

**Supporting Information:**

**NMRlipids IV: Headgroup & glycerol backbone structures, and cation binding in bilayers with PS lipids**

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## **S1 Simulated systems**

### **S1.1 CHARMM36**

To be written by Piggot, Madsen and Ollila

### **S1.2 CHARMM36ua**

To be written by Piggot

### **S1.3 Slipids**

To be written by Piggot and Favela

### **S1.4 Berger**

To be written by Piggot and Ollila Simulations with excess sodium were taken directly from Ref. 1 and simulations with calcium directly from 2. Simulation of POPC at 310 K was taken directly from Ref. 3.

### **S1.5 GROMOS-CKP**

To be written by Piggot

### **S1.6 Lipid17**

To be written by Kav, Miettinen and Meclr.

### **S1.7 MacRog**

To be written by Javanainen and Piggot

## S2 Electrometer concept in PC lipid bilayers mixed with negatively charged lipids

The electrometer concept is based on the empirical observations that the order parameters of  $\alpha$  and  $\beta$  carbons in PC lipid headgroup decrease (increase) proportionally to the bound positive (negative) charge<sup>4-7</sup> (Fig. S1). Therefore, the headgroup order parameters can be used to measure the ion binding affinity to lipid bilayers containing PC lipids.<sup>4-6,8-10</sup> Changes of the headgroup order parameters of other lipids with bound charge, like negatively charged PS and PG lipids, are also systematic, but less well characterized.<sup>8-11</sup> Therefore, the ion binding affinity to negatively charged bilayers can be better characterized measuring the PC headgroup order parameters from mixed bilayers.<sup>8-12</sup>

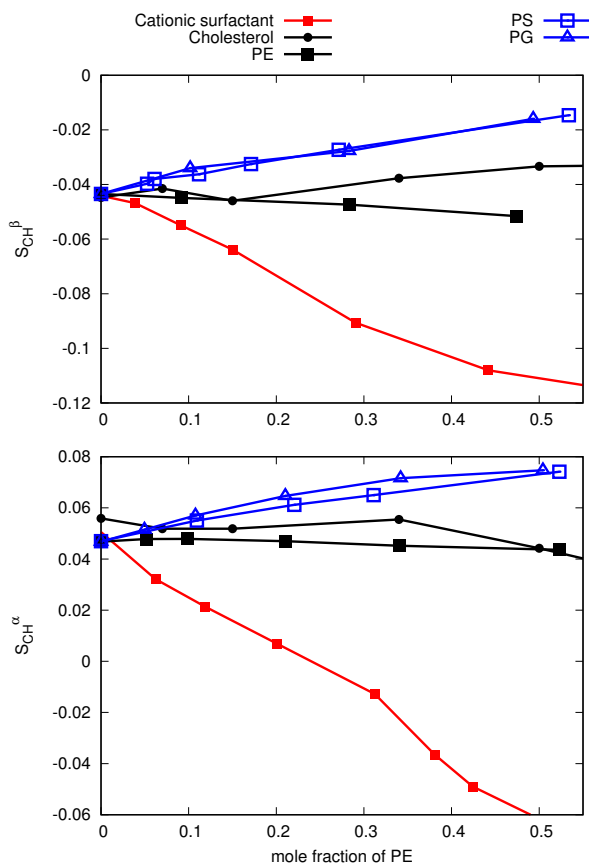


Figure S1: Headgroup order parameters of POPC measured from mixtures with PS (bovine brain), POPG, POPE, cholesterol and cationic surfactant.<sup>7,13,14</sup> Signs are taken from separate experiments.<sup>15,16</sup>

When comparing ion binding measured from the PC headgroup order parameters between neutral and anionic membranes, it is important to note that the order parameters increase due to the addition of negative charged lipids according to the electrometer concept<sup>6,13</sup> (Fig. S1). Therefore, the headgroup order parameters of POPC in mixtures with anionic lipids are larger than in pure POPC lipid bilayer, which is also seen in the ion binding experiments from different mixtures in the absence of added calcium (Fig. S2). Upon addition of  $\text{CaCl}_2$ , the order parameters decrease and reach the values of pure PC bilayer close to the  $\text{CaCl}_2$  concentrations of  $\sim 50\text{-}300\text{mM}$ , depending on the amount of negatively charged lipids in the mixture (Fig. S2). Around these concentrations the positive charge of bound  $\text{Ca}^{2+}$  cancels the negative charge lipids, resulting to a neutral membrane. Above such concentrations, the specific binding of calcium leads to the overcharging of the membrane.

Because the POPC headgroup order parameters in mixtures with different amounts of anionic lipids but without additional salt are not equal, the binding affinity of calcium can be better compared by plotting the changes of order parameters as a function of added calcium. As expected, such a plot reveals more pronounced order parameter decrease in systems with larger fractions of negatively charged lipids (Fig. S3), indicating an increase in the calcium binding affinity with the increasing amount of negatively charged lipids in membranes. In conclusion, the presented empirical comparison of headgroup order parameter changes from various mixtures of POPC and anionic lipids with added calcium gives physically consistent results, suggesting that the electrometer concept can be used to determine the cation binding affinity also to the lipid bilayers containing mixtures of PC and anionic lipids.

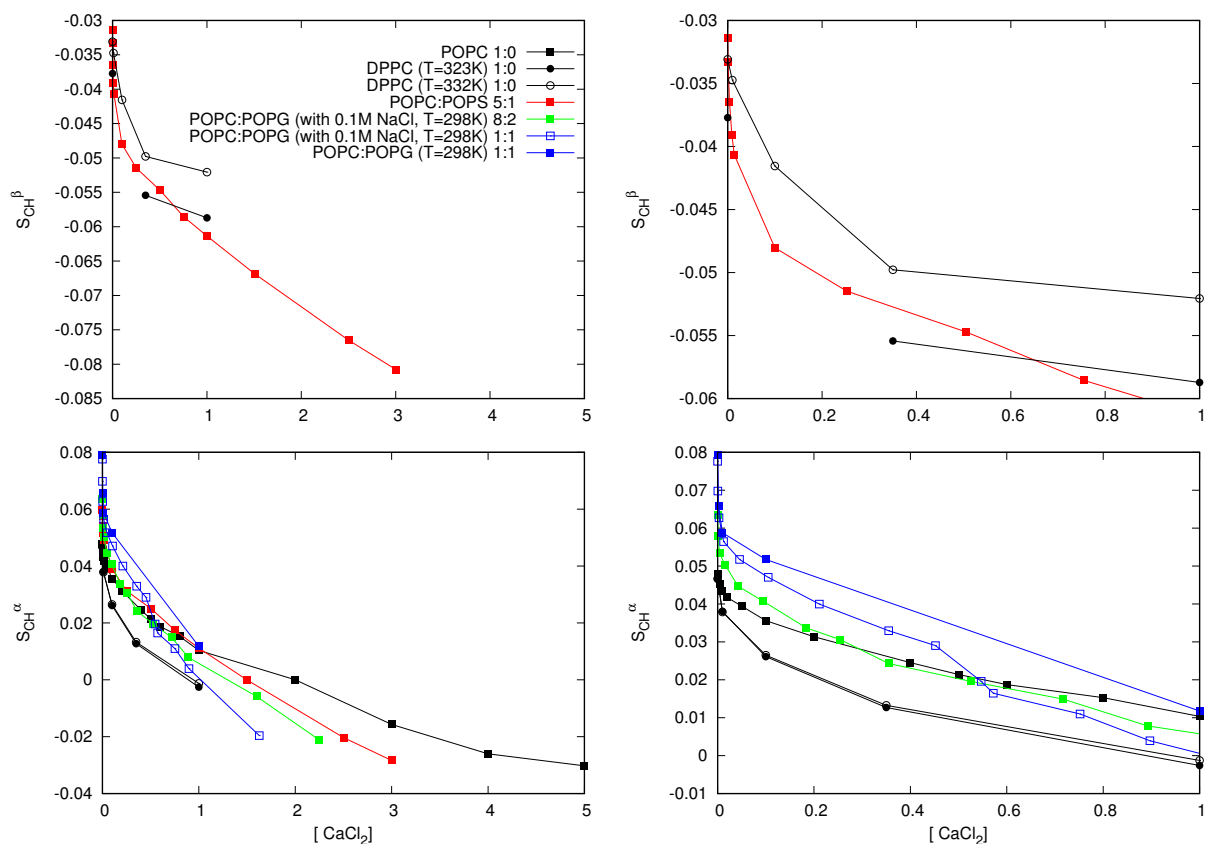


Figure S2: Headgroup order parameters of POPC as a function of  $CaCl$  concentration from experiments different mole fractions of negatively charged lipids (left column). Right column shows the same data zoomed to the concentrations below 1M. Data for Pure DPPC from,<sup>4</sup> for pure POPC from,<sup>5</sup> for POPC:POPS (5:1) mixture from,<sup>10</sup> for POPC:POPG (8:2,1:1) mixtures with 0.1M NaCl from<sup>9</sup> and for POPC:POPG (1:1) mixture data without NaCl from.<sup>8</sup>

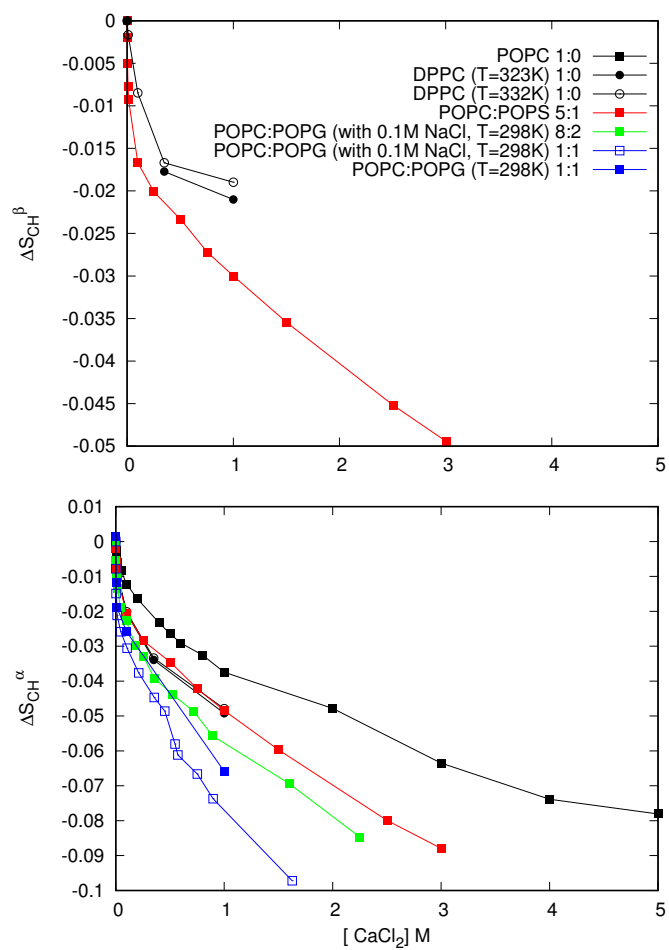


Figure S3: Changes of POPC headgroup order parameters as a function of  $CaCl_2$  measured from mixed bilayers containing different amounts of anionic lipids. The original data is the same as in figure S2.

### S3 Calibration of PC headgroup order parameter response to the bound cations

Before using the headgroup order parameters to compare ion binding affinity between simulations and experiments, the response of the order parameters to the bound charge in simulations should be quantified against experiments.<sup>17,18</sup> In our previous work,<sup>17</sup> the ratio  $\Delta S_{\text{CH}}^{\beta}/\Delta S_{\text{CH}}^{\alpha}$  was in good agreement with the experiments<sup>4</sup> in the Lipid14 model, but was underestimated by other models. In the more recent study,<sup>18</sup> the headgroup order parameter responses were compared more carefully with the experiments of cationic surfactants in POPC bilayer.<sup>7</sup> The comparison reveals that the both headgroup order parameters in the Lipid14 model are too sensitive to the bound charge, while CHARMM36 gives better agreement for the  $\alpha$  carbon (Fig. S4). Therefore, the headgroup order parameter response to the bound charge is actually more realistic in CHARMM36 model than in the Lipid14, where both order parameters are equally too sensitive to the bound cations giving a good result for the  $\Delta S_{\text{CH}}^{\beta}/\Delta S_{\text{CH}}^{\alpha}$  ratio. The ratio was overestimated for the CHARMM36 model because the  $\beta$ -carbon order parameter is relatively more sensitive than the  $\alpha$ -carbon order parameter. These results have to be taken into account when analysing the ion binding affinities using headgroup order parameters in simulations. However, we conclude that the discrepancies arising from the sensitivity of lipid headgroup to bound charge are typically smaller than the discrepancies arising from ion binding affinity.

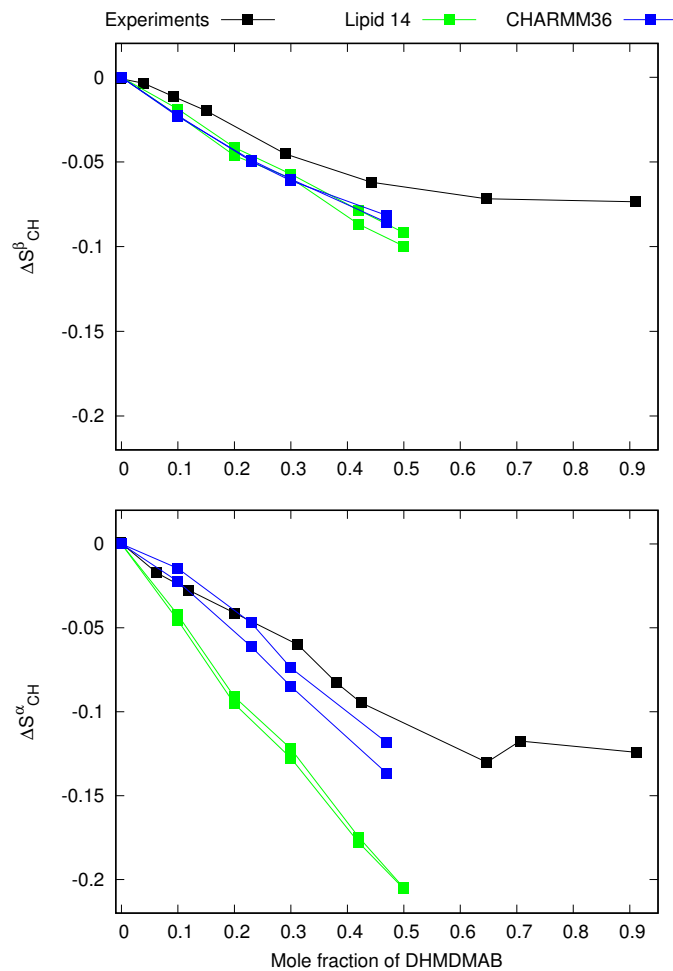


Figure S4: Responses of headgroup order parameters to the fixed amount of cationic surfactants in POPC bilayer from simulations and experiments.<sup>7</sup> The simulation results for Lipid14 are directly from Ref. 18. The CHARMM36 simulation data and details are available from Ref. 19.



## S4 Difference between POPC and OPPS in MacRog model

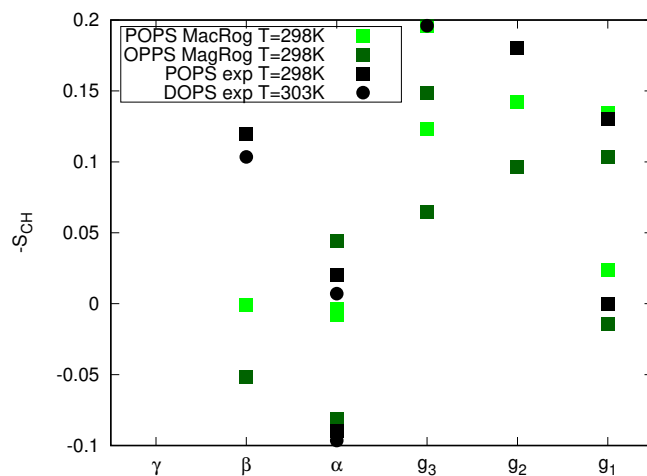


Figure S5: Headgroup order parameters from POPS and OPPS simulations with MacRog model.

## S5 Dihedrals

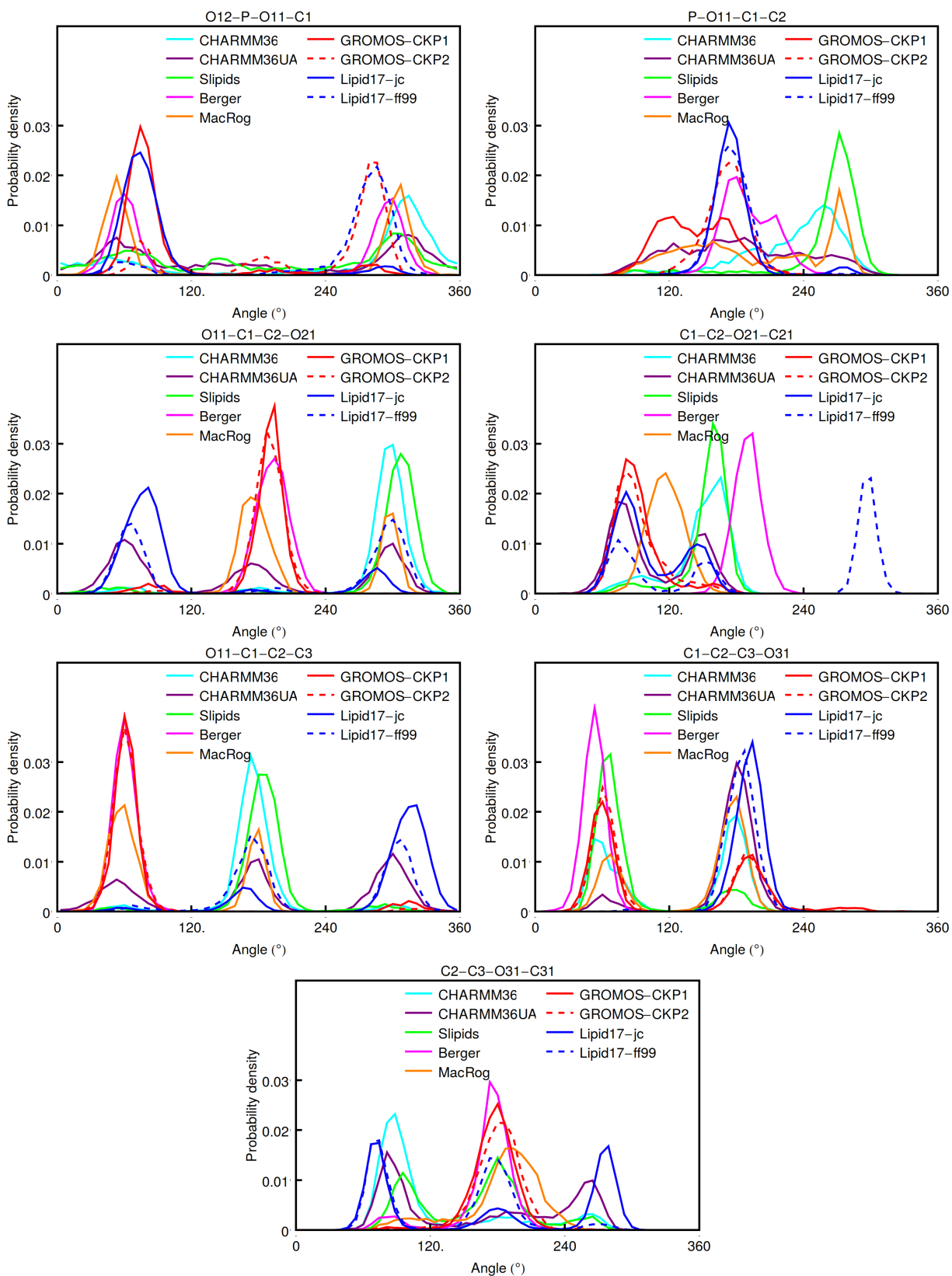


Figure S6: Dihedral angle distributions of bonds from phosphate to acyl chain carbonyls from different simulation models.

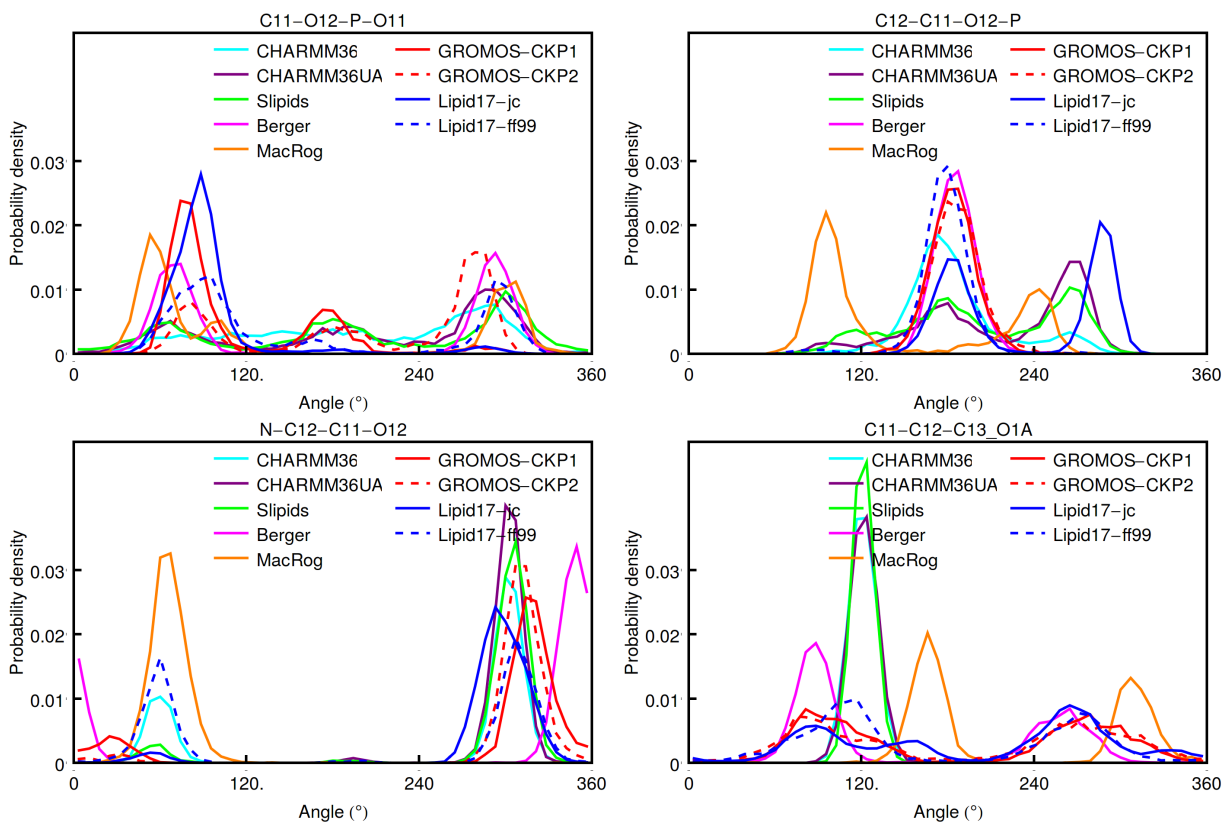


Figure S7: Dihedral angle distributions of bonds from phosphate to headgroup from different simulation models.

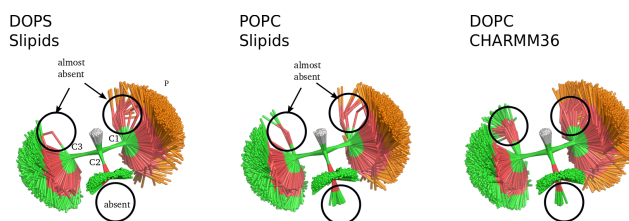


Figure S8: Snapshots overlayed from different simulations for glycerol backbone region by Pavel Buslaev.

## S6 Sodium binding to DMPC:DOPS mixture

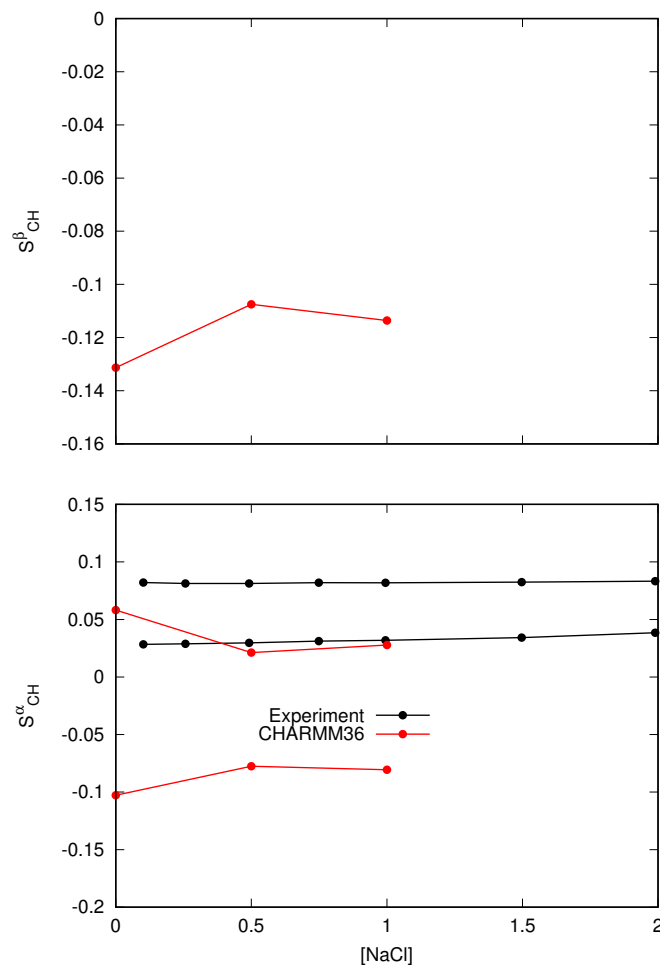


Figure S9: Order parameters of PS headgroup as a function of added NaCl measured from DMPC:DMPS (3:1) mixture.<sup>11</sup>

The experimental results show essentially no changes in the order parameters as a function of added NaCl, while significant changes are observed in simulations. However, the minimum buffer concentration of NaCl in the experimental was 100mM.<sup>11</sup> Therefore, we cannot exclude the possibility that the NaCl induced changes were already saturated with 100mM NaCl concentration, which was the case for  $\text{CaCl}_2$  in Fig. 9.

## S7 Calcium binding to POPC in CHARMM36 simulation with NBfix

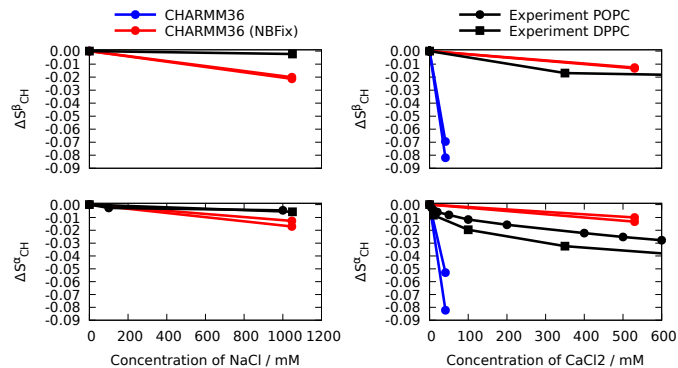


Figure S10: The response of headgroup order parameters to the fixed amount of cationic surfactants in POPC bilayer is compared between simulations and experiments.<sup>7</sup>

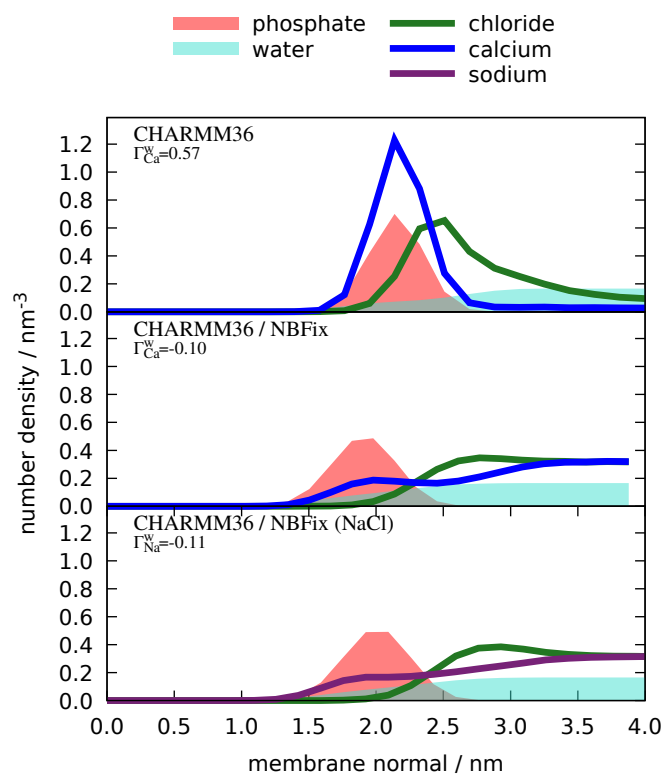


Figure S11: The response of headgroup order parameters to the fixed amount of cationic surfactants in POPC bilayer is compared between simulations and experiments.<sup>7</sup>

## S8 Details of the rough subjective force field ranking (Fig. 4)

The assessment was based fully on the Fig. 3. First, for each carbon (the columns in Fig. 3) in each force field (the rows), we looked separately at deviations in magnitude and forking.

**Magnitude** deviations, i.e., how close to the experimentally obtained C–H order parameters (OPs) the force-field-produced OPs were. For each carbon, the following 5-step scale was used:

**0 ( )**: More than half of all the calculated OPs (that is, of all different hydrogens in all different lipids) were within the *subjective sweet spots* (SSP, blue-shaded areas in Fig. 3).

**1 (Ⓜ)**: All the calculated OPs were  $< 0.03$  units away from the SSP.

**2 (M)**: All the calculated OPs were  $< 0.05$  units away from the SSP.

**3 (M)**: All the calculated OPs were  $< 0.10$  units away from the SSP.

**4 (M)**: Some of the calculated OPs were  $> 0.10$  units away from the SSP.

**Forking** deviations, i.e., how well the difference in order parameters of two hydrogens attached to a given carbon matched that obtained experimentally. Note that this is not relevant for  $\beta$  and  $g_2$ , which have only one hydrogen. For the  $\alpha$  carbon, for which a considerable forking of 0.105 is experimentally seen, the following 5-step scale was used:

**0 ( )**: The distance  $D$  between the dots (that mark the measurement-time-weighted averages in Fig. 3) was  $0.08 < D < 0.13$  units for all the calculated OPs (that is, for all different lipids).

**1 (f)**:  $(0.06 < D < 0.08)$  OR  $(0.13 < D < 0.15)$ .

**2 (F)**:  $(0.04 < D < 0.06)$  OR  $(0.15 < D < 0.17)$ .



**3 (F):**  $(0.02 < D < 0.04)$  OR  $(0.17 < D < 0.19)$ .

**4 (F):**  $(D < 0.02)$  OR  $(0.19 < D)$ .

For the  $g_3$  carbon, for which no forking is indicated by experiments, the following 5-step scale was used:

**0 ( ):**  $D < 0.02$ .

**1 (F):**  $0.02 < D < 0.04$ .

**2 (F):**  $0.04 < D < 0.06$ .

**3 (F):**  $0.06 < D < 0.08$ .

**4 (F):**  $0.08 < D$ .

For the  $g_1$  carbon, for which a considerable forking of 0.13 is experimentally seen, the following 5-step scale was used:

**0 ( ):**  $0.11 < D < 0.15$ .

**1 (F):**  $(0.09 < D < 0.11)$  OR  $(0.15 < D < 0.17)$ .

**2 (F):**  $(0.07 < D < 0.09)$  OR  $(0.17 < D < 0.19)$ .

**3 (F):**  $(0.05 < D < 0.07)$  OR  $(0.19 < D < 0.21)$ .

**4 (F):**  $(D < 0.05)$  OR  $(0.21 < D)$ .

Based on these assessments of magnitude and forking deviations, each carbon was then assigned to one of the following groups: "within experimental error" (magnitude and forking deviations both on step 0 of the scales described above), "almost within experimental error" (sum of the magnitude and forking deviation steps 1 or 2), "clear deviation from experiments" (sum of magnitude and forking deviation steps from 3 to 5), and "major deviation from experiments" (sum of magnitude and forking deviation steps from 6 to 8). These groups are

indicated by colors in Fig. 4. (Note that for  $\beta$  and  $g_2$ , for which there can be no forking, the corresponding group assignment limits were: 0, 1, 2, and 3.)

Finally, the total ability of the force field to describe the headgroup and glycerol structure was estimated. To this end, the groups were given the following weights: 0 (within experimental error), 1 (almost within experimental error), 2 (clear deviation from experiments), 4 (major deviation from experiments), and the weights of the five carbons were summed up. The sum, given in the  $\Sigma$ -column of Fig. 3, was then used to (roughly and subjectively, as should be clear from the above description) rank the force fields.

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