

CMEE lecture – Determining the structure of ecological networks

Monday 2nd March 2015

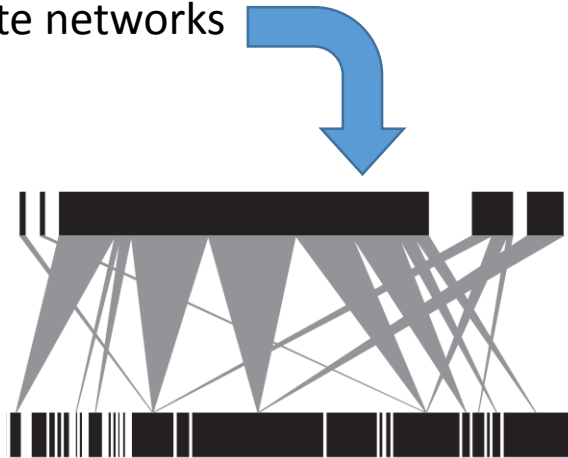
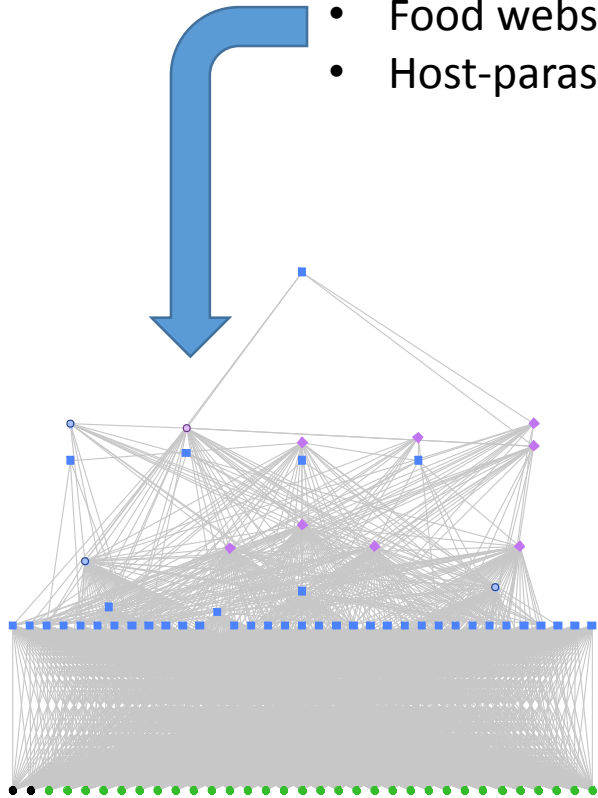
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Ecological networks

Trophic networks

e.g.

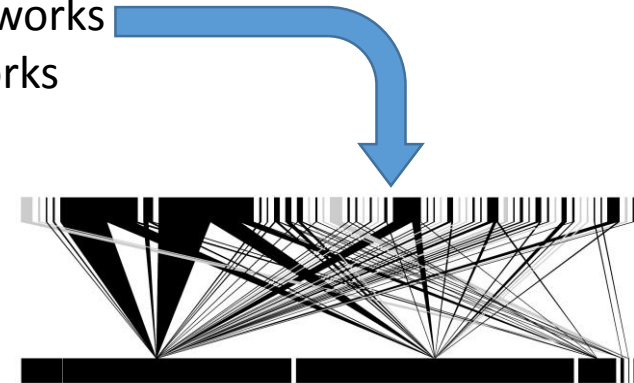
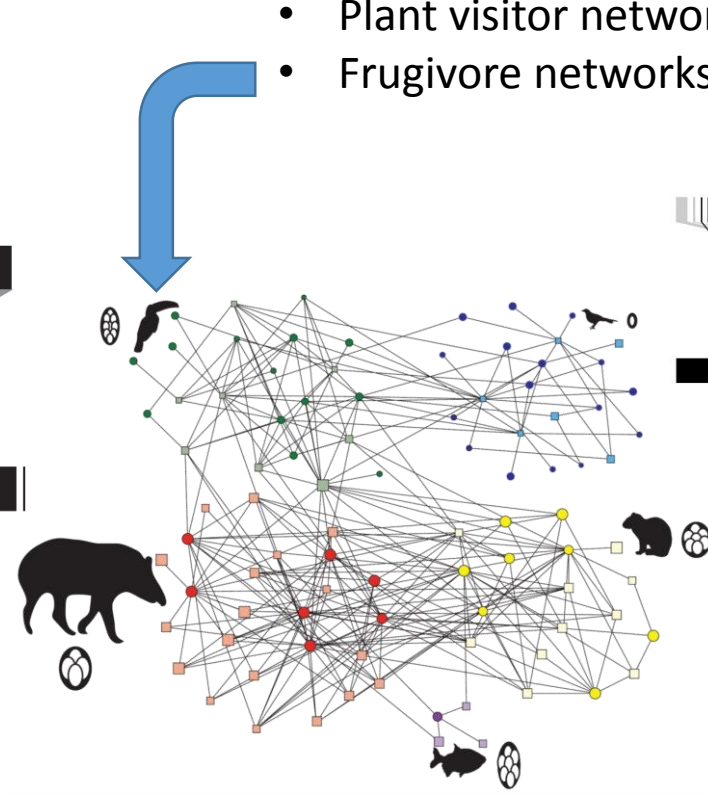
- Food webs
- Host-parasite networks



Mutualistic networks

e.g.

- Plant visitor networks
- Frugivore networks









Methods for constructing food webs

1. Gut contents analysis

- Very slow and labour-intensive.
- Rare interactions are often missed.
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- Can only build summary webs, i.e. behavioural shifts of species between sites or over time are missed.

Extrapolating from the literature

- Usually done by hand, and repeated by individual researchers anew for each study

Extrapolating from the literature

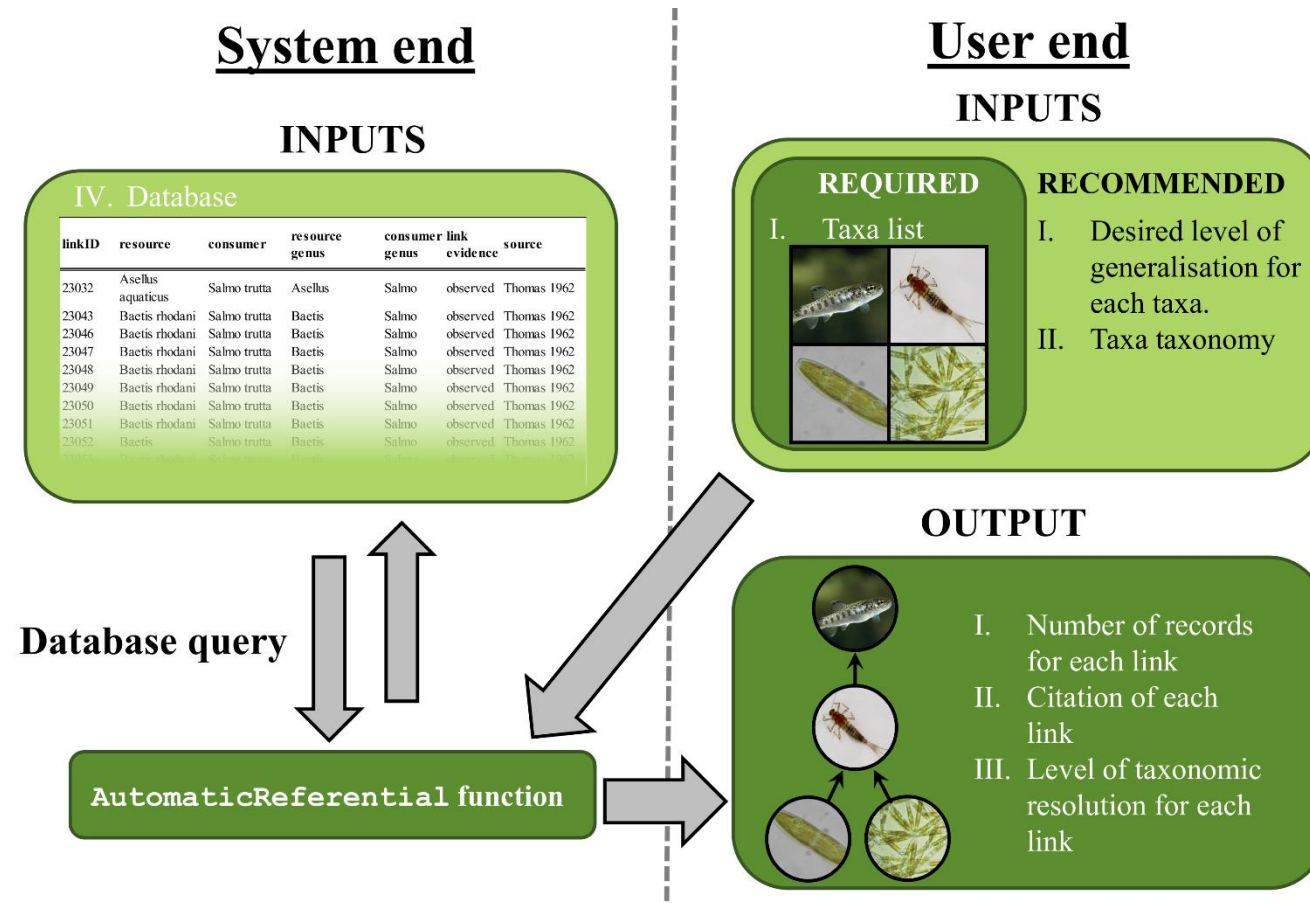
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- Not reported fully – therefor impossible to repeat

Extrapolating from the literature

- Usually done by hand, and repeated by individual researchers anew for each study
- Not reported fully – therefore impossible to repeat
- Needs to be automated, which would allow networks to be repeated and queried by different researchers

The Automatic Referential method:

Searches for matches between a given species list and a given reference dataset.



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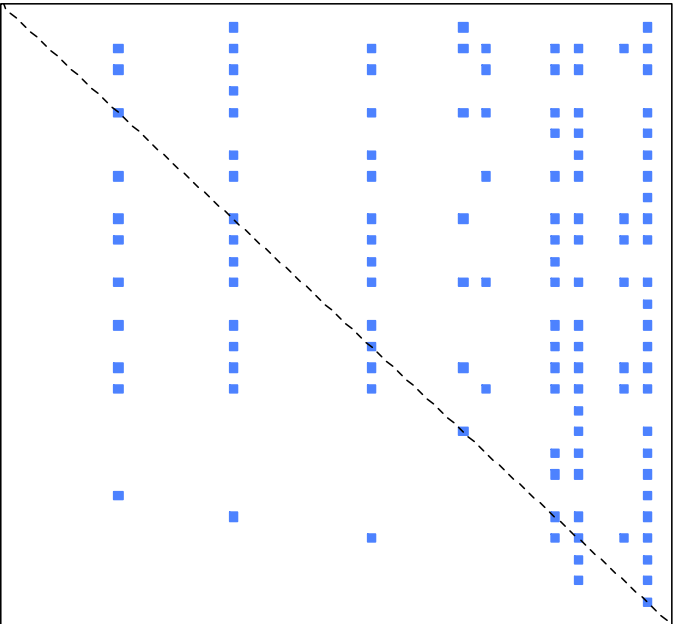
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3. Deterministic models

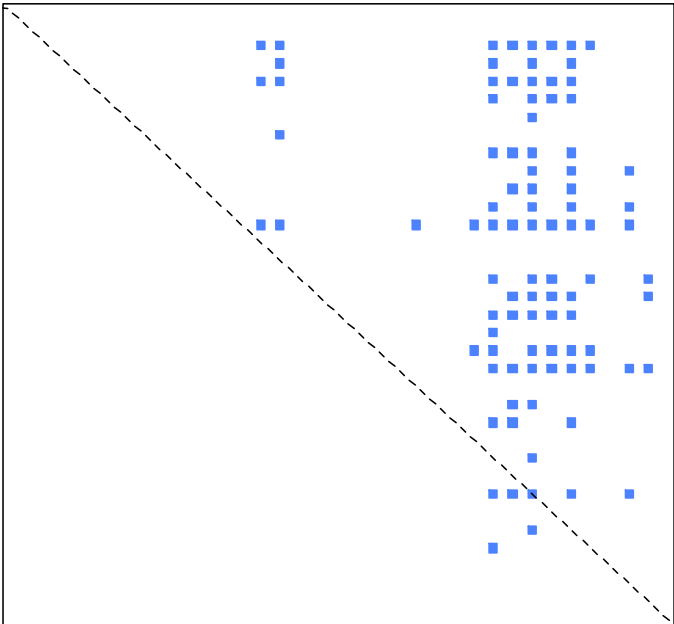
- Require prior knowledge of the structure of the network

Empirical webs

Broadstone

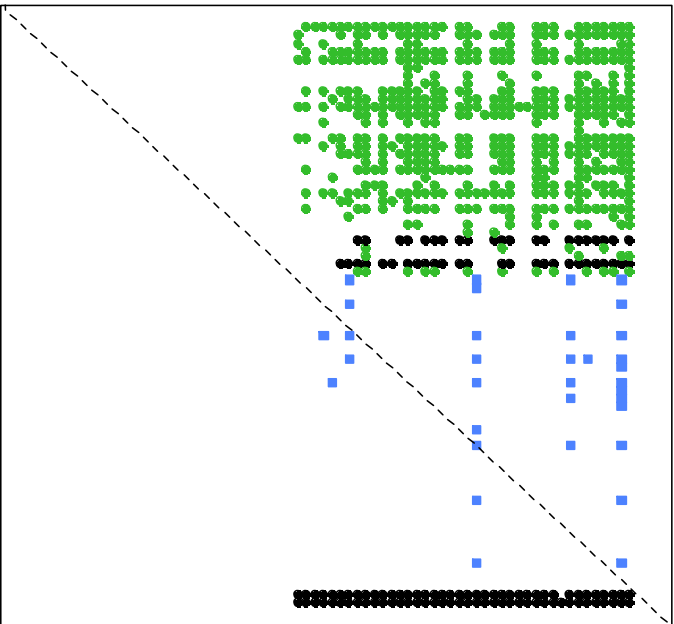


Afon Hirnant



Invertebrate-invertebrate interactions

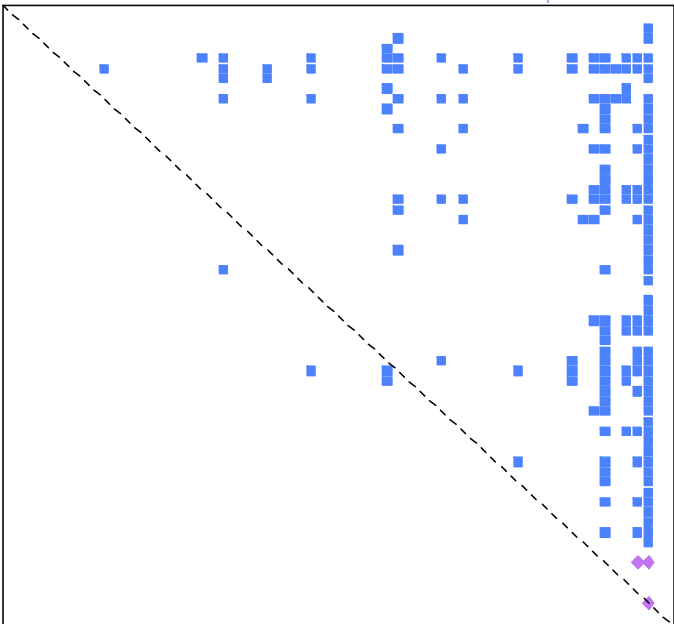
Millstream



Algae-invertebrates

detritus-invertebrates

Tadnoll



Invertebrate-fish

Fish-fish

Interactions from the literature

Constructed a dataset of interactions from the literature:

- 20,823 pairwise trophic interactions
- from 51 different data sources
- 10,348 producer-animal links,
- 9,531 animal-animal links
- 944 detritus-animal links
- Up to date, consistent taxonomy

Deterministic models

- Allometric Diet Breadth Model (Petchey et al 2008)
- 3 simpler models based on body size (Allesina 2011)
 1. $B_j - B_i$ (Difference model)
 2. B_i / B_j (Ratio model)
 3. $(B_j - B_i)(B_i / B_j)$ (Difference/Ratio model)

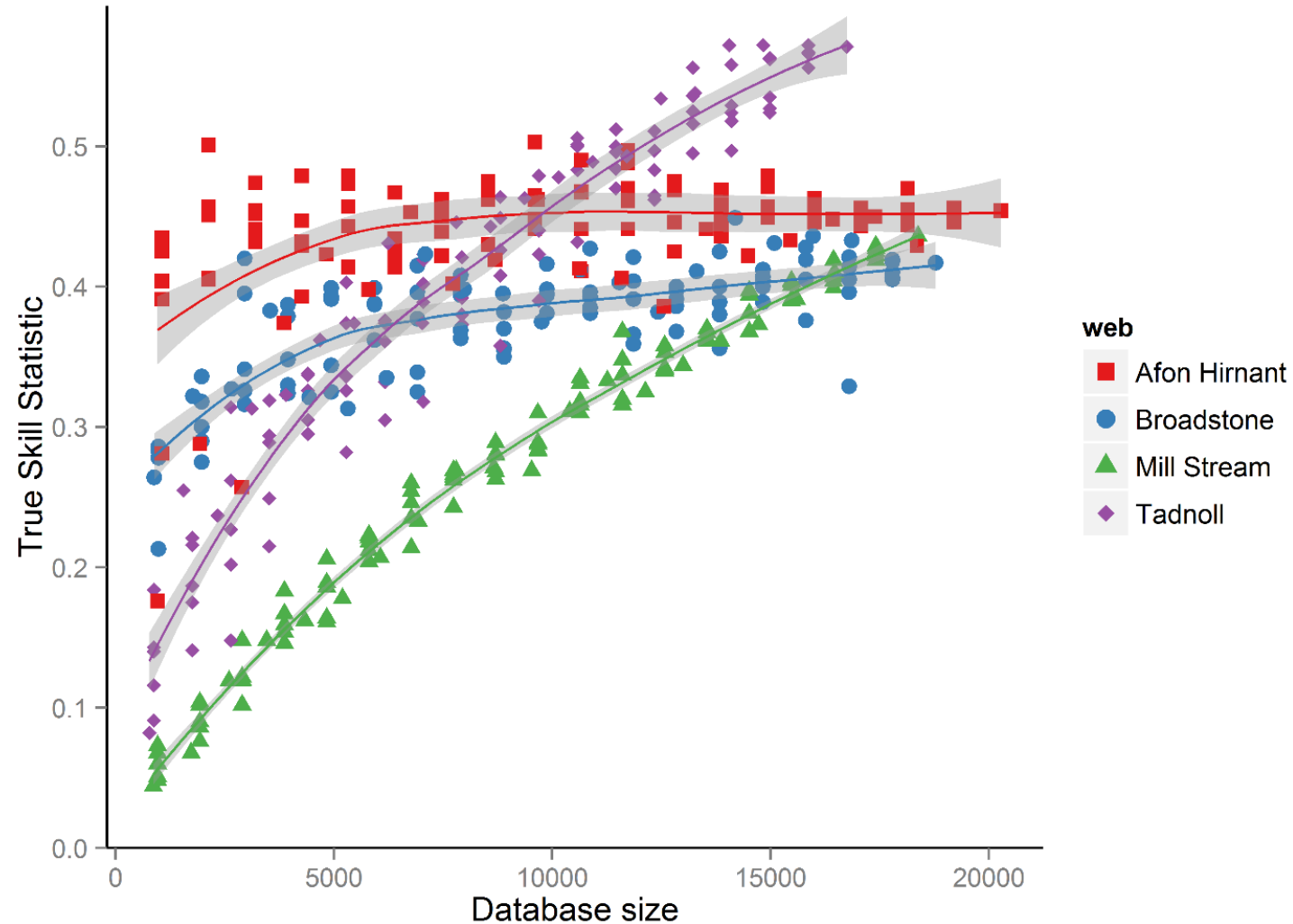
Measures of performance

- True Positive Rate (most commonly used) (a)
 - Links which were found empirically and were also predicted
- False Positive Rate (b)
 - Links which were predicted, but not found empirically
- False Negative Rate (c)
 - Links which were found empirically but not predicted
- True Negative Rate (d)
 - Links which were neither found empirically or predicted

$$TSS = (ad - bc) / [(a + c)(b + d)]$$

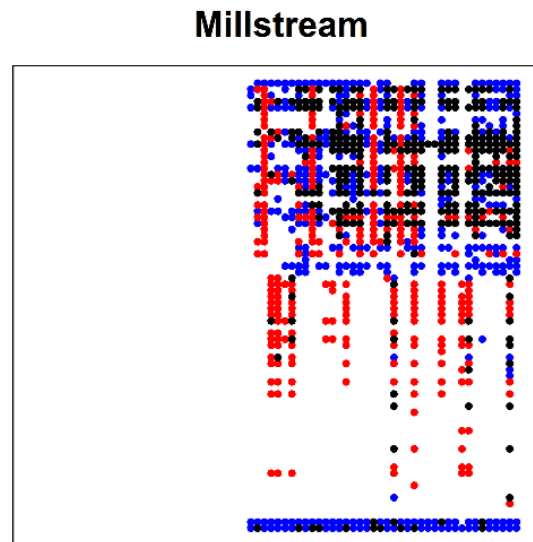
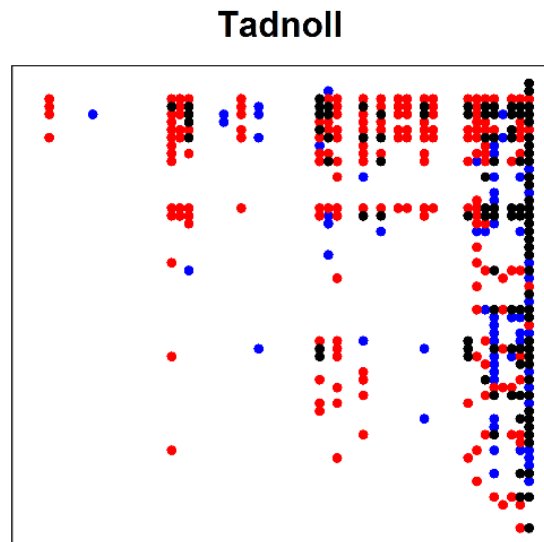
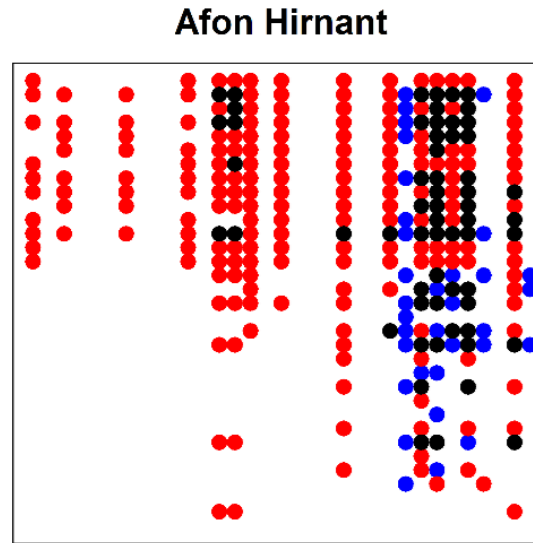
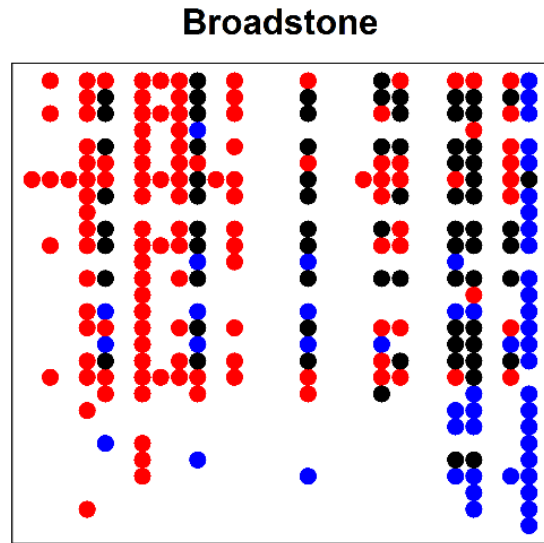
How is the quality of the food webs generated by the AR method impacted by the quality of the reference dataset?

The quality of the generated food webs improved as the quality of the dataset improved



Does the AR method generate realistic food webs compared with the empirical data?

The AR method generated realistic food webs



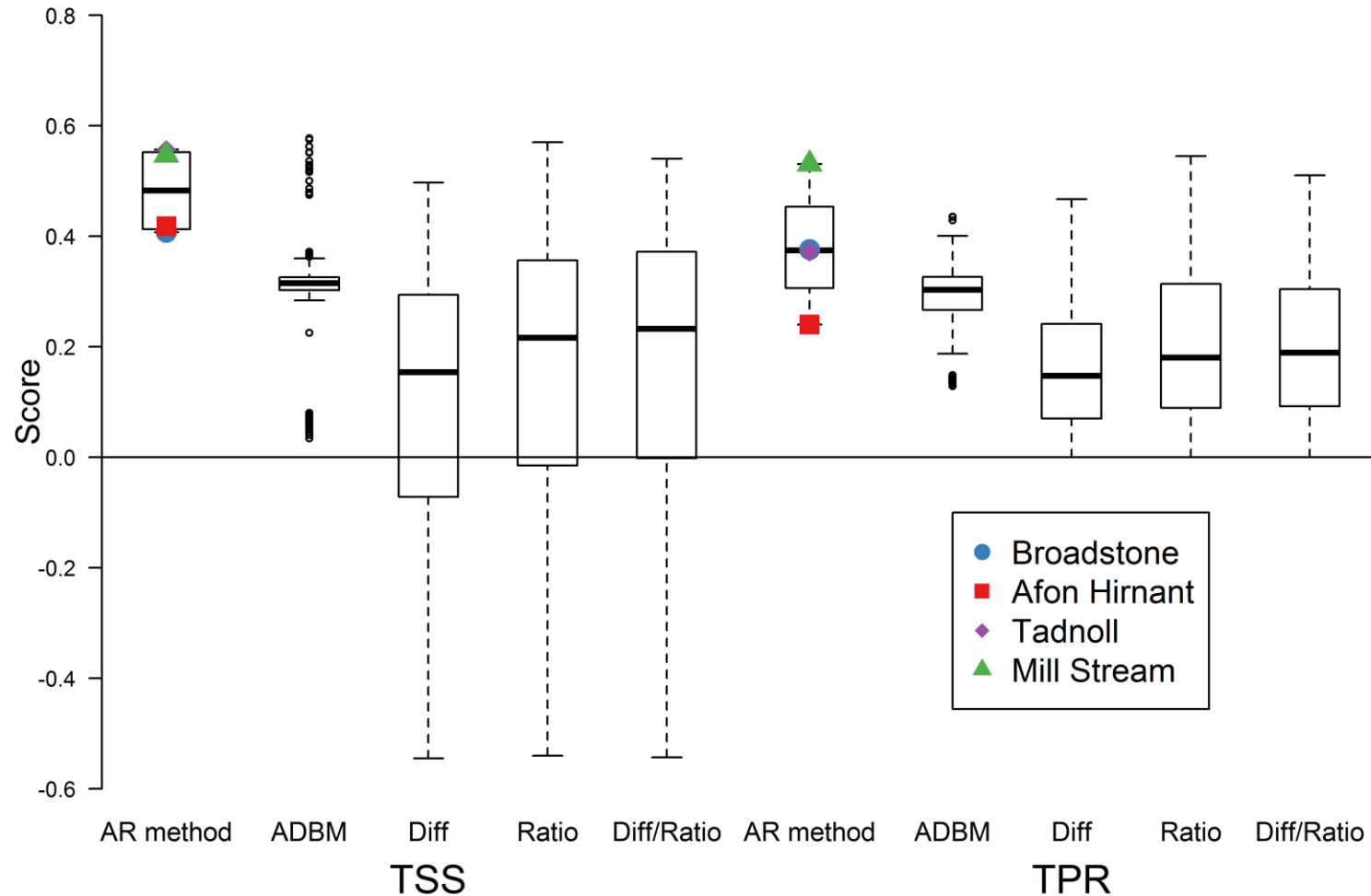
● = correctly generated

● = incorrectly generated

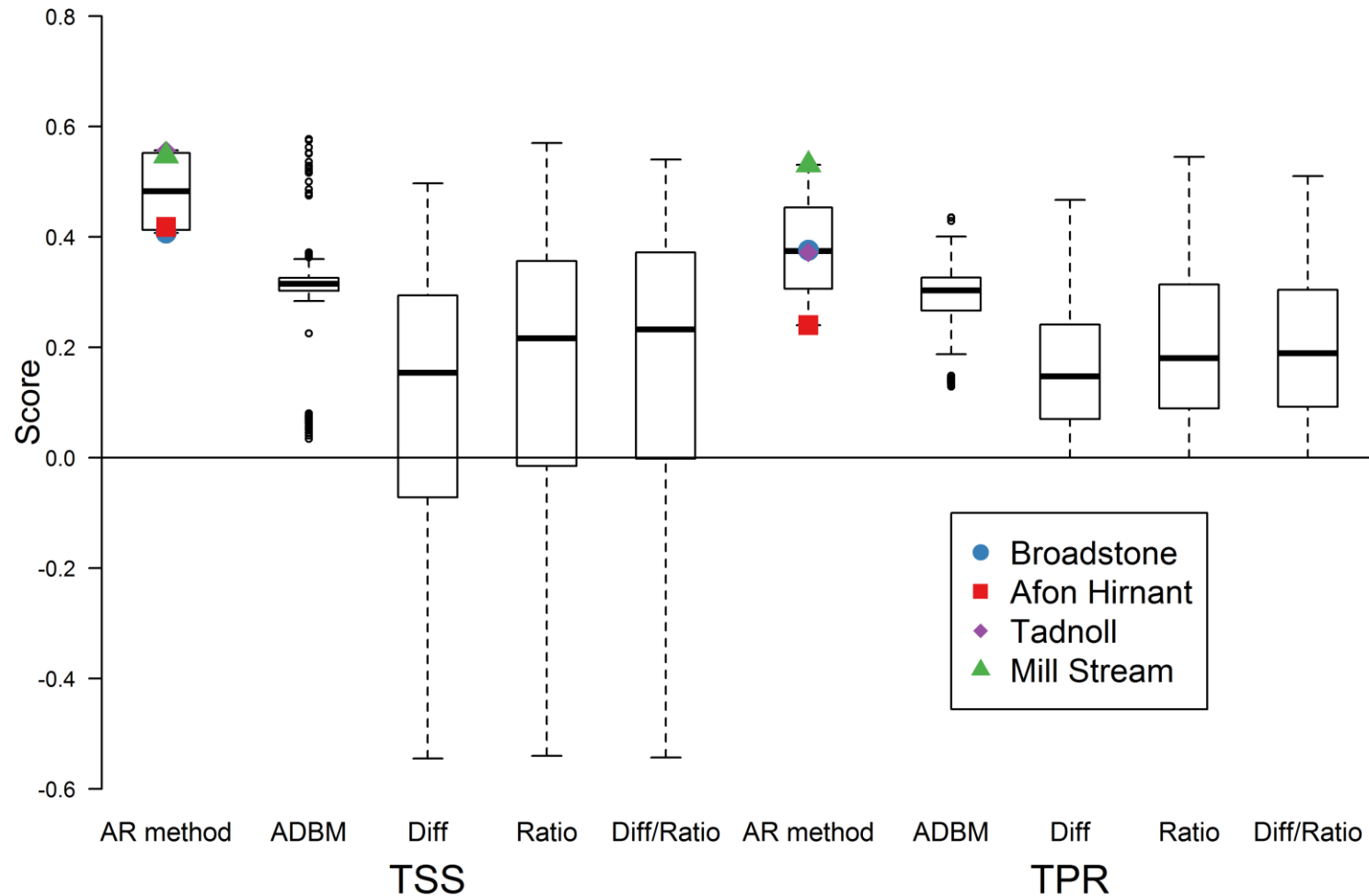
● = not generated but found empirically

How does the performance of the AR method compare to deterministic models?

Without parameter optimisation the AR method outperformed deterministic models



Without parameter optimisation the AR method outperformed deterministic models



And some networks produced were very poor

Conclusions

- For predicting new networks, the AR method is the most appropriate approach
- However, combining this with deterministic models might produce the best quality predictions...

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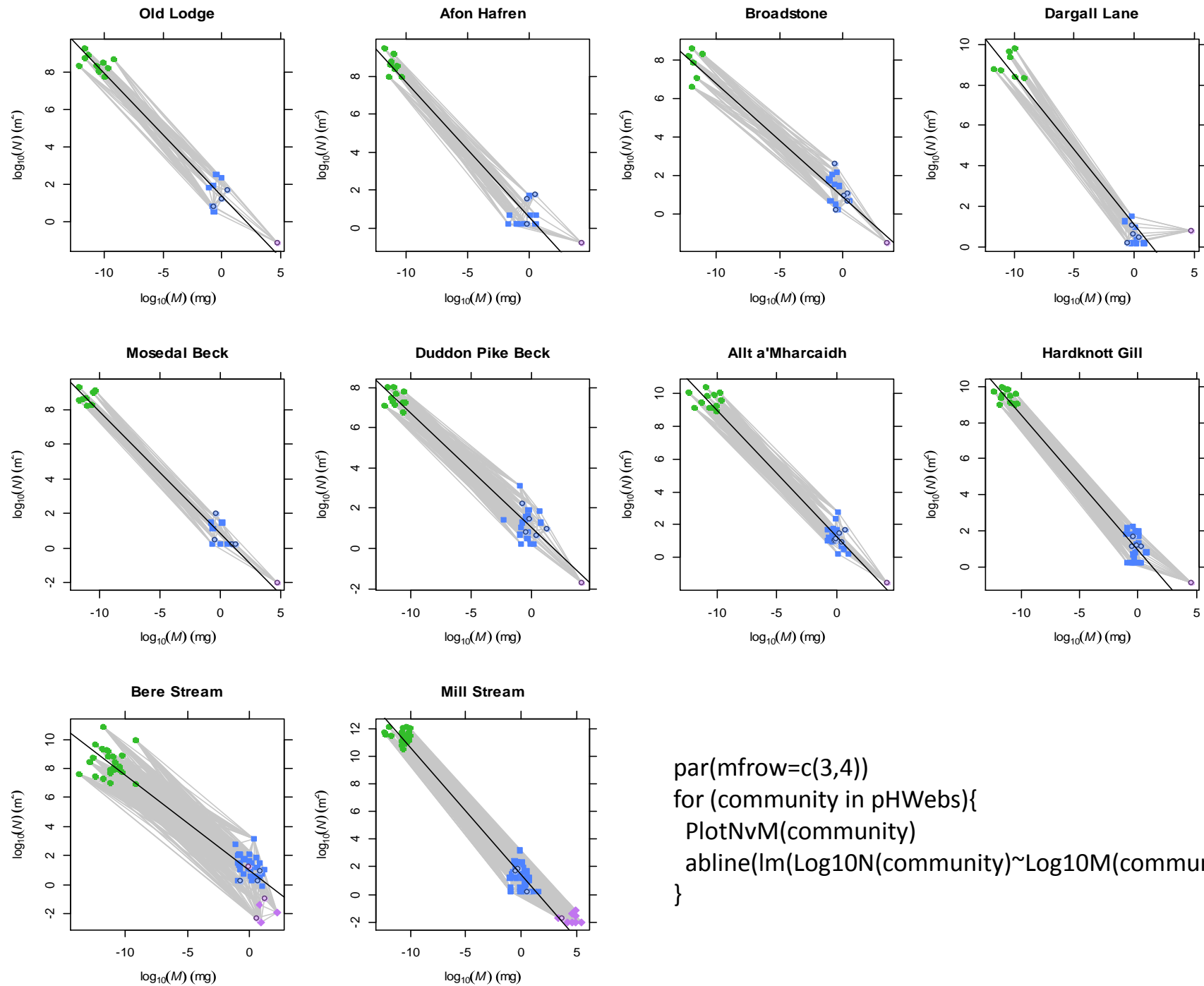
Using the AR method I have now generated an unprecedentedly large collection of freshwater food webs – 455 webs distributed across 22 sites and 25 years

Conclusions

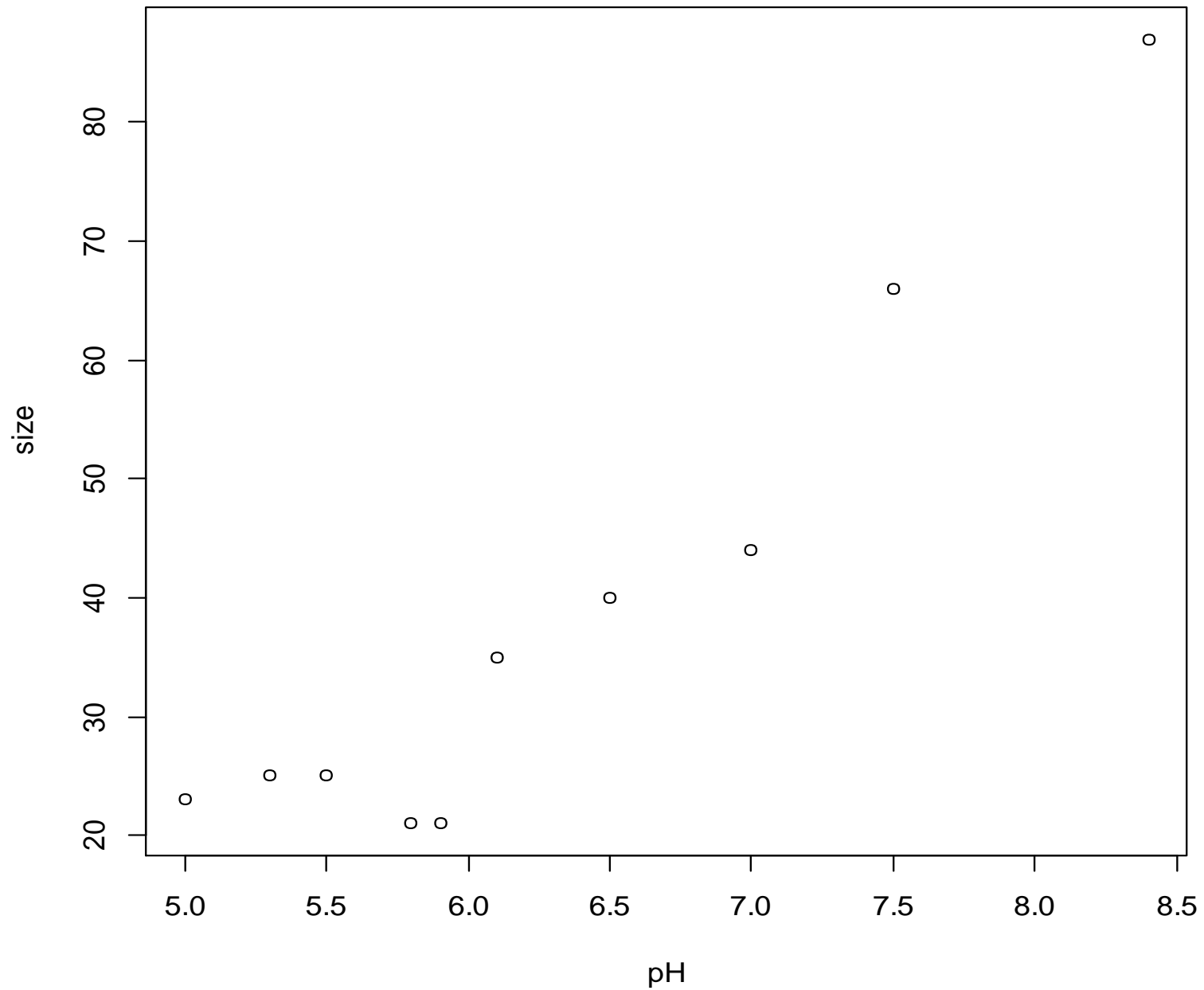
- For predicting new networks, the AR method is the most appropriate approach
- However, combining this with deterministic models might produce the best quality predictions...

Using the AR method I have now generated an unprecedentedly large collection of freshwater food webs – 455 webs distributed across 22 sites and 25 years

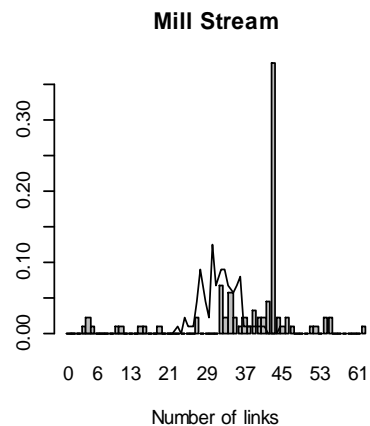
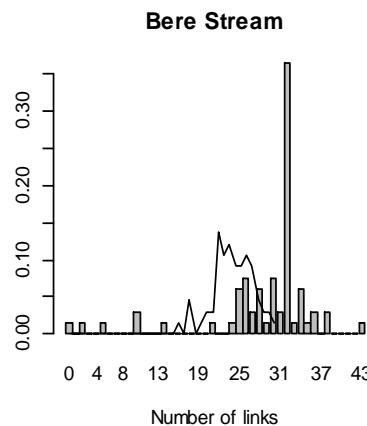
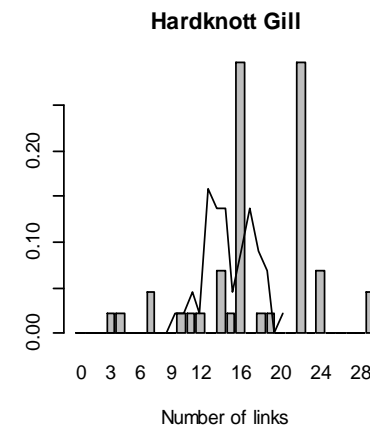
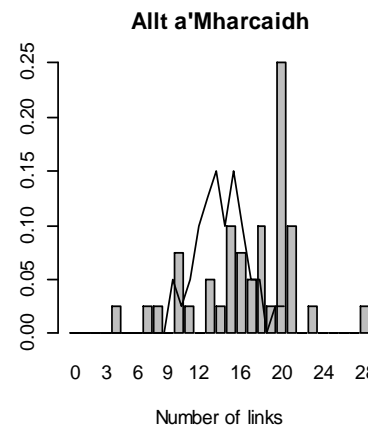
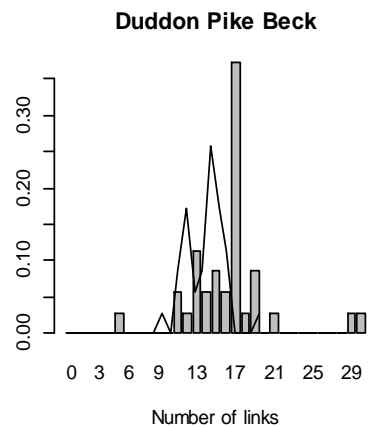
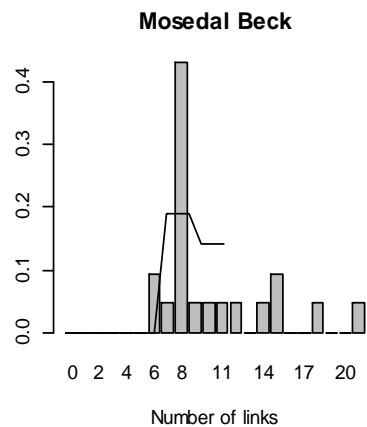
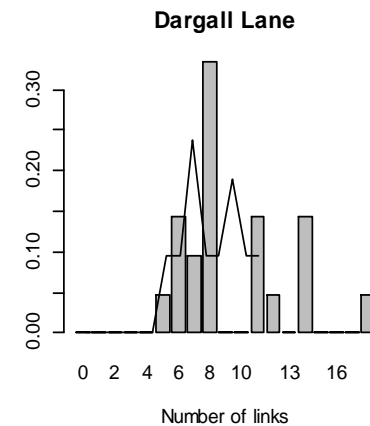
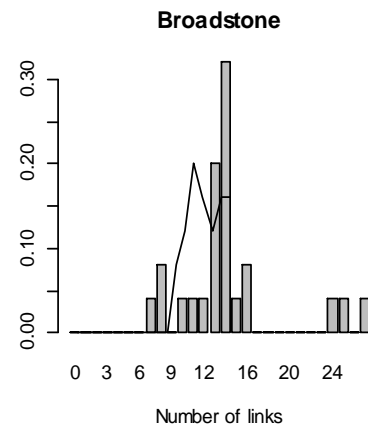
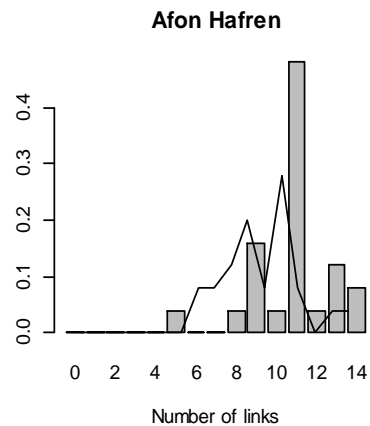
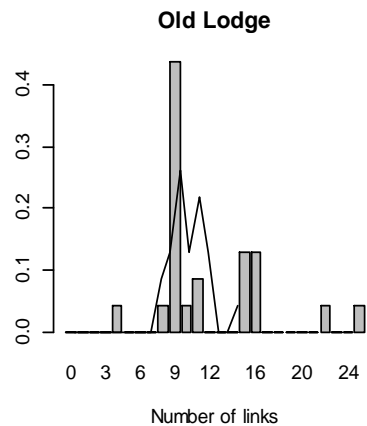
Any Questions?



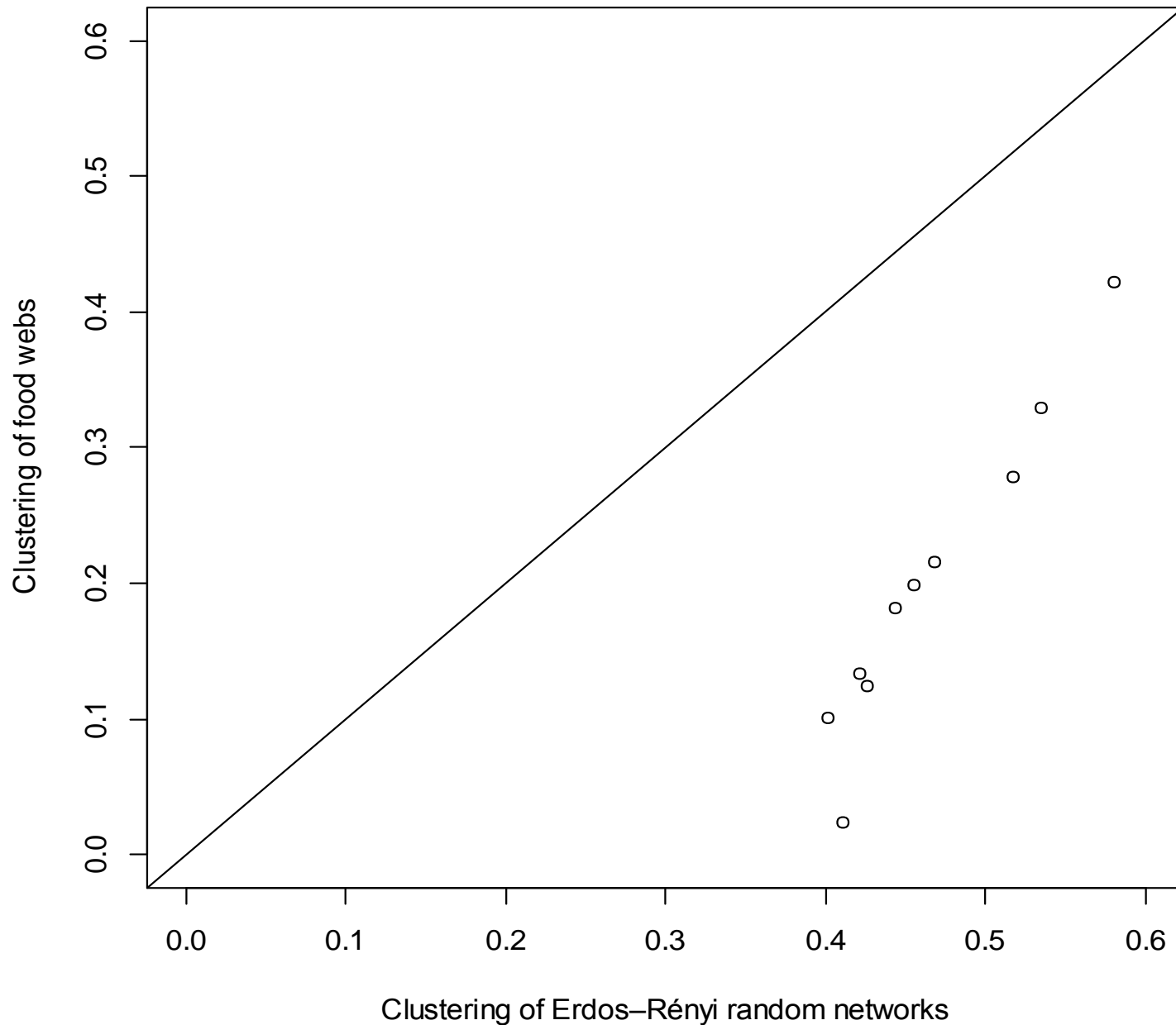
```
par(mfrow=c(3,4))
for (community in pHWebs){
  PlotNvM(community)
  abline(lm(Log10N(community)~Log10M(community)))
}
```



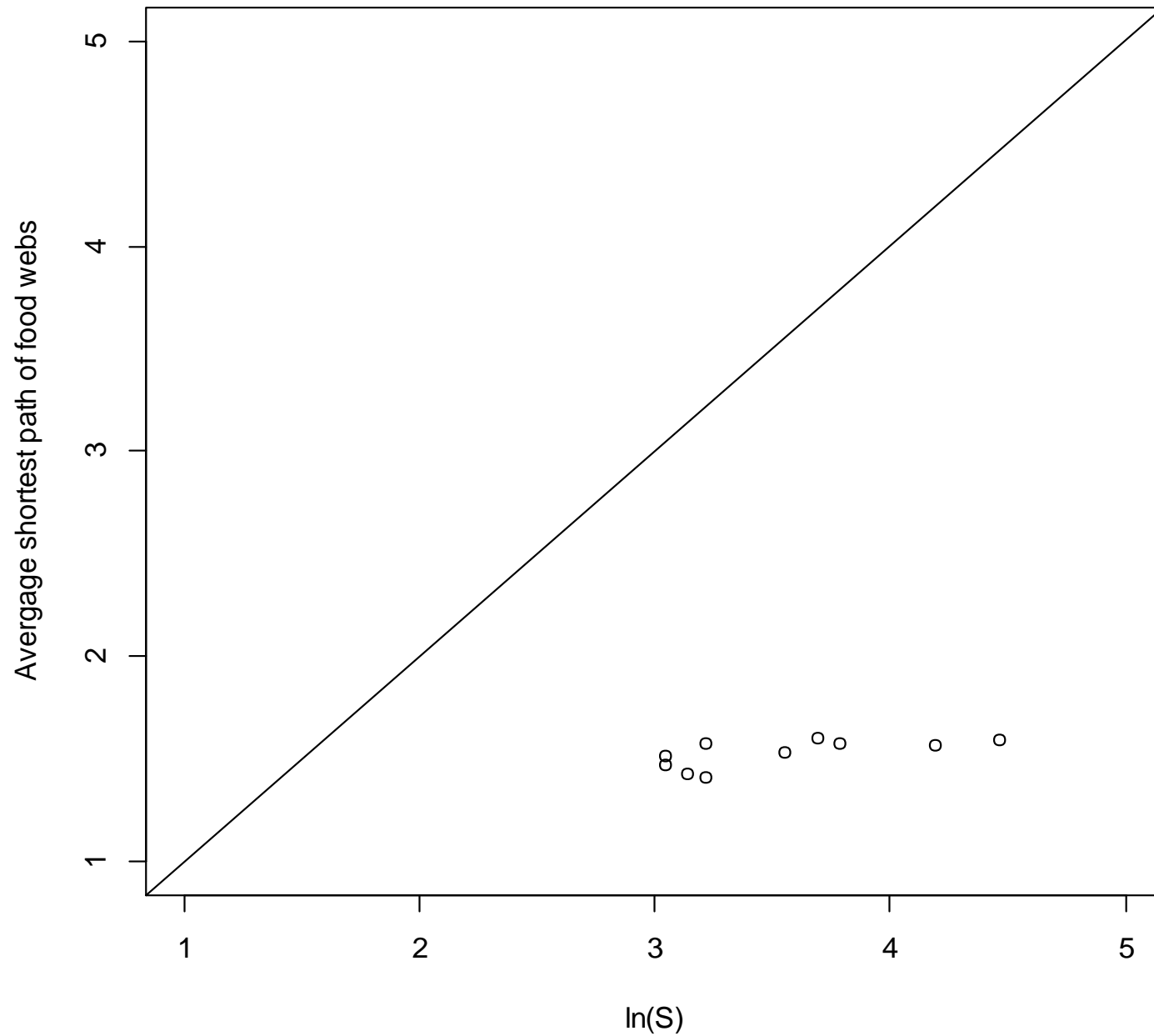
```
pH <- CollectionCPS(pHWebs)$pH  
size <- sapply(pHWebs, 'NumberOfNodes')  
plot(size~pH)
```



```
par(mfrow=c(3,4))
for (community in pHWebs){
  PlotDegreeDistribution(community, main=CPS(community)$title)
  g <- Tolgraph(community)
  n <- NumberOfNodes(community)
  m <- NumberOfTrophicLinks(community)
  ER <- erdos.renyi.game(n=n, p.or.m=m, type="gnm")
  points(degree.distribution(ER), type='l')
}
```



```
CC <- {}  
ER.CC <- {}  
for (community in pHWebs){  
  g <- Tolgraph(community)  
  CC <- c(CC, transitivity(g))  
  n <- NumberOfNodes(community)  
  m <- NumberOfTrophicLinks(community)  
  ER <- erdos.renyi.game(n=n, p.or.m=m, type="gnm")  
  ER.CC <- c(ER.CC, transitivity(ER))  
}  
plot(CC~ER.CC, ylim=c(0,0.6), xlim=c(0,0.6),  
      ylab="Clustering of food webs",  
      xlab="Clustering of Erdős-Rényi random networks")  
abline(0,1)
```



```
SP <- {}  
size <- {}  
for (community in pHWebs){  
  community <- RemoveIsolatedNodes(community)  
  g <- Tolgraph(community)  
  SP <- c(SP, mean(shortest.paths(g)))  
  size <- c(size, NumberOfNodes(community))  
}  
plot(SP~log(size), ylim=c(1,5), xlim=c(1,5),  
      ylab="Avergage shortest path of food webs",  
      xlab="ln(S)")  
abline(0,1)
```

