

Operational modelling for outbreak response

When are genomic data useful?

Thibaut Jombart

31st October 2018

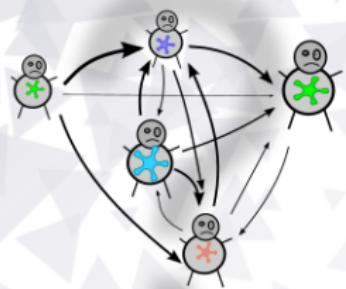
London School of Hygiene and Tropical Medicine
Imperial College London



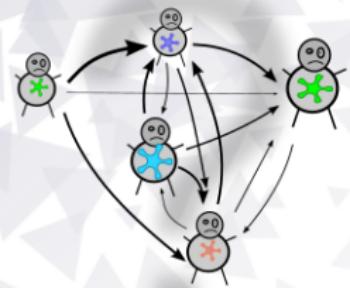
@teebzr

Reconstructing transmission trees

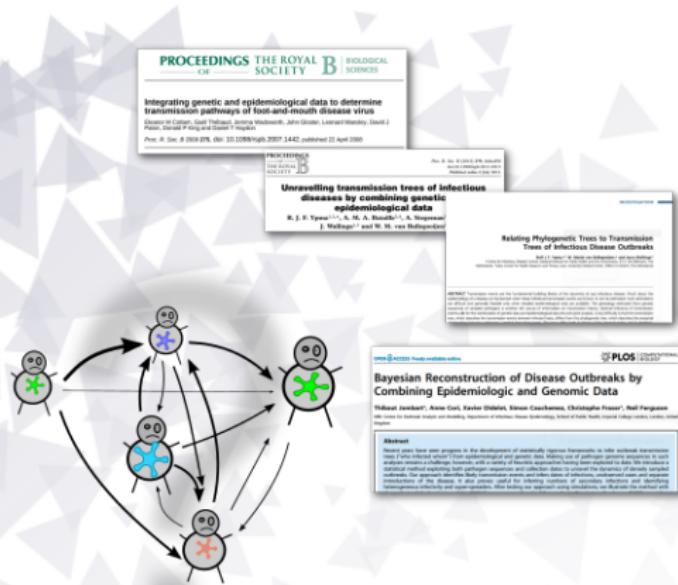
Who infects whom? Many answers for a single question



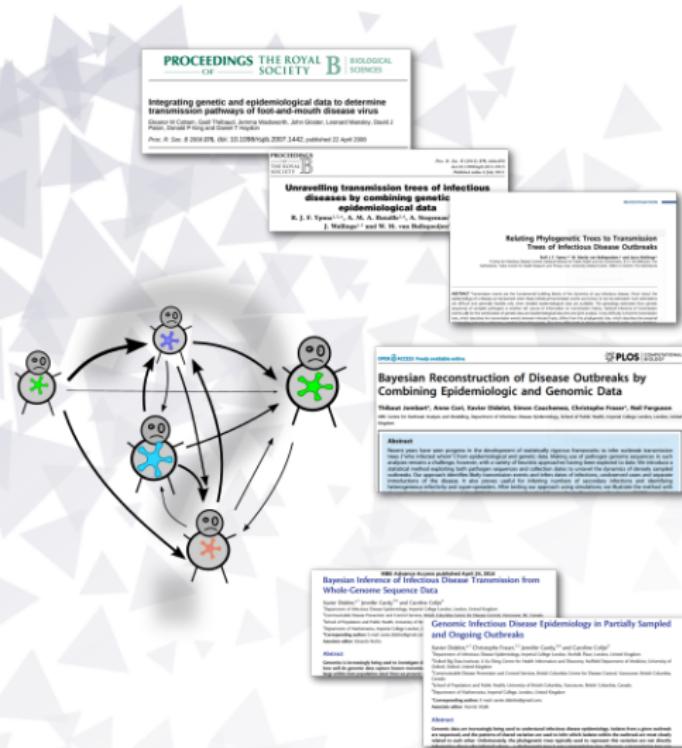
Who infects whom? Many answers for a single question



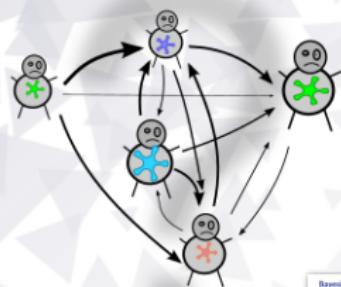
Who infects whom? Many answers for a single question



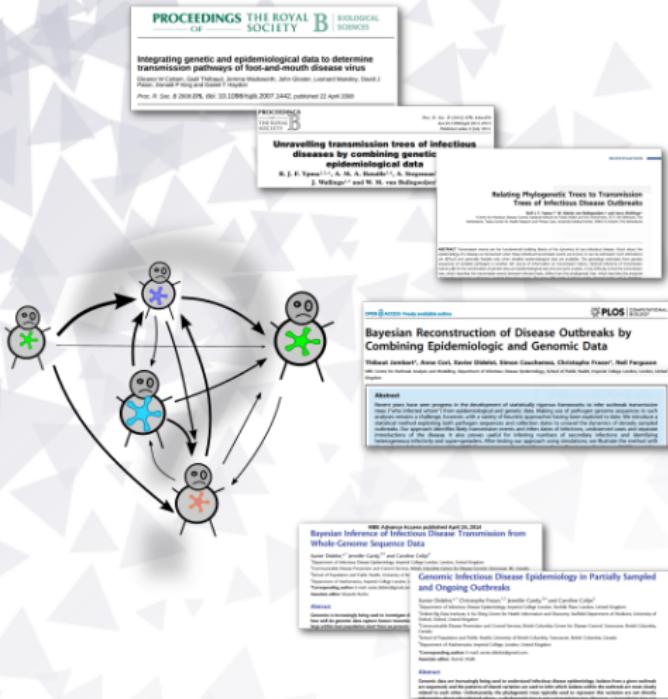
Who infects whom? Many answers for a single question



Who infects whom? Many answers for a single question



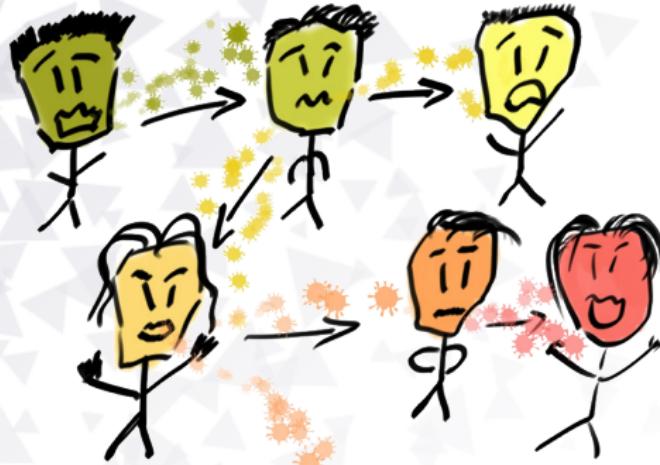
Who infects whom? Many answers for a single question



Methods heavily
rely on whole genome
sequence data

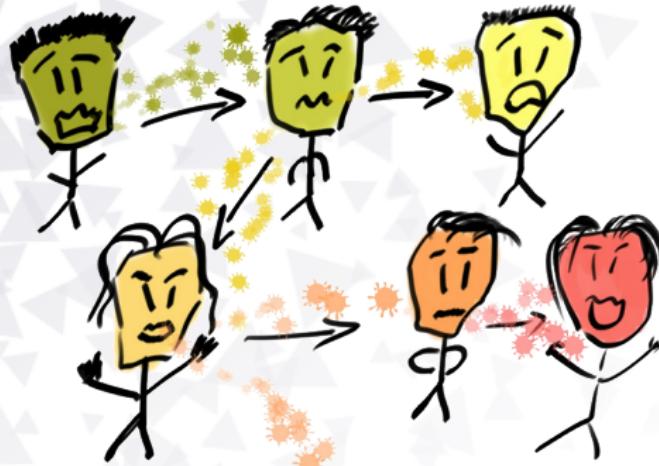


Using WGS to infer who infected whom



Mutations accumulate along transmission chains.

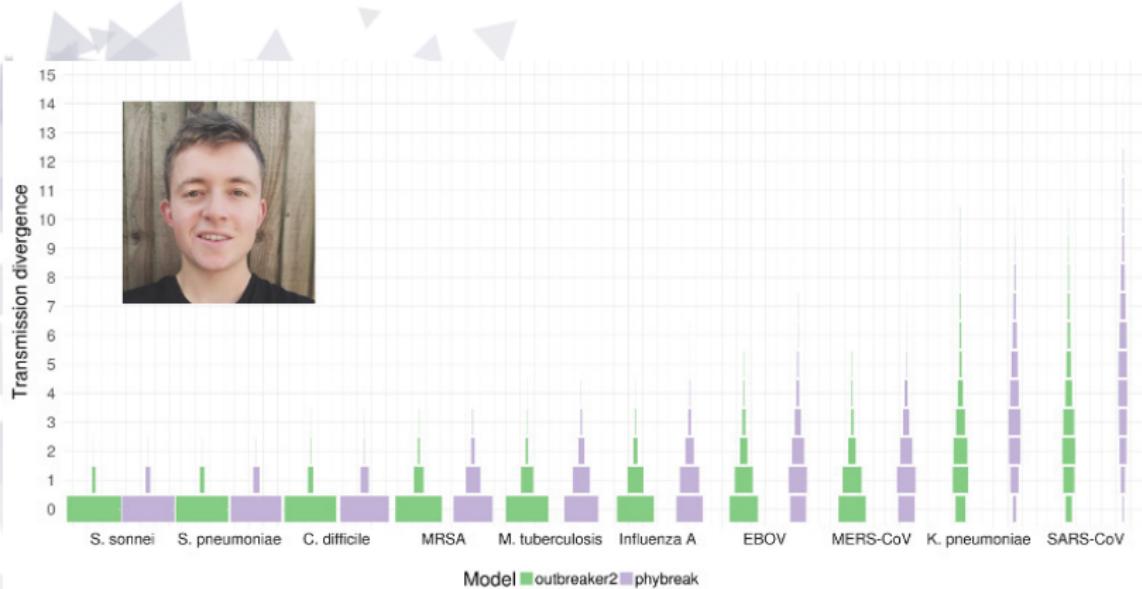
Using WGS to infer who infected whom



Mutations accumulate along transmission chains.

Can be used to reconstruct transmission trees.

How informative are whole genome sequences?



[Campbell et al. (2018) PLoS Computational Biology]

Insufficient diversity for most diseases.

Evidence synthesis approach to outbreak reconstruction



Combine different data to shrink the set of plausible trees.

outbreaker2: evidence synthesis framework for outbreak reconstruction

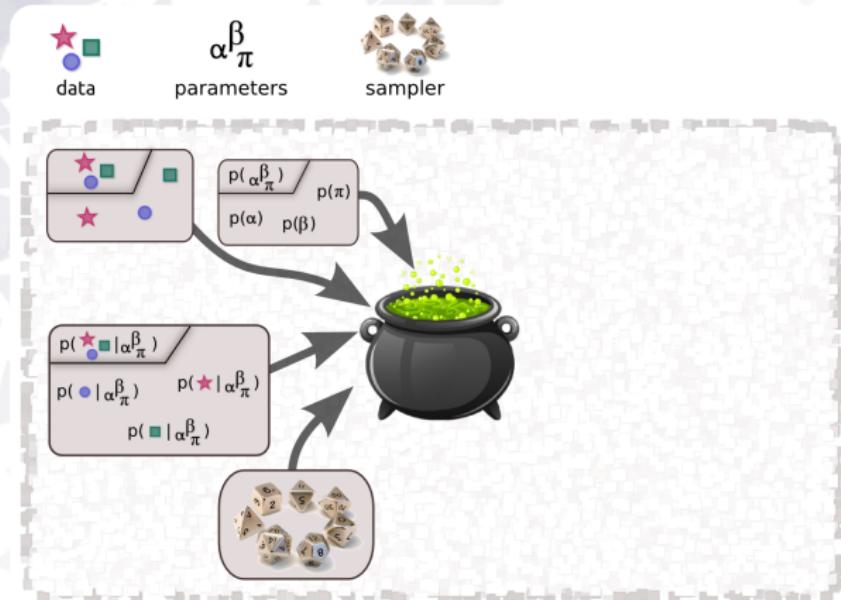
Modularity: customise data, prior, likelihood, MCMC.



[Campbell et al. (2018) BMC Bioinformatics]

outbreaker2: evidence synthesis framework for outbreak reconstruction

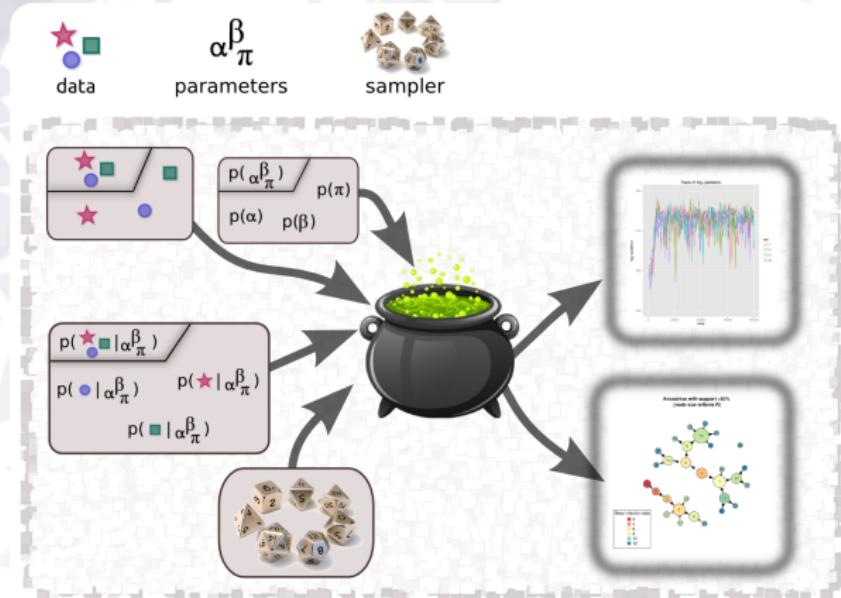
Modularity: customise data, prior, likelihood, MCMC.



[Campbell et al. (2018) BMC Bioinformatics]

outbreaker2: evidence synthesis framework for outbreak reconstruction

Modularity: customise data, prior, likelihood, MCMC.



[Campbell et al. (2018) BMC Bioinformatics]

Example: implementing *TransPhylo* in *outbreaker2*

outbreaker likelihood

- $p(s, t | \alpha, T^{inf}, \kappa, \mu, \pi) = p(T^{inf} | \alpha, \kappa) p(T^{inf} | t) p(s | \alpha, \kappa, \mu) p(\kappa | \pi)$
- i.e. *timing infection* \times *incubation* \times *genetic (simple)* \times *missing cases*

Example: implementing *TransPhylo* in *outbreaker2*

outbreaker likelihood

- $p(s, t | \alpha, T^{inf}, \kappa, \mu, \pi) = p(T^{inf} | \alpha, \kappa) p(T^{inf} | t) p(s | \alpha, \kappa, \mu) p(\kappa | \pi)$
- i.e. *timing infection* \times *incubation* \times *genetic (simple)* \times *missing cases*

TransPhylo likelihood

- $p(G | \beta, \gamma, N_{eg}, \alpha) = p(G | N_{eg}, \alpha) \times p(\alpha | \beta, \gamma)$
- i.e. *phylogeny (coalescent)* \times *SIR*

Example: implementing *TransPhylo* in *outbreaker2*

outbreaker likelihood

- $p(s, t | \alpha, T^{inf}, \kappa, \mu, \pi) = p(T^{inf} | \alpha, \kappa) p(T^{inf} | t) p(s | \alpha, \kappa, \mu) p(\kappa | \pi)$
- i.e. *timing infection* \times *incubation* \times *genetic (simple)* \times *missing cases*

TransPhylo likelihood

- $p(G | \beta, \gamma, N_{eg}, \alpha) = p(G | N_{eg}, \alpha) \times p(\alpha | \beta, \gamma)$
- i.e. *phylogeny (coalescent)* \times *SIR*

Can we combine the two models?

TransPhylo module for *outbreaker2*

3 Custom likelihood

In order to calculate the likelihood under the TransPhylo model, we need to (i) extract the transmission tree from the outbreaker2 parameter, (ii) combine this transmission tree with the phylogenetic tree to form a colored tree, and (iii) calculate the likelihood of this colored tree. Step (i) is easy since transmission tree are encoded almost in the same way in TransPhylo and outbreaker2. For step (ii) we have to write the `combine` function which is tedious but not especially interesting (this function is included in this Rnw file but its code is not shown in the pdf). For step (iii) we only need to call the appropriate function of the TransPhylo package which is `probPTreeGivenTree`. During step (ii) messages can arise indicating that the transmission tree and phylogenetic tree are in fact incompatible, in which case the likelihood is returned as -Inf.

```
lik_TransPhylo <- function(data, param) {
  ttree <- list(ttree = chind(param$t.inf, data$dates, param$alpha),
               nam = data$ptree$nam)
  ttrees$tree(which(is.na(ttrees$ttree[, 3])), 3) <- 0
  txt <- capture.output(ttree <- combine(ttrees, data$ptree))
  if (length(txt) == 0) {
    prob <- probPTreeGivenTree(ttree, neg = 366 * 0.26)
  } else {
    prob <- -Inf
  }
  return(prob)
}

lik_transphylo <- function(date, param, i = NULL, custom_functions = NULL)
{
  ## function (date, param, i = NULL, custom_functions = NULL)

  new_move_tinf <- function(param, data, list_custom_ll = new_model) {
    for (i in 1:date$N) {
      current_ll <- api$cpp_ll_all(data, param, i = NULL, list_custom_ll)
      modif <- sample(c(-100:-1, 1:100), 1)
      param$ll.inf[i] <- param$ll.inf[i] + modif
      new_ll <- api$cpp_ll_all(data, param, i = NULL, list_custom_ll)
      if (log10(modif[i]) > log10(ll.current_ll)) {
        param$ll.inf[i] <- param$ll.inf[i] + modif
      }
    }
    return(param)
  }

  new_moves <- custom_moves(t.inf = new_move_tinf)
  new_moves

  ##
  ## //////////////////////////////////////////////////////////////////
  ## class: outbreaker_moves list
  ## number of items: 8
  ## // movement functions //
  ## See
}
```

[Campbell et al. (2018) BMC Bioinformatics]

TransPhylo module for *outbreaker2*

3 Custom likelihood

In order to calculate the likelihood under the TransPhylo model, we need to (i) extract the transmission tree from the outbreaker2 parameter, (ii) combine this transmission tree with the phylogenetic tree to form a colored tree, and (iii) calculate the likelihood of this colored tree. Step (i) is easy since transmission tree are encoded almost in the same way in TransPhylo and outbreaker2. For step (ii) we have to write the `combine` function which is tedious but not especially interesting (this function is included in this Rnw file but its code is not shown in the pdf). For step (iii) we only need to call the appropriate function of the TransPhylo package which is `probPTreeGivenTree`. During step (ii) messages can arise indicating that the transmission tree and phylogenetic tree are in fact incompatible, in which case the likelihood is returned as -Inf.

```
lik_TransPhylo <- function(data, param) {
  ttree <- list(ttree = chind(param$t.inf, data$dates, param$alpha),
               nam = data$pstree$nam)
  ttrees$tree(which(is.na(ttrees$ttree[, 3])), 3) <- 0
  txt <- capture.output(ttree <- combine(ttree, data$pstree))
  if (length(txt) == 0) {
    prob <- probPTreeGivenTree(ttree, neg = 366 * 0.26)
  } else {
    prob <- -Inf
  }
  return(prob)
}

## Function to calculate the likelihood
## Function (date, param, i = NULL, custom_functions = NULL)
## Function (date, param, i = NULL, list_custom_ll = new_model) {
##   for (i in 1:date$ll) {
##     current_ll <- api$pp_ll_all(data, param, i = NULL, list_custom_ll)
##     modif <- sample(c(-100:-1, 1:100), 1)
##     param$ll.inf[i] <- param$ll.inf[i] + modif
##     new_ll <- api$pp_ll_all(data, param, i = NULL, list_custom_ll)
##     if (log10(modif[i]) > log10(ll - current_ll)) {
##       param$ll.inf[i] <- param$ll.inf[i] + modif
##     }
##   }
##   return(param)
## }

new_moves_tinf <- function(param, data, list_custom_ll = new_model) {
  for (i in 1:ll) {
    current_ll <- api$pp_ll_all(data, param, i = NULL, list_custom_ll)
    modif <- sample(c(-100:-1, 1:100), 1)
    param$ll.inf[i] <- param$ll.inf[i] + modif
    new_ll <- api$pp_ll_all(data, param, i = NULL, list_custom_ll)
    if (log10(modif[i]) > log10(new_ll - current_ll)) {
      param$ll.inf[i] <- param$ll.inf[i] + modif
    }
  }
  return(param)
}

new_moves <- custom_moves(t.inf = new_moves_tinf)
new_moves

## 
## ////////////// outbreaker movement functions ///
## 
## class: outbreaker_moves list
## number of items: 8
## 
## // movement functions //
## 
## 
```

likelihood

[Campbell et al. (2018) BMC Bioinformatics]

TransPhylo module for *outbreaker2*

3 Custom likelihood

In order to calculate the likelihood under the TransPhylo model, we need to (i) extract the transmission tree from the outbreaker2 parameter, (ii) combine this transmission tree with the phylogenetic tree to form a colored tree, and (iii) calculate the likelihood of this colored tree. Step (i) is easy since transmission tree are encoded almost in the same way in TransPhylo and outbreaker2. For step (ii) we have to write the `combine` function which is tedious but not especially interesting (this function is included in this Rnw file but its code is not shown in the pdf). For step (iii) we only need to call the appropriate function of the TransPhylo package which is `probPTreeGivenTree`. During step (ii) messages can arise indicating that the transmission tree and phylogenetic tree are in fact incompatible, in which case the likelihood is returned as -Inf.

```
lik_TransPhylo <- function(data, param) {
  ttree <- list(ttree = chind(param$t.inf, data$dates, param$alpha),
               nam = data$ptree$nam)
  ttree$tree$which(is.na(ttree$tree[,3]))[,3] <- 0
  txt <- capture.output(ctree <- combine(ttree, data$ptree))
  if (length(txt)==0) {
    prob <- probPTreeGivenTree(ctree, neg = 366 * 0.26)
  } else {
    prob <- -Inf
  }
  return(prob)
}
```

likelihood

```
args(api$cpp_ll_all)
## function (date, param, i = NULL, custom_functions = NULL)
## NULL

new_move_tinf <- function(param, data, list_custom_ll = new_model) {
  for (i in 1:date$N) {
    current_ll <- api$cpp_ll_all(data, param, i = NULL, list_custom_ll)
    modif <- sample(c(-100:-1, 1:100), 1)
    param$inf[i] <- param$inf[i] + modif
    new_ll <- api$cpp_ll_all(data, param, i = NULL, list_custom_ll)
    if (log(modif[i]) > log(ll(current_ll))) {
      param$inf[i] <- param$inf[i] + modif
    }
  }
  return(param)
}

new_moves <- custom_moves(t_inf = new_move_tinf)
new_moves

##
## //////////////////////////////////////////////////////////////////
## class: outbreaker_moves list
## number of items: 8
## //////////////////////////////////////////////////////////////////
## movement functions //
## See
```

[Campbell et al. (2018) BMC Bioinformatics]

TransPhylo module for *outbreaker2*

3 Custom likelihood

In order to calculate the likelihood under the TransPhylo model, we need to (i) extract the transmission tree from the outbreaker2 parameter, (ii) combine this transmission tree with the phylogenetic tree to form a colored tree, and (iii) calculate the likelihood of this colored tree. Step (i) is easy since transmission tree are encoded almost in the same way in TransPhylo and outbreaker2. For step (ii) we have to write the `combine` function which is tedious but not especially interesting (this function is included in this Rnw file but its code is not shown in the pdf). For step (iii) we only need to call the appropriate function of the TransPhylo package which is `probPTreeGivenTree`. During step (ii) messages can arise indicating that the transmission tree and phylogenetic tree are in fact incompatible, in which case the likelihood is returned as -Inf.

```
lik_TransPhylo <- function(data, param) {
  ttree <- list(ttree = chind(param$t.inf, data$dates, param$alpha),
               nam = data$ptree$nam)
  ttrees$tree(which(is.na(ttrees$ttree[, 3])), 3) <- 0
  txt <- capture.output(ctree <- combine(ttrees, data$ptree))
  if (length(txt) == 0) {
    prob <- probPTreeGivenTree(ctree, neg = 366 * 0.26)
  } else {
    prob <- -Inf
  }
  return(prob)
}
```

likelihood

Total: 25 lines of R

outbreaker2: 7,500 lines of R/C++

Code difference: 0.3%

movement function

```
args(api$cpp_ll_all)
## function (date, param, i = NULL, custom_functions = NULL)
## NULL

new_move_tinf <- function(param, data, list_custom_ll = new_model) {
  for (i in 1:date$N) {
    current_ll <- api$cpp_ll_all(data, param, i = NULL, list_custom_ll)
    modif <- sample(c(-100:-1, 1:100), 1)
    param$inf[i] <- param$inf[i] + modif
    new_ll <- api$cpp_ll_all(data, param, i = NULL, list_custom_ll)
    if (log(new_ll) > log(current_ll)) {
      param$inf[i] <- param$inf[i] + modif
    }
  }
  return(param)
}

new_moves <- custom_moves(t_inf = new_move_tinf)
new_moves

## 
## ///////////////////////////////////////////////////////////////////
## class: outbreaker_moves list
## number of items: 8
## // movement functions //
## @na
```

[Campbell et al. (2018) BMC Bioinformatics]

TransPhylo module for *outbreaker2*

3 Custom likelihood

In order to calculate the likelihood under the TransPhylo model, we need to (i) extract the transmission tree from the *outbreaker2* parameter, (ii) combine this transmission tree with the phylogenetic tree to form a colored tree, and (iii) calculate the likelihood of this colored tree. Step (i) is easy since transmission tree are encoded as lists in *outbreaker2* and phylogenetic trees are graphs. Step (ii) we have to write a graph function to do this. Step (iii) is especially interesting (this function is in *outbreaker2*) because it is a recursive function that needs to check if there are any loops in the tree. If there are loops, then the likelihood is zero. If there are no loops, then we just need to call the appropriate function in *outbreaker2*. In this case, the likelihood is zero. If there are loops, then (iii) messages can arise indicating that the transmission tree and phylogenetic tree are in fact in loops. In this case the likelihood is returned as -Inf.

```
lik_TransPhylo <- function(data, param) {
  ttree <- list(ttree = chid(param$inf, data$data, param$alpha),
               ann = data$tree$ann)
  ttrees@tree[which(is.na(ttrees@tree[,3]))] <- 0
  ttrees@tree <- capture.output(ttree <- combine(ttrees,data$ptree))
  if (length(ttree) == 0) {
    prob <- 1 - exp(-param$lambda * (param$beta - 0.25) * 0.25)
  } else {
    prob <- 1
  }
  return(prob)
}
```

likelihood

Total: 25 lines of R

outbreaker2: 7,500 lines of R/C++

Code difference: 0.3%

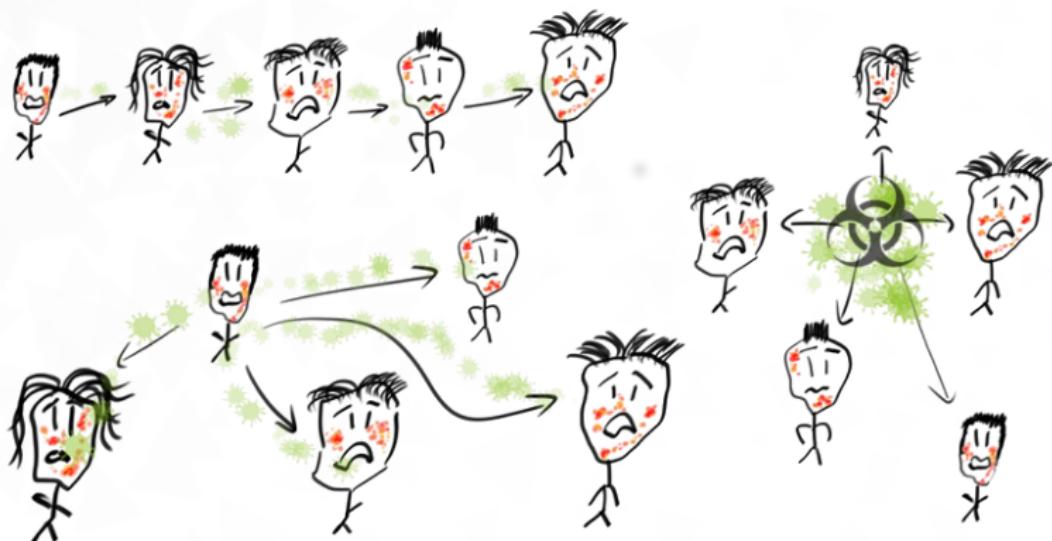
[Campbell et al. (2018) BMC Bioinformatics]



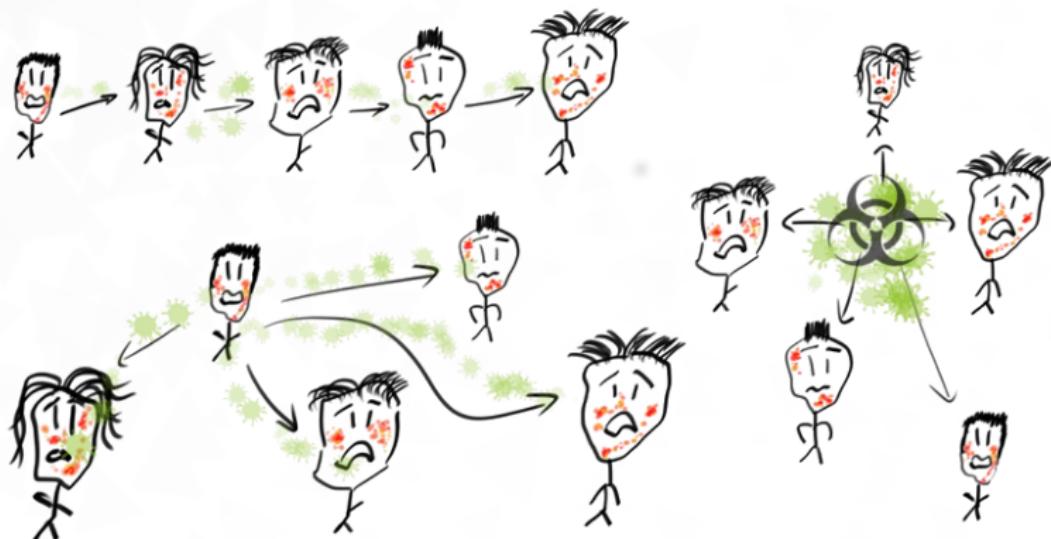
The background of the slide features a large number of small, light-gray triangles of various sizes scattered across the white surface, creating a subtle geometric pattern.

Identifying transmission clusters

Sustained transmission vs repeated introductions



Sustained transmission vs repeated introductions



Different transmission patterns call for different interventions

One outbreak.. how many introductions?



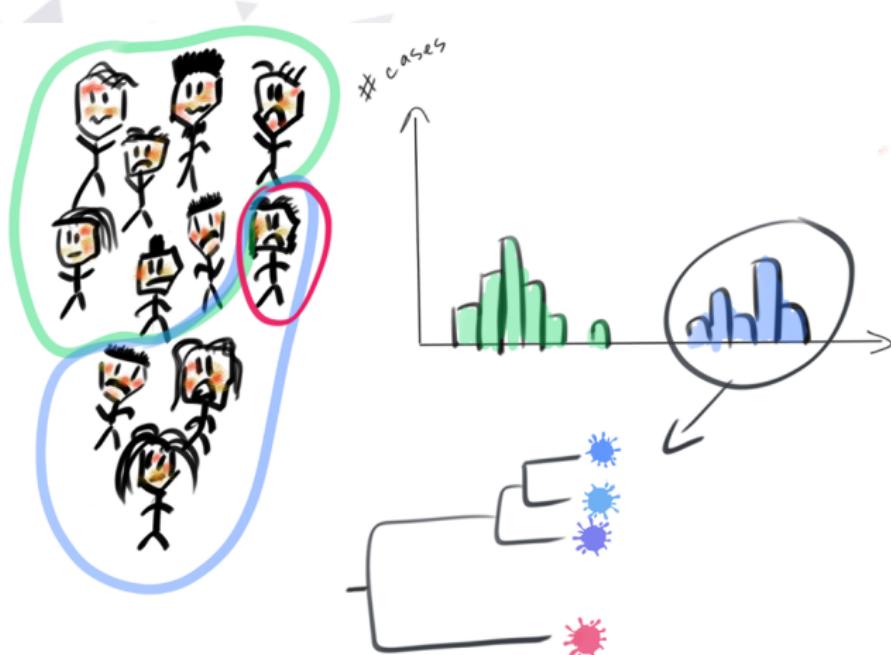
One outbreak.. how many introductions?



One outbreak.. how many introductions?



One outbreak.. how many introductions?



Combined data sources can detect **outbreak clusters**

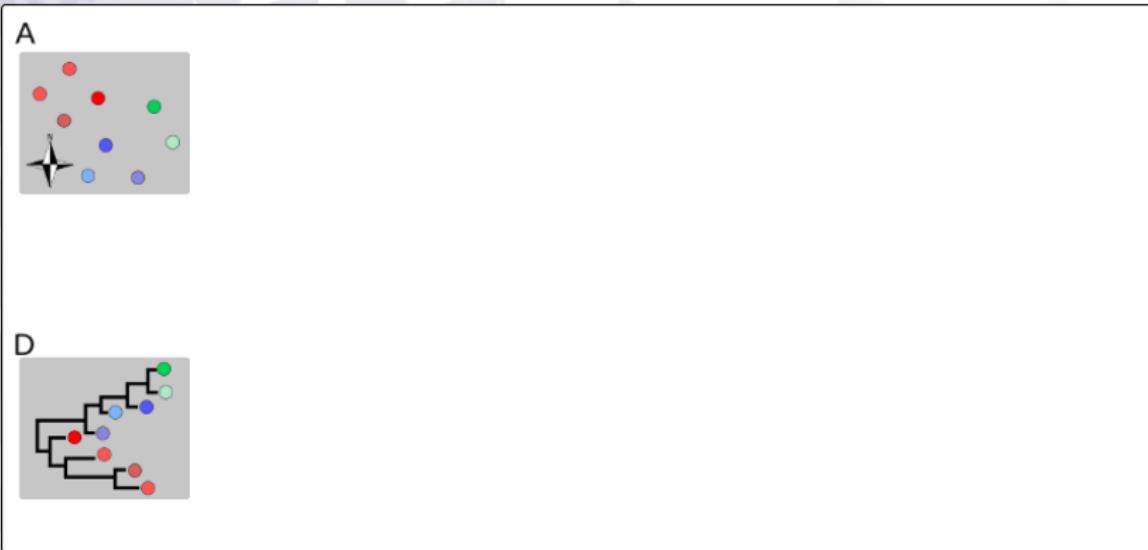


Aims: develop a new method which...

- **detects outbreak clusters:** cases stemming from same introduction (same transmission tree)
- **integrates different data:** temporal, spatial, genetic, etc.
- **works fast, scales well:** so that it can be used for real-time outbreak detection

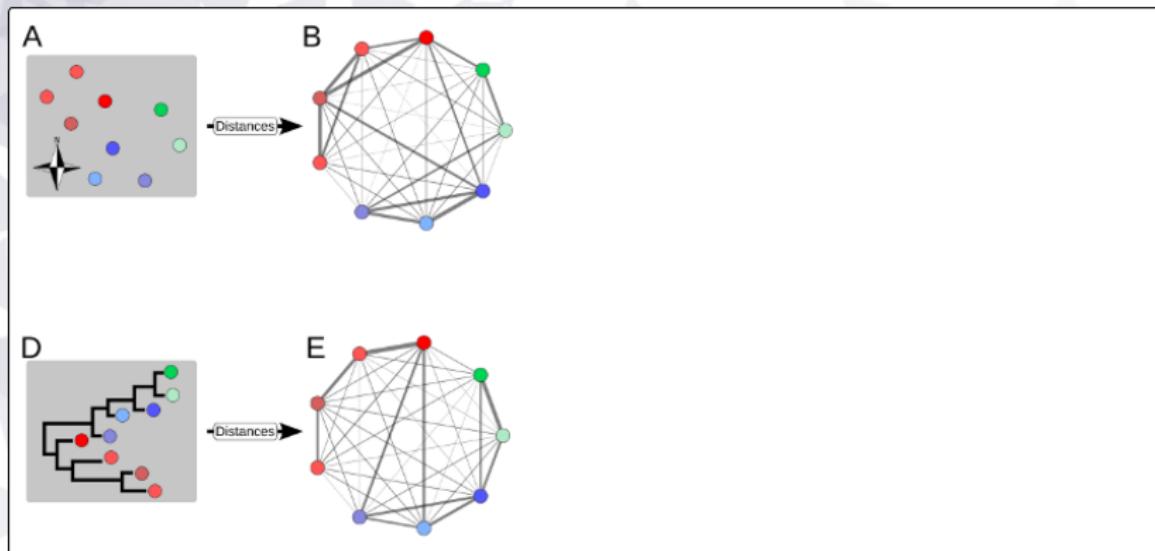
¹well, really, I made that up because I was reading 'Snuff' by Terry Pratchett at the time; incidentally, Pratchett was a huge fan of using long footnotes in his novels, which are often quite entertaining to read; well, this does not apply here: if you are still reading this, you probably missed what I just said

A graph-based evidence synthesis approach



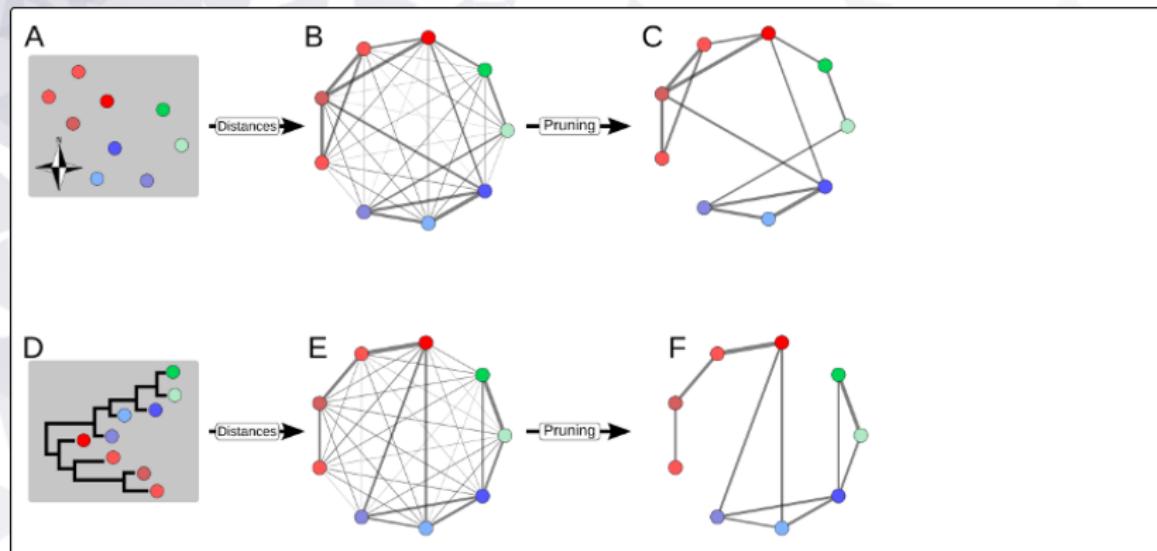
[Cori et al. (2018) PLoS Computational Biology]

A graph-based evidence synthesis approach



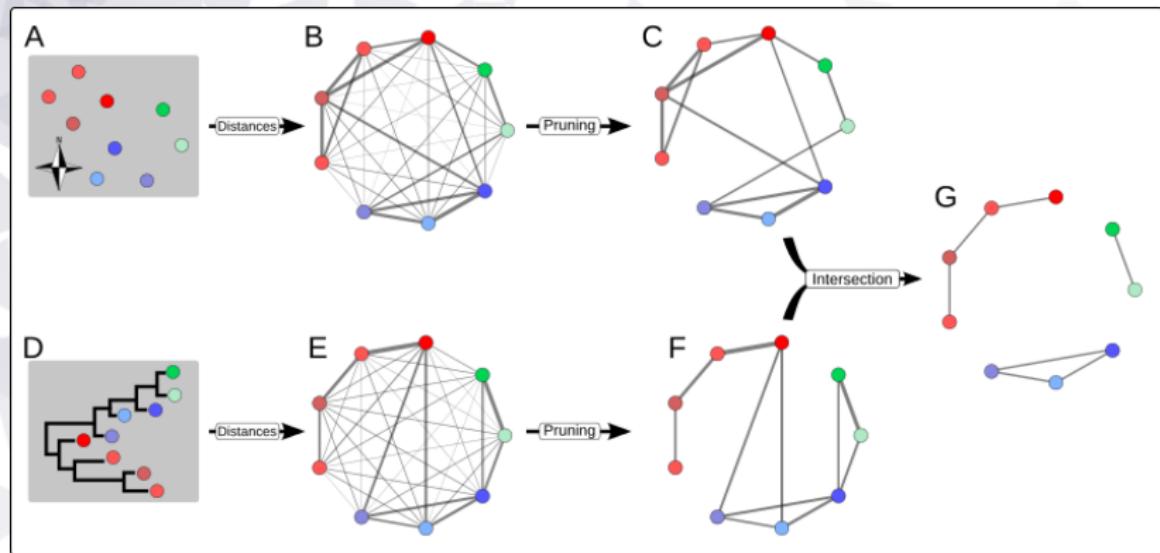
[Cori et al. (2018) PLoS Computational Biology]

A graph-based evidence synthesis approach



[Cori et al. (2018) PLoS Computational Biology]

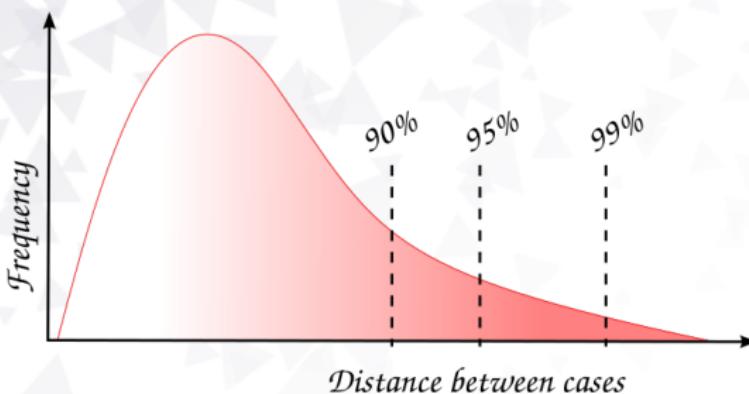
A graph-based evidence synthesis approach



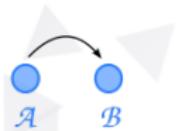
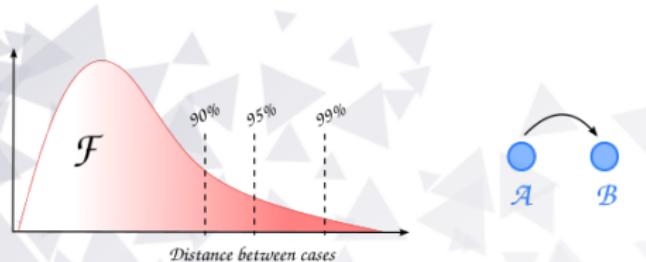
[Cori et al. (2018) PLoS Computational Biology]

Pruning graphs: where to cut?

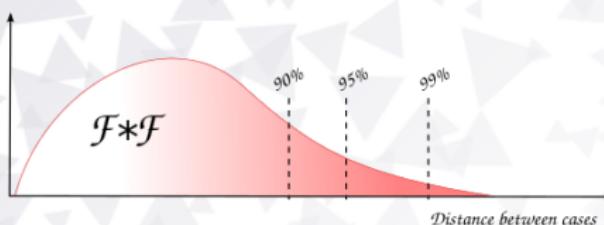
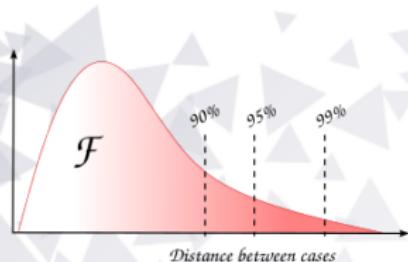
Assuming a known expected distribution between pairs of cases (e.g. serial interval, spatial kernel, molecular clock), different quantiles can be used:



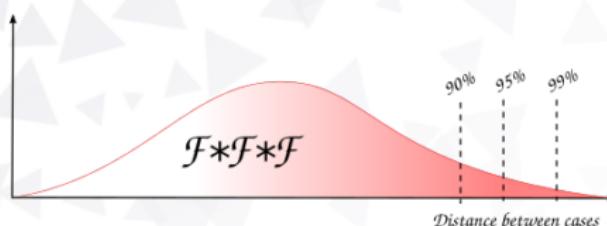
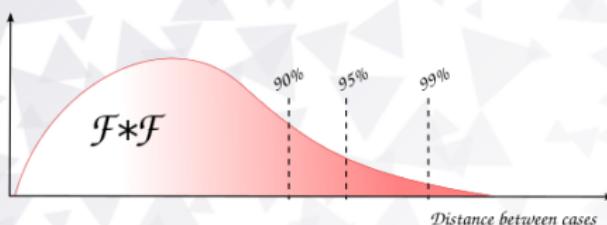
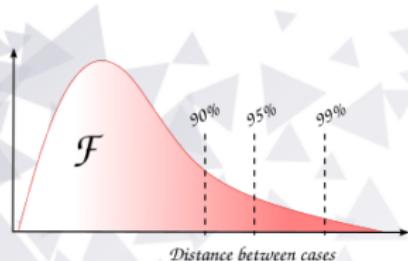
Pruning graphs: where to cut?



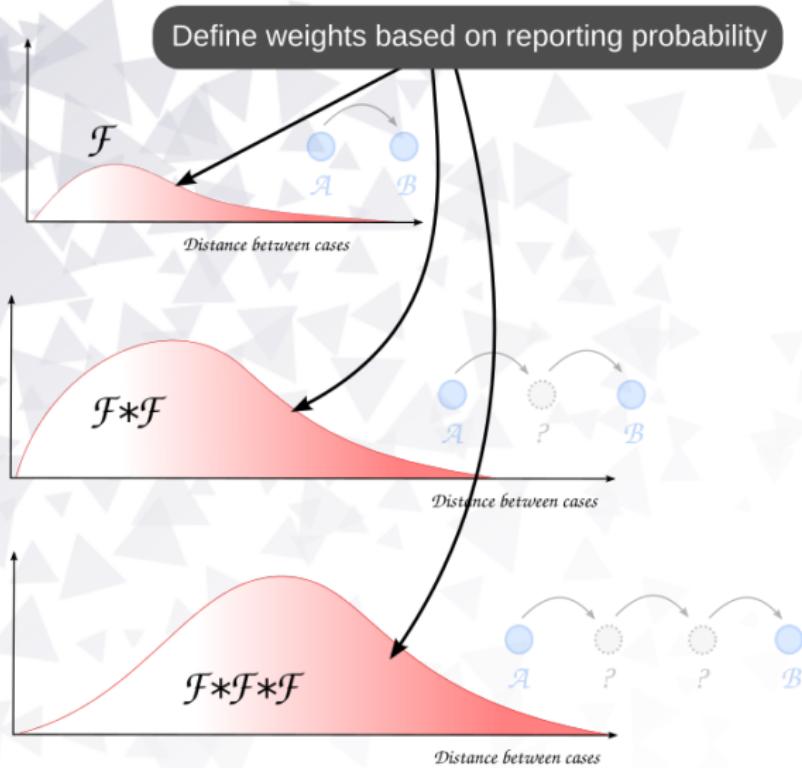
Pruning graphs: where to cut?



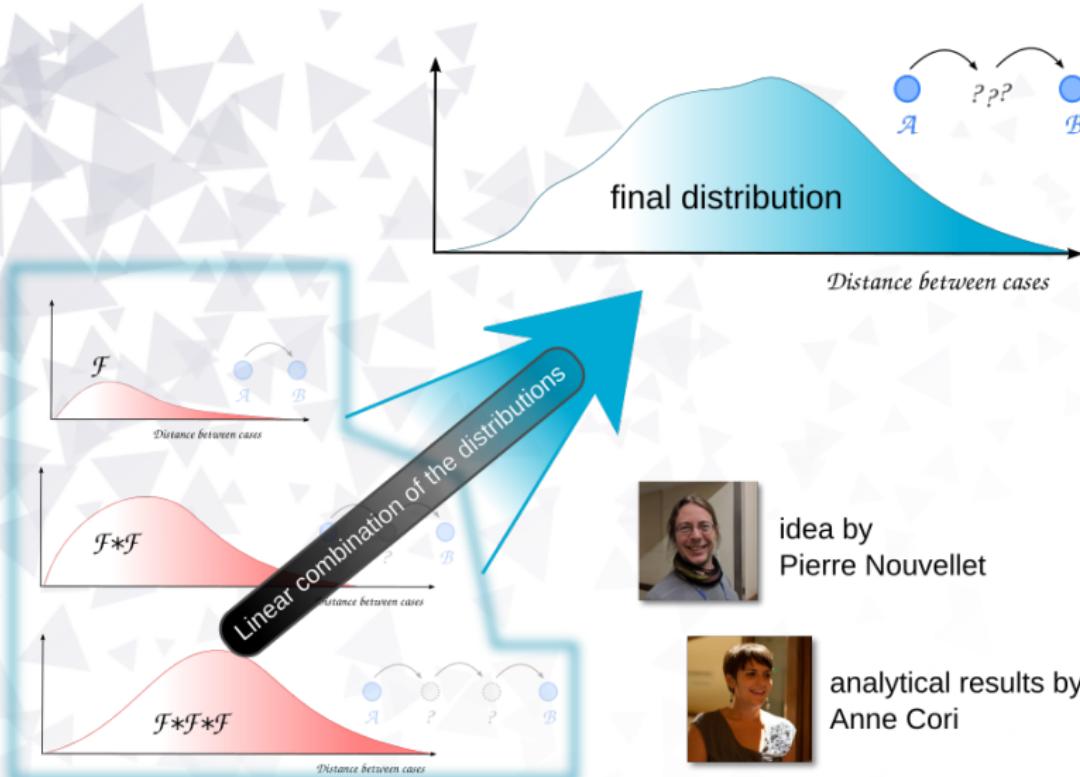
Pruning graphs: where to cut?



Pruning graphs: where to cut?



Pruning graphs: where to cut?



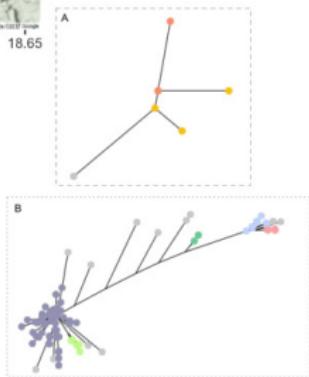
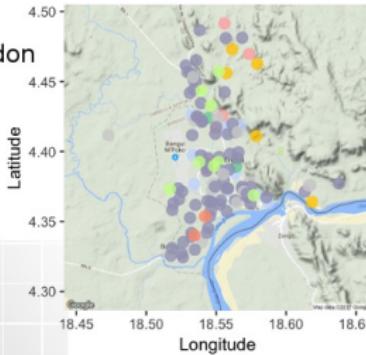
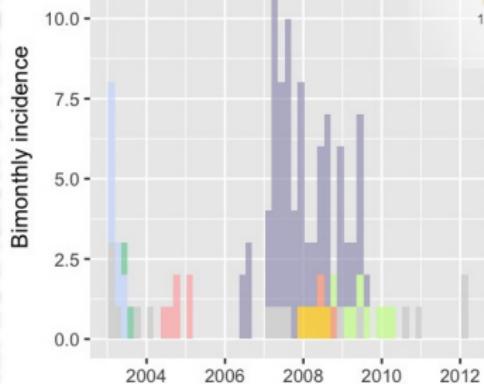
Application: dog rabies epidemics, Central African Republic



Anne Cori
Imperial College London

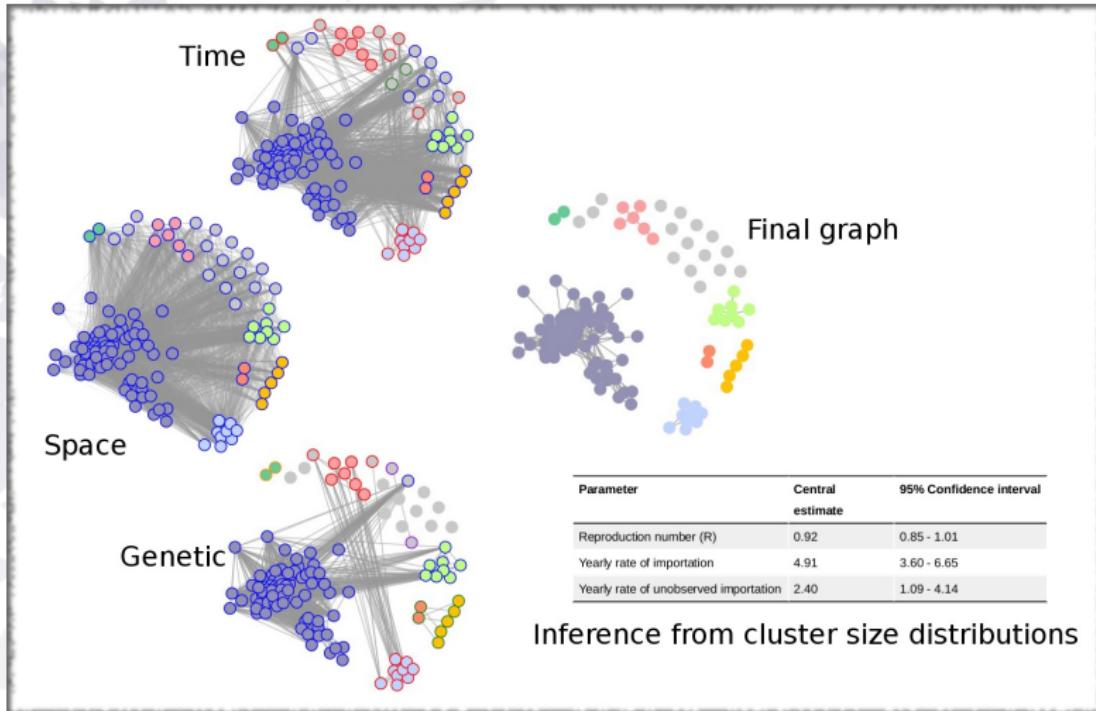


Pierre Nouvellet
University of Sussex

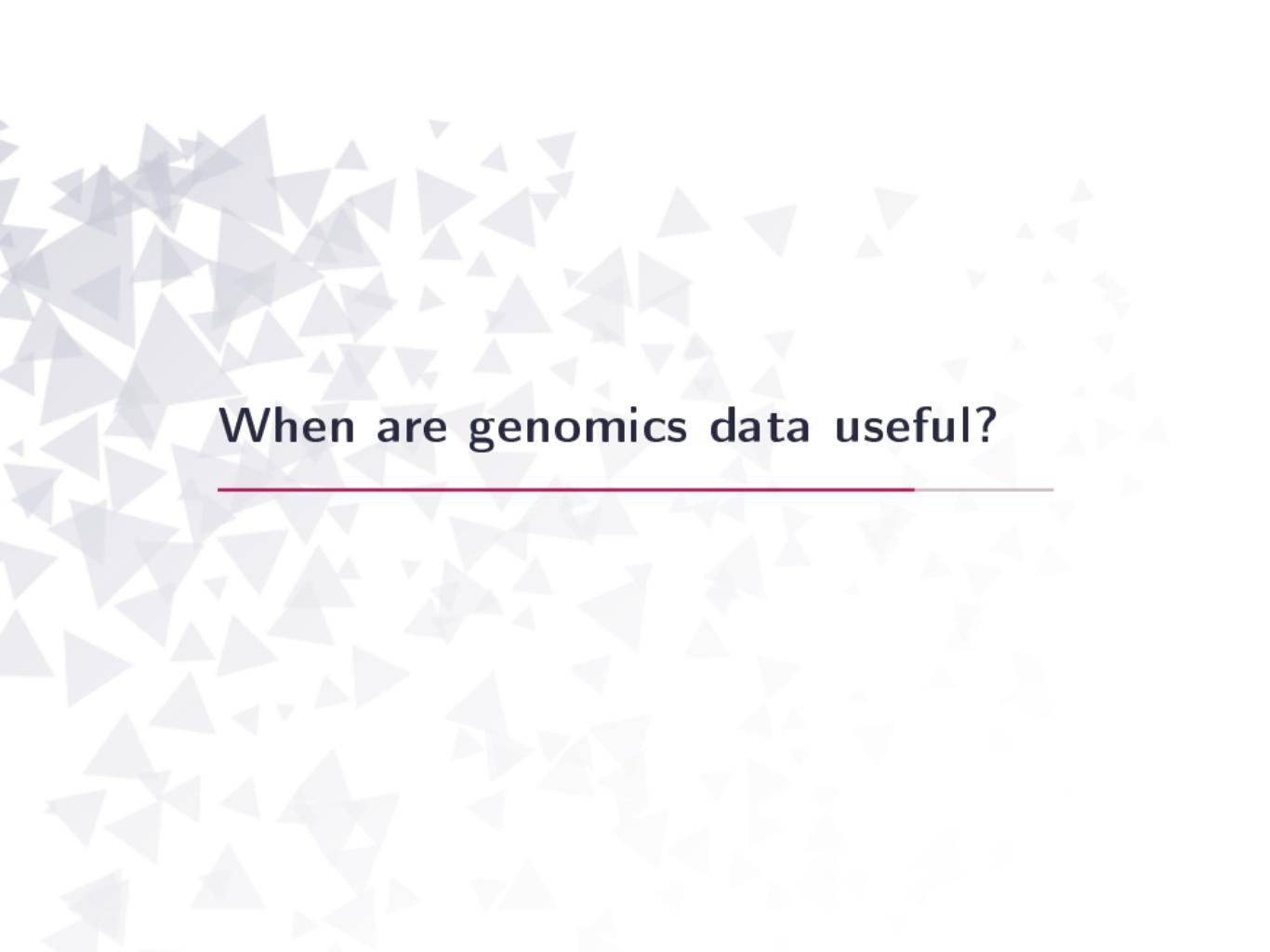


[Cori et al. (2018) PLoS Computational Biology]

Results

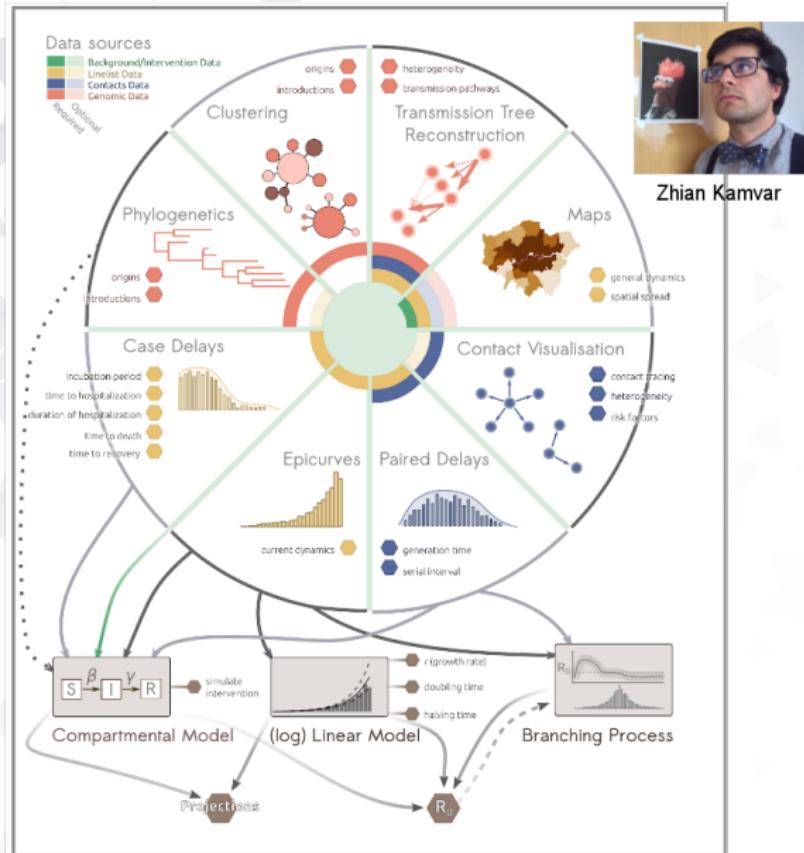


[Cori et al. (2018) PLoS Computational Biology]

The background of the slide features a large number of small, light-gray triangles of various sizes scattered across the frame, creating a subtle geometric pattern.

When are genomics data useful?

The outbreak analytics toolbox



Pathogen genetics in outbreak response



- in general, not needed for forecasting or control

Pathogen genetics in outbreak response



- in general, not needed for forecasting or control
- useful to detect multiple introductions or superspreading

Pathogen genetics in outbreak response



- in general, not needed for forecasting or control
- useful to detect multiple introductions or superspreading
- can complement contact tracing data

Pathogen genetics in outbreak response



- in general, not needed for forecasting or control
- useful to detect multiple introductions or superspreading
- can complement contact tracing data
- WGS are costly: is it worth it?

Thanks to:

- **Session:** John Edmunds
- **Collaborators:** Finlay Campbell, Anne Cori, Pierre Nouvellet, Zhian Kamvar, Amrish Baidjoe, Roz Eggo, Tini Garske, Hervé Bourhy, Emmanuel Nakouné, Jimmy Whitworth, Dan Bausch, Bayard Roberts, Neil Ferguson
- **Groups:** R Epidemics Consortium
- **Funding:** GCRF project RECAP (ES/P010873/1), UK PH RST, HPRU-NIHR, MRC

RECON