User Manual for PARSED (PARticle SEgmentation Detector)

(Version 1.0)

1 Installation Requirements

1.1 Hardware

- A desktop computer or workstation with multi-core CPUs and Linux OS;
- An NVIDIA CUDA compatible GPU with *Kepler* architecture or newer (SM \geq 3.5);
- A fast storage disk with enough I/O speed. We recommend solid state drive (SSD) for high processing speed.

1.2 Software dependency

All required software and packages are listed in Table 1.

Table 1: Software and packages required for running PARSED

Software	Description	Version
python3	Python interpreter version 3	3.6
CUDA	NVIDIA CUDA acceleration library	8.0
docker	Container platform for software deployment	1.10
nvidia-docker	NVIDIA container runtime for docker	2.0
h5py	Python wrapper for HDF5 data format	2.7.1
Keras	Tensorflow API	2.0.8
mrcfile	Python wrapper for MRC file format	1.0.1
pandas	Data frame module	0.20.3
trackpy	A python program of Crocker-Grier algorithm	0.4.1
tensorflow-gpu	GPU accelerated TensorFlow	1.3.0
numba	Python JIT acceleration for trackpy	0.37.0
matplotlib	Python 2D plotting library	2.1.0
cv2	OpenCV library for morphological interfaces	3.3.1

Specific notes:

- python3 interpreter ≥ 3.6. For a system-wide build of PARSED, please refer documents from other Linux distributions (e.g., CentOS 7) to gain python3 support;
- Keras ≥ 2.0 ;
- NVIDIA CUDA library ≥ 8.0 ;
- docker platform ≥ 1.10 (containerized installation, see details in Subsec. 2.3), nvidia-docker (container runtime for NVIDIA CUDA capacity).

2 Software Deployments

We provide two deployment methods:

- System-wide deployment;
- Containerized deployment.

We recommend the second one to avoid dependency problems between different systems. To enable

CUDA functionality, both methods need following *basic system environment* with NVIDIA proprietary driver.

2.1 Basic system configuration

(1) Install NVIDIA proprietary driver

Follow NVIDIA official instructions at https://www.nvidia.com/Download/index.aspx?lang=en.

(2) Check NVIDIA driver

Run nvidia-smi in your shell:

```
$ nvidia-smi
```

2.2 System-wide deployment

(1) Install CUDA libraries

Follow NVIDIA official instructions at https://developer.nvidia.com/cuda-downloads.

(2) Install required packages/software

Use pip3 to fetch python packages and install pip3 with Linux package manager such as apt-get:

```
$ sudo apt-get install pip3
```

Then, install python packages:

```
$ sudo pip install h5py==2.7.1 \
Keras==2.0.8 \
mrcfile==1.0.1 \
numba==0.37.0 \
pandas==0.20.3 \
trackpy==0.4.1 \
matplotlib==2.1.0
```

For TensorFlow and OpenCV, please refer their official documents for complete installation.

- TensorFlow at https://www.tensorflow.org/.
- OpenCV at https://opencv.org/.
- (3) Download our compressed file parsed v1.tar.gz and extract related files, as listed Table 2:

Table 2: Files in parsed_v1.zip

	<u> </u>
Dockerfile	Instruction file for installing docker
<pre>parsed_main.py</pre>	PARSED main program
<pre>pre_train_model.h5</pre>	Pre-trained model of PARSED neural networks
<pre>mic_preprocess.py</pre>	Module for pre-processing raw micrographs
particle_mass.py	Program to generate particle mass distribution

2.3 Containerized deployment

(1) Install docker

Follow official documents and instructions at https://www.docker.com/get-started. Install docker with Linux package manager:

```
$ sudo apt-get install docker.io
```

- (2) Install NVIDIA containerized runtime nvidia-docker Follow official nvidia-docker wiki page at https://github.com/NVIDIA/nvidia-docker.
- (3) Build PARSED locally by downloading our compressed file parsed_v1.tar.gz, and extracting related files, as listed in Table 2. Make sure current user is in docker group and then build the PARSED docker image:

```
$ docker build -t parsed:local .
```

2.4 Installation check

To check system-wide deployment, run PARSED as:

```
$ python3 parsed_main.py -help
```

To check containerized deployment, please run:

```
$ docker run --runtime=nvidia -it --rm parsed:local python3
parsed_main.py --help
```

If everything works well, a built-in help page will print on screen, as shown in Figure 1.

```
imus: parsed $ python3 parsed_main.py --help
Usage: parsed_main.py <pick> [options]
    PARSED (PARticle SEgmentation Detector)
    PARSED is a deep-learning model that reads a list of MRC files of cryo-EM micrographs, and then automatically picks particles of biological macromolecules in these micrographs. The picked particles could be directly imported into 3D reconstruction programs such as
    CryoSPARC or RELION.
    Reference:
    R. Yao, J. Qian, Q. Huang. A universal deep-learning model for automated cryo-EM particle picking (To be published).
Options:
                              show program's version number and exit show this help message and exit
    -version
  -h, --help
  --model=./pre_train_model.h5
                               file name of used model (default pre_train_model.h5)
  --data_path=PATH_OF_MICROGRAPHS
                              path for target micrograph sets
  --output path=PATH OUTPUT
 path for output location
```

Figure 1: Help page for PARSED

3 Picking Particles with PARSED

In this section, we show how to use PARSED to pick particles from cryo-EM micrographs. Parameters for running PARSED are listed in Table 3.

Table 3: Parameters for running PARSED

Parameters	Description	
model	pre-trained segmentation model	
data_path	directory path of input data	
output_path	storage path of output data	
file_pattern	filename of micrographs, also accept Unix bash-like regular expression	
	(RegEx)	
job_suffix	suffix for output filename	
angpixel	sample rate for specified micrographs (in Å/pixel)	
img_size	size of micrographs (the largest one of Width and Height, in pixel)	
edge_cut	cropped edge size (in pixel)	
core_num	parallel running process number	
aperture	diameter of particle-picking aperture (in Å)	
mass_min	minimal picking mass of detected blobs (particle candidates)	
gpu_id	IDs of used GPUs (start from 0)	

3.1 Running PARSED

With micrographs in subdirectory demo, run PARSED to detect particles in these micrographs:

```
$ python3 -W ignore parsed_main.py \
  --model=./pre_train_model.h5 --data_path=./demo \
  --output_path=./output --file_pattern=*.mrc \
  --angpixel=1.34 --img_size=4096 --edge_cut=0 \
  --job_suffix=autopick --core_num=4 -- aperture=160 \
  --mass_min=4 --gpu_id=1
```

Example of program outputs on screen is shown in Figure 2.

```
Choosing mode: pick
Initializing model framework
Using TensorFlow backend.
Coordinates extraction on pl 025 finished 293 found Timespan 11.649s [ 0 / 6 ]
Coordinates extraction on pl 026 finished 301 found Timespan 11.755s [ 1 / 6 ]
Coordinates extraction on pl 027 finished 301 found Timespan 11.755s [ 1 / 6 ]
Coordinates extraction on pl 027 finished 291 found Timespan 11.761s [ 2 / 6 ]
Coordinates extraction on pl 027 finished 291 found Timespan 11.761s [ 2 / 6 ]
Coordinates extraction on pl 027 finished 299 found Timespan 6.194s [ 5 / 6 ]
Coordinates extraction on pl 029 finished 294 found Timespan 6.217s [ 4 / 6 ]
Coordinates extraction finished 1790 TotalPickNum found
```

Figure 2: PARSED running screenshot

The coordinates of the detected particles will be stored in STAR format. Suffix for filenames is given by --job_suffix, and location is given by --output_path.

3.2 Selecting detected particles

PARSED records the properties of detected blobs (particle candidates) in separate files with suffix _param, e.g., their mass (i.e., blob brightness). To reduce falsely positive picking, one could filter the detected particles by using *grep*, *sed*, and *awk*, etc. Also, we offer a python script particle_mass.py to generate blob mass distribution for all the detected particles, and then select some of them for further single-particle analysis (SPA) with a mass selection threshold (see also Methods).

Create the mass distribution of detected particles to tmp hist.jpg:

```
$ python3 particle_mass.py drawmass --pick_output=./output \
    --job_suffix=autopick --tmp_hist=tmp_hist.jpg
```

Two typical mass distributions are illustrated in Figure 3.

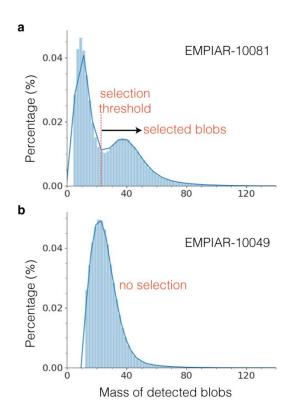


Figure 3: Mass distributions of detected blobs (particle candidates). **a**, A two-peak distribution with a selection threshold at mass of ~21 (from the dataset EMPIAR-10081). In this case, only those detected blobs with mass values greater than the selection threshold are selected as the final picked particles for further investigations. **b**, A one-peak distribution without selection (from the dataset EMPIAR-10049). In this case, all detected blobs are selected as the final picked particles.

Then, determine whether or not a selection is needed:

```
$ python3 particle_mass.py cutoff --pick_output=./output \
   --job_suffix=autopick --output_suffix=checked --thres=28
```

Output results will be generated in the same directory.

4 Importing Picked Particles into 3D Reconstruction Program

In this section, we show how to import the final picked particles into *ab initio* 3D reconstruction program RELION to build 3D density maps with standard single-particle analysis (SPA) procedures. Please see more details about RELION at: http://www2.mrc-lmb.cam.ac.uk/relion/index.php?title=Main_Page.

4.1 Copying particles to RELION work directory

To import the picked particles into RELION, we copy the output results back to RELION work directory. Suppose RELION Import, and CTF estimation for the micrographs have been completed, and work directory is: ~/Relion2Test/demodir, directory for raw micrographs is: ~/Relion2Test/demodir/Micrographs; then, copy the results to the micrograph folder for RELION:

demodir \$ cp ~/parsed/output/*_checked.star Micrographs/

4.2 Importing particles into RELION

Start RELION and select Import option in left, as shown in Figure 4. Here, we designate Import job as parsedDemo, and suffix as checked.



Figure 4: RELION panel for Import job

4.3 Checking imported particles with RELION

Before displaying picked particles, we create a Manual picking job in RELION. Next, as illustrated in Figure 5, we select our Import job, click Display green drop-down menu and then item out:coords_suffix_checked.star, the picked particles will be displayed, as in Figure 6.

After importing the particles, further steps in the SPA workflow, such as particle extraction, 2D classification and 3D reconstruction etc., could be carried out with standard procedures in RELION.

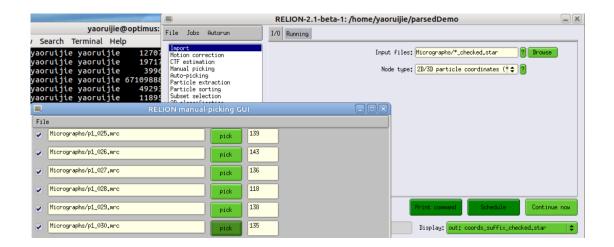


Figure 5: RELION panels for checking imported particles

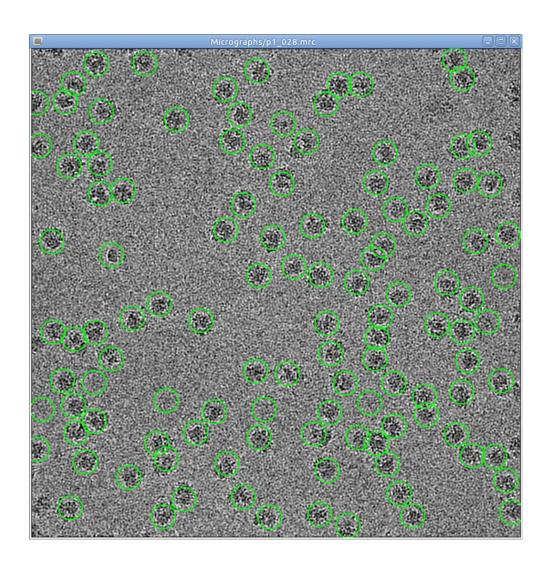


Figure 6: Display of the picked particles imported into RELION