Endovir

A pipeline to identify endogenous viruses in SRA datasets

https://github.com/NCBI-Hackathons/EndoVir

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Foreword

Based on previous work

- ViruSpy ¹
- Logic mostly implemented in Bash and Perl
 - → Complex to expand and adjust
- Initiated during my stay as a visiting bioinformartician as NCBI from Oct.2017 to Dec. 2017.
- Ongoing

 $^{^1}$ https://github.com/NCBI-Hackathons/ViruSpy

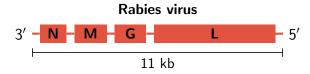
Foreword

Standing on the shoulder of giants

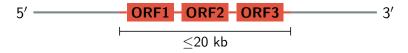
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Exogenous viruses

- ▶ Integration into host genome not required for replication
- ► Hist genome integration occurs occasionally, e.g. Human Herpes virus 6
- ▶ Not part of the host genome
- ► RNA viruses: ≤≈ 40 kb
- ► DNA viruses: ≤≈ 2 Mbp



Endogenous viral element (EVEs)

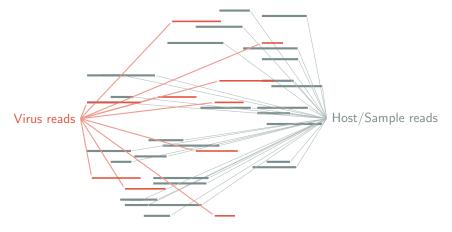


- ▶ Integration into host genome mandatory for replication
- Retroviridae, e.g. HIV
- Part of the host genome
- ▶ Not widely analyzed: complex analysis and missing data ²
- Allowing to date virus evolution
- Characterize the mechanisms that underpin the integration of viruses into host genomes

²Katzourakis, A. and Gifford, R.J. (2010). PLos Genet., 6, e1001191

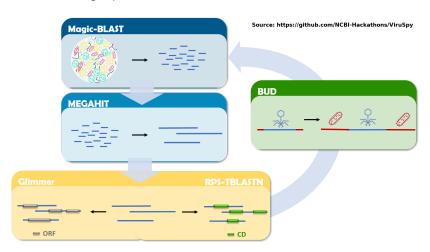
SRA data: a treasure trove for virus genomics

- SRA data contains host and virus sequences
- Exo/Endogenous and putative novel viruses



BUD algorithm

▶ "Building Up Domains"



Endovir

```
Open source MIT licence

Modern Implemented in Python 3 (3.6)

Independent Using only Python standard libraries

Limited disk I/O Using pipes where possible

Modularity Changing external tools, e.g. assembler, without touching Endovir internals

Status Alpha, ongoing development
```

Biological databases

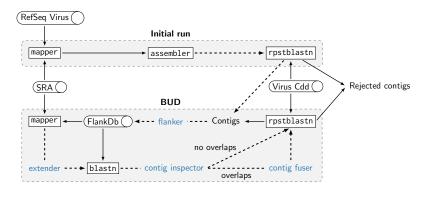
Endovir uses two biological databases:

Sequences Virus genomes sequences from RefSeq

Domains virus-specific Cdd database. We provide a setup script to create the Cdd database

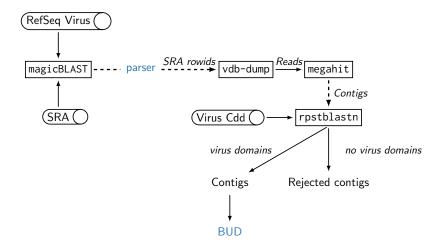
Usage

Workflow

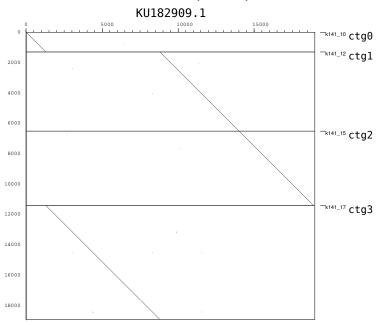




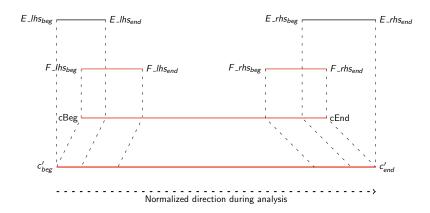
Init: current version



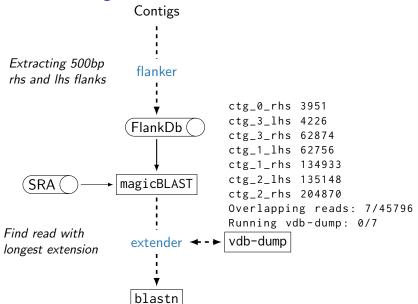
Init: SRR5150787 example (Ebola)



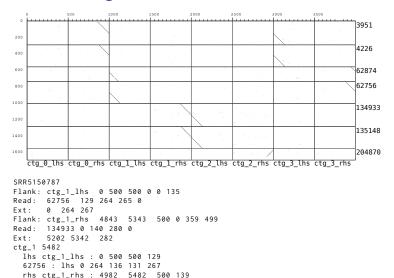
Endovir's contig model



BUD: Extending

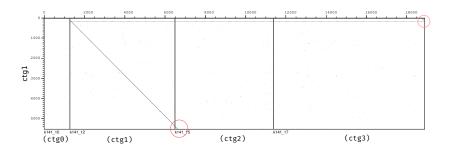


BUD: Extending

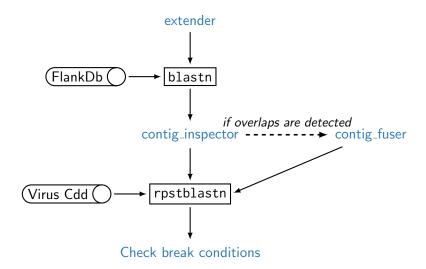


134933 : rhs 5202 5342 141 141 282

BUD: Extending



BUD: Overlap check



BUD: Overlap check

```
Checking flanks for overlaps
Blast results: ctg_0_lhs vs. ctg_0_lhs
Not an overlap
-----
Blast results: ctg_0_lhs vs. ctg_2_rhs
Checking ctg 0 lhs and ctg 2 rhs
ctg_2:rhs + ctg_0:lhs
LHS: 130
RHS 117
oLHS: 129
oRHS 139
Src: ctg 2 7724
RHS_flank: 7224 7724 500
 Extension: 7544 7606 182
Dst: ctg_0 5482 0 126 0
Brk: ctg_2 7597 ctg_0 129
Blast results: ctg_0_rhs vs. ctg_1_lhs
Checking ctg 0 rhs and ctg 1 lhs
 Update: ctg 0 is now ctg 2
ctg_2 ctg_1
ctg 2:rhs + ctg 1:lhs
IHS: 130
RHS 0
ol HS · 134
oRHS 12
Src: ctg_2 7724
RHS_flank: 7224 7724 500
 Extension: 10428 10568 0
Dst: ctg_1 5042 0 134 0
Brk: ctg_2 7589 ctg_1 134
```

BUD: Break conditions

Extending into host genome Empty Cdd database screen
No grow Contig was not extended
No contigs No contigs were identified

Modularity

Facilitating extending and adding other tools using classes which act as messenger between tool and Endovir.

```
class MappingAlignment:
  class Read:
   def init (self, name, start, stop, strand, glen):
  class Flank
   def init (self. name. start. stop. strand):
 def __init__(self, cols):
    self.read = self.Read(cols[0]. cols[6]. cols[7]. cols[13]. cols[15])
    self.flank = self.Flank(cols[1], cols[8], cols[9], cols[14])
    self.pident = float(cols[2])
class MagicblastParser:
  def init (self):
    self.alignments = []
  def parse(self. src):
    self.alignments = []
   for i in src:
      if i[0] != '#':
        self.alignments.append(MagicblastAlignment(i.strip().split('\t')))
```

Outlook

- Adjust tool parameters on the fly
- ► Replace SRA toolkit with NCBI's ngs-python
- ▶ Test on more diverse datasets
- Docker

Acknowledgments

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