

1 Culturable diversity of Arctic phytoplankton during pack ice  
2 melting

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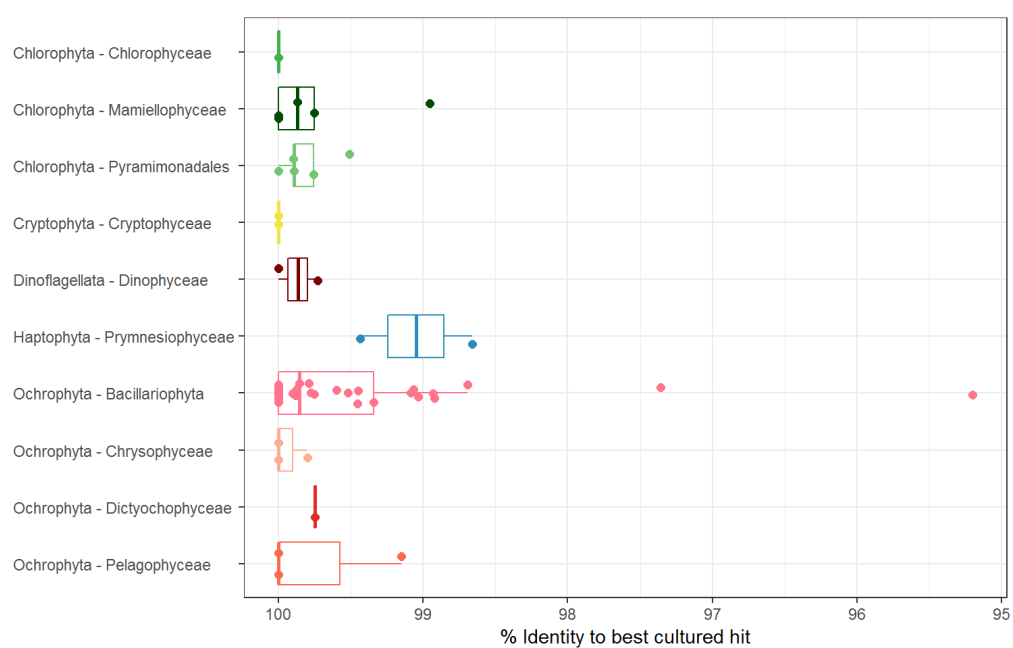
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15 **Supplementary material**

16     Supplementary material is available on GitHub at  
17     <https://github.com/vaulot/Paper-2019-Ribeiro-GE-cultures>

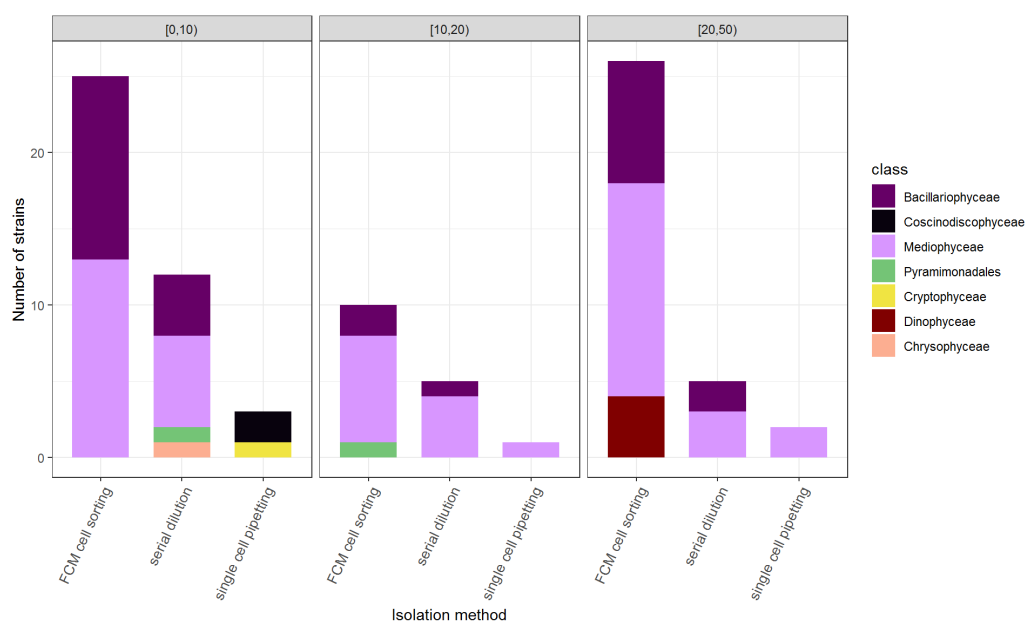
18     Supplementary Data S1: File GE\_cultures\_Tables.xlsx. Sheet Data S1. Strains  
19     collected during GE campaign, including both Amundsen an Ice Camp samples:  
20     RCC and GenBank accession number, taxonomy, respective clusters, sampling  
21     substrate, depth and date, geographic coordinates and isolation method.

22     Supplementary Data S2: File GE\_cultures\_Tables.xlsx. Sheet Data S2. Best  
23     BLAST hit for representative 18S rRNA sequences from each phylotype against  
24     all GenBank sequences, PR<sup>2</sup> sequences and sequences from cultured strains.



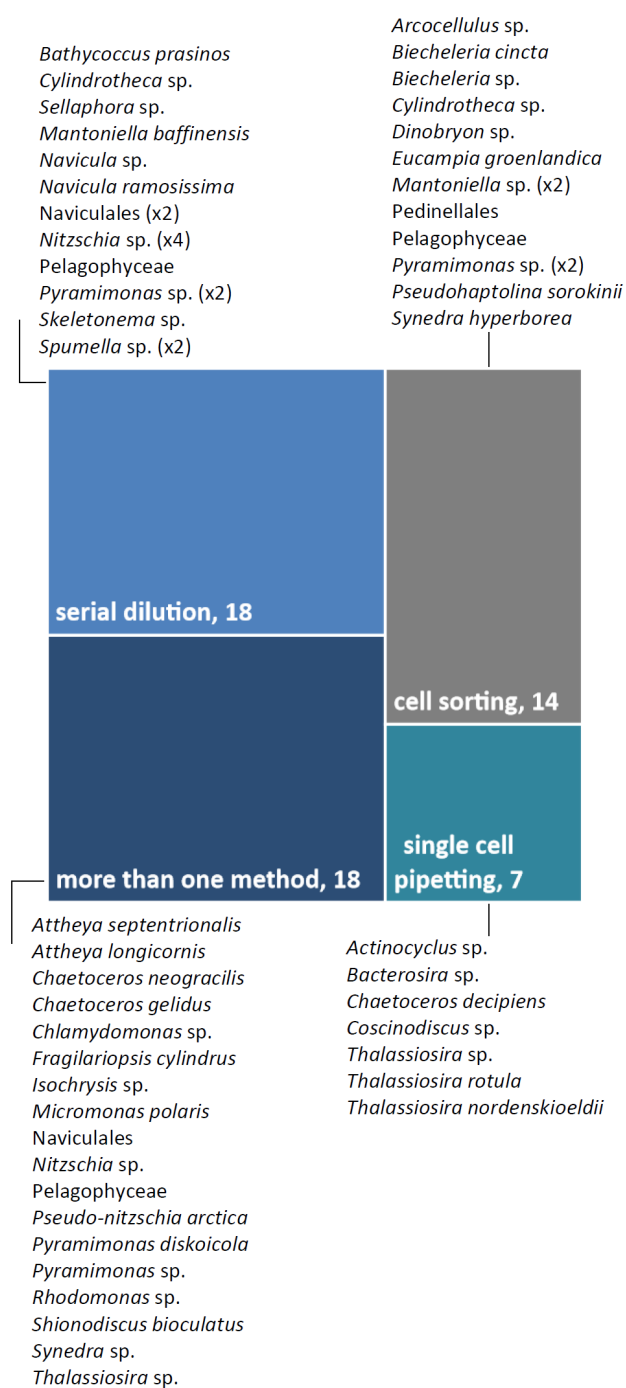
**Figure S1. Novelty of phylotypes.**

Percentage of similarity of phylotype representative 18S rRNA sequence to best BLAST hit from GenBank (see Supplementary Data S2).



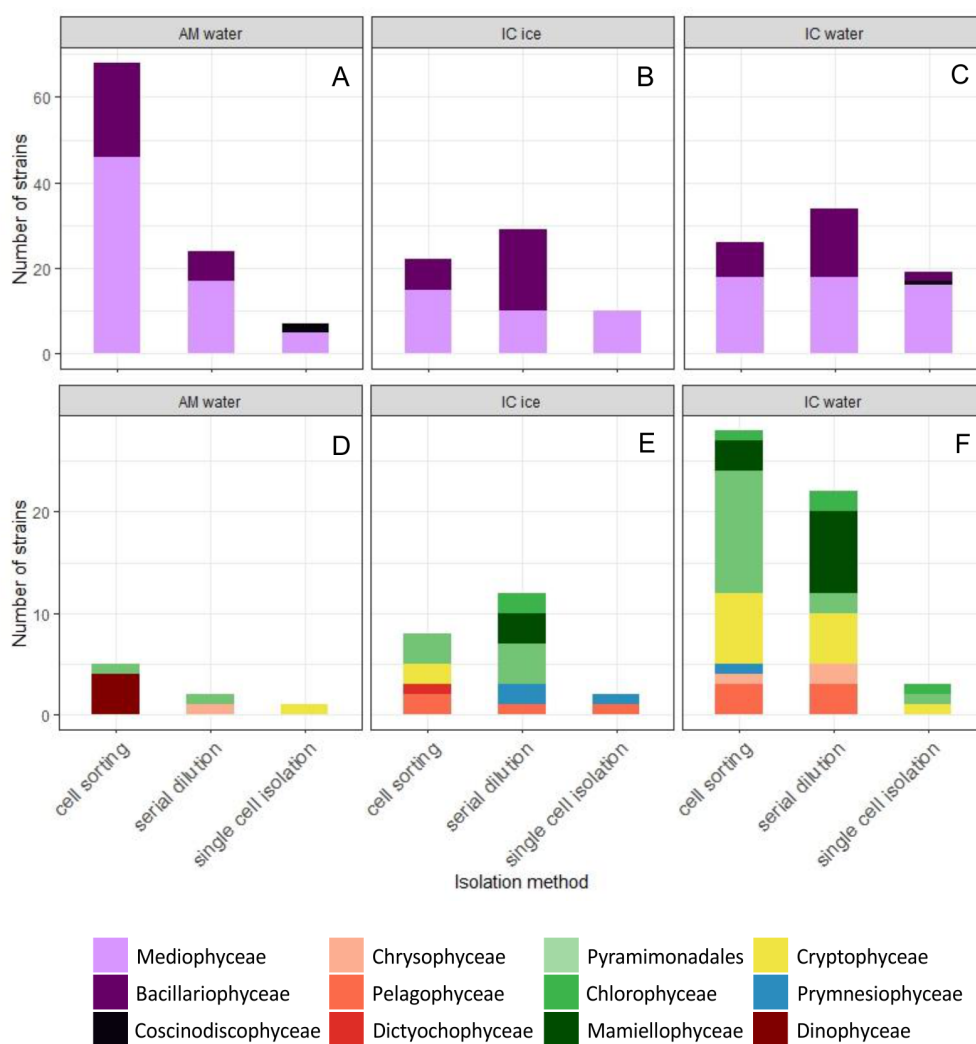
**Figure S2. Strains from Amundsen cruise as a function of isolation method and depth.**

Strain class distribution for the Amundsen cruise separated according to the method of isolation (cell sorting, serial dilution and single cell isolation) and sampling depth range.



**Figure S3. Phylotype as a function of isolation method.**

Treemap of the number of strains isolated as function of the isolation method.



**Figure S4. Strains as a function of isolation method and substrate.**

Strains class distribution separated according to the method of isolation (cell sorting, serial dilution and single cell isolation) and sampling substrate: water samples from the Amundsen cruise, and water and ice samples from the Ice Camp for diatoms (top panels) and non-diatoms (bottom panels).