface maps (only if using .GII input data).

You can also use the *Map*Delineation Tool to define
retinotopic brain areas (refer to
the tutorial document on
delineations).

You can warp a normalised retinotopic atlas into native space to create ROIs.

You can use Eccentricity Plots and Visual Field Coverage to visualise pRF data from several regions of interest.

And there are other options (and more may be added in future...)

■ SamSrf X Analysis GUI v Working Folder ~ Data Files		egion of Interest ~ Model Si	pecification ~ Apertures ~ Hemodynamic Response ~ Connective Fields ~ Model	Fittins	~ Miscellaneous		_		
Standard_2D_Gaussian_			prf_gaussian_rf(x0_y0_Sigma) HRF: de Haas can						
Algorithm: samsrf_fit_prf	f								
Parameter Sc	caled	rositive							
x0	~				(in:				
y0	✓) (°				
Sigma	V	✓			Response (a.u.)	10 15 20 Time (s)	25	30	
Field	Value		With default parameter tolerance	•	Region of Interest:				
Name	pRF_Gai	ussian	Load stimulus apertures		H:\FMHSfiles\X001\prf\occ	label			
Prf_Function	1x1 funct	tion_handle	Loading H:\FMHSfiles\X001\pr\aps_Bars: 100 volumes Warning: Apertures contain negative values! Aperture pixel coordinates undefined. Vectorising apertures Square apertures		H:\FMHSfiles\X001\func\bi_uBars1.gii				
Scaling_Factor		9.5000			H:\FMHSfiles\X001\func\bi				
TR		1	Using SamSrf biophysical model (% pRF overlap)	H:\FMHSfiles\X001\func\bi_uBars3.gii					
Hrf			Surface data input		H:\FMHSfiles\X001\func\bi_uBars4.gii H:\FMHSfiles\X001\func\bi_uBars5.gii H:\FMHSfiles\X001\func\bi_uBars6.gii				
Aperture_File	H:\FMHS	Sfiles\X001\prf\aps_Bars	Reading ROI mask						
Polar_Search_Space		✓	Loaded H:\FMHSfiles\X001\prf\occ.label Loading H:\FMHSfiles\X001\prf\occ: 10 vertices						
Param1		1x36 double							
Param2		1x29 double	Haemodynamic response function Using de Haas canonical HRF						
Param3		1x34 double	Loading pre-defined predictions						
Downsample_Predictions		1	Loading src_pRF_Gaussian.mat						
Noise_Ceiling_Threshold		0							
Seed_Fine_Fit	0x0 char		Convolving predictions with HRF						
Coarse_Fit_Only			Coarse fitting						
Smoothed_Coarse_Fit		0	Using only the maximal coarse fit R^2 as parameter estimate Block size: 10000 vertices						
Fine_Fit_Threshold		0.0100	Coarse fitting completed in 0.096374 minutes.						
Coarse_Fit_Block_Size		10000							
Only_Positive_Coarse_Fits	\$		Parallel computing!						
Replace_Bad_Fits			Fine fitting completed in 0.006886 hours.						
Coarse_Fit_Percentile		100							
Aperture_Mean_Response									
Compressive_Nonlinearity			Tidying up final results structure Computed normalised R^2		Subject surf folder:				
			Compressing surface data file Saving pRF fitting results Saved bi_pRF_Gaussian.mat		H:\FMHSfiles\X001\surf	Normalise	Export		
			Whole analysis completed in 0.0074022 hours.	~	✓	✓			

Finally, once you specified a model, and defined all the files for data and ROIs etc, you may run the exact same analysis for another participant.

17. Use the *Replace String* option under the *Model Fitting* menu to quickly replace any mention of the current subject's pathname to a new one.

For example, here we might want to replace X001 with X002. It will do this for all the data files, for the ROI and surf folders, and any mention of X001 in the table.

Note: This will also replace the string in the working folder!

The idea is that you'll be in the same subfolder for the subject, so if we were in X001/prf before we should now be in X002/prf.

This also means you must be consistent in your naming!
Don't name the folders X001 but some files S01 or some such.

Example Analysis 2D Gaus	sian		prf_gaussian_rf(x0,y0,Sigma)	HF	RF: de Haas canoni	cal	
Algorithm: samsrf_fit_prf							
Parameter Sca	led Positive	1					
к0	✓			(a.u.)			
y0	V						
Sigma	V	✓		sb onse			
				0 5 10	15 20 Time (s)	25	
Field	Value		Hrf	Region of Interest:			
Name	pRF_Gaussian		Char or Vector of scalars	H:\FMHSfiles\X001\prf\occ.labe	ı		
Prf_Function	1x1 function_handle		Which hemodynamic response function (HRF) to use. You can typically use the	U-\EMUQfilos\V004\func\hi_uPo	aro1 dii		
Scaling_Factor		9.5000	canonical HRF (based on the data in de Haas et al., 2014, Curr Biol), in which case				
TR	1		you must leave this empty (e.g. []).	H:\FMHSfiles\X001\func\bi_uBars3.gii			
Hrf			If you estimated the HRF (see Cookbook or by refitting), you can provide here the file name of the estimated HRF. Or you can provide the HRF directly as a vector,	H:\FMHSfiles\X001\func\bi_uBa			
Aperture_File	H:\FMHSfiles\X001\p	orf\aps_Bars	where each component corresponds to one TR. Obviously, this latter option	H:\FMHSfiles\X001\func\bi_uBars5.gii H:\FMHSfiles\X001\func\bi_uBars6.gii			
Polar_Search_Space	✓		requires that the TR is the same in your HRF and your pRF data whereas the fit parameters of a HRF are more flexible.	The wind industrial in			
Param1		1x36 double	In some designs with very long blocks and/or widely tuned pRFs the canonical				
Param2		1x29 double	HRF in SamSrf may be inappropriate because the undershoot does not capture the				
Param3		1x34 double					
Downsample_Predictions		-	use 0 which defaults to SPM's canonical shape (samsrf_doublegamma(Model.TR) would produce the same result).				
Noise_Ceiling_Threshold		(
Seed_Fine_Fit	0x0 char		 It is also possible to fit the HRF parameters as part of the pRF modelling. To do so, set this parameter to Inf. It will then use SPM's canonical shape for the coarse-fit, 				
Coarse_Fit_Only			and fit five constrained HRF parameters during the fine-fit. However, this is computionally expensive & will probably take a long time.				
Smoothed_Coarse_Fit		(
Fine_Fit_Threshold		0.0100	made dot time to 1. Tribatorol you do, you need to openly time, even in tribate [] to do				
Coarse_Fit_Block_Size		10000	the canonical.				
Only_Positive_Coarse_Fits			Note: If you are using the SamSrfAnalysis GUI then you can only enter single	er			
Replace_Bad_Fits			values here, that is, 1, 0, Inf, or []. Other entries will be ignored. You can however load individual subject HRFs or HRF vectors using the menu.				
Coarse_Fit_Percentile		100					
Aperture_Mean_Response							
				Subject surf folder:			
Compressive_Nonlinearity			- 1				
Compressive_Nonlinearity				H:\FMHSfiles\X001\surf			
Compressive_Nonlinearity					rmalise	Export	

18. Related to the previous point, you can use *Run Batch Analysis* to automatically run the same analysis on a set of subjects.

A SamSef V Analysis GIII v0 06

Aperture_Mean_Response

Compressive_Nonlinearity

This will internally replace all the instances of the subject ID with the other ones in the batch. For example, here all instances of X001 would be replaced with X002, X003, etc.

Note: It is therefore crucial that you start this analysis in the working folder of one subject in the batch! All the names must be consistent.

For example, you could specify your model in the working folder *X001/prf*, and then run the batch on X001, X002, X003, etc.

Example Analysis	ZD Gaussian			prf_gaussian_rf(x0,y0,Sigma)	HRF: de Haas canonical			
Algorithm: samsrf	f_fit_prf							
Parameter Scaled		Positive						
к0					(a) /			
y0								
Sigma	5		V		0 5 10 15 20 25 30			
Field		Value		Нц	Time (s) Region of Interest:			
Name Prf_Function		pRF_Gaussian		Char or Vector of scalars Which hemodynamic response function (HRF) to use. You can typically use the canonical HRF (based on the data in de Haas et al., 2014, Curr Biol), in which case you must leave this empty (e.g. []).	H:\FMHSfiles\X001\prf\occ.label			
		1x1 function_handle			H:\FMHSfiles\X001\func\bi_uBars1.gii			
Scaling_Factor		9.5000			H:\FMHSfiles\X001\func\bi_uBars2.gii			
TR			1	1,71,2 11	H:\FMHSfiles\X001\func\bi_uBars3.gii			
Hrf				If you estimated the HRF (see Cookbook or by refitting), you can provide here the file name of the estimated HRF. Or you can provide the HRF directly as a vector,	H:\FMHSfiles\X001\func\bi_uBars4.gii H:\FMHSfiles\X001\func\bi_uBars5.gii			
Aperture_File		H:\FMHSfile	s\X001\prf\aps_Bars	where each component corresponds to one TR. Obviously, this latter option requires that the TR is the same in your HRF and your pRF data whereas the fit	H:\FMHSfiles\X001\func\bi_uBars6.gii			
Polar_Search_Space	ce		✓	parameters of a HRF are more flexible.				
Param1			1x36 double	In some designs with very long blocks and/or widely tuned pRFs the canonical				
Param2			1x29 double	HRF in SamSrf may be inappropriate because the undershoot does not capture the signal modulation accurately. In this case, using a canonical with a less				
Param3			1x34 double	pronounced undershoot may be more appropriate. For example, you could instead use 0 which defaults to SPM's canonical shape (samsrf_doublegamma(Model.TR)				
Downsample_Predi	ictions		1	would produce the same result).				
Noise_Ceiling_Thre	eshold		0	It is also possible to fit the HRF parameters as part of the pRF modelling. To do so,				
Seed_Fine_Fit		0x0 char		set this parameter to Inf. It will then use SPM's canonical shape for the coarse-fit,				
Coarse_Fit_Only				and fit five constrained HRF parameters during the fine-fit. However, this is computionally expensive & will probably take a long time.				
Smoothed_Coarse_	_Fit		0	Finally, if you don't want any HRF to be used (as you might in some situations) you				
Fine_Fit_Threshold			0.0100	must set this to 1. Whatever you do, you need to specify this, even if it just [] to use				
Coarse_Fit_Block_	Size		10000	the canonical.				
Only_Positive_Coa	rse_Fits			Note: If you are using the SamSrfAnalysis GUI then you can only enter single values here, that is, 1, 0, Inf. or []. Other entries will be ignored. You can however				
Replace_Bad_Fits				load individual subject HRFs or HRF vectors using the menu.				
Coarse_Fit_Percen	tile		100					

Subject surf folder:

H:\FMHSfiles\X001\surf

Average	Normalise	Export	
✓	✓		

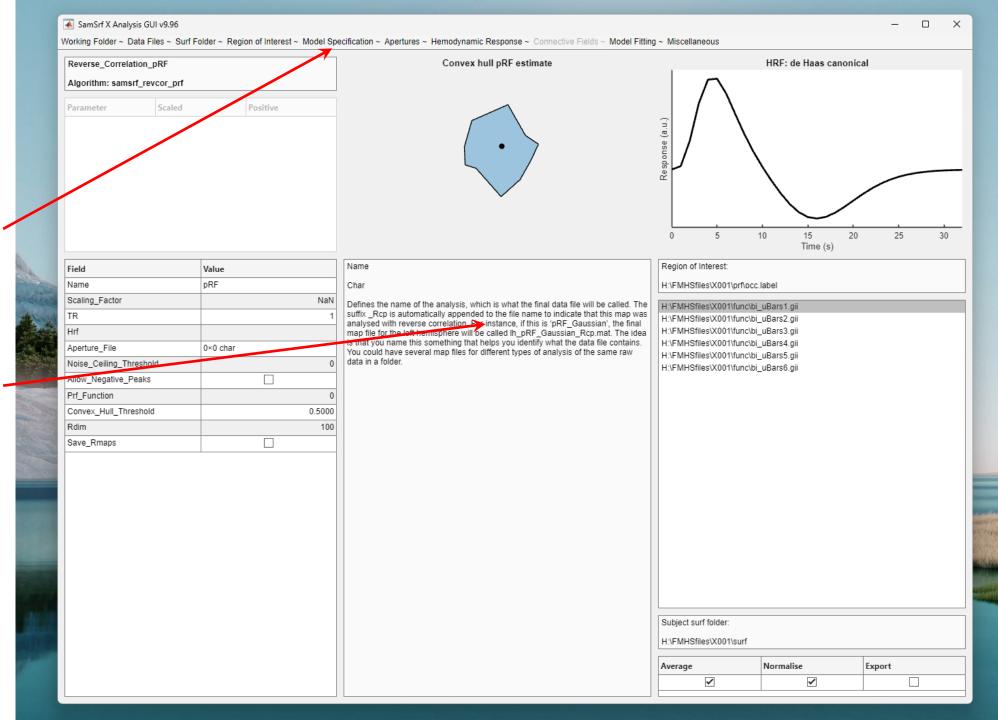
About other analyses

As already explained, there are other algorithms and many model options you can define for each of these.

For example, you could choose the reverse correlation pRF algorithm instead. This has other model options you can define, and the GUI will accordingly look somewhat different.

This manual has no information about these but you can get some more explanation via the *Model Information* option in the *Model Specification* menu.

Finally, you can save and load previously saved model specifications. These files include the data files and ROI labels you defined so you don't have to do this every time. For use with batch analysis, you want to save these .MOD files in the working folder of your subject.



Final Remarks

Default FreeSurfer template

You may want to change the default folder pointing to your *fsaverage* template. You can change this path in the file *SamSrf_defaults.json* in your *SamSrfX* folder. Under a standard *FreeSurfer* installation this will already be correct, but your setup may differ. Obviously, this step only applies if you even use *FreeSurfer*. Moreover, it is only required for optional analyses that involve normalised brain templates.

Default colour schemes

You can also change the default colour schemes for various data types in your maps in the same .JSON file. Colour schemes are defined as .CSV files inside the *Colours* subfolder. You can add or make your own colour schemes. These are simply three-column matrices with the RGB values for each step in the colour map.

Other default settings

There are other default settings (e.g. how much of the brain the Map Display Tool will show). Future versions of this manual may contain more information about these.

Questions / Comments?

SamSrfX is developed by Sam Schwarzkopf at the University of Auckland, New Zealand, with support from his students and collaborators. We strive to make this a user-friendly, easy-to-use tool, making it comparably straightforward for other researchers to run pRF and CF analyses. However, we receive no dedicated funding for this work, and this is not commercial product. Primarily, we developed these methods for our own research. Our capacity to support and troubleshoot other people's research is therefore limited. We cannot guarantee that SamSrf does everything you want or that it is free of bugs and problems.

Nevertheless, please contact Sam if you have questions or suggestions. He or someone from his team will usually reply and – time permitting – we may also be interested in a closer collaboration:

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