Modeling transition/transversion bias of nucleotide substitution over time

Stefan Schmutz
January 2019

Introduction

DNA, the molecule essential for known life, consists of four building blocks. Those blocks (thymine (T), cytosine (C), adenine (A), guanine (G)) are called nucleotides.[1]
Since DNA evolves over time, substitutions (change of nucleotides) occur. For structural reasons, transitions (substitutions between A and G or T and C), are more likely to happen compared to transversions (all other substitutions) in the majority of the cases[2].

The aim of this work is to describe a Markov model of nucleotide substitution over time which consideres nucleotide- and transition/transversion-bias.
The four nucleotides of DNA represent the four different states of the Markov model.
Other parameters needed for the model are the steady-state distribution (π) and state transition probabilities. Those parameters will be estimated in the next section and are based on a pairwise sequence alignment of human and mouse cytochrome b (MT-CYB) which is a mitochondrial gene[3].

Estimations to parameterize the model

There are several different Markov models of nucleotide substitution described in the literature[4]. The HKY85 model[5] was chosen because it fits the problem at hand. The substitution rate matrix Q is defined as follows:

$$Q = \begin{bmatrix} . & \alpha\pi_C & \beta\pi_A & \beta\pi_G \\ \alpha\pi_T & . & \beta\pi_A & \beta\pi_G \\ \beta\pi_T & \beta\pi_C & . & \alpha\pi_G \\ \beta\pi_T & \beta\pi_C & \alpha\pi_A & . \end{bmatrix}.$$

α (transition-rate) and β (transversion rate) can be summarised as the transition/transversion rate ratio $\kappa = \alpha/\beta$. The substitution rate matrix can therefore also be described as follows:

$$Q = \begin{bmatrix} . & \kappa\pi_C & \pi_A & \pi_G \\ \kappa\pi_T & . & \pi_A & \pi_G \\ \pi_T & \pi_C & . & \kappa\pi_G \\ \pi_T & \pi_C & \kappa\pi_A & . \end{bmatrix}.$$

The diagonal of Q is defined by the requirement, that each row of a substitution rate matrix must sum to 0.

Nucleotide frequencies

Estimate nucleotide frequencies from the pairwise alignment of human and mouse cytochrome b gene as given in the file "mt-cyb-human-mouse_cDNAalignment.fasta". Use these values to parameterize the model.

A way to estimate the nucleotide frequencies is to count the occurences and divide them by the total length. Since we're working with a pairwise alignment without indels, the total length of both sequences is the same (1140 nt). The detailed composition is listed in Table 1.

Table 1: Nucleotide frequencies

nucleotide	human	mouse
T	0.2509	0.2868
C	0.3430	0.2737
A	0.2860	0.3167
G	0.1202	0.1228

We assume that the mean of this distribution is the steady-state distribution π.

$$\pi = (\pi_T, \pi_C, \pi_A, \pi_G) = (0.2689, 0.3084, 0.3014, 0.1215)$$

Transition transversion rate ratio

> Propose a simple way of estimating transition transversion rate ratio from the dataset and use this estimate for the parameterization of the model.

The numbers of the 16 possible combinations of the sequence alignment are shown in Table 2 (from mouse in rows to human in columns).

Table 2: Nucleotide comparisons (numbers)

	T	C	A	G
T	232	72	20	3
C	32	249	27	4
A	19	59	264	19
G	3	11	15	111

We get the frequencies (Table 3) when the numbers are divided by the total length (1140 nt).

Table 3: Nucleotide comparisons (frequencies)

	T	C	A	G
T	0.2035	0.0632	0.0175	0.0026
C	0.0281	0.2184	0.0237	0.0035
A	0.0167	0.0518	0.2316	0.0167
G	0.0026	0.0096	0.0132	0.0974

The fraction of sites with transitional differences ($S = 0.1211$) and transversional differences ($V = 0.1281$) can be estimated by taking the sum of the corresponding fields from Table 3.
There are different definitions of the transition transversion rate ratio.[4] It was decided to apply the measure used by Kimura[6] and Hasegawa et al.[5] ($\kappa = \alpha/\beta$). The estimate of transition transversion rate ratio (κ) can be calculated as follows:

$$\kappa = \frac{2ln(1 - 2S - V)}{ln(1 - 2V)} - 1 = 2.1248$$

4

(log-)likelihood

Computing by hand

> By hand, compute the (log-)likelihood for the first 10 alignment positions given that the two
> sequences are separated by evolutionary distance of 0.01 expected substitutions per site.

To compute the likelihood (L) and log-likelihood (ℓ) we're given that the distance of the two sequences from
human and mouse is $d = 0.01$. This means that the predicted actual mutations between the two sequences is
0.01 per position.
We assume that the mouse sequence is ancestral to the human sequence.
Using the following formulae[7], the the transition probabilities can be calculated:

$$\lambda = \frac{1}{2(\pi_A + \pi_G)(\pi_C + \pi_T) + 2\kappa(\pi_A\pi_G + \pi_C\pi_T)}$$

$$P_{AA}(d, \kappa, \pi) = \frac{\pi_A(\pi_A + \pi_G + (\pi_C + \pi_T)e^{-\lambda d}) + \pi_G e^{-(1+(\pi_A+\pi_G)(\kappa-1))\lambda d}}{\pi_A + \pi_G}$$

For this example, likelihood is defined as the product of the likelihoods
of each of the 10 positions:

$$L = \prod_{j=1}^{N} L_{(j)}$$

To avoid underflow (computer issue caused by very small numbers) log-likelihood is often used:

$$\ell = \sum_{j=1}^{N} ln(L_{(j)})$$

Computing using a program

> Write a program to compute the likelihood function of the whole alignment for the same genetic
> distance. Compute the likelihood for the whole alignment for several values of transition-
> transversion ratio around the value you estimated. Can you find a value that gives a better
> likelihood?

Implementing a simulation

> Implement a simulation under your model for two sequences over an arbitrary distance t. Validate
> the program by simulation and show the results of you validation.

Discussion

It's unclear if the (requested) applied model is a good choice for this example. The transition transversion
bias of the given sequence alignment is not very strong. Furthermore more complex models usually require
more data to infer the parameter and are more prone to overfitting.

References

1. Wikipedia contributors. DNA. (2019).

2. Keller, D. A. N., Irene AND Bensasson. Transition-transversion bias is not universal: A counter example from grasshopper pseudogenes. *PLOS Genetics* **3**, 1–7 (2007).

3. Andreu, A. L. *et al.* Exercise intolerance due to mutations in the cytochrome b gene of mitochondrial dna. *New England Journal of Medicine* **341**, 1037–1044 (1999).

4. Yang, Z. Molecular evolution. A statistical approach. (2014) doi:10.1093/acprof:oso/9780199602605.001.0001.

5. Hasegawa, M., Kishino, H. & Yano, T.-a. Dating of the human-ape splitting by a molecular clock of mitochondrial dna. *Journal of Molecular Evolution* **22**, 160–174 (1985).

6. Kimura, M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* **16**, 111–120 (1980).

7. Wikipedia contributors. Models of dna evolution. (2020).