

A tutorial on how to use the Brain Observatory Toolbox

This tutorial will show you how to access the Brain Observatory data using MATLAB. In particular, this tutorial will show you how to:

- 1) Download meta information from brain observatory api and save it as MATLAB datatype
- 2) Build a `brain_observatory_cache` object to:
 - A) Get general information on the brain observatory data
 - B) Filter sessions by specified criteria in different aspects such as brain areas, imaging depth and stimuli type
 - C) Download NWB files of filtered sessions
- 3) Extract imaging data from NWB files to:
 - a) Get and plot fluorescence traces
 - b) Convert fluorescence trace into data that is in "raster format" for further analysis

Organization of the Brain Observatory Data

Before we start, we first want to describe the organization of the Brain Observatory Data.

An *experiment container* contains three *sessions* (also called *experiments* or *ophys_experiments*) where recordings were made on a single mouse, in a single brain region and at a particular imaging depth. Each of these three sessions consists of a series of "subexperiments" where a particular stimulus set was shown. NWB files downloaded from the Allen Institute API each consist of data from a single session. As described below, all data that we extract into "raster format" consists of data from a single "subexperiment" where a particular stimulus set was shown.

Note: within an experiment container the same stimuli might be repeated in different sessions (i.e., different sessions can have the same "subexperiment type"). For example, Natural Movie 1 is shown in all three sessions in an experiment container. For more information see: http://alleninstitute.github.io/AllenSDK/brain_observatory.html

1) Download meta information from brain observatory api and save it as MATLAB datatype

To begin, set up the path to Brain Observatory Toolbox:

```
bot_dir_name = 'Brain-Observatory-Toolbox/';
```

Then, add it to MATLAB's search path

```
addpath(bot_dir_name)
```

Next we will download the meta information which is stored in manifest files on the Allen API. We can do this using the function `get_manifests_info_from_api()`. This function creates a MATLAB structure array with three tables and saves this structure array in a file called `manifests.mat`. `Manifests.mat` will be taken by the `brain_observatory_cache` as shown in the following section.

```
get_manifests_info_from_api()
```

Elapsed time is 0.000814 seconds.

```
load('manifests')
```

There are three tables in `manifest.mat`: `container_manifest` contains metadata of all experiment containers (m containers by n aspects of meta data), `session_manifest` contains metadata of all sessions (m sessions by n aspects of meta data), and `cell_id_mapping` contains mapping between new cell ids and the old cell id of all cells (m cells by number of ids).

As quoted from AllenSDK 's release note (0.13.2): "The cross-session alignment algorithm has been updated and re-run, so all cell specimen IDs have changed".

2) Build a brain_observatory_cache object

`Brain_observatory_cache` is a class that resembles `BrainObservatoryCache` in `allensdk` (https://github.com/AllenInstitute/AllenSDK/blob/master/allensdk/core/brain_observatory_cache.py).

It takes `manifests` and helps you fanthom and access Brain Observatory Data through three stages:

Stage A, it returns general information on the brain observatory data from different aspects through correponding methods whose names all start with `"get_"` ;

Stage B, it filters sessions by different criteria through correponding methods with generic name pattern `"filter_...by_..."`;

Stage C, it downloads NWB file(s) of filtered session(s) through the method named `"download_nwb"`.

To beign, build a `brain_observatory_cache_object`

```
boc = brain_observatory_cache(manifests)
```

```
boc =  
    brain_observatory_cache with properties:  
        session_table: [597×14 table]  
        container_table: [199×13 table]  
        manifests: [1×1 struct]  
        filtered_session_table: [597×14 table]
```

```

        stimuli: {1×17 cell}
targeted_structure: {6×1 cell}
    imaging_depth: [11×1 double]
        container_id: [199×1 double]
            session_id: [597×1 double]
        session_type: {4×1 cell}

```

As you can see here, manifests.mat is stored in property manifests, table session_manifest from manifest.mat is stored in property session_table, and table container_manifest from manifest.mat is stored in property container_table.

The other properties will shine more light during the Stage B: filtering sessions; I will explain them then.

2A) Get general information on the brain observatory data

We implemented several methods whose names all start with "get_" to summarize the brain observatory data from different aspects.

Here shown total number of containers

```
boc.get_total_num_of_containers()
```

```
ans = 199
```

Note: Total number of sessions should be whatever this returns * 3

Here shown all cortical depths that were ever recorded in any experiment container

```
boc.get_all_imaging_depths()
```

```

ans =
    175
    265
    275
    300
    320
    325
    335
    350
    365
    375
    ⋮
    ⋮
    •

```

Notet: Recording didn't take place equally among these depths, only popular depths are shown on homepage of Brain Observatory: <http://observatory.brain-map.org/visualcoding>

Here shown all type of cre driver lines of all mice participated

```
boc.get_all_cre_lines()
```

```
ans = 6x1 cell array
    'Cux2-CreERT2'
    'Emx1-IRES-Cre'
    'Nr5a1-Cre'
    'Rbp4-Cre_KL100'
    'Rorb-IRES2-Cre'
    'Scnn1a-Tg3-Cre'
```

Note: All mice had the same reporter line: Ai93 and tTA driver line: Camk2a-tTA

For more information about transgenic lines: <http://observatory.brain-map.org/visualcoding/transgenic>

Here shown all brain regions that were ever recorded in any experiment container

```
boc.get_all_targeted_structures()
```

```
ans = 6x1 cell array
    'VISal'
    'VISam'
    'VISl'
    'VISp'
    'VISpm'
    'VISrl'
```

For their locations, see homepage of Brain Observatory: <http://observatory.brain-map.org/visualcoding>

Here shown all types of sessions that ever appeared in any experiment container

```
boc.get_all_session_types()
```

```
ans = 4x1 cell array
    'three_session_A'
    'three_session_B'
    'three_session_C'
    'three_session_C2'
```

Note: There are always three sessions in each container: session A, session B, and session C or session C2

Here shown all type of stimuli that were ever used in any of the four types of sessions

```
boc.get_all_stimuli()
```

```
ans = 9x1 cell array
    'drifting_gratings'
    'locally_sparse_noise_eight_degree'
    'locally_sparse_noise_four_degree'
    'natural_movie_one'
    'natural_movie_three'
    'natural_movie_two'
    'natural_scene'
```

```
'spontaneous_activity'
'static_gratings'
```

Note: For mapping between session type and stimulus type, see http://alleninstitute.github.io/AllenSDK/brain_observatory.html

Here shown the number of experiment containers recorded in each brain region

```
boc.get_summary_of_containers_along_targeted_structures()
```

```
VISal    33
VISam    25
VISl     36
VISp     54
VISpm    35
VISrl    16
```

Here shown the number of experiment containers recorded at each cortical depth

```
boc.get_summary_of_containers_along_imaging_depths()
```

```
175      50
265       1
275      72
300       4
320       1
325       3
335       3
350      33
365       1
375      30
435       1
```

Here shown the number of experiment containers recorded at each cortical depth in each brain region

```
boc.get_summary_of_containers_along_depths_and_structures()
```

```
ans = 12x7 table
```

	VISal	VISam	VISl	VISp	VISpm	VISrl	total
	-----	-----	-----	-----	-----	-----	-----
175	8	5	10	11	11	5	50
265	1	0	0	0	0	0	1
275	12	9	15	20	11	5	72
300	1	0	1	1	1	0	4
320	0	1	0	0	0	0	1
325	0	1	1	0	1	0	3
335	0	0	0	2	1	0	3
350	4	4	4	13	4	4	33
365	1	0	0	0	0	0	1
375	6	4	5	7	6	2	30
435	0	1	0	0	0	0	1

total	33	25	36	54	35	16	199
-------	----	----	----	----	----	----	-----

2B) Filter sessions by specified criteria in different aspects such as brain areas, imaging depth and stimuli type

Now we proceed to Stage B: filtering sessions.

Build a new "clean" brain_observatory_cache object first and I will explain right after.

```
boc = brain_observatory_cache(manifests)
```

```
boc =  
  brain_observatory_cache with properties:  
      session_table: [597×14 table]  
      container_table: [199×13 table]  
      manifests: [1×1 struct]  
      filtered_session_table: [597×14 table]  
      stimuli: {9×1 cell}  
      targeted_structure: {6×1 cell}  
      imaging_depth: [11×1 double]  
      container_id: [199×1 double]  
      session_id: [597×1 double]  
      session_type: {4×1 cell}
```

To review, manifests.mat is taken in by brain_observatory_cache and stored in property "manifests", the manifest of all sessions (m sessions by n aspects of meta data) from manifest.mat is stored in property "session_table", and the manifest of all containers (m containers by n aspects of meta data) from manifest.mat is stored in property "container_table".

We implemented several methods with generic name pattern "filter_...by_..." to filter sessions by criteria in different aspects. As methods get called, property filtered_session_table, which started with the same content as session_table, gets eliminated to meet new criteria, and then other properties that each summarize the meta data of all filtered sessions stored in "filtered_session_table" from a different aspect ("meta meta data") get updated. When a brain_observatory_cache object is constructed, the "meta meta data" is simply the "meta meta data" of all sessions.

Here we show an example of searching for sessions that primary visual cortex was recorded at 275 mm deep and during which drifting gratings were shown.

Eliminate all sessions to sessions that only showed drifting gratings

```
boc.filter_sessions_by_stimuli('drifting_gratings')
```

```
ans =  
  brain_observatory_cache with properties:  
      session_table: [597×14 table]
```

```

        container_table: [199×13 table]
        manifests: [1×1 struct]
    filtered_session_table: [199×14 table]
        stimuli: {4×1 cell}
    targeted_structure: {6×1 cell}
    imaging_depth: [11×1 double]
    container_id: [199×1 double]
    session_id: [199×1 double]
    session_type: {'three_session_A'}

```

Note the changes of all properties except the first top three.

Eliminate current filtered sessions to sessions that have posterior Primary Visual Cortex recorded

```
boc.filter_sessions_by_targeted_structure('VISp')
```

```

ans =
    brain_observatory_cache with properties:
        session_table: [597×14 table]
        container_table: [199×13 table]
        manifests: [1×1 struct]
    filtered_session_table: [54×14 table]
        stimuli: {4×1 cell}
    targeted_structure: {'VISp'}
    imaging_depth: [6×1 double]
    container_id: [54×1 double]
    session_id: [54×1 double]
    session_type: {'three_session_A'}

```

Eliminate current filtered sessions to sessions that were recorded at 275 mm deep

```
boc.filter_sessions_by_imaging_depth(275)
```

```

ans =
    brain_observatory_cache with properties:
        session_table: [597×14 table]
        container_table: [199×13 table]
        manifests: [1×1 struct]
    filtered_session_table: [20×14 table]
        stimuli: {4×1 cell}
    targeted_structure: {'VISp'}
    imaging_depth: 275
    container_id: [20×1 double]
    session_id: [20×1 double]
    session_type: {'three_session_A'}

```

As we can see now, there are 20 sessions that met all of the criteria given.

Let's just get one random session out of the 20 sessions, so we don't wait forever when I show you how to download real data at the next stage

```
boc.filter_sessions_by_container_id(527550471)
```

```
ans =
  brain_observatory_cache with properties:
    session_table: [597x14 table]
    container_table: [199x13 table]
    manifests: [1x1 struct]
    filtered_session_table: [1x14 table]
    stimuli: {4x1 cell}
    targeted_structure: 'VISp'
    imaging_depth: 275
    container_id: 527550471
    session_id: 527745328
    session_type: {'three_session_A'}
```

Take a look at metadata of the final filtered session to convince you how friendly my property list ("meta meta data") is

```
boc.filtered_session_table
```

```
ans = 1x14 table
    date_of_acquisition    experiment_container_id    fail_eye_tracking    id    imaging_depth
    -----
    '2016-07-08T15:59:05Z'    5.2755e+08    true    5.2775e+08    275
```

Hint: In future days, when you get lots NWB files downloaded and you lose track of who the heck they are, you can use brain_observatory_cache to look up their meta data and "meta meta data".

2C) Download NWB files of filtered session(s)

Finally comes to Stage C: downading real data stored in NWB files

For more info about NWB,

See its github page: <http://neurodatawithoutborders.github.io/>

See its publication: <http://www.sciencedirect.com/science/article/pii/S0896627315009198>

First, set up the path to NWB directory.

```
nwb_dir_name = [bot_dir_name, 'nwb_files/'];
```

Then, call the method named "download_nwb" to dowload all NWB file(s) corresponding to filtered session(s) stored in brain_observatory_cache and save it under your NWB directory.


```
boc.download_nwb(nwb_dir_name);
```

desired nwb file already exists

Note: The size of a nwb file is at the scale of 100 MB, so you definitely want to move around instead of sitting here and waiting. Once brain_observatory_cache gets the file downloaded, it will spit you a message, so no worries.

3) Import imaging data from nwb files

Now we have acquired an NWB file, we can extract the fluorescence traces from the NWB file and convert them to data in "raster format" (n trials by t times for every cell), which is easier to analyze.

For more info about raster format, see <http://www.readout.info/toolbox-design/data-formats/raster-format/>

To start, add the NWB file to MATLAB searching path.

```
% add path to nwb files  
addpath([bot_dir_name, 'nwb_files/'])
```

3a) Get and plot fluorescence traces

Before we start converting data format, you might just want to plot some fluorescence traces to make sure they look normal or for fun.

To do this you can either

- i) get fluorescence traces of all cells in one session and plot them in your own way or
- ii) use the built-in plot function to plot fluorescence traces of one cell in one session

Here we show an example of i):

Extract and compute all four types fluorescence traces available in the NWB file of session 517745328 and save them to the workspace.

```
session_id = 527745328;  
[raw,demixed,neuropil_corrected,Df0verF] = save_fluorescence_traces(session_id);
```

Elapsed time is 4.392045 seconds.

Note: each type of trace was stored as h5read table (k dimensions of cells by t dimenions of times) and saved as n dimenions of sampling points by k dimensions of cells matlab matrix

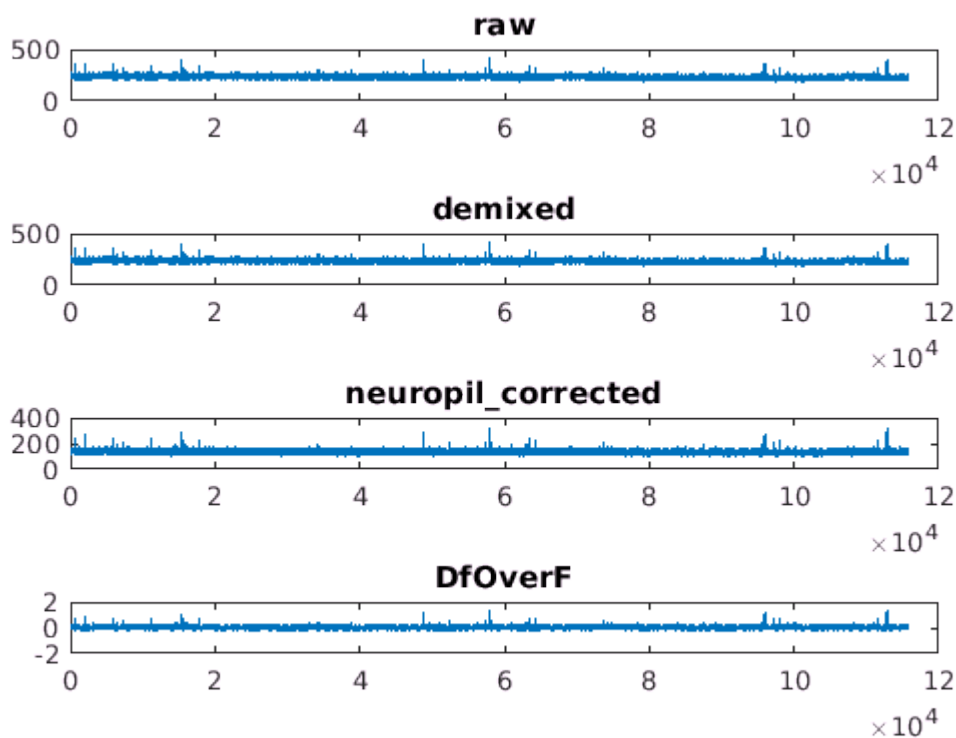
Alternatively, this is an example of ii):

Plot fluorescence traces of a random cell 529022196 in session 527745328 directly

Caution: "plot_fluorecence_traces" takes one new cell id

```
session_id = 527745328;  
cell_id = 529022196;  
plot_fluorecence_traces(session_id, cell_id);
```

Warning: MATLAB has disabled some advanced graphics rendering features by switching to software OpenGL. For more information, [click here](#).



3b) Convert fluorescence trace into data in "raster format"

To start, set up the path to raster directory

```
raster_dir_name = [bot_dir_name, 'raster/'];
```

Tell the "convert" function the session you are working on, the type of stimuli you are interested in, and the type of fluorescence trace you want to use, and then set it off.

```
session_id = 527745328;  
stimuli = 'drifting_gratings';
```

```
fluorescence_trace = 'Df0verF';  
  
current_raster_dir_name = convert_fluorescence_trace_into_raster_format(fluorescence_trace, ..  
    session_id, stimuli, raster_dir_name);
```

Brain-Observatory-Toolbox/raster/drifting_gratings_527745328/ already exists

Note: This doesn't take long.

4) Get Masks...

