

Machine Learning Methods for Neural Data Analysis

Lecture 4: Calcium imaging

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STATS 220/320 (*NBIO220, CS339N*). Winter 2021.

Announcements

- Lab 2 due Thursday, 11:59pm.
- We created a Piazza course where you can discuss with one another. Don't share answers directly, but hints, clarifications, and course discussions are fine.
- Canvas will remain the main source for announcements and course content.
- See Han's announcement about how to save notebooks to PDF.

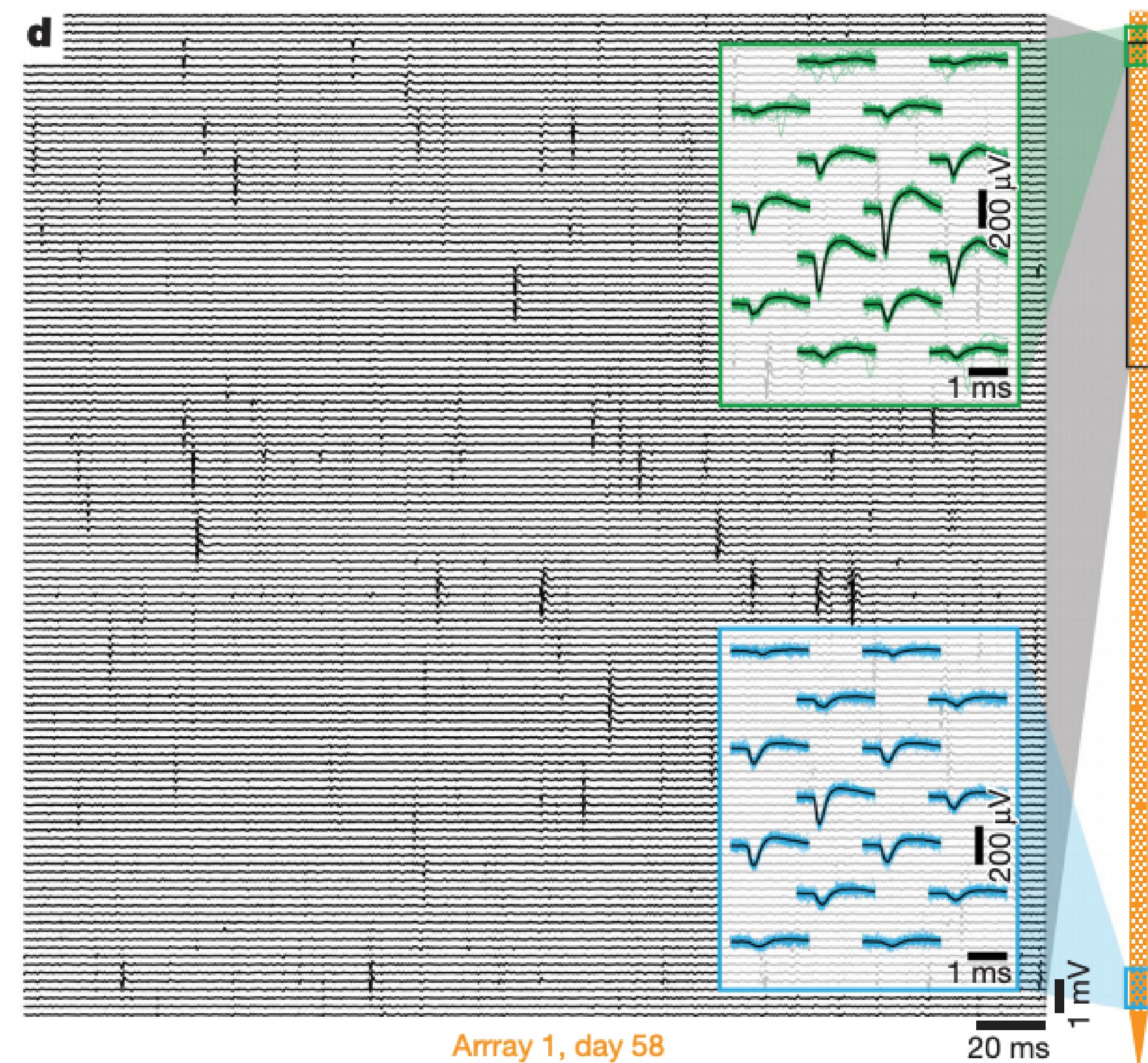
Agenda

1. Optical physiology
2. Constrained Non-negative Matrix Factorization (CNMF)

Recap

Electrophysiology

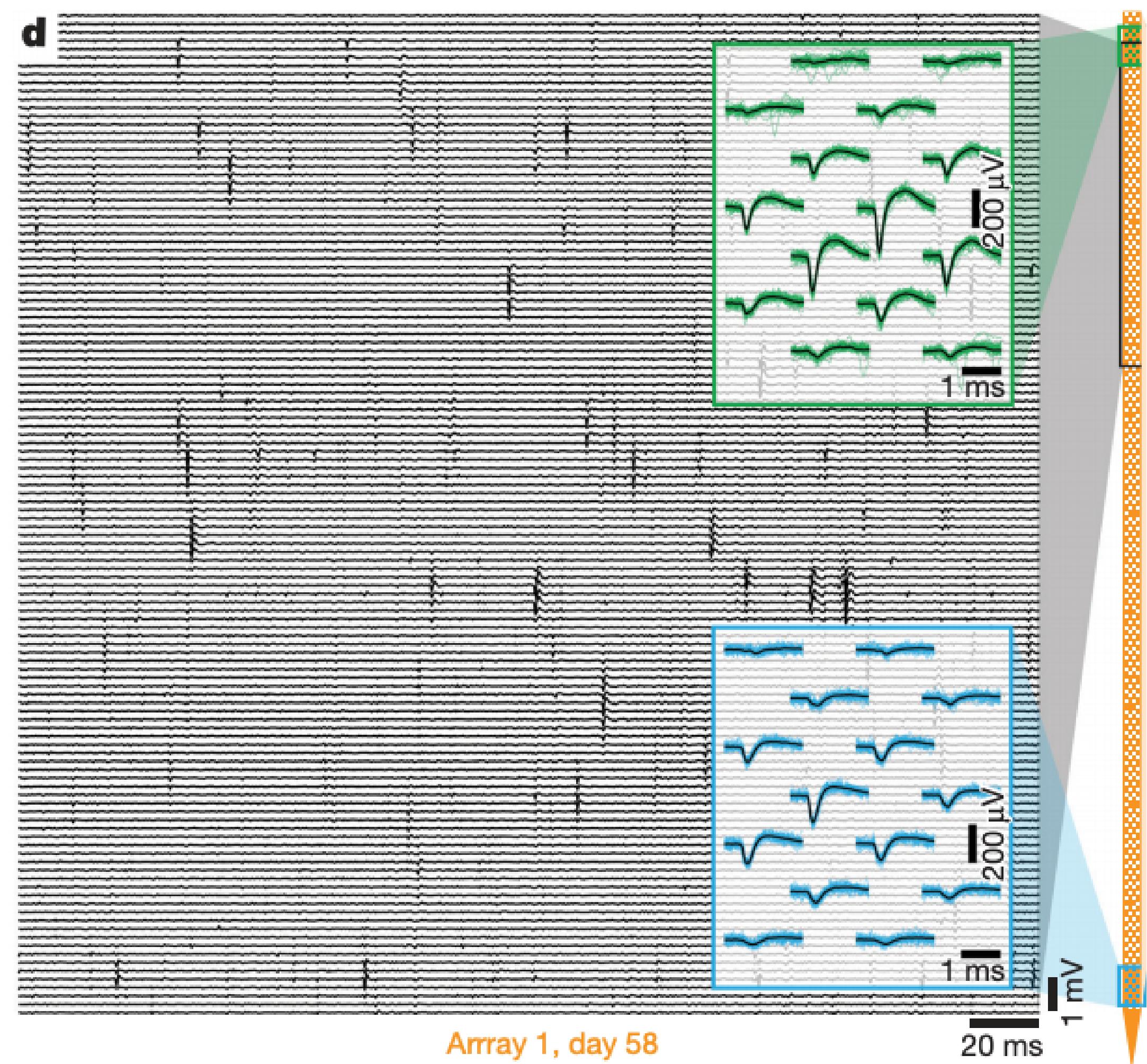
- So far, we've study electrophysiological ("ephys") recordings with high density probes.
- The raw data is a **multidimensional time series of voltage measurements**, one for each recording site on the probe.
- When neurons near the probe fire an **action potential**, it registers a **spike in the voltage** on nearby channels.
- Typical recordings detect spikes from **O(100) neurons**.



Recap

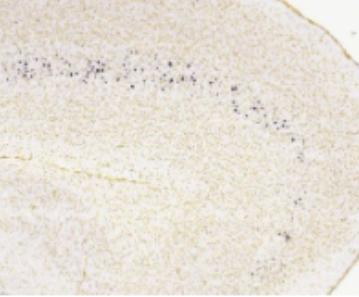
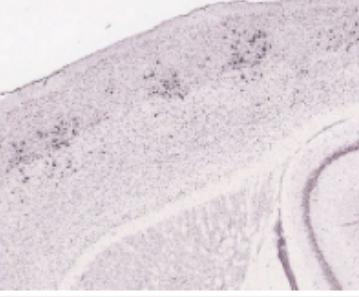
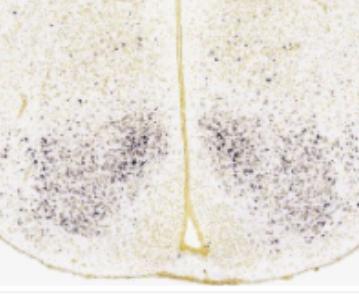
Electrophysiology Limitations

- It's hard to detect neurons that fire rarely and produce low amplitude EAPs.
- More generally, you only pick up cells that happen to be close to the narrow probe.
- No cell-type specificity.
- In particular, ephys does not leverage our powerful genetic toolkits for certain model organisms.



Genetic tools

Cre driver lines in mice

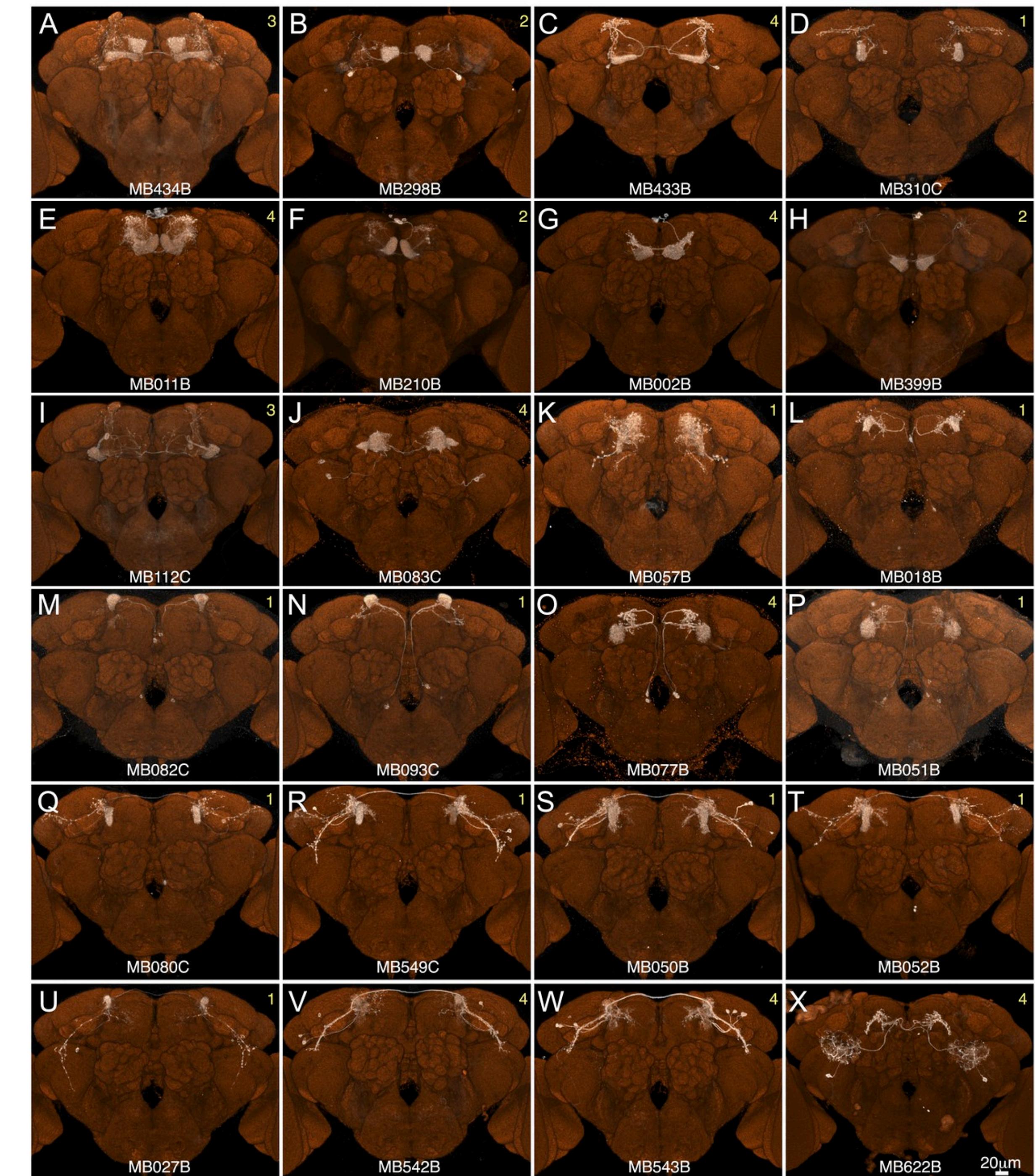
Drivers	Reporters
Data detailing transgene expression in Cre and other driver lines for adult and developing brain. Experiments include colorimetric in situ hybridization, fluorescent in situ hybridization and other histological methods.	
Line Name	
A930038C07Rik-Tg1-Cre Allen Institute for Brain Science	
A930038C07Rik-Tg4-Cre Allen Institute for Brain Science	
Adcyap1-2A-Cre Allen Institute for Brain Science	
Agp-IRES-Cre Bradford Lowell	
Avp-IRES2-Cre Allen Institute for Brain Science	

Genetic tools GAL4 lines in flies

<https://www.janelia.org/node/45217>



Split-GAL4 lines for MBONs



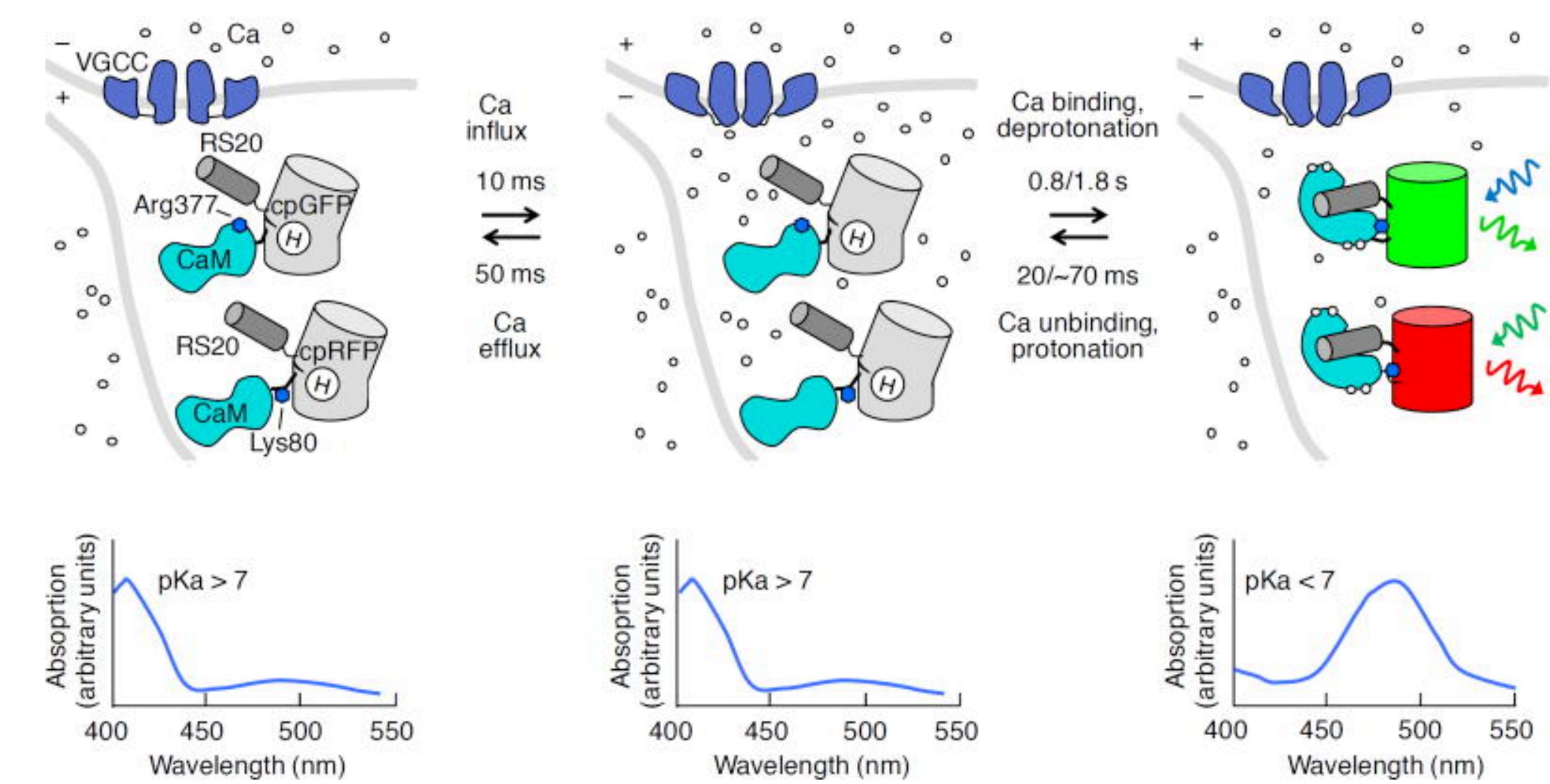
Genetically encoded indicators of neural activity

How can we make cells fluoresce only when they spike?

1. Look for a side effect of spiking.
2. Engineer a protein that fluoresces when that side effect is detected.
3. Modify the DNA of (subsets of) neurons to produce that protein.
4. Use a microscope to measure fluorescence in the genetically modified organism.

Genetically encoded calcium indicators (GECIs)

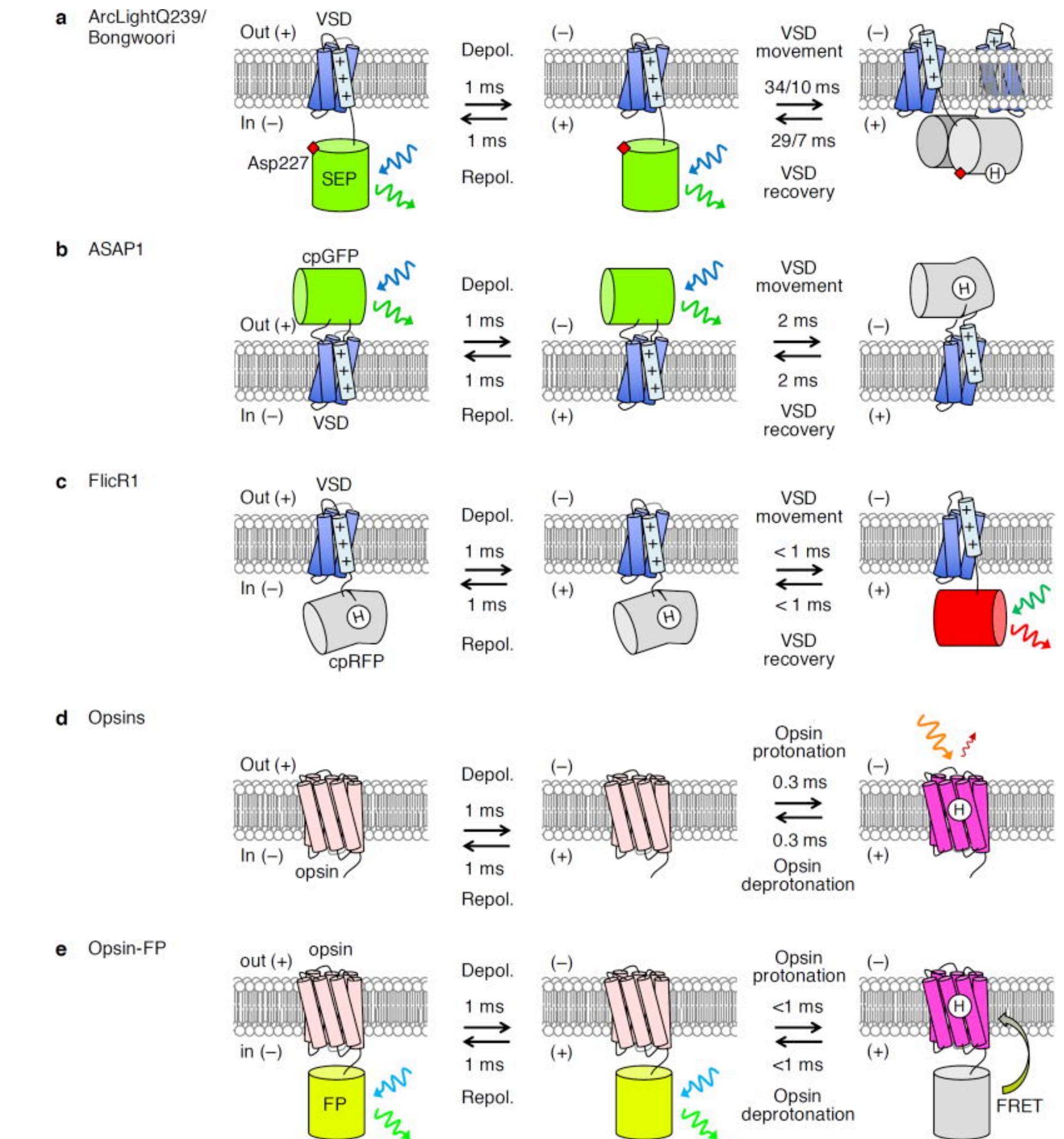
- When neurons spike, voltage gated calcium channels (VGCCs) open and allow a rapid **influx of calcium ions (Ca^{2+})**.
- Genetically encoded calcium indicators (GECIs) like **GCaMP** bind to these calcium ions and become fluorescent.
- The increased fluorescence decays as the calcium unbinds, producing a transient fluorescence indicative of neural spiking.
- Using driver lines, **GECIs can be targeted to specific cell types**.
- In some cases, **multiple GECIs** with different fluorescence wavelengths can be encoded simultaneously in **different subpopulations**.



Lin, Michael Z., and Mark J. Schnitzer. 2016. "Genetically Encoded Indicators of Neuronal Activity." *Nature Neuroscience* 19 (9): 1142–53.

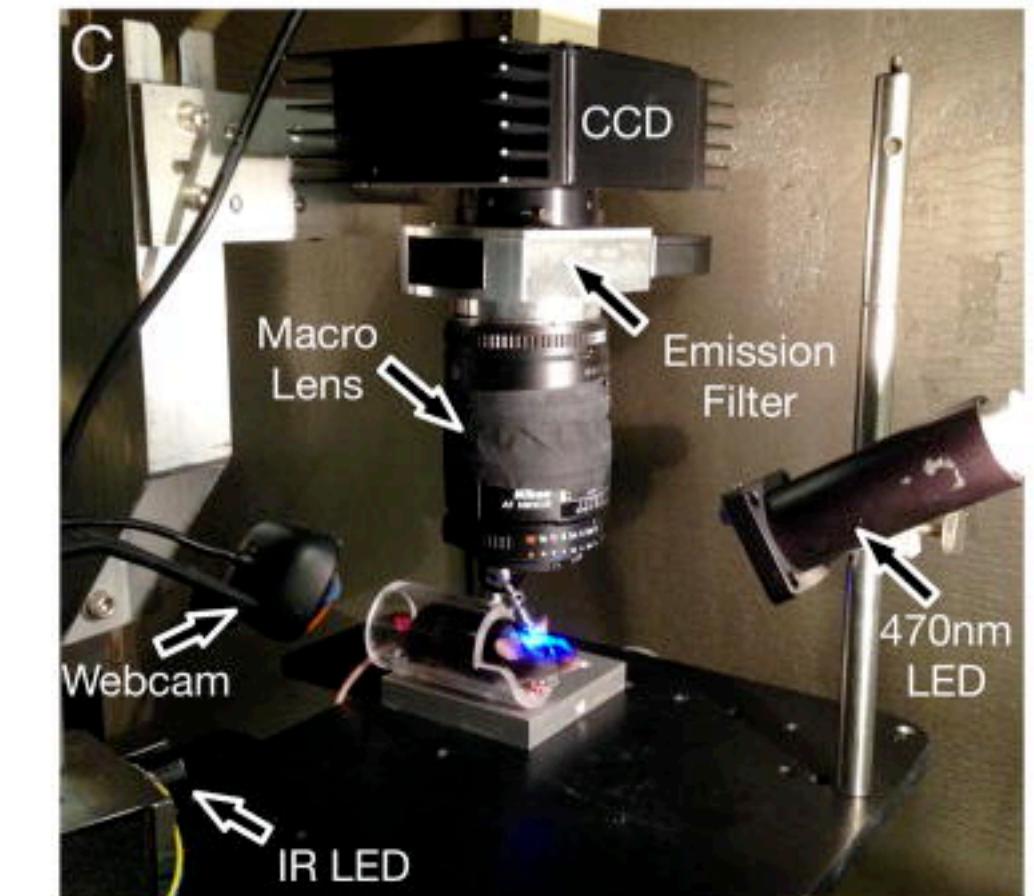
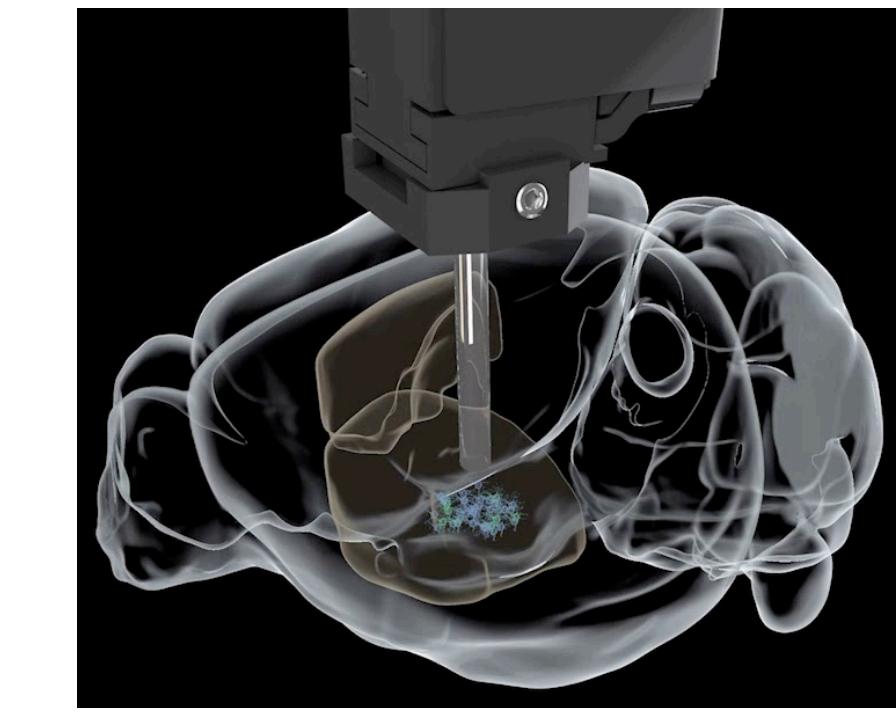
Genetically encoded voltage indicators (GEVIs)

- Calcium is an indirect measure of spiking. Genetically encoded **voltage indicators** modulate fluorescence as a function of membrane potential.
- **Lots of designs:** fusing voltage sensing domains (e.g. from voltage-gated ion channels) to fluorescent proteins; harnessing natural opsins from microbes or algae.
- GECIs are much more established, but great progress in GEVIs has been made in recent years.

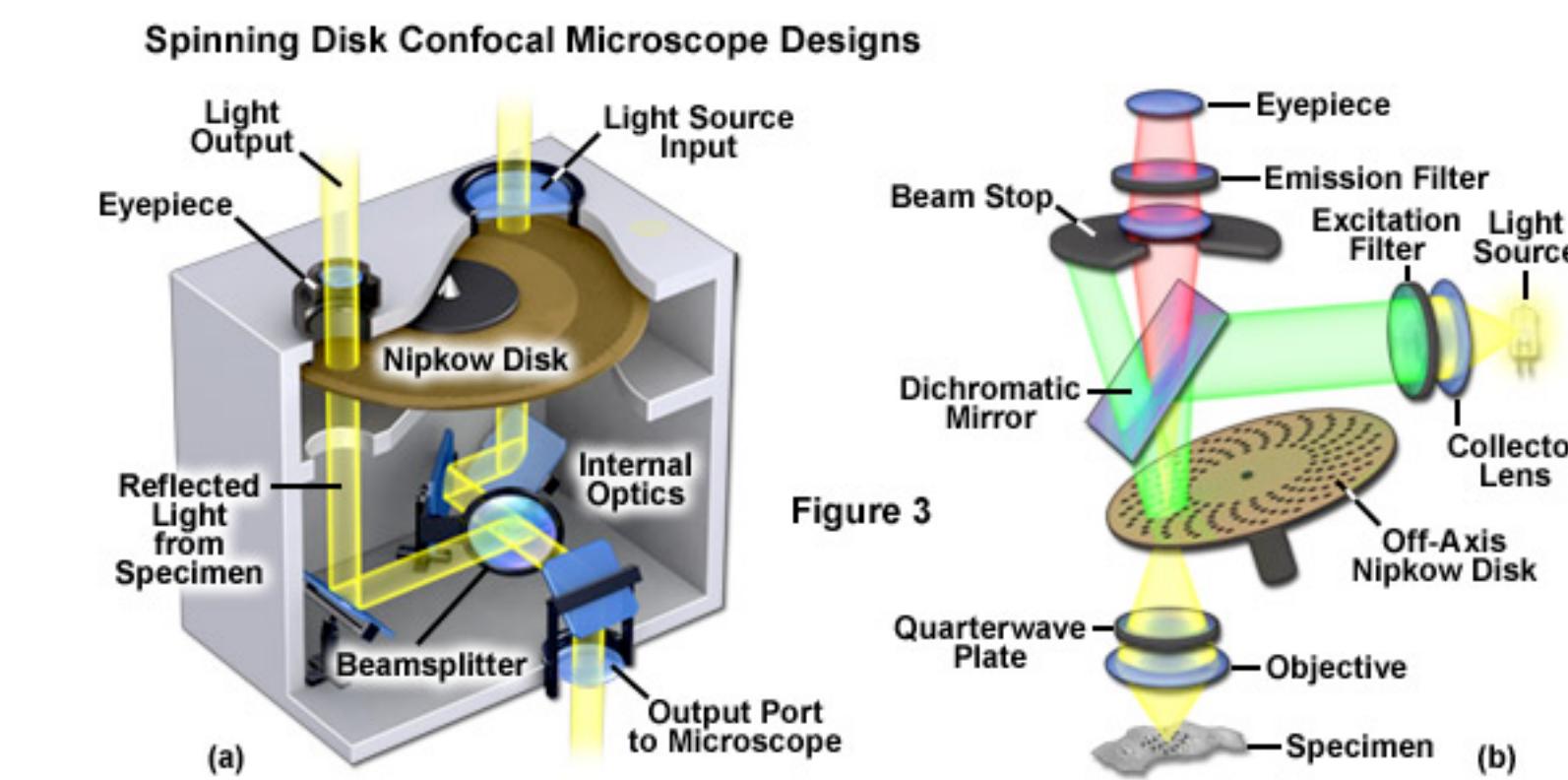


Microscopy

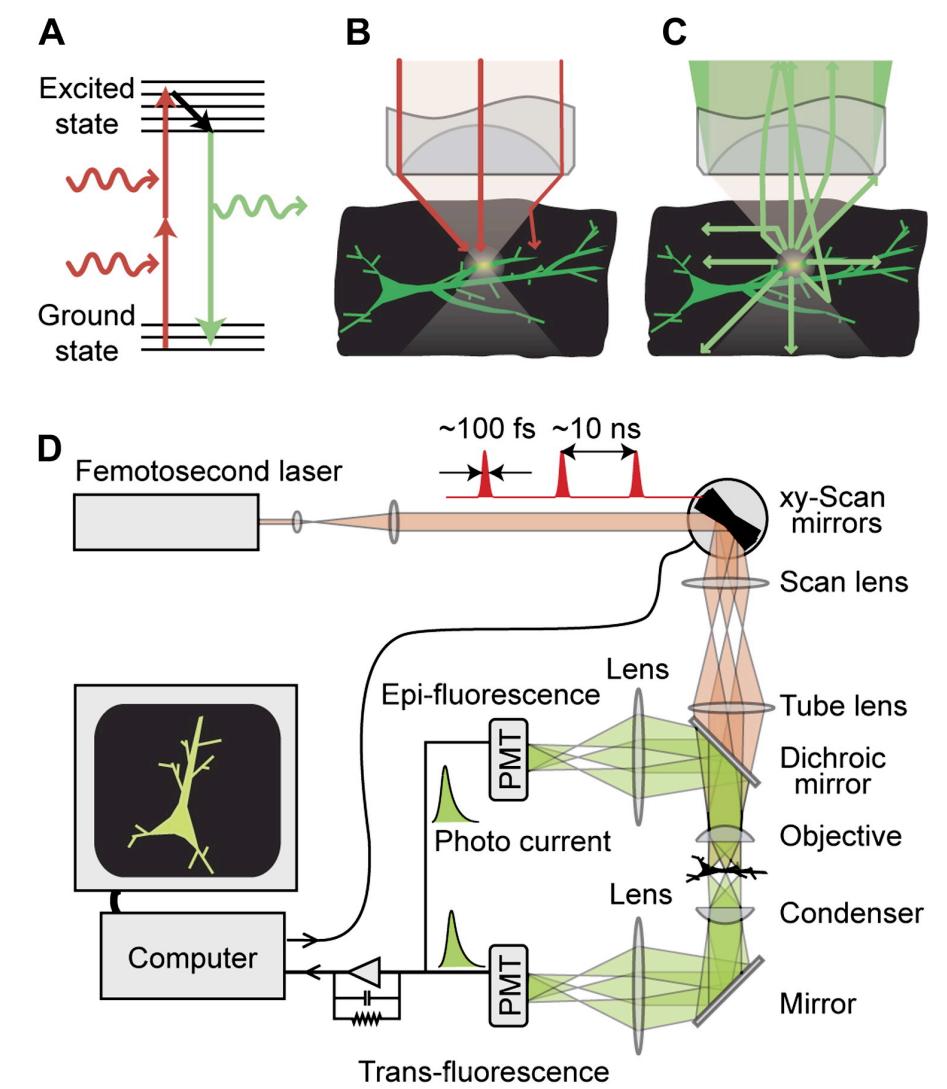
- Expressing the genetically encoded indicator is only half the battle.
- You still need to stimulate the cells with a light source and measure the resulting fluorescence.
- Again, there are lots of approaches: wide-field imaging, **2-photon microscopy**, laser scanning and spinning disk confocal microscopy, miniaturized GRIN lenses, (tapered) fiber photometry.



Silasi et al. J Neurosci Methods. 2016

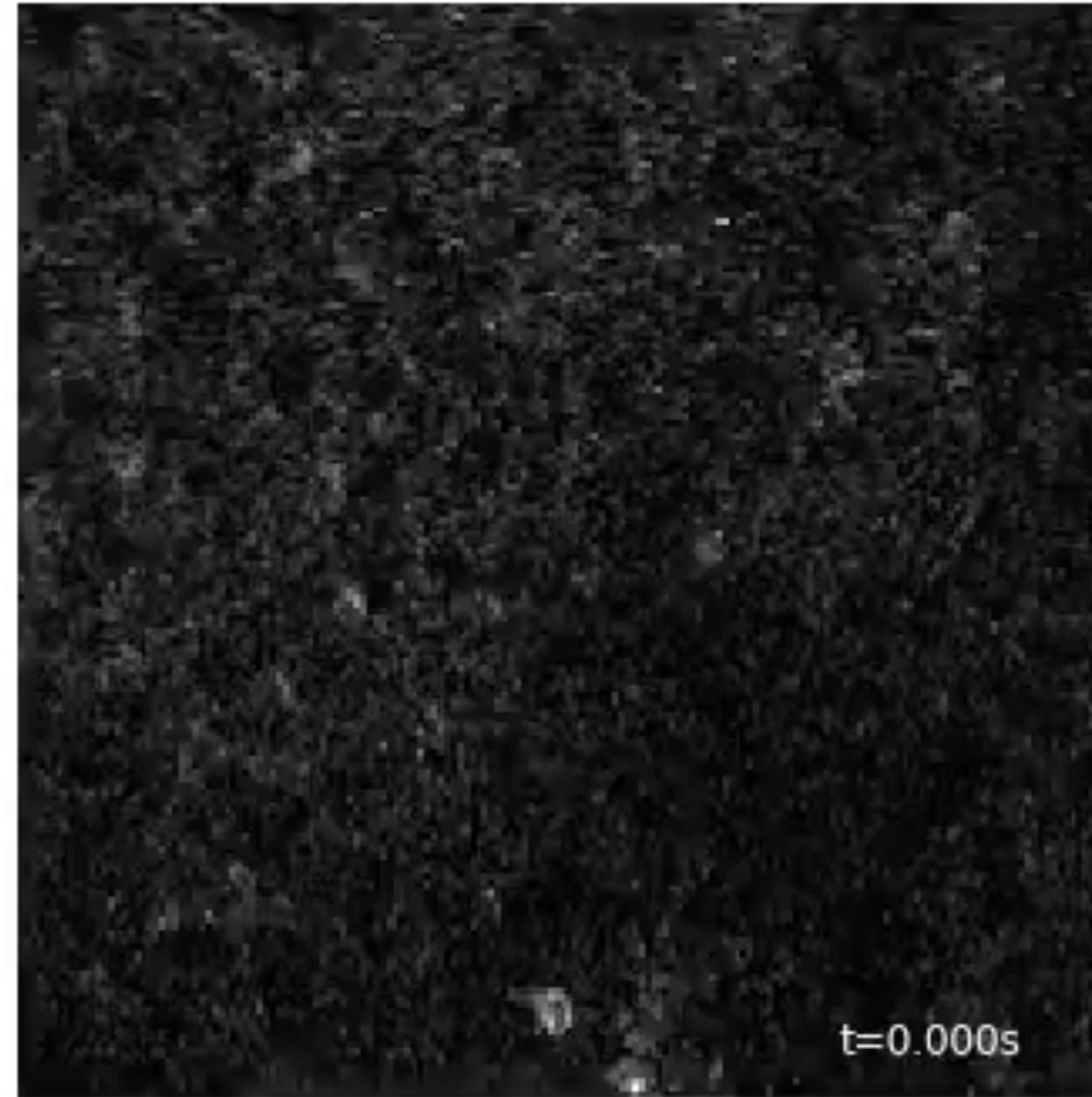


<http://zeiss-campus.magnet.fsu.edu/articles/spinningdisk/introduction.html>



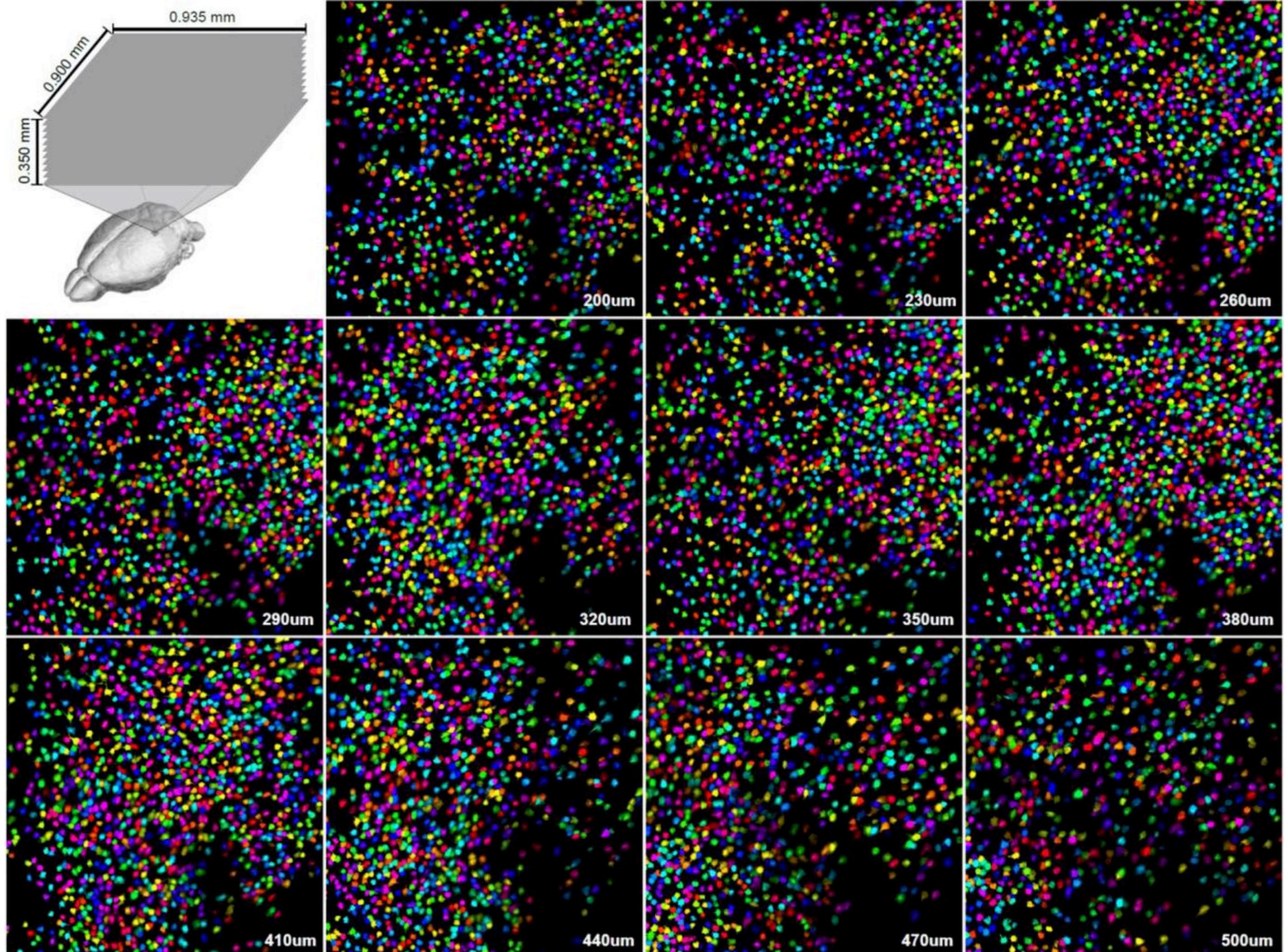
Svoboda and Yasuda. Neuron, 2006.

2 photon calcium imaging



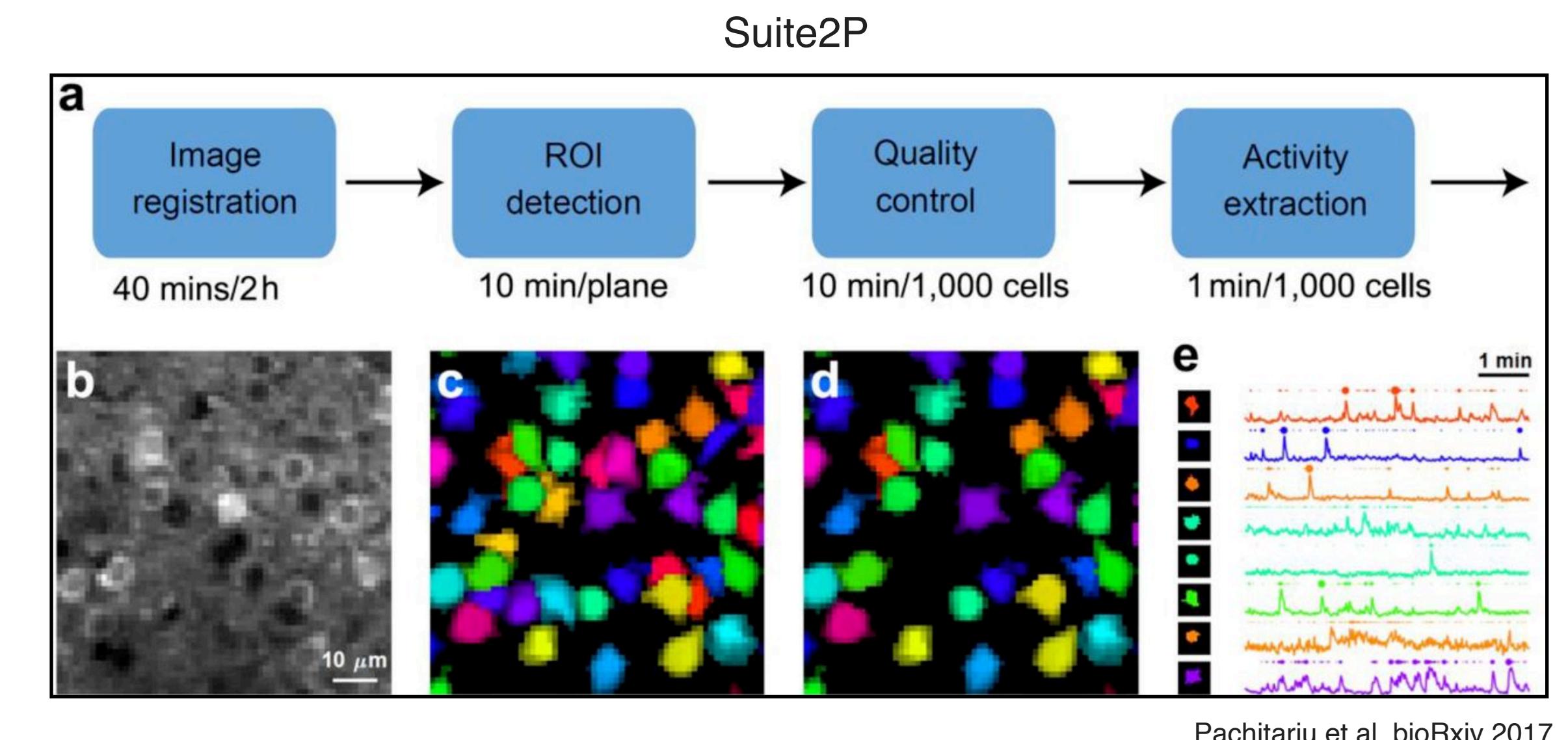
2 photon calcium imaging

Over 10,000 cells

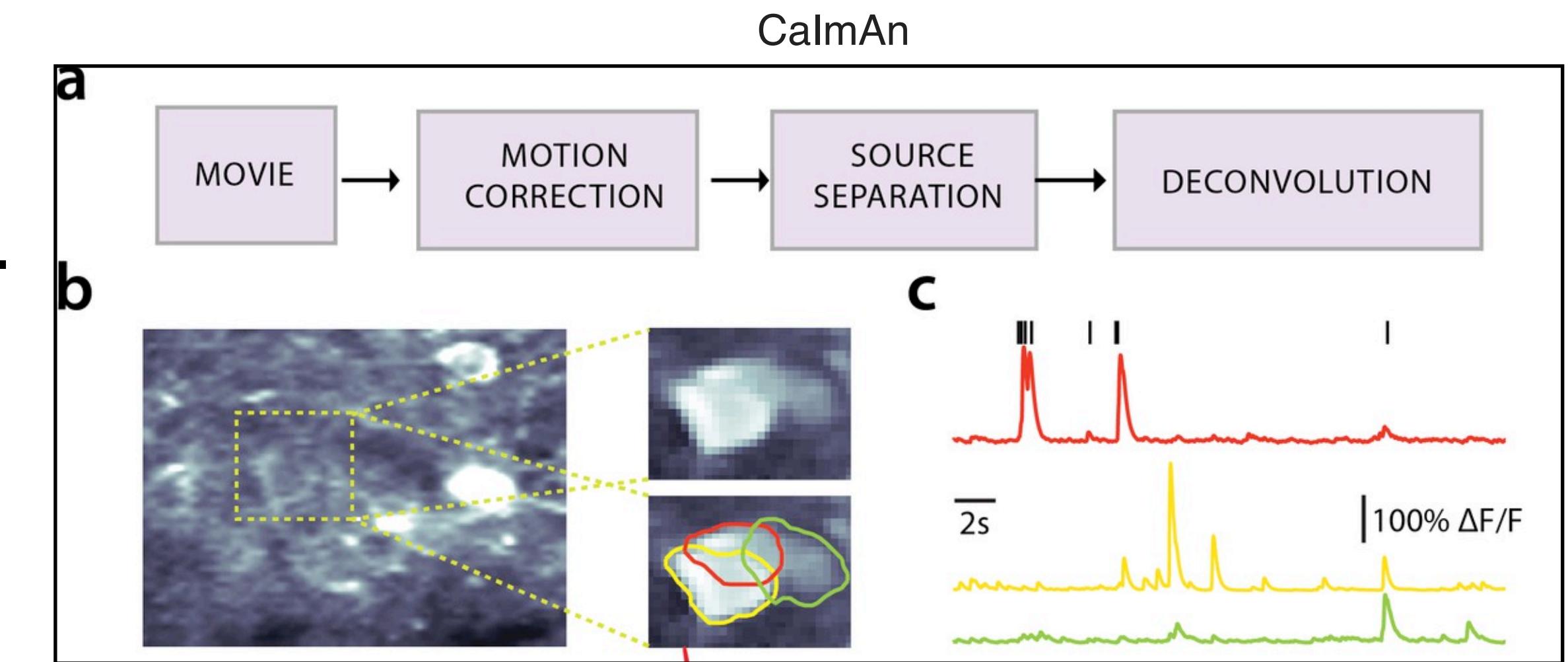


Data analysis pipelines for 2P imaging

- Modern packages like Suite2P and CalmAn go through a few key steps to extract fluorescence traces.
- The key challenges are:
 - Correcting for motion artifacts.
 - Separating overlapping cells.
 - Accounting for background fluorescence.
 - Deconvolving spikes from fluorescence traces.



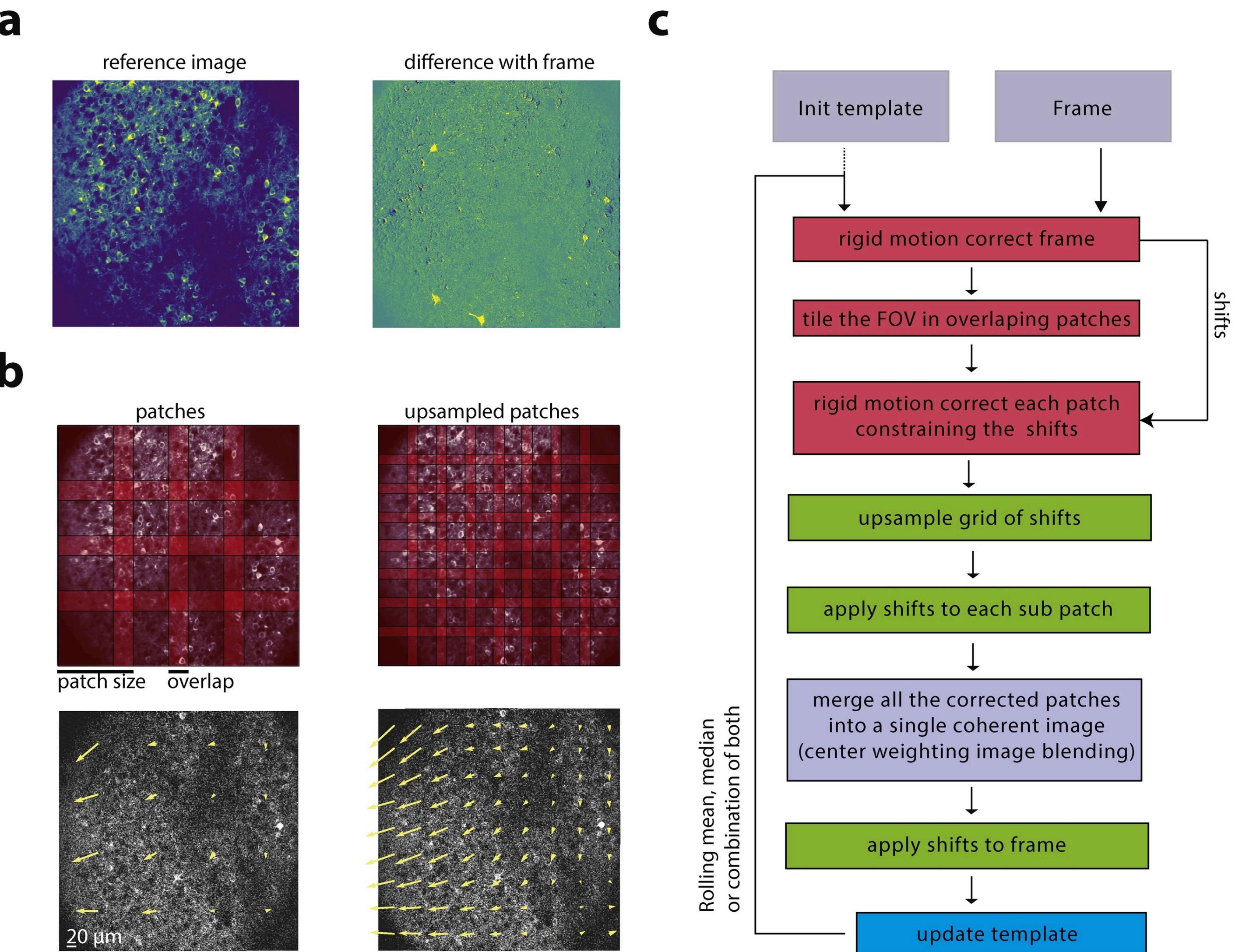
Pachitariu et al, bioRxiv 2017



Giovanucci et al, eLife 2017

Motion correction

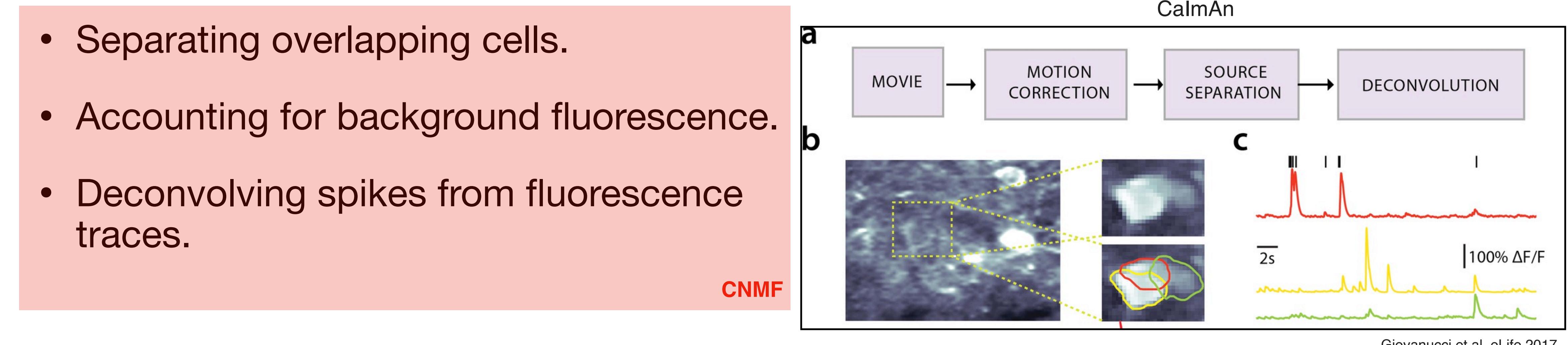
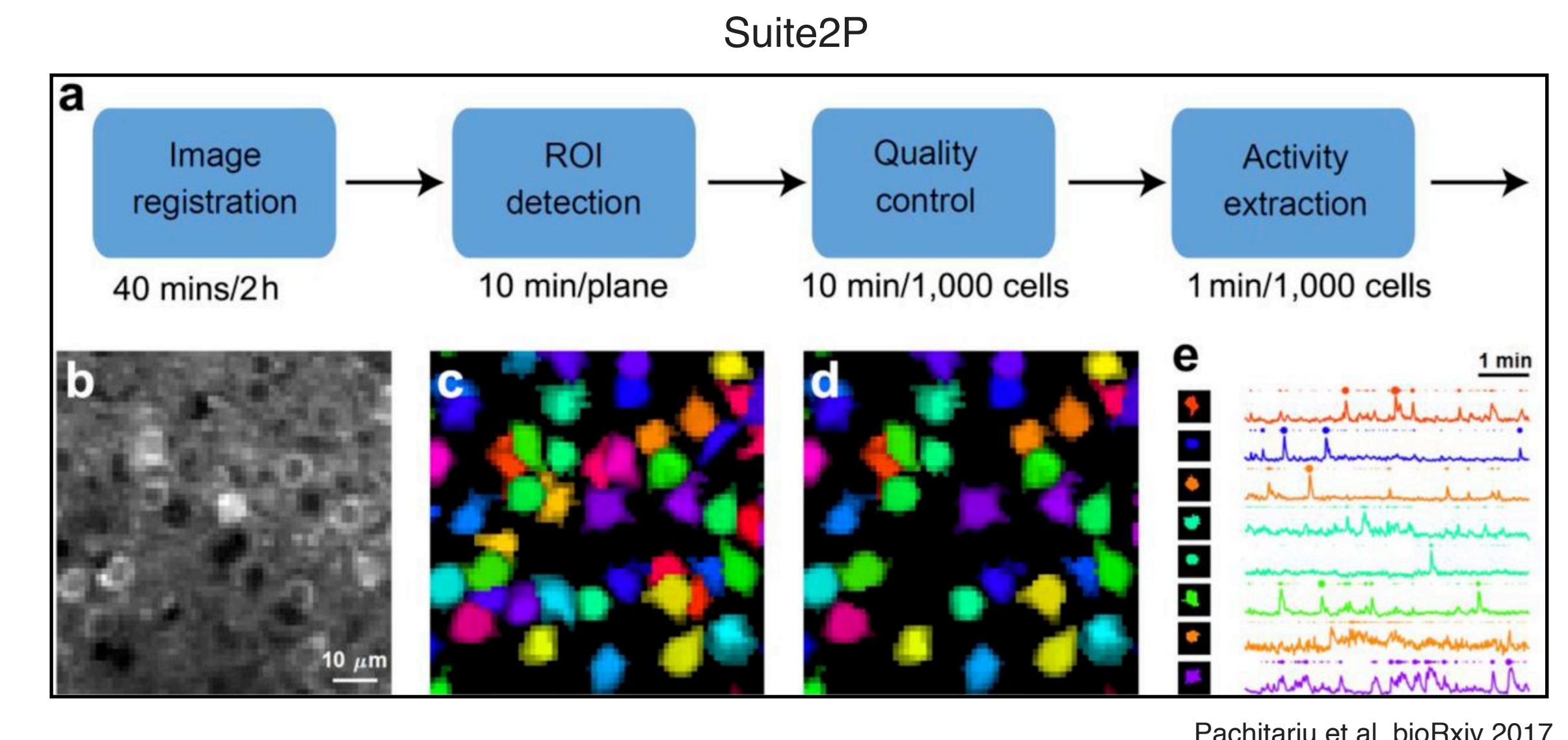
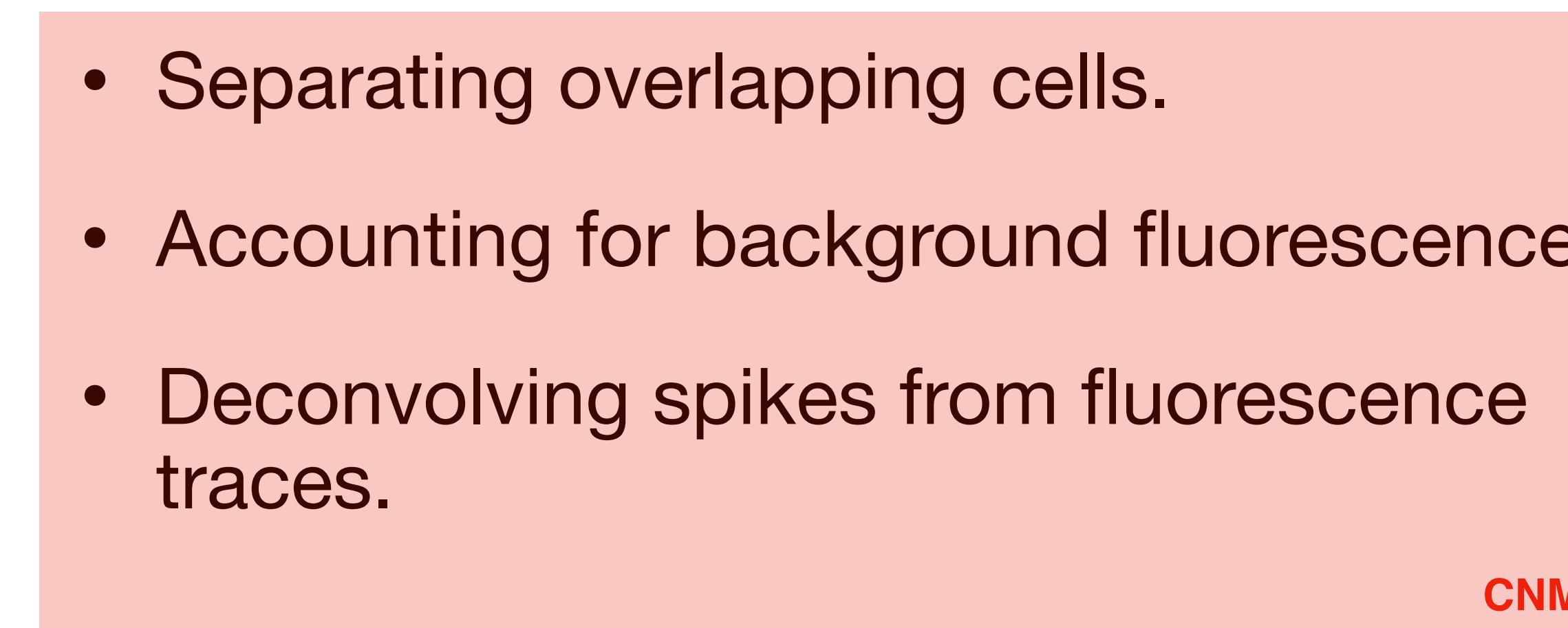
- The brain is squishy and it moves in non-rigid ways in 3D during experiments.
- A variety of non-rigid motion correction algorithms have been proposed:
 - NoRMCorre (Pnevmatikakis and Giovannucci, 2017), used in CalmAn.
 - Phase correlation + kriging (Pachitariu et al, 2017) used in Suite2P.



Pnevmatikakis and Giovannucci, 2017.

Data analysis pipelines for 2P imaging

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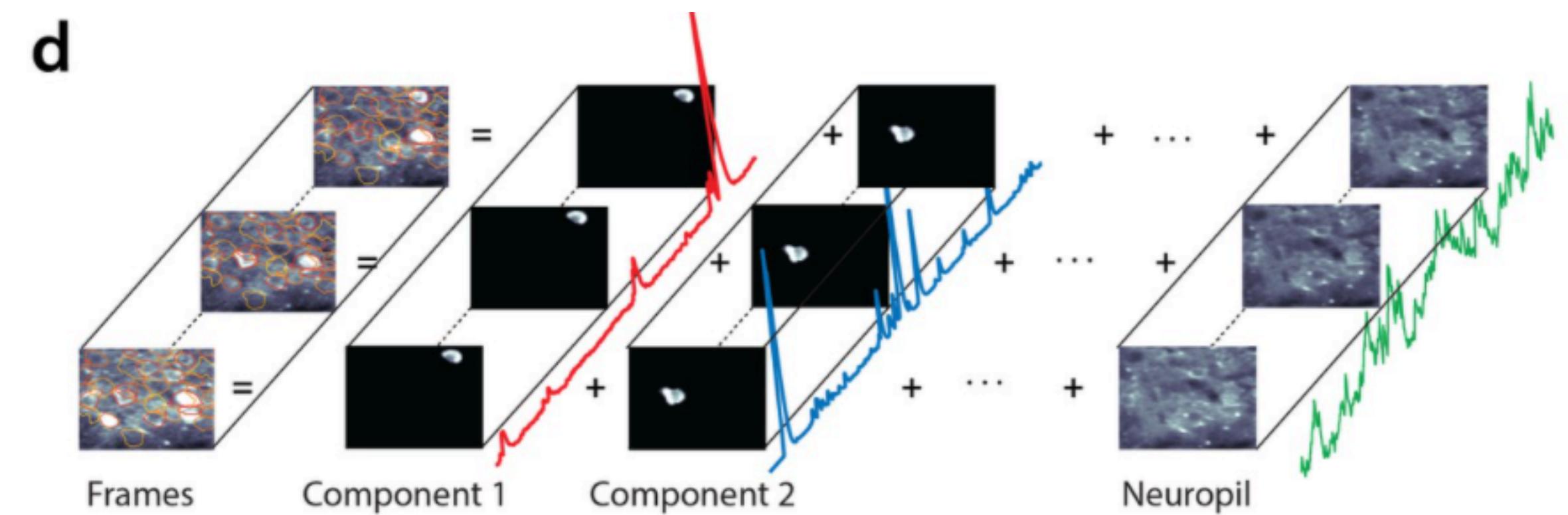


Constrained Non-negative Matrix Factorization (CNMF)

Pnevmatikakis et al, Neuron 2016.

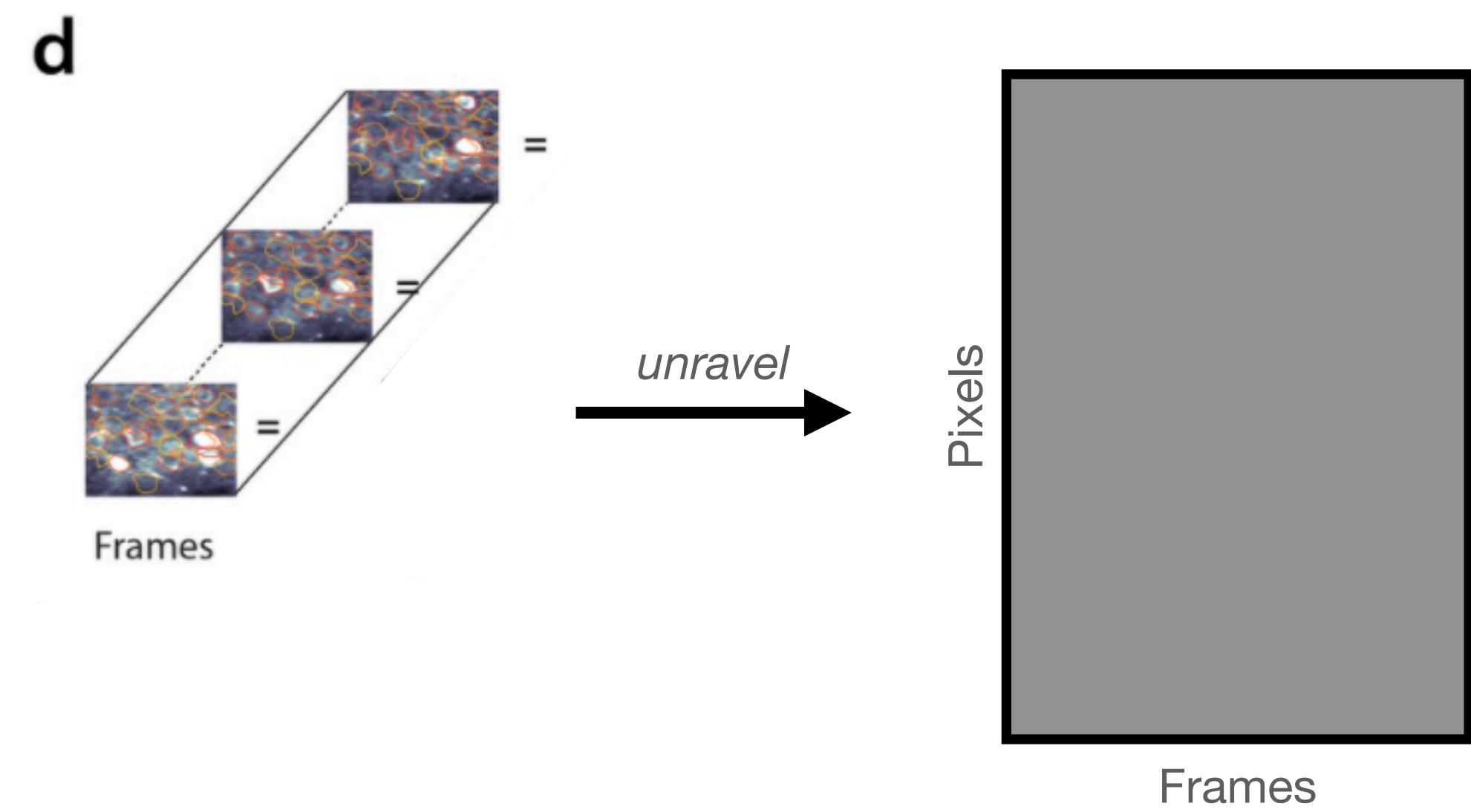
CNMF

- Model the motion corrected movie as a superposition of fluorescence traces from multiple neurons, plus background.
- We can pose this as another convolutional matrix factorization problem.
- **Punchline:** *it's nearly the same as what we did for spike sorting!*



Constants

- Let T denote the number of **frames** in the movie.
- P denote the number of **pixels**.
- D denote the **duration** (in frames) of a calcium spike.
- N denote the (unknown) number of **neurons** that generated the spikes.



Data and Latent Variables

- **Data:**

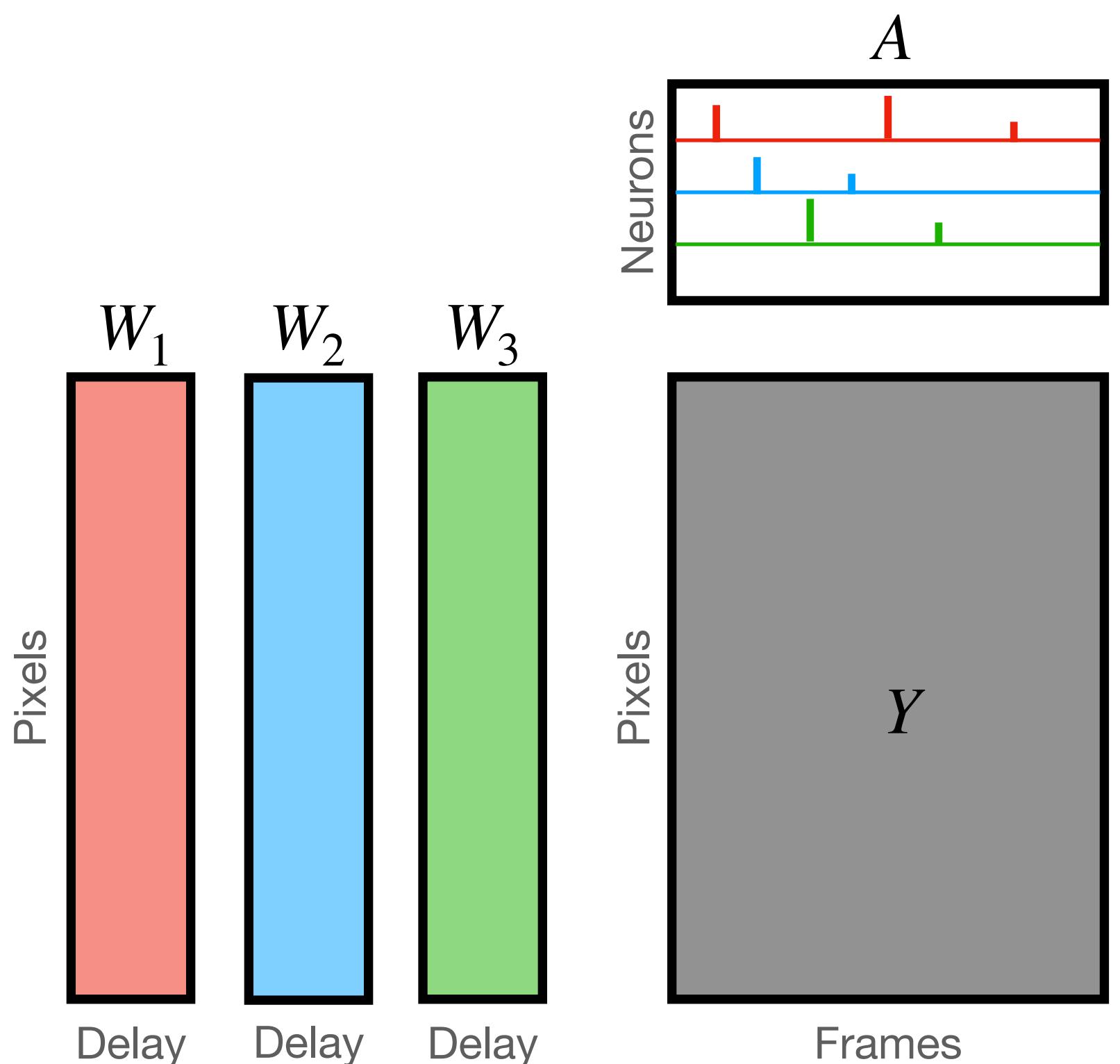
- Let $Y \in \mathbb{R}^{P \times T}$ denote the motion corrected and unraveled video.

- **Latent Variables:**

- Let $A \in \mathbb{R}_+^{N \times T}$ denote the time series of spike amplitudes for each neuron.

- **Parameters:**

- Let $W \in \mathbb{R}^{N \times P \times D}$ denote the array of calcium responses for each neuron.

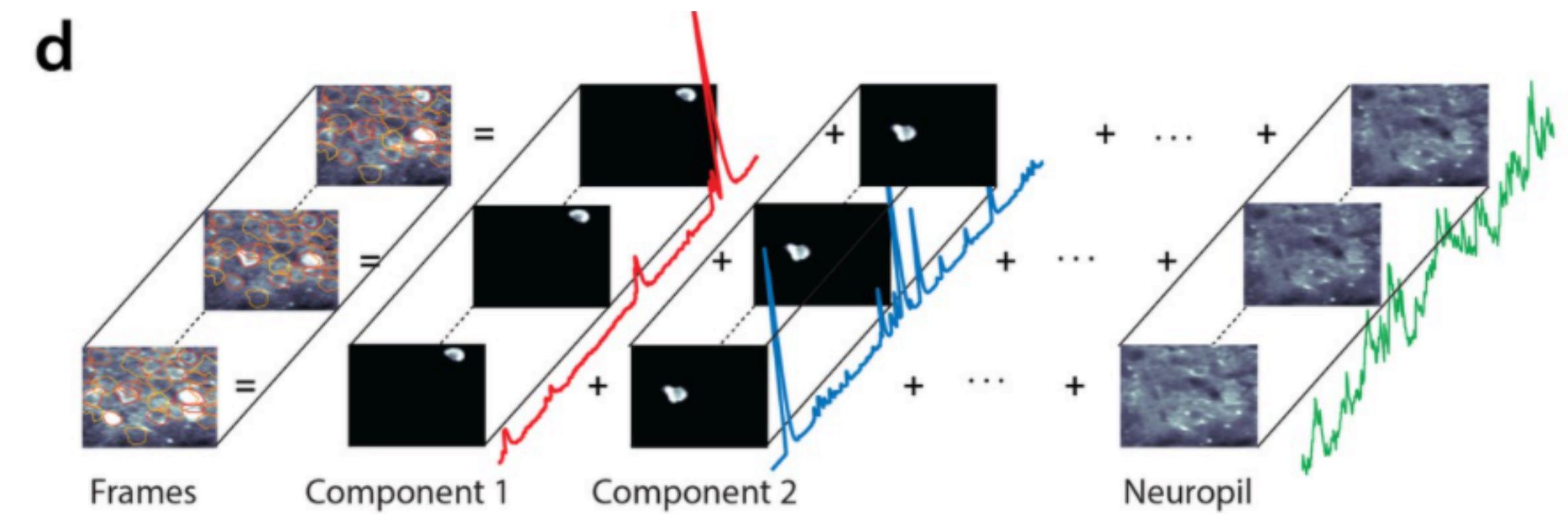


Probabilistic Model

Likelihood

Like last week, assume each spike induces a scaled calcium response in the video.

$$p(Y | A, W, u_0, c_0, \sigma^2) = \prod_{t=1}^T \mathcal{N}\left(y_t | \sum_{n=1}^N [a_n \circledast W_n]_t + u_0 c_{0t}, \sigma^2 I\right)$$



Calcium response model

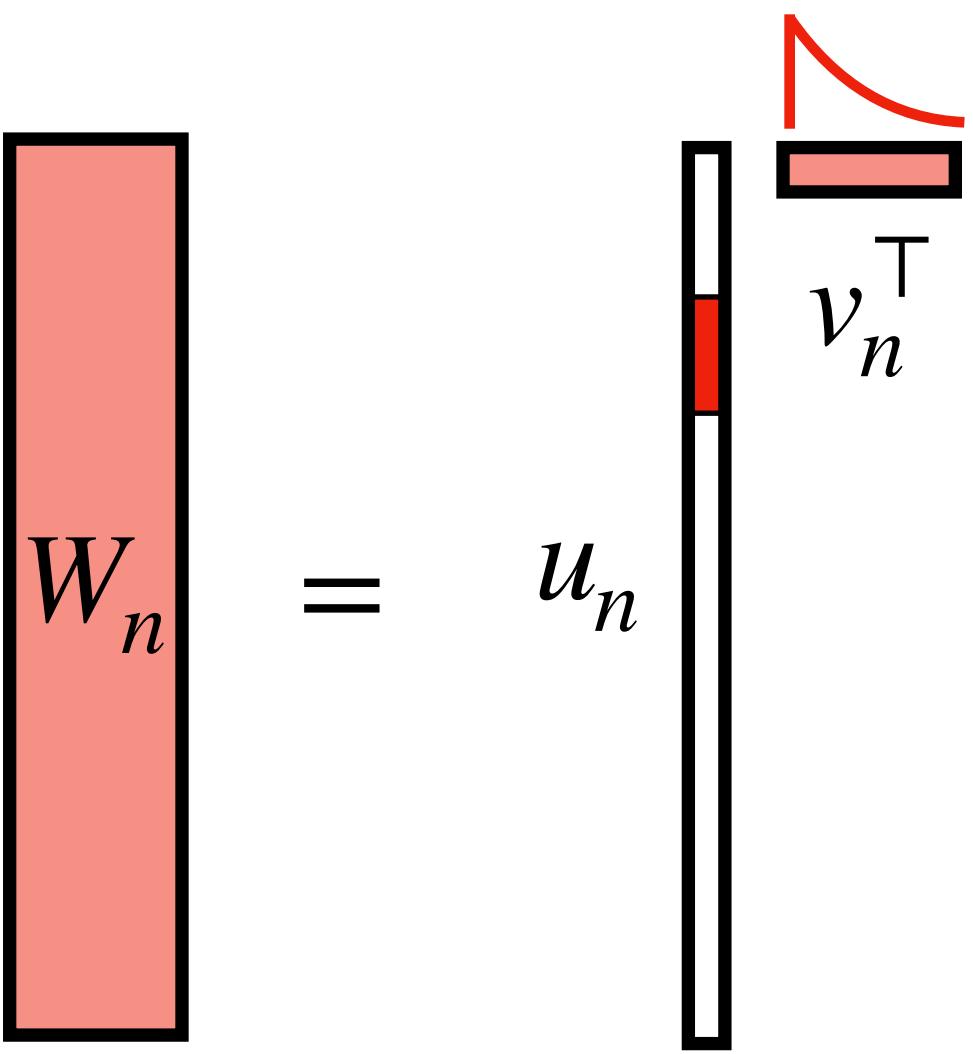
- Assume the calcium responses factor into spatial and temporal components.

$$W_n = u_n v_n^\top$$

- Spatial factor u_n specifies which pixels correspond to neuron n .
- Constrain the temporal components to be exponential decays.

$$v_{nd} = e^{-d/\tau}$$

- Time constant of the decay is a function of the indicator; O(100ms).



Calcium response model

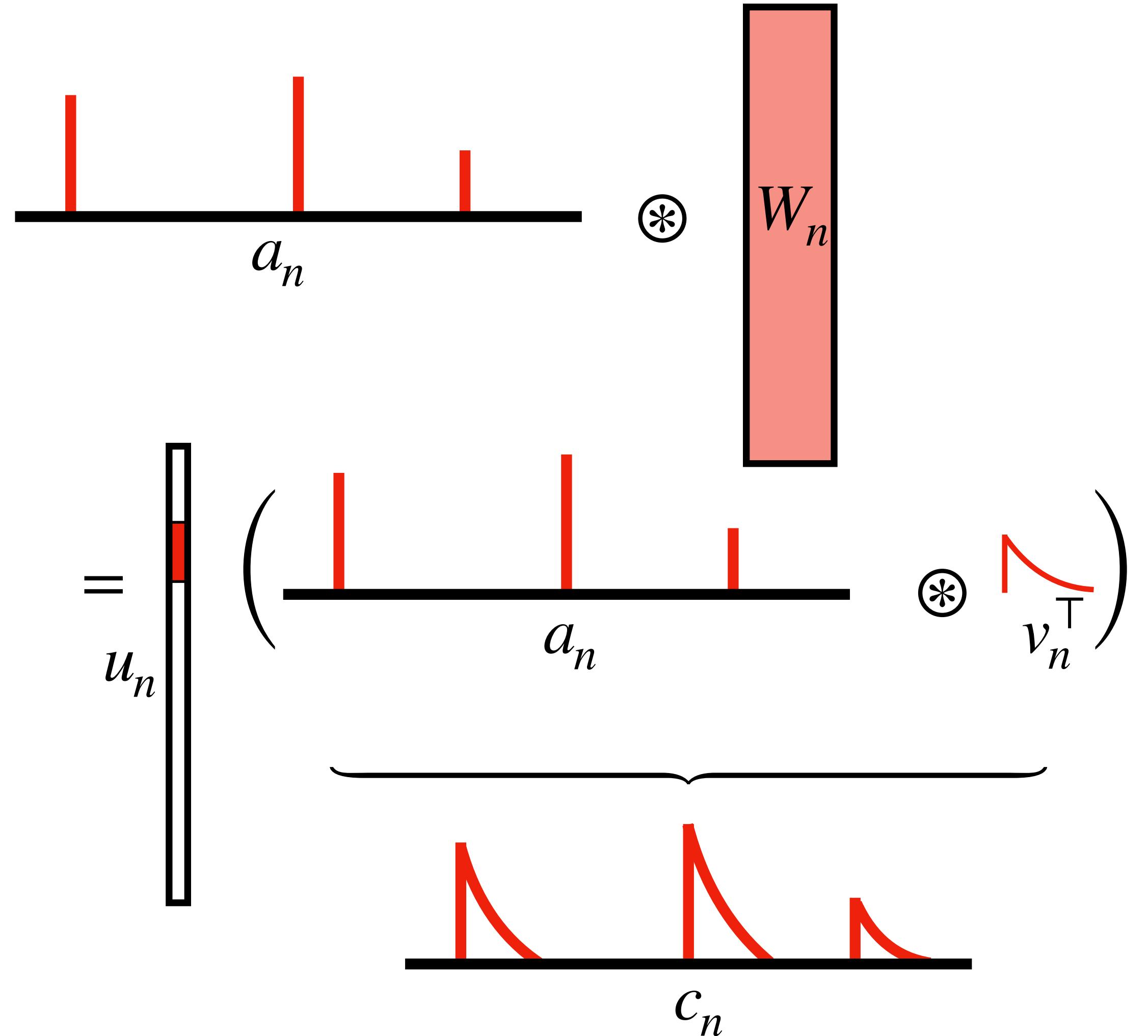
Then

$$[a_n \circledast W_n]_t = u_n [a_n \circledast v_n]_t \triangleq u_n c_{nt},$$

where

$$c_{nt} \triangleq [a_n \circledast v_n]_t = \sum_{d=0}^D a_{n,t-d} v_{nd} = \sum_{d=0}^D a_{n,t-d} e^{-d/\gamma}$$

is the calcium trace of neuron n



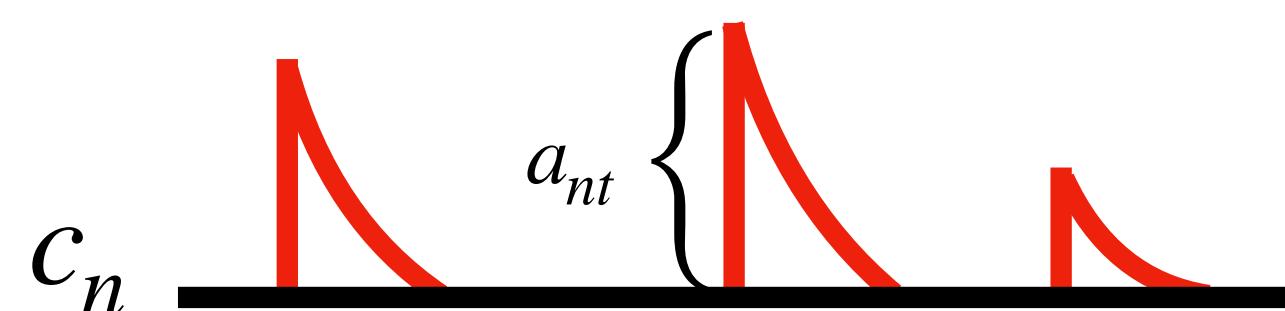
Recursive formulation

The calcium response can be written recursively, thanks to the **exponential response**:

$$\begin{aligned} c_{nt} &= \sum_{d=0}^D a_{n,t-d} e^{-d/\gamma} \\ &= a_{nt} + \sum_{d=1}^{t-1} a_{n,t-d} e^{-d/\gamma} \\ &= a_{nt} + \sum_{d=0}^{t-2} a_{n,t-(d+1)} e^{-(d+1)/\gamma} \\ &= a_{nt} + e^{-1/\gamma} c_{n,t-1}, \end{aligned}$$

Equivalently, $a_{nt} = c_{nt} - e^{-1/\gamma} c_{n,t-1}$.

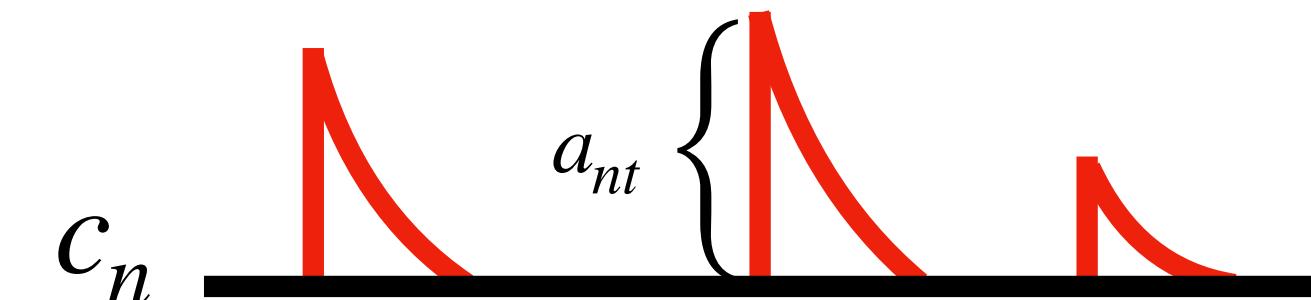
(Note, we took $D \rightarrow \infty$ and zero-padded a_n on the left.)



Recursive formulation

In matrix form,

$$a_n = G c_n \quad G = \begin{bmatrix} 1 & & & \\ e^{-1/\gamma} & 1 & & \\ & e^{-1/\gamma} & 1 & \\ & & \ddots & \ddots \end{bmatrix}.$$



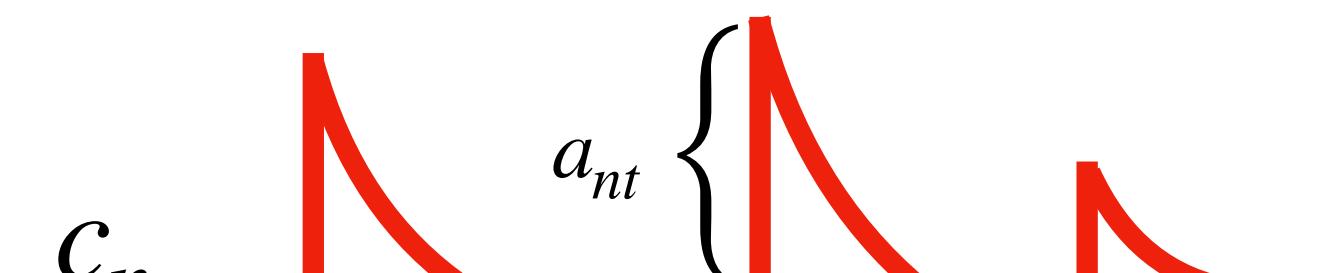
Prior on calcium traces

Via a prior on amplitudes

Suppose $a_{nt} \sim \text{Exp}(\lambda_n)$, as in the spike sorting model.

We can derive the probability of c_n using the change of measure formula,

$$\begin{aligned} p(c_n) &= \left| \frac{da_n}{dc_n} \right| \prod_{t=1}^T \text{Exp}(c_{nt} - e^{-1/\gamma} c_{n,t-1}; \lambda_n) \\ &= |G| \prod_{t=1}^T \text{Exp}(c_{nt} - e^{-1/\gamma} c_{n,t-1}; \lambda_n) \end{aligned}$$



Prior on calcium traces

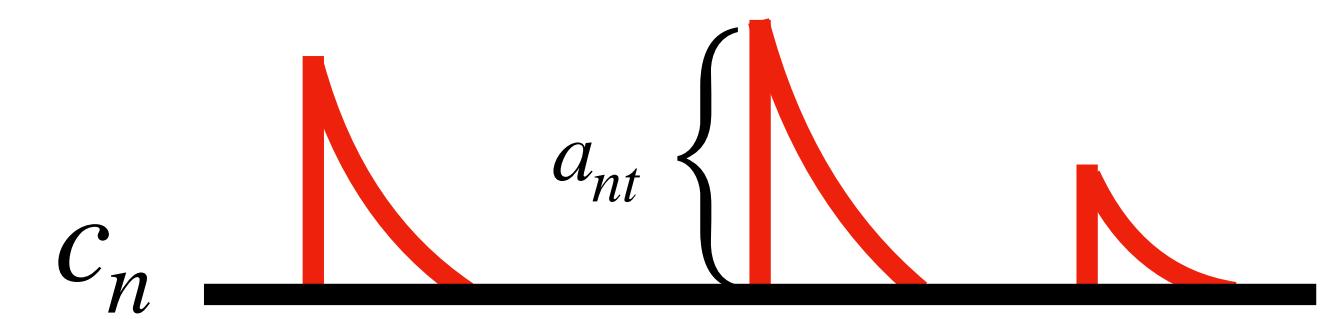
Via a prior on amplitudes

Since G is lower triangular, its determinant is the product of its diagonal; i.e. 1.

Thus,

$$p(c_n) = \prod_{t=1}^T \text{Exp}(c_{nt} - e^{-1/\gamma} c_{n,t-1}; \lambda_n)$$

It's just the probabilities of the “jumps” a_{nt} .



Optimizing the calcium traces

Following the same steps as last week, we end up with the following objective for optimizing the calcium traces:

$$\mathcal{L}(c_n) = -\frac{1}{2\sigma^2} \|c_n - \mu_n\|_2^2 + \lambda_n \sum_{t=1}^T (c_{nt} - e^{-1/\gamma} c_{n,t-1}),$$

where

$$\mu_n = R_n^\top u_n$$

is the residual projected onto the spatial factor for this neuron.

Optimizing the calcium traces

More compactly,

$$\mathcal{L}(c_n) = -\frac{1}{2\sigma^2} \|c_n - \mu_n\|_2^2 + \lambda_n \sum_{t=1}^T (c_{nt} - e^{-1/\gamma} c_{n,t-1})$$

$$= -\frac{1}{2\sigma^2} \|c_n - \mu_n\|_2^2 + \lambda_n \|Gc_n\|_1.$$

We still have that pesky hyperparameter λ_n ...

Optimizing the calcium traces

Dual formulation

- Maximizing $\mathcal{L}(c_n)$ is equivalent to solving the following convex optimization problem,

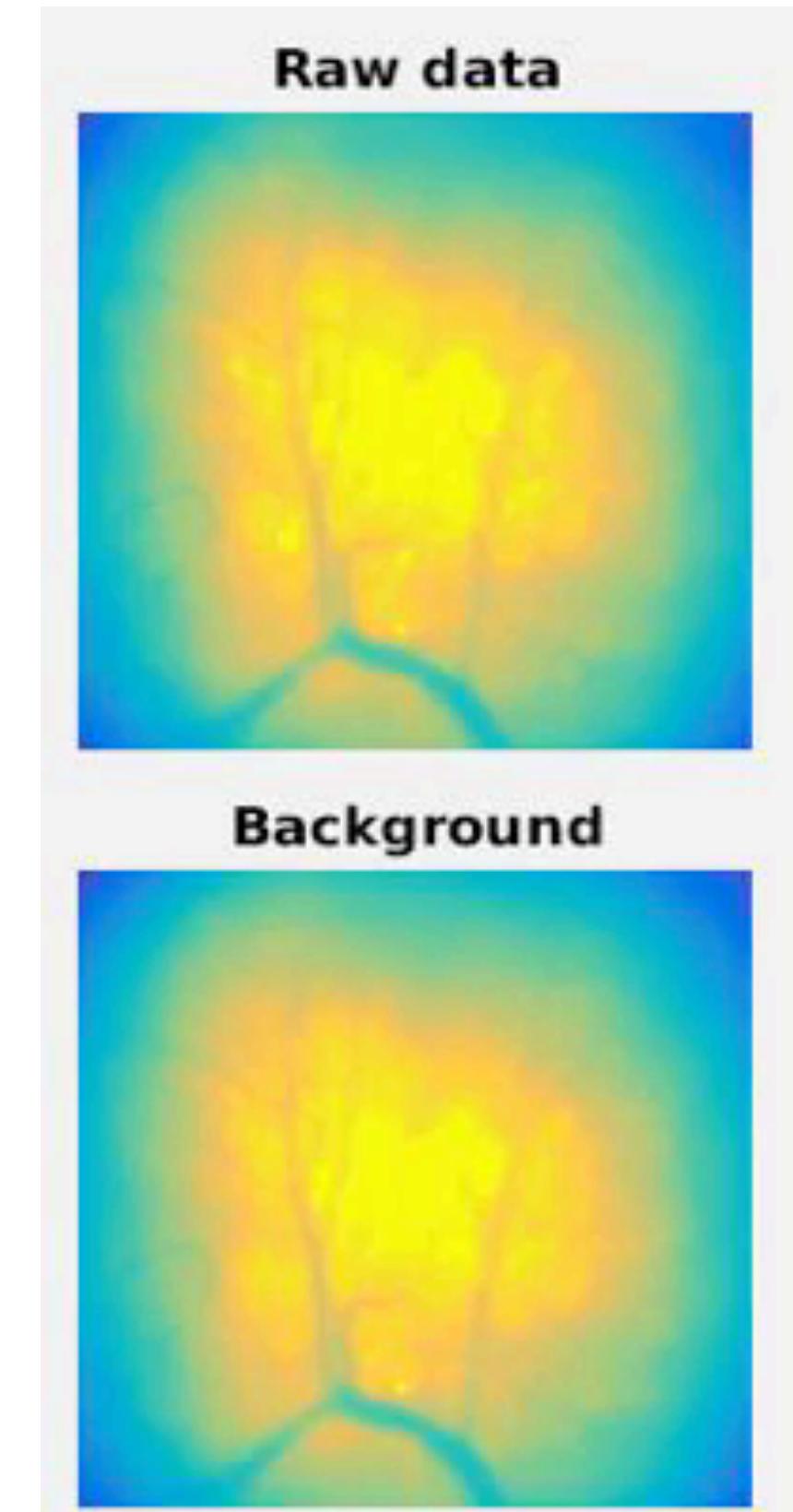
$$\hat{c}_n = \arg \min_{c_n} \|Gc_n\|_1 \quad \text{subject to} \quad \|c_n - \mu_n\|_2 \leq \theta, \quad c_n \geq 0,$$

for some threshold θ .

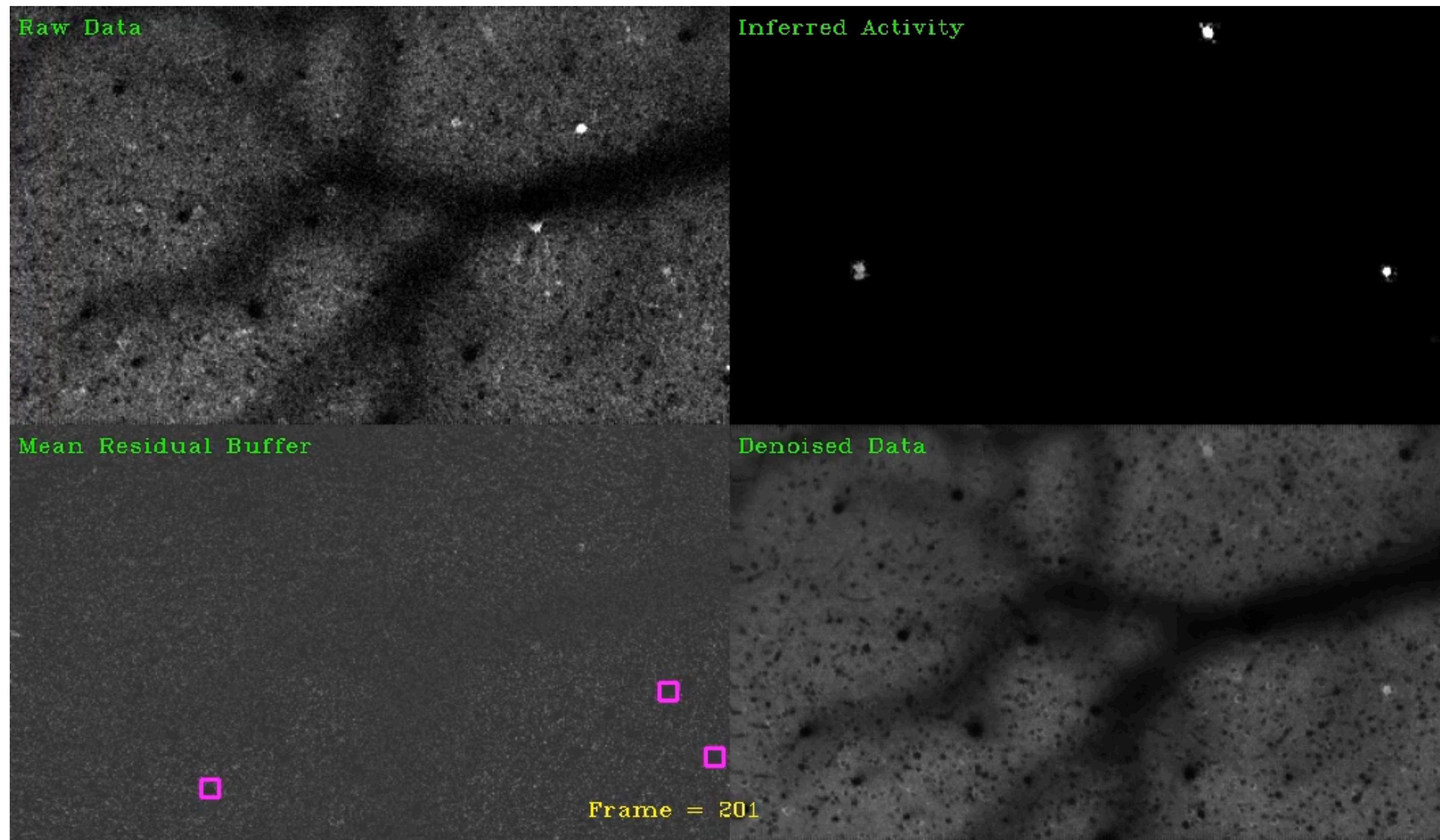
- Under the model, $c_{nt} - \mu_{nt} \sim \mathcal{N}(0, \sigma^2)$.
- By the law of large numbers, $\|c_n - \mu_n\|_2 \rightarrow \sigma\sqrt{T}$.
- **Idea:** set $\theta = (1 + \epsilon)\sigma\sqrt{T}$.
- Since G is a sparse banded matrix, we can solve this in $O(T)$ time with CVX.

Miscellanea

- We typically constrain the **spatial factors to be non-negative** too, unlike in spike sorting.
- We need to account for **background fluorescence** from out-of-focus cells.
- Typically, assume **rank-1** or **spatially smooth** background. See notes.
- As always, **preprocessing is important** for finding candidate neurons and characterizing noise.
More on this in the lab.

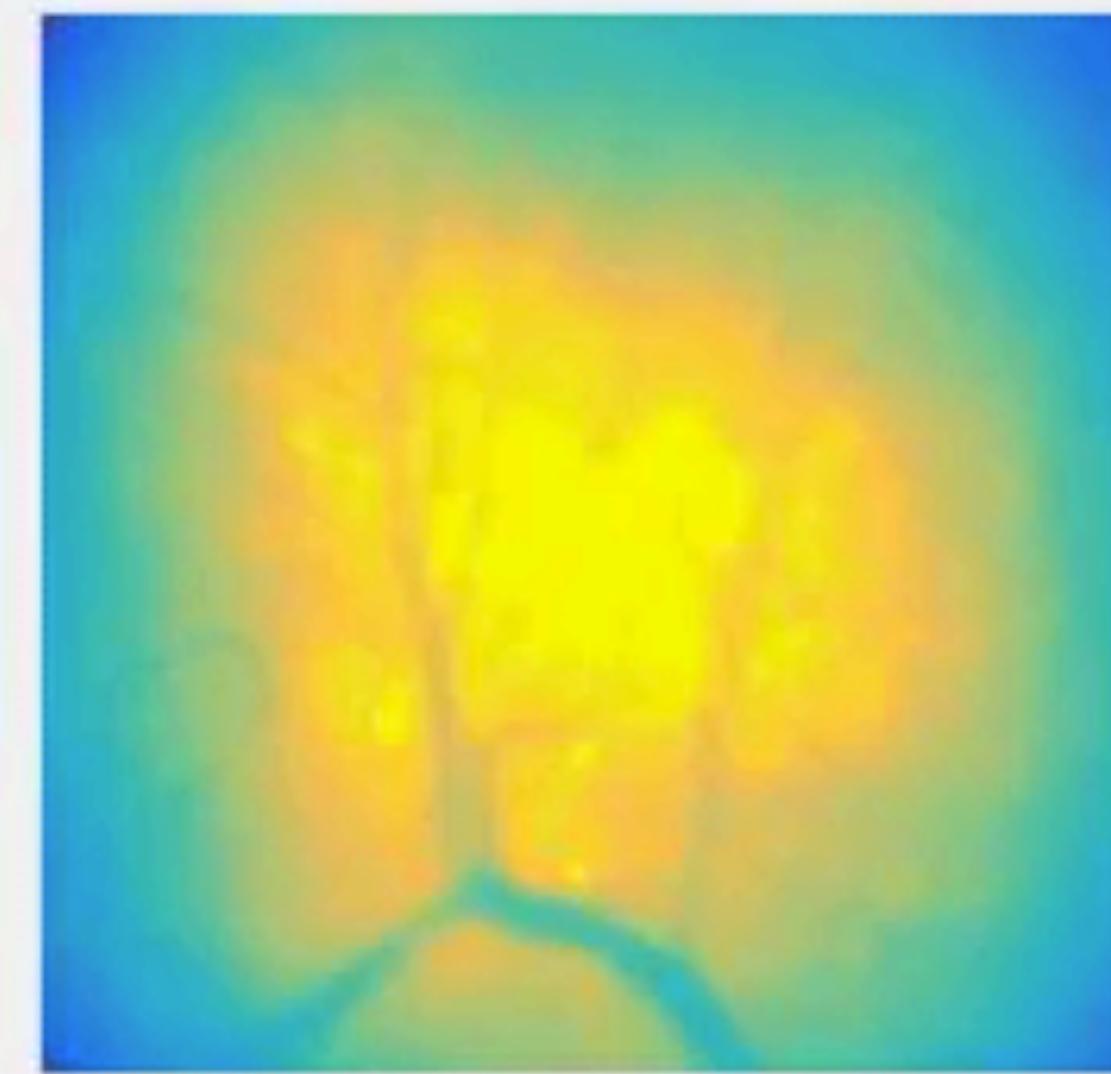


CNMF-E; Zhou et al, eLife 20:

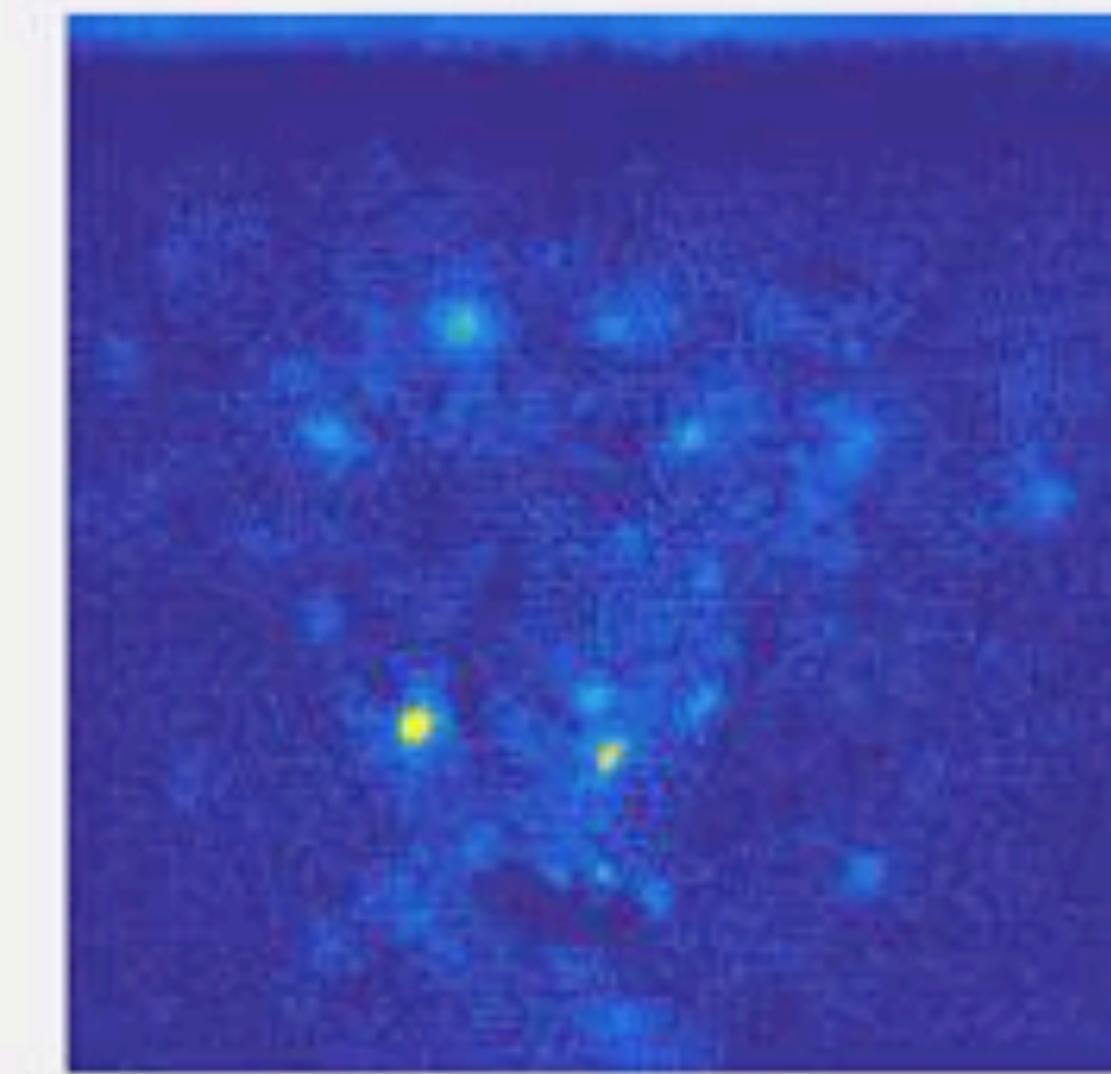


OnACID; Giovanucci et al, NIPS 2017. Mesoscope data from A. Tolias lab

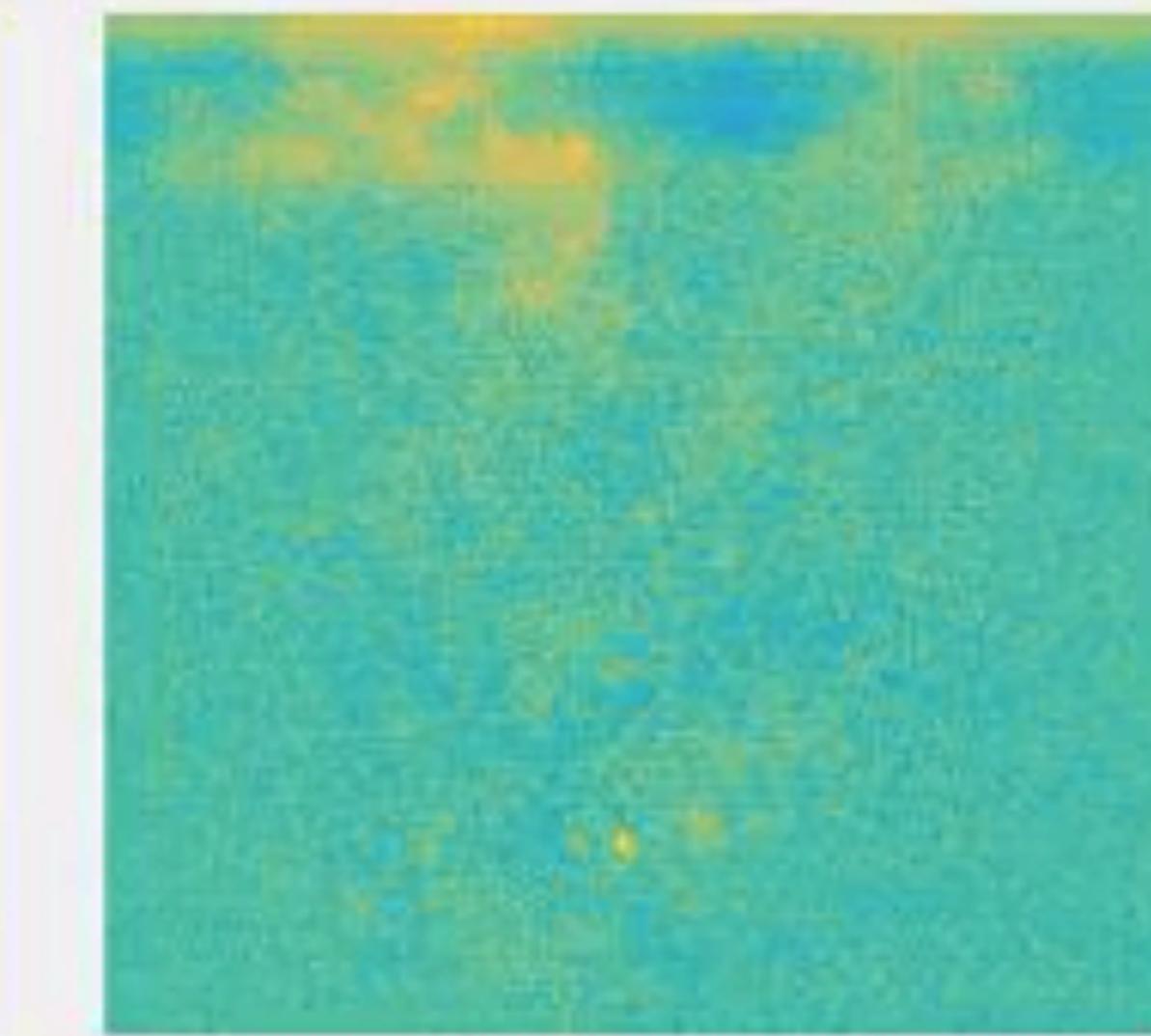
Raw data



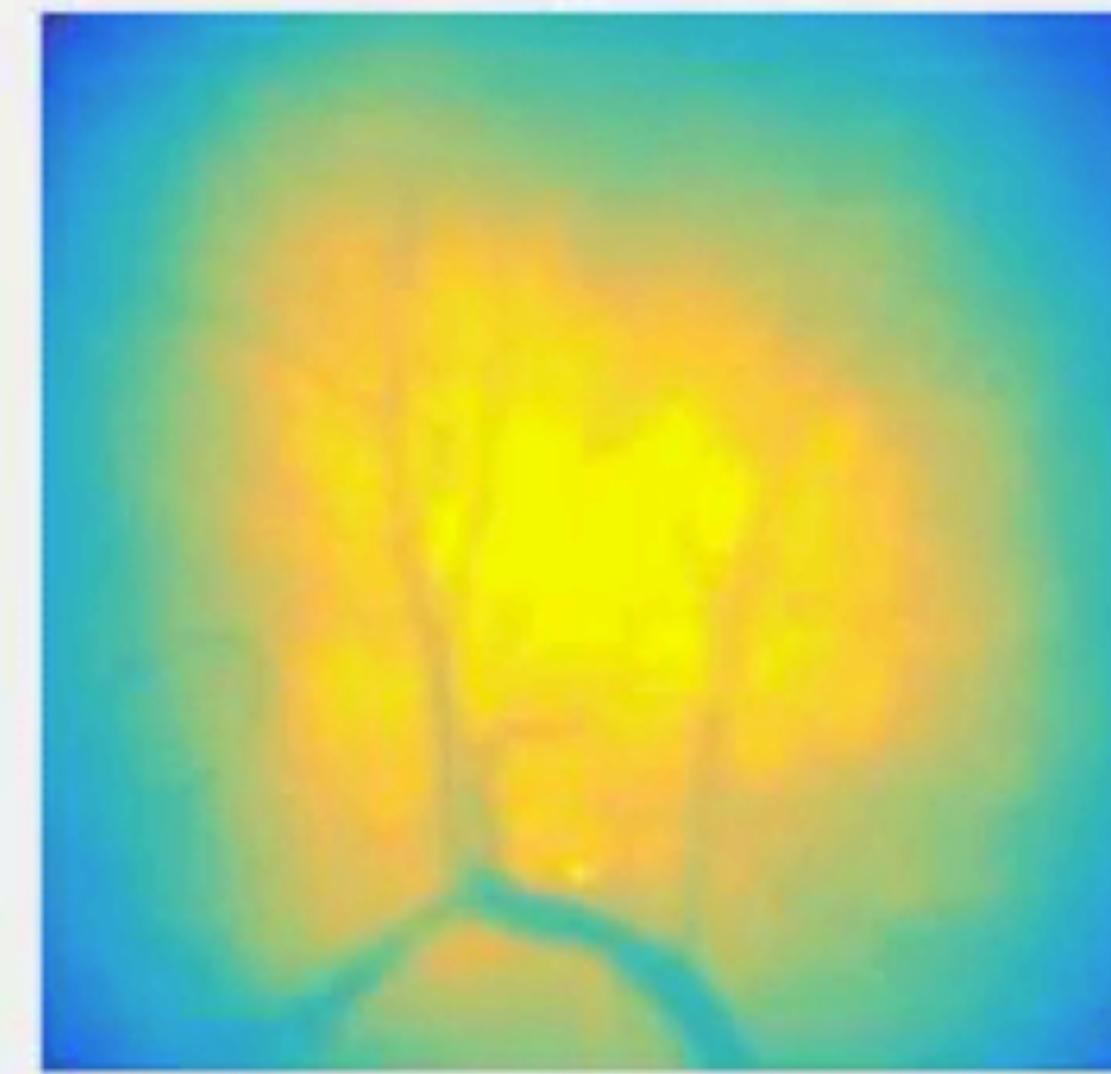
(Raw-BG) X 8



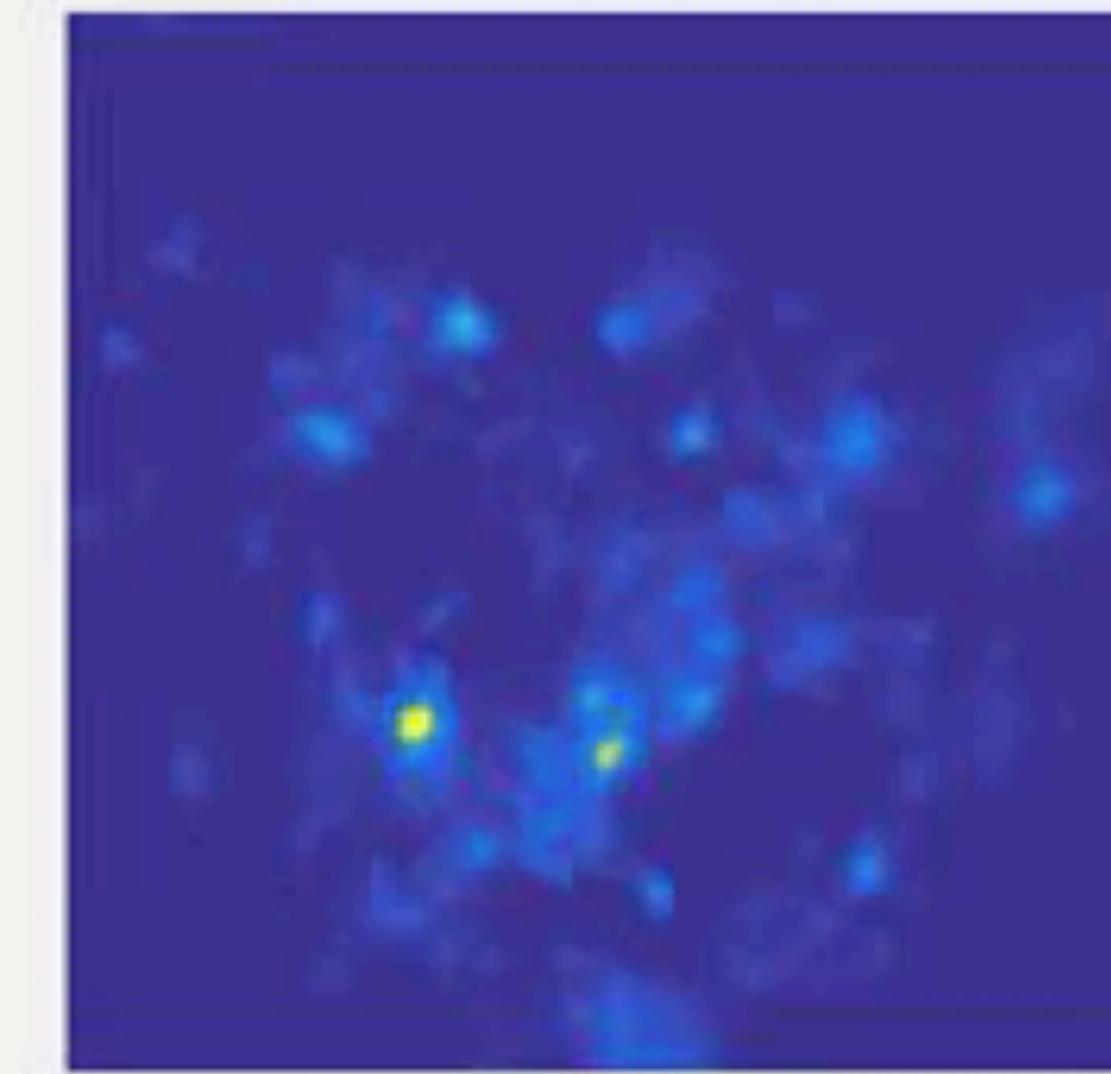
Residual X 8



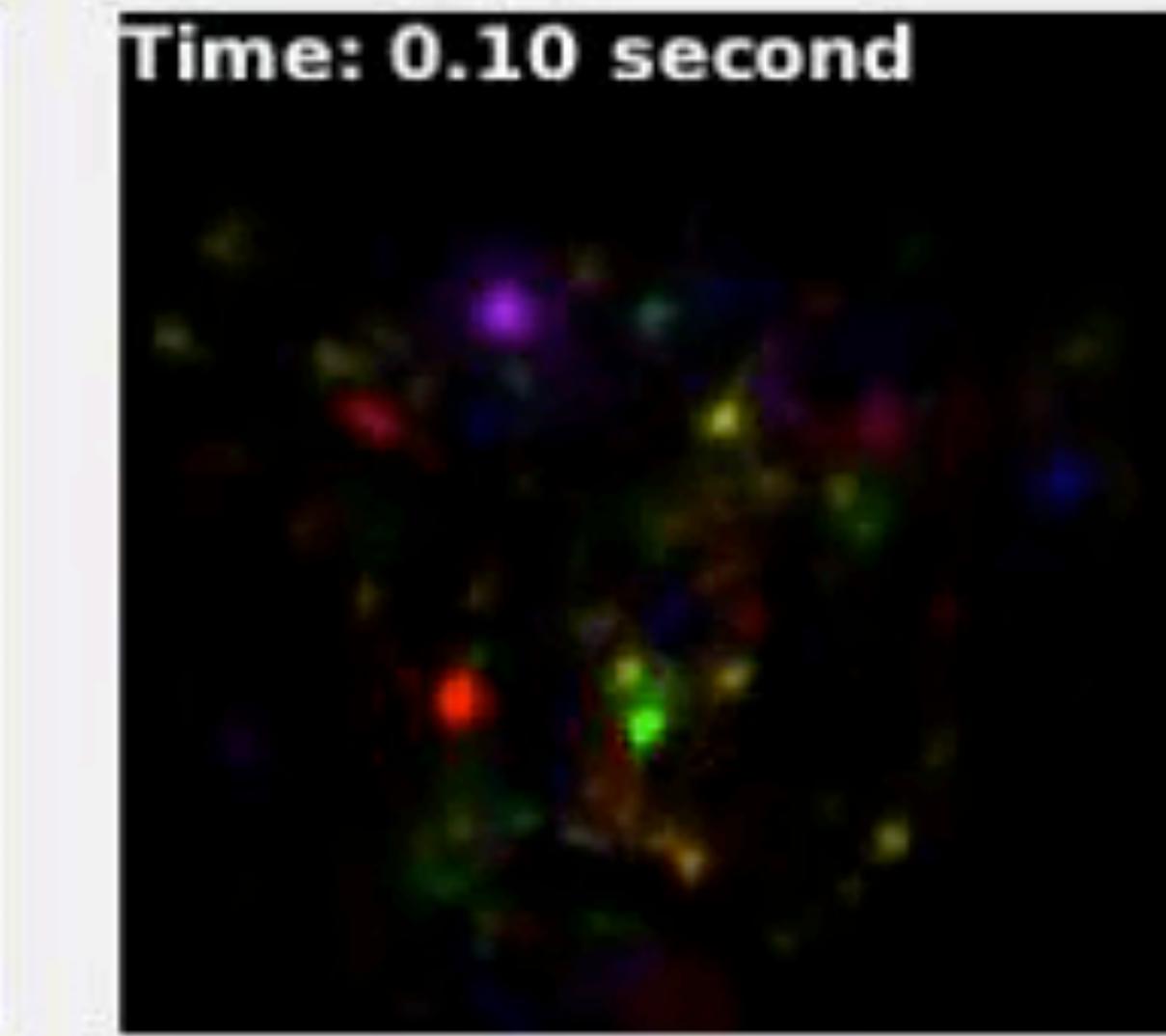
Background



Denoised X 8



Demixed



CNMF-E; Zhou et al, eLife 2018. Very different background model required for 1p data

Conclusion

- **Optical physiology** offers a powerful and complementary toolkit for measuring neural activity in genetically defined cells.
- Methods for extracting calcium fluorescence traces are very similar to those for spike sorting. It's all **convolutional matrix factorization with constraints**.
- If we have an estimate of the noise, we can use it to **set hyper parameters (i.e. thresholds) automatically**.
- **Next time:** we'll dive deeper into the deconvolution problem of inferring spike times and amplitudes from calcium traces.

Further reading

- Ch 3 of the course notes (coming later today)
- Lin, Michael Z., and Mark J. Schnitzer. 2016. “Genetically Encoded Indicators of Neuronal Activity.” *Nature Neuroscience* 19 (9): 1142–53.
- Pnevmatikakis EA, Soudry D, Gao Y, et al. Simultaneous Denoising, Deconvolution, and Demixing of Calcium Imaging Data. *Neuron*. 2016;89(2):285-299. doi:10.1016/j.neuron.2015.11.037
- Pachitariu, Marius, Carsen Stringer, Mario Dipoppa, Sylvia Schröder, L. Federico Rossi, Henry Dalgleish, Matteo Carandini, and Kenneth D. Harris. 2017. “Suite2p: Beyond 10,000 Neurons with Standard Two-Photon Microscopy.” Cold Spring Harbor Laboratory. <https://doi.org/10.1101/061507>.
- Zhou, Pengcheng, Shanna L. Resendez, Jose Rodriguez-Romaguera, Jessica C. Jimenez, Shay Q. Neufeld, Andrea Giovannucci, Johannes Friedrich, et al. 2018. “Efficient and Accurate Extraction of in Vivo Calcium Signals from Microendoscopic Video Data.” *eLife* 7 (February): e28728.
- Giovannucci, Andrea, Johannes Friedrich, Pat Gunn, Jérémie Kalfon, Brandon L. Brown, Sue Ann Koay, Jiannis Taxidis, et al. 2019. “CalmAn an Open Source Tool for Scalable Calcium Imaging Data Analysis.” *eLife* 8 (January). <https://doi.org/10.7554/eLife.38173>.