





# SeqScreen Software Overview

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# **IARPA Fun GCAT Program**



Home Research Office of Collection Fun GCAT

# **FUN GCAT**

### FUNCTIONAL GENOMIC AND COMPUTATIONAL ASSESSMENT OF THREATS



#### INTELLIGENCE VALUE

The Fun GCAT is developing methods to rapidly assess the function of DNA sequences to determine if they pose a threat. Used to automatically process large datasets or to supplement subject matter expert review. Fun GCAT technology will enable improved detection of bio-error or bio-terror.

Current screening methods to flag dangerous DNA sequences are inadequate—they do not consider DNA function, cannot process short or highly engineered sequences, and often require follow-up analysis by an expert. Fun GCAT is developing smart, Al-driven threat screening software to replace current look-up tablebased screening. Fun GCAT researchers developed computational pipelines to analyze DNA and answer three questions per sequence: What organism does it come from? What biological functions does it have? How dangerous is it? By using neural networks and other powerful bioinformatic techniques to learn the common patterns of sequences with similar origins and functions, Fun GCAT tools are demonstrating high predictive accuracy against increasingly challenging test sets. Benchmarking has demonstrated significant performance increases beyond top winners in a closely related bioinformatic software development global challenge. It resulted in 500x improvement in computational efficiency over state-of-the-art and stable performance on even short (<50 base pairs) sequences. This enabled a range of Intelligence Community-relevant missions from rapid screening of very large datasets to field-based, targeted analysis.

Fun GCAT is also developing new approaches to meet the demand for rapid, relatively high-throughput experimental assessment of DNA function. The program has developed optical, cell-based tests to identify disruptors of the immune system hardwired within cells. New functions for dozens of virus genes have been discovered and the technologies are being leveraged to develop threat prediction capabilities for faster

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#### RESEARCH AREA(S)

Machine learning, Synthetic biology, Threat determination, Taxonomic classification, Biosecurity, Bioinformatics, DNA screening, Gene function, Viral, Toxins, Innate immunity, Microscopy, DNA synthesis

## BROAD AGENCY ANNOUNCEMENT

LINK(S) TO BAA

■ IARPA-BAA-16-08

SOLICITATION STATUS

## Disclaimer for our Fun GCAT work:

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# Software Developed under IARPA Fun GCAT Program

## **SeqScreen Fast and Sensitive Modes:**

Input = Short Sequences
Processing Goal = Linux Server

## **SeqScreen ONT Mode:**

Input = Nanopore Sequences
Processing Goal = Laptop









https://gitlab.com/treangenlab/seqscreen

Machine Learning (ML) Database

Open-Source ML-Based Functional Annotation Software



S2FAS







# **SeqScreen Publication and GitLab Repo**

Balaji et al. Genome Biology (2022) 23:133 https://doi.org/10.1186/s13059-022-02695-x

Genome Biology

SOFTWARE Open Access

# SeqScreen: accurate and sensitive functional screening of pathogenic sequences via ensemble learning



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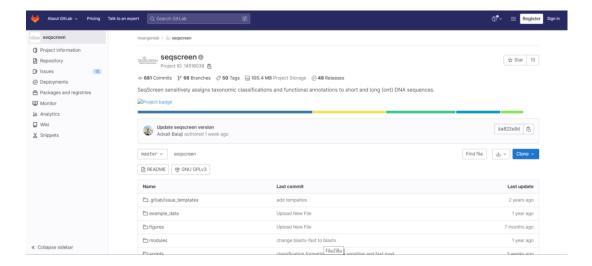
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#### Abstract

The COVID-19 pandemic has emphasized the importance of accurate detection of known and emerging pathogens. However, robust characterization of pathogenic sequences remains an open challenge. To address this need we developed SeqScreen, which accurately characterizes short nucleotide sequences using taxonomic and functional labels and a customized set of curated Functions of Sequences of Concern (FunSoCs) specific to microbial pathogenesis. We show our ensemble machine learning model can label protein-coding sequences with



https://gitlab.com/treangenlab/seqscreen

https://link.springer.com/article/10.1186/s13059-022-02695-x







# **Installation Tips**

- SeqScreen is easiest to install via conda or mamba, or as a conda-pack or Docker/Singularity container if you are working on an air-gapped system
- There is a --check\_install option to check that the required command line tools, python imports, and database files are present before you run the software
- Please see our wiki documentation for additional information: <a href="https://gitlab.com/treangenlab/seqscreen/">https://gitlab.com/treangenlab/seqscreen/</a> /-/wikis/Home

## Home

- 01. SeqScreen Overview
- 02. SeqScreen Dependencies
- · 03. Installation and Execution
- 04. Initialization Workflow
- 05. SeqMapper Workflow
- 06. Taxonomic Identification
   Workflow
- 07. Functional Annotation Workflow
- 08. Identifying Functions of Sequences of Concern
- 09. Report Generation Workflow
- 10. HTML Report
- 11. Frequently Asked Questions







# **Three Different SeqScreen Modes**

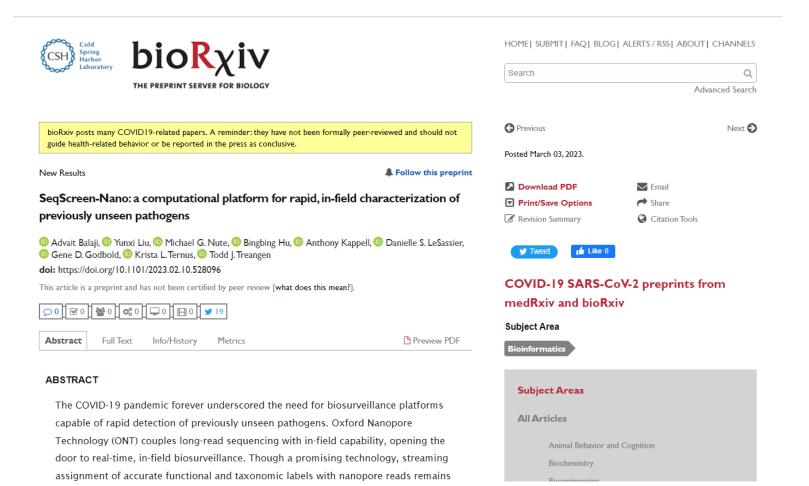
- Fast mode (default)
  - Good choice for large datasets, uses DIAMOND instead of BLAST
  - Only one protein-coding region expected per read
- Sensitive mode
  - Ideal for deeply characterizing each read, uses BLASTX and optionally BLASTN
  - Only one protein-coding region expected per read
- ONT mode
  - Suitable when memory is limited, like in today's course
  - More than one protein-coding region is expected per read (e.g., nanopore sequences, contigs, vector sequences)
- All modes are run within Nextflow







# **SeqScreen-Nano Preprint (i.e., ONT Mode)**



Manuscript in prep

https://www.biorxiv.org/content/10.1101/2023.02.10.528096v2.abstract







# **Many Other Execution Options**

--fasta Path to the input NT FASTA file Path to the databases directory containing centrifuge, blast, go etc --databases --working Path to the output working directory Number of threads to use (Default=1) --threads You can see a full list of Use SegScreen sensitive mode (old default mode) --sensitive Enable SegScreen to run ONT reads --ont options with the --help Cutoff to use for blastx/diamond --evalue --hmmscan Run hmmscan on input sequences command or in our software Tiebreak across all proteins within this % of the top bitscore --bitscore Include all ancestral GO terms in output --ancestral --splitby Max number of sequences in an input chunk to diamond --includecc Include cellular component go terms --blastn Run blastn in addition to blastx (sensitive mode only) --taxlimit Maximum number of multi-taxIDs to output for a single query in fast mode Have pipeline modules run on SLURM execution nodes (Default = run locally) --slurm --report\_prefix Add prefix to beginning of segscreen\_report.tsv and segscreen\_html\_report.zip. The prefix will --skip\_report Skip\_report generation step and only generate intermediate files --report\_only Remove intermediate output and only save the results in {output\_dir}/report\_generation Format type: [1] Original, [2] Hits only, [3] FunSoC only, [4] Gene-Centric, [5] Gene-Centric F --format --online Pull reference genomes from NCBI for reference\_inference [Needs web access] --filter\_taxon Filter comma separated list of taxon --keep\_taxon Keep comma separated list of taxon --taxonomy\_confidence\_threshold Confidence threshold for multi-taxids (Average) [Default 0.0] --keep\_html\_ont Keep html report in ont mode [Takes additional memory] --check\_install Check for required command line tools, python imports, and database files Display the version and exit --version







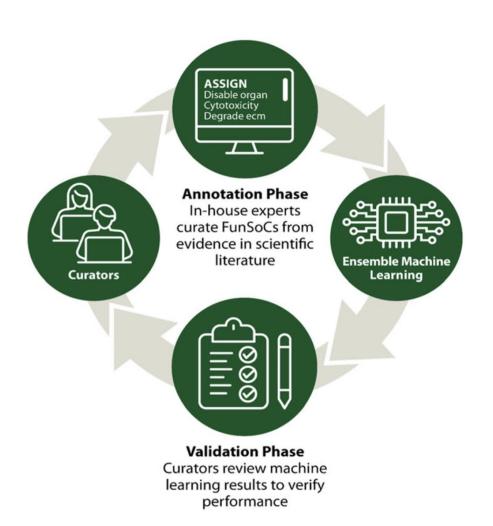
documentation

--help

Print this help message out

# **Human In-the-Loop Pathogen Prediction**

Predict FunSoCs on targeted sets of sequences contained in database



Have biocurators review, provide feedback on good predictions and bad ones

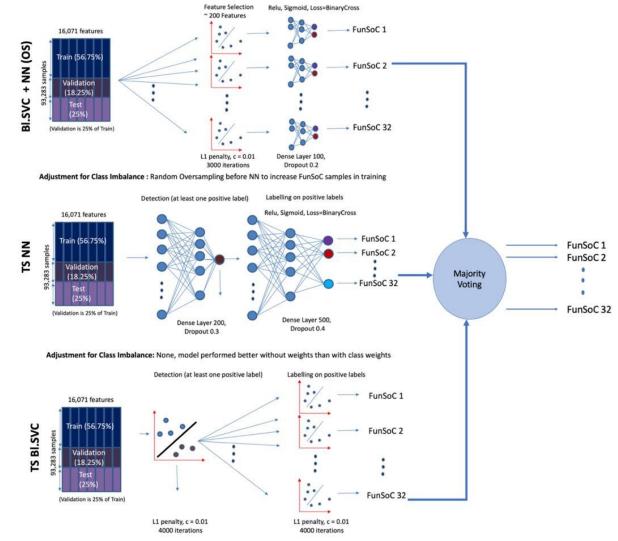








# Human In-the-Loop Pathogen Prediction



Adjustment for Class Imbalance: Class weights passed to SVC via the class\_weights ="balanced" parameter

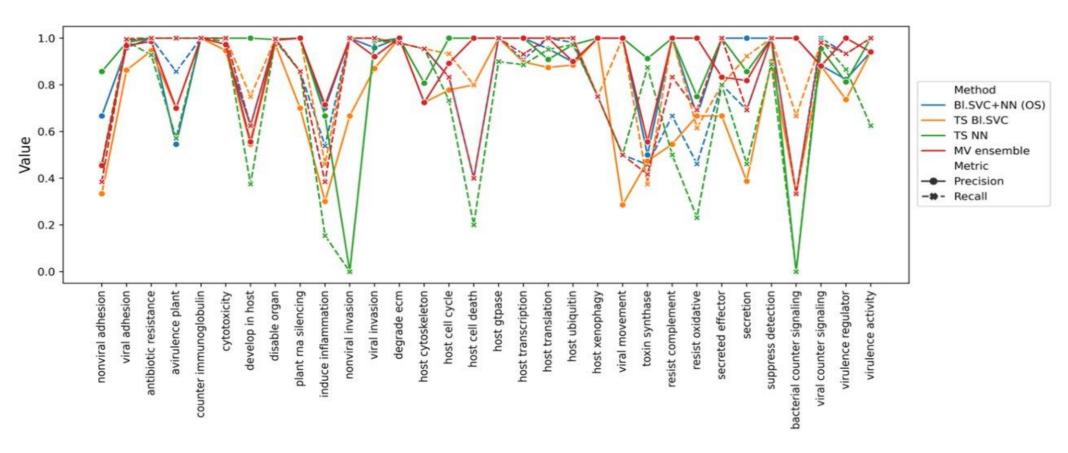






## **Peformance across FunSoCs**

Top Performing models: Positive Label Precision and Recall per FunSoC

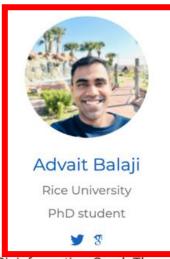


FunSoCs









Bioinformatics, Graph Theory, Pathogen Detection and Analysis, Machine Learning, Microbial genomics and Metagenomics



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String algorithms, Pangenomics, High performance computing, Cheminformatics



Kristen Curry Rice University PhD student



Computational Biology, Bioinformatics, Microbial genomics and Metagenomics



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Computational Biology, Bioinformatics, Microbial genomics and Metagenomics



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Computational Biology, Bioinformatics, Microbial genomics and Metagenomics, Graph algorithms and data structures, Deep learning



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