

# Package ‘st4gi’

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**Title** Statistical tools for genetic improvement

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**Description** Statistical tools for the analysis of experimental data for crop genetic improvement

**Depends** R (>= 3.1.2)

**Imports** lme4, shiny

**License** MIT + file LICENSE

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**LazyData** true

**Suggests** testthat

**NeedsCompilation** no

## R topics documented:

ammi	2
ammigxe	3
aovmet	5
checkdata01	6
checkdata02	6
domeans	7
elston	8
met8x12	9
msdplot	9
mveb	11
mvemet	12
pesekbaker	13
pjpz09	14
rsa	15
rts1	16
rts2	16
rts3	17
rts4	17

spconsis . . . . .	18
spg . . . . .	21
tai . . . . .	21

<b>Index</b>	<b>23</b>
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ammi	<i>AMMI or GGE with data at plot level</i>
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---

## Description

This function runs AMMI (Gollob, H. R., 1968) or GGE biplot (Yan , W. et al., 2000) with data at plot level.

## Usage

```
ammi(trait, geno, env, rep, data, method = "AMMI", f = 0.5, biplot = 2,
     biplot1 = "effects", title = NULL, xlab = NULL,
     color = c("darkorange", "black", "gray"), ...)
```

## Arguments

trait	The trait to analyze.
geno	The genotypes.
env	The environments.
rep	The replications or blocks. A RCBD is assumed.
data	The name of the data frame containing the data.
method	AMMI or GGE.
f	Scaling factor, defaults to 0.5.
biplot	Choose 1 for the trait-PC1 biplot and 2 for the PC1-PC2 biplot.
biplot1	Choose "effects" or "means" for biplot1.
title	Main title for biplot1 or biplot2.
xlab	Xlab for biplot1.
color	Color for lines, symbols and/or labels for environments, genotypes and axes.
...	Additional graphic parameters.

## Details

Significance of PCs are evaluated only with method = "AMMI" and if the data are balanced.

## Value

It returns the first and second PC values for genotypes and environments, a table with the contribution of each PC, a dispersion plot of means or effects against the first PC, or a dispersion plot of PC1 against PC2. Significance of PCs are included in the contributions table only if method is set to AMMI.

## Author(s)

Raul Eyzaguirre

## References

Gollob, H. R. (1968). A Statistical Model which combines Features of Factor Analytic and Analysis of Variance Techniques, *Psychometrika*, Vol 33(1): 73-114.

Yan, W. et al. (2000). Cultivar evaluation and mega-environment investigation based on the GGE biplot, *Crop Sci.*, Vol 40: 597-605.

## See Also

svd

## Examples

```
# The data
head(met8x12)
str(met8x12)

# Run AMMI for trait y, biplot1 by default
ammi("y", "geno", "env", "rep", met8x12)

# Run AMMI for trait y, biplot2
ammi("y", "geno", "env", "rep", met8x12, biplot = 2)
```

---

ammigxe

*AMMI or GGE with data from an interaction means matrix*

---

## Description

This function runs AMMI (Gollob, H. R., 1968) or GGE biplot (Yan , W. et al., 2000) with data from an interaction means matrix.

## Usage

```
ammigxe(int.mean, trait = NULL, rep.num = NULL, rdf = NULL, rms = NULL,
        method = "AMMI", f = 0.5, biplot = 2, biplot1 = "effects",
        title = NULL, xlab = NULL, color = c("darkorange", "black", "gray"),
        ...)
```

## Arguments

int.mean	GxE means matrix, genotypes in rows, environments in columns.
trait	Name of the trait.
rep.num	Number of replications.
rdf	Residual degrees of freedom.
rms	Residual mean square.
method	AMMI or GGE.
f	Scaling factor, defaults to 0.5.
biplot	1 for the trait-PC1 biplot and 2 for the PC1-PC2 biplot.
biplot1	Choose "effects" or "means" for biplot1.
title	Main title for biplot1 or biplot2.

xlab	Xlab for biplot1.
color	Color for lines, symbols and/or labels for environments, genotypes and axes.
...	Additional graphic parameters.

### Details

Significance of PCs are evaluated only with `method = "AMMI"` and if `rep.num`, `rms` and `rdf` are specified.

### Value

It returns the first and second PC values for genotypes and environments, a table with the contribution of each PC, a dispersion plot of means or effects against the first PC, or a dispersion plot of PC1 against PC2. Significance of PCs are included in the contributions table only if `rep.num`, `rms` and `rdf` are specified and `method` is set to AMMI.

### Author(s)

Raul Eyzaguirre

### References

Gollob, H. R. (1968). A Statistical Model which combines Features of Factor Analytic and Analysis of Variance Techniques, *Psychometrika*, Vol 33(1): 73-114.

Yan, W. et al. (2000). Cultivar evaluation and mega-environment investigation based on the GGE biplot, *Crop Sci.*, Vol 40: 597-605.

### See Also

`svd`

### Examples

```
# The data
head(met8x12)
str(met8x12)

# Compute GxE means
int.mean <- tapply(met8x12$y, list(met8x12$geno, met8x12$env), mean, na.rm = TRUE)

# Run AMMI with GxE means matrix, biplot2
ammigxe(int.mean, trait = "y", biplot = 2)

# Run GGE with GxE means matrix, biplot2
ammigxe(int.mean, trait = "y", method = "GGE", biplot = 2)
```

---

aovmet	<i>ANOVA for MET</i>
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---

## Description

Fit an analysis of variance model for a multi environment trial (MET).

## Usage

```
aovmet(trait, geno, env, rep, data, maxp = 0.05)
```

## Arguments

trait	The trait to analyze.
geno	The genotypes.
env	The environments.
rep	The replications or blocks.
data	The name of the data frame containing the data.
maxp	Maximum allowed proportion of missing values to estimate, defaults to 5%.

## Details

If data is unbalanced, missing values are estimated up to an specified maximum proportion, 5% by default. Genotypes and environments are considered as fixed factors while the blocks are considered as random and nested into the environments.

## Value

It returns the ANOVA table.

## Author(s)

Raul Eyzaguirre

## Examples

```
# The data
head(met8x12)
str(met8x12)

# Run ANOVA for MET
aovmet("y", "geno", "env", "rep", met8x12)
```

---

checkdata01	<i>Check data for a RCBD</i>
-------------	------------------------------

---

**Description**

This function checks the frequencies of genotypes in a RCBD.

**Usage**

```
checkdata01(trait, geno, data)
```

**Arguments**

trait	The trait to analyze.
geno	The genotypes.
data	The name of the data frame.

**Details**

This function checks if there is more than one replication in a RCBD, if there is any genotype without data, and if the design is balanced.

**Value**

c1, c2, c3, three control values.

**Author(s)**

Raul Eyzaguirre.

---

checkdata02	<i>Check data for a MET in a RCBD</i>
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---

**Description**

This function checks the frequencies of genotypes in each environment in a RCBD.

**Usage**

```
checkdata02(trait, geno, env, data)
```

**Arguments**

trait	The trait to analyze
geno	The genotypes
env	The environments
data	The name of the data frame

**Details**

This function checks if there is more than one replication in a RCBD in several environments, if there is any genotype without data for some specific environments, and if the design is balanced.

**Value**

c1, c2, c3, three control values

**Author(s)**

Raul Eyzaguirre

---

domeans	<i>Compute means over some factors</i>
---------	--

---

**Description**

Compute means for several traits for some specific factors.

**Usage**

```
domeans(traits, factors, addcol = NULL, data)
```

**Arguments**

traits	List of traits to compute means.
factors	List of factors.
addcol	Additional columns to keep.
data	The name of the data frame containing the data.

**Details**

This function computes means for all the traits for each level's combination of the factors. Additional columns can be kept if specified in addcol.

**Value**

It returns a data frame with the means.

**Author(s)**

Raul Eyzaguirre

**Examples**

```
# The data
head(spg)
str(spg)

# Compute means for all the traits across the two replications
# for each genotype and location.
traits <- c('rytha', 'bc', 'dm', 'star', 'nocr')
factors <- c('geno', 'loc')
domeans(traits, factors, data=spg)

# Save the output in a data.frame with name 'output1'
# and compute means for each genotype across the two locations.
output1 <- domeans(traits, factors, data=spg)
domeans(traits, 'geno', data=output1)
```

---

elston	<i>Elston Index</i>
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---

**Description**

Function to compute the Elston index (Elston, R. C., 1963).

**Usage**

```
elston(traits, geno, data, lb = 1)
```

**Arguments**

traits	List of traits.
geno	The genotypes.
data	The name of the data frame containing the data.
lb	Lower bound. 1 for $k = \min(x)$ and 2 for $k = (n * \min(x) - \max(x)) / (n - 1)$

**Details**

The Elston index is a weight free index.

**Value**

It returns the Elston index value and the Elston index value sorted in descending order.

**Author(s)**

Raul Eyzaguirre

**References**

Elston, R. C. (1963). A weight-free index for the purpose of ranking or selection with respect to several traits at a time. *Biometrics*. 19(1): 85-97.



## Examples

```
# The data
head(spg)
str(spg)

# Run Elston index with all the traits
elston(c("rytha", "bc", "dm", "star", "nocr"), "geno", spg)
```

---

met8x12	<i>Yields for a multi-environment trial (MET)</i>
---------	---

---

## Description

This data set gives the yields per plot for 8 genotypes in 12 environments with a RCBD with 3 blocks in each environment.

## Usage

```
met8x12
```

## Format

A data frame with 4 columns and 288 rows.

## Source

International Potato Center, sweetpotato experimental data.

---

msdplot	<i>Plot means and standard deviations with a dotplot</i>
---------	--

---

## Description

Function to plot means and confidence limits.

## Usage

```
msdplot(trait, groups, data, conf = 0.95, nmax = 10, dotplot = "TRUE",
  sort.means = "none", main.title = NULL, x.title = "groups",
  y.title = "", col = "black", bg = "darkorange", col.lines = "black")
```

**Arguments**

<code>trait</code>	The trait to plot.
<code>groups</code>	The grouping factor.
<code>data</code>	The name of the data frame containing the data.
<code>conf</code>	Probability for the confidence limits or number of standard deviations.
<code>nmax</code>	Maximum number of points for the compulsory dotplot, default is 10.
<code>dotplot</code>	Logical. If TRUE, a dotplot is shown. If FALSE it will only suppress the dots if the number of data points is larger than <code>nmax</code> .
<code>sort.means</code>	Sort for means. Options are "none", "increasing", and "decreasing", "none" by default.
<code>main.title</code>	Main title.
<code>x.title</code>	Title for x axis.
<code>y.title</code>	Title for y axis.
<code>col</code>	Line color for circles.
<code>bg</code>	Background color for circles.
<code>col.lines</code>	Line color for confidenced interval lines.

**Details**

An alternative to the controversial dynamite plots. If `conf` is set to a value greater than or equal to 1, then it is interpreted as number of standard deviations.

**Value**

It returns a plot with the means represented by circles, a line representing a confidence limit or a number of standard deviations, and alternatively the individual data points.

**Author(s)**

Raul Eyzaguirre

**Examples**

```
# Simulate some data
mydata <- data.frame(y = rnorm(50, sample(40:60, 5), sample(5:10, 5)),
                    g = rep(1:5, 10))

# Draw the plot
msdplot("y", "g", mydata)
```

mveb

*Estimation of missing values for a RCBD***Description**

Function to estimate missing values for a Randomized Complete Block Design (RCBD) by the least squares method.

**Usage**

```
mveb(trait, geno, rep, data, maxp = 0.05, tol = 1e-06)
```

**Arguments**

trait	The trait to estimate missing values.
geno	The genotypes.
rep	The replications or blocks. A RCBD is assumed.
data	The name of the data frame.
maxp	Maximum allowed proportion of missing values to estimate, defaults to 5%.
tol	Tolerance for the convergence of the iterative estimation process.

**Details**

A data.frame with data for a RCBD with at least two replications and at least one datum for each treatment must be loaded. Experimental data with only one replication, any treatment without data, or more missing values than specified in maxp will generate an error message.

**Value**

It returns a data frame with name new.data and the number est.num and proportion est.prop of estimated missing values. The new.data data frame contains the experimental layout and columns trait and trait.est with the original data and the original data plus the estimated values.

**Author(s)**

Raul Eyzaguirre.

**Examples**

```
# The data
head(met8x12)
str(met8x12)

# Choose one environment
temp <- subset(met8x12, env=="TM80N")

# Missing value in the first row
head(temp)

# Estimate the missing value
mveb("y", "geno", "rep", temp)
```

mvemet

*Estimation of missing values for a MET in a RCBD***Description**

Function to estimate missing values for a Multi Environment Trial (MET) with a Randomized Complete Block Design (RCBD) by the least squares method.

**Usage**

```
mvemet(trait, geno, env, rep, data, maxp = 0.05, tol = 1e-06)
```

**Arguments**

trait	The trait to estimate missing values.
geno	The genotypes.
env	The environments.
rep	The replications or blocks. A RCBD is assumed.
data	The name of the data frame.
maxp	Maximum allowed proportion of missing values to estimate, defaults to 5%.
tol	Tolerance for the convergence of the iterative estimation process.

**Details**

A data.frame with data for a MET in a RCBD with at least two replications and at least one datum for each treatment must be loaded. Experimental data with only one replication, any treatment without data, or more missing values than specified in maxp will generate an error message.

**Value**

It returns a data frame with name new.data and the number est.num and proportion est.prop of estimated missing values. The new.data data frame contains the experimental layout and columns trait and trait.est with the original data and the original data plus the estimated values.

**Author(s)**

Raul Eyzaguirre.

**Examples**

```
# The data
head(met8x12)
str(met8x12)

# Estimate the missing values
mvemet("y", "geno", "env", "rep", met8x12)
```

pesekbaker

*Pesek-Baker Index***Description**

Function to compute the Pesek-Baker index (Pesek, J. and R.J. Baker., 1969).

**Usage**

```
pesekbaker(traits, geno, env, rep, data, dgg = NULL, units = "sdu",
           sf = 0.1)
```

**Arguments**

traits	List of traits.
geno	The genotypes.
env	The environments.
rep	The replications or blocks.
data	The name of the data frame containing the data.
dgg	Desired genetic gains, defaults to one standard deviation for each trait.
units	Units for dgg, "actual" or "sdu". See details for more information.
sf	Selected fraction, defaults to 0.1.

**Details**

The Pesek-Baker is an index where relative economic weights have been replaced by desired gains. If dgg is not specified, the standard deviations of the traits are used. It means that the desired genetic gains are equal to one standard deviation for each trait. dgg can be specified in actual units (units = "actual") or in standard deviations (units = "sdu"), defaults to "sdu". For example, if you have a trait which is expressed in kilograms and with a standard deviation of 5 kilograms, typing dgg = 2 means a desired genetic gain of 2 standard deviations that corresponds to 10 kilograms. If you type dgg = 2 and units = "actual" then this means a desired genetic gain of 2 kilograms. If dgg = NULL then the desired genetic gain will be one standard deviation, no matter if units is set as "actual" or "sdu". To compute the index the package lme4 is needed.

**Value**

It returns:

- \$Desired.Genetic.Gains, the desired genetic gains in actual units,
- \$Standard.Deviations, the estimated standard deviations,
- \$Genetic.Variances, the estimated genetic variances,
- \$Correlation.Matrix, the estimated correlation matrix,
- \$Index.Coefficients, the index coefficients,
- \$Response.to.Selection, the response to selection,
- \$Std.Response.to.Selection, the standardized response to selection,
- \$Pesek.Baker.Index, the Pesek-Baker index value, and
- \$Sorted.Pesek.Baker.Index the Pesek-Baker index value sorted in descending order.

**Author(s)**

Raul Eyzaguirre

**References**

Pesek, J. and R.J. Baker.(1969). Desired improvement in relation to selection indices. Can. J. Plant. Sci. 9:803-804.

**Examples**

```
# The data
head(spg)
str(spg)

# Run Pesek-Baker index with all the traits
pesekbaker(c("rytha", "bc", "dm", "star", "nocr"), "geno", "loc", "rep", spg)

# Use different desired genetic gains for each trait,
# more weight on bc and dm, less on star and nocr.
pesekbaker(c("rytha", "bc", "dm", "star", "nocr"), "geno", "loc", "rep", spg,
           dgg = c(1, 1.5, 1.5, 0.8, 0.8))
```

---

pjpz09

*Data for a yield trial*

---

**Description**

This data set contains data for several traits for a one location experiment.

**Usage**

pjpz09

**Format**

A data frame with 10 columns and 204 rows.

**Source**

International Potato Center, sweetpotato experimental data.

---

**rsa***Regression Stability Analysis*

---

**Description**

Function to run the regression stability analysis (Yates and Cochran, 1938, Finlay and Wilkinson, 1963). This implementation follows the formulas of Eberhart and Russell (1966).

**Usage**

```
rsa(trait, geno, env, rep, data, maxp = 0.05)
```

**Arguments**

trait	The trait to analyze.
geno	The genotypes.
env	The environments.
rep	The replications or blocks.
data	The name of the data frame containing the data.
maxp	Maximum allowed proportion of missing values to estimate, defaults to 5%.

**Details**

The regression stability analysis is evaluated with a balanced data set. If data is unbalanced, missing values are estimated up to an specified maximum proportion, 5% by default. For the ANOVA table, genotypes and environments are considered as fixed factors while the blocks are considered as random and nested into the environments. To run a regression stability analysis you need a set of genotypes evaluated in a set of environments. At least 3 genotypes or environments are needed. In a regression stability analysis for genotypes grown at several environments, for each genotype a simple linear regression of individual yield (Y) on the mean yield of all genotypes for each environment (X) is fitted. In a similar way, for each environment, a simple linear regression of individual yield (Y) on the mean yield of all environments for each genotype (X) is fitted.

**Value**

It returns the regression stability analysis for genotypes and environments.

**Author(s)**

Raul Eyzaguirre

**References**

- Eberhart, S. A. and Russell, W. A. (1966). Stability Parameters for Comparing Varieties. *Crop Sci.* 6: 36-40.
- Finlay, K. W., and Wilkinson, G. N. (1963). The Analysis of Adaption in a Plant-Breeding Programme. *Aust. J. Agric. Res.* 14: 742-754.
- Yates, F., and Cochran, W. G. (1938). The Analysis of Group Experiments. *J. Agric. Sci.* 28: 556-580.

**Examples**

```
# The data
head(met8x12)
str(met8x12)

# Run regression stability analysis
rsa("y", "geno", "env", "rep", met8x12)
```

---

rts1*Response to selection for a single experiment*

---

**Description**

It finds the optimum number of replications to get the maximum response to selection for a single experiment for a given plot capacity, number of selected genotypes, genotypic variance and error variance.

**Usage**

```
rts1()
```

**Details**

It uses package shiny for the web layout. Type `rts1()` in the R console to run the app.

**Value**

It returns a plot of response to selection versus number of replications and computes the optimum number of replications and the response to selection at this optimum value.

**Author(s)**

Raul Eyzaguirre.

---

rts2*Response to selection with several locations*

---

**Description**

It finds the optimum number of replications to get the maximum response to selection with several locations for a given plot capacity, number of locations, number of selected genotypes, genotypic variance, genotypic by location variance, and error variance.

**Usage**

```
rts2()
```

**Details**

It uses package shiny for the web layout. Type `rts2()` in the R console to run the app.



**Value**

It returns a plot of response to selection versus number of replications and computes the optimum number of replications and the response to selection at this optimum value.

**Author(s)**

Raul Eyzaguirre.

---

 rts3

---

*Response to selection with several locations and years*


---

**Description**

It finds the optimum number of replications to get the maximum response to selection with several locations and years for a given plot capacity, number of locations, number of years, number of selected genotypes, genotypic variance, genotypic by location variance, genotypic by year variance, genotypic by location by year variance, and error variance.

**Usage**

```
rts3()
```

**Details**

It uses package shiny for the web layout. Type `rts3()` in the R console to run the app.

**Value**

It returns a plot of response to selection versus number of replications and computes the optimum number of replications and the response to selection at this optimum value.

**Author(s)**

Raul Eyzaguirre.

---

 rts4

---

*Response to selection with several locations in two steps (two years)*


---

**Description**

It computes the response to selection for each step in a two steps selection with several locations for a given number of genotypes at step 1, the number of locations, replications and selected genotypes at step 1, the number of locations, replications and selected genotypes at step 2, and the genotypic, genotypic by location, genotypic by year, genotypic by location by year, and error variances.

**Usage**

```
rts4()
```

**Details**

It uses package shiny for the web layout. Type `rts4()` in the R console to run the app.

**Value**

It returns the response to selection at step 1 and 2.

**Author(s)**

Raul Eyzaguirre.

---

spconsis

*Check consistency for sweetpotato experimental data*


---

**Description**

Set of rules to check for consistency of sweetpotato experimental data. Data labels must be defined as specified in the PROCEDURES FOR THE EVALUATION AND ANALYSIS OF SWEET-POTATO TRIALS document.

**Usage**

```
spconsis(data, plot.size, width = 240)
```

**Arguments**

<code>data</code>	The name of the data frame.
<code>plot.size</code>	Plot size in square meters.
<code>width</code>	Number of columns for the output file.

**Details**

The data frame must use the following labels:

- L : Locations
- Y : Year
- S : Season
- G : Genotypes
- NAME : Names for genotypes
- E : Environments
- R : Replications or blocks
- NOPS : Number of plants planted
- NOPE : Number of plants established
- VIR1 : Virus symptoms (1-9), first evaluation
- VIR2 : Virus symptoms (1-9), second evaluation
- VIR3 : Virus symptoms (1-9), third evaluation
- ALT1 : Alternaria symptoms (1-9), first evaluation

- ALT2 : Alternaria symptoms (1-9), second evaluation
- VV1 : Vine vigor (1-9), first evaluation
- VV2 : Vine vigor2 (1-9), second evaluation
- VW : Vine weight
- NOPH : Number of plants harvested
- NOPR : Number of plants with roots
- NOCR : Number of commercial roots
- NONC : Number of non commercial roots
- CRW : Commercial root weight
- NCRW : Non commercial root weight
- RFCP : Root primary flesh color
- RFCS : Root secondary flesh color
- SCOL : Storage root skin color
- FCOL : Storage root flesh color
- RS : Root size (1-9)
- RF : Root form (1-9)
- DAMR : Root defects (1-9)
- RSPR : Root sprouting (1-9)
- WED1 : Weevil damage (1-9), first evaluation
- WED2 : Weevil damage2 (1-9), second evaluation
- DMF : Fresh weight of roots for dry matter assessment
- DMD : Dry weight of DMF samples
- DMVF : Fresh weight vines for dry matter assessment
- DMVD : Dry weight of DMVF samples
- DM : Storage root dry matter content (
- DMFY : Dry matter foliage yield
- DMRY : Dry matter root yield
- FRAW1 : Root fiber (1-9), first determination
- SURAW1 : Root sugar (1-9), first determination
- STRAW1 : Root starch (1-9), first determination
- COOF1 : Cooked fiber (1-9), first evaluation
- COOSU1 : Cooked sugars (1-9), first evaluation
- COOST1 : Cooked starch (1-9), first evaluation
- COOT1 : Cooked taste (1-9), first evaluation
- COOAP1 : Cooked appearance (1-9), first evaluation
- FRAW2 : Root fiber (1-9), second determination
- SURAW2 : Root sugar (1-9), second determination
- STRAW2 : Root starch (1-9), second determination
- COOF2 : Cooked fiber (1-9), second evaluation
- COOSU2 : Cooked sugars (1-9), second evaluation

- COOST2 : Cooked starch (1-9), second evaluation
- COOT2 : Cooked taste (1-9), second evaluation
- COOAP2 : Cooked appearance (1-9), second evaluation
- PROT : Protein
- FE : Iron in dry weight
- ZN : Zinc in dry weight
- CA : Calcium in dry weight
- MG : Magnesium in dry weight
- BC : Beta-carotene in dry weight (NIRS)
- BC.CC : Beta-carotene with color charts
- TC : Total carotenoids in dry weight (NIRS)
- STAR : Starch
- FRUC : Fructose
- GLUC : Glucose
- SUCR : Sucrose
- MALT : Maltose
- TRW : Total root weight
- CYTHA : Commercial root yield t/ ha
- RYTHA : Total root yield t/ha
- ACRW : Average commercial root weight = CRW/NOCR
- NRPP : Number of roots per plant
- YPP : Yield per plant Kg
- CI : Percent marketable roots (commercial index)
- HI : Harvest index
- SHI : Harvest sowing index (survival)
- BIOM : Biomass yield
- FYTHA : Foliage total yield t/ha
- RFR : Root foliage ratio

### Value

It returns a file with name checks.txt with a list of all rows with some kind of inconsistency and all rows with outliers.

### Author(s)

Raul Eyzaguirre.

### Examples

```
# The data
head(pjpz09)
str(pjpz09)

# Check the data
spconsis(pjpz09, 4.5)
```

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spg	<i>Some traits for a multi-environment trial (MET)</i>
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---

**Description**

This data set has data for root yield in tons per hectare (rytha), beta-carotene (bc), dry matter (dm), starch (star) and number of commercial roots (nocr) for 8 genotypes in 2 locations with a RCBD with 2 blocks in each location.

**Usage**

```
spg
```

**Format**

A data frame with 8 columns and 32 rows.

**Source**

International Potato Center, sweetpotato experimental data.

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tai	<i>Tai's stability analysis</i>
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**Description**

This function runs Tai's stability analysis (Tai, G. C. C., 1971). It assumes a RCBD with fixed effects for genotypes and random effects for environments.

**Usage**

```
tai(trait, geno, env, rep, data, conf = 0.95, title = NULL,
    color = c("darkorange", "black", "gray"), ...)
```

**Arguments**

trait	The trait to analyze.
geno	The genotypes.
env	The environments.
rep	The replications or blocks. A RCBD is assumed.
data	The name of the data frame containing the data.
conf	Probability for the Tai limits.
title	Main title for plot.
color	Color for symbols, labels and lines.
...	Additional graphic parameters.

**Details**

The limits for alpha and lambda are computed using the mean squares from an ANOVA table for a RCBD with blocks nested into environments. If the data set is unbalanced, a warning is produced.

**Value**

It returns the Tai graph for stability analysis and the values of alpha and lambda for each genotype.

**Author(s)**

Raul Eyzaguirre

**References**

Tai, G. C. C. (1971). Genotypic Stability Analysis and Its Application to Potato Regional Trials, Crop Science, Vol 11.

**Examples**

```
# The data
head(met8x12)
str(met8x12)

# Run Tai for trait y
tai("y", "geno", "env", "rep", met8x12)
```

# Index

ammi, [2](#)  
ammigxe, [3](#)  
aovmet, [5](#)  
  
checkdata01, [6](#)  
checkdata02, [6](#)  
  
domeans, [7](#)  
  
elston, [8](#)  
  
met8x12, [9](#)  
msdplot, [9](#)  
mveb, [11](#)  
mvemet, [12](#)  
  
pesekbaker, [13](#)  
pjpz09, [14](#)  
  
rsa, [15](#)  
rts1, [16](#)  
rts2, [16](#)  
rts3, [17](#)  
rts4, [17](#)  
  
spconsis, [18](#)  
spg, [21](#)  
  
tai, [21](#)