# Package 'mSigTools'

August 27, 2022

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Type Package
Title Mutational Signature Analysis Tools
Version 1.0.3
Description Utility functions for mutational signature analysis.  This package provides two groups of functions. One is for dealing with mutational signature ``exposures" (i.e. the counts of mutations in a sample that are due to each mutational signature). The other group of functions is for matching two sets of mutational signatures. The match minimizes the total distance between paired signatures by using the ``Hungarian algorithm" described in:  Kuhn, H. W. (1955) <doi:10.1002 nav.3800020109="">.  'mSigTools' stands for mutational Signature analysis Tools.</doi:10.1002>
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<pre>URL https://github.com/Rozen-Lab/mSigTools</pre>
<pre>BugReports https://github.com/Rozen-Lab/mSigTools/issues Encoding UTF-8</pre>
Language en-US
RoxygenNote 7.2.1
Imports clue, philentropy
<b>Suggests</b> spelling, testthat (>= 3.0.0)
Config/testthat/edition 3
NeedsCompilation no
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R topics documented:
match_two_sig_sets

2 match\_two\_sig\_sets

Index		11
	write_exposure	10
	TP_FP_FN_avg_sim	9
	sort_exposure	8
	sig_dist_matrix	7

 ${\tt match\_two\_sig\_sets}$ 

Find an optimal matching between two sets of signatures subject to a maximum distance.

## **Description**

Find an optimal matching between two sets of signatures subject to a maximum distance.

## Usage

```
match_two_sig_sets(
   x1,
   x2,
   method = "cosine",
   convert.sim.to.dist = function(x) {
      return(1 - x)
   },
   cutoff = 0.9
)
```

#### **Arguments**

x1 A numerical-matrix-like object with columns as signatures.

A numerical-matrix-like object with columns as signatures. Needs to have the

same number of rows as x1.

method As for the distance function in package philenropy.

convert.sim.to.dist

If method specifies a similarity rather than a distance, use this function to convert

the similarity to a distance.

cutoff A maximum distance or minimum similarity over which to pair signatures be-

tween x1 and x2.

#### **Details**

Match signatures between x1 and x2 using the function solve\_LSAP, which uses the "Hungarian" (a.k.a "Kuhn-Munkres") algorithm https://en.wikipedia.org/wiki/Hungarian\_algorithm, which optimizes the total cost associated with the links between nodes. This function generates a distance matrix between the two sets of signatures using method and, if necessary, convert.sim.to.dist. It then sets distances > cutoff to very large values and then applies solve\_LSAP to the resulting matrix to compute a matching between x1 and x2 that minimizes the sum of the distances.

plot\_exposure 3

#### Value

A list with the elements

• table Table of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the distance of the match.

- orig.matrix The matrix of numeric distances between x1 and x2.
- modified.matrix The argument orig.matrix with distances > cutoff changed to very large values.

#### **Examples**

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2) colnames(ex.sigs) <- c("ex1", "ex2", "ex3") ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2) colnames(ref.sigs) <- c("ref1", "ref2") match_two_sig_sets(ex.sigs, ref.sigs, cutoff = .9)
```

plot\_exposure

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of samples.

#### **Description**

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of samples.

# Usage

```
plot_exposure(
   exposure,
   samples.per.line = 30,
   plot.proportion = FALSE,
   xlim = NULL,
   ylim = NULL,
   legend.x = NULL,
   legend.y = NULL,
   cex.legend = 0.9,
   cex.yaxis = 1,
   cex.xaxis = NULL,
   plot.sample.names = TRUE,
   yaxis.labels = NULL,
   ...
)
```

# Arguments

exposure

Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use sort\_exposure. Do not use column names that start with multiple

4 plot\_exposure

underscores. The exposures will often be mutation counts, but could also be e.g. mutations per megabase.

samples.per.line

Number of samples to show in each plot.

plot.proportion

Plot exposure proportions rather than counts.

xlim, ylim Limits for the x and y axis. If NULL(default), the function tries to do something

reasonable.

legend.x, legend.y

The x and y co-ordinates to be used to position the legend.

cex.legend A numerical value giving the amount by which legend plotting text and symbols

should be magnified relative to the default.

cex.yaxis A numerical value giving the amount by which y axis values should be magnified

relative to the default.

cex.xaxis A numerical value giving the amount by which x axis values should be magni-

fied relative to the default. If NULL(default), the function tries to do something

reasonable.

plot.sample.names

Whether to plot sample names below the x axis. Default is TRUE.

yaxis.labels User defined y axis labels to be plotted. If NULL(default), the function tries to do

something reasonable.

Other arguments passed to barplot. If ylab is not included, it defaults to a value depending on plot.proportion. If col is not supplied the function tries

to do something reasonable.

#### Value

An **invisible** list. The first element is a logical value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of the bar x-axis midpoints drawn, useful for adding to the graph.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
old.par <- par(mar = c(8, 5, 1, 1))
plot_exposure(exposure[, 1:30],
   main = "Liver-HCC exposure", cex.yaxis = 0.8,
   plot.proportion = TRUE
)
par(old.par)</pre>
```

plot\_exposure\_to\_pdf 5

## **Description**

Plot exposures in multiple plots to a single PDF file, with each plot showing exposures for a manageable number of samples.

# Usage

```
plot_exposure_to_pdf(
  exposure,
  file,
  mfrow = c(2, 1),
  mar = c(6, 4, 3, 2),
  oma = c(3, 2, 0, 2),
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
  ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
  plot.sample.names = TRUE,
  yaxis.labels = NULL,
 width = 8.2677,
  height = 11.6929,
)
```

## **Arguments**

exposure	Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use sort_exposure. Do not use column names that start with multiple underscores. The exposures will often be mutation counts, but could also be e.g. mutations per megabase.
file	The name of the PDF file to be produced.
mfrow	A vector of the form c(nr,nc). Subsequent figures will be drawn in an nr-by-nc array on the device by rows.
mar	A numerical vector of the form c(bottom,left,top,right) which gives the number of lines of margin to be specified on the four sides of the plot.
oma	A vector of the form c(bottom,left,top,right) giving the size of the outer margins in lines of text.
samples.per.li	ne

Number of samples to show in each plot.

6 read\_exposure

1				
Plot exposure proportions rather than counts.				
Limits for the x and y axis. If NULL(default), the function tries to do something reasonable.				
legend.x, legend.y				
The x and y co-ordinates to be used to position the legend.				
A numerical value giving the amount by which legend plotting text and symbols should be magnified relative to the default.				
A numerical value giving the amount by which y axis values should be magnified relative to the default.				
A numerical value giving the amount by which x axis values should be magnified relative to the default. If NULL(default), the function tries to do something reasonable.				
plot.sample.names				
Whether to plot sample names below the x axis. Default is TRUE.				
User defined y axis labels to be plotted. If NULL(default), the function tries to do something reasonable.				
The width and height of the graphics region in inches.				
Other arguments passed to barplot. If ylab is not included, it defaults to a value depending on plot.proportion. If col is not supplied the function tries to do something reasonable.				

#### Value

An **invisible** list. The first element is a logical value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of the bar x-axis midpoints drawn, useful for adding to the graph.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
plot_exposure_to_pdf(exposure,
   file = file.path(tempdir(), "Liver-HCC.exposure.pdf"),
   cex.yaxis = 0.8, plot.proportion = TRUE
)</pre>
```

read\_exposure

Read an exposure matrix from a file.

## **Description**

Read an exposure matrix from a file.

## Usage

```
read_exposure(file, check.names = FALSE)
```

sig\_dist\_matrix 7

#### **Arguments**

file File path to a CSV file containing an exposure matrix, i.e. the numbers of mu-

tations due to each mutational signature. Each row corresponds to a mutational signature an each column corresponds to a tumor or other biological sample.

check.names Passed to read.csv. **IMPORTANT**: If TRUE this will replace the double colon

in identifiers of the form <tumor\_type>::<sample\_id> with two periods (i.e. <tumor\_type>..<sample\_id>. If check.names is true, generate a warning if double

colons were present.

#### Value

Numerical matrix of exposures, with the same shape as the contents of file.

#### **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)</pre>
```

sig\_dist\_matrix

Compute a matrix of distances / similarities between two sets of signatures.

# Description

Compute a matrix of distances / similarities between two sets of signatures.

# Usage

```
sig_dist_matrix(x1, x2, method = "cosine")
```

# **Arguments**

x1 The first set of signatures (a numerical matrix-like object in which each column

is a signature).

x2 The second set of signatures, similar data type to x1, and must have the same

number of rows as x1.

method As for the distance function in package philenropy.

# Value

A numeric matrix with dimensions  $ncol(x1) \times ncol(x2)$ . Each element represents the distance or similarity (depending on method) between a column in x1 and a column in x2.

8 sort\_exposure

#### **Examples**

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.4, 0.6), nrow = 2) colnames(ex.sigs) <- c("ex1", "ex2", "ex3") ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2) colnames(ref.sigs) <- c("ref1", "ref2") sig_dist_matrix(ex.sigs, ref.sigs)
```

sort\_exposure

Sort columns of an exposure matrix based on the number of mutations in each sample (column).

## **Description**

Sort columns of an exposure matrix based on the number of mutations in each sample (column).

# Usage

```
sort_exposure(exposure, decreasing = TRUE)
```

#### **Arguments**

exposure

Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column

names are taken as the sample IDs.

decreasing

If TRUE, sort from largest to smallest.

#### Value

The original exposure with columns sorted.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
exposure.sorted <- sort_exposure(exposure)</pre>
```

TP\_FP\_FN\_avg\_sim

TP_FP_FN_avg_sim Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.
--

#### **Description**

Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.

#### Usage

```
TP_FP_FN_avg_sim(extracted.sigs, reference.sigs, similarity.cutoff = 0.9)
```

# **Arguments**

extracted.sigs Mutational signatures discovered by some analysis. A numerical-matrix-like object with columns as signatures.

reference.sigs A numerical-matrix-like object with columns as signatures. This matrix should contain the reference mutational signatures. For example, these might be from a synthetic data set or they could be from reference set of signatures, such as the signatures at the COSMIC mutational signatures web site. See CRAN package cosmicsig.

similarity.cutoff

A signature in reference.sigs must be matched by >= similarity.cutoff by a signature in extracted.sigs to be considered detected.

# **Details**

Match signatures in extracted.sigs to signatures in reference.sigs using match\_two\_sig\_sets based on cosine similarity.

#### Value

A list with the elements

- TP The number of true positive extracted signatures.
- FP The number of false positive extracted signatures.
- FN The number of false negative reference signatures.
- avg.cos.sim The average cosine similarity of true positives to their matching reference signatures.
- table A data frame of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the cosine similarity of the match
- sim.matrix The numeric distance or similarity matrix between extracted.sigs and reference.sigs as returned from sig\_dist\_matrix.
- unmatched.ex.sigs The identifiers of the extracted signatures that did not match a reference signature.
- unmatched.ref.sigs The identifiers of the reference signatures that did not match an extracted signature.

10 write\_exposure

#### **Examples**

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2)
colnames(ex.sigs) <- c("ex1", "ex2", "ex3")
ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2)
colnames(ref.sigs) <- c("ref1", "ref2")
TP_FP_FN_avg_sim(
    extracted.sigs = ex.sigs,
    reference.sigs = ref.sigs,
    similarity.cutoff = .9
)</pre>
```

write\_exposure

Write an exposure matrix to a file.

# Description

Write an exposure matrix to a file.

## Usage

```
write_exposure(exposure, file, row.names = TRUE)
```

# Arguments

exposure Exposures as a numerical matrix (or data.frame) with signatures in rows and

samples in columns. Rownames are taken as the signature names and column

names are taken as the sample IDs.

file File to which to write the exposure matrix (as a CSV file).

row.names Either a logical value indicating whether the row names of exposure are to be

written along with exposure, or a character vector of row names to be written.

#### Value

No return value, called for side effects.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
write_exposure(exposure, file = file.path(tempdir(), "Liver-HCC.exposure.csv"))</pre>
```

# **Index**

```
barplot, 4, 6

distance, 2, 7

match_two_sig_sets, 2, 9

plot_exposure, 3

plot_exposure_to_pdf, 5

read_exposure, 6

sig_dist_matrix, 7, 9

solve_LSAP, 2

sort_exposure, 3, 5, 8

TP_FP_FN_avg_sim, 9

write_exposure, 10
```