The $E.\ coli$ molecular phenotype under different growth conditions

Supplementary materials

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#### Supplementary Figures

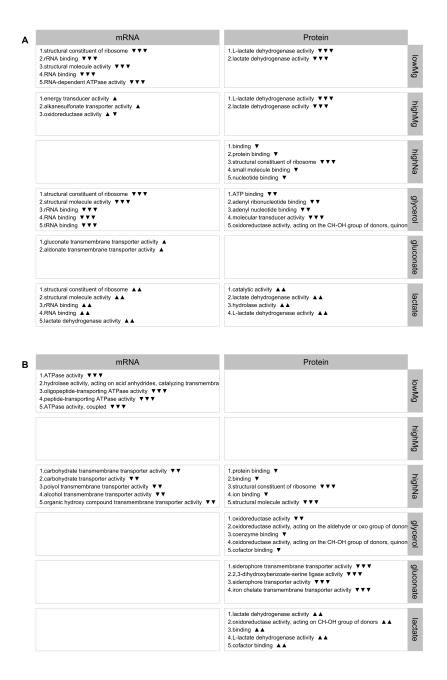


Figure S1: Significantly differentially expressed molecular functions, as determined by GO annotations. For each condition, we show the top-5 differentially expressed molecular functions according to either mRNA or protein abundances. Empty boxes indicate that no differentially expressed pathways were found. The arrows next to pathway names indicate the proportion of up- and down-regulated genes among the significantly differentially expressed genes in this pathway. One up arrow indicates that 60% or more of the genes are up-regulated, two arrows correspond to 80% or more genes, and three arrows correspond to 95% or more genes being up-regulated. Similarly, down arrows indicate the proportion of down-regulated genes. (A) Exponential phase. (B) Stationary phase.

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Figure S2: Significantly differentially expressed KEGG pathways and associated genes with glycerol as carbon source, as determined by mRNA abundances in exponential phase. The top 2 differentially expressed KEGG pathways are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed pathways and for each pathway we show up to 15 of the most significantly changing genes.

Log2 Fold Change

-3

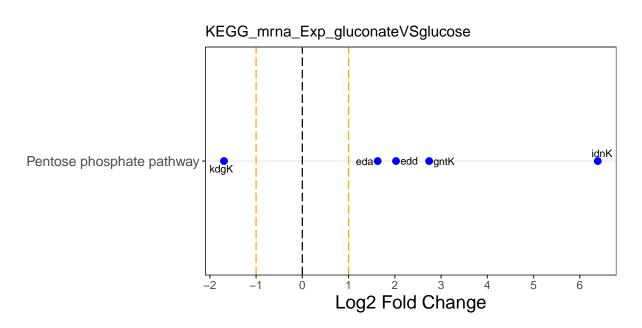


Figure S3: Significantly differentially expressed KEGG pathway and associated genes with gluconate as carbon source, as determined by mRNA abundances in exponential phase. The top differentially expressed KEGG pathway is shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed pathways and for each pathway we show up to 15 of the most significantly changing genes.

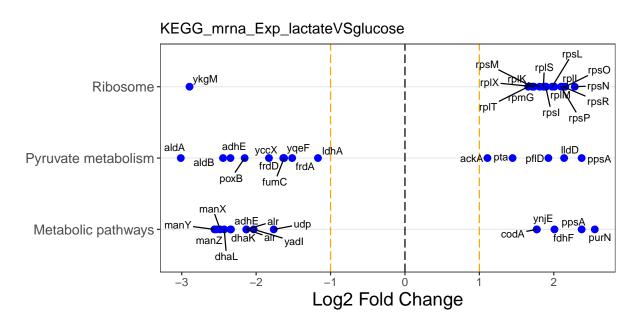


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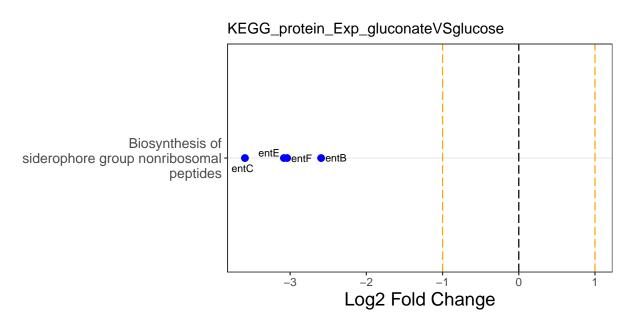


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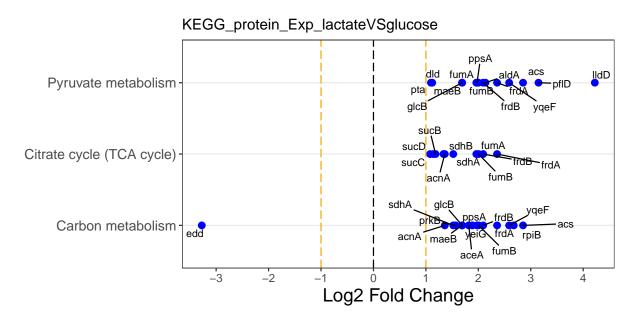


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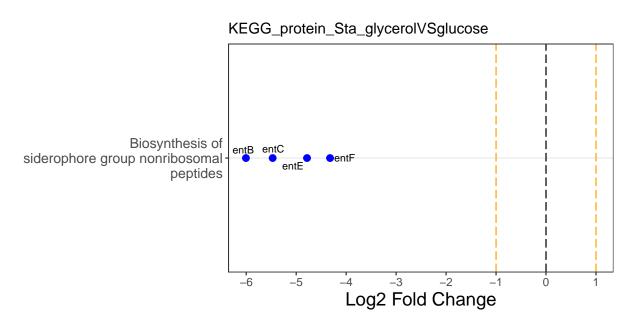


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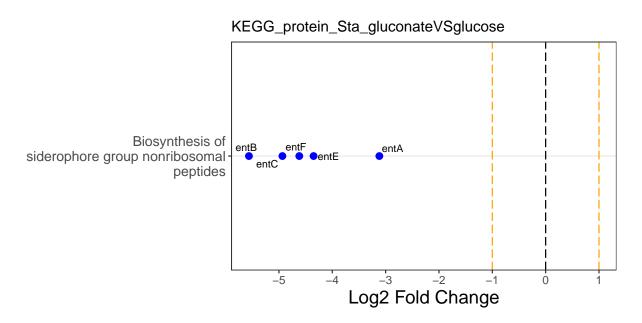


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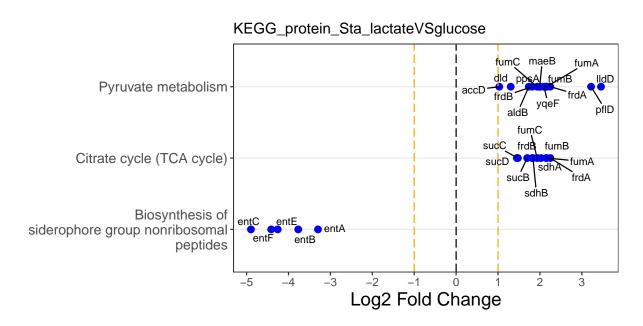


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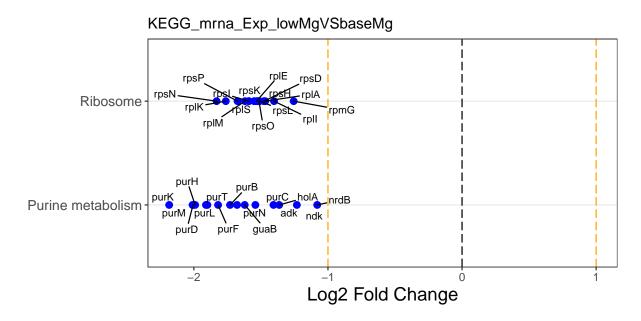


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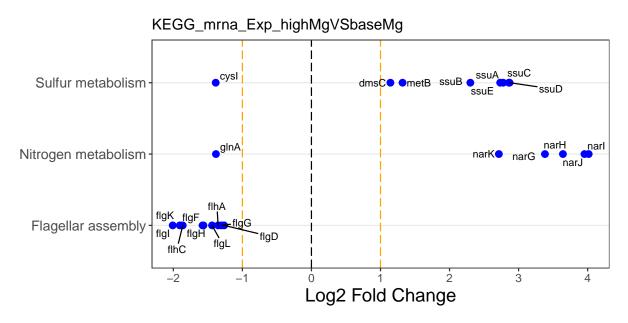


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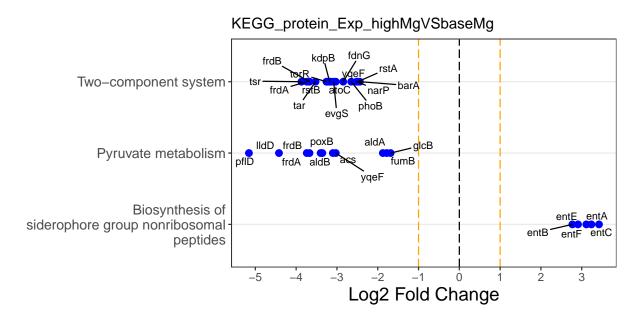


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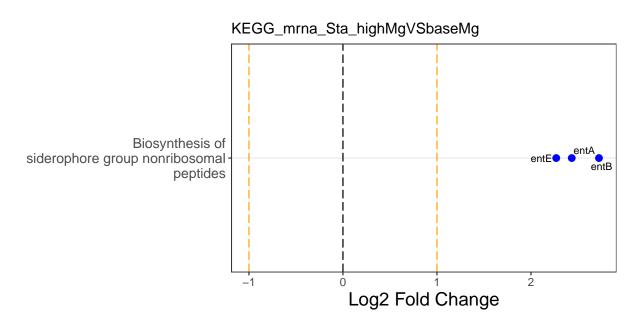


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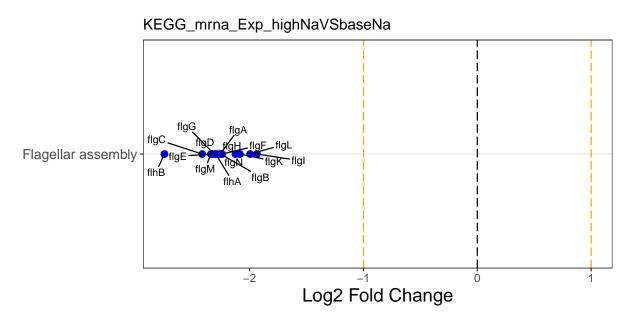


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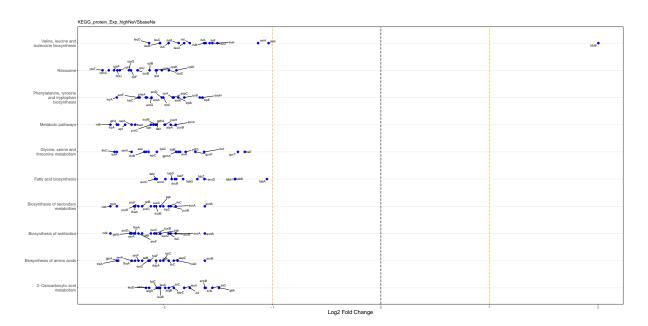


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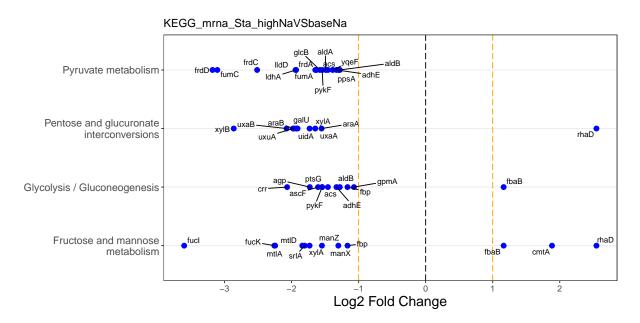


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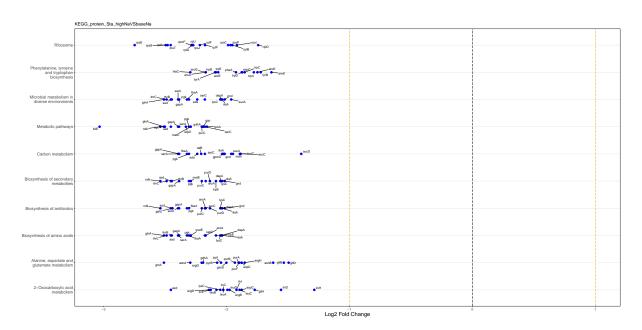


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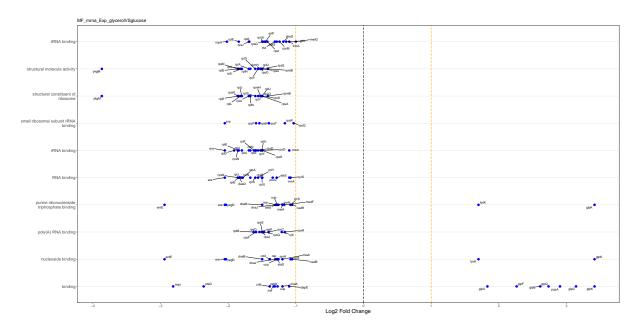


Figure S18: Significantly differentially expressed GO annotations related with molecular functions and associated genes with glycerol as carbon source, as determined by mRNA abundances in exponential phase. The top 10 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

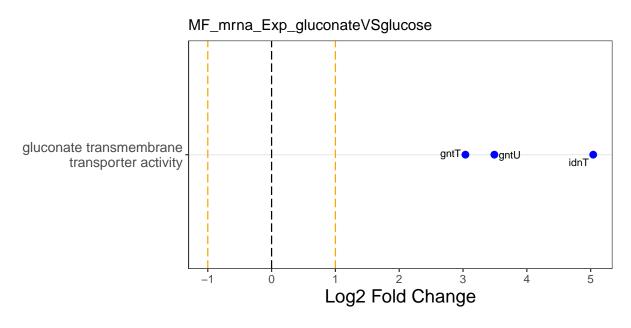


Figure S19: Significantly differentially expressed GO annotations related with molecular functions and associated genes with gluconate as carbon source, as determined by mRNA abundances in exponential phase. The top 2 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

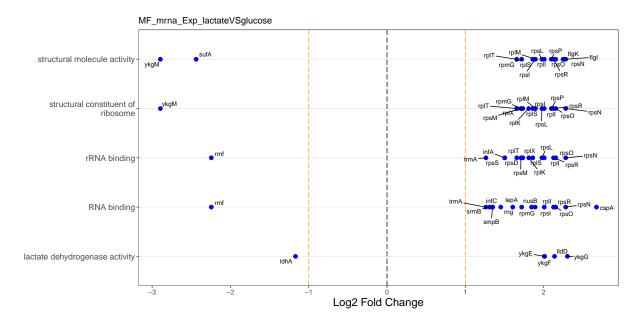


Figure S20: Significantly differentially expressed GO annotations related with molecular functions and associated genes with lactate as carbon source, as determined by mRNA abundances in exponential phase. The top 5 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

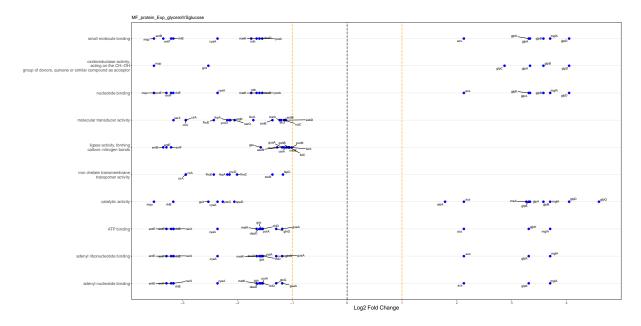


Figure S21: Significantly differentially expressed GO annotations related with molecular functions and associated genes with glycerol as carbon source, as determined by protein abundances in exponential phase. The top 10 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

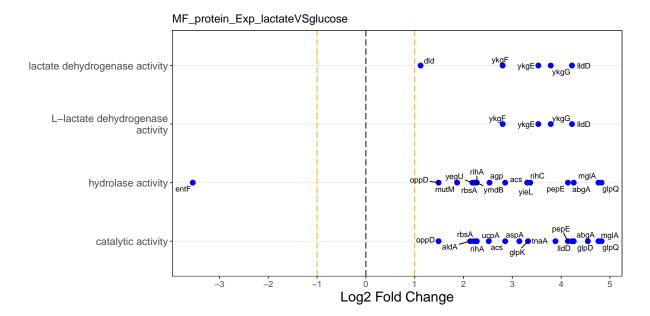


Figure S22: Significantly differentially expressed GO annotations related with molecular functions and associated genes with lactate as carbon source, as determined by protein abundances in exponential phase. The top 4 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

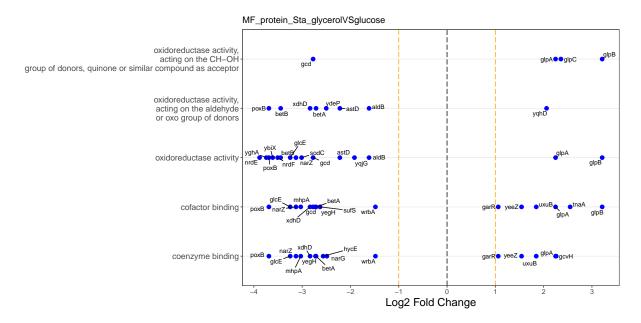


Figure S23: Significantly differentially expressed GO annotations related with molecular functions and associated genes with glycerol as carbon source, as determined by protein abundances in stationary phase. The top 5 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

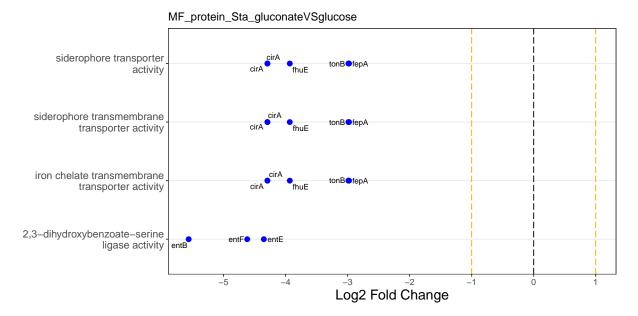


Figure S24: Significantly differentially expressed GO annotations related with molecular functions and associated genes with gluconate as carbon source, as determined by protein abundances in stationary phase. The top 4 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

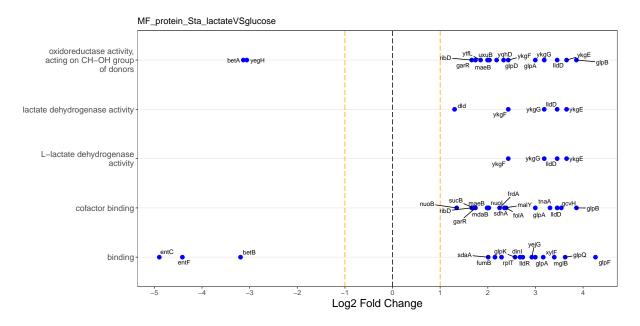


Figure S25: Significantly differentially expressed GO annotations related with molecular functions and associated genes with lactate as carbon source, as determined by protein abundances in stationary phase. The top 5 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

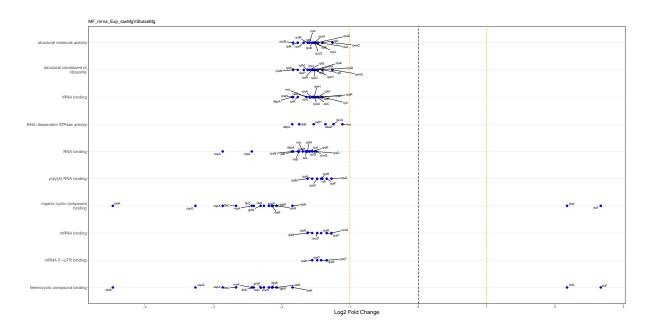


Figure S26: Significantly differentially expressed GO annotations related with molecular functions and associated genes with low  $Mg^{2+}$  levels, as determined by mRNA abundances in exponential phase. The top 10 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

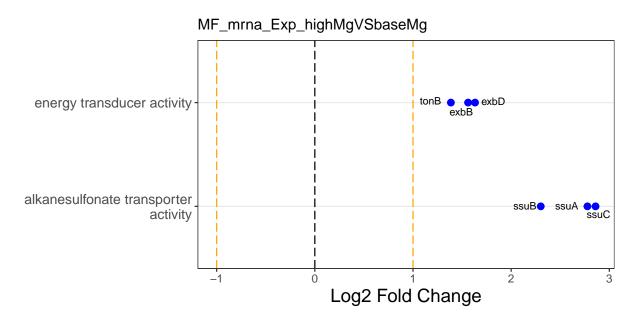


Figure S27: Significantly differentially expressed GO annotations related with molecular functions and associated genes with high  $\mathrm{Mg^{2+}}$  levels, as determined by mRNA abundances in exponential phase. The top 3 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

# lactate dehydrogenase activity L-lactate dehydrogenase activity

Figure S28: Significantly differentially expressed GO annotations related with molecular functions and associated genes with low  $Mg^{2+}$  levels, as determined by protein abundances in exponential phase. The top 2 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

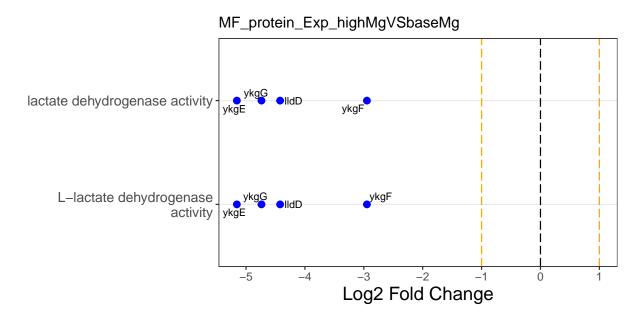


Figure S29: Significantly differentially expressed GO annotations related with molecular functions and associated genes with high  $Mg^{2+}$  levels, as determined by protein abundances in exponential phase. The top 2 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

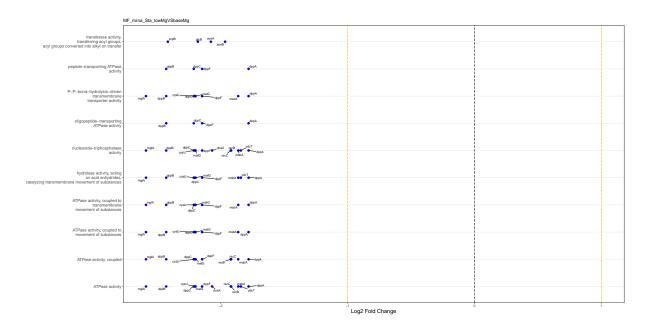


Figure S30: Significantly differentially expressed GO annotations related with molecular functions and associated genes with low  $Mg^{2+}$  levels, as determined by mRNA abundances in stationary phase. The top 10 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

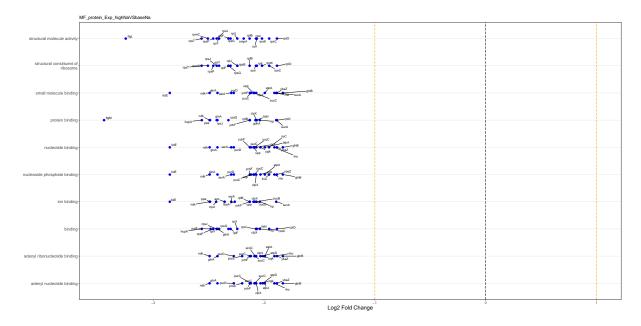


Figure S31: Significantly differentially expressed GO annotations related with molecular functions and associated genes with high  $Na^+$  levels, as determined by protein abundances in exponential phase. The top differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

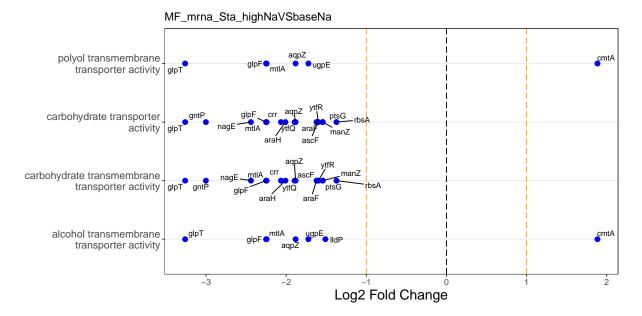


Figure S32: Significantly differentially expressed GO annotations related with molecular functions and associated genes with high  $Na^+$  levels, as determined by mRNA abundances in stationary phase. The top 5 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

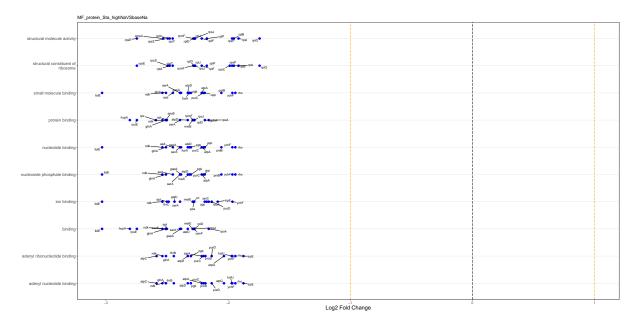


Figure S33: Significantly differentially expressed GO annotations related with molecular functions and associated genes with high  $Na^+$  levels, as determined by protein abundances in stationary phase. The top 10 differentially expressed molecular functions are shown along the y axis, and the relative fold change of the corresponding genes is shown along the x axis. We show up to 10 of the most significantly changed molecular functions and for each molecular function, we show up to 15 of the most significantly changing genes.

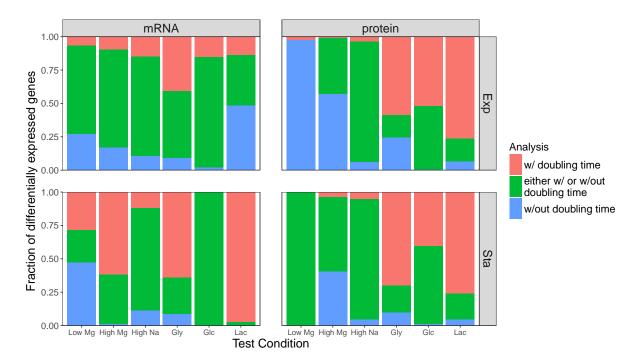


Figure S34: Fraction of differentially expressed genes that are found in analyses with or without controlling for doubling time. Shown are the fractions of genes identified as differentially expressed only when controlling for doubling time (red), only when not controlling for doubling time (blue), or in both cases (green). Combined with the absolute numbers of differentially expressed genes in the various conditions (Figure 5), we can see that the main differences in analyses with or without doubling time arise for protein abundances analyzed with respect to different carbon sources.

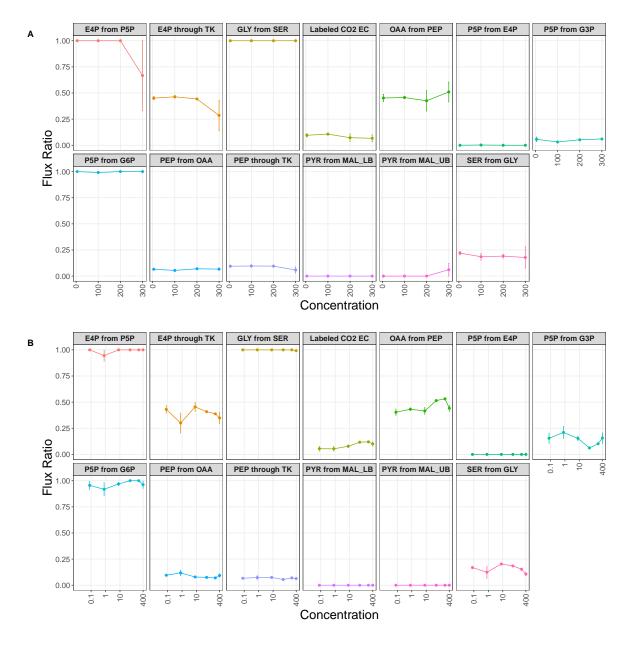


Figure S35: Flux ratios versus ion concentrations. 13 different flux ratios were measured with respect to four different  $\mathrm{Na^+}$  and five different  $\mathrm{Mg^{2+}}$  concentrations. (A) Concentrations with respect to changing  $\mathrm{Na+}$  concentrations. (B) Concentrations with respect to changing  $\mathrm{Mg^{2+}}$  concentrations. There was no significant trend of increase or decrease in flux ratios with respect to either  $\mathrm{Na^+}$  or  $\mathrm{Mg^{2+}}$  concentrations (Supplementary Table 13).

#### List of Supplementary Tables

Supplementary Table S1: Meta information for each sample. Includes information about sample numbers, experiment, the growth time at which the sample was collected, harvest date, number of RNA samples (technical replicates), number of protein samples (technical replicates), batch number,  $Mg^{2+}$  and  $Na^{+}$  concentrations, growth phase, doubling time (mean,  $\pm 95\%$  confidence interval,  $r^{2}$  from the linear fit to OD600 values).

File name: tableS1 meta data.csv

**Supplementary Table S2:** Normalized mRNA counts. Includes data for 4196 distinct proteins each for 152 samples.

File name: tableS2 mRNA normalized raw data.csv

**Supplementary Table S3:** Normalized protein counts. Includes data for 4196 distinct proteins each for 105 samples.

File name: tableS3 protein normalized raw data.csv

**Supplementary Table S4:** Mean flux ratios for 13 branches each, measured for varying Mg²⁺ and Na⁺ concentrations in exponential and stationary phase.

 $File \ name: \ table S4_flux Data.csv$ 

**Supplementary Table S5:** Doubling time measurements in exponential phase. Includes the mean,  $\pm 95\%$  confidence interval, and  $r^2$  from the linear fit to OD600 values.

File name: tableS5 doubling times.csv

Supplementary Table S6: z-scores obtained from tests for significant clustering of mRNA counts.

File name: tableS6_clustering_mrna_cophenetic.csv

Supplementary Table S7: z-scores obtained from tests for significant clustering of protein counts.

File name: tableS7 clustering protein cophenetic.csv

**Supplementary Table S8:** Combined results from tests for differential expression for all genes and all distinct tests considered.

The table contains the following information:

- Gene id (ECB number for mRNA and YP number for proteins), and corresponding gene name
- Results from DeSeq2 calculation, including base mean value, log2FoldChange, ifcSE, stat, pvalue, padj.
- Direction of change relative to base level ("+1" for increase, "-1" for decrease)
- Data type (mRNA or protein)
- Growth phase
- What is tested; base value and contrast.
- Individual output file name

- Carbon Source, Mg²⁺ and Na⁺ levels and growth phase of test data
- Control parameters of the test (batch only or batch plus doubling time)

File name: tableS8 combinedOutputDF DeSeq.csv

**Supplementary Table S9:** Filtered version of Supplementary Table S8, retaining only genes with P < 0.05 and log2FoldChange > 2.

File name: tableS9 combinedDifferentiallyExpressedGenes DeSeq.csv

**Supplementary Table S10:** Complete results from DAVID enrichment analysis for KEGG pathways and molecular functions.

File name: tableS10 combinedResultList DAVID.csv

**Supplementary Table S11:** List of the additional proteins identified as differentially expressed when controlling for doubling time, tested for different carbons sources. Results are provided for both exponential and stationary phases.

File name: tableS11 changed protein carbonSource ExpSta.csv

**Supplementary Table S12:** Enriched KEGG pathways and molecular functions based on the genes listed in Supplementary Table S11.

 $File \ name: \ table S12_changed_DAVID_P05.csv$ 

**Supplementary Table S13:** Results from linear regressions of flux ratios against ion concentrations  $(Mg^{2+} \text{ and } Na^{+}).$ 

File name: tableS13_flux_vs_conc_Pvalues.csv

**Supplementary Table S14:** Results from linear regressions of flux ratios against doubling times  $(Mg^{2+} \text{ and } Na^{+})$ .

File name: tableS14 flux vs doublingTime Pvalues tog.csv