Multiple Sequence Alignment & Phylogeny

Multiple Sequence Alignment

What Is a Multiple Sequence Alignment?

- The alignment of more than two sequences
- MSAs = multiple-sequence alignments
- The goal of an MSA is twofold:
 - Aligning corresponding regions of the sequences
 - Revealing positions that are conserved
- The main steps to a useful MSA require
 - Choosing the right sequences
 - Choosing the right MSA method
 - Interpreting the alignment

Evolution

- Amino acids mutate randomly
- Mutations are then selected (accepted) or counter-selected (rejected)
- If a mutation is harmful, it is counter-selected
 - It disappears from the genome
 - You never see it
- Mutations of important positions (such as active sites) are almost always harmful
- You can recognize important positions because they never mutate!

MSAs reveal these conserved positions

An Example of Conserved Positions: The Serine Proteases Active Site

```
CLPP ECOLI E.col(40) ERVIFLTGOV----EDHMANLIVAOMLFLEAENPEKDIYLYINSPGGVITAGMSIYDTMOFIKPD----VSTIC (105)
CLPI MYXXA M.xan(26) DRIIMLGTPV----NDDVANIIVAOLLFLESEDPDKGINLYINSPGGSVTAGLAIYDTMOYVKCP----VSTIC (91)
                                                                                                       Active Site
21228980 M.maz(27) MISLFGLPAYQSIDEEDAEQVLRWIRKY-----RDYPLELILHTPGGQLHASIQIARALKNHPKK----TRVLI (92)
15643678 T.mar(58) SISFLGFPVRRYIDIEDSEEILRAIKLTP----SDMPIDLILHTPGGLVLAAEQIARALKMHKGK----VTVFV (123)
          M.jan(64) SIGLFGIPVYKFITIEDSEEILRAIRAAP----KDKPIDLIIHTPGGLVLAATQIAKALKAHPAE----TRVIV
          P.fur(59) SIGFFGIPVYKFISIEDSEEVLRAIRMAP----KDKPIDLIIHTPGGLVLAATOIAKALKDHPAE-
22972030 C.aur (53) TMSLLGFPLVRYINIEDSEAVLRAIKMTD----RDIPIDLILHTPGGLVLAAEQIARALTKHAAK-
23050732 M.bar(75) AISLFGIPAYOYIDEEDAEOILRWIRKY-----KDYPLELILHTPGGOLHSSIOJARALRRHS
          S.mel(50) HVARVAVTGLIQ---DD RELVERLERIAD N--QSVKALIVTISSPGGTTYGGEVIYKA
          A.tum(27) AIMAGGNQFRPALNLASYAPLLEKAFAVKDA----PAVAISLNSPGGSPVOAD YNRIRQLAF
17934547
CLPP ECOLI E.col (106) MGQAASMGAFLLTAGAKGKRFCLPNSRVMI
                                                                                                 (147)
CLP1 MYXXA M.xan( 92) VGQAASMGALLLLAGAKGKRYALPNSP WIHOPLGGA---
          M.maz(93)PHYSMSGGTIIALAADE-IV
          T.mar (124) PHYAMSGGTLIALAADE
15668307
          M. jan (130) PHYAMSGGTLL AADK-IIMDENAVLGPVD-POLGO-
          C.aur (119) PHYAMSGGTLIALAADE-IVMDENAVLGPVD-POLGO----HPAASILSVLERKPLSEIDDET
          M.bar (140) PHYSMSGGTIIALAANE-IVMDRDAVIGPID-PQIGDFIRGMYPAPSWIYAAETKK-EKADDT LVMS----
23050732
          S.mel (117) RTLAASAGYLIALAGDR-IVAGETSITGSIG-VIFOY----POVKTLMDKLGVSLESIKSTELKAEPSPFHPPS (184)
          A.tum(97)EDVAASGGYMIALAGDE-IIADPTSIVGSIG-VVSGG----FGFPEMLRKIGVERRVYTZGENKVILDPFOPEK (164)
CLPP ECOLI E.col (148) ----EIHAREILKVKGRMNELMALHT-----GOSLEGIERDT-----ERD-RFLS
CLP1 MYXXA M.xan(134)----DIQAKEILRLRSYINGLIVKHT-----GHTIERIEKDT-----ERD-YFM
21228980 M.maz(157)----DISRKALRLTRNVAKELLEGKIQPD-GKEDRLEEVVEKLVSG-ENIHSTY
         T.mar(185)----DIAEKAIRQVKEFVVEILSDKV----SKEKAEKIADKLCSG-YWTHD
15668307
          M.jan(191)----DIAKKAINQVQNFVYNLLKDKY-----GEEKAKELSKILTEG-RWTHDYPITVEEAKEL (243)
18976612
          P.fur (186) ---- DVAKKAI KQVQDFLYDLLKDKY----- GEEKARELAQILTEG-RWTHDYPITVEHAREL (238)
22972030
          C.aur(180)----DIAEKAIROVKRTVCELLRDKM-----PVERAEEVAHTLASG-VWTHDYPITVSEAREL (232)
23050732
          M.bar (205) ---- DVSRKALKFTRNVAKELLEGKIQPGPAGESRLDEVVEKLVSG-EMIMSTPLSAGEAKKI (262)
15964138
          S.mel (185) DEARAMIQAMIDDSYGWFVDLVAERRK------LPRPEALALADGRIFTGRQALEGKLVDEL (240)
          A.tum(165)EGDIDYLKSLQVEIHNVFIDMVKMRRG----SKLK--GDDALFSGLFWTGMRGLDLGLIDGL (220)
```

MSA – Example Applications

| Application | Procedure |
|------------------------|---|
| Extrapolation | Determine the function of your protein |
| Phylogenetic analysis | Build a Phylogenetic tree |
| Pattern identification | Discover important positions |
| Domain identification | Turn your alignment into a domain profile |

MSA – Example Applications

| DNA regulatory elements identification | Use your alignment to discover promoters |
|--|--|
| Structure prediction | Predict the secondary structure of proteins and RNA molecules |
| nsSNP analysis | Discover important allelic variations in human and other animals (nsSNP: non-synonymous single-nucleotide polymorphisms) |
| PCR analysis | Select your PCR primers |

Choosing the Right Sequences

- When building an alignment, it is your job to select the sequences
- Two main factors when selecting sequences:
 - Number of sequences
 - Nature of the sequences
- A reasonable number of sequences: 20 to 50
 - Ideal for most methods
 - Small alignments are easy to display and analyze
- Types of sequences
 - Well-selected sequences ⇔ informative alignment

Choosing Sequences that are Different Enough

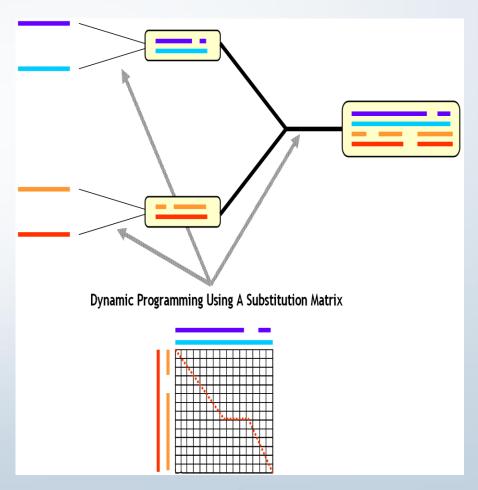
- An alignment is useful if . . .
 - The sequences are correctly aligned
 - It can be used to produce trees, profiles, and structure predictions
- To obtain this result, the sequences must be
 - Not too similar
 - Not too different
- Sequences that are very similar . . .
 - Are easy to align correctly
 - Are not informative ⇔ useless trees and profiles, bad predictions
- Sequences that are very different . . .
 - Are difficult to align
 - Are very informative \Leftrightarrow good trees and profiles, good predictions

DNA or Proteins?

- DNA sequences are harder to align than proteins
 - DNA-comparison models are less sophisticated
- Most methods work for both DNA and proteins
 - The results are less useful for DNA
- If your DNA is coding, work on the translated proteins
- If sequences are homologous . . .
 - Along their entire length ⇔ use progressive alignment methods
 - In terms of local similarity \Leftrightarrow use motif-discovery methods

MSA Methods: Progressive Algorithm

- Sequences are grouped by similarity (guide tree)
- Sequences are aligned 2 by 2
- The intermediate alignments are then aligned 2 by 2
- You align 2 sequences by using dynamic programming



MSA Methods: Progressive Algorithm

- Its main strength is its speed
- Its main weakness is its greed
 - Sequences aligned at the beginning are never realigned
 - Early mistakes cannot be corrected
- Assemble datasets with lots of intermediate sequences
- Imagine each sequence is part of a stone bridge across a river:
 - Doesn't matter how wide the river is, if the stones are close enough together
 - Doesn't matter how diverse your sequences are, if each sequence has a close relative



https://www.tes.com

Selecting a Method

- Many alternative methods exist for MSAs
- Most of them use the progressive algorithm
- They all are approximate methods
- None is guaranteed to deliver the best alignments
- All existing methods have pros and cons
 - ClustalW is the most popular
 - T-Coffee/M-Coffee and ProbCons are more accurate but slower
 - MUSCLE is very fast, ideal for very large datasets

Aligning Your Sequences Correctly

- It can be difficult to align sequences correctly
 - They evolve too fast
- For proteins, the best alternative is to use 3D structure
 - 3D Structures change slower than sequences
- Unfortunately few sequences have a known structure

• Expresso lets you find the structures that correspond to your sequences and use them to build an MSA

Using the EXPRESSO Web server

The EXPRESSO Web server aligns sequences using 3D information

- The EXPRESSO tool . . .
 - Looks in PDB for the structure of your sequences
 - Aligns your sequences using structural information
 - Returns a multiple-sequence alignment based on structure

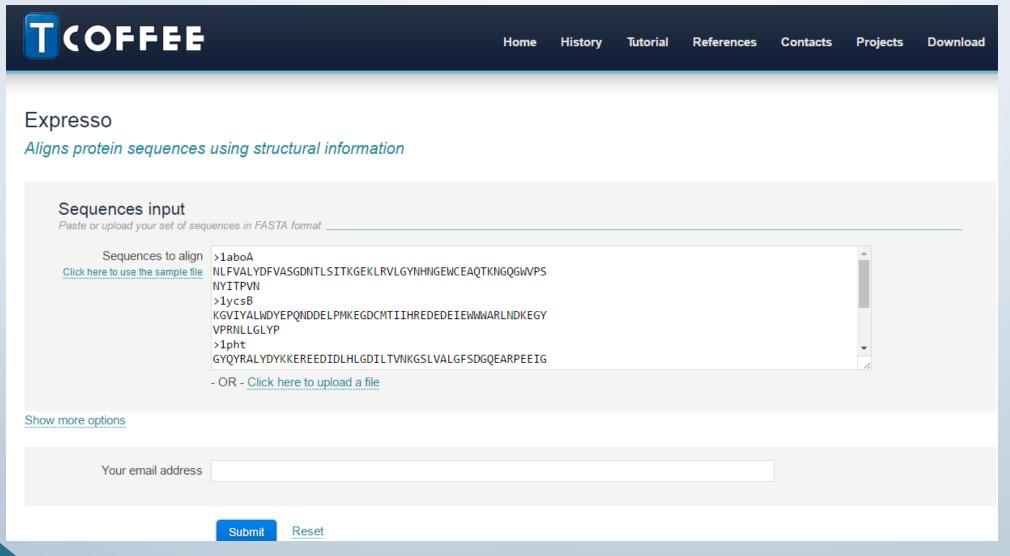
• If your sequences have a known structure, EXPRESSO is the most accurate MSA method

EXPRESSO Web server – Colored Output

- Red and Orange residues are probably well aligned
- Yellow should be treated with caution
- Green and blue are probably incorrectly aligned



EXPRESSO Web server - Example



EXPRESSO Web server - Example

Expresso alignment result

```
MSA
The multiple sequence alignment result as produced by T-coffee.
T-COFFEE, Version 11.00.d625267 (2016-01-11 15:25:41 - Revision d625267 - Build 507)
Cedric Notredame
SCORE=91
 BAD AVG GOOD
cons
1aboA
          -NLFVALYDFVASGDNTLSITKGEKLRVL------GYNHN---GEWCEAOTK---NGOGWVPSNY
1ycsB
1pht
         -DRVRKKSG-------AAW0G0IV------
NFRVYYRDS------RDPVWKGPAKLL------
                                                          -GWYCTNLTPEGYAVESEAHPGSV0IYPVAALERIN
-WKG----EGAVVIQDN---SDIKVVPRRKAKIIR
lvie
cons
```

Local Multiple Comparison Methods

 Most MSA programs assume your sequences are related along their whole length

• When this assumption is not true, the progressive approach will not work

• The only alternative is to compare multiple sequences locally

Local Multiple-Comparison Methods

- Gibbs Sampler
 - Will make a local multiple alignment
 - Will ignore unrelated segments of your sequences
 - Ideal for finding DNA patterns such as promoters
- Motif discovery methods
 - Will look for motifs conserved in your sequences
 - The sequences do not need to be aligned
- The most popular motif-discovery methods:
 - TEIRESIAS, MEME, SMILE, PRATT

MSA - Summary

- Assembling MSAs is a bit of an art
- Experience is a key factor

Most methods are now available online

- Make sure you know which method to use:
 - ClustalW-like method to align homologous sequences
 - Motif method to look for conserved regions

Phylogeny

Why Build a Phylogenetic Tree?

 Phylogenetic trees reconstruct the evolutionary history of your sequences

• They tell you who is closer to whom in the big tree of life

 Phylogenetic trees are based on sequence similarity rather than morphologic characters

3 applications to use your constructed Tree

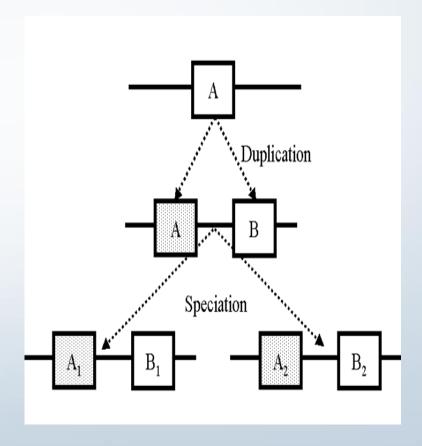
- Finding the closest relative of your organism
 - Usually done with a tree based on the ribosomal RNA

- Discovering the function of a gene
 - Finding the orthologues of your gene

- Finding the origin of your gene
 - Finding whether your gene comes from another species

Orthology and Paralogy

- Orthologous genes
 - Separated by speciation
 - Often have the same function
- Paralogous genes
 - Separated by duplications
 - Can have different functions
- In the graph:
 - A is paralogous with B
 - A1 is orthologous with A2



Bioinformatics: Life Sciences, Robert Lessick

Working on the Right Data

• Garbage in ⇒ Garbage out

• The quality of your tree depends on the quality of the data

Your first task is to assemble a very accurate MSA

Again: DNA or Proteins

- Most phylogenetic methods work on Proteins and DNA sequences
- If possible, always compute a multiple-sequence alignment on the protein sequences
 - Translate the sequences if the DNA is coding
 - Align the sequences
 - Thread the DNA sequences back onto the protein MSA
- If your DNA sequences are coding and have more than 70% identity . . .
 - Compute the **tree** on the **DNA** multiple-sequence alignment
- If your DNA sequences are coding and have less than 70% identity . . .
 - Compute the **tree** on the **protein** multiple-sequence alignment

Which Sequences?

- Orthologous sequences
 - Produce a species tree
 - Show how the considered species have diverged

- Paralogous sequences
 - Produce a gene tree
 - Show the evolution of a protein family

Building the Right Tree

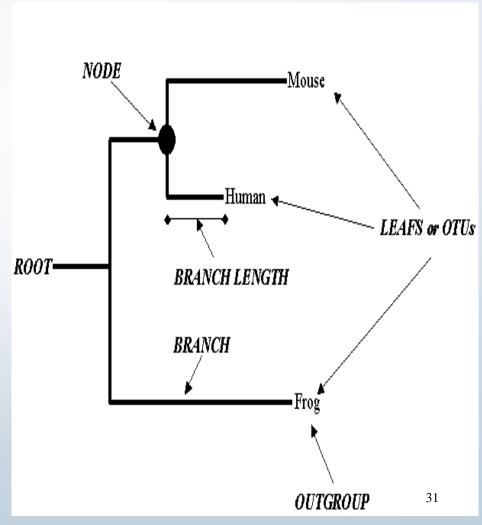
- There are two types of tree-reconstruction methods
 - Distance-based methods
 - Statistical methods
 - O Statistical methods are the most accurate
 - Maximum likelihood of success
 - Parsimony
 - O Statistical methods take more time
 - Limited to small datasets

Distance-based Methods for Tree Reconstruction

- Distance-based methods are the most popular
 - Neighbor Joining (NJ)
 - UPGMA
- Distance-based methods involve 2 steps:
 - Measure the distances between pairs of sequences in the MSA
 - Transform the distance matrix into a tree
- The two most popular packages for making trees are
 - Clustalw: very simple, not very sophisticated
 - **Phylip**: very powerful, less convivial

Reading Your Tree

- There's a lot of vocabulary in a tree
- Nodes correspond to common ancestors
- **Root** is the oldest ancestor
 - Often artificial
 - Only meaningful with a good outgroup
- Trees can be un-rooted
- Branch lengths are only meaningful when the tree is scaled
 - Cladograms have the same branch lengths
 - Phylograms have real branch lengths



Bioinformatics: Life Sciences, Robert Lessick

Bootstrapping

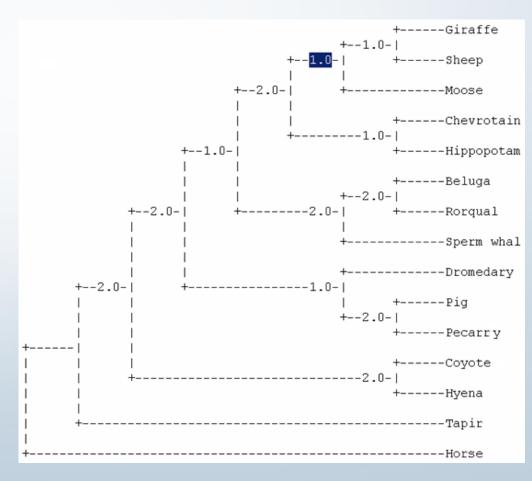
- Use bootstrapping to verify the solidity of each node
- ClustalW and Phylip do bootstrap operations automatically
- Bootstrapping involves these steps:
 - Select a subset of your MSA
 - Redo the tree
 - Repeat this operation N times (100 or 1000 times if you can)
 - Compute a consensus tree of the N trees
 - Measure how many of the N trees agree with the consensus tree on each node
- Each node gets a bootstrap figure between 0 and N
- High bootstrap ⇔ good node

A Bootstrapped Tree

• This tree was produced with 2 bootstrap cycles

• It shows some nodes as more robust than others

• In practice, always use more than 100 cycles



Practical Session

EMBL-EBI

EMBL-EBI

Research

Training About us



Multiple Sequence Alignment





Tools > Multiple Sequence Alignment

Multiple Sequence Alignment (MSA) is generally the alignment of three or more biological sequences (protein or nucleic acid) of similar length. From the output, homology can be inferred and the evolutionary relationships between the sequences studied.

By contrast, Pairwise Sequence Alignment tools are used to identify regions of similarity that may indicate functional, structural and/or evolutionary relationships between two biological sequences.

Clustal Omega @

New MSA tool that uses seeded quide trees and HMM profile-profile techniques to generate alignments. Suitable for medium-large alignments.

Launch Clustal Omega

Kalign @

Very fast MSA tool that concentrates on local regions. Suitable for large alignments.

Launch Kalign

MAFFT @

MSA tool that uses Fast Fourier Transforms. Suitable for medium-large alignments.

MUSCLE @

Accurate MSA tool, especially good with proteins. Suitable for medium alignments.

Launch MUSCLE

MView @

Transform a Sequence Similarity Search result into a Multiple Sequence Alignment or reformat a Multiple Sequence Alignment using the MView program.

Launch MView

Consistency-based MSA tool that attempts to mitigate the pitfalls of progressive alignment methods. Suitable for small alignments.

Example: Human TNF-alpha orthologous

TNF tumor necrosis factor [Homo sapiens (human)]

Gene ID: 7124, updated on 6-Dec-2016





Official Symbol TNF provided by HGNC

Official Full Name tumor necrosis factor provided by HGNC

Primary source HGNC:HGNC:11892

See related Ensembl:ENSG00000232810 MIM:191160; Vega:OTTHUMG00000031194

Gene type protein coding
RefSeq status REVIEWED
Organism Homo sapiens

Lineage Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates;

Haplorrhini; Catarrhini; Hominidae; Homo

Also known as DIF; TNFA; TNFSF2; TNLG1F; TNF-alpha

Summary This gene encodes a multifunctional proinflammatory cytokine that belongs to the tumor necrosis factor (TNF) superfamily.

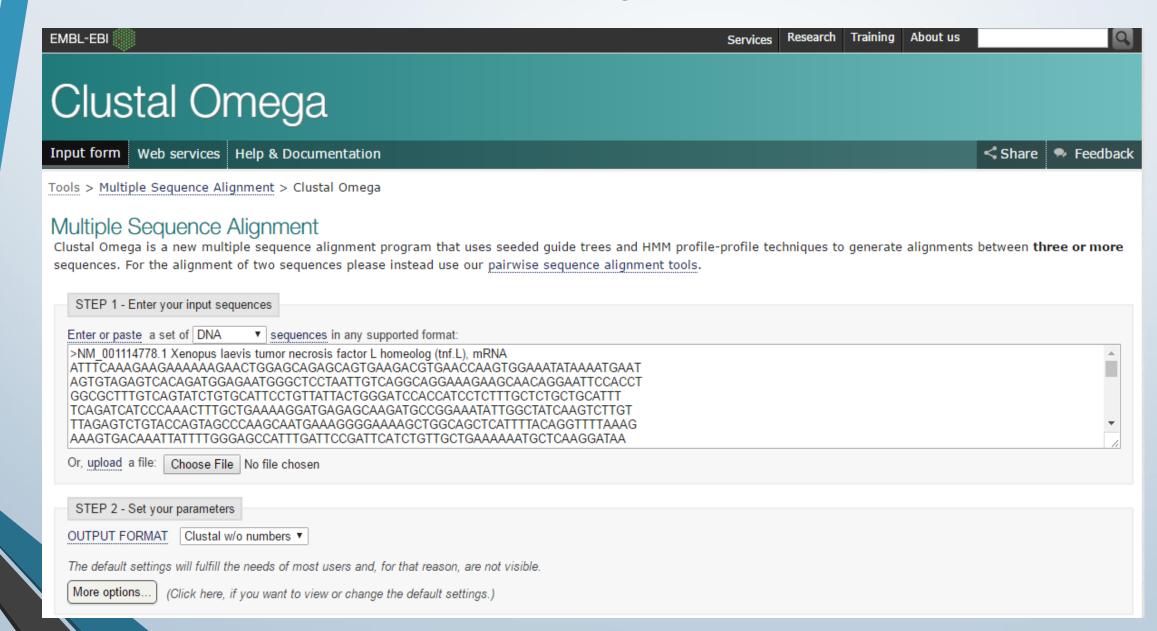
This cytokine is mainly secreted by macrophages. It can bind to, and thus functions through its receptors

TNFRSF1A/TNFR1 and TNFRSF1B/TNFBR. This cytokine is involved in the regulation of a wide spectrum of biological processes including cell proliferation, differentiation, apoptosis, lipid metabolism, and coagulation. This cytokine has been implicated in a variety of diseases, including autoimmune diseases, insulin resistance, and cancer. Knockout studies in mice

also suggested the neuroprotective function of this cytokine. [provided by RefSeq, Jul 2008]

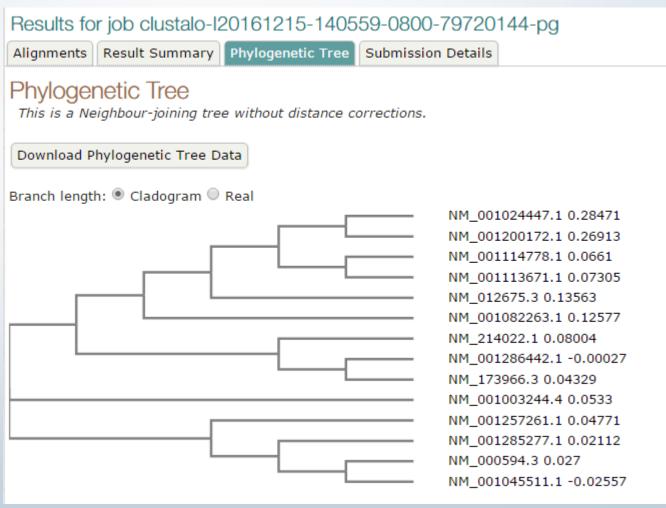
Orthologs mouse all

Clustal Omega / DNA

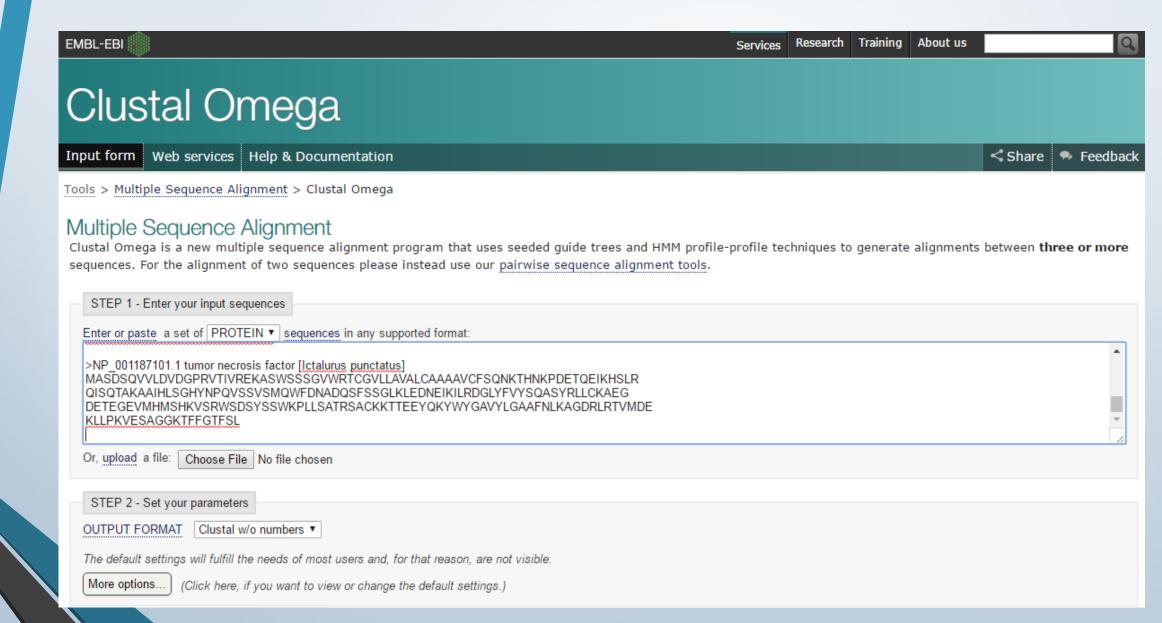


Results

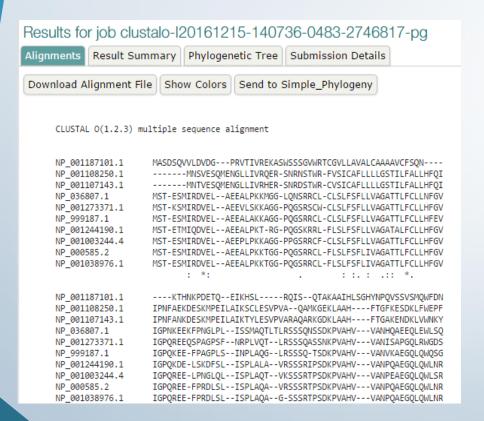


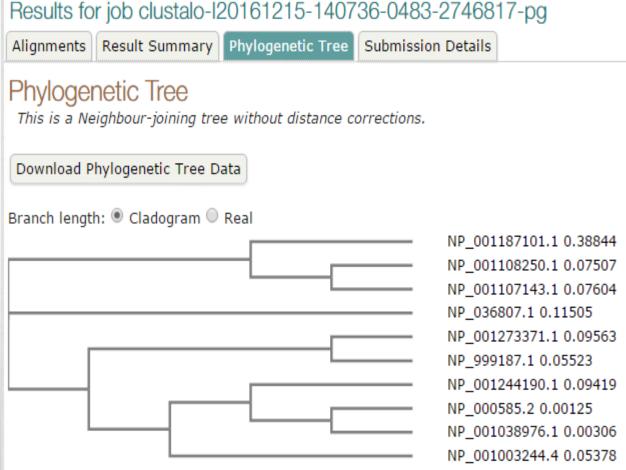


Clustal Omega / Protein

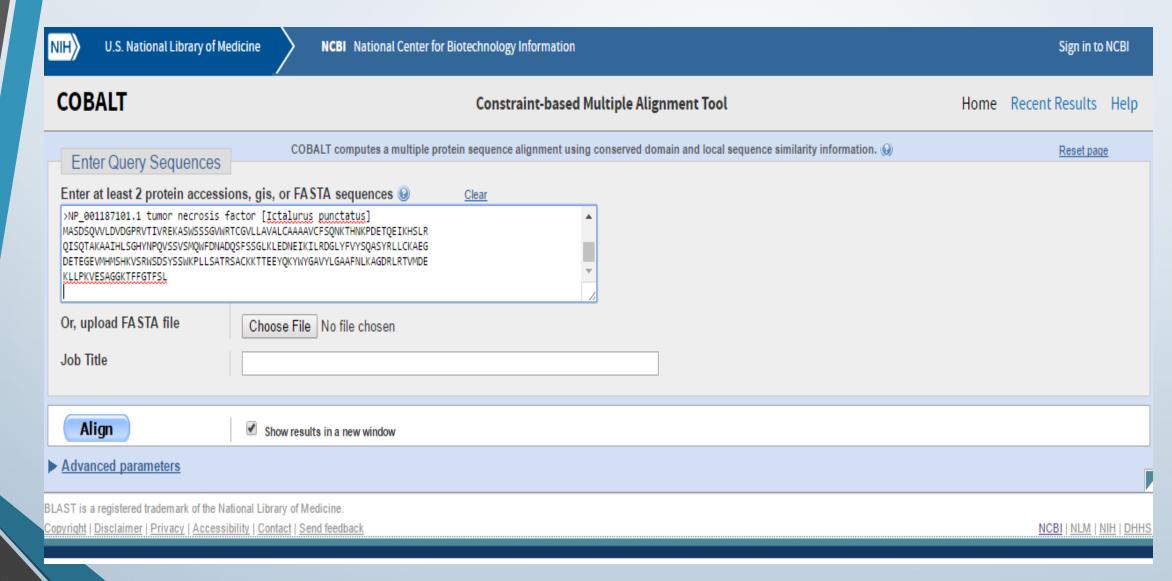


Results



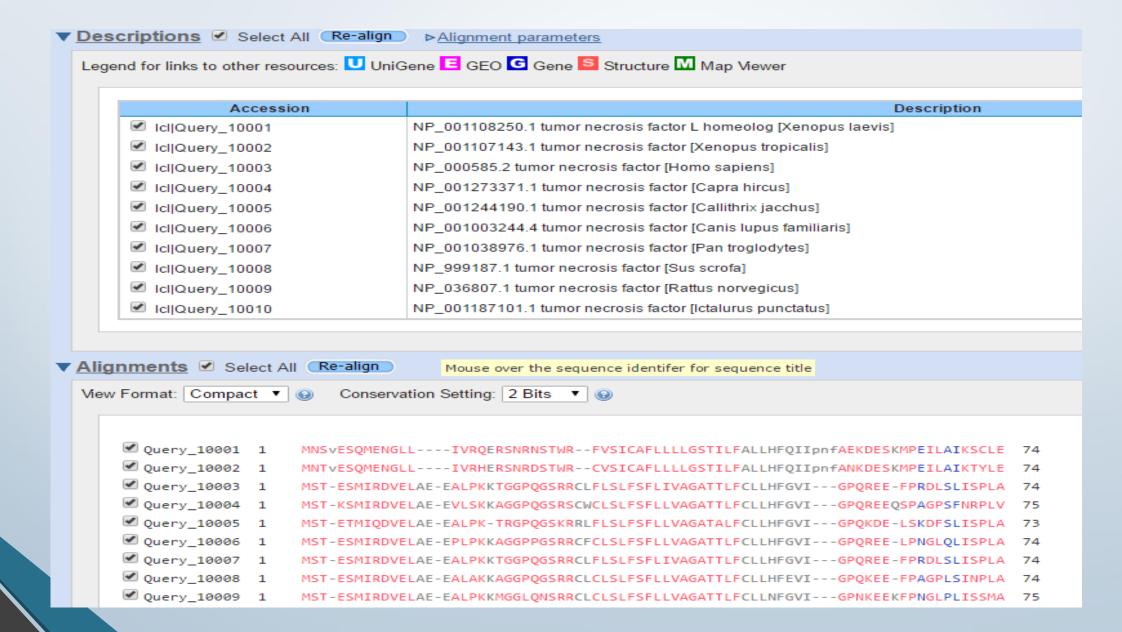


NCBI

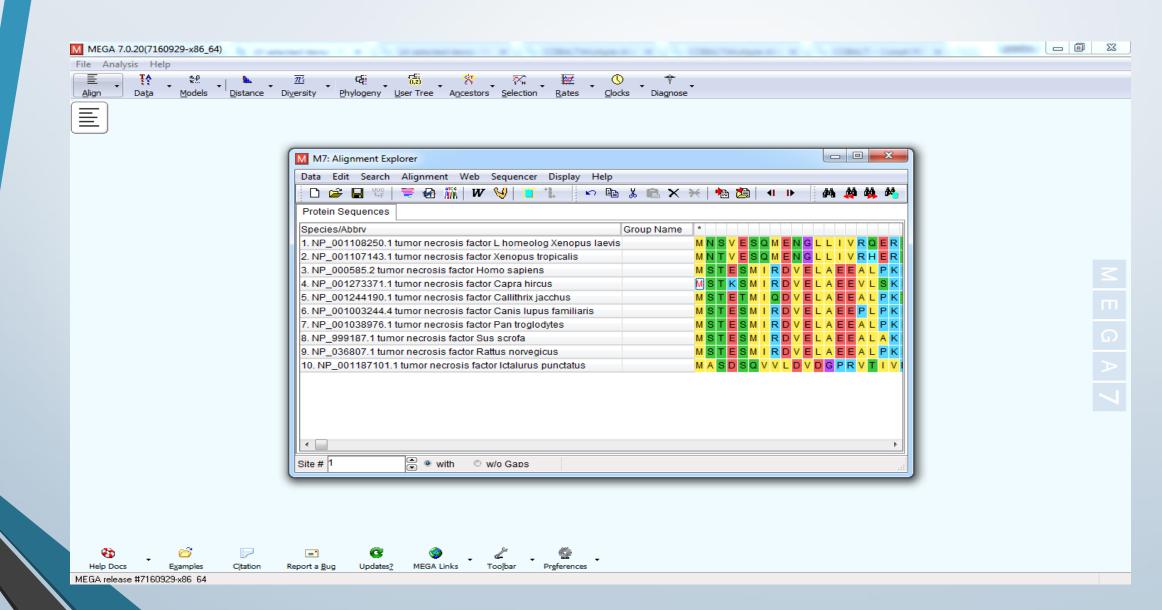


https://www.ncbi.nlm.nih.gov/tools/cobalt/re_cobalt.cgi

Results



MEGA Software



Results

