

Endovir

A pipeline to identify endogenous viruses in SRA datasets

<https://github.com/NCBI-Hackathons/EndoVir>

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Foreword

Based on previous work

- ▶ ViruSpy ¹
- ▶ Logic mostly implemented in Bash and Perl
→ Complex to expand and adjust
- ▶ Initiated during my stay as a visiting bioinformatician at NCBI from Oct.2017 to Dec. 2017.
- ▶ Ongoing

¹<https://github.com/NCBI-Hackathons/ViruSpy>

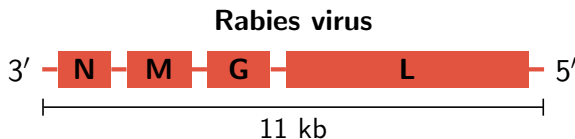
Foreword

Standing on the shoulder of giants

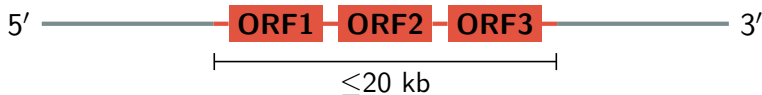
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Exogenous viruses

- ▶ Integration into host genome not required for replication
- ▶ Hist genome integration occurs occasionally, e.g. Human Herpes virus 6
- ▶ Not part of the host genome
- ▶ RNA viruses: $\leq \approx 40$ kb
- ▶ DNA viruses: $\leq \approx 2$ Mbp



Endogenous viral element (EVEs)

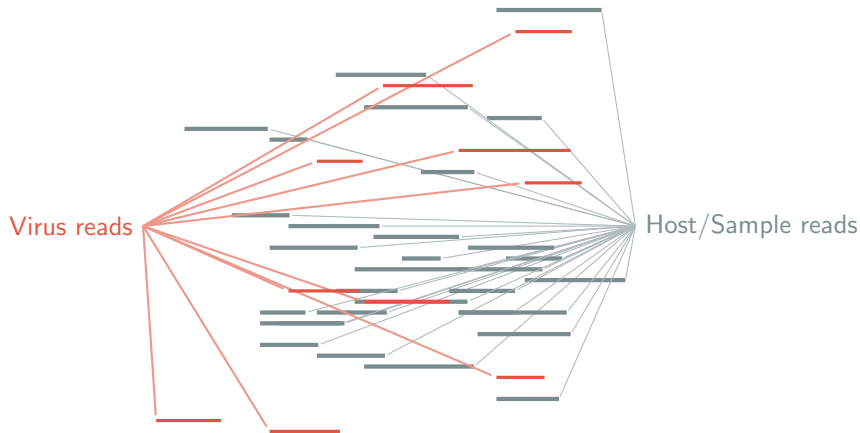


- ▶ Integration into host genome mandatory for replication
- ▶ Retroviridae, e.g. HIV
- ▶ Part of the host genome
- ▶ Not widely analyzed: complex analysis and missing data ²
- ▶ Allowing to date virus evolution
- ▶ Characterize the mechanisms that underpin the integration of viruses into host genomes

²Katzourakis, A. and Gifford, R.J. (2010). PLoS Genet., 6, e1001191

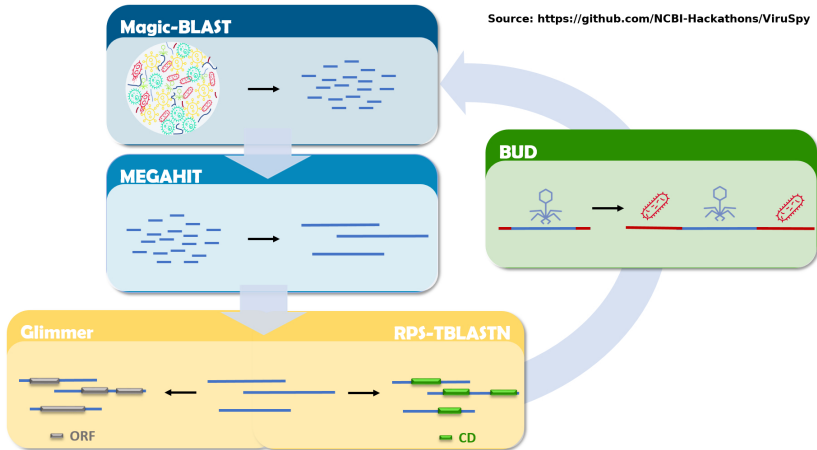
SRA data: a treasure trove for virus genomics

- ▶ SRA data contains host and virus sequences
- ▶ Exo/Endogenous and putative novel viruses



BUD algorithm

► "Building Up Domains"



Endovir

Open source MIT license

Modern Implemented in Python 3 (3.6)

Independent Using only Python standard libraries

Limited disk I/O Using pipes where possible

Modularity Changing external tools, e.g. assembler, without touching Endovir internals

Status Alpha, ongoing development

Biological databases

Endovir uses two biological databases:

Sequences Virus genomes sequences from RefSeq

Domains virus-specific Cdd database. We provide a setup script to create the Cdd database

Usage

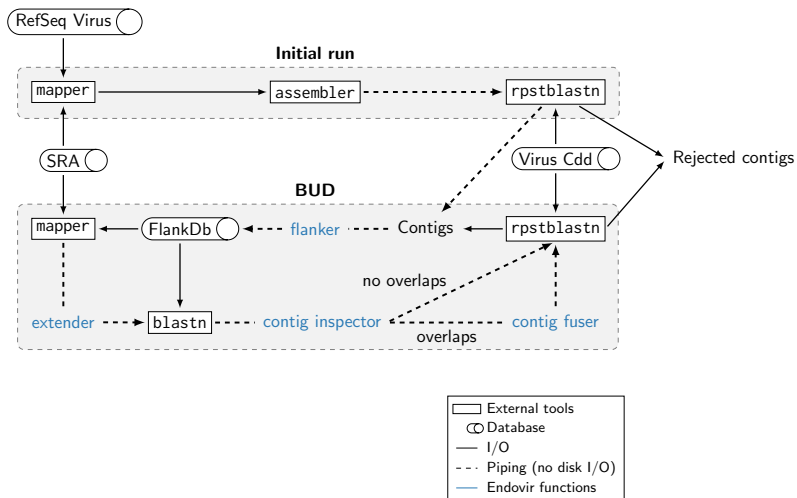
```
usage: endovir.py [-h] [-srr SRR] [--wd WD] [--max_cpu MAX_CPU]
```

Endovir

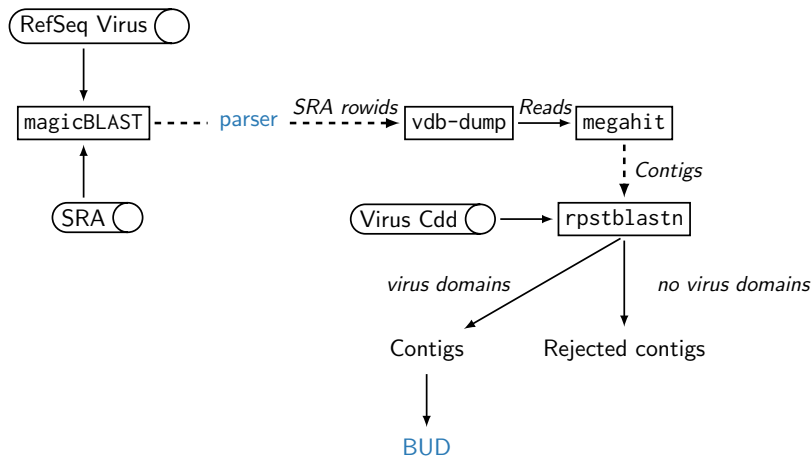
optional arguments:

-h, --help	show this help message and exit
-srr SRR	SRR number, e.g. SRR5150787
--wd WD	Working directory for analysis
--max_cpu MAX_CPU, -p MAX_CPU	Max number of cores to use. NOT YET IMPLEMENTED

Workflow

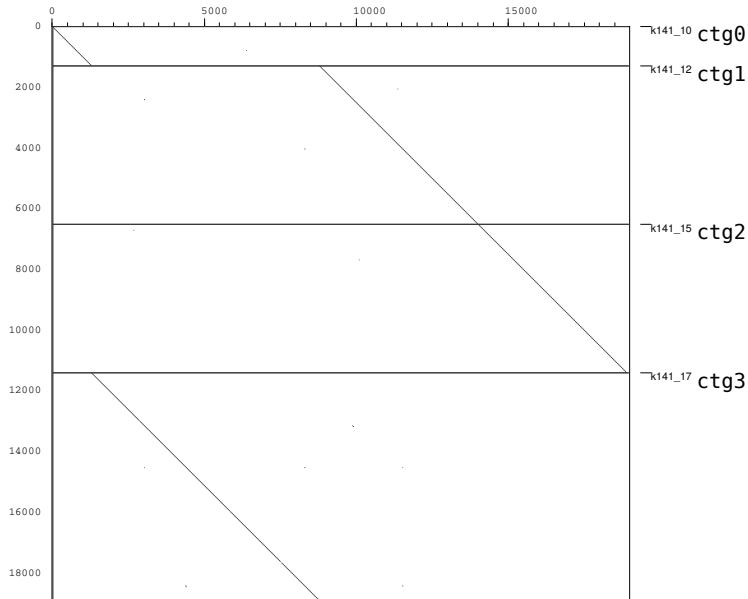


Init: current version

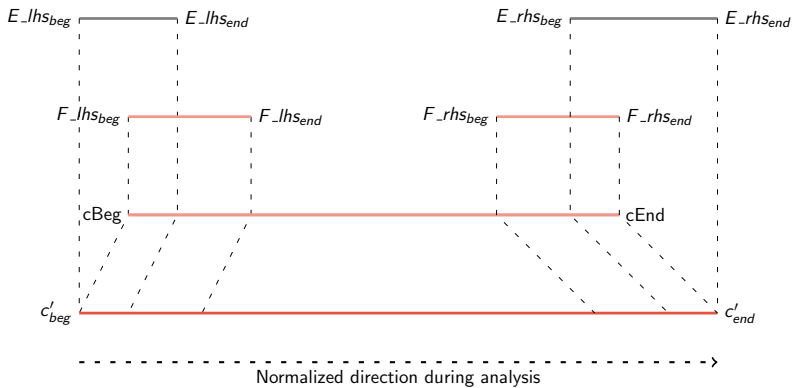


Init: SRR5150787 example (Ebola)

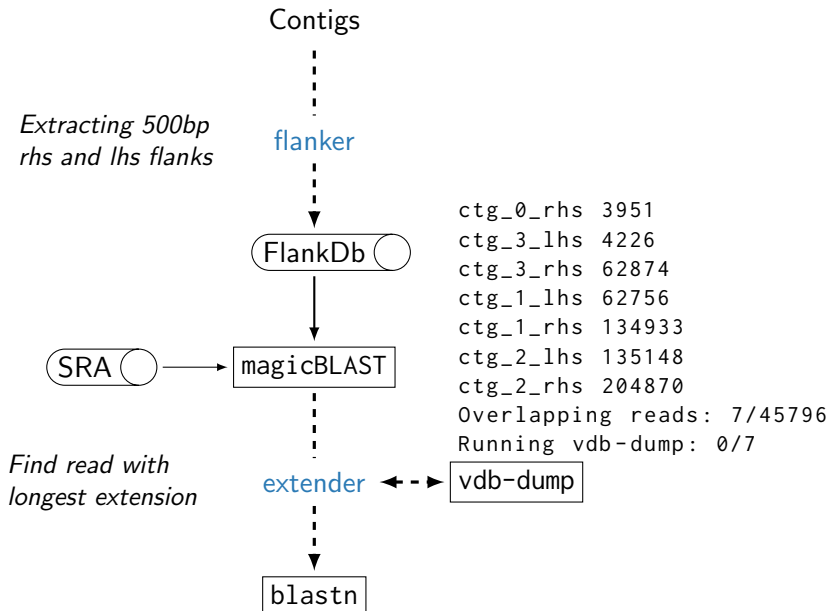
KU182909.1



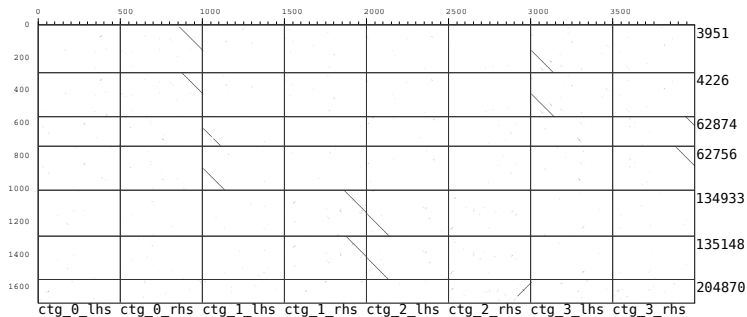
Endovir's contig model



BUD: Extending



BUD: Extending



SRR5150787

Flank: ctg_1_lhs 0 500 500 0 0 135

Read: 62756 129 264 265 0

Ext: 0 264 267

Flank: ctg_1_rhs 4843 5343 500 0 359 499

Read: 134933 0 140 280 0

Ext: 5202 5342 282

ctg_1 5482

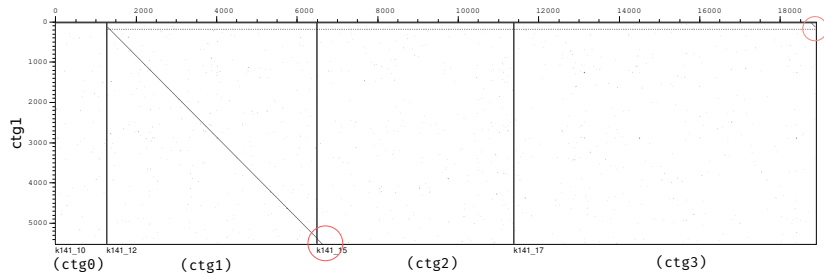
lhs ctg_1_lhs : 0 500 500 129

62756 : lhs 0 264 136 131 267

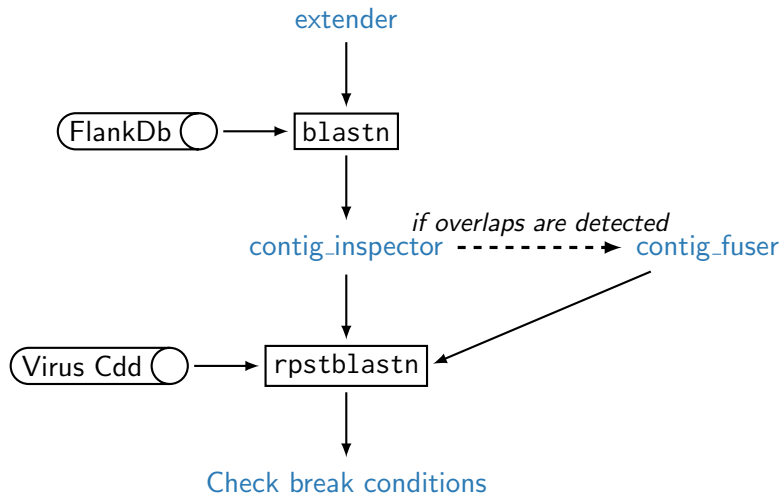
rhs ctg_1_rhs : 4982 5482 500 139

134933 : rhs 5202 5342 141 141 282

BUD: Extending



BUD: Overlap check



BUD: Overlap check

```
Checking flanks for overlaps
Blast results: ctg_0_lhs vs. ctg_0_lhs
Not an overlap
-----
Blast results: ctg_0_lhs vs. ctg_2_rhs
Checking ctg_0_lhs and ctg_2_rhs
ctg_2:rhs + ctg_0:lhs
LHS: 130
RHS 117
oLHS: 129
oRHS 139
Src: ctg_2 7724
    RHS_flank: 7224 7724 500
      Extension: 7544 7606 182
Dst: ctg_0 5482 0 126 0
Brk: ctg_2 7597 ctg_0 129
-----
Blast results: ctg_0_rhs vs. ctg_1_lhs
Checking ctg_0_rhs and ctg_1_lhs
Update: ctg_0 is now ctg_2
ctg_2 ctg_1
ctg_2:rhs + ctg_1:lhs
LHS: 130
RHS 0
oLHS: 134
oRHS 12
Src: ctg_2 7724
    RHS_flank: 7224 7724 500
      Extension: 10428 10568 0
Dst: ctg_1 5042 0 134 0
Brk: ctg_2 7589 ctg_1 134
```

BUD: Break conditions

Extending into host genome Empty Cdd database screen

No grow Contig was not extended

No contigs No contigs were identified

Modularity

- Facilitating extending and adding other tools using classes which act as messenger between tool and Endovir.

```
class MappingAlignment:

    class Read:
        def __init__(self, name, start, stop, strand, qlen):

    class Flank:
        def __init__(self, name, start, stop, strand):

    def __init__(self, cols):
        self.read = self.Read(cols[0], cols[6], cols[7], cols[13], cols[15])
        self.flank = self.Flank(cols[1], cols[8], cols[9], cols[14])
        self.pident = float(cols[2])

class MagicblastParser:

    def __init__(self):
        self.alignments = []

    def parse(self, src):
        self.alignments = []
        for i in src:
            if i[0] != '#':
                self.alignments.append(MagicblastAlignment(i.strip().split('\t')))
```

Outlook

- ▶ Adjust tool parameters on the fly
- ▶ Replace SRA toolkit with NCBI's ngs-python
- ▶ Test on more diverse datasets
- ▶ Docker

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