

go_networks_plus_trifid

2023-12-15

Figure 2 G, H

```
library(dplyr)
library(tidyr)
library(ggplot2)
library(enrichplot)
library(clusterProfiler)
library(here); i_am("R/17_go_networks_plus_trifid.Rmd")
```

```
figs = here("R/plots")
load(file.path(figs, "Themogenesis_object.RData"))
head(specific)
```

```
##
## HALLMARK_ADIPOGENESIS HALLMARK_ADIPOGENESIS
## HALLMARK_FATTY_ACID_METABOLISM HALLMARK_FATTY_ACID_METABOLISM
## WP_THERMOGENESIS WP_THERMOGENESIS
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE
## GOBP_ADAPTIVE_THERMOGENESIS GOBP_ADAPTIVE_THERMOGENESIS
##
## HALLMARK_ADIPOGENESIS HALLMARK_ADIPOGENESIS
## HALLMARK_FATTY_ACID_METABOLISM HALLMARK_FATTY_ACID_METABOLISM
## WP_THERMOGENESIS WP_THERMOGENESIS
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE
## GOBP_ADAPTIVE_THERMOGENESIS GOBP_ADAPTIVE_THERMOGENESIS
##
## GeneRatio BgRatio
## HALLMARK_ADIPOGENESIS 9/35 181/1124
## HALLMARK_FATTY_ACID_METABOLISM 7/35 129/1124
## WP_THERMOGENESIS 4/35 80/1124
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS 3/35 58/1124
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE 1/35 13/1124
## GOBP_ADAPTIVE_THERMOGENESIS 4/35 100/1124
##
## pvalue p.adjust
## HALLMARK_ADIPOGENESIS 0.09506160 0.4806583
## HALLMARK_FATTY_ACID_METABOLISM 0.09613165 0.4806583
## WP_THERMOGENESIS 0.23428577 0.5027113
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS 0.26843887 0.5027113
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE 0.33866058 0.5027113
## GOBP_ADAPTIVE_THERMOGENESIS 0.37939955 0.5027113
##
## qvalue
## HALLMARK_ADIPOGENESIS 0.4806583
```

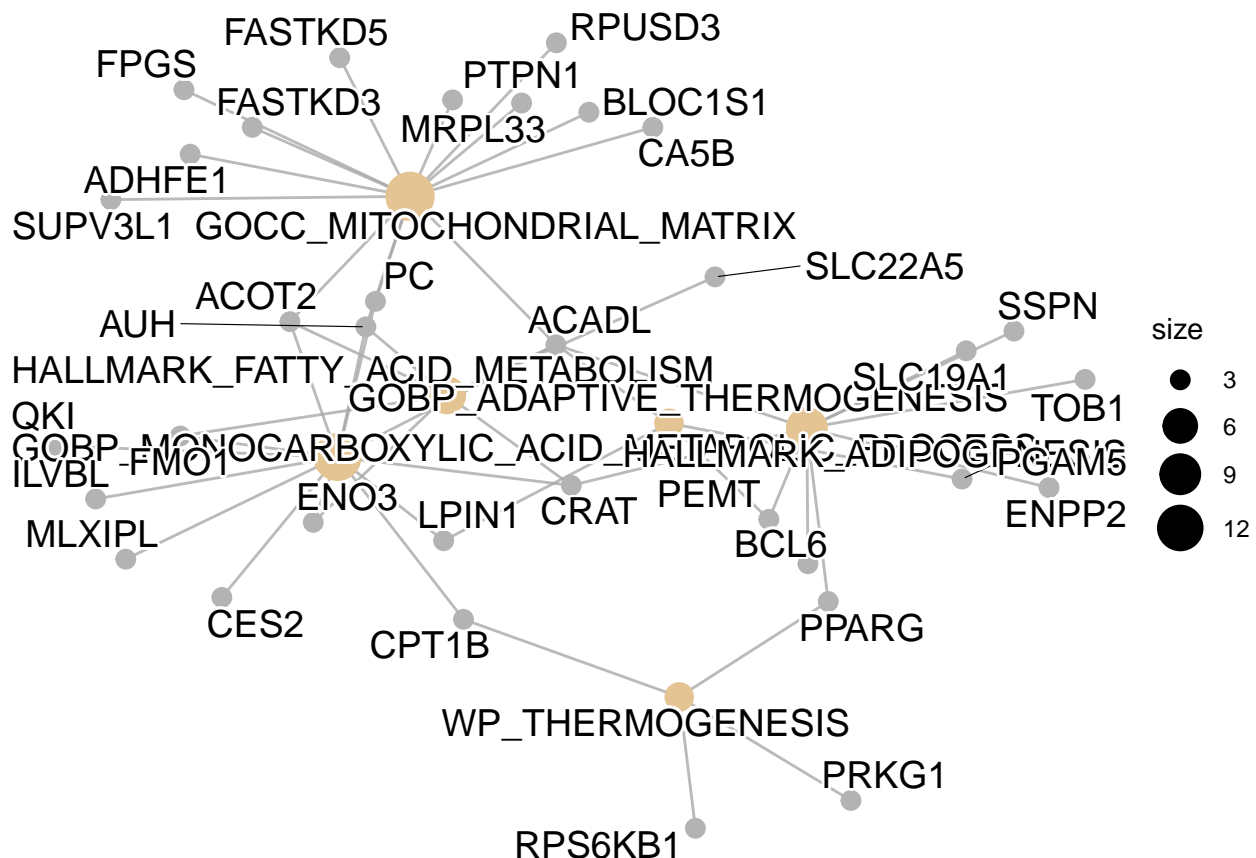
```

## HALLMARK_FATTY_ACID_METABOLISM 0.4806583
## WP_THERMOGENESIS 0.5027113
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS 0.5027113
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE 0.5027113
## GOBP_ADAPTIVE_THERMOGENESIS 0.5027113
##
## HALLMARK_ADIPOGENESIS PEMT/PPARG/ENPP2/TOB1/BCL6/ACADL/CRAT/SLC19A1/
## HALLMARK_FATTY_ACID_METABOLISM FMO1/ACADL/CRAT/SLC22A5/AUH/ACOT2/
## WP_THERMOGENESIS PPARG/PRKG1/CPT1B/RPS6KB1/
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS PEMT/LPIN1/
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE
## GOBP_ADAPTIVE_THERMOGENESIS PEMT/LPIN1/PGAM5/
##
## Count
## HALLMARK_ADIPOGENESIS 9
## HALLMARK_FATTY_ACID_METABOLISM 7
## WP_THERMOGENESIS 4
## GOBP_POSITIVE_REGULATION_OF_COLD_INDUCED_THERMOGENESIS 3
## WP_DIFFERENTIATION_OF_WHITE_AND_BROWN_ADIPOCYTE 1
## GOBP_ADAPTIVE_THERMOGENESIS 4

```

```
specific = arrange(specific, desc(Count))
```

```
cnetplot(specific, showCategory=6, categorySize="geneNum")
```



depicts the linkages of genes and biological concepts (e.g. GO terms or KEGG pathways) as a network.
to plot the trifold score, we just need a sorted list of these with names of genes

```
trifid = read.delim(here("31_leafcutter", "trifid_DIFFERENCE_with_Alt_introns.tsv"))
diff = trifid$trifid_diff
names(diff) = trifid$gene_beige
diff = diff[order(diff, decreasing=T)]
summary(diff)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.     NA's
## -1.10000 -0.30421  0.00000 -0.01699  0.30189  1.10000         2
```

```
shorten = function(ont) {
  no_beg = gsub("^(GO..|HALLMARK|WP|REACTOME)_", "", ont)
  abb = abbreviate(stringr::str_to_title(gsub("_", " ", no_beg)), minlength=40, dot=T, named = F)
  return(abb)
}
```

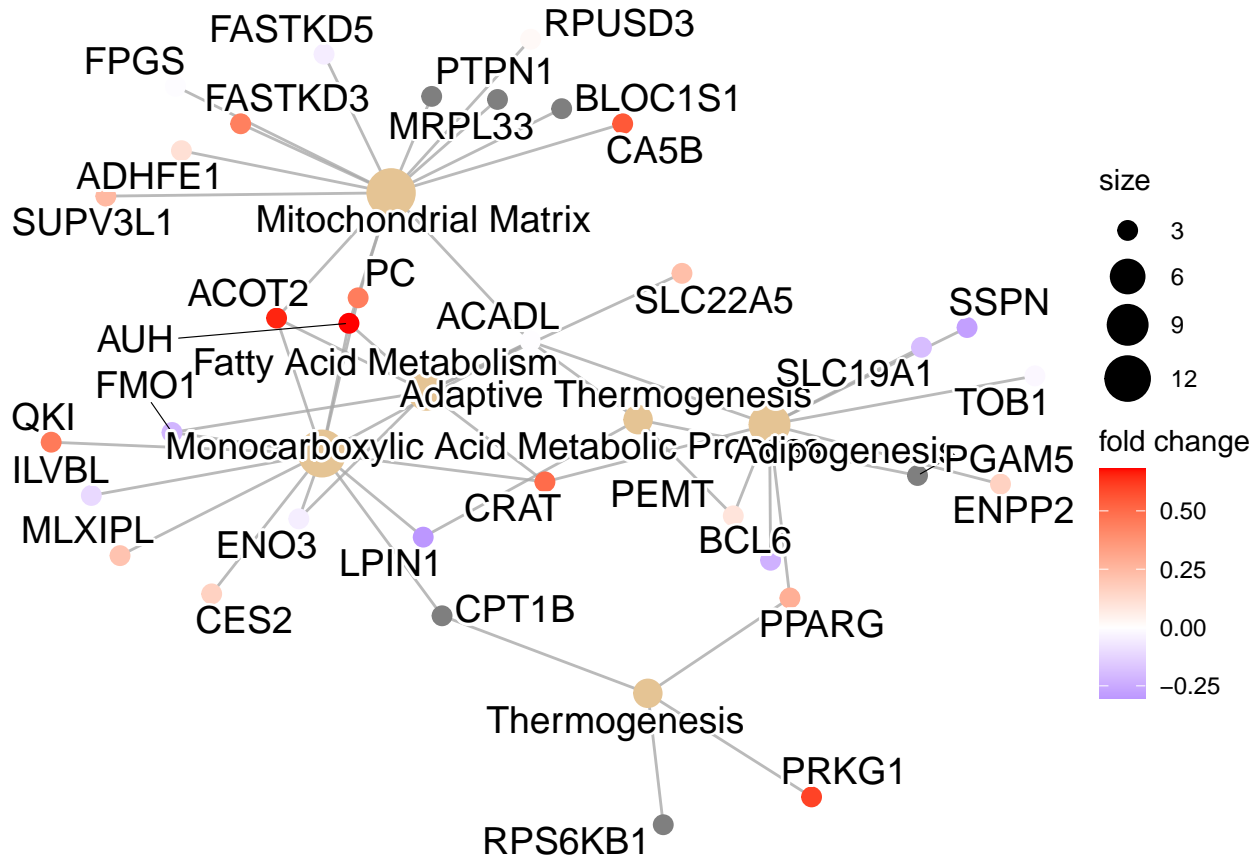
```
specific = mutate(specific, Description = shorten(ID) )
```

```
cnetplot(specific, showCategory=6, categorySize="geneNum", foldChange=diff)
```

```
## Warning in cnetplot.enrichResult(x, ...): Use 'color.params = list(foldChange = your_value)' instead
## The foldChange parameter will be removed in the next version.
```

```
## Scale for size is already present.
```

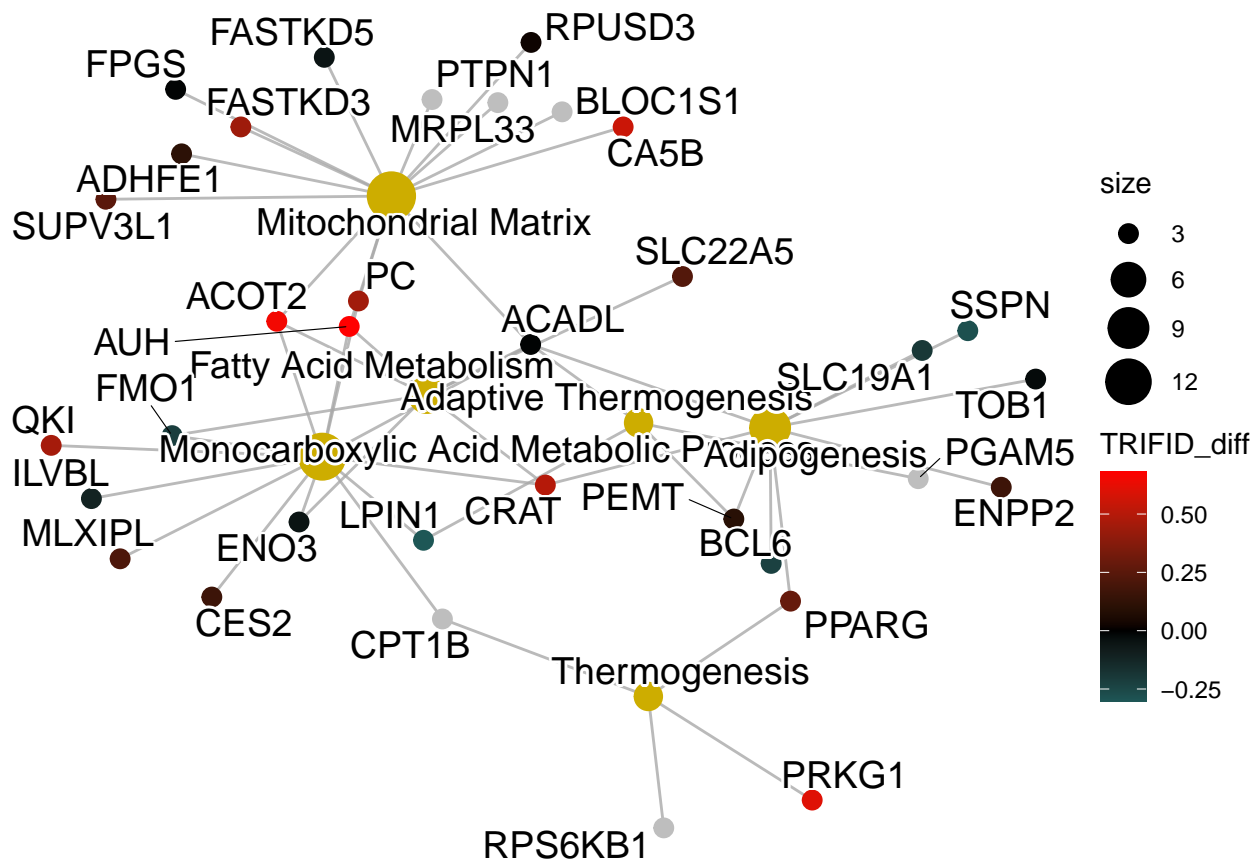
```
## Adding another scale for size, which will replace the existing scale.
```



```
cnetplot(specific, showCategory=6, categorySize="geneNum", color.params = list(foldChange = diff),
        color_category='gold3' ) + scale_colour_gradient2(high="red", low="cyan3", mid="black", na.val
```

```
## Warning in cnetplot.enrichResult(x, ...): Use 'color.params = list(category = your_value)' instead of
## The color_category parameter will be removed in the next version.
```

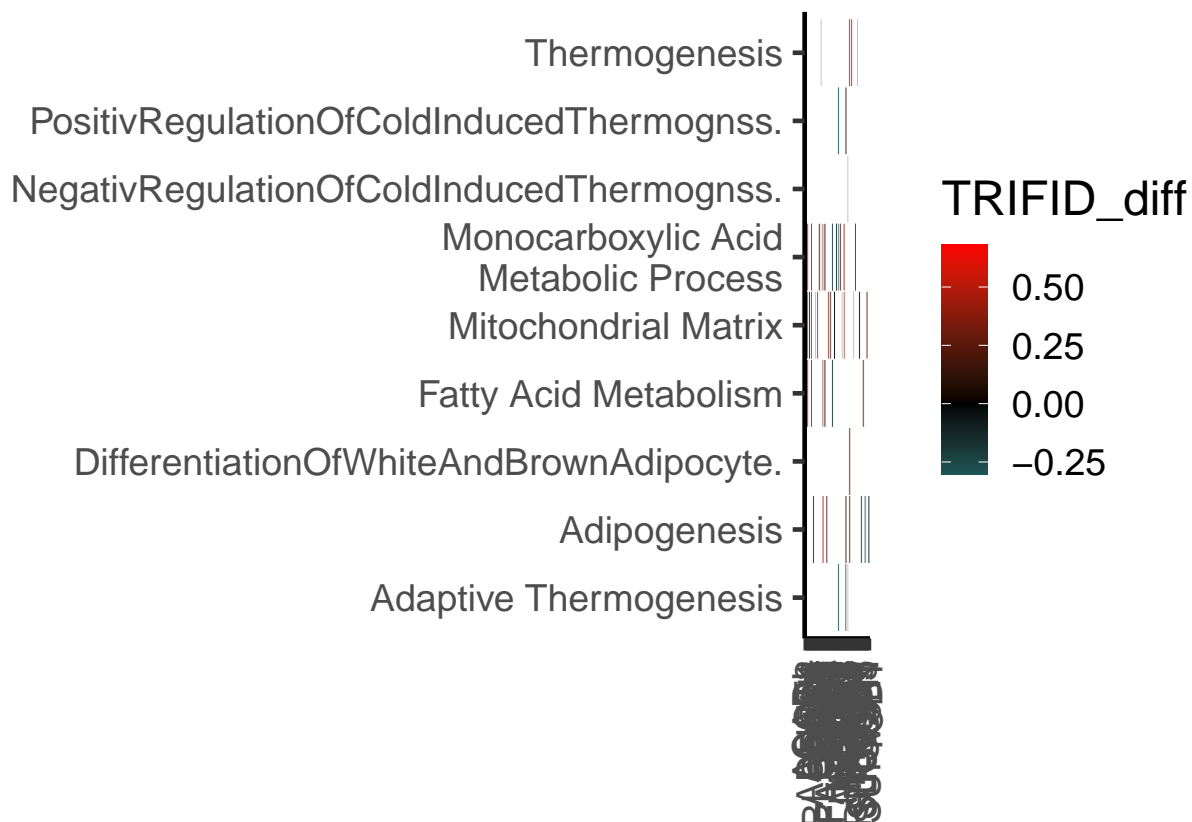
```
## Scale for size is already present.
## Adding another scale for size, which will replace the existing scale.
## Scale for colour is already present.
## Adding another scale for colour, which will replace the existing scale.
```



```
ggsave(file.path(figs, "thermogenesis_network_w_trifid.pdf"), width=9, height=7)
```

```
heatplot(specific, foldChange = diff) + theme_classic(base_size=18) +
  theme(axis.text.x = element_text(angle=90, vjust = 0.5, hjust=1)) +
  scale_fill_gradient2(high="red", low="cyan3", mid="black", na.value="grey", name="TRIFID_diff")
```

```
## Scale for fill is already present.
## Adding another scale for fill, which will replace the existing scale.
```



```
ggsave(file.path(figs, "thermogenesis_nheatplot_w_trifid.pdf"), width=15, height=4)
```

```
load(file.path(figs, "GO_object.RData"))
head(sig_ob)
```

```
##                                                    ID
## GOBP_ACTIN_FILAMENT_BASED_PROCESS GOBP_ACTIN_FILAMENT_BASED_PROCESS
## GOCC_CELL_LEADING_EDGE GOCC_CELL_LEADING_EDGE
## GOCC_GOLGI_CIS_CISTERNA GOCC_GOLGI_CIS_CISTERNA
## GOCC_CELL_CORTEX GOCC_CELL_CORTEX
## GOCC_CELL_PROJECTION_MEMBRANE GOCC_CELL_PROJECTION_MEMBRANE
## GOCC_RUFFLE GOCC_RUFFLE
## Description GeneRatio
## GOBP_ACTIN_FILAMENT_BASED_PROCESS GOBP_ACTIN_FILAMENT_BASED_PROCESS 38/344
## GOCC_CELL_LEADING_EDGE GOCC_CELL_LEADING_EDGE 24/292
## GOCC_GOLGI_CIS_CISTERNA GOCC_GOLGI_CIS_CISTERNA 5/292
## GOCC_CELL_CORTEX GOCC_CELL_CORTEX 17/292
## GOCC_CELL_PROJECTION_MEMBRANE GOCC_CELL_PROJECTION_MEMBRANE 16/292
## GOCC_RUFFLE GOCC_RUFFLE 13/292
## BgRatio pvalue p.adjust qvalue
## GOBP_ACTIN_FILAMENT_BASED_PROCESS 484/9243 9.306001e-06 9.306001e-06 0.03123878
## GOCC_CELL_LEADING_EDGE 286/7873 1.465656e-04 1.465656e-04 0.04969881
## GOCC_GOLGI_CIS_CISTERNA 17/7873 2.905469e-04 2.905469e-04 0.04969881
## GOCC_CELL_CORTEX 184/7873 4.595973e-04 4.595973e-04 0.05241022
## GOCC_CELL_PROJECTION_MEMBRANE 175/7873 7.541362e-04 7.541362e-04 0.05946280
```

```

## GOCC_RUFFLE                                128/7873 8.858972e-04 8.858972e-04 0.05946280
##
## GOBP_ACTIN_FILAMENT_BASED_PROCESS ADD3/KANK1/DIXDC1/SVIL/PRKG1/MY01C/INPP5K/MY06/DPYSL3/SH3KBP1/ARHG
## GOCC_CELL_LEADING_EDGE
## GOCC_GOLGI_CIS_CISTERNA
## GOCC_CELL_CORTEX
## GOCC_CELL_PROJECTION_MEMBRANE
## GOCC_RUFFLE
##
##                                     Count setSize
## GOBP_ACTIN_FILAMENT_BASED_PROCESS      38      344
## GOCC_CELL_LEADING_EDGE                  24      292
## GOCC_GOLGI_CIS_CISTERNA                  5      292
## GOCC_CELL_CORTEX                        17      292
## GOCC_CELL_PROJECTION_MEMBRANE           16      292
## GOCC_RUFFLE                             13      292

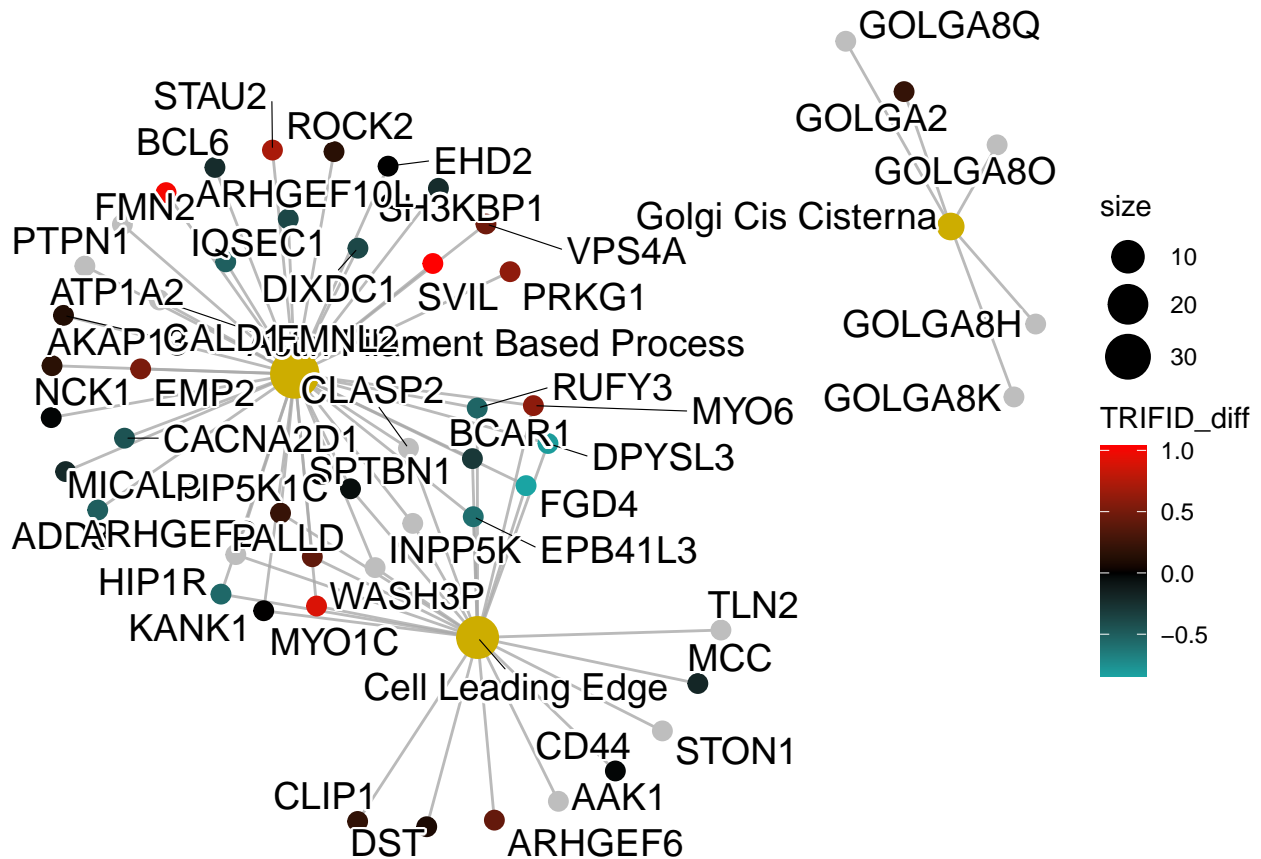
sig_ob = mutate(sig_ob, Description = shorten(ID) )

cnetplot(sig_ob, showCategory=3, categorySize="geneNum", color.params = list(foldChange = diff),
          color_category='gold3' ) + scale_colour_gradient2(high="red", low="cyan3", mid="black", na.val

## Warning in cnetplot.enrichResult(x, ...): Use 'color.params = list(category = your_value)' instead of
## The color_category parameter will be removed in the next version.

## Scale for size is already present.
## Adding another scale for size, which will replace the existing scale.
## Scale for colour is already present.
## Adding another scale for colour, which will replace the existing scale.

```



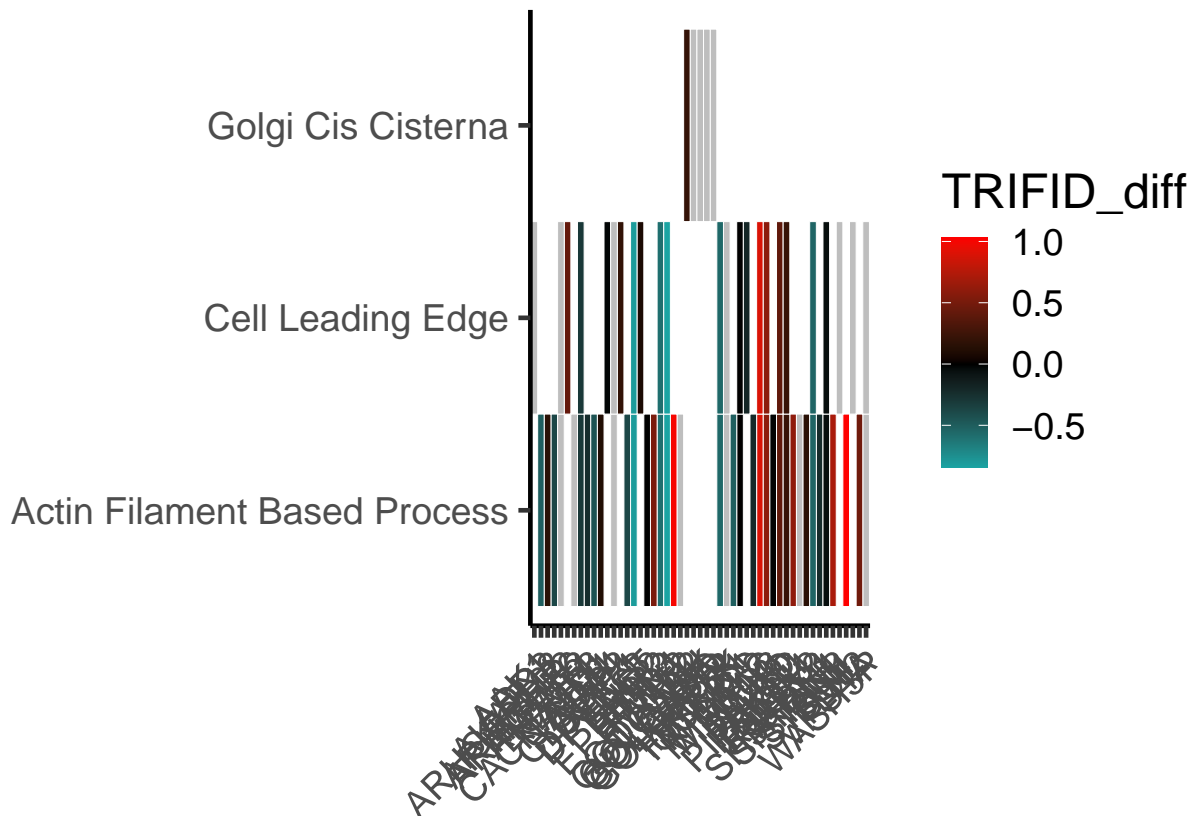
```

heatplot(sig_ob, foldChange = diff, showCategory = 3) + theme_classic(base_size=18) +
  theme(axis.text.x = element_text(angle=45, hjust=1)) + guides(fill = guide_colourbar(reverse=F)) +
  scale_fill_gradient2(high="red", low="cyan3", mid="black", na.value="grey", name="TRIFID_diff")

```

```
## Scale for fill is already present.
```

```
## Adding another scale for fill, which will replace the existing scale.
```



```
ggsave(file.path(figs, "go_heatplot_w_trifid.pdf"), width=15, height=3)
```

Average TRIFID scores per GO term

```
actin_genes = sig_ob@result$geneID[1]
actin_genes = stringr::str_split_1(actin_genes, "/")

mean = filter(trifid, gene_beige %in% actin_genes) %>% summarise(mean(trifid_diff))
head(mean)
```

```
##   mean(trifid_diff)
## 1      0.007550428
```

```
as.numeric(mean)
```

```
## [1] 0.007550428
```

```
average_trifid = function(geneID){
  score = c()
  for (i in 1:length(geneID)){
    genes = stringr::str_split_1(geneID[i], "/")
    trifid_score = filter(trifid, gene_beige %in% genes) %>% summarise(mean(trifid_diff))
```



```

    score = c(score, as.numeric(trifid_score))
  }
  return (score)
}

with_trifid = mutate(filter(sig_ob, qvalue < 0.05), trifid = average_trifid(geneID))
head(with_trifid)

```

```

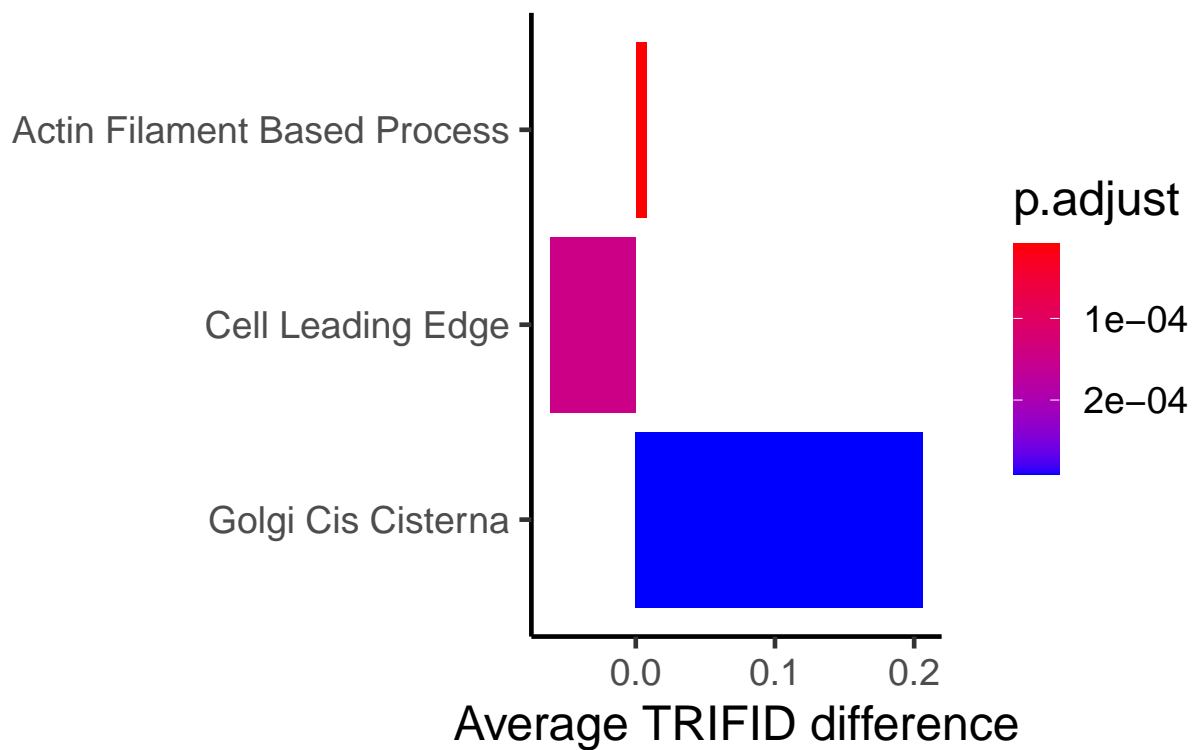
##
## GOBP_ACTIN_FILAMENT_BASED_PROCESS GOBP_ACTIN_FILAMENT_BASED_PROCESS
## GOCC_CELL_LEADING_EDGE GOCC_CELL_LEADING_EDGE
## GOCC_GOLGI_CIS_CISTERNA GOCC_GOLGI_CIS_CISTERNA
##
## Description GeneRatio
## GOBP_ACTIN_FILAMENT_BASED_PROCESS Actin Filament Based Process 38/344
## GOCC_CELL_LEADING_EDGE Cell Leading Edge 24/292
## GOCC_GOLGI_CIS_CISTERNA Golgi Cis Cisterna 5/292
##
## BgRatio pvalue p.adjust qvalue
## GOBP_ACTIN_FILAMENT_BASED_PROCESS 484/9243 9.306001e-06 9.306001e-06 0.03123878
## GOCC_CELL_LEADING_EDGE 286/7873 1.465656e-04 1.465656e-04 0.04969881
## GOCC_GOLGI_CIS_CISTERNA 17/7873 2.905469e-04 2.905469e-04 0.04969881
##
## GOBP_ACTIN_FILAMENT_BASED_PROCESS ADD3/KANK1/DIXDC1/SVIL/PRKG1/MYO1C/INPP5K/MYO6/DPYSL3/SH3KBP1/ARHG
## GOCC_CELL_LEADING_EDGE
## GOCC_GOLGI_CIS_CISTERNA
##
## Count setSize trifid
## GOBP_ACTIN_FILAMENT_BASED_PROCESS 38 344 0.007550428
## GOCC_CELL_LEADING_EDGE 24 292 -0.061614454
## GOCC_GOLGI_CIS_CISTERNA 5 292 0.206333333

```

```

barplot(with_trifid, x="trifid") + labs(x="Average TRIFID difference")+ theme_classic(base_size=18)

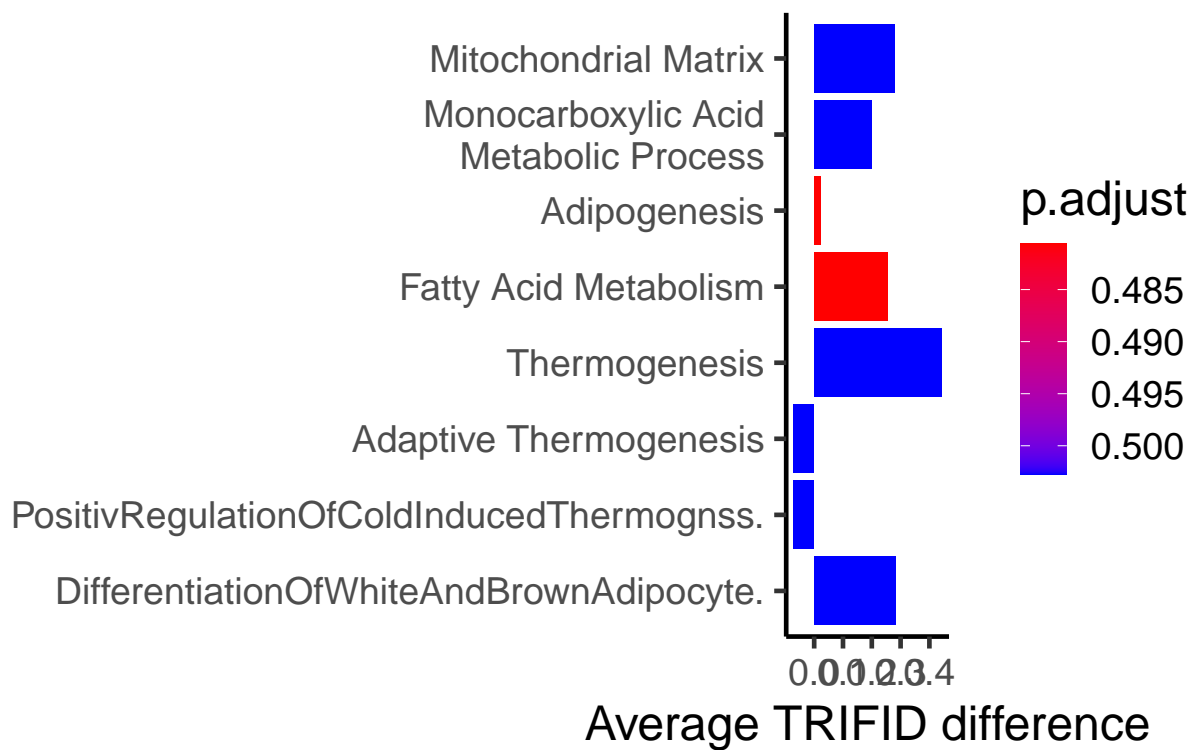
```



```
#dotplot(with_trifid, x="trifid") + labs(x="Average TRIFID difference")
ggsave(file.path(figs, "actin_trifid_average.pdf"))
```

```
## Saving 6.5 x 4.5 in image
```

```
thermo_trifid = mutate(specific, trifid = average_trifid(geneID))
barplot(thermo_trifid, x="trifid") + labs(x="Average TRIFID difference") + theme_classic(base_size=18)
```



```
ggsave(file.path(figs, "thermogenesis_trifid_average.pdf"), width=9)
```

```
## Saving 9 x 4.5 in image
```