# Package 'sceptre'

September 28, 2020
Type Package
Title Conditional resampling differential expression analysis for pooled CRISPR screens
Version 0.1.0
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<b>Description</b> Functions for performing differential expression analyses in single-cell pooled CRISPR screens based on conditional resampling.
License GPL-3
Encoding UTF-8
LazyData true
RoxygenNote 7.1.1
Depends sn, MASS, VGAM, tidyverse, R (>= 3.10)
Suggests knitr, rmarkdown
VignetteBuilder knitr
R topics documented:
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```
run_gene_precomputation
```

Run gene precomputation

## **Description**

This function runs the precomputation for a given gene. In particlar, it fits an NB regression of expression against covariates. The estimate of theta (i.e., the NB dispersion parameter) is obtained from glm.nb function. This is sensible as, under the null hypothesis, the NB model without the gRNA indicator is true. Offsets are obtained by log-transforming the fitted values.

# Usage

```
run_gene_precomputation(
  expressions,
  covariate_matrix,
  gene_precomp_dispersion,
  gene_precomp_offsets
)
```

## **Arguments**

```
expressions the vector of gene expressions

covariate_matrix

the cell-specific covariate matrix

gene_precomp_dispersion

the pre-computed dispersion parameter (NULL if none)

gene_precomp_offsets

the pre-computed gene offsets (NULL of none)
```

## Value

a named list containing two items: offsets and dispersion.

## **Examples**

```
sim_dat <- simulate_crispr_screen_data(num_cells = 1000,
grna_mean_prob = 0.2,
covariate_sampler = list(cell_size = rnorm, cell_cycle = runif),
mRNA_mean_expression = 40,
gRNA_effect = 0,
covariate_effects = c(0.5, 1),
zero_inflation = 0,
neg_binom_size = 2)
expressions <- sim_dat$Y
covariate_matrix <- sim_dat$covariate_df
gene_precomp <- run_gene_precomputation(expressions, covariate_matrix, NULL, NULL)</pre>
```

```
run_gRNA_precomputation
```

Run gRNA precomputation

## **Description**

This function runs the precomputation for a given gRNA.

## Usage

```
\verb"run_gRNA_precomputation" (gRNA_indicators, covariate\_matrix")
```

#### **Arguments**

```
gRNA_indicators a\ vector\ of\ gRNA\ indicators covariate\_matrix the\ cell-specific\ covariate\ matrix
```

#### Value

the fitted probabilities

## **Examples**

```
sim_dat <- simulate_crispr_screen_data(num_cells = 1000,
grna_mean_prob = 0.2,
covariate_sampler = list(cell_size = rnorm, cell_cycle = runif),
mRNA_mean_expression = 40,
gRNA_effect = -4,
covariate_effects = c(0.5, 1),
zero_inflation = 0,
neg_binom_size = 2)
gRNA_indicators <- sim_dat$X
covariate_matrix <-sim_dat$covariate_df
fitted_probs <- run_gRNA_precomputation(gRNA_indicators, covariate_matrix)</pre>
```

```
run_sceptre_gRNA_gene_pair
```

Run sceptre on a gRNA-gene pair

## **Description**

This function runs the sceptre algorithm on a single gRNA-gene pair. It requires as arguments the gene expression vector, the gRNA indicator vector, and the covariate matrix. Users optionally can pass the gRNA precomputation or gene precomputation as arguments.

#### Usage

```
run_sceptre_gRNA_gene_pair(
  expressions,
  gRNA_indicators,
  covariate_matrix,
  gRNA_precomp = NULL,
  gene_precomp_dispersion = NULL,
  gene_precomp_offsets = NULL,
  B = 500,
  seed = NULL
)
```

#### **Arguments**

```
expressions
                  a vector a gene expressions
gRNA_indicators
                  a vector of gRNA inicators
covariate_matrix
                  the matrix of cell-specific covariates (e.g., library size, batch effect, cell cycle,
                  (optional) the gRNA precomputation (a vector of gRNA presence conditional
gRNA_precomp
                  probabilities)
gene_precomp_dispersion
                  (optional) the pre-computed gene dispersion
gene_precomp_offsets
                  (optional) the pre-computed gene offsets
                  number of resamples (default 500)
В
                  (optional) seed to the random number generator
seed
```

## Value

a p-value of the null hypothesis of no gRNA effect on gene expression

#### **Examples**

```
# An example in which the alternative is true.
sim_dat <- simulate_crispr_screen_data(num_cells = 1000,</pre>
grna_mean_prob = 0.2,
covariate_sampler = list(cell_size = rnorm, cell_cycle = runif),
mRNA_mean_expression = 40,
gRNA\_effect = -4,
covariate_effects = c(0.5, 1),
zero_inflation = 0,
neg\_binom\_size = 2)
expressions <- sim_dat$Y</pre>
gRNA_indicators <- sim_dat$X</pre>
covariate_matrix <-sim_dat$covariate_df</pre>
run_sceptre_gRNA_gene_pair(expressions = expressions,
gRNA_indicators = gRNA_indicators,
covariate_matrix = covariate_matrix)
# An example in which the null is true.
```

```
sim_dat <- simulate_crispr_screen_data(num_cells = 1000,
grna_mean_prob = 0.2,
covariate_sampler = list(cell_size = rnorm, cell_cycle = runif),
mRNA_mean_expression = 40,
gRNA_effect = 0,
covariate_effects = c(0.5, 1),
zero_inflation = 0,
neg_binom_size = 2)
expressions <- sim_dat$Y
gRNA_indicators <- sim_dat$X
covariate_matrix <-sim_dat$covariate_df
run_sceptre_gRNA_gene_pair(expressions = expressions,
gRNA_indicators = gRNA_indicators,
covariate_matrix = covariate_matrix)</pre>
```

run\_sceptre\_using\_precomp

Run sceptre using precomputations for gRNAs and genes.

#### **Description**

This function is the workhorse function of the sceptre package. It runs a distilled CRT using a negative binomial test statistic based on an expression vector, a gRNA indicator vector, an offset vector (from the distillation step), gRNA conditional probabilities, an estimate of the negative binomial dispersion parameter, and the number of resampling replicates.

# Usage

```
run_sceptre_using_precomp(
  expressions,
  gRNA_indicators,
  gRNA_precomp,
  gene_precomp_dispersion,
  gene_precomp_offsets,
  B,
  seed
)
```

## Arguments

#### **Details**

This currently is a one-tailed, left-sided test. Thus, it is suitable for up-regulatory elements like enhancers and promoters but not down-regulatory elements like silencers.

#### Value

a p-value of the null hypothesis of no gRNA effect on gene expression

```
simulate_crispr_screen_data

Simulate CRISPR screen data.
```

## Description

A function to simulate pooled CRISPR screen data.

## Usage

```
simulate_crispr_screen_data(
  num_cells,
  grna_mean_prob,
  covariate_sampler,
  mRNA_mean_expression,
  gRNA_effect,
  covariate_effects,
  zero_inflation,
  neg_binom_size,
  seed = NULL
)
```

## **Arguments**

```
number of cells in experiment
grna_mean_prob mean number of cells perturbed (absent covariates)
covariate_sampler
                  a list of functions. Each function in this list takes as an argument num_cells and
                  returns num_cells random variates.
mRNA_mean_expression
                  mean mRNA expression of the cell (absent covariates)
gRNA_effect
                  effect of the gRNA perturbation on mRNA expression
covariate_effects
                  a numeric vector indicating the covariate effects on mRNA expression
zero_inflation a numeric scalar between 0 and 1 (inclusive) giving the mean fraction of cell
                  expressions set to zero
neg_binom_size size parameter for the negative binomial model
                  a seed to the random number generator
seed
```

#### Value

a list containing (i) the expression vector Y, (ii) the data frame of covariates, and (iii) the gRNA presence indicator vector.

## **Examples**

```
num_cells <- 1000
grna_mean_prob <- 0.2</pre>
covariate_sampler <- list(</pre>
cell_size = runif,
cell_cycle = function(x) \{runif(n = x, min = 0, max = 1)\}
mRNA_mean_expression <- 40
gRNA_effect <- 4
covariate_effects <- c(2, 1)</pre>
zero_inflation <- 0</pre>
neg_binom_size <- 2</pre>
simulated_data <- simulate_crispr_screen_data(num_cells,</pre>
 grna_mean_prob,
 covariate_sampler,
 mRNA_mean_expression,
 gRNA_effect,
 covariate_effects,
 zero_inflation,
 neg_binom_size)
```

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