#### **MINIREVIEWS**

# Ocean Warming and Spread of Pathogenic Vibrios in the Aquatic Environment

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**Abstract** Vibrios are among the most common bacteria that inhabit surface waters throughout the world and are responsible for a number of severe infections both in humans and animals. Several reports recently showed that human Vibrio illnesses are increasing worldwide including fatal acute diarrheal diseases, such as cholera, gastroenteritis, wound infections, and septicemia. Many scientists believe this increase may be associated with global warming and rise in sea surface temperature (SST), although not enough evidence is available to support a causal link between emergence of Vibrio infections and climate warming. The effect of increased SST in promoting spread of vibrios in coastal and brackish waters is considered a causal factor explaining this trend. Field and laboratory studies carried out over the past 40 years supported this hypothesis, clearly showing temperature promotes Vibrio growth and persistence in the aquatic environment. Most recently, a long-term retrospective microbiological study carried out in the coastal waters of the southern North Sea provided the first experimental evidence for a positive and significant relationship between SST and Vibrio occurrence over a multidecadal time scale. As a future challenge, macroecological studies of the effects of ocean warming on Vibrio persistence and spread in the aquatic environment over large spatial and temporal scales would conclusively support evidence acquired to date combined with studies of the impact of global warming on epidemiologically relevant variables, such as host susceptibility and exposure.

Assessing a causal link between ongoing climate change and enhanced growth and spread of vibrios and related illness is expected to improve forecast and mitigate future outbreaks associated with these pathogens.

### Introduction

Warming of the climate system is unequivocal, with evidence from observations of increases in global average air and ocean temperatures, widespread melting of snow and ice, and rising global average sea level [1]. Global average temperatures have risen by nearly 0.8 °C since the late nineteenth century and approximately 0.2 °C/decade over the past 25 years [1]. It has been estimated that since the 1960s, about 90 % of the excess heat added to the Earth's climate system has been stored in the oceans [2]. In Europe, warming of regional seas has accelerated at an unprecedented rate over the last 25 years, with the Baltic, North, and Black Seas showing greatest increasing temperature [3]. At the global scale, temperature fluctuations in surface waters have been related to large-scale climate patterns such as the North Atlantic Oscillation (NAO) and El-Niño Southern Oscillation (ENSO) [4]. Altered patterns of precipitation and run-off related to global warming are also driving reduction in salinity in estuaries and coastal wetlands [5].

Vibrio spp. are naturally occurring bacteria in riverine, coastal, and estuarine ecosystems around the world and include several human and animal pathogens that can negatively impact human health and ecosystem services (e.g., recreational water use and aquaculture) [6]. Over 80 species have now been described, including at least 12 capable of causing infection in humans (e.g., Vibrio cholerae, Vibrio parahaemolyticus, and Vibrio vulnificus) [7]. V. cholerae, the causative agent of cholera, is by far the most important pathogenic species of the genus, accounting for about three million cases of human infections each year, with a case

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fatality rate of about 2.4 % [8]. Overall, *Vibrio* diseases deriving from both water and seafood are increasing worldwide. The number of cholera cases reported to WHO continues to rise, and for 2011 alone, a total of 589,854 cases and 7,816 deaths were reported by 58 countries. Surveillance data from both COVIS and FoodNet indicate the incidence of vibriosis (excluding toxigenic *V. cholerae* O1 and O139) increased in the USA from 1996 to 2010 [9].

Vibrios grow preferentially in warm (>18 °C), low salinity (<25 ppt NaCl) seawater and brackish waters. Ongoing warming and reduced salinity of coastal regions are expected to support the spread of these bacteria at a global scale, especially in northern latitudes. Increased seawater temperature over the past decade has been linked to outbreaks of Vibrio-associated human illness caused by V. cholerae, V. parahaemolyticus, and V. vulnificus including Chile [10], Peru [11], Alaska [12], Israel [13], Spain [14], and Europe [15]. A geographical transition in terms of disease into non-endemic regions has also been observed especially in temperate and cold regions of the world [15] which are the most affected by global warming [16]. Warming patterns have also been correlated with an increase in cholera cases in cholera-endemic countries of sub-Saharan Africa [17] and the Bay of Bengal [18, 19].

In addition to human illnesses, prevalence and severity of a wide range of diseases of marine organisms, e.g., corals, bivalves, and fish, have been linked to elevated sea surface temperature (SST) and Vibrio infections. Coral bleaching is the most serious disease affecting coral reefs, and of the eight coral pathogens implicated in the onset of coral diseases worldwide, half belong to the Vibrionaceae family [20, 21]. In the Mediterranean Sea, Vibrio corallilyticus infections were shown to trigger mass mortality events of the purple gorgonian Paramuricea clavata, a key structuring species of coralligenous assemblages, related to SST anomalies registered during recent years [22]. Other Vibrio species, such as Vibrio splendidus and Vibrio aestuarianus, are believed to play a role in the abnormal mortality of Pacific oysters (Crassostrea gigas), which has increased since 2008 in warmer seasons in some areas of Ireland, France, the Netherlands, and the UK [23, 24].

Notwithstanding the increasing number of reports showing an unequivocal positive correlation between increasing environmental temperature and spread of *Vibrio* diseases, conclusive evidence linking emergence of infections with climate change is lacking. The main concern is that insufficient experimental evidence has been accumulated to date to support a causal link. Contrasting reports such as the recent observation that vibriosis has increased significantly in the USA in the last decade [9, 25] also in areas where long-term coastal temperature trends are weakly cooling or not significant (South Atlantic Bight  $[-0.1\pm0.3~^{\circ}\text{C}~(100~\text{years})]$  and off Florida  $[-0.3\pm0.2~^{\circ}\text{C}~(100~\text{years})]$  [26] might be also

taken into account suggesting that factors additional than climate are likely to be responsible for driving disease.

More effort should be devoted in the future to study the long-term climate change effects on *Vibrio* pathogen development and survival in the aquatic environment. In addition, for full comprehension of climate change effects on *Vibrio* disease transmission, the influence of a changing climate on exposure patterns (access to drinking and recreational water) and host susceptibility (health status) must be taken into account. Several reports have been published recently that deal with these issues [27, 28]. This review provides succinct update and discussion of the current state of knowledge regarding effects of increasing SST may have on *Vibrio* persistence and spread in the aquatic environment.

# **Ecology of Vibrios in a Warming Ocean: Knowledge and Gaps**

Understanding the ecology of vibrios, namely the role played by environmental factors in survival, growth, and dispersion of these bacteria, is crucial to understand fully their interaction with humans and animals and, consequently, epidemiology of the associated diseases. Vibrios are natural inhabitants of the brackish and marine environment throughout the world and, although our knowledge has increased substantially in recent years [29, 30], we are still far from having a comprehensive view of their global ecology. Most studies have focused on the population ecology of single species (mainly *V. cholerae*) and generally following a reductionist approach, namely lab-scale experiments, and examining the role of a single/few environmental factors affecting Vibrio growth and survival [31–33]. Findings from these studies have provided important information about the ecology of vibrios at the molecular, cell, and population levels, namely conditions and resources regulating their metabolism and determining their ecological niche. Conditions include physicochemical parameters, such as temperature, salinity, and pH, whilst environmental resources include carbon and energy needed for metabolism and consumption by microorganisms. Several studies have shown that vibrios are chemoheterotrophic halotolerant microorganisms (optimal NaCl concentration is between 0.2 and 3.0 %), with an optimal temperature of growth ranging from 30 to 40 °C and optimal pH of ca. 8.0 [7].

Furthermore, studies conducted both in the laboratory and field showed that, together with environmental conditions and resources, interactions of vibrios with other living organisms and abiotic substrates are the main contributors in shaping the ecological niche of these species [34, 35]. For example, in the aquatic environment, *V. cholerae* has been reported to be associated with a variety of living organisms, including animals with an exoskeleton of chitin, aquatic



plants, protozoa, bivalves, waterbirds, as well as abiotic substrates (e.g., sediments) which represent environmental reservoirs (living or nonliving substrate favoring bacterial persistence and replication in the environment) and/or hosts (living organism that temporarily harbors the bacteria, generally providing nourishment and shelter) [36]. Both the reservoir and host largely influence the ecology of vibrios by favoring their survival and dispersion in the environment and also serving as a vector of their associated disease. Interaction between Vibrio and environmental surfaces, namely chitin and the exoskeleton of marine crustaceans, was investigated extensively for *V. cholerae* and exemplifies a successful bacteria-substrate interaction with complex and significant influence on the lifestyle of the bacterium [35]. In fact, the *V. cholerae*—chitin interaction has been shown to positively influence food availability, adaptation to environmental nutrient gradients, tolerance to stress, and protection from predators [35]. Trophic interactions also play a significant role in controlling Vibrio proliferation in coastal waters and these include bottom upregulation by the food resource, such as dissolved organic carbon and nutrients [37], as well as top downregulation by protozoan predation [38] and lysis by bacteriophage [39].

In spite of their importance, these studies are limited in not having addressed the influence of environmental variables at the global ecosystem scale. Extreme weather events, human-induced eutrophication, and pollution, as well as large-scale climatic processes (i.e., ENSO and NAO) affect both ecological resources and conditions and the ecological interactions within a biological community. These factors are expected to directly influence multiple levels of organization in an aquatic system, as well as trigger indirect cascading effects, from ecosystem to single cells, all of which are difficult to measure, especially simultaneously. For example, global warming is known to affect the physiology of *Vibrio* cells directly, but also to influence abundance and structure of their main environmental reservoirs [3].

Studies aimed at addressing the challenging field of *Vibrio* global ecology are difficult to design and perform; they are carried out mainly in field investigations of *Vibrio* populations, that is, temporal and spatial occurrence of *Vibrio* populations and their relation to detected ecosystem variables [40–42]. Although very useful, such studies tend to be descriptive and based on correlation and not "cause and effect." Ecological modeling is a promising method to achieve better understanding of *Vibrio* global ecology, but it is largely constrained by the lack of sufficient experimental observations.

In conclusion, the reductionist approach taken to date, although successful in providing a detailed understanding of *Vibrio* species, underestimates the complexity of the ecosystem, notably that of climate change effects on *Vibrio* 

populations (Fig. 1 provides a schematic overview of the direct and indirect influence of ocean warming on vibrios). It should be pointed out that holistic studies are ambitious and difficult to perform in a rigorous scientific way. As a future challenge, macroecological studies investigating statistical patterns of abundance, distribution, and activity of *Vibrio* bacteria over large spatial and temporal scales can help close the gaps providing the basis for dealing with threats that climate linked *Vibrio* proliferation may pose to human health.

# Temperature Effects on Vibrio Viability, Growth, and Pathogencity

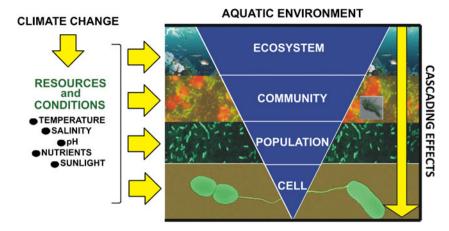
It has long been established that temperature strongly influences metabolism and growth rates of *Vibrio* species. Early studies of *V. cholerae* showed that by increasing the water temperature up to 30 °C in laboratory microcosms, a pronounced effect on the multiplication of these bacteria is observed [32]. The same phenomenon has been observed for other *Vibrio* species, including those pathogenic for humans, namely *V. parahaemolyticus* [43] and *V. vulnificus* [44].

Most vibrios grow well on culture media when the temperature is above 17 °C. The optimal temperature for growth may vary among different species, but is generally in the range of 30–40 °C [7]. Vibrios are sensitive to cold temperatures below 10 °C when they may enter the viable but nonculturable (VBNC) state. The VBNC state, firstly described for the human pathogen *V. cholerae*, is a dormant condition in which bacteria remain viable, but are not culturable in conventional laboratory media [45]. Temperature upshift was shown to promote regrowth of several VBNC *Vibrio* cells which regain culturability and normal phenotypic traits [46].

Recent studies have shown that temperature has an effect on *Vibrio* pathogenicity. Using proteomic analysis and bioassays, Kimes et al. [47] demonstrated direct temperature regulation of virulence in the coral pathogen *V. coralliilyticus*, including upregulation at 27 °C of several virulence factors involved in motility, host degradation, secretion, and antimicrobial resistance [47]. Increased expression of known and putative virulence-associated traits was also correlated with increased temperature in clinical reference isolates of *V. vulnificus* and *V. parahaemolyticus* [48, 49].

Ultimately, for *V. cholerae*, temperature also appears to play a role in conversion of *V. cholerae* from non-O1 to O1 serogroup [50]. In addition, it regulates transcription of *toxT*, a gene encoding the regulatory protein ToxT which directly activates transcription of virulence factors, including cholera toxin (CT) and the toxin-coregulated pilus (TCP) [51]. The expression of CT and TCP via ToxT is downregulated in *V.* 





**Figure 1** Simplified overview of the impact of climate change on vibrios in the aquatic environment. Climate change is affecting ecological resources and conditions required by biological species. These are directly influencing *Vibrio* cells and populations (e.g., *Vibrio* metabolism and growth) as well as higher levels of organization of the aquatic system (e.g., living communities and the

physicochemical environment). The impact of climate change on the upper levels of ecosystem organization may ultimately trigger indirect cascading effects that could have profound repercussion on the vibrios (e.g., by affecting *Vibrio* environmental reservoirs and trophic interactions)

cholerae at 37 °C and upregulated at 30 °C. It was suggested that negative regulation of ToxT-dependent transcription by temperature and other environmental signals prevents incorrect temporal and spatial expression of virulence factors during the host phase of the *V. cholerae* life cycle [51, 52].

### Temperature Effects on *Vibrio* Interaction with Environmental Reservoirs and Hosts

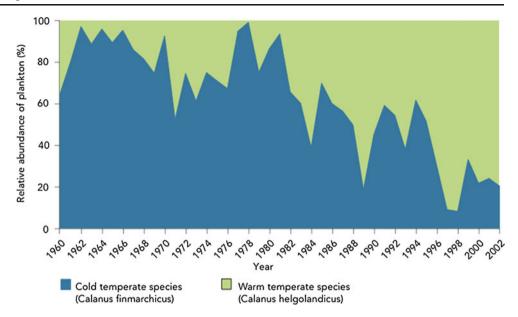
Although much is known about direct effects of temperature on the physiology of Vibrio cells, there is very limited information available on the indirect effects that an increase in environmental temperature may have on Vibrio populations by altering the abundance and structure of Vibrio reservoirs and hosts in the aquatic environment. Vibrios have a high affinity for colonization of chitin surfaces and chitinous organisms, especially zooplankters, which are considered perhaps the main environmental reservoir of these bacteria in the aquatic environment [36]. The copepod exoskeleton has been shown to support large populations of vibrios, including the pathogenic species, V. cholerae [53]. Due to its clinical importance, V. cholerae has been used as a model to study Vibrio-plankton interaction. The exoskeleton surface of a single colonized copepod has been shown to contain up to 10<sup>4</sup> cells of V. cholerae, thus providing the required infectious dose for clinical cholera (ranging from 10<sup>4</sup> to 10<sup>11</sup> depending on the bacterial strain and its host [54]). The importance of copepods in cholera transmission was further demonstrated in a study in which the number of cholera cases in Bangladesh villages was reduced significantly when copepods were removed from drinking water [55].

The role of temperature in *V. cholerae*–plankton interactions was studied by Huq et al. [56] who showed that temperatures ranging from 25 to 30 °C significantly promote attachment of *V. cholerae* to copepods. More recently, Stauder et al. [57] analyzed the molecular basis of the attachment and suggested increased ambient temperature significantly enhanced expression of two colonization factors involved in *V. cholerae* interaction with environmental chitin surfaces, namely *N*-acetylglucosamine-binding protein A (GbpA) and mannose-sensitive hemagglutinin [58, 59]. According to their findings, ocean warming may favor *Vibrio* colonization of plankton, amplifying the role of the plankton reservoir in supporting larger populations of vibrios in the aquatic environment.

Ocean warming affects abundance, diversity, and distribution of plankton communities. In the eastern North Atlantic and on the European shelf, strong biogeographical shifts in copepod assemblages occurred during the last 50 years, with a poleward extension of more than 10° latitude (more than 1,000 km) for warm-water species and a decrease in the number of colder-water species [60] (Fig. 2). This coincided with an observed increase in planktonic biodiversity and parallel reduction in phytoplankton [61] and zooplankton size [3]. Little is known about the effect that ongoing dramatic changes in the plankton community may have on Vibrio populations or, as yet unknown environmental reservoirs or hosts of these bacteria [36]. Gaps in our knowledge of Vibrio ecology will need to be addressed to answer questions concerning environmental reservoirs of these bacteria including species-specific interaction between vibrios and hosts [62].



Figure 2 Long-term changes in plankton assemblages in the North Atlantic and North Sea spanning four decades. Plankton data recorded by the CPR survey in the North Atlantic and North Sea show the increase in dominance of warm-water copepod species (Calanus helgolandicus) (green) and concomitant decrease of cold-water species (Calanus finmarchicus) over the period 1960-2002. Copyright© 2010 Sir Alister Hardy Foundation for Ocean Science. All Rights Reserved. **SAHFOS** 



### Field Studies of Seasonal Effect of SST on *Vibrio* Abundance and Distribution

Most of the studies cited above employed laboratory model systems to investigate the effect of increased environmental temperatures on *Vibrio* populations. However laboratory experiments are simplified representations of the natural ecosystem and need to be integrated with observational studies in the field.

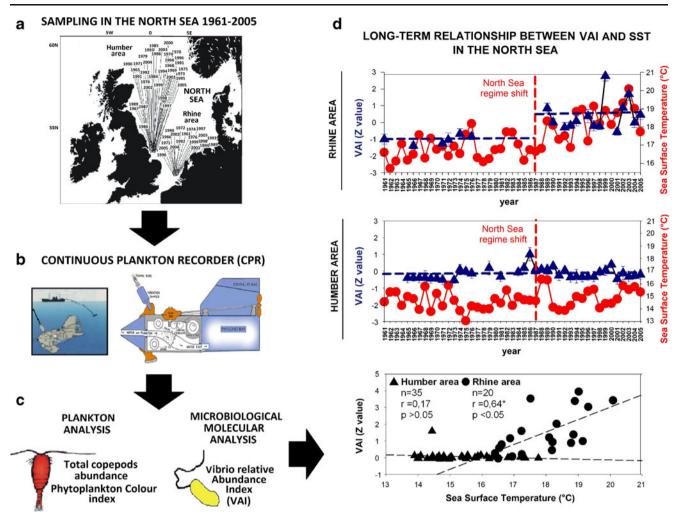
Field studies conducted worldwide over the last 40 years have clearly showed the number of Vibrio in estuarine and coastal waters is strongly associated with seasonal temperature [6]. For example, in temperate regions of the world, the abundance of V. cholerae and V. parahaemolyticus, both of which occupy similar ecological niches, show a strong seasonal oscillation with higher abundances generally observed during the warmer season of the year, that is when the water temperature typically exceeds 20 °C [63]. Similar temperature-driven dynamics have been observed for V. vulnificus [64] and other species of Vibrio [22, 65]. Thompson et al. [66] observed that temperature influences the overall structure of Vibrio populations in field study conducted in the North Atlantic Ocean by identifying distinct warm-water and year-round populations of vibrios over a 15-month period. Seasonal changes in temperature are known to directly influence both bacteria and their aquatic reservoirs and hosts. In situ data collected in analyses of zooplankton and water samples from coastal waters of the NW Mediterranean Sea over an annual cycle showed that the percentage of plankton-associated V. cholerae was positively correlated with SST, increasing dramatically at temperatures above 22 °C [57]. These data are consistent with the laboratory observations showing elevated temperatures promote attachment of *V. cholerae* to copepods [32]. Whether the decline in Vibrio abundance during cold seasons of the year is the result of temperature-mediated cell mortality or induction of the VBNC state or to impact on the bacterial host or vector remains open to debate [67].

Seasonal temperature affects the incidence of Vibriorelated diseases and epidemiology [68]. Analysis of cholera outbreaks has shown that temperature and other variables, such as salinity and plankton concentration, influence seasonal transmission of cholera in those regions of the world where the human population relies on untreated water as a source of drinking water [69]. In addition, ENSO events have been found to be related to anomalous patterns in cholera dynamics and other Vibrio infections [29, 68]. Using remote sensing, Lobitz et al. [19] were the first to explore the relationship between SST and cholera incidence over a multi-seasonal scale, finding changes in coastal ecosystems to be significantly related to the seasonal pattern of cholera epidemics. Based on those studies, a hierarchical model has been proposed for cholera epidemiology, defining the role of environment, weather, and climate-related variables in cholera outbreaks [29].

# **Long-term Effects of Increasing SST on** *Vibrio* **Populations**

Long-term effects of ocean warming on abundance and distribution of vibrios in the aquatic environment are difficult to assess because of the lack of historical data. Most recently, an ingenious retrospective study of formalin-fixed plankton samples collected by the Continuous Plankton Recorder (CPR) survey in the North Sea has provided the first evidence for a positive and significant relationship between SST and *Vibrio* abundance over a multidecadal scale (Fig. 3). In the region around the North Sea sampling sites, an unprecedented increase in bathing infections





**Figure 3** Retrospective assessment of long-term relative abundance of vibrios in the North Sea. Formalin-fixed plankton samples were collected by the CPR survey off the Rhine and Humber estuaries, in August, over the period 1961 to 2005 (a). CPR samples were collected by the continuous plankton recorder (b). Back in the laboratory, the silk containing the formalin-fixed plankton was cut into blocks and microbiological molecular analyses were carried out to assess *Vibrio*-relative abundance (see Vezzulli et al. [70] for details) (c). Phytoplankton and zooplankton analysis was also carried out following standard CPR

methodology [71] (c). Long-term variation in standardized (Z) VAI (triangles; error bars indicate standard deviation, n=5) and SST (circles) for 1961–2005 off the Rhine and Humber estuaries in the North Sea (d).  $Vertical\ line=$ regime shift step change in temperature after 1987 [72].  $Horizontal\ lines=$ average VAI values. The Pearson correlation analysis between VAI and SST in the North Sea (Pearson's correlation on pooled data; n=55; r=0.27\*; P<0.05) (d). Z values are obtained by subtracting the population mean and dividing the difference by the standard deviation. Modified from Vezzulli et al. [70]

associated with *Vibrio* species (*V. cholerae* non O1-O139, *V. vulnificus*, and *Vibrio alginolyticus*) have been reported (Baker-Austin et al. [15] and references therein).

The CPR survey is one of the longest running marine biological monitoring programs in the world and provides a long-term archive of formalin-preserved plankton samples (http://www.sahfos.ac.uk). In their work, Vezzulli et al. [70] exploited the well-known association between vibrios and plankton, which is considered to be one of the largest reservoirs of these bacteria in nature, to assess possible linkage between *Vibrio* occurrence in the sea and environmental variables (SST and phytoplankton and zooplankton abundance) over a decadal scale by molecular analysis of the microbial community on these historical CPR samples. To this end, they

recovered environmental DNA from the CPR samples that had been stored for up to ~50 years in formalin-fixed format, which is suitable for molecular analyses of the associated prokaryotic community. An unbiased index of abundance for *Vibrio* quantification in the CPR samples, termed "*Vibrio*-relative abundance index—VAI," was developed. This index measures the relative proportion of plankton-associated vibrios in comparison to the total number of associated bacterial cells. 16S rDNA pyrosequencing analysis was also carried out to assess the relative contribution of the *Vibrio* genus to the total plankton-associated bacterial community in comparison to other bacterial groups. Using this approach, it was shown that vibrios, including the species *V. cholerae*, have increased in prevalence during the past 50 years in the coastal North Sea



and that this increase is correlated significantly with increasing SST during that time. These findings provide support for the view that global warming is having a strong impact on the composition of marine prokaryotic communities, with potential important implications for human and animal health.

#### **Conclusions**

Whether or not ocean warming is significantly promoting growth and distribution of vibrios in the aquatic environment globally is a matter of debate. An increasing number of studies and a significant amount of data support this hypothesis, although there are gaps that preclude conclusive evidence. Implementation of long-term ecological studies that incorporate historical as well as newly produced field data, such as has been gathered for eukaryotic communities (e.g., http://www.lternet.edu/; http://www.sahfos.ac.uk), is needed. In addition to the biology and ecology of vibrios, epidemiological studies of their associated diseases would be useful, especially those investigating changes in disease transmission patterns (e.g., pathways of exposure and susceptibility of host populations) in a time of climate changes. If a causal link between the spread of Vibrio illness and climate change could be proven definitively, it would greatly improve our chance to forecast and mitigate future outbreaks associated with these pathogens.

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### References

- 1. IPCC (2007) Climate change 2007: synthesis report. IPCC, Geneva
- Roemmich D, Gould WJ, Gilson J (2012) 135 years of global ocean warming between the Challenger expedition and the Argo Programme. Nat Clim Change 2:425–428
- Reid PC, Gorick G, Edwards M (2011) Climate change and European Marine Ecosystem Research. Sir Alister Hardy Foundation for Ocean Science, Plymouth, 53 p
- Kirov B, Georgieva K (2001) Long-term variations and interrelations of ENSO, NAO and solar activity. Phys Chem Earth 27(6–8):441–448
- Hakkinen S (2002) Surface salinity variability in the northern North Atlantic during recent decades. J Geophys Res 107:8003
- Pruzzo C, Huq A, Colwell RR, Donelli G (2005) Pathogenic Vibrio species in the marine and estuarine environment. In: Belkin S, Colwell RR (eds) Ocean and health pathogens in the marine environment. Springer, Heidelberg, pp 217–252
- Oliver JD, Pruzzo C, Vezzulli L, Kaper JB (2013) Vibrio species. In: Doyle MP, Buchanan RL (eds) Food microbiology: fundamentals

- and frontiers, 4th edn. ASM, Washington, pp 401–440. doi:10.1128/9781555818463.ch16
- World Health Organization: Cholera, 2010. Weekly Epidemiological Record 2011, 86:325–340
- Newton A, Kendall M, Vugia DJ, Henao OL, Mahon BE (2012) Increasing rates of vibriosis in the United States, 1996–2010: review of surveillance data from 2 systems. Clin Infect Dis 54(5):391–395
- González-Escalona N, Cachicas V, Acevedo C, Rioseco ML, Vergara JA, Cabello F et al (2005) Vibrio parahaemolyticus diarrhea, Chile, 1998 and 2004. Emerging Infect Dis 11:129–131
- Martinez-Urtaza J, Huapaya B, Gavilan RG, Blanco-Abad V, Ansede-Bermejo J, Cadarso-Suarez C et al (2008) Emergence of Asiatic *Vibrio* diseases in South America in phase with El Niño. Epidemiology 19:829–837
- CDC (Centers for Disease Control and Prevention) (2008)
  Outbreak of Vibrio parahaemolyticus infections associated with eating raw oysters—Pacific Northwest, 1997. Morb Mort Week Rep 47:457–462
- Paz S, Bisharat N, Paz E, Kidar O, Cohen D (2007) Climate change and the emergence of *Vibrio vulnificus* disease in Israel. Environ Res 103:390–396
- Baker-Austin C, Stockley L, Rangdale R, Martinez-Urtza J (2010) Environmental occurrence and clinical impact of *Vibrio vulnificus* and *Vibrio parahaemolyticus*: a European perspective. Environ Micro Rep 2:7–18
- Baker-Austin C, Trinanes JA, Taylor NGH, Hartnell R, Siitonen A, Martinez-Urtaza J (2012) Emerging Vibrio risk at high latitudes in response to ocean warming. doi:10.1038/nclimate1628
- Lima FP, Wethey DS (2012) Three decades of high-resolution coastal sea surface temperatures reveal more than warming. Nat Commun 3:704
- 17. Fernandez MAL, Bauernfeind A, Jimenez JD, Gil CL, El Omeiri N, Guibert DH et al (2009) Influence of temperature and rainfall on the evolution of cholera epidemics in Lusaka, Zambia, 2003–2006: analysis of a time series. T Roy Soc Trop Med H 103(2):137–143
- Pascual M, Rodó X, Ellner SP, Colwell RR, Bouma MJ (2000) Cholera dynamics and El Niño-Southern oscillation. Science 289 (5485):1766–1769
- Lobitz B, Beck L, Huq A, Wood B, Fuchs G, Faruque AS et al (2000) Climate and infectious disease: use of remote sensing for detection of *Vibrio cholerae* by indirect measurement. Proc Nat Acad Sci USA 97:1438–1443
- Kushmaro A, Bani E, Loya Y, Stackebrandt E, Rosenberg E (2001)
  Vibrio shiloi sp nov the causative agent of bleaching of the coral Oculina patagonica. Int J Syst Evol Micro 51:1383–1388
- Ben-Haim Y, Thompson FL, Thompson CC, Cnockaert MC, Hoste B, Swings J et al (2003) Vibrio coralliilyticus sp nov., a temperaturedependent pathogen of the coral Pocillopora damicornis. Int J Syst Evol Micr 53:309–315
- Vezzulli L, Previati M, Pruzzo C, Marchese A, Bourne DG, Cerrano C (2010) Vibrio infections triggering mass mortality events in a warming Mediterranean sea. Environ Microbiol 12:2007–2019
- Sugumar G, Nakai T, Hirata Y, Matsubara D, Muroga K (1998) Vibrio splendidus biovar II as the causative agent of bacillary necrosis of Japanese oyster Crassostrea gigas larvae. Dis Aquat Organ 33:111–118
- 24. Garnier M, Labreuche Y, Garcia C, Robert A, Nicolas JL (2007) Evidence for the involvement of pathogenic bacteria in summer mortalities of the Pacific oyster *Crassostrea gigas*. Microb Ecol 53:187–196
- Weis KE, Hammond RM, Hutchinson R, Blackmore CG (2011)
  Vibrio illness in Florida, 1998–2007. Epidemiol Infect 139 (4):591–598
- Shearman RK, Lentz SJ (2012) Long-term sea surface temperature variability along the U.S. East Coast. J Phys Oceanogr 40:1004– 1017



 Harvell CD, Mitchell CE, Ward JR, Altizer S, Dobson AP, Ostfeld RS et al (2002) Ecology—climate warming and disease risks for terrestrial and marine biota. Science 296:2158–2162

- 28. Semenza JC, Menne B (2009) Climate change and infectious diseases in Europe. Lancet Infect Dis 9(6):365–375
- Lipp EK, Huq A, Colwell RR (2002) Effects of global climate on infectious disease: the cholera model. Clin Microbiol Rev 15:757– 770
- Colwell RR (2005) Global microbial ecology of *Vibrio cholerae*.
  In: Belkin S, Colwell RR (eds) Ocean and health pathogens in the marine environment. Springer, Heidelberg, pp 297–305
- Singleton FL, Attwell R, Jangi S, Colwell RR (1982) Effects of temperature and salinity on *Vibrio cholerae* growth. Appl Environ Microb 44(5):1047–1058
- 32. Huq A, West PA, Small EB, Huq MI, Colwell RR (1984) Influence of water temperature, salinity, and pH on survival and growth of toxigenic *Vibrio cholerae* serovar 01 associated with live copepods in laboratory microcosms. Appl Environ Microbiol 48:420–424
- Pfeffer CS, Hite MF, Oliver JD (2003) Ecology of Vibrio vulnificus in estuarine waters of eastern North Carolina. Appl Environ Microbiol 69(6):3526–3531
- Huq A, Small EB, West PA, Huq MI, Rahman R, Colwell RR (1983)
  Ecological relationships between *Vibrio cholerae* and planktonic crustacean copepods. Appl Environ Microbiol 45:275–283
- Pruzzo C, Vezzulli L, Colwell RR (2008) Global impact of Vibrio cholerae interactions with chitin. Environ Microbiol 10(6):1400– 1410
- Vezzulli L, Pruzzo C, Huq A, Colwell RR (2010) Environmental reservoirs of *Vibrio cholerae* and their role in cholera. Env Microbiol Rep 2:27–33
- 37. Kirschner AKT, Schlesinger J, Farnleitner AH, Hornek R, Suss B, Golda B et al (2008) Rapid growth of planktonic *Vibrio cholerae* non-O1/non-O139 strains in a large alkaline lake in Austria: dependence on temperature and dissolved organic carbon quality. Appl Environ Microbiol 74(7):2004–2015
- Worden AZ, Seidel M, Smriga S, Wick A, Malfatti F, Bartlett D et al (2006) Trophic regulation of *Vibrio cholerae* in coastal marine waters. Environ Microbiol 8:21–29
- Jensen MA, Faruque SM, Mekalanos JJ, Levin BR (2006) Modelling the role of bacteriophage in the control of cholera outbreaks. Proc Natl Acad Sci USA 103:4652–4657
- Huq A, Sack RB, Nizam A, Longini IM, Nair GB, Ali A et al (2005) Critical factors influencing the occurrence of *Vibrio cholerae* in the environment of Bangladesh. Appl Environ Microbiol 71(8):4645–4654
- 41. Vezzulli L, Pezzati E, Moreno M, Fabiano M, Pane L, Pruzzo C (2009) Benthic ecology of *Vibrio* spp. and pathogenic *Vibrio* species in a coastal Mediterranean environment (La Spezia Gulf, Italy). Microb Ecol 58:808–818
- Martinez-Urtaza J, Blanco-Abad V, Rodriguez-Castro A, Ansede-Bermejo J, Miranda A, Rodriguez-Alvarez MX (2012) Ecological determinants of the occurrence and dynamics of *Vibrio parahaemolyticus* in offshore areas. ISME J 6(5):994–1006
- 43. Miles DW, Ross T, Olley J, McMeekin TA (1997) Development and evaluation of a predictive model for the effect of temperature and water activity on the growth rate of *Vibrio parahaemolyticus*. Int J Food Microbiol 38:133–142
- 44. Kaspar CW, Tamplin ML (1993) Effects of temperature and salinity on the survival of *Vibrio vulnificus* in seawater and shellfish. Appl Environ Microbiol 59:2425–2429
- 45. Xu HS, Roberts N, Singleton FL, Attwell RW, Grimes DJ, Colwell RR (1982) Survival and viability of nonculturable *Escherichia coli* and *Vibrio cholerae* in the estuarine and marine environment. Microb Ecol 8:313–323
- Coutard F, Crassous P, Droguet M, Gobin E, Colwell RR, Pommepuy M, Hervio-Heath D (2007) Recovery in culture of

- viable but nonculturable *Vibrio parahaemolyticus*: regrowth or resuscitation? ISME J 1(2):111–120
- Kimes NE, Grim CJ, Johnson WR, Hasan NA, Tall BD, Kothary MH et al (2012) Temperature regulation of virulence factors in the pathogen *Vibrio corallilyticus*. ISME J 6:835–846
- Oh MH, Lee SM, Lee DH, Choi SH (2009) Regulation of the Vibrio vulnificus hupA gene by temperature alteration and cyclic AMP receptor protein and evaluation of its role in virulence. Infect Immun 77(3):1208–1215
- Mahoney JC, Gerding MJ, Jones SH, Whistler CA (2010) Comparison of the pathogenic potentials of environmental and clinical *Vibrio parahaemolyticus* strains indicates a role for temperature regulation in virulence. Appl Environ Microbiol 76 (22):7459–7465
- Montilla R, Chowdhury MA, Huq A, Xu B, Colwell RR (1996) Serogroup conversion of *Vibrio cholerae* non-O1 to *Vibrio cholerae* O1: effect of growth state of cells, temperature, and salinity. Can J Microbiol 42(1):87–93
- Schuhmacher DA, Klose KE (1999) Environmental signals modulate ToxT-dependent virulence factor expression in *Vibrio cholerae*. J Bacteriol 181(5):1508–1514
- Peterson KM (2002) Expression of Vibrio cholerae virulence genes in response to environmental signals. CIIM 3(2):29–38
- Tamplin ML, Gauzens AL, Huq A, Sack DA, Colwell RR (1990) Attachment of *Vibrio cholerae* serogroup O1 to zooplankton and phytoplankton of Bangladesh waters. Appl Environ Microbiol 56 (6):1977–1980
- Nelson EJ, Harris JB, Morris JG Jr, Calderwood SB, Camilli A (2009) Cholera transmission: the host, pathogen and bacteriophage dynamic. Nat Rev Microbiol 7:693–702
- Colwell RR, Huq A, Islam MS, Aziz KMA, Yunus M, Huda Khan N et al (2003) Reduction of cholera in Bangladeshi villages by simple filtration. Proc Natl Acad Sci USA 100(3):1051–1055
- Huq A (1984) The role of planktonic copepods in the survival and multiplication of *Vibrio cholerae* in the aquatic environment. University of Maryland, College Park
- 57. Stauder M, Vezzulli L, Pezzati E, Repetto B, Pruzzo C (2010) Temperature affects *Vibrio cholerae* O1 El Tor persistence in the aquatic environment via an enhanced expression of GbpA and MSHA adhesins. Env Microbiol Rep 2:140–144
- Chiavelli DA, Marsh JW, Taylor RK (2001) The mannosesensitive hemagglutinin of *Vibrio cholerae* promotes adherence to zooplankton. Appl Environ Microbiol 67:3220–3225
- Kirn TJ, Jude BA, Taylor RK (2005) A colonization factor links *Vibrio cholerae* environmental survival and human infection. Nature 438:863–866
- Beaugrand G, Reid PC, Ibañez F, Alistair Lindley J, Edwards M (2002) Reorganization of North Atlantic marine copepod biodiversity and climate. Science 296(5573):1692–1694
- Moran XAG, Lopez-Urrutia A, Calvo-Diaz A, Li WKW (2010) Increasing importance of small phytoplankton in a warmer ocean. Global Change Biol 16(3):1137–1144
- Rawlings TK, Ruiz GM, Colwell RR (2007) Association of Vibrio cholerae serogroups O1 El Tor and O139 Bengal with the copepods Acartia tonsa and Eurytemora affinis. Appl Environ Microbiol 73:7926–7933
- Hsieh JL, Fries JS, Noble RT (2008) Dynamics and predictive modelling of *Vibrio* spp. in the Neuse river estuary, North Carolina, USA. Environ Microbiol 10(1):57–64
- Randa MA, Polz MF, Lim E (2004) Effects of temperature and salinity on *Vibrio vulnificus* population dynamics as assessed by quantitative PCR. Appl Environ Microbiol 70(9):5469–5476
- 65. Oberbeckmann S, Fuchs BM, Meiners M, Wichels A, Wiltshire KH, Gerdts G (2012) Seasonal dynamics and modeling of a *Vibrio* community in coastal waters of the North Sea. Microb Ecol 63 (3):543–551



- Thompson JR, Randa MA, Marcelino LA, Tomita-Mitchell A, Lim E, Polz MF (2004) Diversity and dynamics of a North Atlantic coastal *Vibrio* community. Appl Environ Microbiol 70:4103–4110
- 67. Colwell RR (2000) Viable but nonculturable bacteria: a survival strategy. J Infect Chemother 6:121–125
- 68. Martinez-Urtaza J, Bowers JC, Trinanes J, DePaola A (2010) Climate anomalies and the increasing risk of *Vibrio parahaemolyticus* and *Vibrio vulnificus* illnesses. Food Res Int 43(7):1780–1790
- De Magny GC, Colwell RR (2009) Cholera and climate: a demonstrated relationship. Trans Am Clin Climatol Assoc 120:119–128
- Vezzulli L, Brettar I, Pezzati E, Reid PC, Colwell RR, Höfle MG, Pruzzo C (2012) Long-term effects of ocean warming on the prokaryotic community: evidence from the vibrios. ISME J 6:21–30
- Reid PC, Colebrook JM, Matthews JBL, Aiken J (2003) The continuous plankton recorder: concepts and history, from plankton indicator to undulating recorders. Prog Oceanogr 58:117–173
- Kirby RR, Beaugrand G, Lindley JA, Richardson AJ, Edwards M, Reid PC (2007) Climate effects and benthic-pelagic coupling in the North Sea. Mar Ecol Prog Ser 330:31–38

