

# NMRLipids IV: Headgroup & glycerol backbone structures, and cation binding in bilayers with PS lipids

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Primarily measured but also simulated NMR order parameters will be collected also for other than phosphatidylcholine (these are discussed in NMRLipids I) headgroup. The information will be used to understand structural differences between different lipid molecules in bilayers.

## INTRODUCTION

In NMRLipids I and II project we were looking for a MD model which would correctly reproduce headgroup and glycerol backbone structures and cation binding for PC lipid bilayers [1, 2]. Here we extend the same goal for lipids with negatively charged PS headgroup. Chemical structure of PS headgroup together with other common biological lipids is shown in Fig. 1.

Absolute values of experimental order parameters for different lipid headgroups are collected from the literature in Fig. 2. Since order parameter signs are known only for PC, only absolute values are shown. Main conclusions regarding the structure of different common lipid headgroups in the literature are

- 1) glycerol backbone structures are largely similar irrespectively of the headgroup [3],
  - 2) glycerol backbone and headgroup structure and behaviour are similar in model membranes and in bacteria [3–5],
  - 3) headgroup structures are similar in PC, PE and PG lipids, while headgroup is more rigid in PS lipids [6, 7].
- Careful discussion and analysis of structural details of PE, PG or PS headgroups is not available, in contrast to PC lipids (see [1] and references therein).

As shown in Fig. 3, order parameters of PC headgroup

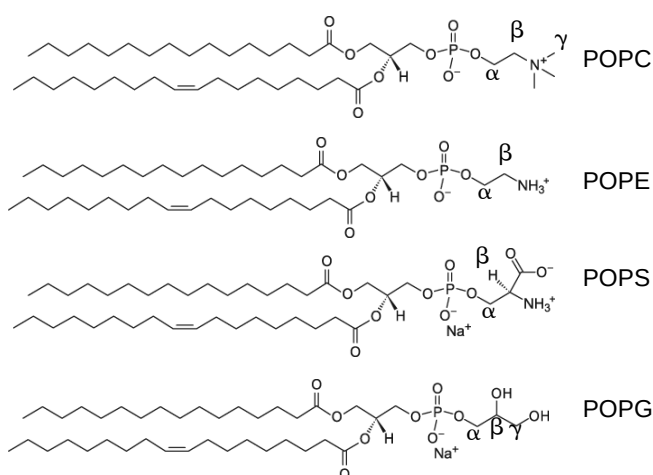


FIG. 1: Chemical structures and labels for the headgroup carbons.

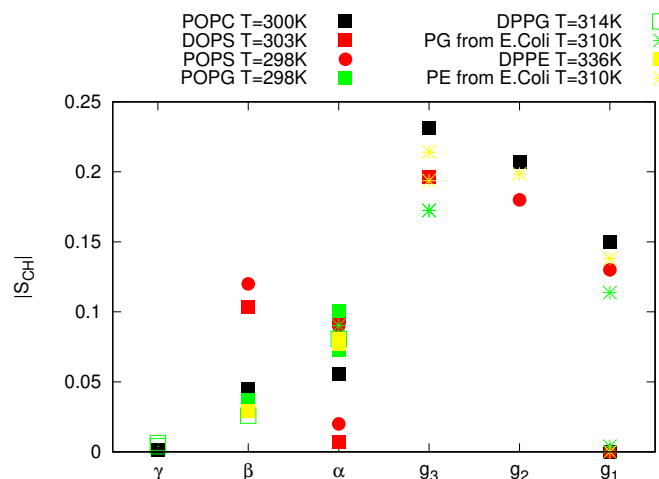


FIG. 2: Absolute values of order parameters for headgroup and glycerol backbone with different headgroups from experiments. POPC values are from [8], DOPS from [9] contains 0.1M of NaCl, POPG from [10] contains 10nM PIPES, DPPG from [6] contains 10mM PIPES and 100mM NaCl, DPPE from [11], E.coliPE and E.coliPG are from [3].

behave in various lipid mixtures as expected from the electrometer concept [4, 12], i.e., order parameters increase when anionic lipids are mixed with PC and decrease with cationic surfactants. The changes with the addition of neutral lipids is significantly smaller.

## RESULTS AND DISCUSSION

### PS headgroup and glycerol backbone structure in simulations and experiments

The headgroup order parameters of DOPS and POPS bilayers from different experiments and simulations are shown in Figs. 4. None of the tested models gives satisfactory agreement with experiments for order parameters in headgroup  $\alpha$  and  $\beta$  carbons.

Glycerol backbone order parameters seems similar in all models, except in Slipids. Even though glycerol backbone order parameter values are not yet experimentally available for PS lipids, the comparison with the results for PC lipids

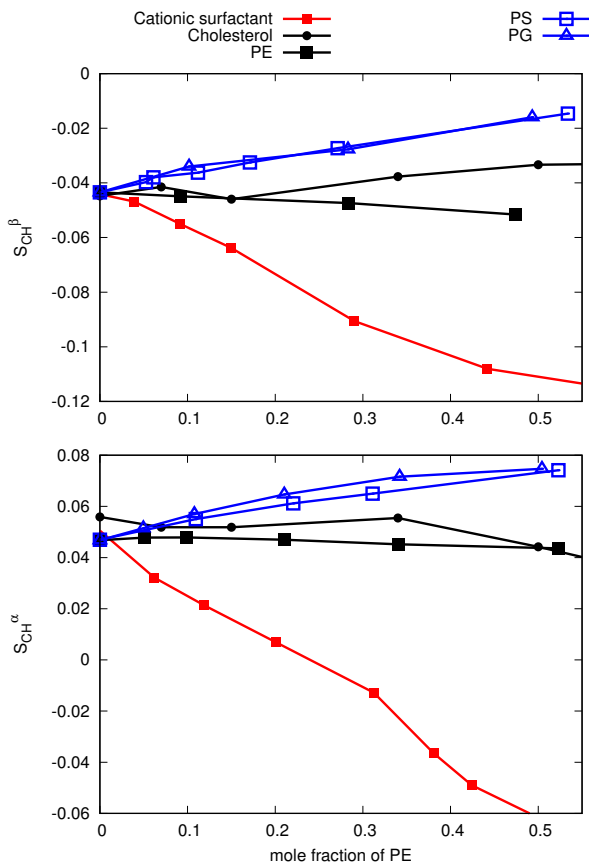


FIG. 3: PC headgroup order parameters from experiments of mixtures with PE, PS, PG and cholesterol [4, 8, 13]. Signs are determined as discussed in [1, 14].

suggest that Slipid model do not correctly capture the glycerol backbone structure [1]. The glycerol backbone structures between PC and PS lipids simulated with CHARMM36 are compared with the structures simulated with CHARMM36 in Fig. 5. The differences in sampled conformation leading to the order parameter differences are clearly visible in the figure.

3. Dihedral angle distributions in Fig. 13 should be included in the discussion.

### Effect of PS on PC headgroup

The headgroup order parameters of POPC mixed with PS lipids are shown in Fig. 6 from different simulation model and experiments [4] with different mole fractions. As already discussed previously, the PC lipid headgroup behaviour follows the electrometer concept in experiments when mixed with other lipids, i.e., the order parameters increase when mixed with negatively charged lipids (PS, PI, CL, PA and PG) remains almost unchanged when mixed with neutral lipids (PE and SM) [4]. This is not the case in simulation data shown in Fig. 6.

5. More data to be collected before discussion.

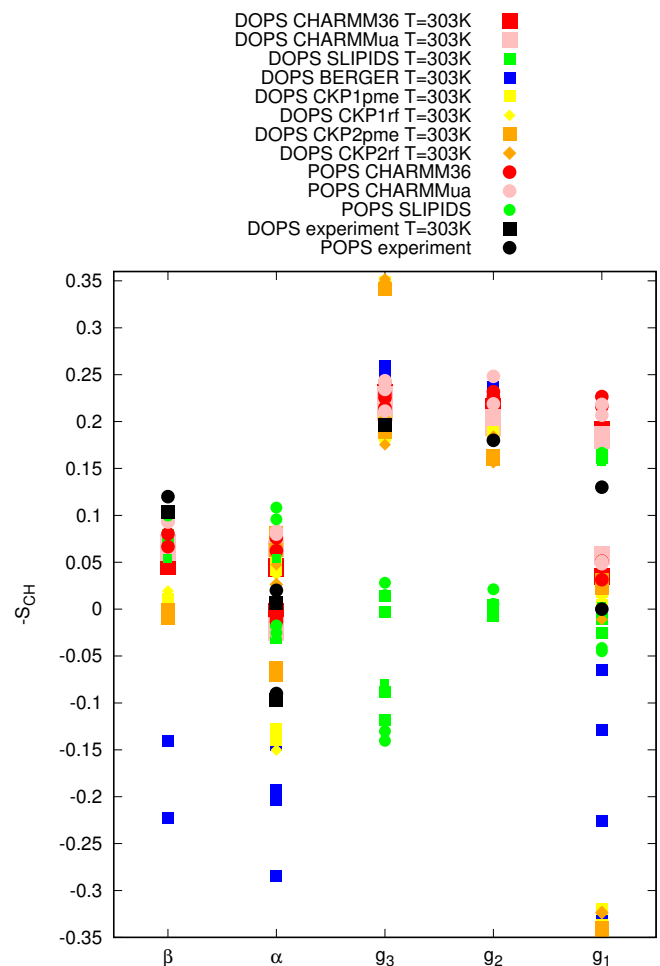


FIG. 4: Order parameters for DOPS headgroup and glycerol backbone from simulations with different models and experiments without  $\text{CaCl}_2$ . Experimental data from [9] contains 0.1M of NaCl. Signs are taken from experiments for POPS described in Supplementary Information.

1. Check and report all the counterions. 2. Glycerol backbone order parameters should be available from the spectra measured by Tiago Ferreira.

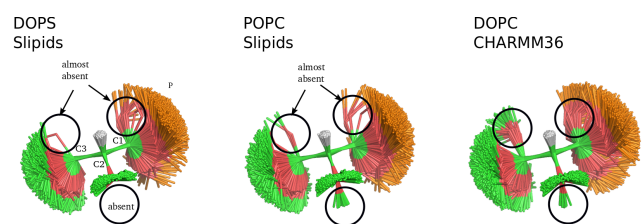


FIG. 5: Snapshots overlaid from different simulations for glycerol backbone region by Pavel Buslaev.

### Effect of PC on PS headgroup

The headgroup order parameters of PS mixed with varying amounts of PC from simulations and experiments [10, 15] are shown in Fig. 7. The effect of increasing amount of PC to PS

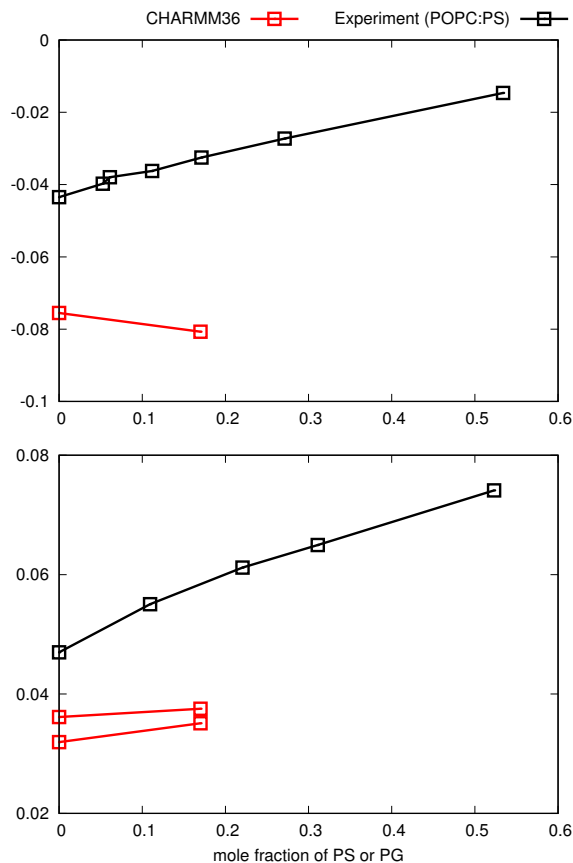


FIG. 6: PC headgroup order parameters from mixtures with PE, PS and PG lipids with various mole fractions from different simulation models and experiments [4]. Signs are determined as discussed in [1, 14].

4. Simulation of CHARMM36 at 298K should be maybe rerun with Gromacs 5.

headgroup seems to qualitatively incorrect in CHARMM36 simulations. The  $\beta$ -carbon order parameter increases in experiment, but decreases in simulations with both tested counterions ( $Na^+$  and  $K^+$ ). Larger  $\alpha$ -carbon order parameter decreases with the addition of PC in experiment, while the lower remains unchanged. In simulations the larger increases and the lower decreases. Interestingly, the  $\alpha$ -carbon order parameters are closer to experiments in pure PS system with  $K^+$  counterions than with  $Na^+$ .

#### $Ca^{2+}$ binding affinity in bilayers with negatively charged PS lipids

PC lipid headgroup order parameters can be used to measure ion binding affinity, because their magnitude is proportional to the amount of bound charge in bilayer [2, 12]. The molecular electrometer concept can be used also for bilayers containing PC lipids mixed with charged lipids [10, 15, 16]. This is demonstrated in Fig. 8, showing the changes of PC headgroup order parameters as a function of  $CaCl_2$  concentration in the presence of different amounts of negatively charged PS

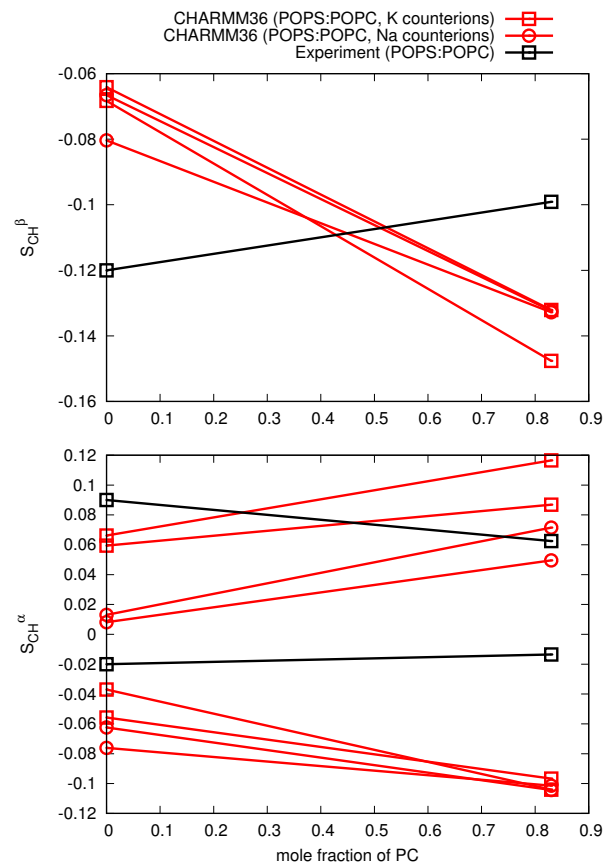


FIG. 7: PS order parameters from mixtures with PC lipids with various mole fractions from different simulation models and experiments [10, 15]. Signs for PS are measures as described in SI.

6. Some simulations contain potassium as counterions, while some sodium. All experiments here contain some amount of sodium salt. The best ion concentrations for comparison should be figured out.

7. Why there is difference between CHARMM36 simulation results from POPS:POPC mixture and pure POPS? Discussion in <https://github.com/NMRLipids/NMRLipidsIVotherHGs/issues/1>

or PG lipids. The decrease of order parameters with  $CaCl_2$  is more pronounced for systems with more negatively charged lipids. Order parameters reach the values of pure PC bilayer close to  $CaCl_2$  concentrations of  $\sim 50-300mM$ . At this point the  $Ca^{2+}$  binding presumably fully cancels the charge from negative lipids and overcharging occurs above these concentrations. The interpretation of this data and some other results has been that [5]

- ”(i)  $Ca^{2+}$  binds to neutral lipids (phosphatidylcholine, phosphatidylethanolamine) and negatively charged lipids (phosphatidylglycerol) with approximately the same binding constant of  $K = 10-20 M^{-1}$ ;
- (ii) the free  $Ca^{2+}$  concentration at the membrane interface is distinctly enhanced if the membrane carries a negative surface charge, either due to protein or to lipid;

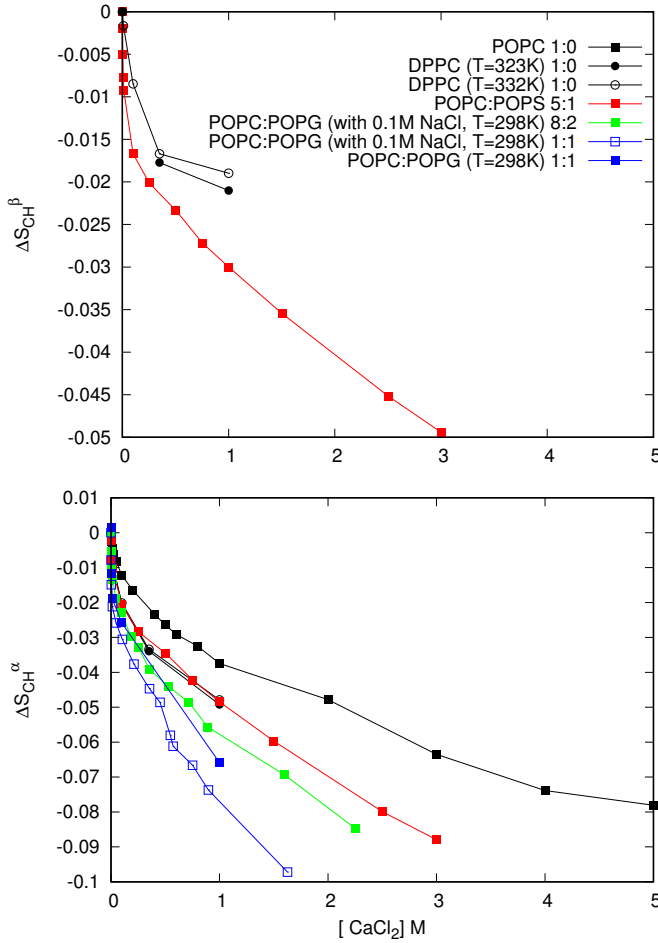


FIG. 8: The change of PC headgroup order parameters in the presence of different amount of negatively charged lipids respect to the values without added  $\text{CaCl}_2$ . The original data is the same as in Fig. ??.

- (iii) increased inter-facial  $\text{Ca}^{2+}$  also means increased amounts of bound  $\text{Ca}^{2+}$  at neutral and charged lipids;
- (iv) the actual binding step can be described by a Langmuir adsorption isotherm with a 1 lipid:1  $\text{Ca}^{2+}$  stoichiometry, provided the interfacial concentration  $C_M$ , is used to describe the chemical binding equilibrium."

8. When we have more data for Ca binding to PS containing bilayers, the discussion will be updated and PG results moved to other manuscript. Comparison of  $\text{Ca}^{2+}$  binding in PG between CHARMM36 simulations and experiments [10] is shown in Fig. 9. The decrease of  $\alpha$  order parameter is in agreement with experiments, while decrease of  $\beta$  order parameter is overestimated. The result is very similar to the results with PC in NMRlipids II publication [2]. It should be, however, noted that the  $\beta$ -order parameters are not actually measured for PG, but they are calculated from empirical relation  $\Delta S_\beta = 0.43\Delta S_\alpha$  [17]. Anyway, the data presented in NMRlipids II project and in Fig.

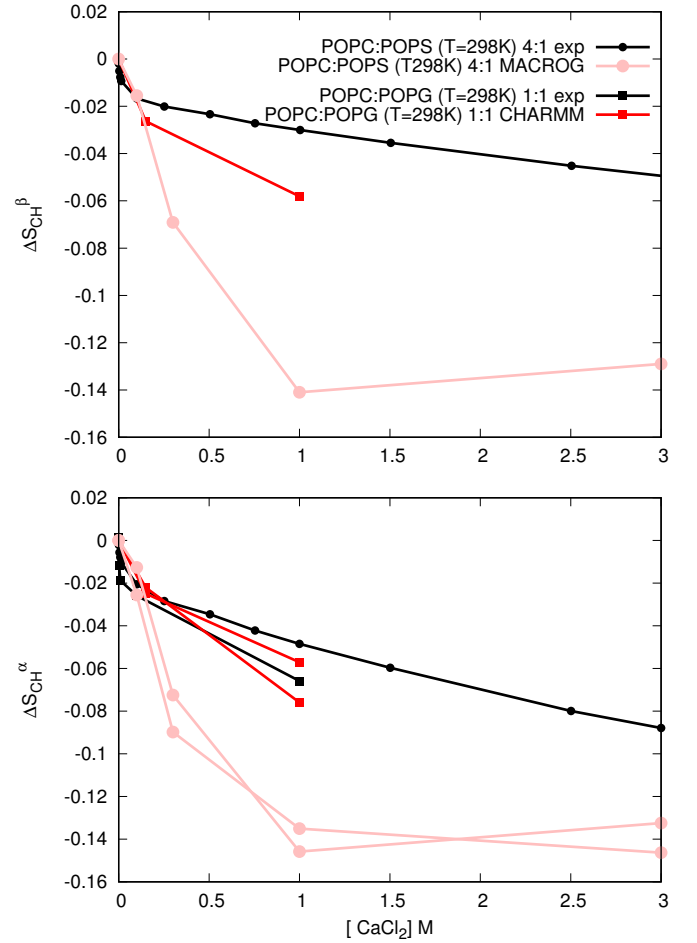


FIG. 9: PG order parameters as a function  $\text{CaCl}_2$  concentration from experiments [10] and CHARMM36 simulations. Note that beta order parameter is calculated from empirical relation  $\Delta S_\beta = 0.43\Delta S_\alpha$  [17], not actually measured.

9 together suggest that Calcium binding is similarly overestimated by CHARMM36 model in pure POPC bilayers and mixtures with POPG. The good agreement of  $\alpha$  carbon would be explained by too weak dependence of its order parameter of bound charge 9. The response of CHARMM36 against cationic surfactant experiments [13] is to be checked. I have already ran the simulations, analysis yet to be done.

Also dependence of  $\beta$ -carbon of PG on  $\text{CaCl}_2$  concentration is compared with experiments [10] in Fig. 10. Absolute value of the order parameter is too large without ions, but rapid decrease due to addition of  $\text{CaCl}_2$  is observed in agreement with experiments for systems with 1:1 mixture of POPC and POPG. In addition, absolute value in systems with  $\text{CaCl}_2$  is in agreement with experiments. However, system with 4:1 mixture of POPC and POPG behaves differently, but experimental data is not available for comparison for this mixture.

10. More simulation data for systems with negatively charged lipids and  $\text{CaCl}_2$  to be collected

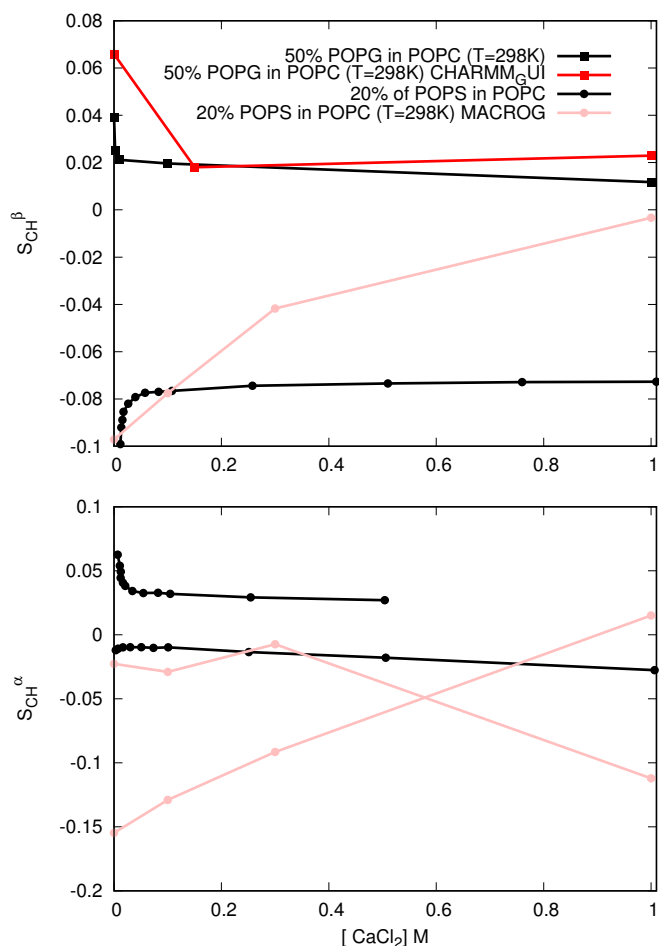


FIG. 10: PG and PS order parameters as a function of  $\text{CaCl}_2$  concentration taken from [10] and [15], respectively.

12. Get the small concentration data from the inserts

#### Effect of $\text{Ca}^{2+}$ binding to PS headgroup

Also the experimental order parameters for PS and PG headgroups as a function of  $\text{CaCl}_2$  concentration are shown in Fig. 10. These should be compared to simulations for potential structural interpretation of the changes.

## CONCLUSIONS

## SUPPLEMENTARY INFORMATION

### Simulated systems

#### Measurements of order parameter sign

Fig. 11 summarizes the experimental results on the order parameter sign measurement for POPS sample. The experimental protocol is the same used in Ref. 27. In (a) you see the headgroup region of the INEPT spectrum where alpha and beta are identified. In (b) you have the R-PDLF slices for alpha and beta where you see one single splitting for beta (which gives an order parameter equal to 0.12), and for alpha a superposition of a large splitting (order parameter equal to 0.09) and a very small splitting which cannot be calculated. On the bottom you have the S-DROSS slices of these two carbons. The grey lines show a random collection of slices from noise such that it gets clear what is significant. The S-DROSS slice for beta clearly shows that the order parameter is negative. The slice for alpha shows that the higher order parameter is positive and suggests that the smaller order parameter is negative (from the deviation towards negative values in the longer  $t_1$  times).

The results updated with SIMPSON simulations for the S-DROSS profiles are shown in Fig. 12. The value for the smaller alpha order parameter is taken from Fig 3 in Ref. 28, because resolution in  $^{13}\text{C}$  NMR experiments was not high enough to determine numerical value for this. The plots in Fig. 12 (c) show the following. The error bars and points are the experimental S-DROSS data. The thick lines are SIMPSON simulations. The simulations were done by using the order parameter for beta equal to -0.12 and for alpha one order parameter equal to 0.09 and the other equal to -0.02 (black) or 0.02 (grey). Since the black lines agree with experimental data, we conclude that the order parameters for  $\beta$  carbon are -0.12 and for  $\alpha$  order parameters are 0.09 and -0.02.

### Dihedrals

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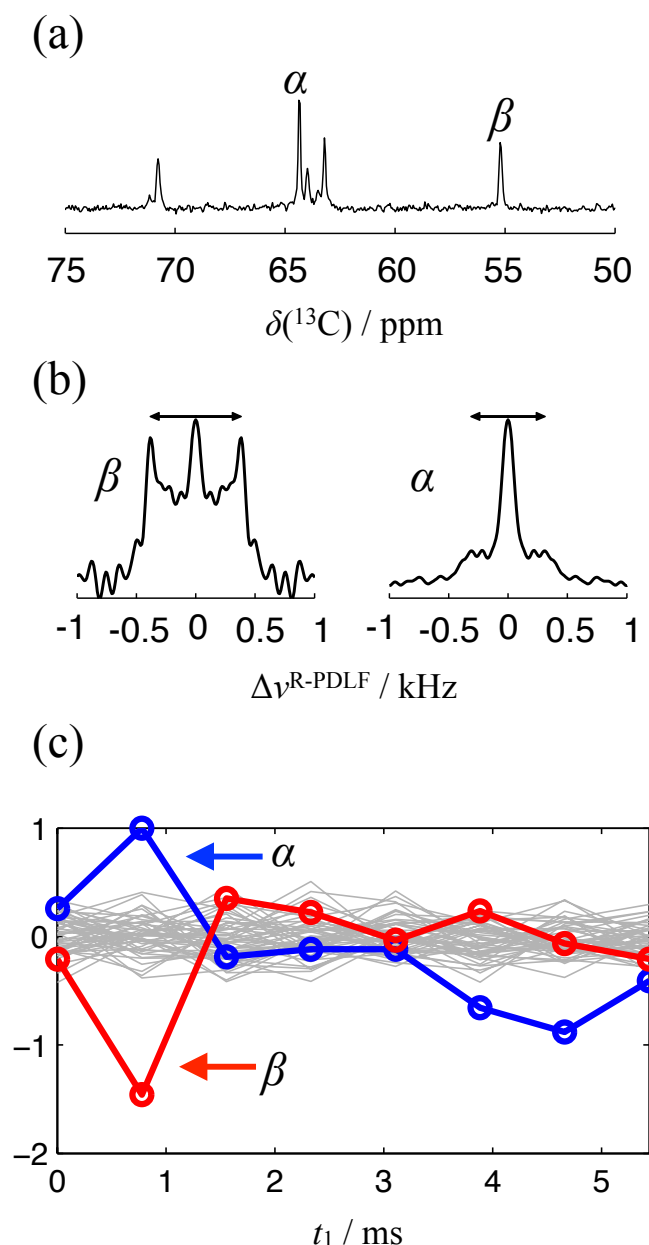


FIG. 11: Experimental results for sign measurement for POPS sample

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## ToDo

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|--|----|
| 3. Dihedral angle distributions in Fig. 13 should be included in the discussion. . . . .   | 2  |
| 5. More data to be collected before discussion. . . . .  | 2  |
| 1. Check and report all the counterions. . . . .   | 2  |
| 2. Glycerol backbone order parameters should be available from the spectra measured by Tiago Ferreira. . . .   | 2  |
| 4. Simulation of CHARMM36 at 298K should be maybe rerun with Gromacs 5. . . . .  | 3  |
| 6. Some simulations contain potassium as counterions, while some sodium. All experiments here contain some amount of sodium salt. The best ion concentrations for comparison should be figured out. . . . .  | 3  |
| 7. Why there is difference between CHARMM36 simulation results from POPS:POPC mixture and pure POPS? Discussion in <a href="https://github.com/NMRLipids/NMRLipidsIVotherHGs/issues/1">https://github.com/NMRLipids/NMRLipidsIVotherHGs/issues/1</a> | 3  |
| 8. When we have more data for Ca binding to PS containing bilayers, the discussion will be updated and PG results moved to other manuscript. . . . .   | 4  |



9. The response of CHARMM36 against cationic surfactant experiments [13] is to be checked. I have already ran the simulations, analysis yet to be done . . .	4	19. Correct citation(s) for CKP. . . . .	8
10. More simulation data for systems with negatively charged lipids and $\text{CaCl}_2$ to be collected . . . . .	4	20. Delivered by Piggot. We need to decide between RF and PME or discuss this somehow. Data to be uploaded in Zenodo? . . . . .	8
12. Get the small concentration data from the inserts . . .	5	21. Correct citation for CHARMM POPS . . . . .	8
11. These should be compared to simulations for potential structural interpretation of the changes. . . . .	5	22. Delivered by Piggot. We need to decide the switching version or discuss this somehow. Data to be uploaded in Zenodo? . . . . .	8
13. Correct citation for CHARMM DOPS . . . . .	8	23. Correct citation for CHARMMua DOPS . . . . .	8
14. By Piggot: <a href="http://nmrlipids.blogspot.com/2017/03/nmrlipids-headgroup-glycerol.html?showComment=1491425687561#c4932902612512697301">http://nmrlipids.blogspot.com/2017/03/nmrlipids-headgroup-glycerol.html?showComment=1491425687561#c4932902612512697301</a> We need to decide the switching version or discuss this somehow. . . . .	8	24. Delivered by Piggot. We need to decide the switching version or discuss this somehow. Data to be uploaded in Zenodo? . . . . .	8
15. Correct citation for CHARMMua DOPS . . . . .	8	25. Delivered by Piggot. We need to decide the cut-off version or discuss this somehow. Data to be uploaded in Zenodo? . . . . .	8
16. Delivered by Piggot. We need to decide the switching version or discuss this somehow. Data to be uploaded in Zenodo? . . . . .	8	26. Correct citation for CHARMM POPS . . . . .	8
17. Delivered by Piggot. We need to decide the cut-off version or discuss this somehow. Data to be uploaded in Zenodo? . . . . .	8	27. Details to be filled and data to be uploaded in Zenodo by Ollila. . . . .	8
18. Delivered by Piggot. Data to be uploaded in Zenodo? . . . . .	8	28. Correct citation for CHARMM POPG . . . . .	8
		29. Details to be filled and data to be uploaded in Zenodo by Ollila. . . . .	8

TABLE I: List of MD simulations. The salt concentrations calculated as  $[\text{salt}] = N_c \times [\text{water}] / N_w$ , where  $[\text{water}] = 55.5 \text{ M}$ .

lipid/counter-ions	force field for lipids / ions	NaCl (mM)	CaCl <sub>2</sub> (mM)	<sup>a</sup> N <sub>l</sub>	<sup>b</sup> N <sub>w</sub>	<sup>c</sup> N <sub>c</sub>	<sup>d</sup> T (K)	<sup>e</sup> t <sub>sim</sub> (ns)	<sup>f</sup> t <sub>anal</sub> (ns)	<sup>g</sup> files
DPPE	Slipids [18]	0		0 288	9386	0	336	200	100	[19]
DOPS/Na <sup>+</sup>	CHARMM36 [?] ] 13.	0		0 128	4480	0	303	500	100	[?] ] 14.
DOPS/Na <sup>+</sup>	CHARMM36ua [?] ] 15.	0		0 128	4480	0	303	500	100	[?] ] 16.
DOPS/Na <sup>+</sup>	Slipids [20]	0		0 128	4480	0	303	500	100	[?] ] 17.
DOPS/Na <sup>+</sup>	Slipids [20]	0		0 288	11232	0	303	200	100	[21]
DOPS/Na <sup>+</sup>	Berger [22]	0		0 128	4480	0	303	500	100	[?] ] 18.
DOPS/Na <sup>+</sup>	GROMOS-CKP [?] ] 19.	0		0 128	4480	0	303	500	100	[?] ] 20.
POPS/Na <sup>+</sup>	CHARMM36 [?] ] 21.	0		0 128	4480	0	298	500	100	[?] ] 22.
POPS/Na <sup>+</sup>	CHARMM36ua [?] ] 23.	0		0 128	4480	0	298	500	100	[?] ] 24.
POPS/Na <sup>+</sup>	Slipids [20]	0		0 128	4480	0	298	500	100	[?] ] 25.
POPC:POPS (5:1)/Na <sup>+</sup>	CHARMM36 [23?] ] 26.	0		0 ?	? ?	0	?	?	?	[?] ] 27.
POPG/Na <sup>+</sup>	CHARMM36 [?] ] 28.	0		0 ?	? ?	0	?	?	?	[?] ] 29.
POPG/Na <sup>+</sup>	Slipids [20]	0		0 288	10664	0	298	250	100	[24]
DPPG/Na <sup>+</sup>	Slipids [20]	0		0 288	11232	0	314	200	100	[25]
DPPG/Na <sup>+</sup>	Slipids [20]	0		0 288	11232	0	298	400	100	[26]

<sup>a</sup>Number of lipid molecules with largest mole fraction<sup>b</sup>Number of water molecules<sup>c</sup>Number of additional cations<sup>d</sup>Simulation temperature<sup>e</sup>Total simulation time<sup>f</sup>Time used for analysis<sup>g</sup>Reference for simulation files



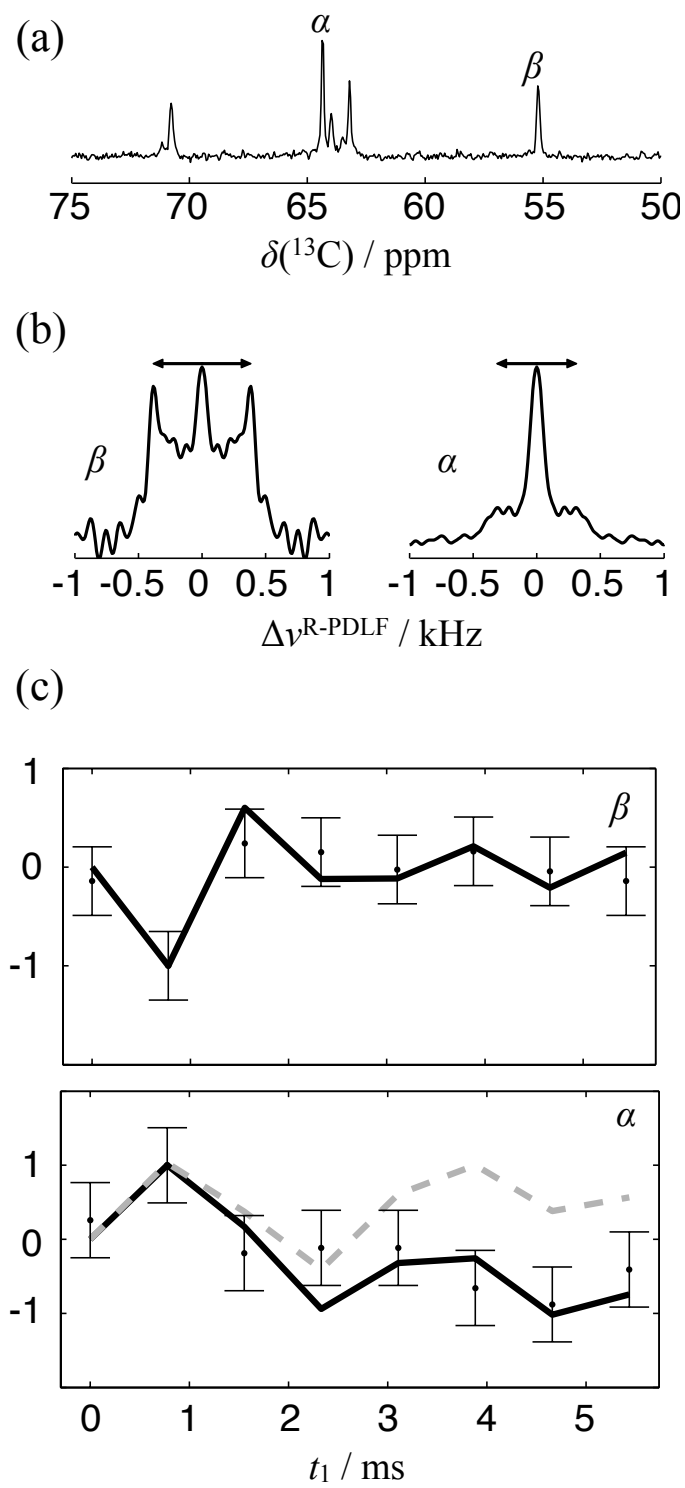


FIG. 12: Experimental results for sign measurement for POPS sample

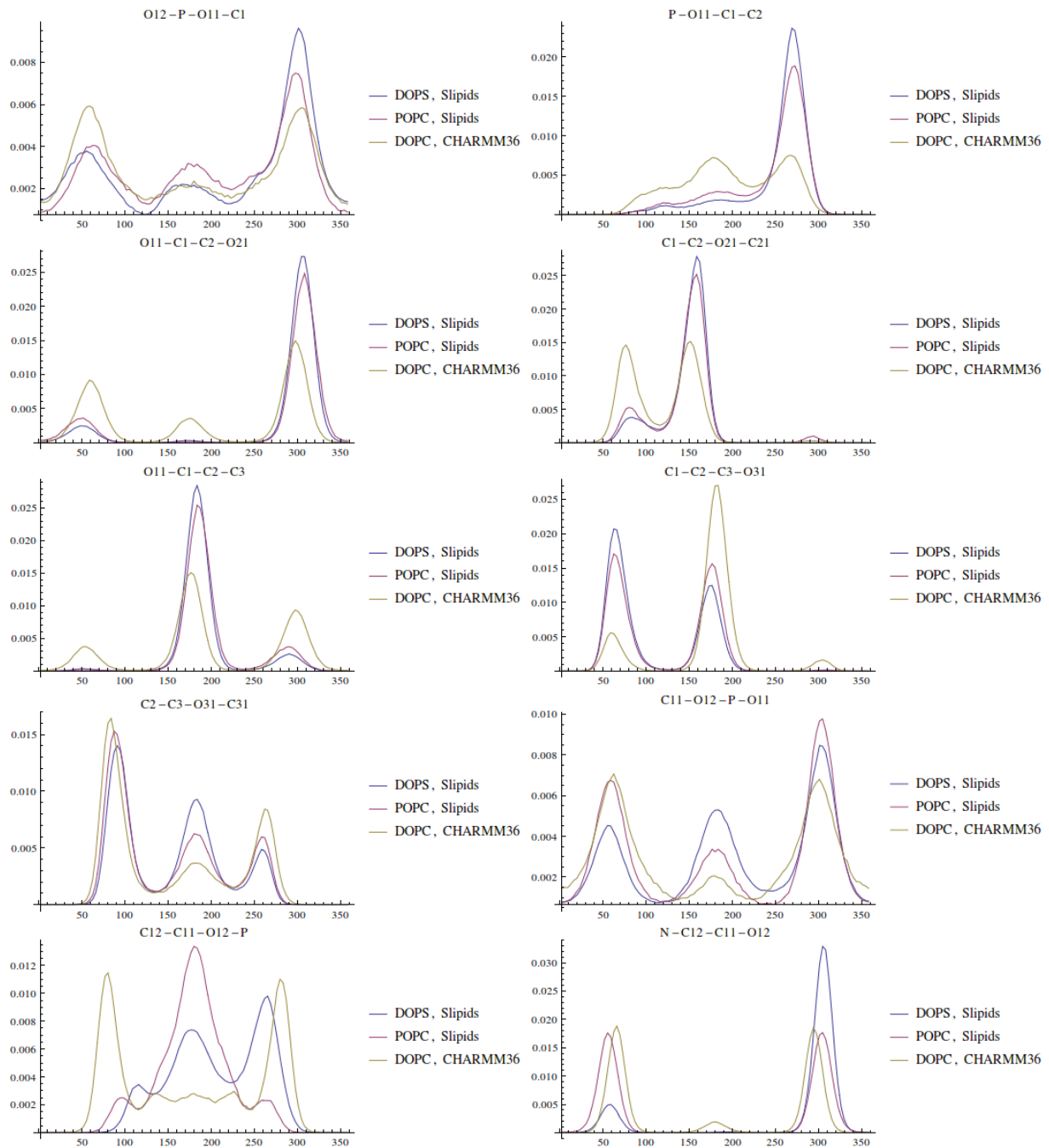


FIG. 13: Experimental results for sign measurement for POPS sample