Celligner_demo

October 27, 2021

0.1 Loading packages

```
[]: from genepy.utils import helper as h

# to comment in your case
from taigapy import TaigaClient
tc = TaigaClient()

from celligner import Celligner
import pandas as pd
# to comment in your case
from depmapomics import tracker as track
#autoreload
%load_ext autoreload
%autoreload 2
#output
from bokeh.plotting import output_notebook
output_notebook()

from celligner.params import TISSUE_COLOR
```

0.2 Loading expression files

0.3 Managing annotations

TCGA_expression = TCGA_expression[list(common)]

```
[]: # loading annotations

CCLE_annotation = track.getTracker() # the function uses pygsheets to load this:

→ REFSHEET_URL=https://docs.google.com/spreadsheets/d/

→ 1Pgb5fIClGnErEqzxpU7qqX6ULpGTDjvzWwDN8XUJKIY

# Sheets.from_files(MY_ID, MYSTORAGE_ID).get(REFSHEET_URL).sheets[0].

→ to_frame(index_col=0)

# you can also get it from pd.read_csv('gs://ccle_default_params/celligner_ex/

→ CCLE_annotation.csv.gz', index_col=0)

# can be loaded from

# pd.read_csv('gs://ccle_default_params/celligner_ex/TCGA_annotation.csv.gz', usindex_col=0)

TCGA_annotation = tc.get(name='celligner-input-9827', file='tumor_annotations') # generated manually
```

```
[13]: rename = {np.nan: "unknown", "adrenal cortex": "adrenal", "colorectal":

¬"unknown", 'brain': "central_nervous_system"}

 []: # some name are not consistent between the two datasets
     CCLE_annotation = CCLE_annotation.replace({"tissue_type": rename})
     TCGA annotation = TCGA annotation.replace({"tissue type": rename})
     0.4 Fitting celliner with the CCLE dataset
 []: # issues when rerunning celligner
 []: my_alligner = Celligner(make_plots=True, priotize_fit=True)
     my alligner.fit(CCLE expression, CCLE annotation)
 [4]: my_alligner.method = "mnn_marioni"
     my_alligner.priotize_fit=True
     my_alligner.mnn_kwargs = {'k1': 5, 'k2': 50, 'cosine_norm': False, "fk":5}
     _ = my_alligner.transform()
     reducing dimensionality...
     doing differential expression analysis on the clusters..
     regressing out the cPCA components..
     doing the MNN analysis using Marioni et al. method..
      Looking for MNNs...
      Found 12278 mutual nearest neighbors.
     reducing dimensionality...
     making plot ...
 []: # using the marioni mnn method
     my_alligner.method = "mnn_marioni"
     my_alligner.priotize_fit=True
     my_alligner.mnn_kwargs = {'k1': 5, 'k2': 50, 'cosine_norm': True, "fk":5}
     _ = my_alligner.transform(_rerun=False)
 [7]: my_alligner.umap_kwargs
 [7]: {'n_neighbors': 15, 'min_dist': 0.2, 'metric': 'cosine', 'n_components': 2}
 [8]: my_alligner.plot(color_column="tissue_type", colortable=TISSUE_COLOR,_
      -umap_kwargs={'n_neighbors': 10,'min_dist': 0.3, 'metric': 'euclidean'})
     reducing dimensionality...
     making plot ...
```

```
[8]: Figure(id='1762', ...)
[]: my_alligner.plot(rerun=False)
[]: # running with regular mnn
     my_alligner.method = "mnn"
     _ = my_alligner.transform(TCGA_expression, TCGA_annotation)
[4]: my_alligner.save('../temp/demo/')
    0.5 adding other datasets to celligner
[2]: # you can load the dataset from gcp: (you can do so by hand or by installing
     \rightarrow gsutil)
     # (make sure you have the right folder and then do:
     # ! qsutil cp qs://celliqner/model.pkl ../temp/demo/
     my_alligner = Celligner()
     my_alligner.load('../temp/demo/')
    fetching gene names from biomart cache
    using only usefull genes
[9]: # met500
    met500_ann = tc.get(name='met500-fc3c', file='met500_ann')
     met500_meta = tc.get(name='met500-fc3c', file='met500_meta')
     met500_TPM = tc.get(name='met500-fc3c', file='met500_TPM') #20,979x868 matrix
     #Novartis PDX
     Novartis_PDX_ann = tc.get(name='pdx-data-3d29', file='Novartis_PDX_ann')
     Novartis_PDX_TPM = tc.get(name='pdx-data-3d29', file='Novartis_PDX_TPM').T #__
     \rightarrow 38,087x445
     #pediatric PDX
     pediatric_PDX_ann = tc.get(name='pdx-data-3d29', file='pediatric_PDX_ann')
     pediatric_PDX_TPM = tc.get(name='pdx-data-3d29', file='pediatric_PDX_TPM')__
      \rightarrow #80,000x250
    No dataset version provided. Using version 1.
    No dataset version provided. Using version 1.
    No dataset version provided. Using version 1.
    No dataset version provided. Using version 2.
    No dataset version provided. Using version 2.
```

No dataset version provided. Using version 2. No dataset version provided. Using version 2.

0.5.1 Managing annotations

```
[14]: met500_meta["primary_site"] = met500_ann['primary_site'].values
    met500 ann = met500 meta.rename(columns={"Sample id": 'sample id', 'tissue':
     ⇒set index('sample id', | |

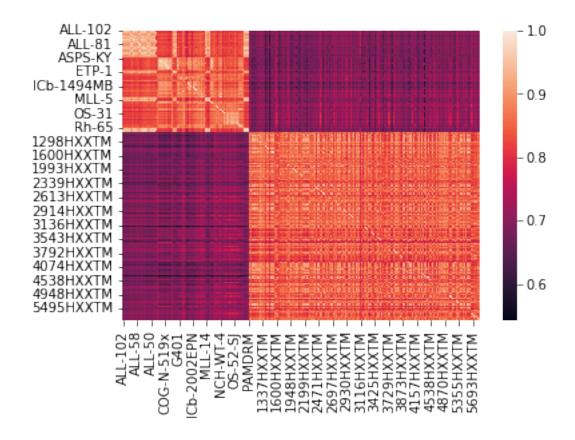
drop=True)[["tissue_type","disease_type","cell_type"]].

     →replace({"tissue type":rename, "cell type": {"tumor": "met500 tumor"}})
[15]: pediatric PDX ann = pediatric PDX ann.rename(columns={"sampleID": 'sample id', |
     →'lineage': 'tissue_type', 'subtype': "disease_type", "type": "cell_type"}).
     →set_index('sample_id', drop=True)[['cell_type', 'disease_type', |
     [16]: | Novartis_PDX_ann = Novartis_PDX_ann.rename(columns={"sampleID": 'sample_id',__

→set_index('sample_id', drop=True)[['cell_type', 'disease_type',

     [17]: # Looking at dataset similarity
    import seaborn as sns
    sns.heatmap(pd.concat([pediatric_PDX_TPM.loc[:,set(pediatric_PDX_TPM.columns) &__
     →set(Novartis PDX_TPM.columns)], Novartis PDX_TPM.loc[:,set(pediatric_PDX_TPM.
     →columns) & set(Novartis_PDX_TPM.columns)]]).T.corr())
```

[17]: <AxesSubplot:>



```
[18]: Novartis_PDX_TPM = Novartis_PDX_TPM.loc[:,set(Novartis_PDX_TPM)&_

→set(pediatric_PDX_TPM)]

pediatric_PDX_TPM = pediatric_PDX_TPM.loc[:,set(Novartis_PDX_TPM)&_

→set(pediatric_PDX_TPM)]
```

```
[19]: pediatric_PDX_ann['cell_type'] = "ped PDX"
```

```
[20]: Novartis_PDX_ann = Novartis_PDX_ann.loc[Novartis_PDX_TPM.index]
pediatric_PDX_ann = pediatric_PDX_ann.loc[pediatric_PDX_TPM.index]
```

```
# if you want to align to both CCLE and TCGA, you can ask celligner to consider the two (fit + _pre-transformed_ transform datasets) as a fit dataset by calling:

# my_alligner.putAllToFit()

# you can add your dataset as a dataset to be aligned to, by puting it in fit:

# my_alligner.addToFit(yourdataset).transform()

# /!\ need to already have a transform dataset (if you loaded the example → model, this is TCGA)

# you can add your dataset as one to align, by putting it in transform:
```

```
# my_alliquer.addToTransform(yourdataset)
     \# /!\ need to already have a fit dataset (if you loaded the example model, this
     \rightarrow is CCLE)
     # if your dataset is small enough it might actually not work well to put it in_
     → transform it seems!
     # if your dataset is small and similar enough, you can add the parameter_
     →dotransform=False (or dofit=False) so that it doesn't fully retransforms or
     →refit but uses cached computation instead.
     my alligner.priotize fit=False
     my_alligner.putAllToFit(redo_diff=False)
     _ = my_alligner.transform(met500_TPM, met500_ann, recompute_contamination=False)
    clustering...
    WARNING: You're trying to run this on 29593 dimensions of `.X`, if you really
    want this, set `use rep='X'`.
             Falling back to preprocessing with `sc.pp.pca` and default params.
    done
    looking at 868 samples.
    found 18218 common genes
    creating a transform input..
    clustering..
    WARNING: You're trying to run this on 18218 dimensions of `.X`, if you really
    want this, set `use_rep='X'`.
             Falling back to preprocessing with `sc.pp.pca` and default params.
    reducing dimensionality...
    doing differential expression analysis on the clusters..
    running differential expression on 35 clusters
    running limmapy on the samples
    you need to have R installed with the limma library installed
    there is 0.235 overlap between the fit and transform dataset in their most
    variable genes
    regressing out the cPCA components..
    doing the MNN analysis using Marioni et al. method..
      Looking for MNNs...
      Found 3416 mutual nearest neighbors.
    done
    reducing dimensionality...
    making plot ...
[]: my_alligner.putAllToFit(redo_diff=False)
[]: _= my_alligner.transform(pd.concat([Novartis_PDX_TPM, pediatric_PDX_TPM]), pd.
```

concat([Novartis_PDX_ann, pediatric_PDX_ann]), recompute_contamination=False)

```
[]: from celligner.params import TISSUE_COLOR

[]: my_alligner.plot(color_column="tissue_type", colortable=TISSUE_COLOR)
```