Package 'mSigTools'

October 9, 2022

Oct	0001 7, 2022
Type Package	
Title Mutational Signature Analysis Tools	
Version 1.0.6	
Description Utility functions for mutational s This package provides two groups of fur for dealing with mutational signature `` (i.e. the counts of mutations in a sample due to each mutational signature). The o functions is for matching or comparing s mutational signatures. mSigTools' stands mutational Signature analysis Tools.	exposures" that are ther group of sets of
License GPL-3	
URL https://github.com/Rozen-Lab/mSi	gTools
BugReports https://github.com/Rozen-L	ab/mSigTools/issues
Encoding UTF-8	
Language en-US	
RoxygenNote 7.2.1	
Imports clue, philentropy, quadprog	
Suggests spelling, testthat (>= 3.0.0), sets, con	smicsig, ICAMS
Config/testthat/edition 3	
NeedsCompilation no	
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best_reconstruction_QP

Find "best" reconstruction of a target signature or spectrum from a set of signatures.

Description

Find "best" reconstruction of a target signature or spectrum from a set of signatures.

Usage

```
best_reconstruction_QP(
   target.sig,
   sig.universe,
   max.subset.size = NULL,
   method = "cosine",
   trim.less.than = 1e-10
)
```

Arguments

target.sig The signature or spectrum to reconstruct; a non-negative numeric vector or 1column matrix-like object. sig.universe The universe of signatures from which to reconstruct target.sig; a non-negative numeric matrix-like object with nrow(sig.universe) == length(target.sig). The sums of each column must be 1. Must not contain duplicate columns or have other non-unique quadratic programming solutions (not checked, but will generate an error from solve. QP in package quadprog). max.subset.size Maximum number of signatures to use to reconstruct target.sig. method As in dist_one_one in package philentropy, and used only to find the final "best" reconstruction. The optimized exposures from which to selected the "best" reconstruction are calculated using optimize_exposure_QP, which uses solve.QP in package quadprog.

trim.less.than After optimizing exposures with optimize_exposure_QP, remove exposures less than trim.less.than and then re-optimize.

Details

This function should be fast if you do not specify max.subset.size, but it will be combinatorially slow if max.subset.size is large and trim.less.than is small or negative. So do not do that. If max.subset.size is NULL, then the function just uses optimize_exposure_QP and then excludes exposures < trim.less.than, and then re-runs optimize_exposure_QP. Otherwise, after excluding exposures < trim.less.than, then the function runs optimize_exposure_QP on subsets of signatures of size <= max.subset.size, removes exposures < trim.less.than, reruns optimize_exposure_QP, calculates the reconstruction and similarity between the reconstruction and the target.sig and returns the information for the exposures that have the greatest similarity.

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Value

A list with elements:

• optimized.exposure A numerical vector of the exposures that give the "best" reconstruction. This vector is empty if there is an error.

- similarity The similarity between the reconstruction (see below) and target.sig according to the distance or similarity provided by the method argument.
- method The value specified for the method argument, or an error message if optimize. exposure is empty.
- reconstruction The reconstruction of target.sig according to optimized.exposure.

Examples

match_two_sig_sets

Find an optimal matching between two sets of signatures subject to a maximum distance.

Description

Find an optimal matching between two sets of signatures subject to a maximum distance.

```
match_two_sig_sets(
    x1,
    x2,
    method = "cosine",
    convert.sim.to.dist = function(x) {
        return(1 - x)
    },
    cutoff = 0.9
)
```

Arguments

x1 A numerical-matrix-like object with columns as signatures.

x2 A numerical-matrix-like object with columns as signatures. Needs to have the

same number of rows as x1.

method As for the distance function in package philenropy.

convert.sim.to.dist

If method specifies a similarity rather than a distance, use this function to convert

the similarity to a distance.

cutoff A maximum distance or minimum similarity over which to pair signatures be-

tween x1 and x2.

Details

Match signatures between x1 and x2 using the function solve_LSAP, which uses the "Hungarian" (a.k.a "Kuhn-Munkres") algorithm https://en.wikipedia.org/wiki/Hungarian_algorithm, which optimizes the total cost associated with the links between nodes. This function generates a distance matrix between the two sets of signatures using method and, if necessary, convert.sim.to.dist. It then sets distances > cutoff to very large values and then applies solve_LSAP to the resulting matrix to compute a matching between x1 and x2 that minimizes the sum of the distances.

Value

A list with the elements

- table Table of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the distance of the match.
- orig.matrix The matrix of numeric distances between x1 and x2.
- modified.matrix The argument orig.matrix with distances > cutoff changed to very large values.

Examples

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2) colnames(ex.sigs) <- c("ex1", "ex2", "ex3") ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2) colnames(ref.sigs) <- c("ref1", "ref2") match_two_sig_sets(ex.sigs, ref.sigs, cutoff = .9)
```

optimize_exposure_QP Quadratic programming optimization of signature activities

Description

Quadratic programming optimization of signature activities

```
optimize_exposure_QP(spectrum, signatures)
```

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Arguments

spectrum Mutational signature or mutational spectrum as a numeric vector or single col-

umn data frame or matrix.

signatures Matrix or data frame of signatures from which to reconstruct spectrum. Rows

are mutation types and columns are signatures. Should have column names for interpretable results. Cannot be a vector because the column names are needed.

Details

Code adapted from SignatureEstimation::decomposeQP and uses solve.QP in package quadprog.

Value

A vector of exposures with names being the colnames from signatures.

Examples

plot_exposure

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of samples.

Description

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of samples.

```
plot_exposure(
  exposure,
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
  ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
  plot.sample.names = TRUE,
  yaxis.labels = NULL,
  ...
)
```

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Arguments

exposure

Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use sort_exposure. Do not use column names that start with multiple underscores. The exposures will often be mutation counts, but could also be e.g. mutations per megabase.

samples.per.line

Number of samples to show in each plot.

plot.proportion

Plot exposure proportions rather than counts.

xlim, ylim Limits for the x and y axis. If NULL(default), the function tries to do something reasonable.

legend.x, legend.y

The x and y co-ordinates to be used to position the legend.

cex.legend A numerical value giving the amount by which legend plotting text and symbols should be magnified relative to the default.

cex.yaxis A numerical value giving the amount by which y axis values should be magnified

relative to the default.

cex.xaxis A numerical value giving the amount by which x axis values should be magni-

fied relative to the default. If NULL(default), the function tries to do something

reasonable.

plot.sample.names

Whether to plot sample names below the x axis. Default is TRUE. Ignored if there are no column names on exposure.

yaxis.labels User defined y axis labels to be plotted. If NULL(default), the function tries to do

something reasonable.

Other arguments passed to barplot. If ylab is not included, it defaults to a value depending on plot.proportion. If col is not supplied the function tries to do something reasonable.

Value

An **invisible** list. The first element is a logical value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of the bar x-axis midpoints drawn, useful for adding to the graph.

Examples

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
old.par <- par(mar = c(8, 5, 1, 1))
plot_exposure(exposure[, 1:30],
   main = "Liver-HCC exposure", cex.yaxis = 0.8,
   plot.proportion = TRUE
)
par(old.par)</pre>
```

plot_exposure_to_pdf

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Description

Plot exposures in multiple plots to a single PDF file, with each plot showing exposures for a manageable number of samples.

Usage

```
plot_exposure_to_pdf(
  exposure,
  file,
  mfrow = c(2, 1),
  mar = c(6, 4, 3, 2),
  oma = c(3, 2, 0, 2),
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
  ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
  plot.sample.names = TRUE,
  yaxis.labels = NULL,
 width = 8.2677,
  height = 11.6929,
)
```

Arguments

exposure	Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use sort_exposure. Do not use column names that start with multiple underscores. The exposures will often be mutation counts, but could also be e.g. mutations per megabase.
file	The name of the PDF file to be produced.
mfrow	A vector of the form c(nr,nc). Subsequent figures will be drawn in an nr-by-nc array on the device by rows.
mar	A numerical vector of the form c(bottom,left,top,right) which gives the number of lines of margin to be specified on the four sides of the plot.
oma	A vector of the form c(bottom,left,top,right) giving the size of the outer margins in lines of text.
samples.per.lin	ne

Number of samples to show in each plot.

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plot.proportion	n
	Plot exposure proportions rather than counts.
xlim, ylim	Limits for the x and y axis. If NULL(default), the function tries to do something reasonable.
legend.x, legen	nd.y
	The x and y co-ordinates to be used to position the legend.
cex.legend	A numerical value giving the amount by which legend plotting text and symbols should be magnified relative to the default.
cex.yaxis	A numerical value giving the amount by which y axis values should be magnified relative to the default.
cex.xaxis	A numerical value giving the amount by which x axis values should be magnified relative to the default. If NULL(default), the function tries to do something reasonable.
plot.sample.nar	mes
	Whether to plot sample names below the x axis. Default is TRUE. Ignored if there are no column names on exposure.
yaxis.labels	User defined y axis labels to be plotted. If NULL(default), the function tries to do something reasonable.
width, height	The width and height of the graphics region in inches.
	Other arguments passed to barplot. If ylab is not included, it defaults to a value depending on plot.proportion. If col is not supplied the function tries to do something reasonable.

Value

An **invisible** list. The first element is a logical value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of the bar x-axis midpoints drawn, useful for adding to the graph.

Examples

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
plot_exposure_to_pdf(exposure,
   file = file.path(tempdir(), "Liver-HCC.exposure.pdf"),
   cex.yaxis = 0.8, plot.proportion = TRUE
)</pre>
```

read_exposure

Read an exposure matrix from a file.

Description

Read an exposure matrix from a file.

```
read_exposure(file, check.names = FALSE)
```

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Arguments

file File path to a CSV file containing an exposure matrix, i.e. the numbers of mu-

tations due to each mutational signature. Each row corresponds to a mutational signature an each column corresponds to a tumor or other biological sample.

check.names Passed to read.csv. **IMPORTANT**: If TRUE this will replace the double colon

in identifiers of the form <tumor_type>::<sample_id> with two periods (i.e. <tumor_type>..<sample_id>. If check.names is true, generate a warning if double

colons were present.

Value

Numerical matrix of exposures, with the same shape as the contents of file.

Examples

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)</pre>
```

sig_dist_matrix

Compute a matrix of distances / similarities between two sets of signatures.

Description

Compute a matrix of distances / similarities between two sets of signatures.

Usage

```
sig_dist_matrix(x1, x2, method = "cosine")
```

Arguments

x1 The first set of signatures (a numerical matrix-like object in which each column

is a signature).

x2 The second set of signatures, similar data type to x1, and must have the same

number of rows as x1.

method As for the distance function in package philenropy.

Value

A numeric matrix with dimensions $ncol(x1) \times ncol(x2)$. Each element represents the distance or similarity (depending on method) between a column in x1 and a column in x2.

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Examples

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.4, 0.6), nrow = 2) colnames(ex.sigs) <- c("ex1", "ex2", "ex3") ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2) colnames(ref.sigs) <- c("ref1", "ref2") sig_dist_matrix(ex.sigs, ref.sigs)
```

sort_exposure

Sort columns of an exposure matrix based on the number of mutations in each sample (column).

Description

Sort columns of an exposure matrix based on the number of mutations in each sample (column).

Usage

```
sort_exposure(exposure, decreasing = TRUE)
```

Arguments

exposure

Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column

names are taken as the sample IDs.

decreasing

If TRUE, sort from largest to smallest.

Value

The original exposure with columns sorted.

Examples

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
exposure.sorted <- sort_exposure(exposure)</pre>
```

TP_FP_FN_avg_sim

_	d best matches (by cosine similarity) of a set of mutational signa- es to a set of reference mutational signatures.
ture	es to a set of reference mutational signatures.

Description

Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.

Usage

```
TP_FP_FN_avg_sim(extracted.sigs, reference.sigs, similarity.cutoff = 0.9)
```

Arguments

extracted.sigs Mutational signatures discovered by some analysis. A numerical-matrix-like object with columns as signatures.

reference.sigs A numerical-matrix-like object with columns as signatures. This matrix should contain the reference mutational signatures. For example, these might be from a synthetic data set or they could be from reference set of signatures, such as the signatures at the COSMIC mutational signatures web site. See CRAN package cosmicsig.

similarity.cutoff

A signature in reference.sigs must be matched by >= similarity.cutoff by a signature in extracted.sigs to be considered detected.

Details

Match signatures in extracted.sigs to signatures in reference.sigs using match_two_sig_sets based on cosine similarity.

Value

A list with the elements

- TP The number of true positive extracted signatures.
- FP The number of false positive extracted signatures.
- FN The number of false negative reference signatures.
- avg.cos.sim The average cosine similarity of true positives to their matching reference signatures.
- table A data frame of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the cosine similarity of the match
- sim.matrix The numeric distance or similarity matrix between extracted.sigs and reference.sigs as returned from sig_dist_matrix.
- unmatched.ex.sigs The identifiers of the extracted signatures that did not match a reference signature.
- unmatched.ref.sigs The identifiers of the reference signatures that did not match an extracted signature.

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Examples

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2)
colnames(ex.sigs) <- c("ex1", "ex2", "ex3")
ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2)
colnames(ref.sigs) <- c("ref1", "ref2")
TP_FP_FN_avg_sim(
    extracted.sigs = ex.sigs,
    reference.sigs = ref.sigs,
    similarity.cutoff = .9
)</pre>
```

write_exposure

Write an exposure matrix to a file.

Description

Write an exposure matrix to a file.

Usage

```
write_exposure(exposure, file, row.names = TRUE)
```

Arguments

exposure Exposures as a numerical matrix (or data.frame) with signatures in rows and

samples in columns. Rownames are taken as the signature names and column

names are taken as the sample IDs.

file File to which to write the exposure matrix (as a CSV file).

row.names Either a logical value indicating whether the row names of exposure are to be

written along with exposure, or a character vector of row names to be written.

Value

No return value, called for side effects.

Examples

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
write_exposure(exposure, file = file.path(tempdir(), "Liver-HCC.exposure.csv"))</pre>
```

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