Possible Error Message Bug in DESeq2

This report highlights a potential bug in an error message in DESeq2's DESeqDataSetFromMatrix. While it causes no harm to the data, it can mislead an inexperienced user in such a way as to compromise the results of an analysis.

TL;DNR

When using DESeq2 Passing a data.frame containing a column of type factor results in an error message stating the user has entered a column of type character. A character can be safely cast as an integer, but a factor cannot; R will silently change them to an ordered sequence, completely corrupting and invalidating the data (ie, the sequence 1, 100, 1000 is changed to 1, 2, 3). If a user forgets to set stringsAsFactors = FALSE and depends too heavily om the error message, they can essentially randomized thier results without R or DESeq2 warning them. The user is ultimately responsible for their types. However given that R promotes the accidental use of factors, has default behavior that corrupts such data silently, and that R users are often investigators from other disciplines who may not have experience coding, steps to reduce the chance of error may be helpful.

After being sent some bad data, I observed what I think is a bug in which DESeq2 misdiagnoses user input containing factors. The error does no harm in and of itself, but if a user takes what seems like a reasonable next step based on the error message, R's type system will silently invalidate the results. We will need a basic counts file to demonstrate and reproduce the issues, so we will recreate the one from the vignette:

Steps to Re-create

Obtain a counts file

```
library(DESeq2)
                    # qo-to DE analysis package
library(pasilla)
                    # sample data used in tutorial
library(tidyverse) # duh
# get the counts information from the pasilla package
pasCts <- system.file("extdata", "pasilla_gene_counts.tsv",</pre>
                       package = "pasilla", mustWork = TRUE)
# load the sample annotation file from the pasilla package
pasAnno <- system.file("extdata", "pasilla_sample_annotation.csv",</pre>
                       package = "pasilla", mustWork = TRUE)
# create a matrix of counts
cts <- as.matrix(read.csv(pasCts, sep = "\t", row.names = "gene id"))
# read in the sample data.
coldata <- read.csv(pasAnno, row.names = 1)</pre>
# select the desired features (just following the tutorial)
coldata <- coldata[, c("condition", "type")]</pre>
# clean/standardize the rownames
```

```
rownames(coldata) <- sub("fb", "", rownames(coldata))</pre>
# sort/reorder the columns to match samples
cts <- cts[, rownames(coldata)]</pre>
```

Introduce a character

8599 FBgn0037429

171

118

I found the error after reading in a bad counts matrix file that was given to me. Somewhere in pre-processing a header of some kind had gotten duplicated and nestled a few thousand rows into the counts file. It looked something like this:

sampleName sampleName sampleName sampleName sampleName sampleName

Perhaps tables had been stacked on top of one another to make the counts file and there was an off-by-one error, I don't know, but is an easy error to imagine making in Bioinformatics and data-related computing in general. It can be reproduced like this:

```
ctsBad <- cts
ctsBad[8600,] <- c("here", "there", "and", "everywhere", "yeah", "whoo!", "oops!")
ctsBad[8597:8602, ]
                                                         untreated2 untreated3
##
               treated1 treated2 treated3 untreated1
                                                                     "0"
## FBgn0037427 "0"
                         "0"
                                  "0"
                                            "0"
                                                         "0"
## FBgn0037428 "2"
                         "0"
                                  "0"
                                            "0"
                                                         "0"
                                                                     "0"
                                                         "204"
                                                                     "85"
## FBgn0037429 "171"
                         "118"
                                  "101"
                                            "121"
## FBgn0037430 "here"
                         "there"
                                  "and"
                                            "everywhere" "yeah"
                                                                     "whoo!"
                         "2"
                                            "3"
                                                                     "1"
## FBgn0037431 "9"
                                  "1"
                                                         "7"
                         "2"
                                  "6"
                                            "6"
                                                         "17"
                                                                     "4"
## FBgn0037432 "9"
               untreated4
## FBgn0037427 "0"
## FBgn0037428 "0"
## FBgn0037429 "103"
## FBgn0037430 "oops!"
## FBgn0037431 "3"
## FBgn0037432 "7"
write.csv(ctsBad, "badCounts.csv")
```

When we write the data to a file, the well-know-but-still-menacing factor default issues comes in to play. However, DESeq2 appears to misdiagnose the types. To see how, we first read in the file.

```
ctsBadFile <- read.delim("badCounts.csv", sep = ",")</pre>
sapply(ctsBadFile, class)
##
                 treated1
                            treated2
                                        treated3 untreated1 untreated2 untreated3
##
     "factor"
                 "factor"
                             "factor"
                                        "factor"
                                                    "factor"
                                                                "factor"
                                                                            "factor"
## untreated4
##
     "factor"
ctsBadFile[8597:8602, ]
                   X treated1 treated2 treated3 untreated1 untreated2 untreated3
## 8597 FBgn0037427
                            0
                                      0
                                                0
                                                           0
                                                                       0
                                                                                   0
## 8598 FBgn0037428
                            2
                                      0
                                               0
                                                           0
                                                                       0
                                                                                   0
                                              101
                                                         121
                                                                     204
```

85

```
## 8600 FBgn0037430
                                  there
                                              and everywhere
                                                                               whoo!
                         here
                                                                    veah
## 8601 FBgn0037431
                                                                       7
                            9
                                      2
                                                1
                                                            3
                                                                                   1
                                      2
## 8602 FBgn0037432
                             9
                                                6
                                                            6
                                                                       17
                                                                                    4
##
        untreated4
## 8597
## 8598
                  0
                103
## 8599
## 8600
              oops!
## 8601
                  3
                  7
## 8602
```

Call DESeqDataSetFromMatrix()

Though we're passing factors. we're told we are passing characters.

```
# make the input the corrected sized matrix
ctsBadFile$X <- NULL</pre>
# demonstrate that we're passing factors
sapply(ctsBadFile, class)
##
     treated1
                treated2
                            treated3 untreated1 untreated2 untreated3 untreated4
     "factor"
                "factor"
##
                            "factor"
                                       "factor"
                                                   "factor"
                                                              "factor"
                                                                          "factor"
error <- tryCatch(
  {
    # try to use the bad one
    dds <- DESeqDataSetFromMatrix(countData = ctsBadFile,</pre>
                                   colData = coldata, design = ~ condition)
  },
  # "upon error 'e', use this function to show a message of 'e'"
  error = function(e) { return(e)}
# show the error messagge (just splitting it because it is long)
errorMessage <- unlist(str_split(error, ":"))</pre>
## Warning in stri_split_regex(string, pattern, n = n, simplify = simplify, :
## argument is not an atomic vector; coercing
cat("\n", errorMessage[2], errorMessage[3])
##
```

Suspected Cause

##

The real issue is that characters can be safely coerced to integers, but factors cannot. Moreover, they fail silently and return invalid results. We demonstrate below for the sake of completeness:

counts matrix should be numeric, currently it has mode character

```
# a vector of integers
x <- c("1", "10", "1000")
as.integer(x)</pre>
```

```
## [1] 1 10 100 1000
```

When you do that to a list of factors however, you get a deeply problematic and counter-intuitive result:

```
# the same, as a factor
x <- factor(c("1", "10", "100", "1000"))
as.integer(x)
## [1] 1 2 3 4
The output is effectively unrelated to the input. Given the error message, it's tempting to think one should
```

just change the characters to integers (or numerics), which is a normal and safe operation:

```
# apply the fix insinutated by the error message
ctsBadFileAsInt <- ctsBadFile %>%
 mutate_all(as.integer)
ctsBadFileAsNumeric <- ctsBadFile %>%
 mutate_all(as.numeric)
ctsBadFile %>% head()
```

##		treated1	treated2	treated3	untreated1	untreated2	untreated3	untreated4
##	1	0	0	1	0	0	0	0
##	2	140	88	70	92	161	76	70
##	3	4	0	0	5	1	0	0
##	4	1	0	0	0	2	1	2
##	5	6205	3072	3334	4664	8714	3564	3150
##	6	722	299	308	583	761	245	310

ctsBadFileAsInt %>% head()

##		${\tt treated1}$	${\tt treated2}$	${\tt treated3}$	${\tt untreated1}$	${\tt untreated2}$	${\tt untreated3}$	${\tt untreated 4}$
##	1	1	1	2	1	1	1	1
##	2	485	2285	2139	2847	712	2011	2033
##	3	2133	1	1	2144	2	1	1
##	4	2	1	1	1	1046	2	813
##	5	2772	1281	1413	2066	3368	1343	1290
##	6	2966	1251	1326	2325	3172	978	1273

ctsBadFileAsNumeric %>% head()

##		treated1	treated2	treated3	untreated1	${\tt untreated2}$	untreated3	untreated4
##	1	1	1	2	1	1	1	1
##	2	485	2285	2139	2847	712	2011	2033
##	3	2133	1	1	2144	2	1	1
##	4	2	1	1	1	1046	2	813
##	5	2772	1281	1413	2066	3368	1343	1290
##	6	2966	1251	1326	2325	3172	978	1273

We see that the

However, we don't actually have characters, we have factors, so we coerce our data to something only vaguely based on the actual values:

```
# observe results
message("original")
## original
```

```
cts %>% head(10)
```

```
treated1 treated2 treated3 untreated1 untreated2 untreated3
##
## FBgn000003
                      0
                                0
                                         1
                                                     0
                                                                0
                                                                            0
                                        70
                                                    92
                                                              161
                                                                           76
## FBgn0000008
                     140
                               88
## FBgn0000014
                       4
                                0
                                         0
                                                     5
                                                                1
                                                                            0
## FBgn000015
                       1
                                         0
                                                     0
                                                                2
                                                                            1
                                0
## FBgn000017
                    6205
                             3072
                                      3334
                                                  4664
                                                             8714
                                                                         3564
## FBgn000018
                     722
                              299
                                       308
                                                   583
                                                              761
                                                                          245
## FBgn0000022
                                                                            0
                      0
                                0
                                         0
                                                     0
                                                                1
## FBgn0000024
                      10
                                7
                                         5
                                                    10
                                                                11
                                                                            3
## FBgn0000028
                       0
                                         1
                                                     0
                                                                            0
                                1
                                                                1
## FBgn0000032
                    1698
                              696
                                       757
                                                  1446
                                                             1713
                                                                          615
##
               untreated4
## FBgn000003
                        70
## FBgn0000008
## FBgn000014
                         0
                         2
## FBgn000015
## FBgn0000017
                      3150
## FBgn000018
                       310
## FBgn0000022
                         0
## FBgn0000024
                         3
## FBgn0000028
                         0
## FBgn0000032
                       672
```

message("bad file")

bad file

ctsBadFile %>% head(10)

##		treated1	${\tt treated2}$	${\tt treated3}$	${\tt untreated1}$	${\tt untreated2}$	${\tt untreated3}$	untreated4
##	1	0	0	1	0	0	0	0
##	2	140	88	70	92	161	76	70
##	3	4	0	0	5	1	0	0
##	4	1	0	0	0	2	1	2
##	5	6205	3072	3334	4664	8714	3564	3150
##	6	722	299	308	583	761	245	310
##	7	0	0	0	0	1	0	0
##	8	10	7	5	10	11	3	3
##	9	0	1	1	0	1	0	0
##	10	1698	696	757	1446	1713	615	672

message("after coercion")

after coercion

ctsBadFileAsInt %>% head(10)

##		treated1	${\tt treated2}$	${\tt treated3}$	${\tt untreated1}$	${\tt untreated2}$	${\tt untreated3}$	${\tt untreated4}$
##	1	1	1	2	1	1	1	1
##	2	485	2285	2139	2847	712	2011	2033
##	3	2133	1	1	2144	2	1	1
##	4	2	1	1	1	1046	2	813
##	5	2772	1281	1413	2066	3368	1343	1290
##	6	2966	1251	1326	2325	3172	978	1273
##	7	1	1	1	1	2	1	1
##	8	3	2061	1824	3	142	1176	1237
##	9	1	2	2	1	2	1	1

10 767 2055 2213 489 809 1823 1999

DESeq2 has excellent documentation, and during routine QC check presented in the vignette, I identified the problem. In the case presented here, the plotMA function makes it clear that something is wrong, but on my initial dataset, the graphs looked passable.

Inspecting the source code of DESeqDataSetFromMatrix, we see that there is a coercion of input to type matrix, which is reasonable given that most users will pass a dataframe. However, passing a factor to as.matrix will results in another silent conversion of factor to character. Because DESeq2 checks the types after coercion of the inputs, the message given to users the current state after modification, not the nature of the input as it was given. This can be illustrated with a modification to DESeq2 source code, presented below (my own comments are denoted ### ---> to distinguish them from those of the original author).

```
DESeqDataSetFromMatrixDebug <- function( countData, colData, design, tidy=FALSE, ignoreRank=FALSE, ...
{
  ### ---> a function to observe types at a given time point
  printDebugOutput <- function(tag)</pre>
    ### ---> so we can pass an identifier of some kind
    message(tag)
    ### ---> what is the class of the count data at this point?
    print(paste("class of countData: ", class(countData)))
    ### ---> what are the classes of each of the columns? (truncated so it isn't too long)
    print(sapply(countData, class)[1:5])
  ### ---> inital types
  printDebugOutput("initial")
  if (tidy) {
    stopifnot(ncol(countData) > 1)
    rownms <- as.character(countData[,1])</pre>
    countData <- countData[,-1,drop=FALSE]</pre>
    rownames(countData) <- rownms</pre>
  }
  # check that these agree in number
  stopifnot(ncol(countData) == nrow(colData))
  ### ---> there is a coercion to type matrix
  # we expect a matrix of counts, which are non-negative integers
  countData <- as.matrix( countData )</pre>
  ### ---> reobserve the types
  printDebugOutput("after coercion to matrix")
  ### ---> the code is unchanged after this point
  if (is(colData, "data.frame"))
    colData <- as(colData, "DataFrame")</pre>
```

```
# check if the rownames of colData are simply in different order
  # than the colnames of the countData, if so throw an error
  # as the user probably should investigate what's wrong
  if (!is.null(rownames(colData)) & !is.null(colnames(countData))) {
   if (all(sort(rownames(colData)) == sort(colnames(countData)))) {
      if (!all(rownames(colData) == colnames(countData))) {
        stop(paste("rownames of the colData:
  ",paste(rownames(colData),collapse=","),"
  are not in the same order as the colnames of the countData:
  ",paste(colnames(countData),collapse=",")))
   }
  }
  if (is.null(rownames(colData)) & !is.null(colnames(countData))) {
   rownames(colData) <- colnames(countData)</pre>
  se <- SummarizedExperiment(assays = SimpleList(counts=countData), colData = colData, ...)</pre>
  object <- DESeqDataSet(se, design = design, ignoreRank)</pre>
 return(object)
We can use this function to check the types:
tryCatch(
  {
    # try to use the bad one
   dds <- DESeqDataSetFromMatrixDebug(countData = ctsBadFile,</pre>
                                       colData = coldata, design = ~ condition)
 },
  # "upon error 'e', use this function to show a message of 'e'"
  error = function(e) { print(e) }
)
## initial
## [1] "class of countData: data.frame"
    treated1
              treated2 treated3 untreated1 untreated2
     "factor"
                "factor"
                           "factor" "factor"
                                                 "factor"
##
## after coercion to matrix
## [1] "class of countData: matrix"
            0
                 140
                                                           6205
## "character" "character" "character" "character"
## <simpleError in DESeqDataSet(se, design = design, ignoreRank): counts matrix should be numeric, curr
This behavior can be confirmed in base R.
class(iris[,5])
## [1] "factor"
class(as.matrix(iris)[,5])
## [1] "character"
```

Potential Solution

The actual warning is raised in a separate function called DESeqDataSet, which is wrapped by DESeqDataSetFromMatrix. DESeqDataSetFromMatrix, however, could check for factors (or do whatever type-checking desired) before passing the input to DESeqDataSet. An example might look like this:

```
DESeqDataSetFromMatrixDebugFactorSafe <- function( countData, colData, design, tidy=FALSE, ignoreRank=F
  ### ---> checking for factors initially
  if(isFALSE(length(which(sapply(countData, is.factor))) == 0)){
    print("Error: inputs of type factor cannot be safely coerced to type integer.",
          "Please inspect input, and convert factors to characters before casting as integer")
    stop(
      paste("Error: inputs of type factor cannot be safely coerced to type integer.",
            "Please inspect input, and convert factors to characters before casting as integer")
  ### ---> the code is unchanged after this point
  if (tidy) {
    stopifnot(ncol(countData) > 1)
    rownms <- as.character(countData[,1])</pre>
    countData <- countData[,-1,drop=FALSE]</pre>
    rownames(countData) <- rownms</pre>
  }
  # check that these agree in number
  stopifnot(ncol(countData) == nrow(colData))
  # we expect a matrix of counts, which are non-negative integers
  countData <- as.matrix( countData )</pre>
  if (is(colData, "data.frame"))
    colData <- as(colData, "DataFrame")</pre>
  # check if the rownames of colData are simply in different order
  # than the colnames of the countData, if so throw an error
  # as the user probably should investigate what's wrong
  if (!is.null(rownames(colData)) & !is.null(colnames(countData))) {
    if (all(sort(rownames(colData)) == sort(colnames(countData)))) {
      if (!all(rownames(colData) == colnames(countData))) {
        stop(paste("rownames of the colData:
  ",paste(rownames(colData),collapse=","),"
  are not in the same order as the colnames of the countData:
  ",paste(colnames(countData),collapse=",")))
  }
  if (is.null(rownames(colData)) & !is.null(colnames(countData))) {
    rownames(colData) <- colnames(countData)</pre>
  se <- SummarizedExperiment(assays = SimpleList(counts=countData), colData = colData, ...)</pre>
```

```
object <- DESeqDataSet(se, design = design, ignoreRank)
    return(object)
}
This should prevent the error without interfering with the normal function of the program:
# do we catch the bad one?</pre>
```

```
# do we catch the bad one?
tryCatch(
 {
    # try to use the bad one
   dds <- DESeqDataSetFromMatrixDebugFactorSafe(countData = ctsBadFile,</pre>
                                                    colData = coldata, design = ~ condition)
 },
  error = function(e) { print(e) }
## Warning in print.default("Error: inputs of type factor cannot be safely coerced
## to type integer.", : NAs introduced by coercion
## <simpleError in print.default("Error: inputs of type factor cannot be safely coerced to type integer
dds <- DESeqDataSetFromMatrixDebugFactorSafe(countData = cts,</pre>
                                              colData = coldata, design = ~ condition)
# do we allow the good one?
DESeq(dds)
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
## class: DESeqDataSet
## dim: 14599 7
## metadata(1): version
## assays(4): counts mu H cooks
## rownames(14599): FBgn0000003 FBgn0000008 ... FBgn0261574 FBgn0261575
## rowData names(22): baseMean baseVar ... deviance maxCooks
## colnames(7): treated1 treated2 ... untreated3 untreated4
## colData names(3): condition type sizeFactor
```