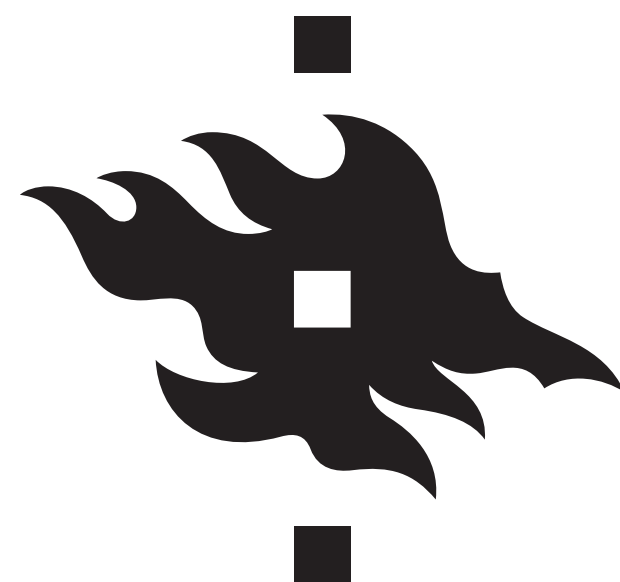


Genome Assembly

From theory to practice (and back)



UNIVERSITY OF HELSINKI

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Lecture outline

1. The problem
2. Practical issues
3. Theoretical problem formulations
4. Practical genome assembly
 - Contig assembly
 - Scaffolding
 - Gap filling
5. And back: a more "practical" theoretical formulation

LECTURE
Theory + Abstract view



ASSIGNMENT
Practical + Hands-on view

The problem

(A general description of the input and output)

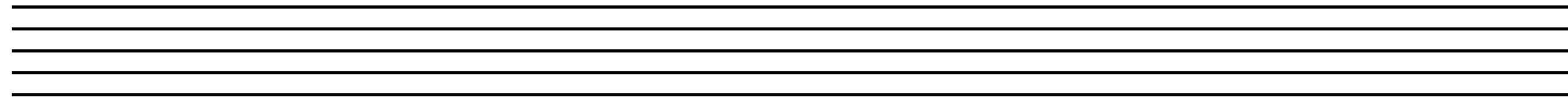
Short-read sequencing

(Second-generation sequencing)

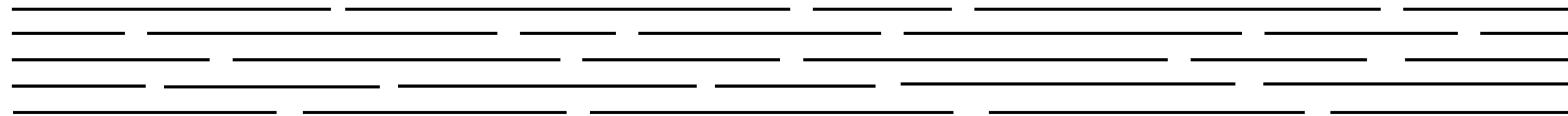
DNA



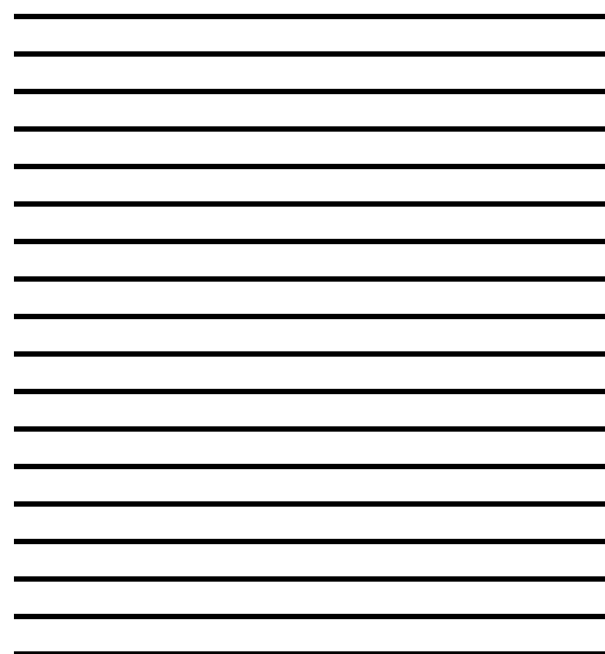
Amplification



Fragmentation

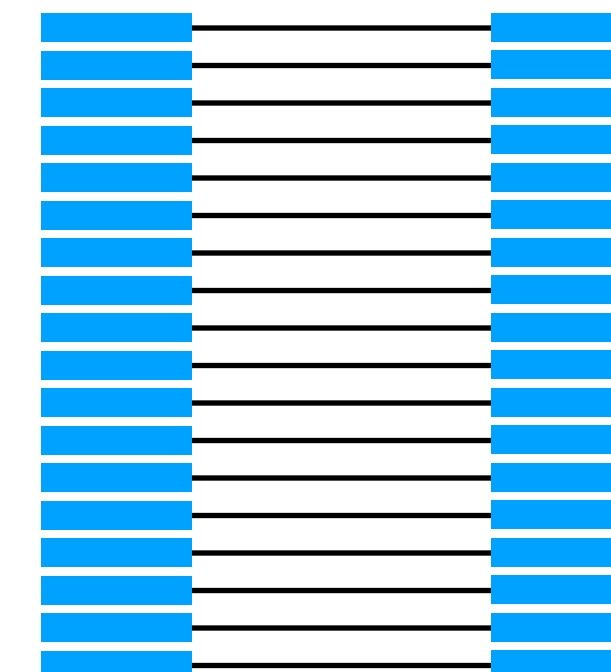
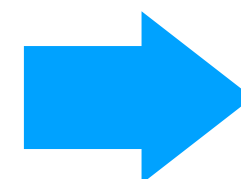


Size selection



length ~450

Sequencing



length 100 ~250 100

The assembly problem

INPUT: A collection of paired-end reads

OUTPUT: The genome from which they were sequenced

(We will see precise computational formulations later)

The assembly problem

INPUT: A collection of paired-end reads

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(We will see precise computational formulations later)

ACTAGTGCTAGATGCTCGAGCTAGCTAGCT

The assembly problem

INPUT: A collection of paired-end reads

OUTPUT: The genome from which they were sequenced

(We will see precise computational formulations later)

The diagram illustrates the assembly problem using paired-end reads. It shows a collection of reads of varying lengths, each represented by a sequence of nucleotides (A, C, G, T) and dots indicating gaps. The reads are arranged in a way that suggests they are overlapping and can be used to reconstruct a longer genome sequence. At the bottom, the reconstructed genome sequence is shown in blue text: **ACTAGTGCTAGATGCTCGAGCTAGCTAGCT**.

ACTAGTGCTAGATGCTCGAGCTAGCTAGCT

The assembly problem

INPUT: A collection of paired-end reads

OUTPUT: The genome from which they were sequenced

(We will see precise computational formulations later)

T	G	C	T	C	C	T	A	G	C
T	G	C	T	C	C	T	A	G	C
T	G	C	T	A	G	G	A	G	C
C	T	A	G	T	G	C	T	C	G
A	T	G	C	T	A	G	C	T	A
G	A	T	G	C	T	C	G	A	G
A	G	A	T	G	T	A	G	C	T
T	A	G	A	T	C	T	A	G	C
T	T	G	C	T	C	T	A	G	C
T	T	G	C	T	G	A	G	C	T
G	T	G	C	T	C	G	A	G	C
G	T	G	C	T	C	G	A	G	C
C	T	A	G	T	T	A	G	C	T
A	C	T	A	G	G	T	C	C	G
					T	G	C	T	A

???

Practical issues

(several of which are not covered in this course)

Repeats longer than read length

MAIN TOPIC

- If every substring of the genome of length = **read length - 1** is unique \rightarrow trivial

AATTGAATTACACCAC

AATTG

ATTGA

TGAAT

TGAAT

GAATT

AATTT

ATTTA

TTTAC

TTACA

TACAC

ACACC

CACCA

ACCAC

read length - 1

Repeats longer than read length

MAIN TOPIC

- If every substring of the genome of length = **read length - 1** is unique \rightarrow trivial
- Otherwise \rightarrow ambiguity

Repeats longer than read length

MAIN TOPIC

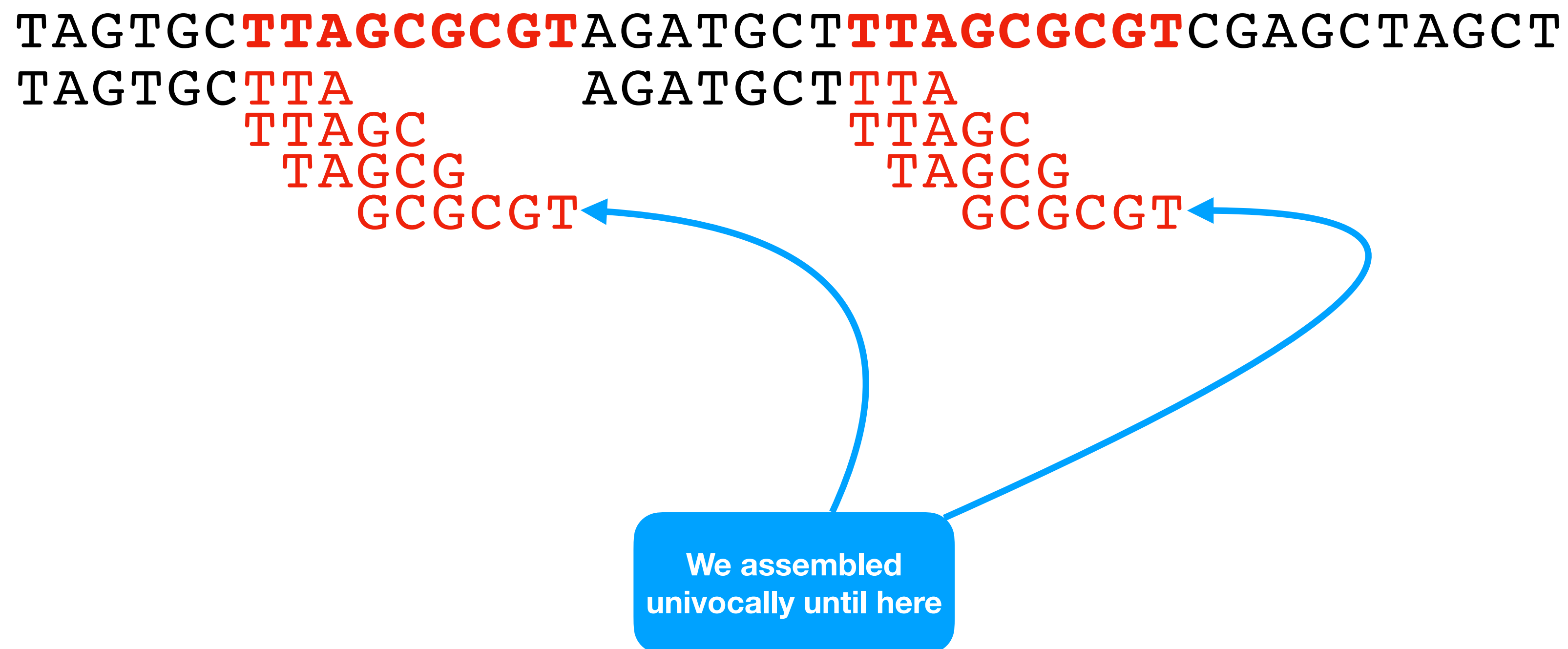
- If every substring of the genome of length = **read length - 1** is unique → trivial
- Otherwise → ambiguity

TAGTGCT**TTAGCGCGT**AGATGCT**TTAGCGCGT**CGAGCTAGCT
TAGTGCT**TTA**AGATGCT**TTA**
TAGTGCT**TTAGC**
TAGTGCT**TAGCG**
TAGTGCT**GCGCGT**

Repeats longer than read length

MAIN TOPIC

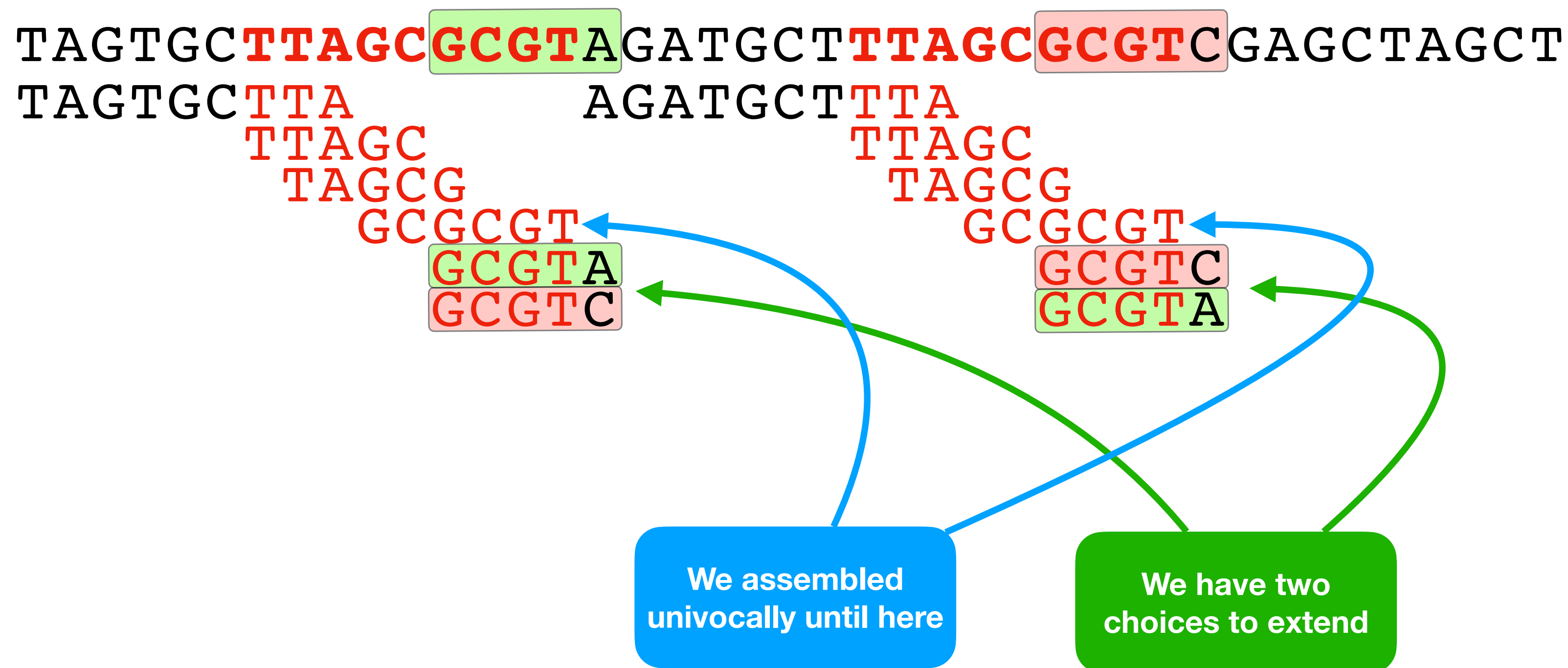
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Repeats longer than read length

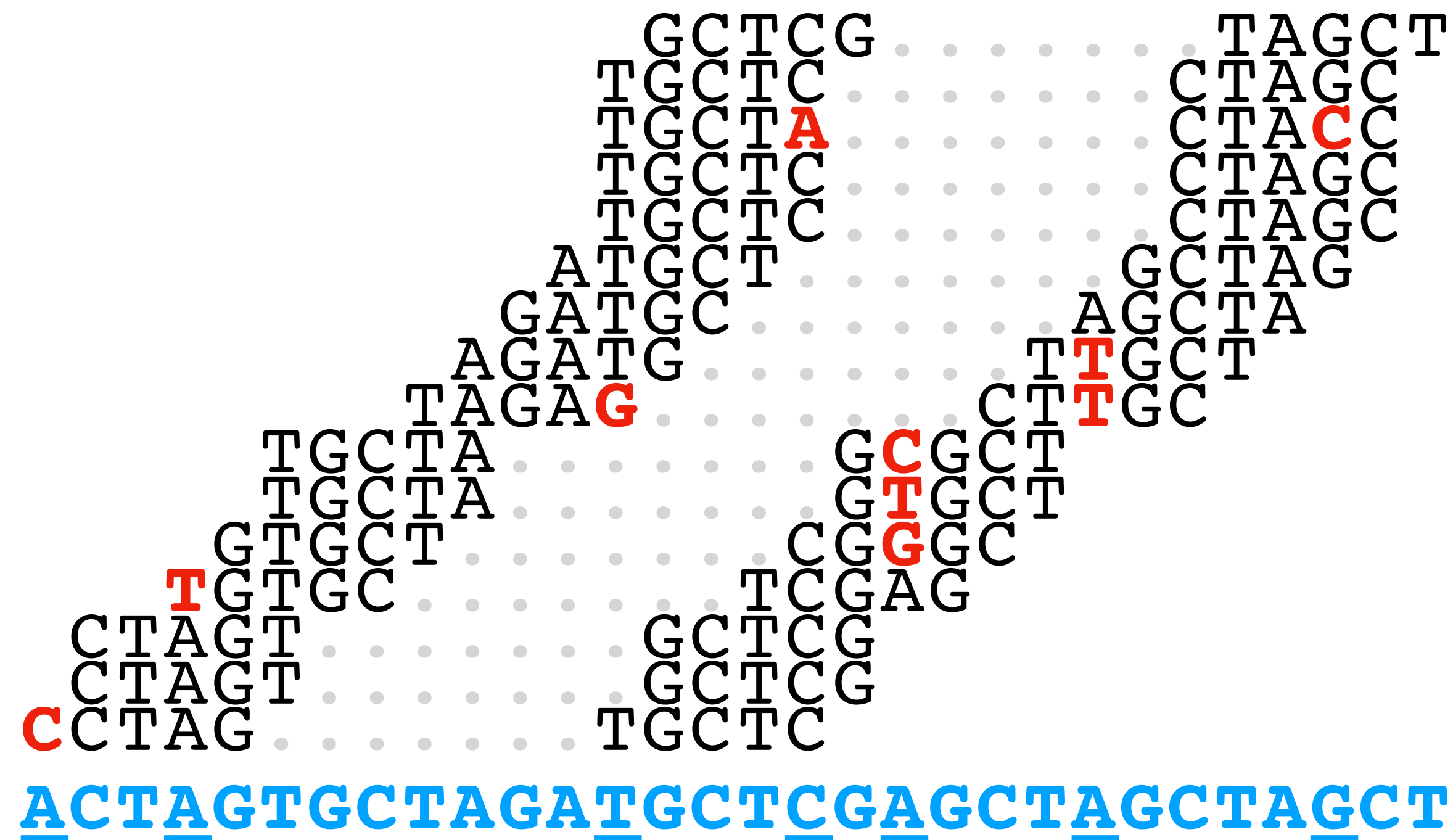
MAIN TOPIC

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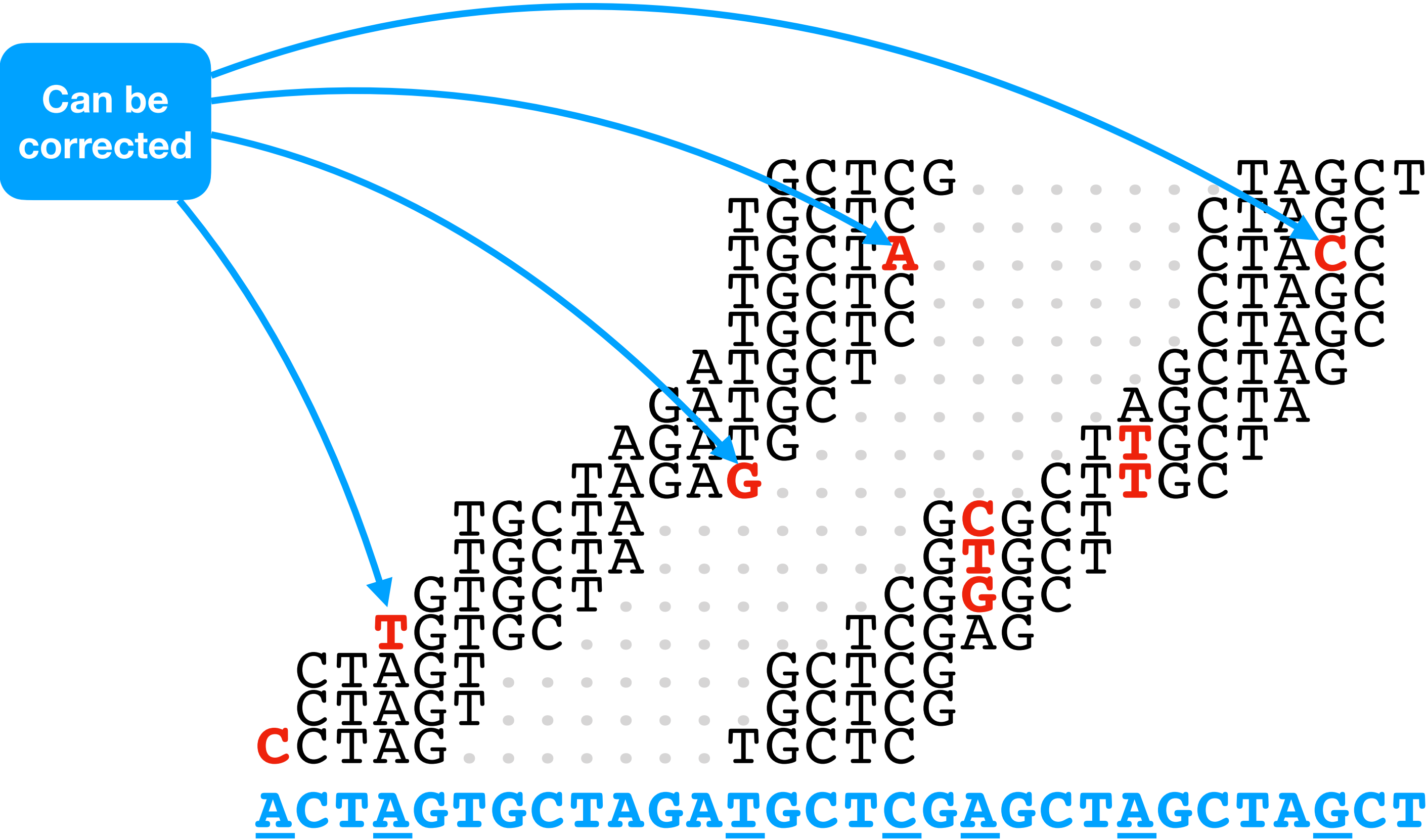
Sequencing errors

ASSIGNMENT



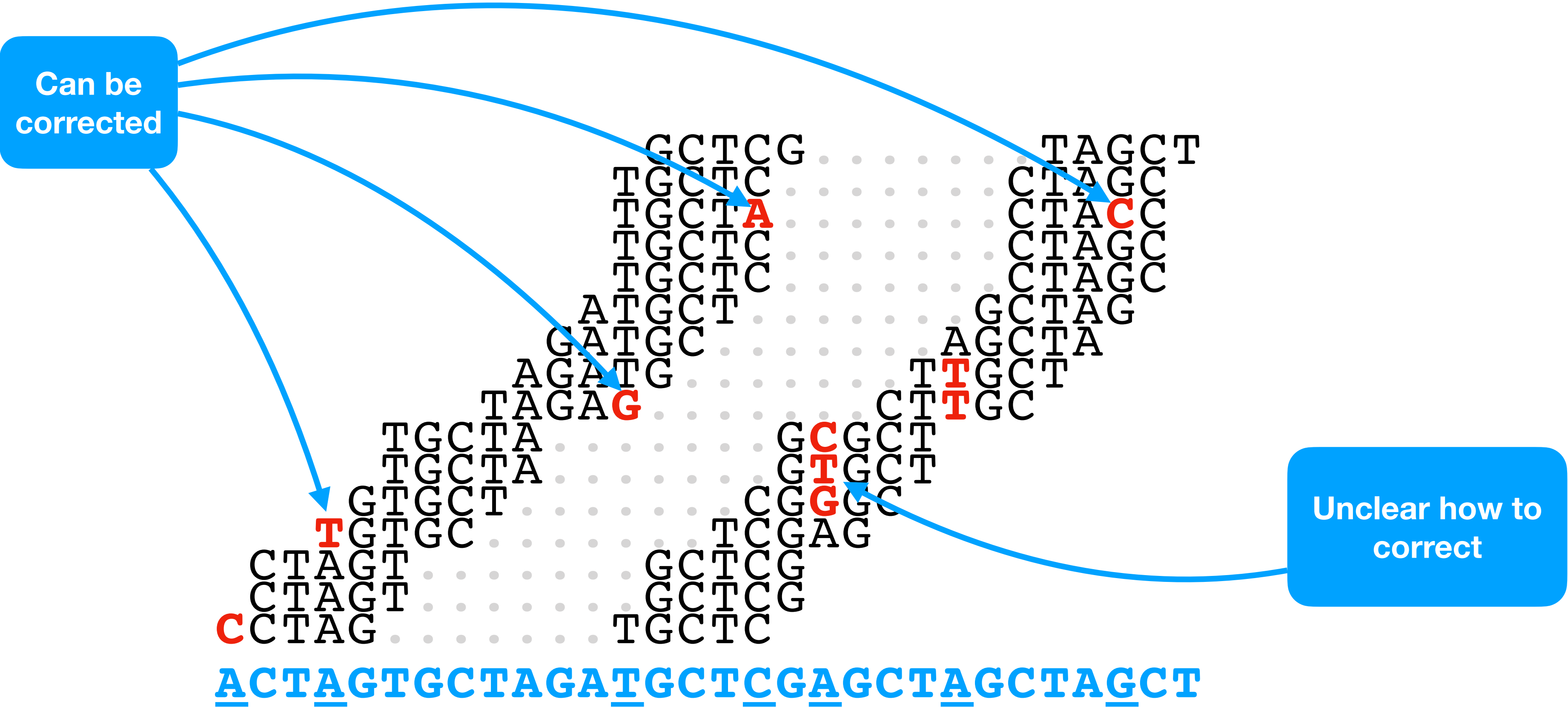
Sequencing errors

ASSIGNMENT

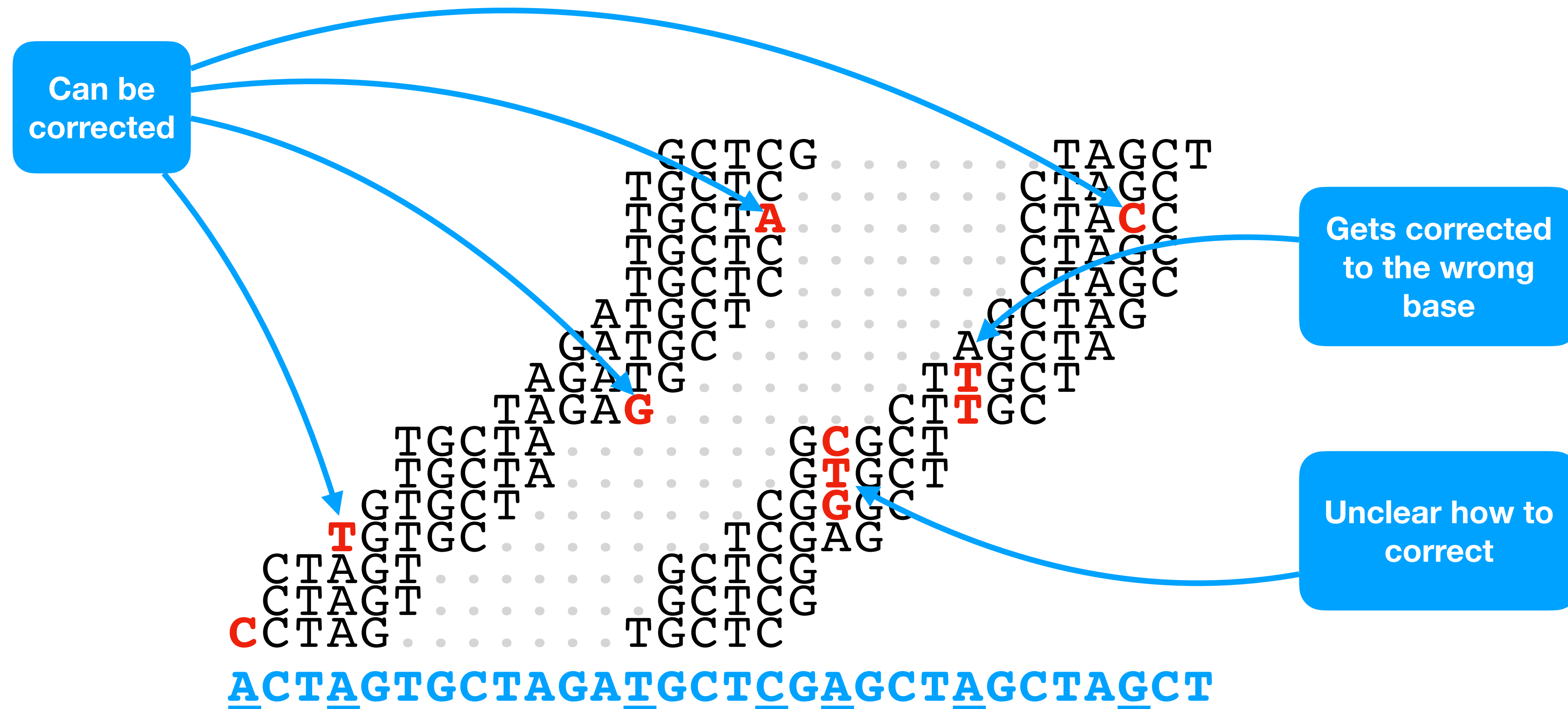


Sequencing errors

ASSIGNMENT

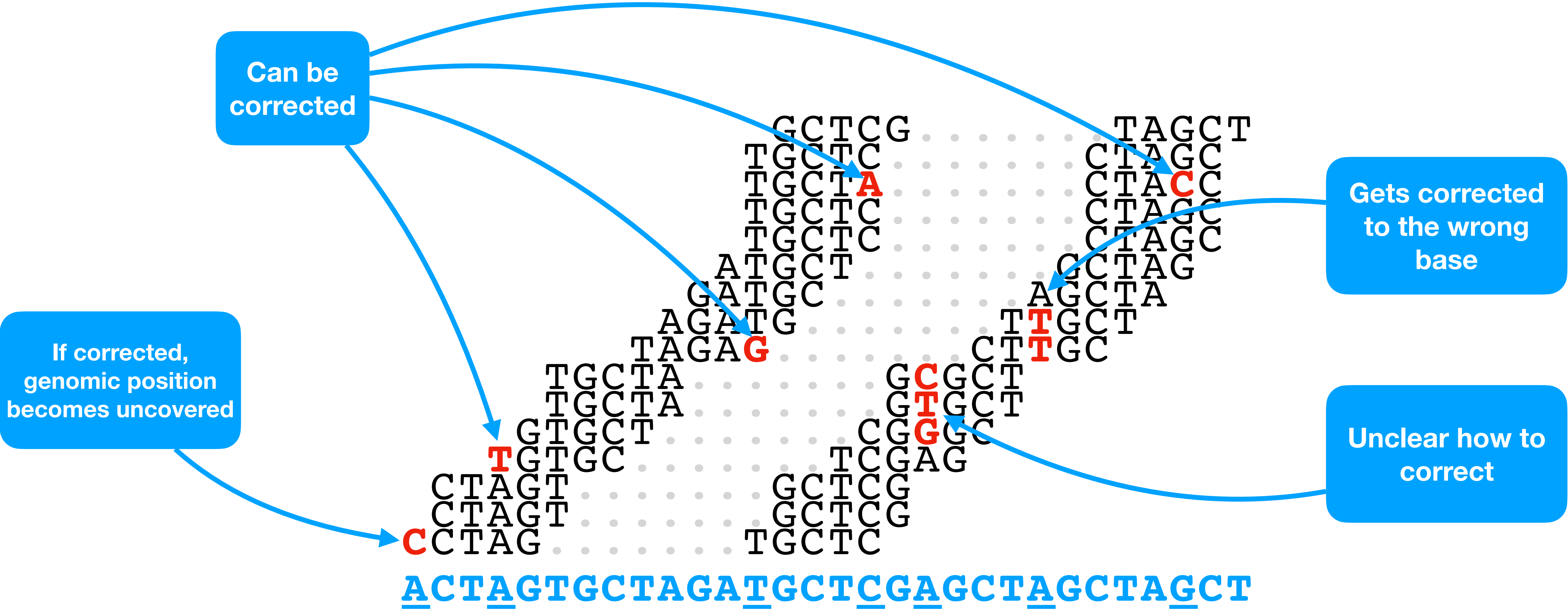


ASSIGNMENT



Sequencing errors

ASSIGNMENT



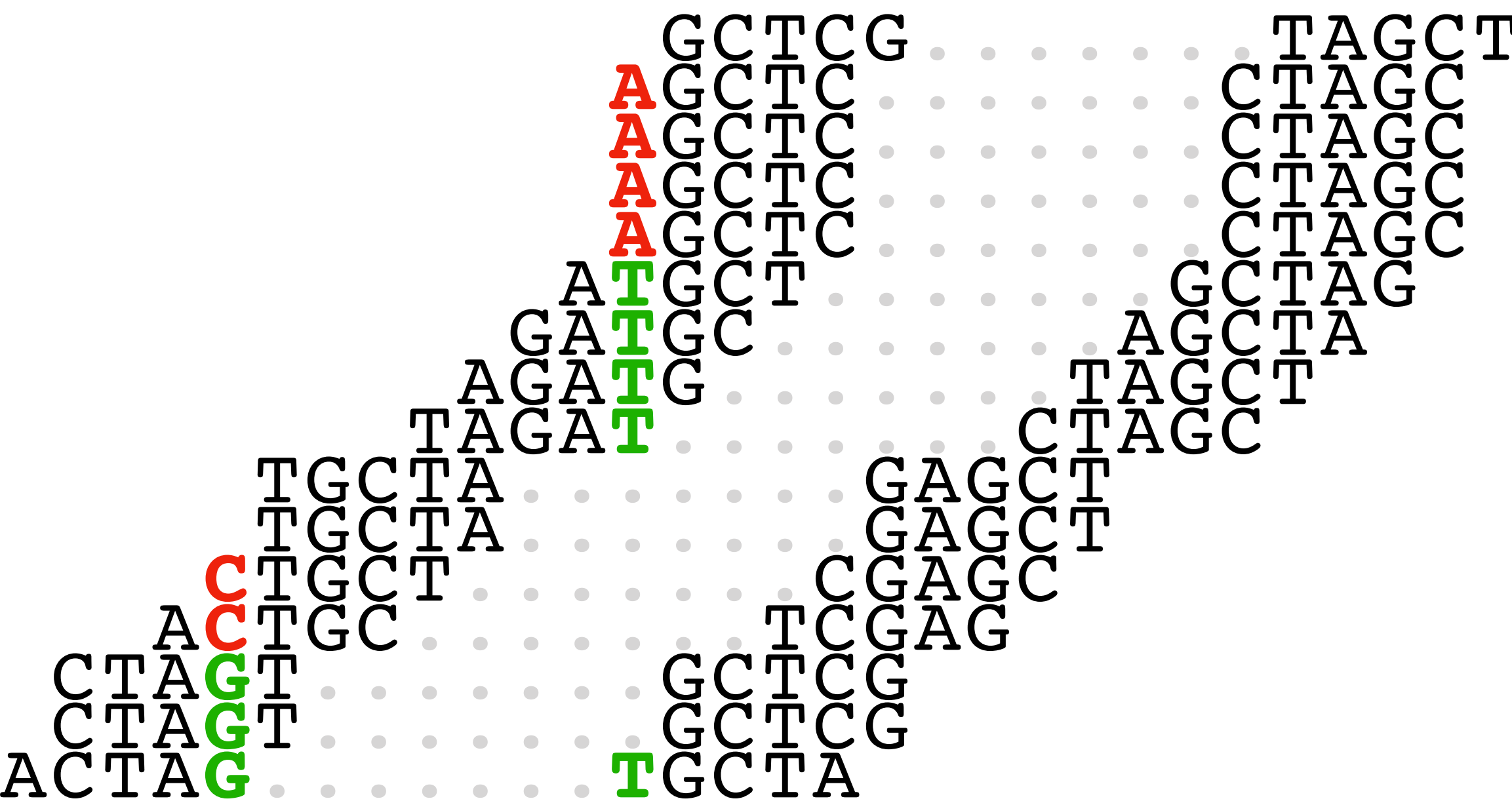
Polyploidy

LATER IN THE LECTURE

Mother	ACTA C TGCTAGAA A GCTCGAGCTAGCTAGCT
Father	ACTA G TGCTAGAT T GCTCGAGCTAGCTAGCT

Polyploidy

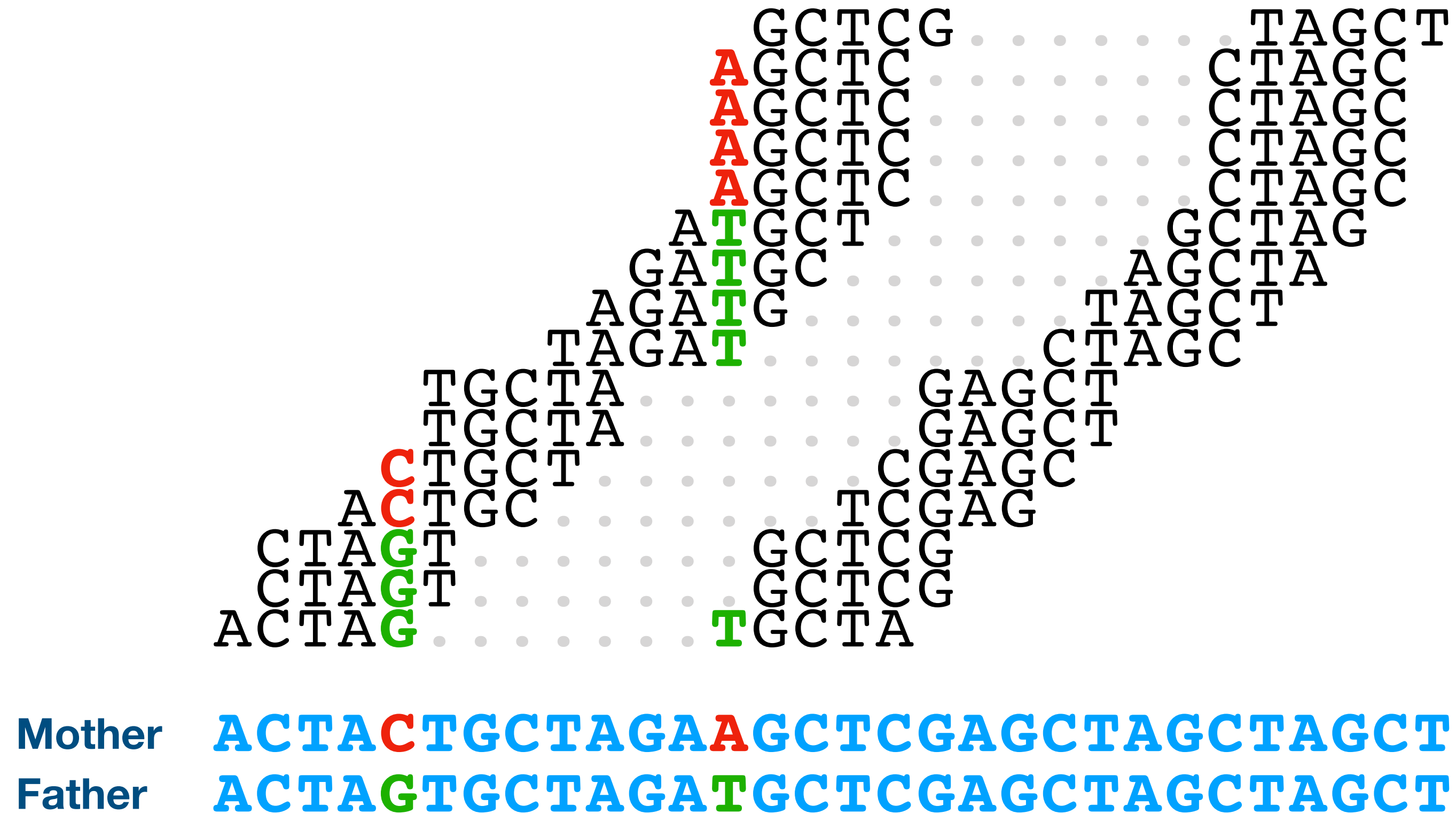
LATER IN THE LECTURE



Mother **ACTA****C****TGCTAGAA****A****GCTCGAGCTAGCTAGCT**
Father **ACTA****G****TGCTAGAT****G****GCTCGAGCTAGCTAGCT**

Polyploidy

LATER IN THE LECTURE



- Sequencing errors + polyploidy at the same time
- Phasing SNPs (**C** and **A** from same haplotype, **NOT** e.g. **C** and **T**) is a separate problem, called *haplotype assembly* or *haplotype phasing*

Unsequenced areas

T
A
G
A
T
G
G
G
C
C
T
T
C
G

C
T
A
G
C
C
C
C
C
C
T
T
A
G
G
C
C
T

C T A G T G C T C G
 C T A G T G C T C G
 A C T A G T G C T A
ACTAGT . . TAGATGCTCG . . CTAGCTAGCT

Non uniform paired-end distance

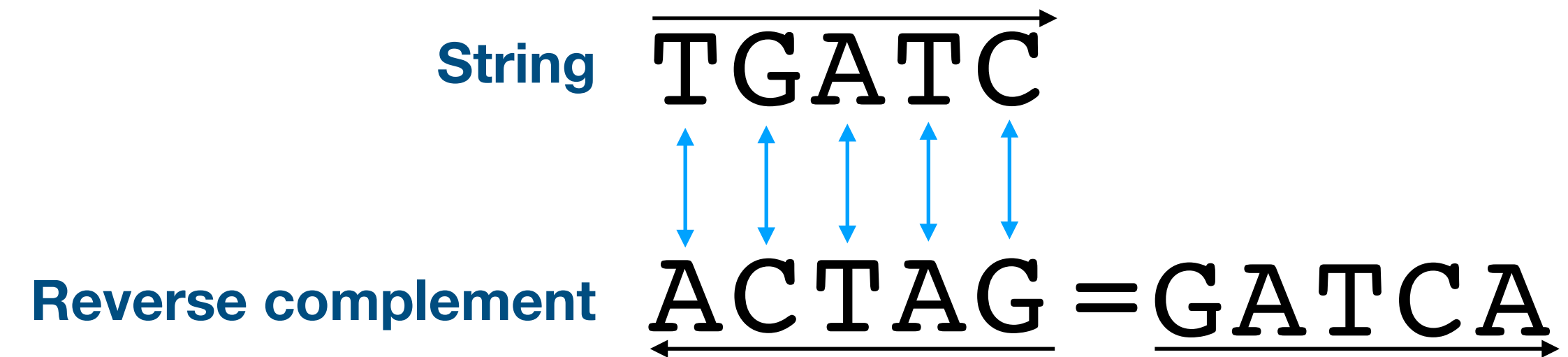
Diagram illustrating the process of transcription. The DNA double helix is shown with a template strand (3' to 5') and a coding strand (5' to 3'). A single-strand binding protein (SSB) is bound to the template strand. RNA polymerase (RNAP) is shown transcribing the template strand into a complementary RNA strand (5' to 3'). The RNA sequence shown is 5'-ACTAGTGCTAGATGCTCGAGCTAGCTAGCT-3'.

- Distance between each pair not known precisely from the sequencer

Double-stranded DNA

NOT IN THE LECTURE

- Reads consist of strings and their reverse complements:



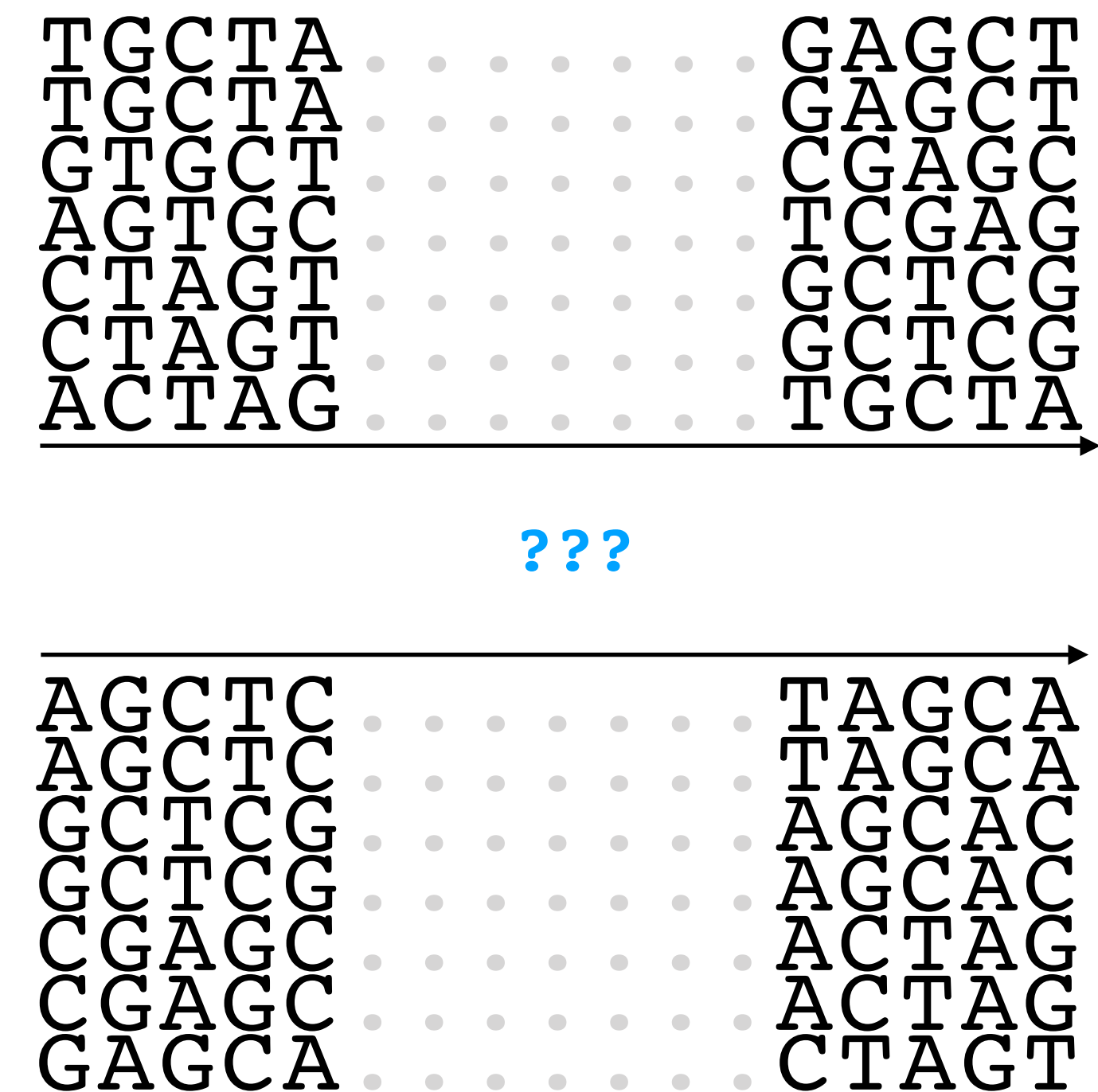
NOT IN THE LECTURE

-
- String **TGATC**
- lement **ACTAG = GATCA**



NOT IN THE LECTURE

-
- String **TGATC**
- Reverse complement **ACTAG = GATCA**



Large amount of data

NOT IN THE LECTURE

	Genome length	Total bases at 30x coverage	Size if each base takes 2 bits
E. coli	$4.6 \cdot 10^6$	$138 \cdot 10^6$	34 MBytes
Human	$3.2 \cdot 10^9$	$96 \cdot 10^9$	24 GBytes
Spruce	$25 \cdot 10^9$	$750 \cdot 10^9$	187.5 GBytes
Axolotl	$32 \cdot 10^9$	$960 \cdot 10^9$	240 GBytes

Theoretical problem formulations

("Classical" computational formulations of how to
obtain the output from the input)

Shortest superstring

INPUT: A collection of strings (the reads)

OUTPUT: A string S such that every given string is a substring of S (S is a *superstring*),
and S is shortest

Shortest superstring

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OUTPUT: A string S such that every given string is a substring of S (S is a *superstring*), and S is shortest

TAGA
ATAG
CATA
TCAT

Input

TCATAGA

Output S

Shortest superstring

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TAGA
ATAG
CATA
TCAT

Input

TAGA
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TCAT
TCATAGA

Output S

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TAGA	TAGA
ATAG	ATAG
CATA	CATA
TCAT	TCAT
	TCATAGA
Input	Output S

- NP-hard to compute (i.e. it cannot be solved efficiently)
- Not practical: it collapses repeats (main drawback)

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TCAT	TCAT
	TCATAGA
Input	Output S

- NP-hard to compute (i.e. it cannot be solved efficiently)
- Not practical: it collapses repeats (main drawback)

The genome [**TCATATATAGA**

The reads [TCAT
CATA
ATAT
TATA
ATAT
TATA
ATAG
TAGA

Shortest superstring

INPUT: A collection of strings (the reads)

OUTPUT: A string S such that every given string is a substring of S (S is a *superstring*), and S is shortest

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ATAG	ATAG
CATA	CATA
TCAT	TCAT
	TCATAGA
Input	Output S

- NP-hard to compute (i.e. it cannot be solved efficiently)
- Not practical: it collapses repeats (main drawback)

The genome	[TCATATAGA	The shortest superstring S	[TCATATAGA	
The reads	[TCAT]	The input	[TCAT
		CATA				CATA
		ATAT				ATAT
		TATA				TATA
		ATAT				ATAT
		TATA				TATA
		ATAG				ATAG
		TAGA				TAGA

Overlap graphs + Hamiltonian path

ACTAGACTAGACC

ACTA
CTAG
TAGA
AGAC
GACT
ACTA
CTAG
TAGA
AGAC
GACC

Overlap graphs + Hamiltonian path

ACTAGACTAGACC
ACTA
CTAG
TAGA
AGAC
GACT
ACTA
CTAG
TAGA
AGAC
GACC

INPUT: Overlap graph of order t :

- Every read is a node
- Every suffix-prefix overlap of length $\geq t$ is an edge

OUTPUT: A path going through every node (i.e. read) exactly one (*Hamiltonian*)

Overlap graphs + Hamiltonian path

ACTAGACTAGACC

ACTA
CTAG
TAGA
AGAC
GACT
ACTA
CTAG
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GACC

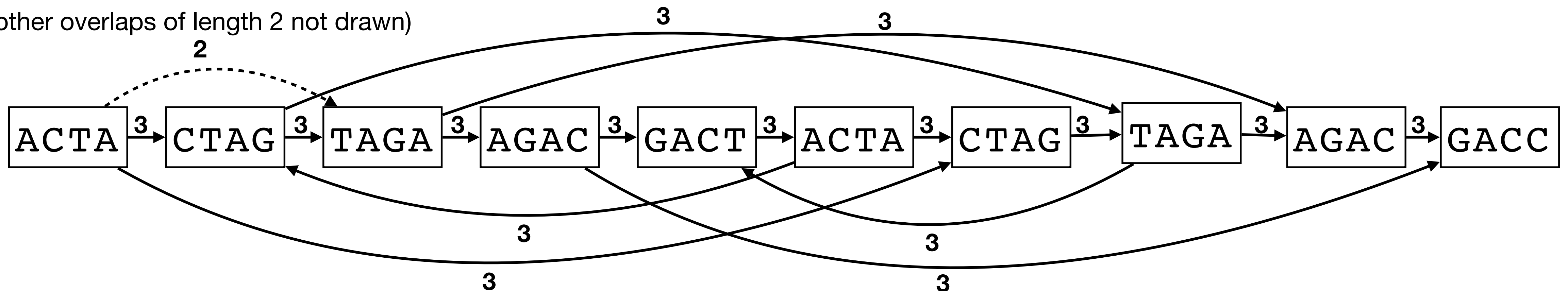
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Overlap graph of order 2:

(other overlaps of length 2 not drawn)



Overlap graphs + Hamiltonian path

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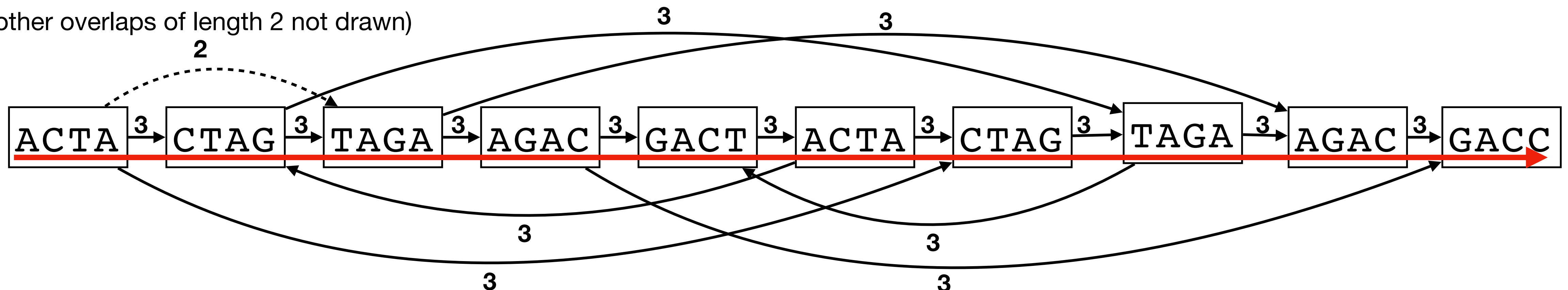
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Overlap graphs + Hamiltonian path

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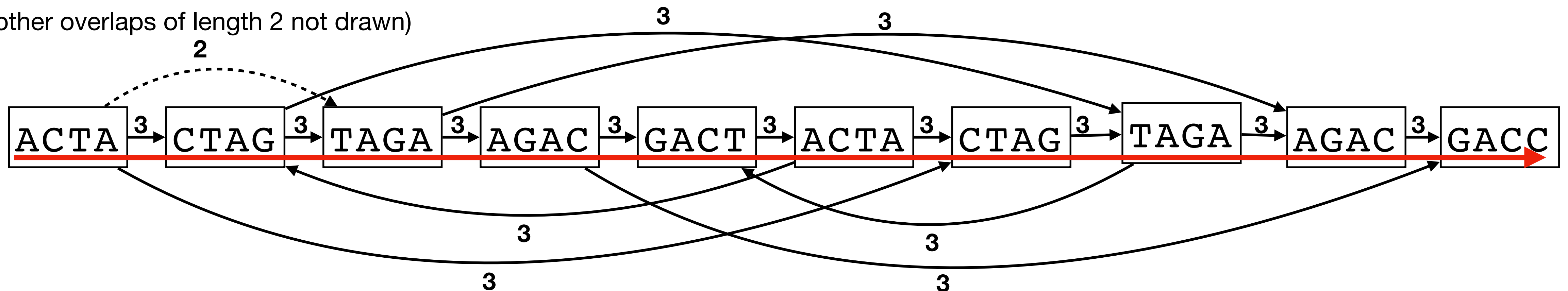
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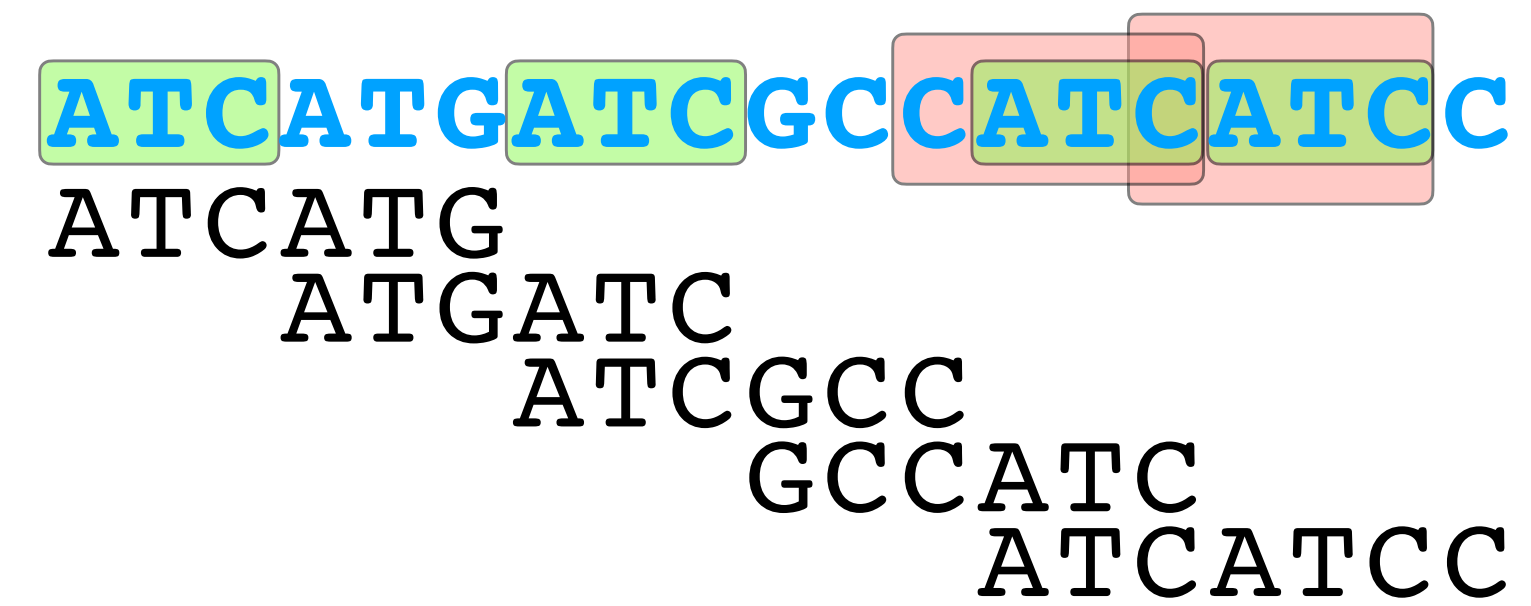
- NP-hard to compute
- Not practical: usually graph has no Hamiltonian path (missing coverage, errors)

Overlap graph of order 2:

(other overlaps of length 2 not drawn)



De Bruijn graphs + Eulerian path



De Bruijn graphs + Eulerian path



INPUT: De Bruijn graph of order k :

- Every k -mer (substring of length k) in the reads is a ***single*** node
- Every $(k + 1)$ -mer is a ***different*** arc from its length- k prefix to its length- k suffix

ASSUMPTION: Every length- $(k + 1)$ interval of the genome appears the same number of times in the reads (*uniform coverage*)

OUTPUT: A path going through every ***edge*** (i.e. $(k + 1)$ -mer) exactly one (*Eulerian*)

De Bruijn graphs + Eulerian path

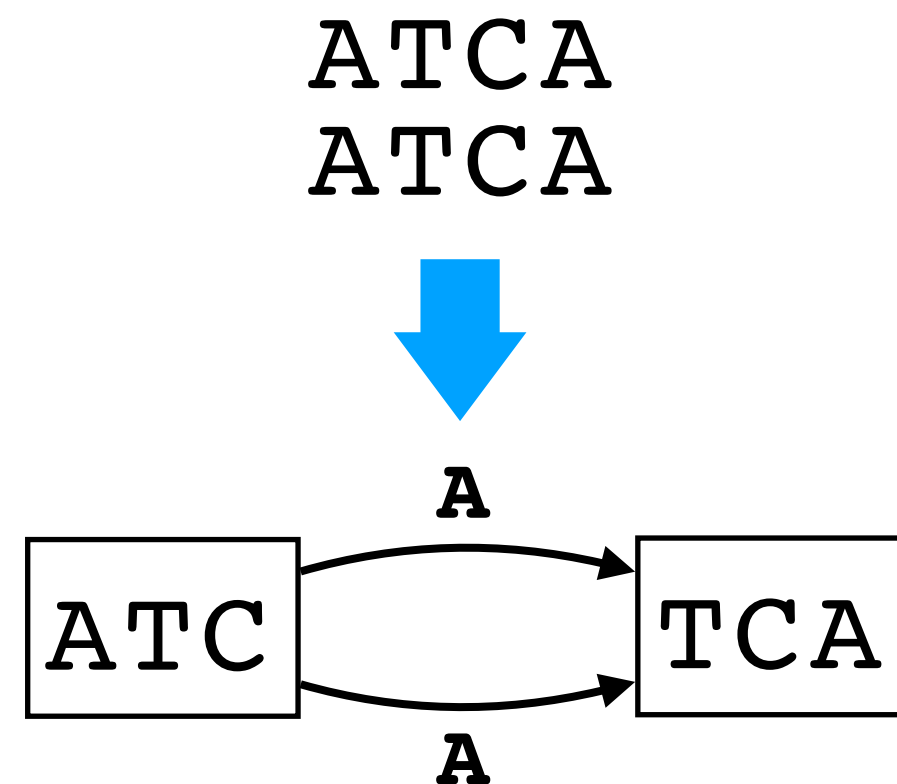


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De Bruijn graphs + Eulerian path



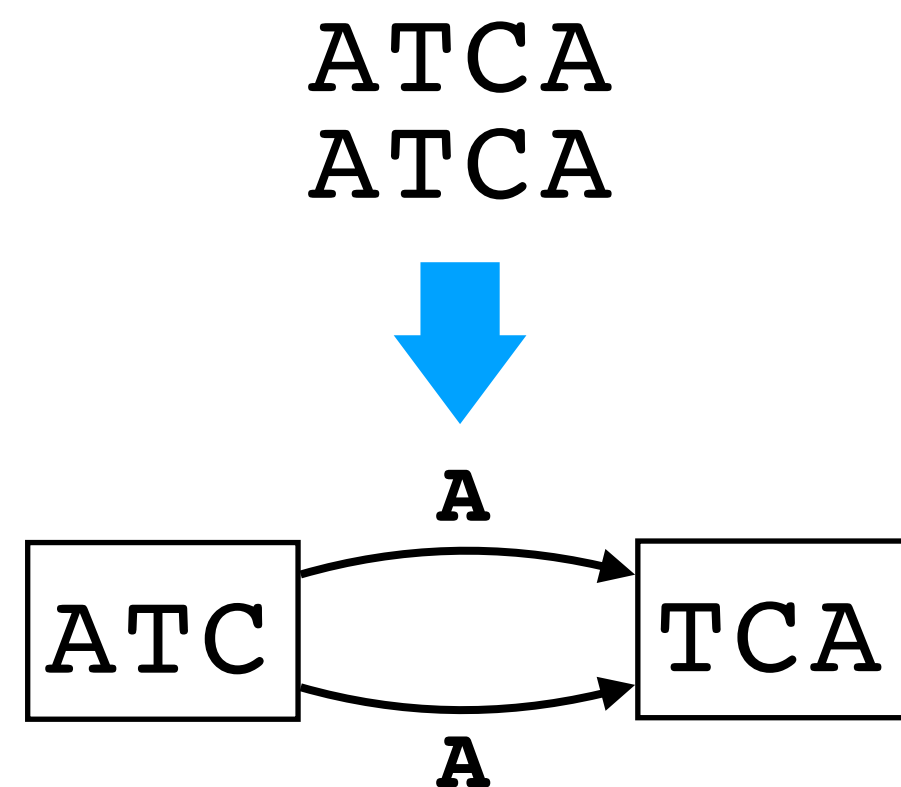
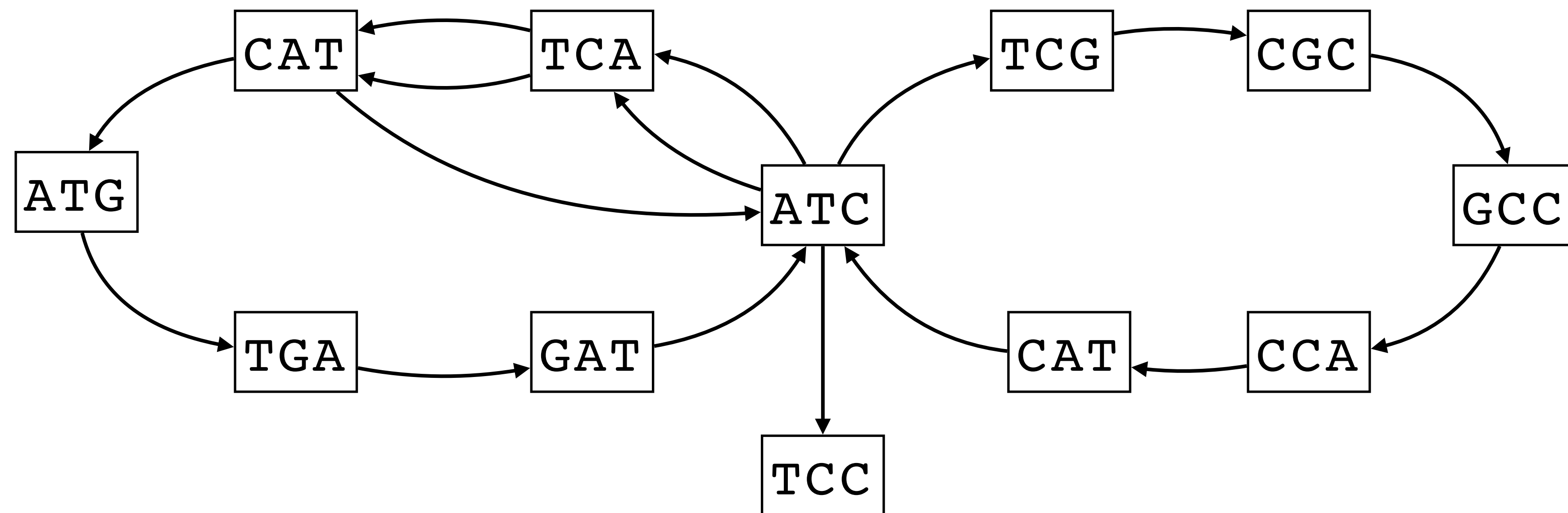
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De Bruijn graph of order 3:



De Bruijn graphs + Eulerian path



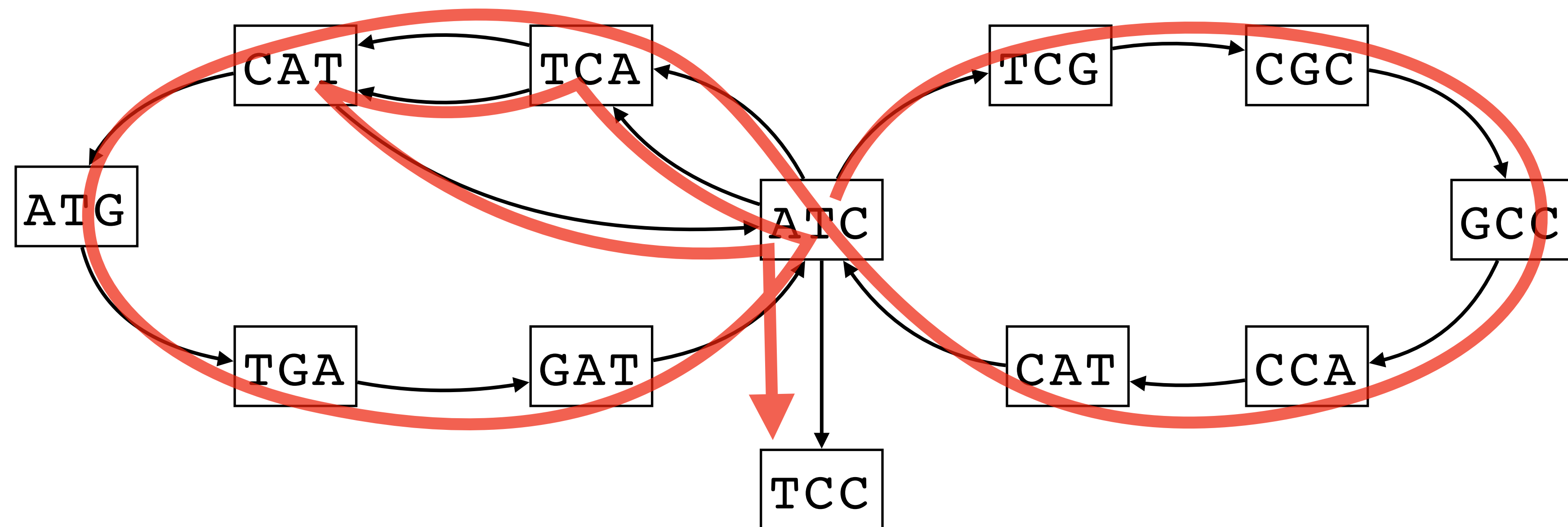
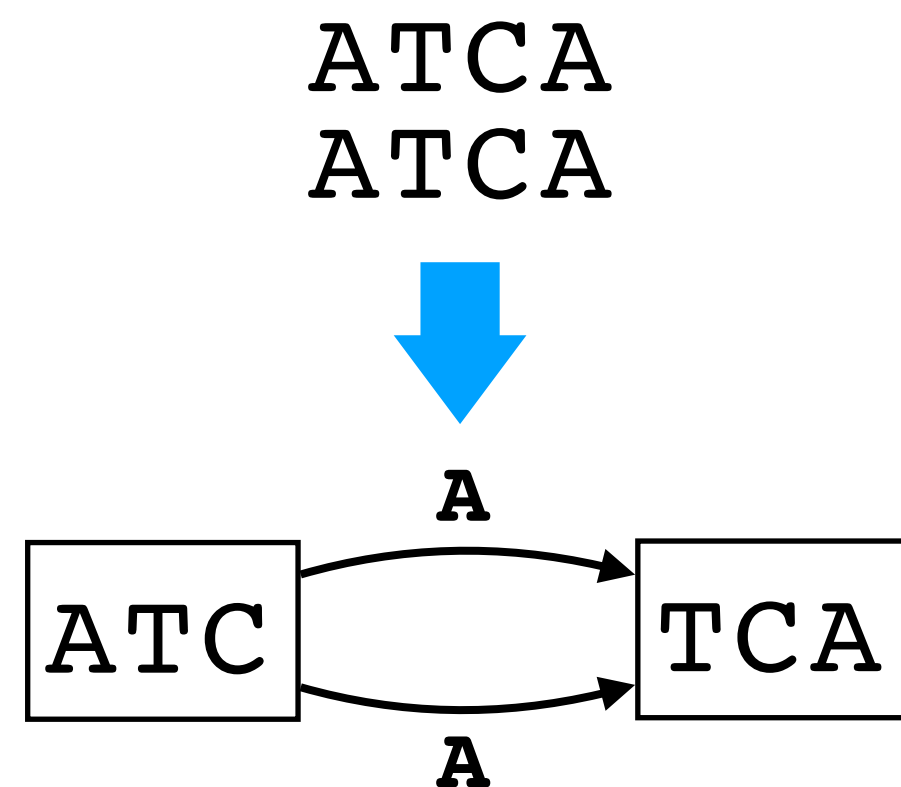
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De Bruijn graph of order 3:



De Bruijn graphs + Eulerian path



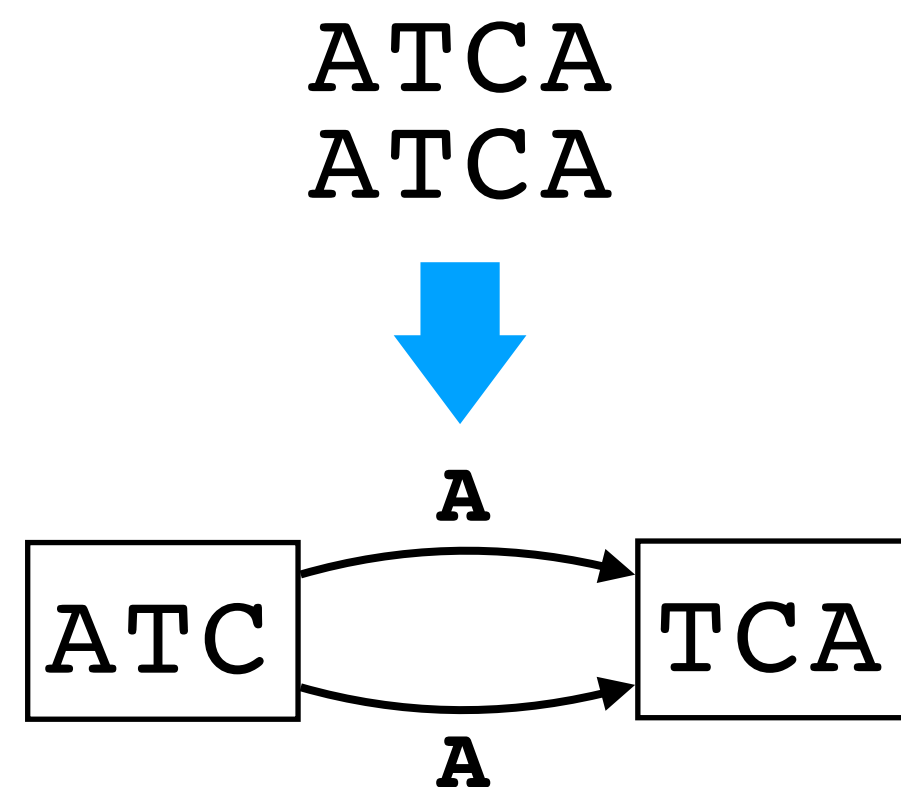
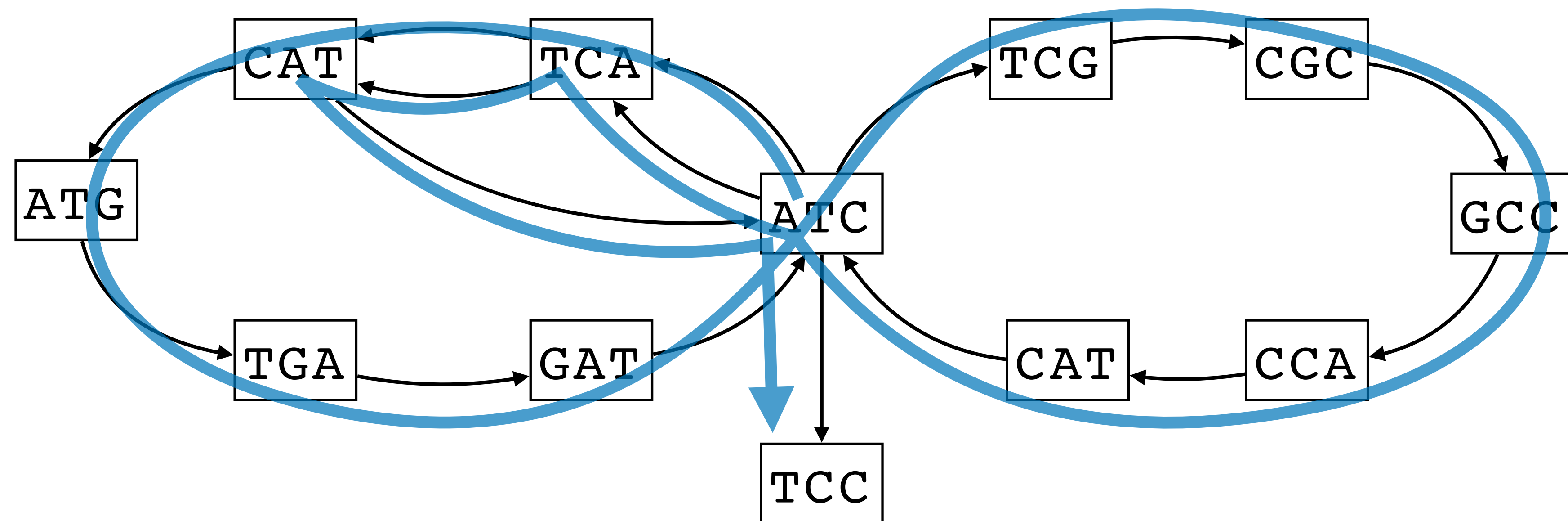
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De Bruijn graphs + Eulerian path



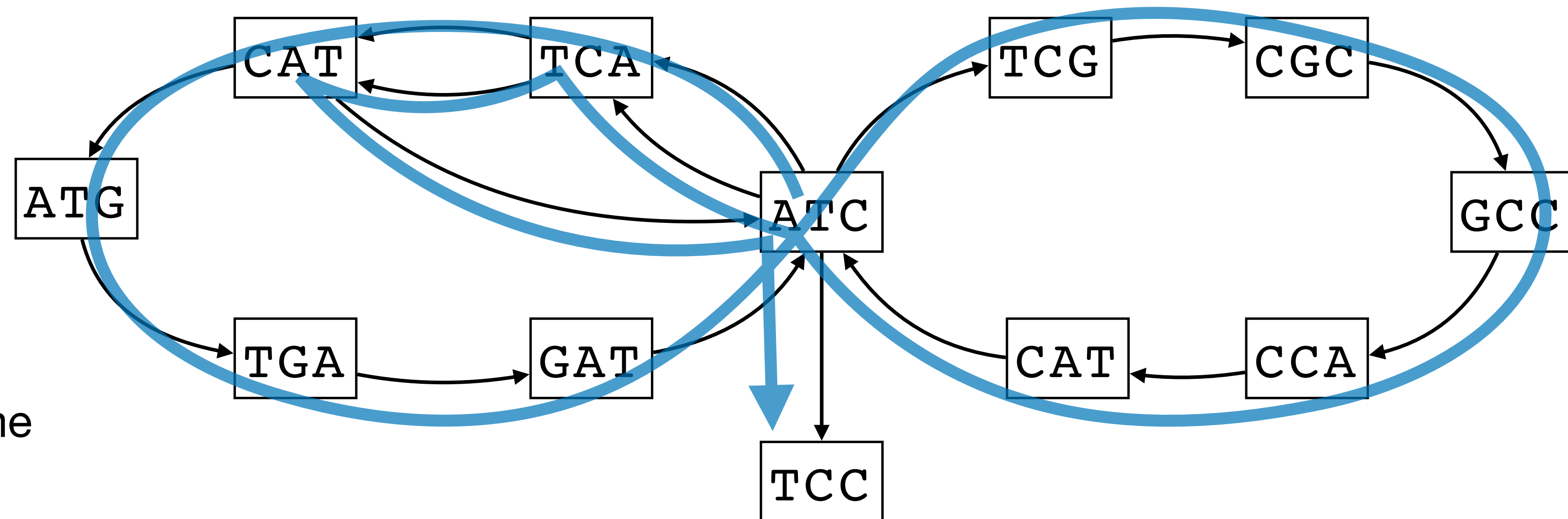
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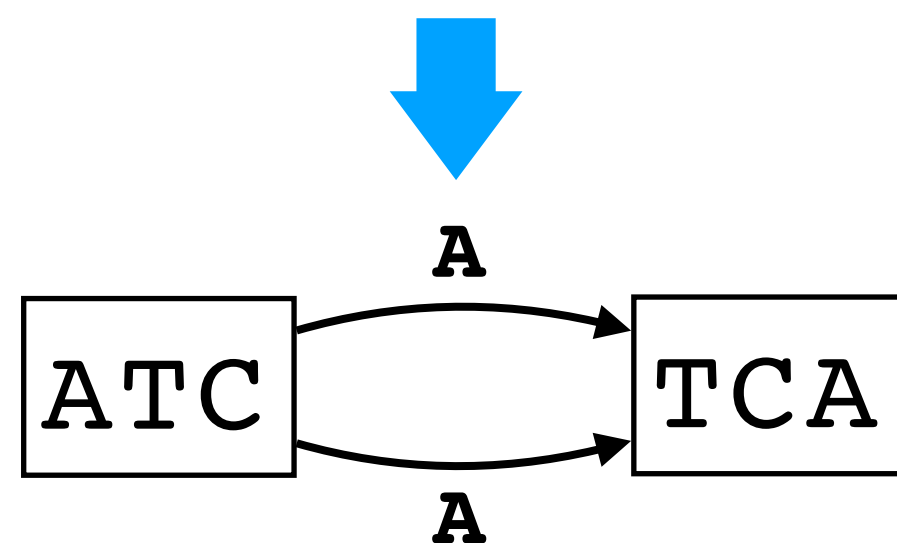
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De Bruijn graph of order 3:



ATCA
ATCA



- Can be solved in $O(|edges|)$ time
- Too restrictive assumption

Section summary

- Modeling the genome assembly problem evolved over time (and still does)

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- Computational complexity ranges from NP-hard to linear

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- Modeling the genome assembly problem evolved over time (and still does)
- Computational complexity ranges from NP-hard to linear
- Not robust to practical issues, and hard to integrate them into the formulations
- Most importantly (even if the above are solved):
 - Many solutions, which one is the true genome?
 - We will see a different "theoretical" approach, closer to practice

AT END OF LECTURE

Practical genome assembly

(The sequence of algorithmic steps behind
"real" genome assemblers)

Contig assembly

Focus on single-end reads
Assembling a full genomic
sequence in one shot is
hopeless



Forget about the assembly
model (i.e. the problem
formulation)
Assemble only parts about
which we are sure (contigs →
contiguous sequences)

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- Focus on ***unitigs*** =_{def} "non-branching" path

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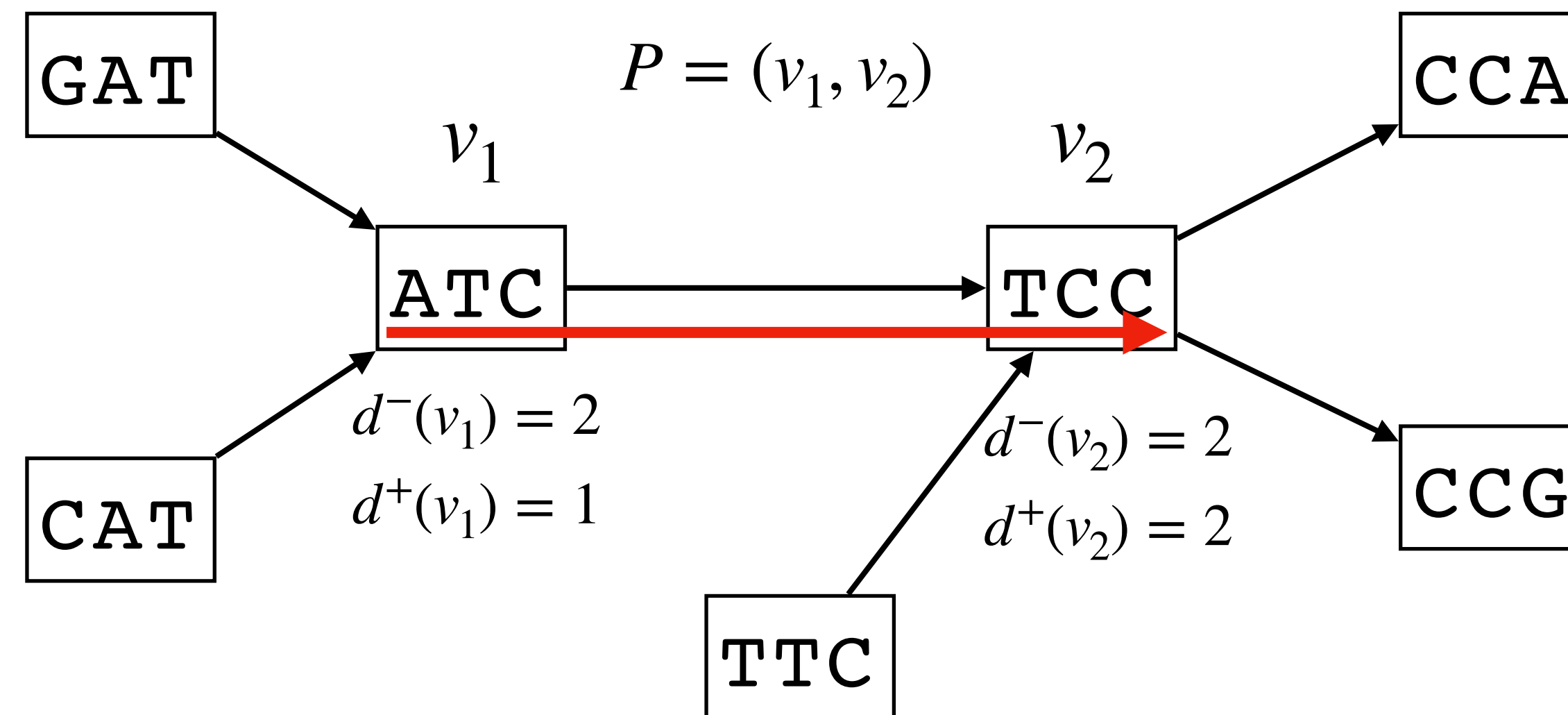
- Assume an assembly graph (here de Bruijn with parallel edges collapsed into one)
- Focus on **unitigs** =_{def} "non-branching" path
- (Usually) Contig =_{def} unitig in a graph "corrected" for polyploidy

Unitigs

- Let $P = (v_1, v_2, \dots, v_{t-1}, v_t)$ be a path. We say that P is a ***unitig*** if either:

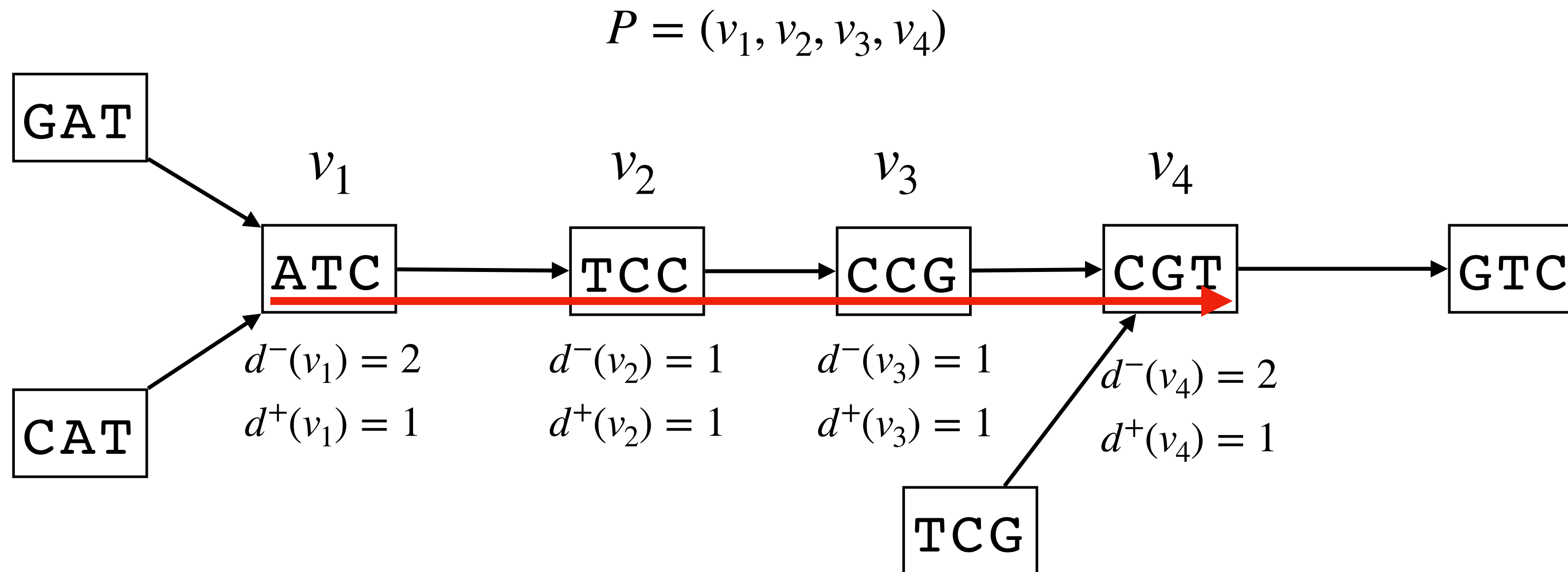
Unitigs

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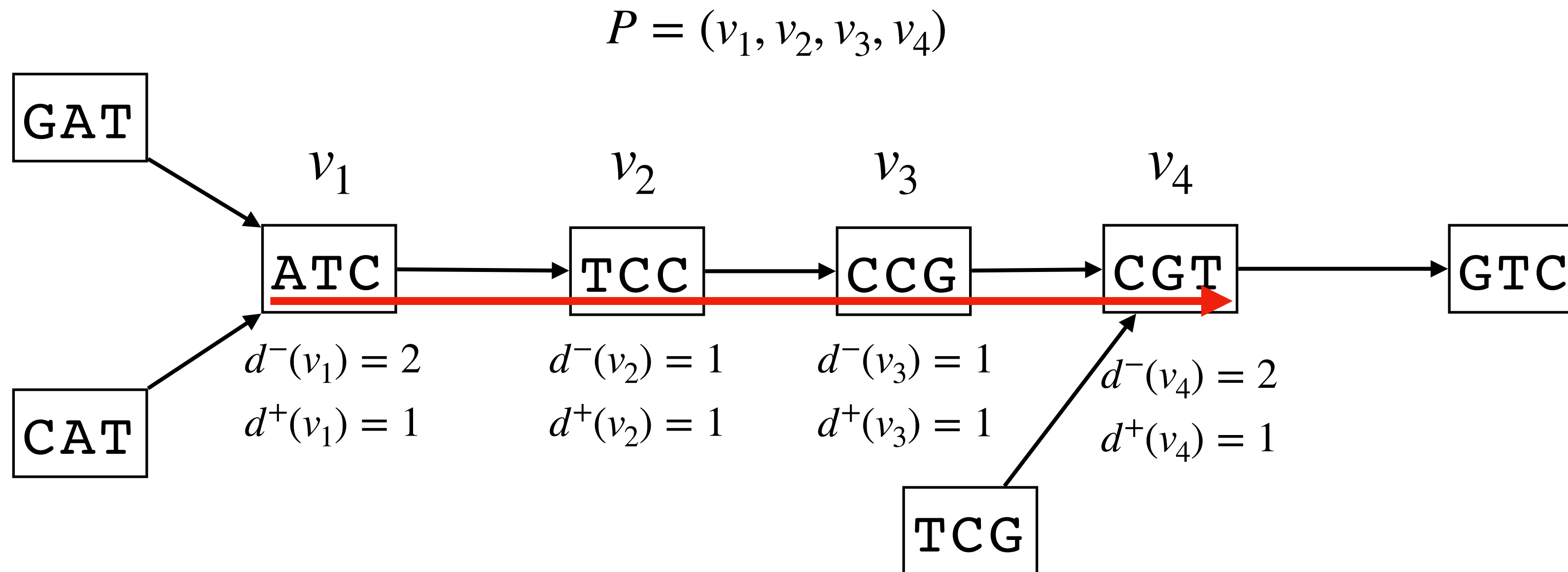
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 - ▶ for every $i \in \{2, \dots, t-1\}$, we have $d^-(v_i) = d^+(v_i) = 1$,
(every internal node has exactly one in-neighbor and one out-neighbor)



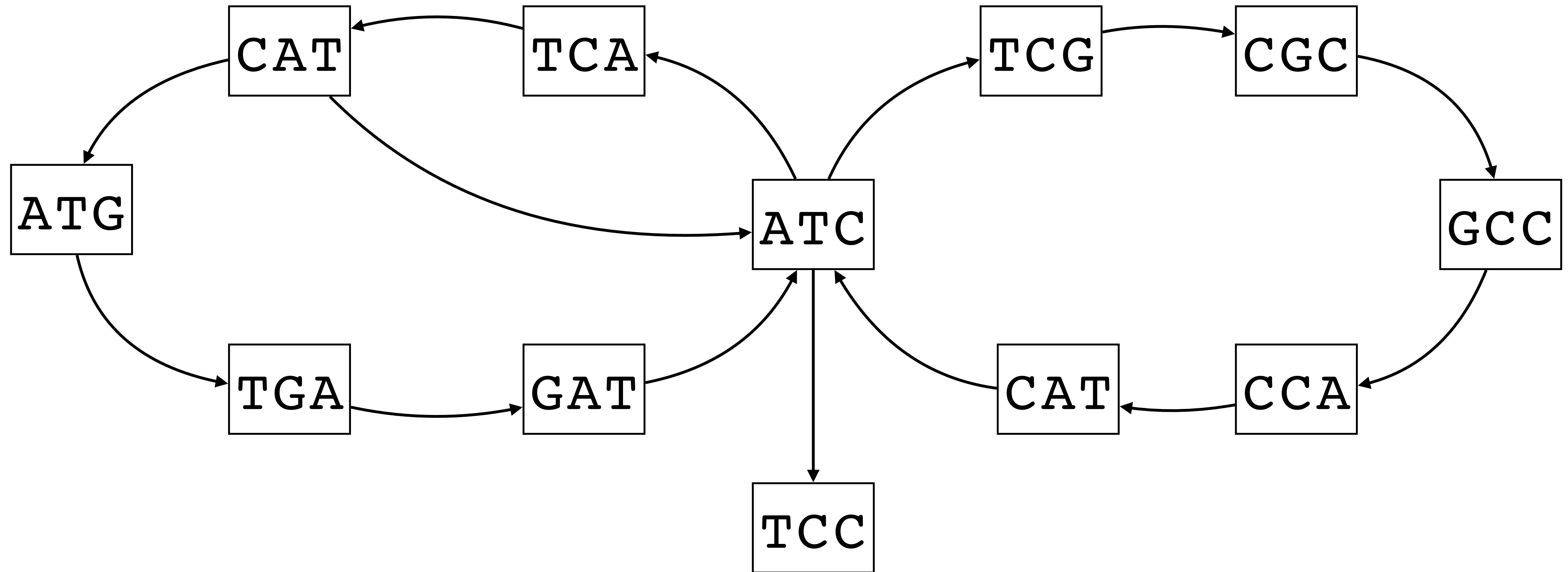
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 - for every $i \in \{2, \dots, t-1\}$, we have $d^-(v_i) = d^+(v_i) = 1$,
(every internal node has exactly one in-neighbor and one out-neighbor)

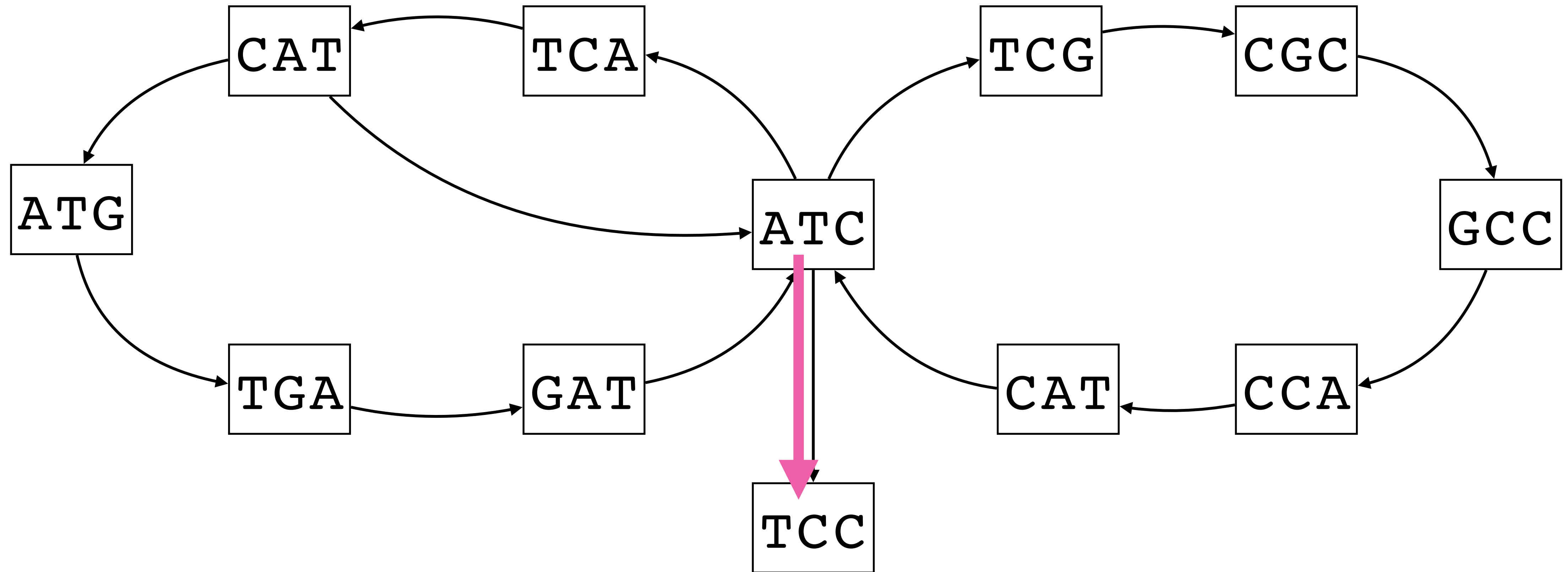


- We want maximal (longest) unitigs

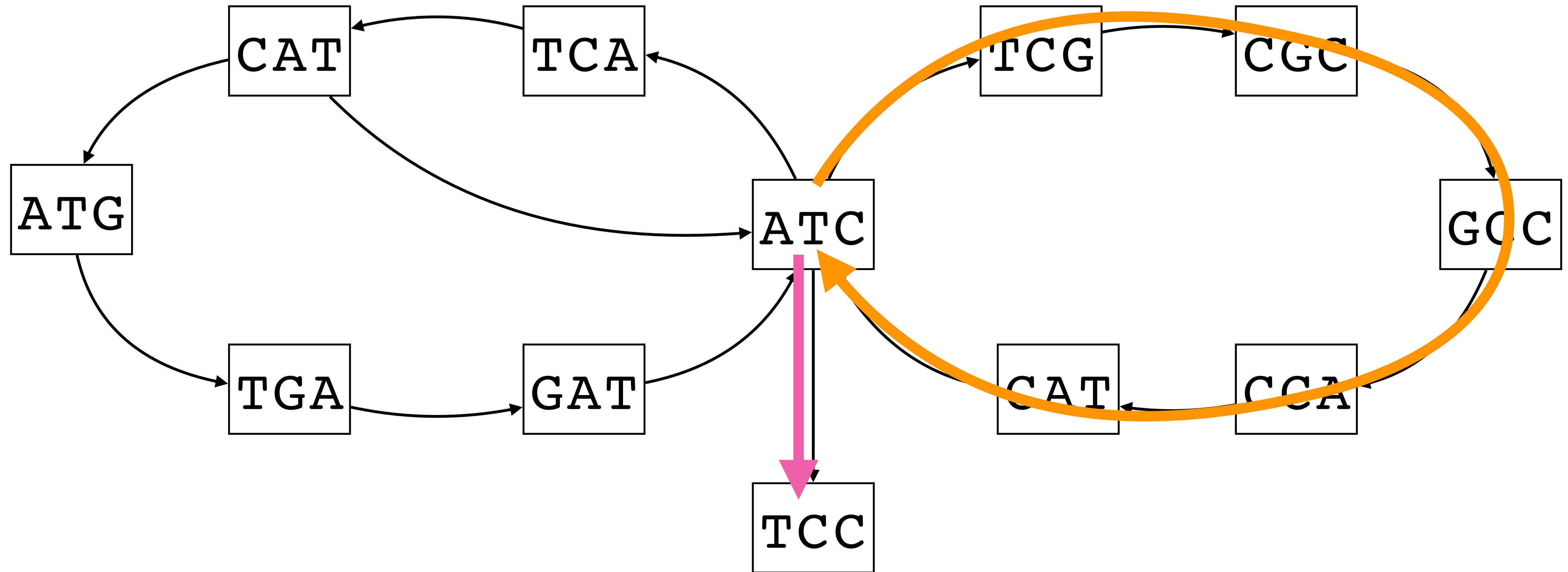
Example



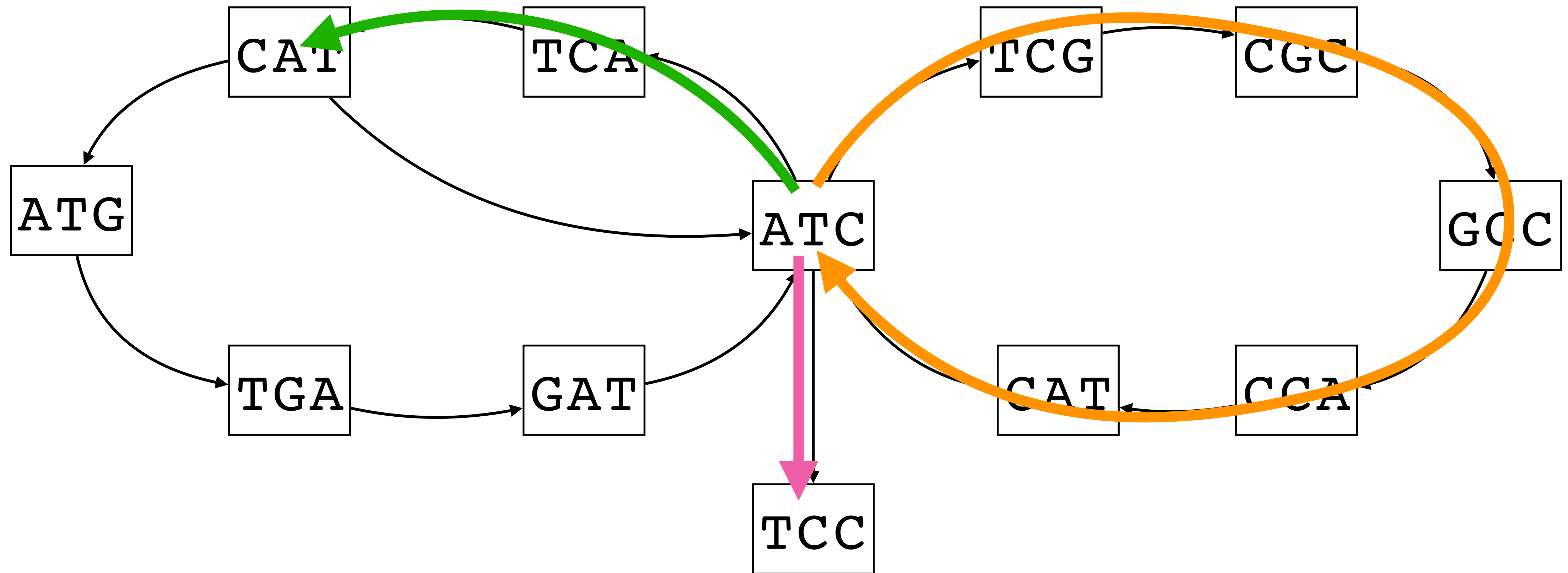
Example



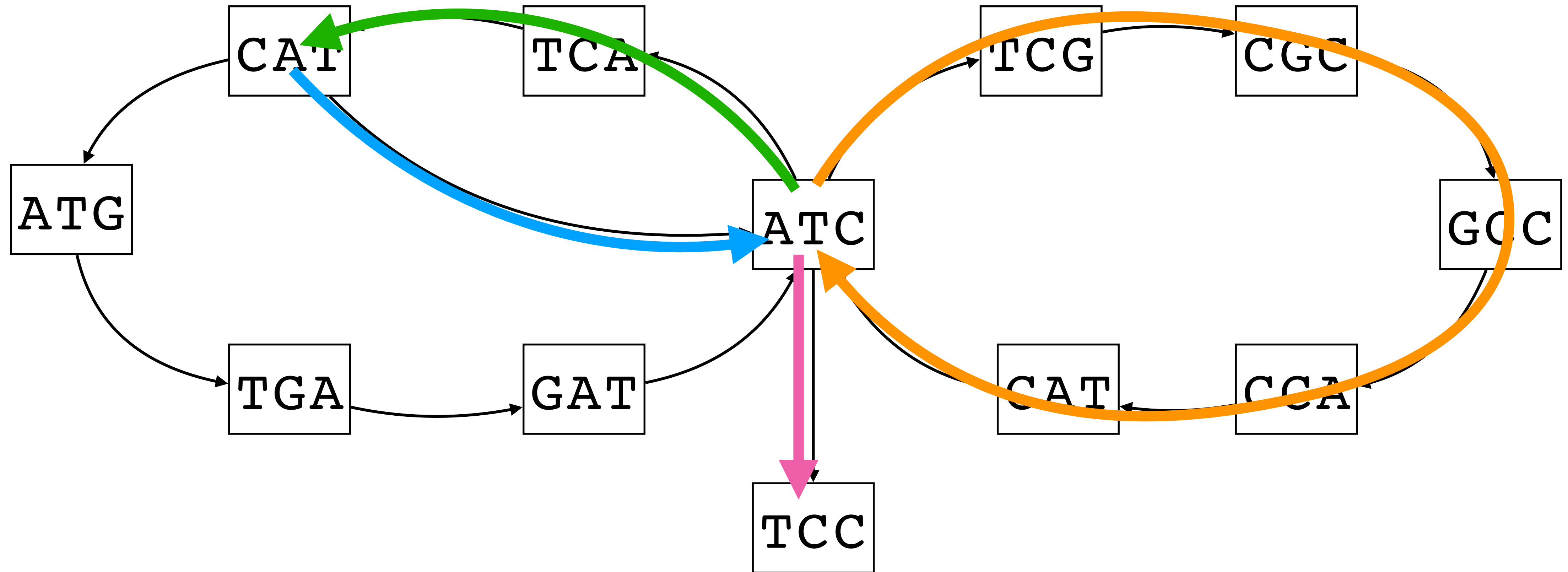
Example



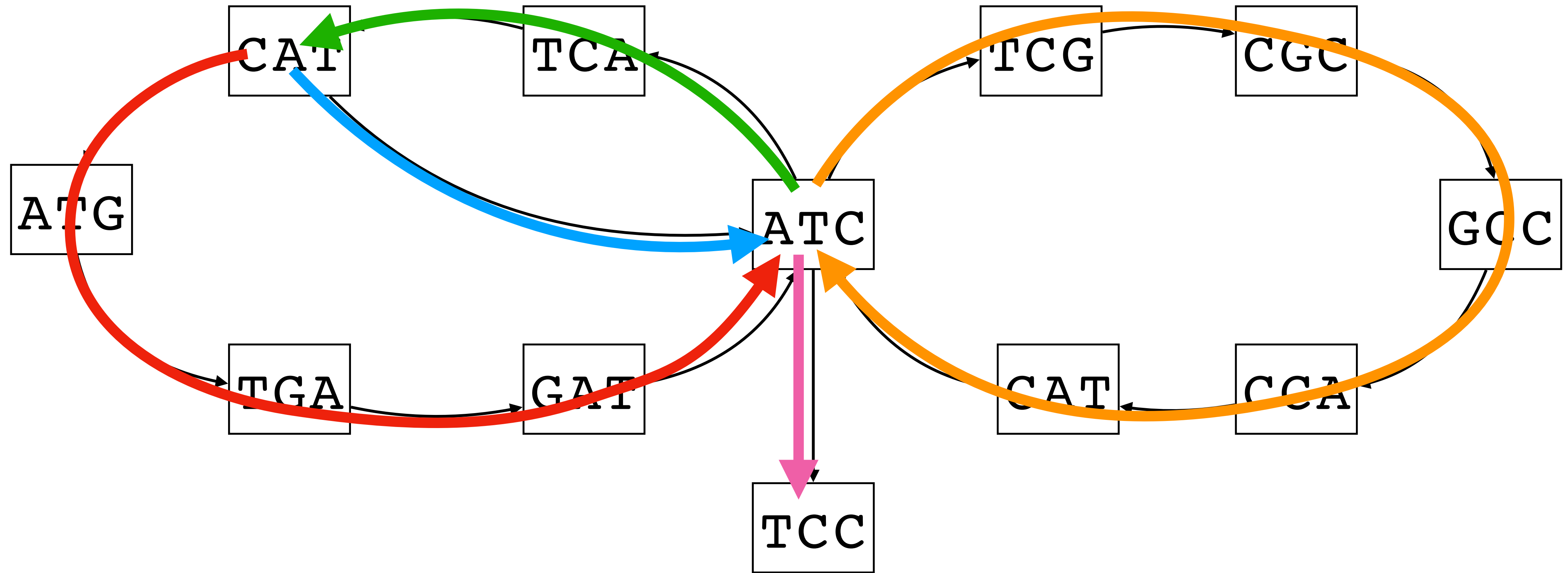
Example



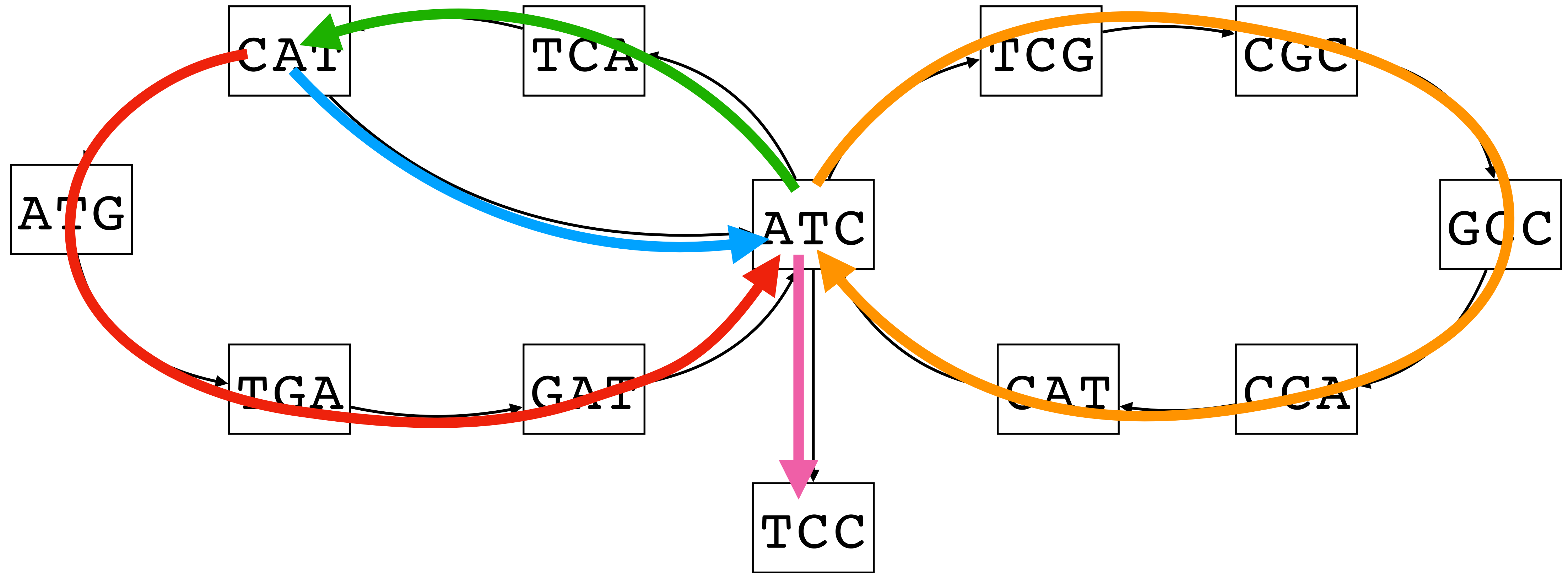
Example



Example



Example



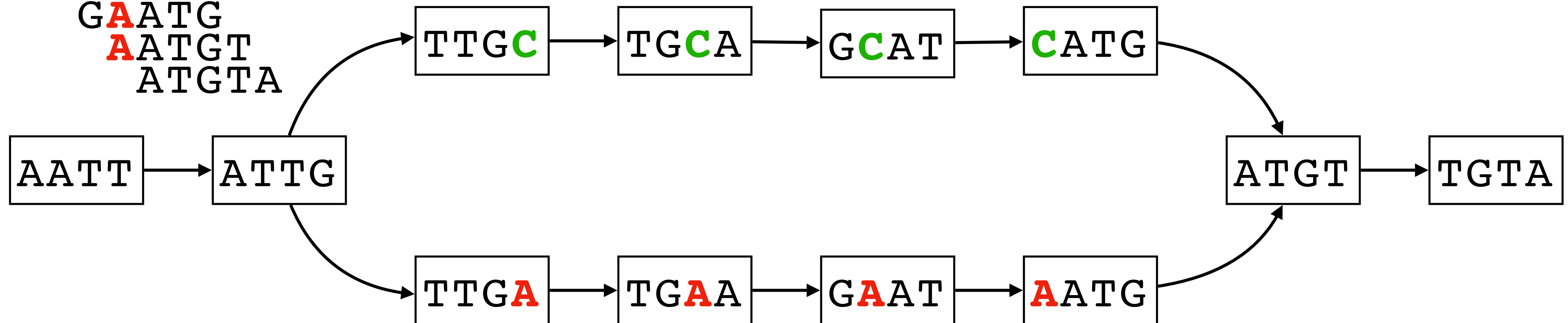
- Unitigs can be found in $O(|\text{edges}|)$ time

Bubble popping

ATGTA
CATGT
GCATG
TGCAT
TTGCA
ATTGC
AATTG
Mother **AATTGCATGTA**
Father **AATTGAATGTA**
AATTG
ATTG**A**
TTG**A**A
TG**A**AT
G**A**ATG
AATGT
ATGTA

Bubble popping

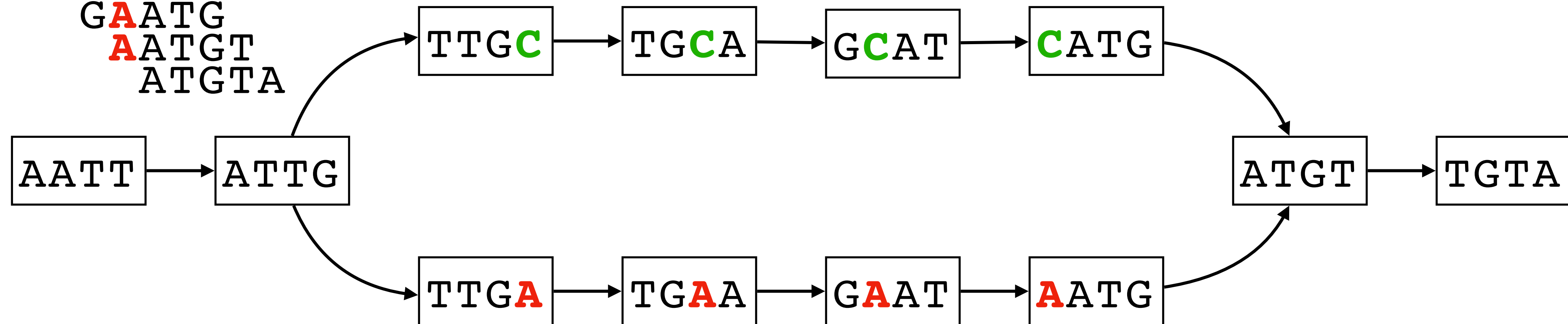
ATGTA
CATGT
GCATG
TGCA
TTGCA
ATTGC
AATTG
Mother **AATTGCATGTA**
Father **AATTGAATGTA**
AATTG
ATTG**A**
TTG**AA**
TG**AA**T
G**A**ATG
AATGT
ATGTA



Bubble popping

- Bubble =_{def} two paths of same length with same start and end node
 - Caused by a SNP in haplotype not present in the other

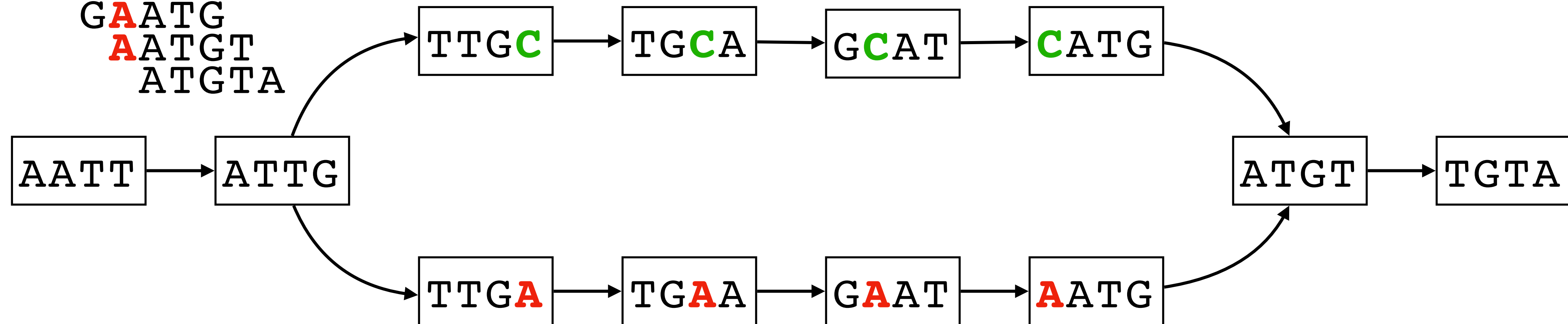
ATGTA
CATGT
GCATG
TGCA
TTGC
ATTG
AATTG
Mother **AATTGCATGTA**
Father **AATTGAATGTA**
AATTG
ATTG**A**
TTG**AA**
TG**AA**T
G**A**ATG
AATGT
ATGTA



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- Leads to shorter unitigs (poliploidy can be solved later)

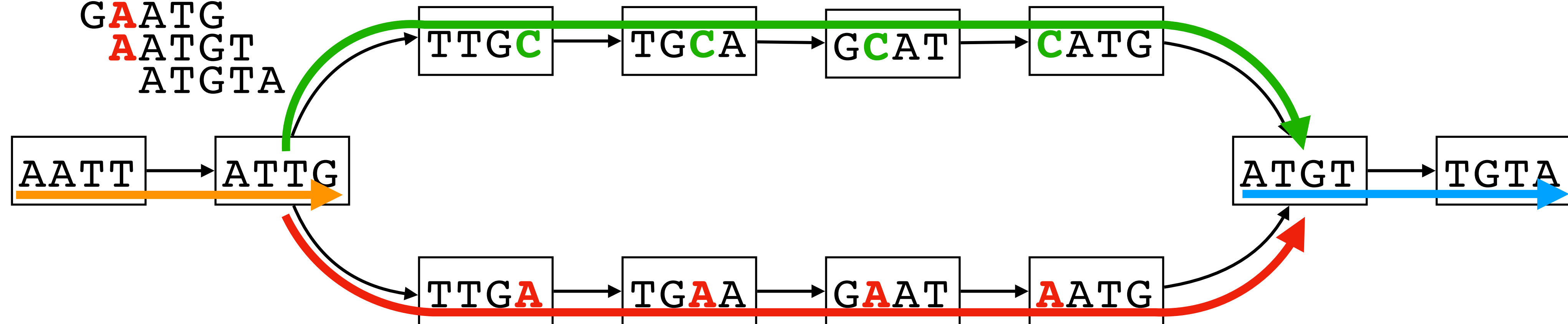
ATGTA
CATGT
GCATG
TGCA
TTGC
ATTG
AATTG
Mother AATTGCATGTA
Father AATTGAATGTA
AATTG
ATTGA
TTGAA
TGAA
GAATG
AATGT
ATGTA



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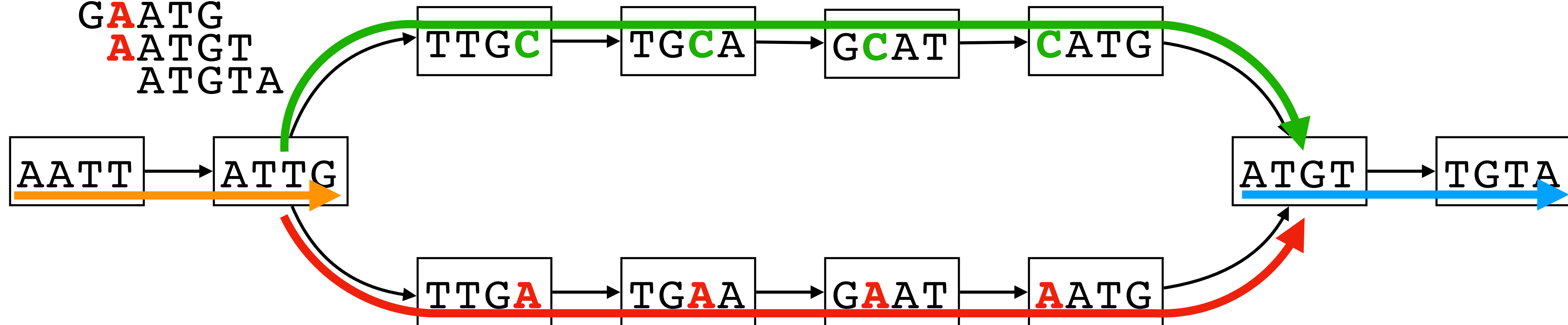
ATGTA
CATGT
GCATG
TGCA
TTGC
ATTG
AATTG
Mother **AATTGCATGTA**
Father **AATTGAATGTA**
AATTG
ATTGA
TTGAA
TGAA
GAATG
AATGT
ATGTA



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 - Taku Onodera, Kunihiro Sadakane, Tetsuo Shibuya:
Detecting Superbubbles in Assembly Graphs. WABI 2013: 338-348

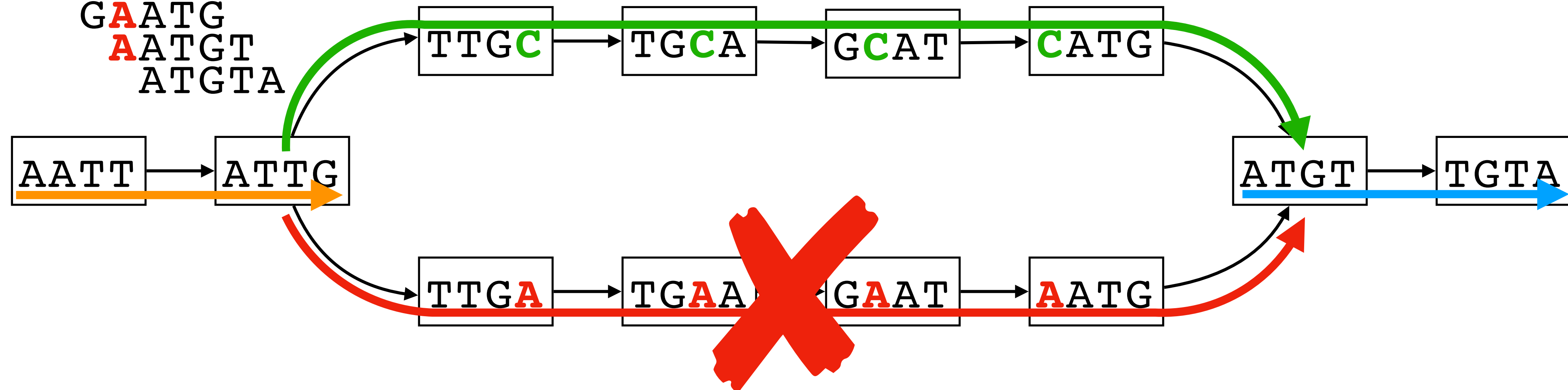
ATGTA
 CATGT
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 TGCAT
 TTGCA
 ATTGC
 AATTG
 Mother **AATTGCATGTA**
 Father **AATTGAATGTA**
 AATTG
 ATTGA
 TTGAA
 TGAAT
 GAATG
 AATGT
 ATGTA



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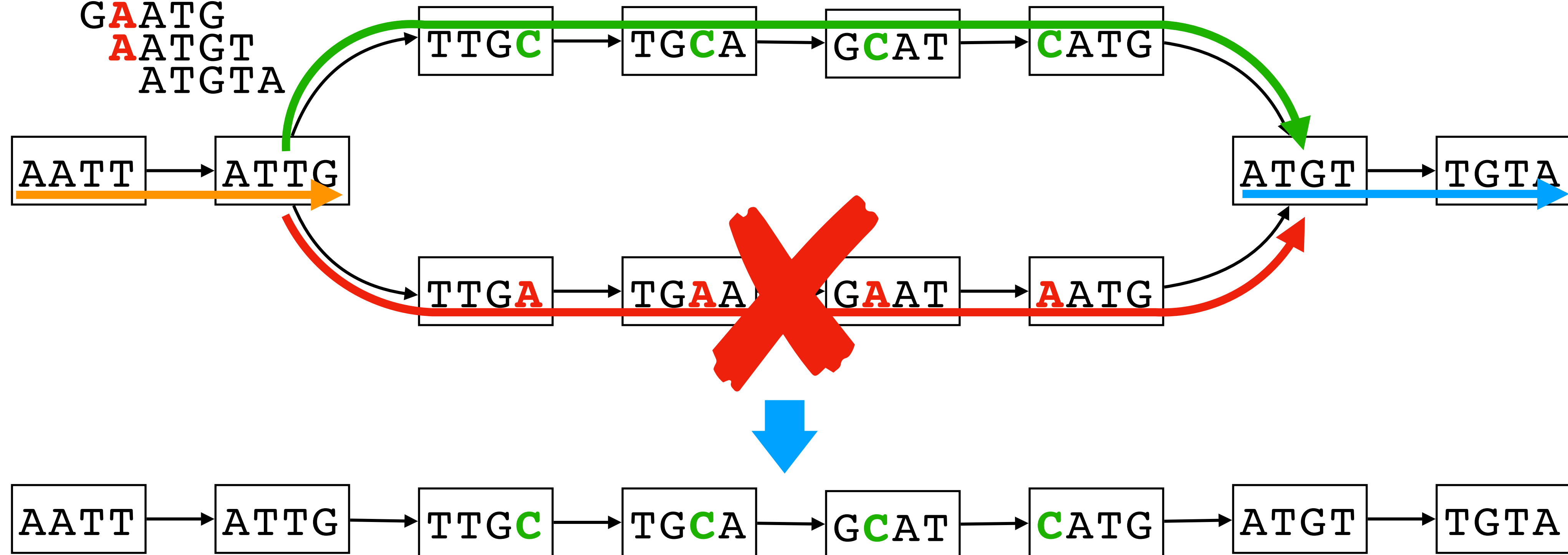
ATGTA
CATGT
GCATG
TGCA
TTGC
ATTG
AATTG
Mother **AATTGCATGTA**
Father **AATTGAATGTA**
AATTG
ATTGA
TTGAA
TGAA
GAATG
AATGT
ATGTA



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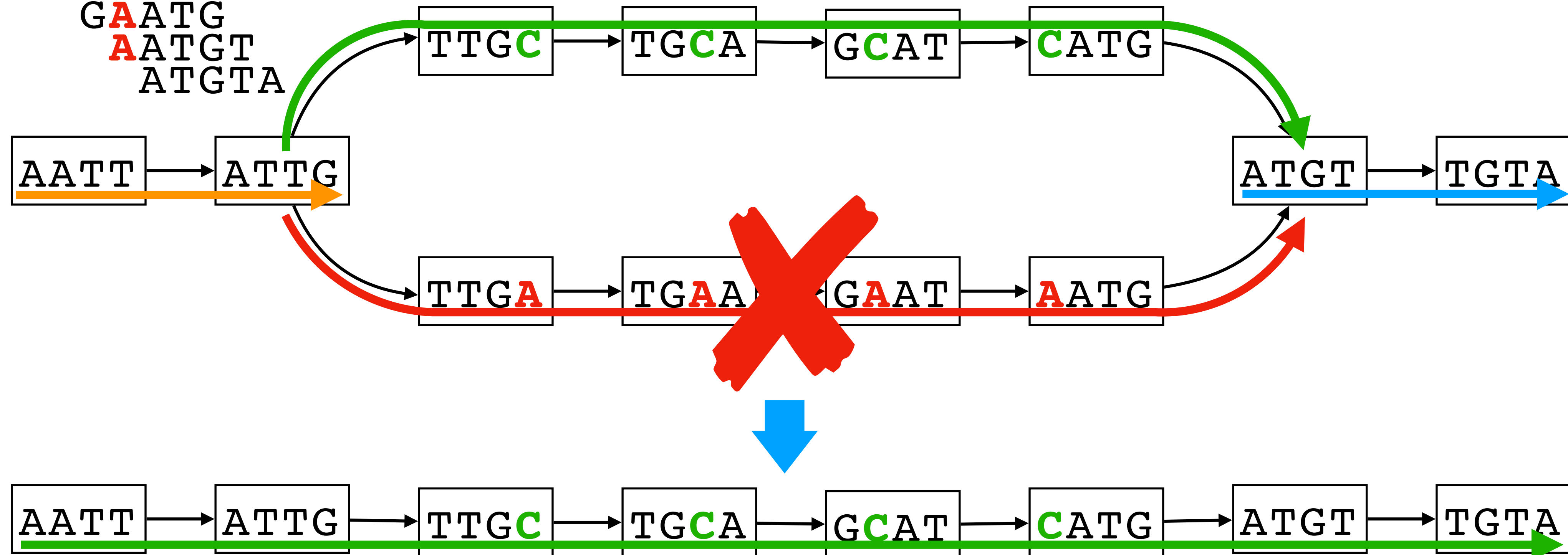
ATGTA
CATGT
GCATG
TGCAT
TTGCA
ATTGC
AATTG
Mother AATTGCATGTA
Father AATTGAATGTA
AATTG
ATTGA
TTGAA
TGAAAT
GAATG
AATGT
ATGTA



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ATGTA
CATGT
GCATG
TGCA
TTGC
ATTG
AATTG
Mother **AATTGCATGTA**
Father **AATTGAATGTA**
AATTG
ATTGA
TTGAA
TGAA
GAATG
AATGT
ATGTA

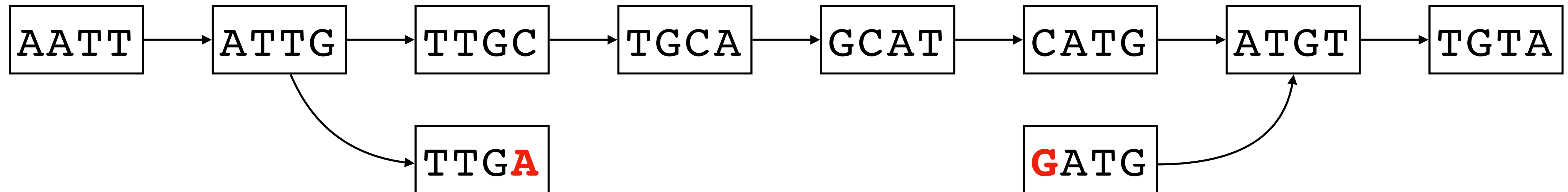


Tip removal

ATGTA
GATGT
GCATG
TGCAT
TTGCA
ATTGC
ATTGA
AATTG
AATTGCATGTA

Tip removal

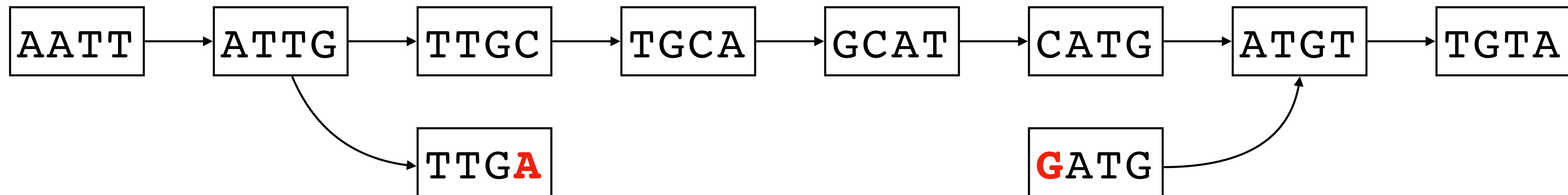
ATGTA
GATGT
GCATG
TGCAT
TTGCA
ATTGC
ATTGA
AATTG
AATTGCATGTA



Tip removal

- Tip =_{def} a "short" path that either:
 - ends in a *sink* v (i.e. $d^+(v) = 0$)
 - starts in a *source* v (i.e. $d^-(v) = 0$)

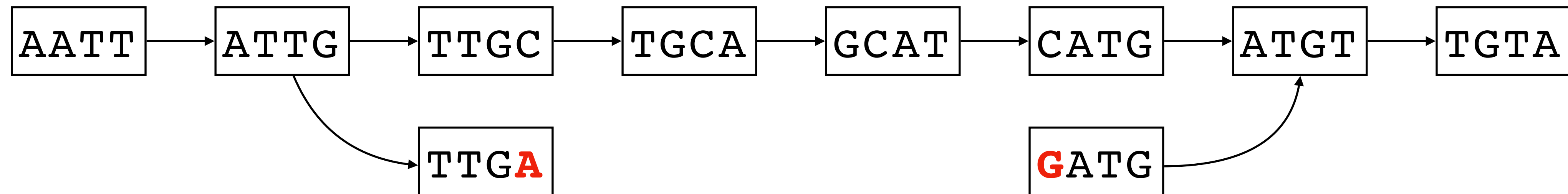
ATGTA
 GATGT
 GCATG
 TGCAT
 TTGCA
 ATTGC
 ATTG**A**
AATTG
AATTGCATGTA



Tip removal

ATGTA
GATGT
GCATG
TGCAT
TTGCA
ATTGC
ATTGA
AATTG
AATTGCATGTA

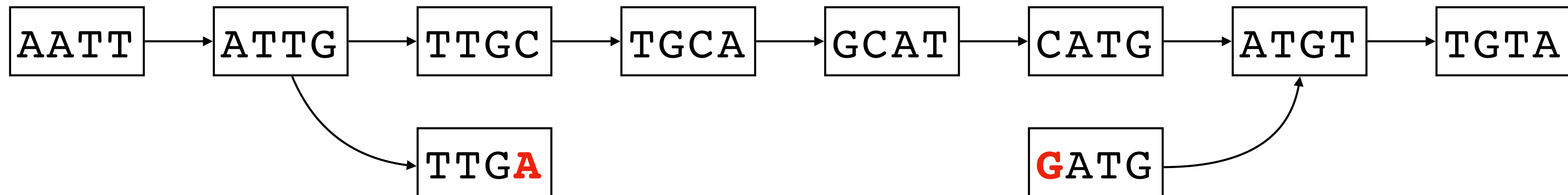
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Tip removal

ATGTA
GATGT
GCATG
TGCAT
TTGCA
ATTGC
ATTGA
AATTG
AATTGCATGTA

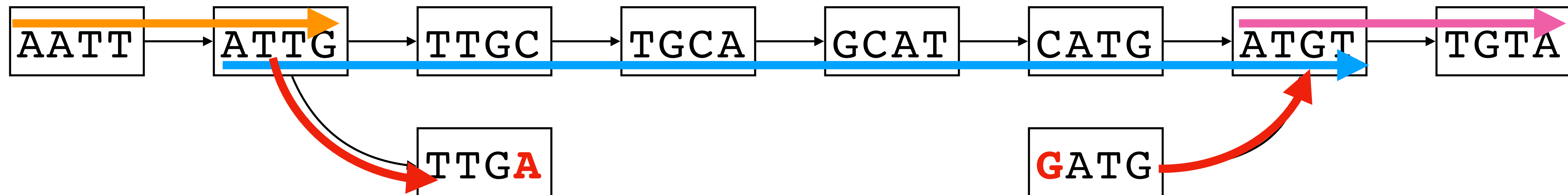
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GCATG
TGCAT
TTGCA
ATTGC
ATTGA
AATTG
AATTGCATGTA

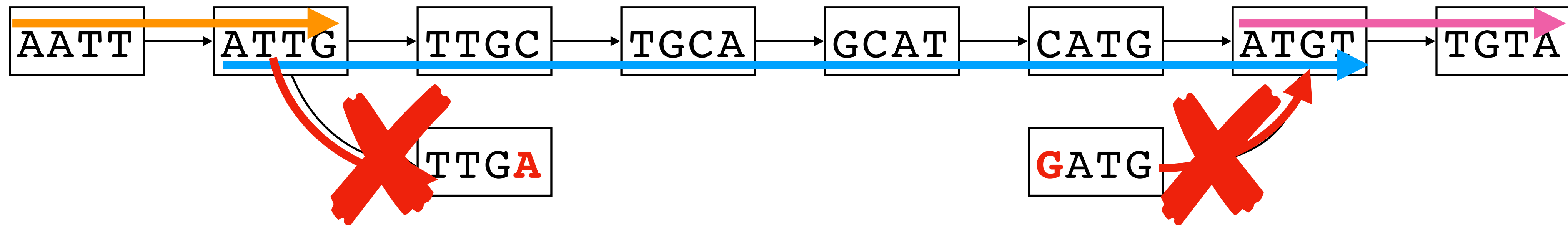
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ATGTA
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TGCAT
TTGCA
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ATTGA
AATTG
AATTGCATGTA

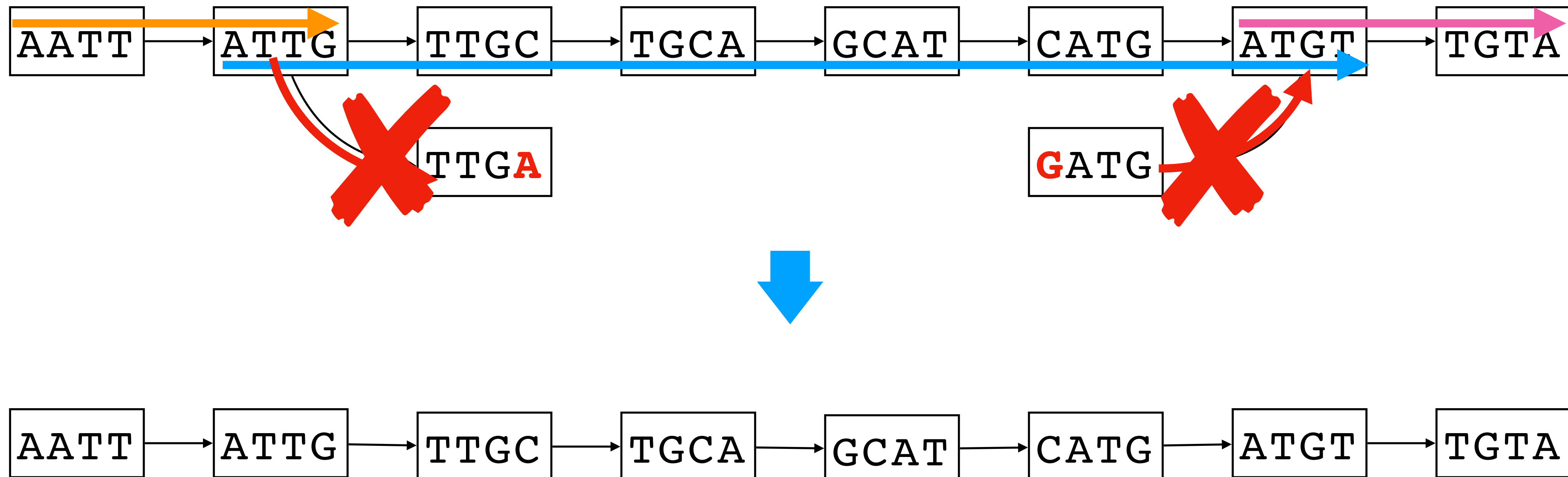
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ATGTA
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GCATG
TGCAT
TTGCA
ATTGC
ATTGA
AATTG
AATTGCATGTA

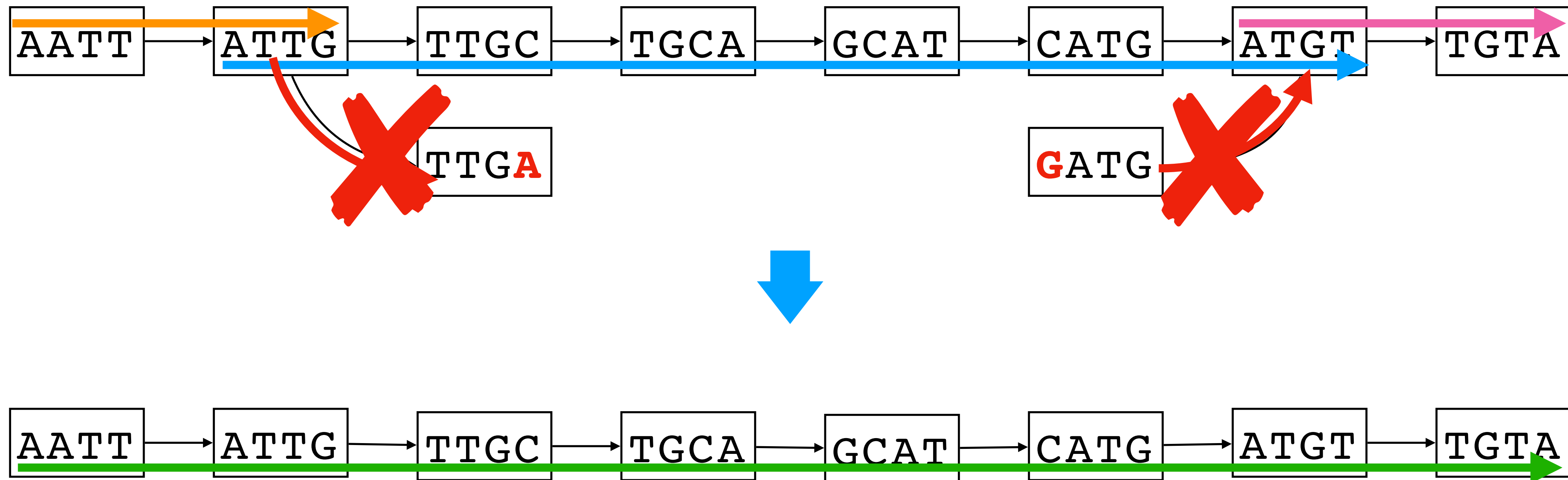
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Tip removal

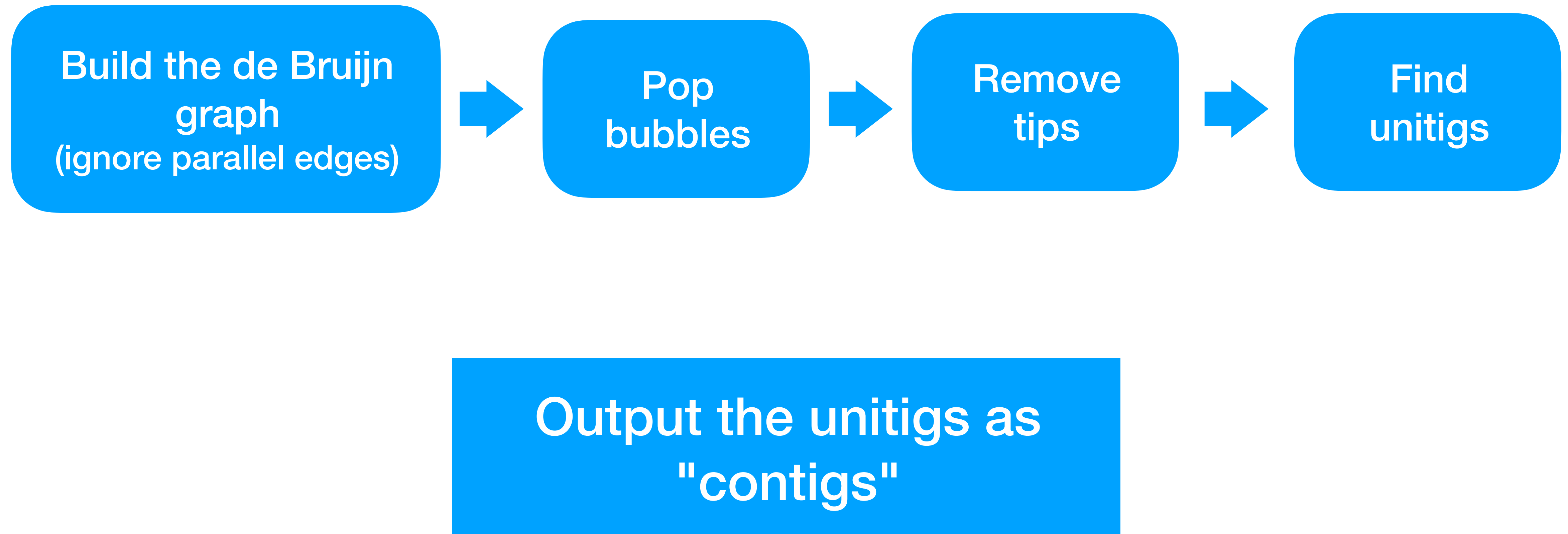
ATGTA
GATGT
GCATG
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Contigs assembly

(simplified to ignore some practical issues e.g. errors, reverse complements)



Scaffolding

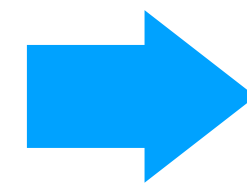
Bring in paired-end
information
Align reads to contigs



Chain (order) the contigs
Output chains of contigs
with "gaps" (NNNN...)
between them

Contigs

TCGATAGCTAAAA
AATTGT
ATAGAGATATTT
ATATCGCTAGA



Scaffolds

TCGATAGCTAAAA**NNNNNNNNNN**AATTGT**NNN**ATAGAGATATTT
ATATCGCTAGA

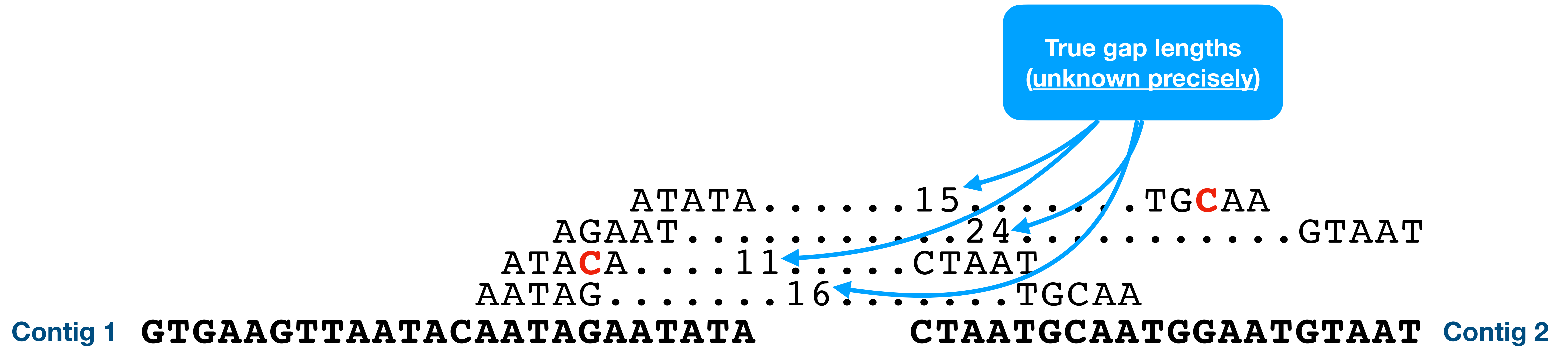
Align paired-end reads to contigs

ATATA 15 TGCAA
AGAAT 24 GTAAT
ATACAA 11 CTAAT
AATAG 16 TGCAA

Contig 1 **GTGAAGTTAATACAATAGAATATA** **CTAATGCAATGGAATGTAAT** Contig 2

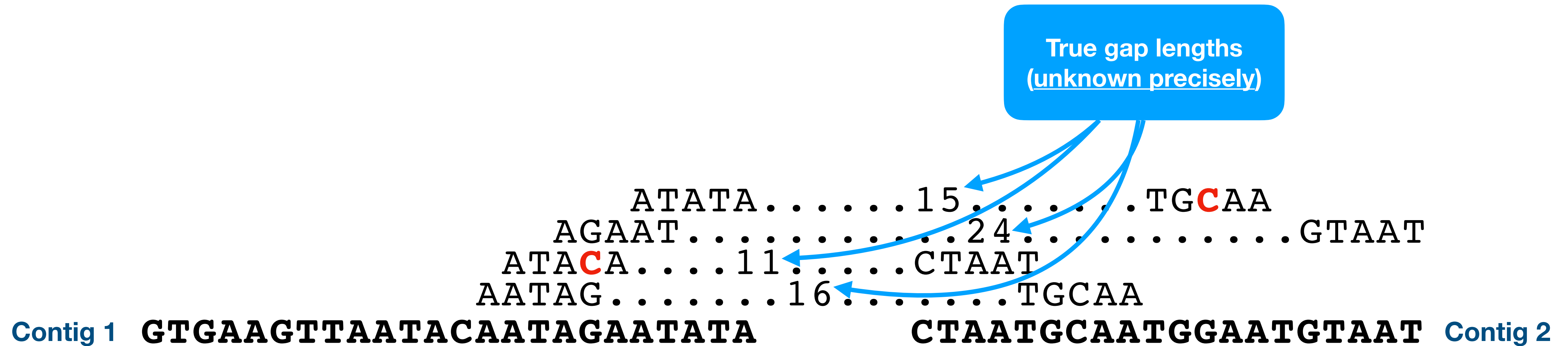
- Align paired-end reads to contigs, focus on read pairs aligning to different contigs

Align paired-end reads to contigs



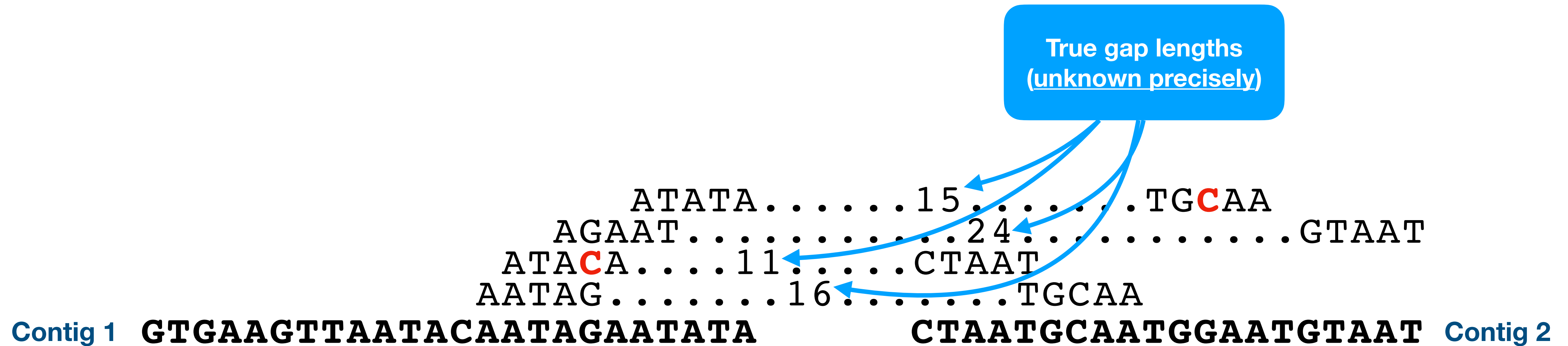
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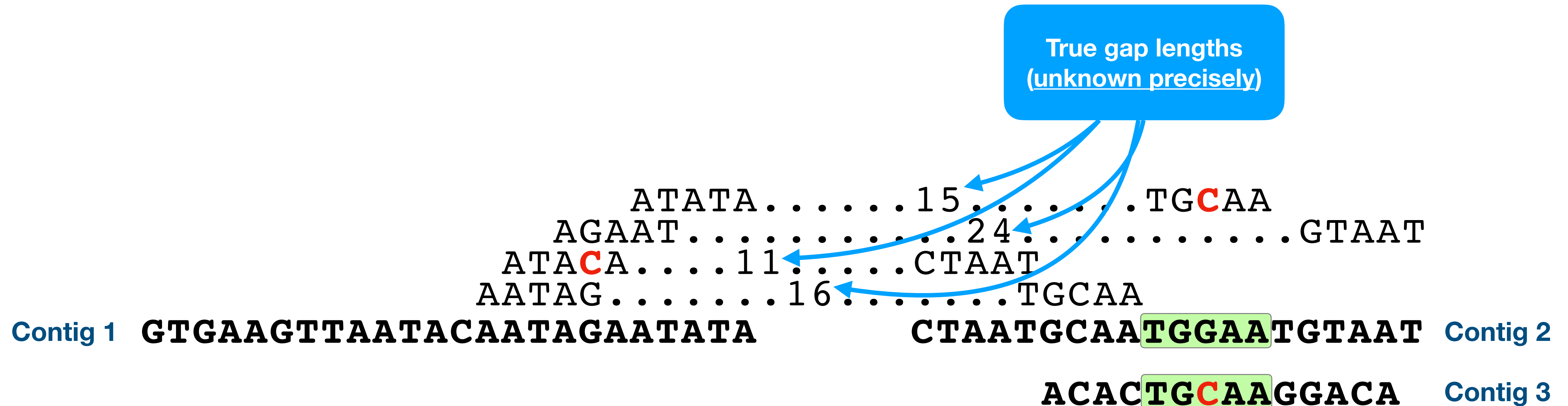
- Align paired-end reads to contigs, focus on read pairs aligning to different contigs
 - Even if we know two contigs are "consecutive" in a genome, it is not easy to estimate the gap length between them, see e.g.

Align paired-end reads to contigs



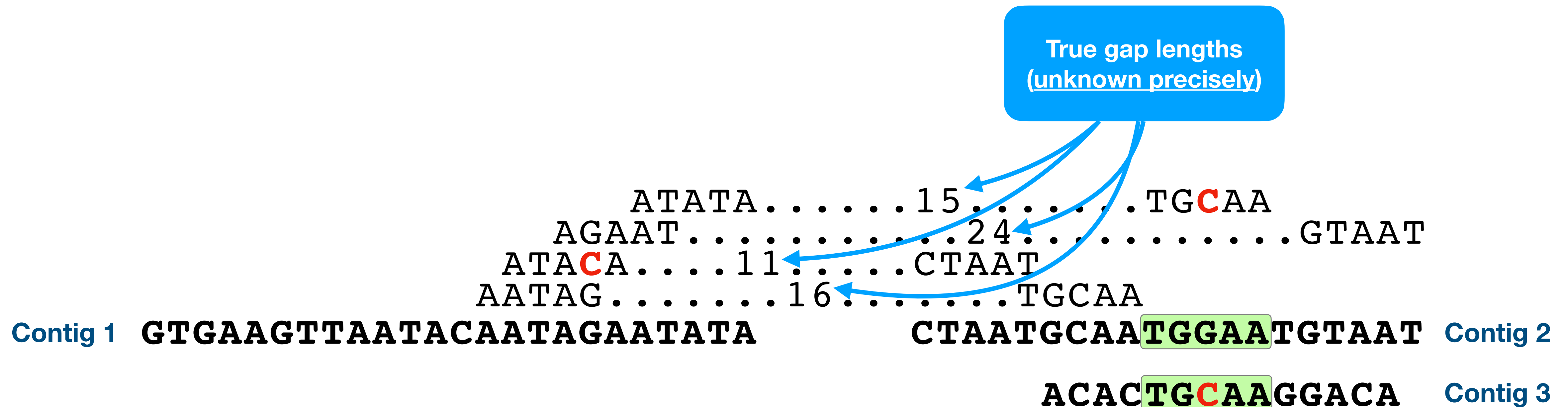
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 - ▶ Kristoffer Sahlin, Nathaniel Street, Joakim Lundeberg, Lars Arvestad:
Improved gap size estimation for scaffolding algorithms. *Bioinformatics* 28(17): 2215-2222 (2012)

Align paired-end reads to contigs



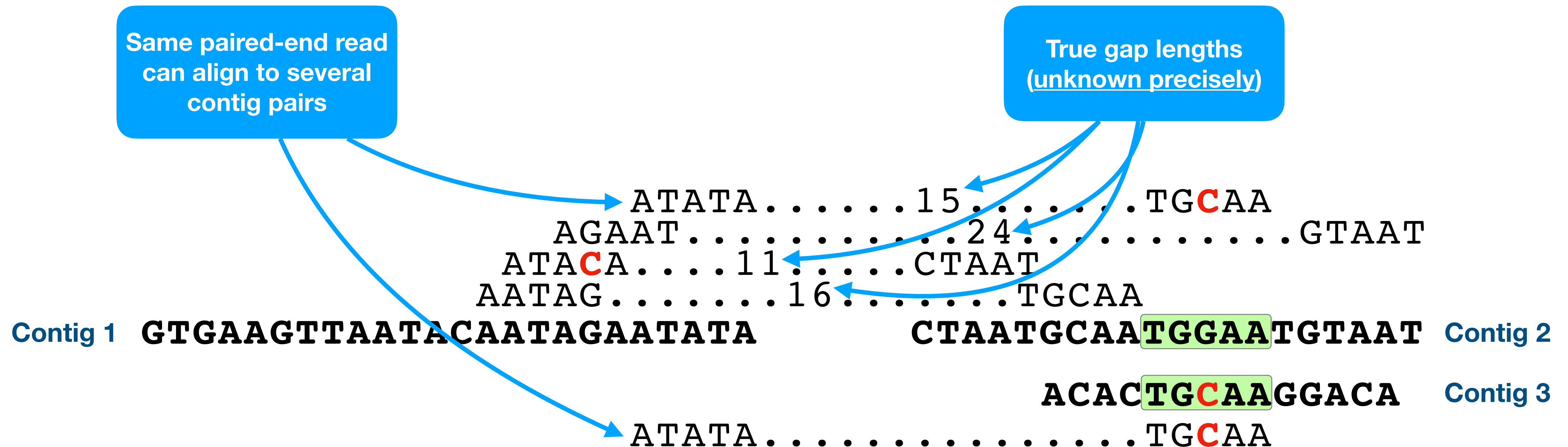
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Align paired-end reads to contigs



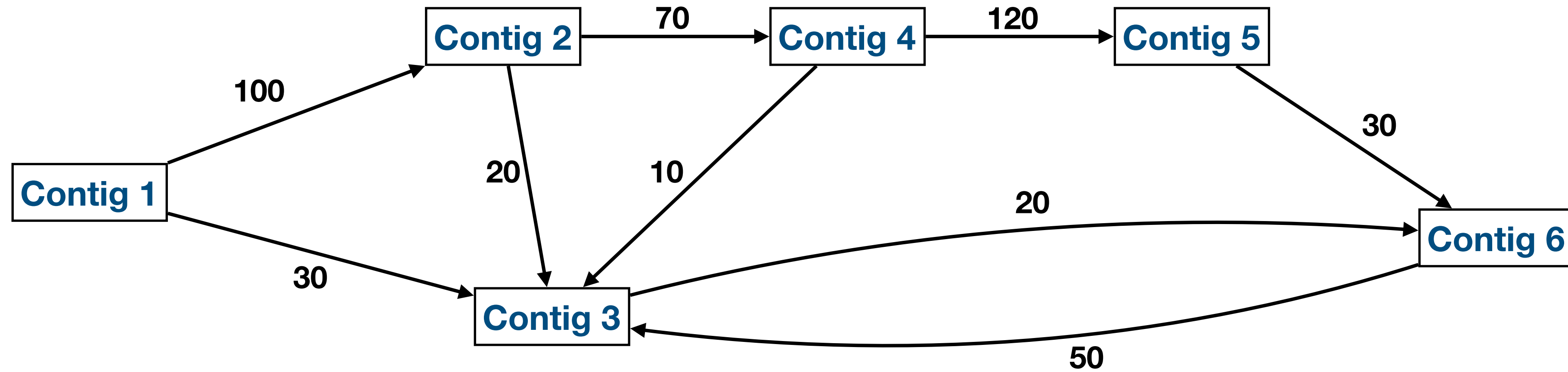
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Align paired-end reads to contigs

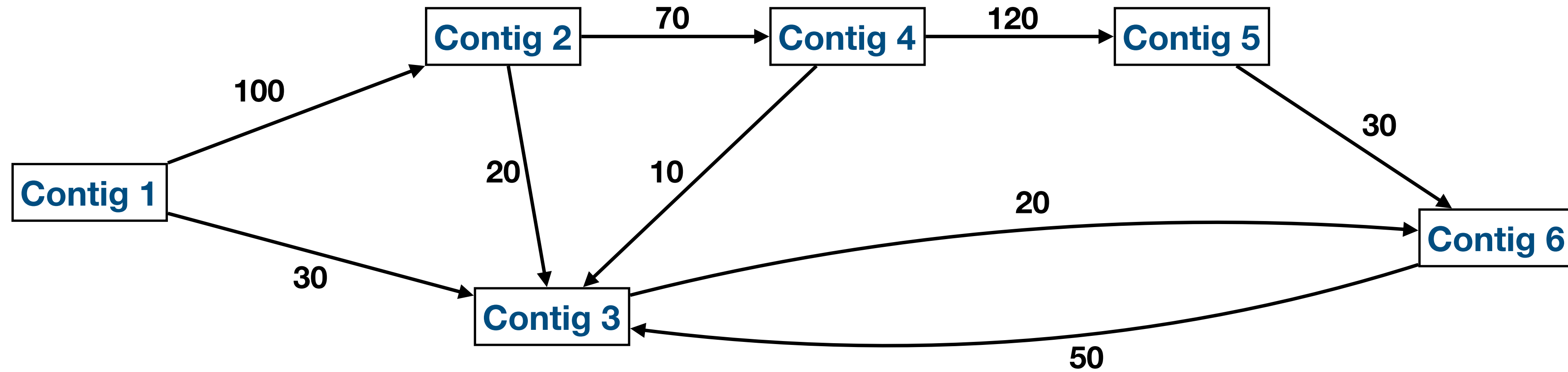


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Chaining (scaffolding) contigs

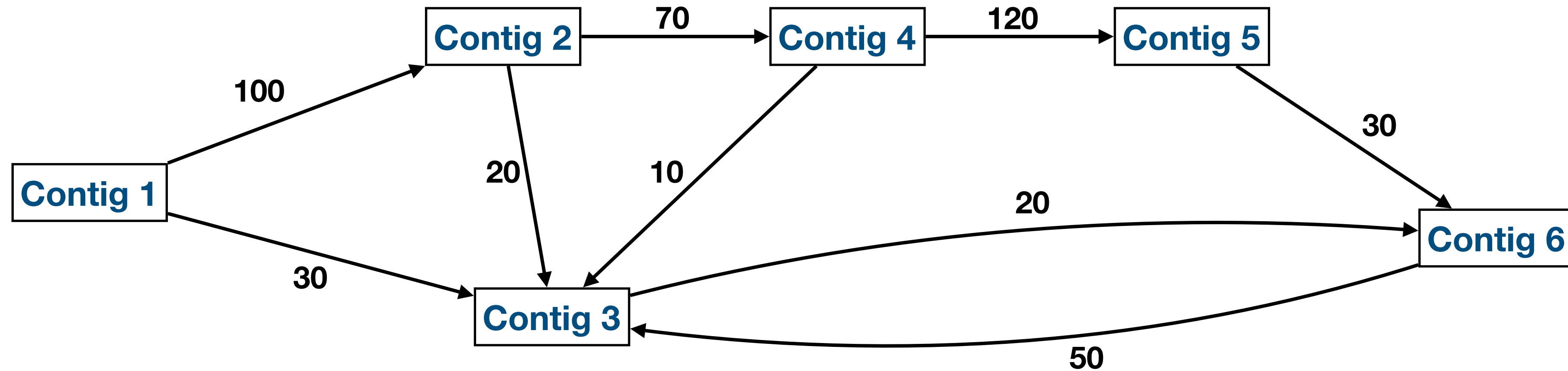


Chaining (scaffolding) contigs



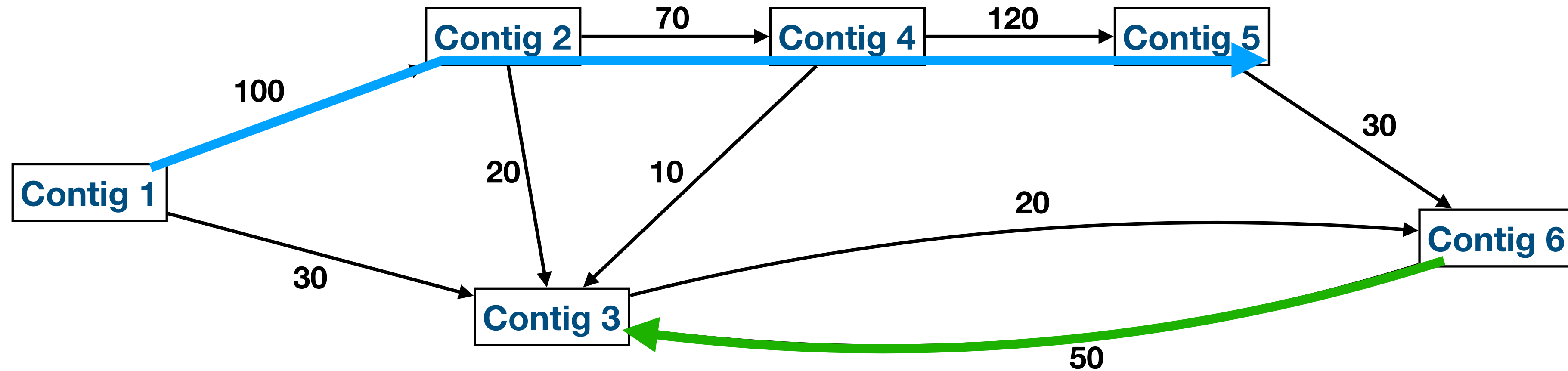
- Another contig assembly-like problem

Chaining (scaffolding) contigs



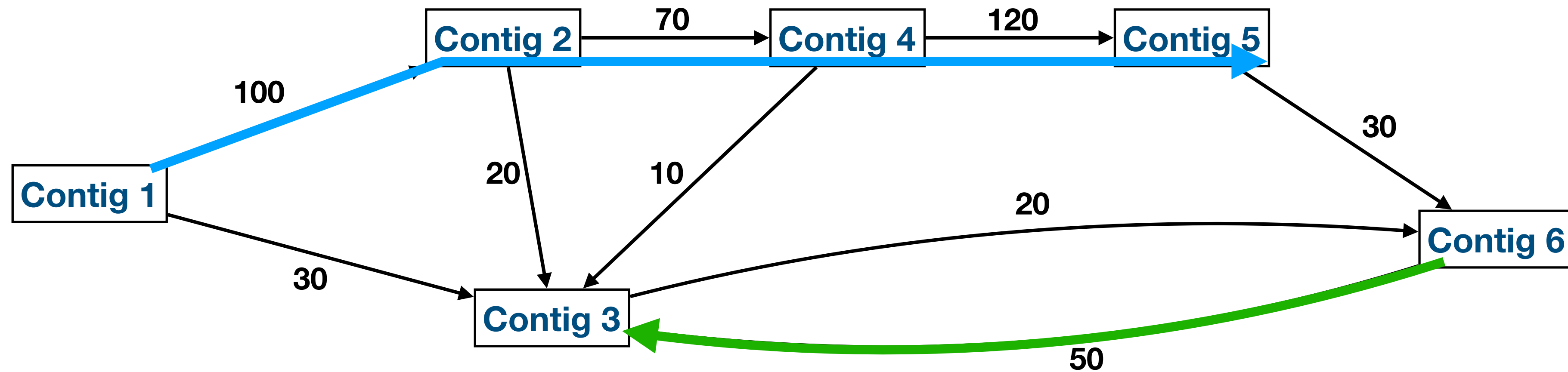
- Another contig assembly-like problem
- Now weights on edges (how much "evidence" there is): new problem formulations

Chaining (scaffolding) contigs



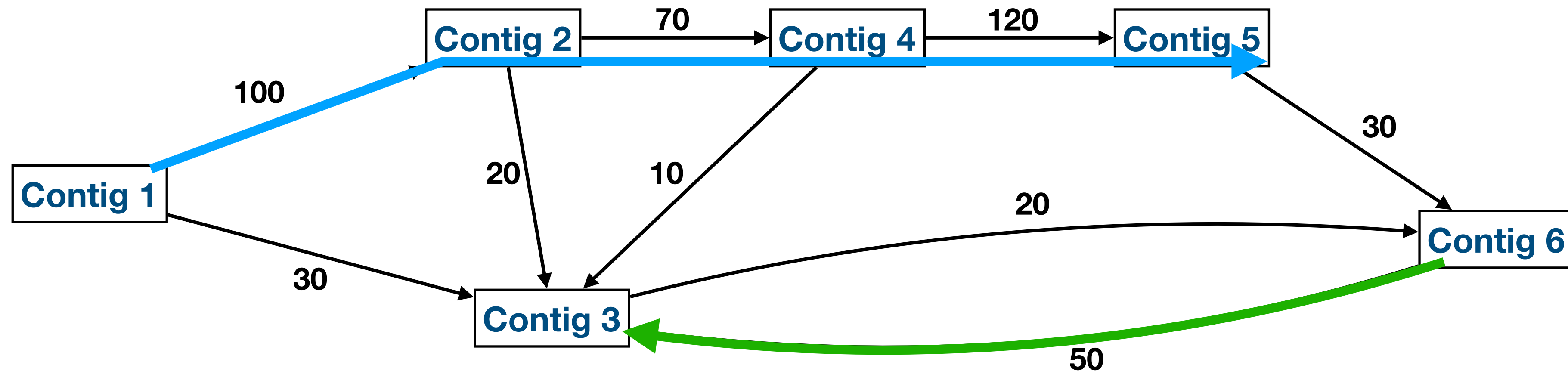
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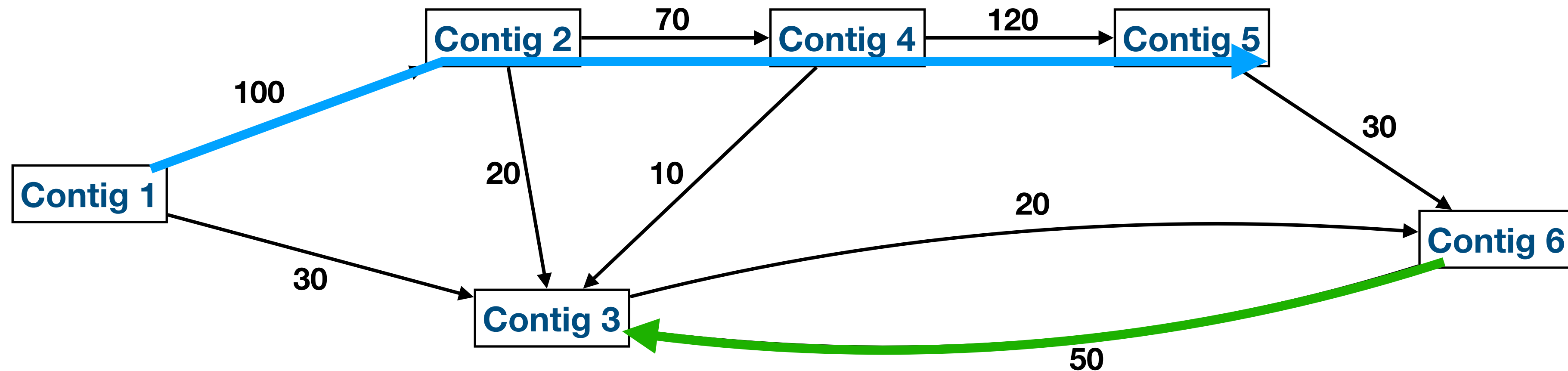
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Chaining (scaffolding) contigs



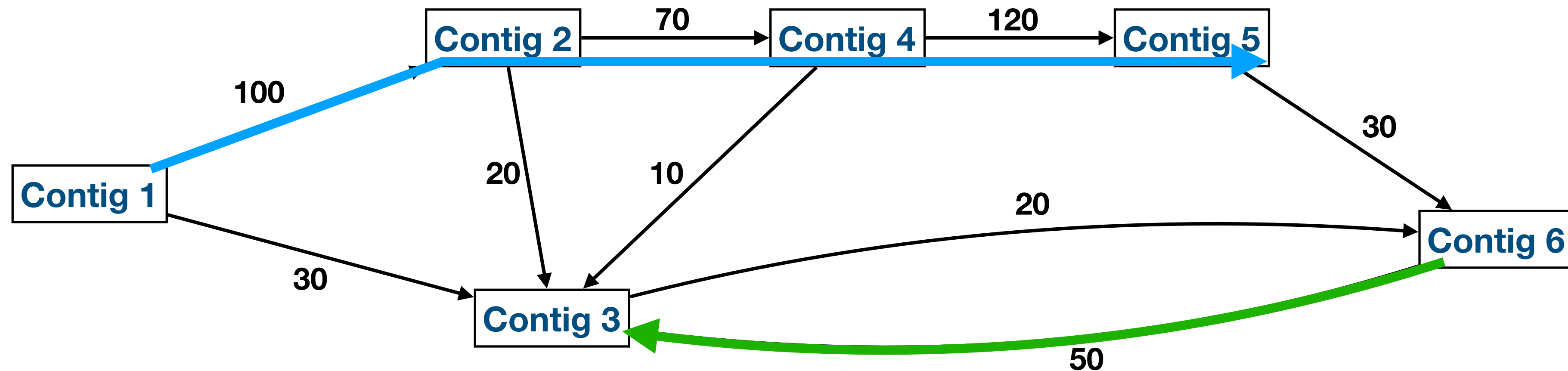
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Chaining (scaffolding) contigs



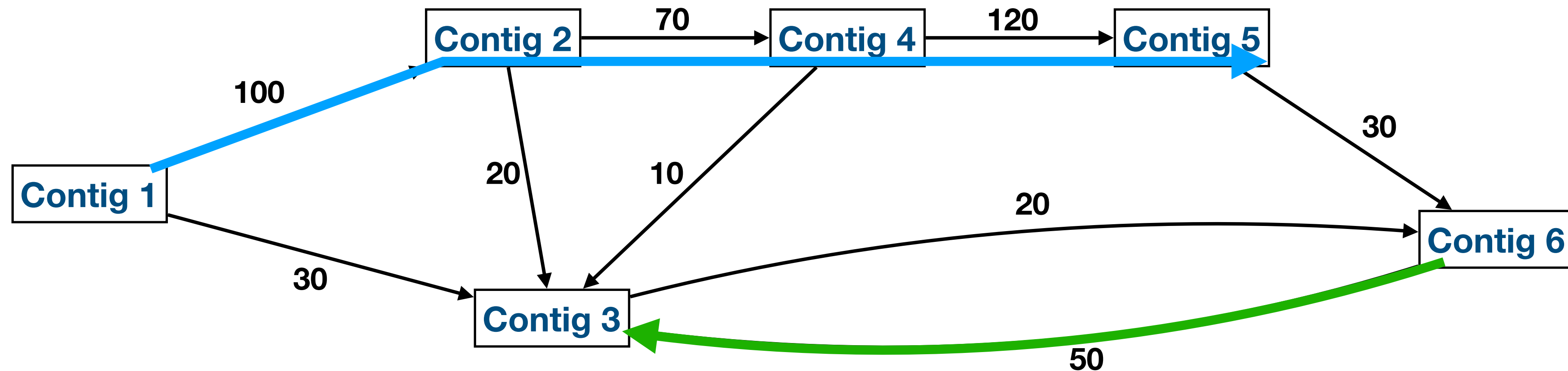
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Chaining (scaffolding) contigs



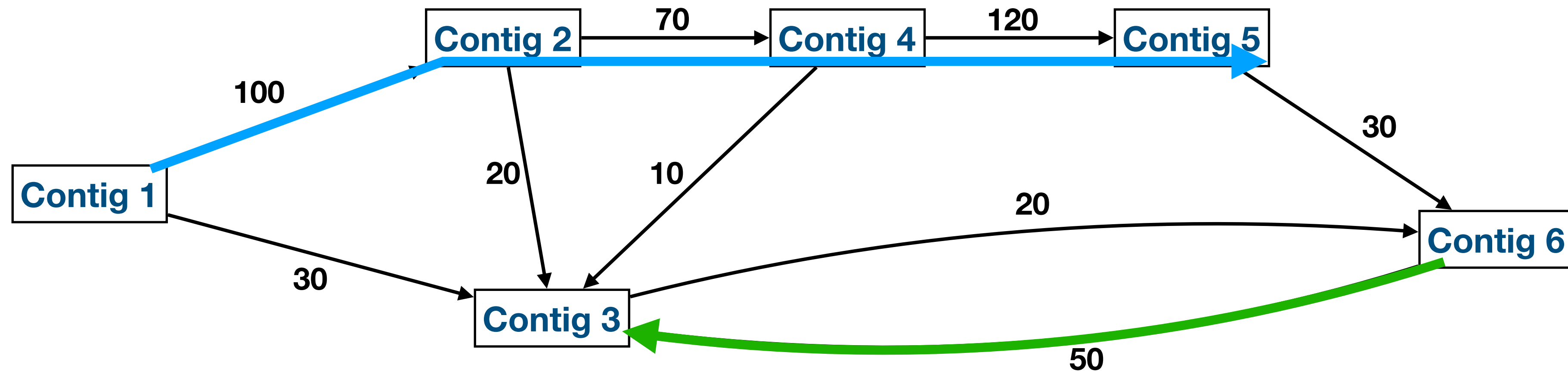
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Chaining (scaffolding) contigs



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Chaining (scaffolding) contigs



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Fast scaffolding with small independent mixed integer programs. Bioinformatics 27(23): 3259-3265 (2011)
 - Igor Mandric, Alex Zelikovsky:
ScaffMatch: Scaffolding Algorithm Based on Maximum Weight Matching. RECOMB 2015: 222-223

Gap filling

Scaffolds contain gap
length estimates
(number of Ns)

Bring back all reads



Find filling paths from the
assembly graph

Output the scaffolds in
which some gaps are "filled"

TCGATAGCTAAAANNNNNNNNNNAATTGTNNNATAGAGATATTT



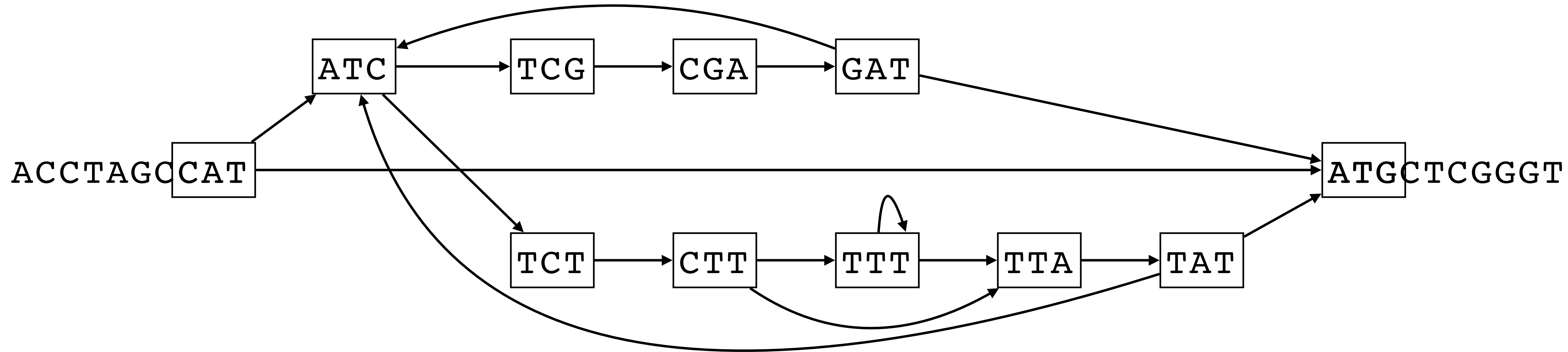
TCGATAGCTAAAATGCCGTTCGGAATTGTNNNATAGAGATATTT

Find path of given length

ACCTAGCCAT ← **Gap length 5** → ATGCTCGGGT

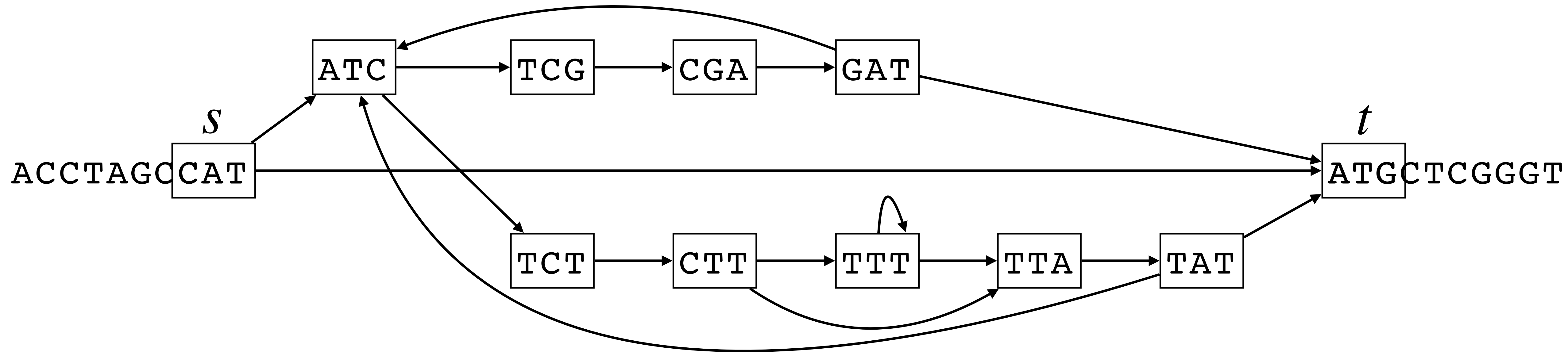
A horizontal red line with arrowheads at both ends connects the two DNA sequences. The text "Gap length 5" is written in red above the line.

Find path of given length



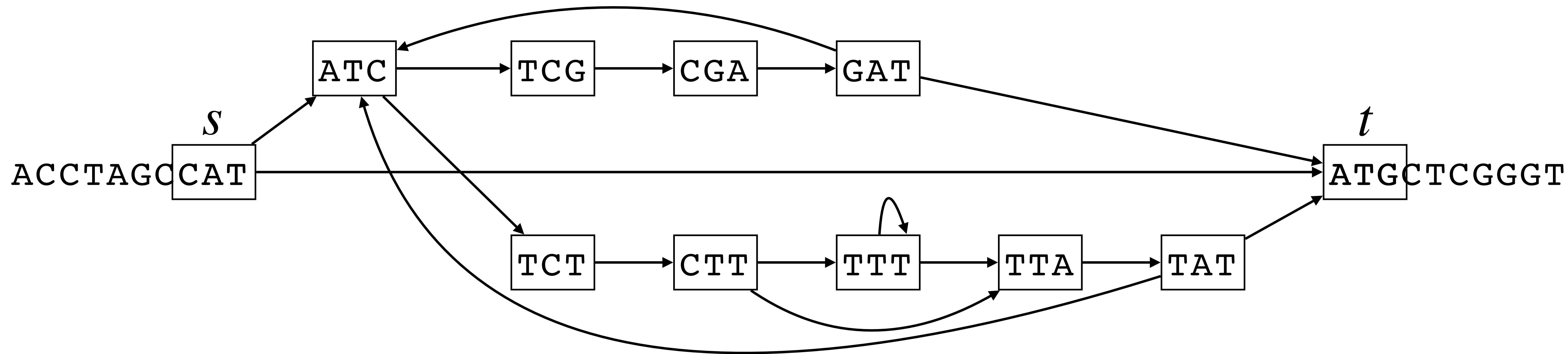
- Build the de Bruijn graph of all reads

Find path of given length



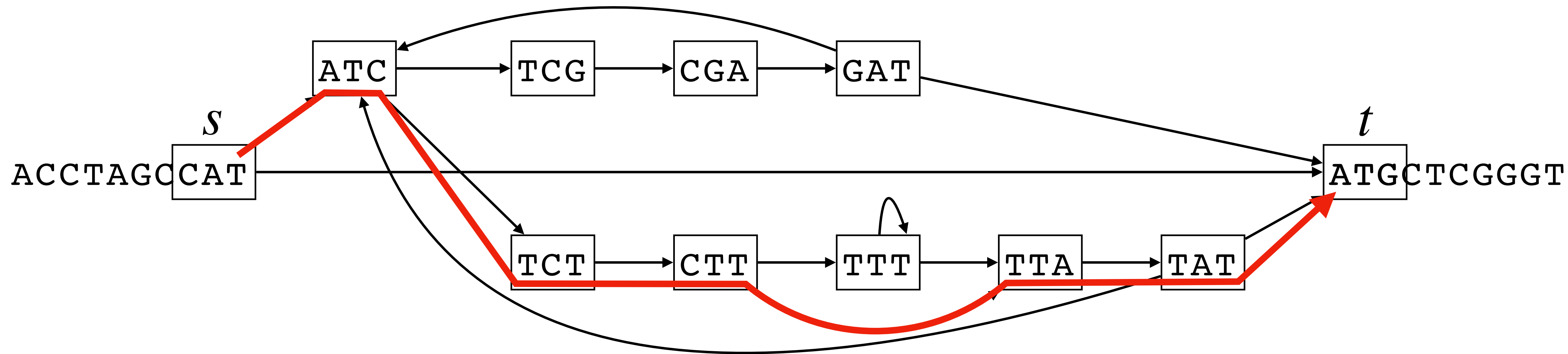
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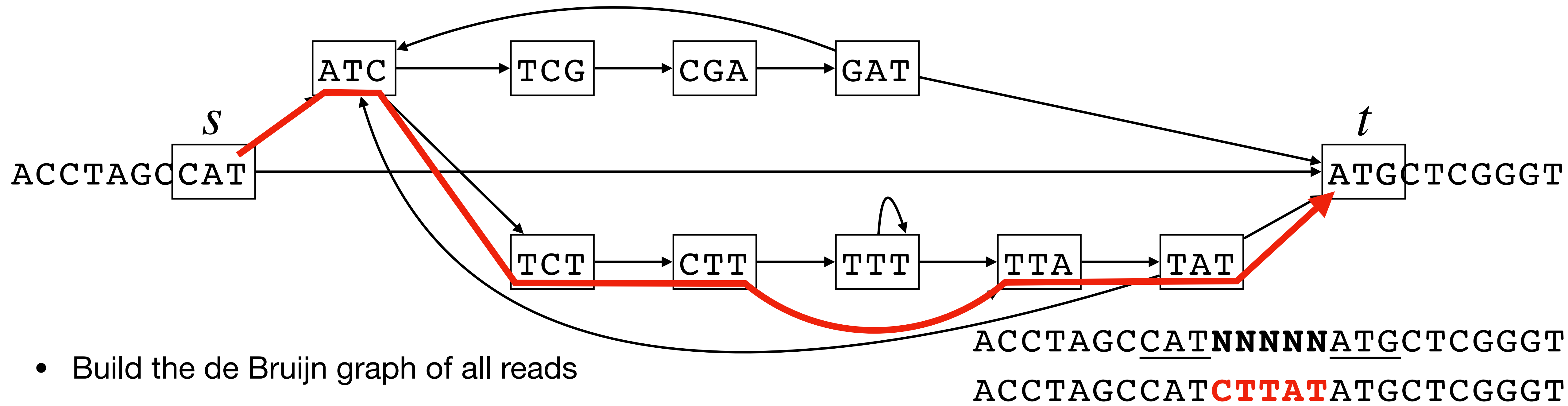
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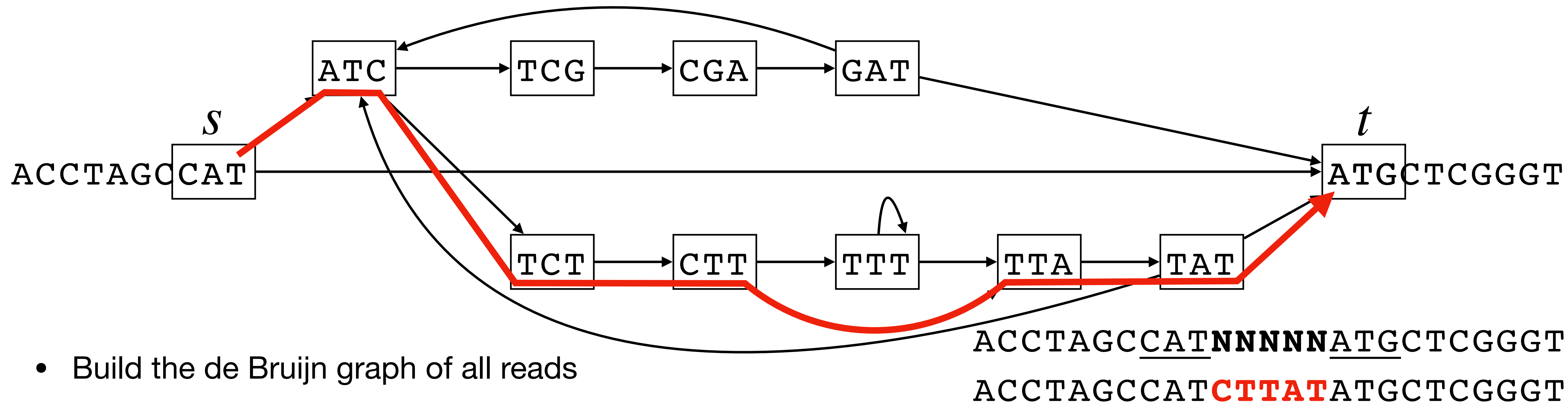
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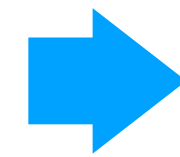
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- Can be solved by dynamic programming in time $O(d \mid \text{edges} \mid)$
 - Leena Salmela, Kristoffer Sahlin, Veli Mäkinen, Alexandru I. Tomescu:
Gap Filling as Exact Path Length Problem. RECOMB 2015: 281-292

Section summary

Contig assembly

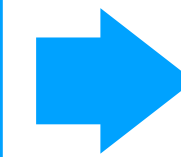
Forget about the assembly model (i.e. the problem formulation)

Assemble only parts about which we are sure (contigs → contiguous sequences)



Scaffolding

Chain (order) the contigs
Output chains of contigs with "gaps" (NNNN...) between them



Gap filling

Find filling paths from the assembly graph

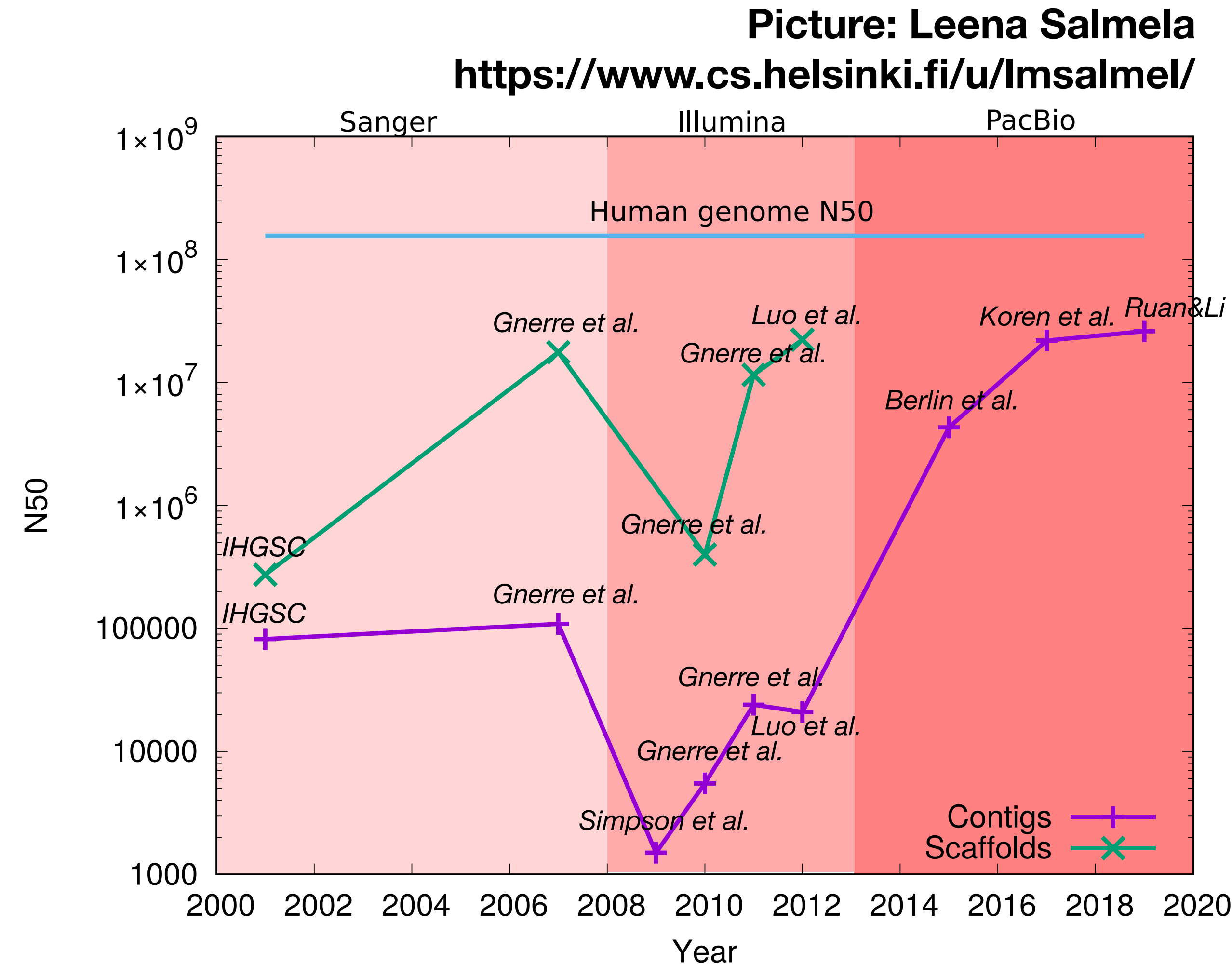
Output the scaffolds in which some gaps are "filled"

- A natural decomposition into subproblems based on the available paired-end information
- One can improve each step individually, thus improving the overall result

Long-read sequencing

(Third-generation sequencing)

- No paired-end reads (focus is on contig assembly)
- Higher error rate: 15% compared to 0.1% for short reads
 - ▶ Still developing: accurate PacBio HiFi reads
- No "clear" best strategy
- Short reads still relevant for some scenarios (e.g. metagenomic sequencing)



N50 measure → ASSIGNMENT

A more "practical" theoretical formulation

(A principled approach to contig assembly)

Back to contig assembly

- Goal: obtain sequences that are "guaranteed" to occur in the genome

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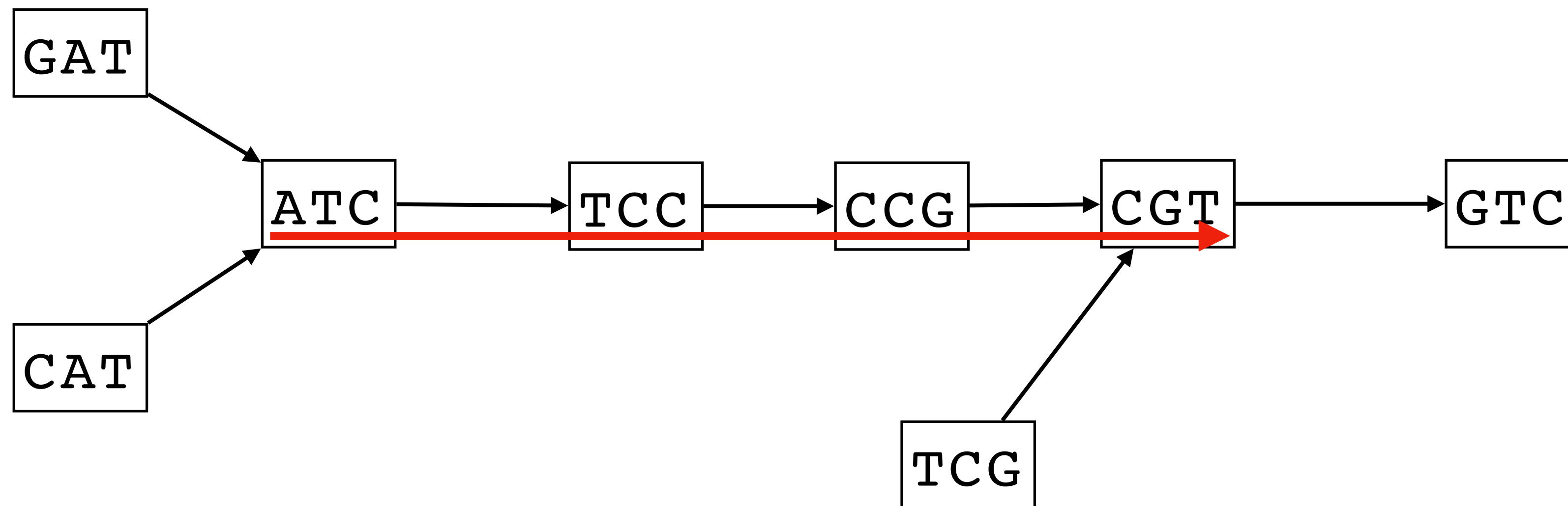
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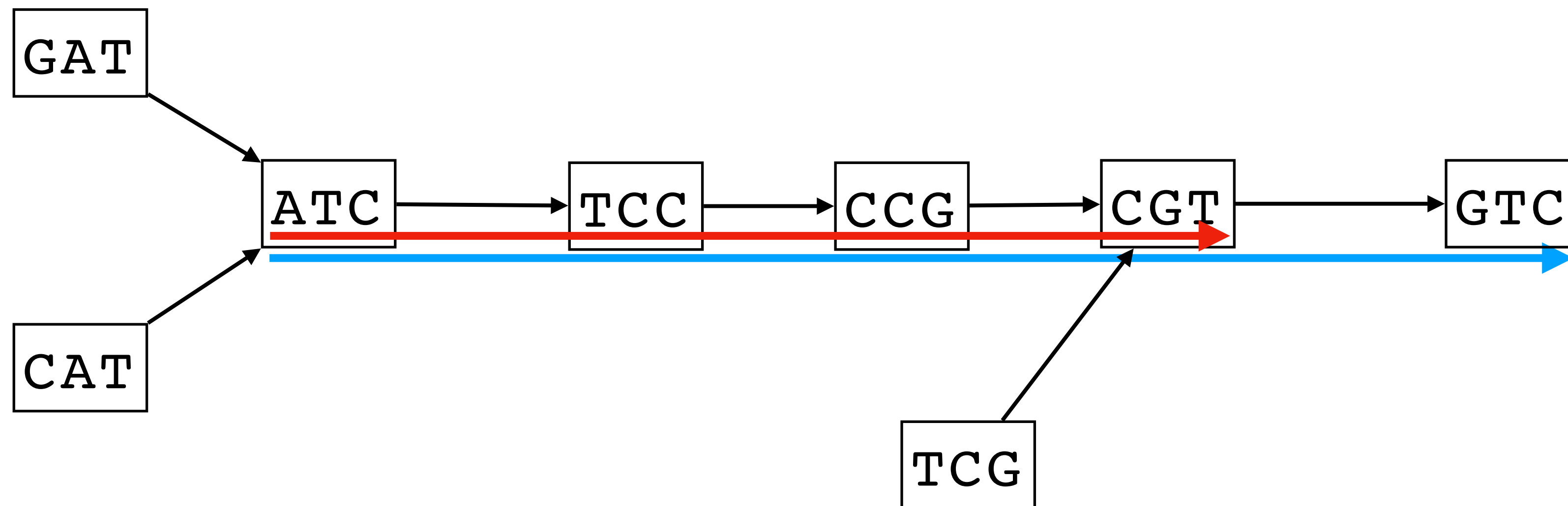
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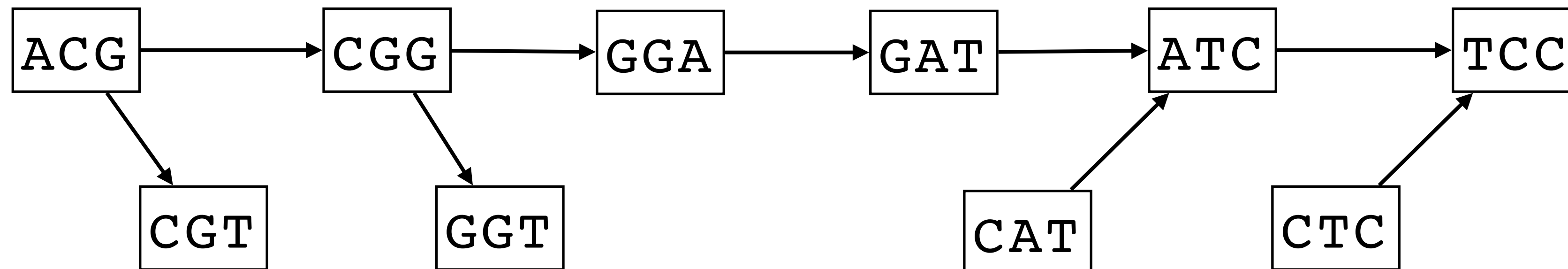


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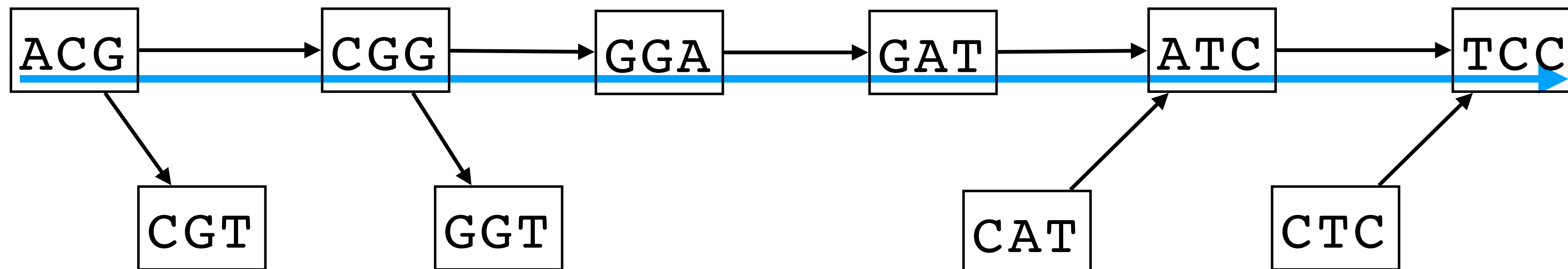
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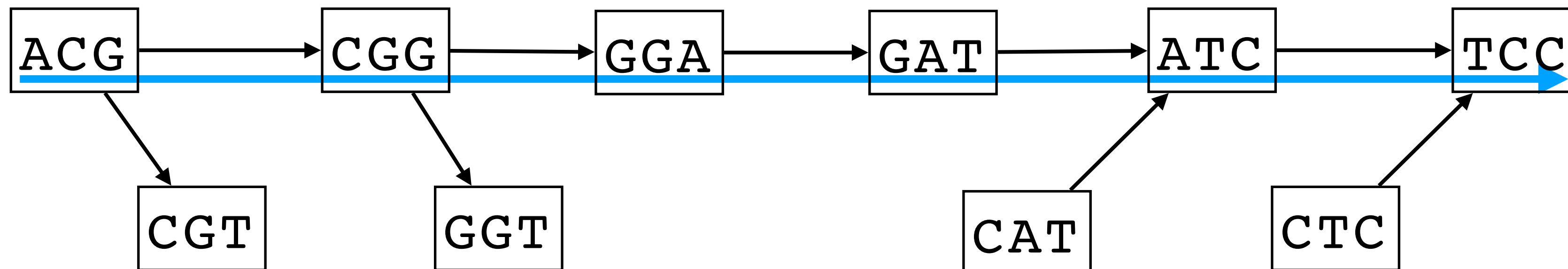
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- Assume that the "genome assembly solution" is a circular walk covering every edge at least once (walk can repeat nodes)
 - ▶ Trivial to find one, exponential to find all
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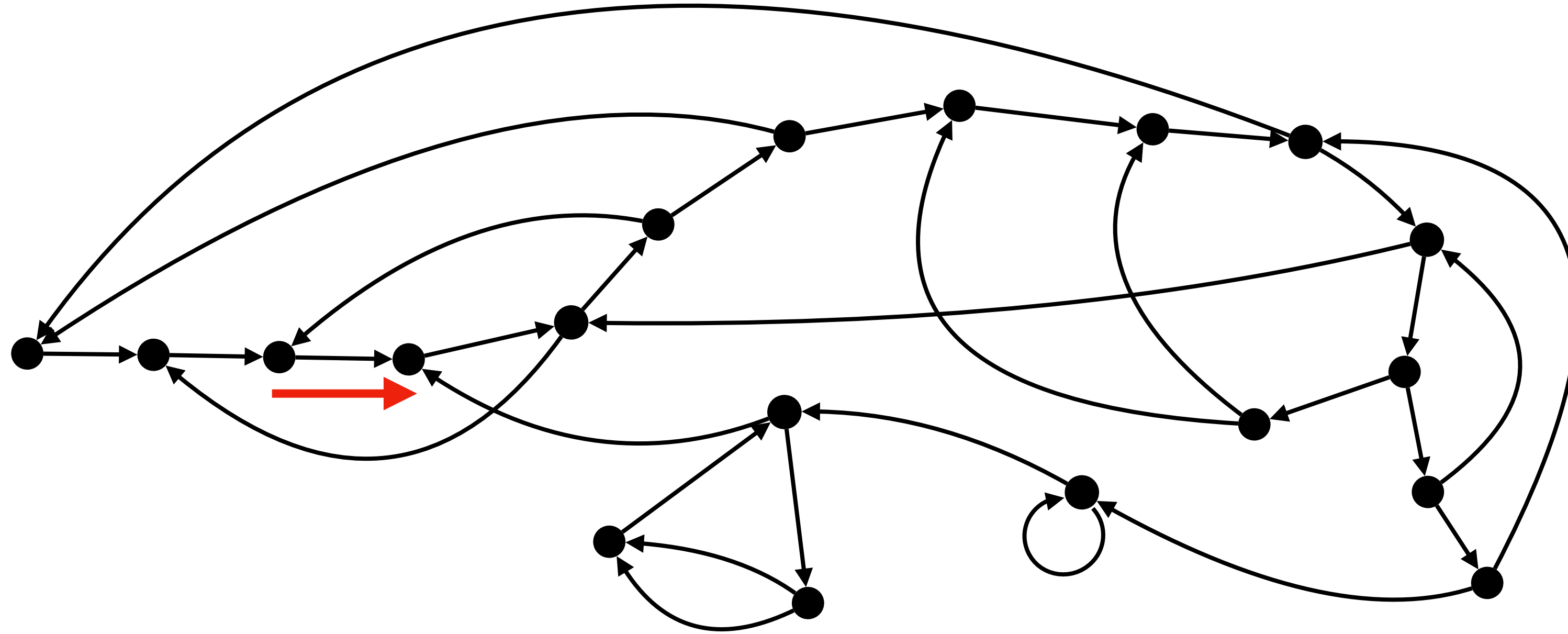
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An Optimal $O(nm)$ Algorithm for Enumerating All Walks Common to All Closed Edge-covering Walks of a Graph. ACM Trans. Algorithms 15(4): 48:1-48:17 (2019)

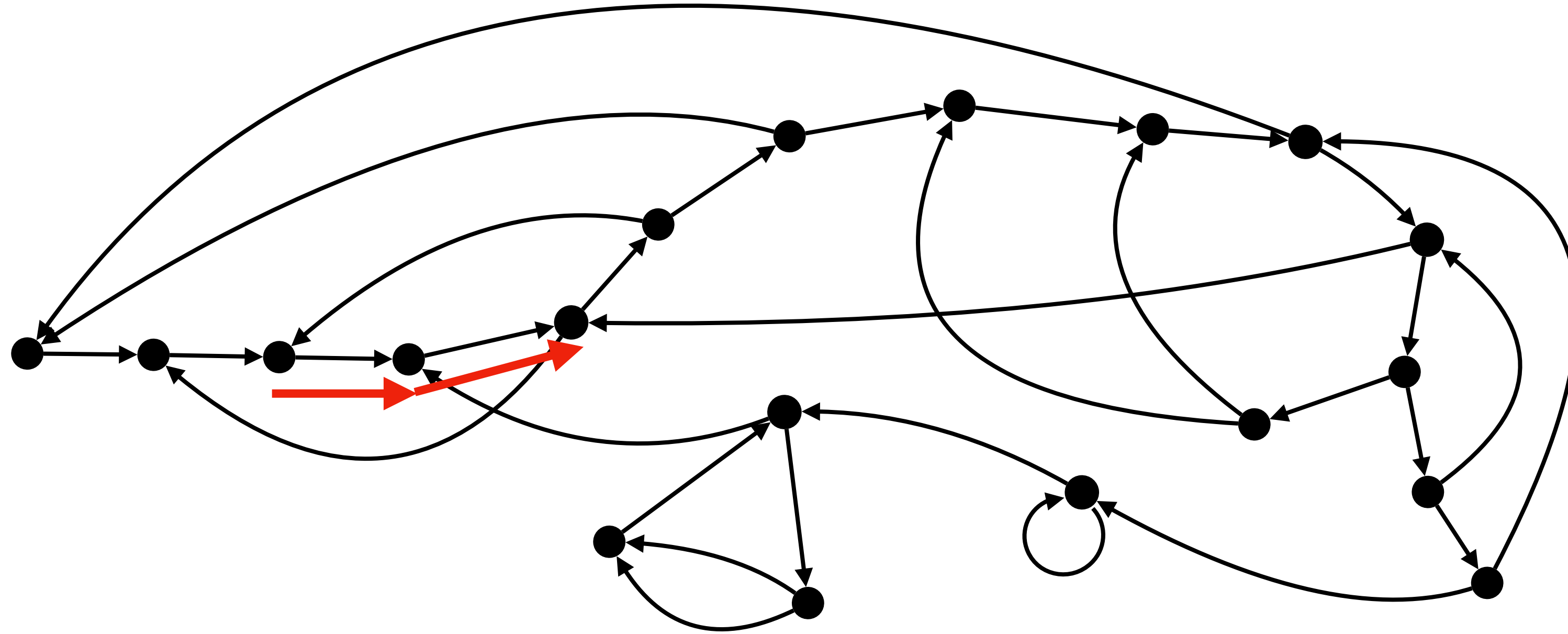
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- Can be adapted to deal with practical issues (ongoing work)
 - **Subprojects available as Master thesis topics**

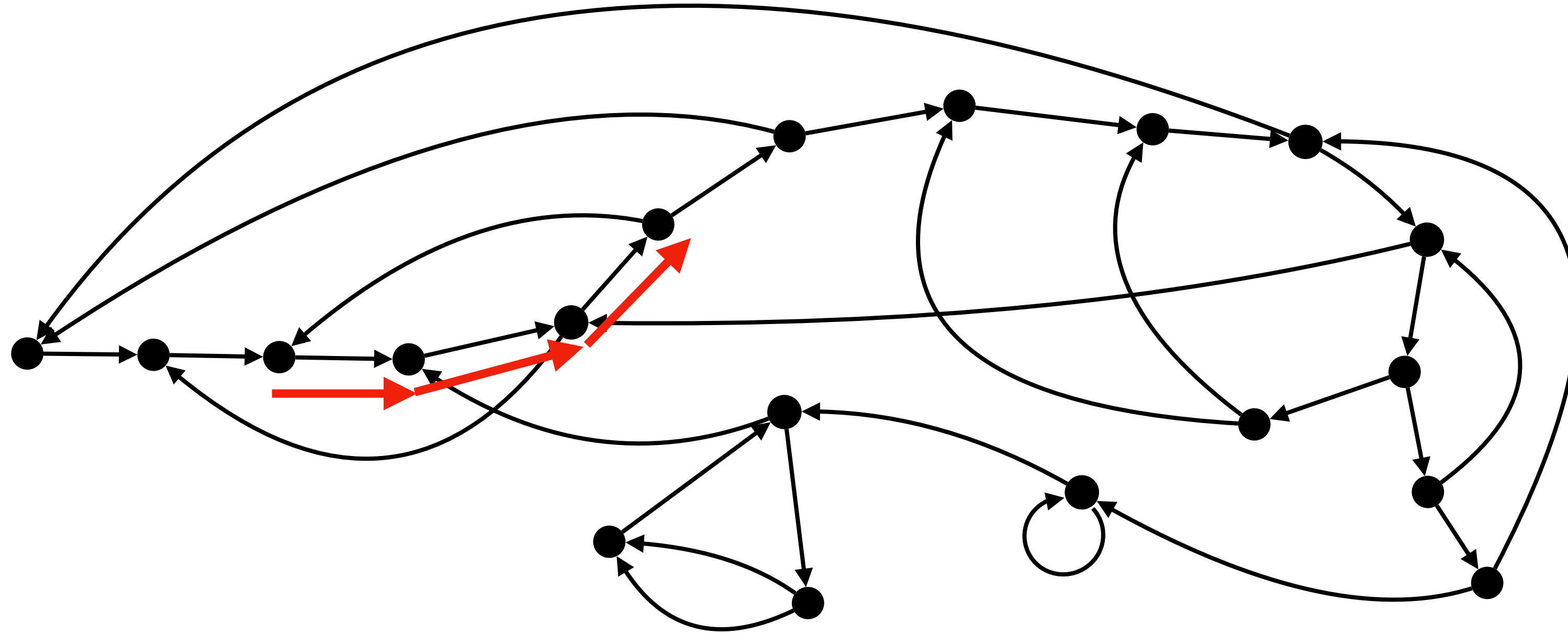
Example + Results on perfect data



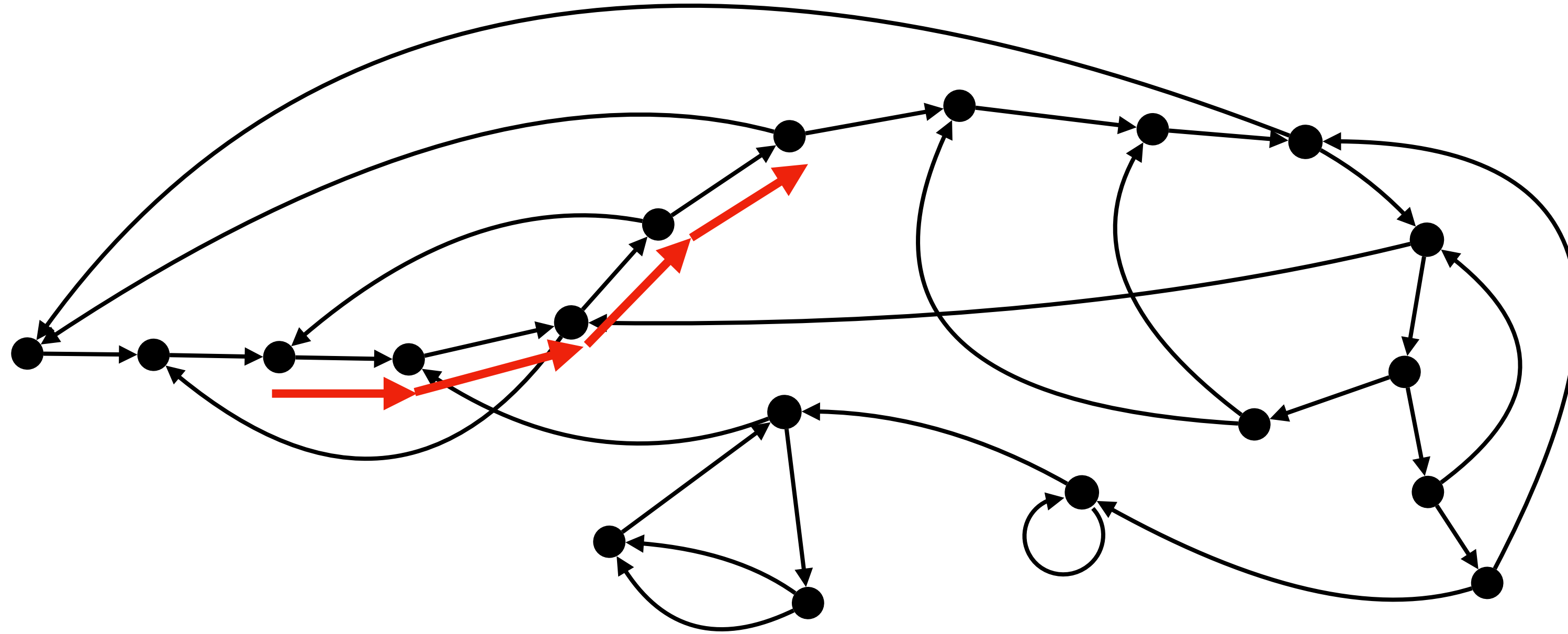
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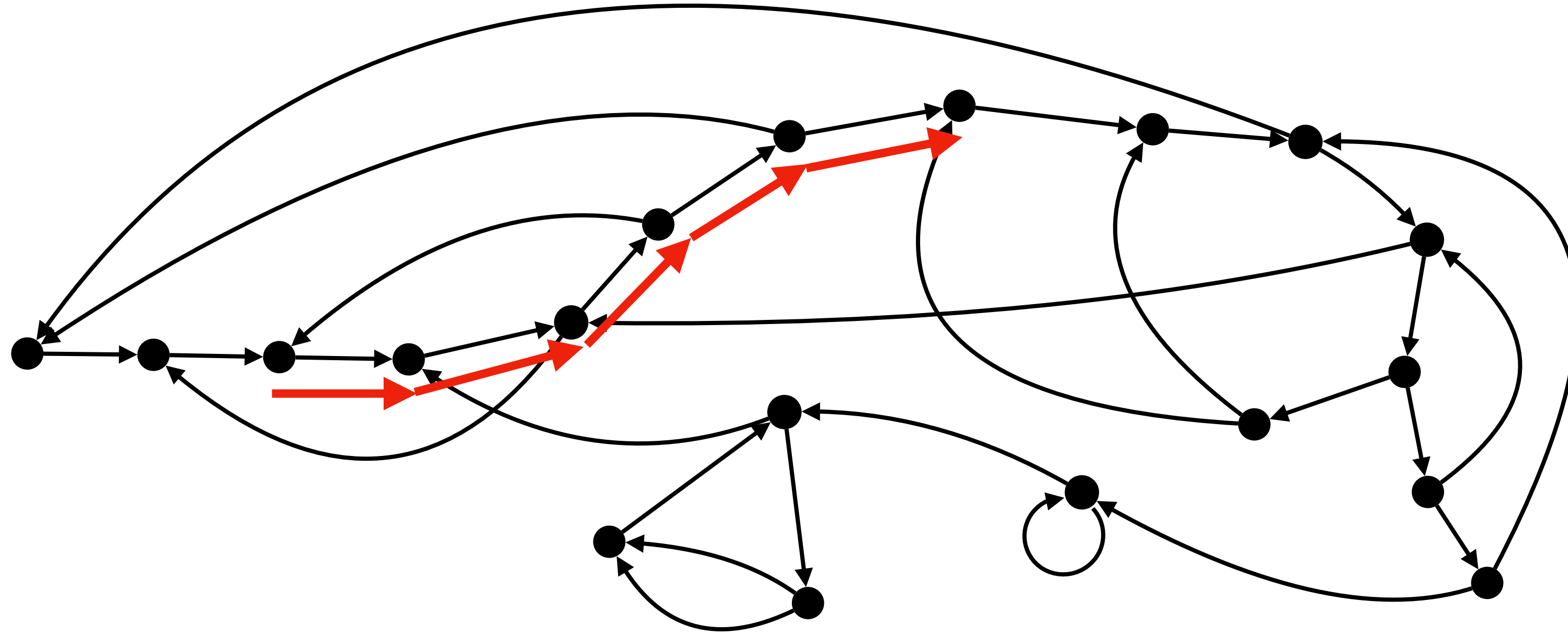
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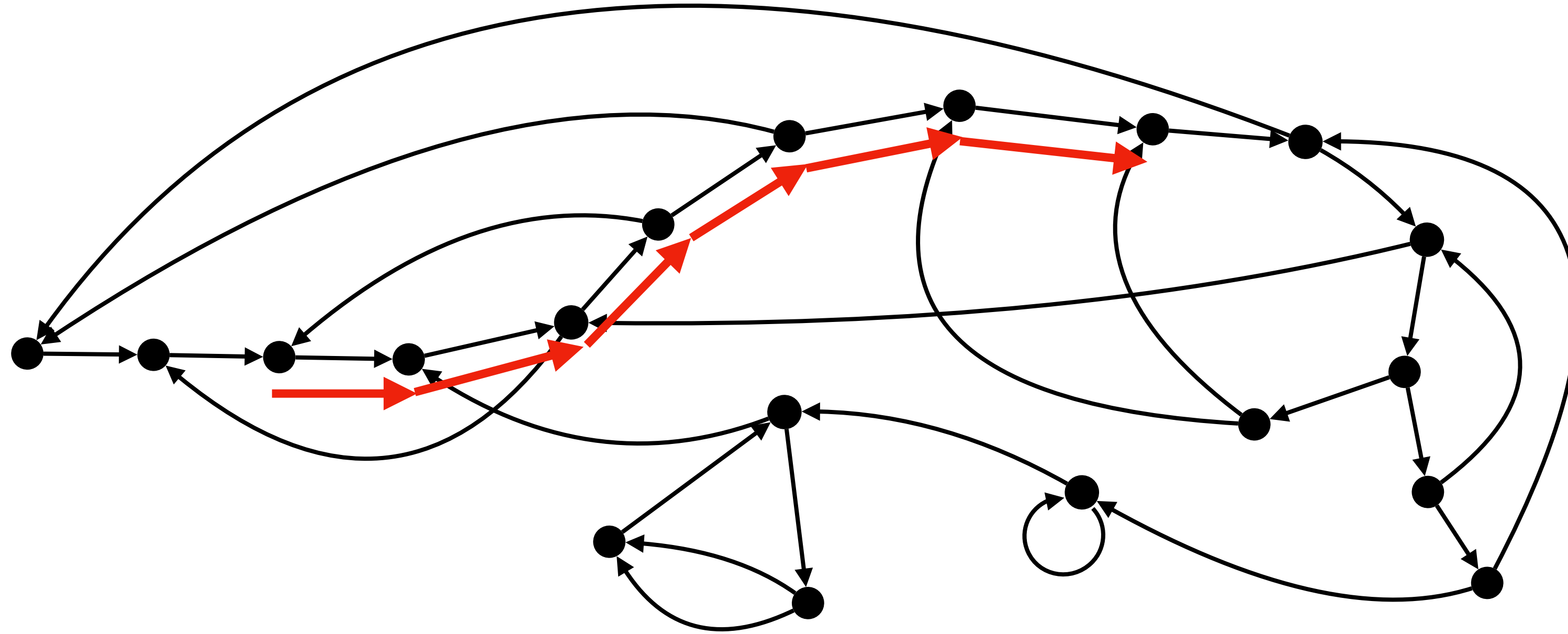
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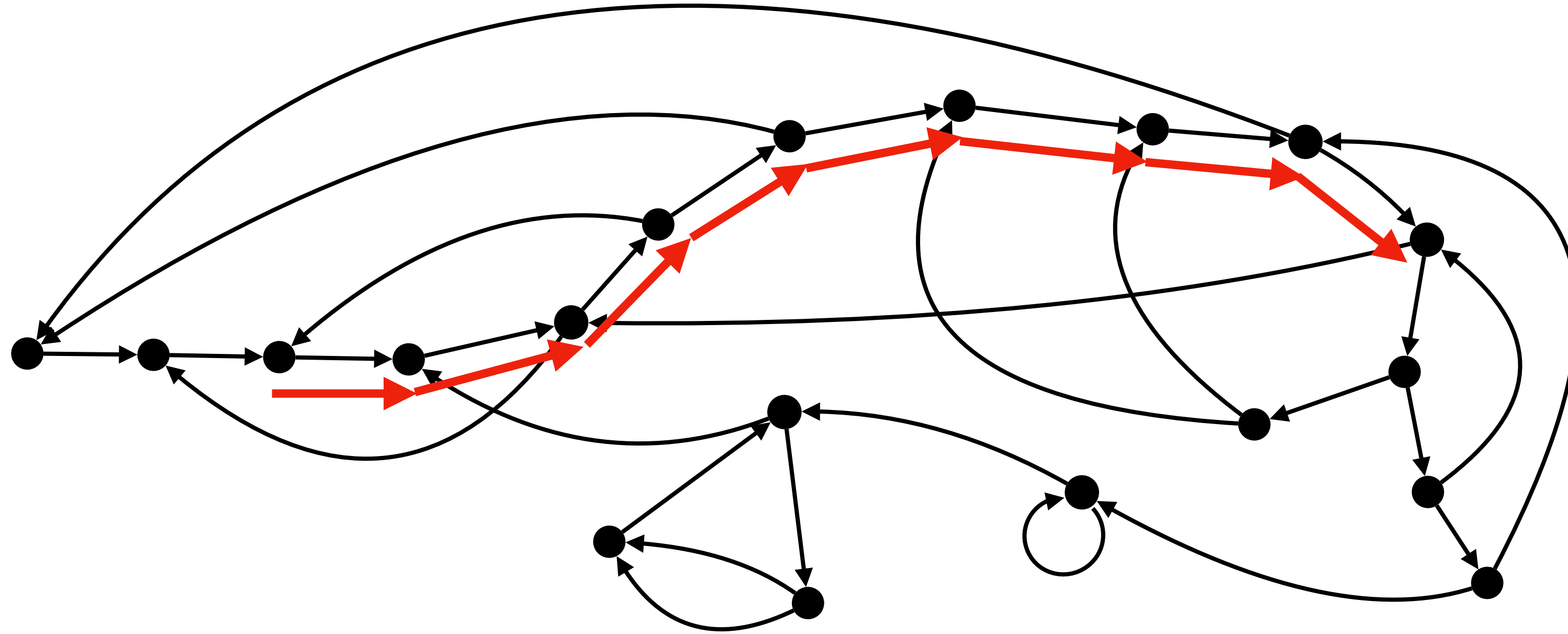
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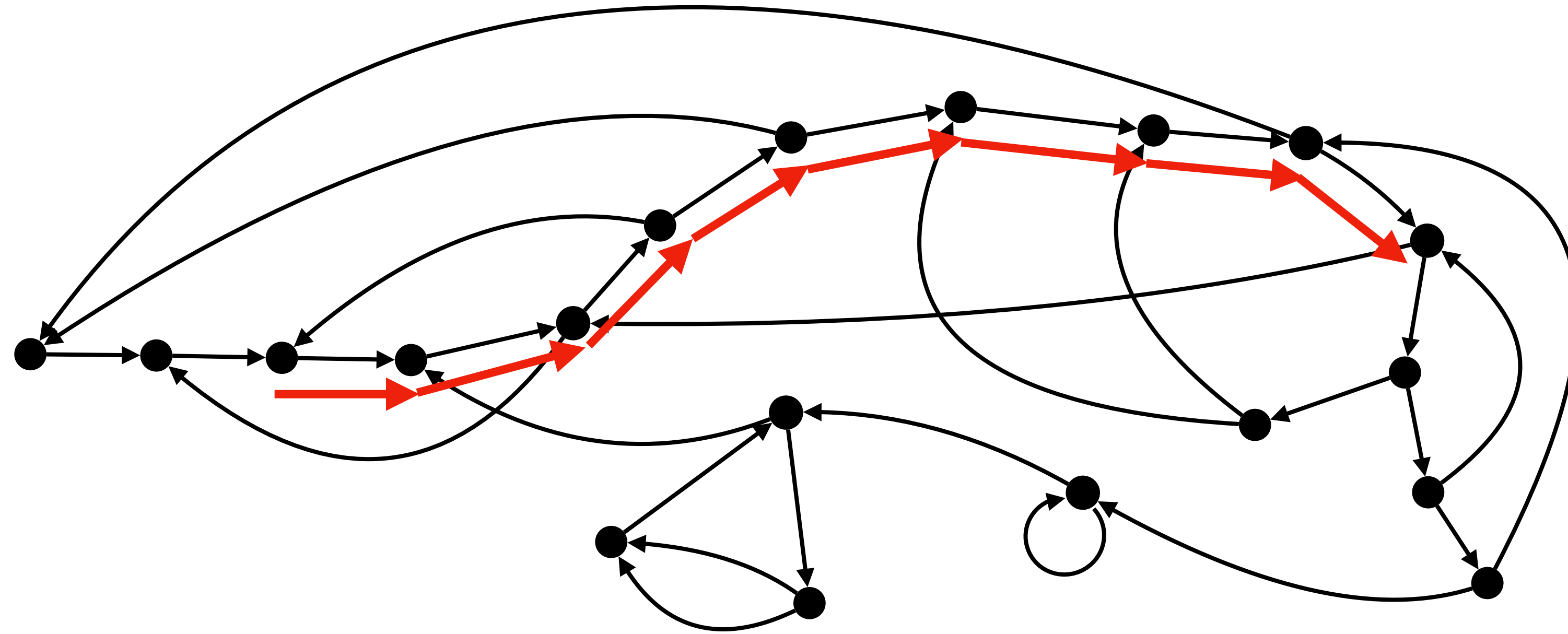
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Example + Results on perfect data



chr10, length 135M	#Strings	Avg. length	Avg. #SNPs / string
unitigs	260K	546	26
omnitigs	158K (-40%)	887 (+62%)	41 (+58%)

Section summary

Theory is important,
but more so when
it is motivated by practice