

six_donor_DGE_limma

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```
library(limma)
library(edgeR)
library(biomaRt)
library(ggplot2)
library(ggrepel)
library(tidyr)
library(RUVSeq)
library(clusterProfiler)

library(here)

## here() starts at /projects/imb-pkbphil/sp/rnaseq/six_donor_trans/splicing_paper
i_am("R/01_six_donor_DGE_limma.Rmd")

## here() starts at /projects/imb-pkbphil/sp/rnaseq/six_donor_trans/splicing_paper
knitr::opts_chunk$set(echo = TRUE, dev = c("pdf"), fig.path = "plots/six_donor_DGE_limma/")

Load experiment 1; select relevant samples

la = read.delim(here("02featureCounts/late_adipo_s4_s6.nommm.feature.counts"), skip=1)
colnames(la) = gsub("output.01hisat.", "", gsub(".sorted.bam", "", colnames(la)))

# select the bulk day15 samples
la = la[,c(1:6,grep("^19|2[01234]", colnames(la)))]
head(la)

##          Geneid      Chr
## 1 ENSG00000223972.5 chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1
## 2 ENSG00000227232.5 chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1
## 3 ENSG00000278267.1           chr1
## 4 ENSG00000243485.5           chr1;chr1;chr1;chr1;chr1
## 5 ENSG00000284332.1           chr1
## 6 ENSG00000237613.2           chr1;chr1;chr1;chr1;chr1
##                                     Start
## 1                 11869;12010;12179;12613;12613;12975;13221;13221;13453
## 2 14404;15005;15796;16607;16858;17233;17606;17915;18268;24738;29534
## 3                               17369
```

```

## 4                               29554;30267;30564;30976;30976
## 5                               30366
## 6                               34554;35245;35277;35721;35721
##                                         End
## 1           12227;12057;12227;12721;12697;13052;13374;14409;13670
## 2 14501;15038;15947;16765;17055;17368;17742;18061;18366;24891;29570
## 3                                         17436
## 4                               30039;30667;30667;31109;31097
## 5                                         30503
## 6                               35174;35481;35481;36073;36081
##             Strand Length 19.21423_S40 20.21424_S47 21.21425_S54
## 1     +;+;+;+;+;+;+;+    1735      0      0      0
## 2   -;-;-;-;-;-;-;-;-    1351      5     15     27
## 3           -       68      6      6      5
## 4     +;+;+;+;+    1021      0      0      0
## 5           +     138      0      0      0
## 6   -;-;-;-;-    1219      1      2      1
## 22.21426_S3 23.21427_S10 24.21428_S17
## 1         0      0      0
## 2         13     13     37
## 3         8      3      9
## 4         0      0      0
## 5         0      0      0
## 6         0      0      0

```

Load experiment 2; select relevant samples

```

fc = read.delim(here("02featureCounts/beige_rnaseq.nommm.feature.counts"), skip=1)
colnames(fc) = gsub("output.01hisat.", "", gsub(".sorted.bam", "", colnames(fc)))
head(fc)[1:9]

```

```

##          Geneid                         Chr
## 1 ENSG00000223972.5      chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1
## 2 ENSG00000227232.5  chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1
## 3 ENSG00000278267.1                         chr1
## 4 ENSG00000243485.5      chr1;chr1;chr1;chr1;chr1
## 5 ENSG00000284332.1                         chr1
## 6 ENSG00000237613.2      chr1;chr1;chr1;chr1;chr1
##                                         Start
## 1           11869;12010;12179;12613;12613;12975;13221;13221;13453
## 2 14404;15005;15796;16607;16858;17233;17606;17915;18268;24738;29534
## 3                                         17369
## 4                               29554;30267;30564;30976;30976
## 5                                         30366
## 6                               34554;35245;35277;35721;35721
##                                         End
## 1           12227;12057;12227;12721;12697;13052;13374;14409;13670
## 2 14501;15038;15947;16765;17055;17368;17742;18061;18366;24891;29570
## 3                                         17436
## 4                               30039;30667;30667;31109;31097
## 5                                         30503
## 6                               35174;35481;35481;36073;36081
##             Strand Length 1.22589_S146 2.22590_S149 3.22591_S154

```

```

## 1      +;+;+;+;+;+;+;+;+  1735      0      0      0
## 2 -;-;-;-;-;-;-;-;-;-  1351     323    193    171
## 3          -       68      45      39      26
## 4      +;+;+;+;+  1021      0      0      0
## 5          +     138      0      0      0
## 6      -;-;-;-;-  1219      0      0      0

```

Match experiments

```
nrow(la); nrow(fc)
```

```
## [1] 60668
```

```
## [1] 60668
```

```

counts = merge(fc, la, by=colnames(fc)[1:6])
#head(counts)
colnames(counts)

```

```

## [1] "Geneid"         "Chr"           "Start"        "End"
## [5] "Strand"         "Length"        "1.22589_S146" "2.22590_S149"
## [9] "3.22591_S154"   "4.22592_S128"  "5.22593_S132"  "6.22594_S136"
## [13] "7.22595_S140"   "8.22596_S143"  "9.22597_S147"  "10.22598_S150"
## [17] "11.22599_S155"  "12.22600_S129" "13.22601_S133" "14.22602_S137"
## [21] "15.22603_S141"  "16.22604_S144" "17.22605_S148" "18.22606_S151"
## [25] "19.22607_S156"  "20.22608_S130" "21.22609_S134" "22.22610_S138"
## [29] "23.22611_S142"  "24.22612_S145" "25.22613_S153" "26.22614_S152"
## [33] "27.22615_S157"  "28.22616_S131" "29.22617_S135" "30.22618_S139"
## [37] "19.21423_S40"   "20.21424_S47"  "21.21425_S54"  "22.21426_S3"
## [41] "23.21427_S10"   "24.21428_S17"

```

```
nrow(counts)
```

```
## [1] 60668
```

Save this to file for GEO submission later. [Reanalysis of s4/s6 white]

```
write.table(counts, here("02featureCounts/six_donors.merged.nomm.counts"), sep="\t", quote=F, row.names=F)
```

formatting

```

counts$Geneid = gsub("\\..*", "", counts$Geneid)
counts[counts$Geneid == "ENSG00000132170", 7:ncol(counts)] # double check gene matching

```

```

##      1.22589_S146 2.22590_S149 3.22591_S154 4.22592_S128 5.22593_S132
## 6545      11069      13502      17008      7451      14564
##      6.22594_S136 7.22595_S140 8.22596_S143 9.22597_S147 10.22598_S150
## 6545      8291      11652      14165      17663      20856
##      11.22599_S155 12.22600_S129 13.22601_S133 14.22602_S137 15.22603_S141
## 6545      12708      10109      16121      17043      12563

```

```

##      16.22604_S144 17.22605_S148 18.22606_S151 19.22607_S156 20.22608_S130
## 6545      7407       8276       16290       10702       12934
## 21.22609_S134 22.22610_S138 23.22611_S142 24.22612_S145 25.22613_S153
## 6545     13346      10671      9431       12668       12939
## 26.22614_S152 27.22615_S157 28.22616_S131 29.22617_S135 30.22618_S139
## 6545     7333       5670       8552       5035       9837
## 19.21423_S40 20.21424_S47 21.21425_S54 22.21426_S3 23.21427_S10
## 6545     10822      12847      13837      8180       16506
## 24.21428_S17
## 6545     18386

```

Load info

```

info = readxl::read_xlsx(here("sample_info/publication_ids.xlsx"))
info$frac_assigned_to_genes = as.double(gsub("%", "", info$percent_assigned_to_genes))/100
info$donor.condition = paste(info$donor, info$condition, sep=".")  

head(info)

```

```

## # A tibble: 6 x 11
##   sample      time    rep donor condition name      dataset fc_file
##   <chr>      <chr> <dbl> <chr>   <chr>   <chr>   <chr>   <chr>
## 1 19-21423_S40 day15     1 subject4 white  day15_subject4_wh~ late_a~ beige_~
## 2 20-21424_S47 day15     2 subject4 white  day15_subject4_wh~ late_a~ beige_~
## 3 21-21425_S54 day15     3 subject4 white  day15_subject4_wh~ late_a~ beige_~
## 4 22-21426_S3  day15     1 subject6 white  day15_subject6_wh~ late_a~ beige_~
## 5 23-21427_S10 day15     2 subject6 white  day15_subject6_wh~ late_a~ beige_~
## 6 24-21428_S17 day15     3 subject6 white  day15_subject6_wh~ late_a~ beige_~
## # i 3 more variables: percent_assigned_to_genes <chr>,
## #   frac_assigned_to_genes <dbl>, donor.condition <chr>

```

```

counts$Geneid = gsub("\\..*", "", counts$Geneid)
counts[counts$Geneid == "ENSG00000132170", 7:ncol(counts)] # double check gene matching

```

```

##      1.22589_S146 2.22590_S149 3.22591_S154 4.22592_S128 5.22593_S132
## 6545     11069       13502       17008       7451       14564
## 6.22594_S136 7.22595_S140 8.22596_S143 9.22597_S147 10.22598_S150
## 6545     8291       11652       14165       17663       20856
## 11.22599_S155 12.22600_S129 13.22601_S133 14.22602_S137 15.22603_S141
## 6545     12708       10109       16121       17043       12563
## 16.22604_S144 17.22605_S148 18.22606_S151 19.22607_S156 20.22608_S130
## 6545     7407       8276       16290       10702       12934
## 21.22609_S134 22.22610_S138 23.22611_S142 24.22612_S145 25.22613_S153
## 6545     13346      10671      9431       12668       12939
## 26.22614_S152 27.22615_S157 28.22616_S131 29.22617_S135 30.22618_S139
## 6545     7333       5670       8552       5035       9837
## 19.21423_S40 20.21424_S47 21.21425_S54 22.21426_S3 23.21427_S10
## 6545     10822      12847      13837      8180       16506
## 24.21428_S17
## 6545     18386

```

Make object

```

rownames(info) = gsub("-", ".", info$sample)

## Warning: Setting row names on a tibble is deprecated.

info = info[colnames(counts)[7:ncol(counts)],] #make sure order is the same
summary(rownames(info) == colnames(counts)[7:ncol(counts)])

```

```

##      Mode   FALSE
## logical      36

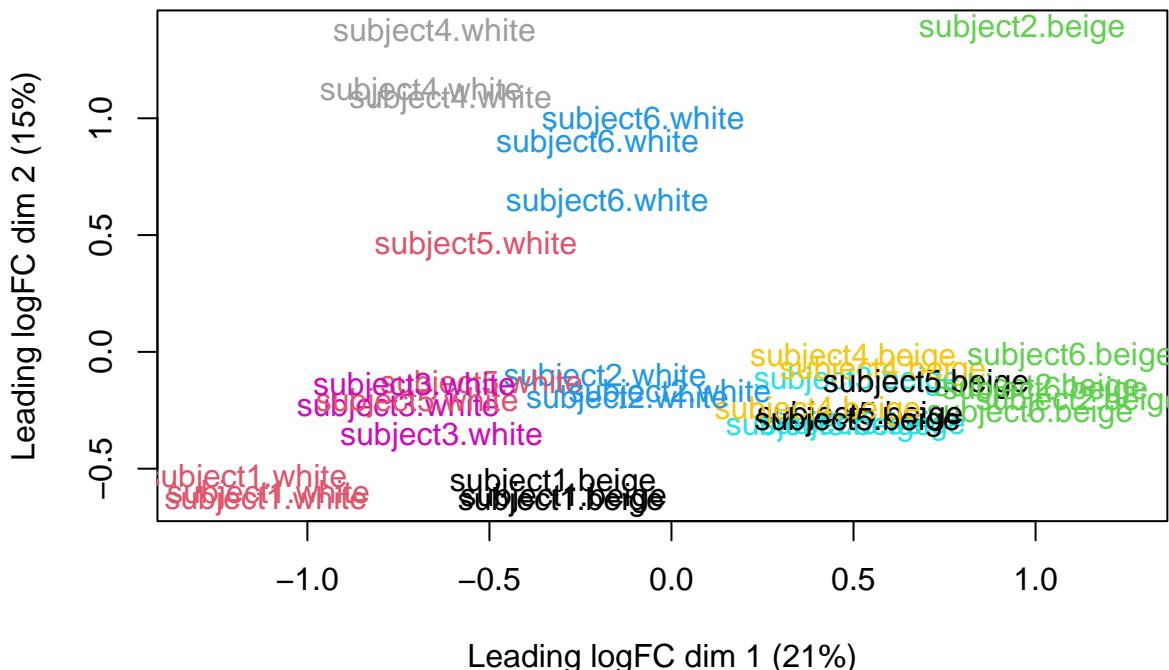
ob = DGEList(counts = data.matrix(counts[7:ncol(counts)]),
             samples = info,
             group = info$donor.condition,
             genes = counts[c(1,6)])
rownames(ob$counts) = ob$genes$Geneid
#head(ob)

summary(ob$counts[grep("ENSG00000132170", rownames(ob$counts)),c(1,5:ncol(ob$counts))]) ##PPARG check

##      Min. 1st Qu. Median     Mean 3rd Qu.    Max.
##      5035    9431   12563   12075   14165   20856

plotMDS(ob, top = 5000, labels = ob$samples$group, col = c(1:12)[ob$samples$group])

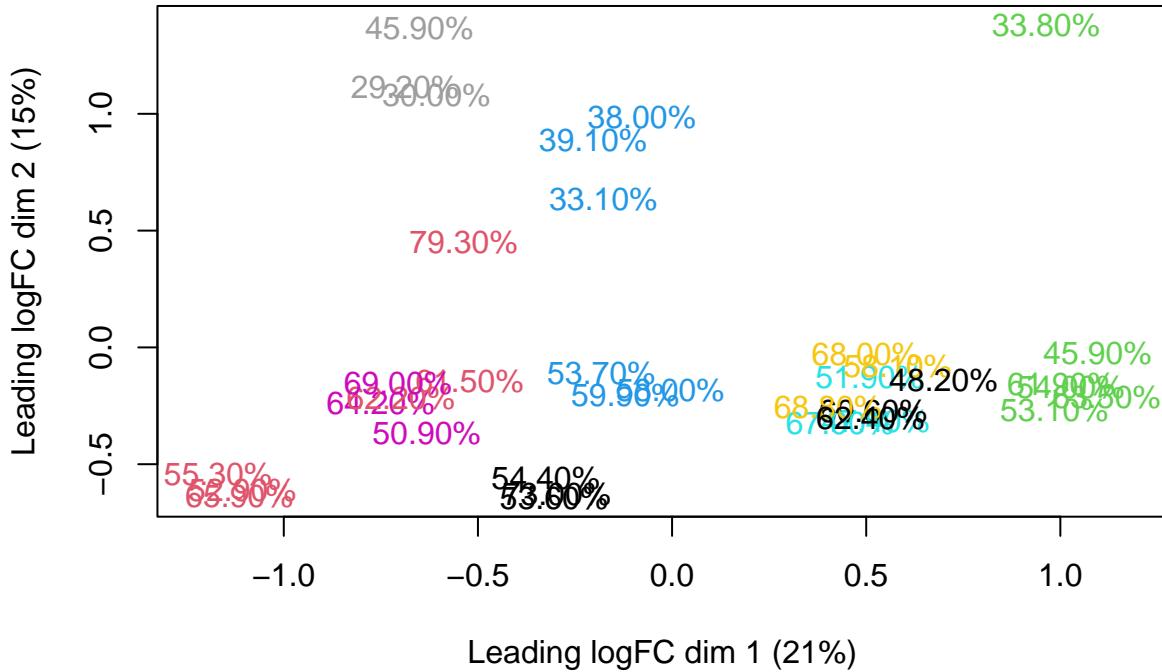
```



```

library(RColorBrewer)
to_col = colorRampPalette(c("blue","red"))(25)
plotMDS(ob, top = 5000, labels = ob$samples$percent_assigned_to_genes, col = c(1:12)[ob$samples$group])

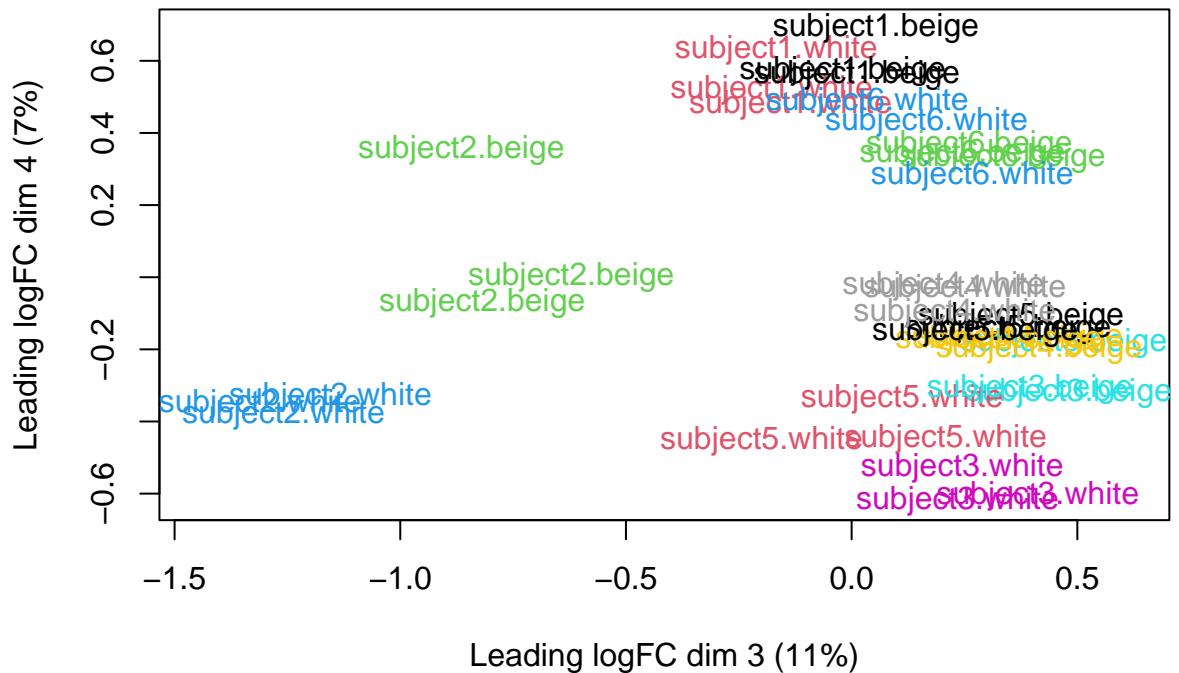
```



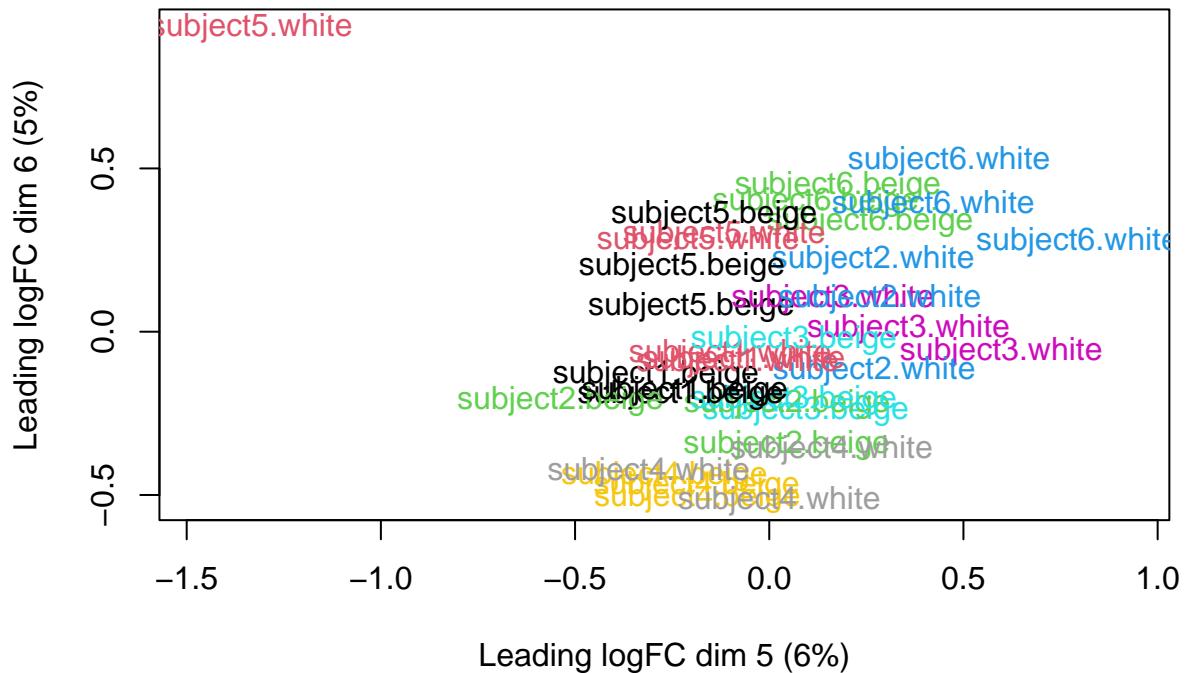
```

plotMDS(ob, top = 5000, labels = ob$samples$group, col = c(1:12)[ob$samples$group] ,
dim.plot = c(3,4))

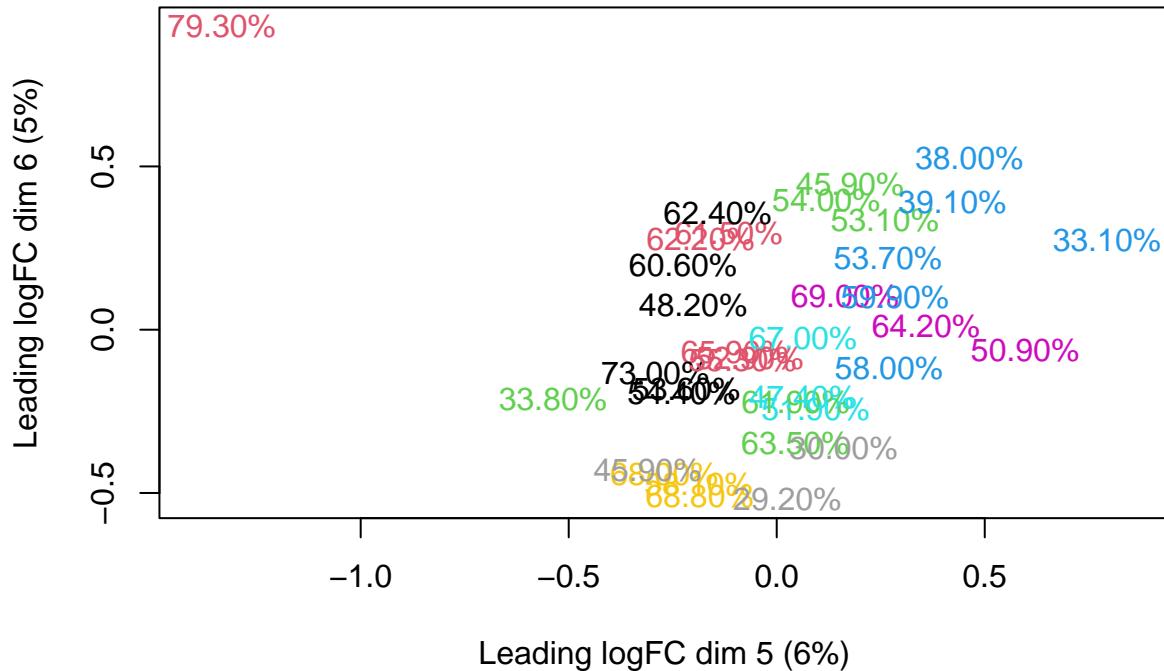
```



```
plotMDS(ob, top = 5000, labels = ob$samples$group, col = c(1:12)[ob$samples$group],
        dim.plot = c(5,6))
```



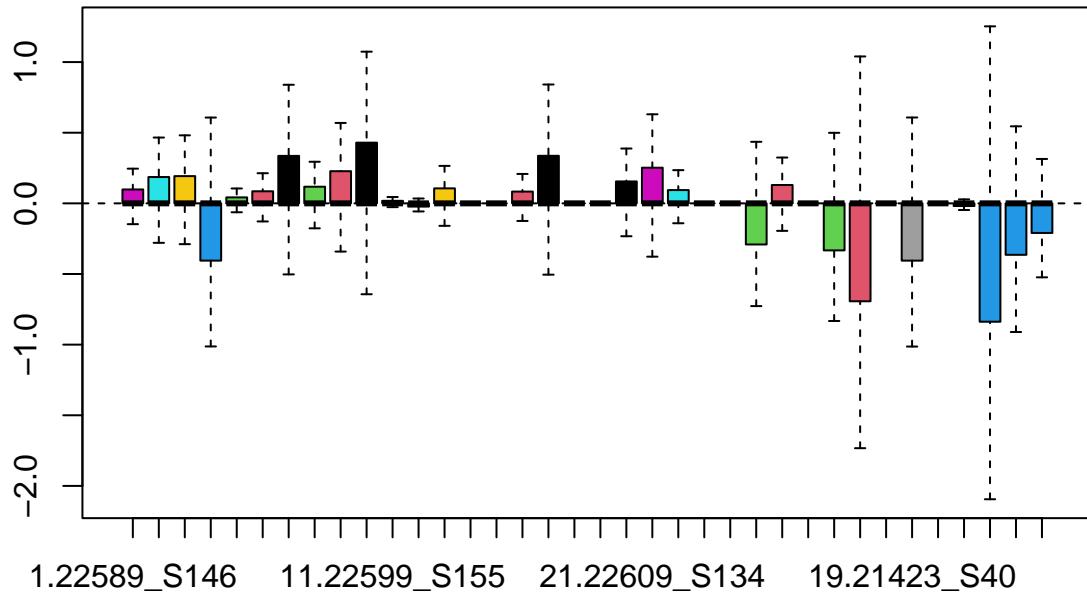
```
plotMDS(ob, top = 5000, labels = ob$samples$percent_assigned_to_genes, col = c(1:12)[ob$samples$group],  
dim.plot = c(5,6))
```



Filter

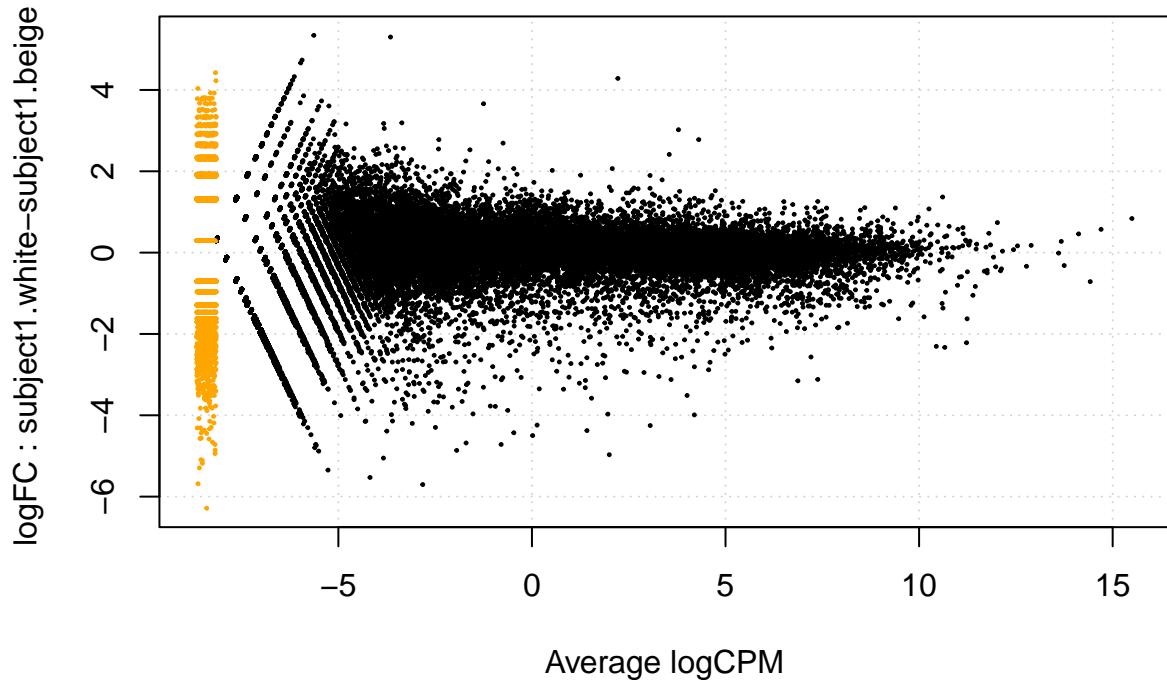
```
plotRLE(ob$counts, outline=FALSE, col=ob$samples$group, main="Before Filtering")
```

Before Filtering



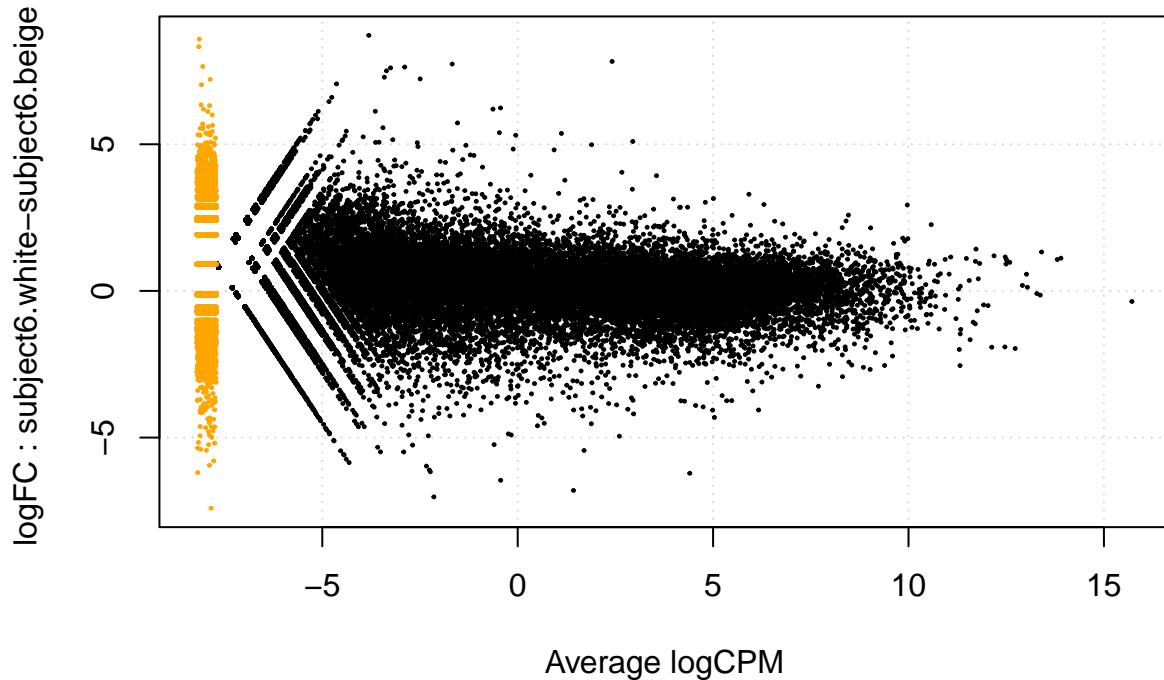
```
plotSmear(ob, main = "Before Filtering")
```

Before Filtering



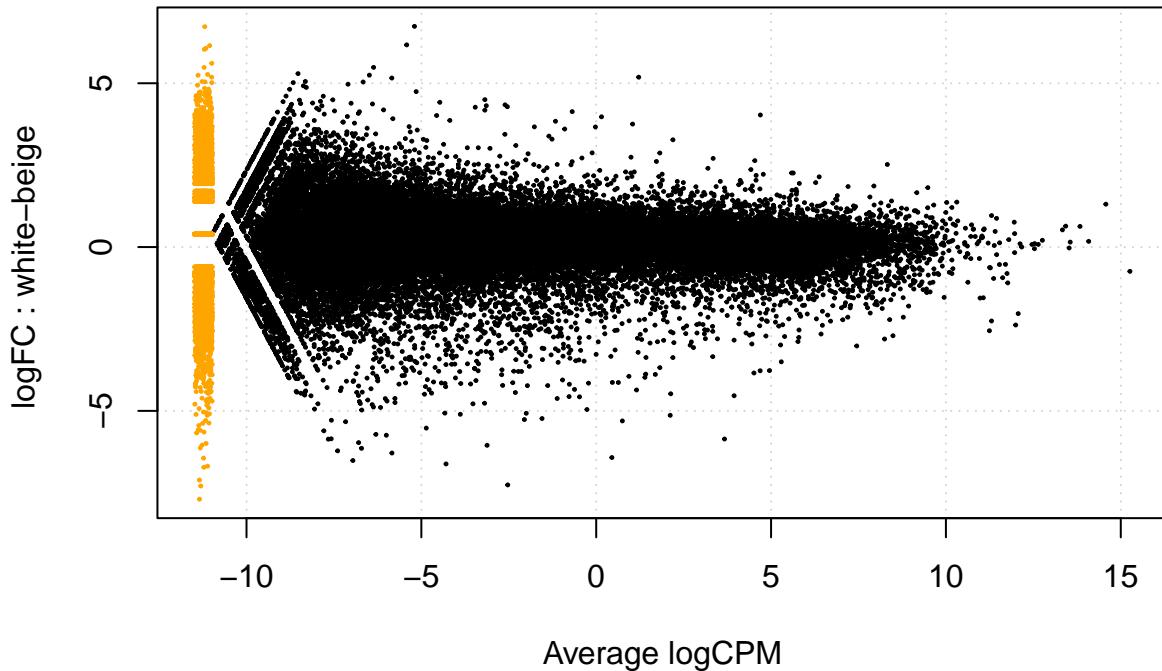
```
plotSmear(ob, main = "Before Filtering", pair=c("subject6.beige","subject6.white"))
```

Before Filtering



```
ob$samples$group = as.factor(ob$samples$condition)
plotSmear(ob, main = "Before Filtering: White vs Beige")
```

Before Filtering: White vs Beige



```
nrow(ob) #60 668 genes before filtering
```

```
## [1] 60668
```

Slight shift in mean between white and beige for s6.

Conduct a filtering using white and beige min.count and min.prop are default I just thought I'd list them here

```
levels(ob$samples$group)
```

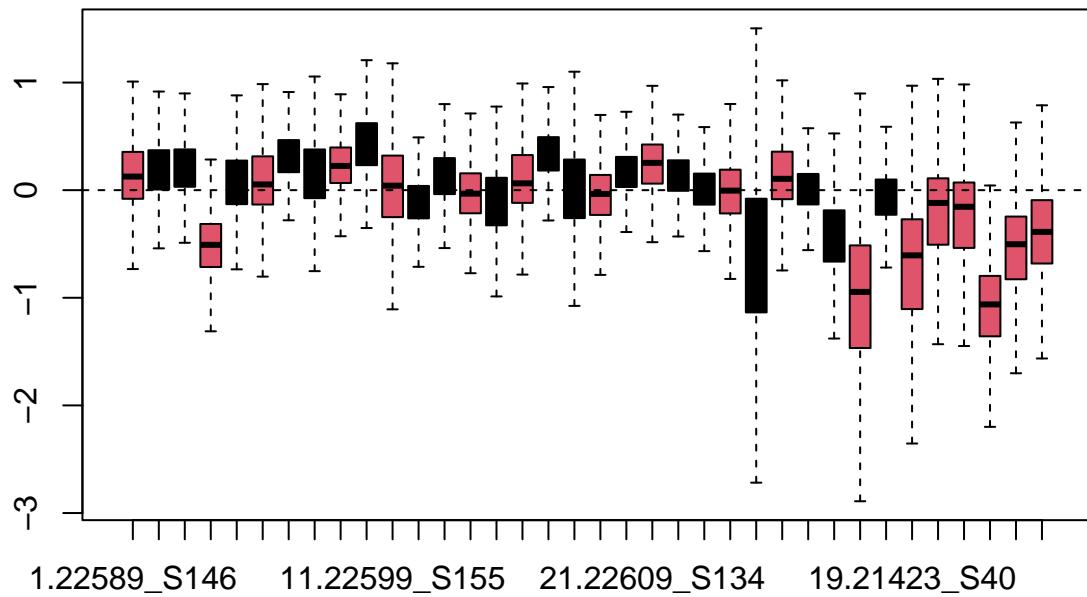
```
## [1] "beige" "white"
```

```
filt = ob$filterByExpr(ob, min.count=10, min.prop=0.7),, keep.lib.sizes=FALSE]
nrow(filt) #18061
```

```
## [1] 18061
```

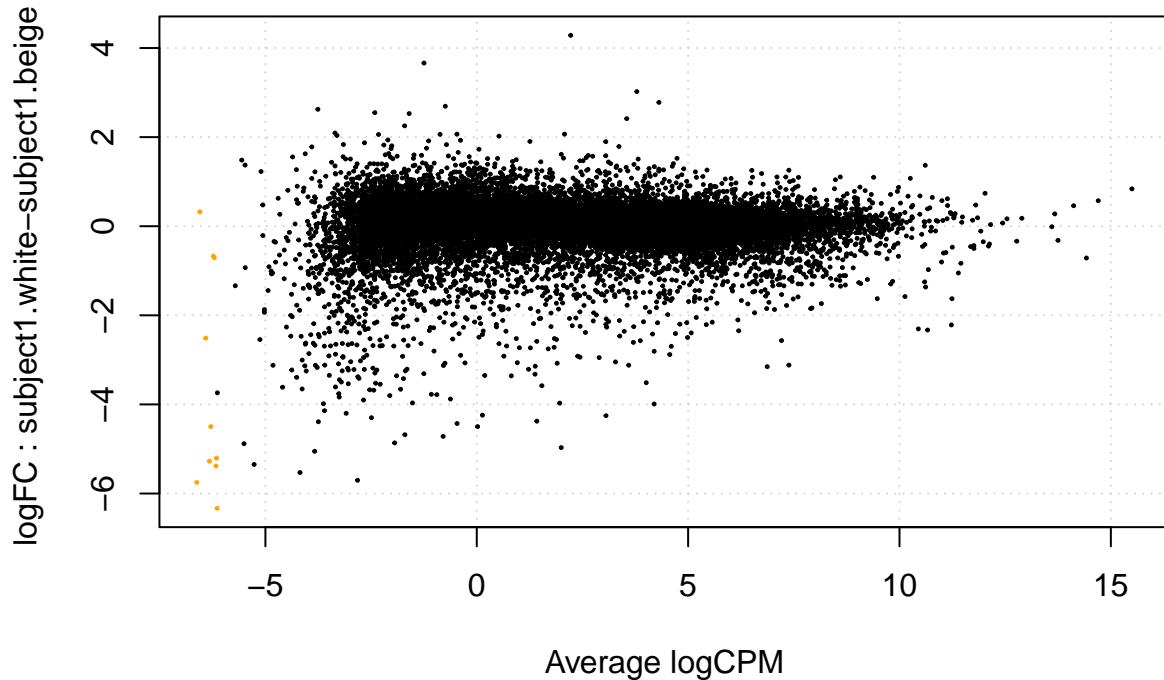
```
plotRLE(filt$counts, outline=FALSE, col=filt$samples$group, main="After Filtering")
```

After Filtering



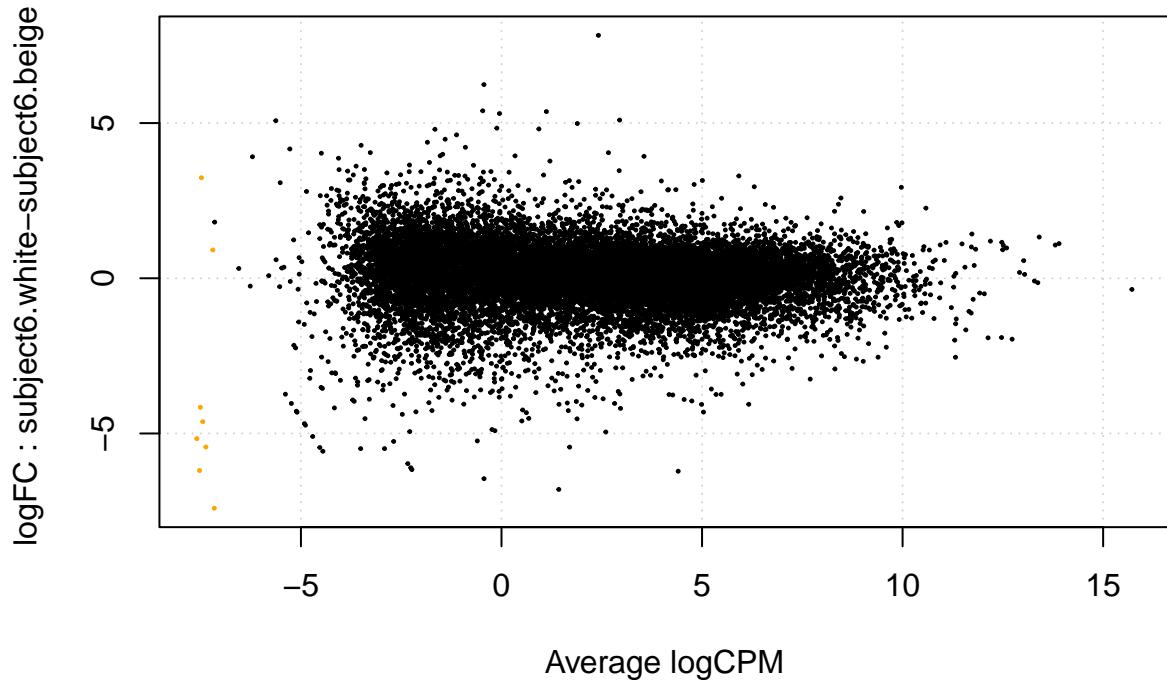
```
filt$samples$group = as.factor(filt$samples$donor.condition)
plotSmear(filt, main = "After Filtering")
```

After Filtering



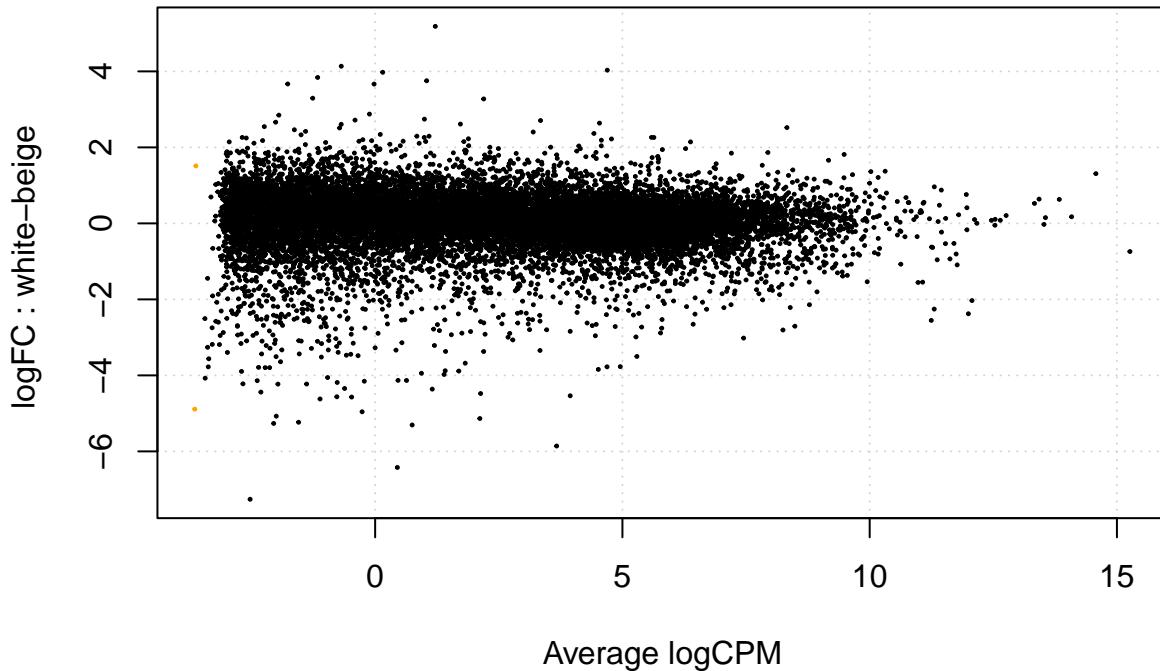
```
plotSmear(filt, main = "After Filtering", pair=c("subject6.beige","subject6.white"))
```

After Filtering



```
filt$samples$group = as.factor(filt$samples$condition)
plotSmear(filt, main = "After Filtering: White vs Beige")
```

After Filtering: White vs Beige



Sanity checks

```
summary(filt$counts[grep("ENSG00000132170", rownames(filt$counts)),]) #PPARG is included and looks high

##      Min. 1st Qu. Median     Mean 3rd Qu.    Max.
##      5035    9211   12616   12123   14265   20856

summary(filt$counts[grep("ENSG00000228630", rownames(filt$counts)),]) #HOTAIR is still detected at low

##      Min. 1st Qu. Median     Mean 3rd Qu.    Max.
##      26.00   91.25  146.50  339.61  454.00 1520.00

rownames(filt$counts)[grep("ENSG00000223972.5", rownames(filt$counts))] #This lowly expr gene is filtered

## character(0)

summary(filt$counts[grep("ENSG00000176194", rownames(filt$counts)),]) #CIDEA at low levels in beige samples

##      Min. 1st Qu. Median     Mean 3rd Qu.    Max.
##      0.0     17.0    46.5   310.0   521.8  1901.0
```

Norm Factors

```
filt = calcNormFactors(filt, method="TMM")
filt$samples[c("norm.factors")]
```

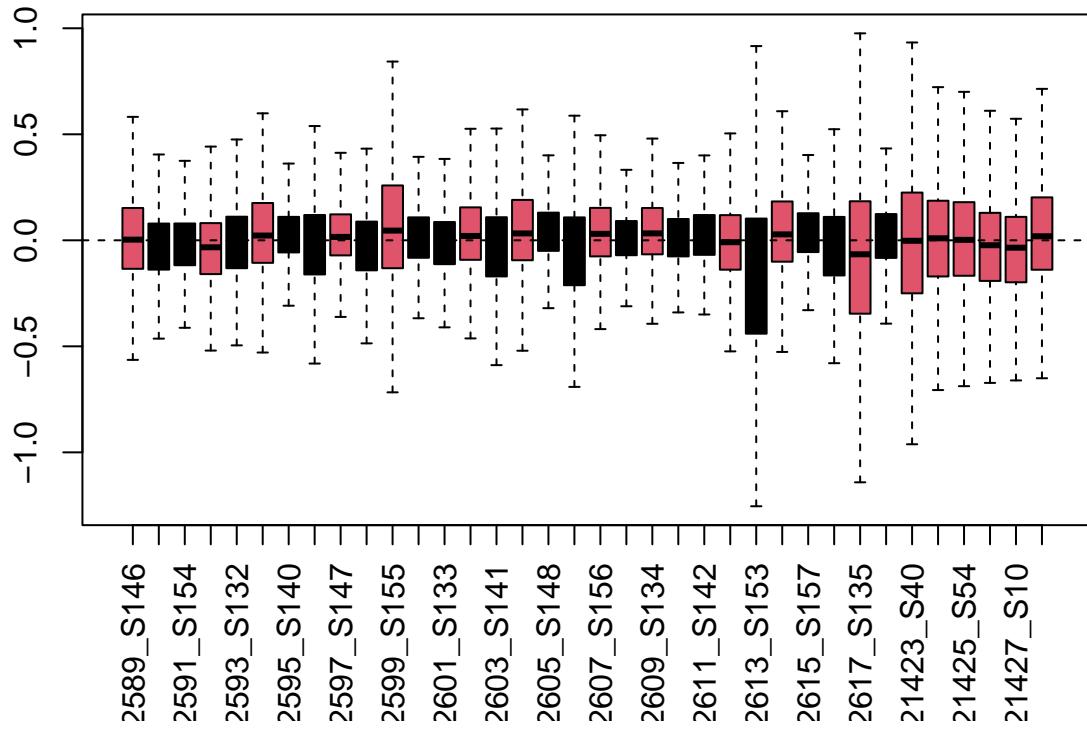
```
##          norm.factors
## 1.22589_S146      1.0048258
## 2.22590_S149      0.9551677
## 3.22591_S154      0.9775318
## 4.22592_S128      0.9512721
## 5.22593_S132      0.9729840
## 6.22594_S136      1.0364256
## 7.22595_S140      1.0386764
## 8.22596_S143      0.9575702
## 9.22597_S147      1.0409671
## 10.22598_S150     0.9547073
## 11.22599_S155     1.1037450
## 12.22600_S129     1.0151597
## 13.22601_S133     0.9888540
## 14.22602_S137     1.0428647
## 15.22603_S141     0.9422844
## 16.22604_S144     1.0496733
## 17.22605_S148     1.0612113
## 18.22606_S151     0.9162526
## 19.22607_S156     1.0529346
## 20.22608_S130     1.0062590
## 21.22609_S134     1.0560136
## 22.22610_S138     1.0151836
## 23.22611_S142     1.0381196
## 24.22612_S145     0.9904936
## 25.22613_S153     0.8176175
## 26.22614_S152     1.0443173
## 27.22615_S157     1.0504565
## 28.22616_S131     0.9522480
## 29.22617_S135     0.9044118
## 30.22618_S139     1.0246099
## 19.21423_S40      1.0362388
## 20.21424_S47      1.0329218
## 21.21425_S54      1.0242432
## 22.21426_S3       0.9804359
## 23.21427_S10      0.9537317
## 24.21428_S17      1.0674308
```

```
summary(filt$samples$norm.factors)
```

```
##      Min. 1st Qu. Median      Mean 3rd Qu.      Max.
## 0.8176  0.9570  1.0152  1.0016  1.0414  1.1037
```

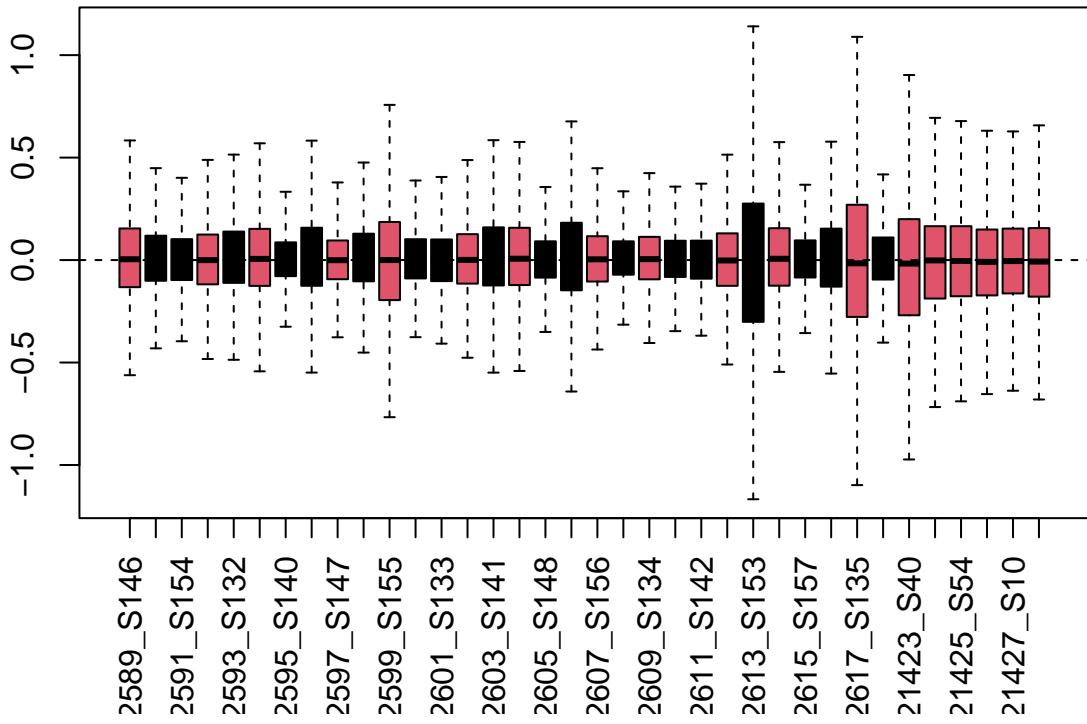
```
plotRLE(cpm(filt, normalized.lib.sizes = F), outline=F, col=filt$samples$group, las=3,
        main="After TMM normalisation (no libsize)")
```

After TMM normalisation (no libsize)



```
plotRLE(cpm(filt, normalized.lib.sizes = T), outline=F, col=filt$samples$group, las=3,  
main="After TMM + libsize normalisation")
```

After TMM + libsize normalisation



Annotate gene lists

```

mart <- biomaRt::useMart(биомарт = "ensembl",
  dataset = "hsapiens_gene_ensembl",
  host = "https://sep2019.archive.ensembl.org")

#searchFilters(mart, pattern="ensembl")

annot = getBM(c("external_gene_name", "description", "gene_biotype", "ensembl_gene_id", "ensembl_gene_id",
  filters = "ensembl_gene_id",
  values = filt$genes$Geneid,
  mart = mart)

head(annot, n=2); dim(annot)

##   external_gene_name                               description
## 1           TSPAN6 tetratspanin 6 [Source:HGNC Symbol;Acc:HGNC:11858]
## 2           TNMD    tenomodulin [Source:HGNC Symbol;Acc:HGNC:17757]
##   gene_biotype ensembl_gene_id ensembl_gene_id_version
## 1 protein_coding ENSG000000000003          ENSG000000000003.15
## 2 protein_coding ENSG000000000005          ENSG000000000005.6

## [1] 18058      5

```

```

#Tidying up the annot table
annot$description = gsub("\\[Source:.+\\]", "", annot$description)
colnames(annot)[1] = "gene_name"

#Add gene names to filt_series
filt$genes = merge(filt$genes, annot, by.x= "Geneid",
                    by.y = "ensembl_gene_id", sort=FALSE)
#head(filt$genes)
#make sure length is numeric
filt$genes$Length = as.numeric(filt$genes$Length)

#match the gene lists from genes and counts
nrow(filt$counts); nrow(filt$genes)

## [1] 18061

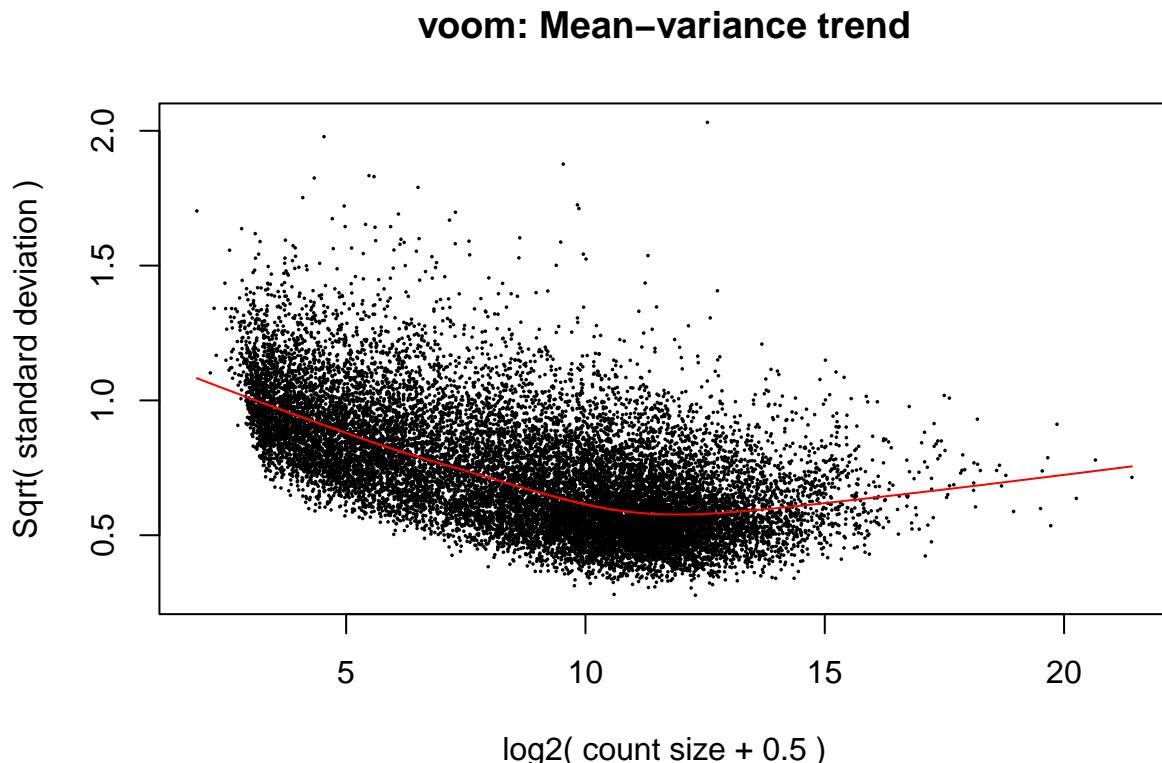
## [1] 18061

save(filt, file = here("03limma/DGElist_ob_limma_filt.RData"))

```

voom normlisation

```
norm = voom(filt, plot=T)
```



Limma

```
#head(norm$design)

#set some factor levels
norm$targets$group = paste(norm$targets$condition, norm$targets$donor, sep=".") #explicit
#simplify names for intercepting
norm$targets$group = gsub("subject","s", norm$targets$group)
norm$targets$group = factor(gsub("eige|hite","", norm$targets$group))
norm$targets$group

## [1] w.s3 b.s3 b.s4 w.s2 b.s2 w.s1 b.s1 b.s6 w.s5 b.s5 w.s3 b.s3 b.s4 w.s2 b.s2
## [16] w.s1 b.s1 b.s6 w.s5 b.s5 w.s3 b.s3 b.s4 w.s2 b.s2 w.s1 b.s1 b.s6 w.s5 b.s5
## [31] w.s4 w.s4 w.s4 w.s6 w.s6 w.s6
## Levels: b.s1 b.s2 b.s3 b.s4 b.s5 b.s6 w.s1 w.s2 w.s3 w.s4 w.s5 w.s6

design = model.matrix(~0 +frac_assigned_to_genes + group, data=norm$targets)
colnames(design) = c("frac_assigned_to_genes",levels(norm$targets$group))
head(design)

##          frac_assigned_to_genes b.s1 b.s2 b.s3 b.s4 b.s5 b.s6 w.s1 w.s2
## 1.22589_S146              0.642  0  0  0  0  0  0  0  0
## 2.22590_S149              0.519  0  0  1  0  0  0  0  0
## 3.22591_S154              0.581  0  0  0  1  0  0  0  0
## 4.22592_S128              0.537  0  0  0  0  0  0  0  1
## 5.22593_S132              0.619  0  1  0  0  0  0  0  0
## 6.22594_S136              0.659  0  0  0  0  0  0  1  0
##          w.s3 w.s4 w.s5 w.s6
## 1.22589_S146    1  0  0  0
## 2.22590_S149    0  0  0  0
## 3.22591_S154    0  0  0  0
## 4.22592_S128    0  0  0  0
## 5.22593_S132    0  0  0  0
## 6.22594_S136    0  0  0  0

explicit_fit = lmFit(norm, design=design)
coi = makeContrasts(s4 = b.s4- w.s4,
                     s3 = b.s3 - w.s3,
                     s2 = b.s2 - w.s2,
                     s1 = b.s1 - w.s1,
                     s6 = b.s6 - w.s6,
                     s5 = b.s5 - w.s5,
                     levels = design)
cfit = contrasts.fit(explicit_fit, coi)
efit = eBayes(cfit, robust=T)

results= decideTests(efit, method="separate", adjust.method = "BH", p.value = 0.01)
summary(results)

##           s4      s3      s2      s1      s6      s5
## Down     1155    798   1941     84   1520    971
```

```

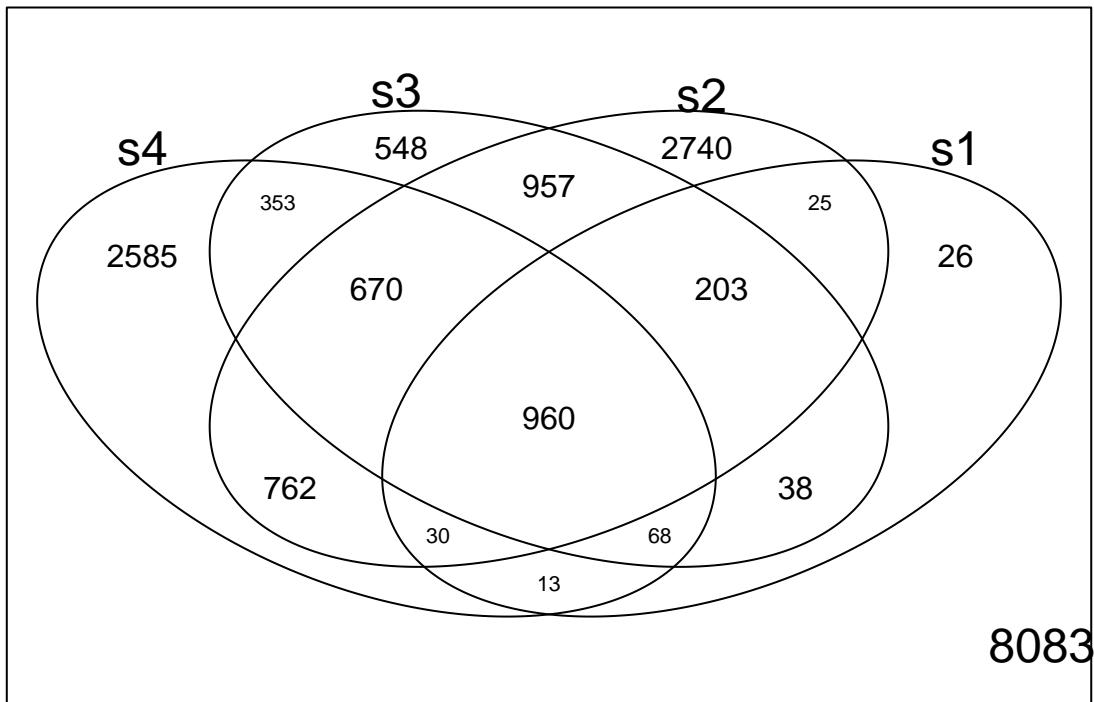
## NotSig 15319 15656 13868 17223 14562 15465
## Up      1587  1607  2252   754  1979  1625

results= decideTests(efit, method="separate", adjust.method = "BH", p.value = 0.05)
summary(results)

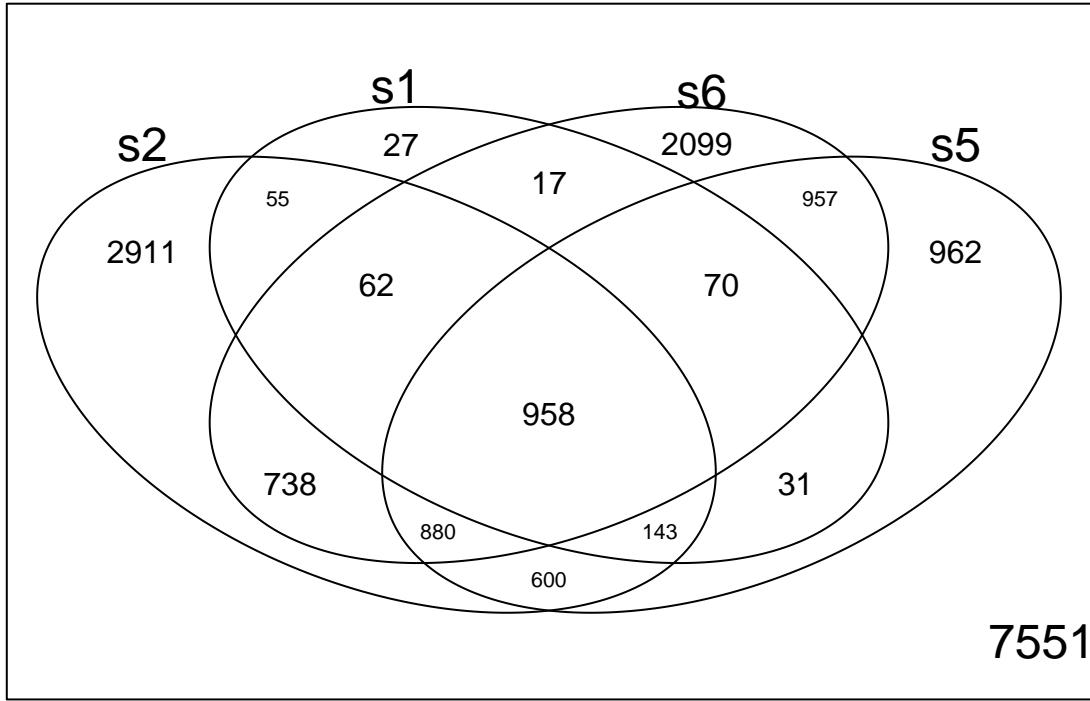
##          s4      s3      s2      s1      s6      s5
## Down    2430   1514   3189   276   2668   2039
## NotSig 12620  14264  11714  16698  12280  13460
## Up     3011   2283   3158   1087   3113   2562

vennDiagram(results[,1:4])

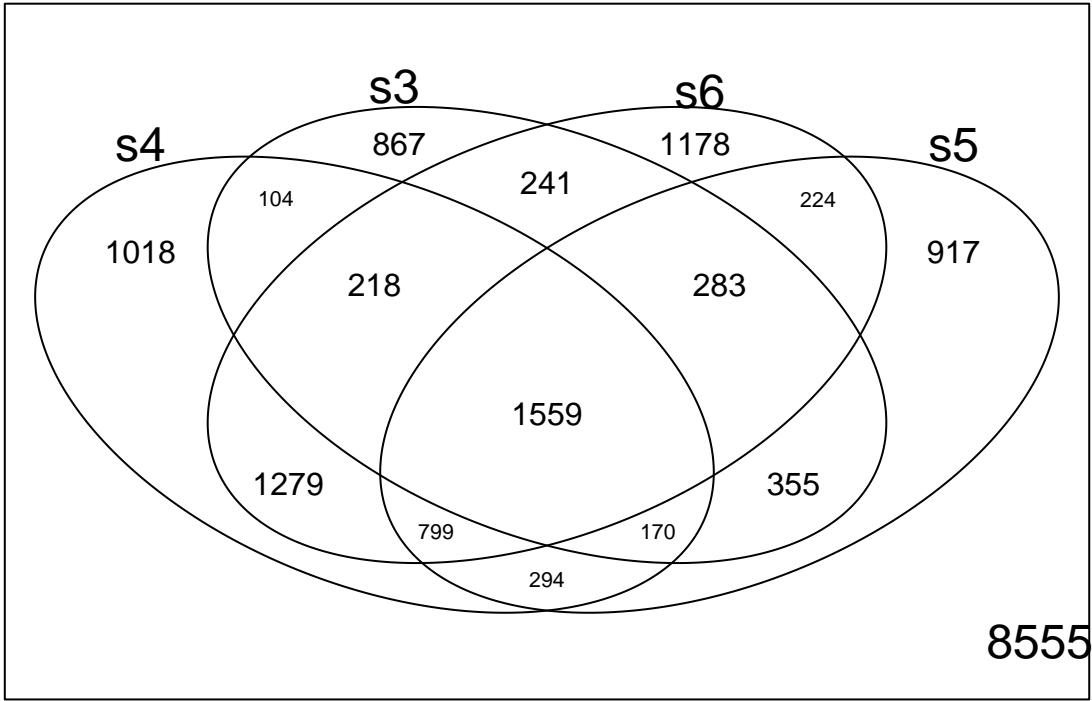
```



```
vennDiagram(results[,c(3:6)])
```



```
vennDiagram(results[,c(1:2,5:6)])
```



s4 has a small amount of independent genes, but not more than subject2.

Upsets

```
make_upset_table = function(res) {
  library(UpSetR)

  res= data.frame(res)
  new_bin = matrix(NA,ncol=ncol(res)*2, nrow=nrow(res))
  colnames(new_bin) = paste(rep(colnames(res),each=2), c("beige","white"), sep= "_")
  rownames(new_bin) = rownames(res)
  for (i in 1:nrow(res)){
    jcount=0
    for (j in 1:length(res[i])){
      nj = j + jcount
      v = res[i,j]
      if (v == 0){
        new_bin[i,nj] = 0
        new_bin[i,nj+1] = 0
      } else if (v == 1){
        new_bin[i,nj] = 1
        new_bin[i,nj+1] = 0
      } else if (v == -1){
        new_bin[i,nj] = 0
      }
    }
  }
}
```

```

        new_bin[i,nj+1] = 1
    } else {print("Error")}

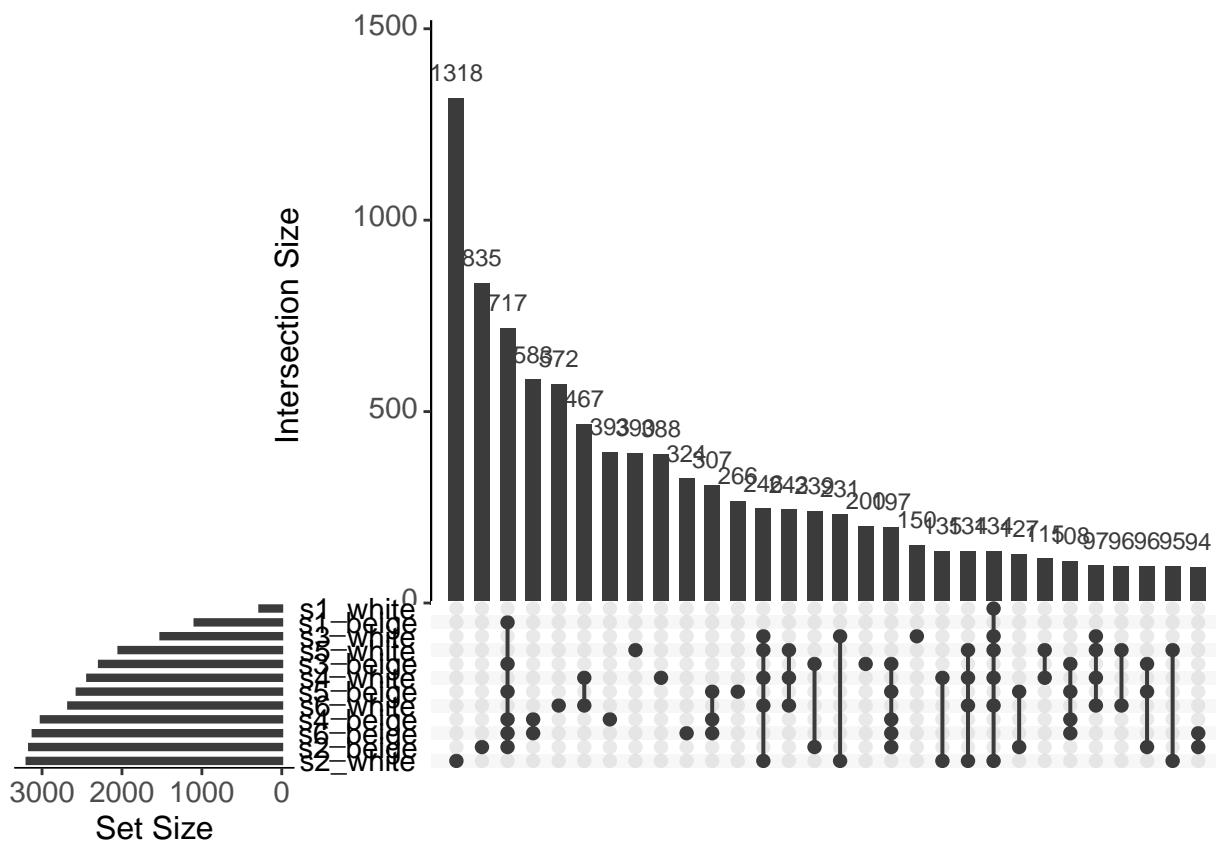
    jcount = jcount + 1
}
}
summary(new_bin)
new_bin = as.data.frame(new_bin)
#print(upset(new_bin,
#  order.by = "freq", nintersects=25))
return(new_bin)
}

```

```

results_bin = make_upset_table(results)
upset(results_bin,nsets=14,
      order.by = "freq", nintersects=30, text.scale=1.5)

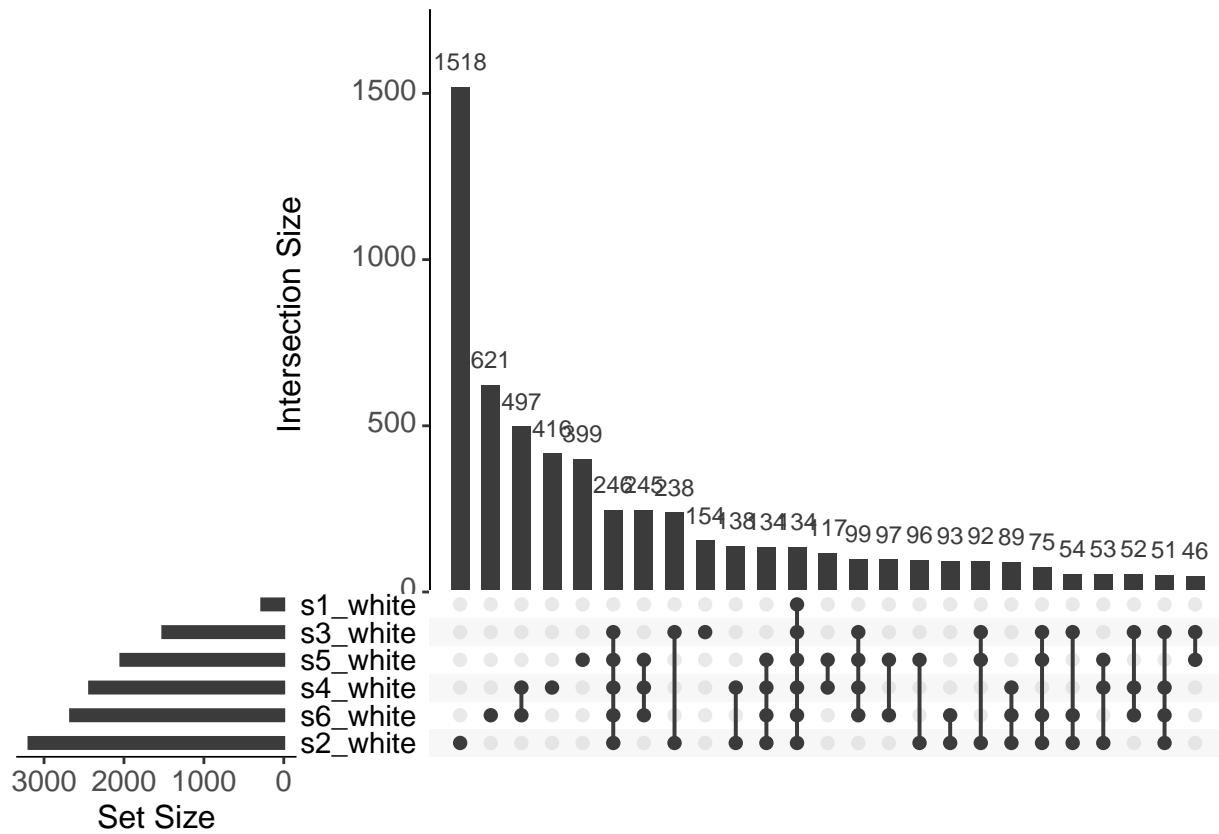
```



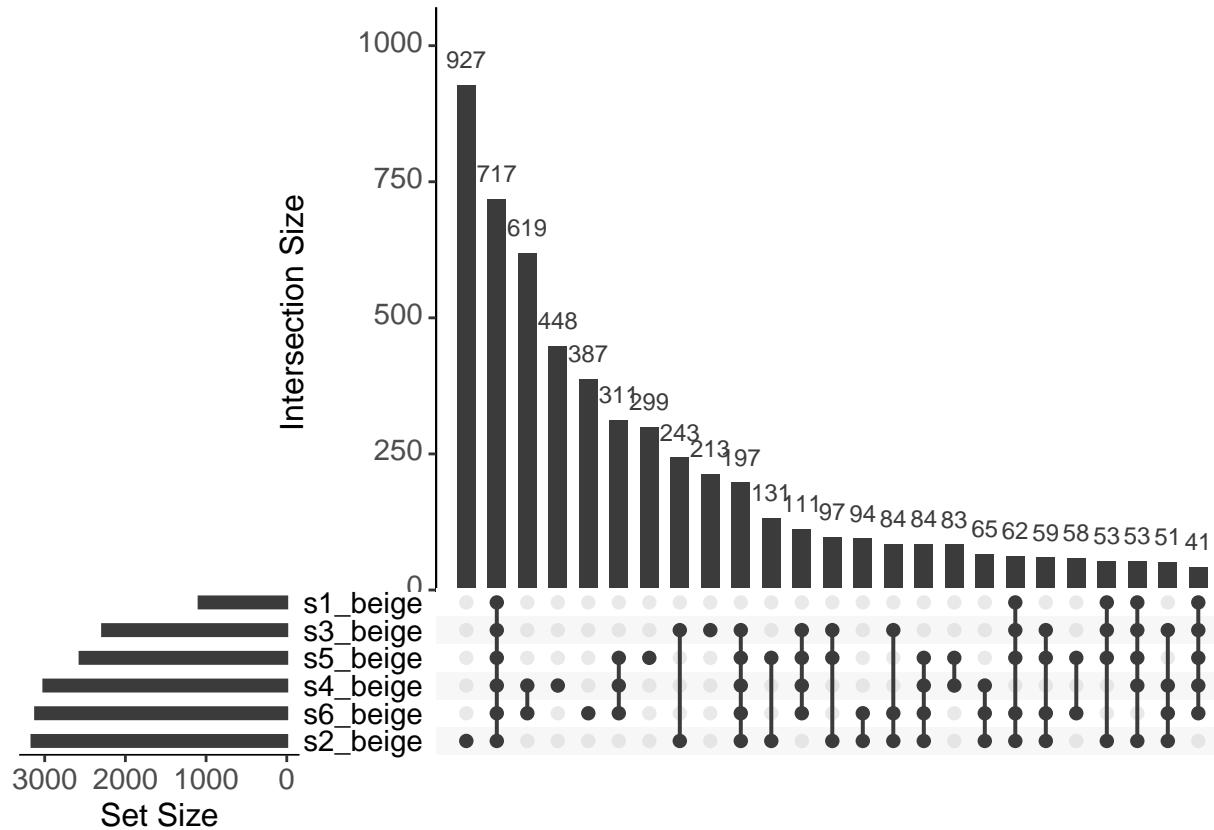
```

upset(results_bin[,grep("white", colnames(results_bin))],nsets=14,
      order.by = "freq", nintersects=25, text.scale=1.5)

```



```
upset(results_bin[,grep("beige", colnames(results_bin))], nsets=14,
      order.by = "freq", nintersects=25, text.scale=1.5)
```



Overall pvalue

We can extract merely like this

```
anytable = topTable(efit, number = Inf, adjust.method = "BH",
                    coef = c('s3','s4','s2','s1','s6','s5'))
anytable$AvelogFC = rowMeans(anytable[c('s3','s4','s2','s1','s6','s5')])
summary(anytable$adj.P.Val < 0.01) #much more conservative than previous tries
```

```
##      Mode     FALSE     TRUE
## logical    10502    7559
```

Donor tables

```
donortabs = list()
donors = c('s3','s4','s2','s1','s6','s5')
base_matrix = matrix(NA, dimnames = list(efit$genes$gene_name,donors),
                     nrow = nrow(efit$genes), ncol=length(donors))
donorlists = list(adj.p.val = base_matrix, logFC = base_matrix,
                  genes= efit$genes )
for ( co in donors){
  onedonor = topTable(efit, number = Inf, adjust.method = "BH",
                      coef = co)
```

```

donortabs[[co]] = onedonor
donorlists$adj.p.val[,co] = onedonor$adj.P.Val[order(onedonor$Geneid)]
donorlists$logFC[,co] = onedonor$logFC[order(onedonor$Geneid)]
}
summary(donortabs)

##      Length Class      Mode
## s3 13    data.frame list
## s4 13    data.frame list
## s2 13    data.frame list
## s1 13    data.frame list
## s6 13    data.frame list
## s5 13    data.frame list

ann_cols = c("Geneid", "gene_name", "description")
alltab = anytable[,!colnames(anytable) %in%c("ensembl_gene_id_version", "ID")]
for (d in donors){
  formatted = donortabs[[d]]
  formatted = formatted[,c(ann_cols,"P.Value","adj.P.Val","AveExpr")]
  alltab = merge(alltab, formatted, by=ann_cols,
                 suffixes = c("",paste(".", d, sep="")))
  alltab = alltab[!duplicated(alltab$gene_name),] #remove duplicates because they tend to propagate
}
head(alltab); nrow(alltab)

##           Geneid gene_name
## 1 ENSG00000000003    TSPAN6
## 2 ENSG00000000005    TNMD
## 3 ENSG00000000419    DPM1
## 4 ENSG00000000457    SCYL3
## 5 ENSG00000000460 C1orf112
## 6 ENSG00000000938    FGR
##                                     description Length
## 1                               tetraspanin 6     4536
## 2                               tenomodulin 1476
## 3 dolichyl-phosphate mannosyltransferase subunit 1, catalytic 1207
## 4                               SCY1 like pseudokinase 3   6883
## 5                               chromosome 1 open reading frame 112 5970
## 6 FGR proto-oncogene, Src family tyrosine kinase 3382
##      gene_biotype      s4      s3      s2      s1      s6
## 1 protein_coding 0.37343465 0.57278654 0.8603088 0.49422154 0.6439646
## 2 protein_coding 1.81772689 2.58660597 -0.2720670 2.38787369 1.9785258
## 3 protein_coding 0.06689829 -0.06185359 0.1194288 -0.11790050 0.1710766
## 4 protein_coding 0.32546768 0.49896973 0.5484816 0.39439050 0.3469994
## 5 protein_coding -0.17866146 0.13017667 0.3071896 -0.05290602 -0.2082301
## 6 protein_coding 3.24138536 2.88724528 2.3497110 1.91496687 2.7442577
##      s5 AveExpr          F      P.Value    adj.P.Val    AvelogFC
## 1 0.56429720 5.212308 31.7140824 2.969288e-11 7.806158e-10 0.5848355
## 2 1.94504039 1.961126 20.9163666 3.809066e-09 5.434087e-08 1.7406176
## 3 -0.01815705 5.314282 0.4712428 8.237264e-01 8.380174e-01 0.0265821
## 4 0.35682721 3.626998 10.0207970 6.300732e-06 3.633461e-05 0.4118560
## 5 -0.36021700 1.367959 1.3456734 2.705375e-01 3.177382e-01 -0.0604414

```

```

## 6 2.88754674 2.951839 95.9233119 2.476214e-17 5.025045e-15 2.6708521
## P.Value.s3 adj.P.Val.s3 AveExpr.s3 P.Value.s4 adj.P.Val.s4 AveExpr.s4
## 1 1.180377e-05 2.253571e-04 5.212308 1.164889e-02 4.171991e-02 5.212308
## 2 1.956120e-06 5.218110e-05 1.961126 1.864017e-03 1.155716e-02 1.961126
## 3 6.755915e-01 8.196868e-01 5.314282 7.245401e-01 8.149446e-01 5.314282
## 4 8.348144e-04 6.877477e-03 3.626998 6.456487e-02 1.441980e-01 3.626998
## 5 5.128644e-01 7.062247e-01 1.367959 4.826271e-01 6.174632e-01 1.367959
## 6 3.418267e-11 6.015586e-09 2.951839 1.506080e-10 5.681113e-08 2.951839
## P.Value.s2 adj.P.Val.s2 AveExpr.s2 P.Value.s1 adj.P.Val.s1 AveExpr.s1
## 1 1.031683e-08 4.254162e-07 5.212308 7.423497e-05 2.280200e-03 5.212308
## 2 5.374467e-01 6.575098e-01 1.961126 4.606466e-06 2.307037e-04 1.961126
## 3 4.196868e-01 5.530398e-01 5.314282 4.216597e-01 7.813318e-01 5.314282
## 4 3.317889e-04 2.112990e-03 3.626998 5.556707e-03 6.606388e-02 3.626998
## 5 1.314679e-01 2.355830e-01 1.367959 7.868527e-01 9.473852e-01 1.367959
## 6 3.346207e-09 1.798686e-07 2.951839 8.493511e-08 8.063249e-06 2.951839
## P.Value.s6 adj.P.Val.s6 AveExpr.s6 P.Value.s5 adj.P.Val.s5 AveExpr.s5
## 1 5.072093e-06 1.086679e-04 5.212308 1.989400e-05 3.539955e-04 5.212308
## 2 1.584704e-04 1.496932e-03 1.961126 1.460798e-04 1.739187e-03 1.961126
## 3 2.804584e-01 4.204665e-01 5.314282 9.043584e-01 9.430495e-01 5.314282
## 4 2.076997e-02 6.094661e-02 3.626998 1.359659e-02 5.250544e-02 3.626998
## 5 3.288567e-01 4.715745e-01 1.367959 8.133352e-02 1.835518e-01 1.367959
## 6 2.754188e-10 5.292092e-08 2.951839 6.097364e-11 1.596007e-08 2.951839

## [1] 18043

summary(rowSums(alltab[,grep("adj.P.Val\\\\.s", colnames(alltab))]) < 0.05) == 6

##      Mode   FALSE    TRUE
## logical 17190     853

```

We get a different number of significant genes than using decideTests because in order to extract the p.adjust for each contrast we have to forgo the p-value adjustment across contrasts.

```

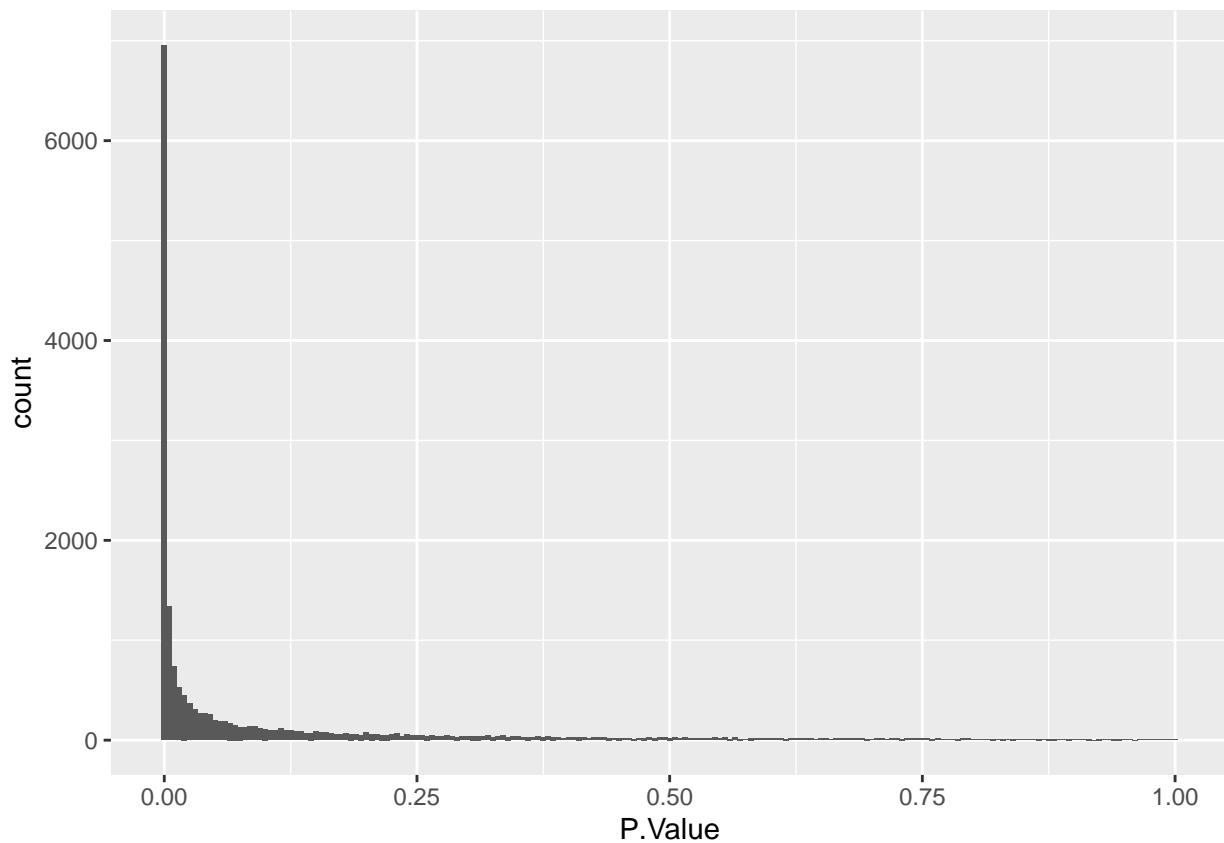
logfc_cols = grep("^s[1-6]$", colnames(alltab))
colnames(alltab)[logfc_cols] = paste0("logFC.", colnames(alltab)[logfc_cols])
alltab = rename(alltab, all.donors.P.Value = P.Value,
               all.donors.adj.P.Val = adj.P.Val,
               all.donors.AvelogFC = AvelogFC)

write.table(alltab, here("O3limma/any_and_all_donor_DGE.tsv"),
            sep="\t", quote=F, row.names = F)

```

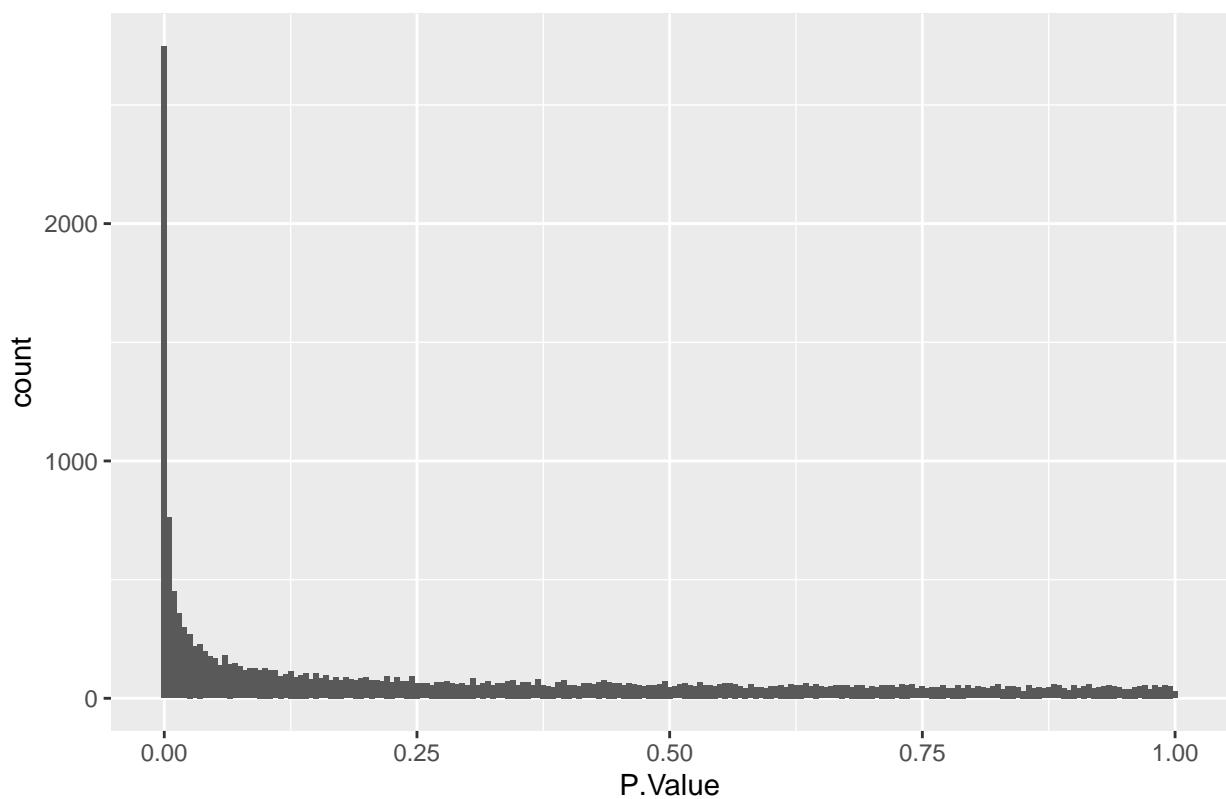
Pvalue histograms

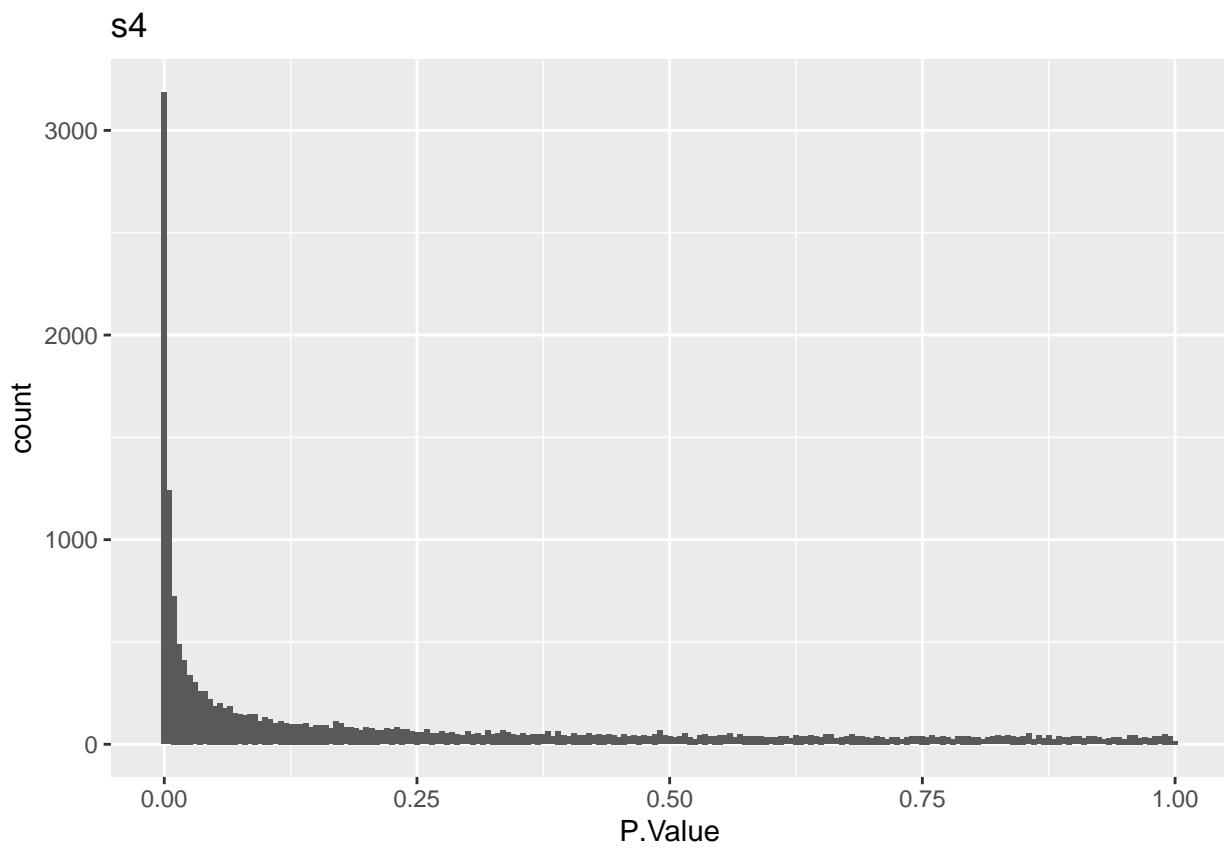
```
ggplot(anytable) + geom_histogram(aes(x=P.Value), binwidth = 0.005)
```

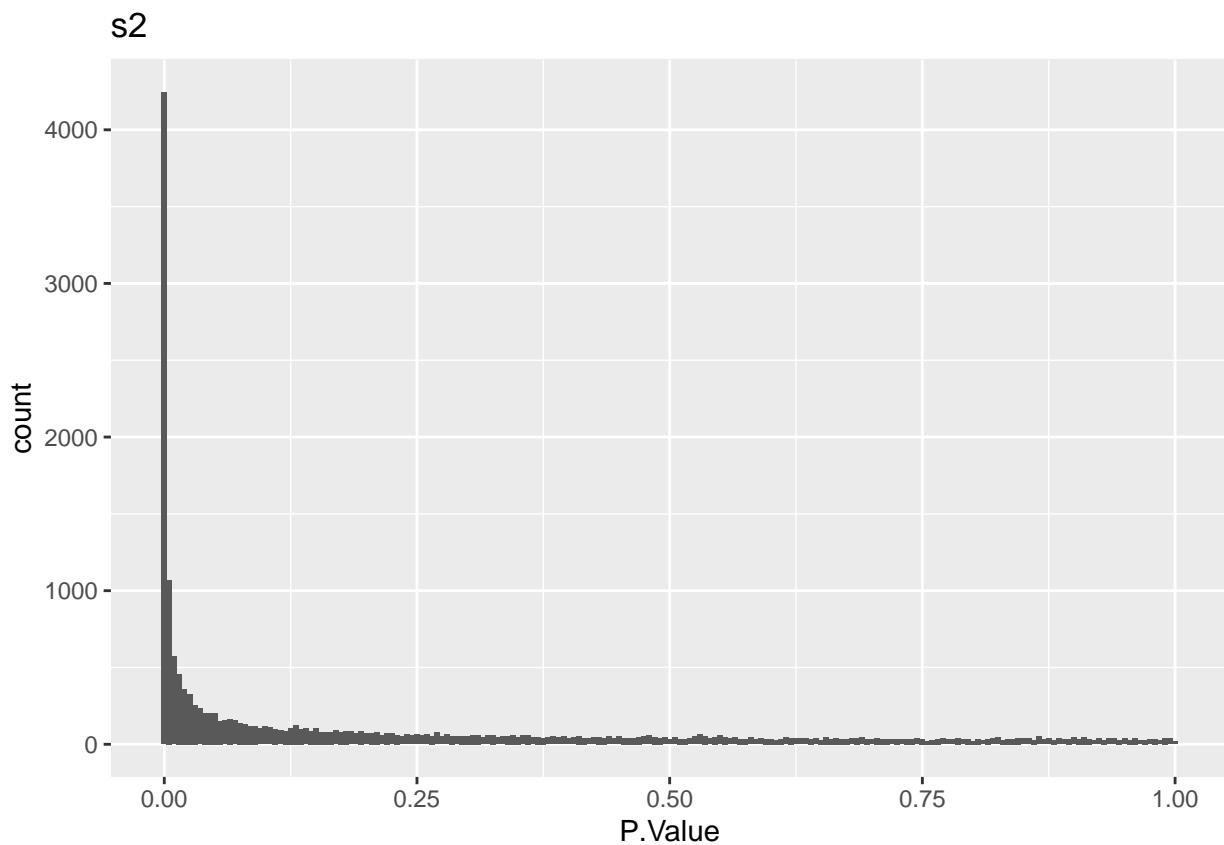


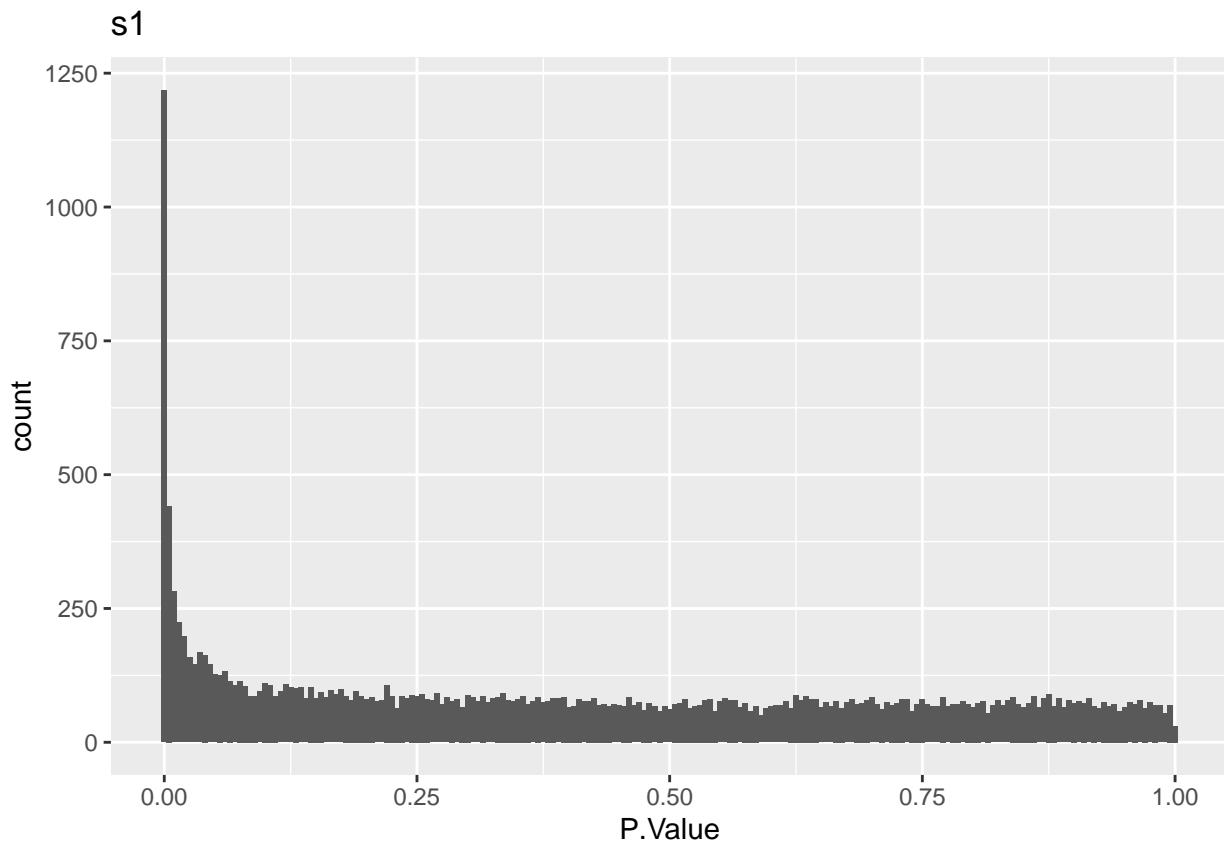
```
for (d in donors){  
  p = ggplot(donortabs[[d]]) + geom_histogram(aes(x=P.Value), binwidth = 0.005) + ggtitle(d)  
  print(p)  
}
```

s3

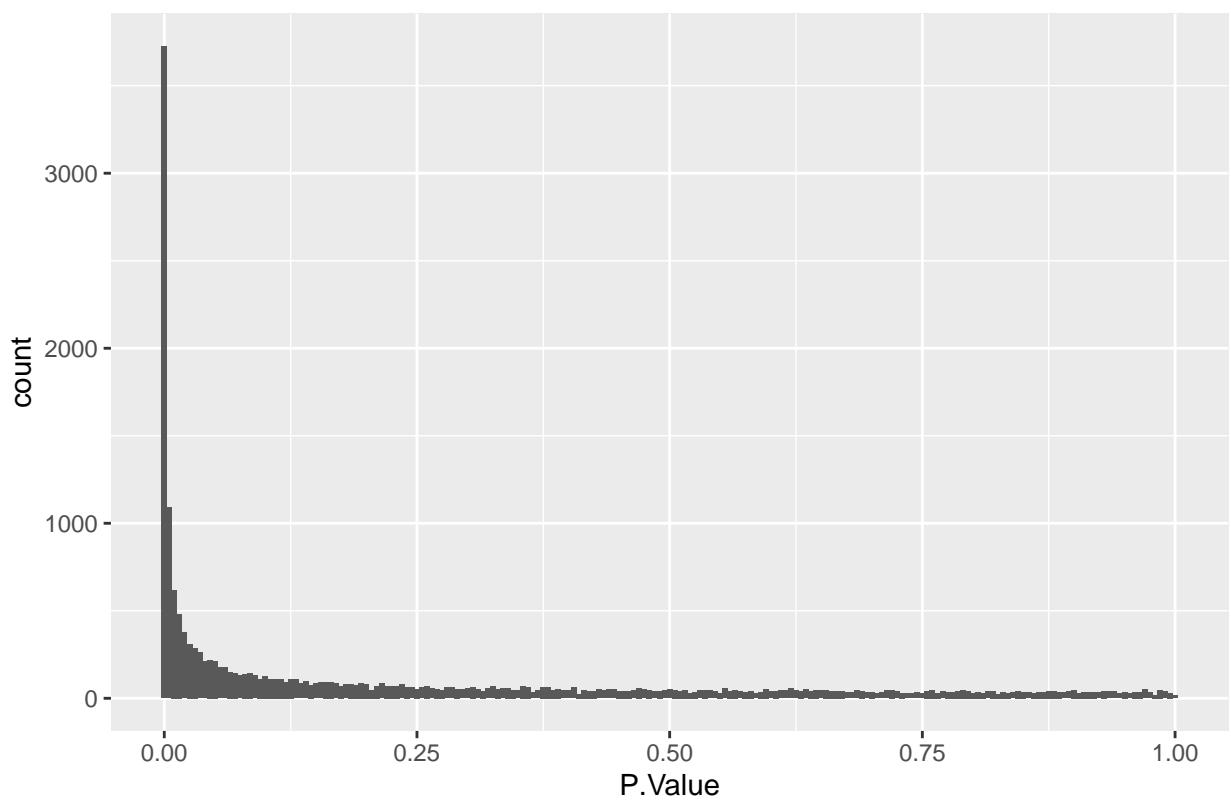


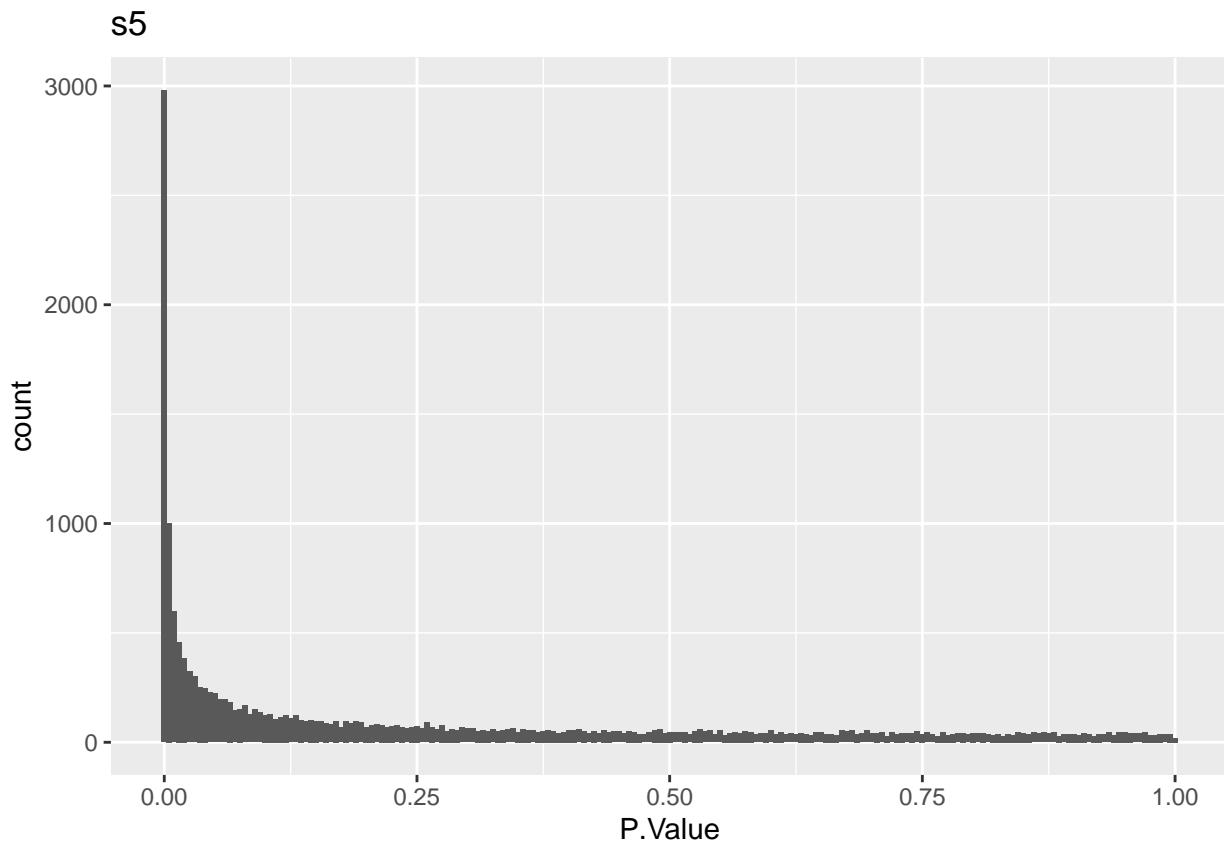






s6

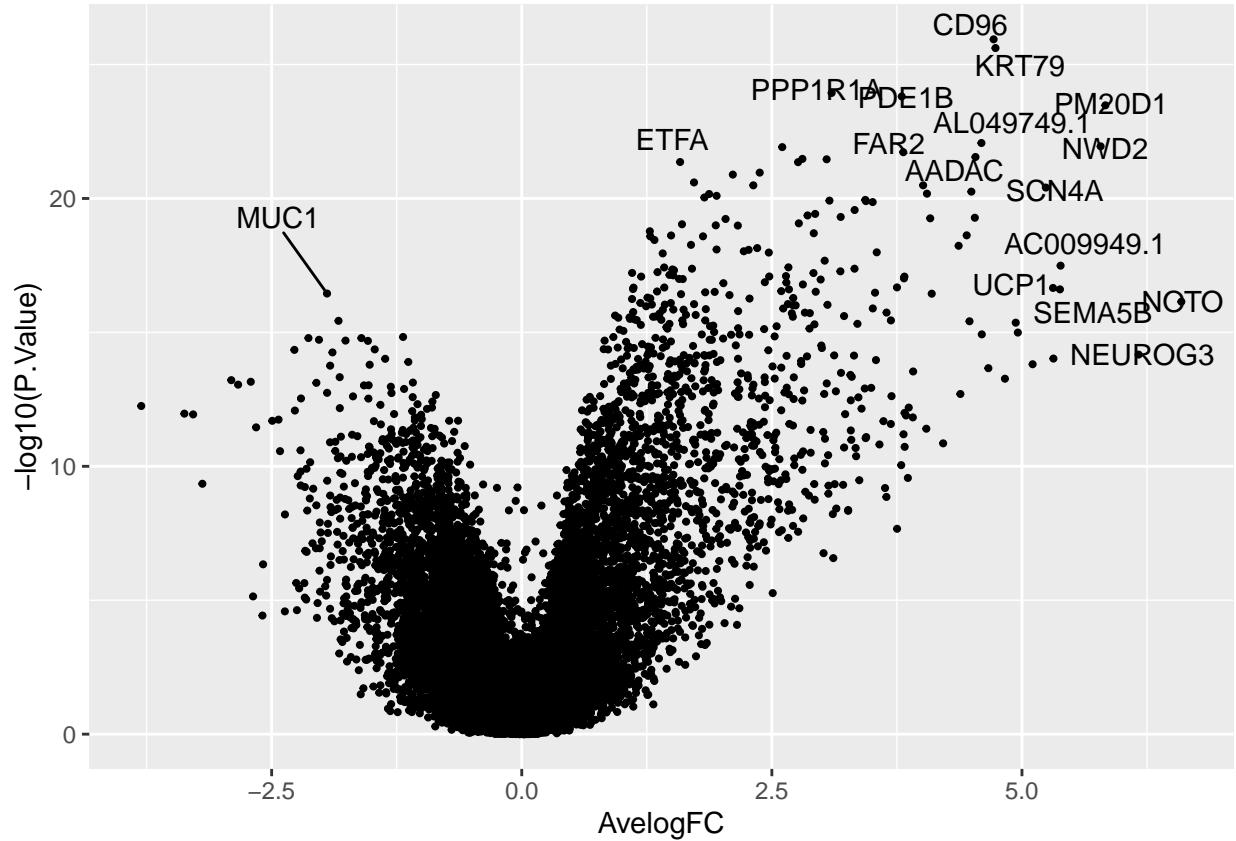




Volcano Plots

```
ggplot(anytable, aes(x=AvelogFC, y=-log10(P.Value))) +
  geom_point(show.legend = F, size=0.75) +
  geom_text_repel(data=anytable[anytable$AvelogFC > 5 |
    anytable$AvelogFC < -2.75 |
    anytable$adj.P.Val < 0.0001,],
    aes(label=gene_name))

## Warning: ggrepel: 3599 unlabeled data points (too many overlaps). Consider
## increasing max.overlaps
```



```

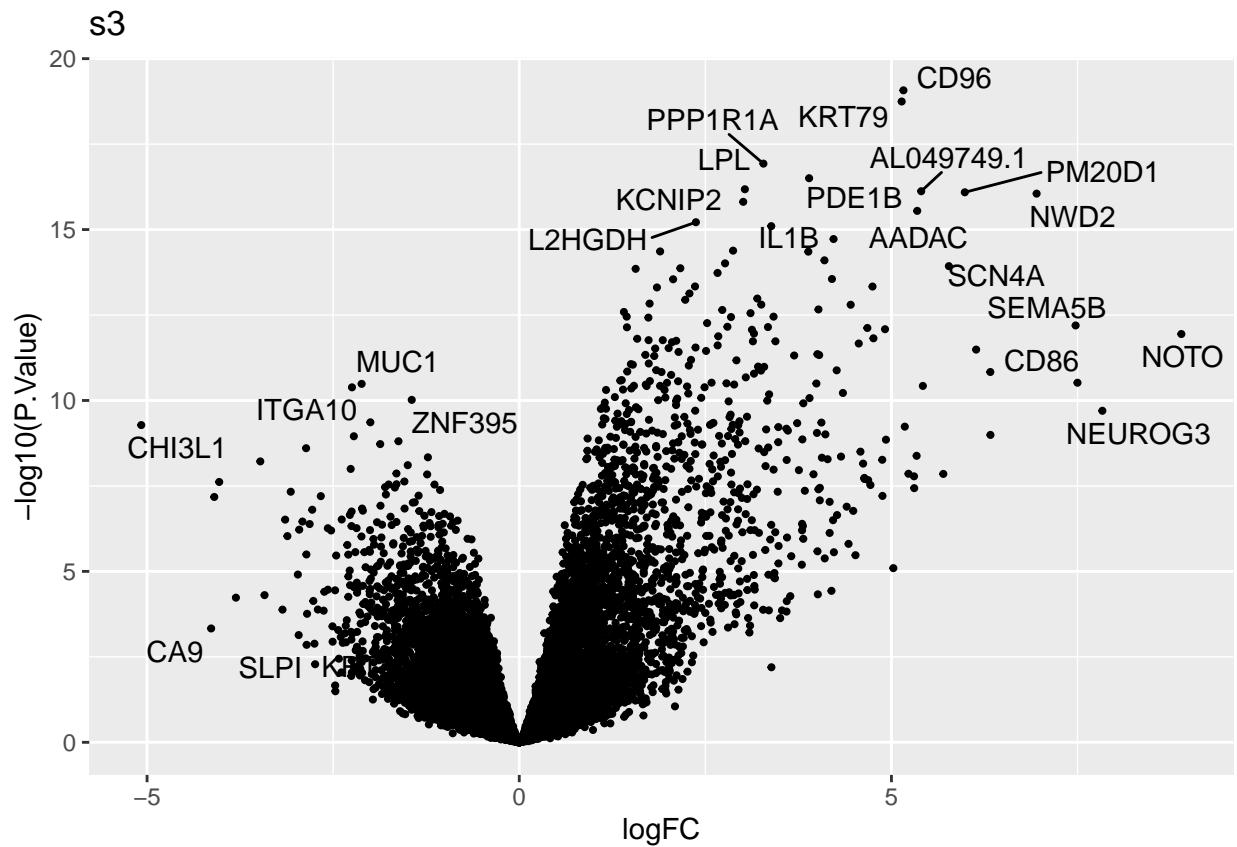
for ( d in donors){
  tab =donortabs[[d]]
  p = ggplot(tab, aes(x=logFC, y=-log10(P.Value))) +
    geom_point(show.legend = F, size=0.75) + ggtitle(d) +
    geom_text_repel(data=tab[tab$logFC > 5 |
                           tab$logFC < -2.75 |
                           tab$adj.P.Val < 0.0001,],
                    aes(label=gene_name))
}
print(p)

```

```

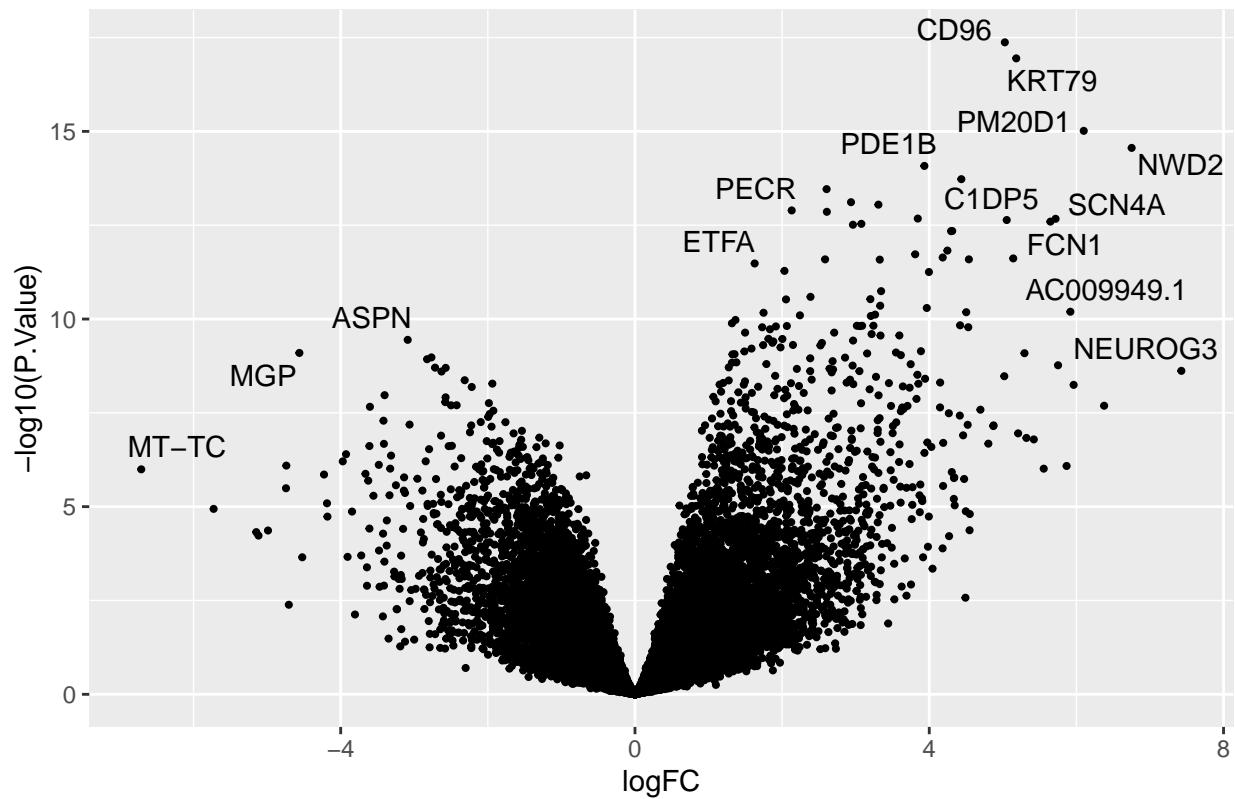
## Warning: ggrepel: 767 unlabeled data points (too many overlaps). Consider
## increasing max.overlaps

```

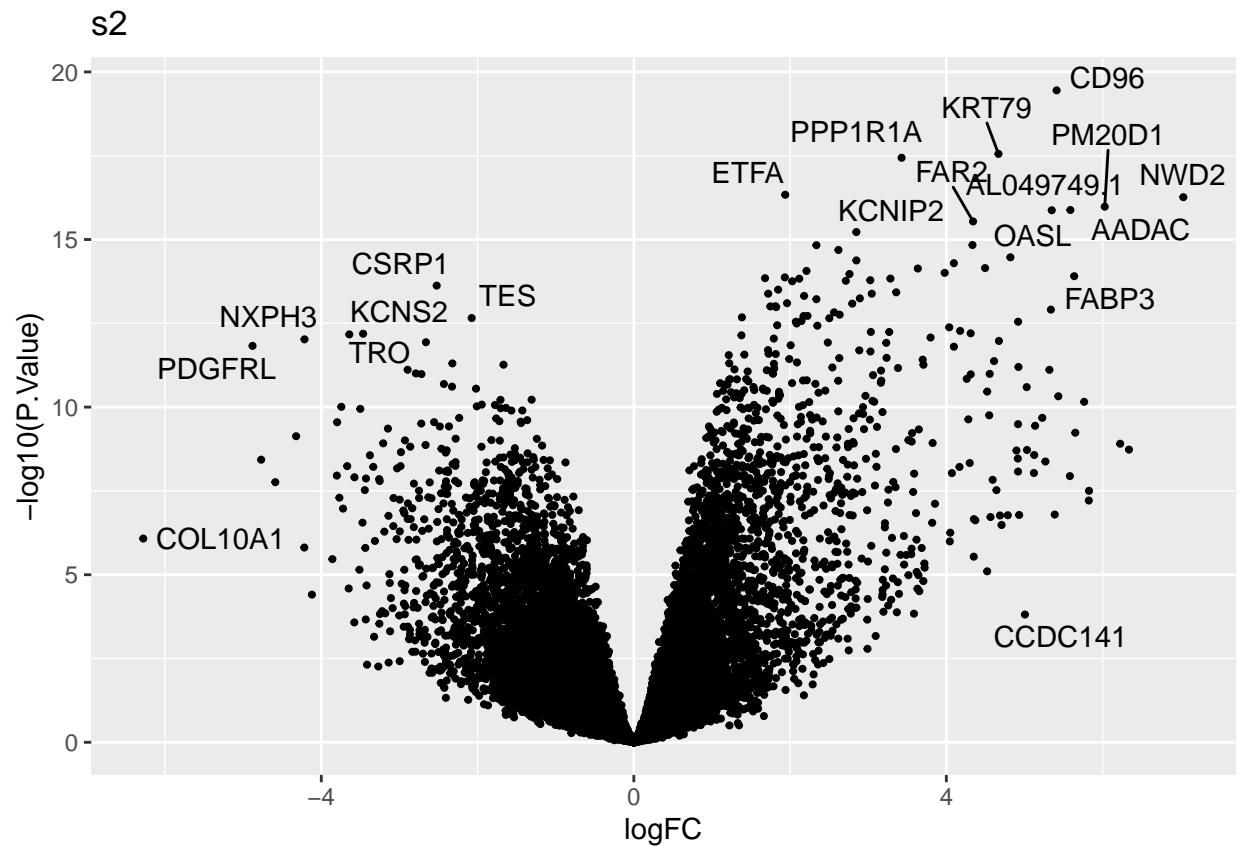


```
## Warning: ggrepel: 581 unlabeled data points (too many overlaps). Consider  
## increasing max.overlaps
```

s4

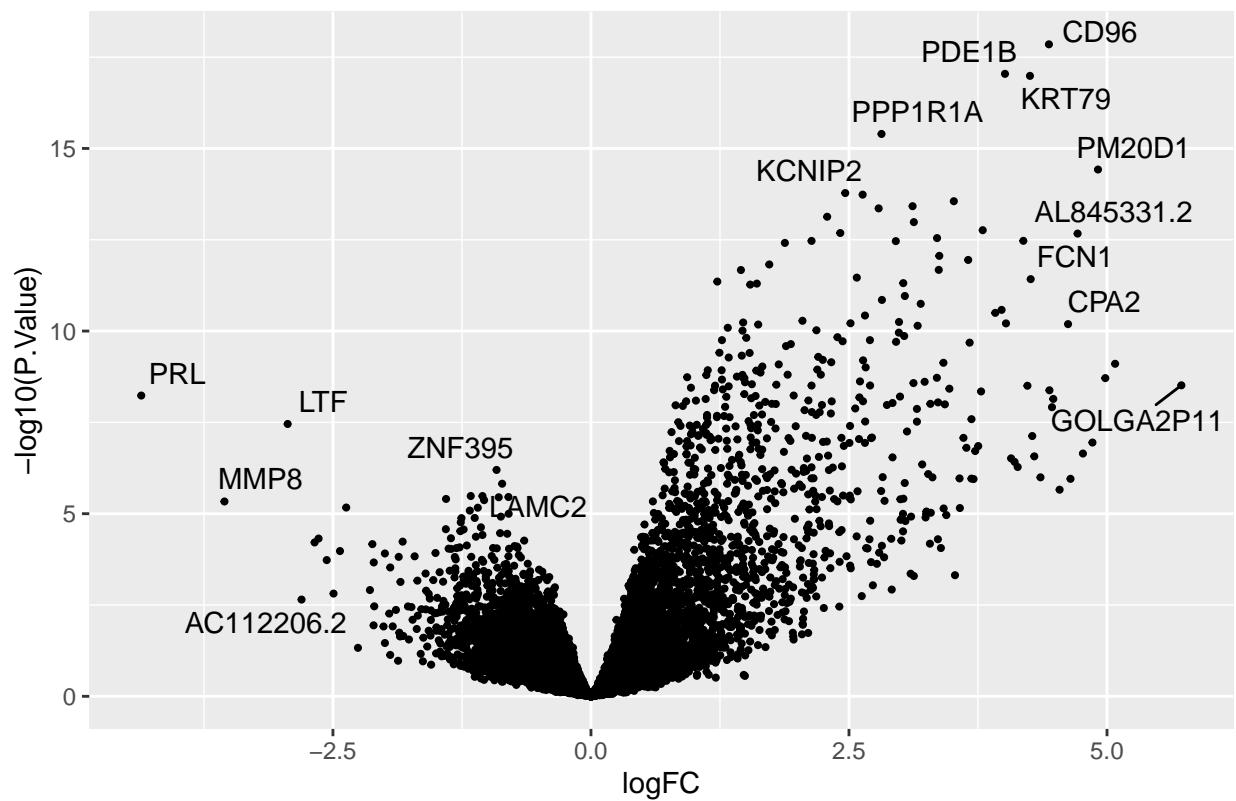


```
## Warning: ggrepel: 1521 unlabeled data points (too many overlaps). Consider  
## increasing max.overlaps
```



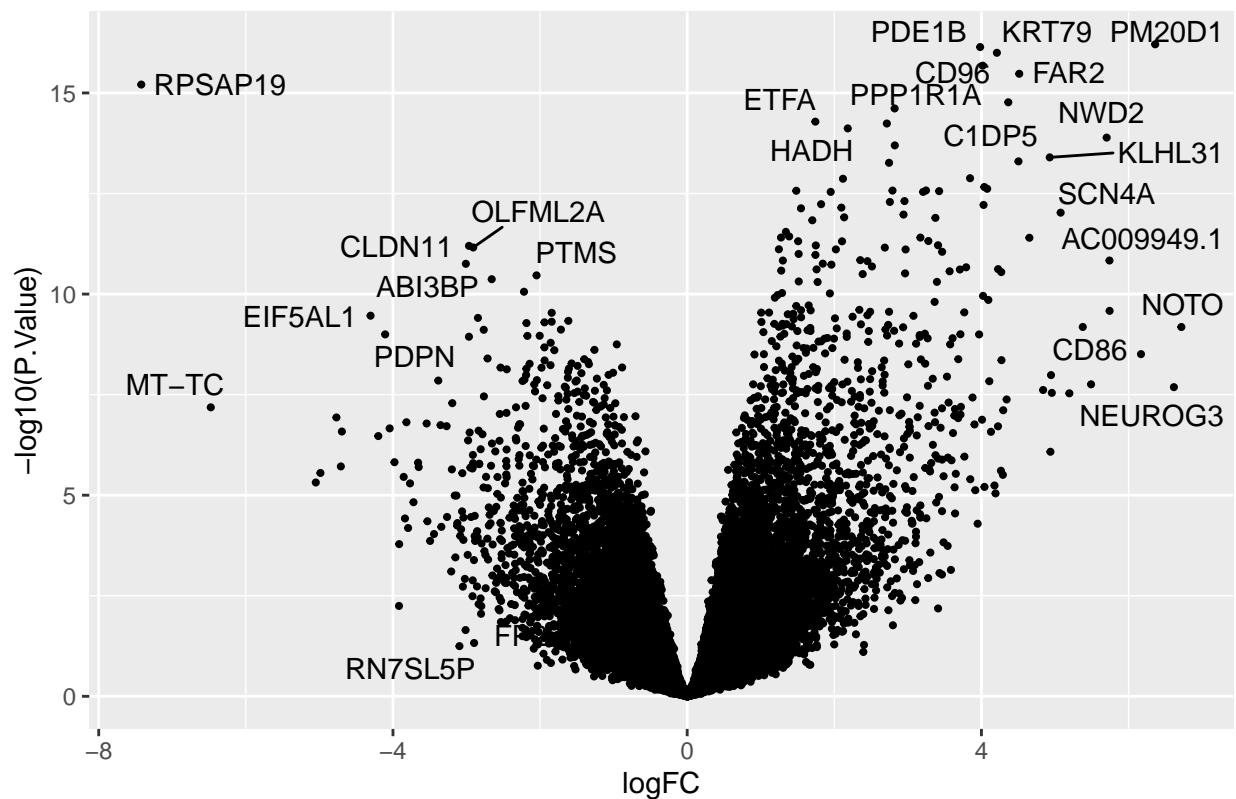
```
## Warning: ggrepel: 290 unlabeled data points (too many overlaps). Consider
## increasing max.overlaps
```

s1



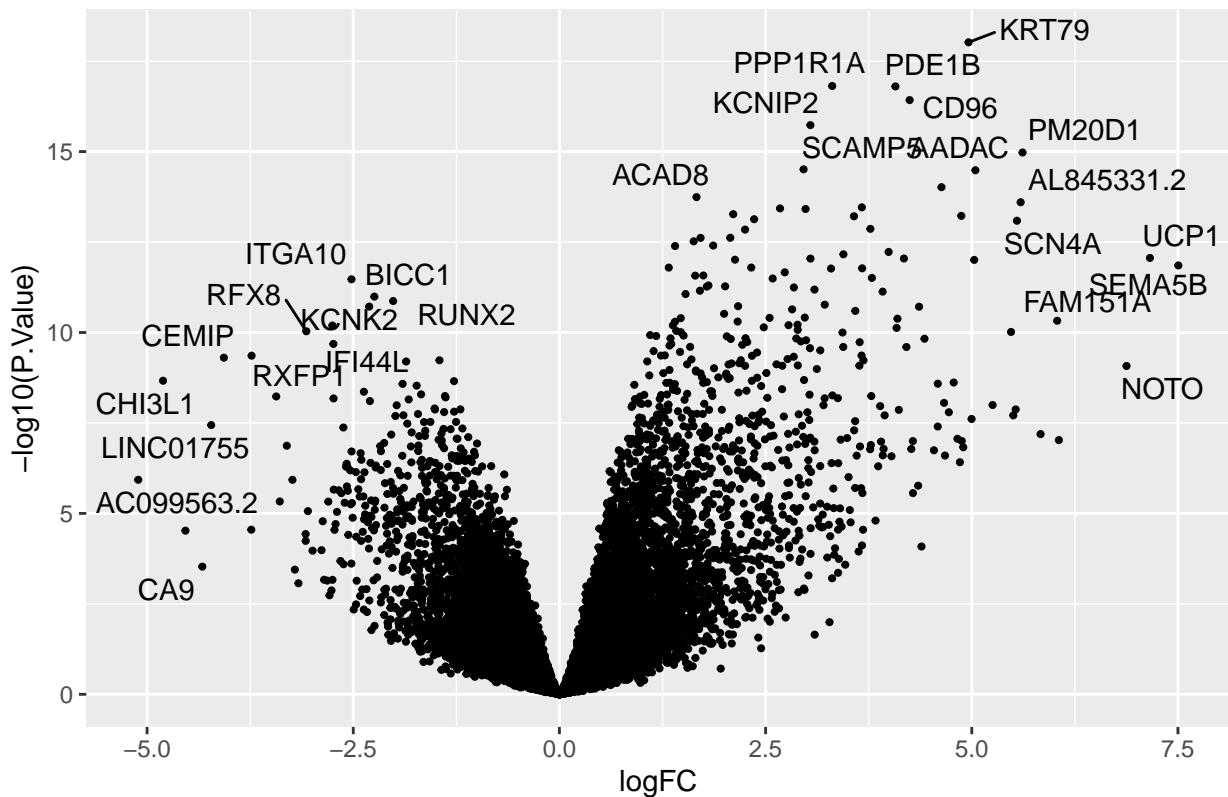
```
## Warning: ggrepel: 852 unlabeled data points (too many overlaps). Consider  
## increasing max.overlaps
```

s6



```
## Warning: ggrepel: 725 unlabeled data points (too many overlaps). Consider
## increasing max.overlaps
```

s5



GO

```
molsig <- clusterProfiler::read.gmt(here("annotations/msigdb.v2023.1.Hs.symbols.gmt"))
head(molsig); nrow(molsig)
```

```
##      term      gene
## 1 chr1p11  LINC02798
## 2 chr1p11  MTIF2P1
## 3 chr1p11  SRGAP2C
## 4 chr1p11 SRGAP2-AS1
## 5 chr1p11  LINC01691
## 6 chr1p11  NBPF26

## [1] 3961711

prefixes = c("HALLMARK", "KEGG", "REACTOME", "WP", "GOBP", "GOCC", "GOMF")
colnames(molsig) = c("term", "gene")
some.molsig = molsig[gsub("_.*","", molsig$term) %in% prefixes,]
some.molsig$term = factor(some.molsig$term)
table(gsub("_.*","", some.molsig$term))

##          GOBP      GOCC      GOMF HALLMARK      KEGG REACTOME      WP
##        642656     98915    108833      7322     12796     92769    31635
```

```

rm(molsig)
shorten = function(ont) {
  abbreviate(gsub("_", " ", tolower(ont)), minlength=40, dot=T, named = F)
}

```

GO: Genes DE in ANY donor (n=5109)

```

genes= anytable$gene_name[anytable$adj.P.Val < 0.001]
any = enricher(genes,
               universe = anytable$gene_name,
               TERM2GENE = some.molsig)
head(any)

```

		GeneRatio	BgRatio	pvalue	p.adjust
##					
## GOCC_MITOCHONDRIAL_MATRIX		275/4094			
## HALLMARK_OXIDATIVE_PHOSPHORYLATION		133/4094			
## HALLMARKADIPOGENESIS		128/4094			
## HALLMARK_EPITHELIAL_MESENCHYMAL_TRANSITION		123/4094			
## GOCC_MITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX		158/4094			
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT	REACTOME_THE_CITRIC_ACID_TCA_C	109/4094			
##					
## GOCC_MITOCHONDRIAL_MATRIX		450/13071			
## HALLMARK_OXIDATIVE_PHOSPHORYLATION		200/13071			
## HALLMARKADIPOGENESIS		197/13071			
## HALLMARK_EPITHELIAL_MESENCHYMAL_TRANSITION		197/13071			
## GOCC_MITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX		282/13071			
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT	173/13071				
##					
## GOCC_MITOCHONDRIAL_MATRIX		4.871789e-40			
## HALLMARK_OXIDATIVE_PHOSPHORYLATION		7.953976e-25			
## HALLMARKADIPOGENESIS		1.464833e-22			
## HALLMARK_EPITHELIAL_MESENCHYMAL_TRANSITION		1.419945e-19			
## GOCC_MITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX		3.907187e-18			
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT	6.394579e-18				
##					
## GOCC_MITOCHONDRIAL_MATRIX		3.964174e-36			
## HALLMARK_OXIDATIVE_PHOSPHORYLATION		3.236075e-21			

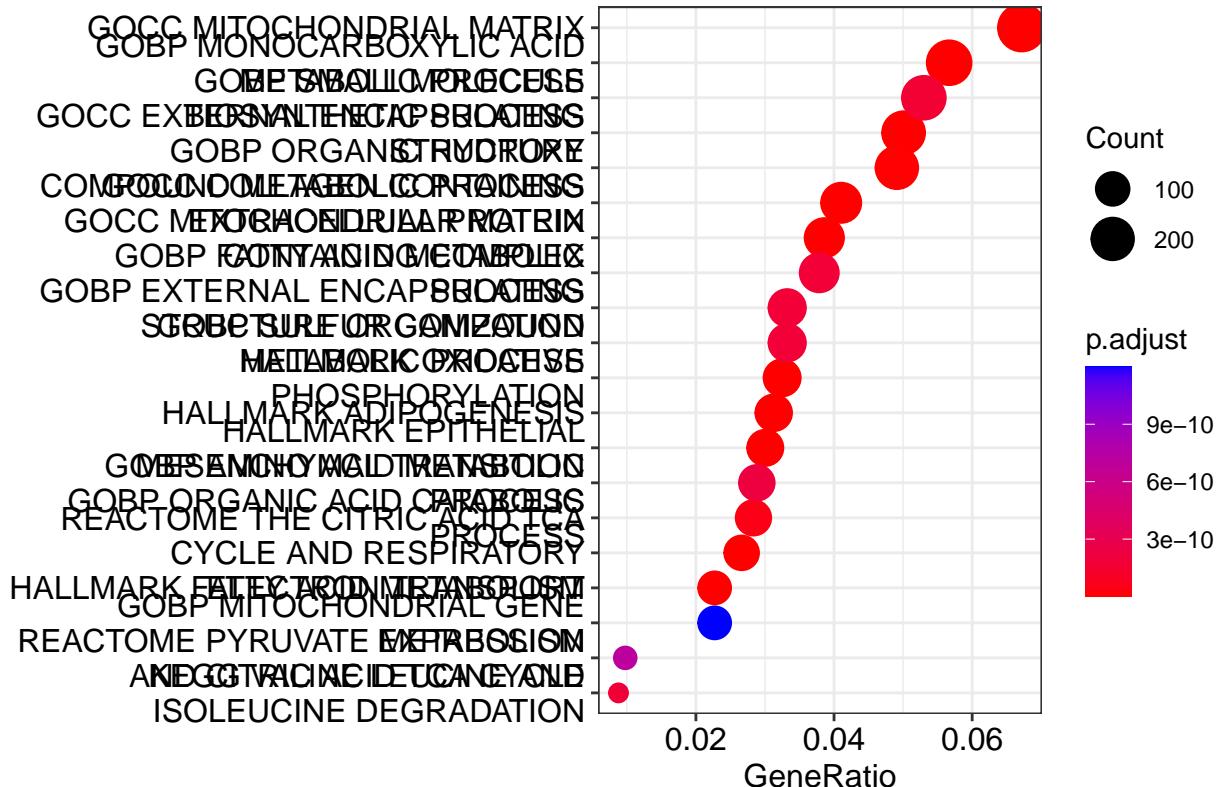
```

## HALLMARKADIPOGENESIS 3.973115e-19
## HALLMARKEPITHELIAL_MESENCHYMAL_TRANSITION 2.888522e-16
## GOCCMITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX 6.358555e-15
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT 8.672115e-15
## qvalue
## GOCCMITOCHONDRIAL_MATRIX 3.307175e-36
## HALLMARKOXIDATIVE_PHOSPHORYLATION 2.699747e-21
## HALLMARKADIPOGENESIS 3.314634e-19
## HALLMARKEPITHELIAL_MESENCHYMAL_TRANSITION 2.409796e-16
## GOCCMITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX 5.304726e-15
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT 7.234849e-15
## ETFA/ACAA2/HADH/MLYCD/ETFDH/ALI
## GOCCMITOCHONDRIAL_MATRIX
## HALLMARKOXIDATIVE_PHOSPHORYLATION
## HALLMARKADIPOGENESIS
## HALLMARKEPITHELIAL_MESENCHYMAL_TRANSITION
## GOCCMITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT
## Count
## GOCCMITOCHONDRIAL_MATRIX 275
## HALLMARKOXIDATIVE_PHOSPHORYLATION 133
## HALLMARKADIPOGENESIS 128
## HALLMARKEPITHELIAL_MESENCHYMAL_TRANSITION 123
## GOCCMITOCHONDRIAL_PROTEIN_CONTAINING_COMPLEX 158
## REACTOME_THE_CITRIC_ACID_TCA_CYCLE_AND_RESPIRATORY_ELECTRON_TRANSPORT 109

```

```
dotplot(any, showCategory = 20) + ggtitle(paste0("Any test; n=", length(genes)))
```

Any test; n=5109



```
head(results)
```

```
## TestResults matrix
##           Contrasts
##           s4 s3 s2 s1 s6 s5
## ENSG00000000003  1  1  1  1  1  1
## ENSG00000000005  1  1  0  1  1  1
## ENSG00000000419  0  0  0  0  0  0
## ENSG00000000457  0  1  1  0  0  0
## ENSG00000000460  0  0  0  0  0  0
## ENSG00000000938  1  1  1  1  1  1

decided = results[,donors]
rownames(decided) = filt$genes$gene_name[order(rownames(decided))]
head(decided); nrow(decided)
```

```
## TestResults matrix
##           Contrasts
##           s3 s4 s2 s1 s6 s5
## TSPAN6      1  1  1  1  1  1
## TNMD       1  1  0  1  1  1
## DPM1        0  0  0  0  0  0
## SCYL3       1  0  1  0  0  0
## C1orf112    0  0  0  0  0  0
## FGR        1  1  1  1  1  1
```

```

## [1] 18061

ddf = as.data.frame(decided)
ddf$str = paste0(ddf$s3,ddf$s4,ddf$s2,ddf$s1,ddf$s6,ddf$s5)

contrasts = list(beige_all = rownames(ddf)[grep("1{6}", ddf$str)],
                 s2_beige = rownames(ddf)[grep("001000", ddf$str)],
                 s4_6_beige = rownames(ddf)[grep("010010", ddf$str)],
                 s4_beige = rownames(ddf)[grep("010000", ddf$str)],
                 s6_beige = rownames(ddf)[grep("000010", ddf$str)],
                 s5_beige = rownames(ddf)[grep("000001", ddf$str)],

                 s4_6_5_beige = rownames(ddf)[grep("010011", ddf$str)],
                 s3_s2_beige = rownames(ddf)[grep("101000", ddf$str)],

                 s3_beige = rownames(ddf)[grep("100000", ddf$str)],
                 beige_not9 = rownames(ddf)[grep("111011", ddf$str)],

                 white_all = rownames(ddf)[grep("(‐1){6}", ddf$str)],
                 s2_white = rownames(ddf)[grep("00‐1000", ddf$str)],
                 s4_6_white = rownames(ddf)[grep("0‐100‐10", ddf$str)],
                 s4_white = rownames(ddf)[grep("0‐10000", ddf$str)],
                 s6_white = rownames(ddf)[grep("0000‐10", ddf$str)],
                 s5_white = rownames(ddf)[grep("00000‐1", ddf$str)],

                 s4_6_5_white = rownames(ddf)[grep("0‐100‐1‐1", ddf$str)],
                 s3_s2_white = rownames(ddf)[grep("‐10‐1000", ddf$str)],
                 white_not9 = rownames(ddf)[grep("‐1‐1‐10‐1‐1", ddf$str)],
                 s3_white = rownames(ddf)[grep("‐100000", ddf$str)])
)

str(contrasts)

## List of 20
## $ beige_all : chr [1:717] "TSPAN6" "FGR" "ICA1" "POLDIP2" ...
## $ s2_beige : chr [1:835] "CFH" "FUCA2" "AK2" "GCFC2" ...
## $ s4_6_beige : chr [1:583] "CYP51A1" "RHBDD2" "SPATA20" "CCDC124" ...
## $ s4_beige : chr [1:393] "LAS1L" "DVL2" "ABCB4" "RPUSD1" ...
## $ s6_beige : chr [1:324] "EEF1AKNMT" "BID" "GRAMD1B" "RIPOR3" ...
## $ s5_beige : chr [1:266] "WNT16" "RHOBTB2" "NUCD3" "BAK1" ...
## $ s4_6_5_beige: chr [1:307] "FAM214B" "PDK2" "NADK" "PSMC4" ...
## $ s3_s2_beige : chr [1:241] "SCYL3" "MAP3K9" "AGPS" "GLRX2" ...
## $ s3_beige : chr [1:350] "CASP10" "SLC7A2" "CDC27" "SCIN" ...
## $ beige_not9 : chr [1:197] "SLC22A16" "HCCS" "ST3GAL1" "POMT2" ...
## $ white_all : chr [1:134] "DCN" "LTF" "SLC7A14" "CLDN11" ...
## $ s2_white : chr [1:1318] "TMEM176A" "SARM1" "PLXND1" "ARHGAP33" ...
## $ s4_6_white : chr [1:467] "HOXA11" "RALA" "MATR3" "BCLAF1" ...
## $ s4_white : chr [1:388] "FBXL3" "ZFX" "FARP2" "PAFAH1B1" ...
## $ s6_white : chr [1:572] "KRIT1" "HSPB6" "IFRD1" "E2F2" ...
## $ s5_white : chr [1:390] "RBM6" "CROT" "AP2B1" "ST7L" ...
## $ s4_6_5_white: chr [1:243] "RECQL" "RANBP9" "PGM3" "CLK1" ...
## $ s3_s2_white : chr [1:231] "IFFO1" "MRC2" "PLAUR" "DNASE1L1" ...
## $ white_not9 : chr [1:246] "ETV1" "RWDD2A" "GLT8D1" "HGF" ...
## $ s3_white : chr [1:150] "CASP10" "STAB1" "CDH1" "DTNBP1" ...

```

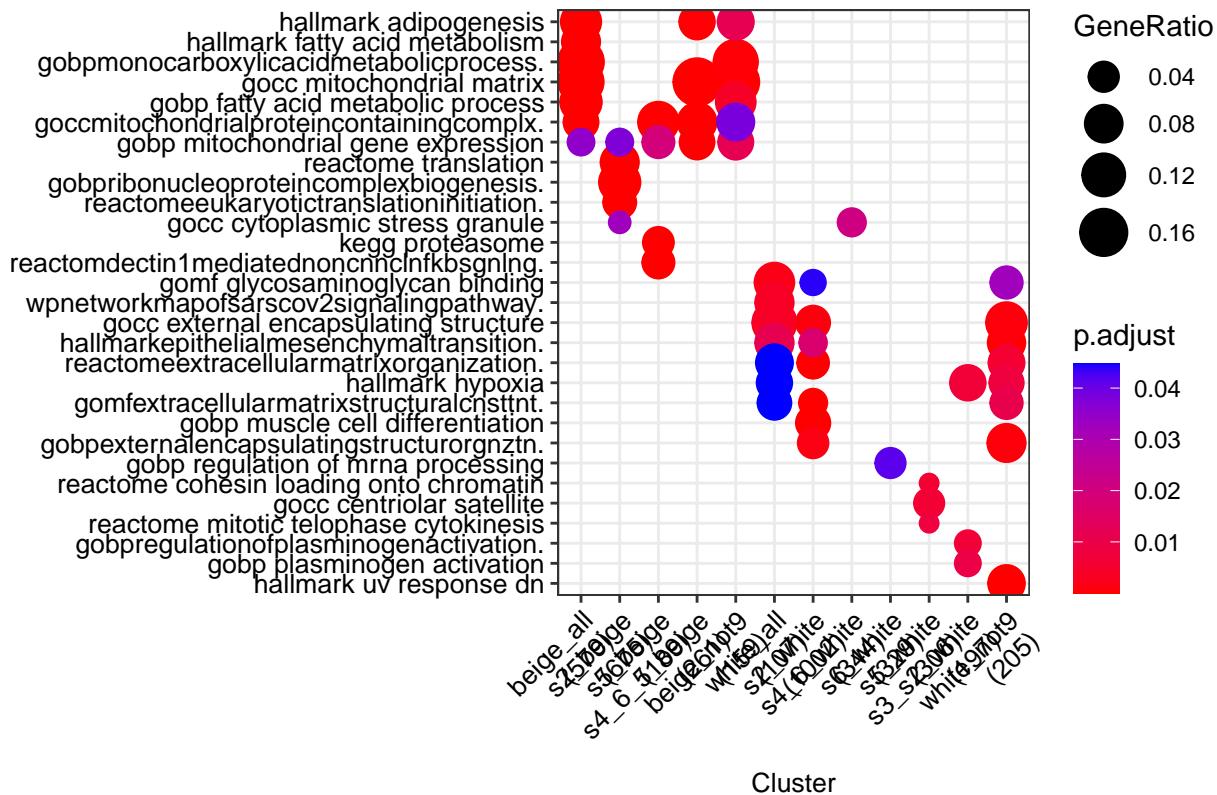
GO for combinations of donors

```
go_all = compareCluster(geneClusters = contrasts,
                        fun = "enricher",
                        TERM2GENE = some.molsig,
                        universe = anytable$gene_name)
head(go_all)

##      Cluster                      ID
## 1 beige_all      HALLMARKADIPOGENESIS
## 2 beige_all      HALLMARKFATTYACIDMETABOLISM
## 3 beige_all GOBP_MONOCARBOXYLICACIDMETABOLICPROCESS
## 4 beige_all      GOCCMITOCHONDRIALMATRIX
## 5 beige_all      GOBP_FATTY_ACID_METABOLIC_PROCESS
## 6 beige_all      GOBP_ORGANIC_ACID_CATABOLIC_PROCESS
##
##                                Description GeneRatio    BgRatio      pvalue
## 1                  HALLMARKADIPOGENESIS 57/579 197/13071 2.920537e-31
## 2      HALLMARKFATTYACIDMETABOLISM 48/579 142/13071 6.681039e-30
## 3 GOBP_MONOCARBOXYLICACIDMETABOLICPROCESS 84/579 474/13071 8.146883e-29
## 4                  GOCCMITOCHONDRIALMATRIX 81/579 450/13071 2.876342e-28
## 5      GOBP_FATTY_ACID_METABOLIC_PROCESS 62/579 303/13071 7.835987e-25
## 6      GOBP_ORGANIC_ACID_CATABOLIC_PROCESS 47/579 206/13071 4.138292e-21
##      p.adjust      qvalue
## 1 1.590817e-27 1.445820e-27
## 2 1.819581e-26 1.653733e-26
## 3 1.479202e-25 1.344379e-25
## 4 3.916859e-25 3.559852e-25
## 5 8.536524e-22 7.758452e-22
## 6 3.756880e-18 3.414454e-18
##
##      1
##      2
## 3 PDK4/ACSM3/ADIPOR2/ABHD5/NR1H3/MSMO1/ACAA1/ME1/ACSL4/MGAT4A/ALDH3A2/ACADVL/ACAT1/ACACB/MCCC1/SLC1A
## 4      POLDIP2/PDK4/SLC25A5/ACSM3/LARS2/ALAS1/MRPS35/CS/ACADVL/ACAT1/MCCC1/BCKDHB/HADHA/DLD/ACO2/M
## 5
## 6
##      Count
## 1    57
## 2    48
## 3    84
## 4    81
## 5    62
## 6    47

p = dotplot(go_all, showCategory =3, font.size=10) + theme(axis.text.x = element_text(angle=45, hjust=1))
p + scale_y_discrete(labels=shorten(levels(p$data$Description)))

## Scale for y is already present.
## Adding another scale for y, which will replace the existing scale.
```



GO beige in all donors

Using the significance table rather than the results test because its more reproducible.

```
beige_all = alltab[rowSums(alltab[grep("adj.P.Val.s[1-6]", colnames(alltab))] < 0.05) == 6,]
nrow(beige_all)
```

```
## [1] 853
```

```
go_beige_all = enricher(beige_all$gene_name,
                        TERM2GENE = some.molsig,
                        universe = anytable$gene_name)
head(go_beige_all)
```

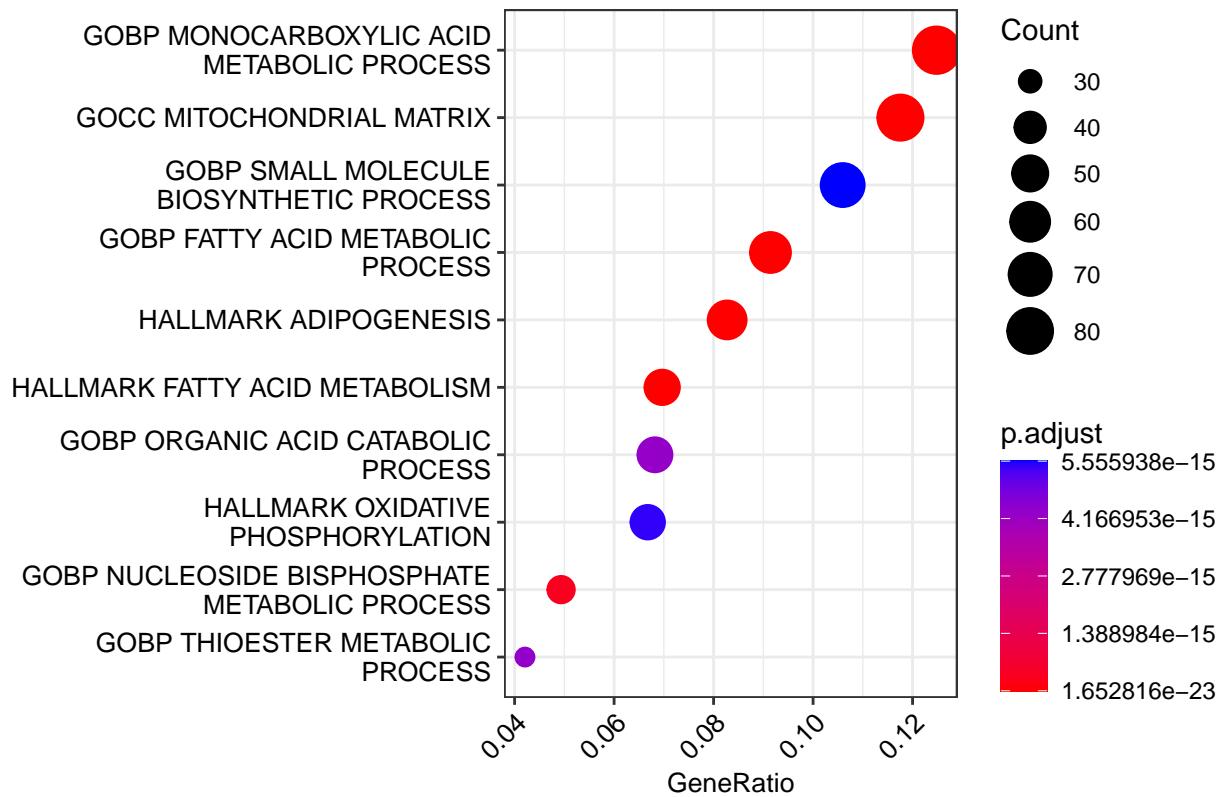
	ID
## HALLMARKADIPOGENESIS	HALLMARKADIPOGENESIS
## HALLMARKFATTYACIDMETABOLISM	HALLMARKFATTYACIDMETABOLISM
## GOBP MONOCARBOXYLIC ACID METABOLIC PROCESS	GOBP MONOCARBOXYLIC ACID METABOLIC PROCESS
## GOCC MITOCHONDRIAL MATRIX	GOCC MITOCHONDRIAL MATRIX
## GOBP FATTY ACID METABOLIC PROCESS	GOBP FATTY ACID METABOLIC PROCESS
## GOBP NUCLEOSIDE BISPHOSPHATE METABOLIC PROCESS	GOBP NUCLEOSIDE BISPHOSPHATE METABOLIC PROCESS
## HALLMARKADIPOGENESIS	Description
## HALLMARKFATTYACIDMETABOLISM	HALLMARKADIPOGENESIS

```

## GOBP_MONOCARBOXYLIC_ACID_METABOLIC_PROCESS GOBP_MONOCARBOXYLIC_ACID_METABOLIC_PROCESS
## GOCC_MITOCHONDRIAL_MATRIX GOCC_MITOCHONDRIAL_MATRIX
## GOBP_FATTY_ACID_METABOLIC_PROCESS GOBP_FATTY_ACID_METABOLIC_PROCESS
## GOBP_NUCLEOSIDE_BISPHEROPHATE_METABOLIC_PROCESS GOBP_NUCLEOSIDE_BISPHEROPHATE_METABOLIC_PROCESS
## GeneRatio BgRatio pvalue
## HALLMARKADIPOGENESIS 57/689 197/13071 2.785333e-27
## HALLMARKFATTY_ACID_METABOLISM 48/689 142/13071 1.734421e-26
## GOBP_MONOCARBOXYLIC_ACID_METABOLIC_PROCESS 86/689 474/13071 8.604599e-25
## GOCC_MITOCHONDRIAL_MATRIX 81/689 450/13071 3.861167e-23
## GOBP_FATTY_ACID_METABOLIC_PROCESS 63/689 303/13071 1.575969e-21
## GOBP_NUCLEOSIDE_BISPHEROPHATE_METABOLIC_PROCESS 34/689 100/13071 3.407372e-19
## p.adjust qvalue
## HALLMARKADIPOGENESIS 1.652816e-23 1.495577e-23
## HALLMARKFATTY_ACID_METABOLISM 5.146027e-23 4.656464e-23
## GOBP_MONOCARBOXYLIC_ACID_METABOLIC_PROCESS 1.701990e-21 1.540072e-21
## GOCC_MITOCHONDRIAL_MATRIX 5.728041e-20 5.183108e-20
## GOBP_FATTY_ACID_METABOLIC_PROCESS 1.870360e-18 1.692424e-18
## GOBP_NUCLEOSIDE_BISPHEROPHATE_METABOLIC_PROCESS 3.369891e-16 3.049299e-16
##
## HALLMARKADIPOGENESIS
## HALLMARKFATTY_ACID_METABOLISM
## GOBP_MONOCARBOXYLIC_ACID_METABOLIC_PROCESS PDK4/ACSM3/ADIPOR2/ABHD5/NR1H3/MSMO1/ACAA1/ME1/PKM/AC
## GOCC_MITOCHONDRIAL_MATRIX POLDIP2/PDK4/SLC25A5/ACSM3/LARS2/ALAS
## GOBP_FATTY_ACID_METABOLIC_PROCESS
## GOBP_NUCLEOSIDE_BISPHEROPHATE_METABOLIC_PROCESS
## Count
## HALLMARKADIPOGENESIS 57
## HALLMARKFATTY_ACID_METABOLISM 48
## GOBP_MONOCARBOXYLIC_ACID_METABOLIC_PROCESS 86
## GOCC_MITOCHONDRIAL_MATRIX 81
## GOBP_FATTY_ACID_METABOLIC_PROCESS 63
## GOBP_NUCLEOSIDE_BISPHEROPHATE_METABOLIC_PROCESS 34

p = dotplot(go_beige_all, showCategory =10, font.size=10) + theme(axis.text.x = element_text(angle=45,

```



FPKM tables

```

lib_rpkm = data.frame(rpkm(filt, normalize.lib.sizes=TRUE))
colnames(lib_rpkm) = paste(filt$samples$donor.condition, "_rep", filt$samples$rep, sep="")
lib_rpkm = lib_rpkm[order(colnames(lib_rpkm))]

format_rpkm = merge(filt$genes, lib_rpkm,
                     by.x="Geneid", by.y = 'row.names', sort=FALSE)
head(format_rpkm)

##           Geneid Length gene_name
## 1 ENSG00000000003    4536   TSPAN6
## 2 ENSG00000000005    1476    TMMD
## 3 ENSG00000000419    1207    DPM1
## 4 ENSG00000000457    6883    SCYL3
## 5 ENSG00000000460    5970  C1orf112
## 6 ENSG00000000938    3382     FGR
##                                         description gene_biotype
## 1                               tetraspanin 6 protein_coding
## 2                               tenomodulin protein_coding
## 3 dolichyl-phosphate mannosyltransferase subunit 1, catalytic protein_coding
## 4                               SCY1 like pseudokinase 3 protein_coding
## 5 chromosome 1 open reading frame 112 protein_coding

```

```

## 6          FGR proto-oncogene, Src family tyrosine kinase  protein_coding
##   ensembl_gene_id_version subject1.beige_rep1 subject1.beige_rep2
## 1      ENSG00000000003.15      6.9254899      7.1041492
## 2      ENSG00000000005.6      2.2801229      1.5381684
## 3      ENSG00000000419.12     33.4367039     31.4487083
## 4      ENSG00000000457.14     1.9757409     1.8447962
## 5      ENSG00000000460.17     0.3906274     0.3652596
## 6      ENSG00000000938.13     3.5126295     2.5525319
##   subject1.beige_rep3 subject1.white_rep1 subject1.white_rep2
## 1      7.0164629      4.7128194      5.0708211
## 2      1.0495540      0.4615557      0.1656141
## 3      27.0153634     33.0186300     32.4906301
## 4      1.7676057      1.4624651      1.4171984
## 5      0.4261506      0.3954615      0.4328556
## 6      2.8674982      0.7050256      0.9189715
##   subject1.white_rep3 subject2.beige_rep1 subject2.beige_rep2
## 1      5.4418345     10.9284653     9.774450
## 2      0.3331110     16.3059522    19.540289
## 3      34.7173572     33.3397649     27.154128
## 4      1.3734576     2.1309163     2.059405
## 5      0.3949397     0.4049026     0.418865
## 6      0.7467195     6.3532910     4.664816
##   subject2.beige_rep3 subject2.white_rep1 subject2.white_rep2
## 1      16.8977361     6.4045447     6.887769
## 2      56.2187609     23.9793882    33.149772
## 3      36.4339823     28.3932325    32.103143
## 4      2.9304897     1.5831367     1.618697
## 5      0.6188623     0.3936126     0.406746
## 6      2.9052807     0.8177922     0.756395
##   subject2.white_rep3 subject3.beige_rep1 subject3.beige_rep2
## 1      6.1627964     10.5844425     9.787839
## 2      33.5831338     2.5010606     2.553310
## 3      26.8767081     39.3488030    31.606617
## 4      1.5491843     2.2084212     2.076156
## 5      0.3295161     0.5057674     0.436492
## 6      1.0818338     9.3804593     6.719158
##   subject3.beige_rep3 subject3.white_rep1 subject3.white_rep2
## 1      8.801945      6.3522863     6.0961385
## 2      3.315264      0.2897050     0.5306761
## 3      31.369481      31.2666494    35.9181825
## 4      2.056277      1.5754197     1.2810999
## 5      0.501716      0.3948589     0.4949906
## 6      5.440378      0.8493867     1.1545008
##   subject3.white_rep3 subject4.beige_rep1 subject4.beige_rep2
## 1      6.2487150      9.2558572     9.7210231
## 2      0.6497380      1.8138929     3.4756272
## 3      37.0786698     33.5048504    37.0758923
## 4      1.6494505      2.1226017     2.2437674
## 5      0.4316152      0.4215859     0.5620584
## 6      0.8506919      7.4686360     5.3233254
##   subject4.beige_rep3 subject4.white_rep1 subject4.white_rep2
## 1      8.9397143      8.7135345     8.7486479
## 2      1.6126162      1.0275705     0.6455009
## 3      27.5773901     36.3470915    34.3252630

```

```

## 4      2.0604610    1.8713614    1.5792702
## 5      0.4256922    0.7014885    0.5754954
## 6      4.7395932    0.5689426    0.6487976
##   subject4.white_rep3 subject5.beige_rep1 subject5.beige_rep2
## 1      9.6065895    10.6304243   9.6054108
## 2      0.2907158    5.7198626    5.4055047
## 3      33.3195527   30.3958272   30.0042303
## 4      1.6832203    1.8713287   1.9437106
## 5      0.4114251    0.3833146   0.3504826
## 6      0.6868834    7.3511831   4.6432371
##   subject5.beige_rep3 subject5.white_rep1 subject5.white_rep2
## 1      9.9677634    6.5873342   6.3455039
## 2      7.5206419    1.3830061   1.5991558
## 3      34.5704866   34.8785961   35.4498533
## 4      1.9163335    1.3438945   1.6117483
## 5      0.3321905    0.4575574   0.3690112
## 6      4.8957811    0.9359907   0.7870949
##   subject5.white_rep3 subject6.beige_rep1 subject6.beige_rep2
## 1      5.9682040    11.1349030  12.557102
## 2      2.1362231    7.3100161   11.403522
## 3      22.5609106   34.2276292  40.814843
## 4      1.5871360    1.8944114   2.187790
## 5      0.5975049    0.3548748   0.360928
## 6      0.4520281    12.7224387  13.732668
##   subject6.beige_rep3 subject6.white_rep1 subject6.white_rep2
## 1      11.6781141   7.902057   8.8622896
## 2      7.8596296    2.052549   2.5683056
## 3      39.7817825   37.952305  35.7383505
## 4      2.1737580    1.803030   1.8126365
## 5      0.4118008    0.440210   0.4430891
## 6      11.8919911   1.662068   2.4080463
##   subject6.white_rep3
## 1      8.3762978
## 2      1.7968096
## 3      33.8286255
## 4      1.3539377
## 5      0.4257265
## 6      1.8079666

```

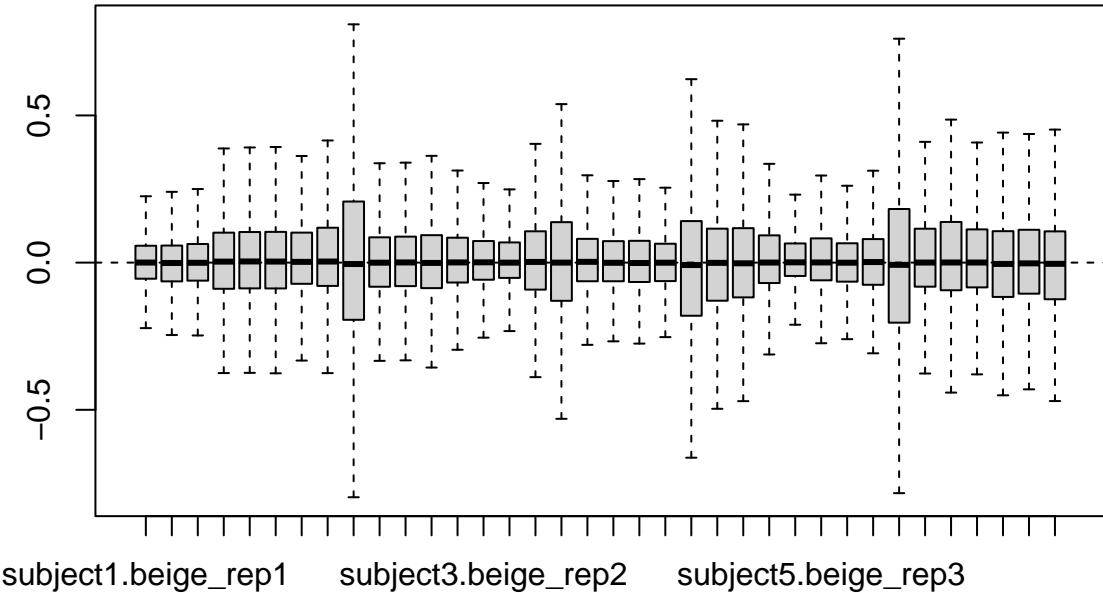
```
rowMeans(format_rpkm[format_rpkm$gene_name == "PPARG",c(7:ncol(format_rpkm))]) #Average RPKM 36
```

```

##      5319
## 36.46861

```

```
plotRLE(data.matrix(format_rpkm[7:ncol(format_rpkm)]), outline=FALSE) #check normalisation
```



```
write.table(format_rpkm, sep='\t', row.names = FALSE, quote = F,
            file=here("03limma/beige_day15_rpkm_tmm.tab"))
```

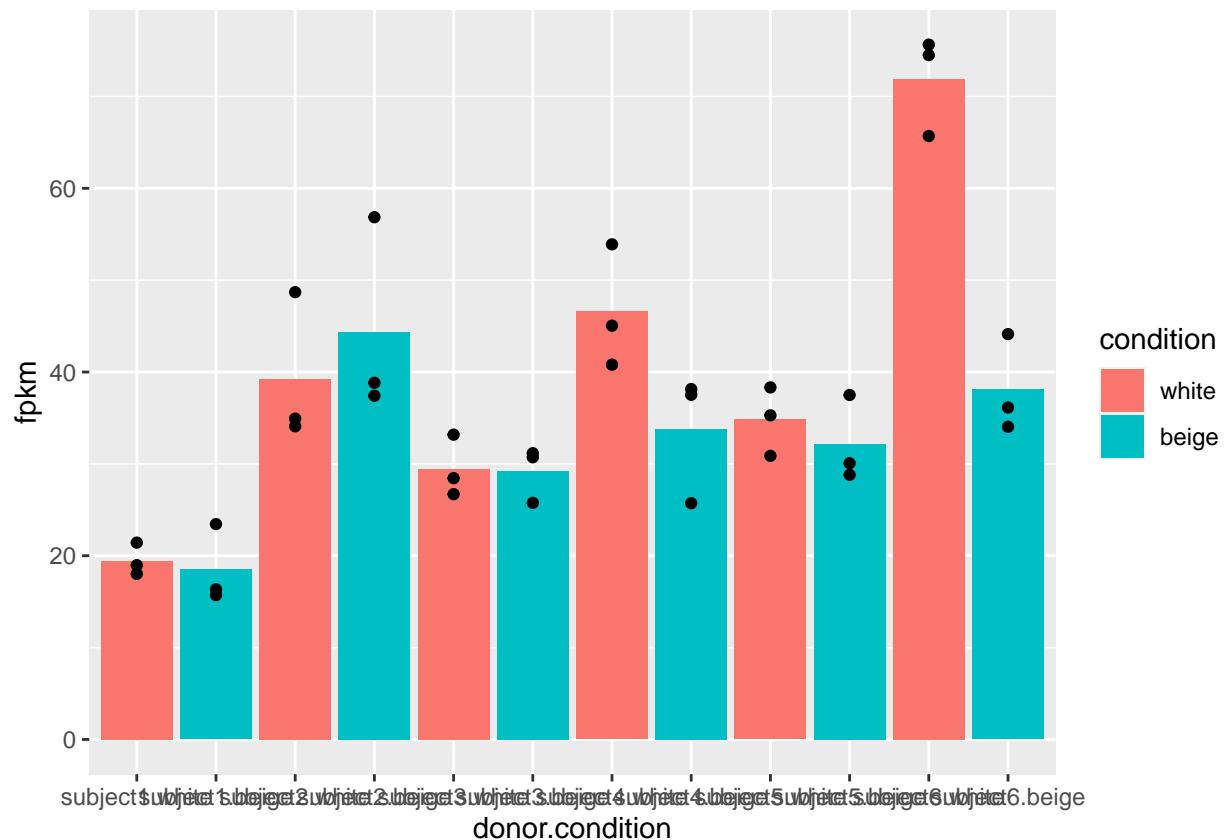
```
library(tidyr)
long = pivot_longer(format_rpkm, 7:ncol(format_rpkm), names_to = "biorep", values_to ="fpkm")
long$donor = factor(gsub("\\..*", "", long$biorep))
long$condition = factor(gsub(".*\\"., "", gsub("_rep.", "", long$biorep)), levels=c("white","beige"))
long$rep = gsub(".*_","", long$biorep)
long$donor.condition = factor(paste(long$donor, long$condition, sep="."),
                               levels = paste(rep(levels(long$donor),each=2), rep(c("white","beige"), 6)))
head(long)

## # A tibble: 6 x 12
##   Geneid Length gene_name description gene_biotype ensembl_gene_id vers~1 biorep
##   <chr>    <dbl>  <chr>      <chr>      <chr>      <chr>
## 1 ENSG0~    4536 TSPAN6    "tetraspan~ protein_cod~ ENSG00000000003.15    subje~
## 2 ENSG0~    4536 TSPAN6    "tetraspan~ protein_cod~ ENSG00000000003.15    subje~
## 3 ENSG0~    4536 TSPAN6    "tetraspan~ protein_cod~ ENSG00000000003.15    subje~
## 4 ENSG0~    4536 TSPAN6    "tetraspan~ protein_cod~ ENSG00000000003.15    subje~
## 5 ENSG0~    4536 TSPAN6    "tetraspan~ protein_cod~ ENSG00000000003.15    subje~
## 6 ENSG0~    4536 TSPAN6    "tetraspan~ protein_cod~ ENSG00000000003.15    subje~
## # i abbreviated name: 1: ensembl_gene_id_version
## # i 5 more variables: fpkm <dbl>, donor <fct>, condition <fct>, rep <chr>,
## #   donor.condition <fct>
```

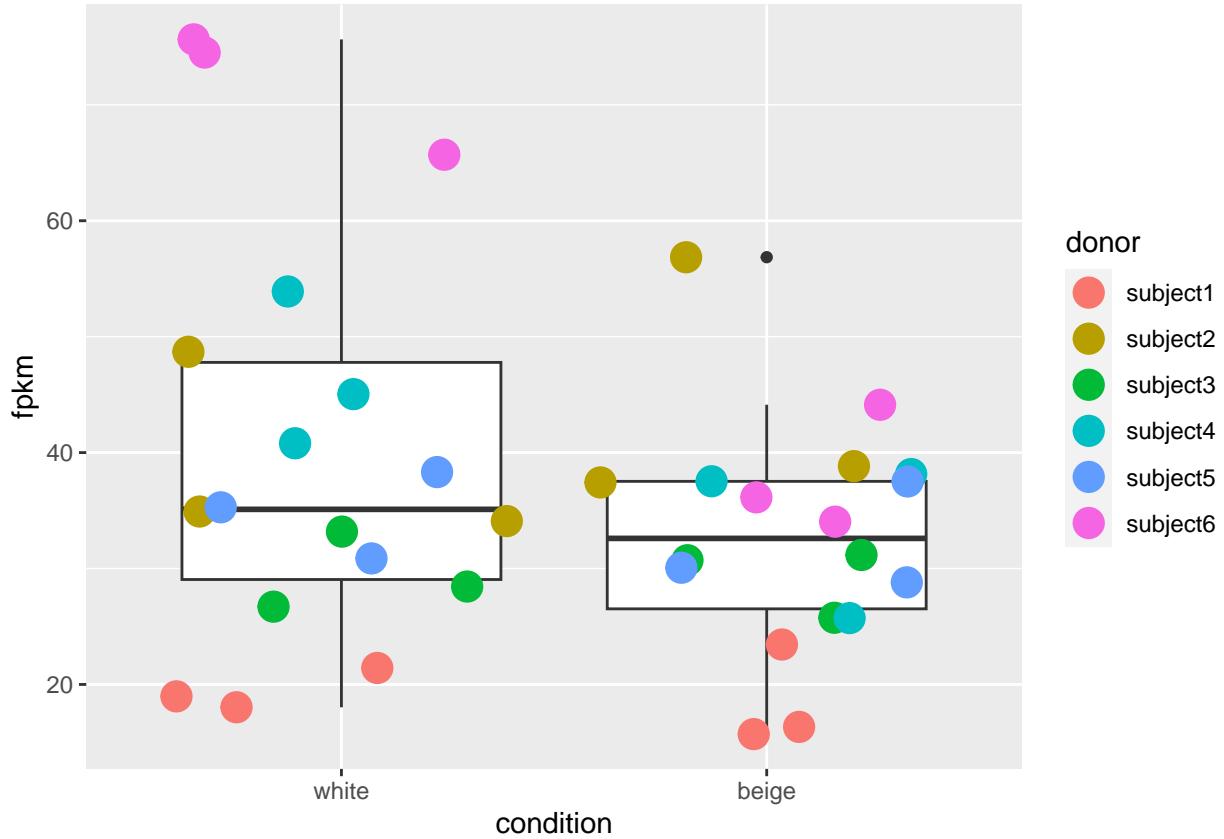
```
long = group_by(long, Geneid) %>% mutate(zscore= scale(fpkm))
head(long)
```

```
## # A tibble: 6 x 13
## # Groups:   Geneid [1]
##   Geneid Length gene_name description gene_biotype ensembl_gene_id_ver~1 biorep
##   <chr>    <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 ENSG0~    4536 TSPAN6 "tetraspan~ protein_cod~ ENSG000000000003.15 subje~
## 2 ENSG0~    4536 TSPAN6 "tetraspan~ protein_cod~ ENSG000000000003.15 subje~
## 3 ENSG0~    4536 TSPAN6 "tetraspan~ protein_cod~ ENSG000000000003.15 subje~
## 4 ENSG0~    4536 TSPAN6 "tetraspan~ protein_cod~ ENSG000000000003.15 subje~
## 5 ENSG0~    4536 TSPAN6 "tetraspan~ protein_cod~ ENSG000000000003.15 subje~
## 6 ENSG0~    4536 TSPAN6 "tetraspan~ protein_cod~ ENSG000000000003.15 subje~
## # i abbreviated name: 1: ensembl_gene_id_version
## # i 6 more variables: fpkm <dbl>, donor <fct>, condition <fct>, rep <chr>,
## #   donor.condition <fct>, zscore <dbl[,1]>
```

```
ggplot(long[long$gene_name == "PPARG",]) + geom_bar(stat="summary",fun="mean", aes(x=donor.condition, y=fpkm)) +
  geom_point(aes(x=donor.condition, y=fpkm))
```



```
ggplot(long[long$gene_name == "PPARG",]) + geom_boxplot(aes(x=condition, y=fpkm)) +
  geom_jitter(aes(x=condition, y=fpkm, colour=donor), size=5)
```



```
save(long, file=here("03limma/rpkm_rep_for_plotting.RData"))
```

```
filt$samples$group = factor(filt$samples$donor.condition)
group_rpkm = data.frame(rpkmByGroup(filt, normalize.lib.sizes=TRUE))
colSums(group_rpkm)
```

```
## subject1.beige subject1.white subject2.beige subject2.white subject3.beige
##      244925.7      228831.0      349289.1      259712.5      296811.1
## subject3.white subject4.beige subject4.white subject5.beige subject5.white
##      263803.7      288923.2      313523.5      279363.2      259112.1
## subject6.beige subject6.white
##      289573.1      327761.0
```

```
format_grpkm = merge(filt$genes, group_rpkm,
                      by.x="Geneid", by.y = 'row.names', sort=FALSE)
head(format_grpkm)
```

	Geneid	Length	gene_name
## 1	ENSG00000000003	4536	TSPAN6
## 2	ENSG00000000005	1476	TNMD
## 3	ENSG00000000419	1207	DPM1
## 4	ENSG00000000457	6883	SCYL3
## 5	ENSG00000000460	5970	C1orf112

```

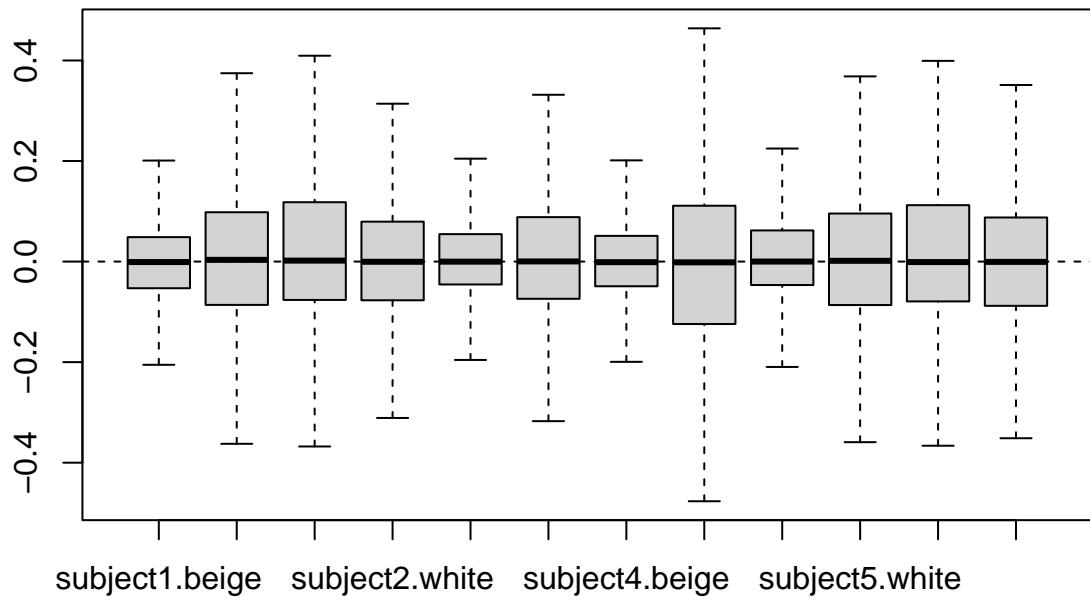
## 6 ENSG00000000938    3382      FGR
##                                         description  gene_biotype
## 1                                         tetraspanin 6 protein_coding
## 2                                         tenomodulin protein_coding
## 3 dolichyl-phosphate mannosyltransferase subunit 1, catalytic protein_coding
## 4                                         SCY1 like pseudokinase 3 protein_coding
## 5                                         chromosome 1 open reading frame 112 protein_coding
## 6 FGR proto-oncogene, Src family tyrosine kinase protein_coding
##   ensembl_gene_id_version subject1.beige subject1.white subject2.beige
## 1     ENSG00000000003.15      7.015370    5.0752183    12.5300941
## 2     ENSG00000000005.6       1.627583    0.3200113    30.6612441
## 3     ENSG00000000419.12    30.635853   33.4089794    32.3055705
## 4     ENSG00000000457.14    1.862880    1.4176901    2.3720554
## 5     ENSG00000000460.17    0.393713    0.4077405    0.4788014
## 6     ENSG00000000938.13    2.977751    0.7902128    4.6470874
##   subject2.white subject3.beige subject3.white subject4.beige subject4.white
## 1     6.4851444   9.7247952   6.2324136   9.3056202   9.0232992
## 2     30.2448972   2.7906896   0.4912616   2.3021246   0.6401144
## 3     29.1252400   34.1100563   34.7548040   32.7208903   34.6618668
## 4     1.5836657   2.1136964   1.5022350   2.1423301   1.7105704
## 5     0.3762281   0.4816479   0.4403470   0.4698593   0.5608791
## 6     0.8865716   7.1807572   0.9509210   5.8448505   0.6362375
##   subject5.beige subject5.white subject6.beige subject6.white
## 1     10.0681020   6.3019890  11.7900221   8.3815053
## 2     6.2119587   1.6856768   8.8600561   2.1400068
## 3     31.6549387  30.9930433  38.2724535  35.8339890
## 4     1.9104027   1.5130770   2.0849212   1.6548458
## 5     0.3557019   0.4699746   0.3751493   0.4361585
## 6     5.6321927   0.7380337  12.7832922   1.9632742

rowMeans(format_grpkm[format_grpkm$gene_name == "PPARG",c(7:ncol(format_grpkm))])#expr matches the repl

##      5319
## 36.46869

plotRLE(data.matrix(format_grpkm[7:ncol(format_grpkm)]), outline=FALSE)

```



```
write.table(format_grpkm, sep='\t', row.names = FALSE, quote=F,  
           file=here("03limma/beige_day15_rpkm_tmm_means.tab"))
```