## NMRlipids IV: Other than PC headgroups

O. H. Samuli Ollila<sup>1,\*</sup>

<sup>1</sup>University of Helsinki
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Primarily measured but also simulated NMR order parameters will be collected also for other than phophatidylcholine (these are discussed in NMRlipids I) headgroup. The information will be used to understand structural differences between different lipid molecules in bilayers.

#### INTRODUCTION

In NMRlipids I and II project we were looking for a MD model which would correctly reproduce headgroup and glycerol backbone structures and cation binding for PC lipid bilayers [1, 4]. Here we extend the same goal for other than PC lipids. Currently the focus is on PE, PG and PS bilayers and their mixtures with PC. Experimental data with different amounts of added salt is now collected and presented in this manuscript. Simulations for bilayers containing PG and PS with low salt conditions are ran with parameters from CHARMM GUI and compared to experiments. Some order parameters from simulations seems to be off from experimental values. Simulation data from other models would be highly useful to see if some of the existing models would reproduce the experimental order parameters and to analyze different conformations predicted by different models respect to experiments.

# EXPERIMENTAL GLYCEROL BACKBONE AND HEADGROUP ORDER PARAMETERS FOR PE, PG AND PS LIPIDS

Absolute values of experimental order parameters for different lipid headgroups are collected in Fig. 1. Signs are measured only for PC as far as I know, thus only absolute values are used for now.

Based on superficial reading, the conclusions in the literature are roughly 1) glycerol backbone structures are largely similar irrespectively of the headroup [10], 2) glycerol backbone and headgroup strucures are similar in model membranes and in bacteria [10?], 3) headgroup structure is phosphatidylserine more rigid than in other phospholipids, which are quite similar to each others [?]. In contrast to PC lipids, extensive discussion on glycerol backbone and headgroup structural details do not exist (as far as I know).

#### GLYCEROL BACKBONE AND HEADGROUP ORDER PARAMETERS FOR PE, PG AND PS LIPIDS IN SIMULATIONS

Several simulations containing PE, PG and PS lipids have been published [? ], however, glycerol backbone and headgroup order parameters are not compared to the experiments

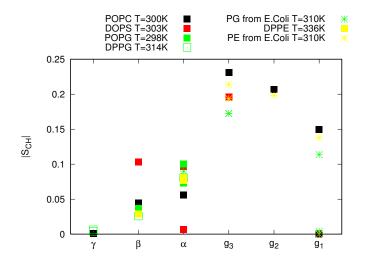


FIG. 1: Absolute values of order parameters for headgroup and glycerol backbone with different headgroups from experiments. POPC values are from [5], DOPS from [6] contains 0.1M of NaCl, POPG from [7] contains 10nM PIPES, DPPG from [8] contains 10mM PIPES and 100mM NaCl, DPPE from [9], E.coliPE and E.coliPG are from [10].

(based on superficial reading of literature).

Glycerol backbone and headgroup order parameters for PS lipids from experiments and simulations are shown in Fig. 2. The preliminary test with CHARMM GUI simulations show suggest that the model overestimates magnitudes in  $\beta$  and  $g_3$  order parameters and forking in  $\alpha$ .

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#### CA2+ BINDING IN BILAYERS WITH NEGATIVELY CHARGED PG AND PS LIPIDS

Ion binding affinity in lipid bilayer containing PC lipids can be measured by using C-H bond order parameters for headgroup  $\alpha$  and  $\beta$  carbons by using the electrometer concept. The decrease of these order parameters is linearly proportional to the amount of bound charge in bilayer [4?]. The electrometer concept can be used also for bilayers containing PC lipids mixed with charged lipids [11?, 12]. This is demonstrated in Figs 4, 5 and 6, where PC headgroup order parameters as a

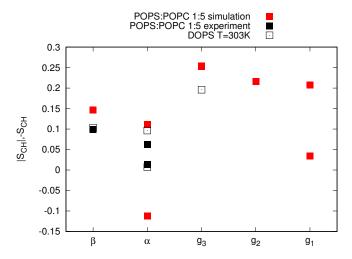


FIG. 2: Order parameters for POPS headgroup and glycerol backbone from simulations and experiments without  $CaCl_2$  (DOPS from [6] contains 0.1M of NaCl, POPC:POPS mixture from [11]). Absolute values are shown for experimental data, because signs are not known. Simulations values are  $-S_{CH}$ 

1.More simulation data for lipids with different headgroups to be collected
2.CHARMM GUI simulation contains only counter ions as potassium. All
experiments here contain some amount of sodium salt. The best ion concentrations
for comparison should be figured out. 3.Experimental signs of the order
parameters would highly useful.

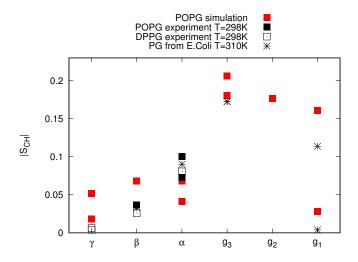


FIG. 3: Order parameters for PG headgroup and glycerol backbone from simulations and experiments without CaCl<sub>2</sub> (POPG from [7] contains 10mM of PIPES, DPPG from [8] contains 10mM PIPES and 100mM CaCl, E.Coli PG results from [10]). Absolute values are shown, because signs are not known for experimental data. Glycerol backbone order parameters are negative in simulations, while alpha,  $\beta$  and smaller  $\gamma$  are positive.

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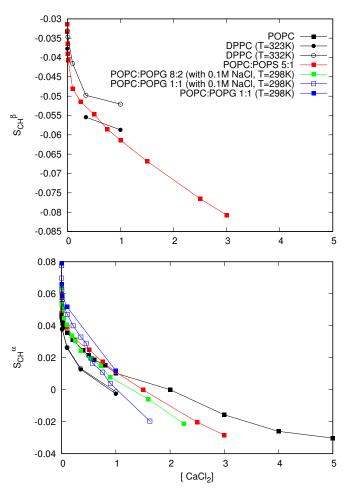


FIG. 4: PC headgroup order parameters as a function of CaCl concentration from experiments containing charged lipids. Pure DPPC data from [13], pure POPC data from [14], POPC:POPS mixture data from [11], POPC:POPG mixture data with 0.1M NaCl from [12] and POPC:POPG mixture data without NaCl from [7].

 ${\bf 7. Check\ the\ NaCl\ concentrations\ in\ the\ samples.}$ 

function of CaCl<sub>2</sub> concentration are shown from experiments with POPC and PS or PG.

Order parameters increase when PS or PG are mixed with PC in the absence of additional ions, as seen from Figs. 4 and 5. In electrometer concept this is explained by the tilting of headgroup more parallel to membrane normal [?]. The order parameter decrease with increasing CaCl<sub>2</sub> concentration is explained by the tilting of headgroup more perpendicular to membrane normal. This decrease is shown to be a good measure for the amount of bound ions in lipid bilayer with PC lipids [4?].

The order parameter decrease as a function of added  $CaCl_2$  for systems with different amount of negative charge are shown in Fig. 6. As expected, the order parameter decrease is more pronounced with increasing amount of negatively charged lipids in bilayer. This is explained by the increase of cation concentration in proximity of bilayer containing nega-

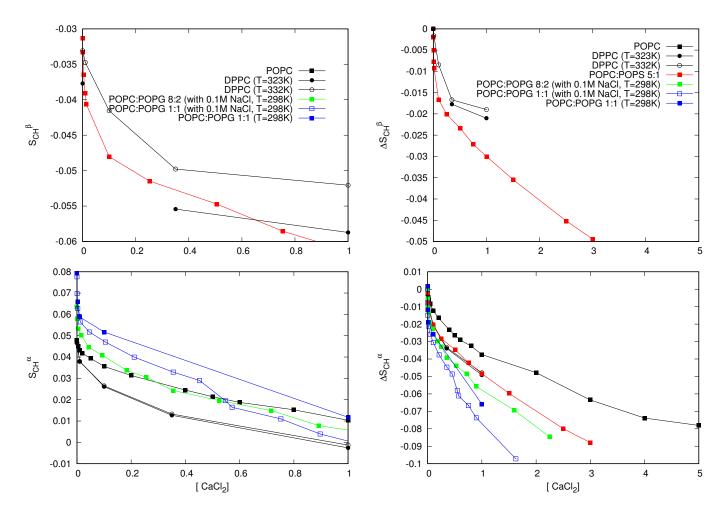


FIG. 5: Figure 4 zoomed to smaller concentrations.

FIG. 6: Changes in order parameters

tively charged lipids [?].

In addition to the changes in PC headgroup order parameters with ion binding, also changes in PS and PG headgroup order parameters are measured.

# CA2+ BINDING IN BILAYERS WITH NEGATIVELY CHARGED PG AND PS LIPIDS IN SIMULATIONS

9. Simulation data for systems with negatively charged lipids and  ${\rm CaCl_2}$  to be collected

### CONCLUSIONS

#### SUPPLEMENTARY INFORMATION

- \* samuli.ollila@helsinki.fi
- [1] A. Botan, F. Favela-Rosales, P. F. J. Fuchs, M. Javanainen, M. Kanduč, W. Kulig, A. Lamberg, C. Loison, A. Lyubartsev, M. S. Miettinen, et al., J. Phys. Chem. B 119, 15075 (2015).
- [2] O. S. Ollila and G. Pabst, Biochimica et Biophysica Acta (BBA)Biomembranes 1858, 2512 (2016).
- [3] T. M. Ferreira, R. Sood, R. Bärenwald, G. Carlström, D. Top-gaard, K. Saalwächter, P. K. J. Kinnunen, and O. H. S. Ollila, Langmuir 32, 6524 (2016).
- [4] A. Catte, M. Girych, M. Javanainen, C. Loison, J. Melcr, M. S. Miettinen, L. Monticelli, J. Maatta, V. S. Oganesyan, O. H. S. Ollila, et al., Phys. Chem. Chem. Phys. 18, 32560 (2016).
- [5] T. M. Ferreira, F. Coreta-Gomes, O. H. S. Ollila, M. J. Moreno, W. L. C. Vaz, and D. Topgaard, Phys. Chem. Chem. Phys. 15, 1976 (2013).
- [6] J. L. Browning and J. Seelig, Biochemistry 19, 1262 (1980).
- [7] F. Borle and J. Seelig, Chemistry and Physics of Lipids 36, 263 (1985).

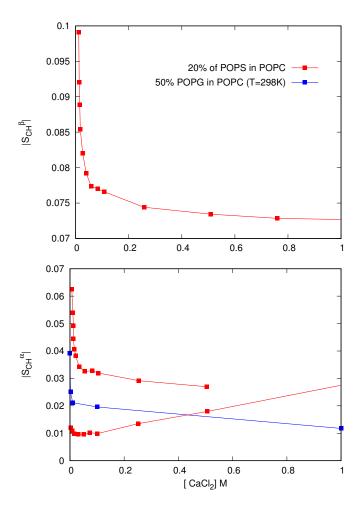


FIG. 7: Changes in order parameters

8.Get the small concentration data from the inserts

[8]	R.	Wo	ohlge	emuth,	N.	Waesp	e-Sarcevio	, and	J.	Seelig,	Bioch	em-
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- [9] J. Seelig and H. U. Gally, Biochemistry 15, 5199 (1976).
- [10] H. U. Gally, G. Pluschke, P. Overath, and J. Seelig, Biochemistry 20, 1826 (1981).
- [11] M. Roux and M. Bloom, Biochemistry 29, 7077 (1990).
- [12] P. M. Macdonald and J. Seelig, Biochemistry 26, 1231 (1987).
- [13] H. Akutsu and J. Seelig, Biochemistry 20, 7366 (1981).
- [14] C. Altenbach and J. Seelig, Biochemistry 23, 3913 (1984).

### ToDo

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7. Check the NaCl concentrations in the samples
9. Simulation data for systems with negatively charged
lipids and $CaCl_2$ to be collected
8. Get the small concentration data from the inserts