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- AAC02295-17 Revised 1
- **Short-form paper** 2

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- 3 An IncX3 epidemic plasmid carrying bla_{NDM-5} in Escherichia coli from swine in multiple
- 4 geographic areas in China
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- 9 Kong, People's Republic of China;
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19 20 Six imported pigs originating from Guangdong, Henan, and Hunan provinces in China during October 21 2015-February 2017 were culture positive for meropenem-resistant Escherichia coli. The samples 22 yielded 9 E. coli isolates of diverse sequence types carrying bla_{NDM-5} on IncX3 (8 isolates from 5 23 farms) or IncFII (1 isolate from 1 farm) plasmids. The mcr-1 gene was co-harboured by four isolates. 24 The IncX3 plasmids (\sim 46 kb) carrying bla_{NDM-5} were identical or nearly identical to each other. 25 (74 words)

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The presence of carbapenemase-producing Enterobacteriaceae (CPE) in livestock animals is concerning as this may facilitate expansion of the gene pool from which pathogenic bacteria can pick up the resistance genes and consumers may be subsequently exposed through the food chain (1,2). For this reason, there is a need to enhance the monitoring of carbapenem resistance in the food supply (1). In Hong Kong, 80% of the food animals are imported from mainland China and involved farm suppliers from multiple provinces in the country (3).

From September 2008 to February 2017, rectal swabs were obtained from randomly selected fresh pig carcasses at a centralized slaughterhouse in Hong Kong by trained veterinary staff, as part of an ongoing surveillance (3). Each swab was collected from a single animal and was inoculated into nutrient broth with 10 mg/L vancomycin and 0.5 mg/L meropenem (4), followed by subculture on MacConkey agar plate supplemented with 2 mg/L meropenem. Five to ten colonies from each selective plate were picked. MALDI-TOF MS was used for bacterial identification. The agar dilution (for colistin) and disc diffusion (for other antibiotics) methods were used to determine antimicrobial susceptibility (5.6). Isolates from the same animal were considered to be unique if the resistance profiles for meropenem and colistin were different.

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In total, 856 pigs were cultured over 263 sampling dates (Table S1). Six pigs originating from six different farms were culture positive for meropenem-resistant E. coli (Table 1). According to the susceptibility patterns, a total of nine isolates were considered to be unique and investigated further. All isolates exhibited positive CarbaNP test result and were resistant to ertapenem, imipenem and meropenem. The presence of carbapenemase genes and mcr-1 were investigated by PCR and sequencing (7-9). The bla_{NDM-5} gene was identified in all nine isolates, of which four were colistin-resistant and co-harbored mcr-1. The isolates were further investigated by MLST and replicon typing (7,9,10). Plasmids carrying bla_{NDM-5} were of X3 (n=8, size ~45 kb) or F36 (n=1, size ~100 kb). Of the four mcr-

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I gene identified, three were harbored on plasmids of different replicon types (X4, FIB and Y). In conjugation experiments (9,10), there was no co-transfer of carbapenem and colistin co-resistance but the plasmids carrying bla_{NDM} or mcr-1 could be separately transferred at frequencies of 10^{-4} to 10^{-5} and 10^{-1} to 10^{-6} transconjugants per donor cell, respectively.

Six isolates (one from each animal, Table 1) were sequenced by an Illumina MiSeq platform at >150-fold coverage. The plasmids were assembled de novo using a CLC Genomics Workbench (Qiagen, Redwood City, United States) and gaps were closed by additional **PCRs** and Sanger sequencing (7,9,10).**ISfinder** (https://wwwis.biotoul.fr/about.php) was used to identify and annotate insertion sequences. In strain P748, bla_{NDM-5} was found in a contig (size ~32 kb) with 100% coverage and 98% identity to p28078-NDM (accession no. MF156713).

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Complete sequences of the five IncX3 plasmids with sizes of ~46 kb were obtained (Table S2). They have a plasmid scaffold typical of IncX3 plasmids (Figure 1a). The genetic load regions in the five plasmids were compared with two reference IncX3 plasmids (pNDM-HN380 and pIncX-SHV) (Figure 1b). In the bla_{NDM}-carrying plasmids, an ISL3 with 8-bp flanking direct repeats (ATATGCAT) was found downstream of the resolvase gene. The umuD gene was split into two fragments ($umuD\Delta 1$ and $umuD\Delta 2$) at the same position as in pIncX-SHV, resulting in a pair of 3-bp direct repeats (TGT). In pNDM-HN380, bla_{NDM} was inserted as an IS26-ISAba125 transposon-like structure (Figure 1b). Subsequently, the upstream ISAba125 was disrupted by IS5 (10). In four plasmids with link to Guangdong and Henan provinces, the sequences inserted between the two umuD fragments were 100% identical (10117 bp in length). This inserted sequence differs from that in pNDM-HN380 by a deletion of 7874 bp (Figure 1b). The remaining plasmid with link to Henan has an additional deletion (616 bp) at the junction between the IS5 and ISAba125 Δ 1 remnant. In the five NDM-5 plasmids, IS5 was inserted at the same position leading to the flanking 4-bp

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direct repeats (CTAA). In pNDM-HN380, IS5 was inserted at a different position in the opposite orientation.

To explore the geographical distribution of potentially related bla_{NDM}-carrying IncX3 plasmids, the complete sequence of pP768-NDM5 (chosen as a representative) was used to query the GenBank. Twenty-two plasmids related to pP768-NDM-5 were identified by a query of the GenBank (Figure 1c). These include 14 plasmids from China, four from Myanmar and one each from Canada, India, Kuwait and Oman (Table S3). The plasmids do not carry resistance genes other than bland. Multiple NDM variants were carried by the plasmids. These include NDM-1 and variants that differed by one to three amino acids, including NDM-4 (M154L), NDM-5 (V88L, M154L), NDM-7 (D130N, M154L) and NDM-17 (V88L, M154L, E170K).

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We identified the occurrence of similar IncX3 plasmids carrying bla_{NDM-5} in pigs originating from multiple farms across three different Chinese provinces. The involvement of IncX3 plasmids (represented by pNDM-HN380) in the dissemination of NDM in multiple geographic areas in China was initially reported by our group in 2012 (7). Subsequently, sporadic reports of pNDM-HN380-like plasmids carrying different NDM variants have also been reported in India, Arabian Pennisula, Europe and Australia (11-14). Recently, a Chinese national survey and several provincial studies revealed that IncX3 plasmids harbouring different bla_{NDM} variants were frequently found among clinical isolates of different multilocus sequence types and species, suggesting that it is an important vector responsible for the wide dissemination of NDM in China (15-17). IncX3 plasmids have a narrow host range and have mainly been found in Enterobacteriaceae (18). Our finding from analysis of complete plasmid sequences indicated that the five IncX3 plasmids originating from pigs are related to pNDM-HN380 as well as plasmids originating from many other geographic areas, thus confirming that this mobile NDM vector is widespread in the ecosystem.

As carbapenems have never been licensed for use in food animals in China, the NDM-producing pig isolates detected in the present study may have been introduced to the farms via human activity or contaminated feeds. It is worrisome that some of the NDMproducing isolates in the present study were found to co-harbour mcr-1 in another plasmid or the chromosome. Nonethelesss, our isolates co-harbouring mcr-1 were recovered before the ban of colistin in animal feeds was implemented in November 2016 in China.

In conclusion, this study identified an epidemic IncX3 plasmid carrying bla_{NDM-5} disseminated among E. coli originating from pigs with epidemiological links to geographically segregated areas in China.

Accession number(s). The complete sequence of the five IncX3 plasmids have been deposited into the GenBank database under accession numbers MF547511 (pP744-NDM5), MF547510 (pP768-NDM5), MF547509 (pP785-NDM5), MF547507 (pP788A-NDM5), and MF547508 (pP855-NDM5).

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Table 1. Sources and characteristics for nine NDM-positive E. coli isolates

Strain	Specimen	Source ^b	Date collected	MLST	bla _{NDM-5}	mcr-1	Replicon type of plasmid harboring		Resistance patterns ^d
							bla _{NDM}	mcr-1	•
P744A	Pig 1	Henan (A1)	Oct 2015	ST 10	+	_	Х3	none	Chl, Nit
P744T ^a	Pig 1	Henan (A1)	Oct 2015	ST 1602	+	+	X3	X4	Chl, Cip, Nit
P748 ^a	Pig 2	Hunan (B)	Jan 2016	ST 167	+	_	F36	none	Gen, Chl, Cip, Nit
P768	Pig 3	Henan (A2)	May 2016	ST 117	+	_	X3	none	Gen, Chl, Cip
P768-11 ^a	Pig 3	Henan (A2)	May 2016	ST 871	+	+	X3	FIB	Cip
P785 ^a	Pig 4	Guangdong (C1)	Jun 2016	ST 7512	+	+	X3	$Chromosomal^{c} \\$	Gen, Chl, Cip, Nit
P788A	Pig 5	Guangdong (C2)	Jun 2016	ST1286	+	_	X3	none	Nit
P788A-32 ^a	Pig 5	Guangdong (C2)	Jun 2016	ST 7510	+	+	X3	Y	Gen, Chl, Nit
P855 ^a	Pig 6	Guangdong (C3)	Feb 2017	ST 7511	+	_	X3	none	Gen, Chl, Cip, Nit

^aThe six isolates were investigated further by genome sequencing.

^bProvince (farm) origin of the pig 195

¹⁹⁶ ^cChromosomal location of mcr-1 in the isolate was confirmed by genome sequencing.

^dResistance patterns for amikacin (Ak), chloramphenicol (Chl), ciprofloxacin (Cip), fosfomycion (Fos), gentamicin (Gen) and nitrofurantoin

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Figure 1. Comparisons of IncX plasmids in this study. (a) Circular map of the plasmid pP768-NDM5. This plasmid was used to illustrate the backbone shared by all the analyzed plasmids and the location of the genetic load region. (b) Comparison of the genetic load region in 5 plasmids harbouring bla_{NDM-5} with two reference plasmids (pIncX-SHV and pNDM-HN380). (c) Alignment of pP768-NDM5 with 22 plasmids identified in the GenBank (last accessed 27 Oct 2017). The circular maps were generated with the BLAST Ring IMAGE Generator and each plasmid was colored by the geographical origin (China, blue; Myanmar, yellow; Oman, orange; India, red; Canada, pink and Kuwait, purple) in the following order (outer to inner circles): pP768-NDM5, pCREC-A6-NDM, pSCE516-2, pNDM5_IncX3, pEc1929, pECNDM101, pAD-19R, pNDM5_WCHEC0215, pK518_NDM5, pK516_NDM5, NUHL24835, pNDM-QD28, pNDM-QD29, pEC50-NDM7, pZHDC40,pJEG027, pM216_X3, pM213_X3 and pM110_X3, pOM26-1, pNDM-MGR194 and pKpN01-NDM7, pKW53T-NDM (full details of plasmids in Table S3).

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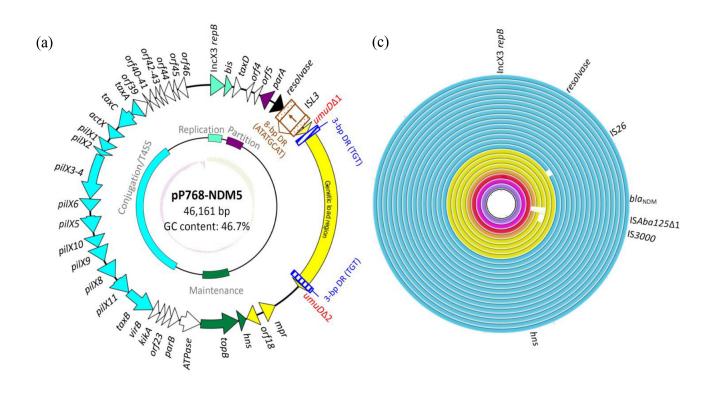
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