# Package 'mSigTools'

August 26, 2022

Type Package
Title Mutational Signature Analysis Tools
Version 1.0.3
Description Utility functions for mutational signature analysis.  This package provides two groups of functions. One is for dealing with mutational signature ``exposures" (i.e. the counts of mutations in a sample that are due to each mutational signature). The other group of functions is for matching two sets of mutational signatures. The match minimizes the total distance between paired signatures by using the ``Hungarian algorithm" described in:  Kuhn, H. W. (1955) <doi:10.1002 nav.3800020109="">.  'mSigTools' stands for mutational Signature analysis Tools.</doi:10.1002>
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 ${\tt match\_two\_sig\_sets}$ 

Find an optimal matching between two sets of signatures subject to a maximum distance.

## **Description**

Find an optimal matching between two sets of signatures subject to a maximum distance.

## Usage

```
match_two_sig_sets(
    x1,
    x2,
    method = "cosine",
    convert.sim.to.dist = function(x) {
        return(1 - x)
    },
    cutoff = 0.9
)
```

## **Arguments**

x1 A numerical-matrix-like object with columns as signatures.

A numerical-matrix-like object with columns as signatures. Needs to have the

same number of rows as x1.

method As for the distance function in package philenropy.

convert.sim.to.dist

If method specifies a similarity rather than a distance, use this function to convert

the similarity to a distance.

cutoff A maximum distance or minimum similarity over which to pair signatures be-

tween x1 and x2.

#### **Details**

Match signatures between x1 and x2 using the function solve\_LSAP, which uses the "Hungarian" (a.k.a "Kuhn-Munkres") algorithm https://en.wikipedia.org/wiki/Hungarian\_algorithm, which optimizes the total cost associated with the links between nodes. This function generates a distance matrix between the two sets of signatures using method and, if necessary, convert.sim.to.dist. It then sets distances > cutoff to very large values and then applies solve\_LSAP to the resulting matrix to compute a matching between x1 and x2 that minimizes the sum of the distances.

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#### Value

A list with the elements

• table Table of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the distance of the match.

- orig.matrix The distance matrix between x1 and x2.
- modified.matrix The orig.matrix with distances > cutoff changed to very large values.

## **Examples**

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2) colnames(ex.sigs) <- c("ex1", "ex2", "ex3") ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2) colnames(ref.sigs) <- c("ref1", "ref2") match_two_sig_sets(ex.sigs, ref.sigs, cutoff = .9)
```

plot\_exposure

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of samples.

#### **Description**

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of samples.

## Usage

```
plot_exposure(
  exposure,
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
  ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
  plot.sample.names = TRUE,
  yaxis.labels = NULL,
  ...
)
```

#### **Arguments**

exposure

Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use sort\_exposure. Do not use column names that start with multiple underscores. The exposures will often be mutation counts, but could also be e.g. mutations per megabase.

```
samples.per.line
                  Number of samples to show in each plot.
plot.proportion
                  Plot exposure proportions rather than counts.
xlim, ylim
                  Limits for the x and y axis. If NULL(default), the function tries to do something
                  reasonable.
legend.x, legend.y
                   The x and y co-ordinates to be used to position the legend.
cex.legend
                   A numerical value giving the amount by which legend plotting text and symbols
                  should be magnified relative to the default.
cex.yaxis
                   A numerical value giving the amount by which y axis values should be magnified
                  relative to the default.
                  A numerical value giving the amount by which x axis values should be magni-
cex.xaxis
                  fied relative to the default. If NULL(default), the function tries to do something
                  reasonable.
plot.sample.names
                   Whether to plot sample names below the x axis. Default is TRUE.
                  User defined y axis labels to be plotted. If NULL(default), the function tries to do
yaxis.labels
                  something reasonable.
                  Other arguments passed to barplot. If ylab is not included, it defaults to a
                  value depending on plot.proportion. If col is not supplied the function tries
                  to do something reasonable.
```

## Value

An **invisible** list whose first element is a logic value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of all the bar midpoints drawn, useful for adding to the graph.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
old.par <- par(mar = c(8, 5, 1, 1))
plot_exposure(exposure[, 1:30],
   main = "Liver-HCC exposure", cex.yaxis = 0.8,
   plot.proportion = TRUE
)
par(old.par)</pre>
```

## **Description**

Plot exposures in multiple plots to a single PDF file, with each plot showing exposures for a manageable number of samples.

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#### **Usage**

```
plot_exposure_to_pdf(
  exposure,
  file,
 mfrow = c(2, 1),
 mar = c(6, 4, 3, 2),
  oma = c(3, 2, 0, 2),
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
 ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
 plot.sample.names = TRUE,
 yaxis.labels = NULL,
 width = 8.2677,
 height = 11.6929,
)
```

## **Arguments**

exposure Exposures as a numerical matrix (or data.frame) with signatures in rows and

samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use sort\_exposure. Do not use column names that start with multiple underscores. The exposures will often be mutation counts, but could also be e.g.

mutations per megabase.

file The name of the PDF file to be produced.

mfrow A vector of the form c(nr,nc). Subsequent figures will be drawn in an nr-by-nc

array on the device by rows.

mar A numerical vector of the form c(bottom,left,top,right) which gives the

number of lines of margin to be specified on the four sides of the plot.

oma A vector of the form c(bottom,left,top,right) giving the size of the outer

margins in lines of text.

samples.per.line

Number of samples to show in each plot.

plot.proportion

Plot exposure proportions rather than counts.

xlim, ylim Limits for the x and y axis. If NULL(default), the function tries to do something

reasonable.

legend.x, legend.y

The x and y co-ordinates to be used to position the legend.

cex.legend A numerical value giving the amount by which legend plotting text and symbols

should be magnified relative to the default.

cex.yaxis A numerical value giving the amount by which y axis values should be magnified

relative to the default.

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cex.xaxis A numerical value giving the amount by which x axis values should be magni-

fied relative to the default. If NULL(default), the function tries to do something

reasonable.

plot.sample.names

Whether to plot sample names below the x axis. Default is TRUE.

yaxis.labels User defined y axis labels to be plotted. If NULL(default), the function tries to do

something reasonable.

width, height The width and height of the graphics region in inches.

.. Other arguments passed to barplot. If ylab is not included, it defaults to a

value depending on plot.proportion. If col is not supplied the function tries

to do something reasonable.

## Value

An **invisible** list whose first element is a logic value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of all the bar midpoints drawn, useful for adding to the graph.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
plot_exposure_to_pdf(exposure,
   file = file.path(tempdir(), "Liver-HCC.exposure.pdf"),
   cex.yaxis = 0.8, plot.proportion = TRUE
)</pre>
```

read\_exposure

Read an exposure matrix from a file.

## **Description**

Read an exposure matrix from a file.

# Usage

```
read_exposure(file, check.names = FALSE)
```

## **Arguments**

file File path to a CSV file containing an exposure matrix, i.e. the numbers of mu-

tations due to each mutational signature. Each row corresponds to a mutational signature an each column corresponds to a tumor or other biological sample.

check.names Passed to read.csv. **IMPORTANT**: If TRUE this will replace the double colon in identifiers of the form stymes types uses male, identifiers of the form stymes types uses male identifiers of the form stymes types used to the form stymes types used to the form stymes type uses male identifiers of the form stymes types used to the form of types used types used to the form of types used to the for

in identifiers of the form <tumor\_type>::<sample\_id> with two periods (i.e. <tumor\_type>..<sample\_id>. If check.names is true, generate a warning if double

colons were present.

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## Value

Numerical matrix of exposures, with the same shape as file.

# **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)</pre>
```

sig\_dist\_matrix

Compute a matrix of distances / similarities between two sets of signatures.

# **Description**

Compute a matrix of distances / similarities between two sets of signatures.

## Usage

```
sig_dist_matrix(x1, x2, method = "cosine")
```

# Arguments

x1	The first set of signatures (a numerical matrix-like object in which each column is a signature).
x2	The second set of signatures, similar data type to $x1$ , and must have the same number of rows as $x1$ .
method	As for the distance function in package philenropy.

## Value

A matrix with dimensions ncol(x1) X ncol(x2) with each element representing the distance or similarity (depending on method) between the column in x1 and a column in x2.

## **Examples**

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.4, 0.6), nrow = 2) colnames(ex.sigs) <- c("ex1", "ex2", "ex3") ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2) colnames(ref.sigs) <- c("ref1", "ref2") sig_dist_matrix(ex.sigs, ref.sigs)
```

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sort_exposure	Sort columns of an exposure matrix based on the number of mutations in each sample (column).

# Description

Sort columns of an exposure matrix based on the number of mutations in each sample (column).

## Usage

```
sort_exposure(exposure, decreasing = TRUE)
```

## **Arguments**

exposure Exposures as a numerical matrix (or data.frame) with signatures in rows and

samples in columns. Rownames are taken as the signature names and column

names are taken as the sample IDs.

decreasing If TRUE, sort from largest to smallest.

## Value

The original exposure with columns sorted.

# **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
exposure.sorted <- sort_exposure(exposure)</pre>
```

TP\_FP\_FN\_avg\_sim

Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.

## **Description**

Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.

# Usage

```
TP_FP_FN_avg_sim(extracted.sigs, reference.sigs, similarity.cutoff = 0.9)
```

TP\_FP\_FN\_avg\_sim

## **Arguments**

extracted.sigs Mutational signatures discovered by some analysis. A numerical-matrix-like object with columns as signatures.

reference.sigs A numerical-matrix-like object with columns as signatures. This matrix should contain the reference mutational signatures. For example, these might be from a synthetic data set or they could be from reference set of signatures, such as the signatures at the COSMIC mutational signatures web site. See CRAN package cosmicsig.

similarity.cutoff

A signature in reference.sigs must be matched by >= similarity.cutoff by a signature in extracted.sigs to be considered detected.

#### **Details**

Match signatures in extracted.sigs to signatures in reference.sigs using match\_two\_sig\_sets based on cosine similarity.

## Value

A list with the elements

- TP The number of true positive extracted signatures.
- FP The number of false positive extracted signatures.
- FN The number of false negative reference signatures.
- avg.cos.sim Average cosine similarity of true positives to their matching reference signatures
- table Table of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the cosine similarity of the match.
- sim.matrix The similarity matrix corresponding to the input signatures.
- unmatched.ex.sigs The identifiers of the extracted signatures that did not match a reference signature.
- unmatched.ref.sigs The identifiers of the reference signatures that did not match an extracted signature.

#### **Examples**

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2)
colnames(ex.sigs) <- c("ex1", "ex2", "ex3")
ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2)
colnames(ref.sigs) <- c("ref1", "ref2")
TP_FP_FN_avg_sim(
    extracted.sigs = ex.sigs,
    reference.sigs = ref.sigs,
    similarity.cutoff = .9
)</pre>
```

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	exposure
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Write an exposure matrix to a file.

## **Description**

Write an exposure matrix to a file.

## Usage

```
write_exposure(exposure, file, row.names = TRUE)
```

## **Arguments**

exposure Exposures as a numerical matrix (or data.frame) with signatures in rows and

samples in columns. Rownames are taken as the signature names and column

names are taken as the sample IDs.

file File to which to write the exposure matrix (as a CSV file).

row.names Either a logical value indicating whether the row names of exposure are to be

written along with exposure, or a character vector of row names to be written.

## Value

No return value, called for side effects.

## **Examples**

```
file <- system.file("extdata",
   "Liver-HCC.exposure.csv",
   package = "mSigTools"
)
exposure <- read_exposure(file)
write_exposure(exposure, file = file.path(tempdir(), "Liver-HCC.exposure.csv"))</pre>
```

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