

Modeling transition/transversion bias of nucleotide substitution over time

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Introduction

DNA, the molecule essential for known life, consists of four building blocks. Those blocks (thymine (T), cytosine (C), adenine (A), guanine (G)) are called nucleotides.¹

Since DNA evolves over time, substitutions (change of nucleotides) occur. For structural reasons, transitions (substitutions between A and G or T and C), are more likely to happen compared to transversions (all other substitutions) in the majority of the cases².

The aim of this work is to describe a Markov model of nucleotide substitution over time which considers nucleotide- and transition/transversion-bias.

The four nucleotides of DNA represent the four different states of the Markov model.

Other parameters needed for the model are the steady-state distribution (π) and state transition probabilities. Those parameters will be estimated in the next section and are based on a pairwise sequence alignment of human and mouse cytochrome b (MT-CYB) which is a mitochondrial gene³.

Estimations to parameterize the model

There are several different Markov models of nucleotide substitution described in the literature⁴. The HKY85 model⁵ was chosen because it fits the problem at hand. The substitution rate matrix Q is defined as follows:

$$Q = \begin{bmatrix} . & \alpha\pi_C & \beta\pi_A & \beta\pi_G \\ \alpha\pi_T & . & \beta\pi_A & \beta\pi_G \\ \beta\pi_T & \beta\pi_C & . & \alpha\pi_G \\ \beta\pi_T & \beta\pi_C & \alpha\pi_A & . \end{bmatrix}.$$

α (transition-rate) and β (transversion rate) can be summarised as the transition/transversion rate ratio $\kappa = \alpha/\beta$. The substitution rate matrix can therefore also be described as follows:

$$Q = \begin{bmatrix} . & \kappa\pi_C & \pi_A & \pi_G \\ \kappa\pi_T & . & \pi_A & \pi_G \\ \pi_T & \pi_C & . & \kappa\pi_G \\ \pi_T & \pi_C & \kappa\pi_A & . \end{bmatrix}.$$

The diagonal of Q is defined by the requirement, that each row of a substitution rate matrix must sum to 0.

Nucleotide frequencies

Estimate nucleotide frequencies from the pairwise alignment of human and mouse cytochrome b gene as given in the file “mt-cyb-human-mouse_cDNAalignment.fasta”. Use these values to parameterize the model.

A way to estimate the nucleotide frequencies is to count the occurrences and divide them by the total length. Since we're working with a pairwise alignment without indels, the total length of both sequences is the same (1140 nt). The detailed composition is listed in Table 1.

Table 1: Nucleotide frequencies

nucleotide	human	mouse
T	0.2509	0.2868
C	0.3430	0.2737
A	0.2860	0.3167
G	0.1202	0.1228

We assume that the mean of this distribution is the steady-state distribution π .

$$\pi = (\pi_T, \pi_C, \pi_A, \pi_G) = (0.2689, 0.3084, 0.3014, 0.1215)$$

Transition transversion rate ratio

Propose a simple way of estimating transition transversion rate ratio from the dataset and use this estimate for the parameterization of the model.

The numbers of the 16 possible combinations of the sequence alignment are shown in Table 2 (from mouse in rows to human in columns).

Table 2: Nucleotide comparisons (numbers)

	T	C	A	G
T	232	72	20	3
C	32	249	27	4
A	19	59	264	19
G	3	11	15	111

We get the frequencies (Table 3) when the numbers are divided by the total length (1140 nt).

Table 3: Nucleotide comparisons (frequencies)

	T	C	A	G
T	0.2035	0.0632	0.0175	0.0026
C	0.0281	0.2184	0.0237	0.0035
A	0.0167	0.0518	0.2316	0.0167
G	0.0026	0.0096	0.0132	0.0974

The fraction of sites with transitional differences ($S = 0.1211$) and transversional differences ($V = 0.1281$) can be estimated by taking the sum of the corresponding fields from Table 3.

There are different definitions of the transition transversion rate ratio.⁴ It was decided to apply the measure used by Kimura⁶ and Hasegawa et al.⁵ ($\kappa = \alpha/\beta$). The estimate of transition transversion rate ratio ($\hat{\kappa}$) can be calculated as follows:

$$\hat{\kappa} = \frac{2\ln(1 - 2S - V)}{\ln(1 - 2V)} - 1 = 2.1248$$

(log-)likelihood

Computing by hand

By hand, compute the (log-)likelihood for the first 10 alignment positions given that the two sequences are separated by evolutionary distance of 0.01 expected substitutions per site.

To compute the likelihood (L) and log-likelihood (ℓ) we're given that the distance of the two sequences from human and mouse is $\hat{d} = 0.01$. This means that the predicted actual mutations between the two sequences is 0.01 per position.

?how do we use that information?

The likelihood at a position is defined as the sum of the probabilities of all possible ancestral nucleotides. For this pairwise alignment we assume one internal node (one common ancestor) which gives us 4 possibilities.

For this example, likelihood is defined as the product of the likelihoods of each of the 10 positions:

$$L = \prod_{j=1}^N L_{(j)}$$

To avoid underflow (computer issue caused by very small numbers) log-likelihood is often used:

$$\ell = \sum_{j=1}^N \ln(L_{(j)})$$

Computing using a program

Write a program to compute the likelihood function of the whole alignment for the same genetic distance. Compute the likelihood for the whole alignment for several values of transition-transversion ratio around the value you estimated. Can you find a value that gives a better likelihood?

Implementing a simulation

Implement a simulation under your model for two sequences over an arbitrary distance t . Validate the program by simulation and show the results of you validation.

References

1. Wikipedia contributors. DNA. (2019).
2. Keller, D. A. N., Irene AND Bensasson. Transition-transversion bias is not universal: A counter example from grasshopper pseudogenes. *PLOS Genetics* **3**, 1–7 (2007).
3. Andreu, A. L. *et al.* Exercise intolerance due to mutations in the cytochrome b gene of mitochondrial dna. *New England Journal of Medicine* **341**, 1037–1044 (1999).
4. Yang, Z. Molecular evolution. A statistical approach. (2014) doi:10.1093/acprof:oso/9780199602605.001.0001.
5. Hasegawa, M., Kishino, H. & Yano, T.-a. Dating of the human-ape splitting by a molecular clock of mitochondrial dna. *Journal of Molecular Evolution* **22**, 160–174 (1985).
6. Kimura, M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* **16**, 111–120 (1980).