



Programme Handbook

2015 – 2016

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Along with this handbook, you will receive a copy of the Student Handbook for the Silwood Park Campus, containing the following important information for all living and working at Silwood.

Introduction to the department

Key contacts and information on the library, IT, safety and seminars.

Academic regulations

The regulations for the EEC course are provided in this handbook, but the Student Handbook provides information about the general regulations. This includes academic integrity, plagiarism, employment during your studies and complaint and appeals procedures.

Welfare and Advice

Imperial has a wide support network for students. The Student Handbook provides details of the available support and key contacts and links.

Student Feedback and Representation

We are very grateful for feedback on the course and will ask you for it at regular intervals! However, there are a range of options for providing feedback and getting support on your academic studies and the Student Handbook provides details.

Electronic copies of both of these handbooks are available on the course website as well as Blackboard. A copy can also be obtained by emailing the Course Administrator Mrs. Amanda Ellis (amanda.ellis@imperial.ac.uk).

1 Course Overview

Welcome to the Masters programmes in Computational Methods in Ecology and Evolution (CMEE) at Silwood Park!

The frontiers of biology are increasingly at the interface between mathematics, computing, and large empirical data sets. Answering important problems about issues ranging from disease dynamics and epidemiology to the effects of climate change and over-fishing on aquatic ecosystems, require computationally sophisticated approaches for management & analysis of "big data", theoretical modelling, and fitting models to data. Indeed, biology is *the* new frontier for applied computer science and mathematics. Donald Knuth, an eminent mathematician and computer scientist has said: "*It is hard for me to say confidently that, after fifty more years of explosive growth of computer science, there will still be a lot of fascinating unsolved problems at peoples' fingertips, that it won't be pretty much working on refinements of well-explored things. Maybe all of the simple stuff and the really great stuff has been discovered. It may not be true, but I can't predict an unending growth. I can't be as confident about computer science as I can about biology. Biology easily has 500 years of exciting problems to work on, it's at that level.*".

MSc and MRes CMEE aim to teach computational approaches towards the empirical and theoretical study of ecological and evolutionary systems. We focus on Computational *Ecology and Evolutionary Biology* in particular because some of the most complex and important dynamics in human-dominated as well as natural environments arise from ecological and evolutionary processes. Therefore, computational skills needed to analyse data and model ecological and evolutionary systems are very relevant to other disciplines. For example, it is now generally recognized that a major new frontier in understanding disease dynamics and epidemiology is the need to consider the *Ecology* of infectious diseases. Furthermore, ecological and evolutionary models and computational tools are also relevant for non-biological disciplines — for example, ecosystem models are increasingly being considered relevant for understanding microeconomic systems.

Both MSc and MRes CMEE run for one year commencing the first week in October through to the end of September. The first nine weeks of the first term is shared between both courses and includes a combination of lectures, workshops and practicals to cover a range of fundamental skills. The MRes option then focuses on a substantial research project for the second and third terms while the MSc option continues with more advanced taught modules followed by a shorter project. Switching between the courses is straightforward during the initial part of the first term, and possible at later stages. MRes students are welcome to choose further MSc lectures that are relevant to their research project.

Research projects are undertaken from the 10th week of the course (MRes) or from the 25th week (MSc). See timetables (section 2.2).

Daily lectures and practicals, unless otherwise stated, commence at 10:00 and would normally finish by 1700, incorporating breaks. Additional independent work is also required. On Wednesday the afternoon is normally, but not always, reserved for private study, sports and leisure activities for students.

In addition to the formal taught and research components of the programme, there are two research seminar series that run at Silwood Park. The first series (running on Mondays 1pm) are internal departmental seminars, presenting research by Imperial PhD students and research staff. The second series (running every other Thursday at 1pm) are external research seminars presented by visiting academics.

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Teaching materials and other course materials will be provided using the online Blackboard virtual learning environment <http://bb.imperial.ac.uk>, and/or for certain modules, using a git version control repository to provide code and data files. Paper copies of lecture notes and handouts are not normally provided but you will receive printing credit for use during the course on your security card.

It is anticipated that reading and coursework will require additional study in your own time. During research projects, you are expected to work full time on the project, but with flexible hours, and in general, students who work extra hours do get more out of the course. Some projects may require out-of-hours work, for example maintaining greenhouse experiments.

The full programme specifications for the MSc and MRes are available on Blackboard and from the course website below, but the following sections provide a summary of the programme and assessment structure for the two courses. Aims, objective and learning outcomes for the course are in section

1.

Course website

<https://www.imperial.ac.uk/life-sciences/postgraduate/masters-courses/-msc-in-computational-methods-in-ecology-and-evolution-cmee/>

1.1 Course Administration

<i>MSc & MRes CMEE Course Director</i>	Dr. Samraat Pawar (ext. 42213, s.pawar@imperial.ac.uk)
<i>MSc & MRes CMEE Course Co-Director</i>	Dr. James Rosindell (ext. 42242, j.rosindell@imperial.ac.uk)
<i>Postgraduate Administrator</i>	Mrs. Amanda Ellis (ext. 42251, amanda.ellis@imperial.ac.uk)
<i>Postgraduate Tutor</i>	Dr. Julia Schroeder (julia.schroeder@imperial.ac.uk)
<i>Director of Postgraduate Studies</i>	Dr. Niki Gounaris (ext. 4 5209, k.gounaris@imperial.ac.uk)
<i>Course Tutor</i>	Mr. Vignesh (Kartik) Chundru (vignesh.chundru14@imperial.ac.uk)
<i>Course Representative</i>	Up to you (see note below)!

(Add 020 759 to extension numbers to call from external phones)

Each Masters course has one or more Course Representatives to represent the student body at Student-Staff meetings and act as a first point of contact if and when issues arise that need to be discussed with the Course Directors. MSc CMEE and MRes CMEE will each have one Representative. The student union will email you about an electronic election for the Reps.

1.2 Course Aims

Both MSc and MRes CMEE aim to:

- Provide training in key biological, statistical, computational, and mathematical topics and how they are integrated.

- Provide students with a background to computational biology with particular focus on theoretical and empirical approaches towards the study of ecological and evolutionary systems.
- Teach state-of-art, scientific computing techniques for the management and analyses of big data and simulation as well as mathematical modelling.
- Provide a thorough understanding of a range of modern techniques in bioinformatics, genomics and ecoinformatics.
- To show how these principles and skills can be applied to solve real world problems and make an informed choice of research topic.
- To prepare for PhD studies and other appropriate career paths in industry or NGOs focusing on any combinations of biology, maths and computing skills.

The main rationale for having an MSc as well as MRes CMEE is to give students the choice to find a right balance between learning technical skills and learning research skills. With this flexibility, students can choose to tailor a course and a project that best suits their personal interests and future plans.

1.3 Learning outcomes

Students will develop:

- Competence in computational methods required to address a range of topical 'real-world' questions in a range of ecological and evolutionary topics, including population biology, population genetics, genomics, and complex systems
- An ability to choose an appropriate quantitative method, in possibly including statistical and mechanistic mathematical modelling, for answering a particular biological question
- An ability to develop, analyse, and numerically simulate theoretical models for ecological and evolutionary systems and fit empirical data to these models

1.4 Transferable Skills

A central objective of the CMEE Masters' is to prepare students for PhD studies and/or a career in computational biology by teaching a suite of transferable skills, be it within academic institutions, government, or industry, by developing a professional approach towards developing and delivering high-quality science along with qualitative, critical thinking and problem solving skills. Students will learn a unique set of transferrable skills in computation relevant not just to biology but also other fields that involve large datasets and complex dynamics and patterns, such as economics, medicine, or sociology. Students will be able to:

- apply computational, statistical and modelling skills;
- communicate effectively through oral presentations, written reports and scientific publications;
- management skills: decision making, problem definition, project design and evaluation, risk management, teamwork and coordination;
- integrate and evaluate information from a variety of sources;
- transfer techniques and solutions from one discipline to another;
- use Information and Communications Technology;
- manage resources and time;
- learn independently with open-mindedness and critical enquiry;
- learn effectively for the purpose of continuing professional development;

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- depending upon choice of taught modules and research project, learn lab and field techniques.

The course objectives will be achieved by providing:

- A course of lectures, seminars and practicals within distinct modules, linked to cutting edge academic research and research groups in Silwood Park and collaborator institutions elsewhere. Both MSc and MRes students will attend the first 9 weeks of modules where they will learn fundamental concepts and techniques in computational biology, with particular focus on ecological and evolutionary theory, data, and systems.
- Hands-on experience of a wide repertoire of methods and techniques involved in the application of computational techniques to ecological and evolutionary systems.
- A research project (~9 months for MRes, ~5.5 months for MSc) based upon theoretical/analytical work, possibly including field and/or laboratory study, on an advanced and original topic from systems biology, ecology, evolution, conservation or another biological field, supervised by academics from either biological, computational or mathematical combinations of these in Silwood Park or elsewhere.
- Strategically timed workshops on publishing scientific manuscripts, selecting a research project, and selecting Job/PhD opportunities and applying to them.

1.5 Course activities and assessment overview

1.5.1 Assessment Overview

Component	MSc CMEE		MRes CMEE	
	% of Course	% of Component	% of Course	% of Component
Coursework				
<i>Computing</i>	13.75	55	13.75	55
<i>Python Long Practical</i>	5	20	5	20
<i>HPC Long Practical</i>	5	20	5	20
<i>Seminar Diary</i>	1.25	5	1.25	5
Coursework Total	25	100	25	100
Exams				
<i>Exam 1</i>	10	40	–	–
<i>Exam 2</i>	15	60	–	–
Exam Total	25	100	–	–
Project				
<i>Proposal defense (presentation + proposal)</i>	2.5	5	–	–
<i>Project progress defense (presentation + report)</i>	–	–	7.5	10
<i>Final Report + Presentation</i>	32.5	65	45	60
<i>Viva</i>	12.5	25	18.75	25
<i>Supervisor mark</i>	2.5	5	3.75	5
Project Total	50	100	75	100

1.5.2 Course structure overview

Activity	MSc CMEE	MRes CMEE
<i>Lectures + practicals, with assessment</i>	Required for first 17 weeks	Required for first 9 weeks, optional attendance in MSc modules within reason thereafter
<i>Exams</i>	Required	Not required
<i>Project report (Dissertation)</i>	Required	Required
<i>Seminars</i>	Required, seminar diary required for a minimum 16 weeks	Required, seminar diary required for a minimum 16 weeks
<i>Workshops</i>	All optional	All optional

1.5.3 MSc CMEE

MSc CMEE students will attend 17 weeks of taught modules over the Autumn and Spring Terms followed by a 5.5-month research project leading to a dissertation (Thesis report). There are three main components to the course assessment:

- i) *The coursework*: 25% of final mark, made up a number of components and modules, assessed by weekly problem sets and practical submissions as well as a seminar diary (see below)
- ii) *Two examinations*: 25% of final mark. January exam covers Autumn material (10%), April exam (15%), mainly covers Spring term material, but assumes knowledge of previous material
- iii) *The research project*: 50% of final mark, including mid-year project proposal defense (oral presentation + proposal) + final viva + thesis

1.5.4 MRes CMEE

MRes CMEE students will attend nine weeks of taught modules over the Autumn Term followed by a 9-month research project leading to a dissertation (thesis report). There are two main components to the course assessment:

- i) *The coursework*: 25% of final mark, made up a number of components and modules, assessed by weekly problem sets and practical submissions as well as a seminar diary (see below)
- ii) *The research project*: 75% of final mark, including project progress viva + project progress report + oral presentation + final viva + thesis report

1.6 External vivas and examiners

All students on both the MSc and MRes programmes will undertake a final 25 minute informal viva with one of the External Examiners, to be held between the internal project viva and the final meeting of the Board of Examiners. The dates are shown above (section 2.2). These external examiner vivas form a part of both the moderation process and oversight of the course by the External Examiners. The current External Examiners are:

Prof. Rob Freckleton	University of Sheffield
Prof. Andrew Leitch	Queen Mary University of London

1.7 Weekly Seminars and Seminar Diary

Two weekly seminars take place most weeks, usually on Monday and Thursday at 1pm in the Hamilton Building. Both MSc and MREs students must attend all the Thursday seminars unless told otherwise, even if one is rescheduled to a day other than Thursday. You will write a half-page on at least 16 of these seminars and create a “seminar journal” for submission as part of their coursework mark (see section on the Coursework Element).

Monday seminars are internal, informal, and intended for work in progress. Attendance is optional. However, they are a good way to practice critical thinking about research, since that is explicitly what the seminar speakers are hoping to get from the audience. Once you start your research project you are not only welcome to give one of these seminars, but also encouraged. It may be sensible to split the time of a one-hour seminar with another MSc/MRes student, effectively dividing two shorter seminars. You can get very good feedback from the community on your project by doing this. Sign-ups are online.

The seminar links are <http://www3.imperial.ac.uk/silwoodparkcampus/research/thursdayseminars> and <http://www3.imperial.ac.uk/silwoodparkcampus/research/mondayseminars>

The Seminar diary is due at the end of the Spring term, the week before your final (project) dissertation submission 2.2.

1.8 Workshops

We will organize a series of general skills workshops on certain Wednesday afternoons (see the timetable in section 2.2), such as those involved in choosing a project, writing skills, choosing and applying for PhD places, etc.

1.9 Summer School on Frontiers in Ecology and Evolution

This two to three-day summer school, practically an extended workshop, will be organized by the CMEE Masters students.

The FrEE Summer School will provide a relaxed set-up to discuss challenges in Ecological and Evolutionary research. It will also be a valuable opportunity to develop your organisational skills, from selecting, inviting and hosting speakers to dealing with the logistics of catering. A few external invitees will run forums on current ‘hot’ topics, such as ecological and evolutionary responses to climate change, deforestation and the global carbon cycle, responses of food webs and plant-pollinator networks to climatic warming, etc.

The focal research themes and techniques will depend on you and the projects you choose, the interests you develop, and anything you all want to learn about. You will learn how to engage and discuss with your peers to prioritize topics for workshops, and to invite speakers when necessary. In most cases, we expect that you will find personnel with the necessary skills right on campus, but you can invite people from outside when needed. Please discuss with the Course Directors and Administrator around the beginning of the Spring term.

1.10 Graduation

The postgraduate graduation ceremony for MSc and MRes students will be held in the Royal Albert Hall in South Kensington, London at the start of the following May. The exact dates are available at <http://www3.imperial.ac.uk/graduation/graduationtimetable>

1.11 Submission of work and penalties for late submission

Project reports and written coursework must be handed in as PDF documents. The method of submission will be announced in class, but will typically involve submission to a version-control (e.g., bitbucket) repository and/or to Blackboard. The final report will additionally have to be emailed to the Postgraduate Administrator and/or Course Director by the deadline. In real life, research reports, grant proposals and other outputs that are submitted late or that do not conform to instructions (e.g. word limits) would not be considered. Therefore, we will dock marks for late projects, using a standard penalty of 5% per day, and for written work not conforming to the stated requirements.

1.12 Teaching facilities

The majority of teaching will take place in the Hamilton building and in these locations. The locations of all teaching activities are given in the timetables below.

1.12.1 Lectures

Most lectures will be held in the Wallace lecture room on the ground floor of the Hamilton Building.

1.12.2 Lab work

The Field Laboratory is on the ground floor of the Hamilton building directly opposite the main entrance.

1.13 Work and desk spaces

Throughout the year, students are welcome to work in the Wallace room of the Hamilton building, which will be kept open 24 hours a day. However, the Hamilton building lobby closes at 7PM. After 7 PM, you can use the computer room — there are enough empty desk spaces to park your laptop and yourself.

Students can also work in the Library and the desks in the CPB common room, though the library has limited desk spaces, and CPB common room is often busy with people doing what people do in common rooms.

Project supervisors may also make desk space available to Masters students at their own discretion. Please feel free to ask your supervisor about a desk space in her/his lab.

1.13.1 Computing facilities and support

There are three levels of computing solutions available to students.

At the lowest and most immediately available level are student laptops, issued to students in the Autumn (and returned in September at the end of the course). A 64 bit Linux operating system (most likely Ubuntu 14.04) will be available on the laptops (that's all you should need for the course). Necessary software will be installed and more can be installed by the students as and when needed. Part of being a good quantitative/computational biologist is achieving a level of mastery of your computer and the software on it – these are your main tools. Much more information will be provided on using your computer in many ways, but you should also take it upon yourself to develop expertise in this area beyond what is taught. In addition, the Hamilton Computer Room is on the first floor of the Hamilton Building and provides 44 desktop computers (with Ubuntu on Virtual machines), charging and network points for laptops and printing facilities. Certain computer practicals will be held in the Hamilton Computer Room. Food and drink – other than drinks in sealed sports drink bottles – are not permitted in the computer room and you must abide by the College's Conditions of Use for IT Services, details of which are here: <http://www3.imperial.ac.uk/ict/newusers>

At the intermediate level are two local, multi-core linux machines in the Pawar lab, called “William” and “Harvey”. These make it relatively easy to learn to run large-ish parallel jobs. If you want access, please ask Samraat Pawar. You would be able to log onto one of them remotely and run single-core or multi-core simulations, statistical fitting, and other computing jobs, with a few rules or constraints to be followed out of respect for other users. William has 16 cores and Harvey has 12. They each have around 50Gb memory. Documentation for these can be found at <https://bitbucket.org/mhasoba/pawarlab/raw/8d814b4571d265d3939b84f9d4fb793e2c716fce/Computing/LabServers/LabServers.pdf>

At the highest level is the Imperial College High Performance Computing (HPC) Cluster, which puts over 10,000 cores at the disposal of the researcher. For most purposes in quantitative biology this is essentially infinite computing power, if you learn to harness it. You will be taught how to harness it during the CMEE HPC module.

ICT Computer support is available in Room 1.12 Hamilton Building (on the 1st floor) from 12.30 to 13.30 Mondays to Fridays during term. Online support is also available: <https://imperial.service-now.com/ict/>

1.13.2 Library facilities

The Michael Way Library at Silwood Park is on the first floor of the Hamilton Building. It maintain a specialised collection of books ranging from entomology, conservation, ecology, evolution and pest management to mathematical and computational biology, and holds a collection of textbooks for the Masters courses at Silwood: <http://www3.imperial.ac.uk/library/usethelibrary/silwood>

While many academic journals are available across the College as electronic journals, the Imperial College Library facilities in South Kensington also houses a wide collection of scientific literature, which may be requested through the Michael Way Library. The full range of library services and e-journals available are described at <http://www.imperial.ac.uk/admin-services/library/>.

1.13.3 Study guides

The College provides a booklet to help with Master's level study. The booklet contains advice, lessons and tips to help you to enhance your learning skills and get the very best from your degree. The booklet explain what you can expect from your study at Imperial, and will provide sources of support and guidance. You should receive a copy of the guide at the start of your course, but you can also download a copy from <http://www3.imperial.ac.uk/students/studyguide>

1.14 Prizes and Awards

1.14.1 The Illumina Harvey Prize

The student with highest final mark in MSc as well as MRes CMEE will be awarded a Illumina Harvey Prize each. Each award comes with a cash award of £500. But more importantly, it is an honour for a job very well done in a difficult course.

The cash prize is sponsored by Illumina, is a global company that develops innovative array-based solutions for DNA, RNA, and protein analysis; they are also some of the world-leaders in Next Generation DNA sequencing Technologies (www.Illumina.com).

The award is partly named after William Harvey (1578–1657), who provided a very early and very compelling example of combined quantitative and biological reasoning, leading to the first proof that blood circulates in the human body, and capillaries must exist connecting the arteries and veins in the circulatory system. Before the invention of the microscope, scientists were aware of arteries and veins but could not see the connections between them (capillaries), and so assumed they were not connected. They knew the heart was a pump, and dominant theories were that blood ebbed and flowed, back and forth like the tides in the arteries and veins, and that blood went out from the heart, was absorbed by the body, was regenerated in the liver which was thought to feed the veins, and the new blood flowed back to the heart. Harvey used biological reasoning, by observing that veins have valves, so blood can only flow through veins toward the heart, showing that the ebb-and-flow theory cannot be correct. Harvey used quantitative reasoning, by calculating the volume of the ventricles of the heart (about 1.5 imperial ounces) and the fraction of blood expelled from the ventricles on each heartbeat (about $\frac{1}{18}$), and thereby calculating that if your heart beats 1000 times every half hour, the liver would have to produce 540 pounds of blood in a day for the dominant theory to be correct. He then developed the hypothesis of capillaries and blood circulation, later confirmed by Harveys experimental work and by the invention of the microscope.

1.15 Background Reading

These books are recommended to all students for wading into ecological and evolutionary ways of thinking.

- Kingsland, S. E. 1995 Modeling nature. Episodes in the history of population ecology. 2nd edn. Chicago, Illinois: University of Chicago Press.
- Otto, S. and Day, T. 2007 A biologist's guide to mathematical modeling in ecology and evolution. New Jersey: Princeton University Press.
- Levins, R. 1966 The strategy of model building in population biology. Am. Sci. 54, 421–431.

Course Overview

- In addition, some might find a essay written by Samraat Pawar when he was a graduate student interesting!: https://dl.dropboxusercontent.com/u/44578955/The_Turbulent_Marriage01.pdf



2 Course details, timetables, and module descriptions

2.1 Teaching Staff

Dr Koenraad Audenaert	Quantum Dynamics and Information Theory (<i>Koenraad.Audenaert@rhul.ac.uk</i>)
Mr Tin-Yu Hui	Statistical genetics (<i>tin-yu.hui11@imperial.ac.uk</i>)
Prof Tim Barraclough	Evolution of species diversity; asexual evolution; experimental evolution. (<i>t.barracough@imperial.ac.uk</i>)
Prof Austin Burt	Evolutionary biology of selfish genetic elements (<i>a.burt@imperial.ac.uk</i>)
Dr Samraat Pawar	Theoretical Ecology; Ecological Systems Biology; Ecological Networks; Ecoinformatics; Metabolic theory and physiological ecology. (<i>s.pawar@imperial.ac.uk</i>)
Dr James Rosindell	Theoretical Ecology; Biodiversity theory; Island biogeography; Ecological neutral theory; Scientific data visualisation. (<i>j.rosindell@imperial.ac.uk</i>)
Dr Mike Tristram	The evolution of retroelements; molecular biology and phylogenetics. (<i>m.tristram@imperial.ac.uk</i>)
Dr Brian Hollis	Sexual selection and genomic conflict using experimental evolution (<i>b.hollis@imperial.ac.uk</i>)
Prof Vincent Jansen	Mathematical biology (01784 443179, <i>Vincent.Jansen@rhul.ac.uk</i>)
Dr Isabel Rosa	Stochastic models of land cover change (<i>i.rosa09@imperial.ac.uk</i>)
Dr Jason Hodgson	Evolutionary genomics and bioinformatics in primate evolution (<i>j.hodgson@imperial.ac.uk</i>)
Dr Marion Pfeifer	Conservation and land management strategies for tropical landscapes (<i>m.pfeifer@imperial.ac.uk</i>)
Dr Rob Ewers	Spatial patterns of forest biodiversity (<i>r.ewers@imperial.ac.uk</i>)
Dr Veronique Lefebvre	Mathematical models of dynamic ecological communities (<i>v.lefebvre@imperial.ac.uk</i>)
Prof Guy Woodward	Impacts of stressors on aquatic ecosystems. (<i>guy.woodward@imperial.ac.uk</i>)

2.2 Outline timetable and important dates

Week	Dates	MSc CMEE	MRes CMEE
Autumn term			
1	5 Oct		Induction
1	6–9 Oct		Foundations of Biological Computing
2	12–16 Oct		Biological Computing in Python I
3	19–23 Oct		Global biodiversity conservation and GIS
4	26–30 Oct		Introduction to R and Statistics
5	2–6 Nov		Biological Computing in R
6	9–13 Nov		Biological Computing in Python II
7	16–20 Nov		Genome evolution + Python Long Practical

Course details, timetables, and module descriptions

8	23–27 Nov	Genomics and Bioinformatics	
9	30 Nov–4 Dec	High Performance Computing	
10	7–11 Dec	Mathematics Primer I	Project and Workshops
11	14–18 Dec	Reading	
Spring term			
15	11–15 Jan	Exam 1	Project and Workshops
16	18–22 Jan	Mathematics Primer II	
17	25–29 Jan	Dynamical Models in Ecology	
18	1–5 Feb	Models in Population Genetics and Genomics	
19	8–12 Feb	Generalised linear modelling	
20	15–19 Feb	Maximum Likelihood Statistics and Model Fitting	
21	22–26 Feb	Introduction to Bayesian Statistics	
22	29 Feb–4 Mar	Networks and Complex Systems in Ecology and Evolution	
23	7–11 Mar	Reading	
24	14–18 Mar	Reading	
25	21–25 Mar	Exam 2	
26	28 Mar–1 Apr	Project and Workshops	
For later weeks and other dates see below			

Other important dates and deadlines:

Date	Activity/Item due
9 December, 1PM	MRes: Project proposal
12 February, 12AM	MSc, MRes: HPC Long Practical submission
8 March, 1PM	MSc, MRes: Python Long Practical submission
4 April, 1PM	MSc, Project proposal; MRes, Project progress report
8 April	MSc, Project proposal presentation; MRes: Project progress presentation
Week 3 of July	FrEE Summer School
29 Aug, 5pm	MSc, MRes: Seminar Diary Submission
1 Sep, 5pm	MSc, MRes: Final project report electronic submission
5 Sep, 5pm	MSc, MRes: Final project report hard copy submission
15–16 Sep	MSc, MRes: Silwood Masters Symposium (final project presentations)
19–21 Sep	MSc, MRes: Internal vivas
22–23 Sept	MSc, MRes: External Examiner vivas

Workshops:

In addition, there are a number of professional skill development workshops, the dates for which are given in the detailed time table below.

2.3 Taught Module Descriptions

All teaching activities are typically scheduled from 10.00am until 5.00pm, except Wednesday afternoon. The morning lectures in the modules/weeks on computing (UNIX/Linux, Python, R, etc.) are interactive, will all require you to use your laptop — so please bring your laptop to every class and session! The afternoon sessions of these modules will involve practicals, and you will typically be assisted by demonstrators (typically one per 6 students).

Also, please note that most if not all the recommended readings and resources in the following module descriptions are available in Central or Silwood Libraries, and often also as e-books.

2.3.1 Course Induction and Foundations of Computing

Week: 1 (*Monday 5th October – Friday 10th October*)

Convenor: Samraat Pawar

This module focuses on the fundamentals of scientific computing, including an introduction to UNIX and Linux, writing elegant documents using L^AT_EX, and version control. Along with technical training, we will introduce you to the philosophy of the course, and have a workshop on choosing research questions and projects. The week begins with a number of important induction events.

All lectures and practicals will be in Wallace unless indicated otherwise, and will be laptop-based.

Timetable:

Monday 5th October	
09:00 – 10:00	<i>Welcome to Silwood Park</i> (Induction, Fisher/Haldane)
10:00 – 11:00	<i>Computer and key issue, meet each other</i> (Wallace)
11:00 – 13:00	<i>Silwood treasure hunt</i> (Induction, Silwood Park, Tim Barraclough) In teams with other new Masters and PhDs, you will scour Silwood for answers to Tim Barracloughs fiendish geographically and ecologically based questions. Meet at the Hamilton building foyer and be prepared in case of bad weather.
13:00 – 14:00	<i>Buffet lunch</i> (Hamilton Foyer)
14:00 – 16:00	<i>Discussion: The Big Picture</i> (Hamilton Foyer, Rob Ewers) In small groups you will discuss and debate big environmental questions
Tuesday 6th October	
10:00 – 11:00	<i>Introduction to CMEE</i> (Wallace, Samraat Pawar and James Rosindell)
11:30 – 12:30	<i>Computer setup</i> (Wallace, Samraat Pawar)
13:30 – 16:30	<i>Intro to UNIX and Linux</i> (Wallace, Samraat Pawar)
17:00 – 20:00	<i>Welcome Reception</i> (CPB common room (floor 1 of CPB Bldg)) Introduction to student activities and people running them + nibbles and drinks. Prizes for the treasure hunt will be presented. Staff and students will attend to meet the new students.
Wednesday 7th October	
10:00 – 11:00	<i>How to get addicted to the terminal and shell scripting</i> (Lecture, Wallace, Samraat Pawar)
11:30 – 12:30	<i>How to get addicted to the terminal and shell scripting</i> (Lecture, Wallace, Samraat Pawar)
14:00 – 15:00	<i>Workshop: Choosing and designing a research project</i> (Wallace, Samraat Pawar)

Course details, timetables, and module descriptions

15:30 – 17:00	<i>Introduction to library services</i> (Introduction, Hamilton Computer Room, Elizabeth Killeen)
Thursday 8th October	
10:00 – 11:00	<i>Version control with Git</i> (Lecture, Wallace, Samraat Pawar)
11:30 – 12:30	<i>Scientific typesetting using L^AT_EX</i> (Lecture, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Git and L^AT_EX catch-up and practicals</i> (Practical, Wallace, Samraat Pawar)
Friday 9th October	
10:00 – 11:00	<i>Provost's Welcome</i> (Haldane, Prof. Alan Spivey)
11:00 – 12:00	<i>Professional skills development programme</i> (Haldane, Graduate School)
14:15 – 15:15	<i>Safety induction</i> (Introduction, Fisher, Stefan Hoyle)
15:30 – 16:30	<i>Project introductions</i> (Induction, Fisher, Joe Tobias)

Readings and Resources:

- There are LOTS OF UNIX tutorials out there. I really like the lectures you can find on <http://software-carpentry.org> (Chapter “shell”). Either watch the video tutorials or read the pdfs of the presentations.
- The Imperial College library provides you with access to several e-books on UNIX, some specific to Mac OSX or Ubuntu, and some more general. Go through the <http://www3.imperial.ac.uk/library> website.
- An extensive list of UNIX commands can be found here (along with their man page): www.oreillynet.com/linux/cmd/.
- <http://www.git-scm.com/book/en/v2> hosts a wonderful book on git. There are also several tutorials on the internet. I really like the approach taken by <http://www.sbf5.com/~cduan/technical/git/> and <https://www.atlassian.com/git/>
- (Health?) benefits of using latex: <http://www.andy-roberts.net/writing/latex/benefits>
- Word vs. L^AT_EX: http://openwetware.org/wiki/Word_vs._LaTeX
- Leslie Lamport, L^AT_EX: A document preparation system, users guide and reference manual, 1994, Addison-Wesley.
- Also see also myriad online resources for L^AT_EX, including [www.http://en.wikibooks.org/wiki/LaTeX/Introduction](http://en.wikibooks.org/wiki/LaTeX/Introduction), www.ctan.org/tex-archive/info/lshort/english/, and <http://ftp.uni-erlangen.de/mirrors/CTAN/info/lshort/english/lshort.pdf>
- Bibliographies in L^AT_EX: <https://schneider.ncifcrf.gov/latex.html>

2.3.2 Biological Computing in Python I

Week: 2 (*Monday 12th October – Friday 16th October*)

Convenor: Samraat Pawar

This is the first of a two-week module on biological computing in the Python language. With the profusion of genomic, environmental and ecological information, the ability to develop automated, reproducible analyses of massive datasets using computer scripts and programs is an essential skill for any aspiring research student. The aim of this module is to introduce the basics of programming using Python, which is a modern, easy-to-write, interpreted (semi-compiled) language that was conceived

with readability of script in mind. It has a feature-rich set of packages that can be used for a wide variety of applications and analyses. The approach will be hands-on and informal, involving lectures interspersed with short exercises in class. There will be longer exercises (and an opportunity for you to catch up!) during the last three hours at the end of each day.

Aims: To learn principles of computer program design and scientific computing, using Python. Specifically,

- Learn to write simple functions and programs in Python.
- Understand of the basics of Python program testing, debugging and documentation.
- Learn to use Python for retrieving, managing, and analyzing data from local and remote databases.
- Learn to automate file handling, string manipulation, and run shell scripts.
- Learn to use Python for efficient numerical analyses.
- Learn to run analyses by patching together R or R + Python scripts and functions.

All lectures and practicals will be in Wallace in the Hamilton Building, and will be laptop-based.

Timetable:

Monday 12th October	
10:00 – 11:00	<i>Why write computer programs?</i> (Lectures + Exercises, Wallace, Samraat Pawar)
11:30 – 12:30	<i>OK, but why program in Python?</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Programming in Python - basics</i> (Lectures + Short Practical, Wallace, Samraat Pawar)
Tuesday 13th October	
10:00 – 11:00	<i>Python basics continued</i> (Lectures + Exercises, Wallace room, Samraat Pawar)
11:30 – 12:00	<i>Python basics continued</i> (Lectures + Exercises, Wallace, Samraat Pawar)
13:30 – 17:00	<i>Python basics continued</i> (Lectures + Short Practical, Wallace, Samraat Pawar)
Wednesday 14th October	
10:00 – 11:00	<i>Writing Python Code</i> (Wallace, Samraat Pawar)
11:30 – 12:30	<i>Writing Python Code</i> (Lectures + Exercises, Wallace room, Samraat Pawar)
Thursday 15th October	
10:00 – 11:00	<i>Writing and testing Python functions</i> (Wallace, Samraat Pawar)
11:30 – 12:30	<i>Writing and testing Python functions</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Writing and testing Python functions</i> (Lectures + Practical, Wallace, Samraat Pawar)
Friday 16th October	
10:00 – 11:00	<i>Writing and testing Python functions</i> (Wallace, Samraat Pawar)
11:30 – 12:30	<i>Writing and testing Python functions</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Writing and testing Python functions</i> (Lectures + Practical, Wallace, Samraat Pawar)

Readings and Resources:

- Browse the Python tutorial: <https://docs.python.org/2/tutorial/>
- For functions and modules: www.learnPythonthehardway.org/book/ex40.html

- For IPython: <http://ipython.org/documentation.html> and <https://github.com/ipython/ipython/wiki?path=Cookbook>

2.3.3 Spatial Analyses and GIS

Week: 3 (*Monday 19th October – Friday 23rd October*)

Convenor: Rob Ewers

This week will teach key skills in using and handling GIS data, along with basic remote sensing to generate GIS data and the use of GIS data in land use change modelling. We will use the open source GIS program **QGIS** along with the land use change modelling software **StocModLCC**. We will look at creating and georeferencing both vector and raster data and how to use GIS tools to create a workflow to carry out simple analyses.

This week is shared with other MSc/MRes courses. Please bring your laptops to all Hamilton computer room sessions and use them instead of the desktops.

Timetable:

Monday 19th October

- | | |
|---------------|--|
| 10:00 – 11:00 | <i>Essentials of remote sensing</i> (Lecture, Fisher/Haldane, Marion Pfeifer) |
| 11:00 – 12:00 | <i>Accessing satellite images and earth observation products</i> (Lecture, Fisher/Haldane, Marion Pfeifer) |
| 13:00 – 16:00 | <i>Downloading and processing imagery</i> (Practicals, Hamilton Computer Room, Marion Pfeifer) |

Tuesday 20th October

- | | |
|---------------|---|
| 10:00 – 11:00 | <i>Validation in earth observation maps</i> (Lecture, Fisher/Haldane, Marion Pfeifer) |
| 11:00 – 12:00 | <i>Linking field data to satellite data</i> (Lecture, Fisher/Haldane, Marion Pfeifer) |
| 14:00 – 17:00 | <i>Upscaling and mapping</i> (Practicals, Hamilton Computer Room, Marion Pfeifer) |

Wednesday 21st October

- | | |
|---------------|---|
| 10:00 – 11:00 | <i>Geographic Information Systems</i> (Lecture, Fisher/Haldane, Rob Ewers) |
| 11:00 – 12:00 | <i>Modelling species distributions</i> (Lecture, Fisher/Haldane, Rob Ewers) |

Thursday 22nd October

- | | |
|---------------|--|
| 10:00 – 11:00 | <i>Essentials of land use change modelling</i> (Lecture, Fisher/Haldane, Isabel Rosa) |
| 11:00 – 12:00 | <i>Types of land use change models</i> (Lecture, Fisher/Haldane, Isabel Rosa) |
| 13:00 – 14:00 | <i>Departmental Seminar</i> (Lecture, Haldane, Visiting staff) |
| 14:00 – 16:00 | <i>Land use change data processing and model calibration</i> (Practicals, Hamilton Computer Room, Isabel Rosa) |

Friday 23rd October

- | | |
|---------------|--|
| 10:00 – 11:00 | <i>Validation of land use change models</i> (Lecture, Fisher/Haldane, Isabel Rosa) |
| 11:00 – 12:00 | <i>Scenario building</i> (Lecture, Fisher/Haldane, Isabel Rosa) |
| 13:00 – 15:00 | <i>Projecting future forest cover change</i> (Practicals, Hamilton Computer Room, Isabel Rosa) |

Readings and Resources:

- *GIS overview*: Longley, PA (2011) Geographical information systems and science. Wiley.
- *Land use change modelling*: Rosa et al. (2013) Predictive modelling of contagious deforestation in the Brazilian Amazon. PLoS ONE 8:e77231.

- *Coordinate systems*: Van Sickle, G (2010) Basic GIS coordinates. CRC Press
<https://www.dawsonera.com/abstract/9781420092325>

2.3.4 Introduction to R and Statistics

Week: 4 (*Monday 26th October – Friday 30th October*)

Convenor: Tim Barraclough

In this module, we will introduce the use of the **open source statistical program R** for research and to review a core set of statistical methods that are of wide use in research projects.

We will cover:

- The difference between response and explanatory variables and between ordinal, categorical and continuous variables;
- The underlying structure of statistical testing using both parametric and non-parametric approaches;
- Tests for assessing differences between samples and correlation between samples;
- Analysis of categorical data;
- Fitting and assessing linear models of continuous response variables.

We will build on the R learnt in this week in a second, more R-focused week (see below).

This week is shared with other Masters courses. Please bring laptops to all computer room sessions and use them instead of the desktops.

Timetable:

Monday 26th October

09:30 – 10:30	<i>Introduction to R</i> (Lecture, Fisher/Haldane, Tim Barraclough)
10:30 – 13:00	<i>Loading and exploring data</i> (Practicals, Hamilton Computer Room, Tim Barraclough)
13:30 – 14:30	<i>Probability and p-values</i> (Lecture, Fisher/Haldane, Tim Barraclough)
14:30 – 17:00	<i>More loading and exploring data</i> (Practicals, Hamilton Computer Room, Tim Barraclough)

Tuesday 27th October

09:30 – 10:30	<i>Categorical data and analysis</i> (Lecture, Fisher/Haldane, Tim Barraclough)
10:30 – 13:00	<i>Chi squared and inter-rater reliability</i> (Practicals, Hamilton Computer Room, Tim Barraclough)
13:30 – 14:30	<i>One sample tests</i> (Lecture, Fisher/Haldane, Tim Barraclough)
14:30 – 17:00	<i>Wilcoxon and t-tests</i> (Practicals, Hamilton Computer Room, Tim Barraclough)

Wednesday 28th October

09:30 – 10:30	<i>Two sample tests and correlations</i> (Lecture, Fisher/Haldane, Tim Barraclough)
10:30 – 13:00	<i>Mann-Whitney, t-tests and correlation coefficients</i> (Practicals, Hamilton Computer Room, Tim Barraclough)
14:00 – 16:00	<i>MasterClass: Stress Management</i> (Workshop, Haldane, Graduate School) This workshop is provided by the Graduate School and you may need to reserve a place on the course through their website http://www3.imperial.ac.uk/graduateschool/currentstudents/professionalskillsmasters

Thursday 29th October

09:30 – 10:30	<i>Simple linear models</i> (Lecture, Fisher/Haldane, Tim Barraclough)
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10:30 – 13:00	<i>Regression and analysis of variance</i> (Practicals, Hamilton Computer Room, Tim Barraclough)
13:00 – 14:00	<i>Departmental Seminar</i> (Lecture, Haldane, Visiting staff)
14:00 – 15:00	<i>Extended linear models</i> (Lecture, Fisher/Haldane, Tim Barraclough)
15:00 – 17:30	<i>Analysis of covariance and more explanatory variables</i> (Practicals, Hamilton Computer Room, Tim Barraclough)

Friday 30th October

09:30 – 10:30	<i>Model criticism and simplification</i> (Lecture, Fisher/Haldane, Tim Barraclough)
10:30 – 13:00	<i>Residuals and the minimum adequate model</i> (Practicals, Hamilton Computer Room, Tim Barraclough)
13:30 – 14:30	<i>Here be dragons - stats in the wild</i> (Lecture, Fisher/Haldane, Tim Barraclough)
14:30 – 17:00	<i>Statistics practice session</i> (Practicals, Hamilton Computer Room, Tim Barraclough)

Readings and Resources:

- Hilborn, R. and Mangel, M., *The Ecological Detective: Confronting Models with Data*, Princeton University Press, 1997.
- Ben Bolker's "Ecological Models and Data in R" is also very good.
- Crawley, MJ (2005) *Statistics: An Introduction Using R*. John Wiley.
- Crawley, MJ (2012) *The R Book*. John Wiley.
<http://imperial.eblib.com/patron/FullRecord.aspx?p=1120574>.
- The Use R! series (the yellow books) by Springer are really good. In particular, consider: "A Beginner's Guide to R", "R by Example", "Numerical Ecology With R", "ggplot2" (we'll see this in another week), "A Primer of Ecology with R", "Nonlinear Regression with R", "Analysis of Phylogenetics and Evolution with R".

2.3.5 Biological Computing in R

Week: 5 (*Monday 2nd November – Friday 6th November*)

Convenor: Samraat Pawar

In this module, we will build upon the introduction to R you received in the Introduction to R and statistics to learn how to use this freely available statistical software with strong programming capabilities. R has become tremendously popular in Biology due to several factors: (i) many packages are available to perform all sorts of statistical and mathematical analysis, (ii) it can produce beautiful graphics, and (iii) it has a very good support for matrix-algebra (you might not know it, but you use it!).

So with R, you have an expanded and versatile suite of biological computing tools at your fingertips, especially for automating statistical analysis and the generation of figures. Therefore, R should become an indispensable component of your biological research workflow.

Aims:

- To learn how to use R for tasks ranging from data exploration and visualization to producing elegant, publication quality graphics.

- To learn R data types and structures and control flows.
- To learn how to write and debug efficient R scripts and functions.
- Learn how to use R packages. and generate elegant graphics.

This module is taught across multiple Silwood and NHM masters courses. Classes will be run in two repeated sessions. The timetable of sessions for different groups will be announced the week before the module – All CMEE students will be in either Group A or B (we won't split up the group!).

Timetable:

Monday 2nd November	
09:00 – 09:30	<i>Why R? Why us? (Group A+B)</i> (Lecture, Fisher/Haldane, Samraat Pawar) Intro to R and the week's module, R vs. other languages
09:30 – 11:00	<i>Easing into R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Easing into R!
11:00 – 12:30	<i>Useful R commands and functions (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Useful R commands and functions
13:30 – 15:00	<i>Easing into R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Easing into R!
15:30 – 17:00	<i>Useful R commands and functions (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Useful R commands and functions
Tuesday 3rd November	
09:00 – 10:30	<i>Programming in R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing functions, programs, debugging in R
11:00 – 12:30	<i>Programming in R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing functions, programs, debugging in R
13:30 – 15:00	<i>Programming in R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing functions, programs, debugging in R
15:30 – 17:00	<i>Programming in R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing functions, programs, debugging in R
Wednesday 4th November	
09:00 – 10:30	<i>Numerical R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Handling Data and Numerical analyses in R
11:00 – 12:30	<i>Numerical R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Handling Data and Numerical analyses in R
Thursday 5th November	
09:00 – 10:30	<i>Efficient R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing efficient programs in R
11:00 – 12:30	<i>Efficient R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing efficient programs in R
13:00 – 14:00	<i>Departmental Seminar</i> (Lecture, Haldane, Visiting staff)
14:00 – 15:30	<i>Efficient R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing efficient programs in R
15:30 – 17:00	<i>Efficient R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Writing efficient programs in R
Friday 6th November	
09:00 – 10:30	<i>Graphics in R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Graphics in R
11:00 – 12:30	<i>Advanced graphics in R (Group B)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) ggplot!

13:30 – 15:00	<i>Graphics in R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) Graphics in R
15:30 – 17:00	<i>Advanced graphics in R (Group A)</i> (Lectures + exercises, Hamilton Computer Room, Samraat Pawar) ggplot!

Readings and Resources:

- The Use R! series (the yellow books) by Springer are really good. In particular, consider: ‘A Beginner’s Guide to R’, ‘R by Example’, ‘Numerical Ecology With R’, ‘ggplot2’ (we’ll see this in another week), ‘A Primer of Ecology with R’, ‘Nonlinear Regression with R’, ‘Analysis of Phylogenetics and Evolution with R’.
- Ben Bolker’s ‘Ecological Models and Data in R’ is also very good.
- For more focus on dynamical models: Soetaert & Herman. 2009 ‘A practical guide to ecological modelling: using R as a simulation platform’.
- There are excellent websites. Besides [CRAN](#) (containing all sorts of guides and manuals), you should check out www.statmethods.net and en.wikibooks.org/wiki/R.Programming and google ‘R Graph Gallery’ for various sites showing graphing options and code.

2.3.6 Biological Computing in Python II

Week: 6 (*Monday 09th – Friday 13th November*)

Convenor: Samraat Pawar

This is the second of the two-week module on biological computing in the Python language. The aims, format, and venue of the lectures and practicals are same as same as *Biological computing in Python I*.

Timetable:

Monday 9th November	
10:00 – 11:00	<i>Profiling and debugging in Python</i> (Lectures + Exercises, Wallace, Samraat Pawar)
11:30 – 12:30	<i>Profiling and debugging in Python</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Profiling and debugging in Python</i> (Lectures + Short Practicals, Wallace, Samraat Pawar)
Tuesday 10th November	
10:00 – 11:00	<i>Useful Python packages and tools</i> (Wallace, Samraat Pawar)
11:30 – 12:30	<i>Useful Python packages and tools</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Useful Python packages and tools</i> (Lectures + Short Practicals, Wallace, Samraat Pawar)
Wednesday 11th November	
10:00 – 11:00	<i>Scipy and scientific computing (number crunching!) in Python</i> (Lectures + Exercises, Wallace, Samraat Pawar)
11:30 – 12:30	<i>Scipy and scientific computing (number crunching!) in Python</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:30 – 16:00	<i>Workshop: Applying for PhD positions / Academic Jobs / Industry Jobs</i> (Haldane room, Samraat Pawar)

Thursday 12th November)

10:00 – 11:00	<i>Databases and Python</i> (Lectures + Exercises, Wallace, Samraat Pawar)
11:30 – 12:30	<i>Databases and Python</i> (Lectures + Exercises, Wallace room, Samraat Pawar)
14:00 – 17:00	<i>Databases and Python</i> (Lectures + Practicals, Wallace room, Samraat Pawar)

Friday 13th November)

10:00 – 11:00	<i>Build and run your workflow with Python</i> (Wallace room, Samraat Pawar)
11:30 – 12:30	<i>Build and run your workflow with Python</i> (Lectures + Exercises, Wallace, Samraat Pawar)
14:00 – 17:00	<i>Build and run... and intro to Python Long Practical</i> (Practicals, Wallace, Samraat Pawar)

Readings and Resources:

- www.docs.python.org/2/howto/regex.html
- Googles short class on regex in Python: www.code.google.com/edu/languages/ and www.google-python-class/regular-expressions.html
- www.regular-expressions.info has a good intro, tips and a great array of canned solutions
- For SciPy and Matplotlib, the official documentation is best: www.docs.scipy.org/doc/scipy/reference/ and www.http://matplotlib.org/
- “The Definitive Guide to SQLite” is a pretty complete guide and freely available from <http://evalenzu.mat.utfsn.cl/Docencia/2012/SQLite.pdf>

2.3.7 Genome evolution and selfish genetic elements + Python Long Practical

Module: 7 (Monday 16th November – Friday 20th November)

Convenor: Mike Tristem and Austin Burt

This module investigates genomes and genome evolution. The first half concentrates on bioinformatic approaches to deal with the huge amounts of sequence data being generated. The second half looks at the evolutionary biology of a particularly interesting category of genes, namely selfish genetic elements that are inherited in a biased manner. This will include a discussion of their possible application in combating human vector-borne diseases.

The computing practicals of this week will not be assessed, but you may participate in them if you want. This module is shared with MSc EEC – you should bring your laptops to the practicals, but you might have to use (god forbid) applications in Windows!

Ideally, you should aim to spend significant periods of time in this week working on your Python Long Practical, which was given to you in the previous week.

Timetable:

Monday 16th November

Course details, timetables, and module descriptions

10:00 – 11:00	<i>Genome-composition and architecture</i> (Lecture, Fisher, Mike Tristem) Genome size and the C-value paradox, genome organisation, highly repetitive DNA, mini and micro satellites, selfish genetic elements and their contribution to total genome size. Evolution of non-genic and repetitive DNA, change in genome size via ployploidy, chromosomal duplication, replication slippage, unequal crossing over, transposition and other mechanisms, origin and evolution of introns.
11:30 – 12:30	<i>Evolution of multi-gene families</i> (Lecture, Fisher, Mike Tristem) Evolution of dispersed and tandemly repeated gene families. Gene duplication, gene conversion and concerted evolution, evolution of globins, rDNA genes and hox genes, evolution and detection of pseudogenes.
14:00 – 15:00	<i>Model genomes and comparative genomics</i> (Lecture, Fisher, Mike Tristem) Model bacterial and eukaryotic genomes. Origin of life, “evolution of the minimum genome”. Uses and applications of comparative genomics.

Tuesday 17th November

10:00 – 11:00	<i>Genomic technologies and genomic resources.</i> (Lecture, Fisher, Mike Tristem) DNA microarrays and gene expression, SNPs and genotyping, proteomics, detecting genetic variation in humans, population markers, pharmacogenomics, Genomic resources, the nature and types of genomic information, the human genome project and its associated databases, genetic maps, cytogenetic maps, morbidity maps, OMIM, Unigene.
11:30 – 12:30	<i>Bioinformatics and genome biology</i> (Lecture, Fisher, Mike Tristem) Genomic scale sequencing and assembly (Sanger sequencing, BACS and YACS, automated sequencers, components and contigs), completed and ongoing genome projects, data mining tools, BLAST, FASTA.
14:00 – 17:00	<i>The human and chimpanzee genome projects</i> (Practicals, Hamilton Computer Room, Mike Tristem) Bioinformatics based practical that will investigate (and then compare and contrast) the human and chimpanzee genome projects at NCBI and ENSEMBL. The practical will include the use of BLAST, map options, and Locuslink to obtain information on specific genes in the human database.

Wednesday 18th November

14:00 – 16:00	<i>MasterClass: Academic Writing Skills</i> (Workshop, Haldane, Graduate School) This workshop is provided by the Graduate School and you may need to reserve a place on the course through their website http://www3.imperial.ac.uk/graduateschool/currentstudents/professionalskillsmasters
16:00 – 17:00	<i>MasterClass: Informational Posters</i> (Workshop, Haldane, Graduate School) This workshop is provided by the Graduate School and you may need to reserve a place on the course through their website http://www3.imperial.ac.uk/graduateschool/currentstudents/professionalskillsmasters

Thursday 19th November

10:00 – 11:00	<i>Transposable elements and retroviruses 1</i> (Lecture, Fisher, Mike Tristem)
11:30 – 12:30	<i>Transposable elements and retroviruses 2</i> (Lecture, Fisher, Mike Tristem)
13:00 – 14:00	<i>Departmental Seminar</i> (Lecture, Haldane, Visiting staff)
14:00 – 16:00	<i>Practical to be confirmed</i> (Practicals, Hamilton Computer Room, Mike Tristem) To be confirmed

Friday 20th November

10:00 – 11:00	<i>Homing endonuclease genes and population genetic engineering 1</i> (Lecture, Fisher, Austin Burt) Homing endonuclease genes (HEGs) are a particularly simple class of selfish genetic element. The potential use of these genes for controlling insect pests will be described, followed by a discussion about the pros and cons of this approach.
11:30 – 12:30	<i>Homing endonuclease genes and population genetic engineering 2</i> (Lecture, Fisher, Austin Burt)
14:00 – 16:00	<i>Ethics of genetic modification and vector control</i> (Discussion, Fisher, Austin Burt)

Readings and resources:

- Lesk, A. M. 2002. Introduction to bioinformatics. Oxford University Press.
- Orengo, C. 2003. Bioinformatics: genes, proteins and computers. Oxford
- Belshaw, R., V. Pereira, A. Katzourakis, G. Talbot, J. Paces, A. Burt, and M. Tristem. 2004. Long-term reinfection of the human genome by endogenous retroviruses. *PNAS* 101:4894-4849.
- Human Genome Consortium. 2001. Initial sequencing and analysis of the human genome. *Nature*. 409:860-921.
- Page, R. D. M. & E. C. Holmes. 1998. Molecular evolution: A phylogenetic approach. Blackwell, Oxford.
- Burt, A. & Trivers, R. 1006. Genes in Conflict. Harvard University press. Cambridge, Mass.
- Camacho, J.P.M. (Ed.) 2004. B Chromosomes in the Eukaryote Genome. Karger, Basel.
- Futuyma, D. 1998 Evolutionary Biology (3rd edition). Sinauer Associates, Inc. Mass.
- Graur, D. & W-H Li. 1999. Fundamentals of Molecular Evolution. Sinauer Associates, Inc. Mass.
- Hurst, L. D., Atlan, A. & Bengtsson, B. O. 1996. Genetic conflicts. *Quarterly Review of Biology*, 71, 317-364.
- <http://www.ncbi.nlm.nih.gov>
- <http://genome.ucsc.edu>

2.3.8 Genomics and Bioinformatics

Week: 8 (*Monday 23rd November – Friday 27th November*)

Convenor: Jason Hodgson

Genetic data contain information about who organisms are, their relationships to other organisms, their population histories, and their histories of adaptation. Thus, genetic data and genetic techniques are central to addressing many questions in evolution, ecology, and conservation. New technologies allow for genetic characterization at the genomic level, and these data allow for an understanding of population processes at resolutions not possible in the past. The goal of this module is to introduce students to the types of questions that can be addressed with genomic data, and the methodologies that are available for answering these questions. Learning will be accomplished through a mix of lectures, computer practicals and group discussions.

Course details, timetables, and module descriptions

This module aims to provide an understanding of:

- genomic data collection methods, and how to choose the data collection technique most appropriate to your question.
- the wealth of data available to biologists in public genomic databases.
- how genetic structure develops within and between populations, how to characterise it, and how to interpret the results of common analyses such as STRUCTURE and PCA.
- how demographic history affects genomic variation, and how to infer past population expansions and contractions from genomic data.
- how migration affects genomic variation, and how patterns of gene flow can be inferred from genomic data.
- how natural selection affects genomic variation, and how selection can be identified from genomic data.
- how phylogenetic relationships among species can be inferred, and what this information can tell us about evolution and conservation efforts.

This week is shared with MSc/MRes EEC. Please bring laptops to all computer room sessions and use them instead of the desktops.

Timetable:

Monday 23rd November

- | | |
|---------------|---|
| 10:00 – 11:00 | <i>Intro</i> (Lecture, Haldane, Jason Hodgson) Introduction to genomics and genomic data |
| 11:30 – 12:30 | <i>Population Structure</i> (Lecture, Haldane, Jason Hodgson) How and why genetic structure develops within species, and how to test for it. |
| 13:30 – 16:30 | <i>Data and databases practical</i> (Practicals, Hamilton Computer Room, Jason Hodgson) Practical introducing students to sources of genomic data, and how to use them. |

Tuesday 24th November

- | | |
|---------------|---|
| 10:00 – 11:00 | <i>Demographic history</i> (Lecture, Haldane, Jason Hodgson) The effects of demography on the genome and methods for inferring demography from genomic data. |
| 11:30 – 12:30 | <i>Gene flow and migration</i> (Lecture, Haldane, Jason Hodgson) The genomic effects of gene flow and migration and methods for inferring migration from genomic data. |
| 13:30 – 16:30 | <i>Population structure and migration practical</i> (Practicals, Hamilton Computer Room, Jason Hodgson) Students will analyze a dataset to assess population structure and test for migration |

Wednesday 25th November

- | | |
|---------------|---|
| 09:30 – 10:30 | <i>Natural Selection</i> (Lecture, Haldane, Jason Hodgson) Genomic signals of natural selection and methods for detecting natural selection |
| 11:00 – 12:00 | <i>Genomic approaches to ecology and conservation</i> (Lecture, Haldane, Jason Hodgson) Genomic methods for understanding ecological and conservation problems. |

Thursday 26th November

- | | |
|---------------|--|
| 10:00 – 11:00 | <i>Phylogenomics</i> (Lecture, Haldane, Jason Hodgson) Introduction to phylogenomics and phylogenomic techniques |
| 11:30 – 12:30 | <i>Detecting selection practical</i> (Practicals, Hamilton Computer Room, Jason Hodgson) Students will analyze a data set to identify regions that may have been subject to recent natural selection |
| 13:00 – 14:00 | <i>Departmental Seminar</i> (Lecture, Haldane, Visiting staff) |

14:30 – 17:00 *Phylogenomics practical* (Practicals, Hamilton Computer Room, Jason Hodgson)
Students will construct and date a phylogenetic tree.

Friday 27th November

10:30 – 11:30 *Study design discussion* (Discussion, Haldane, Jason Hodgson) In small groups students will be given a research question and tasked with designing an appropriate study.

13:30 – 15:30 *Study design presentations* (Discussion, Haldane, Jason Hodgson) Groups will present their study, and field questions from the audience.

Readings and resources: *Population structure*

- Novembre, J. & Ramachandran, S. Perspectives on human population structure at the cusp of the sequencing era. *Annual review of genomics and human genetics* 12, 245-274 (2011).
- Pritchard, J. K., Stephens, M. & Donnelly, P. Inference of population structure using multilocus genotype data. *Genetics* 155, 945-959 (2000).

Demography

- Bertorelle, G., Benazzo, A. & Mona, S. ABC as a flexible framework to estimate demography over space and time: some cons, many pros. *Molecular ecology* 19, 2609-2625 (2010).
- Li, H. & Durbin, R. Inference of human population history from individual whole-genome sequences. *Nature* 475, 493-496 (2011).

2.3.9 High Performance Computing

Week: 9 (*Monday 30th November – Friday 4th December*)

Convenor: James Rosindell

The use of high performance computing is becoming increasingly important in biology. For certain computational tasks we can use large numbers of CPUs in parallel to get numerical results in days that would otherwise have taken years. This module will introduce students to the tools and techniques of high performance computing for biological problems using R. It will also be a good opportunity to develop practical programming skills in the R programming language. However, there will also be a Python HPC session at the end of the week.

This module has a greater proportion of practical than usual because the best way to learn about programming is to try it, the convener will help students individually and be available to answer questions throughout all practical sessions. The biological topics covered will include individual based models, ecological neutral theory and fractals in biology but the techniques learned will be useful much more generally. A significant number of CMEE students typically end up using high performance computing as a tool for their research projects.

Timetable:

Monday 30th November

10:00 – 11:00	<i>Introduction and individual based models</i> (Lecture, Wallace, James Rosindell)
11:30 – 12:30	<i>Introduction and individual based models</i> (Lecture, Wallace, James Rosindell)
14:00 – 17:00	<i>IB models Practical</i> (Practicals, Wallace, James Rosindell)

Tuesday 1st December

Course details, timetables, and module descriptions

10:00 – 11:00	<i>Using HPC</i> (Lecture, Wallace, James Rosindell)
11:30 – 12:30	<i>Using HPC</i> (Practicals, Wallace, James Rosindell)
14:00 – 17:00	<i>Using HPC</i> (Practicals, Wallace, James Rosindell)
Wednesday 2nd December	
10:00 – 11:00	<i>Coalescence methods</i> (Lecture, Wallace, James Rosindell)
11:30 – 12:30	<i>Coalescence methods</i> (Practicals, Wallace, James Rosindell)
Thursday 3rd December	
10:00 – 11:00	<i>Fractal geometry</i> (Lecture, Wallace, Samraat Pawar)
11:30 – 12:30	<i>Fractal geometry</i> (Practicals, Wallace, James Rosindell)
14:00 – 17:00	<i>Fractal geometry</i> (Practicals, Wallace, James Rosindell)
Friday 4th December	
10:00 – 11:00	<i>Fractals in nature</i> (Lecture, Wallace, James Rosindell)
11:30 – 12:30	<i>Fractals in nature</i> (Practicals, Wallace, James Rosindell)
14:00 – 17:00	<i>HPC with Python</i> (Lecture+Practicals, Wallace, Samraat Pawar)

Readings and Resources:

- The best preparation for this module is to be well practiced at programming, especially in R.

2.3.10 Mathematics Primers I – II

Week: 10 (*Monday 7th – Friday 11th December*) and 16 (*Monday 18th – Friday 22nd January*)

Convenor: Koenraad Audenaert

This is a two-week module. The aim is to provide a primer in core mathematical topics that will be of use in the rest of this course and indeed, hopefully, throughout students careers! You will receive an introduction to the most fundamental applied concepts of calculus, linear algebra, probability theory, and dynamical systems, and will become comfortable to receive more in depth training in these fields. The first two parts of the module will be in the Autumn term, and the third part in the Spring.

Scheduling for this module in all three weeks will be (all activities in Wallace):

10:00–11:00: Lecture

11:30–12:30: Lecture

1400–1700: Practicals, except Wednesday

Readings and Resources:

- Sarah P Otto and Troy Day, *A Biologists Guide to Mathematical Modelling in Ecology and Evolution*, Princeton University Press, 2007.
- C. Neuhauser, *Calculus for Biology and Medicine*, 3rd edition (2009)
- D. C. Lay, *Linear Algebra and its Applications*, 4th edition (2010)
- *Linear Algebra*, Schaum's Outlines, 4th edition (2008)
- Any of numerous basic texts in calculus, linear algebra, and probability theory. There are many of these and students prefer different ones, so try a few and choose one. many are available in the Silwood or Central libraries.

2.3.11 Dynamical Models in Ecology

Week: 17 (*Monday 25th – Friday 29th January*)

Convenor: Vincent Jansen

Dynamical systems theory plays a major role in modern theoretical approaches to ecological concepts and phenomena such as competition, predation, metapopulation dynamics, diversity and evolution, and disease spread. This module will introduce some of the key basics of dynamical systems theory in application to these topics. We will look at ordinary differential equations and difference equation models and will use stability analysis and bifurcation analysis as tools to understand the qualitative behaviour of ecological models.

Scheduling for this module will be (all activities in Wallace): 10:00–11:00: Lecture 11:30–12:30: Lecture 1400–1700: Practicals, except Wednesday

Readings and Resources:

- Sarah P Otto and Troy Day, *A Biologists Guide to Mathematical Modelling in Ecology and Evolution*, Princeton University Press, 2007.
- May, R.M. & McLean, A. 2007 *Theoretical Ecology*. Blackwell Scientific. 5 copies in the library.
- Nicholas Gotelli, *A Primer of Ecology*, Sinauer Associates, 4th Edn, 2008.
- Bart Ermentrout: *Simulating, Analyzing, and Animating Dynamical Systems: A Guide to XPPAUT for Researchers and Students*. SIAM. 2002
- Bifurcation analyses using Python <http://www.ni.gsu.edu/~rclewley/PyDSTool/FrontPage.html>

2.3.12 Models in Population Genetics and Genomics

Week: 18 (*Monday 1st February – Friday 5th February*)

Convenors: Austin Burt and Timothy Barraclough

This module will give an introduction to the classic models in population genetics that have been used to study the action and interaction of mutation, drift, migration selection, and transmission ratio distortion. The theory will be integrated with practicals involving microbial population genetics/genomics data.

Scheduling for this module will be (all activities in Wallace): 10:00–11:00: Lecture 11:30–12:30: Lecture 1400–1700: Practicals, except Wednesday

Readings and Resources:

- TBD

2.3.13 Generalised linear modelling

Week: 19 (*Monday 8th February – Friday 12th February*)

Convenor: Brian Hollis

Course details, timetables, and module descriptions

This module builds on the basic linear models and R introduced in the previous term to introduce some key concepts that allow linear models to be applied to a wider range of research problems. This will include using generalised linear models to handle count and binomial data – where residuals are not expected to follow a normal distribution – and the use of structured models to allow for nonindependence in data and to control for known sources of variation in data.

Timetable:

Sunday 8th February	
10:00 – 11:00	<i>Likelihood methods</i> (Lecture, Fisher, Brian Hollis)
11:00 – 13:00	<i>Likelihood, deviance, and AIC</i> (Practicals, Hamilton Computer Room, Brian Hollis)
14:00 – 15:00	<i>Count data and the poisson distribution</i> (Lecture, Fisher, Brian Hollis)
15:00 – 17:00	<i>Generalised linear models</i> (Practicals, Hamilton Computer Room, Brian Hollis)
Monday 9th February	
10:00 – 11:00	<i>Proportions and the binomial distribution</i> (Lecture, Fisher, Brian Hollis)
11:00 – 13:00	<i>Generalised linear models</i> (Practicals, Hamilton Computer Room, Brian Hollis)
14:00 – 15:00	<i>Contrasts in linear models</i> (Lecture, Fisher, Brian Hollis)
15:00 – 17:00	<i>Setting contrast structure</i> (Practicals, Hamilton Computer Room, Brian Hollis)
Tuesday 10th February	
10:00 – 11:00	<i>Dispersion</i> (Lecture, Fisher, Brian Hollis)
11:00 – 13:00	<i>Quasi-poisson and quasi-binomial models</i> (Practicals, Hamilton Computer Room, Brian Hollis)
Wednesday 11th February	
10:00 – 11:00	<i>Variance revisited</i> (Lecture, Fisher, Brian Hollis)
11:00 – 13:00	<i>Nested analysis of variance</i> (Practicals, Hamilton Computer Room, Brian Hollis)
14:00 – 15:00	<i>Fixed and random effects</i> (Lecture, Fisher, Brian Hollis)
15:00 – 17:00	<i>Mixed effects modeling</i> (Practicals, Hamilton Computer Room, Brian Hollis)
Thursday 12th February	
10:00 – 11:00	<i>Generalised linear mixed models</i> (Lecture, Fisher, Brian Hollis)
11:00 – 13:00	<i>Working with GLMMs</i> (Practicals, Hamilton Computer Room, Brian Hollis)
14:00 – 15:00	<i>Generalised linear mixed models</i> (Lecture, Fisher, Brian Hollis)
15:00 – 17:00	<i>Working with GLMMs</i> (Practicals, Hamilton Computer Room, Brian Hollis)

Readings and resources:

- Venables, WN & Ripley, BD (2002) *Modern Applied Statistics with S* (4th Ed). Springer
- Pinheiro, J & Bates, DM (2001) *Mixed Effects Models in S and S-PLUS*. Springer
- Sokal & Rohlf (1995) *Biometry* (3rd Ed). W H Freeman & Co.
- Aitkin et al. (2009) *Statistical Modelling in R*. OUP, Oxford.
- Bolker et al. (2009). Generalized linear mixed models: a practical guide for ecology and evolution. *Trends in Ecology & Evolution* 24: 127-135.

2.3.14 Maximum Likelihood Statistics and Model Fitting

Week: 20 (*Monday 15th February – Friday 19th February*)

Convenor: Tin-Yu Hui

Maximum likelihood estimation (MLE) plays a key role in statistical estimation. It provides a framework to obtain the “best” set of parameters given the observation with an associated statistical model. Many statistical methods used in ecology and evolution, including most of the general and generalised linear models described in the statistics module, are consequences of maximum likelihood under specific applications. This module aims to give a formal definition to MLE, and to apply the technique to problems in ecology. By the end of this module, student will be able to understand the concepts of MLE, and implement MLE in R for standard and non-standard problems.

Timetable:

Monday 15th February	
10:00 – 11:00	<i>Background probability theory</i> (Lecture, Wallace, Tin-Yu Hui) Common discrete/continuous random variables. Probability mass/density function. Expectation of random variables. Central limit theorem.
11:30 – 13:00	<i>Background probability theory continued</i> (Lecture, Wallace, Tin-Yu Hui)
14:00 – 17:00	<i>Background probability theory continued</i> (Practical, Wallace, Tin-Yu HuiHui)
Tuesday 16th February	
10:00 – 11:00	<i>Probability and Likelihoods</i> (Lecture, Wallace, Tin-Yu Hui) Multivariate random variables. Definition of a likelihood function. Maximisation in R.
11:30 – 13:00	<i>Probability and Likelihoods continued</i> (Lecture, Wallace, Tin-Yu Hui)
14:00 – 17:00	<i>Probability and Likelihoods continued</i> (Practical, Wallace, Tin-Yu HuiHui)
Wednesday 17th February	
10:00 – 11:00	<i>Properties of maximum likelihood estimators</i> (Lecture, Wallace, Tin-Yu Hui)
11:30 – 13:00	<i>Properties of maximum likelihood estimators continued</i> (Lecture, Wallace, Tin-Yu Hui)
Thursday 18th February	
10:00 – 11:00	<i>Likelihood-ratio test</i> (Lecture, Wallace, Tin-Yu Hui)
11:30 – 13:00	<i>Confidence interval estimation</i> (Lecture, Wallace, Tin-Yu Hui)
14:00 – 17:00	<i>Likelihood-ratio test and Confidence interval estimation</i> (Practical, Wallace, Tin-Yu HuiHui)
Friday 19th February	
10:00 – 11:00	<i>MLE examples</i> (Practical, Wallace, Tin-Yu Hui)
11:30 – 13:00	<i>MLE examples</i> (Practical, Wallace, Tin-Yu Hui)
14:00 – 17:00	<i>MLE examples</i> (Practical, Wallace, Tin-Yu Hui)

Readings and Resources:

- Millar, R.B., Maximum Likelihood Estimation and Inference – With Examples in R, SAS and ADMB. Wiley-Blackwell, 2011.
- Casella, G. & Berger, R.L., Statistical Inference, second edition, Cengage Learning, 2001.

2.3.15 Introduction to Bayesian Statistics

Week: 20 (Monday 22nd February – Friday 26th February)

Convenor: Koenraad Audenaert

This module will introduce students to background theory and computational practice of using Bayesian statistics. Topics to be covered include the Bayes theorem, fitting by Bayesian methods, MCMC, and other basics of Bayesian statistics as applied to biology.

Scheduling for this module will be (All activities will in Wallace.):

1000–1100: Lecture

1130–1230: Lecture

1400–1700: Practicals, except Wednesday

Readings and Resources:

- R. Christensen et al, Bayesian Ideas and Data Analysis (Chapman & Hall/CRC Texts in Statistical Science)

2.3.16 Networks and Complex Systems in Ecology and Evolution

Week: 21 (*Monday 29th February – Friday 4th March*)

Convenor: Samraat Pawar

In this module a series of lectures and practicals will introduce you to theoretical basics, current topics and empirical approaches on the structure and dynamics of networks arising from (structurally and dynamically complex) ecological and evolutionary systems. The main aim of this module is to introduce students to the key network-oriented ideas, mathematical/computational tools, and data analyses methods. The week will kick off with introductory lectures and practicals on networks in ecology and evolution where you will be given an overview of types of networks that arise in ecology and evolution, why networks are a powerful and necessary representation of such systems, and an overview of the approaches used to study different kinds of networks. Here is a list of topics that will be covered:

All activities will be in Wallace in Hamilton building.

Scheduling for this module will typically be:

1000–1100: Lecture

1130–1230: Lecture

1400–1700: Practicals, except Wednesday

Sub-modules:

Day 1: Introduction to complex systems, scaling and networks (Samraat Pawar), and Modelling stochastic dynamics of biochemical networks (Vahid Shahrezaei)

Day 2: Inference on complex networks (Till Hoffman)

Day 3 – 4: Maximum entropy and interaction networks (Katharina Brinck)

Day 5: Bayesian networks (Veronique Lefebvre)

Readings and Resources:

- Day 1:

- Proulx, S. R., Promislow, D. E. L. & Phillips, P. C. 2005 Network thinking in ecology and evolution. *Trends Ecol. Evol.* 20, 345–53.
- Milo, R., Shen-Orr, S., Itzkovitz, S., Kashtan, N., Chklovskii, D. & Alon, U. Network motifs: simple building blocks of complex networks. *Science* 298, 11–14, 2002.
- Stouffer, D. B. & Bascompte, J. Understanding food-web persistence from local to global scales. *Ecol. Lett.* 13, 154–161, 2010.
- Mileyko, Y., Joh, R. I. & Weitz, J. S. Small-scale copy number variation and large-scale changes in gene expression. *PNAS* 105, 16659–16664, 2008.
- Mark Newman. The structure and function of complex networks. *SIAM Review*, 45:167–256, 2003.
- Mark Newman. *Networks: An Introduction*. OUP, 2010.

- Day 2:

- R. Albert and A.-L. Barabási. Statistical mechanics of complex networks. *Reviews of Modern Physics*, 74:47–97, 2002.
- Albert-László Barabási and Réka Albert. Emergence of scaling in random networks. *Science*, 286(5439):509–512, 1999.
- Paul Erdős and Alfréd Rényi. On the evolution of random graphs. *Publications of the Mathematical Institute of the Hungarian Academy of Sciences*, 5:17–61, 1960.
- Santo Fortunato. Community detection in graphs. *Physics Reports*, 486(3–5):75–174, 2010.
- Paul Holland, Kathryn Blackmond Laskey, and Samuel Leinhardt. Stochastic blockmodels: First steps. *Social Networks*, 5(2):109–137, 1983.
- Mason A. Porter and James P. Gleeson. Dynamical systems on networks: A tutorial. *arXiv*, page 1403.7663, 2014.

- Day 3–4:

- TBA

- Day 5:

- TBA

In addition, on 16th March 15.00–17.00, there will be a workshop “Writing for Masters: Literature Review” in Haldane lecture theatre

This workshop is provided by the Graduate School and you may need to reserve a place on the course through their website <http://www3.imperial.ac.uk/graduateschool/currentstudents/professionalskillsma>

3 Research project and Thesis guidelines

Each MSc and MRes student will carry out an independent research project leading to a written report (dissertation). The project must have at least one supervisor from the Department of Life Sciences at Imperial College, possibly one or more co-supervisors, and two *project committee members*. A list of potential internal supervisors as well as external supervisors and institutions are given below.

3.1 Research project committee

The role of the two committee members will be to mark and provide feedback on the project progress report (MRes), and the Final Project submission, including the Presentation, Viva, and Written Report (MSc and MRes). Project committee members can be anybody from the IC Department of Life Sciences. The student's Supervisor along with the Course Directors will choose the committee members, but the student is encouraged to suggest possible candidates.

3.2 Choosing and proposing your research project

Research project topics are generally part of ongoing active research within the Department and across a range of academic partners. We maintain an online list of available projects to which new project proposals are added throughout the year. The details of available projects are available at:

Webpage <http://goo.gl/GWtgy6>

Tab delimited text file <http://goo.gl/awH7Vf>

This list will provide broad details of research projects but the precise topics of projects will be finalised in discussion between the student and potential supervisors. Project descriptions will appear throughout the year but we aim to provide the majority of project topics by 15th October 2014.

The main difference between MRes and MSc projects will be the scope — keep in mind that the MRes projects have an extra four months or so available. The project can be on any *biological* topic and must entail significant amounts of mathematical theory or statistical analyses of large datasets, or combinations of these.

Both CMEE MRes and MSc students will select a project and turn in a two-page project proposal in pdf format on by the respective deadline given in the summary timetable (section 2.2).

The proposal should be written in latex. It should be on A4 paper with 2cm margins, in 12pt font, double-spaced, with lines numbered continuously. It must not exceed 2 pages *excluding* references. The references can be formatted compactly, but must contain the full title of the paper. The proposal should begin with a title, supervisor name(s) with affiliation(s) and contact details (including email), followed by names of the two Project committee members. Thereafter, it should contain the following sections (all quite brief, of course): (i) Introduction to the project idea and proposed questions, (ii) Proposed methods, (iii) Project feasibility supported by a time line of tasks (Gantt chart), and (iv) Cited references. The proposal will ideally have been prepared in close collaboration with the supervisor(s)!

The report should be pushed, in pdf format, to your git repository into a directory called **Proposal** inside a **Project** directory.

3.3 MRes project progress report and oral presentations

CMEE MRes students will make two project-related presentations (see section 2.2 for dates). The first presentation will follow submission of a project progress report, and the second following submission of the final report (the thesis – see section 3.7). The project progress report and presentation are aimed at providing the Supervisor and the Course Director an account of the status of the project. The presentations will not be marked *per se*, but will play an important role in giving the committee members and supervisors an overview of the project — so please take them seriously!.

3.3.1 The MRes project progress report

The report should give an account of the project's progress and make a convincing case that the project continues to be feasible and approximately on schedule. It is the Project Supervisor's job to give feedback on these aspects and more. The two Project committee members will provide feedback and mark the progress report, as well as provide verbal feedback on the presentation.

The progress report should be written in latex, and formatted along the lines of the project proposal described above (A4 paper with 2cm margins, 12pt font, double-spaced, lines numbered continuously), but can be upto 5 pages long *excluding* figures and tables (including their captions), references and title page (if applicable). Please include the following sections: Introduction, Methods (split up into Theory, Data, Simulations, Statistical analysis etc, as applicable), Preliminary results, and Outlook. The last section (Outlook) must provide a clear assessment of where you and your supervisor think the project is at, and what the next steps are. references should be properly formatted (any style is fine) and cited in the text. All figures and tables should be neatly labelled, captioned, and cited in the text. Please do not embed figures and tables in the text, but include them at the end as you would in the final report along with their captions. Also please do not put captions and figures/tables on separate pages. You may also provide supplementary information, but don't count on people to read it thoroughly. Please cite any supplementary info properly — do not just append a bunch of supplementary pages without referring to it or it's sections in the main text.

The report should be pushed, in pdf format, to your git repository into a directory called **ProgressReport** in the **Project** directory which will presumably already contain your original Proposal.

3.3.2 the MRes project progress presentation

The project progress presentation will be 10 minute long followed by 10 minutes for questions, discussion and changeover. It will be attended by the committee members, project supervisor, at least one of the Course Directors, and all (MSc + MRes) students. You will be penalized for over shooting your allocated presentation time of 10 minutes.

The structure and content of the presentation should match that of the project progress report. The student's responses in the post-presentation discussion will be incorporated into the marking.

All presentations will be on the same day. You will receive a schedule and location in advance.

The most important thing is to get feedback at this stage – so please don't get psyched about how much you have or have not done already while preparing your presentation and project progress report.

3.4 MSc project-related oral presentations

CMEE MSc students will make two project-related presentations (see section 2.2 for dates). The first presentation (Proposal Defense) will coincide with the MRes project progress presentations described above. The second will follow submission of the final report (the thesis – see section 3.7).

3.4.1 the MSc pre-project presentation

The pre-project presentation or defense will aim to put the proposed project and its objectives in a scientific context. You can think of it as a proposal defence of sorts. A good idea might be to focus on a published paper or set of papers on a field or topic that form the background to the proposed Research project. The objectives of the project should be clearly stated and justified in the presentation, and structure of the presentation should be consistent with that of the project proposal. You may have made updates or revisions to the proposal idea and content since you submitted it.

The presentations will not be marked *per se*, but will play an important role in giving the committee members and supervisors an overview of the project — so please take them seriously!

The 10-minute presentation will be followed by 10 minutes for questions, discussion and changeover. It will be attended by the Project committee members, project supervisor, at least one of the Course Directors, and all (MSc + MRes) students. You will be penalized for over shooting your allocated presentation time of 10 minutes.

The most important thing is to get feedback – so please don't get psyched about how much you have or have not done already while preparing your talk.

All presentations will be on the same day. You will receive a schedule and location in advance.

3.5 the final presentation (both MSc and MRes)

The final project presentation will be 10 minutes long followed by 10 minutes for questions and changeover. This presentation, along with the viva, will be part of your dissertation defence, so please take it seriously.

It will be part of the Silwood Masters Symposium wherein all Masters students present their research. Your presentation will usually be attended by the Project committee members, project supervisor, at least one of the Course Directors, and all CMEE (MSc + MRes) students.

The structure and content of the presentation should match that of the final dissertation. The student's responses in the post-presentation discussion will be incorporated into the report marking.

3.6 Lists of potential supervisors

The following sections on potential supervisors and broad research areas are intended to guide you in thinking of the topics you might be interested in and in which staff to approach. All projects must have a Life Sciences supervisor or co-supervisor, but can also be based at external organisations with additional external supervision. Previous external organisations include ZSL, the Met Office,

Royal Botanic Gardens, Kew, English Nature, the Game Conservancy and so on. You are encouraged to look up titles and abstracts of previous year projects from CMEE as well as quantitative projects from other courses such as MSc/MRes EEC, which can be obtained from Mrs. Amanda Ellis (amanda.ellis@imperial.ac.uk). A list of previous CMEE project topics is also provided on Blackboard, and example titles and reports are also available from the course website (google Masters CMEE!).

As and when new supervisors add projects after printing the handbook, details of their research interests may be downloaded from these links:

Please note that this is not an exhaustive list of internal supervisors – there a number of faculty members at South kensington that you cpuld approach. Please have a look at the DoLS website at <http://www.imperial.ac.uk/life-sciences/research/research-themes/>

Webpage <http://goo.gl/ok0vZS>

Tab delimited text file <http://goo.gl/aVGHs4>

3.6.1 Internal supervisor research areas

Cristina Banks-Leite

Understanding bird responses to deforestation in South America and the consequences of species extinction to the loss of functional diversity. An example projects could use data previously collected on birds from the Atlantic Forest and Amazon as well as collate a database on species morphological and behavioural traits. The aim is to address questions such as; which response traits are related to species sensitivity to deforestation; does species extinction lead to loss of functional diversity; and are response and effect traits correlated.

Tim Barraclough

Evolutionary biology and species diversity; speciation in asexual rotifers; evolution in bacterial communities including human gut bacteria.

Tom Bell

Projects are available on microbial ecology and evolution. Projects will involve testing ecological and evolutionary theory using microbial communities as a model system. Projects could typically include laboratory and field work. General areas of interest include:

- Biodiversity and ecosystem functioning. Biodiversity is thought to be an important determinant of ecosystem-level processes (e.g. decomposition). Projects in this area will look at the ecological mechanisms that contribute to biodiversity-ecosystem functioning relationships, and at how biodiversity affects functioning over evolutionary timescales.
- Microbial biogeography. Spatial patterns of microbial community composition and diversity remain understudied. Projects might include, for example, manipulating rates of colonisation to understand how community assembly affects community membership.
- Trophic interactions. Projects will involve examining the role of protists in structuring bacterial community composition. Several hundred protist and bacterial isolates are available to construct food webs and to examine the relative roles of resource competition and predators in determining bacterial community dynamics.

Martin Bidartondo (Royal Botanic Gardens, Kew)

I supervise projects on the ecology and evolution of plant-fungal interactions that may be field,

glasshouse, laboratory and/or computer-based. Current interests include the evolutionary ecology of the fungal symbioses of ancient plant lineages and the ecological drivers of mycorrhizal diversity in trees.

Austin Burt

I supervise computer-based projects on population genetic data analysis and population genetic modelling.

Lauren Cator

Our research aims to improve understanding of mosquito behavior and how behavior mediates interactions with other organisms, the parasites that they transmit, and the dynamic world that they live in. Current areas of research fall under two broad categories: the feeding behaviors of infected mosquitoes and mosquito mating behavior in aerial swarms.

Magda Charalambous

I supervise projects on insect mating behaviour and sexual selection. In the past I have had projects on *Drosophila*, crickets and bulb mites (to look at alternate male mating behaviours). We have video/sound recording facilities available and have used jWatcher software both as an event recorder and to produce transitional probabilities for ethograms. I am happy to discuss any project with a behavioural angle and also to act as internal supervisor for behavioural projects undertaken away from Silwood.

Rob Ewers

I work on spatial patterns of forest and the biodiversity contained within those forests. Work involves investigating and trying to predict patterns of forest cover from local through to global scales, sampling of taxa within selected landscapes, and manipulative field experiments. Most of the work uses invertebrates as a model system, and is focused at the Stability of Altered Forest Ecosystems (SAFE) Project in Malaysian Borneo.

Richard Gill

Social insects (the ants, bees and wasps) have been my focal study system. Their large but intricate societies exhibit efficient cooperative behaviour making them not only interesting for the study of animal behaviour, but also a dominant insect group in the environment that provide crucial ecosystem services. As such my research has followed two primary themes: environmental factors affecting the behaviour and ecology of bees; and the evolution of social strategies. I have recently supervised projects on the impact of neonicotinoids on bee foraging behaviour.

Jason Hodgson

My research focuses on human and primate evolutionary genetics. I use genomic SNP and next-gen sequence data, as well as bioinformatic approaches to test evolutionary hypotheses. I am particularly interested in the way that local adaptation and gene flow have contributed to the distribution of genetic variation. As a globally distributed species, human populations have experienced myriad selective environments and consequently local adaptation has driven population differentiation between regions. Recent migrations associated with the development of agriculture and efficient transportation have spread locally adapted genetic variants across the landscape. I use recently admixed populations to understand the fate of locally adapted alleles when they are introduced to new environments through migration. I am currently working with the people of Madagascar, but also work on populations from the Horn of Africa, Middle East, and Oceania. I am also interested in all aspects of primate evolution. I am particularly interested in integrating molecular phylogenetics with the fossil record to produce a more nuanced understanding of primate evolution.

Samraat Pawar

My group studies dynamics of biological systems across levels of organization, from individuals to communities. We use a combination of mathematical and computational modeling and analyses of massive ecological data sets. We work across organisms and ecosystems, and are particularly interested in the structural and functional differences and similarities between terrestrial and aquatic communities. You can find further details and information on specific projects we are currently involved in by visiting <http://imperial.ac.uk/people/s.pawar>.

Iain Colin Prentice

Understanding and modelling fundamental carbon, water and nutrient cycling processes at the plant and ecosystems levels. Global carbon cycle modelling and climate feedbacks. Assessment of the impacts of global climate and CO₂ changes for natural ecosystems, water resources and crops.

Andy Purvis

Integrative biodiversity science: interests in conservation biology (modelling and projecting responses of species, populations, functional diversity and ecosystem services to human impacts, in both natural and managed systems), macroevolution (primarily in planktonic foraminifera), and phylogenetic comparative biology (especially models of trait evolution)

Ben Raymond

My research interests include the applied evolutionary biology of microbes and microbe-host interactions and the evolution and maintenance of cooperation. I am interesting in offering student projects on the evolution of resistance to biopesticides or antibiotics, plasmid ecology and in the ecology and application of biocontrol agents.

Arkhat Abzhanov

My research group is interested in a variety of topics related to the vertebrate craniofacial (head) development, craniofacial genetic conditions in humans and craniofacial developmental evolution. We use morphometric, molecular, cellular and genetic approaches to study the precise mechanisms of cranial skeletal cell differentiation and skull/face morphogenesis in amniotes. The species we work with range from the laboratory "model" systems, such as chicken embryos and mouse mutants, to the "non-model" species used for evolutionary developmental studies, for example, Darwin's Finches and their relatives from Caribbean Islands, as well as other birds and, more recently, reptiles, both squamates (e.g. Anolis lizards), and archosaurs, such as alligators. This combination of laboratory "model" species with "non-model" species from natural environments allows us to address important conceptual questions, such as the roles of particular developmental genetic mechanisms (e.g. modularity) in evolution of adaptive variation and significant morphological transitions at both small and large evolutionary scales.

Julia Schroeder

I am interested in how genetic variation in traits that are socially interactive can be maintained. The approach is two-fold: I use observations made on wild birds to generate hypotheses which I then test experimentally. I work on two wild bird populations, for both of which I have long-term data, including a pedigree, available. This allows me to study questions in evolutionary biology and behavioural ecology that are otherwise hard to answer in wild study systems. I am also interested in how behaviour of one individual affects other individuals, and whether or not variation in these interactive behaviours has a genetic basis. If that is so, such so-called indirect genetic effects (IGEs) can strongly, and unintuitively, alter predictions of the evolution of said behaviours. For instance, we suspect now that the behaviour of female birds can affect how much parental care her social male will provide for her offspring. He may provide less if she has

cheated on him. However, what cues males use to assess this is unclear, and this may affect how parental care and mating systems can evolve.

Brian Hollis

I study sexual selection and genomic conflict using experimental evolution, primarily with the fruit fly *Drosophila melanogaster*. Many of my experiments involve generating and testing predictions of evolutionary theory using populations adapting to different mating systems. I use both phenotypic and genomic approaches to address interconnected questions about the role of sexual selection in the process of adaptation and the evolutionary impact of conflict between the sexes.

Joe Tobias

My research investigates the processes generating, sustaining and structuring biodiversity. I use a combination of experimental, phylogenetic and spatial mapping approaches to test theory, and to explain microevolutionary and macroevolutionary patterns over a range of spatial and temporal scales. Most of this work uses the world's birds as a study system, but increasingly focuses on other components of biodiversity, from primates and reef fish to insects and plants. I am interested in using these insights to help us predict and manage the response of ecosystems to global change, with applications from biodiversity conservation to environmental policy. Although recent work is global in scope, my main interest lies in tropical systems, especially tropical rainforests. Current areas of interest are: Sensory ecology, signal evolution; speciation and diversification; community assembly and species interactions; ecological and functional impacts of climate and land-use change; sustainable management of ecosystem function and services.

James Rosindell

My main areas of research interest are biodiversity theory, island biogeography, ecological neutral theory and scientific data visualisation. I offer a range of projects on these topics all of which would involve the opportunity to develop programming skills and apply scientific computing to ecology and evolution. The island biogeography projects would involve modeling the richness and abundance of endemic and native species on island archipelagos. Ecological neutral theory is a controversial area of biodiversity theory that uses very simple models; projects here would involve introducing extra realism to these models and fitting them to tropical moist forest tree datasets. Students working on the data visualisation project would be contributing to a wider initiative to provide software to visualize the tree of life and information on each species within it. The software will be for use by scientists and as a tool for public outreach and education on the subjects of ecology and evolution.

Vincent Savolainen

Work in my lab combines field ecology, molecular phylogenetics, population genetics, and genomic approaches to help explain the origin of biodiversity and, where possible, find solutions for its preservation in a rapidly changing world. Most projects will have a wet lab component (genetics and genomics), possibly a field component too (UK, Australia, Africa).

Mike Tristem

I am interested in projects in the following areas:

- Endogenous retrovirus insertional polymorphisms in humans. The published human genome sequence contains over 98,000 endogenous retroviruses (ERVs) but all are defective, containing lethal mutations or major deletions. Only one family of HERVs (human ERVs), termed HERV-K(HML2), has been active since the divergence of humans and chimpanzees approximately 6 mya. The family contains many members that are human-specific as well as several that are insertionally polymorphic, an integrated element being present in some

human individuals but not in others. Possible projects will identify additional insertional polymorphisms via PCR screening of different human individuals. Any polymorphisms will be useful as (i) population markers, (ii) in assessing prehistoric human migration patterns and (iii) investigating the present day activity of HERVs.

- Domestication of human endogenous retroviruses. Some ERVs have become domesticated or co-opted and hence fulfil a function of benefit to the host (e.g. preventing infection by other viruses). Identifying such elements has been very slow to date. However, with the recent sequencing of the human and chimpanzee genomes the process will be much faster and can be accomplished using bioinformatics. Thus possible projects will compare orthologous (in the same location) ERVs in the two genomes. Selection since the divergence of the two hosts will be examined with any evidence of purifying selection indicating domestication of the elements being examined.

Chris Wilson

I am interested in the broad and enduring puzzle of the evolution of sexual reproduction, and the evolutionary and behavioural ecology of host-parasite interactions. My primary study system is a group of bizarre freshwater micro-invertebrates called bdelloid rotifers, and I work in both the field and the laboratory. Methods used include molecular genetics (including next-generation transcriptomics), phylogenetic analysis, microscopy and microphotography (bright field, phase contrast and fluorescence), microbial culturing, aseptic cultivation of fungi and bacteria, experimental bioassays and field surveys. Projects are available focusing on various aspects of this system, using many of the techniques listed above, depending on individual interests.

Magda Charalambous

I supervise projects on insect mating behaviour and sexual selection. In the past I have had projects on *Drosophila*, crickets and bulb mites (to look at alternate male mating behaviours). We have video/sound recording facilities available and have used jWatcher software both as an event recorder and to produce transitional probabilities for ethograms. I am happy to discuss any project with a behavioural angle and also to act as internal supervisor for behavioural projects undertaken away from Silwood.

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Alfried Vogler

My main interest is in the phylogeny and evolution of Coleoptera (beetles), arguably the largest radiation of animal species on Earth. I use DNA sequences and phylogenetic analysis to study a wide range of questions to explain the great diversity of beetles, at all hierarchical levels, from populations to basal relationships of the order. In addition, I develop DNA-based approaches as short-cut for taxonomy, in particular in poorly known tropical groups. This permits novel

analysis of entire insect communities for surveys of total diversity or for monitoring the response to environmental disturbance. I have a joint appointment with the Natural History Museum (NHM) London where I conduct most of my research.

3.6.2 External supervisor and organization research areas

The examples below give outlines of project areas from some recent partners. More projects with external partners are likely to arise during the course. In the first instance, please contact the course directors about possible projects with external organizations.

Projects outside Life Sciences but within Imperial College

Recommended websites to read about research interests of IC staff in other departments and colleges are :

- Mathematics, <http://www3.imperial.ac.uk/mathematics>, their staff list with personal web pages are at <http://www3.imperial.ac.uk/mathematics/about/people>
- Biomathematics Group: <http://www2.imperial.ac.uk/mathematics/php/researchgroups/bms/>
- Statistics <http://www3.imperial.ac.uk/statistics/research>, especially the Statistical Genetics and Bioinformatics Group <http://www3.imperial.ac.uk/statistics/research/statgenbio>.

ZSL Institute of Zoology (IoZ)

<http://www.zsl.org/science/ioz-staff-students>; Contact Dr. Chris Carbone <http://www.zsl.org/users/chris-carbone>

Natural History Museum (NHM)

NHM has a massive collections digitization project going on that are yielding unique and massive datasets on species morphologies, and spatio-temporal distributions, among other things. Contact Andy Purvis Andy.Purvis@nhm.ac.uk or Steve Brooks S.Brooks@nhm.ac.uk for more information and project ideas.

Centre for Environment, Fisheries, and Aquaculture Science (CEFAS) - Simon Jennings and others

A wide range of projects in applied and fundamental aspects of fisheries management and marine ecology and ecosystem management may be available linked with the CEFAS Lowestoft Laboratory in Suffolk. Projects may be located at Silwood or possibly at Lowestoft. For more information about CEFAS and what work goes on there, read <http://www.cefasc.co.uk/> and then initially contact Dan Reuman (d.reuman@imperial.ac.uk) to discuss possible projects and to get CEFAS contacts.

Some specific project areas are:

- Exploring the CPR fish larvae datasets for link with environmental factors and/or fishing effects.

The Continuous Plankton Recorder (CPR) survey, which was started in the North Sea in 1931, is one of only a few long-term biological monitoring programmes and the only one at a spatial scale that gives a systematic coverage of the North Sea in space and time. As well as the zooplankton and phytoplankton, fish larvae were also analysed until the 1970s. At that point cost cuts meant that analysis of fish larvae in the CPR samples was stopped but the samples themselves have been archived. The M1101 Defra funded project allowed for

the analysis of the archived samples for fish larvae. The newly available fish larvae dataset from the CPR covers the entire UK shelf from 1950 to 2004 and comprises about 110,000 data points. This dataset offers a unique opportunity to investigate long term changes over decadal scales in the abundance and distribution of fish larvae in relation to physical and biological factors, as well as fishing effects. Unfortunately only a few species are well sampled by the CPR and can be used for further analysis. Among such taxons we have identified clupeids, sandeels, Atlantic mackerel, dab, Blue whiting. So far work has been done at Cefas on the following use of the dataset:

- The potential for larval fish data from Continuous Plankton Recorder (CPR) surveys to inform the stock assessment of North Sea sandeels (paper in preparation).
- North Sea spawning mackerel and the stock crash in Northern North East Atlantic mackerel (paper in preparation)
- Regime shifts in the Celtic Sea and North Sea and their impacts on fish larvae (paper in preparation)
- Long-term changes in the abundance of clupeids, sandeels, Atlantic mackerel and Dab from the CPR (1960–2004) in the North Sea, in relation to biological (prey fields) and environmental factors and using principal component analysis (temperature, salinity) (paper in preparations).

Clearly there is immense potential for further use of the fish larvae dataset from the CPR. For this particular project I would like to explore the clupeid dataset further. Unfortunately clupeids could not be identified to species level in the CPR samples. We suspect they are mostly herring and sprat, sardine and anchovies might also contribute to the clupeid index in some areas. However initial work on the dataset is encouraging and we have found statistically significant correlations between yearly abundances of clupeids larvae from the CPR and herring spawning stock biomass from stock assessment data from ICES. The idea for this project would be to try and find further links between environmental conditions (both physical and biological), fish larvae and stock assessments in order to:

- Assess the potential of the CPR dataset to be used in support of stock assessment purposes for selected fish.
- And/or explore further the link between environmental conditions and the survival potential of fish larvae.

Contact Sophie Pitois, Plankton ecologist (Cefas), sophie.pitois@cefas.co.uk

Centre for Ecology and Hydrology, Wallingford

Research areas for projects are based around ecological modelling of UK systems, often with practical or policy implications. Previous topic areas include the following:

- Are extinction risks of plant species larger on the edge of distributions than in the centre? Based on Atlas data of the UK and the Netherlands we will look at spatial patterns in range decay of over a wide range of species over the last 50 years. Next to geographical patterns, a range of alternative environmental indicators for correlated extinction will need to be explored. Such information could feed into priorities to set in conservation of declining species.
- Dynamics of butterfly range expansion. Understanding landscape effects on colonisation and extinction can help us to facilitate species movements under changing climates. This project will investigate how landscape structure affects the colonisation of new sites at leading range margins. In addition, local extinctions will be related to landscape characteristics.

- Managing habitats for butterfly populations. Butterfly populations are highly sensitive to land management and understanding the long term effects management regimes can aid conservation. This project will investigate how nature reserves managed in different ways affect butterfly population dynamics. For example, does a heterogeneous management regime lead to more robust and stable populations?
- Floral homogeneity in the UK: the effects of non-native plants and urbanisation. Biotic homogenisation is regarded as a potential cause of biodiversity loss and is strongly linked with the arrival and dominance of non-native plants and animals and urbanisation. Using an extensive dataset on plant distributions at a tetrad scale collected at 2 time periods by the Botanical Society of the British Isles (BSBI) throughout the UK we will calculate similarity indices and look at spatial and temporal trends in similarity for native and non-native plants to investigate whether there is any signs of homogeneity in the British flora and what the mechanisms behind this are addressing questions such as: Are habitats that are more similar associated with higher numbers of alien species? Are urban habitats more similar than habitats in the wider countryside? Have plant communities become more homogeneous over time in the UK?

UNEP-World Conservation Monitoring Centre (UNEP-WCMC)

A wide range of projects addressing global/regional biodiversity and ecosystem services issues, including: international agreements synergies, conservation priority setting, international trade and regional economies, drylands and livelihood support, wildlife trade and socialLaTeX networks, access and benefit sharing, protected area targets and social values, monitoring deforestation and degradation, and more.

Somerset Wildlife Trust

Broad research interests cover the interaction of land use methods and biodiversity on lowland wetlands. A variety of projects are possible from looking at restoration of biodiversity on restored peat extraction sites through to examining the effects of drainage and re-wetting on peat soil integrity and micro-arthropod diversity. Many projects are available concerning the conservation of nationally rare invertebrate species.

3.7 Thesis format and submission

Note that different courses at Silwood have different guidelines and past projects are not all written in the style required here.

Your dissertation should ideally be written using latex. A template can be found here: <http://www.prettyprinting.net/imperial/> (A beamer presentation template is also available there).

The write-up should be in the style of a scientific paper from a journal that it might be appropriate for submission to. By style, we mean the structure and how information is presented – you do not need to make it look like a journal paper, but it should have an abstract, introduction, methods, results, and discussion (possibly conclusions). If the results of your project are publishable, this approach will save much time and will provide valuable experience in paper writing. You may provide appendices if necessary (for example outlining methods in detail).

Always aim to be clear and concise. The size of the thesis will vary according to the student and project, but aim to make it as short as necessary to describe the work done and to discuss it in a general context. However, *do not* omit relevant data and information such as experimental procedures. A CMEE Masters thesis should not exceed 10,000 words in length (excluding figures, tables, references

and appendices). However, a common mistake is to assume that the reader knows the project as well as you do and to leave out clear descriptions of the motivation and structure of your research.

There are a few ways in which your thesis is likely to differ from a typical scientific paper:

1. You should make sure that you clearly state your aims/hypotheses/questions towards the end of your introduction.
2. You should take care to explain everything adequately so that the examiners can see clear evidence of understanding of all the concepts and methods – this might entail providing more detail or background in the introduction and methods than in a typical paper. However, some of this detail can be put in an appendix or supplementary material. For example, a molecular study might state in the Methods section of the main text that you extracted DNA according to a phenol/chloroform extraction protocol according to a particular reference. In the appendix, you should then describe the steps of your lab protocol in sufficient detail that other people could reproduce this procedure by following your description. Another example would be the mathematical derivation of an equation in the main text, or details of a computer algorithm.
3. You should make clear what you did versus what you were provided with. For example, did you collect the data that you analyzed or were they provided to you by someone else? Did you build the mathematical model yourself, or were you given one that you then analyzed or simulated? Did you write all the simulation or analysis code yourself?
4. You should make sure to discuss limitations of your study and what future work you would do to address those limitations or any other questions raised by your work. In your project, most likely you will run out of time to complete everything you want to do. In most papers, the research would not be so time limited so limitations can be addressed by further work.

The class repository/Blackboard has a folder with more advice on scientific writing, and papers from *Ecology Letters*, *Proceedings of the Royal Society B*, *Molecular Ecology*, and *Journal of Animal Ecology*, which we recommend as well-written example papers.

The final thesis must follow these preparation guidelines:

Cover Page

The cover page *must* bear the project title, your name and the month and year of submission. In addition, the following text must appear at the bottom of the cover page:

A thesis submitted in partial fulfilment of the requirements for the degree of Master of Science/Research at Imperial College London

Formatted in the journal style of XXX.

Submitted for the MRes/MSc in Computational Methods in Ecology and Evolution

Obviously, insert your choice of journal and choose the appropriate degree course.

Declaration

The first page inside the cover must provide a brief declaration of the contributions made by you and by others to your project. Key points to address are:

- Was the data provided to you or did you collect or assemble it?
- Were you responsible for data processing or cleaning, if required?
- Were any mathematical models developed by you or by your supervisor?
- What role, if any, did your supervisor play in developing the analyses presented?

Layout

Print your thesis double-sided on A4 paper. The main body of the text should be printed using 1.5 line spacing and page numbering should be used. The thesis margins should be at least 2 cm and the main text font size should not be smaller than 11 point.

Binding

Use a ring binder to bind the thesis. You should include a transparent plastic sheet in front of the cover page and use a plain white A4 sheet of card at the back. You will be provided with printed cover sheets with the college crest and logo.

Number of copies

You *must* supply three bound copies of your thesis: one for the external examiners and Library, and copy of your own, which you *must* take with you to your internal and external vivas.

You must also submit an electronic version of your project report on Blackboard and your class git repository (surely by now we won;t need to tell you into which directory it should go?). The digital copies should be identical to the printed version, including the pagination and cover sheet.

3.7.1 Thesis content guidelines

The following guidelines on content include tips from Andy Purvis, author of over 100 scientific papers and referee of many more.

Introduction

A good introduction should leave the reader with a clear idea of the problem to be tackled and looking forward to the more detailed sections to follow. It should include a section on the general way the problem has been approached. An essential concluding part of the introduction is to clearly define the aims of the research project and any hypotheses tested. Also, think about:

- *What is this paper about? (i.e., the broad area, big picture) Why is that interesting?*
- *Given its so interesting, why dont we know the answer?*
- *So, what is this about, more specifically? What are hypothesised to be the important things?*
Build from the most general and fundamental hypotheses to the most refined or tenuous ones.
- *How, roughly and briefly, will you go about testing these hypotheses? Why are you using this system? What approach will you use?*
- State clearly what your hypotheses are. These are not usually stated explicitly in a paper.

Methods

This should contain details of any methods used extensively during the project, layout of field experiments, theoretical methods, methods of statistical analyses etc. You can use subheadings for different procedures or tests. If field work is done, a general description of the study area may be included here. Extra methodological details can be placed in appendices. The golden rule is that the reader should be able to repeat what you did, should they so wish. The other rule – more important for your project than in a paper – is that you describe in enough detail to show youve understood what you did.

You should feel free to use subheadings in your methods and results to help organise different parts of your project. If so, keep the same order of the different parts of the project in all of your

sections: the methods for testing each hypothesis and the results of those tests are described in the same order as the hypotheses are described in the introduction. Also, think about:

- What is the overall design of the study?
- What are the variables and how do they relate to the hypotheses?
- How did you get the data?
- What are the characteristics of the data set / experiment – how many observations, how many replicates etc.
- General procedures, if any, that are true in all of the analyses (e.g., transformation of data, model checking, how models were compared)
- How did you test the hypotheses, in the logical order outlined in the introduction (i.e., from the general to the specific)? Make sure you show that your tests are appropriate.

Results

Describe your results in a logical order: this may not necessarily be the order in which you did the experiments. Briefly summarise the main results at the end of each main experiment or sequence of associated experiments. Do not duplicate results – put a table or a graph but not both unless the two methods of presentation demonstrate different points of importance. You must refer appropriately to figures or tables in the text and remember to emphasise and perhaps quote significant results. In particular, think about:

- What were the results of your hypothesis tests, in the order you describe them in the Methods?

Discussion

This should attempt to tie together the results, what they indicate in a broader context, the extent to which the original aims have been satisfied and what future work is suggested. Return to and address the ideas raised in the introduction. In particular, think about:

- *Whats the main thing we know now that we didnt know before?*
- *Whats the chain of logic and results that means we know it?*
- *How does this affect our – and other scientists – view of the world? What are the implications?*
- *What are the implications of the intermediate steps in the chain towards the main thing?*
- *What are the caveats that apply to this study? (Leave out caveats that apply to all studies.) What might be done about them? (Very important in a project write-up – What would you do differently if you were doing the project again or had more time?)*
- *What future work could build more broadly on what weve found?*
- *A nice wrap-up, emphasising how this study in this system is of interest to people who work on other things, or other systems.*

Abstract

Now, and only now, write the abstract, making sure it includes the key point from each of Andys top tips above that are *emphasized*. Dont rush the abstract – it is your first opportunity to tell a reader about the research and a clear, concise abstract sets them up to understand your work!

References

Make sure all cited references appear in this list at the end of the thesis using the standard style from your chosen journal. With /LaTeX comes BibTex, which you should use. If, in the horrible scenario, you decide to use something like MS Word, it is *highly* recommended that you use bibliographic software (e.g. Mendeley) to manage and format your citations. In particular, note that you can use Mendeley to output *.bib files that you can use in Latex.

Appendices

Large sets of data (e.g. census results, raw experimental results) should go in an Appendix if these are of value, e.g. indicating an interesting range of variation. All summary tables or graphs and outline results of analysis should be put in the text. Any useful parts of the study not directly relevant to the main theme may also be put in an Appendix (e.g. taxonomic descriptions and drawings in an otherwise ecological study).

Computer Programs

If the program has been published, cite the reference, include it in the reference list and provide a brief outline of the methods it uses. If you are using a program or code generated for the project then a more complete description is needed in the main text. You should provide the code used in an appendix and consider providing a flow chart and usage notes to help interpretation. You should take care to define all the input variables used in the program.

Figures

You should prepare figures to the same standard required for publication. All journals provide advice on preparing figures for publication, so do look at the advice to authors pages for your chosen journal. All figures must be numbered and have a caption that is sufficiently detailed to explain the main features of the content by itself. All figures *must* be referred to in the main text of the thesis. Put the figures in appropriate points in the text, close to the text that refers to them. In particular:

- The resolution of your figures is crucial. For plots, try to use vector image formats (exported as svg, pdf, or eps) and not bitmapped formats like JPG and TIFF. Standard /LaTeX documents typically allow *.eps or *.pdf figures to be inserted. using the freely available (and very capable!) vector graphics program Inkscape to “fine-tune” your figures is often a good idea. Inkscape will also allow svg’s to be exported in a /LaTeX compatible format (see the Inkscape documentation). For RASTER graphics, the freely available GIMP editor works very well. When using Word 2007 on PC (heavens forbid!), figures in Windows Metafile format are the most reliable vector format. For Word 2011 on Mac, figures in PDF format should give a good result. If you do have to use bitmaps, make sure they are at a high resolution (300 dpi or more) – this can be particularly important if you need to present line drawings or photographs of specimens or equipment.
- Plots are all about the data, so reduce margins and maximise the space in the figure for showing the data.
- Create the figure at the right size – when it is included in your thesis are all the axis labels and text going to be clearly legible.
- Avoid ‘chartjunk’ (google Edward Tufte!) – and avoid superfluous lines, legends and titles along with 3D effects.

Tables

Each table should be numbered, have a full descriptive caption and again *must* be referred to in the main text. Column headings should state units of measurement. Avoid large, complicated tables in the main thesis and if you have a large body of numerical data put it in an appendix.

4 Masters Programme regulations

Please see section 1.5.1 for weightages of individual cours components.

4.1 MSc CMEE course regulations

The following notes outline the Department of Biological Science's rules for awarding MSc Degrees (2008 edition – updated September 2011). The University's General Regulations and the College's Special Regulations should be consulted in all cases of doubt.

Course credit requirements

The scheme outlined is that accepted formally by the University and by Imperial College London for the award of *MSc in Computational Methods in Ecology and Evolution*.

[All taught modules listed in section 2.2 are compulsory]

Class Boundaries

The final degree mark will be used to classify MSc degrees according to the following notional boundaries:

Distinction	70% or more
Merit	60% or more (less than 70%)
Pass	50% or more (less than 60%)
Fail	below 50%

These boundaries are moderated at the Examiners' Meeting to take account of your performance over the year and any difficulties you may have experienced (such as illness).

Assessment of Performance

Taking the Course

The word 'take' in the context of these regulations means that you have attended the timetabled parts of a course (unless prevented by illness), sat its examination, submitted the coursework specified for it and completed a research project.

Illness or bereavement

A student who is unable to complete his or her exams because of illness or the death of a near relative must either sit the examination at the next normal time or have a special paper set for them. Original documentation, e.g. doctors letter, should be provided as supporting evidence by the student at the earliest opportunity.

Taught Modules

All modules are assessed by written examinations and coursework, or a combination of the two. Coursework assessment will be in the form of assessed computer practicals, reports or practical write-ups.

Coursework

Receipt of marks for assessed coursework is absolutely dependent upon you delivering the work by the stated deadlines (making due allowance for sickness). Marks for assessed practical class reports can be gained only if you attend and perform the relevant practicals.

Research Projects

Assessment will be by written report and viva voce after the completion the project. Information conveyed in the final project presentation will also be taken into consideration.

- a) The Supervisor will complete an assessment form on your performance during the project, sending the mark direct to the Course Director.

- b) Your project report will be marked by two internal examiners (who are also your Project Committee members – see section 3). Exceptionally, advice may be sought from scientists of equivalent standing from outside the Division;
- c) The two markers will both mark the thesis (report) and agree a mark.
- d) The Supervisor and Markers should each add a written justification of their marks, to inform the External Examiners.
- e) The Markers will viva the student and assign a mark based on the students performance in the viva.
- f) In the case that the two markers differ in 10% or more in their mark, they will provide a written justification on an agreed mark in the assessment form.

External Vivas

Each candidate will be interviewed on his/her project by the External Examiner, the purpose being to reveal any problems the candidate may have had with the project, and to probe their understanding of the research they undertook. There is no mark attached to the viva with the external examiner, but the examiner will take part in moderation discussions and use the feedback from students in assessing the quality of the course.

Resit Examinations a) If you should fail the examination you are entitled to resit it the next two times it is offered;

- b) if the coursework element failed to reach the threshold 50% mark, whether through inadequacy or lateness of submission, the student will normally be asked to repeat the specific failed course components (i.e. take them again);
- c) a candidate who has attended most of a course but fails its coursework element because of ill health or bereavement will normally be allowed to resubmit the relevant coursework by a new deadline;
- d) a candidate who has taken a course but fails to sit its exam because of ill health or bereavement will normally be allowed to carry over their coursework mark for that course to when they next take the exam.

Rules a) All elements of the assessment must be passed before a degree of MSc can be recommended.

- b) To pass the taught course element, the aggregate marks for each, the coursework and the examination components separately must be above 50%, as well as receiving an aggregate mark for the taught course above 50%;
- c) If you fail the examination component of the course, then you would normally resit any failed exam papers the following year. If you fail the coursework component of the taught course you will be allowed to resubmit the failed coursework if it is deemed to be appropriate;
- d) To pass the project element of the course you must obtain a mark of 50%;
- e) If you fail the research project or the coursework component of the taught course then you would normally retake the course the following year.
- f) All students must attend a viva by the External Examiner.
- g) Project reports are normally submitted in early September – vivas with the External Examiner are in late September prior to the Examinations Board Meeting.

- h) Only when a detailed case has been made and accepted by the Board will candidates be moderated into a different degree class from that associated with their aggregate final mark. In such cases, the Board may be guided by a notional Grey Area (up to and including 2% below the threshold mark) within which the degree class might be moderated.
- i) To achieve a Distinction, candidates should normally get 70% overall. Additionally, at least two of the three marks (for the project element, the coursework component of the taught course element, and the exam component of the taught course element) should be at least 70% and none should be less than 60%. To achieve a Merit, candidates should normally get 60% overall, with at least two of the three marks (for the project element, the coursework component of the taught course element, and the exam component of the taught course element) being 60% or more.
- j) Course convenors and project supervisors have a responsibility to notify the Examination Officer of exceptional circumstances known to them that might have affected the course mark given to a candidate; likewise personal tutors must record the existence of any medical or personal problems that might have affected performance in examinations or assessed coursework, and notify the MSc Office. All such circumstances must be made known to the Chair prior to the meeting of the Board of Examiners so that a preliminary assessment can be made of their likely effect, and additional information sought where necessary.

4.2 MRes course regulations

The following notes outline the Department of Biological Science's rules for awarding MRes Degrees (2008 edition – updated September 2011). The University's General Regulations and the College's Special Regulations should be consulted in all cases of doubt.

Course credit requirements

The scheme outlined is that accepted formally by the University and by Imperial College London for the award of *MRes in Computational Methods in Ecology and Evolution*.

[section 2 lists the required modules]

Class Boundaries

The final degree mark will be used to classify MSc degrees according to the following notional boundaries:

Distinction	70% or more
Merit	60% or more (less than 70%)
Pass	50% or more (less than 60%)
Fail	below 50%

These boundaries are moderated at the Examiners' Meeting to take account of your performance over the year and any difficulties you may have experienced (such as illness).

Assessment of Performance

Taking the Course

The word 'take' in the context of these regulations means that you have attended the timetabled parts of a course (unless prevented by illness), sat its examination, submitted the coursework specified for it and completed a research project.

Illness or bereavement

A student who is unable to complete his or her exams because of illness or the death of a near relative must either sit the examination at the next normal time or have a special paper set for them. Original documentation, e.g. doctors letter, should be provided as supporting evidence by the student at the earliest opportunity.

Research Projects

Assessment will be by written report and viva voce after the completion of each of the two projects in March and September.

- a) The Supervisor will complete an assessment form on your performance during the project, sending the mark directly to the Course Director.
- b) Your project report will be marked by two internal examiners. Exceptionally, advice may be sought from scientists of equivalent standing from outside the Division;
- c) The Markers will both mark the thesis (report) and agree a mark.
- d) The Supervisor and two Markers will each add a written justification of their marks on their respective marking forms, to inform the External Examiners.
- e) The Markers will viva the student and assign a mark based on the students performance in the viva.
- f) In the case that the two markers differ in 10% or more in their mark, they will provide a written justification on an agreed mark in the assessment form.

External Viva

After the project submission in September, you will be given a viva by one of the external examiners. External examiners will be given your project reports shortly after your internal viva. This external viva will contribute to moderation of the marks awarded by the Divisions assessors and examiners and determine whether students should be awarded a merit or distinction.

- Rules**
- a) All elements of the assessment must normally be passed before a degree of MRes can be recommended.
 - b) If a research project mark is a fail then the student would normally retake the project the following year.
 - c) All students must attend a viva by the external examiner.
 - d) Project reports are normally to be submitted in September and vivas with the internal and external examiners are in late September prior to the Examinations Board Meeting.
 - e) Only when a detailed case has been made and accepted by the Board will candidates be moderated into a different degree class from that associated with their aggregate final mark. In such cases, the Board may be guided by a notional 'Grey Area' (up to and including 2% below the threshold mark) within which the degree class might be moderated.
 - f) To achieve a Distinction, candidates should normally gain 70% overall, and at least 60% for both projects. To achieve a Merit, candidates should normally gain 60% overall. To achieve a pass, candidates should normally gain 50% overall, and at least 50% on both projects.

- g) Course convenors and project supervisors have a responsibility to notify the Examination Officer of exceptional circumstances known to them that might have affected the course mark given to a candidate; likewise personal tutors must record the existence of any medical or personal problems which might have affected performance in examinations or assessed coursework, and notify the MSc Office. All such circumstances must be made known to the Chair prior to the meeting of the Board of Examiners so that a preliminary assessment can be made of their likely effect, and additional information sought where necessary.

4.3 Regulations for Students

This reproduces the July 2012 version of the Imperial College London Regulations for Students. These regulations, along with Academic and Examination Regulations are available from <http://www3.imperial.ac.uk/registry/proceduresandregulations/regulations>

1. All registered students of the College are subject to the provisions of these Regulations for Students, the College Academic Regulations, and such other Regulations and Instructions for Students as the College may from time to time approve.
2. Any student whose sessional fees or whose residence charges (including charges made by the Du Cane Housing Association) have not been paid in full will not be allowed to proceed to the next year of the course and will be required to withdraw from the College. If any fees or charges are still unpaid at the time when a student enters for the last examination necessary to qualify for the award of a degree/diploma, the award will not be conferred and no certificate in respect of the award will be issued until the debt has been paid in full.
3. Any student wishing to occupy residential accommodation provided by, or on behalf of, Imperial College will be required to abide by the terms and conditions of the Licence. Acceptance of an offer of accommodation will signify acceptance of such terms and conditions.
4. Every registered student of the College is automatically a member of Imperial College Union unless, under the provisions of the Education Act 1994, a student has formally opted out of student union membership by recording that decision with the Academic Registrar in the manner prescribed.
5. Student disciplinary offences of a non academic nature are dealt with under a code of procedure agreed by Imperial College Union and approved by the Governing Body. In the case of serious offences, this may involve the suspension and/or expulsion of the student from the College. Students must not engage in any conduct which causes harm or unreasonable disturbance to students, staff, neighbours or visitors to the College, or damage to any property of the College or its students, staff, neighbours or visitors, or engage in any activity or behaviour which is likely to bring the College into disrepute. Illegal acts on or near College may also constitute offences under these College Regulations for students.
6. Candidates for the PhD or MPhil degrees are required by the College regulations to give conditional authority for their thesis or dissertation to be made available for public reference. Candidates who wish to retain personally, for a limited period, the sole right to grant permission to consult, borrow or copy their work must obtain the agreement of their supervisor and the Graduate School Committee. Approval will be given only in special circumstances and for a period not exceeding two years. Acceptance of a place as a research student at the College is deemed to imply acceptance of these conditions.

7. Undergraduates must inform their Senior Tutor and postgraduates their Postgraduate Tutor if they are absent from College for more than three days during term. If the absence is due to illness a medical certificate must be produced after seven days. If an examination is missed on account of illness a medical certificate must be produced immediately.
8. A student who contracts an infectious or contagious disease may be required to present a medical certificate acceptable to the College Health Service, indicating freedom from infection, before resuming attendance at the College.
9. The College may require a student to be assessed by the College Health Service, or other appropriate medical practitioner approved by the Health Service, if there is reason to believe that the student's state of health makes him/her unable to pursue his/her studies, or causes disruption to other members of the College, or causes or has the potential to cause harm to him/herself or others. If the medical assessment confirms that it is not in the interests of the student or the College that the student should continue his/her programme of study the Head of Department shall consult the College Tutors and, taking into account their advice, may suspend the student until he/she is fit to continue his/her studies or require the student to withdraw from the College. A student who refuses to undergo assessment may be suspended until such time as a medical practitioner acceptable both to the student and the College has assessed the student and confirmed in writing that the student is fit to resume study.

A student who is required to withdraw has the right to appeal against the withdrawal decision but not against the results of the medical assessment on which the decision is based. The student may, however, request that a second medical assessment be obtained from a medical practitioner approved by the College Health Service. The responsibility for hearing and deciding upon appeals is vested in the Senate and is delegated by the Senate to Appeal Committees, whose decisions are final.

A student who is suspended will be regarded as having taken an interruption of studies, and will be required to provide medical evidence as to fitness to return to study in accordance with the conditions attached to the granting of interruption of studies.

10. No work involving ionising radiation may be carried out in any part of the College except in accordance with the current edition of the Imperial College Local Rules for Safe Working Practices with Ionising Radiation (Second Edition Spring 1991).
11. Students who make use of College Computing facilities are required to familiarise themselves with and to abide by the current edition of the Imperial College Information Systems Security Policy and Codes of Practice and Guidelines.
12. Computer misuse will be regarded as a serious offence and will be dealt with under the College Disciplinary Procedure or, where appropriate, under the provisions of the Computer Misuse Act 1990.
13. Students who are authorised, as part of their studies, to make use of data and personal data as defined under the Data Protection Act 1998 are required to familiarise themselves with, and to observe the provisions of, the Act. Further details are available from the College Data Protection Officer.
14. All students must familiarise themselves and comply with the College's Student Intellectual Property Policy.
15. Students must ensure that their personal data on student e-service is kept up to date at all times including any change in their home or term time address or their emergency contact details.

16. Where a student has a pre-existing relationship, or develops a relationship during the course of study, with a member of staff such that there is a potential conflict of interest, especially with regard to the students assessment, the student and member of staff must declare this in confidence to the Head of Department at the start of the course of study or at the point at which the relationship has started if this is during the course.

4.4 Plagiarism and cheating

You are reminded that all work submitted as part of the requirements for any examination (including coursework) of Imperial College must be expressed in your own words and incorporate your own ideas and judgements. Plagiarism, that is, *the presentation of another persons thoughts or words as though they were your own*, must be avoided, with particular care in coursework, essays and reports written in your own time. Note that you are encouraged to read and criticise the work of others as much as possible. You are expected to incorporate this in your thinking and in your coursework and assessments, but you must acknowledge and label your sources.

Direct quotations from the published or unpublished work of others, from the internet or from any other source, must always be clearly identified as such. A full reference to their source must be provided in the proper form and quotation marks used. Remember that a series of short quotations from several different sources, if not clearly identified as such, constitutes plagiarism just as much as a single unacknowledged long quotation from a single source. Equally, if you summarise another persons ideas, judgements, figures, diagrams or software, you must refer to that person in your text and include the work referred to in your bibliography. Departments are able to give advice about the appropriate use and correct acknowledgement of other sources in your own work.

The direct and unacknowledged repetition of your own work which has already been submitted for assessment can constitute self-plagiarism. Where group work is submitted, this should be presented in a way approved by your department. You should therefore consult your tutor or course director if you are in any doubt about what is permissible. You should be aware that you have a collective responsibility for the integrity of group work submitted for assessment.

The use of the work of another student, past or present, constitutes plagiarism. Where work is used without the consent of that student, this will normally be regarded as a major offence of plagiarism. If in doubt, please ask one of the lecturers or course directors for advice on what constitutes plagiarism.

Failure to observe any of these rules may result in an allegation of cheating and may result in a penalty being taken against any student found guilty of plagiarism. Cases of suspected plagiarism and of other forms of cheating will be dealt with under the Colleges Cheating Offences Policy and Procedures: <http://www3.imperial.ac.uk/registry/exams/examoffences>. This link also provides access to the College's statement on Academic Integrity in Examinations and Assessment.

4.5 Student Complaints and Appeals

Imperial College aims to give the highest specialised instruction and service to all its students, however, in some cases it recognises that students may not always be satisfied with the service that they have received. If you wish to raise a concern, you should first seek advice from your student representatives and raise the matter with the individual concerned. If you are not satisfied with the outcome, you should consult the Colleges Registry website which provides clear and consistent

procedures that indicate how you can take your comments further: <http://www3.imperial.ac.uk/registry/proceduresandregulations/policiesandprocedures/complaintsappeals>

4.6 College Principles

The following web page defines the guiding principles of the Imperial community. The principles were developed by academic and support staff in partnership with undergraduate and postgraduate students and will be reviewed annually; see <http://www3.imperial.ac.uk/students/ourprinciples>