

Package ‘CYCLeR’

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Type Package

Title CircRNA transcriptome assembly tool

Version 1.3

Description CYCLeR is a software package for assembly of circRNA transcripts from RNA-seq data. Takes a set of BSJ prediction files and RNA-seq BAM files as an input and outputs circRNA transcripts as FASTA, GTF and flat annotation files. The tools also outputs a padded FASTA to serve as an index for transcript EM abundance estimation.

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LazyData yes

Depends R,
Rsamtools,
SummarizedExperiment,
methods,
tidyverse,
SGSeq,
igraph,
DEXSeq,
polyester

Imports AnnotationDbi,
BiocGenerics,
Biostrings,
GenomicAlignments,
GenomicFeatures,
GenomeInfoDb,
RUnit,
S4Vectors,
grDevices,
graphics,
igraph,
parallel,
rtracklayer,
stats,
tidyverse,
SGSeq,

igraph,
DEXSeq,
polyester
Suggests BiocStyle,
BSgenome.Hsapiens.UCSC.hg38,
TxDb.Hsapiens.UCSC.hg38.knownGene,
knitr,
rmarkdown
VignetteBuilder knitr
biocViews TranscriptomeAssembly, RNASeq, Transcription
RoxygenNote 7.1.1
Encoding UTF-8

R topics documented:

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combine.two.BSJ.tables
<i>combine BSJs</i>

Description

Combine 2 BSJ tables

Usage

combine.two.BSJ.tables(ce_bsjs, ciri_bsjs)

Arguments

ce_bsjs	BSJ table 1
ciri_bsjs	BSJ table 2

Details

Just a combination of BSJ tables to make sure we have a complete set of BSJs. The variable names do not actually matter since the all tables have the same formatting.

Value

Tibble object with combined filtered BSJ coordinate and number of junction spanning reads across sample.

Author(s)

Stefan Stefanov

filter.bam	<i>BAM file filter</i>
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Description

A wrapper function for samtools use to trim the files

Usage

```
## S3 method for class 'bam'
filter(BSJ_gr, sample_table, samtools_prefix)
```

Arguments

BSJ_gr a GRange of BSJ coordinates

sample_table sample table formatted according to the manual, Must contain “sample_name” “treatment” “file_bam” “lib_size” “read_len”; NB the values in column “treatment” can only be “control” and “enriched”

samtools_prefix a string that corresponds to user’s samtools run prefix

Details

This function removes the BAM file reads that do not overlap with the BSJ loci. This significantly speeds up the feature detection and lowers the virtual memory requirements

Value

BAMFileList object with info on the trimmed files

Author(s)

Stefan Stefanov

```
find.depleted.features
```

CircRNA feature selection

Description

CircRNA feature selection

Usage

```
find.depleted.features(circ_fc_adj, sample_table, circ_sg, test = "DEX")
```

Arguments

`circ_fc_adj` count matrix corresponding to the circRNA features

`sample_table` sample table formatted according to the manual, Must contain “sample_name” “treatment” “file_bam” “lib_size” “read_len”; NB the values in column “treatment” can only be “control” and “enriched”

`circ_sg` SGSeq object supplying feature info

`test` either “DEX” for DEXSeq based feature selection or “comparison” simple average comparison

Details

This function works in 2 ways: direct comparison of average quantities or as a wrapper of DEXSeq. In case of dataset with replicates, the suggested approach is the use of DEXSeq statistical test.

Value

vector of featureID

Author(s)

Stefan Stefanov

```
make.BSJ.gr
```

Convert BSJ string to GRanges object

Description

Convert BSJ string to GRanges object

Usage

```
make.BSJ.gr(BSJ_set)
```

Arguments

BSJ_set a list of BSJ ID records procudes by process.BSJ or combine.two.BSJ.tables

Details

Convert BSJ string to GRanges obejct

Value

GRanges object indicating BSJ loci

Author(s)

Stefan Stefanov

make.BSJ.sg	<i>Preparation of the BSJ-specific splice graphs</i>
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Description

Selection of the exons based on BSJ set

Usage

```
make.BSJ.sg(circ_sg, BSJ_gr)
```

Arguments

circ_sg SGSeq prediction object
BSJ_gr a GRange of BSJ coordinates

Details

Selection of the exons based on BSJ set

Value

SGSeq containing exons belonging to BSJ loci

Author(s)

Stefan Stefanov

merge.qics	<i>Merging 2 assemblies</i>
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Description

Pair-wise merging 2 assemblies Pair-wise merging 2 assemblies

Usage

```
## S3 method for class 'qics'  
merge(qics1, qics2)
```

Arguments

qics1	assembly 1
qics2	assembly 2

Value

data.frame of transcript information in flat format

Author(s)

Stefan Stefanov

overlap.SG.BSJ	<i>Overlap of BSJ and a splice graph</i>
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Description

Creates a disjointed set of exons based on a SGSeq object and a BSJ GRanges object

Usage

```
overlap.SG.BSJ(sgfc_pred, BSJ_gr)
```

Arguments

sgfc_pred	SGSeq prediction object
BSJ_gr	a GRange of BSJ coordinates

Details

Creates a disjointed set of exons based on a SGSeq object and a BSJ GRanges object. The function keeps the SGSeq metadata

Value

SGSeq with disjoint exon bins

Author(s)

Stefan Stefanov

parse.files	<i>Parse BSJ input</i>
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Description

Parse BSJ files from CIRI, CIRCexplorer2 or a TSV file

Usage

```
parse.files(file_list, file_path, input_type)
```

Arguments

- | | |
|------------|----------------------------------------------------------------------------------|
| file_list | list with file names |
| file_path | string object with file path, could be an empty string |
| input_type | CIRI for CIRI2 input, CE for CIRCexplorer2 input and tsv for TSV formatted input |

Details

This processes BSJ prediction files and prepares them for the next step of the pipeline. `input_type` is essential for the correct parsing of the files.

Value

Tibble object with combined BSJ coordinate and number of junction spanning reads across sample

Author(s)

Stefan Stefanov

plotRanges2	<i>Plot ranges</i>
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Description

Plots GRanges objects

Usage

```
plotRanges2(...)
```

Details

ggplot of multiple GRanges object. Every object is auto assigned a colour from colorblind friendly scheme

Value

ggplot of multiple GRanges objects

Author(s)

Stefan Stefanov

process.BSJs	<i>Process BSJs</i>
--------------	---------------------

Description

process the BSJ table and select high confidence BSJs

Usage

```
process.BSJs(cdf, sample_table)
```

Arguments

cdf	tibble produced by <code>parse.files</code>
sample_table	sample table formatted according to the manual, Must contain “sample_name” “treatment” “file_bam” “lib_size” “read_len”; NB the values in column “treatment” can only be “control” and “enriched”
file_path	string object with file path, could be an empty string

Details

Filters BSJ based on comparison of the average CPM values of BSJs

Value

Tibble object with combined filtered BSJ coordinate and number of junction spanning reads across sample.

Author(s)

Stefan Stefanov

recount.features	<i>Re-count of the reads per exon bin</i>
------------------	-------------------------------------------

Description

A wrapper function for Rsubread

Usage

```
recount.features(full_sg, sample_table)
```

Arguments

full_sg	a SGSeq object of exon bins
sample_table	sample table formatted according to the manual, Must contain "sample_name" "treatment" "file_bam" "lib_size" "read_len"; NB the values in column "treatment" can only be "control" and "enriched"

Details

This function performs requantification of the exon bins with specifically selected parameters

Value

BAMFileList object with info on the trimmed files

Author(s)

Stefan Stefanov

RPKM.calc

*RPKM calculation for the genomic features***Description**

RPKM calculation for the genomic features

Usage

```
RPKM.calc(
  count_matrix,
  sg,
  bsj_granges,
  bs_genome,
  sample_table,
  feature_type,
  fsj_overhang = 3,
  bsj_overhang = 15,
  eff_length_correction = T,
  gc_correction = F
)
```

Arguments

`count_matrix` count matrix corresponding to the features

`sg` SGSeq object supplying feature info

`bsj_granges` GRange of BSJ coordinates

`bs_genome` a BSGenome object used for extracting the sequences

`sample_table` sample table formatted according to the manual, Must contain “sample_name” “treatment” “file_bam” “lib_size” “read_len”; NB the values in column “treatment” can only be “control” and “enriched”

`feature_type` either “e” for exons or “j” for junctions

`fsj_overhang` the FJS overhang used in the mapping a.k.a. anchor

`bsj_overhang` the BSJ overhang used in the chimeric detection

`eff_length_correction` whether or not to apply effective length correction

`gc_correction` whether or not to apply GC-content correction; requires further testing

Details

This function performs RPKM calculations for the exonic features. The RPKM calculation is performed based on the exact sequences for the exons. For junctions, the sequences are selected based on the exons, flanking the junction. The function takes into account the needed effective length corrections.

Value

BAMFileList object with info on the trimmed files

Author(s)

Stefan Stefanov

`transcripts.per.sample`

Transcript assembly

Description

Transcript assembly per sample Transcript assembly per sample based on sample name in the “sample_table”

Usage

`transcripts.per.sample(i)`

Arguments

`i` name of the sample

Value

`data.frame` of transcript information in flat format

Author(s)

Stefan Stefanov

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