

1 | LOCAL CHROMATIN CONFORMATION

1.1 INTRODUCTION

The Hi-C assay provides a genome-wide overview of chromatin conformation, however this broad scope imposes resolution limits inherent to an all-vs-all assay. For a closer look at chromatin conformation within a region of interest, alternative C-based assays such as 3C, 4C and 5C can be employed alongside classical microscopy techniques like FISH.

Here I discuss two collaborative projects involving the use of 4C and 5C data to “zoom in” on two well-studied regions related to limb development: the ZRS enhancer and HoxD gene cluster.

1.2 CHROMATIN CONFORMATION AT THE SHH LOCUS

Anterior-posterior patterning in the developing limb is regulated in mammals by *Sonic hedgehog* (SHH).^[3] Specifically, the SHH gene is expressed within a confined region named the “zone of polarising activity”. Its expression within this region is known to be regulated by a well-studied enhancer, the “zone of polarising activity regulatory sequence” or ZRS.^[4] ZRS is located almost 1 Mb downstream of its target SHH promoter in humans, and is located in intronic regions of another gene, LMBR1, and is conserved across mammals and fish (Fig. 1).^[4,5] Single point mutations and short insertions within this enhancer have been linked to various limb deformities, including pre- and post-axial polydactyly.^[3,5,6] For example, a heritable point mutation in the ZRS enhancer is the cause of polydactyly in “Hemingway cats”, a large group of domestic cats with extra toes that reside at the former home of Ernest Hemingway.^[6]

Collaborators have developed a model system which allows inducible SHH expression in a non-expressing 14fp cell line derived from the developing limb bud. Application of trichostatin A (TSA) then leads to detectable SHH expression, and increased levels of the histone activation mark H3K27ac at the ZRS (*unpublished data*). However, the question remains whether this TSA treatment is fundamentally altering local chromatin structure, that is, bringing together the ZRS enhancer with its target SHH promoter, or whether ZRS and SHH are in contact in both the active and non-expressing cell lines and SHH expression is blocked through other means. Analysis of the region through FISH implies similar levels of compaction in SHH expressing and non-expressing cells (*data not shown*), suggesting the latter explanation.

My part in this collaboration was to analyse 3C-seq (also known as 4C) data recorded by our collaborators for the SHH–ZRS region in mouse. Additionally, the 4C procedure^[7] was adapted for specific in-house sequencing instruments (Ion Torrent Ion Proton™ sequenced as opposed to Illumina™ technology) and as such required diagnostics to confirm the experimental data was accurate.

1.2.1 Analysis of ZRS interactions

4C experiments were performed by collaborators using the ZRS region as a bait sequence, or “viewpoint”, such that its contacts were measured with all other HindIII restriction fragments genome-wide. 4C was performed in both

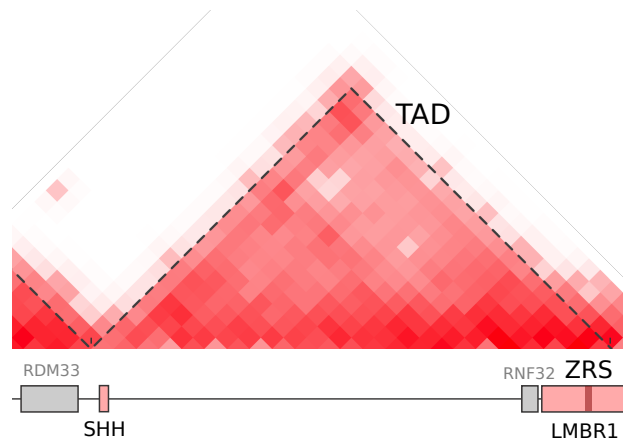


Figure 1: SHH–ZRS contacts occur within a stable TAD. An approximately 1 Mb region of the mouse genome is shown below a Hi-C contact map (derived from previously published data^[1]). A clear TAD can be identified spanning from SHH to ZRS, dashed lines show TAD boundaries called by Dixon *et al.*^[1]. This figure was generated for Anderson *et al.*^[2].

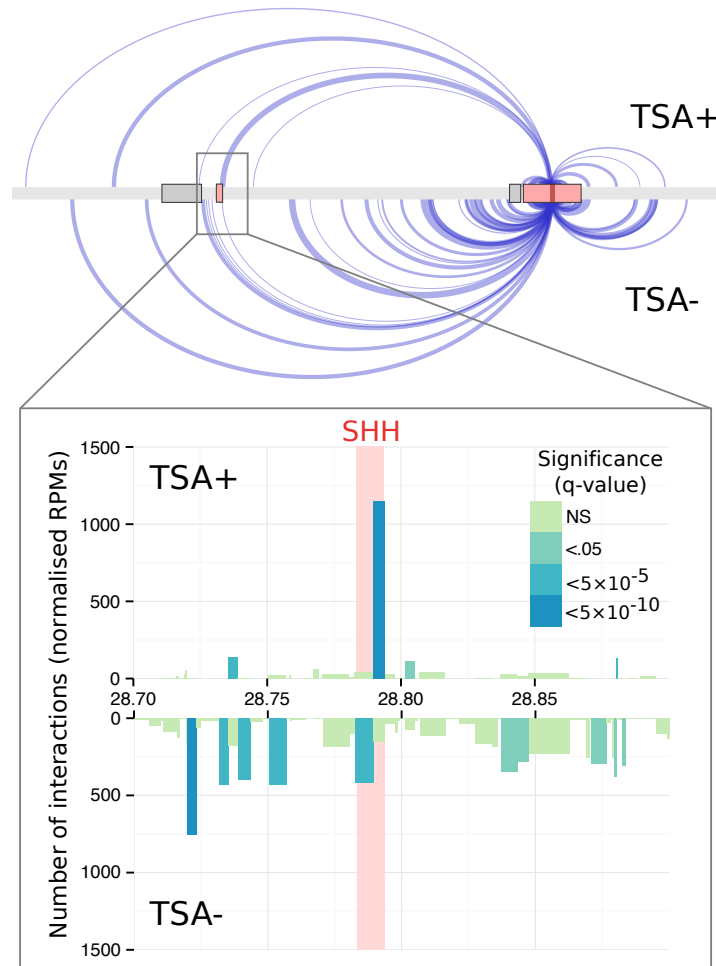


Figure 2: TSA treatment induces a strong ZRS–SHH interaction. 4C interactions are shown as edges from source node (ZRS enhancer bait fragment) to targets along an approximately 2 Mb region of chromosome 5. Edge width is proportional to the number of interactions, only highly significant interactions are shown (FDR q -value $< 5 \times 10^{-5}$). Zoomed region shows the number of interactions of the bait region with SHH in both treated and untreated samples. Each rectangle is a restriction fragment, coloured by FDR q -value indicating the significance of the interaction above expected levels.

untreated and non-SHH expressing cells (*TSA*−) and in cells treated with TSA, thereby causing SHH expression (*TSA*+).

The first stage in analysing these contacts is to convert observed raw sequencing reads to normalised frequencies (Methods ??), these normalised values are then assigned significance scores in the form of *q*-values, with the aim of finding those significantly over-represented relative to expectation (Methods ??).

1.2.2 4C / Hi-C comparison

Hi-C data in mouse cells has been previously published,^[1] so can be compared with this novel 4C data to give broader contextual information about chromatin conformation in the region under study.

1.2.3 Assay diagnostics

The 4C protocol used by our collaborators in this work was that of Stadhouder *et al.*^[7]. In it, the authors advise some statistical tests to ensure the quality of the experiment results. Among these were:^[7]

1. Sequencing reads should be found to have high duplication rates of 95% or greater.
2. 50% or more of all reads should map to the chromosome on which the bait region is located.

1.2.4 3D modelling with 5C data

All-vs-all contacts measured either genome-wide in the case of Hi-C, or over a defined region with 5C, can be used to infer the trajectory of chromatin fibres in three-dimensions through a variety of methods (e.g.^[8–12]). 5C data was generated over this same SHH-ZRS region (Fig. 1) with the aim of developing a multi-point perspective on local chromatin conformation beyond that available from 4C data.

We used this 5C experimental data in combination with a particular three-dimensional inference program (AutoChrom3D^[12]) in an attempt to compare polymer trajectories in TSA treated and untreated 14fp mouse cells.

1.3 5C IN THE HOXD REGION

HoxD is another well-studied genetic system involved in limb development and under the control of known enhancers. In this experiment, our collaborators were interested in the chromatin conformations of HoxD13 loci in both the anterior and posterior developing limb bud, particularly how and where the two differed. To this end, our collaborators performed 5C for two biological replicates in anterior and posterior limb bud cell lines, and my contribution was to call differential contacts between the two conditions.

1.3.1 Differential contacts

1.3.2 5C / Hi-C comparison

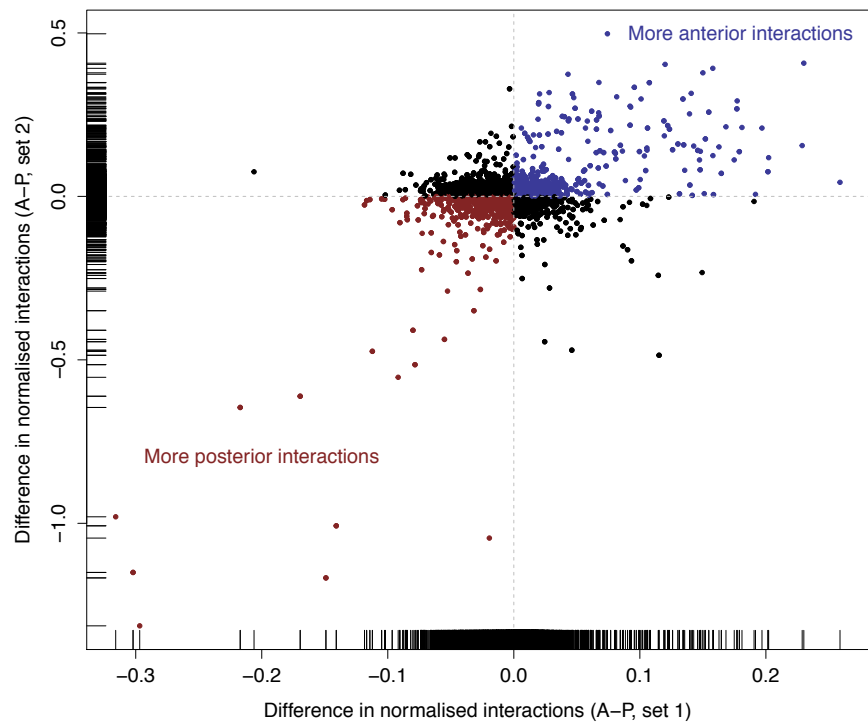


Figure 3: Raw differences between anterior and posterior 5C interactions. Placeholder

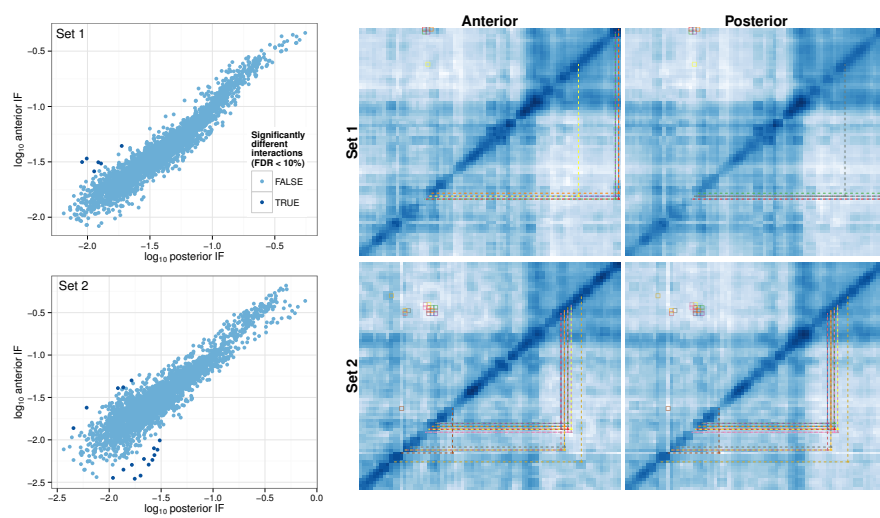


Figure 4: Will we use this stuff? Placeholder

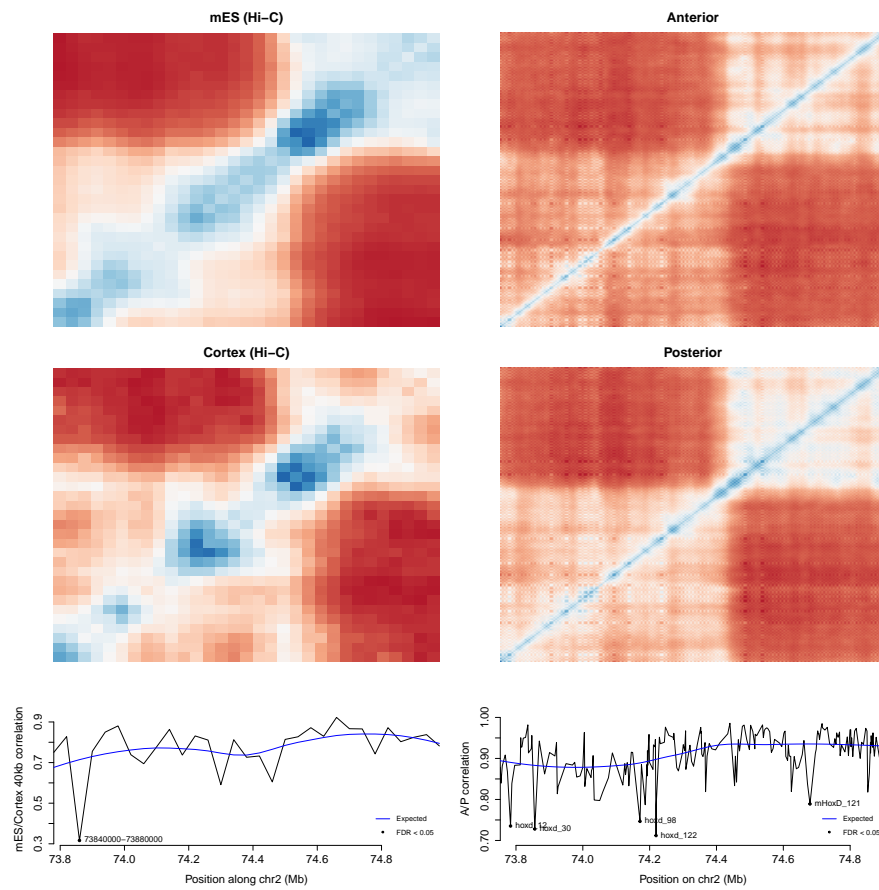


Figure 5: Will we use this stuff? Placeholder

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