

# Package ‘ZINQ’

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**Type** Package

**Title** Zero-Inflated Quantile Approach for Microbiome Association Testing

**Version** 2.0

**Author** Wodan Ling and Michael C. Wu

**Maintainer** Wodan Ling <wol4002@med.cornell.edu>

**Description** This package conducts association testing between individual taxa of microbiome (un-normalized or normalized by any method) and clinical variables (dicrete or quantitative), supporting both unadjusted and adjusted tests.

**License** GPL (>= 3)

**Encoding** UTF-8

**LazyData** true

**NeedsCompilation** no

**Imports** quantreg, MASS, logistf

**RoxygenNote** 7.3.1

**Suggests** knitr,  
rmarkdown

**VignetteBuilder** knitr

## R topics documented:

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Sample_Data	<i>Example data, normalized abundance of two taxa and covariates</i>
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## Description

A dataset containing two taxa which have typical abundance profiles highlighting power of ZINQ, including rarefied abundance of taxon 1, CSS normalized abundance of taxon 2, the clinical variable of interest, and several covariates.

**Usage**

```
Sample_Data
```

**Format**

A data frame with 531 rows and 6 variables:

**rarefied\_taxon1** rarefied abundance of taxon 1.

**CSS\_taxon2** CSS normalized abundance of taxon 2.

**X** the clinical variable of interest, binary variable.

**Z1** covariate 1, continuous variable.

**Z2** covariate 2, continuous variable.

**Z3** covariate 3, continuous variable.

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ZINQ_check	<i>Sanity check before applying ZINQ</i>
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**Description**

Sanity check before applying ZINQ

**Usage**

```
ZINQ_check(tax_tab, metadata, C)
```

**Arguments**

tax_tab	The taxa read count table (un-normalized), sample (row) by taxa (col).
metadata	The metadata, sample (row) by variable (col).
C	The name(s) of clinical variable(s) of interest, e.g., "Condition" or c("Condition", "Batch").

**Details**

- It is recommended to do the sanity check before applying ZINQ. If it is necessary, warnings will be printed to guide the analysis using ZINQ.
- If library size is a confounder of the variable(s) of interest, ZINQ might not control type I error.
- If there are few non-zero read counts, use ZINQ with caution.
- For a perfect separation, e.g., there are all zeroes in one group (case or control), which is an extreme case of imbalance, use ZINQ with caution.
- The sanity check is mainly about zero inflation. Most normalization methods will keep the original zeroes, thus investigating the un-normalized taxa read count table provides sufficient clues to use ZINQ. For normalization methods not retaining the zeroes. e.g., CLR, results of the sanity check is not informative, one can apply ZINQ directly.

**Value**

Print warnings if necessary:

- When library size is a confounder.
- For each taxon, (1) when all read counts are zero, (2) when there are limited non-zero read counts (<30 or <15), (3) when there is a perfect separation w.r.t. the variable(s) of interest.

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ZINQ_combination	<i>Combine the marginal p-values</i>
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**Description**

Combine the marginal p-values

**Usage**

```
ZINQ_combination(
  input,
  method = "MinP",
  taus = c(0.1, 0.25, 0.5, 0.75, 0.9),
  M = 10000
)
```

**Arguments**

input	An output from <a href="#">ZINQ_tests</a> .
method	Combination method, "MinP" for MinP test, "Cauchy" for Cauchy combination test; default is "MinP".
taus	A grid of quantile levels, must be a subset or equal to that from input; default is c(0.1, 0.25, 0.5, 0.75, 0.9).
M	The number of MC draws from the joint distribution of quantile rank-scores when method is "MinP"; default is 10000.

**Details**

- Please choose 'MinP' or 'Cauchy' for method, no other options.
- taus must be a subset or equal to the grid used to produce input.

**Value**

A pvalue, the final p-value of ZINQ.

**References**

- Ling, W. et al. (2021). Powerful and robust non-parametric association testing for microbiome data via a zero-inflated quantile approach (ZINQ). *Microbiome* 9, 181.
- He, Z. et al. (2017). Unified sequence-based association tests allowing for multiple functional annotations and meta-analysis of noncoding variation in metabochip data. *The American Journal of Human Genetics* 101(3), 340–352.

- Lee, S. et al. (2012). Optimal tests for rare variant effects in sequencing association studies. *Biostatistics* 13(4), 762–775.
- Liu, Y., Xie, J. (2019). Cauchy combination test: a powerful test with analytic p-value calculation under arbitrary dependency structures. *Journal of the American Statistical Association*, 1–18.

### Examples

```
n = 300
p <- function(x0, gam0=0.75, gam1=-0.15){
  lc = gam0 + gam1*x0
  exp(lc) / (1 + exp(lc))
}
x = c(rep(0, n), rep(1, n))
w = 0.5 + 1.5*x + (1+0.15*x)*rchisq(2*n,df=1)
b = rbinom(2*n, 1, p(x))
y = w*b
dat = data.frame(y, x)

result = ZINQ_tests(formula.logistic=y~x, formula.quantile=y~x, C="x", data=dat)
ZINQ_combination(result, method="Cauchy")
```

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ZINQ\_tests

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*Marginal tests for the Firth logistic and quantile regression components*


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### Description

Marginal tests for the Firth logistic and quantile regression components

### Usage

```
ZINQ_tests(
  formula.logistic,
  formula.quantile,
  C,
  y_CorD = "C",
  data,
  taus = c(0.1, 0.25, 0.5, 0.75, 0.9),
  seed = 2020
)
```

### Arguments

formula.logistic

The full model of Firth logistic regression, e.g.,  $Y \sim X + Y + Z$ , where  $Y$  is zero-inflated.

formula.quantile

The full model of quantile regression, can be different from formula.logistic.

C

The name(s) of clinical variable(s) of interest, e.g., "Condition" or c("Condition", "Batch").

y_CorD	An indicator: use "D" if Y is count, a perturbation from U(0, 1) will be added to the response; use "C" if Y is continuous; default is "C".
data	A data.frame: better cleaned and processed, use numeric for Y and binary covariates, use factor for multi-class discrete covariates.
taus	A grid of quantile levels, e.g., 0.5 for the median, 0.75 for the 3rd quartile; default is c(0.1, 0.25, 0.5, 0.75, 0.9).
seed	A seed for perturbation when y_CorD is "D"; default is 2020.

### Details

- Compositional data is regarded as continuous, determined by its support.
- taus is a tuning parameter that does not have an efficient selection process yet, try from coarsed to fine grids (e.g., seq(0.1, 0.9, by=0.2) to seq(0.1, 0.9, by=0.1)), or try adding more extreme levels (e.g., c(0.25, 0.5, 0.75) to c(0.1, 0.25, 0.5, 0.75, 0.9)), with a goal to keep type I error controlled and boost the power; for common taxa, start from the default; for rare taxa, start from c(0.25, 0.5, 0.75).
- Quantile rank-score test corrected for zero-inflation is used for the quantile regression component.
- Penalized likelihood-ratio test is used for the Firth logistic regression component.

### Value

A list

- pvalue.logistic - A single p-value from the Firth logistic regression component.
- pvalue.quantile - A length(taus) by 1 vector, a sequence of p-values from the quantile regression component.
- Sigma.hat - A df x length(taus) by df x length(taus) matrix, where df is the dimension of C, the covariance matrix of quantile rank-scores.
- zerorate - The proportion of zeroes in Y.
- taus - The grid of quantile levels used.

### References

- Ling, W. et al. (2021). Powerful and robust non-parametric association testing for microbiome data via a zero-inflated quantile approach (ZINQ). *Microbiome* 9, 181.
- Machado, J.A.F., Silva, J.S. (2005). Quantiles for counts. *Journal of the American Statistical Association* 100(472), 1226–1237.

### Examples

```
n = 300
p <- function(x0, gam0=0.75, gam1=-0.15){
  lc = gam0 + gam1*x0
  exp(lc) / (1 + exp(lc))
}
x = c(rep(0, n), rep(1, n))
w = 0.5 + 1.5*x + (1+0.15*x)*rchisq(2*n,df=1)
b = rbinom(2*n, 1, p(x))
y = w*b
dat = data.frame(y, x)
```

```
ZINQ_tests(formula.logistic=y~x, formula.quantile=y~x, C="x", data=dat)
```

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