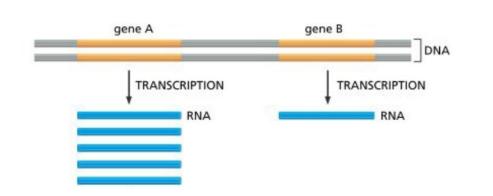
Transcriptomics in Crop Reseach

Introduction to RNAseq technology

Mary-Ann Blätke JJ Szymanski

Mon 6th	Tue 7th	Wed 8th	Fri 17th
Gene to transcript	Quantification	Quality check	Catching up
Sequencing technologies	Getting data	Expression units	Q&A
Intro to bash & setup	Mapping	Normalization	

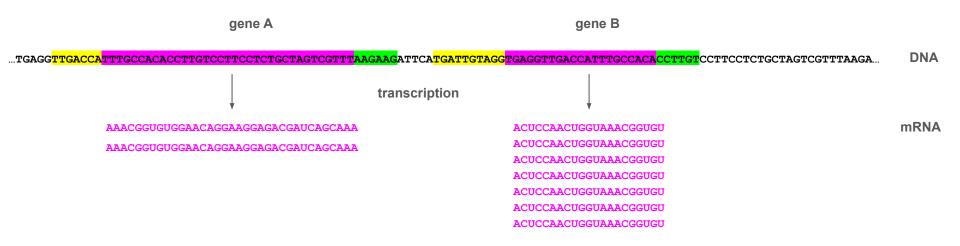


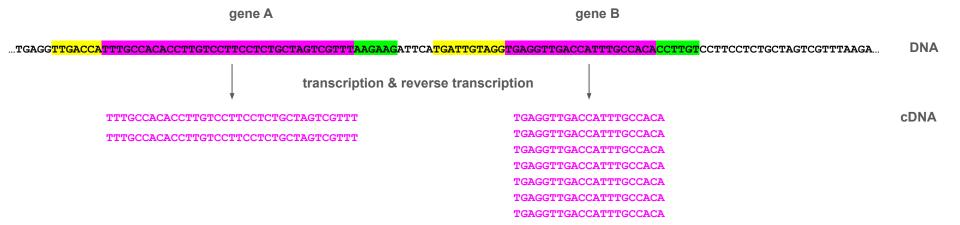
DNA \dots TGAGGTTGACCATTTGCCACACCTTGTCCTCTCCTCGCTAGTCGTTTAAGAAGATTCATGATTGTAGGTGAGGTTGACCATTTGCCACACCTTGTCCTCTCCTCGCTAGTCGTTTAAGA \dots

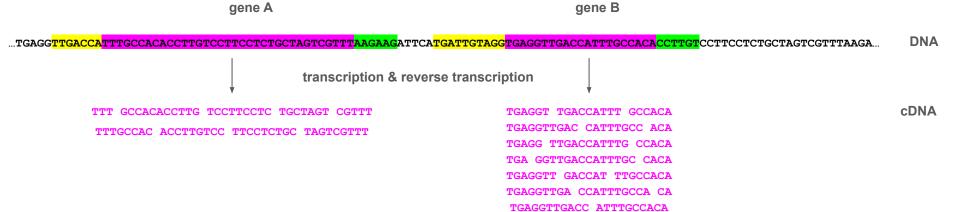
gene A gene B

...TGAGG<mark>TTGACCATTTGCCACACCTTGTCCTTCCTCTGCTAGTCGTTT</mark>AAGAAGATTCA<mark>TGATTGTAGGTGAGGTTGACCATTTGCCACACCTTGT</mark>CCTTCCTCTGCTAGTCGTTTAAGA... DNA









gene A gene B

...TGAGG<mark>TTGACCATTTGCCACACCTTGTCCTTCCTCTGCTAGTCGTTTAAGAAG</mark>ATTCA<mark>TGATTGTAGGTGAGGTTGACCATTTGCCACA<mark>CCTTGT</mark>CCTTCCTCTGCTAGTCGTTTAAGA...</mark>

cDNA

DNA

A bag of sequenced reads

TGAGGT TGACCATTT GCCACA

TGAGGTTGACC ATTTGCCACA

TGAGG TTGACCATTTG CCACA

TGAGGTTGACC ATTTGCCACA

TTTGCCAC ACCTTGTCC TTCCTCTGC TAGTCGTTT

TGAGGTT GACCAT TTGCCACA

TTT GCCACACCTTG TCCTTCCTC TGCTAGT CGTTT

gene A gene B

...TGAGG<mark>TTGACCA<mark>TTTGCCACACCTTGTCCTTCCTCTGCTAGTCGTTT</mark>AAGAAG</mark>ATTCA<mark>TGATTGTAGG</mark>TG<mark>AGGTTGACCATTTGCCACA<mark>CCTTGT</mark>CCTTCCTCTGCTAGTCGTTTAAGA...</mark>

cDNA

DNA

A library!

TGAGGT TGACCATTT GCCACA

TGAGGTTGAC CATTTGCCA CA

TGAGG TTGACCATTTG CCACA TGAGGTTGAC CATTTGCC ACA

TTTGCCAC ACCTTGTCC TTCCTCTGC TAGTCGTTT

TGAGGTT GACCAT TTGCCACA TGA GGTTGACCATTTGC CACA

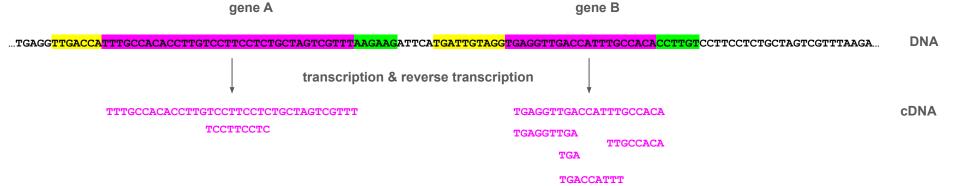
TTT GCCACACCTTG TCCTTCCTC TGCTAGT CGTTT

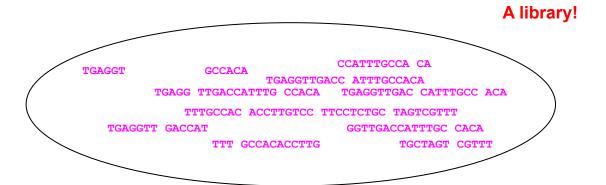
gene B DNA ...TGAGG<mark>TTGACCATTTGCCACACCTTGTCCTTCCTCTGCTAGTCGTTTAAGAAG</mark>ATTCA<mark>TGATTGTAGGTGAGGTTGACCATTTGCCACA<mark>CCTTGT</mark>CCTTCCTCTCGCTAGTCGTTTAAGA...</mark> transcription & reverse transcription **cDNA** TGAGGTTGACCATTTGCCACA TTTGCCACACCTTGTCCTTCCTCTGCTAGTCGTTT

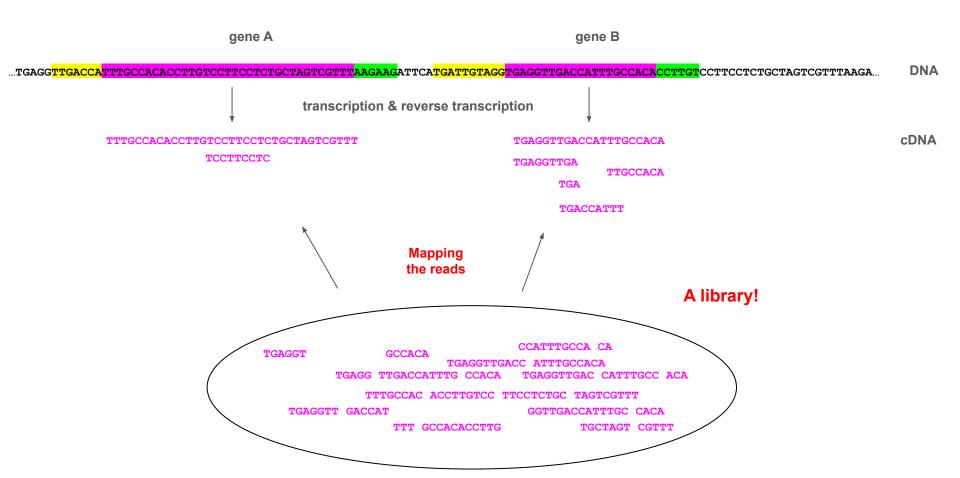
gene A

A library!

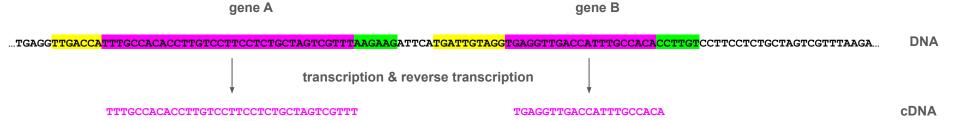
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What data do we need to start an RNAseq project?



A library

TGAGGT TGACCATTT GCCACA

TGAGGTTGAC CATTTGCCA CA

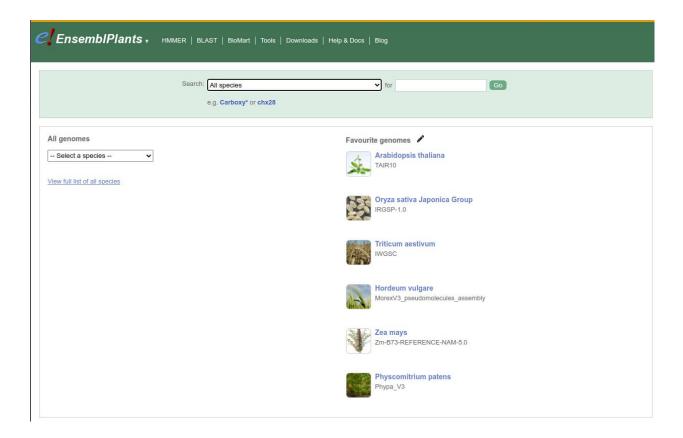
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TTTGCCAC ACCTTGTCC TTCCTCTGC TAGTCGTTT

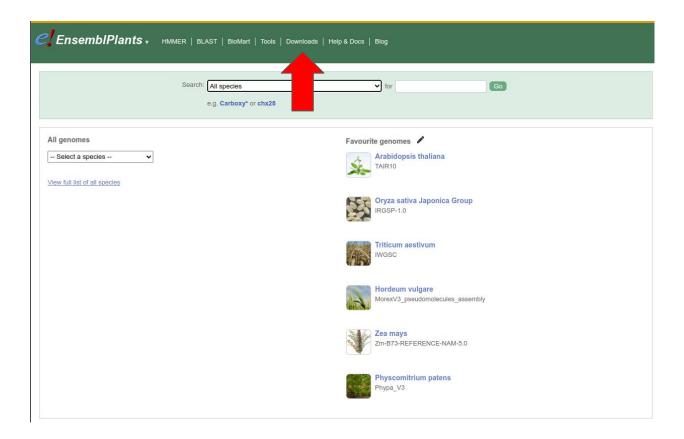
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TTT GCCACACCTTG TCCTTCCTC TGCTAGT CGTTT

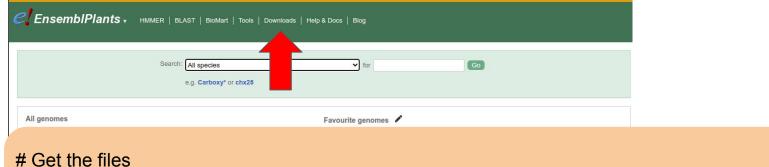
Get the reference genome



Get the reference genome



Get the reference genome



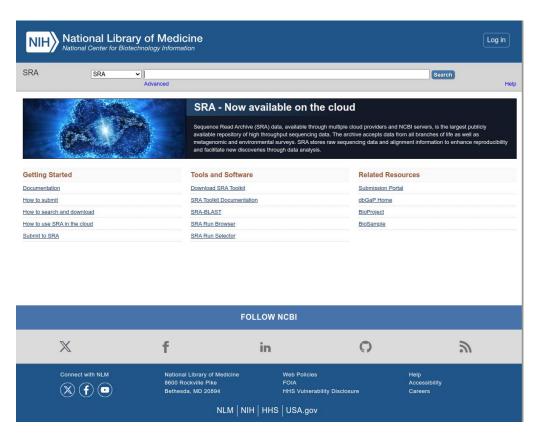
wget https://ftp.ensemblgenomes.ebi.ac.uk/pub/plants/release-57/fasta/solanum_lycopersicum/cdna/ # unzip

gunzip Solanum_lycopersicum.SL3.0.cdna.all.fa.gz



```
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       - get_and_index_genome.sh
       - quality_check.sh
      - map_reads.sh
    genome
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       - cDNA_Solanum_lycopersicum.index
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       - SRR15245013_1.fastq.gz
       - SRR15245013_2.fastq.gz
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      – etc....
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           - abundance.tsv
           run_info.json
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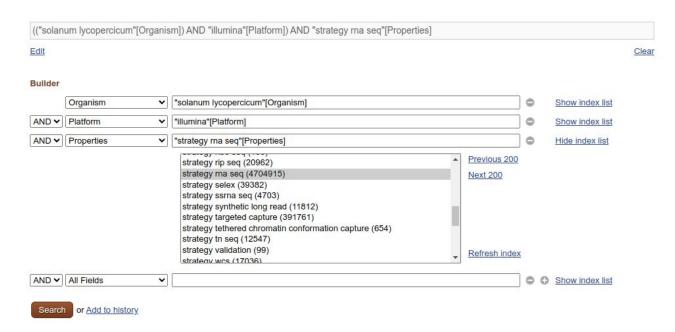
From SRA search



https://www.ncbi.nlm.nih.gov/sra

From SRA search

SRA Advanced Search Builder



From SRA search

AND ▼ | All Fields

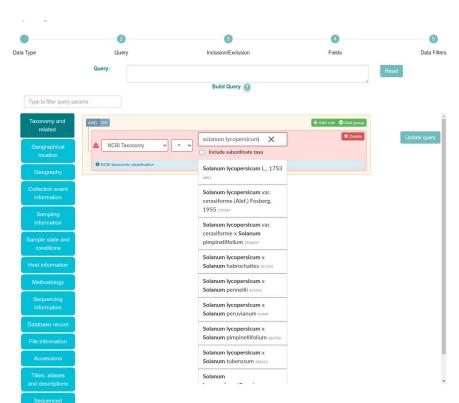
Search or Add to history

SRA Advanced Search Builder (("solanum lycopercicum"[Organism]) AND "illumina"[Platform]) AND "strategy rna seq"[Properties] Edit Clear Builder # Get the files wget https://sra-downloadb.be-md.ncbi.nlm.nih.gov/sos3/sra-pub-zq-22/SRR010/056/SRR10056916.sralite.1 # Convert SRA format to fastq.gz fastq-dump --split-files --gzip -A SRR10056916 SRR10056916.sra strategy validation (99) Refresh index strategy wcs (17036)

Show index list

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Source data of a manuscript





AoB PLANTS 2	20, Vol. 12, No. 5
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doi:10.1093/aobpla/plaa041 Advance Access Publication August 19, 2020 Studies

STUDIES

Transcriptome analysis in osmo-primed tomato seeds with enhanced longevity by heat shock treatment

Thiago Barbosa Batista¹, Geysson Javier Fernandez², Tiago Alexandre da Silva¹, Júlio Maia¹ and Edvaldo Aparecido Amaral da Silva¹

³Department of Plant Production, Sao Paulo State University (UNESP), Botucatu, Sao Paulo, Brazil, ²Institute of Biology, Antioquia University, Medellín, Antioquia, Colombia

*Corresponding author's e-mail address: amaral.silva@unesp.br

Associate Editor: Gabriela Auge

Form & Function. Chief Editor: Kate McCulloh

Abstract

Seed priming is widely used in commercial seeds and its main function is to accelerate and synchronize seed germination. Undesirably, primed seeds show reduced longevity and treatments like heat shock have been shown to improve longevity in primed seeds. Nonetheless, the effect of heat shock treatment on primed seeds at the mRNA level is not known. Thus, the aim of this work was to investigate the effect of heat shock treatment on the longevity of primed tomato (Solanum lycopersicum) seeds at the physiological and transcriptome levels. Tomato seeds were primed and dried (control). Alternatively, primed seeds were subjected to heat shock treatment (38 °C/32 % relative humidity) before drying. Germination, vigor and longevity were evaluated. Transcriptome analysis was performed by RNA sequencing (RNA-seq) from biological samples collected immediately after priming and another samples collected from primed seeds followed by the heat shock treatments. The gene expression was validated by quantitative real time PCR (RT-qPCR). We showed that applying heat shock treatment after priming increased germination speed, enhanced seed longevity and preserved the vigor during storage of primed tomato seeds. Through transcriptome analysis, 368 differentially expressed genes were identified, from which 298 genes were up-regulated and 70 were down-regulated. We showed the increase of mRNA levels of HEAT SHOCK FACTOR-like and HEAT SHOCK PROTEIN-like chaperone genes, suggesting the involvement of the proteins coded by these transcripts in the enhancement of longevity in primed tomato seeds. The heat shock treatment after priming enhances and preserves the vigor of tomato primed seeds during storage. In addition, improves seed longevity through the increase in the expression of transcripts related to protection by response to stress.

Keywords: Chaperone molecules; improved longevity; primed seed; seed conservation; seed quality; Solanum lycopersicum L., storage.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7494243/

Source data of a manuscript





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Keywords: Chaperone molecules; improved longevity; primed seed; seed conservation; seed quality; Solanum lycopersicum

Physiological assays

Seed germination and vigor. Four replications of 50 seeds were germinated in 9 cm Petri dishes with substrate of paper towel moistened with distilled water equivalent to 2.5 times its weight, at 25 °C, under 8 h of light and 16 h in the dark. The length of the primary root, 22mm was used as the germination criterion. Data collection was done in different times after sowing; and ended when the germination rate reached 100 % or at 14 days. Seed vigor was determined by the calculation of the time to 50 % of germination (t50) through the analysis of cumulative germination data using the curve fitting module of the Germinator software package (Joosen et al. 2010).

Longevity. We used ageing protocol to assess seed in which the seeds were placed in a support over a saturated solution of NaCl (75 % RH) at 35 °C in glass bottles hermetically sealed. During storage, the water content of S. lycopersicum seeds stabilized at 0.10 ± 0.007 g H₂O/g DW⁻¹, corresponding to ±9.5 % on wet basis. At different time spans, seeds were imbibed and viability was assessed using the germination assay as described earlier. The different time spans were carried out considering the viability loss behaviour of each treatment group during storage. The viability data were transformed into probit to

libraries were 100 base pair (bp) paired-end sequenced. The data output in fastq file format contained sequence information, including the sequencing quality (Phred quality score). Average Phred scores of ≥20 per position were used for the alignment.

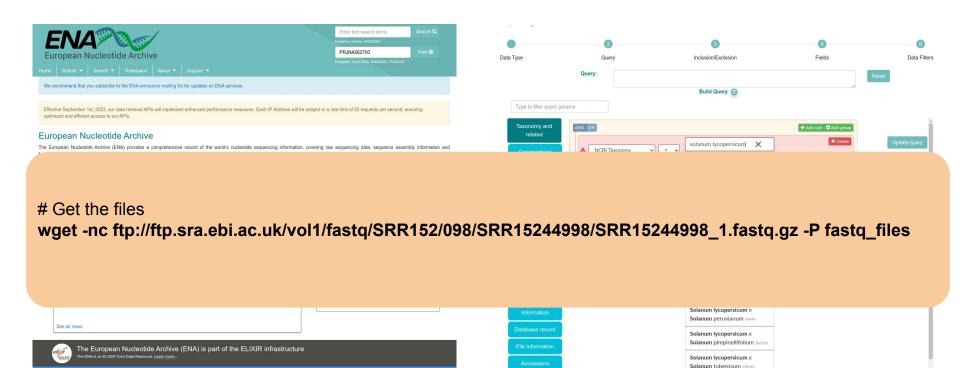
Read alignment and differentially expressed genes. Paired-end reads for mRNA were mapped to the Solanum lycopersicum release 39 reference genome using the default parameters of TopHat2 (Kim et al. 2013). Counts for RefSeq genes were obtained using HTSeq (Anders et al. 2015) and DESeq2 (Love et al. 2014) was used to normalize expression counts. The changes in gene expression were considered statistically significant when fold change ≥2 and P-values ≤ 0.05. The RNAseq data was deposited in NCBI (BioProject PRJNA562700: https://trace.ncbi.nlm.nih.gov/Traces/sra/?study=SRP220280).

The analysis of principal components was made using all the genes expressed on the RNA seq data. The normalized count per gene was used and transformed to Z-score. This matrix was used was used to perform the PCA. For plotting the PCA results, we used the principal component one and two. The heatmap was generated using the normalized counts of the differentially expressed genes. Then we transformed it to z-score and plotted it using the package pheatmaps of R.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7494243/

From ENA search

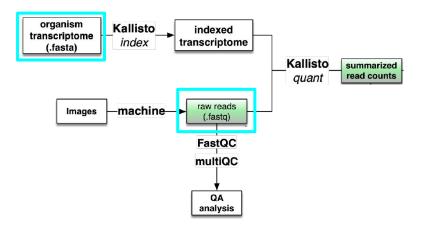
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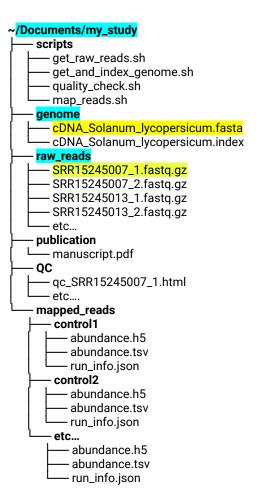


Solanum

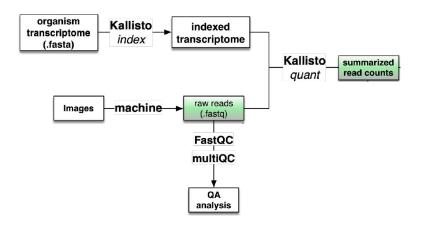
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       SRR15245013_1.fastq.gz
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          abundance.tsv
          - run_info.json
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RNAseq analysis pipeline

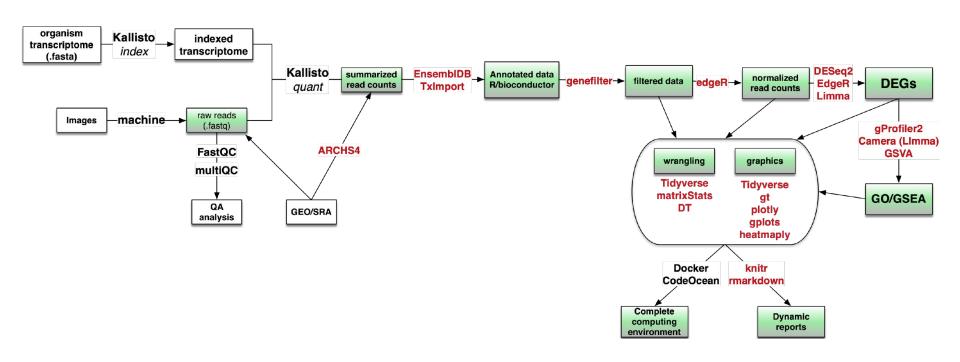




The RNAseq analysis pipeline



The RNAseq analysis pipeline



The End