

Biomolecular Simulation: A Computational Microscope for Molecular Biology

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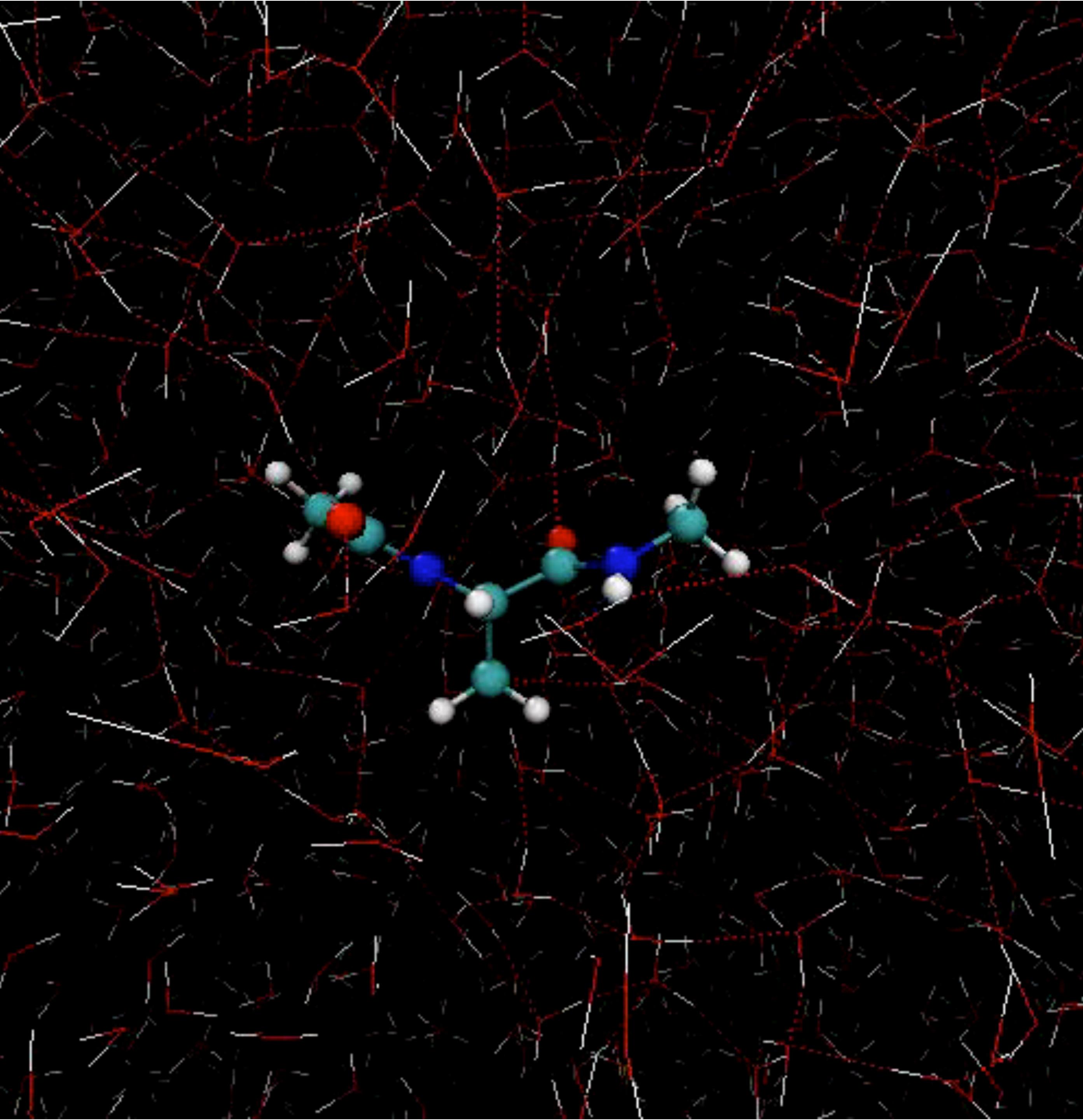
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New York 10032

Structural Bioinformatics

Molecular Dynamics Simulations

Ideally, what should a computational microscope do?

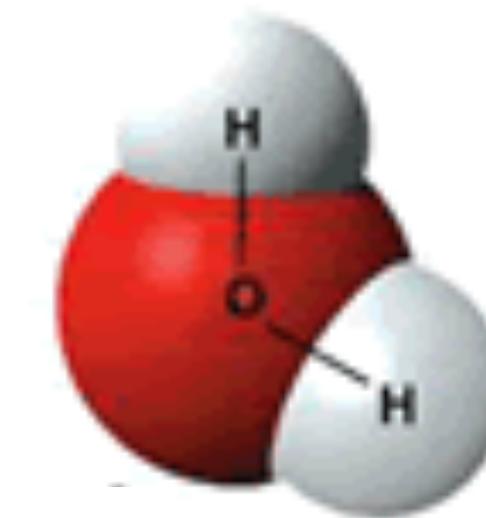
- Observe the time evolution of molecules at high spatial and time resolution
- Observe them for very long time scales
- Be aware of their energy
- Be able to set different experimental conditions (Temperature, Pressure, solution conditions)
- Be accurate
- Be interpretable, that is find suitable macrostates to compare with experiments
- ...



What is a good representation of a molecule?

Structural experiments allow us to see molecules at atomic resolution and in a fixed chemical configuration, this is a molecular perspective (i.e. a molecule is given)

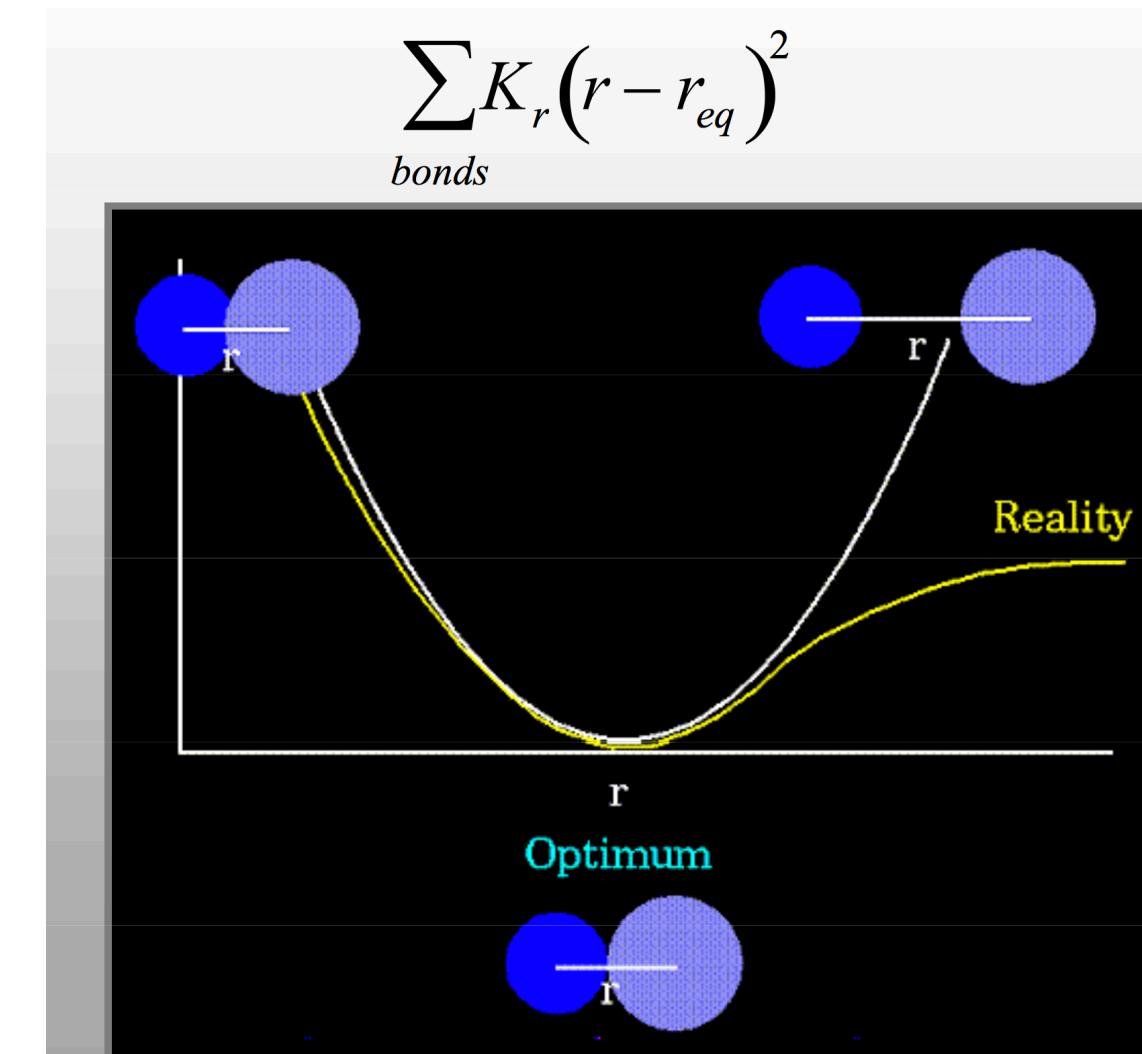
Molecules have covalent properties and non-covalent interactions



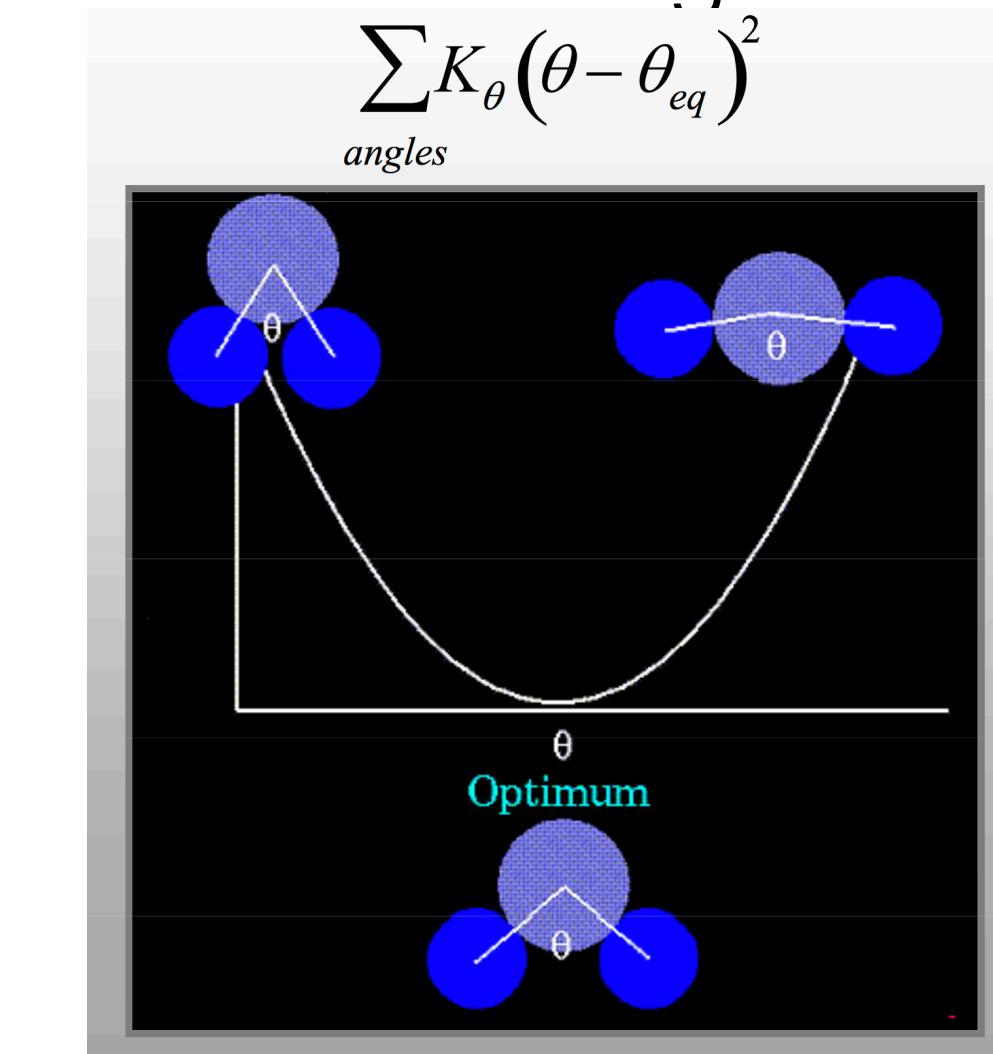
=

Covalent (Geometrical)
O-H distance: 0.9572 Å
H-O-H angle: 104.52

With fluctuations around these average values



+



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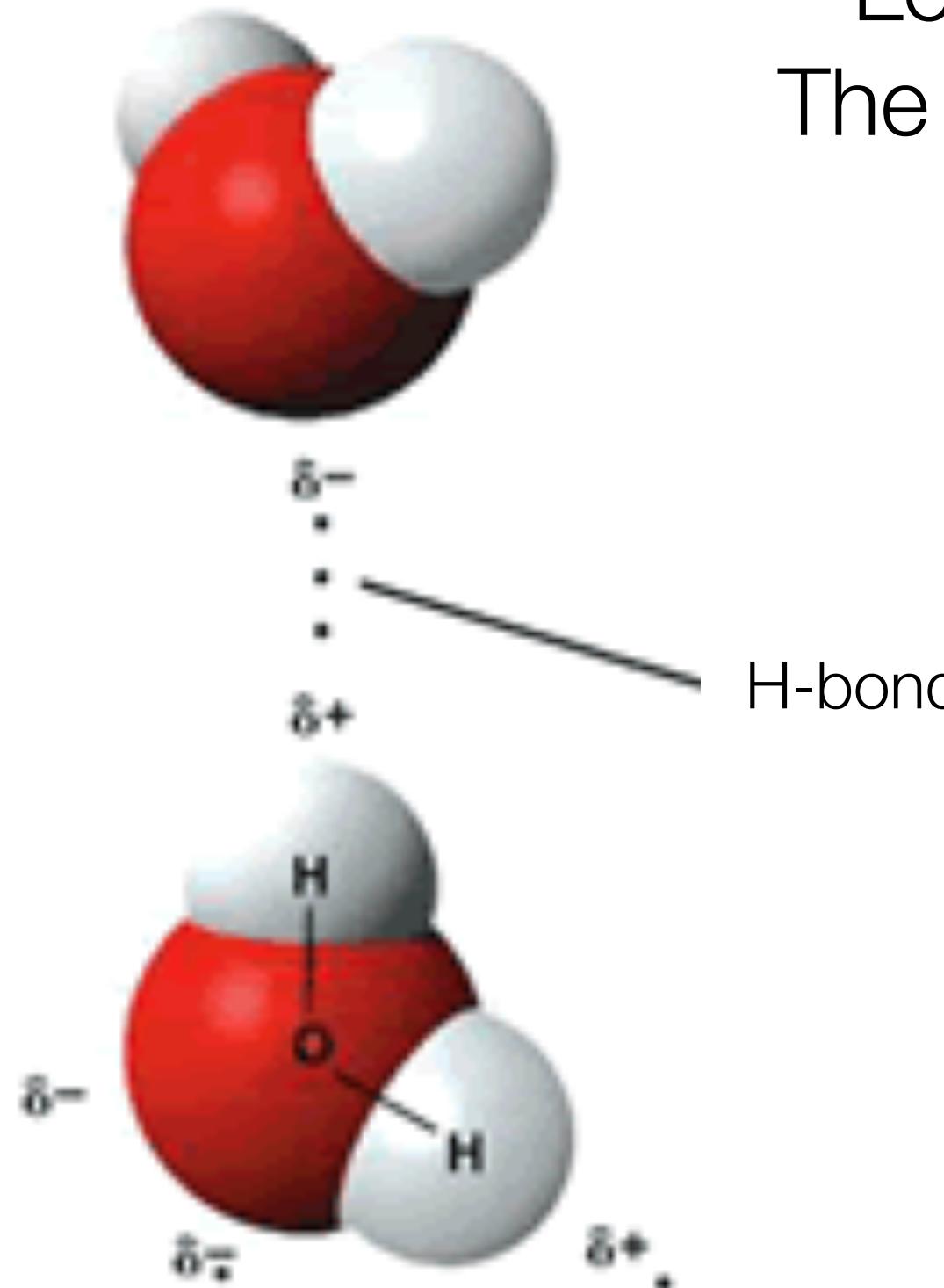
$$E(d_{OH1}, d_{OH2}, \theta_{H1\hat{O}H2}) = 1000 * (d_{OH1} - 0.9572)^2 + 1000 * (d_{OH2} - 0.9572)^2 + 1000 * (\theta_{H1\hat{O}H2} - 104.52)^2$$

So the energy is 0 if the geometry is correct and is >0 if not.

For now 1000 is an arbitrary number. This is called the bond strength and its value can be found studying the infrared vibrations of water for example.



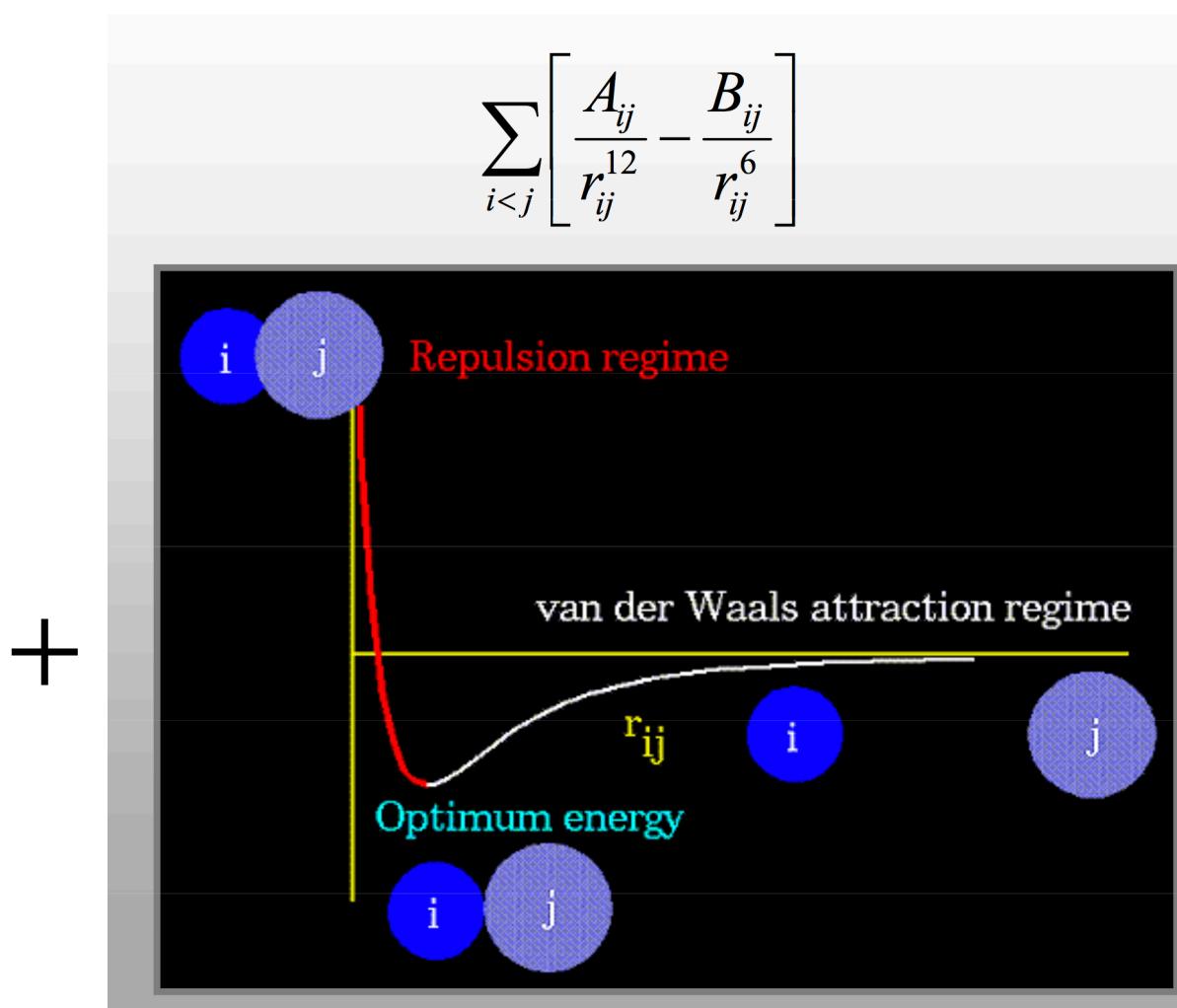
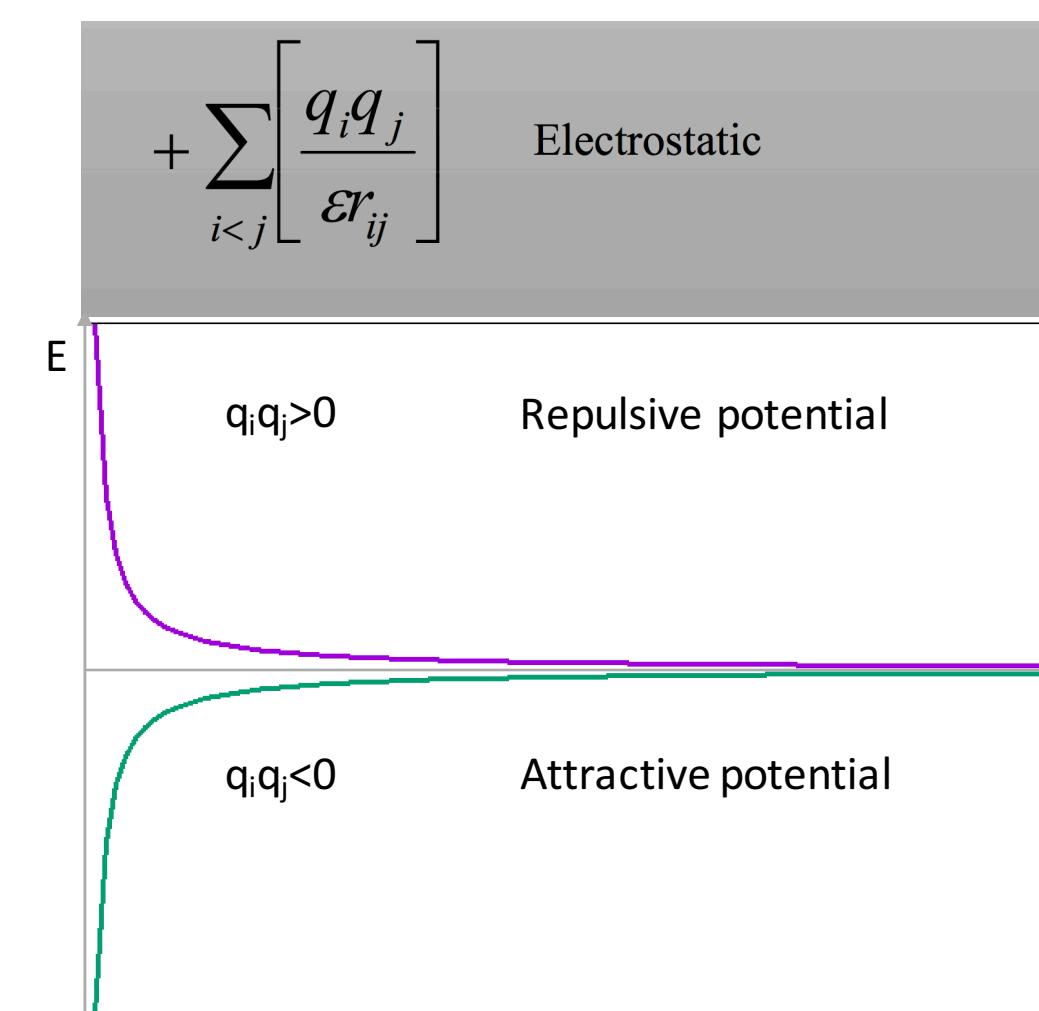
What is a good representation of a molecule?



Non-covalent interactions

Lone pair electrons that can form hydrogen bonds

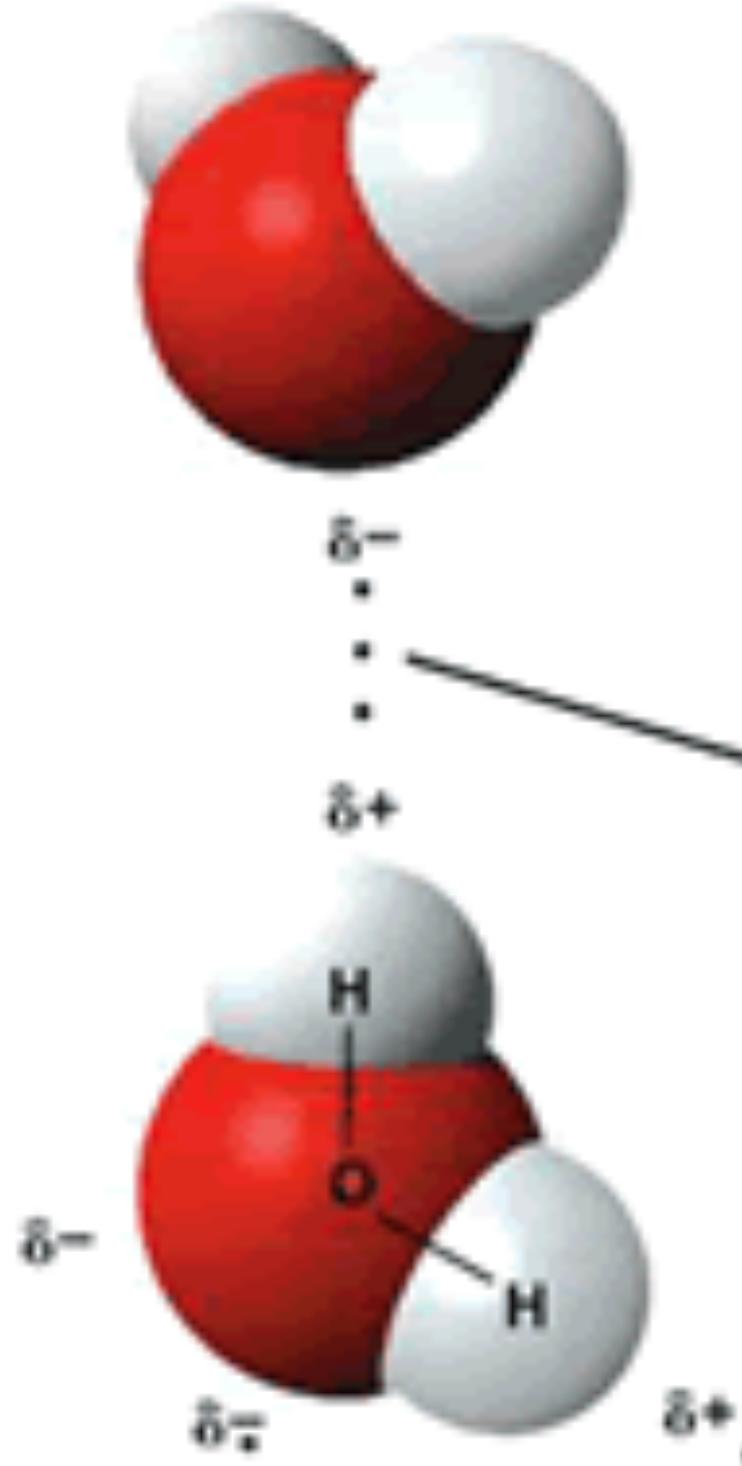
The oxygen is negatively charged while hydrogens are positively charged



Otherwise an O can overlap with an H



What is a good representation of a molecule?



Non-covalent interactions

Lone pair electrons that can form hydrogen bonds

The oxygen is negatively charged while hydrogens
are positively charged

$$E(d_{O1O2}, d_{O1H21}, d_{O1H22}, d_{O2H11}, d_{O2H12}, d_{H11H21}, d_{H11H22}, d_{H12H21}, d_{H12H22}) =$$

$$\frac{q_O q_O e^2}{d_{O1O2}} + \frac{q_O q_H e^2}{d_{O1H21}} + \frac{q_O q_H e^2}{d_{O1H22}} + \frac{q_O q_H e^2}{d_{O2H11}} + \frac{q_O q_H e^2}{d_{O2H12}} + \frac{q_H q_H e^2}{d_{H11H21}} + \frac{q_H q_H e^2}{d_{H11H22}} + \frac{q_H q_H e^2}{d_{H12H21}} + \frac{q_H q_H e^2}{d_{H12H22}} + \frac{A_{OO}}{d_{O1O2}^{12}} - \frac{B_{OO}}{d_{O1O2}^6}$$

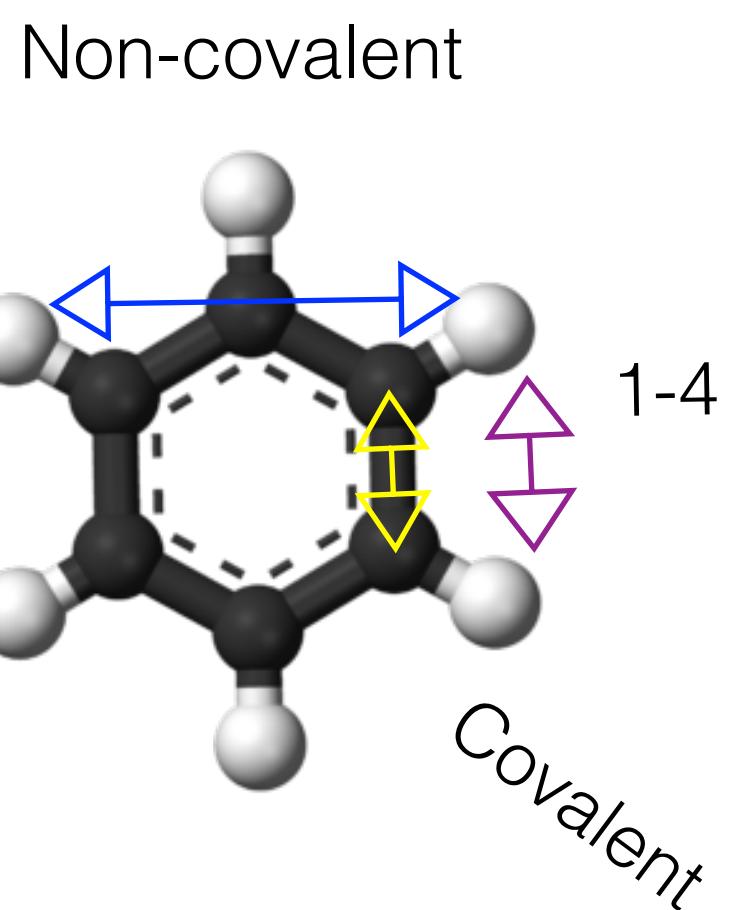
Here the energy is more complicated and the interaction between two water molecules will be the results of the relative orientation of all their atoms

What is a good representation of a molecule?

Are distances and angles enough to describe the geometry?

For example in the case
of benzene this is not
enough

We use dihedral interactions to
add information about the relative
orientation of two couples of
atoms



$$\sum_{\text{dihedrals}} K_\phi (1 + \cos(n\phi))$$

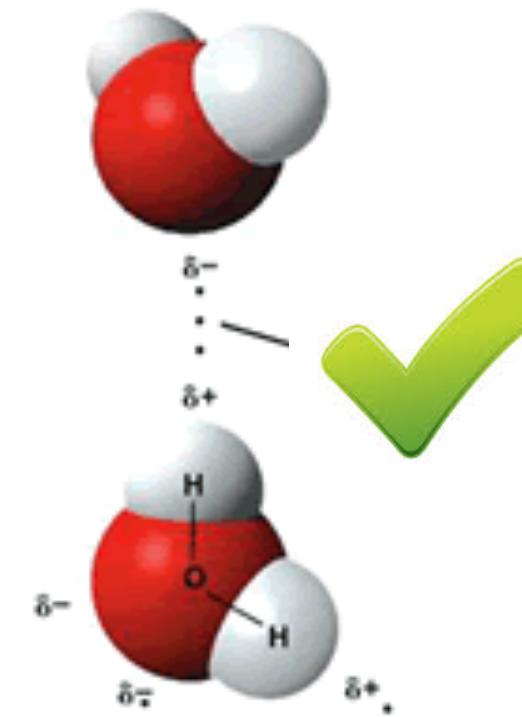


What is a good representation of a molecule?

Are Coulomb and Lennard-Jones interactions enough for non-covalent interactions?

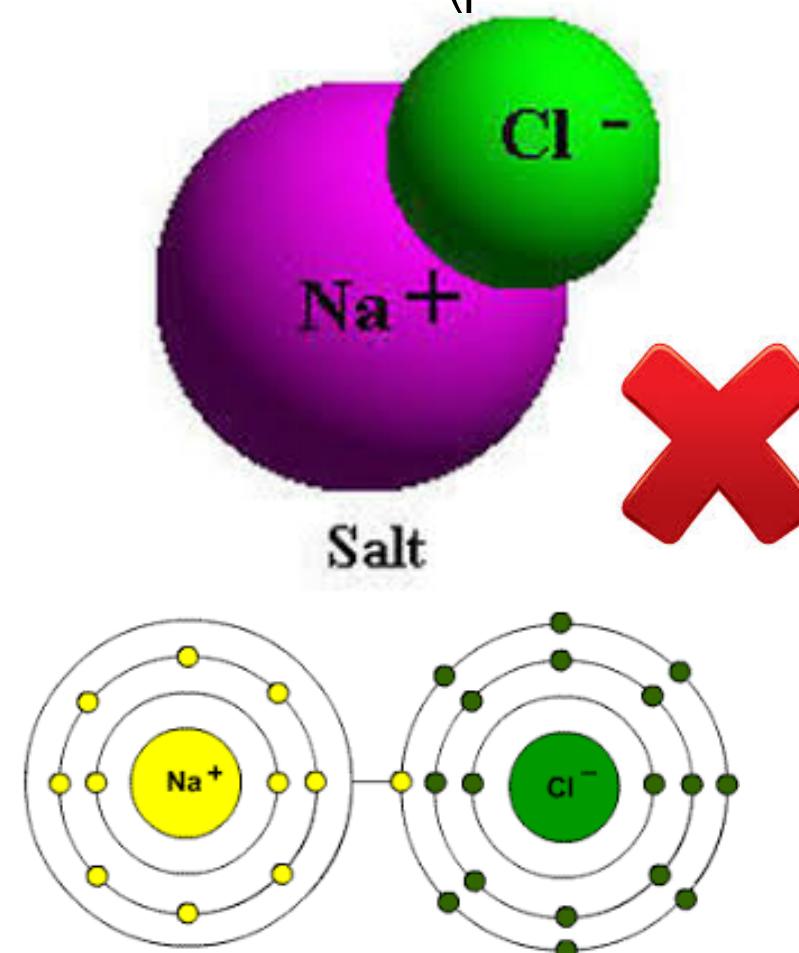
Neutral molecules

Yes, interaction energies between neutral molecules can be reproduced rather accurately (dipolar interaction)



Ionic interactions

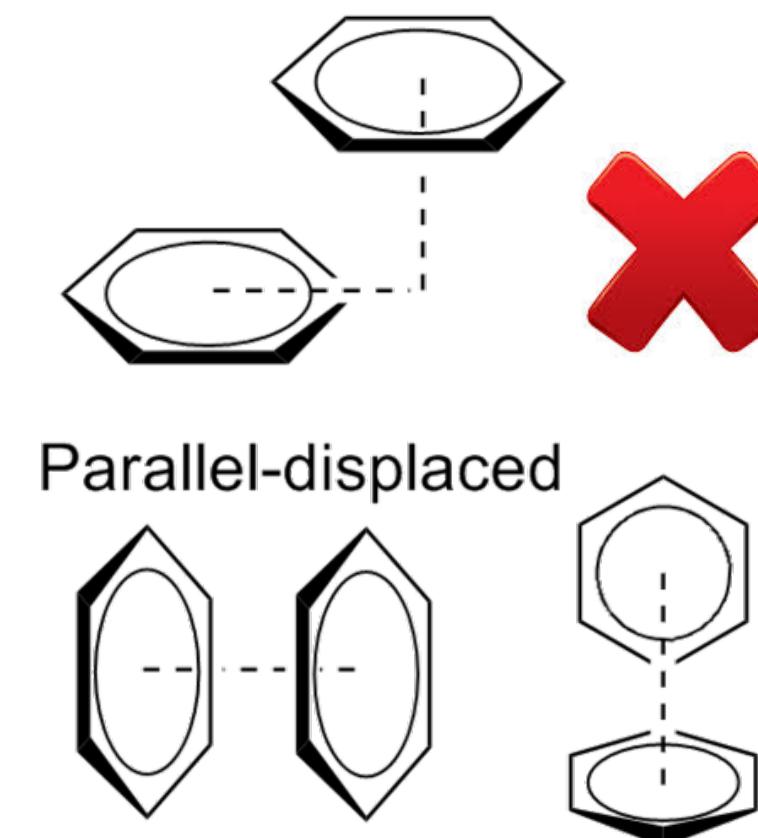
No because the charge of an atom is fixed, while in reality it reacts to the environment (polarisation)



Almost OK for monovalent, bad for others

Pi-stacking

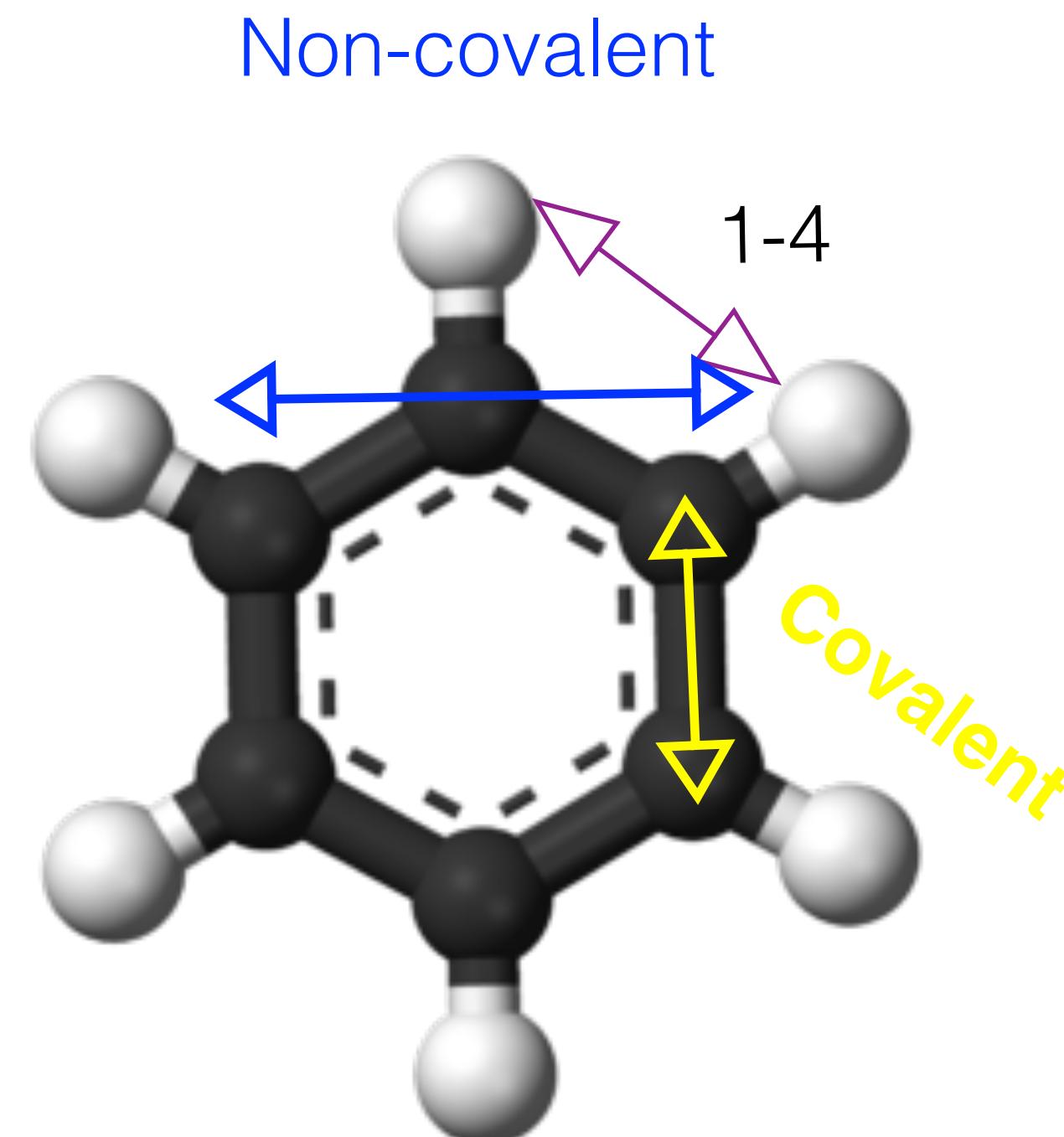
Weakly, because the charge of a ring is distributed over and below the ring (multipole)



Sandwich T-shaped



Molecular Mechanics Force Fields



first approx for vibrations
(anharmonic potential can be used for
more accurate vibrations)

$$V(r) = \sum_{bonds} k_b(b - b_0)^2 + \sum_{angles} k_\theta(\theta - \theta_0)^2 + \sum_{torsions} k_\phi[\cos(n\phi + \delta) + 1]$$

$$+ \sum_{nonbond\ pairs} \left[\frac{q_i q_j}{r_{ij}} + \frac{A_{ij}}{r_{ij}^{12}} - \frac{C_{ij}}{r_{ij}^6} \right]$$

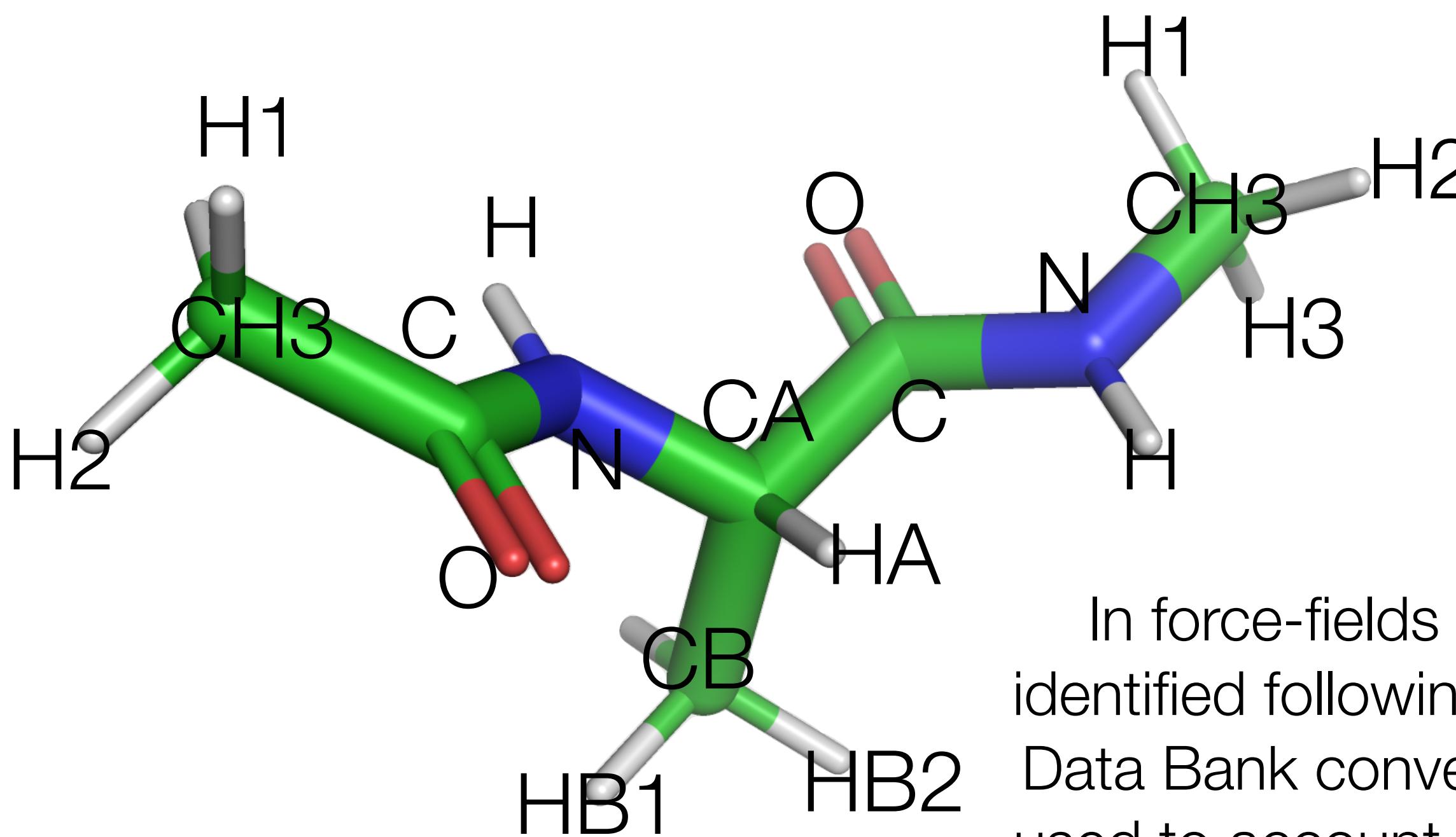
point charge
Coulomb

Dispersions
(interactions of
neutral molecules)
excluded volume

geometrical
consideration
pi-bonds, etc

Force Fields

a force field has ~5000-10000 parameters to describe all the possible bonds, angles, torsion, Coulomb and Van der Waals interactions between atoms.



In force-fields atoms are identified following the Protein Data Bank convention, this is used to account for their local chemical environment and thus allow to define different force-field terms:

[atoms]										
;	nr	type	resnr	residue	atom	cgnr	charge	mass	typeB	char
;	residue	1	ACE	rtp	ACE	q	0.0			
	1	CT	1	ACE	CH3	1	0	12.01		
	2	HC	1	ACE	HH31	2	0.02	1.008		
	3	HC	1	ACE	HH32	3	0.02	1.008		
	4	HC	1	ACE	HH33	4	0.02	1.008		
	5	C	1	ACE	C	5	0.533742	12.01		
	6	O	1	ACE	O	6	-0.593742	16		; qtot 0
;	residue	2	ALA	rtp	ALA	q	0.0			
	7	N	2	ALA	N	7	-0.278592	14.01		
	8	H	2	ALA	H	8	0.305959	1.008		
	9	AA	2	ALA	CA	9	0.0337	12.01		
	10	H1_H1B	2	ALA	HA	10	0.0823	1.008		
	11	CT_CT	2	ALA	CB	11	-0.1825	12.01		
	12	HC	2	ALA	HB1	12	0.0603	1.008		
	13	HC	2	ALA	HB2	13	0.0603	1.008		
	14	HC	2	ALA	HB3	14	0.0603	1.008		
	15	C	2	ALA	C	15	0.451975	12.01		
	16	O	2	ALA	O	16	-0.593742	16		; qtot 0
;	residue	3	NME	rtp	NME	q	0.0			
	17	N	3	NME	N	17	-0.278592	14.01		
	18	H	3	NME	H	18	0.305959	1.008		
	19	CT	3	NME	CH3	19	-0.087367	12.01		
	20	H1	3	NME	HH31	20	0.02	1.008		
	21	H1	3	NME	HH32	21	0.02	1.008		
	22	H1	3	NME	HH33	22	0.02	1.008		; qtot 0



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[bonds]			[pairs]			[angles]			[dihedrals]			[bondtypes]			[angletypes]															
	ai	aj	func	;	ai	aj	funct	;	ai	aj	ak	funct	;	i	j	func	b0	kb	;	i	j	k	func	th0	cth					
;	1	2	1		1	8	1		2	1	3	1		2	1	5	6	9	YY	CT	1	0.1526000	259408.00	;	YY	C	OH	1	110.000	669.440 ; Junmei et al.
1	3	1			1	9	1		2	1	4	1		2	1	5	7	9	C	YY	1	0.1522000	265265.60	;	YY	C	O	1	120.400	669.440 ;
1	4	1			2	6	1		3	1	5	1		3	1	5	6	9	YY	H1	1	0.1090000	284512.00	;	YY	C	O2	1	117.000	585.760 ;
1	5	1			4	6	1		3	1	5	1		4	1	5	6	9	YY	HP	1	0.1090000	284512.00	;	YY	C	N	1	116.600	585.760 ;
5	6	1			4	7	1		4	1	5	1		1	5	7	8	9	WW	CT	1	0.1526000	259408.00	;	H1	YY	H1	1	109.500	292.880 ;
5	7	1			5	10	1		1	5	6	1		6	5	7	8	9	C	WW	1	0.1522000	265265.60	;	H1	YY	N	1	109.500	418.400 ;
5	8	1			5	11	1		1	5	7	1		6	5	7	9	9	WW	H1	1	0.1090000	284512.00	;	HP	YY	HP	1	109.500	292.880 ;
5	9	1			5	15	1		6	5	7	1		5	7	9	10	9	WW	HP	1	0.1090000	284512.00	;	HP	YY	N3	1	109.500	418.400 ;
7	8	1			6	8	1		5	7	9	1		5	7	9	11	9	WW	N	1	0.1449000	282001.60	;	H1	YY	N3	1	109.500	418.400 ;
7	9	1			6	9	1		5	7	9	1		5	7	9	15	9	WW	N3	1	0.1471000	307105.60	;	C	YY	H1	1	109.500	418.400 ;
7	10	1			7	12	1		8	7	9	1		8	7	9	10	9	VV	CT	1	0.1526000	259408.00	;	C	YY	HP	1	109.500	418.400 ;
7	11	1			7	13	1		7	9	10	1		8	7	9	11	9	C	VV	1	0.1522000	265265.60	;	C	YY	HP	1	109.500	418.400 ;
9	10	1			7	14	1		7	9	11	1		8	7	9	15	9	VW	H1	1	0.1090000	284512.00	;	C	YY	N3	1	111.200	669.440 ;
9	11	1			7	16	1		7	9	15	1		7	9	11	12	9	VW	HP	1	0.1090000	284512.00	;	C	YY	N3	1	121.900	418.400 ;
9	15	1			8	10	1		10	9	11	1		10	9	11	12	9	VW	N	1	0.1449000	282001.60	;	C	N	YY	1	118.040	418.400 ;
11	12	1			8	11	1		10	9	15	1		10	9	11	13	9	VW	N3	1	0.1471000	307105.60	;	YY	N	H	1	118.040	418.400 ;
11	13	1			8	15	1		11	9	15	1		10	9	11	13	9	TT	CT	1	0.1526000	259408.00	;	YY	N3	H	1	109.500	418.400 ;
11	14	1			9	18	1		9	11	12	1		10	9	11	14	9	C	TT	1	0.1522000	265265.60	;	YY	N3	CT	1	118.000	418.400 ;
11	12	1			9	19	1		9	11	13	1		15	9	11	12	9	TT	H1	1	0.1090000	284512.00	;	YY	N3	CT	1	109.500	418.400 ;
11	13	1			10	12	1		9	11	14	1		15	9	11	13	9	TT	HP	1	0.1090000	284512.00	;	CT	YY	N3	1	111.200	669.440 ;
11	14	1			10	13	1		12	11	13	1		15	9	11	14	9	TT	N	1	0.1449000	282001.60	;	CT	YY	HP	1	109.500	418.400 ;
11	14	1			10	14	1		12	11	14	1		7	9	15	16	9	TT	N3	1	0.1471000	307105.60	;	YY	CT	CT	1	109.500	334.720 ;
15	16	1			10	17	1		13	11	14	1		10	9	15	16	9	SS	CT	1	0.1526000	259408.00	;	YY	CT	HC	1	109.500	418.400 ;
15	17	1			11	16	1		9	15	16	1		10	9	15	17	9	C	SS	1	0.1522000	265265.60	;	YY	CT	N	1	109.700	669.440 ;
15	17	1			11	17	1		9	15	17	1		11	9	15	16	9	SS	H1	1	0.1090000	284512.00	;	CT	YY	N	1	111.100	527.184 ;
17	18	1			12	15	1		16	15	17	1		11	9	15	17	9	SS	HP	1	0.1090000	284512.00	;	C	YY	CT	1	111.100	527.184 ;
17	18	1			13	15	1		15	17	18	1		9	15	17	18	9	SS	N	1	0.1449000	282001.60	;						
17	19	1			14	15	1		15	17	19	1		9	15	17	19	9	SS	N3	1	0.1471000	307105.60	;						
19	20	1			15	20	1		18	17	19	1		16	15	17	18	9	RR	CT	1	0.1526000	259408.00	;						
19	20	1			15	21	1		17	19	20	1		15	17	19	20	9	C	RR	1	0.1522000	265265.60	;						
19	21	1			16	18	1		17	19	22	1		15	17	19	22	9	RR	HP	1	0.1090000	284512.00	;						
19	22	1			16	19	1		20	19	21	1		18	17	19	20	9	RR	N	1	0.1449000	282001.60	;						
18	20	1			18	20	1		20	19	22	1		18	17															

Force Fields

In the end once you have a configuration of your system and you choose a force-field you can calculate the energy of that configuration as parameterised for the specific force-field you are using, then you can get a different configuration and again calculate its force-field energy.

Over time there have been multiple force-fields that have been parameterised and now we have four main force-field families:

AMBER

CHARMM

GROMOS

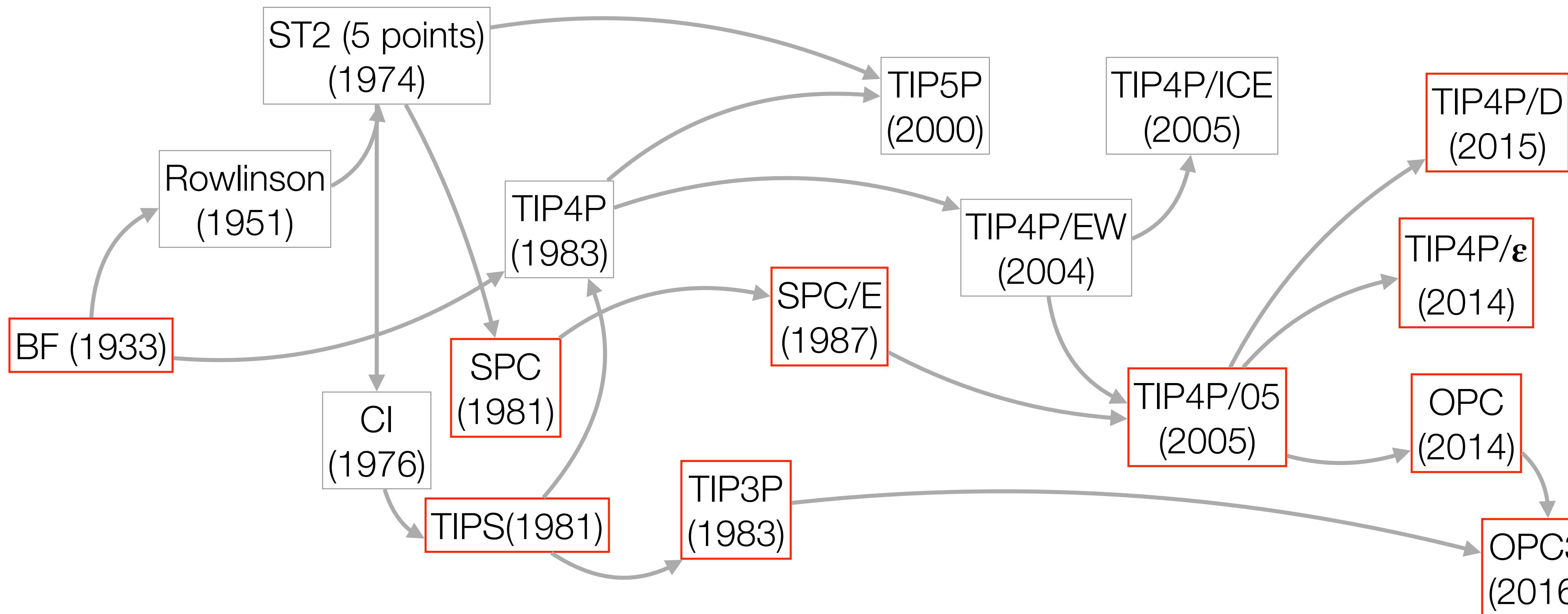
OPLS

And for each family there are multiple generations and variants.

A force-field is then an approximation of the energy of all chemically reasonable configurations of atoms in a biomolecule.



Digression: a brief history of rigid models for water



A Theory of Water and Ionic Solution, with Particular Reference to Hydrogen and Hydroxyl Ions



J. D. BERNAL AND R. H. FOWLER, *University of Cambridge, England*

(Received April 29, 1933)

In 1933 Bernal and Fowler tried to make a model of water to explain the tetrahedral structure of ice and other properties as observed by scattering. They observe that:

"If we take the spectroscopic model and place a charge e at the H positions and -2e on the O nucleus we arrive at a molecule of dipole moment 5.6 D instead of the observed 1.87 D. It is clear that the discrepancy is due to having made no allowance of the effect of the two H⁺ on the O₂⁻ ion. This will be twofold. The centre of the negative electronic charge will be moved from the O nucleus to some position intermediate between it and the H nuclei, and a certain concentration of negative charge will screen the H charges. The simplest assumption is to assume effective charges e' (<e) at the H positions and a charge -2e' situated at x Å from the O nucleus on the bisector of the H⁺O⁻H angle. With an electric dipole of 2D these are connected by the relation:

$$(0.58 - x)\epsilon' = 0.21\epsilon.$$

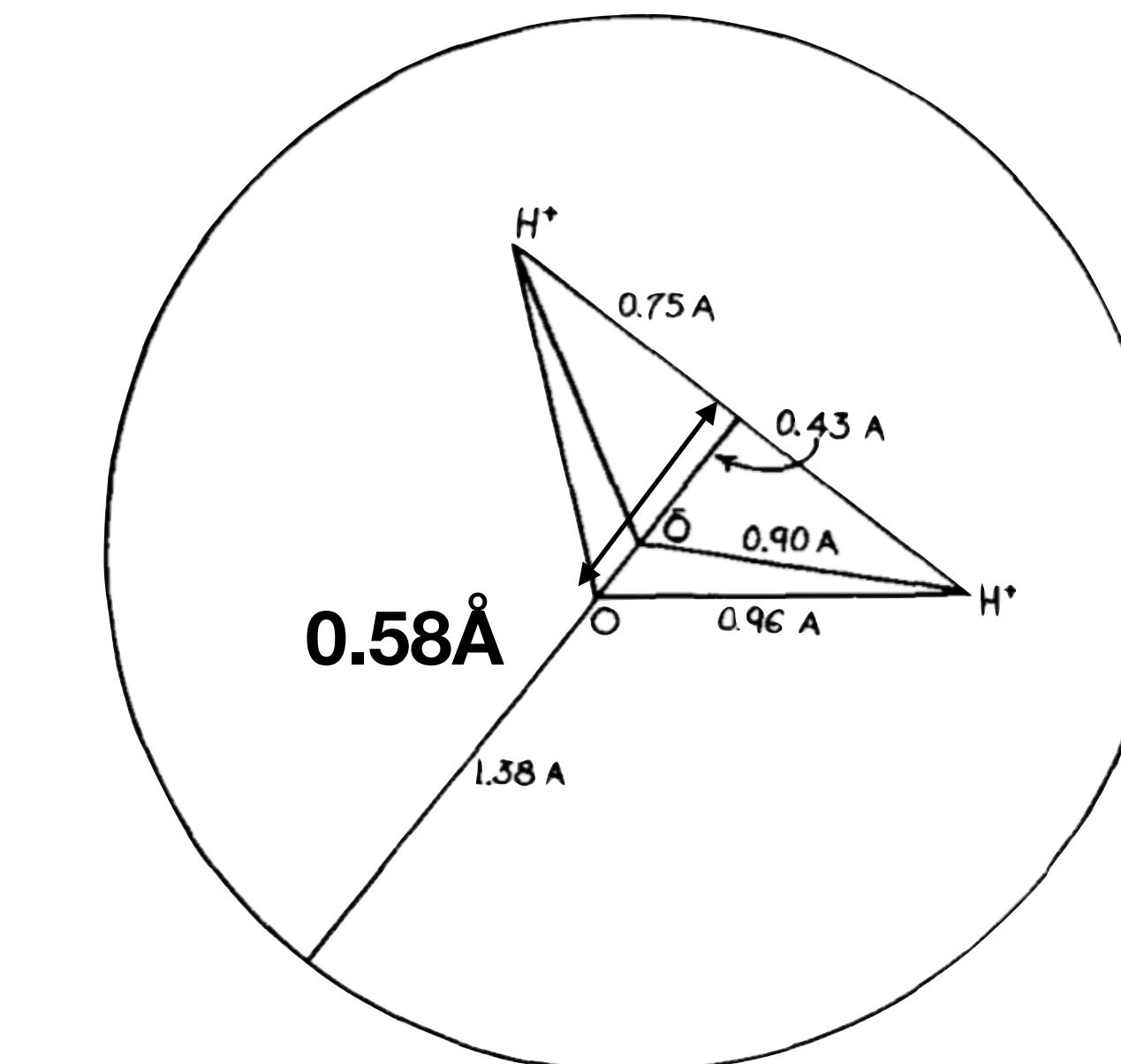


FIG. 10. The water molecule model. H⁺H⁺ are the hydrogen nuclei; O, the oxygen nucleus; Ō is the centre of negative charge and of the molecule.

$$\begin{aligned} p &= qd = 2e \cdot 0.58 \quad (\text{dipole } e = 4.803 \cdot 10^{-18} \text{ (cgs)}) \\ (0.58 - x)2e' &= (2/4.803)e \\ (0.58 - x) &= 1/4.803 e = 0.21e \end{aligned}$$

A Theory of Water and Ionic Solution, with Particular Reference to Hydrogen and Hydroxyl Ions



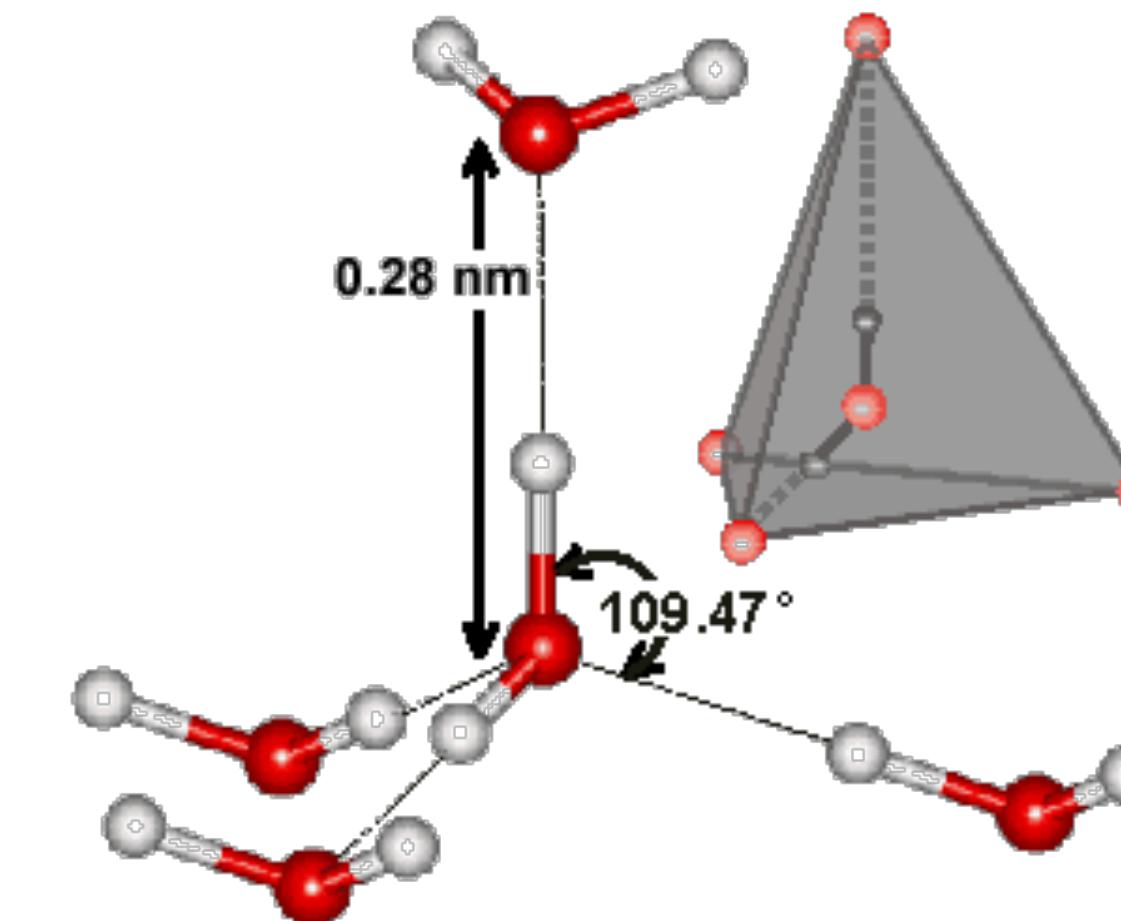
J. D. BERNAL AND R. H. FOWLER, *University of Cambridge, England*

(Received April 29, 1933)

$$(0.58 - x)\epsilon' \approx 0.21\epsilon.$$

...This leaves only the choice of x arbitrary between 0 and 0.37\AA (otherwise ϵ' becomes >1). It is plainly nearer the former because for 6 of the 10 electrons of the system the negative centre must be close to the O nucleus. A value 0.15 for x , giving $\epsilon'=0.49\epsilon$ would not be unreasonable and has the advantage for calculation of giving a tetrahedral arrangement of + and - charges".

$$\begin{aligned}\hat{H}\hat{O}H &= 102.75 \\ \hat{H}\hat{O}'H &= 109.9\end{aligned}$$



TIPS & TIP3P (Jorgensen et al, 1981 - 1983)

J. Am. Chem. Soc. **1981**, *103*, 335–340

Transferable Intermolecular Potential Functions for Water, Alcohols, and Ethers. Application to Liquid Water¹

William L. Jorgensen²

Contribution from the Department of Chemistry, Purdue University, West Lafayette, Indiana 47907. Received May 20, 1980

Rotation-Vibration Spectra of Deuterated Water Vapor*

W. S. BENEDICT, *The Johns Hopkins University, Baltimore, Maryland*

AND

N. GAILAR AND EARLE K. PLYLER, *National Bureau of Standards, Washington 25, D. C.*
(Received August 5, 1955)

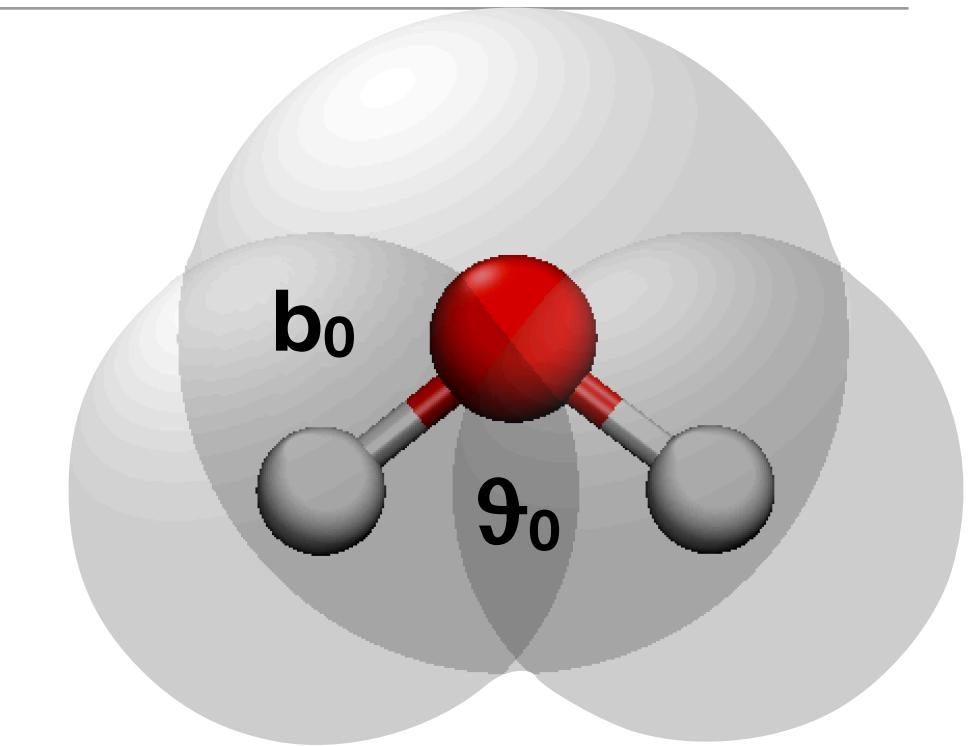
Spectra of heavy water have been obtained under high resolution between $1.25\text{--}4.1\mu$ ($2400\text{--}8000\text{ cm}^{-1}$). Approximately 4500 lines have been measured, and the majority of them analyzed into ten bands of D_2O and nine bands of HDO . The analysis is described in some detail, spectra of all bands are shown and a partial table of lines and a complete table of energy levels are presented. The vibration-rotation constants are derived and compared with those of H_2O .

Step 1: Internal geometry

$$b_0 = 0.9572 \text{ \AA}$$

$$\theta_0 = 104.52$$

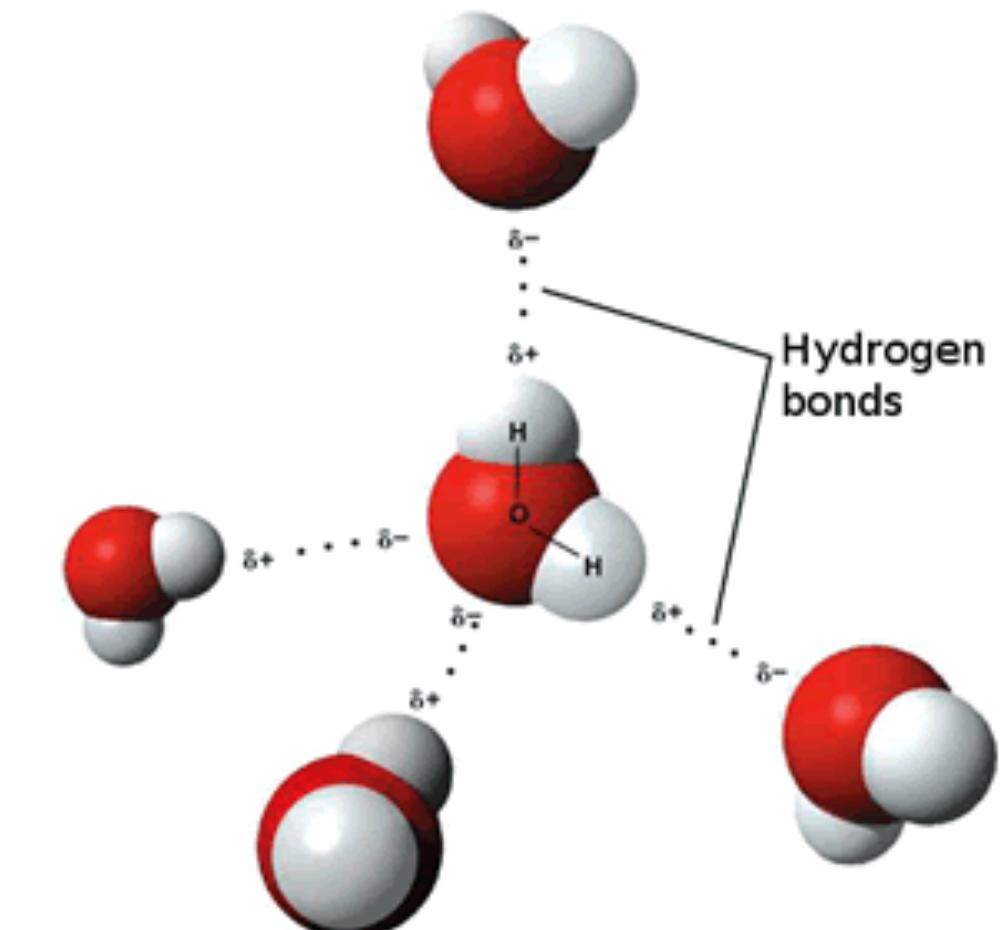
rigid



Step 2: Intermolecular interactions

Key idea: reproduce the energy of a water dimer.

- 1. Basic interacting unit of water**
- 2. Hydrogen bonding**
- 3. Recent (1973) measures of the structure of the dimer**



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DI MILANO



TIPS & TIP3P (Jorgensen et al, 1981 - 1983)

$$E_{m,n} = \sum_{i \in m} \sum_{j \in n} \frac{q_i q_j e^2}{r_{ij}} + \frac{A}{r_{OO}^{12}} - \frac{C}{r_{OO}^6}$$

$q_O = -2q_H$
LJ only for oxygen
3 parameters

$q_O = -0.8, -0.834$
 $A^2 = 580 \times 10^3, 582 \times 10^3$
 $C^2 = 525, 595$

$R_{OO} = 2.98 \pm 0.04 \text{ \AA}$
 $\Theta = 60^\circ \pm 10^\circ$
Gas Dipole: 1.85D

Microwave spectrum and structure of hydrogen bonded water dimer

Thomas R. Dyke and J. S. Muenter

Department of Chemistry, University of Rochester, Rochester, New York 14627
(Received 27 December 1973)

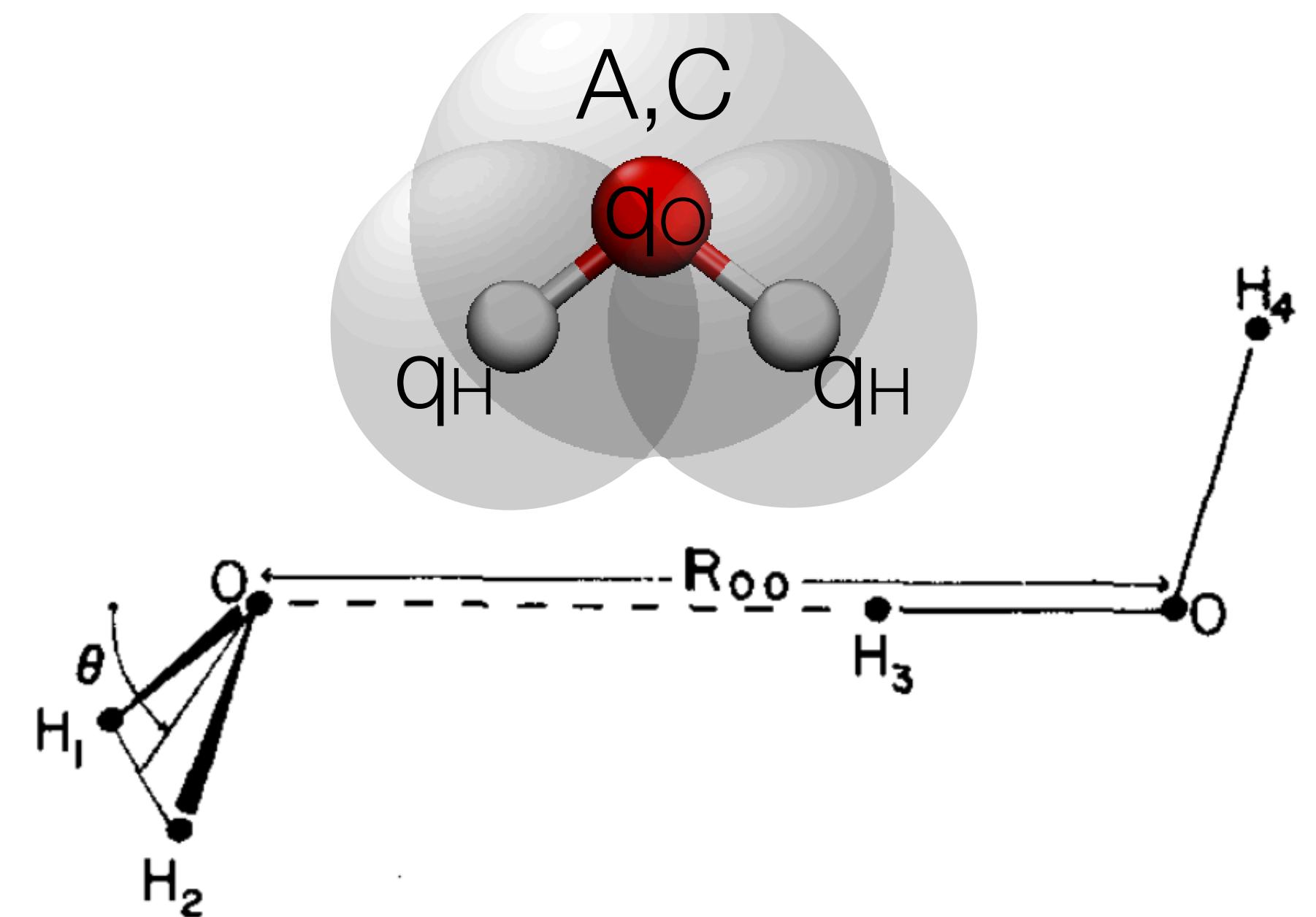


FIG. 1. Structure of water dimer.



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TIPS & TIP3P

(Jorgensen et al, 1981 - 1983)

Dimer	TIPS	EXPT
R_{OO}	2.78	$2.98 \pm 0.04 \text{ \AA}$
θ	27	$60^\circ \pm 10^\circ$
Energy	5.70	$5.44 \pm 0.7 \text{ kcal/mol}$

Are the results transferable
to liquid water?

Table IV. Computed and Experimental Properties of Liquids at 25 °C^a

	water	TIP	exptl ^b
ΔH_V°		8.9	10.7
C_V		15.0	17.9

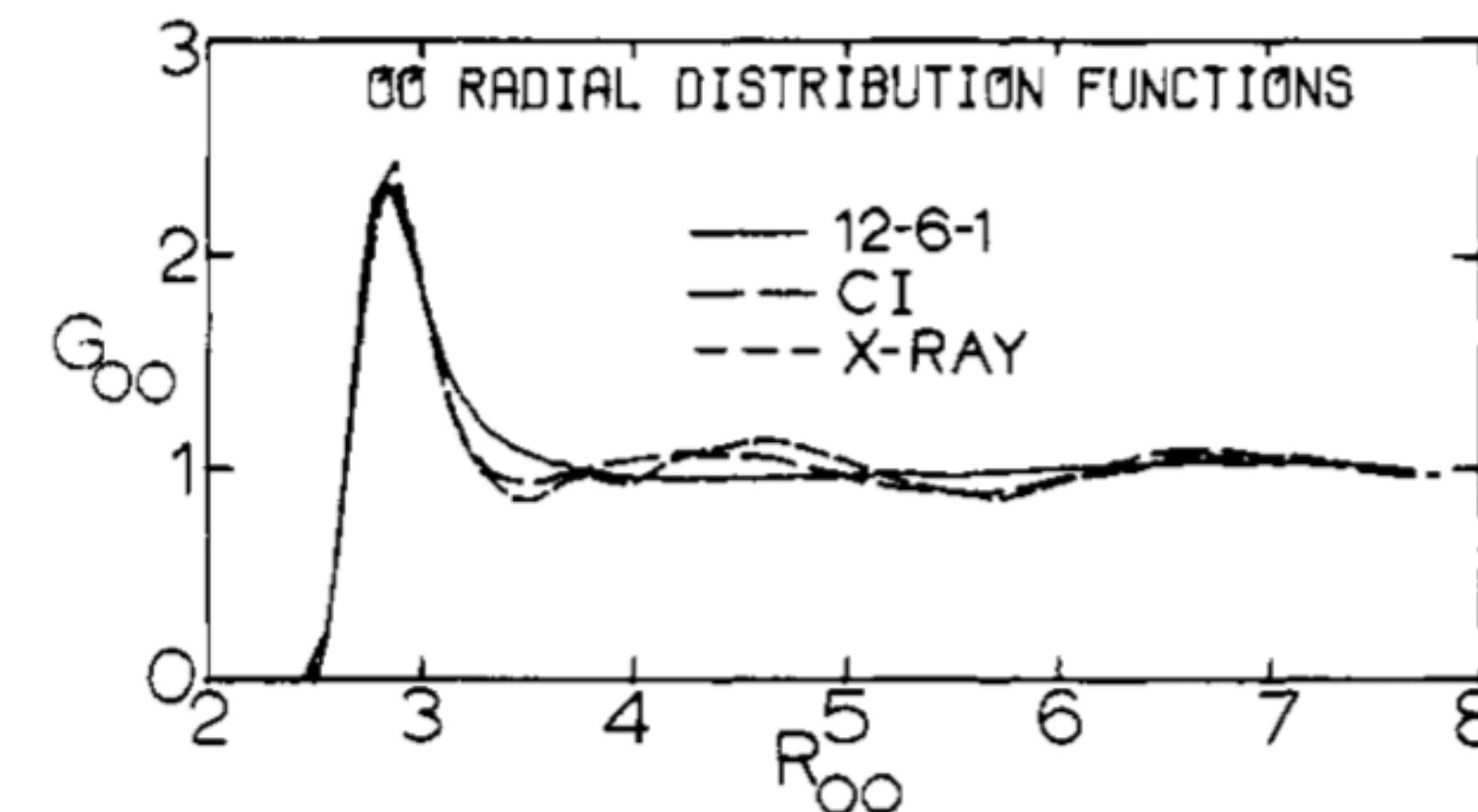


Figure 7. OO radial distribution functions for liquid water at 25 °C from X-ray data (ref 33) and the TIP and CI potential functions.

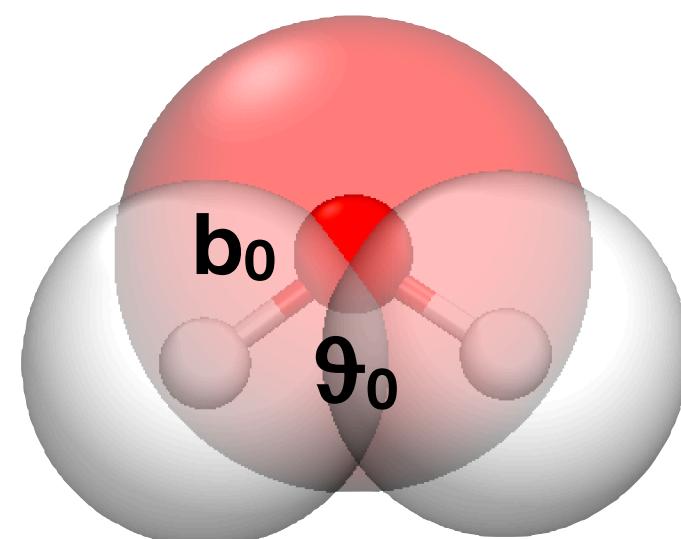


SPC: Berendsen et al 1981

SPC, key idea: try to reproduce the second peak in the g(r) and reproduce the Enthalpy of Vaporisation, 2 restraints for 2 parameters.

Step 1: Internal geometry

$b_0 = 1.0 \text{ \AA}$
 $\theta_0 = 109.28$ rigid

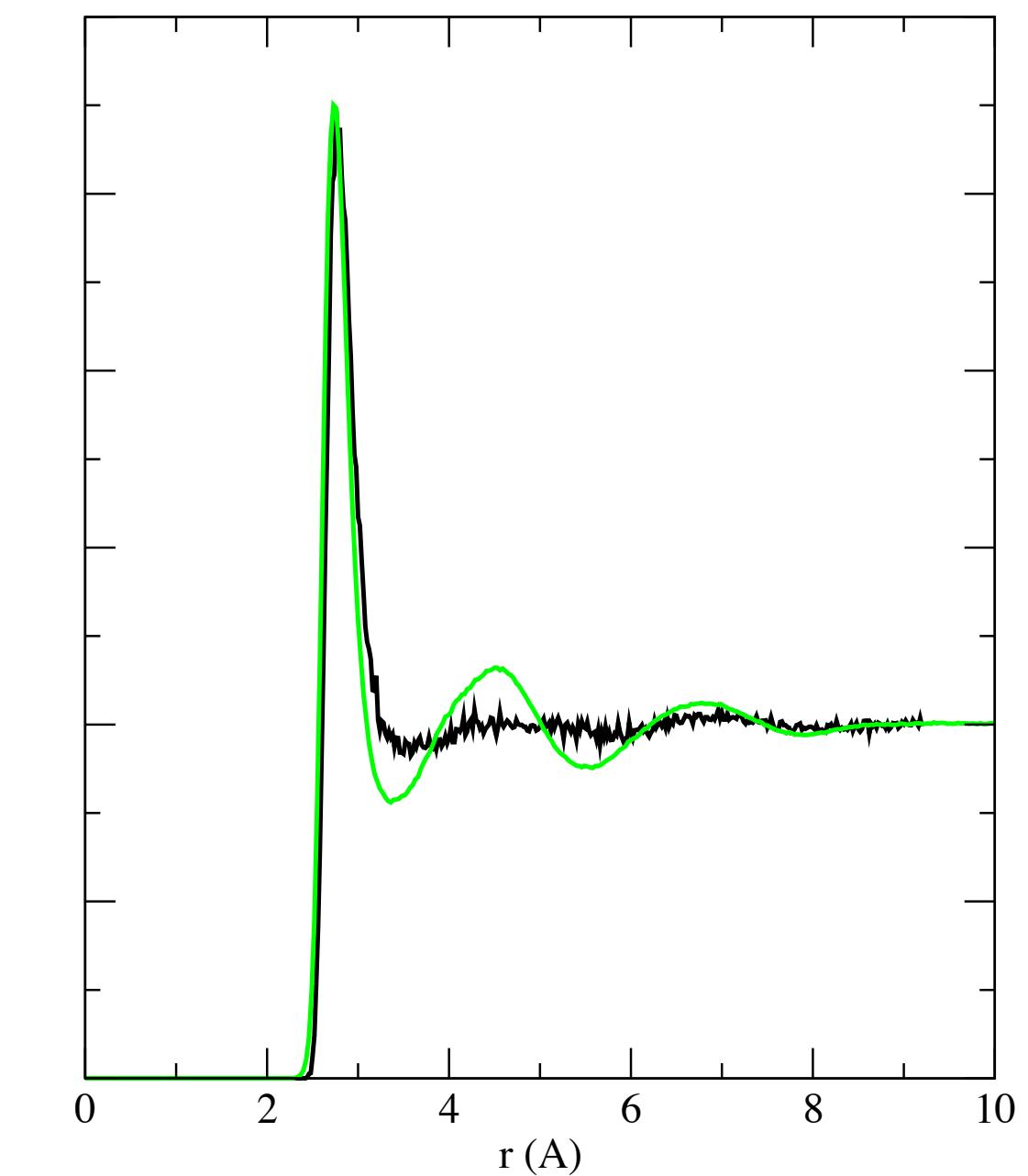
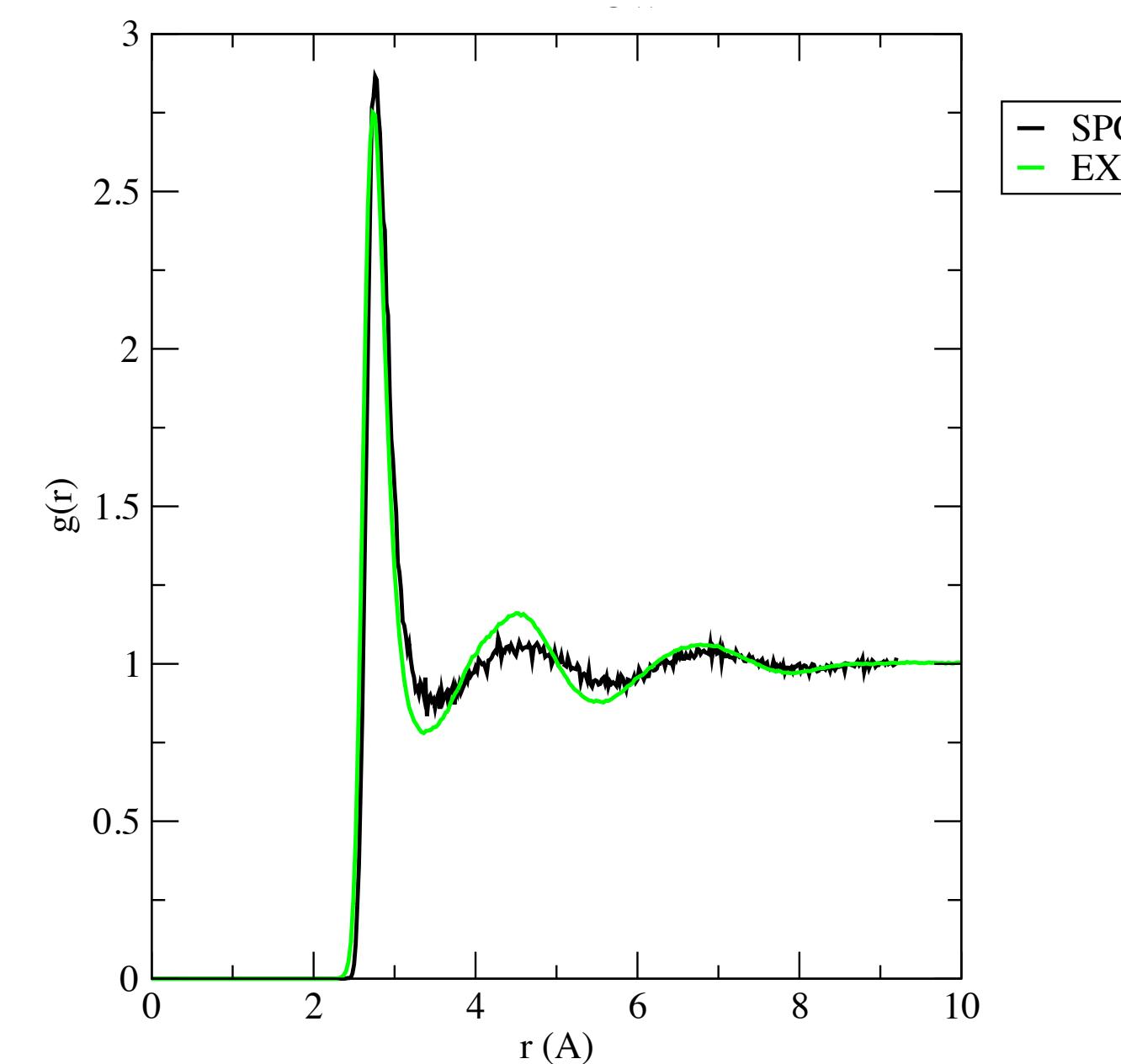


Step 2: Intermolecular interactions

$C = 0.37122 \text{ nm} (\text{kJ.mol}^{-1})^{1/6}$ This was derived by London in ~1920

$$E_{m,n} = \sum_{i \in m} \sum_{j \in n} \frac{q_i q_j e^2}{r_{ij}} + \frac{A}{r_{OO}^{12}} - \frac{C}{r_{OO}^6}$$

$q_0 = -0.82$
 $A = 0.3428$



SPC water can indeed better reproduce the structure of water in solution



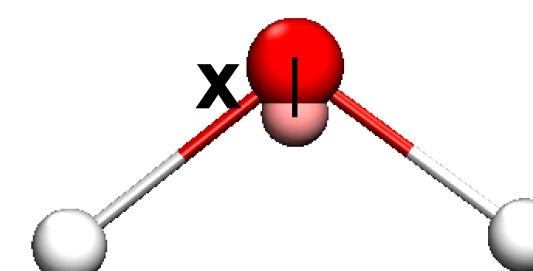
A general purpose model for the condensed phases of water: TIP4P/2005

J. L. F. Abascal^{a)} and C. Vega

*Departamento de Química Física, Facultad de Ciencias Químicas, Universidad Complutense,
28040 Madrid, Spain*

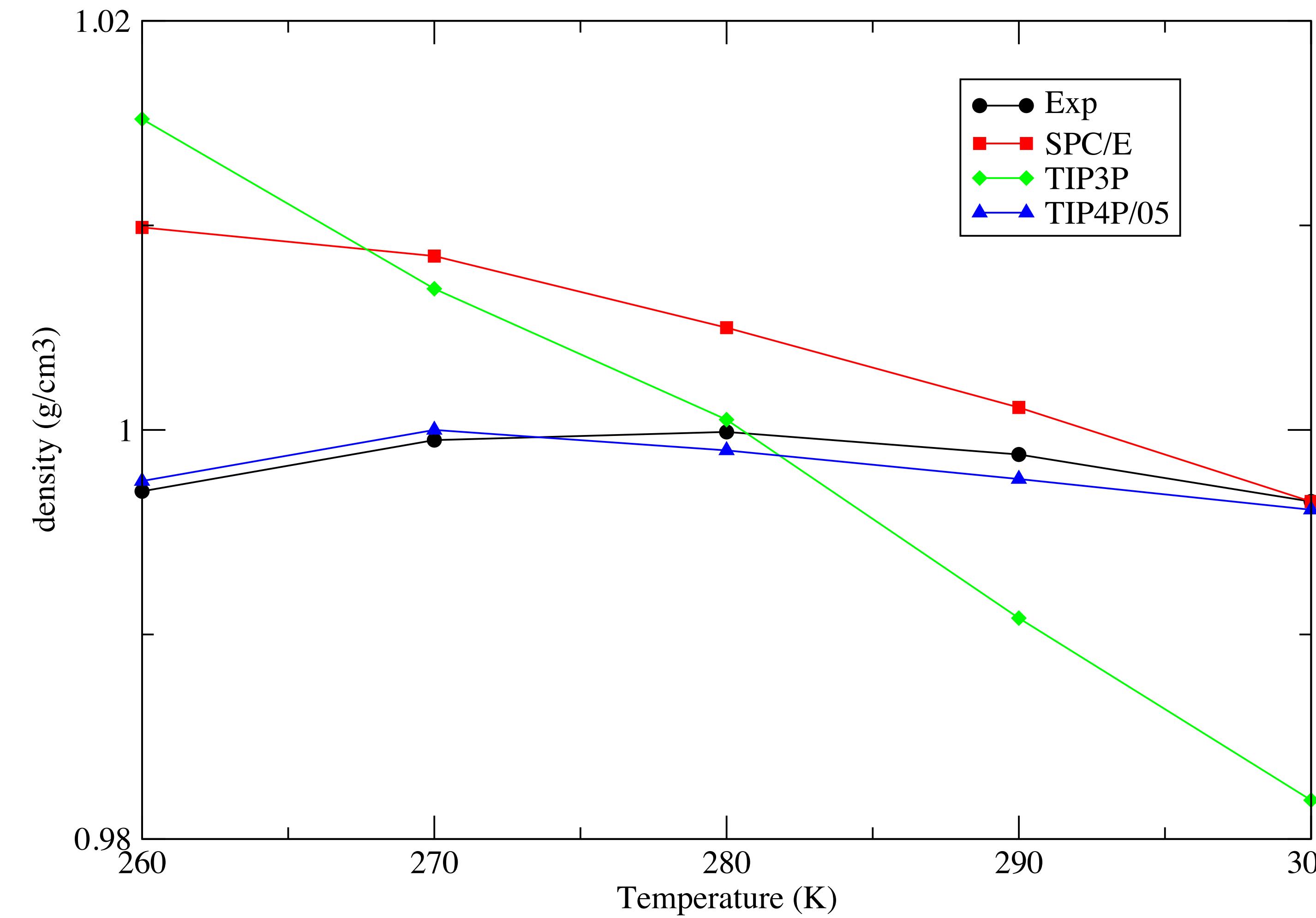
Key ideas: 1) use the water geometry (as in TIPS) 2) use a dislocated positive charge following Bernal - Fowler to better account for the electronic density. 3) use the correction for the enthalpy of vaporisation as in SPC/E; 4) use also properties from ice to train the model.

Systematic optimisation: parameters (q_0 , A , C , and x -> this the displacement of the additional particle with respect to the symmetry axis) and properties (T_{md} , H_v , $\rho(298K)$, density of ice II at 123 K and 0 MPa, and of ice V at 223 K and 530 MPa, and the range of temperatures at which ice III is the thermodynamically stable ice at a pressure of 300 MPa



$b_0 = 0.967 \text{ \AA}$	$q_0 = -1.1128$
$\theta_0 = 104.52^\circ$	$A = 3.06010 \times 10^6$
$x = 0.1546 \text{ \AA}$	$C = 307978$

Water Density as a function of the Temperature



Some comparisons of water models properties

Property (298K, 1bar)	TIP3P	SPC/E	TIP4P/05	TIP4P/ ϵ	EXP
Hv (kcal/mol)	10.2 (9.0)	11.6 (10.4)	11.9 (10.8)	11.8 (10.7)	10.7
Cp (cal/mol/K)	18.5	19.0	21	21	17.9
Density (g/cm ³)	0.984	0.998	0.996	0.995	0.997
Diffusion (cm ² /s)	5.6	2.7	2.3	2.3	2.3
Dielectric constant	94	68	58	78.5	78.5



Force-Fields: from energy to forces

In physics forces are defined as the negative gradient of the energy, so if we know an energy function, a function that given an atomic configuration gives us an energy, we can calculate the forces on the atoms for that configuration:

$$\mathbf{F}(\mathbf{R}) = -\nabla V(\mathbf{R})$$

Here **bold** characters denote a Vector or more general a list of vectors, for example in the case of a force-field applied to alanine dipeptide it will be a list of 22 vectors, each one pointing to an atom, while standard characters denote a Scalar (a number)

How to:

$$\nabla f = \frac{\partial f}{\partial x} \mathbf{i} + \frac{\partial f}{\partial y} \mathbf{j} + \frac{\partial f}{\partial z} \mathbf{k},$$

$$f(x, y, z) = 2x + 3y^2 - \sin(z)$$

$$\nabla f = 2\mathbf{i} + 6y\mathbf{j} - \cos(z)\mathbf{k}.$$

The gradient is the direction of maximum growth of the curve.

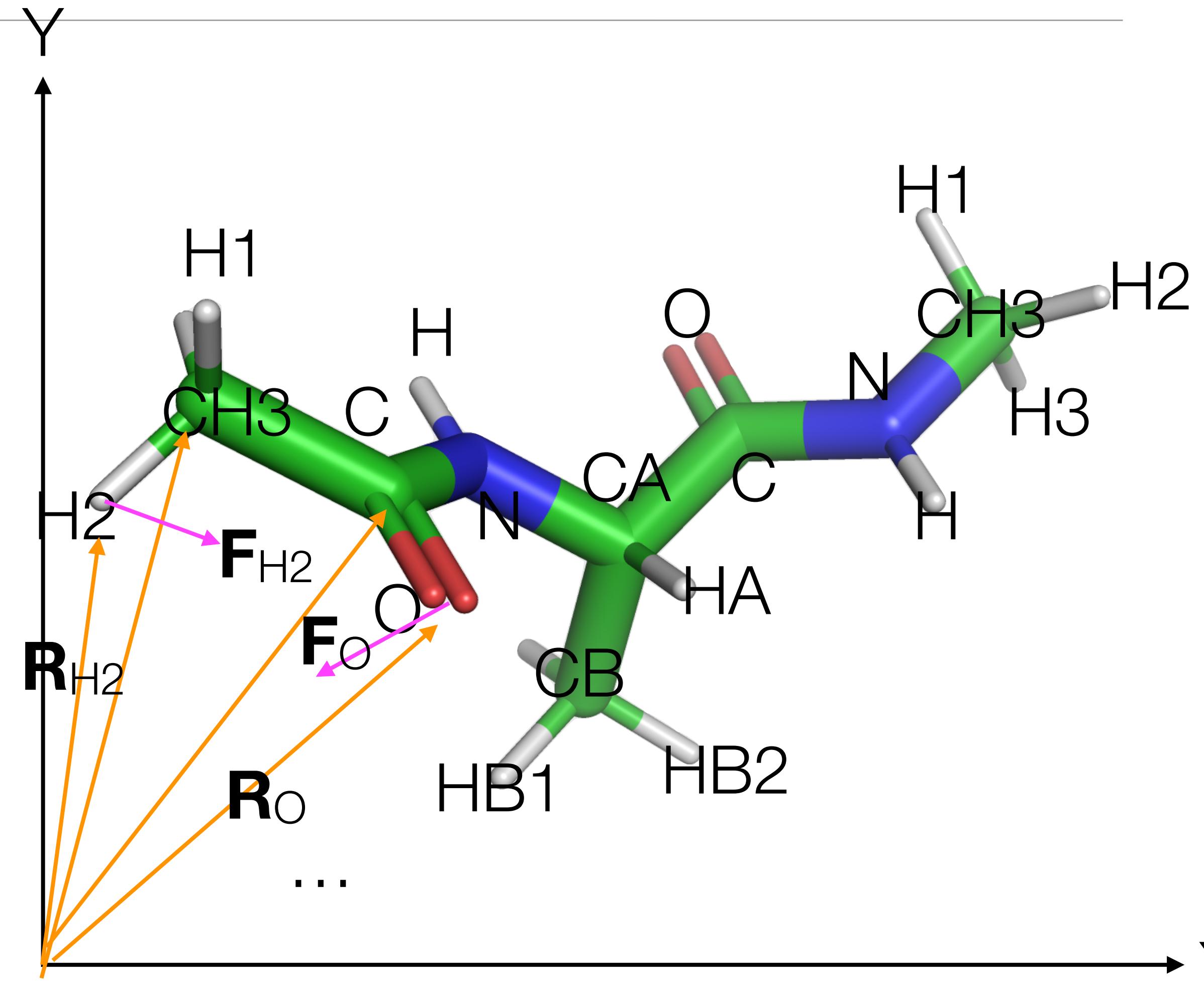


Force-Fields: from energy to forces

$$\mathbf{F}(\mathbf{R}) = -\nabla V(\mathbf{R})$$

The gradient is the direction of maximum growth of the curve, so negative gradient will result in one vector per atom, centered on the atom and pointing in the direction of maximum decrease of the energy. The forces will indicate in which direction the atom should move to lower the overall energy of the complete system.

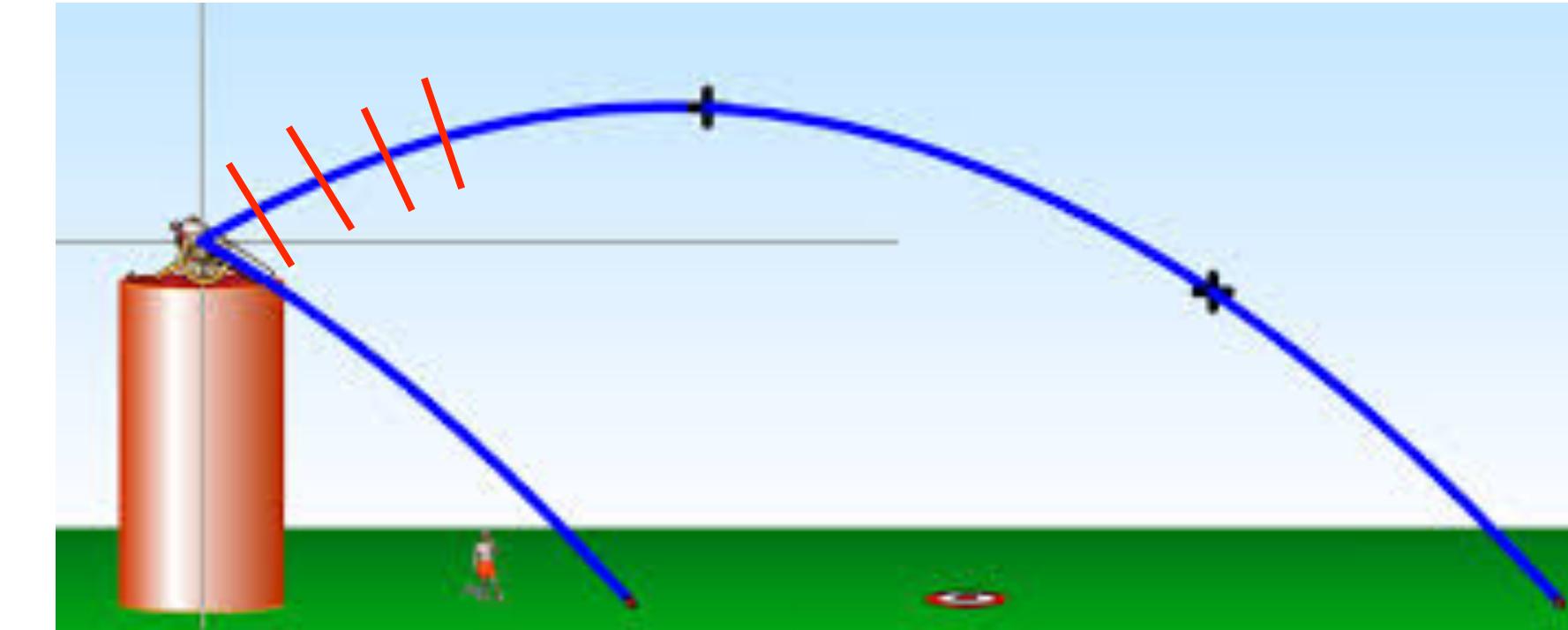
In the figure **orange arrows** represent **positions** vectors for some of the atoms, while **magenta arrows** represent **force** vectors for some of the atoms.



Molecular Dynamics: from forces to motion

Second Newton's Law: the action of the total force \mathbf{F} on a body produces an acceleration \mathbf{a} equal to the total force divided by the mass m of the body: $\mathbf{a}=\mathbf{F}/m$

In principle my forces will change continuously with the motions of the atoms, but we want to solve an iterative problem where atoms are moved by small variations, or alternatively, we want to move atoms forward in time only for a very small time step, how small? Somehow compatible with the time scale of atom motions \sim fs (10^{-15} s), on this time scale we can consider the forces as constant.



Would be like solving simple physics problems for small steps

When forces are constant we know how bodies move: **uniformly accelerated motion**



Classical Mechanics: from forces to motion

$$\mathbf{F}=m\mathbf{g};$$

$$a_x(t)=F_x/m=0;$$

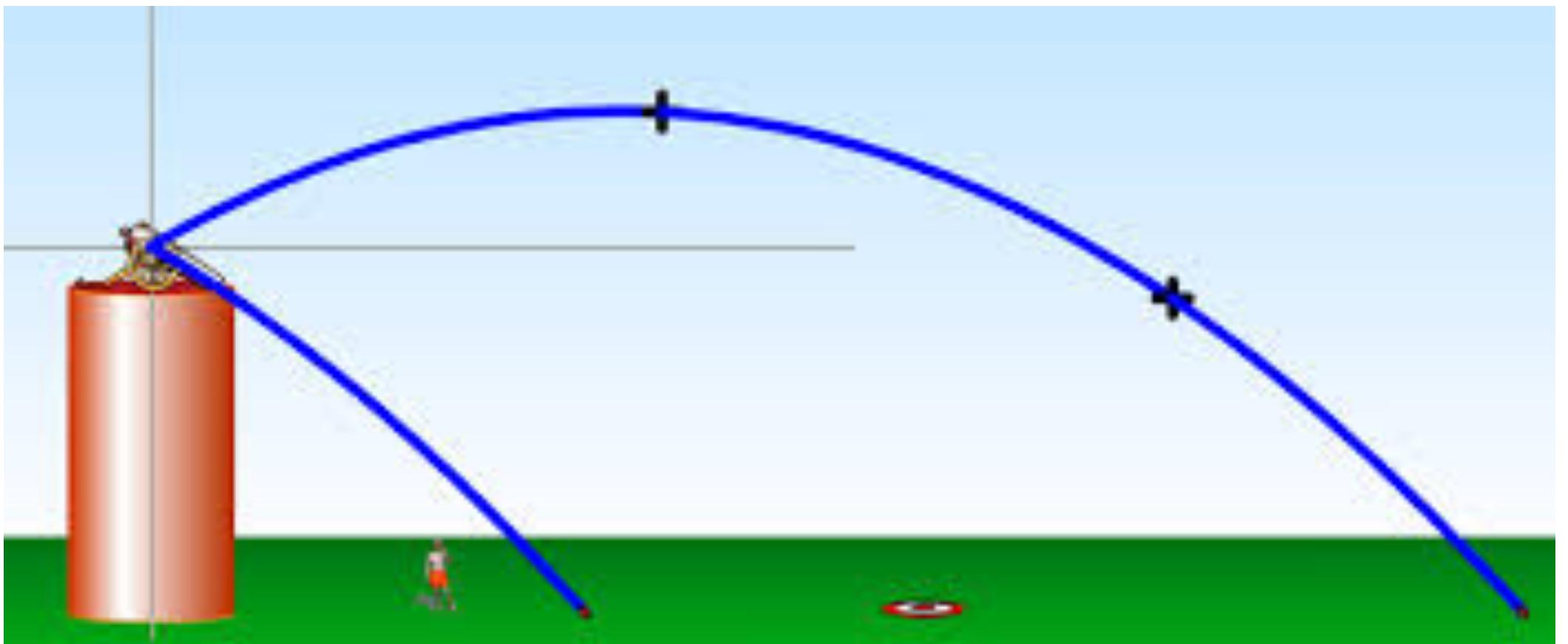
$$a_y(t)=F_y/m=-g;$$

$$v_x(t)=v(0);$$

$$v_y(t)=a_y(t)*t+v_y(0)=-g*t;$$

$$x(t)=v(0)*t;$$

$$y(t)=-0.5*a_y(t)*t^2+v_y(0)*t+y(0)=-0.5*g*t^2+y(0);$$



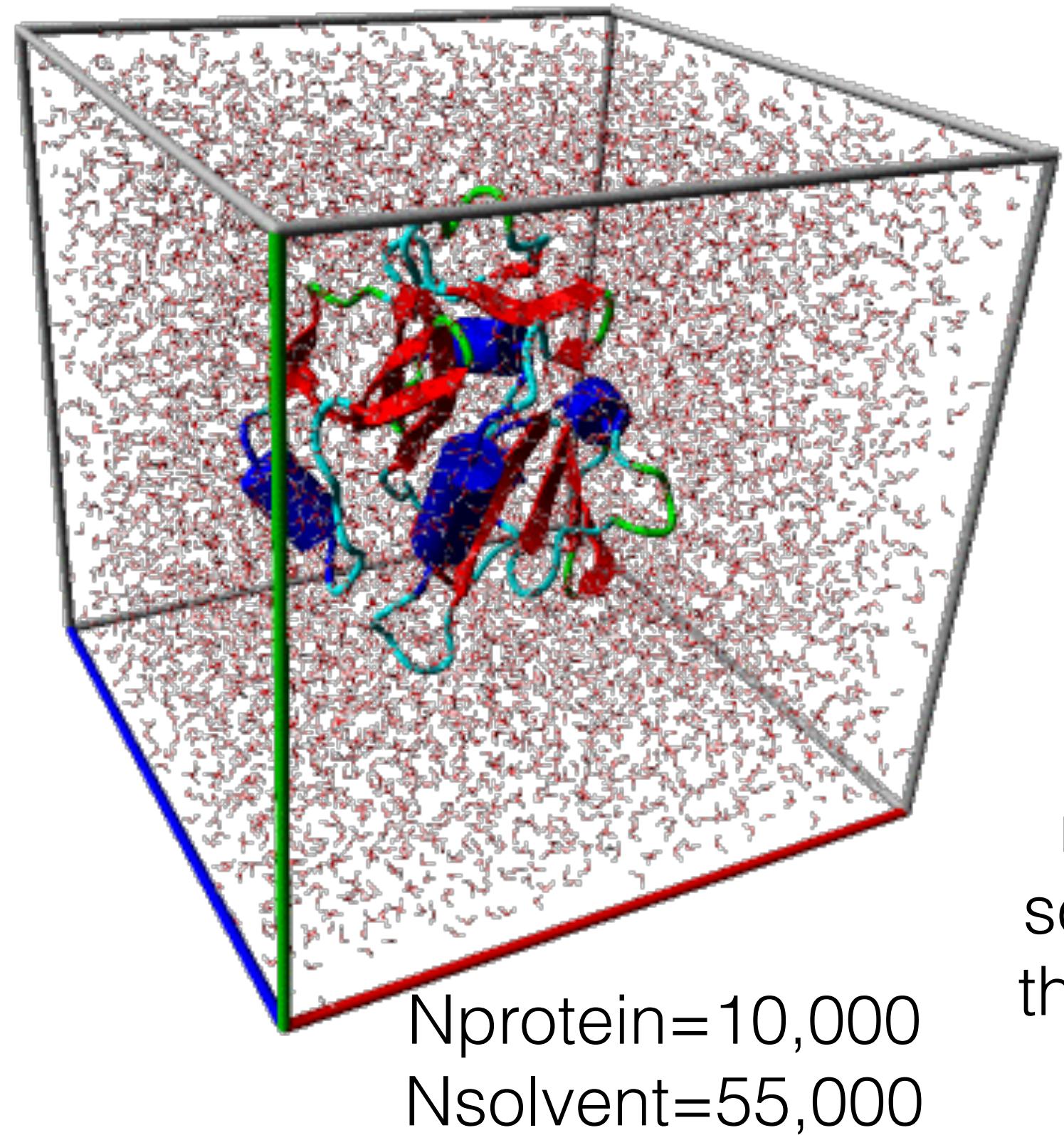
So how can we implement this
as a computer algorithm?



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The simulation box determines the size so the speed of your MD



$$V(r) = \sum_{bonds} k_b(b - b_0)^2 + \sum_{angles} k_\theta(\theta - \theta_0)^2 + \sum_{torsions} k_\phi[\cos(n\phi + \delta) + 1] \\ + \sum_{\substack{\text{nonbond} \\ \text{pairs}}} \left[\frac{q_i q_j}{r_{ij}} + \frac{A_{ij}}{r_{ij}^{12}} - \frac{C_{ij}}{r_{ij}^6} \right]$$

To calculate the energy and thus the forces we need to calculate distances between all pairs of atoms (N^2 calculations)

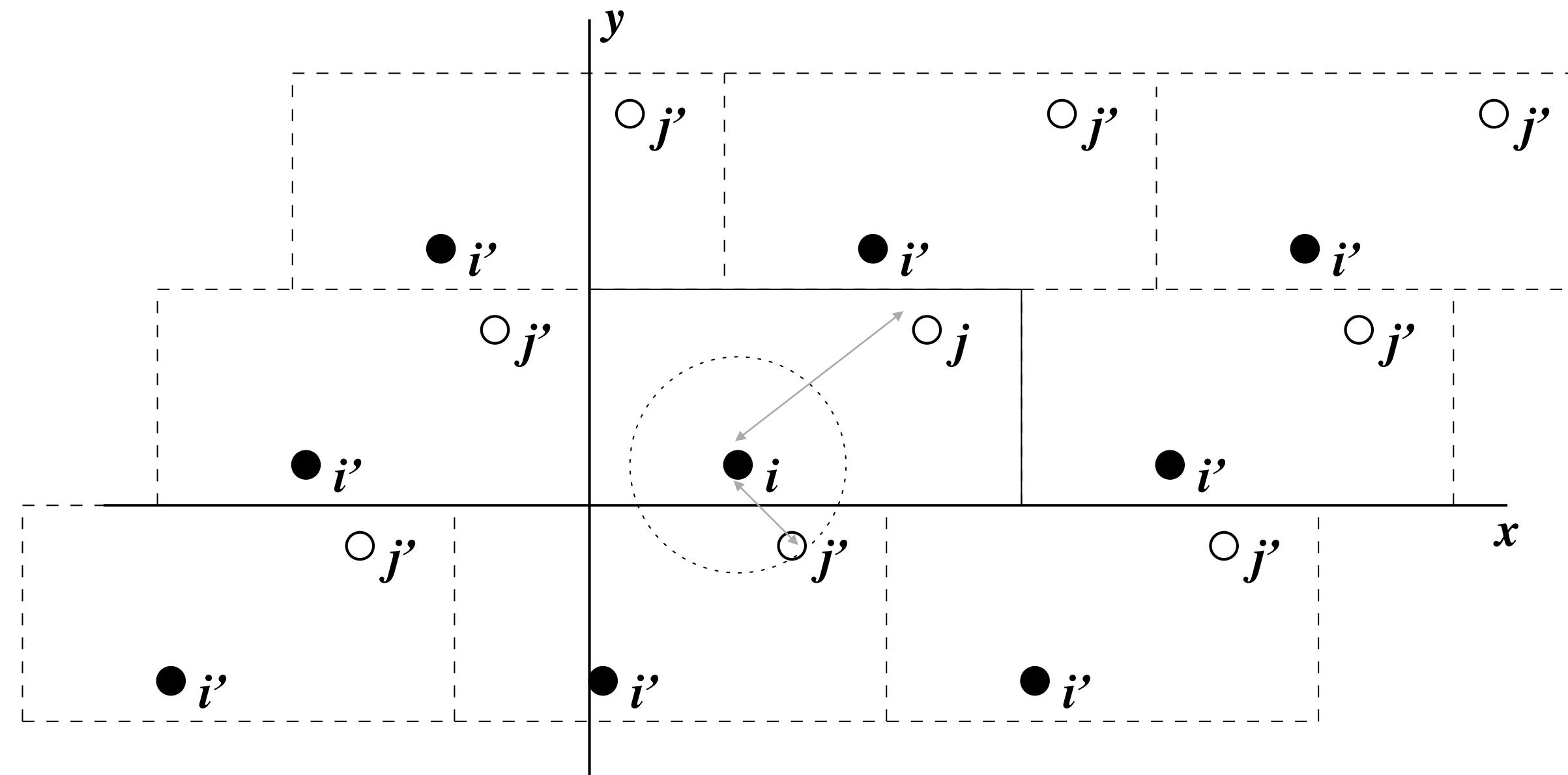
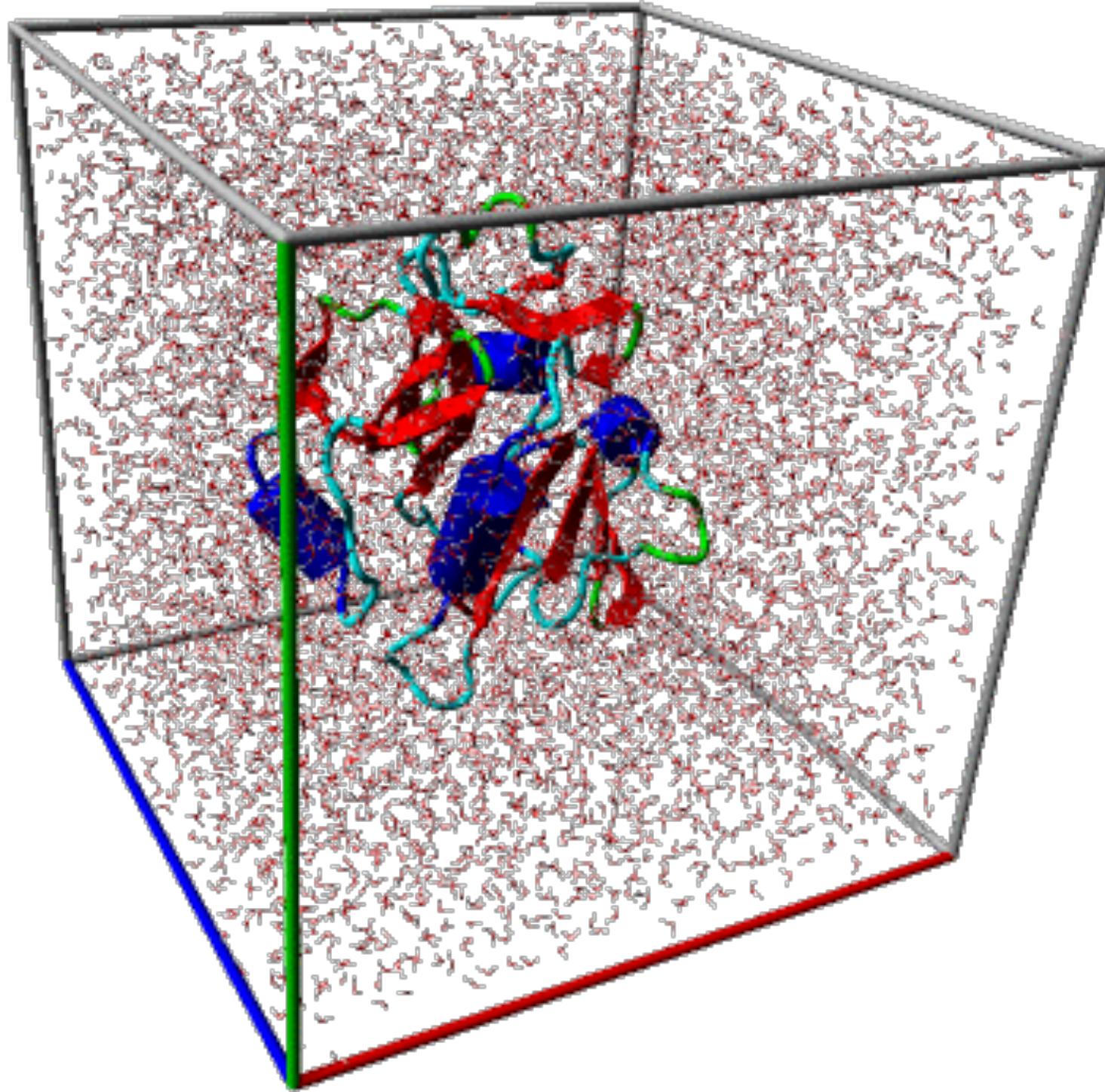
So how big should we set the box?
Consider that we want to simulate molecules in their solution environment so the number of atoms is not only that of the molecule of interest but also that of all the solvent atoms.

Furthermore the size of the box, if the box is like a cuvette can generate artefacts due to the protein-box sides interactions.

Does the protein have the space to experience conformational changes?



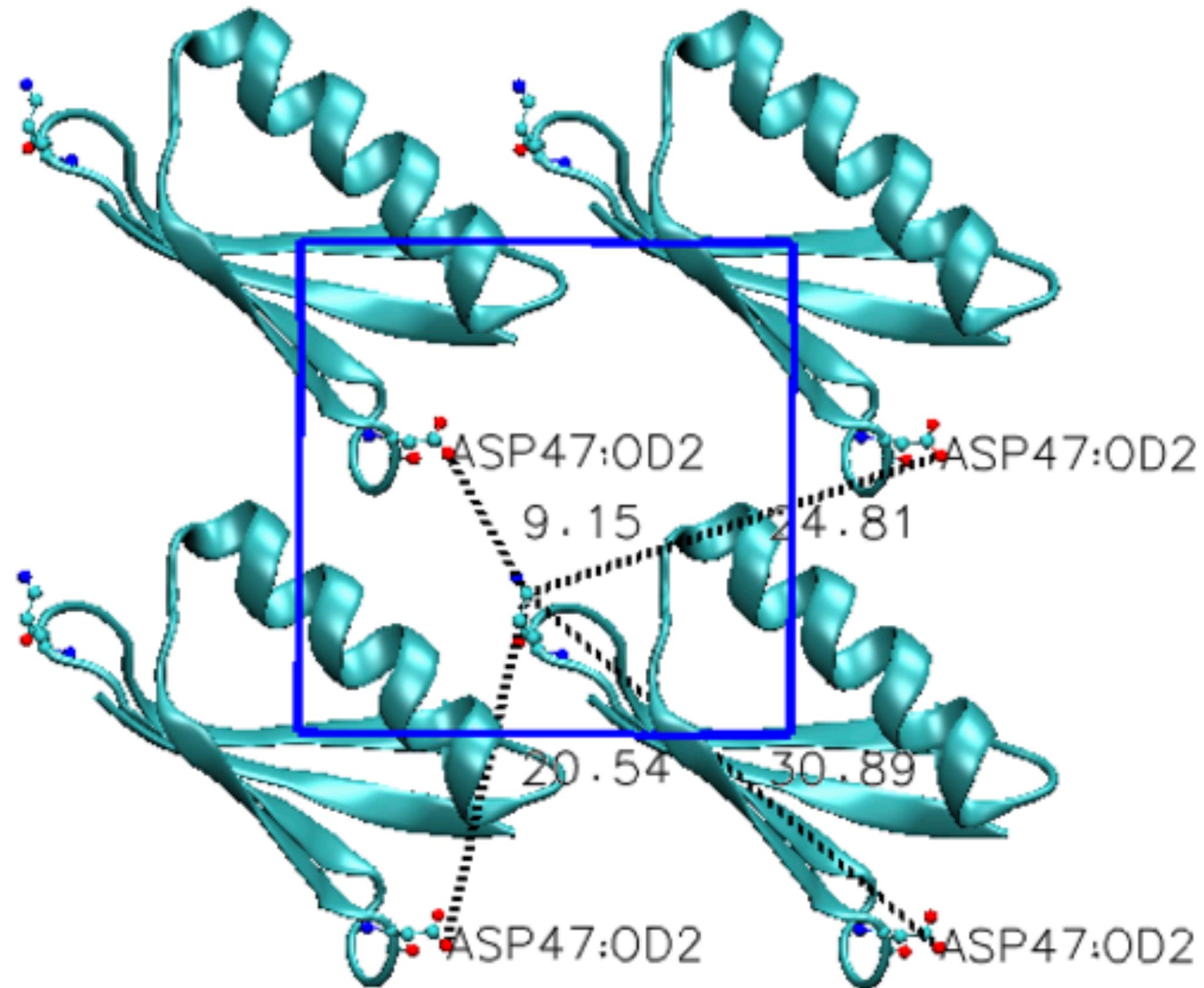
Periodic Boundary Conditions allow to simulate boxes without borders



By defining PBC at least in principle we can simulate a molecule like it is ideally diluted, but if the box is periodic the distance between two atoms can assume an infinite number of values, what is the “correct” distance?

Minimum Image Convention: the distance between two atoms is always the minimum with respect to all the neighbour images interactions can only be calculated up to a distance equal to half of the shortest side of the box

Minimum Image Convention requires a minimum box size



For example one can try to make the box large enough so that the intramolecular distance is generally shorter than the corresponding intermolecular distance. This is not the case of what is shown on the left. Nonetheless the above approach may be too computationally demanding.

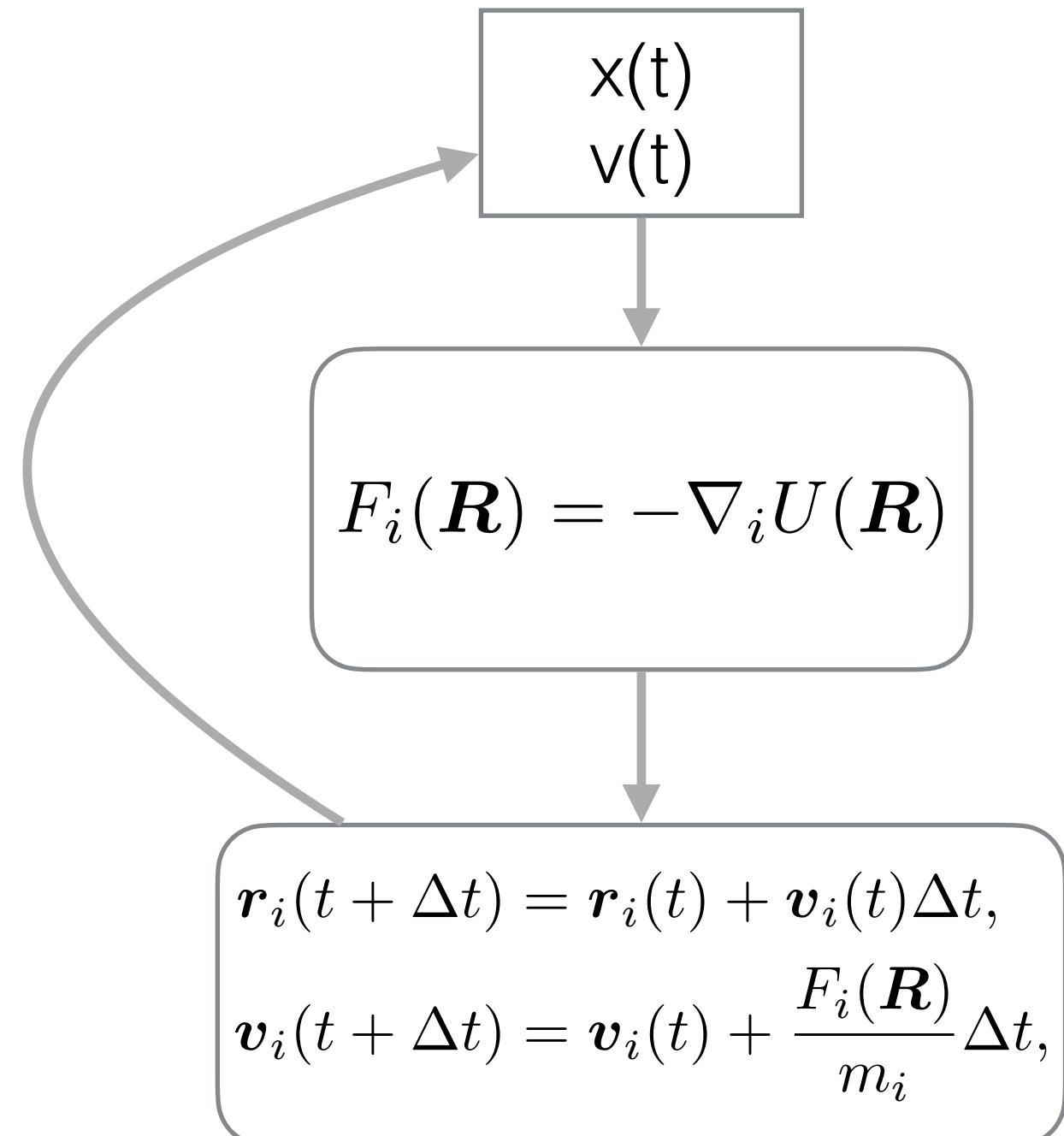


A simple MD algorithm: The Euler algorithm

$$\mathbf{v}_i(t + \Delta t) = \mathbf{v}_i(t) + \frac{\mathbf{F}_i(\mathbf{R})}{m_i} \Delta t,$$

$$\mathbf{r}_i(t + \Delta t) = \mathbf{r}_i(t) + \mathbf{v}_i(t) \Delta t$$

For each atom i , velocities are updated using forces and are used to update positions, and each iteration of the algorithm is a step forward in time of size Δt



1. starting position and velocities
2. calculate forces
3. calculate new positions and velocities

The microscopic world:
Distances in nm
Times in ps
Energies in kJ/mol

To initiate the algorithm you need some initial position and velocities and a force-field.



Classical Physics: Important Properties

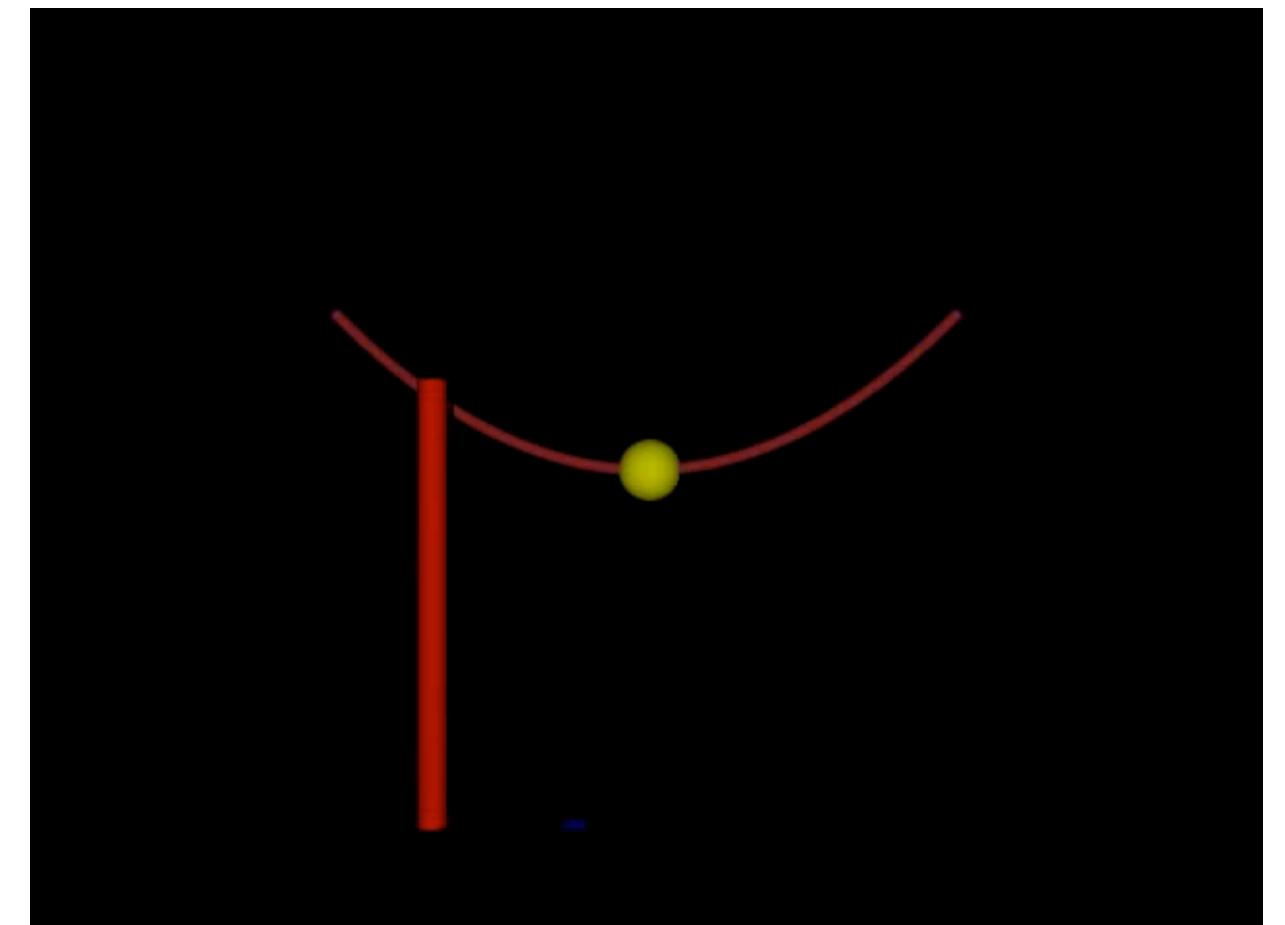
1. If a force does not depend explicitly on time (is conservative) then **the Total Energy of the system is conserved.**

$$E_k = \frac{1}{2}mv^2$$

Kinetic Energy (is the results of all masses and velocities)

$$E=U(x)+E_K(v)$$

Total energy is the sum of the Potential Energy (Force-Field energy) and the Kinetic energy.

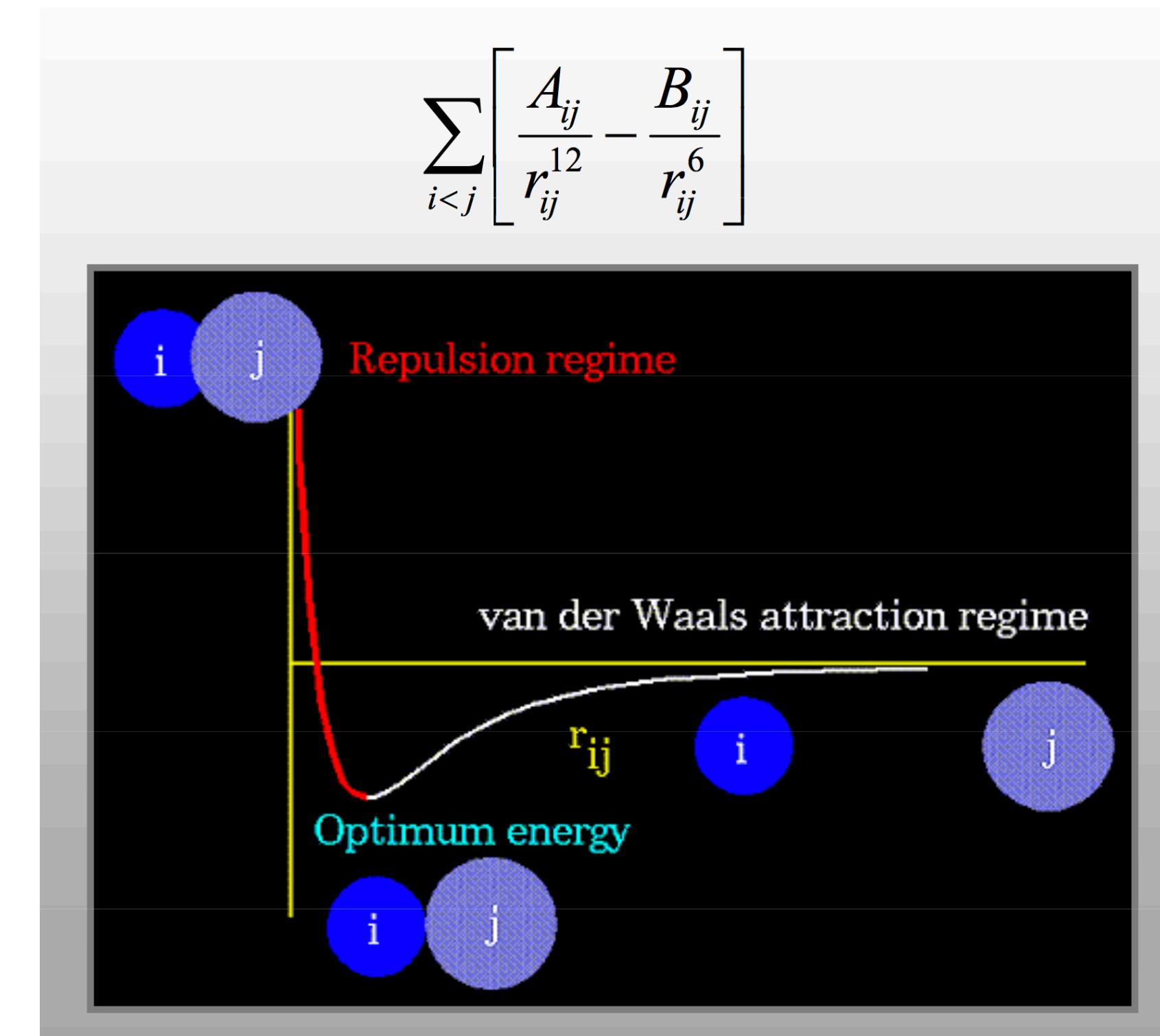
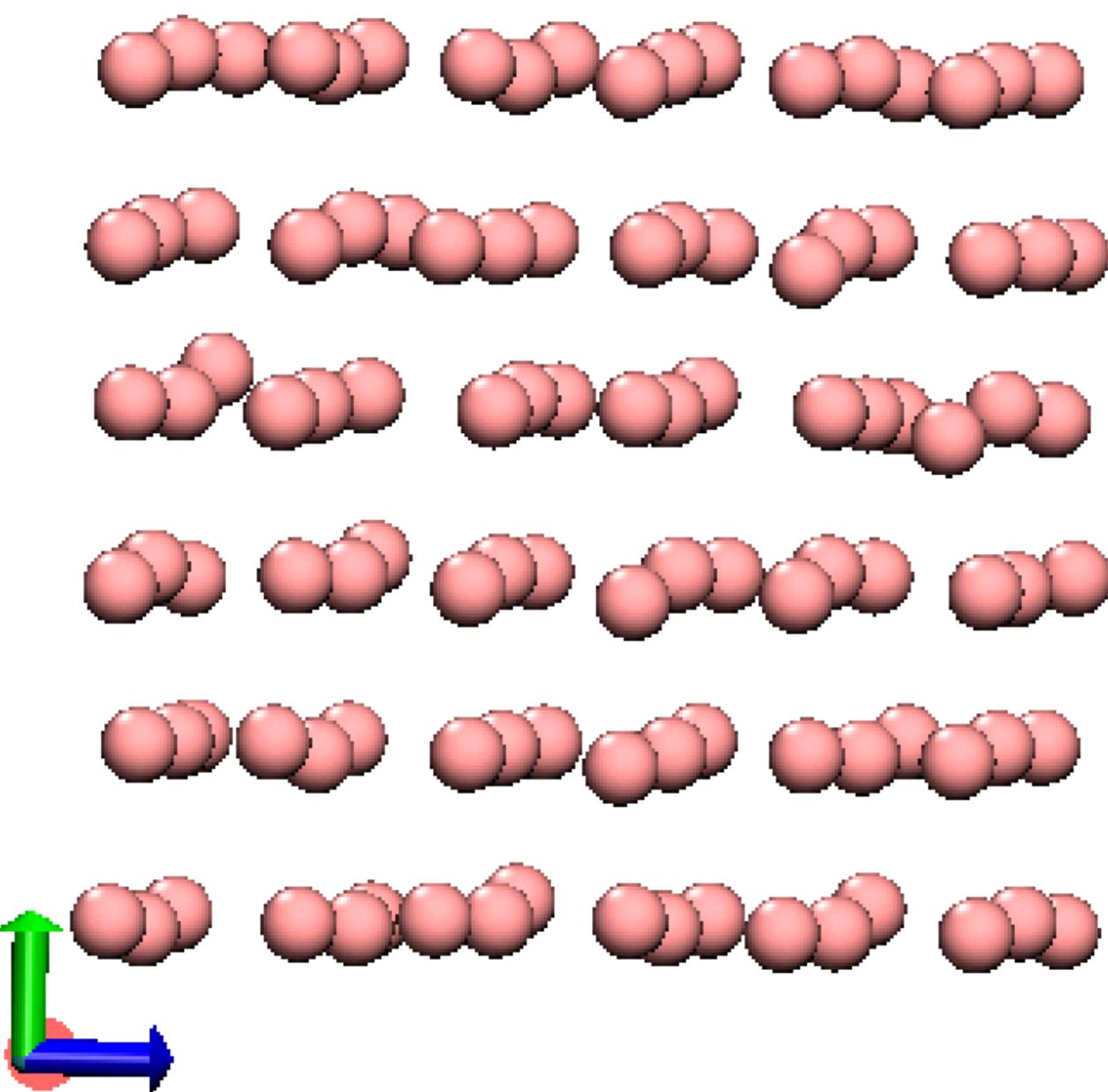


2. If a force does not depend explicitly on time (is conservative) then there is a symmetry for time inversion, i.e. going back in time is equivalent to invert the velocities.



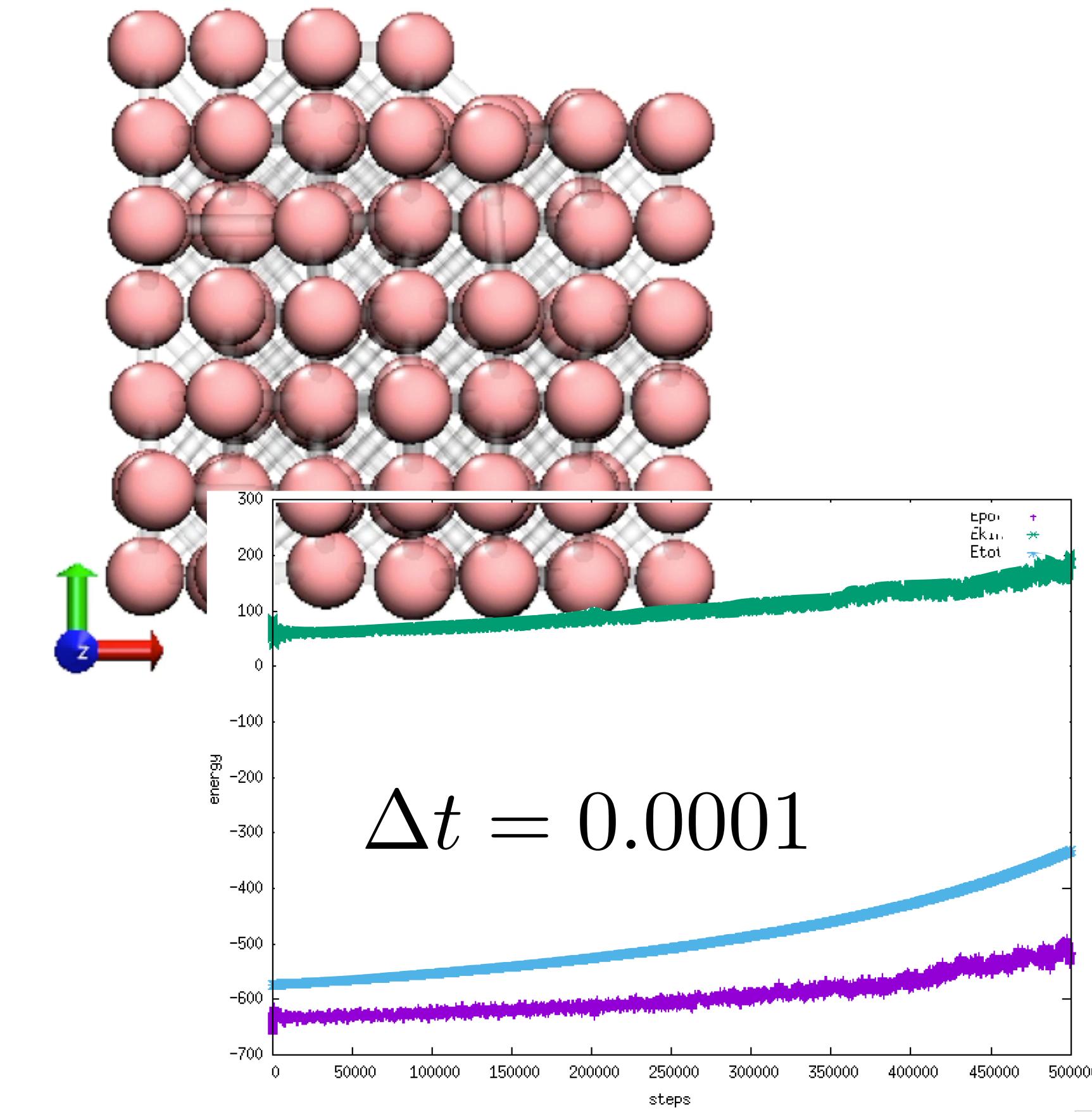
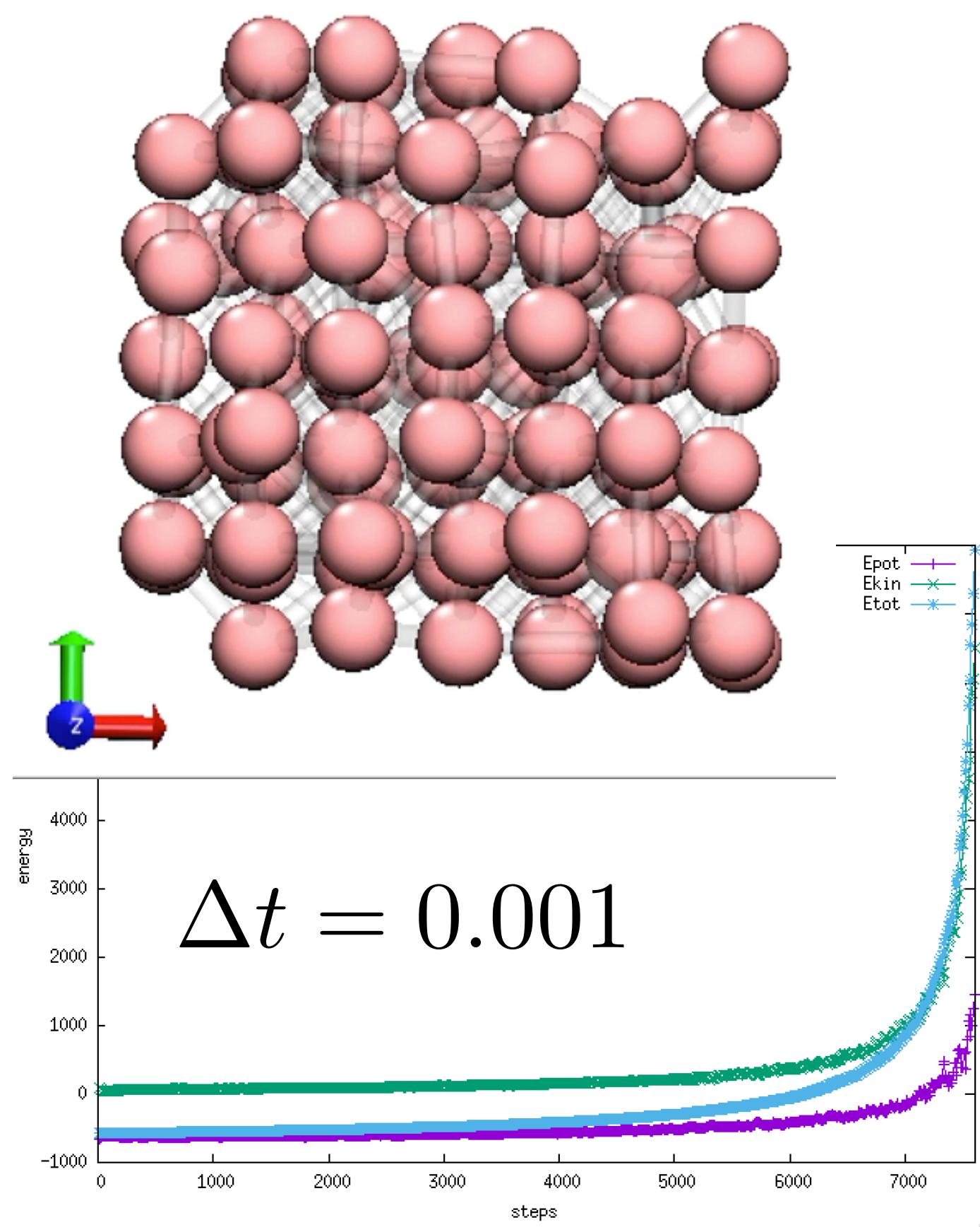
Molecular Dynamics using the Euler algorithm

let's take for example a set of 108 particles with LJ interactions (these can represent for example He atoms)

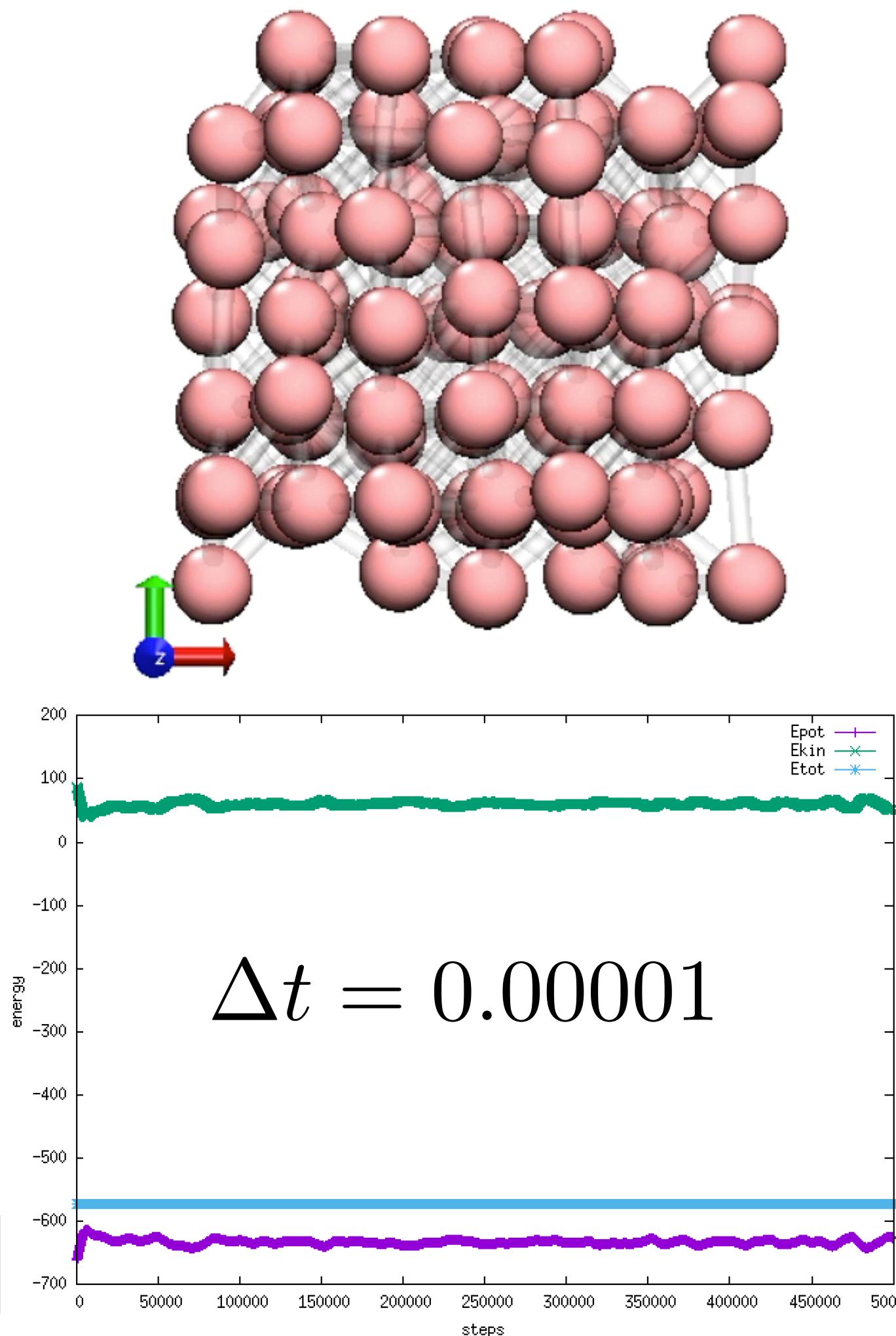


To **test if** the Euler algorithm is a good algorithm we could for example check whether the **total energy is conserved.**

Energy Conservation is violated using Euler algorithm



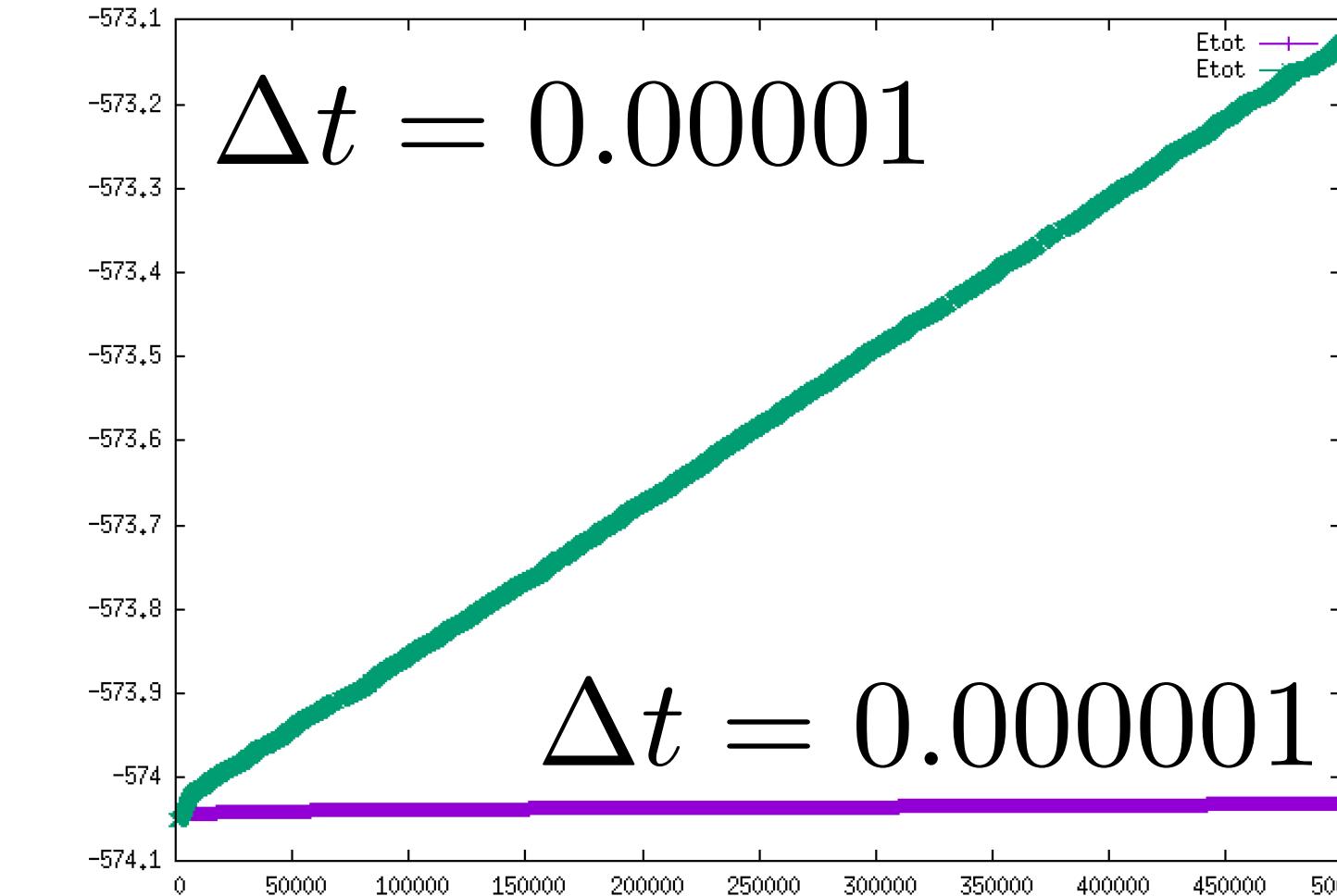
Energy Conservation is violated using Euler algorithm



The Euler algorithm cannot conserve the energy, there is not a time step for which the algorithm can work correctly.

Can we explain why?

there is still a systematic drifts



Euler algorithm does not conserve energy because it is not time-reversible

The Euler algorithm cannot conserve the energy, there is not a time step for which the algorithm can work correctly. Can we explain why? There is a second important property in classical physics: time reversibility.

What happen if we start in $r(t+dt)$ and we go back in time by dt ?

$$\begin{aligned} r(t + \Delta t - \Delta t) &= r(t + \Delta t) - v(t + \Delta t)\Delta t = \\ &= r(t) + v(t)\Delta t - v(t + \Delta t)\Delta t = \\ &= r(t) + v(t)\Delta t - v(t)\Delta t - a(t)\Delta t^2 = \\ &= r(t) - a(t)\Delta t^2 \neq r(t) \end{aligned}$$

It happen that we are not back in $r(t)$! So this algorithm is not time-reversible!



A better Molecular Dynamics algorithm: Velocity Verlet

$$\vec{v}(t + \frac{1}{2} \Delta t) = \vec{v}(t) + \frac{1}{2} \vec{a}(t) \Delta t.$$

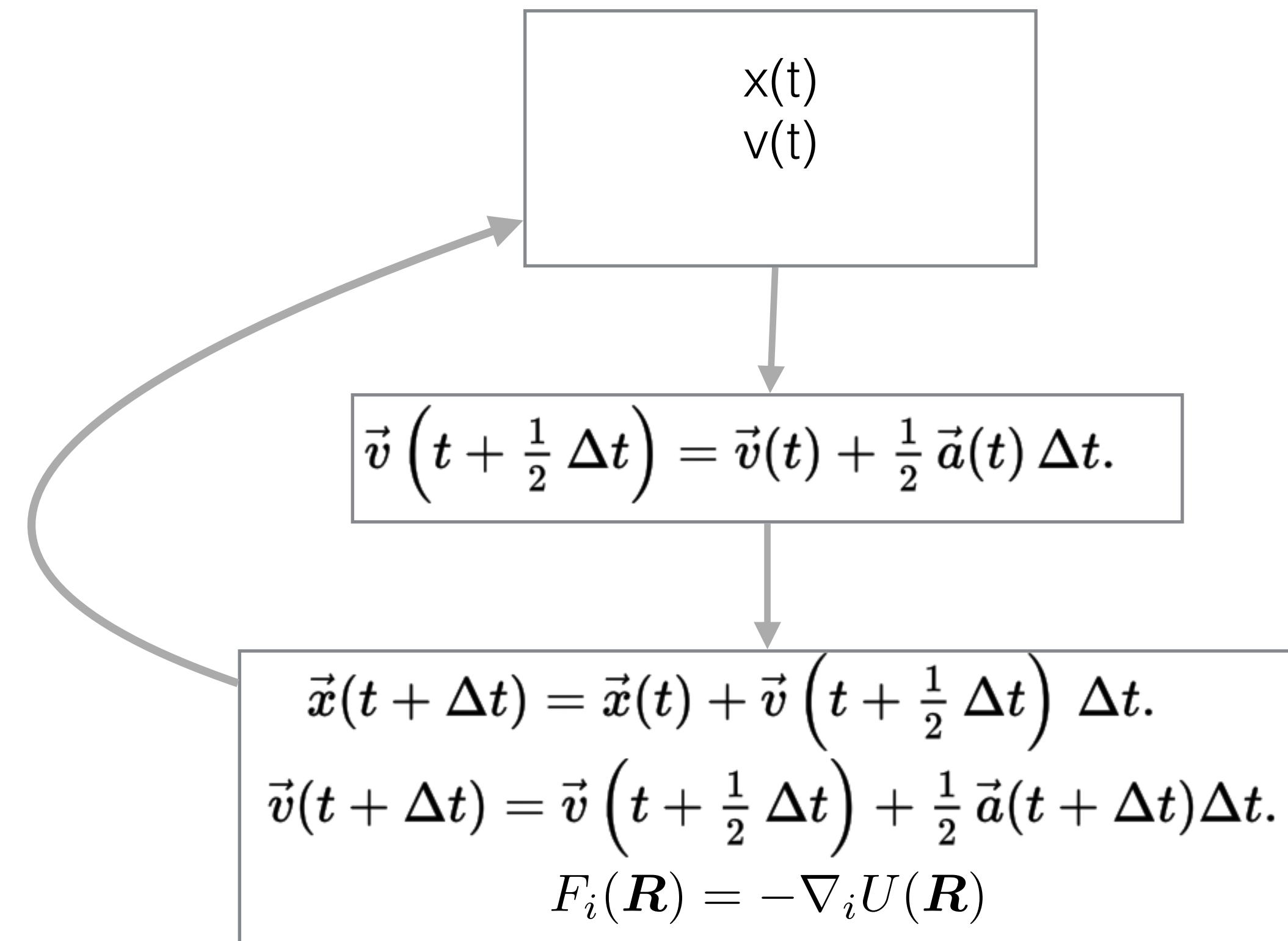
$$\vec{x}(t + \Delta t) = \vec{x}(t) + \vec{v}(t + \frac{1}{2} \Delta t) \Delta t.$$

The idea is to calculate more accurate velocities to be more precise in calculating positions

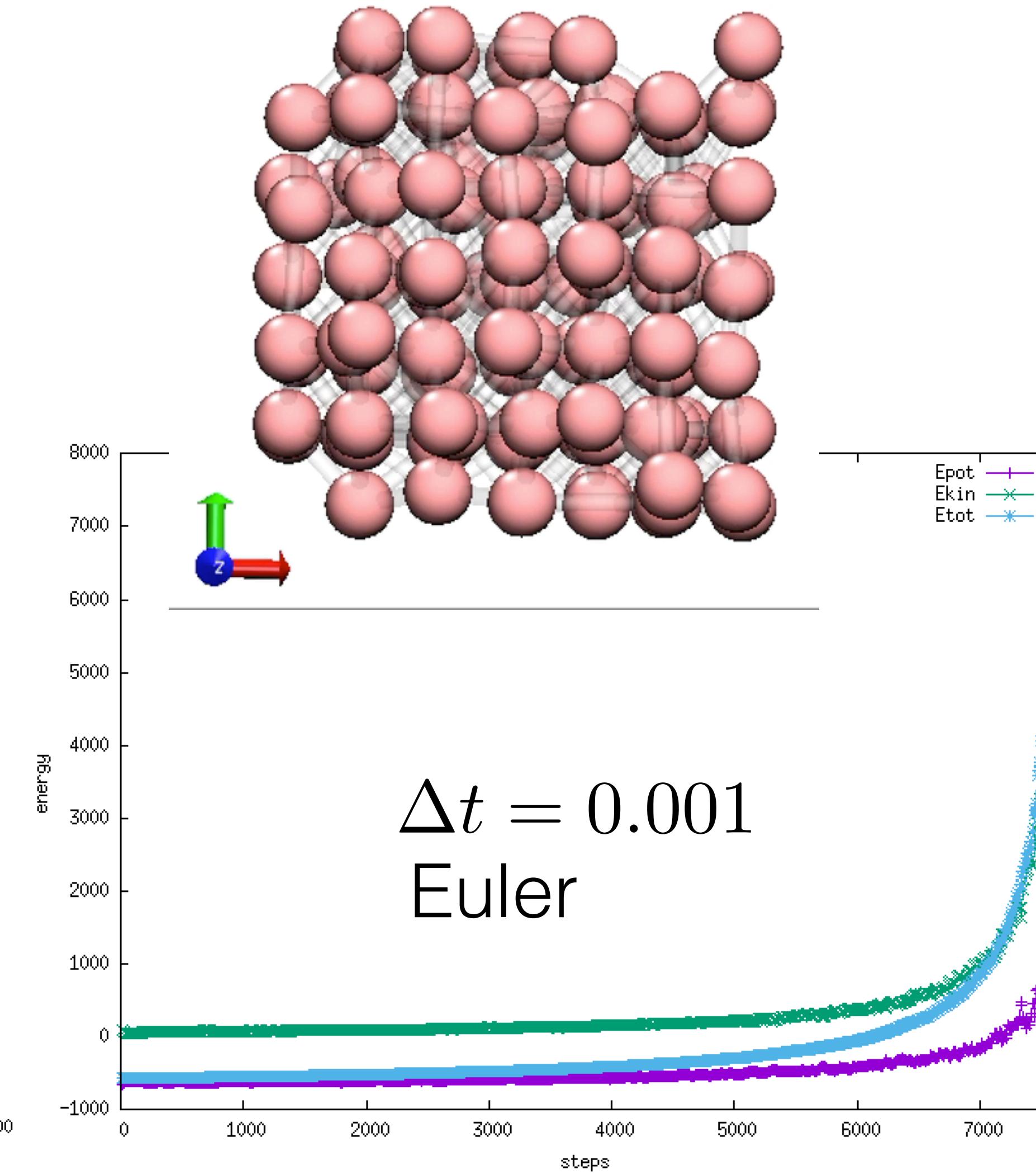
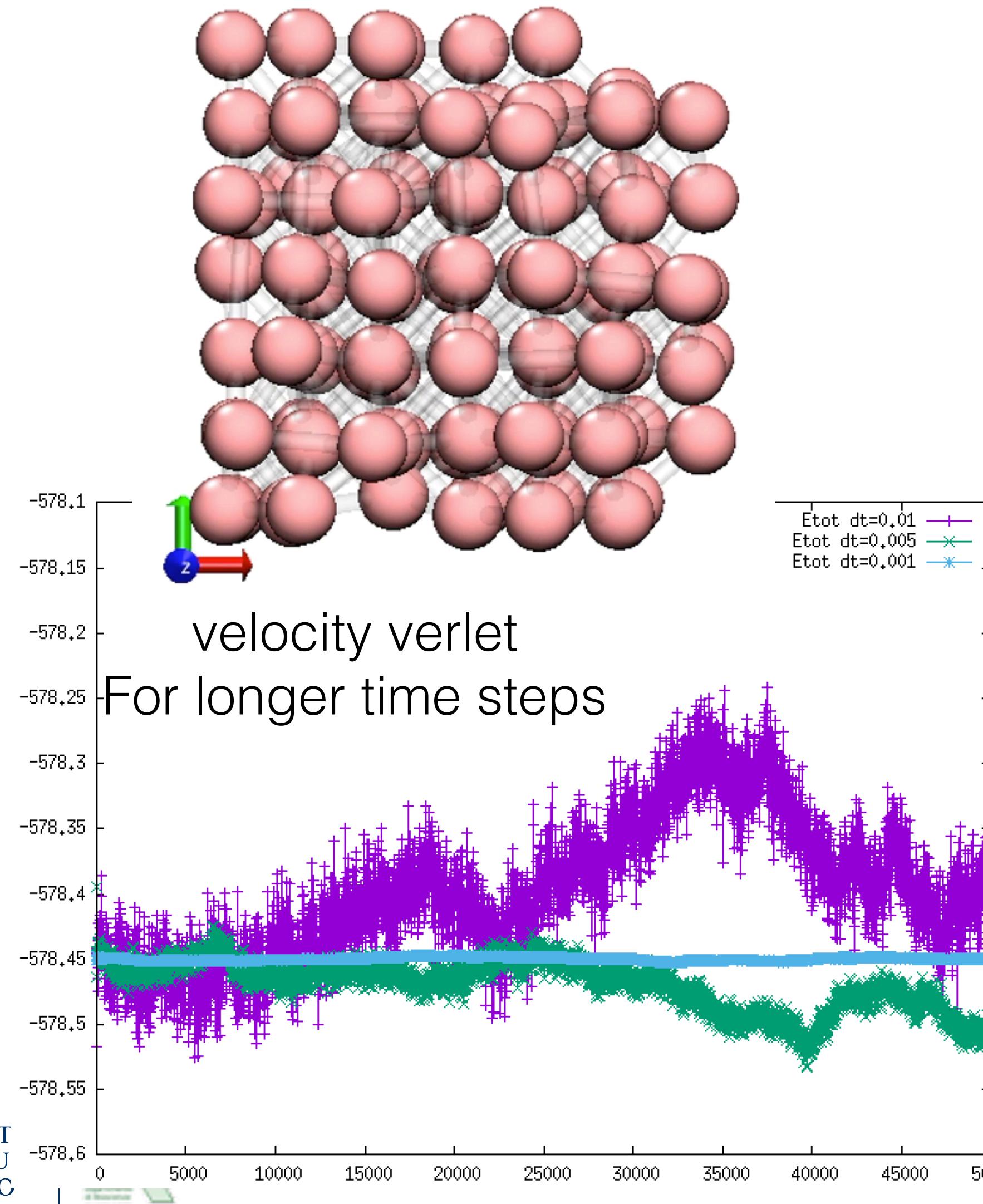
The algorithm

Start: $x(0)$, $v(0)$, $f(0)$

1. calculate velocities at time $t+0.5dt$: $v(t+0.5dt)$
2. calculate positions at time $t+dt$: $x(t+dt)$
3. calculate forces at time $t+dt$: $f(t+dt)$
4. calculate velocities at time $t+dt$: $v(t+dt)$



MD by Velocity Verlet can conserve the total energy



Velocity Verlet is a time-reversible integrator

$$\begin{aligned} r(t + \Delta t - \Delta t) &= r(t + \Delta t) - v(t + \Delta t)\Delta t + \frac{1}{2}a(t + \Delta t)\Delta t^2 = \\ &= r(t) + v(t)\Delta t + \frac{1}{2}a(t)\Delta t^2 - v(t)\Delta t + \\ &\quad - \frac{1}{2}a(t)\Delta t^2 - \frac{1}{2}a(t + \Delta t)\Delta t^2 + \frac{1}{2}a(t + \Delta t)\Delta t^2 = \\ &= r(t) \end{aligned}$$



MD: setting the temperature

If the goal of MD is to calculate an average quantity it does not matter if we calculate it at constant energy, or temperature, or other thermodynamic ensembles. In the thermodynamic limit average quantities do not depend on this choice. Fluctuation do, so if we are interested in probability distributions we need to chose the experimental conditions.

Equipartition Theorem gives us a definition for the temperature: If you set the temperature of a system its average kinetic energy should be equal:

$$\langle K \rangle = \frac{3}{2} N k T$$

We want an algorithm to obtain the correct average kinetic energy with an accuracy that is the standard error of the mean over our number of particles.



MD: the stochastic velocity rescaling algorithm

$$K_0 = \frac{3}{2} N_f k T,$$

$$K(t + \Delta t) = K(t) + (K_0 - K(t)) \frac{\Delta t}{\tau_T} + 2 \sqrt{\frac{K(t) K_0}{N_f}} \frac{dW}{\sqrt{\tau_T}},$$

$$v(t + \Delta t) = v(t + \Delta t) \sqrt{\frac{K(t + \Delta t)}{K(t)}}$$

By setting the temperature we know what should be on average the value of the kinetic energy of the system and this has to do with the system's velocities. So we modify all the velocities to keep them close to the desired target using an Ornstein-Uhlenbeck process: a dynamic process whose equilibrium distribution is a gaussian.

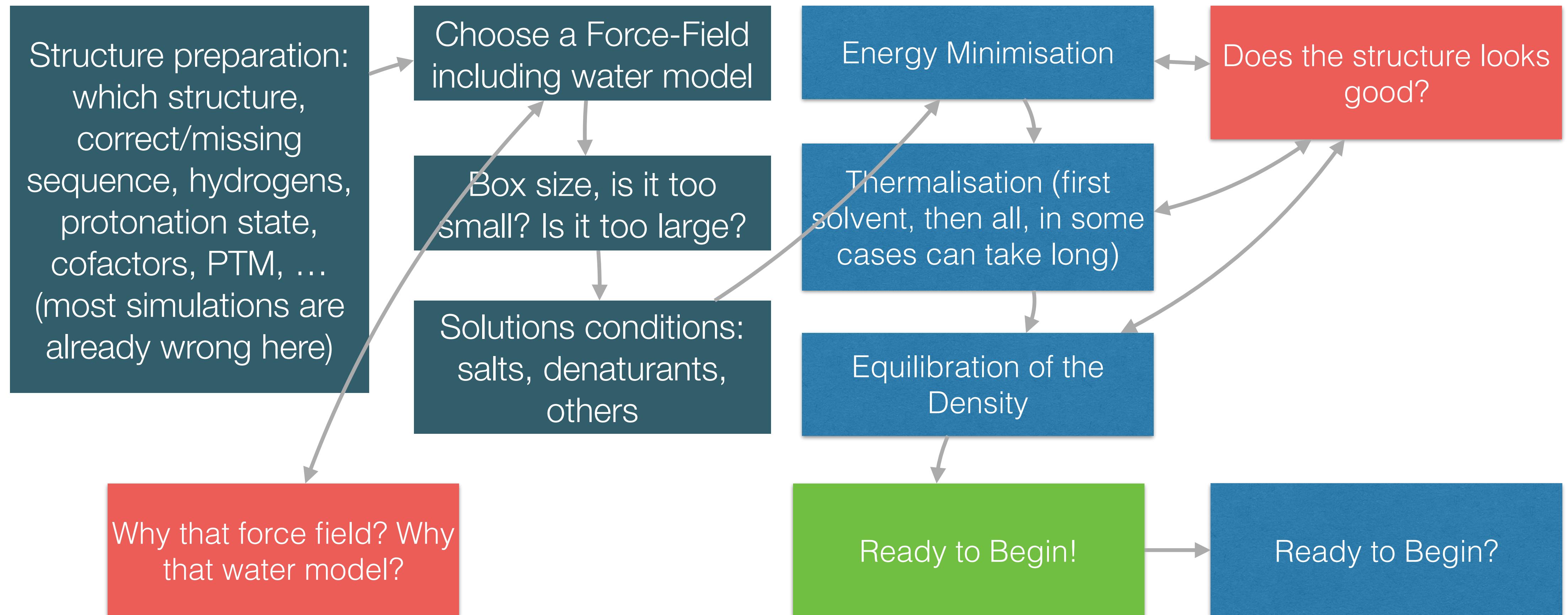
At step 0 compute forces **{x(0), v(0), f(0)}**

1. update kinetic energy
2. update velocities
3. update positions
4. calculate forces
5. update velocities
6. update kinetic energy

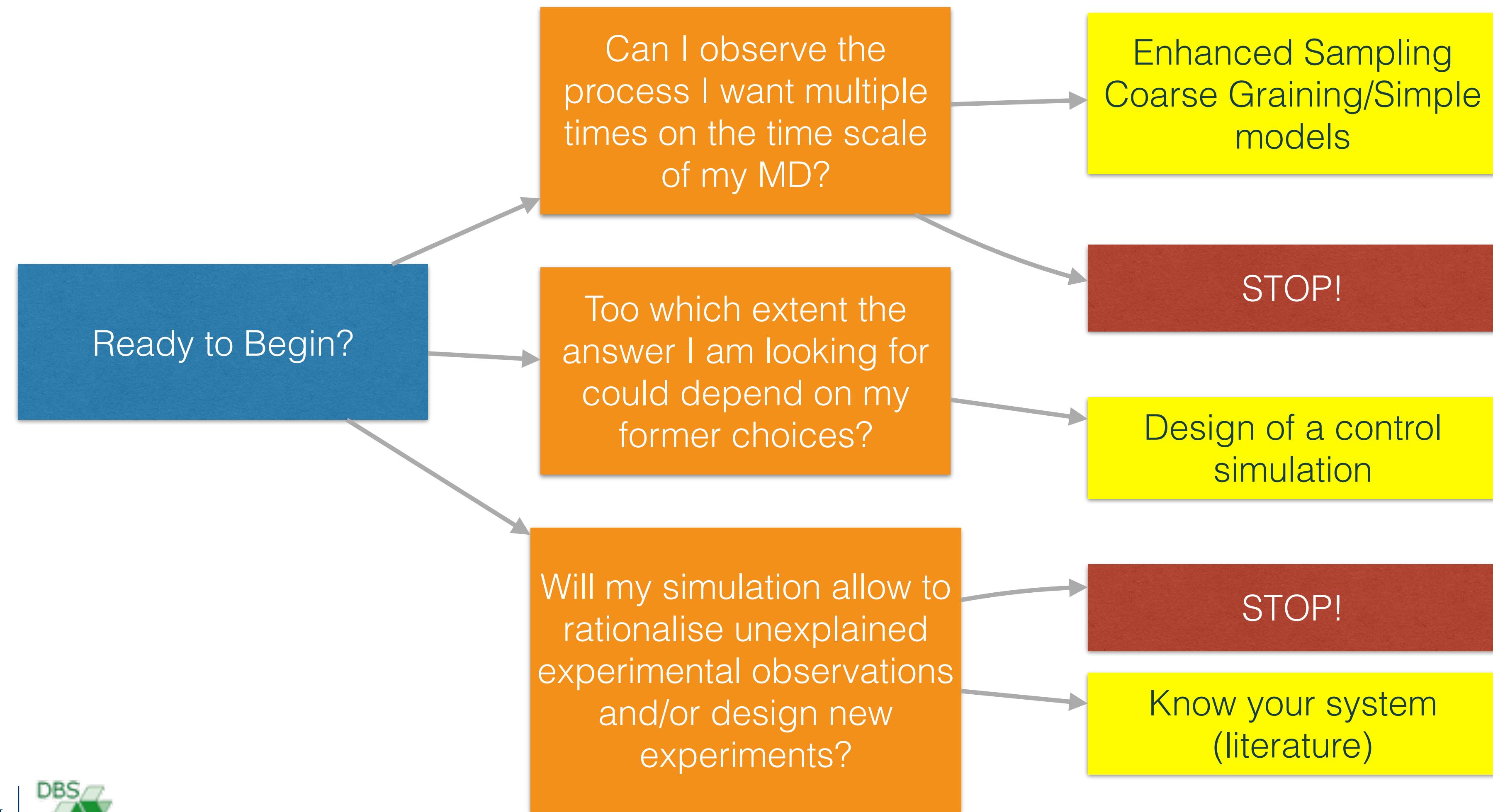
A similar approach can be used to also set the pressure



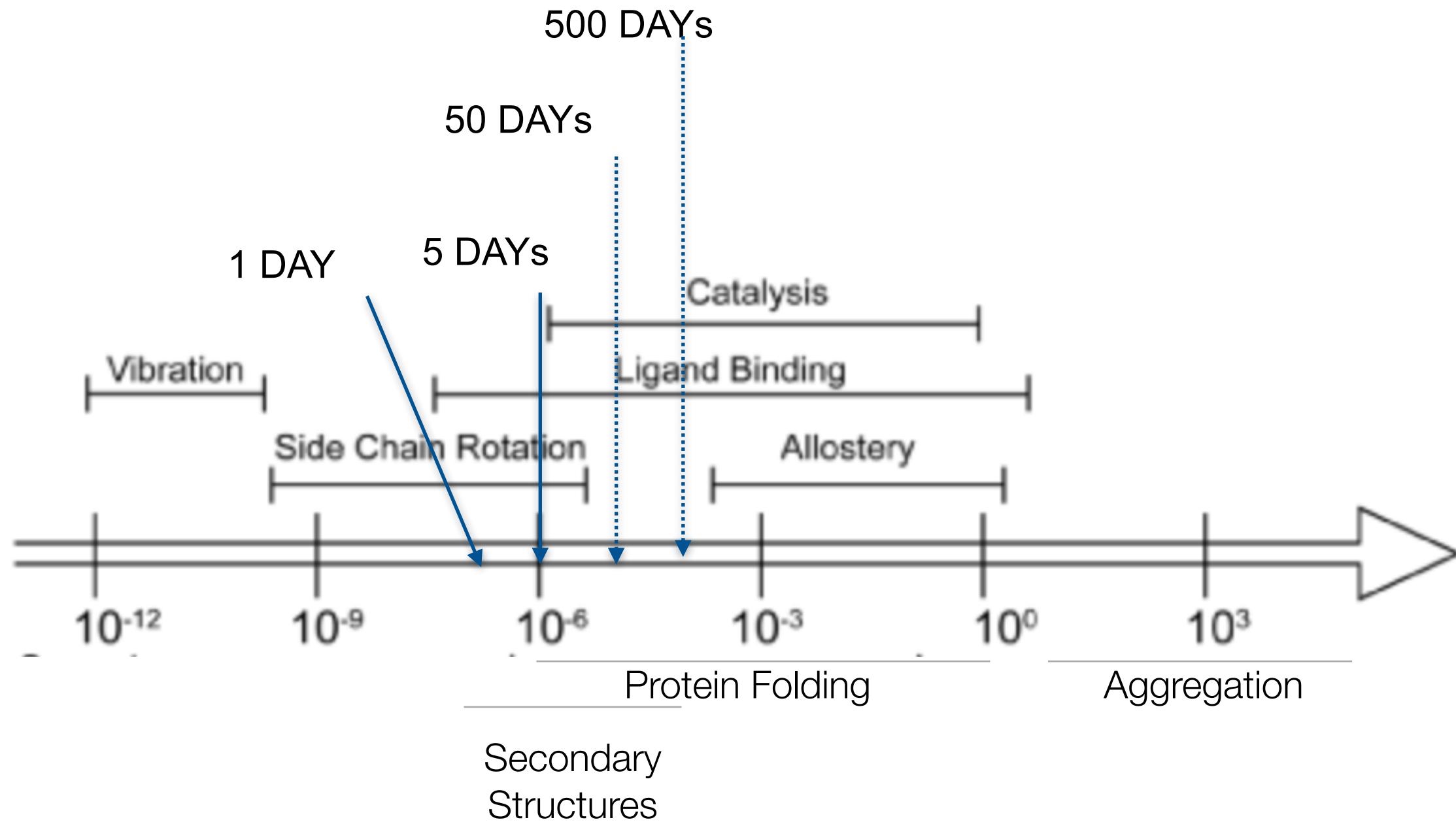
A general scheme to setup MD simulations:



A general scheme to setup MD simulations:



MD simulations: time step, time scales and probabilities



time-step $\sim 10^{-15}$ s

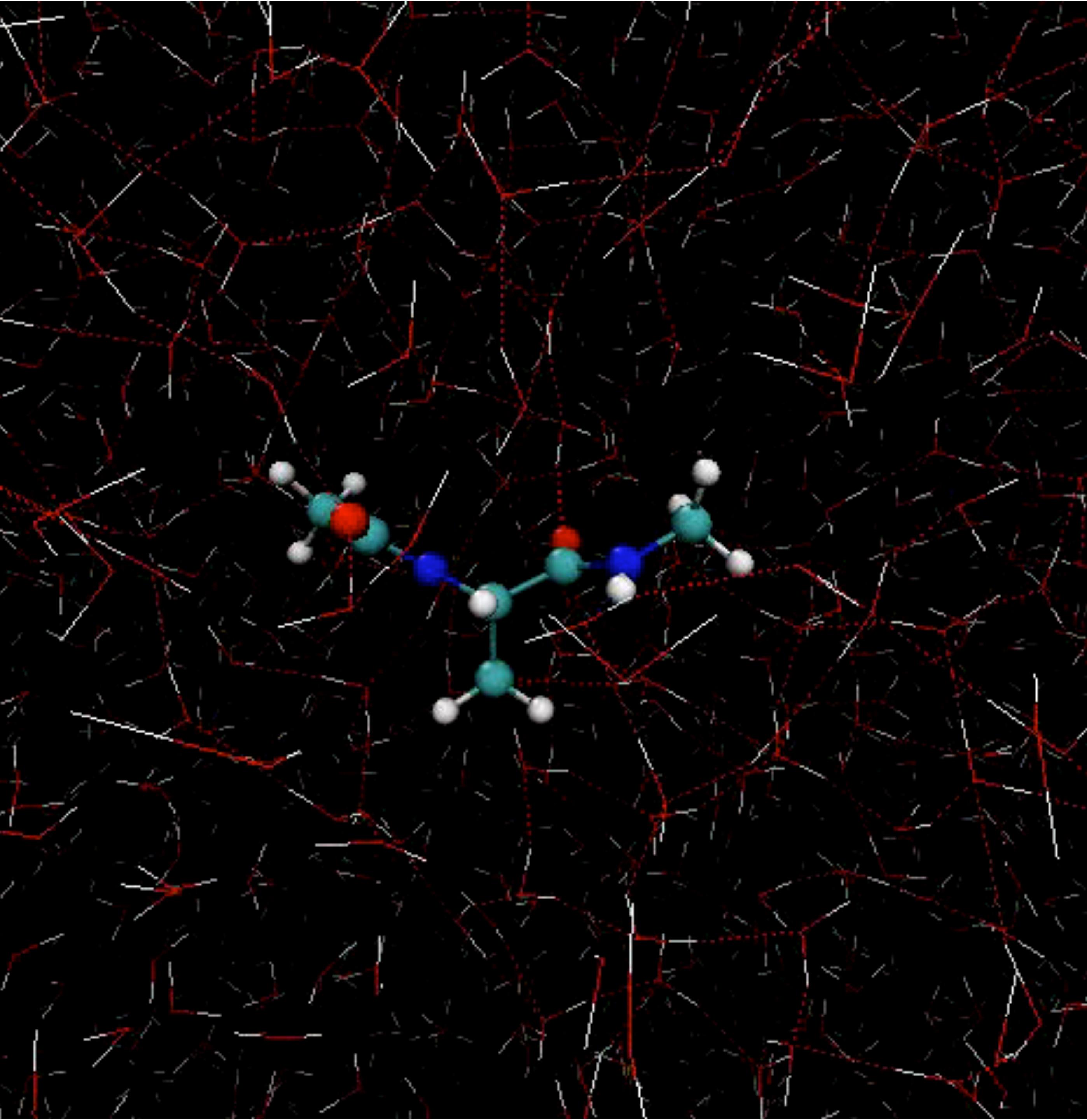
an MD simulation for a 500 AA protein can run at ~ 200 ns/day

the probability of observing an event with a rate of 1 ms with a simulation of 10 us is $\sim 1\%$ in the case of a two state kinetics

How many simulations can we run in parallel?

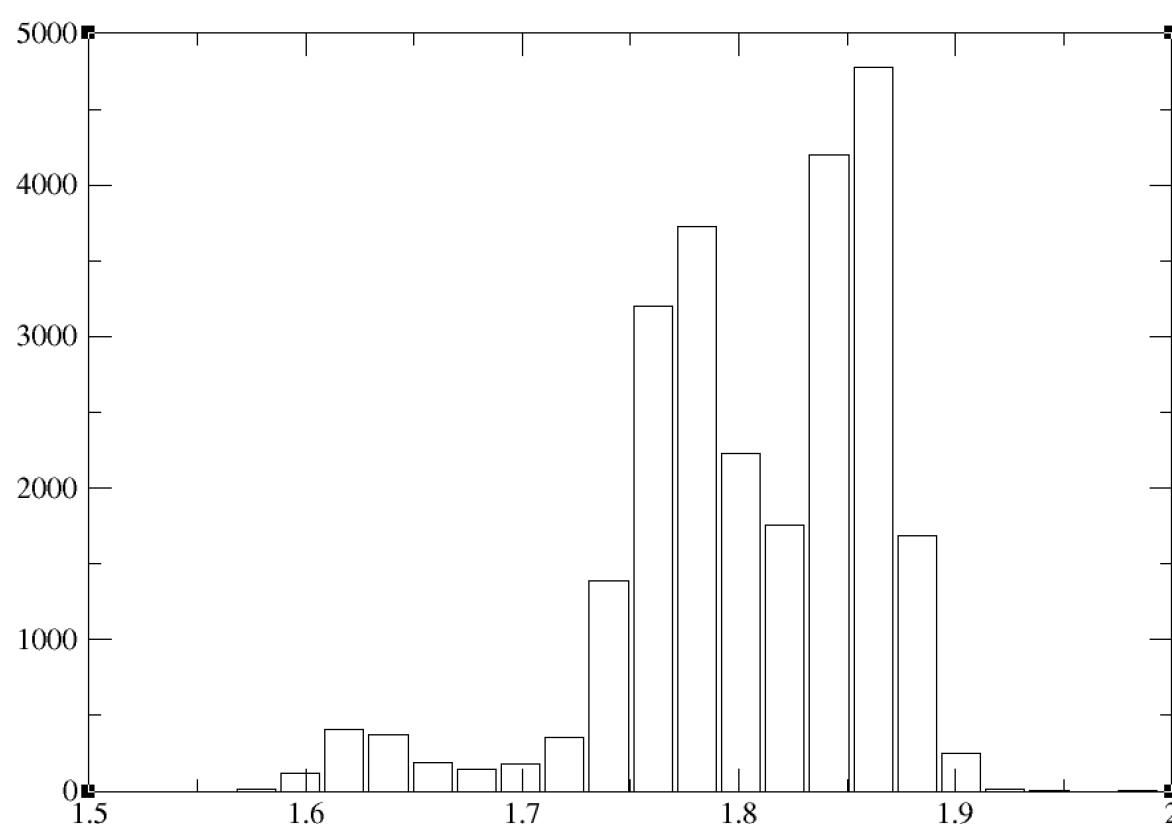
Ideally, what should a computational microscope do?

- Observe the time evolution of molecules at high spatial and time resolution ✓
- Observe them for very long time scales ⚠ (Time scale depends on size, improving...)
- Be aware of their energy ✓
- Be able to set different experimental conditions (Temperature, Pressure, solution conditions) ✓
- Be accurate ⚠ (Force-field dependence, but improving...)
- Be interpretable, that is find suitable macrostates to compare with experiments ⚠ (see later)
- ...

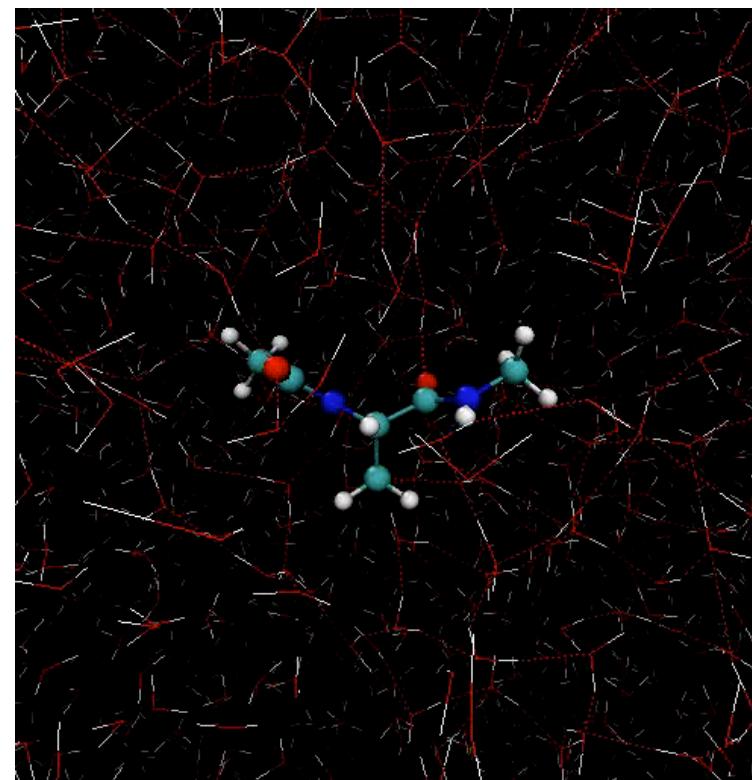


Analysis and Interpretability of MD simulations

A key difference between a simulation and an experiment is that when you do an experiment you have already selected which property of the system to observe, in a simulation instead you set up the conditions of the simulation but then you need to work on how/what to look for.



This is the histogram of the x coordinate of the Ca carbon of the alanine



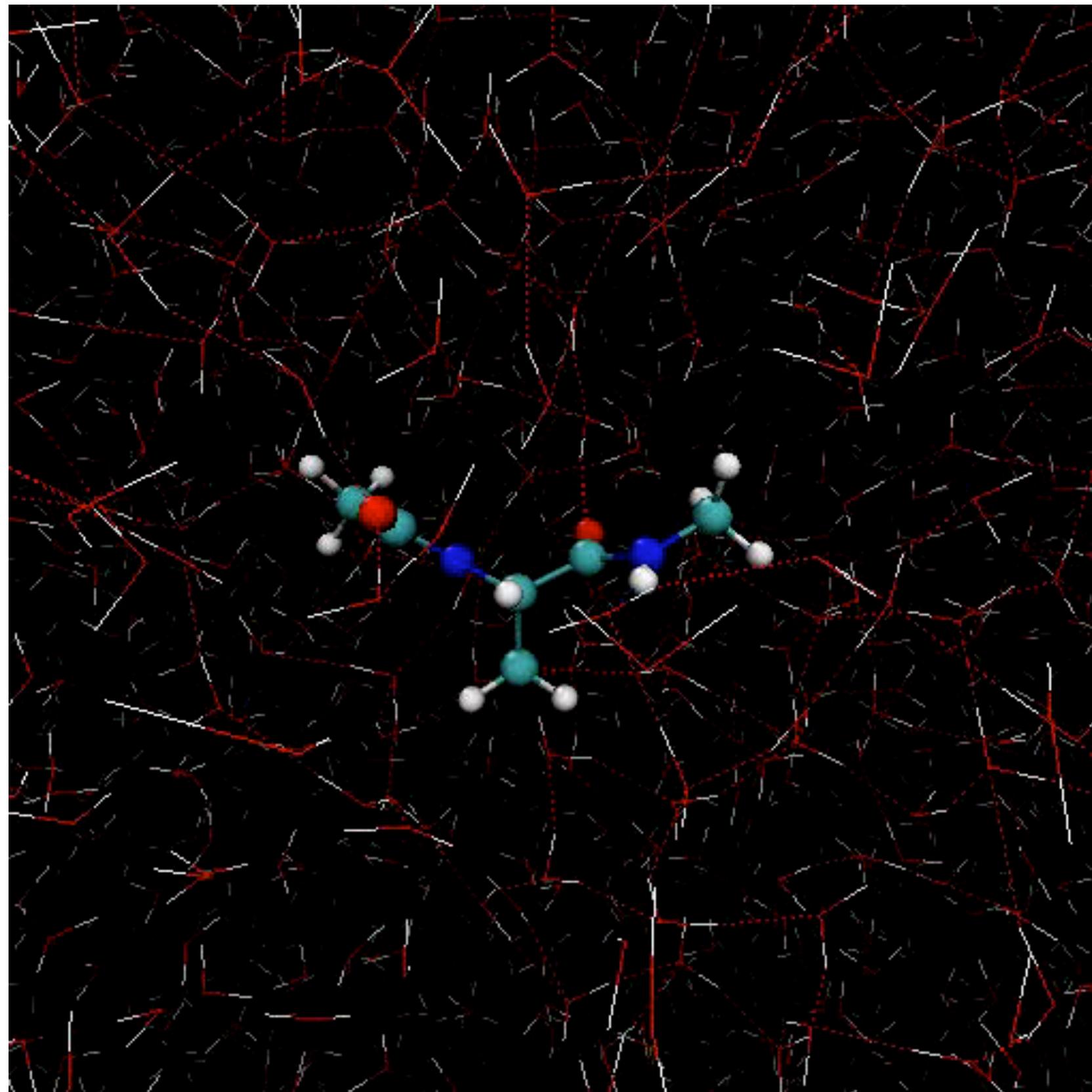
In principle we can look at any atomic quantity, but they are not necessarily interesting, for example the position of an atom in the box...

A very important feature of MD simulations once you decide what to look for is that configurations evolve following the energy gradient and this means that they already appear following Boltzmann statistics.

And a key point is that we have the TIME EVOLUTION of the quantities we look like in a single-molecule experiment.



Analysis and Interpretability of MD simulations



Then, given that we have positions, velocities and forces for all the atoms this means that we can analyse any property that can be defined using these quantities:

Geometric quantities like: distances, dihedral angles, H-bonds, Volume, Density, Secondary Structures, RMSD root mean square deviation of the positions with respect to a reference configuration, RMSF root mean square fluctuations of the positions with respect to a reference configuration, Gyration radius, the radius of the sphere including the system atoms, ...

Force, Velocity related quantities like: Energy, Pressure, Diffusion, ...

Experimental Observables like: NMR quantities as J-Coupling, NOE and chemical shifts can be calculated using approximate functions, SAXS/SANS, FRET, CD, ...



Analysis and Interpretability of MD simulations

Choosing the quantities to analyse get into two common statistical approaches:

- 1) **clustering techniques**; and
- 2) **dimensionality reduction techniques**

Clustering techniques are techniques whose goal is to classify data in sets with a label, dimensionality reduction techniques are instead techniques that looks for optimal projections of highly dimensional data into a limited number of dimensions. The two approaches can also be combined together.

If we take as an example the alanine dipeptide simulation in water, there we have 22 atoms belonging to aladp and 1554 atoms of water (518 water molecule), for a total of 1576 atoms and 4728 coordinates. This means that the problem can be consider as a problem in 4728 spatial dimensions.



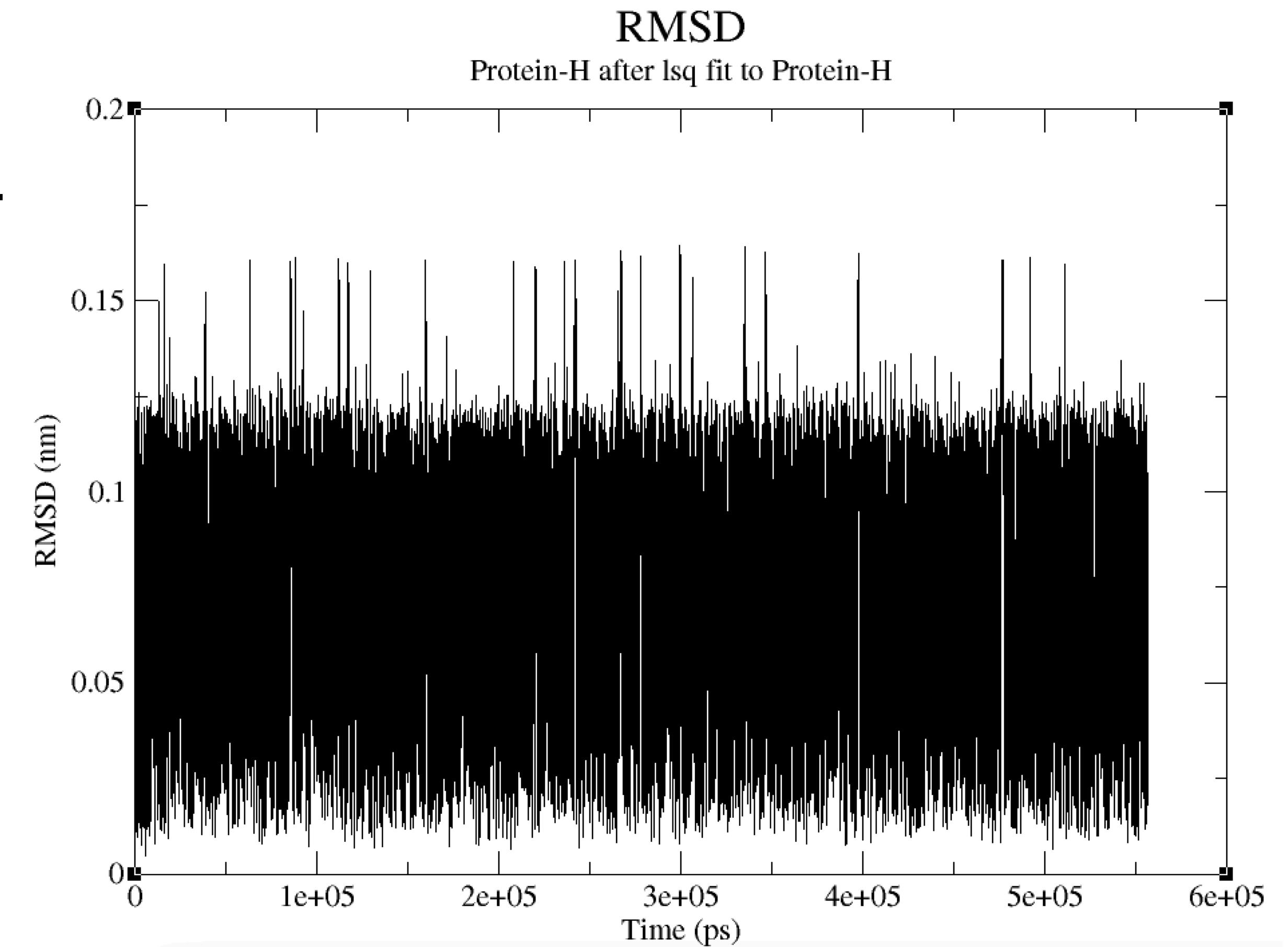
A simple case: Alanine Dipeptide

To do an analysis we should in general first have done a simulation to answer some questions, this should already guide us in the analysis to perform.

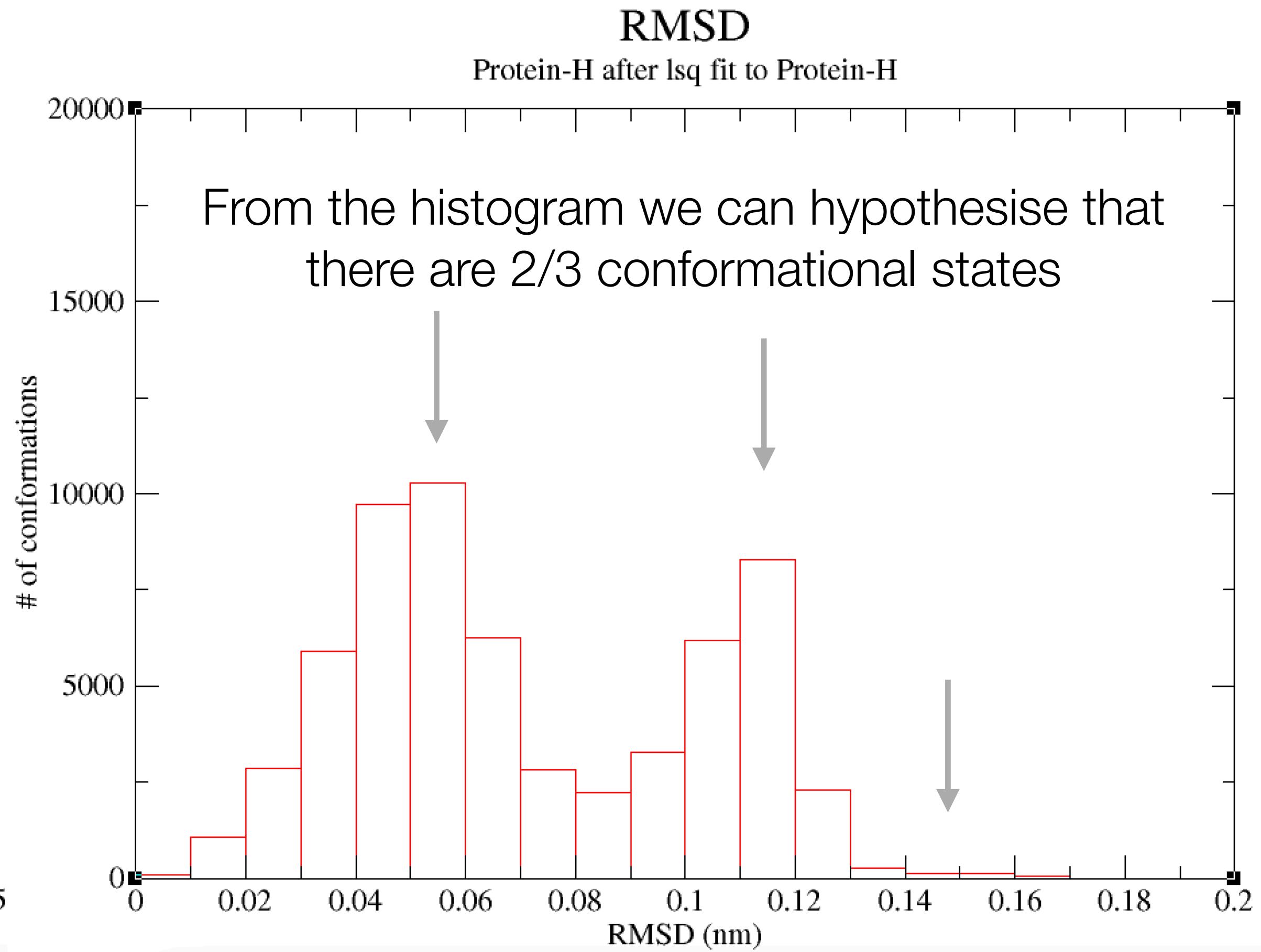
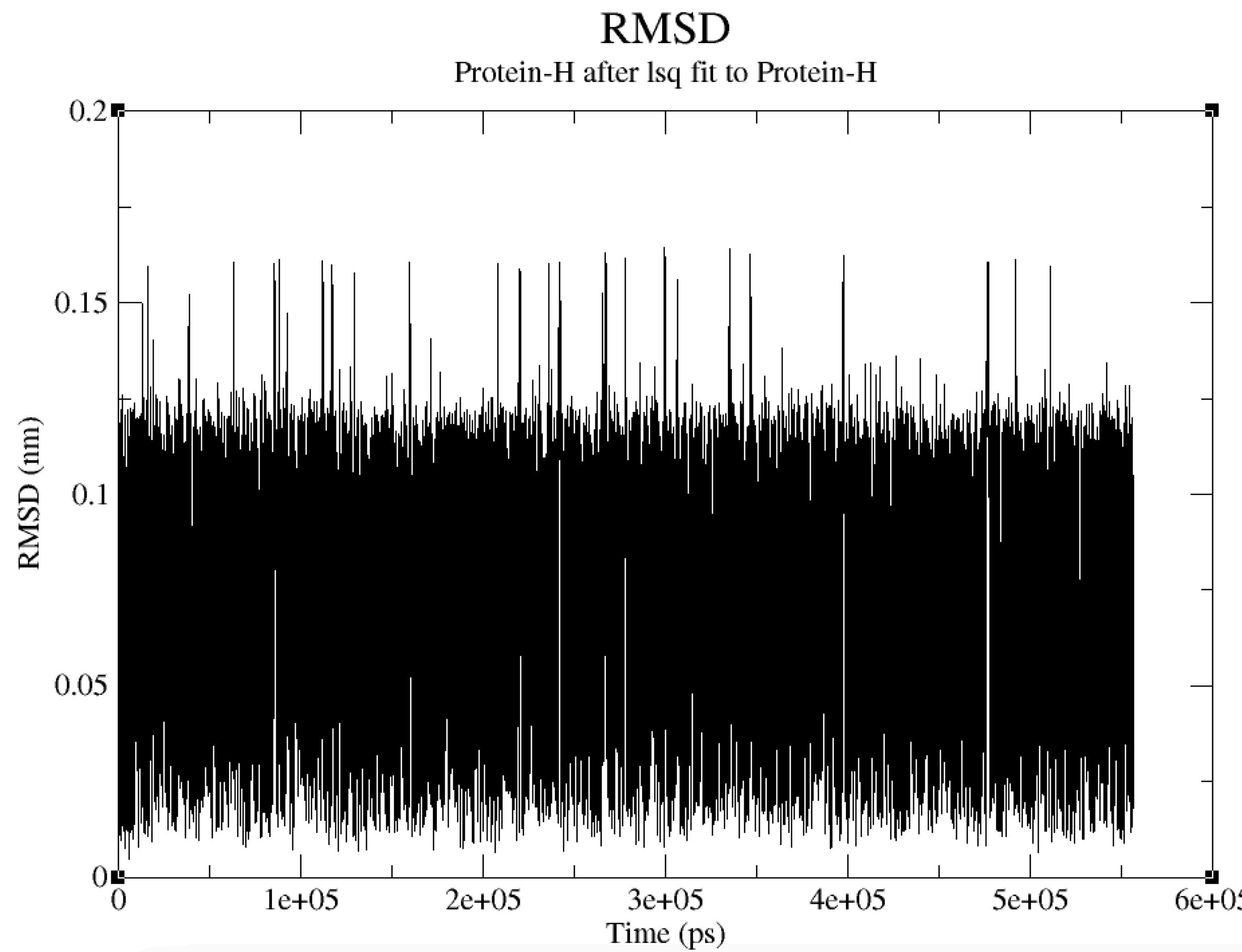
Let's say that we want to learn about the conformational freedom of alanine dipeptide.

We may start looking at the **RMSD** of the molecule in time. The RMSD measures how much each conformation in time differ from a reference configuration (for example the first frame of the trajectory) by comparing the difference in the atomic position after optimally superimposing the two configurations.

https://en.wikipedia.org/wiki/Root-mean-square_deviation_of_atomic_positions



A simple case: Alanine Dipeptide



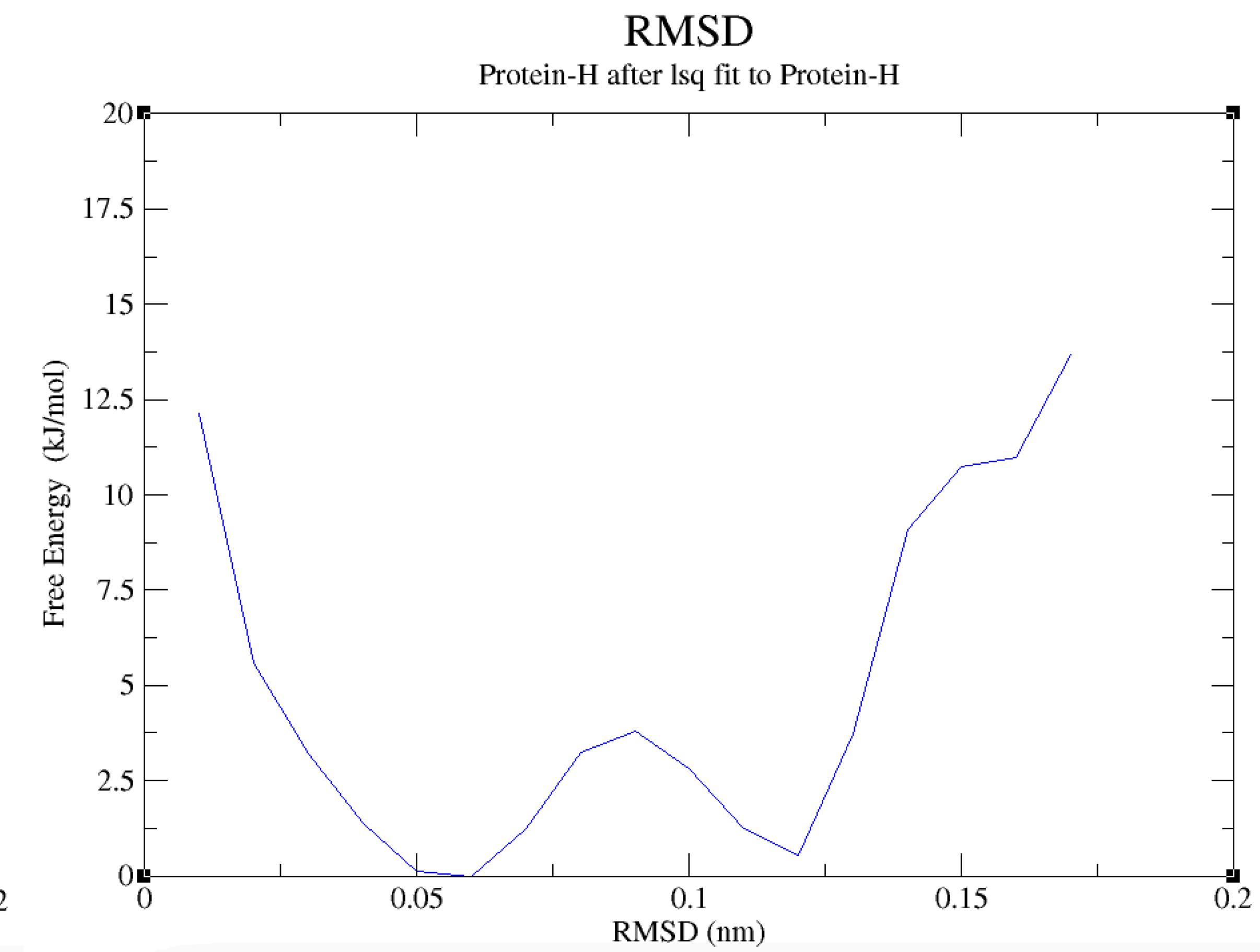
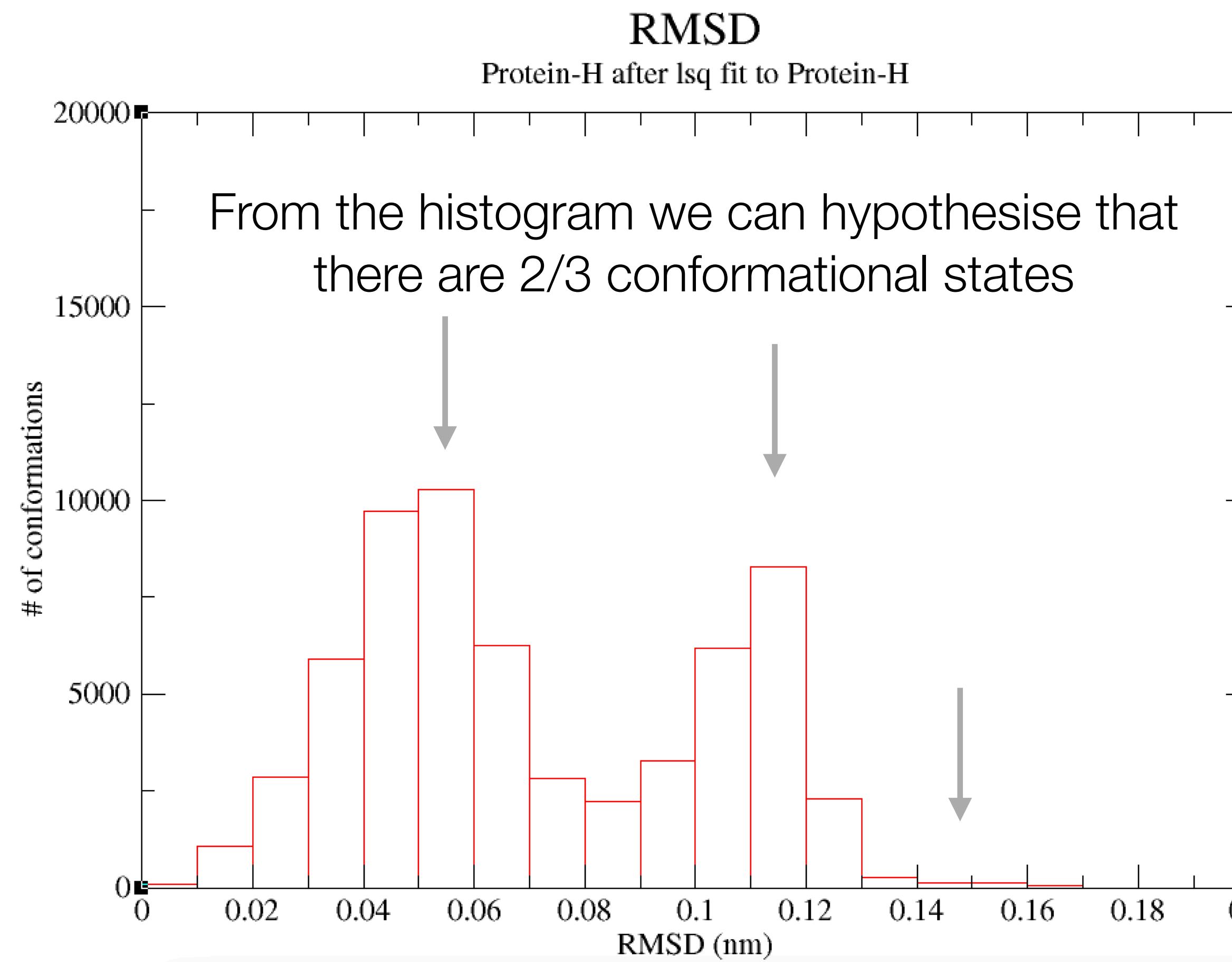
From our time-resolved data we can make an histogram counting each conformation as 1 because in MD conformations appear following Boltzmann probability.



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A simple case: Alanine Dipeptide

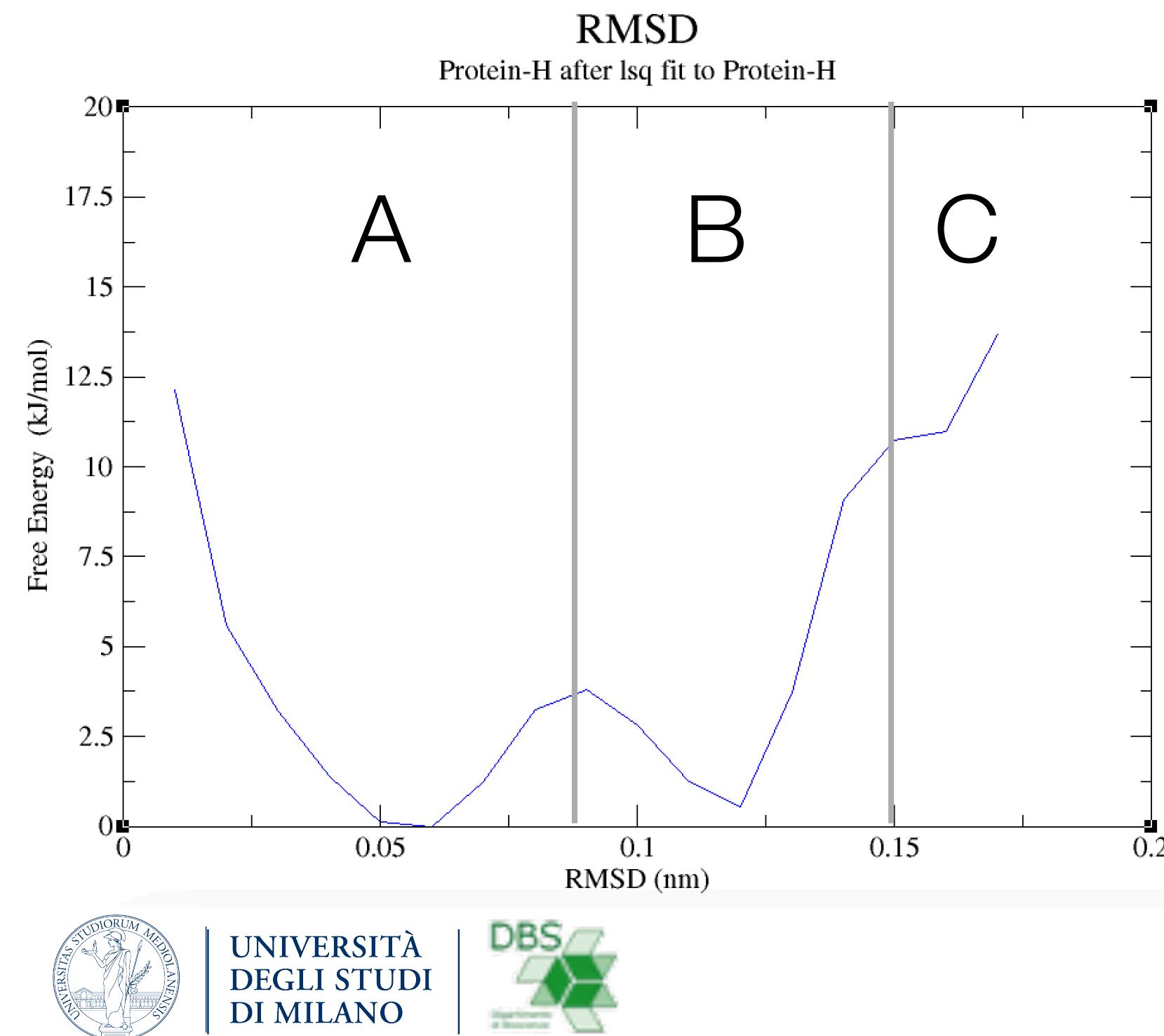


From the histogram we can estimate the Free Energy profile along the RMSD variable by taking $F(\text{rmsd}) = -k\text{bT} \ln(\text{count}(\text{rmsd}))$



AlanineDP: From dimensionality reduction to clustering

What we have done is a dimensionality reduction of the conformational space of our simulation from the many dimension of the coordinate to a single dimension, that is also collective variable. This may or may not be important to describe the conformational dynamics of ALADP, but allows to identify 2/3 states. In principle we can assign to all conformations belonging to the three state a label and so clusterize our trajectory:



Usually we say that the resulting clusterisation is better if the structures belonging to the cluster are overall homogenous with respect to some property.

Furthermore, the number of configurations per state, divided by the total number of configuration will give us a state population:

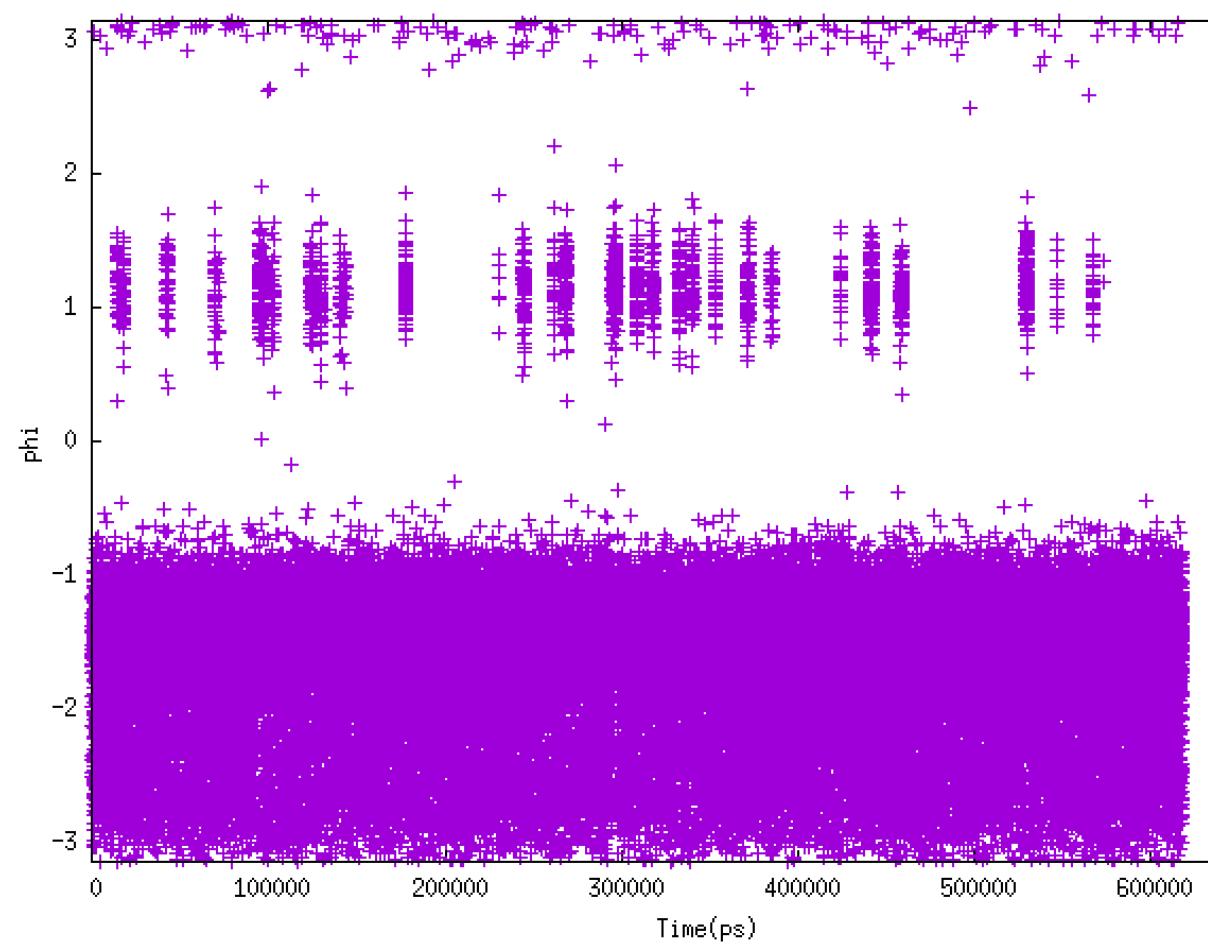
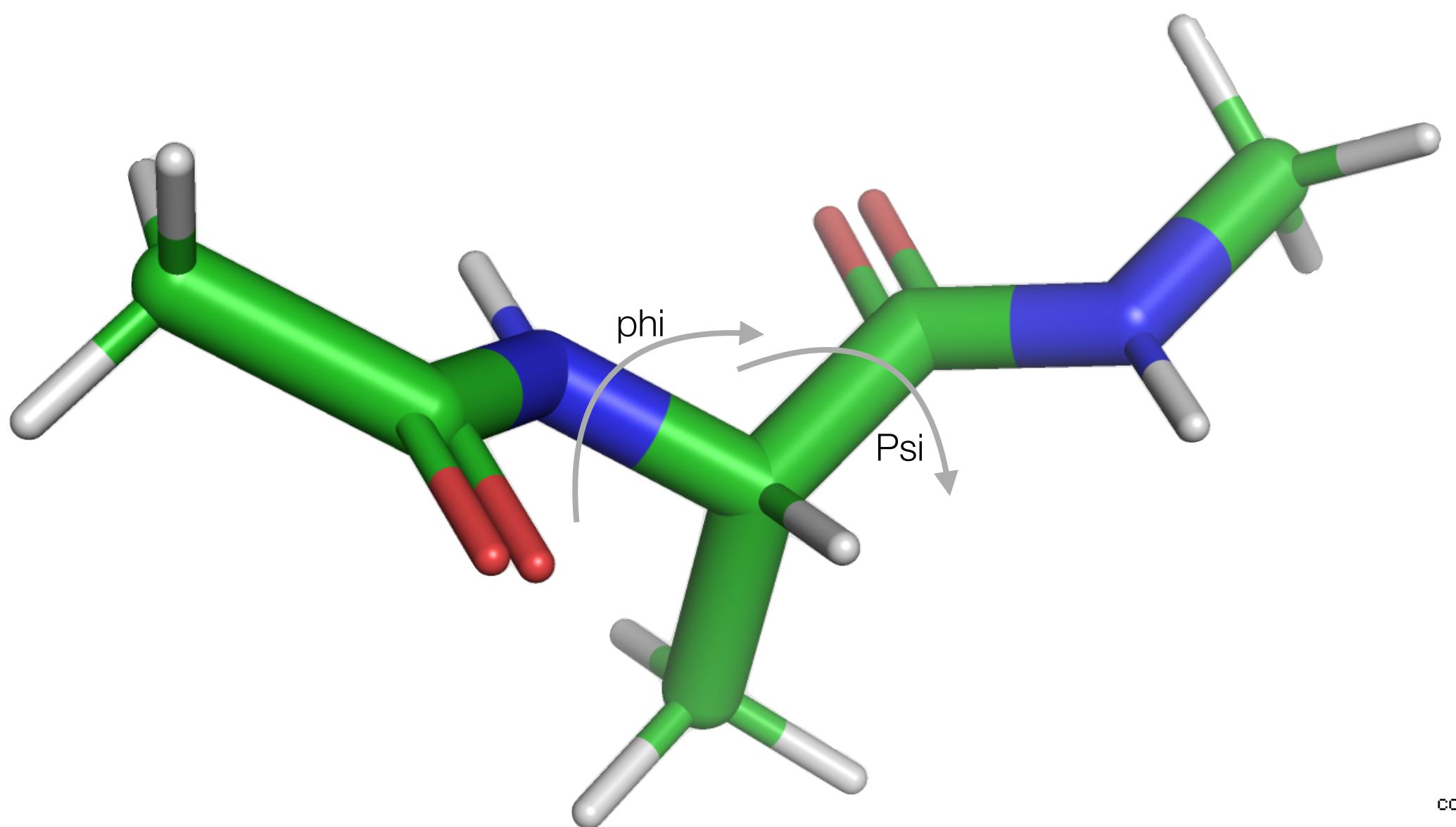
$$\begin{aligned} p_A &= \#A / (\#A + \#B + \#C) & F_A &= -k_B T \ln(p_A) \\ p_B &= \#B / (\#A + \#B + \#C) & F_B &= -k_B T \ln(p_B) \\ p_C &= \#C / (\#A + \#B + \#C) & F_C &= -k_B T \ln(p_C) \end{aligned}$$

$$\Delta F_{AB} = F_A - F_B = -k_B T \ln \left(\frac{p_A}{p_B} \right)$$

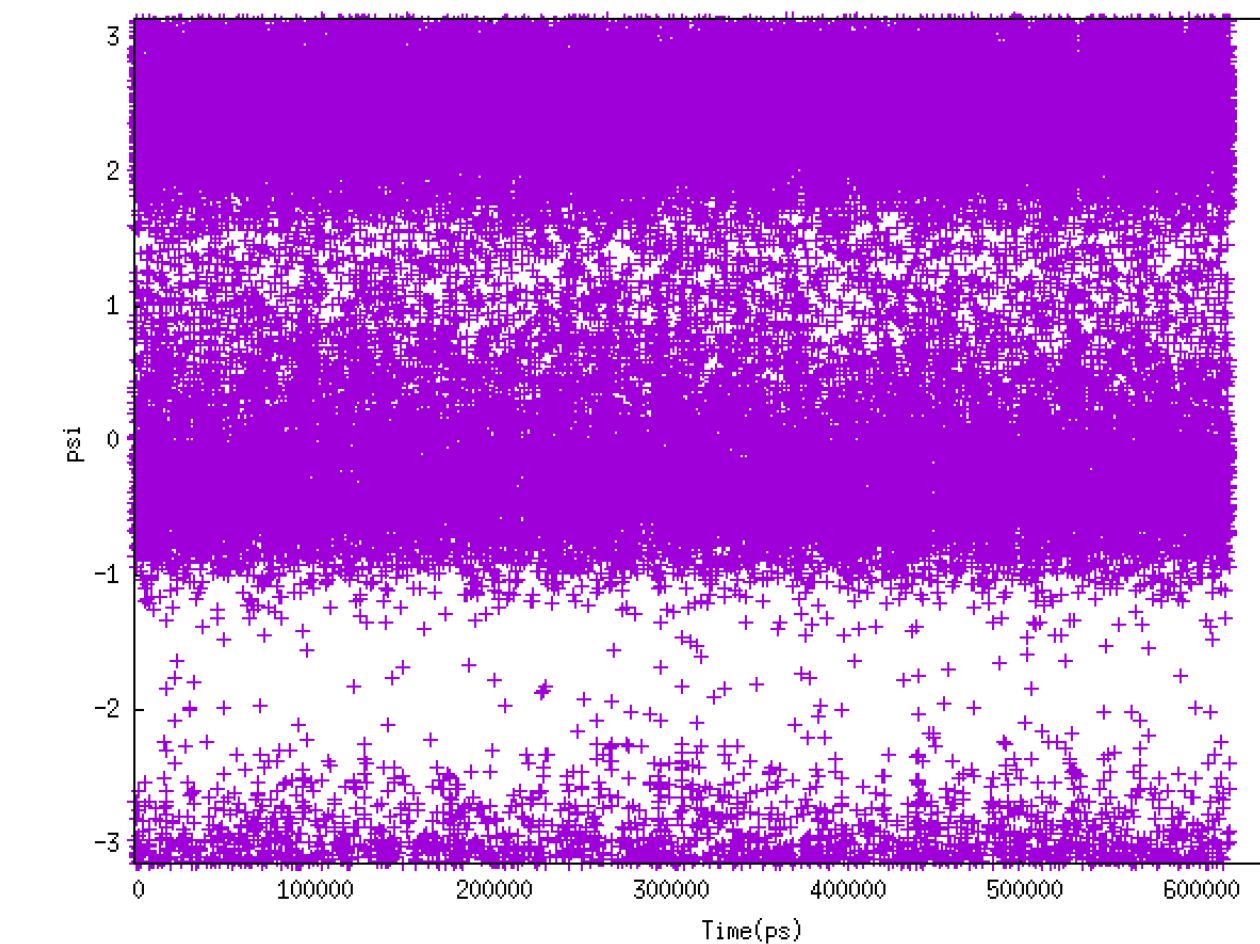
...

Another point of view:

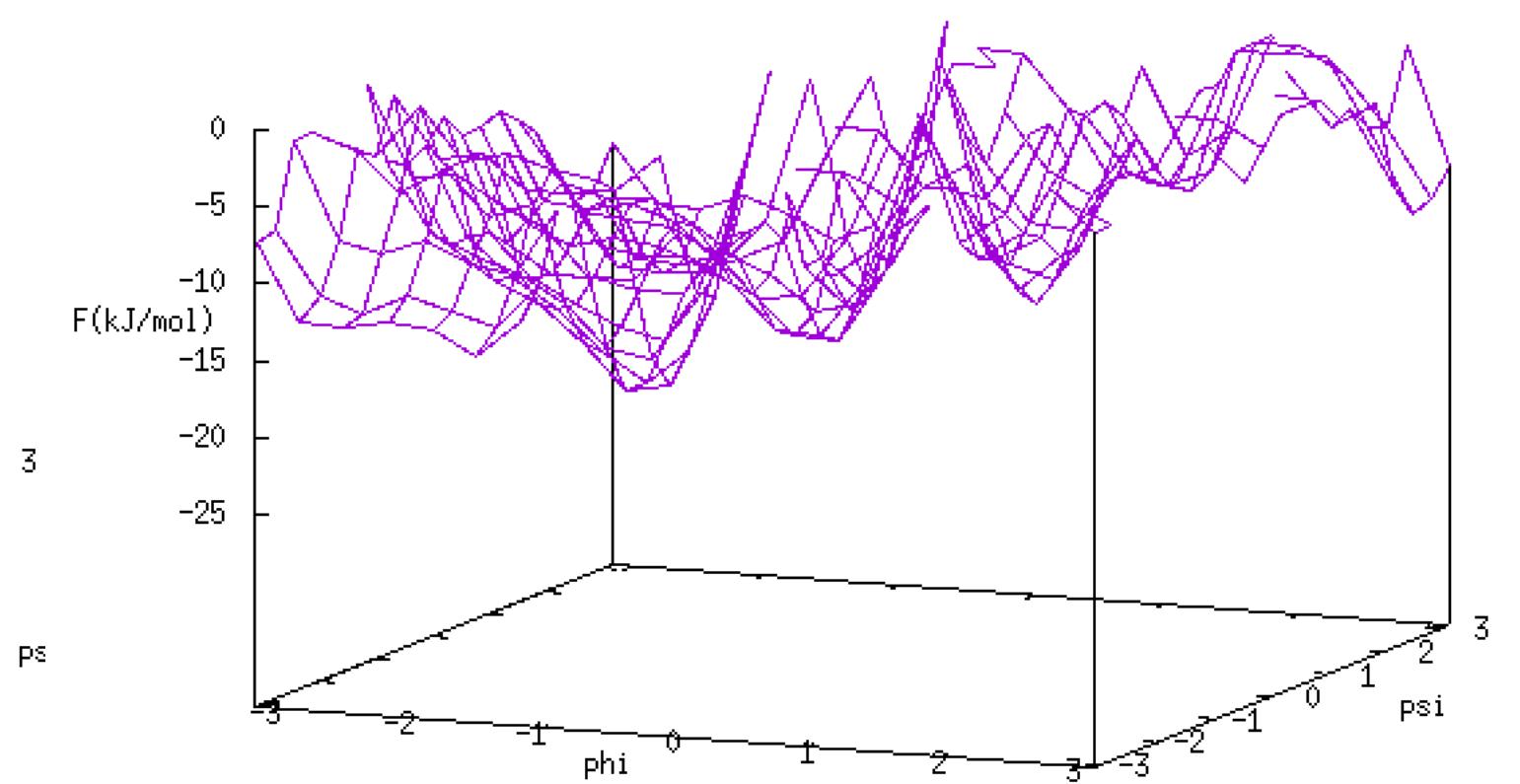
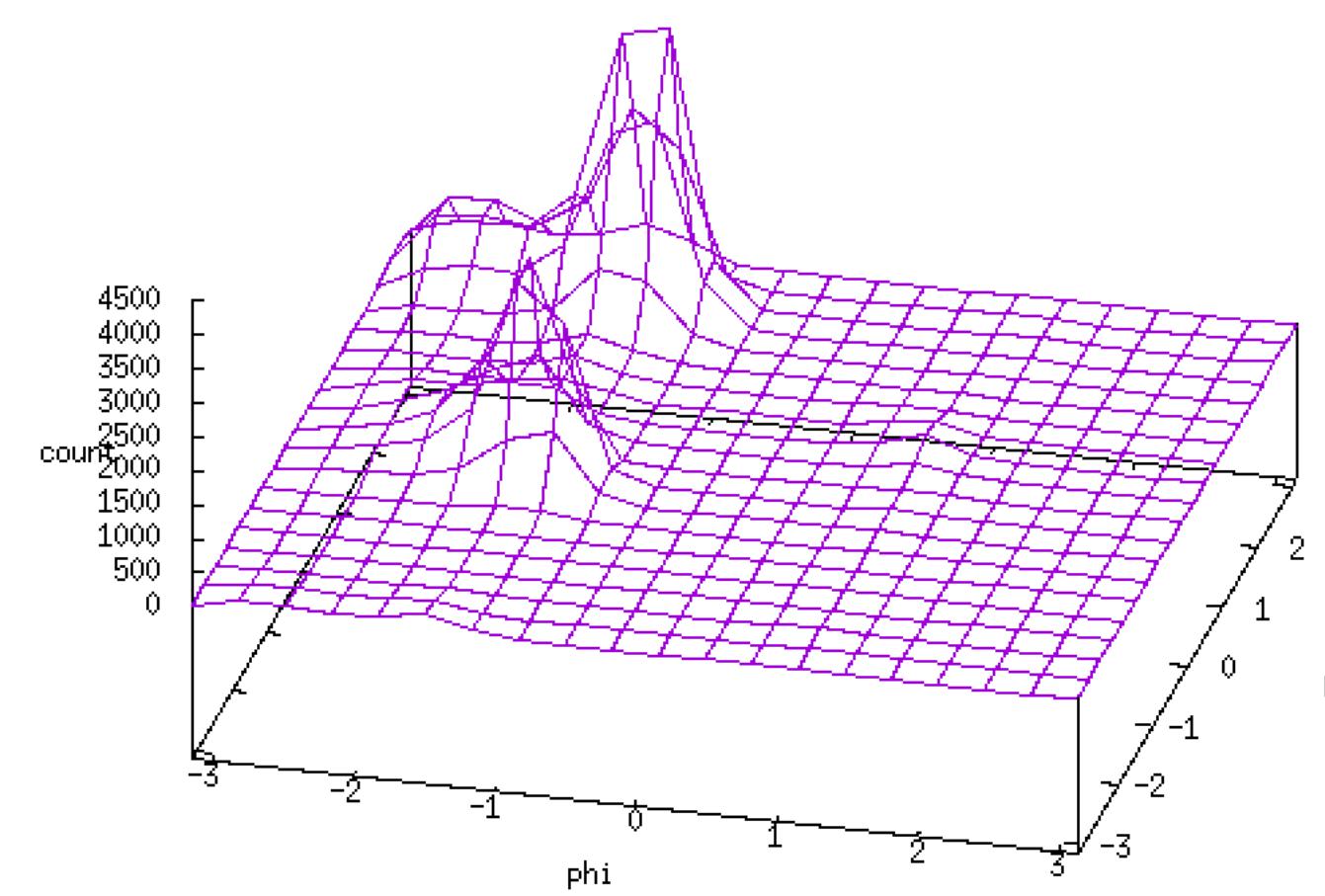
Going on looking and learning about your system you may decide to analyse it in terms of backbone dihedral angles



Histogram in 2D

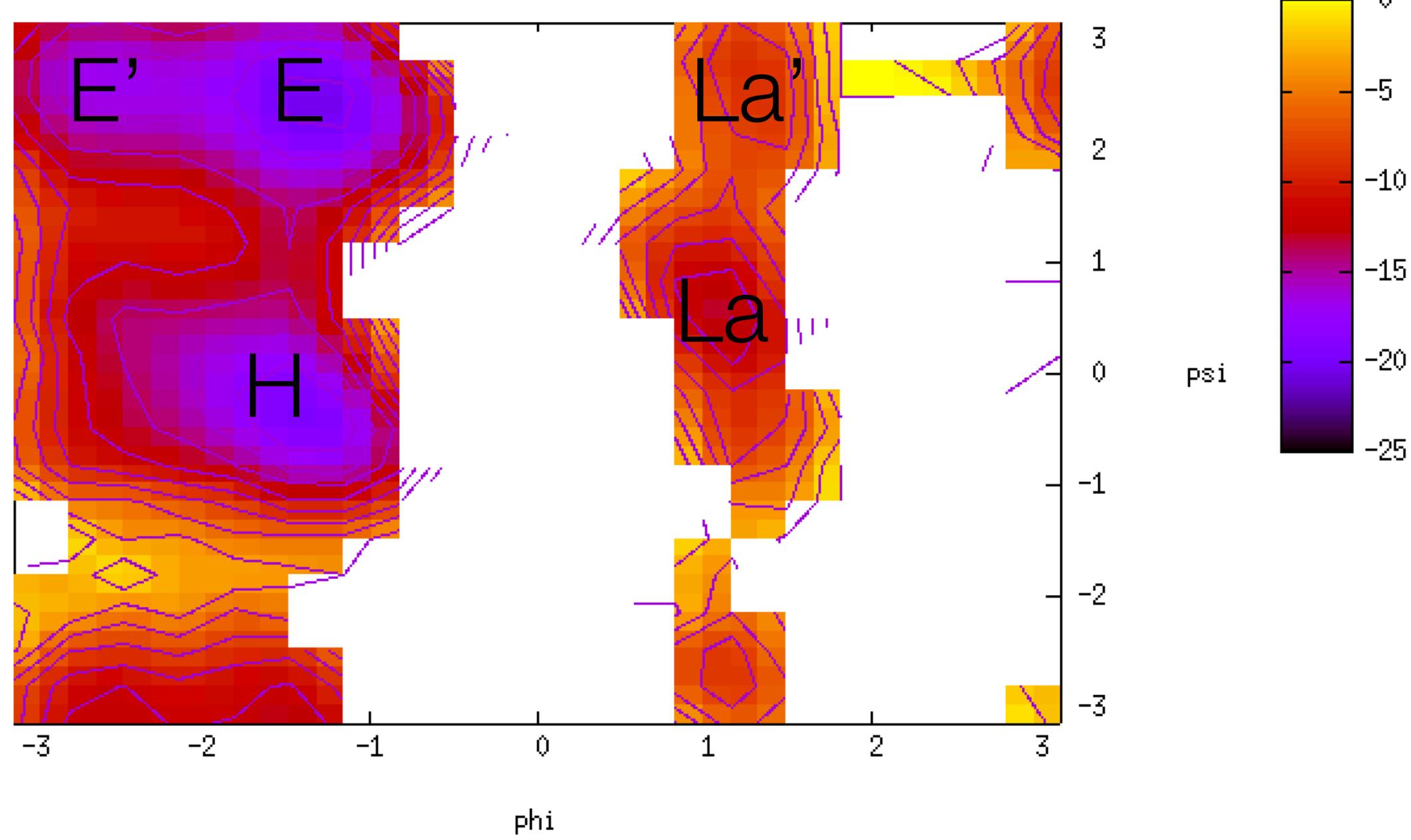


Free Energy Surface



Another point of view:

This is again the free energy as a contour plot, we may identify 3/5 states.



Dimensionality reduction -> Clustering,
I am using different labels because they may
or may not correspond to the formers.

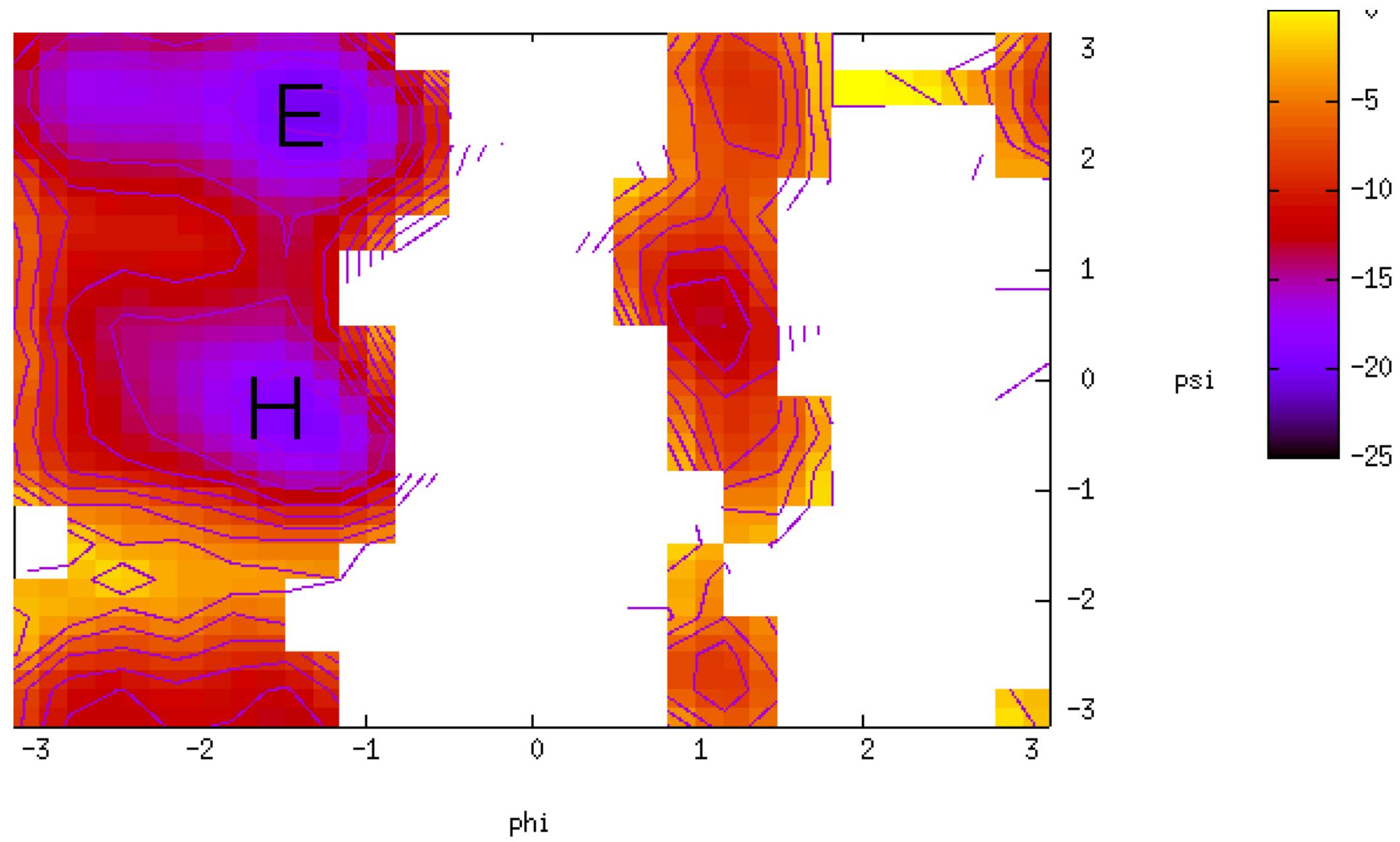
A further analysis of the conformations assigned to the different states may allow to rationalise the conformational space of alanine dipeptide in the way that we call Ramachandran Plot and its connection with the different secondary structures

You may already guess that the more complex the system you work with the more difficult/arbitrary become this analysis.

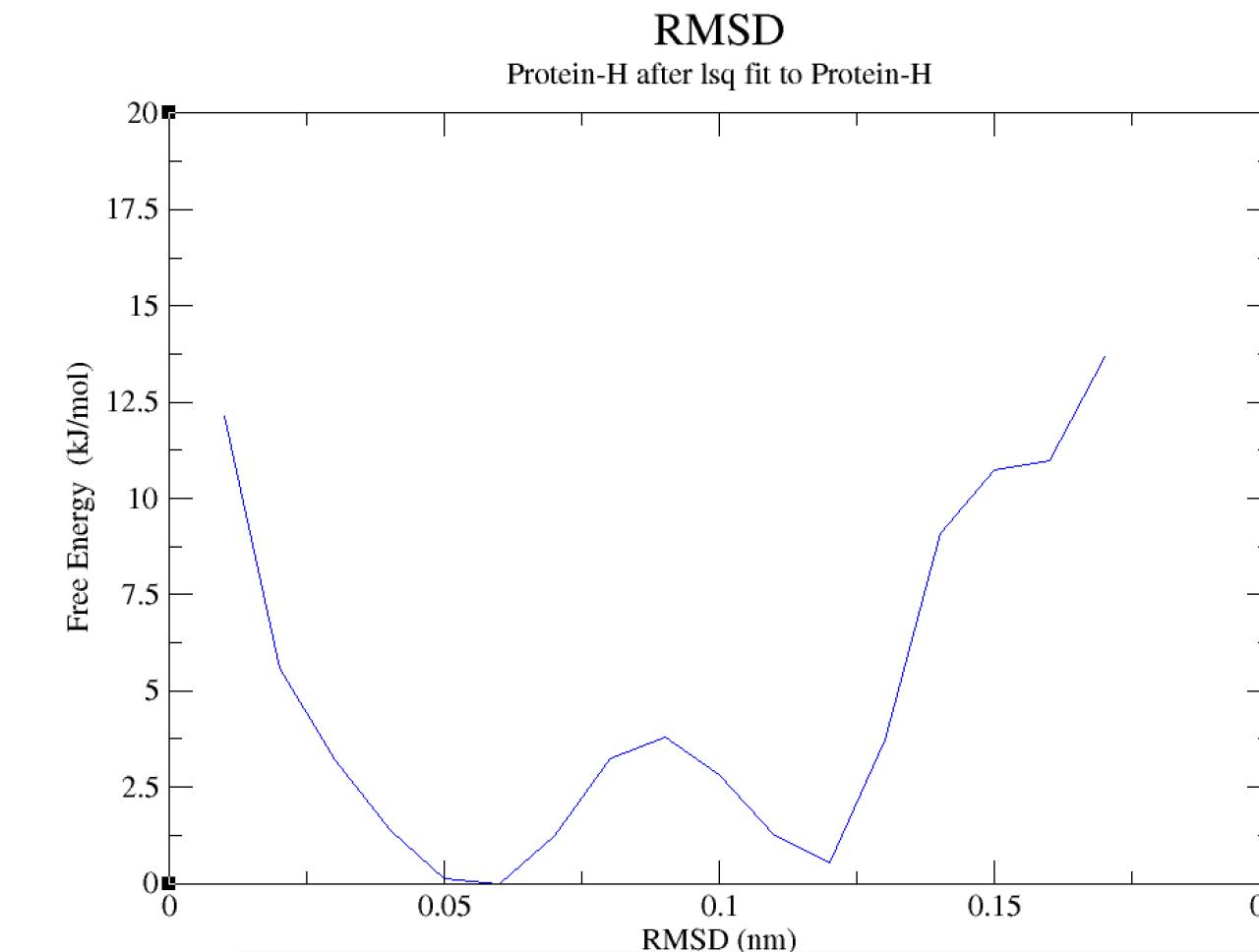
There are also machine learning techniques that try to find ‘optimal’ dimensionality reductions, the most used is Principal Component Analysis.

Equilibrium != Kinetics

By looking at histograms of simulation observables we are analysing equilibrium properties of the simulation, indeed we are removing the time-resolved information. In principle we have seen that kinetic rates are associated to free energy barriers, but while it is possible to go from rates -> barriers the opposite is not trivial:



On the left the barrier between the two main states H and E is ~8 kJ/mol (between 3-4 isolines), in the RMSD Free Energy the barrier between A and B is ~4 kJ/mol



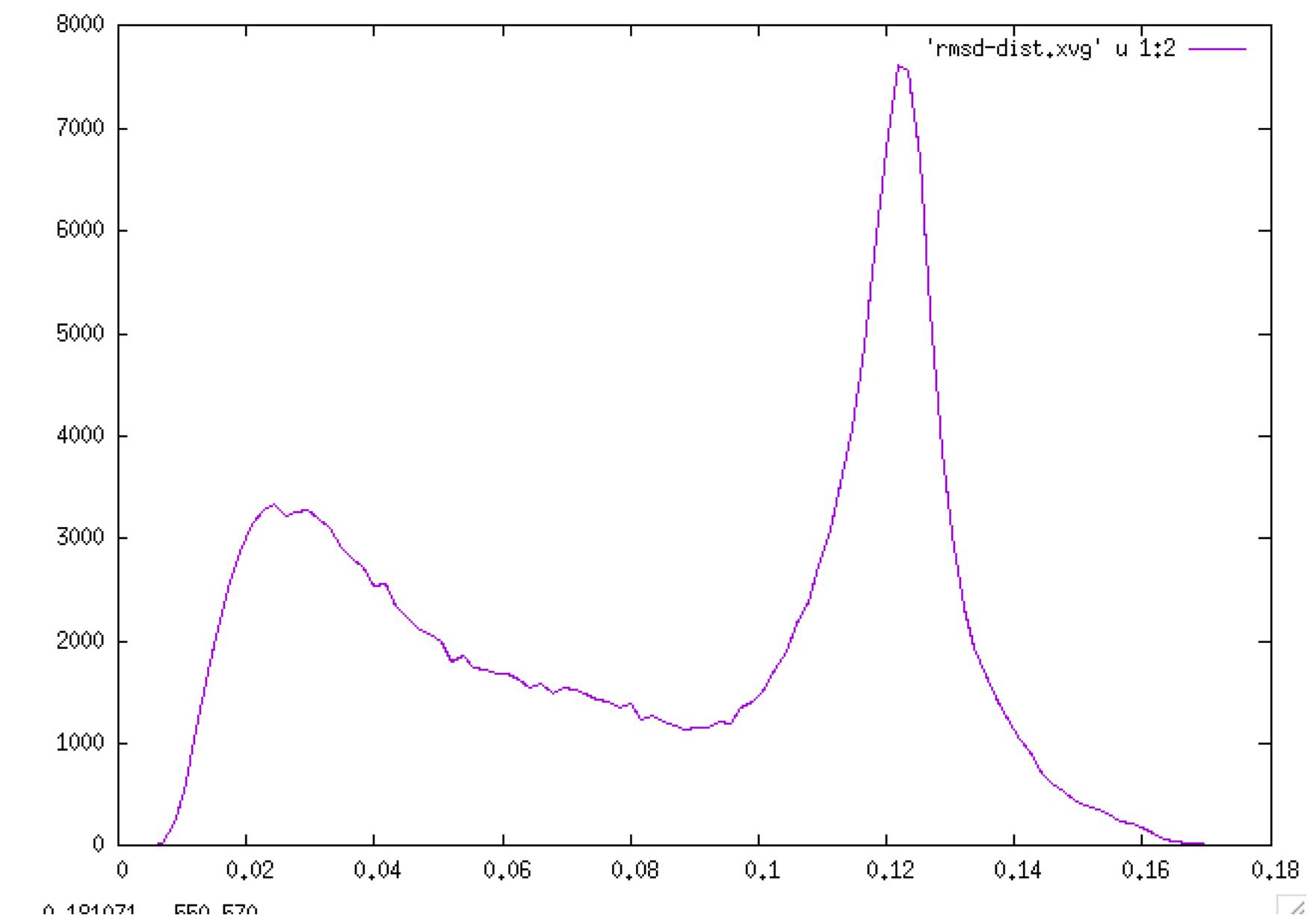
A direct clustering approach

There are multiple clustering algorithm in literature, clustering is a subfield of the statistical (or machine) learning field.

https://en.wikipedia.org/wiki/Cluster_analysis

For example a common approach in MD is that of calculating the RMSD of all pairs of conformations in the simulation, for example if we have 4000 conformation this would result in a 4000x4000 distance matrix. Then one can make an histogram of all RMSD value

This suggest that the conformations are either at 0.02 nm from each other or at 0.12 nm. Then we can consider two conformations to belong to the same cluster if they are closer than 0.06 nm.



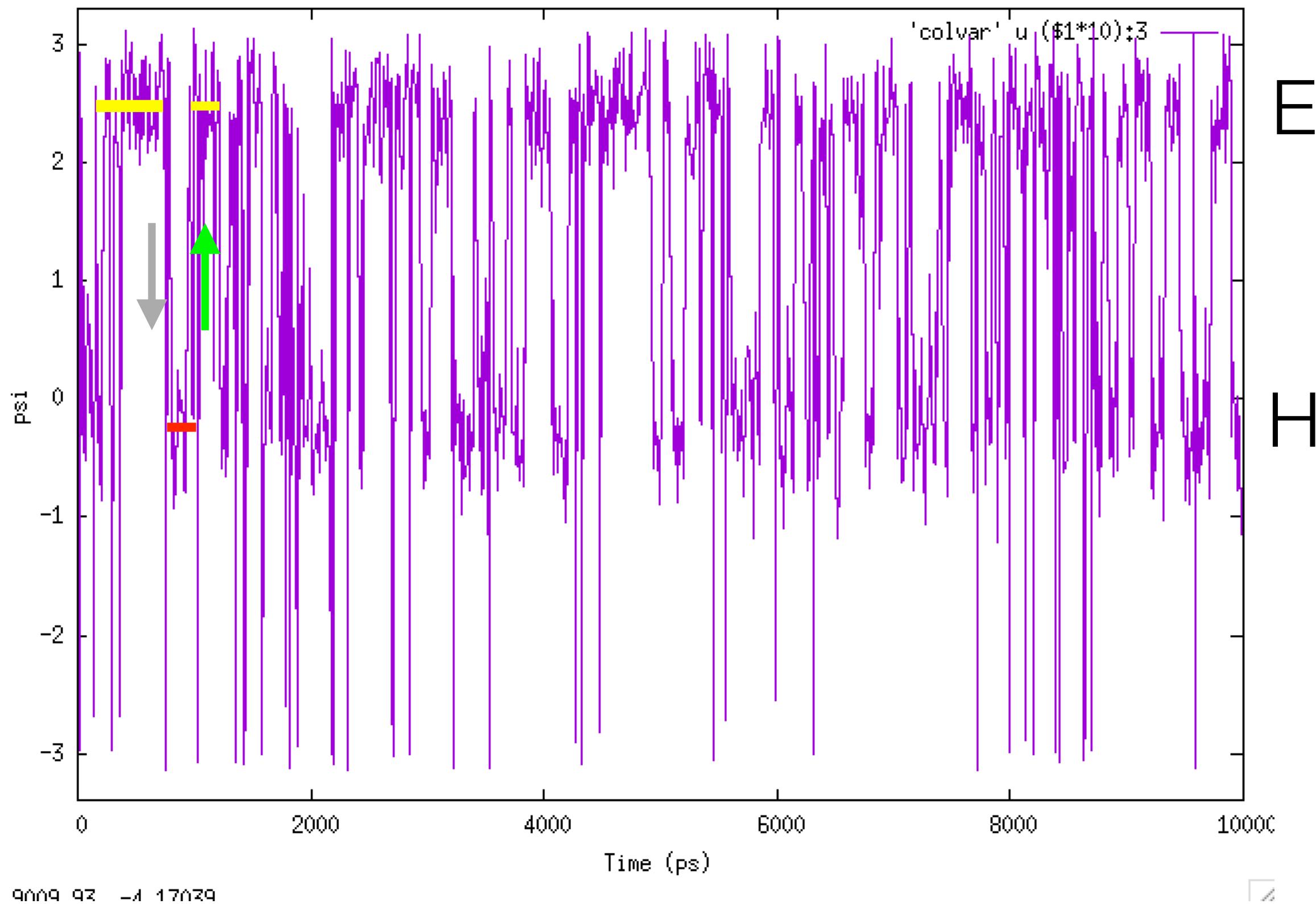
As a result we get 2 clusters: one including 97% of the population and one if the reminder 3%, this is again a different result from the previous clusterings. Again we should investigate whether this clustering is useful to say something on the system or not.



Kinetics > Equilibrium

To perform a kinetic analysis of the simulation we still need to perform a dimensionality reduction + clustering step, that is we need to look at the time-resolved trace of some observable connecting two or more states.

From the previous work we should work in the phi/psi space. We will learn more on this in the “Markov State models” lecture. Here to make it simple we focus on the psi collective variable.



We can zoom in the time resolved plot, from this we can learn a lot:

- **Residence times:** this will be the average time spent in a state
- **Transition rates:** the number of transitions per time unit
- **Equilibrium properties:** remember that from kinetics you can get free energies

Errors estimates: block averaging and replicates

The standard approach to calculate errors is to repeat an observation multiple time to get its average and standard error of the mean. Molecular Dynamics simulations allow to add an additional way to calculate statistical errors that is block averaging:

In this second case you consider your simulation has multiple experiments, multiple blocks, and then you calculate averages and standard errors of the means over the block. But then you can try to divide your simulations in 3, 4, 5, ... blocks

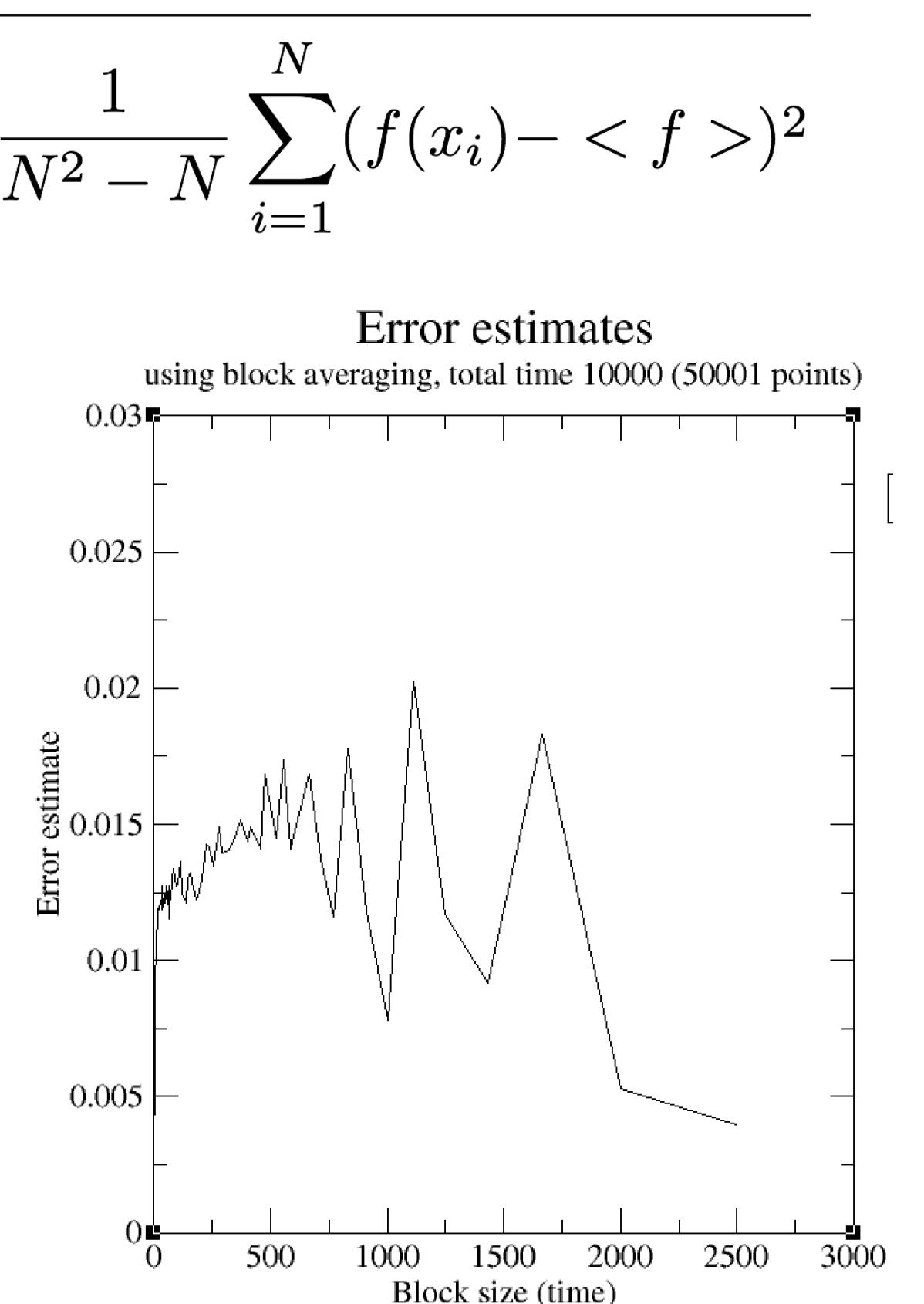
In practice we **estimate** averages using sums:

Variance and Standard deviation:

$$\sigma^2 = \text{var}(f) = \langle (f - \langle f \rangle)^2 \rangle = \int dx (f(x) - \langle f \rangle) \rho(x) = \frac{1}{N-1} \sum_{i=1}^N (f(x_i) - \langle f \rangle)^2$$

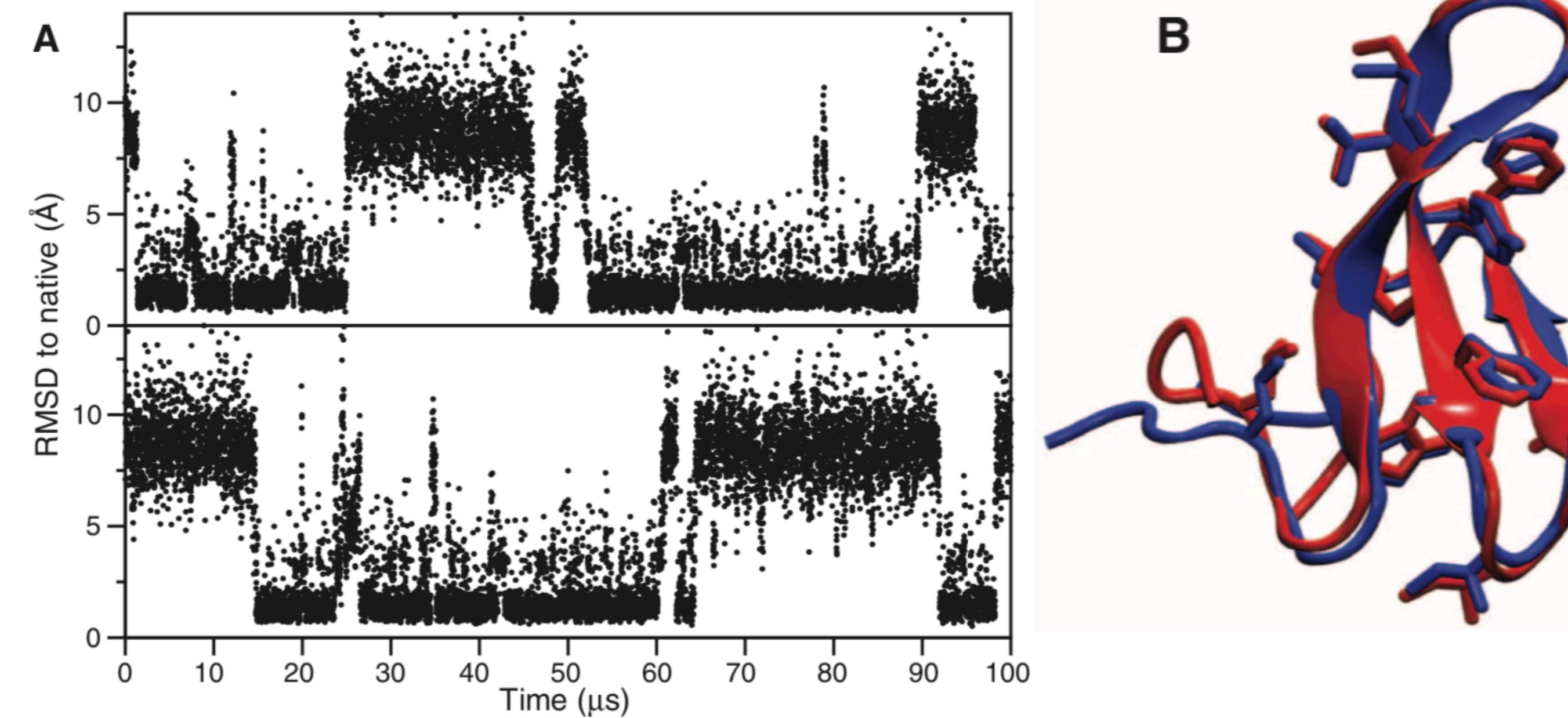
$$\text{Std-err} = \sqrt{\frac{\text{var}(f)}{N}} = \frac{\sigma}{\sqrt{N}} = \sqrt{\frac{1}{N^2 - N} \sum_{i=1}^N (f(x_i) - \langle f \rangle)^2}$$

Let's say that your simulation is 10,000 ps long, then you can split it into 4 blocks by 2500 ps, 5 by 2000 ps etc, when you have only few blocks your error estimate is noise, but there is a regime range, here around 500-600 ps.



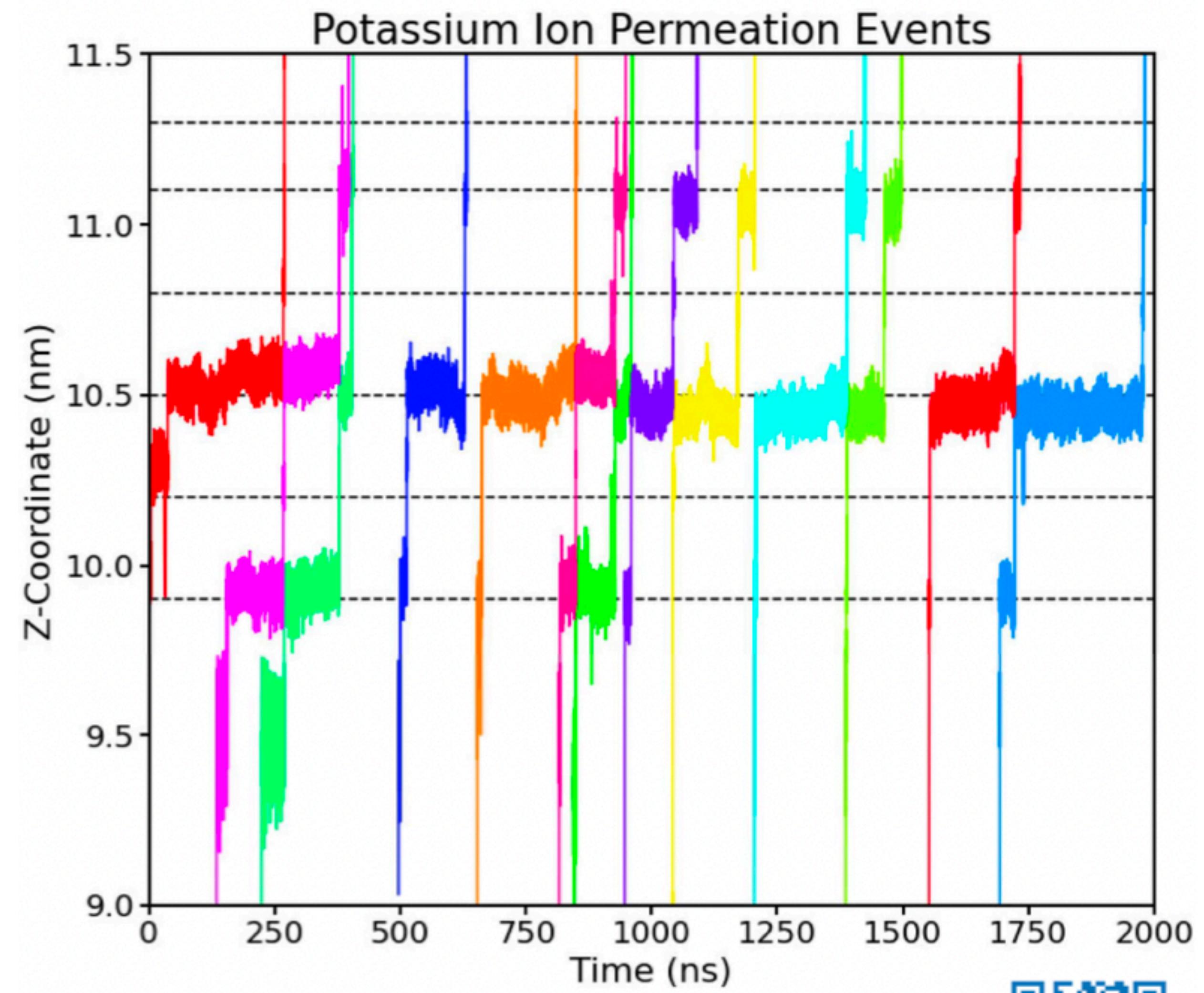
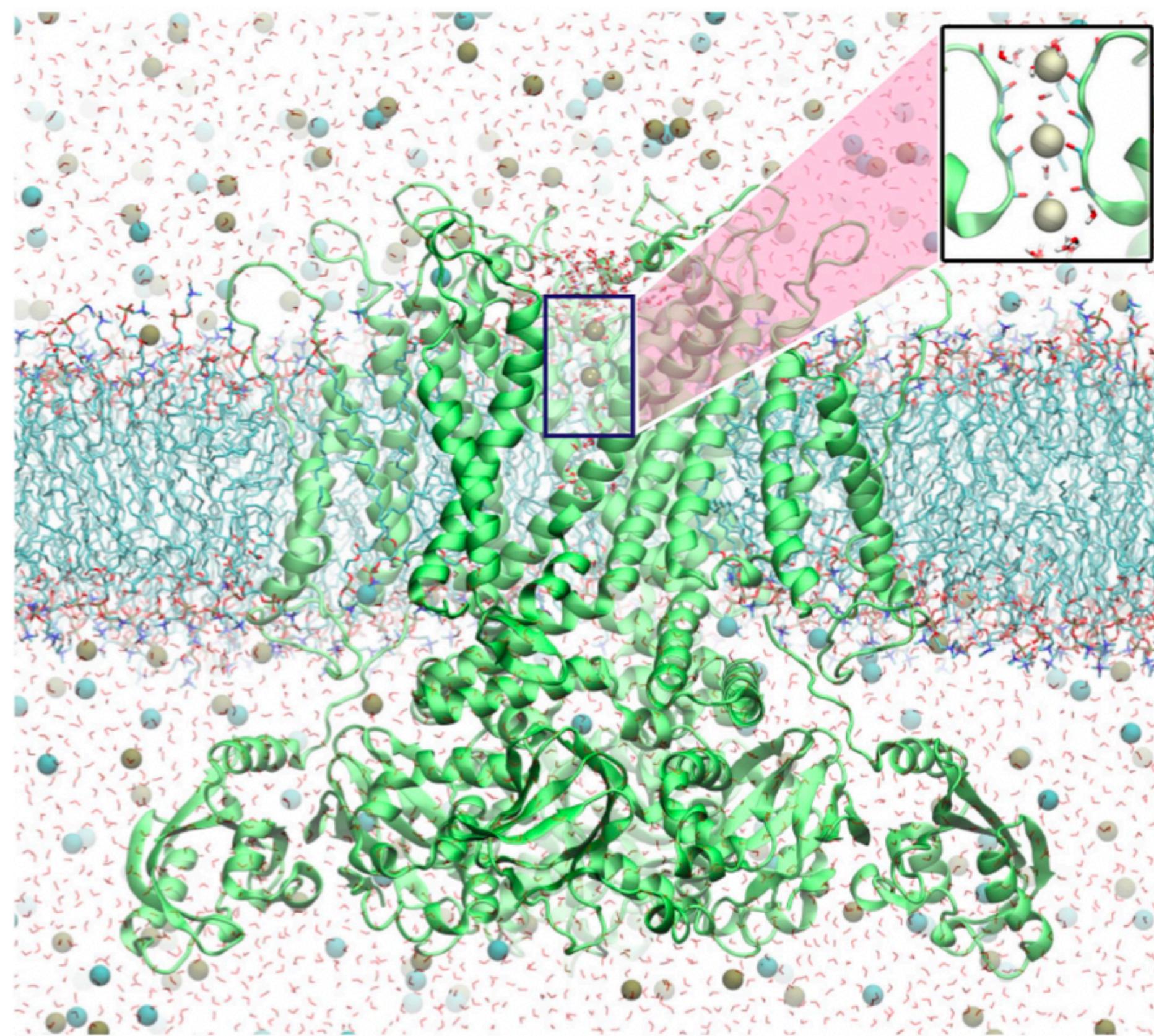
MD examples: Protein Folding

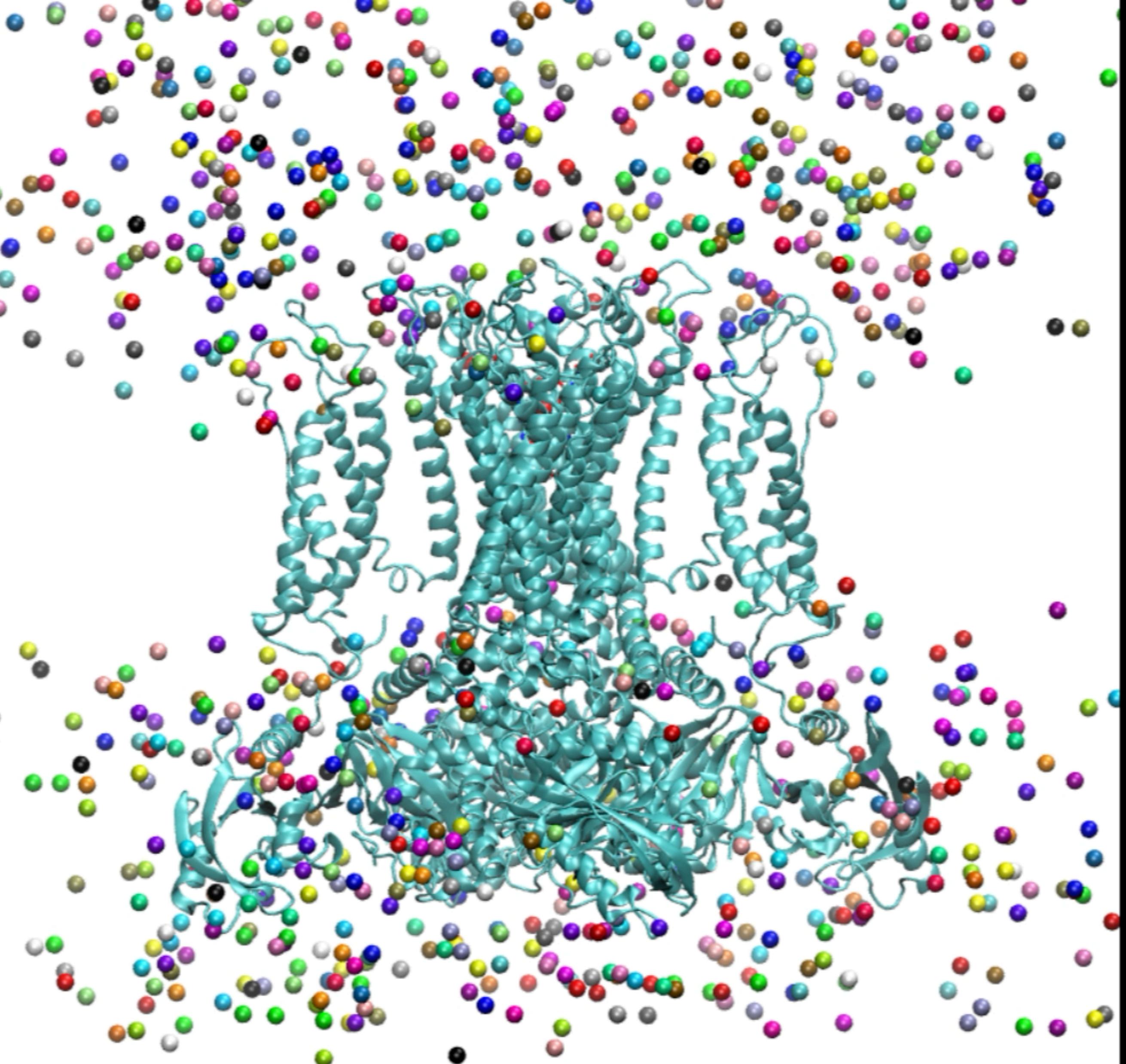
Here we want to study the reversible folding/unfolding of small protein. We take two different unfolded conformations (high RMSD from native) and we let them fold. These are very long MD and you can see multiple events. The presence of multiple events allow to calculate properties by block average and then compare them with the second simulation.



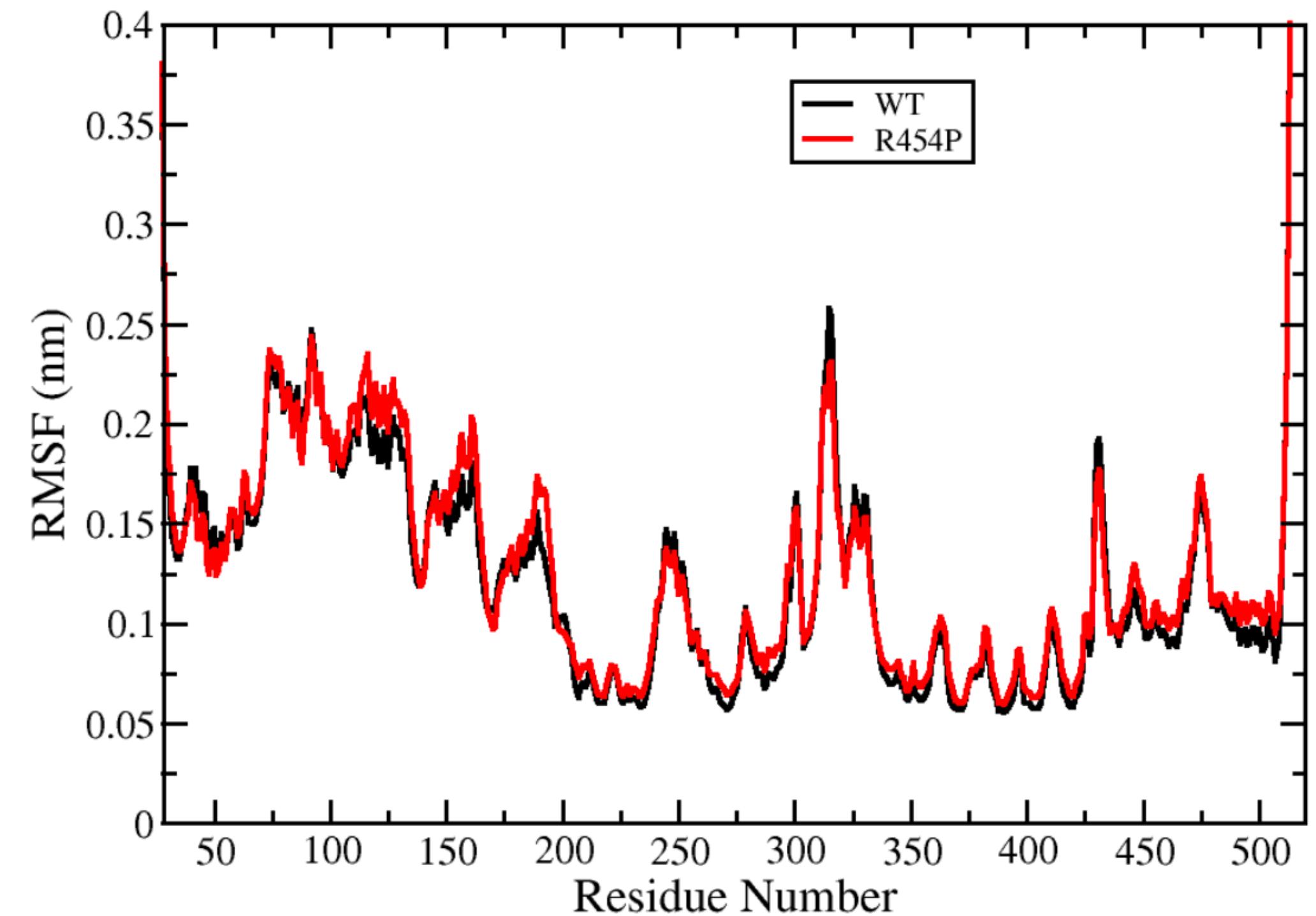
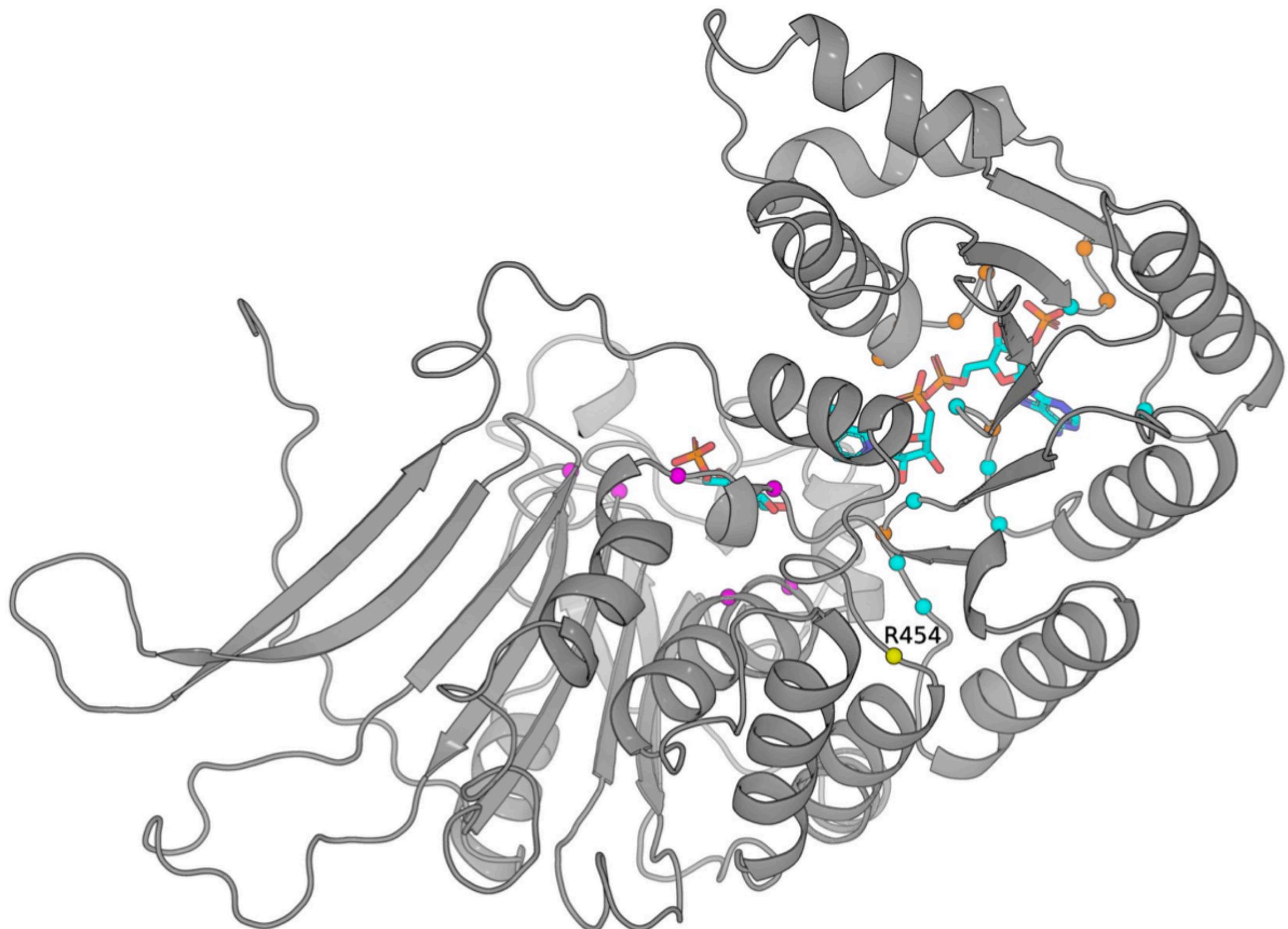
MD examples: Protein Function

Here we observe the mechanisms by which K⁺ ions go through this channel



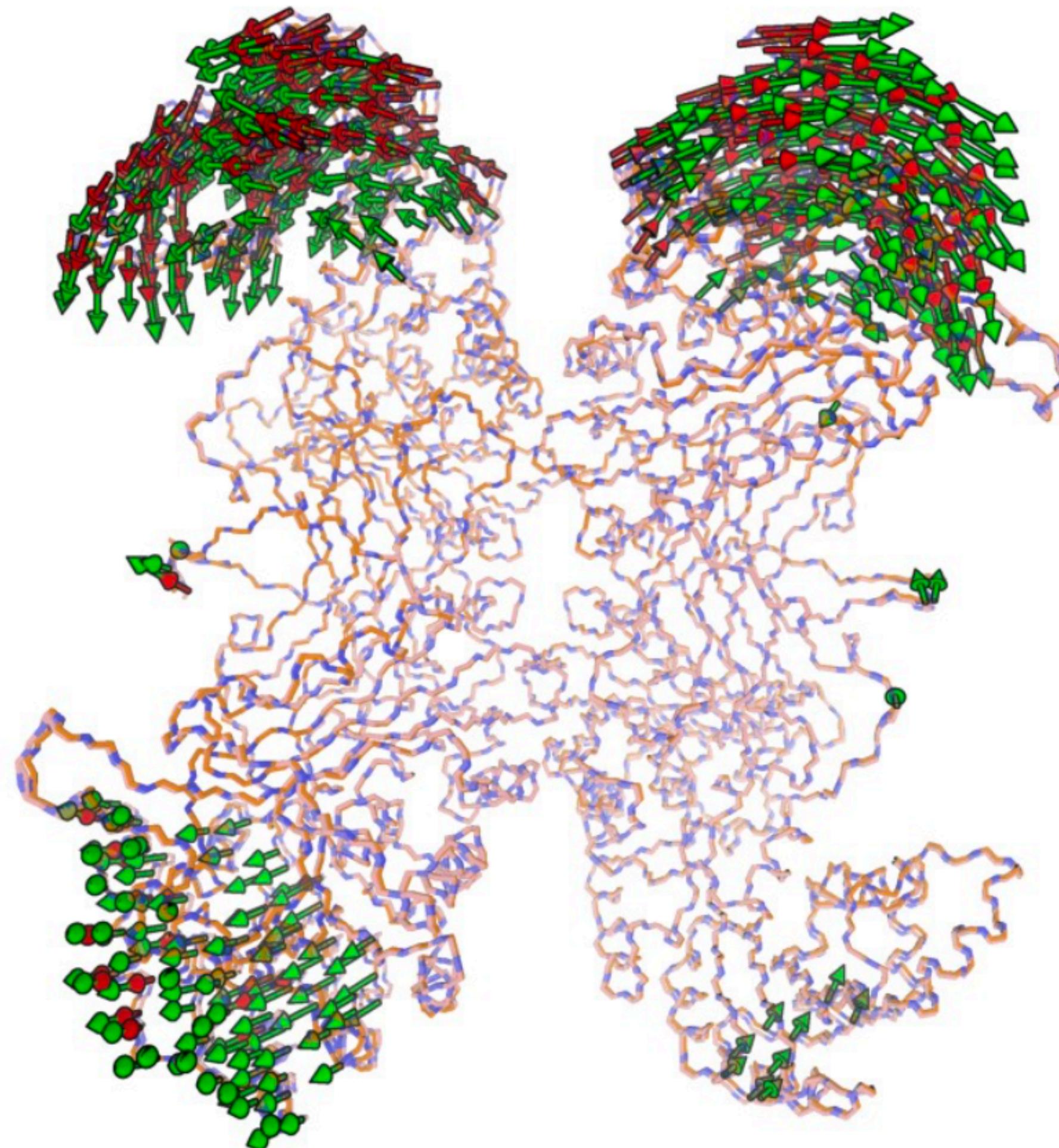


MD examples: Conformational Dynamics

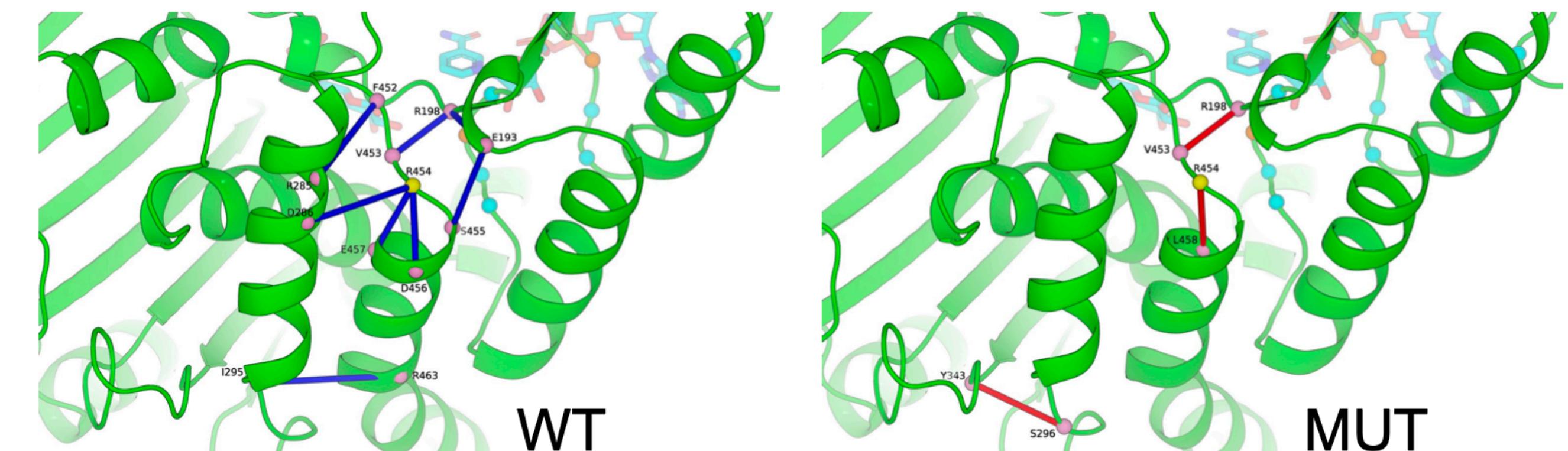


Often MD is used to investigate the different flexibility of different protein regions: here for example we show that a mutation does not seem to change the local flexibility of this protein

MD examples: Conformational Dynamics



Nonetheless, a more global view, for example provided by principal component analysis, show the presence of a global motion, that is stronger in the case of the mutation.



And then this can be associated for example to a difference in the hydrogen bond network around the site of the mutation