

Lecture 10: Single-Cell RNA Sequencing

BIOINF3005/7160: Transcriptomics Applications

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Background

scRNA Protocols

Data Analysis

QC

Quantification

Normalisation

Clustering

DE Analysis

Trajectory Analysis

Spatial Transcriptomics

Background

Introduction

- scRNA-Seq is the 'latest and greatest' transcriptomic technique
- Previously all our analysis involved multiple cells per sample
- All were combined during tissue extraction, library preparation etc.
- Most experiments have **highly** heterogeneous cell populations, e.g.

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- Most experiments have **highly** heterogeneous cell populations, e.g.
 - Different regions of the brain contain highly specialised cells
 - The immune system is highly complex
 - Cancer samples have both infiltrating and tumour cells

Introduction

- If a gene is increased 2-fold in expression:
 - Is this 2-fold in 100% of cells?
 - Or is it 4-fold in 50% of cells?
 - Or is it down 2-fold in 25% and up 8-fold in 25% and unchanged in 50%?
- Changes in gene expression can be highly specific to individual cell-types
- In general, determining heterogeneity of our samples is challenging

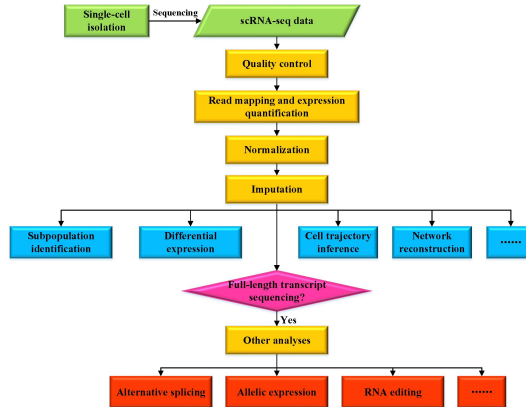
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- Reality is much trickier than this
- How do we characterise which cell is which cell-type?
- How do we capture as many transcripts from each cell as we can?
 - Missing values are a huge issue in scRNA-seq
- How do we compare within the same cell-types between experimental groups?
 - E.g., treated and untreated cell types may not be assigned to the same cluster/cell-type

Workflow Outline

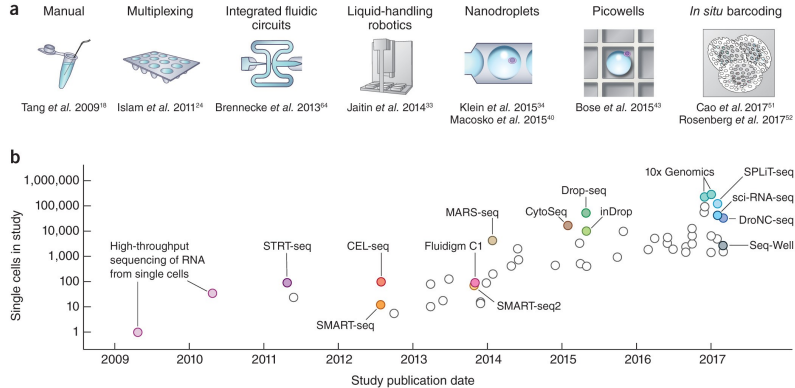


scRNA Protocols

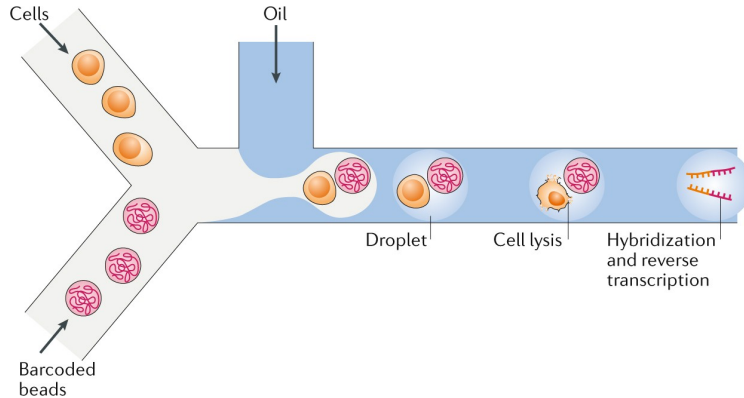
Isolating Individual Cells

- Early protocols used a dilution series or manual isolation with a microscope (*micromanipulation*)
- Laser Capture Micro-dissection (LCM)
- Fluorescence-Activated Cell Sorting (FACS)
 - Labelled antibodies to specific surface markers
 - MACS is a magnetic-based approach
- Microfluidics/Droplet-based approaches

Protocol Timeline



Droplet-based Approaches



Data Analysis

Spatial Transcriptomics