Lost API documentation

Lost Members

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Contents

Chapter 1

An introduction to the lost framework

1.1 The lost framework

The lost framework is a collection of utilities and models for working with biological sequences written in PRISM/B-Prolog. The framework tries to unify all these bits and pieces and to provide a way to integrate them.

1.2 Obtaining a copy of the lost framework

The lost framework can be obtained using *git* if you have an account on the *mox* server. Assuming that you have *git* installed on your local machine, to get a copy you need to clone the central repository:

```
$ git clone ssh://your-username@mox.ruc.dk/var/git/lost.git
```

It is also possible to clone the repository directly from mox if you want to work there, rather than on your local machine. To do this, just type

```
$ git clone /var/git/lost.git
```

In the following we will use `$LOST` to refer to top-most directory of your copy of the framework.

1.3 Configuring your copy of the lost framework

After obtaining the lost framework, a little configuration is needed to get started. You will need to edit the file `$LOST/lost.pl`. In the beginning of the file there are two important facts you may need to change,

```
lost_config(prism_command,'prism').
lost_config(lost_base_directory, '/change/to/local/lost/dir/').
lost_config(platform, windows_or_unix).
```

The option `prism_command` should point to a the main PRISM executable binary. If it is in your *$PATH* then you can usually leave it unchanged. `lost_base_directory` should be the full path of the directory (including trailing /) containing the `lost.pl` file. Note, that even on windows platforms you should use forward slash rather than backslash in the path specification. The value of `platform` should be either `windows` or `unix`.

To get started you can examine and run `example.pl` which is located in the in the `$LOST/scripts/` directory.

1.3.1 File structure layout of the lost framework

The lost framework is divided into several parts:

- `$LOST/scripts/`: Contains scripts that run models. These scripts are just small Prolog programs. The scripts does not consult models directly, but instead use predicates that the framework provides.
- `$LOST/models/`: Contains all the biological sequence models. Each model is located in its own subdirectory of `$TLOST/models/` the name of which is also used as name of the model.
- `$LOST/lib/`: Contains shared libraries used multiple models.
- `$LOST/data/`: Contains sequences and data to be used by the models. Annotations etc. generated by model invoked by the framework stored in this directory and given the extension `.gen`
- `$LOST/tmp/`: Is used for temporary storage of files bt models.

Chapter 2

Creating lost models

2.1 Lost model conventions

Each model is located in its own subdirectory of the of the `$LOST/models/` directory, henceforth called `$MODELS`. So for instance, the sample model called `sample_model1` is located in `$MODELS/sample_model1/`. In the following, we will refer to the directory where a particular model resides as `$MODEL`.

Models are allowed to consult files with paths relative to the `$MODEL` directory, but should under normal circumstances only directly consult file which are located in the `$MODEL` directory or a subdirectory of it. The exception to this is the file `$LOST/lost.pl`. Consulting this file gives access to all the shared APIs (See chapter 3).

To integrate into the framework each model must provide a file called `interface.pl`, which must be located in the same directory as the model. `interface.pl` can then implement various predefined predicates which serves as an entry point of using the model.

The supported interface predicates which a model can provide are:

- `lost_best_annotation/3`.
- `lost_learn/3`

The general form of these predicates is that they take a list of input files, a list of options and the name of an output file. They generally read the input files, do some processing and then write a result to the output file. The type of processing depends on the predicate. The predicate `lost_best_annotation/3` is meant to be used when running the model in prediction mode, so for instance, it might produce a file containing the viterbi path for a given input sequence. The predicate `lost_learn` is meant to be used when training the model and the output file would usually be the learned parameters of the trained model.

Models should declare the *type* associated to the arguments of these predicates. These include,

- The formats of input files, see section **??**.
- The format of the output file, see section **??**.
- Options and default values for options, see section 2.1.2.

2.1.1 Model interface predicates

This section describes predicates, that when implemented by the `interface.pl` provided by a model, allows the model provide functionalities that integrate into the general framework.

`lost_best_annotation(+InputFileNames,+Options,+OutputFilename)`: The framework calls this predicate to obtain a "best annotation" from the model. The model is free to provide this annotation in any way it sees fit. It is the responsibility of the model to save the annotation to `OutputFilename`, before the completion of `lost_best_annotation`.

`InputFileNames`: Is a list of filenames (with absolute paths), each containing an input to the model.

There is no restriction on the format of the files, but the model should declare what file formats it expects. Predicates to parse a wide range of fileformats are supplied in the *io* API (see section 3.2).

`Options`: Is a list of facts on the form, where the functor works as key and the first argument serves as the value of the option, egg. `key(Value)`. This list is use to parameterize the model in various ways. For convenience, option values can be checked an extracted using the predicates `get_option` (see section **??**).

The options used must be declared as described in section .

`OutputFilename`: Full filename which the resulting "best annotation" should be saved to. The model is expected to save the resulting annotation to this file before the completion of `lost_best_annotation`. The *io* API contains some common predicates for saving annotations (see section 3.2).

`lost_learn(+InputFileNames,+Options,+OutputFilename)`

This predicate is used for training models. The model is expected to save the result of the training session (e.g. a switch parameter file or similar) to `OutputFilename`.

`InputFileNames`: Is a list of filenames (with absolute paths), each containing an input to the model. These are used for providing the traning data. There is no restriction on the format of the files, but the model should declare what file formats it expects. Predicates to parse a wide range of fileformats are supplied in the *io* API (see section 3.2).

`Options`:Is a list of facts on the form, where the functor works as key and the first argument serves as the value of the option, egg. `key(Value)`. This list is use to parameterize the model in various ways. For convenience, option values can be checked an extracted using the predicates `get_option` (see section **??**).

`OutputFilename`: Full filename which the resulting switch parameters or similar should be saved to. The model is expected to save the result to this file before the completion of `lost_learn`. The *io* API contains some common predicates for saving annotations (see section 3.2).

2.1.2 Declaration of options for interface predicates

Options to interface predicates should also be declared in the `interface.pl` file. An option is declared by adding a fact `lost_option/4` to the file:

```
lost_option(InterfaceGoal, OptionName, DefaultValue, Description).
```

- `InterfaceGoal`: InterfaceGoal is one of
  - `lost_best_annotation`
  - `lost_learn`
- `OptionName`: An atom for the name of key. This is the functor identifying this particular option.
- `DefaultValue`: If the option is left unspecified then it will take this value.
- `Description`: A textual description of the purpose of the option. Should be in single quotes.

As example, consider an option declaration:

```
lost_option(lost_best_annotation,,default,'A parameter filename').
```

In this declaration, we say that the goal `lost_best_annotation` take the option `parameter_file`. If this option is not a member of the `Options` list, then it will be added before calling `lost_best_annotation`. For instance, assume the model is `sample_model1`, then we might call

```
get_annotation_file(sample_models1,['/some/path/infile.seq'],[],FileOut),
```

which will start a new PRISM process and invoke,

```
lost_best_annotation(['/some/path/infile.seq'],[parameter_file(default)],FileOut),
```

where the option `parameter_file` has been inserted with the default value. A default value is only inserted if the option is left unspecified.

2.1.3 Declaration of file formats for interface predicates

Input file formats

For each model interface predicate the format of input files and output files should be declared as facts in the `interface.pl` file for the model. To declare the format of the input files, a fact,

```
lost_input_format(PredicateName, [ Format1, .., FormatN ]).
```

where `PredicateName` is the name of the predicate to which the declaration belong. The second argument is a list of format identifiers as described in chapter **??**. For instance, we may have a declaration,

```
lost_input(lost_best_annotation,
    [ text(prolog(sequence(dna))), text(prolog(sequence(amino_acids)))]).
```

The last entry of the list of formats may have the special star quantifier, which works similar to a kleene star: It states that the last entry is a placeholder for between zero or more entries of the specified type. For instance,

```
lost_input(lost_best_annotation,
    [ text(prolog(sequence(dna))), star(text(prolog(sequence(dna))))]).
```

specifies that `lost_best_annotation` takes one or more input files of the type `text(prolog(sequence(dna)))`.

Output file formats

Contrary to the input file formats, the output file format can depend on the options given to the model. That means, the model may produce files in different output formats depending on the values of the options it is called with.

The output format is thus declared with a predicate,

```
lost_output_format(PredicateName,Options, Format)
```

This predicate should specify the relation between a set of `Options` and a particular `Format` for the interface predicate `PredicateName`. `Format` should unify to one of the formats described in chapter **??**.

Chapter 3

Lost shared APIs

To use the lost APIs, the file `$LOST/lost.pl` must be consulted. To make the models independent of the absolute path of the `$LOST` directory, they should consult it with a path relative to the model path (e.g. `:- ['../../lost.pl'].`). Then, APIs, which are located in the `$LOST/lib` directory can be consulted using the goal,

```
lost_include_api(+APIName)
```

where `APIName` is the name of a Prolog file located in the `$LOST/lib/` directory except the `.pl` extension.

3.1 The interface API: `interface`

The API provides the interface to lost models following the conventions described in section 2.1.

get_annotation_file(Model, Inputs, Options, Filename)

This API provides `get_annotation_file/4` which is used to retrieve the best annotation generated by a specified model with specified parameters and input sequences. If no such file currently exists, then the model will be run (e.g. the `lost_best_annotation/3` provided by the model will be called).

The generated annotation files are named according to a convention. All annotation files will be placed in the `$LOST/data/` directory. The `Filename` is construed according to the following convention:

```
{Modelname}_{Id}.gen
```

The first time an annotation is generated the file `annotation.idx` will be created in this directory. This file serves as a database to map filenames of the generated annotation files to the (models ,inputs,probability parameters) that generated the particular annotations. This database file contains Prolog facts on the form,

```
fileid(Id,Filename,Model,Options,InputFiles).
```

The annotation index is automatically maintained by `get_annotation/4` and should normally not be edited by hand.

If annotation for a particular run of a model is not present then `get_annotation_file/4` will start a new PRISM process that invokes the `lost_best_annotation` predicate provided be the model `interface.pl` file. By the contract of model conventions, the model will generate the annotation and save it to the file indicated by the provided filename.

3.2 Input-Output API: `io`

In this module (`io.pl`), severals predicates are defined to manipulate `*.seq` files :

- loading information from files that extracts from a file data information used as input of models (sequence annotation for example);
- saving information into a file;
- and maybe more.

3.2.1 Loading information from files

- `load_annotation_from_file(++Type_Info,++Options,++File,--Annotation)`: Generate from `File` a sequence of `Annotation`. It is assumed that `File` is composed of terms. `Type_Info` is used to specify what format of information into file
  - `sequence` means that information is stored into a list. For example,

    ```
    data(Key_Index,1,10,[a,t,c,c,c..]).
    ```

  - `db` means that information is represented by a set of range that specified specific zone (coding region for example)

    ```
    gb(Key_Index,1,10).
    ```

  For each `Type_Info`, several options are available represented by the list `Options`. Options available for `sequence`:

  - [] (default): data list is the 2^{th} argument of the terms and these lists of data are appended;
  - data_position(Num) specified that data list is `Num`th argument of term;
  - `range(Min,Max)` extracts from the list of the complete annotation the sublist from position `Min` to position `Max`;
  - `all_lists`: generate a list of each data list by term. Warning: `range(Min,Max)` is not support by this option.

File Example `toto.seq`:

```
data(Key_Index,1,5,[1,2,3,4,5]).
data(Key_Index,6,10,[6,7,8,9,10]).
data(Key_Index,11,15,[11,12,13,14,15]).
```

Results of request are:

```
| ?- load_annotation_from_file(sequence,[data_position(4)],'toto.seq',R).
R = [1,2,3,4,5,6,7,8,9,10,11,12,13,14,15] ?
| ?- load_annotation_from_file(sequence,[data_position(4),range(4,10)],'toto.seq',R).
R = [4,5,6,7,8,9,10] ?
load_annotation_from_file(sequence,[data_position(4),all_lists],'toto.seq',R).
R = [[1,2,3,4,5],[6,7,8,9,10],[11,12,13,14,15]] ?
```

Options available for `db`:

- [] (default): first and the second element of the term defined a range. A list of 0-1 values is generated, 0 when you are outside ranges and 1 you are inside at least one;
- `in_db(Letter)` replaces the default value 1 by `Letter`;
- `out_db(Letter)` replaces the default value 0 by `Letter`;
- `range_position(Min,Max)` allows to specify the position argument number of a term of the minimal and maximal value of term;
- `range(Min,Max)` extracts from the list of the complete annotation the sublist from position `Min` to position `Max`;

File Example `toto.seq`:

```
gb(3,5).
gb(7,9).
gb(8,11). % Overlap ;)
```

Results of request are:

```
| ?- load_annotation_from_file(db,[],'toto.seq',R).
R = [0,0,1,1,1,0,1,1,1,1,1] ?
| ?- load_annotation_from_file(db,[in_db(c),out_db(nc)],'toto.seq',R).
R =  [nc,nc,c,c,c,nc,c,c,c,c,c]?
| ?- load_annotation_from_file(db,[range(3,7)],R).
R = [1,1,1,0,1] ?
| ?- load_annotation_from_file(db,[in_db(c),out_db(nc),range(8,16)],'toto.seq',R).
R =  [c,c,c,nc,nc,nc,nc,nc]?
```

3.2.2 Saving information to files

The io API also contains some predicates for saving sequences to a file.

- `save_annotation_to_sequence_file(+KeyIndex,+ChunkSize,+Annotation,+File)`: Saves the data in the list given by `Annotation` to the file `File` in *sequence* format which is number of Prolog facts, on the form:

  ```
  data(KeyIndex,1,5, [a,t,c,c,g]).
  ```

  The first argument `KeyIndex` is used as an identifier. `ChunkSize` is the number of elements from `Annotation` to store with each fact. The second and third argument of a stored fact, corresponds to the start and end position in `Annotation`. The fourth argument of the fact is a list containing the relevant elements of the `Annotation` list. Note that if the the length of `Annotation` is not a multiple of `ChunkSize`, then the last fact stored will have a shorter range.

3.3 The Stats API: `stats`

In this module (`stats.pl`), several predicates are defined to automatically compute frequencies, probabilities of occurrences of nucleotides, codons ... This computation is based on a simple counting method given a simple input data.

Variable `Data_Type` is used to specify the type of the input data: Data type available are:

- `nucleotid` where input data are composed of letter from $\{a, c, g, t\}$;
- `codon` where a codon is represented by a list of three letters from $\{a, c, g, t\}$;
- `animo_acid` where input data are composed of letter from $\{a, c, d, e, f, g, h, i, k, l, m, n, p, q, r, s, t, v, w, y\}$;
- `length`. This type is used to compute frequencies and statistics about length of specific region of a genome.

Here is the description of main predicates of `stats.pl`

- stats(++Data_Type,++Options,++Data,++Input_Counting,–Result)
  `Data_Type` setting define on which counting procedure is initialized. User must take care then to give the right `Data`. For {`nucleotid,codon,animo_acid`} data type, `Data` must be a Prolog list with the right element, For `length` data type, `Data` must be a list of ranges represented by a list of two integer values ([Min,Max]). Two options are available:
  - `order(Num)` defines the size of the past stored to perform the counting
  - `past(List)` allows to give as input of the computation a previous past. This option is useful to make connexion between two counting computations.

Input_Counting is used as well to perform a counting computation on an input data divided into a set of lists. By default, counting computation is initialized when is a variable Input_Counting. However, the counting can restart from Input_Counting if this variable is unified to the result of a previous result. Finally, Result has the following format:

```
[(Past1,[(Elt1,Count11),(Elt2,Count12),...]),
 (Past2,[(Elt1,Count21),(Elt2,Count22),...]),
 ...]
```

For example let consider this following data:

```
data('U00096',1,5,[a,g,c,t,t]).
data('U00096',6,10,[c,c,c,c,c]).
```

Here is the result of this following request

```
| ?- stats(nucleotid,[],[a,g,c,t,t],_,R).
R = [([],[(a,1),(c,1),(g,1),(t,2)])] ?
| ?- stats(nucleotid,[order(1)],[a,g,c,t,t],_,R).
R = [([a],[(a,0),(c,0),(g,1),(t,0)]),
     ([c],[(a,0),(c,0),(g,0),(t,1)]),
     ([g],[(a,0),(c,1),(g,0),(t,0)]),
     ([t],[(a,0),(c,0),(g,0),(t,1)])]?
| ?- stats(nucleotid,[order(2)],[a,g,c,t,t],_,R1),
     stats(nucleotid,[order(2),past([t,t])],[c,c,c,c,c],R1,R).
R = [([a,a],[(a,0),(c,0),(g,0),(t,0)]),
     ([a,c],[(a,0),(c,0),(g,0),(t,1)]),
     ([a,g],[(a,0),(c,1),(g,0),(t,0)]),
     ([a,t],[(a,0),(c,0),(g,0),(t,0)]),
     ([c,a],[(a,0),(c,0),(g,0),(t,0)]),
     ([c,c],[(a,0),(c,3),(g,0),(t,1)]),
     ([c,g],[(a,0),(c,1),(g,0),(t,0)]),
     ([c,t],[(a,0),(c,0),(g,0),(t,1)]),
     ([g,a],[(a,0),(c,0),(g,0),(t,0)]),
     ([g,c],[(a,0),(c,0),(g,0),(t,1)]),
     ([g,g],[(a,0),(c,1),(g,0),(t,0)]),
     ([g,t],[(a,0),(c,0),(g,0),(t,1)]),
     ([t,a],[(a,0),(c,0),(g,0),(t,0)]),
     ([t,c],[(a,0),(c,0),(g,0),(t,1)]),
     ([t,g],[(a,0),(c,0),(g,0),(t,0)]),
     ([t,t],[(a,0),(c,1),(g,0),(t,0)]),
     ]?
```

- stats(++Data_Type,++Options,++Data,++Input_Counting,–Past,–Result) stats/6 computes exactly the same counting. This predicate allows only

to record into `Past` the last past of the computing. This predicate is usefull to compute statistics on a series of data input. For example, this following request

```
| ? - stats(nucleotid,[order(2)],[a,g,c,t,t],_,Past,R1),
      stats(nucleotid,[order(2),past(Past)],[c,c,c,c,c],R1,Past2,R).
Past = [t,t],
Past2 = [c,c],
R = ... ?
```

can be used to obtain counting information on the previous data.

- normalize(++Data_Type,++List_Counting,–Probabilities) From the result of a counting procedure, `normalize`/3 modified `List_Counting` to compute domains and probabilities distributions.

For example, here is the result of several requests:

```
| ?- stats(nucleotid,[],[a,g,c,t,t],_,R),
     normalize(nucleotid,R,S).
S = [([],([a,c,g,t],[0.2,0.2,0.2,0.4]))]
| ?- stats(nucleotid,[order(1)],[a,g,c,t,t],_,R),
     normalize(nucleotid,R,S).
S = [([a],([a,c,g,t],[0,0,1,0])),
     ([c],([a,c,g,t],[0,0,0,1])),
     ([g],([a,c,g,t],[0,1,0,0])),
     ([t],([a,c,g,t],[0,0,0,1]))] ?
```

Chapter 4

Models

Pre-defined models are introduced. These models are used to build more complex models.

4.1 Parsers of Biological Data

To extract information from different Biological database, several parsers have been designed to parse report of analyses available on different web-servers (Easygene, Genemark) and database (Genbank). These parsers generated a series of Prolog terms that can be used after that input of different probabilistic models. `script_parser.pl` collects different scripts to generate different data files.

4.1.1 Parser_fna

*.fna file of Genbank is composed of a complete genome in the FASTA format. Parser_fna permits to parse this *fna.file from Genbank and generate list of terms. These terms store the genome into a Prolog list composed of $\{a, c, g, t\}$. Two scripts are implemented:
`parser_fna(++Name_FNA_File,++Name_GBK_File,++Options)` and
`parser_fna(++Name_FNA_File,++Name_GBK_File,++Options,--OutputFile)` Note that *.gbk file is necessary as well. This file is used to automatically extract genome information (Genbank key and size of genome).

Output format of the generated terms are:
`data(Genebank_Key,Start,End,List_of_Data)`. Option `list(Number)` can be used to divide the complete genome into several lists with a length defined by the parameter `Number`. Example with the E.Coli K12 genome:

```
>gi|48994873|gb|U00096.2| Escherichia coli .....
AGCTTTTCATTCTGACTGCAACGGGCAATATGTCTCTGTGTGG.....
.....
```

Result by default:

```
%>gi|48994873|gb|U00096.2| Escherichia coli ....
data('U00096',1,4639675,[a,g,c,t,t,t, ...]).
```

Result when `list(280)` option is used

```
%>gi|48994873|gb|U00096.2| Escherichia .....
data('U00096',1,280,[a,g,c,t,...]).
data('U00096',281,560,[c,c,c,...]).
....
```

4.1.2 Parser_ptt

*.ptt file of Genbank is composed of information about known or predicted genes in a genome. Parser_ptt parses this *.pt file from Genbank and generate list of terms. Two scripts are implemented:
`parser_ptt(++Name_PTT_File)` and
`parser_fna(++Name_PTT_File,,--OutputFile)`

Output format of the generated terms are:
`gb(Genebank_Key,Start,End)`.

4.1.3 Parser_Easygene

4.1.4 Parser_Genemark

4.2 Models for measurement and statistical reports

4.2.1 accuracy_report

The model `accuracy_report` can be used the produce a report of various measures of the accuracy of particular gene predictions compared with a golden standard such a genebank.

To use the the model, you need to call `get_annotation_file/4` with the following arguments,

```
get_annotation_file(accuracy_report,
    [ReferenceFile,PredictionFile],
    [
     option(reference_functor,RefFunctor),
     option(prediction_functor,PredFunctor),
     option(start,StartPos),
     option(end,EndPos)
    ],
    OutputFile),
```

`ReferenceFile` must be the full path to a file with facts representing the "correct predictions". `PredictionFile` must be the full path to a file with facts representing the predictions. Both files must be a a `db` type format, with facts on the following form,

```
functor(To, From, Strand, ReadingFrame, Name).
```

Each such fact represent a prediction in the `PredictionFile` or correct gene in the in `ReferenceFile`. The `functor` is a any given functor, but the `ReferenceFile` and the `PredictionFile` should use different functors. The *To* argument represents the position in the genome where the prediction begins (inclusive) and the `From` argument represents the end position of the prediction `ReadingFrame` is a integer in the range $\{1, 2, 3\}$ `Strand` is either `+` for the forward strand or `-` for the reverse strand.

`get_annotation_file` for the `accuracy_report` model must be called with four mandatory options:

- `reference_functor`: The functor used in the `ReferenceFile`
- `prediction_functor`: The functor used in the `PredictionFile`
- `start`: An integer corresponding to the beginning of the range on which accuracy should be measured.
- `end`: An integer corresponding to the end of the range (inclusive) on which accuracy should be measured.

Chapter 5

File formats

The chapter documents the different types of file formats used in the lost framework. The naming of file formats follow a simple scheme using nested Prolog functors where the outermost functor is the most general and the innermost is the most specific description of the file format. For instance, consider the naming for a DNA sequence expressed as a Prolog file,

```
text(prolog(sequence(dna)))
```

The outermost functor `text` specifies that the file is a text file, and the next functor `prolog` says that the text file contains Prolog code. The next one, `sequence`, specifies the type of data (sequence data) we expect to be expressed in the prolog facts finally, the innermost functor `dna` specifies the type of sequence that we are dealing with.

5.1 text(prolog(_))

`text(prolog(_))` formats contain Prolog facts. The functor and arity of those facts cannot be determined by knowing that it is `text(prolog(_))`, but needs specification using further embbed functors.

5.1.1 text(prolog(sequence(_)))

This format should be specified for sequence data expressed as prolog facts. For instance, a file of this format may contain facts like the ones below, which specifies the alphabet,

```
data(alphabet,1,10, [a,b,c,d,e,f,g,h,i,j]).
data(alphabet,11,20],[k,l,m,n,o,p,q,r,s,t]).
data(alphabet,21,27,[u,v,w,x,y,z]).
```

The form of these facts are,

```
functor(Identifier,To,From,SequenceElementList).
```

The format does not dictate the `functor` of the facts (e.g. `data`), nor the size of the data list in the fourth argument. However, the range expressed by `To` and `From` should correspond to the numer of elements in `SequenceElementList`.

`text(prolog(sequence(dna)))`

Like the above format but restricts the alphabet of the data elements in the `SequenceElementList` to the set {`a,g,c,t`}.

`text(prolog(sequence(rna)))`

Like the above format but restricts the alphabet of the data elements in the `SequenceElementList` to the set {`a,g,c,u`}.

`text(prolog(sequence(amino_acids)))`

Like the above format but restricts the alphabet of the data elements in the `SequenceElementList` to the set {`a,c,d,e,f,g,h,i,k,l,m,n,p,q,r,s,t,v,w,y`}.

5.1.2 `text(prolog(ranges(_)))`

The format consists of Prolog facts, each of which contain an annotation for a particular range of some sequence

```
F =.. [ Functor, LeftEnd, RightEnd | _ ]
```

The functor may vary depending on the type of annotation but is expected to be same within a file. The first two arguments, `LeftEnd` and `RightEnd` are positive integers which specifies the range relative to the sequence. Both are inclusive.

`text(prolog(ranges(gene)))`

The format consists of Prolog facts, each of which contain an gene annotation for a particular range of some DNA sequence (all facts refer to the same sequence).

The facts are on the form,

```
functor(LeftEnd,RightEnd,Strand,Frame,ExtraList).
```

The functor may vary depending on the type of annotation but is expected to be same within a file. The first two arguments, `Start` and `End` are positive integers which specifies the range of the annotation relative to the DNA sequence. The `Strand` argument is `+` for the forward strand or`-` for the reverse strand. The `Frame` argument is one of {`1,2,3`}. The final argument, `ExtraList` is a list possibly containing extra information.

5.1.3 `text(prolog(prism_switches))`

This is the format used by PRISM to save and load parameter files.

Chapter 6

Notes and stuff

6.1 Feature wish list

- A `data` directory instead of a `sequences` directory.
- Division of models into models (probabilistic models) and nodes (which are just data processing).
- kind of `make.pl` file that manages the consulting of all the useful file (lost, /lib/*.*, /scripts/*.*). Just let to the user to do consulting of model file. If it is possible, have a very simple interaction with the user to set the lost location, set platform, or list models or data available.
- Naming stuff: `load_annotation_from_file -> load_data_from_file`
- Utility for removing generated sequence files
- Document gene database file format and make sure that all models adhere to this format.
- Default option value declarations for models.