

Species Loss Due to Nutrient Addition Increases with Spatial Scale in Global Grasslands

Eric W. Seabloom^{1*}, Evan Batzer², Jonathan M. Chase^{3,4}, W. Stanley Harpole^{3,5,6}, Peter B. Adler⁷, Sumanta Bagchi⁸, Jonathan D. Bakker⁹, Isabel C. Barrio¹⁰, Lori Biederman¹¹, Elizabeth H. Boughton¹², Miguel N. Bugalho¹³, Maria C. Caldeira¹⁴, Jane A. Catford¹⁵, Pedro Daleo¹⁶, Nico Eisenhauer^{3,17}, Anu Eskelinen^{3,5,18}, Sylvia Haider^{3,19}, Lauren M. Hallett²⁰, Ingibjörg Svala Jónsdóttir²¹, Kaitlin Kimmel²², Marirose Kuhlman²³, Andrew MacDougall²⁴, Cecilia D. Molina²⁵, Joslin L. Moore²⁶, John W. Morgan²⁷, Ranjan Muthukrishnan²⁸, Timothy Ohlert²⁹, Anita C. Risch³⁰, Christiane Roscher^{3,5}, Martin Schütz³⁰, Grégory Sonnier¹², Pedro M. Tognetti²⁵, Risto Virtanen³¹, Peter A. Wilfahrt¹, Elizabeth T. Borer¹

¹. Dept. of Ecology, Evolution, and Behavior. University of Minnesota. St. Paul, MN 55108 USA

². Dept. of Plant Sciences, University of California, Davis, Davis CA 95616 USA

³. German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig, Leipzig, Germany

⁴. Dept. of Computer Sciences, Martin Luther University, Halle (Saale), Germany

⁵. Helmholtz Center for Environmental Research – UFZ, Dept. of Physiological Diversity, Permoserstrasse 15, 04318 Leipzig, Germany.

⁶. Martin Luther University Halle-Wittenberg, am Kirchtor 1, 06108 Halle (Saale), Germany."

⁷. Dept. of Wildland Resources and the Ecology Center, Utah State University, Logan, UT 84322 USA

⁸. Centre for Ecological Sciences, Indian Institute of Science, Bangalore 560012, India

⁹. School of Environmental and Forest Sciences, University of Washington, Box 354115, Seattle WA 98195-4115 USA

¹⁰. Faculty of Environmental and Forest Sciences, Agricultural University of Iceland, Árleyni 22, 112 Reykjavík, Iceland

¹¹. Dept. of Ecology, Evolution, and Organismal Biology, Iowa State University, Ames Iowa 50010 USA

¹². Archbold Biological Station, Venus, FL 33960 USA

¹³. Centre for Applied Ecology "Prof. Baeta Neves" (CEABN-InBIO), School of Agriculture, University of Lisbon, Portugal

¹⁴. Forest Research Centre, School of Agriculture, University of Lisbon, Portugal

¹⁵. Dept. of Geography, King's College London, 30 Aldwych, WC2B 4BG, UK

¹⁶. Instituto de Investigaciones Marinas y Costeras (IIMyC), CONICET – UNMDP, CC 1260 Correo Central, B7600WAG, Mar del Plata, Argentina

¹⁷. Institute of Biology, Leipzig University, Deutscher Platz, 5e, 04103 Leipzig, Germany

¹⁸. Dept. of Ecology and Genetics, University of Oulu, Oulu, Finland

¹⁹. Institute of Biology / Geobotany and Botanical Garden, Martin Luther University Halle-Wittenberg, Halle, Germany

²⁰. Dept. of Biology and Environmental Studies Program, University of Oregon, Eugene, Oregon 97403 USA

²¹. Institute of Life and Environmental Sciences, University of Iceland, Sturlugata 7, 102 Reykjavík, Iceland

²². Dept. of Earth and Planetary Sciences, Johns Hopkins University, Baltimore, MD 21218, USA

- ²³. MPG Ranch, Missoula, MT, 59801 USA
- ²⁴. University of Guelph, Guelph, ON N1G 2W1, Canada
- ²⁵. Facultad de Agronomia – IFEVA, UBA – CONICET, Av San Martin 4453, Buenos Aires, Argentina
- ²⁶. Monash University, Clayton VIC 3800, Australia
- ²⁷. Dept. of Ecology, Environment & Evolution, La Trobe University, Bundoora 3086, Victoria, Australia
- ²⁸. Environmental Resilience Institute, Indiana University, Bloomington IN, 47401 USA
- ²⁹. University of New Mexico, Dept. of Biology, MSC03 2020, 1 University of New Mexico, Albuquerque, NM, 87031, USA
- ³⁰. Swiss Federal Institute for Forest, Snow and Landscape Research WSL, Zuercherstrasse 111, 8903 Birmensdorf, Switzerland
- ³¹. Dept. of Ecology and Genetics, University of Oulu, Finland

* Correspondence: seabloom@umn.edu

Keywords: Community Ecology, Biodiversity, Grasslands, Nutrients, Herbivores

Running Head: Species loss in global grasslands

Statement of authorship: EWS wrote the first draft of the manuscript, and all authors contributed substantially to revisions. EWS analyzed the data with contributions from PBA, EB, SB, JMC, LMH, WSH, JLM, and TO. Detailed author contributions are presented in Table S8.

Data accessibility statement: Data supporting the results are archived at the Dryad Digital Repository (DOI supplied upon acceptance).

67 **Article Type:** Letters

68 **Abstract Word Count:** 144

69 **Main Text Word Count:** 5759

70 **Acknowledgments Word Count:** 88

71 **References Word Count:** 1814

72 **Table and Figure Legends Word Count:** 514

73 **Number of References:** 71

74 **Number of Figures:** 4

75 **Number of Tables:** 0

76 **Number of Text Boxes:** 0

77

Abstract The effects of human-induced changes to nutrient supplies and herbivore density on species diversity vary with spatial scale because coexistence mechanisms are scale dependent. This scale dependence may alter the species-area relationship (SAR), which describes changes in species richness (S) as a power function of the sample area (A): $S=cA^z$, where c and z are constants. We analyzed the effects of experimental manipulations of nutrient supply and herbivore density on species richness across a range of scales ($0.01 - 75 \text{ m}^2$) at 30 grasslands in 10 countries. We found that nutrient addition reduced the number of species that could coexist locally, indicated by the SAR intercepts ($\log c$), but did not affect the SAR slopes (z). As a result, proportional species loss due to nutrient enrichment was constant across sampling scales, and total species loss increased over threefold across our range of sampling scales.

What determines the number of species at a location? This question lies at the core of community ecology. The answer is inherently scale dependent (Arrhenius 1921; Godwin 1923; Gleason 1926; MacArthur & Wilson 1967; Grace *et al.* 2011; Chase *et al.* 2018), because different mechanisms influence diversity at different spatial scales (Chesson 2000; Leibold *et al.* 2004; Hart *et al.* 2017; Leibold & Chase 2017; Thompson *et al.* 2020). For example, non-spatial coexistence mechanisms that depend on tradeoffs (e.g., in resource use efficiency or susceptibility to consumers) or temporal variability (e.g., temporal storage effects) can lead to coexistence at very small spatial scales (Hutchinson 1961; Tilman 1982; Holt *et al.* 1994; Chesson 2000). In contrast, coexistence mechanisms that depend on spatial variability, e.g., dispersal limitation or competition-colonization tradeoffs influence diversity at larger spatial scales (Godwin 1923; Gleason 1926; MacArthur & Wilson 1967; Hastings 1980; Chesson 2000; Leibold *et al.* 2004; Vellend 2010). Furthermore, the size of individuals and the spatial heterogeneity of the environment will determine the scales at which species interact and the minimum possible scale of coexistence (Goldberg & Miller 1990; Oksanen 1996; Seabloom *et al.* 2005). For these reasons, differences in diversity observed in field studies, across space or in response to environmental changes induced by ecological and anthropogenic drivers, will reflect both sampling scale and the scale-dependence of coexistence mechanisms. Nevertheless, most empirical studies sample and compare diversity at a single scale, leading to an incomplete understanding of diversity responses to ecological drivers (Chalcraft *et al.* 2008; Lan *et al.* 2015; Chase *et al.* 2018).

A variety of approaches have been used to incorporate scale explicitly into measures of diversity, the most canonical of which is the species-area relationship (SAR)(Arrhenius 1921; MacArthur & Wilson 1967; Flather 1996; Chase *et al.* 2018). While a wide variety of SAR models exist (Flather 1996; Dengler *et al.* 2020), a simple model which has been found to be applicable in many systems is

$$S = cA^z \quad (1a)$$

or equivalently

$$\log(S) = \log(c) + z \log(A) \quad (1b)$$

in which S is the number of species and A is the area sampled (Arrhenius 1921; Flather 1996; Fridley *et al.* 2005; Drakare *et al.* 2006; Dengler *et al.* 2020).

In this formulation, the y-intercept, $\log(c)$ is a measure of local diversity (proportional to α diversity when $A=1$), and the slope (z) is a measure of spatial heterogeneity in community composition (proportional to some measures of β diversity)(Crist & Veech 2006; Grace *et al.* 2011; Scheiner *et al.* 2011). While the SAR only attains a true asymptote at a global scale (Williamson *et al.* 2001), saturation within sampling confines can be taken as a measure of the available species pool (γ diversity)(Grace *et al.* 2011; Chao *et al.* 2014). The x-intercept indicates

the Minimal Insular Area (A_{min}) (sensu, Heatwole 1975), the area at which only a single species is found ($S=1$ or $\log(S)=0$) and is a nonlinear function of c and z :

$$A_{min} = c^{-\frac{1}{z}} \quad (2)$$

A_{min} can be thought of as the minimal area of coexistence, the area above which more than one species can co-occur (Heatwole 1975; Connor & McCoy 1979; Seabloom *et al.* 2005).

Understanding the patterns and determinants of diversity across scales has gained increased relevance as human domination of the biosphere has altered many of the controls on species diversity, leading to scale-dependent changes in diversity (Chase *et al.* 2019). For example, humans have increased the supply of biologically-limiting nutrients and have changed the density of herbivores in many ecosystems (Foley *et al.* 2005; Ripple *et al.* 2015; Steffen *et al.* 2015), both of which can alter plant diversity at a range of spatial scales (Chaneton & Facelli 1991; Olff & Ritchie 1998; Crawley *et al.* 2005; Bakker *et al.* 2006; Hillebrand *et al.* 2007; Chalcraft *et al.* 2008; Leps 2014; Lan *et al.* 2015).

The effects of environmental change on diversity may shift with spatial scale, and this scale-dependence may be reflected in diversity-scaling relationships such as the parameters of the SAR (Chaneton & Facelli 1991; Olff & Ritchie 1998; Bakker *et al.* 2006; Chalcraft *et al.* 2008; Lan *et al.* 2015; Chase *et al.* 2018). For example, in grassland ecosystems, increasing the supply rates of biologically limiting nutrients like nitrogen (N) and phosphorus (P) often leads to reduced plant diversity at local scales (i.e., lower $\log c$ and higher A_{min} ; Figure 1) by reducing the opportunity for coexistence through tradeoffs in soil resource use efficiency and increasing competition for light (Goldberg & Miller 1990; Dybzinski & Tilman 2007; Harpole & Tilman 2007; Hautier *et al.* 2009; Borer *et al.* 2014b; Harpole *et al.* 2016; Midolo *et al.* 2019). While the effects of nutrient addition and herbivory on local coexistence in grasslands have been examined experimentally, it remains unclear how these effects will change with spatial scale. Importantly, if environmental changes alter the SAR slope, diversity change measured at a single scale may overestimate or underestimate diversity change at larger spatial scales (Figure 1)(Lan *et al.* 2015).

Changes in the scaling relationship will depend on the specific coexistence mechanisms (e.g., spatial versus non-spatial mechanisms) affected by nutrient enrichment or herbivore exclusion. Furthermore, different mechanisms will be associated with the total biomass, light availability, and size of the species pool (Bakker *et al.* 2006; Chalcraft *et al.* 2008; Borer *et al.* 2014b; Lan *et al.* 2015; Harpole *et al.* 2016). For example, nutrient enrichment or herbivore exclusion may increase biomass which can lead to thinning due to light competition, which would reduce the SAR slope and intercept simply due to the presence of fewer larger individuals in each sample (Lan *et al.* 2015). These treatments also may increase the dominance of a few species, and this reduced evenness would be expected to reduce the SAR intercept but increase the SAR slope (Lan *et al.* 2015)(Figure 1E). Treatments may also cause the extinction of specific species leading to reduced species pool and lower SAR slope (Figure 1A) (Lan *et al.* 2015).

We have more specific expectations for cross-scale effects of nutrient addition than for herbivore exclusion, because nutrient addition has more consistent effects on grassland plant diversity, and nutrient effects have been examined across scales (Chalcraft *et al.* 2008; Borer *et al.* 2014b; Lan *et al.* 2015). For example, if nutrient-induced light limitation reduces coexistence opportunities for a consistent set of species through non-spatial processes such as reduced niche dimensionality (Harpole & Tilman 2007; Harpole *et al.* 2016), then the local diversity loss ($\log c \downarrow$) will be accompanied by a reduced total species pool (γ diversity \downarrow) (Harpole & Tilman 2007; Chalcraft *et al.* 2008; Lan *et al.* 2015)(Figure 1C & D). These effects would also be associated with increased biomass and reduced light availability. In this case, the effects on the SAR slope depend on the relative rate of species loss across scales (Lan *et al.* 2015). If there is a constant proportional loss of species with increasing area, then the slope of the SAR would remain constant (Figure 1C & D). In contrast, the slope would decline if there is greater proportional loss at larger spatial scales ($z \downarrow$)(Figure 1A & B), resulting in a more spatially homogeneous community. In a less extreme case, nutrient addition could increase the dominance of species that favor high nutrient conditions without causing species extinctions. In this case, nutrient addition would increase the SAR slope ($z \uparrow$) due to local diversity loss ($\log c \downarrow$), but the total species pool would remain unchanged (Figure 1A & B). This effect also would be reflected in reduced species evenness (Lan *et al.* 2015).

Nutrient addition also could influence spatial coexistence mechanisms, such as competition-colonization tradeoffs or mass effects by reducing dispersal and local recruitment (Hastings 1980; Tilman *et al.* 1994; Leibold *et al.* 2004; Vellend 2010). Reduced dispersal and colonization would lower local diversity ($\log c \downarrow$) but leave the total species pool unchanged, resulting in an increased SAR slope ($z \uparrow$) in communities with low to moderate dispersal (Mouquet & Loreau 2003; Lan *et al.* 2015)(Figure 1A & B), although very high rates of dispersal may reduce both local richness and the total species pool (Mouquet & Loreau 2003). Finally, nutrient addition could increase individual plant size (Goldberg 1987; Oksanen 1996), thereby reducing local diversity ($\log c \downarrow$). As with reduced dispersal, increased plant size would not affect the total species pool but would increase the SAR slope ($z \uparrow$) (Lan *et al.* 2015) (Figure 1E & F).

In any of these cases, a constant SAR slope indicates constant proportional change in species with increasing spatial scale, whereas a change in the SAR slope indicates an increasing or decreasing proportional change in the numbers of species with increasing spatial scale (Lan *et al.* 2015). Importantly, if there is a constant proportional loss (or gain) of species, this means that there is an increase in the total number of species lost (or gained) at larger spatial scales (Lan *et al.* 2015) (Figure 1C & D).

In summary, as we expect nutrient addition to decrease local diversity in grasslands ($\log c \downarrow$) (Borer *et al.* 2014b; Midolo *et al.* 2019), the SAR slope will either increase ($z \uparrow$) or decrease ($z \downarrow$) depending on the change in diversity at larger scales (Chalcraft *et al.* 2008; Lan *et al.* 2015)(Figure 1). While there are few specific predictions for the minimal area of coexistence (A_{min}), we expect this to be negatively correlated with local diversity ($\log c$) via the relationship in Equation 2, such that the expected nutrient-induced reduction in c should lead to an increase in A_{min} depending on the change in z .

While the predictions for herbivore effects on diversity scaling are less well developed, the effects should be mediated through changes in the same core processes governing nutrient effects. For example, herbivores may reduce diversity if the community becomes dominated by a few unpalatable species that are resistant to grazing ($\log c \downarrow$), or they may increase diversity if they reduce the abundance of dominant, competitive, or fast-growing species ($\log c \uparrow$) (Olf & Ritchie 1998; Viola *et al.* 2010; Lind *et al.* 2013; Koerner *et al.* 2018). Herbivores also may increase diversity by increasing availability of a limiting resource (e.g., light) or increasing seed dispersal and colonization rates (Olf & Ritchie 1998; Borer *et al.* 2014b). The presence of herbivores also may change environmental variation, for example through localized deposition of feces or urine (Olf & Ritchie 1998), which may increase the SAR slope ($z \uparrow$). As is the case with nutrient effects, we expect herbivore effects on plant diversity to be related to evenness, light availability, and plant biomass, such that herbivores will likely have positive effects on diversity at light-limited, productive sites that are dominated by a few plant species (Bakker *et al.* 2006; Borer *et al.* 2014b; Koerner *et al.* 2018).

Here we analyze species richness data from 30 grasslands or low-stature shrublands (hereafter grasslands) spanning spatial scales of three orders of magnitude ($0.01 - 75 \text{ m}^2$) in the context of the Nutrient Network Distributed Experiment (NutNet, www.nutnet.org), a globally-replicated experiment manipulating nutrient supply and herbivore density (Borer *et al.* 2014a; Borer *et al.* 2017). These sites are located in 10 countries on five continents and represent a wide range of environmental conditions and ecosystem types including annual grasslands, deserts, tundra, montane meadows, semi-arid and mesic grasslands, and old fields. We use these data to address the long-standing gap in our understanding of how environmental drivers affect diversity across spatial scales (Chalcraft *et al.* 2008; Chase *et al.* 2018). Specifically, we quantify variability among sites in the slope and intercept of the species-area relationship (SAR) and test the effects of nutrient addition and herbivore exclusion on the SAR. In addition, we test whether among-site differences in the SAR are correlated with evenness, light availability, plant biomass, the size of the total species pool, and whether these covariates affect the strength of the nutrient or consumer reduction treatments (Chalcraft *et al.* 2008; Lan *et al.* 2015).

Materials and Methods

Experimental Design and Data Collection

We include data from 30 sites in 10 countries which are part of the Nutrient Network (NutNet) distributed experiment (Borer *et al.* 2014a; Borer *et al.* 2017) (Table S1). All sites were dominated by herbaceous or low-statured vegetation and spanned wide gradients including elevation (6 to 3500 m), latitude (52° S to 69° N), mean annual precipitation (249 to 1877 mm yr^{-1}), mean annual temperature (-3 to 23° C), and mean aboveground live biomass (34 to 900 g m^{-2}). Local richness (plot scale richness ranged from 4 to 43 species m^{-2}) and total site richness (richness pooled across all sampled plots ranged from 18 to 142 species site^{-1}) were highly variable among sites. We used data from control plots at 30 sites and from two experiments at subsets of the sites: the Multiple-nutrient experiment (21 sites) and the Consumer-nutrient experiment (16 sites). Experimental duration at the time of sampling varied from 3-11 years

(Table S1), but inclusion of duration in statistical models did not qualitatively change results. While other studies have used this experiment to examine the influence of nutrients and herbivores on patterns of local diversity and coexistence (Borer *et al.* 2014b; Harpole *et al.* 2016; Borer *et al.* 2017), there has been no examination of the potential influence of scaling relationships and scale-dependence of treatment effects.

Multiple-nutrient Experiment: At 21 of the 30 sites, we replicated an experiment that factorially combined three nutrient-addition treatments each at two levels (Control or Fertilized): Nitrogen addition ($10 \text{ g N m}^{-2} \text{ yr}^{-1}$ as timed-release urea), Phosphorus addition ($10 \text{ g P m}^{-2} \text{ yr}^{-1}$ as triple-super phosphate), and Potassium and Micronutrient addition ($10 \text{ g K m}^{-2} \text{ yr}^{-1}$ as potassium sulfate and $100 \text{ g m}^{-2} \text{ yr}^{-1}$ of a micronutrient mix (6% Ca, 3% Mg, 12% S, 0.1% B, 1% Cu, 17% Fe, 2.5% Mn, 0.05% Mo, and 1% Zn). N, P, and K were applied annually, and the micronutrient mix was applied once at the start of the study.

Consumer-nutrient Experiment: At 16 of the 30 sites, we added an additional two treatments using herbivore fencing: Fenced with no nutrients added and Fenced with all nutrients added. Fences were 2.1 m tall and excluded aboveground, non-climbing, vertebrate herbivores. The lower 0.9 m was composed of 1 cm woven wire mesh with a 0.3 m outward-facing flange stapled to the ground to exclude digging animals. The top 1.2 m was composed of five equally spaced rows of woven wire to prohibit large animals from entering the plots. In combination with the unfenced control and unfenced plots with all nutrients added, we had a full factorial combination of nutrient addition (Control or Fertilized) and vertebrate consumer presence (Control or Fenced).

Data Collection: We estimated SARs using data collected at five spatial scales: 0.01 m^2 , 1 m^2 , 6.25 m^2 , mean 25 m^2 , and 75 m^2 . To do this, we sampled plant community composition in $1 \text{ m} \times 1 \text{ m}$ quadrats by visually estimating the areal cover of each species. The cover data allowed us to estimate richness and Simpson's evenness at an area of 1 m^2 (Smith & Wilson 1996). We also recorded the presence of all species in the entire $5 \times 5 \text{ m}$ plot (25 m^2), the $2.5 \times 2.5 \text{ m}$ subplot in which the cover plot was located (6.25 m^2), and four $0.1 \times 0.1 \text{ m}$ quadrats placed at the corners of the 1 m^2 cover quadrat (0.01 m^2). We used the mean richness of the four 0.01 m^2 in our analyses. We excluded a small number of the 0.01 m^2 quadrats in which the mean species richness was zero as log richness was undefined (0.1% of samples). We aggregated species lists across the three replicate $5 \times 5 \text{ m}$ plots to estimate species richness at the 75 m^2 scale. This among block estimate of richness was not nested and incorporates among block variability. Nested and non-nested SAR's have similar slopes in non-forested terrestrial habitats (Drakare *et al.* 2006), and our results were qualitatively similar when we only used the fully nested samples with a maximum area of 25 m^2 . One site did not collect species richness data at the 75 m^2 scale, so the maximum scale was at this site (chilcas.ar) was 18.75 m^2 (area of three 2.5 m^2 plots). Inclusion or exclusion of this site did not qualitatively change results. We calculated site richness (i.e., site species pool) as the total number of species found across all sampled plots and years at the site (Table S1).

We sampled aboveground plant biomass by clipping all aboveground biomass (live and dead) in two 0.1 m x 1 m strips, sorting current year's biomass (live biomass) from previous year's biomass (dead biomass), drying the biomass to a constant mass at 60 °C, and weighing it to the nearest 0.01 g. Within each 1 m² quadrat, we measured the proportion of photosynthetically active radiation (PAR) reaching the ground level.

Statistical Analyses

All analyses were conducted using R version 4.0.2 (R Development Core Team 2010). We fit a SAR for each treatment at each site using Equation 1b (Figures S1 – S4) with the lmer function in the lmer4 R library (version 1.1-23), which fits linear models to subgroups of data (e.g., different sites). We used the site-level estimates of z , c , and A_{min} in subsequent analyses. A_{min} was natural log transformed due to a highly skewed distribution. We tested whether among site differences in the SAR parameters were correlated with evenness, light availability, plant biomass, the size of the total species pool.

While we focus on the SAR model in Equation 1, there are a wide range of potential models for SARs (e.g., Flather 1996). As our goal was to compare parameters across sites, we fit the same model to all sites. However, we also tested whether an alternate model could provide a better fit to the data. To do this, we used a general three parameter model (a , b , c) which is recommended in the absence of a specific theoretical basis (Flather 1996):

$$S = \frac{a+bA}{1+cA} \quad (2)$$

We fit this model to 30 sites with observational data, and compared the AIC and root mean square error (RMSE) between the two SAR models (Equations 1 and 2). We also fit this model separately to the 4 treatments (Control, Fertilized, Fenced, Fertilized & Fenced) at the 16 sites that implemented the Consumer-nutrient Experiment.

For sites that implemented the Multiple-Nutrient or Consumer Nutrient experiments, we tested whether experimental treatments altered the SAR parameters with Mixed Effects Models (MEMs) using the lmer function in the lme4 R library with p-values generated using Satterthwaite's degrees of freedom method using the lmerTest R library (version 3.1-2). We included site as a random effect in these models, and model specifications are included in Tables S2-S4. Inclusion of experimental duration as a random effect in these models did not change any results and occasionally prevented models from fitting due to singularities. We tested for effects of differential errors associated with the estimates of c and z at each site by running weighted regression in which weights were the inverse of the standard error of site-level estimate of c and z . Weighted regressions were nearly identical to unweighted regressions and did not change interpretation of any results presented here. Here we present results of unweighted regressions in the interest of parsimony and to give equal weight to each site.

In testing for interactions between treatment effects and covariates (evenness, light availability, plant biomass, the size of the total species pool), we used site level treatment mean of evenness, light availability, and plant biomass, while the total species pool has only a single

measurement per site. We note here that evenness (1 m^2), light (1 m^2), and plant biomass (0.2 m^2) are measured at different scales within each plot, while the total species pool is the summed number of species across 30 1 m^2 plots. In these analyses, the correlation among these covariates could lead to a suite of comparable models rather than a single best model. To address this, we used a multi-model approach, as described in Grueber et al. (2011) using the dredge and model.avg functions in the MuMIn library (version 1.43.17). We standardized the input variables using the arm library (version 1.11-1). In the model averaging, we included all models within 4 AIC_c units of the best model.

Results

We started by testing if the appropriateness of the two-parameter SAR formulation (Equation 1) relative to a more general three parameter model (Equation 2). In paired t-test, the two-parameter model fit the observational data (30 sites) better than the three-parameter model based on AIC ($t = -2.343$, $p=0.026$) and RMSE ($t = -2.231$, $p = 0.033$). We also compared these models using the 16 sites that implemented the Consumer-nutrient Experiment. As was the case for observational sites, the three-parameter model did not provide a better fit based on AIC and RMSE. Based on these analyses, we concluded that the two-parameter model was appropriate for our analyses and our subsequent descriptions and discussion focus only on that model.

Overall, sites showed more variability in SAR intercept (c) than in the SAR slope (z) (Figure S1). The coefficient of variation ($\text{CV} = \text{standard deviation} / \text{mean}$) for z (0.19) was about 3 times lower than for c (0.58). Across all sites ($N=30$) under ambient conditions (Control Plots), SAR slopes ranged from 0.12 to 0.33 ($\text{mean}=0.23 \log(\text{species})/\log(\text{m}^2)$) and the intercept ranged from 2.6 to 33.0 species m^{-2} ($\text{mean}=10.7$). The x-intercept (A_{\min}) had a mean of $1.2 \times 10^{-3} \text{ m}^2$ (12 cm^2) and was highly variable ranging from near 0 to 0.02 m^2 (200 cm^2) with a CV of 2.5. We used natural log transformed values of A_{\min} in our analyses, which ranged from -14.5 to -4.6 $\log(\text{m}^2)$ (Figures 2 & 3).

In the observational data, the SAR slope (z) and ($\log c$) were uncorrelated ($r=0.11$, $p=0.548$), slope (z) and x-intercept ($\log(A_{\min})$) were positively correlated ($r=0.51$, $p=0.004$), and ($\log c$) and x-intercept ($\log(A_{\min})$) were negatively correlated ($r=-0.73$, $p<0.001$). As would be expected, the SAR intercept was highly correlated with species richness at 1 m^2 when $\log(A)=0$ ($r=0.97$, $p<0.001$).

Among site variability in the SAR slope increased with site richness but was unrelated to any other of our covariates in the observational data (evenness, live biomass, or proportion of light at ground level) (Table S2; Figure 4). As predicted, local richness ($\log c$) increased with light availability and total site richness (Table S2; Figure 4), and the minimal area of coexistence (A_{\min}) declined with light availability (Table S2; Figure 4). There were no significant correlations among the site means of the covariates ($p > 0.05$).

Nutrient addition reduced local diversity ($\log c$) and increased the minimal coexistence area (A_{\min}) but did not affect the slope of the SAR (Figures 2 & 3; Tables S3 & S4). The lack of a

treatment effect on the SAR slope may reflect either a lack of change in the slope or high variability in the estimates. As noted above, slopes did not vary widely among sites, and variance among slope estimates among sites was not large within treatments (Figures 4 & 5). For example, in our analyses of the experimental data, the standard error in the slope estimates and treatment effects were close to 0.01 (Tables S3 & S4), suggesting that we could detect small differences in slopes among sites and treatments.

Because slopes remained constant and species loss was proportional across scales, addition of all nutrients in combination caused more absolute species loss at the largest scale (mean of 3.2 species lost at 75 m²) than at the smallest scale (mean of 0.9 species lost at 0.01 m²). The nutrient effect on species loss was driven by the effects of N addition (Figure 3; Table S4). Fencing did not have a consistent effect on any of the SAR parameters (Figure 2; Table S3).

There were significant interactions between experimental treatments and the environmental covariates. For example, the interaction between site richness and the effects of nutrient enrichment on local richness (log c), was such that sites with more species had higher rates of species loss in the experimental plots (Figure 4, Table S5). Fencing effects on local richness were strongly affected by light transmission, with higher species loss at sites in which fencing reduced light availability (Figure 4, Table S5).

Some of the focal environmental covariates that we measured at the plot scale, live biomass and light transmission, were affected by experiment treatments. Nutrient addition, primarily N & P addition, reduced light transmission and increased live biomass (Figure 5; Tables S6 & S7). Evenness was unaffected by the experimental treatments (Figure 5; Tables S6 & S7). The covariate, site richness is measured at the site scale, so does not vary among plot or treatments within a site.

Discussion

We found that experimental addition of nutrients, and nitrogen in particular, reduced the SAR intercept (log c), but did not have a consistent effect on the SAR slopes (z) across sites. As a result, proportional species loss was constant across spatial scales, and total species loss increased over 3.6-fold with spatial scale within individual sites (Figure 1C & D, 3, & 4). Furthermore, we found that nutrient-induced loss of species was highest at sites with larger species pools (i.e., site-level species richness)(Harpole *et al.* 2016), and that the effects of fencing were mediated by light availability (Borer *et al.* 2014b)(Figure 5; Table S5). While our maximum sample area was rather small relative to other studies, the SAR slopes in our data (mean=0.23) were similar to those spanning much larger spatial scales in other terrestrial, non-forested habitats (Drakare *et al.* 2006; Dengler *et al.* 2020).

Our study allowed us to examine small-scale patterns as reflected in the intercept of the SAR (log c) and the minimal area of coexistence (A_{min}). At this scale, our results showed wide variation in local diversity (log c) and the minimal area of coexistence (A_{min}) among sites. This among-site variation was related to light availability and total site richness. Across all sites, we found that nutrient addition reduced local diversity (log c↓) (Borer *et al.* 2014b; Harpole *et al.*

2016; Midolo *et al.* 2019) and also increased the minimal area of coexistence (A_{min}), which has not been previously reported. Nutrient induced changes in local richness were strongest at sites with a larger number of species, as has been shown by Harpole *et al.* (2016) using many of these same sites. Finally, we found no consistent relationship in the influence of herbivore exclusion on local diversity measures, which in itself is consistent with other studies that find that herbivore effects on diversity depend on site context (Proulx & Mazumder 1998; Borer *et al.* 2014b; Koerner *et al.* 2018). In our case, the effects of herbivores depended on light transmission, supporting the evidence for light as an important mechanism underpinning herbivore effects on richness (Borer *et al.* 2014b).

As opposed to the local-scale patterns, we found much less variation in the scaling relationships indicated by SAR slopes (z). While local diversity varied widely in unmanipulated control plots, SAR slopes were similar across sites. It is important to note that a wide of array of processes govern SARs, and the consistent slopes do not necessarily indicate that the same processes govern diversity across these sites. We found mixed results in our tests of local environmental conditions predicted by theory to mediate SAR slopes. While we did not find the expected negative correlation between SAR slope and evenness in our larger observational data set (30 sites), we found this in the subset of 16 sites at which we conducted the Consumer-nutrient experiment. We did find some evidence for increased SAR slope at sites with large numbers of species in the observational data set. These mixed results mirror the literature. Some studies have found relationships between environmental parameters (e.g., productivity) and slopes of the SAR (e.g., Chiarucci *et al.* 2006; Moradi *et al.* 2020), while other multi-site studies failed to find strong relationships between SAR parameters and environmental characteristics (e.g., DeMalach *et al.* 2019; Dengler *et al.* 2020). Furthermore, relationships between diversity and environmental factors may vary in complex ways across scales and among different diversity metrics (Chalcraft *et al.* 2004; Chalcraft *et al.* 2008; Chalcraft *et al.* 2009). In summary, while slopes varied among sites, these scaling relationships were less variable than local diversity and minimal area of coexistence.

Changes in the scaling relationship (i.e., the slope of the SAR) due to external factors, such as nutrient addition or herbivory, also can provide important insights into the nature by which biodiversity responds to experimental treatments (reviewed in Chase *et al.* 2018). Across our study sites, nutrient addition did not systematically change the SAR slope (z), suggesting a constant proportional loss of species, and as a result, total species loss increased with area (Figure 1C & D). For example, addition of all nutrients in combination caused a mean loss of 0.9 species at the smallest scale (0.01 m²) and 3.2 species at the largest scale (75 m²). This is consistent with results from other studies that have manipulated nutrients (Lan *et al.* 2015), but others have shown either increases or decreases in z (and in some cases, β diversity, which is related to z) with nutrient addition (Chalcraft *et al.* 2008; Sandel & Corbin 2012; Leps 2014; Lan *et al.* 2015; Zhou *et al.* 2018). Likewise, we found no effect of herbivore exclusion on the scaling of diversity with area (z), whereas other studies have found positive, negative or neutral effects (Bakker *et al.* 2006; de Bello *et al.* 2007; Fernández-Lugo *et al.* 2011; Li *et al.* 2015; Godo *et al.* 2017). Taken in total, the variable results from other studies and the lack of a consistent

directional shift in the SAR slope in our analysis indicates that nutrients and herbivore effects on diversity scaling will be highly variable and context dependent.

Changes in z (or lack thereof) can be influenced by at least three different features in a regional community (He and Legendre 2002, McGill 2011, Chase and Knight 2013, Chase *et al.* 2018): (1) the density or abundance of individuals (including their size), (2) the relative abundance (evenness) of species in the community, (3) the spatial clustering of species in the region (i.e., intraspecific aggregation, influencing beta diversity). Furthermore, each of these can be altered by a number of environmental features, as well as experimental manipulations (e.g., nutrients and herbivory), but not necessarily with the same effect on z . For example, nutrient addition might simultaneously increase the size of individual plants, which may lead to higher z by reducing local diversity (Lan *et al.* 2015), while concurrently reducing the size of species pool resulting in a lower z (Lan *et al.* 2015); with the net result being no effect. Likewise, grasslands differ considerably in their ambient conditions of the three components that may influence z (DeMalach *et al.* 2019), and thus experimentally changing the biotic or abiotic environment could, for example, lead to higher or lower spatial clustering depending on where they started, which may obscure a general influence of experimental treatments on z . We did not collect the spatially-explicit abundance data needed to fully resolve the importance of these processes (Powell *et al.* 2013; McGlinn *et al.* 2019); making these measurements at a large number of locations would provide greater mechanistic insight into the processes we describe here.

The lack of a consistent change in the diversity-scaling relationships (i.e., the slope of the SAR) to nutrient addition and herbivore exclusion could arise if these treatments primarily reduce non-spatial (local) rather than spatial coexistence mechanisms (but see Chalcraft *et al.* 2008). Non-spatial mechanisms, such as tradeoffs among different resource use efficiency or susceptibility to consumers, should have consistent effects across scales. For example, nutrient addition has frequently been implicated in a loss of non-spatial coexistence mechanisms; the addition of limiting resources leads to a reduction in niche dimensions or switch to single factor limitation such as light (Goldberg & Miller 1990; Dyzinski & Tilman 2007; Hautier *et al.* 2009; Borer *et al.* 2014b; Harpole *et al.* 2016). With respect to spatial mechanisms, a lack of consistent response in z one of three possibilities. First, spatial coexistence mechanisms may not be particularly strong in these systems, at least at the scales as which we sampled ($< 75 \text{ m}^2$). Second, nutrient additions may not have a strong effect on these mechanisms. For example, dispersal limitation and local feedbacks may be more likely to structure coexistence of rare species, and these dynamics may be less strongly influenced by the effect of dominant species. Finally, there may be a lag in diversity responses at larger spatial scales, as has been shown in models with a tradeoff between competition and dispersal (e.g., the extinction debt; Tilman *et al.* 1994).

We note here that SARs arose out of empirical and theoretical work at biogeographical scales (Arrhenius 1921; Godwin 1923; MacArthur & Wilson 1967), especially focused on variation among islands or large habitat patches; however, these relationships have been conceptually useful across a range of other spatial scales (Drakare *et al.* 2006; Dengler *et al.* 2020). Here, our focus is on smaller scale variation within contiguous habitat, and our total species diversity

represents the species pool in a single grassland or meadow. Nevertheless, the slopes we estimated are quite similar to SARs reported in similar habitats but spanning much larger spatial scales (Drakare *et al.* 2006; Dengler *et al.* 2020). Despite this similarity in slopes, processes governing diversity scaling at larger regional and biogeographic scales are different than those acting at the scales we address here.

Understanding and measuring diversity is inherently scale-dependent (Godwin 1923; Gleason 1926; MacArthur & Wilson 1967; Vellend 2010; Grace *et al.* 2011; Chase & Knight 2013; Chase *et al.* 2018), and we have shown that in grassland ecosystems this scaling is remarkably robust to environmental gradients and experimental manipulations of nutrient supplies and herbivore pressure. This result does not contradict the many studies that have shown the strong impacts of nutrients in particular on grassland diversity (Borer *et al.* 2014b; Harpole *et al.* 2016; Midolo *et al.* 2019). Rather, we build on this insight in showing that nutrient-induced diversity loss often causes constant proportional loss across spatial scales. As a result, more species will be lost at larger spatial scales, and existing estimates of nutrient-induced diversity loss are likely too low, because they are typically based on a single, relatively small scale of sampling. More generally, embracing the scale-dependence of diversity, and diversity loss in response to different types of environmental change, is critical if we are to understand the impacts human activities on the biodiversity of the Earth's ecosystems.

References

1. Arrhenius, O. (1921). Species and Area. *Journal of Ecology*, 9, 95-99.
2. Bakker, E.S., Ritchie, M.E., Olff, H., Milchunas, D.G. & Knops, J.M.H. (2006). Herbivore impact on grassland plant diversity depends on habitat productivity and herbivore size. *Ecology Letters*, 9, 780-788.
3. Borer, E.T., Grace, J.B., Harpole, W.S., MacDougall, A.S. & Seabloom, E.W. (2017). A decade of insights into grassland ecosystem responses to global environmental change. *Nature Ecology & Evolution*, 1, 0118.
4. Borer, E.T., Harpole, W.S., Adler, P.B., Lind, E.M., Orrock, J.L., Seabloom, E.W. *et al.* (2014a). Finding generality in ecology: a model for globally distributed experiments. *Methods in Ecology and Evolution*, 5, 65-73.
5. Borer, E.T., Seabloom, E.W., Gruner, D.S., Harpole, W.S., Hillebrand, H., Lind, E.M. *et al.* (2014b). Herbivores and nutrients control grassland plant diversity via light limitation. *Nature*, 508, 517-520.
6. Chalcraft, D.R., Cox, S.B., Clark, C., Cleland, E.E., Suding, K.N., Weiher, E. *et al.* (2008). Scale-dependent responses of plant biodiversity to nitrogen enrichment. *Ecology*, 89, 2165-2171.
7. Chalcraft, D.R., Williams, J.W., Smith, M.D. & Willig, M.R. (2004). Scale dependence in the species-richness-productivity relationship: The role of species turnover. *Ecology*, 85, 2701-2708.
8. Chalcraft, D.R., Wilsey, B.J., Bowles, C. & Willig, M.R. (2009). The relationship between productivity and multiple aspects of biodiversity in six grassland communities. *Biodiversity and Conservation*, 18, 91-104.

9. Chane-ton, E.J. & Facelli, J.M. (1991). Disturbance effects on plant community diversity - spatial scales and dominance hierarchies. *Vegetatio*, 93, 143-155.
10. Chao, A., Gotelli, N.J., Hsieh, T.C., Sander, E.L., Ma, K.H., Colwell, R.K. *et al.* (2014). Rarefaction and extrapolation with Hill numbers: a framework for sampling and estimation in species diversity studies. *Ecological Monographs*, 84, 45-67.
11. Chase, J.M. & Knight, T.M. (2013). Scale-dependent effect sizes of ecological drivers on biodiversity: why standardised sampling is not enough. *Ecology Letters*, 16, 17-26.
12. Chase, J.M., McGill, B.J., McGlinn, D.J., May, F., Blowes, S.A., Xiao, X. *et al.* (2018). Embracing scale-dependence to achieve a deeper understanding of biodiversity and its change across communities. *Ecology Letters*, 21, 1737-1751.
13. Chase, J.M., McGill, B.J., Thompson, P.L., Antão, L.H., Bates, A.E., Blowes, S.A. *et al.* (2019). Species richness change across spatial scales. *Oikos*, 128, 1079-1091.
14. Chesson, P. (2000). Mechanisms of maintenance of species diversity. *Annual Review of Ecology and Systematics*, 31, 343-366.
15. Chiarucci, A., Viciani, D., Winter, C. & Diekmann, M. (2006). Effects of productivity on species–area curves in herbaceous vegetation: evidence from experimental and observational data. *Oikos*, 115, 475-483.
16. Connor, E.F. & McCoy, E.D. (1979). The statistics and biology of the species-area relationship. *American Naturalist*, 113, 791-833.
17. Crawley, M.J., Johnston, A.E., Silvertown, J., Dodd, M., de Mazancourt, C., Heard, M.S. *et al.* (2005). Determinants of species richness in the park grass experiment. *American Naturalist*, 165, 179-192.
18. Crist, T.O. & Veech, J.A. (2006). Additive partitioning of rarefaction curves and species–area relationships: unifying α -, β - and γ -diversity with sample size and habitat area. *Ecology Letters*, 9, 923-932.
19. de Bello, F., Lepš, J. & Sebastià, M.-T. (2007). Grazing effects on the species-area relationship: Variation along a climatic gradient in NE Spain. *Journal of Vegetation Science*, 18, 25-34.
20. DeMalach, N., Saiz, H., Zaady, E. & Maestre, F.T. (2019). Plant species–area relationships are determined by evenness, cover and aggregation in drylands worldwide. *Global Ecology and Biogeography*, 28, 290-299.
21. Dengler, J., Matthews, T.J., Steinbauer, M.J., Wolfrum, S., Boch, S., Chiarucci, A. *et al.* (2020). Species–area relationships in continuous vegetation: Evidence from Palaearctic grasslands. *Journal of Biogeography*, 47, 72-86.
22. Drakare, S., Lennon, J.J. & Hillebrand, H. (2006). The imprint of the geographical, evolutionary and ecological context on species-area relationships. *Ecology Letters*, 9, 215-227.
23. Dybzinski, R. & Tilman, D. (2007). Resource use patterns predict long-term outcomes of plant competition for nutrients and light. *American Naturalist*, 170, 305-318.
24. Fernández-Lugo, S., de Nascimento, L., Mellado, M. & Arévalo, J.R. (2011). Grazing effects on species richness depends on scale: a 5-year study in Tenerife pastures (Canary Islands). *Plant Ecology*, 212, 423-432.
25. Flather, C. (1996). Fitting species–accumulation functions and assessing regional land use impacts on avian diversity. *Journal of Biogeography*, 23, 155-168.

26. Foley, J.A., DeFries, R., Asner, G.P., Barford, C., Bonan, G., Carpenter, S.R. *et al.* (2005). Global consequences of land use. *Science*, 309, 570-574.
27. Fridley, J.D., Peet, R.K., Wentworth, T.R. & White, P.S. (2005). Connecting fine- and broad-scale species-area relationships of Southeastern US Flora. *Ecology*, 86, 1172-1177.
28. Gleason, H.A. (1926). The individualistic concept of the plant association. *Bulletin of Torrey Botanical Society*, 53, 7-26.
29. Godo, L., Orsolya, V., Bela, T., Torok, P., Kelemen, A. & Deak, B. (2017). Scale-dependent effects of grazing on the species richness of alkaline and sand grasslands. *Tuexenia*, 229-246.
30. Godwin, H. (1923). Dispersal of pond flora. *Journal of Ecology*, 11, 160-164.
31. Goldberg, D.E. (1987). Neighborhood competition in an old-field plant community. *Ecology*, 68, 1211-1223.
32. Goldberg, D.E. & Miller, T.E. (1990). Effects of different resource additions on species-diversity in an annual plant community. *Ecology*, 71, 213-225.
33. Grace, J.B., Harrison, S. & Damschen, E.I. (2011). Local richness along gradients in the Siskiyou herb flora: R. H. Whittaker revisited. *Ecology*, 92, 108-120.
34. Harpole, W.S., Sullivan, L.L., Lind, E.M., Firn, J., Adler, P.B., Borer, E.T. *et al.* (2016). Addition of multiple limiting resources reduces grassland diversity. *Nature*, 537, 93-96.
35. Harpole, W.S. & Tilman, D. (2007). Grassland species loss resulting from reduced niche dimension. *Nature*, 446, 791-793.
36. Hart, S.P., Usinowicz, J. & Levine, J.M. (2017). The spatial scales of species coexistence. *Nature Ecology & Evolution*, 1, 1066-1073.
37. Hastings, A. (1980). Disturbance, coexistence, history, and competition for space. *Theoretical Population Biology*, 18, 363-373.
38. Hautier, Y., Niklaus, P.A. & Hector, A. (2009). Competition for Light Causes Plant Biodiversity Loss After Eutrophication. *Science*, 324, 636-638.
39. Heatwole, H. (1975). Biogeography of reptiles on some of the islands and cays of eastern Papua-New Guinea. *Atoll Research Bulletin*, 180.
40. Hillebrand, H., Gruner, D.S., Borer, E.T., Bracken, M.E.S., Cleland, E.E., Elser, J.J. *et al.* (2007). Consumer versus resource control of producer diversity depends on ecosystem type and producer community structure. *Proceedings of the National Academy of Sciences of the United States of America*, 104, 10904-10909.
41. Holt, R.D., Grover, J. & Tilman, D. (1994). Simple Rules for Interspecific Dominance in Systems with Exploitative and Apparent Competition. *American Naturalist*, 144, 741-771.
42. Hutchinson, G.E. (1961). The paradox of the plankton. *The American Naturalist*, 95, 137-145.
43. Koerner, S.E., Smith, M.D., Burkepile, D.E., Hanan, N.P., Avolio, M.L., Collins, S.L. *et al.* (2018). Change in dominance determines herbivore effects on plant biodiversity. *Nature Ecology & Evolution*, 2, 1925-1932.
44. Lan, Z.C., Jenerette, G.D., Zhan, S.X., Li, W.H., Zheng, S.X. & Bai, Y.F. (2015). Testing the scaling effects and mechanisms of N-induced biodiversity loss: evidence from a decade-long grassland experiment. *Journal of Ecology*, 103, 750-760.
45. Leibold, M.A. & Chase, J.M. (2017). *Metacommunity Ecology*. Princeton University Press.

46. Leibold, M.A., Holyoak, M., Mouquet, N., Amarasekare, P., Chase, J.M., Hoopes, M.F. *et al.* (2004). The metacommunity concept: a framework for multi-scale community ecology. *Ecology Letters*, 7, 601-613.
47. Leps, J. (2014). Scale- and time-dependent effects of fertilization, mowing and dominant removal on a grassland community during a 15-year experiment. *Journal of Applied Ecology*, 51, 978-987.
48. Li, W., Zhan, S., Lan, Z., Ben Wu, X. & Bai, Y. (2015). Scale-dependent patterns and mechanisms of grazing-induced biodiversity loss: evidence from a field manipulation experiment in semiarid steppe. *Landscape Ecology*, 30, 1751-1765.
49. Lind, E.M., Borer, E., Seabloom, E., Adler, P., Bakker, J.D., Blumenthal, D.M. *et al.* (2013). Life-history constraints in grassland plant species: a growth-defence trade-off is the norm. *Ecology Letters*, 16, 513-521.
50. MacArthur, R.H. & Wilson, E.O. (1967). *The theory of island biogeography* Princeton University Press, Princeton, NJ, USA.
51. McGlinn, D.J., Xiao, X., May, F., Gotelli, N.J., Engel, T., Blowes, S.A. *et al.* (2019). Measurement of Biodiversity (MoB): A method to separate the scale-dependent effects of species abundance distribution, density, and aggregation on diversity change. *Methods in Ecology and Evolution*, 10, 258-269.
52. Midolo, G., Alkemade, R., Schipper, A.M., Benitez-Lopez, A., Perring, M.P. & De Vries, W. (2019). Impacts of nitrogen addition on plant species richness and abundance: A global meta-analysis. *Global Ecol Biogeogr*, 28, 398-413.
53. Moradi, H., Fattorini, S. & Oldeland, J. (2020). Influence of elevation on the species–area relationship. *J Biogeogr*, n/a.
54. Mouquet, N. & Loreau, M. (2003). Community patterns in source-sink metacommunities. *American Naturalist*, 162, 544-557.
55. Oksanen, J. (1996). Is the humped relationship between species richness and biomass an artefact due to plot size? *Journal of Ecology*, 84, 293-295.
56. Olff, H. & Ritchie, M.E. (1998). Effects of herbivores on grassland plant diversity. *Trends Ecol Evol*, 13, 261-265.
57. Powell, K.I., Chase, J.M. & Knight, T.M. (2013). Invasive Plants Have Scale-Dependent Effects on Diversity by Altering Species-Area Relationships. *Science*, 339, 316-318.
58. Proulx, M. & Mazumder, A. (1998). Reversal of grazing impact on plant species richness in nutrient-poor vs. nutrient-rich ecosystems. *Ecology*, 79, 2581-2592.
59. R Development Core Team (2010). R: A language and environment for statistical computing R Foundation for Statistical Computing, Vienna, Austria.
60. Ripple, W.J., Newsome, T.M., Wolf, C., Dirzo, R., Everatt, K.T., Galetti, M. *et al.* (2015). Collapse of the world's largest herbivores. *Science Advances*, 1.
61. Sandel, B. & Corbin, J.D. (2012). Scale-dependent responses of species richness to experimental manipulation of productivity and disturbance in Californian coastal grasslands. *Journal of Vegetation Science*, 23, 906-918.
62. Scheiner, S.M., Chiarucci, A., Fox, G.A., Helmus, M.R., McGlinn, D.J. & Willig, M.R. (2011). The underpinnings of the relationship of species richness with space and time. *Ecological Monographs*, 81, 195-213.

63. Seabloom, E.W., Bjornstad, O.N., Bolker, B.M. & Reichman, O.J. (2005). The spatial signature of environmental heterogeneity, dispersal, and competition in successional grasslands. *Ecological Monographs*, 75, 199-214.
64. Smith, B. & Wilson, J.B. (1996). A consumer's guide to evenness indices. *Oikos*, 76, 70-82.
65. Steffen, W., Richardson, K., Rockström, J., Cornell, S.E., Fetzer, I., Bennett, E.M. *et al.* (2015). Planetary boundaries: Guiding human development on a changing planet. *Science*.
66. Thompson, P.L., Guzman, L.M., De Meester, L., Horváth, Z., Ptacnik, R., Vanschoenwinkel, B. *et al.* (2020). A process-based metacommunity framework linking local and regional scale community ecology. *Ecology Letters*, 23, 1314-1329.
67. Tilman, D. (1982). *Resource competition and community structure*. Princeton University Press, Princeton, NJ.
68. Tilman, D., May, R.M., Lehman, C.L. & Nowak, M.A. (1994). Habitat destruction and the extinction debt. *Nature (London)*, 371, 65-66.
69. Vellend, M. (2010). Conceptual Synthesis in Community Ecology. *Quarterly Review of Biology*, 85, 183-206.
70. Viola, D.V., Mordecai, E.A., Jaramillo, A.G., Sistla, S.A., Albertson, L.K., Gosnell, J.S. *et al.* (2010). Competition-defense tradeoffs and the maintenance of plant diversity. *Proceedings of the National Academy of Sciences of the United States of America*, 107, 17217-17222.
71. Zhou, X., Liu, X., Zhang, P., Guo, Z. & Du, G. (2018). Increased community compositional dissimilarity alleviates species loss following nutrient enrichment at large spatial scales. *Journal of Plant Ecology*, 12, 376-386.

Acknowledgments

This work was generated using data from the Nutrient Network (<http://www.nutnet.org>) experiment, funded at the site scale by individual researchers. Coordination and data management have been supported by funding to E. Borer and E. Seabloom from the National Science Foundation Research Coordination Network (NSF-DEB-1042132) and Long Term Ecological Research (NSF-DEB-1234162 & DEB-1831944 to Cedar Creek LTER) programs, and the University of Minnesota's Institute on the Environment (DG-0001-13). We also thank the Minnesota Supercomputer Institute for hosting project data and the Institute on the Environment for hosting Network meetings.

Figure 1: Hypothetical effects of experimental treatments (e.g., nutrient addition or herbivore exclusion) on species richness across spatial scales. The solid black line is constant in all panels and shows the species-area relationship (SAR) in control plots plotted in logged and untransformed units. Panel A shows the slope (z), y-intercept ($\log c$), and x-intercept (A_{min}), which are the focus of the analyses in this paper. The black dotted line shows the SAR in the treated plots, and the red arrows show the change in richness at small or large scales. The blue dashed lines show the x and y intercepts in log space ($\log(A)=0$ and $\log(S)=0$). Left panels show $\log(\text{Area})$ and right panels show the same relationship with area untransformed. The dotted lines in panels A and B show the effects of a reduced species pool, which reduces the slope but leaves the y-intercept unchanged. In this case, species loss increases with increasing spatial scale. Panels C and D show the effects of a constant proportional loss of species with increasing area (C), which leads to increasing total species loss with area (D). In this case, the y-intercept is reduced, while the slope is held constant. Panels E and F show the effects of reduced local richness without a reduction in the total species pool. In this case, the y-intercept is decreased but the slope is increased, such that species loss declines with area.

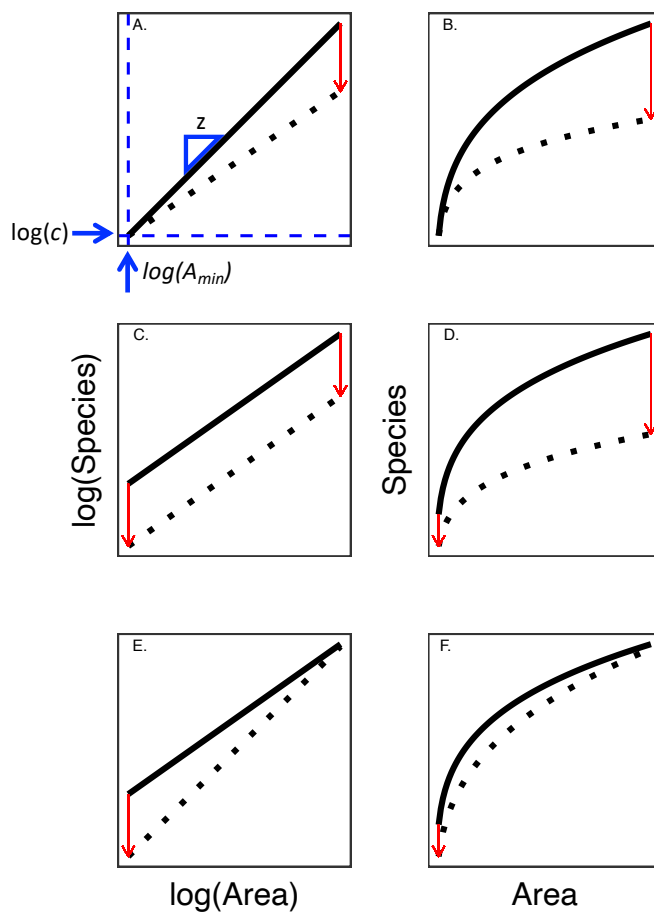


Figure 2: Effects of consumers and nutrient addition on slope (z), ($\log c$), and x-intercept (a) of the species-area relationships (SAR) in grasslands (16 sites). SAR plots (Panels A & B) use the mean parameter value for each treatment (Panels C - E). Open circle shows the values in control plots at the larger set of 30 observational sites which includes the 16 experimental sites (solid circles). Error bars represent 1 standard error of the mean (SEM). Full analysis is shown in Table S3, which includes estimates of treatment effects sizes and significance.

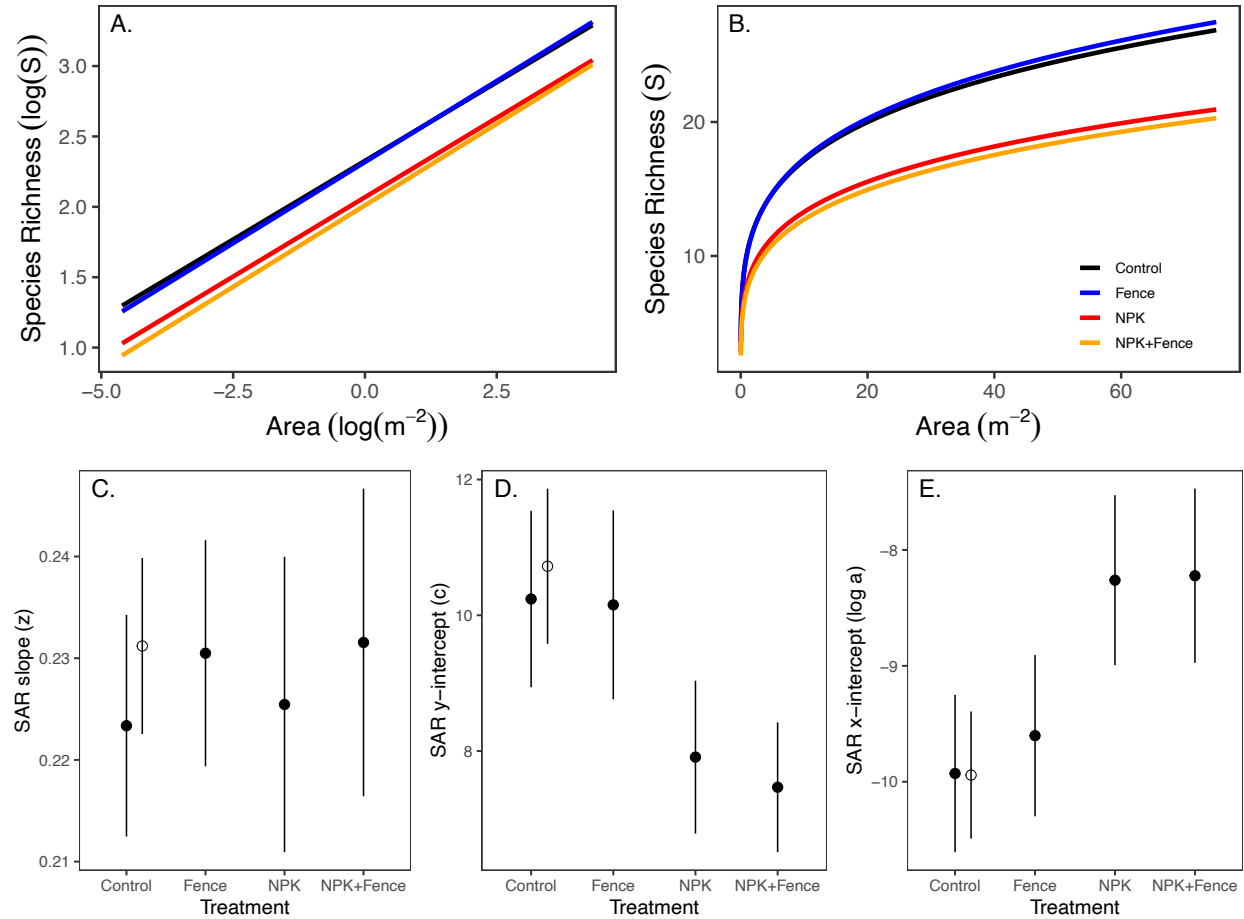


Figure 3: Effects of nutrient addition on slope (z), ($\log c$), and x-intercept (a) of the species-area relationships (SAR) in grasslands (21 sites). SAR plots (Panels A & B) use the mean parameter value for each treatment (Panels C - E). Open circles show the values in Control plots at the larger set of 30 observational sites which includes the 21 experimental sites (solid circles). Error bars represent 1 standard error of the mean (SEM). Full analysis is shown in Table S4, which includes estimates of treatment effects sizes and significance.

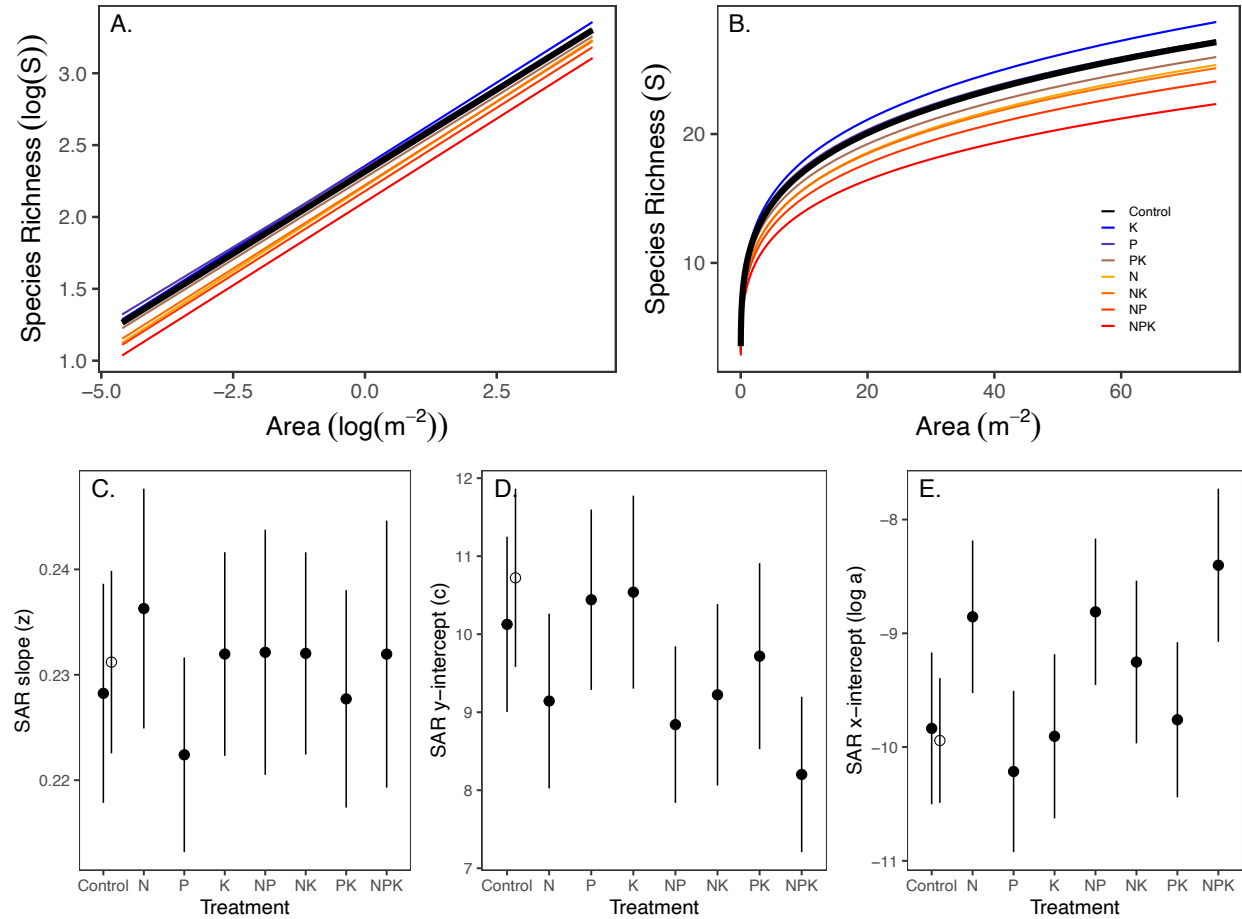


Figure 4: Effects of proportion light transmission, aboveground live biomass, site richness, and evenness on the slope (z), (log c), and x-intercept (a) of the species-area relationships (SAR) in grasslands. Solid points show SAR parameters for the subset of sites with all fencing and nutrient addition treatments (16 sites). Open circles and dashed lines show SAR parameters from sites with control plots (black lines, 30 sites) and the subset of these with nutrient addition treatments but not fencing (red lines, 21 sites). Lines are shown only for significant regressions. Full analysis is presented in Table S5.

