

Package ‘screenr’

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Type Package

Title Construction of Binary Test-Screening Rules

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Imports glmpath, pROC, dplyr, scales, stringr, epiR

Description Package screenr enables easy development and validation of diagnostic test screening tools. It is designed to enable those with only a basic familiarity with R to develop, validate and implement screening tools for diagnostic tests. Consider the situation where a definitive test for some condition is relatively expensive, and the condition is rare. In that case, universal testing would not be efficient in terms of the yield of positive results per test performed. Now suppose that responses to a set of simple diagnostic questions or observations may be predictive of the definitive test result. Package screenr enables estimation of thresholds for making decisions about when to perform the definitive test on newly observed subjects based on Receiver Operating Characteristics (ROC) estimated from an initial sample. The choice of a particular screening threshold is left to the user, and should be based on careful consideration of application-specific tradeoffs between sensitivity (true positive fraction) and specificity (true negative fraction).

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URL <https://github.com/sgutreuter/screenr/>

Encoding UTF-8

RoxygenNote 7.1.2

BugReports <https://github.com/sgutreuter/screenr/issues>

LazyData true

Suggests rmarkdown,
knitr

VignetteBuilder knitr

R topics documented:

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coef.lasso_screenr	<i>An S3 Method to Extract Coefficients from lasso_screenr Objects</i>
--------------------	------------------------------------------------------------------------

Description

coef.lasso_screenr extracts the logistic model parameter estimates from lasso_screenr-class objects.

Usage

```
## S3 method for class 'lasso_screenr'
coef(object, ..., intercept = TRUE, or = FALSE)
```

Arguments

object	an object of class lasso_screenr.
...	optional arguments passed to predict methods.
intercept	(logical) retain (TRUE, default) or drop (FALSE) the intercept coefficients.
or	return odds ratios if TRUE; logit-scale coefficients are the default.

Details

coef.lasso_screenr extracts the estimated coefficients from lasso_screenr objects.

Value

a $p \times 2$ matrix containing the estimated coefficients from the AIC- and BIC-best logistic regression models, where p is the number of coefficients.

Examples

```
attach(uniobj1)
coef(uniobj1)
```

coef.logreg_screenr	<i>An S3 Method to Extract Coefficients from logreg_screenr Objects</i>
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Description

coef.logreg_screenr extracts the logistic model parameter estimates from logreg_screenr-class objects.

Usage

```
## S3 method for class 'logreg_screenr'
coef(object, ..., intercept = TRUE, or = FALSE)
```

Arguments

<code>object</code>	an object of class <code>logreg_screenr</code> .
<code>...</code>	optional arguments passed to <code>predict</code> methods.
<code>intercept</code>	(logical) retain (TRUE, default) or drop (FALSE) the intercept coefficients.
<code>or</code>	return odds ratios if TRUE; logit-scale coefficients are the default.

Details

`coef.logreg_screenr` extracts the estimated coefficients from `logreg_screenr` objects.

Value

A numeric vector containing the estimated coefficients on the logit scale.

Examples

```
attach(uniobj2)
class(uniobj2)
coef(uniobj2)
```

easy_tool	<i>Simplifying Screening from lasso_screenr or logreg_screenr Objects</i>
-----------	---------------------------------------------------------------------------

Description

`easy_tool` rescales model coefficients to whole numbers ranging from 1 to `max(QuestionWeights)`. Those rescaled and rounded coefficients can be used as weights for each screening question in a simplified model-based screening tool. The test screening score is the sum of the weights for each subject.

Usage

```
easy_tool(object, max = 3, model = c("minAIC", "minBIC"), crossval = TRUE, ...)
```

Arguments

<code>object</code>	an object of class <code>lasso_screenr</code> or <code>logreg_screenr</code> .
<code>max</code>	(numeric) the desired maximum value for the response weights (default is 3).
<code>model</code>	(for <code>lasso_screenr</code> objects only) the desired basis model. Valid options are "minAIC" (the default) and "minBIC".
<code>crossval</code>	a (logical) indicator for cross-validated (TRUE) or in-sample (FALSE) performance evaluation.
<code>...</code>	additional arguments passed to <code>coef.lasso_screenr</code> or <code>coef.logreg_screenr</code>

Details

The `QuestionWeights` (see `Value`, below) are the foundation for easy screening. For example, the screening tool could consist of a simple questionnaire followed by the weight for each question, expressed as a small whole number (1, ..., max) and/or an equal number of open circles. The person doing the screening need only circle the numerical weight and/or fill in the circles if and only if the subject provides a "yes" response to a particular question. The person doing the screening then obtains the final score for that subject by adding up the circled numbers or counting the total number of filled-in circles. Testing is mandatory for consenting subjects for whom that final score equals or exceeds the chosen threshold based on the receiver-operating characteristics of `CVresults`.

The value chosen for max involves a trade-off between the ease of manual scoring and the degree to which the ROC from the re-scaling matches the ROC from the model. Small values of max make manual scoring easy, and sufficiently large values will match the screening performance of the model fit. A value of 3 may be a reasonable compromise. It is prudent to compare the ROCs from a few values of max with the ROC from the model and base the final choice on the trade-off between ease of manual scoring and the desired combination of sensitivity and specificity.

Value

`easy_tool` returns (invisibly) an object of class `easy_tool` containing:

`QuestionWeights` Weights for the screening questions obtained by rescaling the non-zero-valued logistic regression coefficients to whole numbers ranging from 1 to max.

`Type` The type of test performance evaluation ("cross-validated" or "in-sample").

`Scores` A data frame containing the testing outcomes (response) and cross-validated scores obtained as the sums of the weighted responses to the set of screening questions (score).

`ROC` An object of class `roc` containing the receiver-operating characteristic produced by ``pROC::roc``.

Note

Execute `methods(class = "easy_tool")` to see available methods.

See Also

[rescale_to_int](#), [ntpp.easy_tool](#), [plot.easy_tool](#), [print.easy_tool](#) and [summary.easy_tool](#)

Examples

```
attach(uniobj1)
tool <- easy_tool(uniobj1, max = 3)
class(tool)
```

get_what

S3 Methods for Extraction of Object Components

Description

get_what extracts components from objects.

Usage

```
get_what(from, what, ...)
```

Arguments

from	an object from which to extract what.
what	the element to extract from from.
...	additional arguments.

See Also

[get_what.easy_tool](#), [get_what.lasso_screenr](#), [get_what.logreg_screenr](#) and [get_what.simple_screenr](#).

get_what.easy_tool

An S3 Method for Extraction of Components from easy_tool Objects

Description

get_what.easy_tool extracts components from easy_tool-class objects.

Usage

```
## S3 method for class 'easy_tool'
get_what(
  from = NULL,
  what = NULL,
  ...,
  bootreps = 2000,
  conf.level = 0.95,
  se.min = 0.7
)
```

Arguments

from	the easy_tool-class object from which to extract the component.
what	the (character) name of the component to extract. Valid values are "Call", "QuestionWeights", "ROCci", "ROC" and "Scores".
...	optional arguments to get_what methods.
bootreps	the number of bootstrap replications for estimation of confidence intervals for what = "ROCci".
conf.level	(optional) confidence level for what = ROCci
se.min	minimum value of sensitivity printed for what = ROCci (default = 0.7).

Details

get_what is provided to enable easy extraction of components that are not provided by the plot, predict, print or summary methods.

Valid values of what are:

"Call" returns the function call that created from.

"QuestionWeights" returns the screening question weights, which are the re-scaled logistic-regression coefficients.

ROCci returns a data frame containing sensitivities, specificities and their confidence limits, and thresholds

"Scores" returns the screening scores for each subject, which are the sums of the products of the binary question responses and their QuestionWeights

"ROC" returns the receiver-operating characteristic for the Scores

Value

The selected component is returned invisibly.

Examples

```
## Not run:
attach(uniobj1)
tool <- easy_tool(uniobj1, max = 3, crossval = TRUE)
## Get and print sensitivities and specificities at thresholds for the
## local maxima of the ROC curve
ROCci <- get_what(from = tool, what = "ROCci")
print(ROCci)

## End(Not run)
```

get_what.lasso_screenr

An S3 Method for Extraction of Components from lasso_screenr Objects

Description

get_what.lasso_screenr extracts components from lasso_screenr-class objects.

Usage

```
## S3 method for class 'lasso_screenr'
get_what(
  from = NULL,
  what = c("glmpathObj", "ROCci", "cvROC", "isROC"),
  ...,
  model = c("minAIC", "minBIC"),
  conf.level = 0.95,
  bootreps = 2000,
  se.min = 0.7
)
```

Arguments

from	the lasso_screenr-class object from which to extract the component.
what	the (character) name of the component to extract. Valid values are "glmpathObj", "ROCci", "cvROC" and "isROC".
...	optional arguments to get_what methods.
model	the (character) name of the model for which the component is desired. Valid values are "minAIC" and "minBIC".
conf.level	(optional) confidence level for what = "ROCci"
bootreps	the number of bootstrap replications for estimation of confidence intervals for what = "ROCci".
se.min	minimum value of sensitivity printed for what = ROCci (default = 0.7).

Details

get_what is provided to enable easy extraction of components that are not provided by the coef, plot, predict, print or summary methods.

The following values of what return:

"glmpathObj" the entire glmpath-class object produced by [glmpath](#).

ROCci a data frame containing cross-validated sensitivities, specificities and their confidence limits, and thresholds.

"cvROC" the roc-class object produced by `roc` containing the k -fold cross-validated receiver-operating characteristic.

"isROC" the roc-class object produced by `roc` containing the in-sample (overly optimistic) receiver-operating characteristic.

Value

The selected component is returned invisibly.

Examples

```
## Not run:
attach(uniobj1)
## Plot the coefficient paths
pathobj <- get_what(from = uniobj1, what = "glmpathObj", model = "minAIC")
plot(pathobj)
## Get and print cross-validated sensitivities and specificities at
## thresholds for the local maxima of the ROC curve
cvROCCi <- get_what(from = uniobj1, what = "ROCCi", model = "minBIC")
print(cvROCCi)

## End(Not run)
```

get_what.logreg_screenr

An S3 Method for Extraction of Components from logreg_screenr Objects

Description

`get_what.logreg_screenr` extracts components from `logreg_screenr`-class objects.

Usage

```
## S3 method for class 'logreg_screenr'
get_what(
  from = NULL,
  what = c("ModelFit", "ROCCi", "cvROC", "isROC"),
  ...,
  conf.level = 0.95,
  bootreps = 2000,
  se.min = 0.7
)
```

Arguments

from	the logreg_screenr-class object from which to extract the component.
what	the (character) name of the component to extract. Valid values are "ModelFit", "ROCci", "cvROC" and "isROC".
...	optional arguments to get_what methods.
conf.level	(optional) confidence level for what = "ROCci"
bootreps	the number of bootstrap replications for estimation of confidence intervals for what = "ROCci".
se.min	minimum value of sensitivity printed for what = ROCci (default = 0.7).

Details

get_what is provided to enable easy extraction of components for those who wish to perform computations that are not provided by the coef, plot, predict, print or summary methods.

The following values of what return:

"ModelFit" the entire glm-class object produced by `glm`.

ROCci a data frame containing cross-validated sensitivities, specificities and their confidence limits, and thresholds.

"cvROC" the roc-class object produced by `roc` containing the k -fold cross-validated receiver-operating characteristic.

"isROC" the roc-class object produced by `roc` containing the in-sample (overly optimistic) receiver-operating characteristic.

Value

The selected component is returned invisibly.

Examples

```
## Not run:
attach(uniobj2)
## Get and print cross-validated sensitivities and specificities at
## thresholds for the local maxima of the ROC curve
myROCci <- get_what(from = uniobj2, what = "ROCci")
print(myROCci)

## End(Not run)
```

get_what.simple_screenr

An S3 Method for Extraction of Components from simple_screenr Objects

Description

get_what.simple_screenr extracts components from simple_screenr-class objects.

Usage

```
## S3 method for class 'simple_screenr'
get_what(
  from = NULL,
  what = c("ROCci", "isROC"),
  ...,
  conf.level = 0.95,
  bootreps = 2000,
  se.min = 0.6
)
```

Arguments

from	the simple_screenr-class object from which to extract the component.
what	the (character) name of the component to extract. Valid values are "ROCci" and "isROC".
...	optional arguments to get_what methods.
conf.level	(optional) confidence level for what = "ROCci"
bootreps	the number of bootstrap replications for estimation of confidence intervals for what = "ROCci".
se.min	minimum value of sensitivity printed for what = ROCci (default = 0.6).

Details

get_what is provided to enable easy extraction of components for those who wish to perform computations that are not provided by the plot, predict, print or summary methods.

The following values of what return:

"isROC" the roc-class object produced by [roc](#) containing the in-sample (overly optimistic) receiver-operating characteristic.

"ROCci" a data frame containing cross-validated sensitivities, specificities and their confidence limits, and thresholds.

Value

The selected component is returned invisibly.

Examples

```
## Not run:
data(unicorns)
too_simple <- simple_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7,
                             data = unicorns)
too_simple_roc <- get_what(from = too_simple, what = "isROC" )
plot(too_simple_roc)

## End(Not run)
```

inverse_link

Compute the Inverses of Binomial Link Functions

Description

inverse_link returns the inverse of logit, cloglog and probit link functions for a linear predictor

Usage

```
inverse_link(lp = NULL, link = c("logit", "cloglog", "probit"))
```

Arguments

lp	numeric vector containing the estimated link.
link	(character) name of the link function (one of "logit", "cloglog" or "probit").

Details

inverse_link returns the inverses of logit, cloglog and probit link functions, and is provided as a (laborious) way to compute predicted values from the ModelFit component of logreg_screenr-class objects. The predict methods are a better way to obtain predicted values.

Value

A numeric vector containing the inverse of the link function for the linear predictor.

Note

inverse_link may not be included in future versions of the screenr package.

See Also

[predict.logreg_screenr](#)

Examples

```
## Make predictions of probability of infection from new observations
attach(uniobj2)
new_corns <- data.frame(ID = c("Alice D.", "Bernie P."),
                        testresult = c(NA, NA), Q1 = c(0, 0), Q2 = c(0, 0),
                        Q3 = c(0, 0), Q4 = c(0, 0), Q5 = c(0, 1), Q6 = c(0, 1 ),
                        Q7 = c(0, 1))
mfit <- get_what(from = uniobj2 , what = "ModelFit")
coefs <- mfit$coefficients
lp <- as.matrix(cbind(rep(1, nrow(new_corns)), new_corns[, 3:9])) %*%
      as.matrix(coefs, ncol = 1)
(preds <- inverse_link(lp, link = "logit"))
## Note that only the predicted values are returned.
```

keepfirst

Return Data Frame Rows Having Unique Values in Selected Columns

Description

keepfirst extracts those rows of a data frame which have unique values in selected columns.

Usage

```
keepfirst(x, colnames, data = NULL)
```

Arguments

x	character-valued column name along which the dataframe is sorted.
colnames	a character vector of column names to identify uniqueness.
data	a data frame.

Details

The dataframe data is sorted, and then only those rows which are unique with respect to the values of selected columns.

Value

A data frame consisting of the rows of data which are unique with respect to colnames

lasso_screenr	<i>Fitting Screening Tools Using Lasso-Like Regularization of Logistic Regression</i>
---------------	---------------------------------------------------------------------------------------

Description

lasso_screenr is a convenience function which integrates logistic regression using $L1$ regularization, k -fold cross-validation and estimation of the receiver-operating characteristic. The in-sample and out-of-sample performance is estimated from the models which produced the minimum AIC and minimum BIC. Execute `methods(class = "lasso_screenr")` to identify available methods.

Usage

```
lasso_screenr(
  formula,
  data = NULL,
  Nfolds = 10,
  L2 = TRUE,
  seed = Sys.time(),
  ...
)
```

Arguments

formula	an object of class <code>stats::formula</code> defining the testing outcome and predictor covariates.
data	a dataframe containing the variables defined in formula.
Nfolds	the number of folds used for k -fold cross validation (default = 10, minimum = 2, maximum = 100).
L2	(logical) switch controlling penalization using the $L2$ norm of the parameters (default = TRUE).
seed	random number generator seed for cross-validation data splitting.
...	additional arguments passed to glmpath or roc .

Details

lasso_screenr uses the $L1$ path regularizer of Park and Hastie (2007), as implemented in the `glmpath` package. Park-Hastie regularization is similar to the conventional lasso and the elastic net. It differs from the lasso with the inclusion of a very small, fixed ($1e-5$) penalty on the $L2$ norm of the parameters, and differs from the elastic net in that the $L2$ penalty is fixed. Like the elastic net, the Park-Hastie regularization is robust to highly correlated predictors. The $L2$ penalization can be turned off (`L2 = FALSE`), in which case the regularization is similar to the conventional lasso. Like all $L1$ regularizers, the Park-Hastie algorithm automatically selects covariates.

The receiver-operating characteristics are computed using the `pROC` package.

Out-of-sample performance is estimated using k -fold cross-validation. For a gentle but Python-centric introduction to k -fold cross-validation, see <https://machinelearningmastery.com/k-fold-cross-validation/>

Value

Return (invisibly) an object of class `lasso_screenr` containing the elements:

`Call` The function call.

`Prevalence` Prevalence of the binary response variable.

`glmPathObj` An object of class `glmPath` returned by `glmPath::glmPath`. See `help(glmPath)` and `methods(class = "glmPath")`.

`Xmat` The matrix of predictors.

`isResults` A list structure containing the results from the two model fits which produced the minimum AIC and BIC values, respectively. The results consist of `Coefficients` (the logit-scale parameter estimates, including the intercept), `isPreds` (the in-sample predicted probabilities) and `isROC` (the in-sample receiver-operating characteristic (ROC) of class `roc`).

`RNG` Specification of the random-number generator used for k-fold data splitting.

`RNGseed` RNG seed.

`cvResults` A list structure containing the results of *k*- fold cross-validation estimation of out-of-sample performance.

The list elements of `cvResults` are:

`Nfolds` the number folds *k*

`X_ho` the matrix of held-out predictors for each cross-validation fold

`minAICcvPreds` the held-out responses and out-of-sample predicted probabilities from AIC-best model selection

`minAICcvROC` the out-of-sample ROC object of class `roc` from AIC-best model selection

`minBICcvPreds` the held-out responses and out-of-sample predicted probabilities from BIC-best model selection

`minBICcvROC` the corresponding out-of-sample predicted probabilities and ROC object from BIC-best model selection

References

Park MY, Hastie T. L1-regularization path algorithm for generalized linear models. *Journal of the Royal Statistical Society Series B*. 2007;69(4):659-677. <https://doi.org/10.1111/j.1467-9868.2007.00607.x>

Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Müller M. pROC: An open-source package for R and S+ to analyze and compare ROC curves. *BMC Bioinformatics*. 2011;12(77):1-8. <http://doi.org/10.1186/1471-2105-12-77>

See Also

[glmPath](#), [roc](#)

Examples

```
data(unicorns)
help(unicorns)
uniobj1 <- lasso_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7,
                        data = unicorns, Nfolds = 10)
summary(uniobj1)
```

logreg_screenr

Fitting Screening Tools Using Ordinary Logistic Regression

Description

logreg_screenr is a convenience function which integrates ordinary logistic regression k -fold cross-validation and estimation of the receiver-operating characteristic.

Usage

```
logreg_screenr(
  formula,
  data = NULL,
  link = c("logit", "cloglog", "probit"),
  Nfolds = 10,
  seed = Sys.time(),
  ...
)
```

Arguments

formula	an object of class <code>stats::formula</code> defining the testing outcome and predictor covariates, which is passed to <code>stats::glm()</code> .
data	the "training" sample; a data frame containing the testing outcome and predictive covariates to be used for testing screening. The testing outcome must be binary (0,1) indicating negative and positive test results, respectively, or logical (TRUE/FALSE). The covariates are typically binary (0 = no, 1 = yes) responses to questions which may be predictive of the test result, but any numeric or factor covariates can be used.
link	the character-valued name of the link function for logistic regression. Choices are "logit" (default), "cloglog" or "probit".
Nfolds	number of folds used for k -fold cross validation (default = 10, minimum = 2, maximum = 100).
seed	random-number generator seed for cross-validation data splitting.
...	additional arguments passed to or from other <code>stats::glm</code> or <code>pROC::roc</code> .

Details

The results provide information from which to choose a probability threshold above which individual out-of-sample probabilities indicate the need to perform a diagnostic test. Out-of-sample performance is estimated using k -fold cross validation.

The receiver operating characteristics are computed using the pROC package. See References and package documentation for additional details.

For a gentle but python-centric introduction to k -fold cross-validation, see <https://machinelearningmastery.com/k-fold-cross-validation/>.

Value

An object of class logreg_screenr containing the elements:

Call The function call.

formula The formula object.

Prevalence Prevalence (proportion) of the test condition in the training sample.

ModelFit An object of class glm (See [glm](#)) containing the results of the model fit.

ISroc An object of class [roc](#) containing the "in-sample" (overly-optimistic) receiver operating characteristics, and additional functions for use with this object are available in the pROC package.

CVpreds An object of class cv.predictions containing the data and cross-validated predicted condition y .

CVroc An object of class [roc](#) containing the k -fold cross-validated "out-of-sample" receiver operating characteristics, and additional functions for use with this object are available in the pROC package.

CVcoef the estimated coefficients from cross-validation

X_ho the matrix of held-out predictors for each cross-validation fold

Note

logreg_screenr is intended mainly for comparison with lasso_screenr. Careful manual model selection is required with logreg_screenr. lasso_screenr is easier and should generally produce better results.

References

Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Müller M. pROC: An open-source package for R and S+ to analyze and compare ROC curves. BMC Bioinformatics. 2011;12(77):1-8. <http://doi.org/10.1186/1471-2105-12-77>

See Also

[glm](#)

Examples

```
data(unicorns)
help(unicorns)
uniobj2 <- logreg_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7,
                          data = unicorns, link = "logit", Nfolds = 10)
summary(uniobj2)
```

nnt_	<i>Compute the Ratio of Total Tests Performed Per Postive Result</i>
------	----------------------------------------------------------------------

Description

nnt_ computes the anticipated average number of tests performed in order to observe a positive test result.

Usage

```
nnt_(dframe)
```

Arguments

dframe a data frame containing columns sensitivity, specificity and prev.

ntpp	<i>An S3 Method to Compute the Ratio of Total Tests to Positive Results</i>
------	-----------------------------------------------------------------------------

Description

ntpp computes the ratio of the total number of tests performed per positive test result.

Usage

```
ntpp(object, ...)
```

Arguments

object an object from which to compute the number of tests per test positive test results.
 ... additional arguments.

Details

The anticipated number of tests required to detect a single positive *nttp* is given by

$$nntp = (SeP + (1 - Sp)(1 - P))/SeP$$

where *Se* is sensitivity, *P* is prevalence and *Sp* is specificity. The anticipated prevalence among those screened out is given by

$$P_{untested} = ((1 - Se)P)/((1 - Se)P + Sp(1 - P))$$

See Also

[ntpp.lasso_screenr](#) [ntpp.logreg_screenr](#) [ntpp.data.frame](#) [ntpp.simple_screenr](#)

ntpp.data.frame

Compute the Ratio of Total Tests to Positive Results from a Data Frame

Description

ntpp.data.frame computes the ratio of the total number of tests performed per positive test result from data frames.

Usage

```
## S3 method for class 'data.frame'
ntpp(object, ...)
```

Arguments

object a dataframe containing columns sensitivity, specificity and prev.
... optional arguments to ntp methods.

Details

The anticipated number of tests required to detect a single positive *ntpp* is given by

$$ntpp = (SeP + (1 - Sp)(1 - P))/SeP$$

where *Se* is sensitivity, *P* is prevalence and *Sp* is specificity. The anticipated prevalence among those screened out is given by

$$P_{untested} = ((1 - Se)P)/((1 - Se)P + Sp(1 - P))$$

Value

a data frame containing the following columns:

sensitivity the sensitivity (proportion)

specificity the specificity (proportion)

prev prevalence proportion of the test condition

ntpp anticipated total tests required per positive result

prev_untested anticipated prevalence proportion among the untested

ntpp.easy_tool	<i>Compute the Ratio of Total Tests to Positive Results from easy_tool Objects</i>
----------------	------------------------------------------------------------------------------------

Description

ntpp.easy_tool computes the ratio of the total number of tests performed per positive test result from easy_tool-class objects.

Usage

```
## S3 method for class 'easy_tool'
ntpp(object, ..., prev = NULL)
```

Arguments

object	an easy_tool-class object produced by easy_tool.
...	optional arguments to ntp methods.
prev	an optional prevalence proportion for the test outcome; if missing the prevalence is obtained from object.

Details

The anticipated number of tests required to detect a single positive *nttp* is given by

$$nntp = (SeP + (1 - Sp)(1 - P))/SeP$$

where *Se* is sensitivity, *P* is prevalence and *Sp* is specificity. The anticipated prevalence among those screened out is given by

$$P_{untested} = ((1 - Se)P)/((1 - Se)P + Sp(1 - P))$$

Value

A data frame containing the following columns:

sensitivity	The sensitivity (proportion) of the screener.
specificity	The specificity (proportion) of the screener.
ntpp	the number of tests required to discover a single positive test result.
prev_untested	The prevalence proportion of the test condition among those who are screened out of testing.

See Also

[ntpp.lasso_screenr](#) [ntpp.logreg_screenr](#) [ntpp.data.frame](#) [ntpp.simple_screenr](#)

Examples

```
attach(uniobj1)
tool <- easy_tool(uniobj1, max = 3, crossval = TRUE)
ntpp(tool)
```

ntpp.lasso_screenr	<i>Compute the Ratio of Total Tests to Positive Results from lasso_screenr Objects</i>
--------------------	----------------------------------------------------------------------------------------

Description

ntpp.lasso_screenr computes the ratio of the total number of tests performed per positive test result from lasso_screenr-class objects.

Usage

```
## S3 method for class 'lasso_screenr'
ntpp(
  object,
  ...,
  model = c("minAIC", "minBIC"),
  type = c("cvResults", "isResults"),
  prev = NULL
)
```

Arguments

object	a lasso_screenr-class object produced by lasso_screenr.
...	optional arguments to ntp methods.
model	(character) select the model which produced the minimum AIC ("minAIC", the default) or minimum BIC ("minBIC").
type	(character) one of "cvResults" (the default) or "isResults" to specify k -fold cross-validated or in-sample receiver-operating characteristics, respectively.
prev	an optional prevalence proportion for the test outcome; if missing the prevalence is obtained from object.

Details

The anticipated number of tests required to detect a single positive ntp is given by

$$nntp = (SeP + (1 - Sp)(1 - P))/SeP$$

where Se is sensitivity, P is prevalence and Sp is specificity. The anticipated prevalence among those screened out is given by

$$P_{untested} = ((1 - Se)P)/((1 - Se)P + Sp(1 - P))$$

Value

A data frame containing the following columns:

sensitivity The sensitivity (proportion) of the screener.

specificity The specificity (proportion) of the screener.

ntpp the number of tests required to discover a single positive test result.

prev_untested The prevalence proportion of the test condition among those who are screened out of testing.

Examples

```
attach(uniobj1)
ntpp(uniobj1)
```

ntpp.logreg_screenr	<i>Compute the Ratio of Total Tests to Positive Results from logreg_screenr Objects</i>
---------------------	-----------------------------------------------------------------------------------------

Description

ntpp.logreg_screenr computes the ratio of the total number of tests performed per positive test result from logreg_screenr-class objects.

Usage

```
## S3 method for class 'logreg_screenr'
ntpp(object, ..., type = c("cvResults", "isResults"), prev = NULL)
```

Arguments

object	a logreg_screenr-class object produced by logreg_screenr.
...	optional arguments to ntp methods.
type	(character) one of "cvResults" (the default) or "isResults" to specify k -fold cross-validated or in-sample receiver-operating characteristics, respectively.
prev	an optional prevalence proportion for the test outcome; if missing the prevalence is obtained from object.

Details

The anticipated number of tests required to detect a single positive $ntpp$ is given by

$$ntpp = (SeP + (1 - Sp)(1 - P))/SeP$$

where Se is sensitivity, P is prevalence and Sp is specificity. The anticipated prevalence among those screened out is given by

$$P_{untested} = ((1 - Se)P)/((1 - Se)P + Sp(1 - P))$$

Value

A data frame containing the following columns:

sensitivity The sensitivity (proportion) of the screener.

specificity The specificity (proportion) of the screener.

ntpp the number of tests required to discover a single positive test result.

prev_untested The prevalence proportion of the test condition among those who are screened out of testing.

Examples

```
attach(uniobj2)
ntpp(uniobj2)
```

ntpp.simple_screenr	<i>Compute the Ratio of Total Tests to Positive Results from simple_screenr Objects</i>
---------------------	-----------------------------------------------------------------------------------------

Description

ntpp.simple_screenr computes the ratio of the total number of tests performed per positive test result from simple_screenr-class objects.

Usage

```
## S3 method for class 'simple_screenr'
ntpp(object, ..., prev = NULL)
```

Arguments

object a simple_screenr-class object produced by simple_screenr.

... optional arguments to ntp methods.

prev an optional prevalence proportion for the test outcome; if missing the prevalence is obtained from object.

Details

The anticipated number of tests required to detect a single positive *nttp* is given by

$$nttp = (SeP + (1 - Sp)(1 - P))/SeP$$

where *Se* is sensitivity, *P* is prevalence and *Sp* is specificity. The anticipated prevalence among those screened out is given by

$$P_{untested} = ((1 - Se)P)/((1 - Se)P + Sp(1 - P))$$

Value

A data frame containing the following columns:

sensitivity The sensitivity (proportion) of the screener.

specificity The specificity (proportion) of the screener.

ntpp the number of tests required to discover a single positive test result.

prev_untested The prevalence proportion of the test condition among those who are screened out of testing.

plot.easy_tool	<i>Plot ROC Curves from easy_tool-Class Objects</i>
----------------	-----------------------------------------------------

Description

plot.easy_tool plots the k -fold cross-validated receiver-operating characteristics, including confidence intervals on the combinations of the local maxima of sensitivity and specificity.

Usage

```
## S3 method for class 'easy_tool'
plot(x, ..., plot_ci = TRUE, conf_level = 0.95, bootreps = 2000)
```

Arguments

x	an object of class easy_tool.
...	any additional arguments passed to pROC::plot.roc or pROC::lines.roc.
plot_ci	(logical) plot confidence intervals if TRUE.
conf_level	confidence level
bootreps	the number of bootstrap replications for estimation of confidence intervals.

Details

plot.easy_tool is an enhanced convenience wrapper for pROC::plot.roc.

Value

This function produces a plot as a side effect and (optionally) returns a dataframe containing sensitivities, specificities and their lower and upper confidence limits for threshold values of $\Pr(\text{response} = 1)$.

References

Fawcett T. An introduction to ROC analysis. Pattern Recognition Letters. 2006. 27(8):861-874. <https://doi.org/10.1016/j.patrec.2005.10.010>

Linden A. Measuring diagnostic and predictive accuracy in disease management: an introduction to receiver operating characteristic (ROC) analysis. Journal of Evaluation in Clinical Practice. 2006; 12(2):132-139. <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1365-2753.2005.00598.x>

Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Muller M. pROC: an open-source package for R and S+ to analyze and compare ROC curves. BMC Bioinformatics 2011; 12:77. <https://www.biomedcentral.com/1471-2105/12/77>

Examples

```
attach(uniobj1)
tool <- easy_tool(uniobj1, max = 3, crossval = TRUE)
plot(tool)
```

plot.lasso_screenr	<i>Plot ROC Curves from lasso_screenr-Class Objects</i>
--------------------	---------------------------------------------------------

Description

plot.lasso_screenr plots the k -fold cross-validated receiver-operating characteristic, including confidence intervals on the combinations of the local maxima of sensitivity and specificity.

Usage

```
## S3 method for class 'lasso_screenr'
plot(
  x,
  ...,
  plot_ci = TRUE,
  model = c("minAIC", "minBIC"),
  conf_level = 0.95,
  bootreps = 2000
)
```

Arguments

x	an object of class lasso_screenr.
...	any additional arguments passed to pROC::plot.roc or pROC::lines.roc.
plot_ci	(logical) plot confidence intervals if TRUE.
model	(character) select either the model which produced the minimum AIC ("minAIC") or minimum BIC ("minBIC").
conf_level	confidence level
bootreps	the number of bootstrap replications for estimation of confidence intervals.

Details

Plot cross-validated (out-of-sample) ROC curve with pointwise confidence intervals along with the overly optimistic in-sample ROC curve. `plot.lasso_screenr` is an enhanced convenience wrapper for `pROC::plot.roc`.

Value

This function produces a plot as a side effect.

References

Fawcett T. An introduction to ROC analysis. *Pattern Recognition Letters*. 2006. 27(8):861-874. <https://doi.org/10.1016/j.patrec.2005.10.010>

Linden A. Measuring diagnostic and predictive accuracy in disease management: an introduction to receiver operating characteristic (ROC) analysis. *Journal of Evaluation in Clinical Practice*. 2006; 12(2):132-139. <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1365-2753.2005.00598.x>

Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Muller M. pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC Bioinformatics* 2011; 12:77. <https://www.biomedcentral.com/1471-2105/12/77>

Examples

```
attach(uniobj1)
plot(uniobj1, model = "minAIC")
```

`plot.logreg_screenr` *Plot ROC Curves from logreg_screenr-Class Objects*

Description

`plot.logreg_screenr` plots the k -fold cross-validated receiver-operating characteristic, including confidence intervals on the combinations of the local maxima of sensitivity and specificity.

Usage

```
## S3 method for class 'logreg_screenr'
plot(x, ..., plot_ci = TRUE, conf_level = 0.95, bootreps = 2000)
```

Arguments

<code>x</code>	an object of class <code>logreg_screenr</code> .
<code>...</code>	additional arguments passed to plot.roc and friends.
<code>plot_ci</code>	logical indicator for plotting point-wise confidence intervals at the locally maximum subset of coordinates for on sensitivity and specificity (default = TRUE). See also ci.thresholds .

conf_level	confidence level in the interval (0,1). Default is 0.95 producing 95% confidence intervals.
bootreps	number of bootstrap replications for estimation of confidence (default = 2000).

Details

Plot cross-validated (out-of-sample) ROC curve with pointwise confidence intervals along with the overly optimistic in-sample ROC curve. `plot.lasso_screenr` is an enhanced convenience wrapper for `pROC::plot.roc`.

Value

This function produces a plot as a side effect.

References

- Fawcett T. An introduction to ROC analysis. *Pattern Recognition Letters*. 2006. 27(8):861-874. <https://doi.org/10.1016/j.patrec.2005.10.010>
- Linden A. Measuring diagnostic and predictive accuracy in disease management: an introduction to receiver operating characteristic (ROC) analysis. *Journal of Evaluation in Clinical Practice*. 2006; 12(2):132-139. <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1365-2753.2005.00598.x>
- Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Muller M. pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC Bioinformatics* 2011; 12:77. <https://www.biomedcentral.com/1471-2105/12/77>

Examples

```
attach(uniobj2)
plot(uniobj2)
```

`plot.simple_screenr` *Plot ROC Curves from simple_screenr-Class Objects*

Description

`plot.simple_screenr` plots the k -fold cross-validated receiver-operating characteristic, including confidence intervals on the combinations of the local maxima of sensitivity and specificity.

Plot ROC curve with pointwise 95 intervals on sensitivity and specificity and (optionally) returns a dataframe containing numerical values.

Usage

```
## S3 method for class 'simple_screenr'
plot(x, ..., plot_ci = TRUE, conf_level = 0.95, bootreps = 2000)
```

Arguments

<code>x</code>	an object of class <code>simple_screenr</code> .
<code>...</code>	additional arguments for <code>\link{plot}</code> or passed to <code>\link{plot.roc}</code> and friends.
<code>plot_ci</code>	logical indicator for plotting point-wise confidence intervals at the locally maximum subset of coordinates for on sensitivity and specificity (default = TRUE). See also ci.thresholds .
<code>conf_level</code>	confidence level in the interval (0,1). Default is 0.95 producing 95% confidence intervals.
<code>bootreps</code>	numeric-valued number of bootstrap replication for estimation of 95% confidence intervals.

Value

This function produces a plot as a side effect, and (optionally) returns a dataframe containing medians and bootstrap confidence limits of sensitivity and specificity.

References

- Fawcett T. An introduction to ROC analysis. *Pattern Recognition Letters*. 2006. 27(8):861-874. <https://doi.org/10.1016/j.patrec.2005.10.010>
- Linden A. Measuring diagnostic and predictive accuracy in disease management: an introduction to receiver operating characteristic (ROC) analysis. *Journal of Evaluation in Clinical Practice*. 2006; 12(2):132-139. <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1365-2753.2005.00598.x>
- Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Muller M. pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC Bioinformatics* 2011; 12:77. <https://www.biomedcentral.com/1471-2105/12/77>

Examples

```
data(unicorns)
too_simple <- simple_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7,
                             data = unicorns)
plot(too_simple)
```

`predict.lasso_screenr` *A Prediction Method for lasso_screenr-Class Objects*

Description

`predict.lasso_screenr` computes predicted probabilities of positive test results from new data.

Usage

```
## S3 method for class 'lasso_screenr'
predict(object = NULL, ..., newdata = NULL)
```

Arguments

object	an object of class lasso_screenr produced by <code>`lasso_screenr`</code> .
...	optional arguments to predict methods.
newdata	new dataframe from which predicted probabilities of positive test results are desired. The dataframe must contain values of the same response variables and covariates that were used to obtain obj.

Details

This method is a convenience wrapper for ``glmpath::predict.glmpath``.

Value

`predict.lasso_screenr` returns (invisibly) a dataframe augmenting the complete cases in `newdata` with the predicted probabilities of positive test results `phat_minAIC` and `phat_minBIC` from the models that produced the minimum AIC and BIC, respectively.

Examples

```
attach(uniobj1)
## Get some new observations
new_corns <- data.frame(ID = c("Alice D.", "Bernie P."),
                        testresult = c(NA, NA),
                        Q1 = c(0, 0), Q2 = c(0, 0), Q3 = c(0, 1), Q4 = c(0, 0),
                        Q5 = c(0, 1), Q6 = c(0, 1), Q7 = c(0, 1))
## Predict the probabilities of testing positive for the new subjects
predict(uniobj1, newdata = new_corns)
```

predict.logreg_screenr

A Prediction Method for logreg_screenr-Class Objects

Description

`predict.logreg_screenr` computes predicted probabilities of positive test results from new data.

Usage

```
## S3 method for class 'logreg_screenr'
predict(object = NULL, ..., newdata = NULL)
```

Arguments

object	an object of class logreg_screenr produced by <code>`logreg_screenr`</code> .
...	optional arguments to predict methods.
newdata	new dataframe from which predicted probabilities of positive test results are desired. The dataframe must contain values of the same response variables and covariates that were used to obtain object.

Details

This method is a convenience wrapper for ``stats::predict.glm``.

Value

`predict.logreg_screenr` returns (invisibly) a dataframe augmenting newdata with the predicted probabilities of positive test results `phat`.

Examples

```
attach(uniobj2)
## Get some new observations
new_corns <- data.frame(ID = c("Alice D.", "Bernie P."),
                        testresult = c(NA, NA), Q1 = c(0, 0), Q2 = c(0, 0),
                        Q3 = c(0, 0), Q4 = c(0, 0), Q5 = c(0, 1), Q6 = c(0, 1 ),
                        Q7 = c(0, 1))
## Predict the probabilities of testing positive for the new subjects
predict(uniobj2, newdata = new_corns)
```

print.easy_tool

A Print Method for easy_tool-Class Objects

Description

`print.easy_tool` is a print method.

Usage

```
## S3 method for class 'easy_tool'
print(x, ...)
```

Arguments

```
x          an object of class easy_tool.
...        optional arguments to print methods.
```

See Also

`get_what.easy_tool(from,what)` for `what = "ROCci"`.

Examples

```
attach(uniobj1)
print(uniobj1)
```

print.lasso_screenr *A Print Method for lasso_screenr-Class Objects*

Description

print.lasso_screenr is a print method for lasso_screenr-class objects.

Usage

```
## S3 method for class 'lasso_screenr'  
print(x, ...)
```

Arguments

x	an object of class lasso_screenr
...	optional arguments to print methods.

See Also

get_what.lasso_screenr(from, what) for what = "ROCci".

Examples

```
attach(uniobj1)  
print(uniobj1)
```

print.logreg_screenr *A Print Method for logreg_screenr-Class Objects*

Description

print.logreg_screenr is a print method for logreg_screenr-class objects.

Usage

```
## S3 method for class 'logreg_screenr'  
print(x, ..., quote = FALSE)
```

Arguments

x	an object of class logreg_screenr.
...	optional arguments to print methods.
quote	logical indicator for whether or not strings should be printed.

Nothing. Thresholds, specificities and sensitivities are printed as a side effect.

get_what.logreg_screenr(from,what) for what = "ROCci".

```
attach(uniobj2)
print(uniobj2)
```

`print.simple_screenr` *A Print Method for simple_screenr-Class Objects*

`print.simple_screenr` is print method for `simple_screenr` objects.

```
## S3 method for class 'simple_screenr'
print(x, ...)
```

x	an object of class <code>simple_screenr</code> .
...	optional arguments to print methods.

Nothing. Thresholds, specificities and sensitivities are printed as a side effect.

```
get_what.simple_screenr(from,what) for what = "ROCci".
```

[illegible]

rescale_to_int

*Rescale Positive Vectors or Matrices to Integers***Description**

rescale_to_int rescales the *non-zero* elements of real-valued numeric vectors or matrices to integers in the closed interval $[1, \text{max}]$. Any zero-valued elements are left unchanged.

Usage

```
rescale_to_int(x, max, colwise = TRUE)
```

Arguments

x	numeric matrix or vector of non-negative real numbers.
max	the value of largest element in the rescaled integer-valued vector.
colwise	(logical) rescale the matrix by column if TRUE (the default) or by row if FALSE.

Value

A matrix of integers corresponding to x in which smallest *non-zero* element in each column/row is 1 and the largest element is max. Any elements having value zero are unchanged. If x is a vector then the result is a $r \times 1$ matrix, where r is the number of elements in x. Otherwise the result is a $r \times c$ matrix where c is the number of columns in x.

See Also

[rescale](#)

Examples

```
x <- c(0.55, 1.21, 0.94, 0, 0.13)
rescale_to_int(x, max = 5)
```

roc_ci

*Compute Bootstrap Confidence Limits for Sensitivities and Specificities***Description**

roc_ci computes bootstrap confidence intervals from objects of class roc, as produced by the pROC package. roc_ci is simply a convenience wrapper for pROC::ci.thresholds re-formatted for screenr.

Usage

```
roc_ci(
  object,
  bootreps = 2000,
  conf.level = 0.95,
  progress = "none",
  thresholds = "local maximas",
  se.min = 0.5
)
```

Arguments

<code>object</code>	an object of class <code>roc</code> .
<code>bootreps</code>	number of bootstrap replicates (default = 2000).
<code>conf.level</code>	confidence level for uncertainty intervals (default = 0.95).
<code>progress</code>	type of progress display (see <code>help(pROC::ci.thresholds)</code>).
<code>thresholds</code>	type of thresholds (see <code>help(pROC::ci.thresholds)</code>).
<code>se.min</code>	minimum value of sensitivity returned.

Value

a data frame containing thresholds with sensitivities, specificities and uncertainty intervals.

See Also

[ci.thresholds](#)

screenr

screenr Package

Description

The `screenr` package enables construction of binary test-screening tools. It is designed to enable those with only a basic familiarity with R to develop, validate and implement screening tools for diagnostic tests.

Consider the situation where a diagnostic test for some condition is relatively expensive, and the condition is rare. In that case, universal testing would not be efficient in terms of the yield of positive results per test performed. Now suppose that responses to a set of simple questions may be predictive of the condition. Package `screenr` enables estimation of thresholds for making decisions about when to test in order to screen in/out individuals based on Receiver Operating Characteristics (ROC) estimated from an initial sample. The choice of a particular screening threshold is left to the user, and should be based on careful consideration of application-specific tradeoffs between sensitivity and specificity. `screenr` also enables easy construction of screening tools.

Details

The high-level functions in the screenr package are:

`lasso_screenr` Test-screening based on GLM path regularization of logistic regression models

`logreg_screenr` Test-screening based maximum-likelihood estimation of logistic regression models

`easy_tool` Easy implementation of test-screening tools

`simple_screenr` Simple un-optimized test-screening

`rescale_to_int` Rescale a strictly positive vector of real numbers to integers

`sens_spec_plus` Sensitivity, specificity and friends

There are plot, print, summary, predict, get_what, and ntp methods for the objects produced by `lasso_screenr`, `logreg_screenr`, `simple_screenr` and `easy_tool`. In addition, there is a `coef` method for `lasso_screenr` and `logreg_screenr` objects.

Note

A tutorial is available from `vignette("screenr_Tutorial", package = "screenr")`

The canonical source for screenr is <https://github.com/sgutreuter/screenr>

Author(s)

Steve Gutreuter: <sgutreuter@gmail.com>

sens_spec_plus	<i>Compute Sensitivity, specificity and a few friends</i>
----------------	-----------------------------------------------------------

Description

`sens_spec_plus` computes sensitivity, specificity and a few friends from a gold standard and testing results. `sens_spec_plus` is a convenience wrapper for `epiR::epi.tests`.

Usage

```
sens_spec_plus(
  test = NULL,
  gold = NULL,
  data = NULL,
  method = c("exact", "wilson", "agresti", "clopper-pearson", "jeffreys"),
  conf.level = 0.95
)
```

Arguments

<code>test</code>	character-valued name of the variable containing testing results, code as 0 for negative and 1 for positive.
<code>gold</code>	character-valued name of the variable containing gold standard, code as 0 for negative and 1 for positive.
<code>data</code>	data frame containing <code>test</code> and <code>gold</code> .
<code>method</code>	type of confidence interval ("exact", "wilson", "agresti", "clopper-pearson" or "jeffreys"). The default is "exact".
<code>conf.level</code>	confidence level, a numeric value between 0 and 1.

Value

a list containing components `table` and `ests`:

`table` a 2 x 2 cross-tabulation which is the anti-transpose of the result produced by `base::table(gold, test)`.

`ests` a dataframe containing the apparent and true positive proportions, sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), and the lower and upper confidence limits for each.

See Also

[epi.tests](#)

Examples

```
Gold <- rbinom(20, 1, 0.50)
Test <- Gold; Test[c(3, 5, 9, 12, 16)] <- 1 - Test[c(3, 5, 9, 12, 16)]
dat <- data.frame(Gold = Gold, Test = Test)
sens_spec_plus(test = "Test", gold = "Gold", data = dat)
```

simple_screenr

An Overly Simple Approach to Test Screening

Description

`simple_screenr` implements the method described in Bandason et al. (2016).

Usage

```
simple_screenr(formula, data)
```

Arguments

formula	an object of class <code>formula</code> defining the testing outcome and predictor covariates.
data	the "training" sample; a data frame containing the testing outcome and predictive covariates to be used for testing screening. The testing outcome must be binary (0,1) indicating negative and positive test results, respectively, or logical (TRUE/FALSE), and the screening scores are the row-wise sums of the values of those covariates. The covariates are typically binary (0 = no, 1 = yes) responses to questions, but the responses may also be ordinal numeric values.

Details

`simple_screenr` computes the in-sample (*overly optimistic*) performances for development of a very simple test screening tool based on the sums of affirmative questionnaire responses. `simpleScreenr` is not optimized and is intended only for comparison with `lasso_screenr` or `logreg_screenr`, either of which will almost certainly out-perform `simple_screenr`.

Value

An object of class `simple_screenr` containing the elements:

Call The function call.

Prevalence Prevalence of the test condition in the training sample.

ISroc An object of class **roc** containing the "in-sample" (overly-optimistic) receiver operating characteristics, and additional functions for use with this object are available in the **pROC** package.

Scores The training sample, including the scores.

References

Bandason T, McHugh G, Dauya E, Mungofa S, Munyati SM, Weiss HA, Mujuru H, Kranzer K, Ferrand RA. Validation of a screening tool to identify older children living with HIV in primary care facilities in high HIV prevalence settings. *AIDS*. 2016;30(5):779-785 <http://dx.doi.org/10.1097/OAD.0000000000000959>

Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez J-C, Müller M. pROC: An open-source package for R and S+ to analyze and compare ROC curves. BMC Bioinformatics. 2011;12(77):1-8. <http://doi.org/10.1186/1471-2105-12-77>

See Also

[easy_tool](#) for a better approach to simplification using the results from `lasso_screenr` or `logreg_screenr`.
[lasso_screenr](#), [logreg_screenr](#)

Examples

[illegible]

summary.easy_tool	<i>A Summary Method for easy_tool-Class Objects</i>
-------------------	-----------------------------------------------------

Description

summary.easy_tool provides a summary method for easy-tool-class objects.

Usage

```
## S3 method for class 'easy_tool'
summary(object, ...)
```

Arguments

object	an easy_tool object.
...	optional arguments passed to summary methods.

Details

This is essentially a wrapper for `glmpath::summary.glmpath` provided for `lasso_screenr` objects.

Value

a dataframe containing the summary, including the Df, Deviance, AIC and BIC for each step along the GLM path for which the active set changed.

Examples

```
attach(uniobj1)
summary(uniobj1)
```

summary.lasso_screenr	<i>A Summary Method for lasso_screenr-Class Objects</i>
-----------------------	---------------------------------------------------------

Description

summary.lasso_screenr provides a summary method for lasso_screenr-class objects.

Usage

```
## S3 method for class 'lasso_screenr'
summary(object, ...)
```

Arguments

object a lasso_screenr object
 ... optional arguments passed to summary methods.

Details

This is essentially a wrapper for `glmpath::summary.glmpath` provided for `lasso_screenr` objects.

Value

a dataframe containing the summary, including the Df, Deviance, AIC and BIC for each step along the GLM path for which the active set changed.

Examples

```
attach(uniobj1)
summary(uniobj1)
```

```
summary.logreg_screenr
```

A Summary Method for logreg_screenr-Class Objects

Description

`summary.logreg_screenr` provides a summary method for `logreg_screenr`-class objects.

Usage

```
## S3 method for class 'logreg_screenr'
summary(object, ..., diagnostics = FALSE)
```

Arguments

object an object of class `logreg_screenr` produced by function `logreg_screenr`.
 ... optional arguments passed to summary methods.
 diagnostics a logical value; plot model diagnostics if TRUE.

Value

Nothing. Summaries are printed as a side effect.

Examples

```
attach(uniobj2)
summary(uniobj2)
```

```
summary.simple_screenr
```

A Summary Method for simple_screenr-Class Objects

Description

`summary.simple_screenr` provides a summary method for `simple_screenr`-class objects.

Usage

```
## S3 method for class 'simple_screenr'
summary(object, ...)
```

Arguments

`object` an object of class `simple_screenr`.
`...` optional arguments passed to summary methods.

Value

Nothing. Thresholds, specificities and sensitivities are printed as a side effect.

Examples

```
data(unicorns)
toosimple <- simple_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7,
                           data = unicorns)
summary(toosimple)
```

```
unicorns
```

UIV Testing Training Data on Unicorns

Description

A preliminary study was conducted in which a random sample of 6,000 properly consented [unicorns](<https://www.britannica.com/topic/unicorn>) were recruited from 20 clinics. Each unicorn was asked seven questions about their behavior and health. Unicorns responded by stomping a hoof once to indicate "no", and twice to indicate "yes". A sample of venous blood was drawn from each, and was subsequently tested for the presence of antibodies to Unicorn Immunodeficiency Virus (UIV) using a standard assay algorithm.

Usage

```
data(unicorns)
```


Format

A data frame with eight columns:

ID Patient ID

Q1 Response to screening question 1 (0 = "no", 1 = "yes")

Q2 Response to screening question 2 (0 = "no", 1 = "yes")

Q3 Response to screening question 3 (0 = "no", 1 = "yes")

Q4 Response to screening question 4 (0 = "no", 1 = "yes")

Q5 Response to screening question 5 (0 = "no", 1 = "yes")

Q6 Response to screening question 6 (0 = "no", 1 = "yes")

Q7 Response to screening question 7 (0 = "no", 1 = "yes")

testresult UIV status, where 0 and 1 denote negative and positive test results, respectively.

Note

In reality, the question responses and test results were generated using Bernoulli random-number generators.

Examples

```
## Not run:
head(unicorns)

## End(Not run)
```

uniobj1

A lasso_screenr object

Description

The result of `uniobj1 <- lasso_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7, data = unicorns, Nfolds = 10, seed = 123)`

Usage

```
uniobj1
```

Format

An object of class `lasso_screenr`

Examples

```
## Not run:
summary(uniobj1)

## End(Not run)
```

uniobj2	A logreg_screenr object
---------	-------------------------

Description

The result of `uniobj2 <- logreg_screenr(testresult ~ Q1 + Q2 + Q3 + Q4 + Q5 + Q6 + Q7, data = unicorns, link = "logit", Nfolds = 10, seed = 123)`

Usage

```
uniobj2
```

Format

An object of class `logreg_screenr`

Examples

```
## Not run:
summary(uniobj2)

## End(Not run)
```

val_data	<i>UIV Test Validation Data on Unicorns</i>
----------	---------------------------------------------

Description

A follow-up study was conducted in which a random sample of 3,000 properly consented unicorns were recruited from 20 additional clinics. Each unicorn was asked six questions about their behavior and health. Unicorns responded by stomping a hoof once to indicate "no", and twice to indicate "yes". A sample of venous blood was drawn from each, and was subsequently tested for the presence of antibodies to Unicorn Immunodeficiency Virus (UIV) using a standard assay algorithm.

Usage

```
val_data
```

Format

A data frame with eight columns:

```
ID Patient ID
Q1 Response to screening question 1 (0 = "no", 1 = "yes")
Q2 Response to screening question 2 (0 = "no", 1 = "yes")
Q3 Response to screening question 3 (0 = "no", 1 = "yes")
```

Q4 Response to screening question 4 (0 = "no", 1 = "yes")

Q5 Response to screening question 5 (0 = "no", 1 = "yes")

Q6 Response to screening question 6 (0 = "no", 1 = "yes")

Q7 Response to screening question 7 (0 = "no", 1 = "yes")

testresult UIV status, where 0 and 1 denote negative and positive test results, respectively.

Examples

```
## Not run:
```

```
head(val_data)
```

```
## End(Not run)
```

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