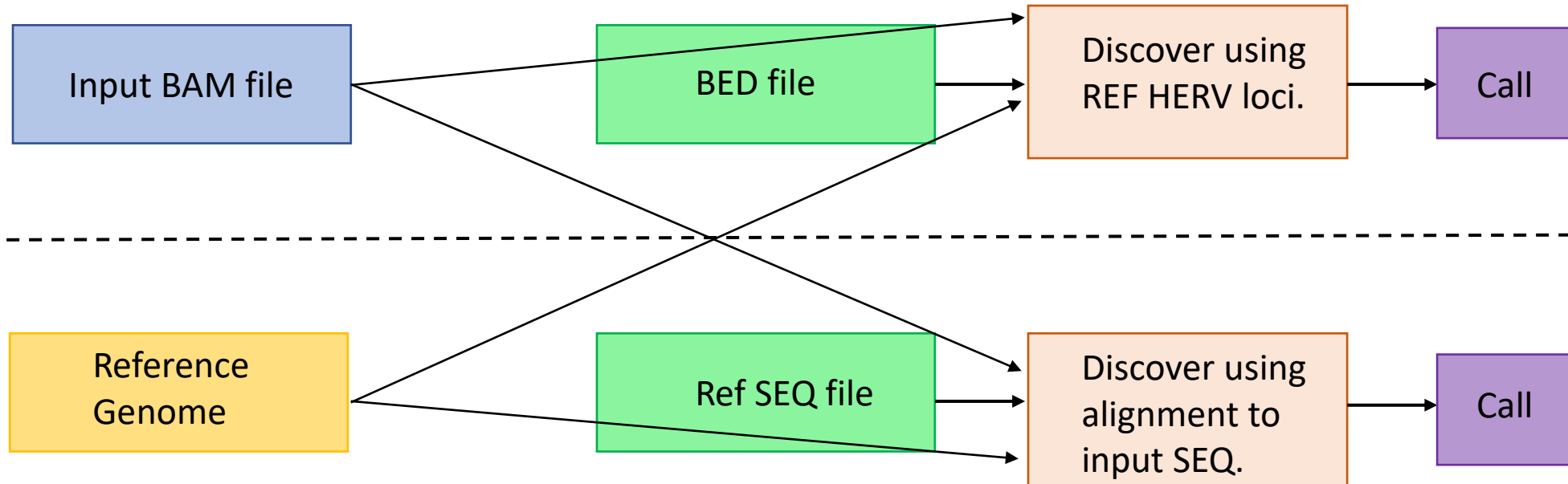


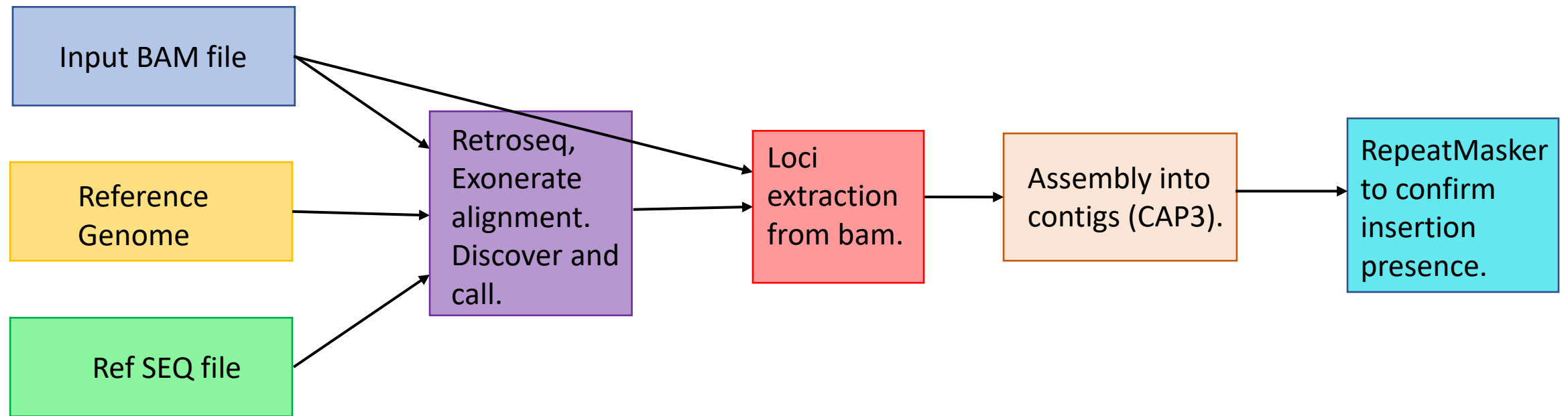
Flow diagrams for tested
tools

Retroseq



Retroseq can be implemented either using alignment to a reference fasta of a target sequence (e.g. LTR5_Hs) or instead by mapping discordant reads to reference loci – assuming that for a non-reference insertion the reads are aligned to reference copies. Alignment requires exonerate and takes longer to run. More information can be found here: <https://github.com/tk2/RetroSeq>

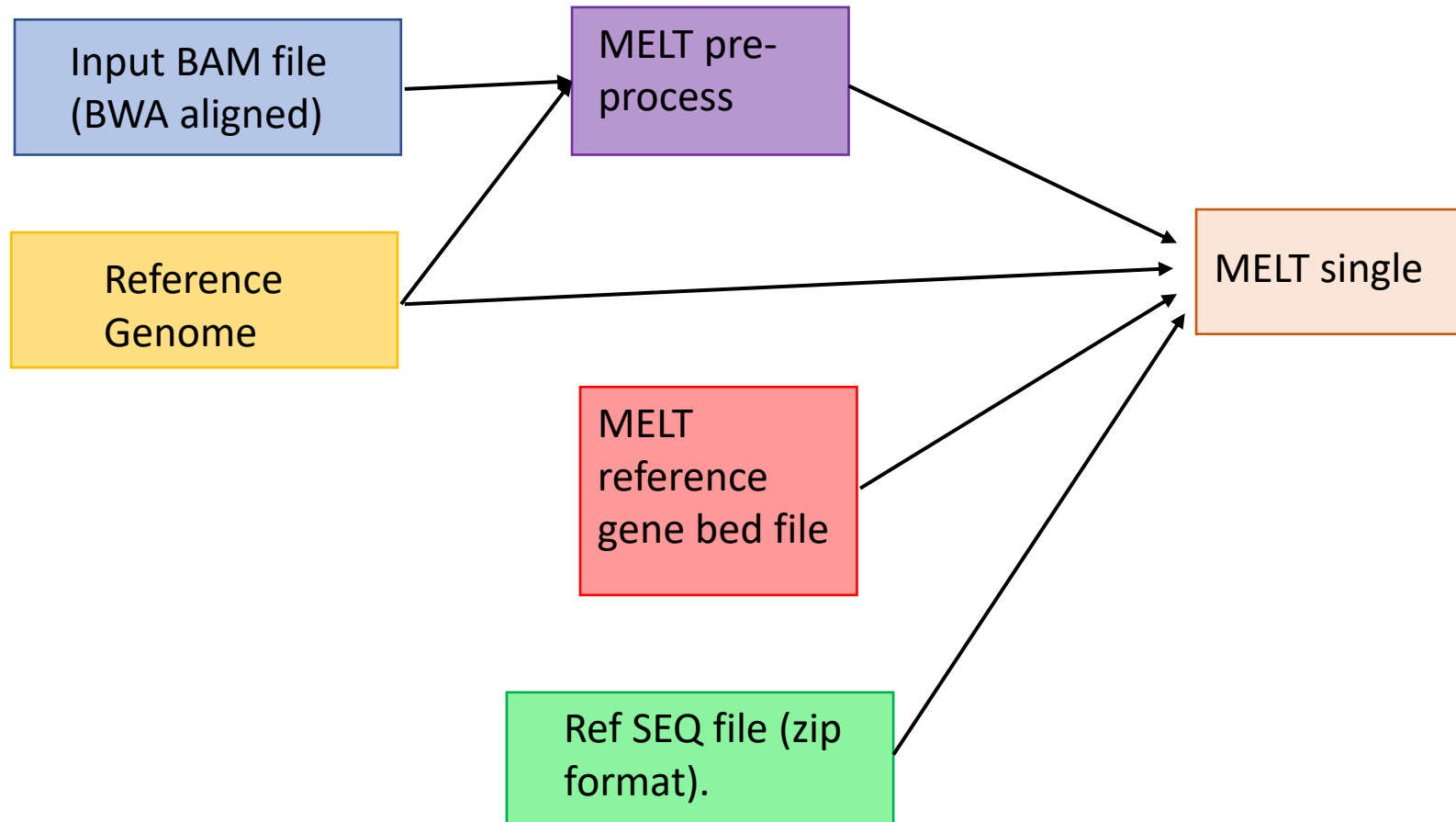
Retroseq+



Retroseq+ first applies Retroseq to an input bam file, high confidence loci are filtered. Reads at these loci are extracted from the input bam file and assembled into continuous sequences. These sequences are input to repeatmasker which identifies repetitive elements. If contigs are positive for target sequence in contigs up and downstream from the locus, they are output into the final result. More information can be found here:

<https://www.pnas.org/doi/10.1073/pnas.1602336113>

MELT



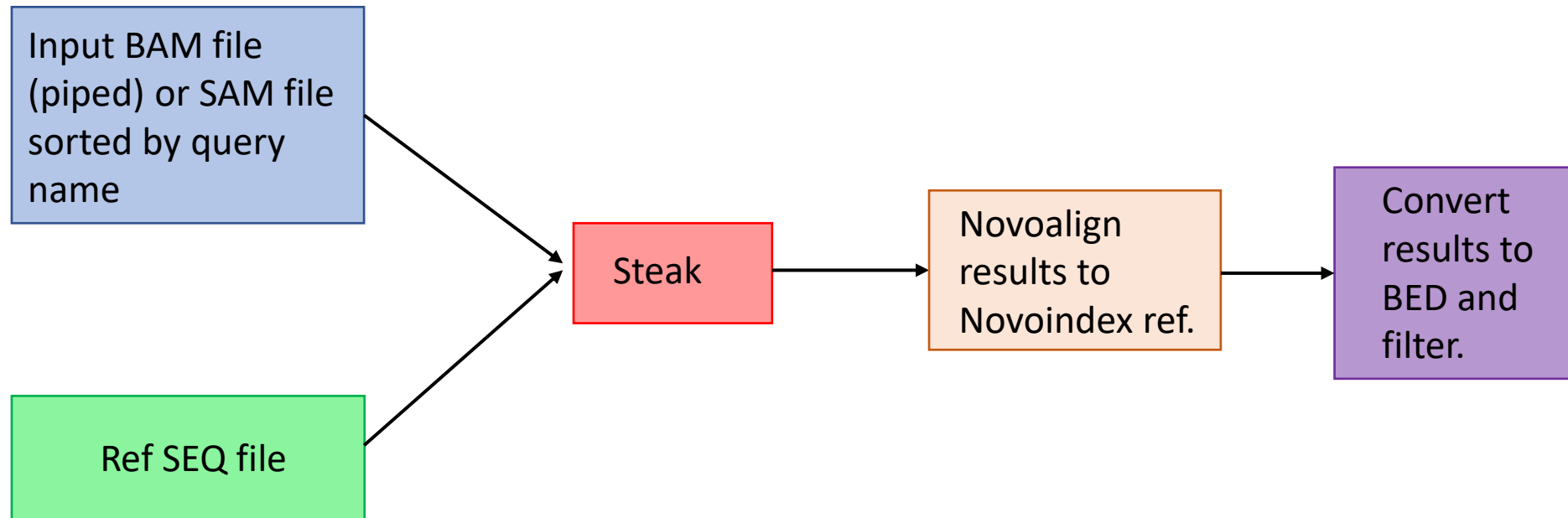
MELT has an initial pre-processing step to speed up run-time.

This step produces 3 files related to discordant read pairs.

Following this, MELT 'single' is run to identify HERV-K loci. More information can be found here:

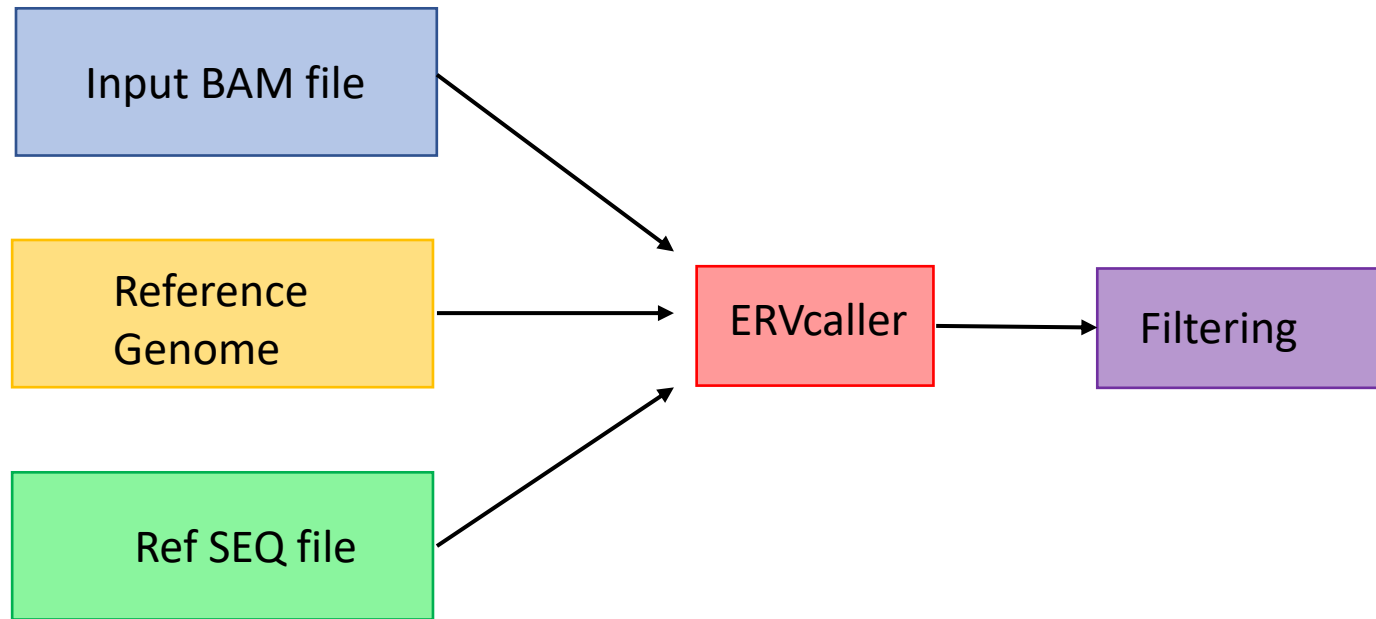
<https://melt.igs.umaryland.edu/manual.php>

Steak



For our analysis, inputs were in SAM format. Steak takes in a genome sequence file and a fasta of the target insertion. Following the Steak command, post-processing with Novoalign and filtering is applied as outlined on the Steak github page. More information can be found here: <https://github.com/applevir/STEAK>

ERVcaller



ERVcaller is the simplest tool to run, with no pre-processing. Recommended filtering options for ERVcaller results are outlined in the ERVcaller github page and we used these recommended filters for our analysis. More information can be found here: <https://github.com/xunchen85/ERVcaller>