Isolation of pathogenic *Escherichia coli* from buffalo meat sold in Parbhani city, Maharashtra, India

C. S. Shekh, V. V. Deshmukh, R. N. Waghamare, N. M. Markandeya and M. S. Vaidya

College of Veterinary and Animal Sciences, MAFS University, Parbhani - 413 402, Maharashtra, India Corresponding author: R. N. Waghamare email: rupeshwaghmare@gmail.com Received: 18-07-2012, Accepted: 29-09-2012, Published online: 23-02-2013

How to cite this article:

Shekh CS, Deshmukh VV, Waghamare RN, NM Markandeya and Vaidya MS (2013) Isolation of pathogenic *E. coli* from buffalo meat sold in Parbhani city, Maharashtra, India, *Vet. World* 6(5): 277-279, doi: 10.5455/vetworld.2013.277-279

Abstract

Aim: Isolation, characterization, in-vitro pathogenicity and antibiogram study of E.coli from buffalo meat sold in Parbhani city.

Materials and Methods: Meat samples were collected from buffalo immediately after slaughter. Isolation, identification and enumeration of *E. coli* were done by following standard methods and protocols. Hemolysin test and Congo red binding assay were used to study *in-vitro* pathogenicity of *E. coli* isolates. Disc diffusion method was used to study antibiogram of pathogenic *E. coli* isolates.

Results: A total of 250 buffalo meat samples were collected and processed. A total of 22 (8.80 percent) *E. coli* isolates were isolated with average differential count of $1.231 \pm 0.136 \log_{10} \text{ cfu/g}$ on EMB agar. All the *E. coli* isolates were confirmed by Grams staining, biochemical reactions and sugar fermentation and motility tests. A total of 9 (3.6 percent) *E. coli* isolates were found to be pathogenic by *in-vitro* pathogenicity testing. Antibiogram studies of pathogenic *E. coli* isolates showed that all 9 isolates were sensitive to gentamycin ($20 \pm 1.49 \text{ mm}$) while 7 isolate showed resistance to enrofloxacin ($18.22 \pm 3.58 \text{ mm}$) and tetracycline ($11.44 \pm 2.04 \text{ mm}$).

Conclusion: Buffalo meat sold in Parbhani city is an important source of *E. coli* infection to human population. A total of 9 pathogenic *E. coli* were isolated from buffalo meat immediately after slaughter. All isolates were characterized and confirmed pathogenic by *in-vitro* pathogenicity tests. Antibiogram studies of all isolates revealed sensitivity to gentamicin and resistance to tetracycline and enrofloxacin.

Keywords: antibiogram, buffalo meat, *E. coli*, pathogenicity

Introduction

India is a leading exporter of buffalo (Bubalus bubalis) meat earning foreign exchange. The production of buffalo meat (beef) is increasing by 4.5 percent per annum [1]. According to Animal Husbandary statistical database of Department of Animal Husbandary, Dairying and Fisheries (DADF), there are a total of 5,520 recognized and 4,707 unrecognized slaughter houses in the country. The buffalo carcass has less fat, less bone and higher proportion of muscle than cattle [2]. Sanitary indices refer to certain organisms or group of organisms which indicates presence of potentially pathogenic or spoilage organisms in food. Coliform organisms in food indicate poor sanitary practices and presence of other hazardous organisms. This group includes E. coli and enumeration of E. coli counts is important to determine sanitary indices [2]. Presence of E. coli in food is an indicator of poor sanitary conditions during processing. Isolation of E. coli from meat and meat products is a common phenomenon [3, 4]. Studies showed that normal intestinal microflora specially commmensal E.coli strains under specific condition, might serve as reservoir of resistance genes that could be acquired by pathogenic bacteria [5]. Since commensal bacterial strains are exposed to the same selective pressure as pathogenic strains, they might be used as an indicator of trends in antimicrobial resistance [6]. Also E. coli being a member of Enterobacteriacae is being exposed to various antibiotics and

antimicrobial agents in maintenance of hygiene in animal husbandry practices as well as during processing of meat. This results into development of drug resistant condition. Antibiotic resistance to beef borne *E. coli* is well documented [8,9].

The growing problem of antibiotic resistant has become a significant public health concern world wide. All uses of antimicrobial in human medicine and animal husbandry create selective pressure that favors emergence of antimicrobial resistance among microorganism [7].

Keeping in view the public health importance and hazards caused by pathogenic *E. coli*, the present study was undertaken to isolate and identify these pathogenic and drug resistant organisms from buffalo meat sold in Parbhani city.

Materials and Methods

Sample collection and processing: Buffalo beef samples were collected from Parbhani Municipal Council Abattoir. A total of 250 meat samples approximately weighing 50 gm were collected from neck site of slaughtered animal in sterile polyethylene schachets for the period from December 2011 to May 2012. These meat samples collected were brought to the laboratory on ice within one hour and processed immediately.

Isolation, identification and enumeration of *E. coli*: Isolation of *E. coli* was done as per the method

www.veterinaryworld.org 277

Table-1. Results of sensitivity pattern of E. coli isolates

Antibiotics	S	I	R	Average zone of Inhibition in mm
Gentamycin	9	0	0	20.00 ± 1.49
Enrofloxacin	2	0	7	18.22 ± 3.58
Tetracycline	1	1	7	11.44 ± 2.04

S - Sensitive, I - Intermediate sensitive, R - Resistant

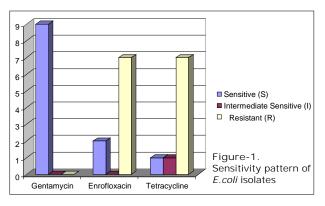
described in BAM [10]. A quantity of 10gm of beef sample was minced with the help of sterile scissors and mixed in 90 ml normal saline solution (pH 7.2) in screw cap bottle. Ten fold serial dilutions were made up to 10⁻⁵ dilution in normal saline solution (pH 7.2). A quantity of 0.1 ml inoculam from 10⁻³ and 10⁻⁴ dilutions was used by spread plate technique on Eosin Metheline Blue agar (EMB) (Himedia Laboratories, Mumbai). Incubation was done at 37°C for 24 hrs and colony counts were taken. Typical characteristics colony of *E. coli* on EMB agar as greenish metallic sheen was enumerated and isolated. Identification of *E. coli* was done by biochemical test i.e IMViC, catalase, oxidase & nitrate reduction and sugar fermentation and motility tests. [11].

In-vitro pathogenicity test of E. coli: In-vitro pathogenicity test of confirmed E. coli isolates was done by heamolysis and Congo red binding assay [12,13]. Colonies showing hemolytic zone are considered as heamolytic positive. While isolates, producing intense orange or brick red colour on Congo red medium were considered as positive and those producing grayish white colonies recorded as negative.

Antibiogram of *E. coli*: Antibiotic sensitivity of all pathogenic *E. coli* isolates was done by disc diffusion method [14] on Muller-Hinkton agar. Three antimicrobial discs were used with different concentrations viz, Gentamycin (10 mcg), Tetracycline (30mcg) and Enrofloxacin (10mcg). Zones of complete inhibition were measured in millimeter with rule.

Results

250 buffalo meat samples were screened for isolation of $E.\ coli$ on EMB agar. A total of $22\ E.\ coli$ isolates were obtained. The isolates were identified based upon colony characteristics comprising of bluish green metallic sheen. The percentage of $E.\ coli$ isolation observed was 8.80 percent. Having average differential count of 22 isolates of $E.\ coli$ on EMB agar observed was $1.231 \pm 0.136 \log_{10} \text{cfu/gm}$. A total of $22\ E.\ coli$ isolates were subjected to identification based upon Grams staining, biochemical characters, sugar fermentation and motility tests. All the 22 isolates were subjected to in-vitro pathogenecity test that is hemolysin test and Congo red binding assay. A total of $9\ E.\ coli$ isolates were found positive for pathogenecity in both the in-vitro pathogenecity tests. The percent



positivity of hemolysin test and Congo red binding assay observed was 3.60 percent. All 9 pathogenic *E. coli* isolates were subjected to antibiotic sensitivity test against gentamycin, tetracycline and enrofloxacin. The zones of inhibition of antibiotic were recorded. The average zone of inhibition seen were gentamicin (20.00 ±1.49mm), enrofloxacin (18.22±3.58mm) and tetracycline (11.44±2.04 mm). All 9 isolates were sensitive to gentamicin while 7 isolates showed resistance to enrofloxacin and Tetracycline. A complete sensitivity of all 9 isolates (100 percent) was observed against gentamicin.

Discussion

In present study, very low percentage (08.80 percent) of E. coli was isolated. The findings of present study are in agreement with earlier reports [3,15]. The E. coli counts are in the of range 6.85 to 7.40 \log_{10} cfu/g [16]. Scanning of available literature reveals that percentage of E. coli isolation and identification varies considerably [17,18]. The variation in percentage of E. coli isolation and identification may be due to difference in hygienic conditions at different slaughter houses

All the 22 *E. coli* isolates shown typical Grams staining reaction, biochemical reactions, motility and sugar fermentation reactions described earlier [11]. Many workers successfully used staining characters, biochemical reactions, sugar fermentation reactions and motility patterns for confirmation of *E. coli* isolated from meat [19].

Hemolysis of sheep RBC is an important criterion for identification of pathogenic strains of *E. coli. Invitro* pathogenecity studies of hemolytic *E. coli* were done by using 5 percent sheep blood agar [20]. Many workers successfully used Congo red binding assay in tryptose soya agar for identification of pathogenic *E. coli* [20-22]. The observations are in agreement with earlier workers [20,21].

Antimicrobial sensitivity testing of the pathogenic *E. coli* isolated from beef samples confirms that all 9 pathogenic isolates were highly sensitive to Gentamycin. Out of nine isolates 7 isolates shown resistance to Enrofloxacin and Tetracycline. Similar reports in previous studies showed resistance of meat borne *E. coli* to Gentamycin and Enrofloxacin [23], Tetracycline and Ciprofloxacin [24,25]. In present study sensitivity of *E. coli* to Tetracyclin was observed

www.veterinaryworld.org 278

in 2 isolates but at low level.

The resistance of pathogenic *E. coli* to the antibiotics may be due to indiscriminate use of antibiotics in animal husbandry practices. Pathogenic *E. coli* of meat origin are always having grater potential to enter into food chain. Buffalo meat, beef and other types of meat are cheap source of human infection [4,26].

Conclusion

Buffalo meat sold in Parbhani city is an important source of *E. coli* infection to human population. All isolates were characterized and confirmed pathogenic by *in-vitro* pathogenicity tests. Antibiogram studies of all isolates revealed sensitivity to gentamicin and resistance to tetracycline and enrofloxacin.

Authors' contribution

The present work was carried out by CSS under the guidance of VV with the technical support of RW. RW drafted and revised the manuscript. All authors read and approved the final manuscript.

Acknowledgements

The authors are thankful to the Associate Dean, College of Veterinary and Animal Sciences, Parbhani, Maharashtra for providing the facilities to pursue this work.

Competing interests

Authors declares that they have no competing interest.

References

- USDA, (2012) Livestock and poultry: world market and trade. Annual report of the year 2012.
- Sherikar A. T., V. N. Bachil and D. C. Thaplial (2011) Text book of Elements of Veterinary Public Health. Indian Council of Agriculture Research, New Delhi.
- 3. Elimali Mehmet., Zeynep Ulukanli, Hilmi Yaman, Mehmet Tuzcu, Kenan Genctav and Perihan Cavli (2005) A Seven Month Survey for the Detection of *E. coli* O157:H7 from Ground Beef Samples in the Markets of Turkey, *Pakistan J. Nutrition* 4(3): 158-161.
- 4. Islam M. A., Abdus S. Mondol, Enne de Boer, Rijkelt R. Beumer, Marcel H. Zwietering, Kaisar A. Talukder and Annet E. Heuvelink (2008) Prevalence and Genetic Characterization of Shiga Toxin-Producing *Escherichia coli* isolates from Slaughtered Animals in Bangladesh. *J. Appl. and Environ. Microbio.*, 74 (17) 5414–5421.
- Ajiboye R., O. Solberg, B. Lee, E. Raphael, C. DebRoy and L. Riley (2009) Global spread of mobile antimicrobial drug resistance determinant in human and animal Escherichia Coli and Salmonella strains causing community-Acquired infections. Clin. Infect Dis. 49(3): 365-371.
- Bywater R., H. Delyker, E. Deroover, A. D. Jong, H. Marion, M. MacConville, T. Rowans, T. Shryock, D. Suster, V. Thomas, M. Valle and J. Walters (2004) A European survey of antimicrobial susceptibility among zoonotic and commensal bacteria isolated from food producing animals. *J. Antimicro. Chemo.* 54(4):744-754.
- 7. Acar J. F. and G. Moulin (2006) Antimicrobial resistance at farm level. *Sci Tech Rev*. 25(2): 775-792.
- Schroeder C. M., D. G. White, Ge B, Y. Zhang, P. Fmcdermott, S. Ayers, S. Zhao and Meng J. (2003) Isolation of antimicrobial resistant *Escherichia coli* from retail meats purchased in Grater Washington DC, USA. *Int. J. Food Microbiol.* 85(1-2): 197-202.
- Rao T. S., J. P. S. Gill, G. V. V. P. S. Ravikumar and Sandeep Ghatak (2011) Multidrug resistance pattern of shiga-toxin

- producing *Escherichia coli* (STEC) and non STEC isolates from meat foods, drinking water and human diarrhoeic samples of Panjab, India. *Archiv. Clin. Microbiol.* 2:2.
- BAM (1998) Bacteriological Analytical Manual, 8th edition publication by FDA, U.S.
- Cowan S. T. and K. J. Steele (1993) Characters of Gram positive bacteria. In Cowan and Steel's manual for Identification of Medical bacteria 3rd edn. Cambridge Univ. Press.
- Agarwal R. K., K. N. Bhilegaokar, D. K. Singh, A. Kumar and R. S. Ratore (2003) Laboratory manual for the isolation and identification of food borne Pathogens. 1st Edn. ICAR, New Delhi, pp. 55-69.
- 13. Ishiguro E. E., T. Anisworth, J. T. Trust and W. W. Kay (1985). Congo red, a differential medium for *Aeromonas salmonicida*, detects the presence of the cell surface protein array involved in virulence *J. of Bacteriol*. 164 (3): 1233.
- Anonymous (2001) Performance standards for antimicrobial susceptibility testing; National Committee of Clinical Laboratory Standards (NCCLS) Document M100-S11, NCCLS, Pennsylvania, 19087 USA.
- Hazarika R. A., D. K Singh, K. N. Kapoor, R. K. Agarwal (2004a) Prevalence of different serotypes of *Escherichia coli* in Buffalo meat and its product. *Ind. J. Comparative Microbio., Immunology and Infec. Dises*. 25 (1) 19-22.
- Clavero M., S. Rocelle; Beuchat, R. Larry and M. P. Doyle (1998) Thermal Inactivation of *Escherichia coli* O157:H7 Isolated from Ground Beef and Bovine Feces, and Suitability of Media for Enumeration, J. Food Protc. 61(3) 285-289.
- Hazarika R. A., D. K. Singh, K. N. Kapoor, R. K. Agrawal, A.B. Pandey and D. N. Rajkumar (2004b) Detection and characterization of verotoxin producing Escherichia coli (VTEC) isolated from buffalo meat., *J. food Safety* 24 (4): 281-190.
- 18. Rahimi Ebrahim., Hamid Reza Kazemeini and Mohammad Salajegheh (2012) *E. coli* O157:H7/NM prevalence in raw beef, camel, sheep, goat, and water buffalo meat in Fars and Khuzestan provinces, Iran *Vet. Res. Forum* 3(1) 13-17.
- Leclercq Alexandre., Bernard Lambert, Denis Pierard and Jacques Mahillon (2001) Particular Biochemical Profiles for Enterohemorrhagic *Escherichia coli* O157:H7 Isolates on the ID 32E System, *J. Clin. Microbiol.* 39(3):1161-1164.
- Raji M. A., J. O. Adekeye, J. K. P. Kwaga and J. O. O. Bale (2003) In vitro and in vivo pathogenicity studies of *Escherichia coli* isolated from poultry in Nigeria, *Israel J. Vet. Med.* 58(1).
- Chavan Somnath K., D. R. Kalore and A. A. Nagdire (2012).
 Pathogenic attributes of *E. coli* isolated from commercial broilers, *Indian Vet. J.* 89(1): 39-40.
- Roy P., V. Purushothaman, A. Koteeswaran, and A. S. Dhillon (2006) Isolation, Characterization, and Antimicrobial Isolated from Japanese Quail Drug Resistance Pattern of *Escherichia coli* and their Environment, *J. Appl. Poult. Res.* 15:442–446.
- Osterblada M., E. Kilpia, A. Hakanena, L. Palmub and P. Huovinena (1999) Antimicrobial resistance level of enterobacteria isolated from minced meat. J. Antimicrob. Chemother. 44(2): 298-299.
- 24. Khan Asis, S. C Das, T. Ramamurthy, A. Sikdar, J. Khanam, S. Yamasaki, Y. Takeda and G. Balakrishna Nair. (2002). Antibiotic resistance, virulence gene & molecular profile of shiga toxine producing *E. coli* isolated from diverse source in Calcutta, India. *J. Clin. Microbiol.* 40 (6): 2009-2015.
- Adetunji V. O. and Tajudeen O. I. (2011). Antibiotic resistance of *E. coli, Listeria & Salmonella* isolates from retail meat tables in Ibdan municipal abattoir, Nigeria. *American J. of Biotech*. 10 (30): 5795-5799.
- 26. Xia Xiaodong., Jianghong Meng, Patrick F. McDermott Sherry Ayers, Karen Blickenstaff, Thu-Thuy Tran, Jason Abbott, Jie Zheng and Shaohua Zhao (2010) Presence and Characterization of Shiga Toxin-Producing Escherichia coli and Other Potentially Diarrheagenic E. coli Strains in Retail Meats, Appl. Environ. Microbiol. 76(6):1709-1717.

Copyright of Veterinary World is the property of Veterinary World and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.