R documentation

of all in '.'

August 19, 2021

R topics documented:

`combine.two.BSJ.tables` *combine BSJs*

Description

Combine 2 BSJ tables

Usage

```
combine.two.BSJ.tables(ce_bsjs, ciri_bsjs)
```

Arguments

`ce_bsjs`	BSJ table 1
`ciri_bsjs`	BSJ table 2

Details

Just a combination of BSJ tables to make sure we have a complete set of BSJs. The variable names do not actually matter since the all tables have the same formatting.

Value

`Tibble` object with combined filtered BSJ coordinate and number of junction spanning reads across sample.

Author(s)

Stefan Stefanov

`filter.bam` *BAM file filter*

Description

A wrapper function for samtools use to trim the files

Usage

```
## S3 method for class 'bam'
filter(BSJ_gr, sample_table, samtools_prefix)
```

Arguments

`BSJ_gr`	a GRange of BSJ cooredinates
`sample_table`	sample table formatted according to the manual, Must contain "sample_name" "treatment" "file_bam" "lib_size" "read_len"; NB the values in column "treatment" can only be "control" and "enriched"
`samtools_prefix`	a string that corresponds to user's samtools run prefix

Details

This function removes the BAM file reads that do not overlap with the BSJ loci. This significantly speeds us the feature detection and lowers the virtual memory requirements

Value

`BAMFileList` object with info on the trimmed files

Author(s)

Stefan Stefanov

find.depleted.features *CircRNA feature selection*

Description

CircRNA feature selection

Usage

```
find.depleted.features(circ_fc_adj, sample_table, circ_sg, test = "DEX")
```

Arguments

circ_fc_adj	count matrix corresponding to the circRNA features
sample_table	sample table formatted according to the manual, Must contain "sample_name" "treatment" "file_bam" "lib_size" "read_len"; NB the values in column "treatment" can only be "control" and "enriched"
circ_sg	SGSeq object supplying feature info
test	either "DEX" for DEXSeq based feature selection or "comparison" simple average comaparison

Details

This function works in 2 ways: direct comparison of average quantities or as a wrapper of DEXSeq. In case of dataset with replicates, the suggested approach is the use of DEXSeq statistical test.

Value

vector of featureID

Author(s)

Stefan Stefanov

make.BSJ.gr *Convert BSJ string to GRanges obejct*

Description

Convert BSJ string to GRanges obejct

Usage

```
make.BSJ.gr(BSJ_set)
```

Arguments

BSJ_set	a list of BSJ ID records procudes by process.BSJs or combine.two.BSJ.tables

Details

Convert BSJ string to GRanges obejct

Value

GRanges object indicating BSJ loci

Author(s)

Stefan Stefanov

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make.BSJ.sg *Preparation of the BSJ-specific splice graphs*

Description

Selection of the exons based on BSJ set

Usage

```
make.BSJ.sg(circ_sg, BSJ_gr)
```

Arguments

circ_sg	SGSeq prediction object
BSJ_gr	a GRange of BSJ coordinates

Details

Selection of the exons based on BSJ set

Value

SGSeq containing exons belonging to BSJ loci

Author(s)

Stefan Stefanov

merge.qics *Merging 2 assemblies*

Description

Pair-wise merging 2 assemblies Pair-wise merging 2 assemblies

Usage

```
## S3 method for class 'qics'
merge(qics1, qics2)
```

Arguments

qics1	assembly 1
qics2	assembly 2

Value

data.frame of transcript information in flat format

Author(s)

Stefan Stefanov

overlap.SG.BSJ *Overlap of BSJ and a splice graph*

Description

Creates a disjointed set of exons based on a SGSeq obejct and a BSJ GRanges object

Usage

```
overlap.SG.BSJ(sgfc_pred, BSJ_gr)
```

Arguments

sgfc_pred	SGSeq prediction object
BSJ_gr	a GRange of BSJ cooredinates

Details

Creates a disjointed set of exons based on a SGSeq obejct and a BSJ GRanges object. The function keeps the SGSeq metadata

Value

`SGSeq` with disjoint exon bins

Author(s)

Stefan Stefanov

`parse.files` *Parse BSJ input*

Description

Parse BSJ files from CIRI, CIRCexplorer2 or a TSV file

Usage

```
parse.files(file_list, file_path, input_type)
```

Arguments

`file_list`	list with file names
`file_path`	string object with file path, clould be an empty string
`input_type`	`CIRI` for CIRI2 input, `CE` for CIRCexplorer2 input and `tsv` for TSV formatted input

Details

This processes BSJ prediction files and prepares them for the next step of the pipeline. `input_type` is essential for the correct parsing of the files.

Value

`Tibble` object with combined BSJ coordinate and number of junction spanning reads across sample

Author(s)

Stefan Stefanov

plotRanges2 *Plot ranges*

Description

Plots GRanges objects

Usage

```
plotRanges2(...)
```

Details

ggplot of multiple GRanges object. Every object is auto assigned a colour from colorblind friendly scheme

Value

ggplot of multiple GRanges objects

Author(s)

Stefan Stefanov

process.BSJs *Process BSJs*

Description

process the BSJ table and select high confidence BSJs

Usage

```
process.BSJs(cdf, sample_table)
```

Arguments

cdf	tibble produced by `parse.files`
sample_table	sample table formatted according to the manual, Must contain "sample_name" "treatment" "file_bam" "lib_size" "read_len"; NB the values in column "treatment" can only be "control" and "enriched"
file_path	string object with file path, clould be an empty string

Details

Filters BSJ based on comparison of the average CPM values of BSJs

Value

`Tibble` object with combined filtered BSJ coordinate and number of junction spanning reads across sample.

Author(s)

Stefan Stefanov

`recount.features` *Re-count of the reads per exon bin*

Description

A wrapper function for Rsubread

Usage

```
recount.features(full_sg, sample_table)
```

Arguments

`full_sg`	a `SGSeq` obeject of exon bins
`sample_table`	sample table formatted according to the manual, Must contain "sample_name" "treatment" "file_bam" "lib_size" "read_len"; NB the values in column "treatment" can only be "control" and "enriched"

Details

This function performs requantification of the exon bins with specifically selected parameters

Value

`BAMFileList` object with info on the trimmed files

Author(s)

Stefan Stefanov

RPKM.calc *RPKM calculation for the genomic features*

Description

RPKM calculation for the genomic features

Usage

```
RPKM.calc(
  count_matrix,
  sg,
  bsj_granges,
  bs_genome,
  sample_table,
  feature_type,
  fsj_overhang = 3,
  bsj_overhang = 15,
  eff_length_correction = T,
  gc_correction = F
)
```

Arguments

count_matrix	count matrix corresponding to the features
sg	SGSeq object supplying feature info
bsj_granges	GRange of BSJ cooredinates
bs_genome	a BSGenome object used for extracting the sequences
sample_table	sample table formatted according to the manual, Must contain "sample_name" "treatment" "file_bam" "lib_size" "read_len"; NB the values in column "treatment" can only be "control" and "enriched"
feature_type	either "e" for exons ot "j" for junctions
fsj_overhang	the FJS overhand used in the mapping a.k.a. anchor
bsj_overhang	the BSJ overhand used in the chimeric detection
eff_length_correction	whether or not to apply effective length correction
gc_correction	whether or not to apply GC-content correction; requires further testing

Details

This function performs RPKM calculations for the exonic features. The RPKM calculation is performed based on the exact sequences for the exons. For junctions, the sequences are selected based on the exons, flanking the junction. The function takes into account the needed effective length corrections.

Value

`BAMFileList` object with info on the trimmed files

Author(s)

Stefan Stefanov

`transcripts.per.sample` *Transcript assembly*

Description

Transcript assembly per sample Transcript assembly per sample based on sample name in the "sample_table"

Usage

```
transcripts.per.sample(i)
```

Arguments

`i`	name of the sample

Value

`data.frame` of transcript information in flat format

Author(s)

Stefan Stefanov

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