

1.1 Introduction

1.1.1 Motivation

This work has been motivated by recent advances of molecular genetics. The human genome has been sequenced in 2001. Also mouse, drosophila, etc. Nowadays # reference model genomes are available in genbank.

Next-generation sequencing has been the second revolution. NGS produces billions of reads for 1000\$ dollars. Why should one re-sequence a known genome? Resequencing applications include variant calling, etc. So NGS impacts biomedicine.

Given a set of reads, two approaches are possible: assembly and mapping.

Assembly methods are based on overlaps, de brujin graphs, or...

Read mapping methods work on a previously assembled reference genome.

The typical SNPs analysis pipeline 1.1 consists of...

In this work we focus on read mapping, although many core algorithms considered are also applicable to assembly, as well as to later pipeline stages.

Figure 1.1: NGS pipeline.

- Plan A: de-novo assembly
- Plan B: reference mapping
- Plan C: reference guided de-novo assembly

1.1.2 Organization of this manuscript

1.2 Stringology preliminaries

We now introduce fundamental definitions and problems of stringology, in order to keep the manuscript self-contained. The reader familiar with basic stringology can skip this section and proceed to section ??.

1.2.1 Definitions

Let us start by defining primitive objects of stringology: alphabets and strings. An alphabet is a finite set of symbols (or characters); a string (or word) over an alphabet is a finite sequence of symbols from that alphabet. We denote the length of a string s by $|s|$, and by ϵ the empty string s.t. $|\epsilon| = 0$. Given an alphabet Σ , we define $\Sigma^0 = \{\epsilon\}$ as the set containing the empty string, Σ^n as the set of all strings over Σ of length n , and $\Sigma^* = \bigcup_{n=0}^{\infty} \Sigma^n$ as the set of all strings over Σ . Finally, we call any subset of Σ^* a language over Σ .

We now define concatenation, the most fundamental operation on strings. The concatenation operator of two strings is denoted with \cdot and defined as $\cdot : \Sigma^* \times \Sigma^* \rightarrow \Sigma^*$. Given two strings, $x \in \Sigma^m$ with $x = x_1x_2 \dots x_m$, and $y \in \Sigma^n$ with $y = y_1y_2 \dots y_n$, their concatenation $x \cdot y$ (or simply denoted xy) is the string $z \in \Sigma^{m+n}$ consisting of the symbols $x_1x_2 \dots x_my_1y_2 \dots y_n$.

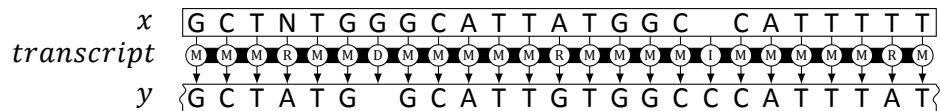
From concatenation we can derive the notion of prefix, suffix, and substring. A string x is a prefix of y iff there is some string z s.t. $y = x \cdot z$. Analogously, x is a suffix of y iff there is some string z s.t. $y = z \cdot x$. Moreover, x is a substring of y iff there is some string w, z s.t. $y = w \cdot x \cdot z$, and then we say that x occurs within y at position $|w|$.

Example 1.1. These definitions allow us to model basic biological sequences. Let us consider the alphabet consisting of DNA bases: $\Sigma = \{A, C, G, T\}$. Examples of strings over Σ are $x = A$, $y = AGGTAC$, $z = TA$. For instance, $y \in \Sigma^6$ and $|y| = 6$. Moreover, the concatenation $x \cdot z$ produces ATA . The string x is a prefix of y , and the string z is a substring of y occurring at position 4 in y .

1.2.2 Alignments

The next step is to define the minimal set of edit operations to transform one string into another: substitutions, insertions and deletions. Given two strings x, y of equal length n , the string x can be transformed into the string y by substituting (or replacing) all symbols x_i s.t. $x_i \neq y_i$ into y_i , for $1 \leq i \leq n$. If the given strings have different lengths, insertion and deletion of symbols from x become necessary to transform it into y . Therefore, given any two strings x, y , we define as edit transcript for x, y any finite sequence of substitutions, insertions and deletions transforming x into y . See Figure 1.2 for an example.

Figure 1.2: Example of edit transcript transforming the string $x = AAAA$ into $y = CCCC$. The transcript character M indicates a match, R a replacement, I an insertion, and D a deletion.



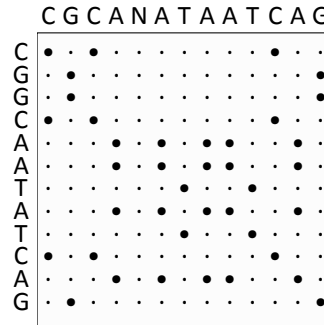
An alignment is an alternative yet equivalent way of visualizing a transformation between strings. While an edit transcript provides an explicit sequence of edit operations

transforming one string into another, an alignment relates pairs of corresponding symbols between two strings. Because some symbols in one string are not related to any symbol in the other string, i.e. some symbols are inserted or removed, we first need to introduce a gap symbol $-$, which is not part of the string alphabet Σ . Subsequently, we can define the alignment of two strings of length m, n over Σ to be a string of length between $\min\{m, n\}$ and $m + n$ over the pair alphabet $(\Sigma \cup \{-\}) \times (\Sigma \cup \{-\})$.

Example 1.2. An alignment of the strings $x = AAAAA$ and $y = CCCCC$ is given by the string $z = \begin{pmatrix} A \\ A \end{pmatrix} \begin{pmatrix} A \\ - \end{pmatrix} \begin{pmatrix} A \\ C \end{pmatrix} \begin{pmatrix} G \\ G \end{pmatrix}$

A dotplot is a way to visualize any alignment between two strings and highlight their similarities. Given two string x, y of length m, n , a dotplot is a $m \times n$ matrix containing a dot at position (i, j) iff the symbol x_i matches symbol y_j . We define a dotplot trace to be a monotonical path in the matrix connecting non-decreasing positions of the matrix. A dotplot trace corresponds to an alignment and vice versa. In a trace, match and mismatch columns of the corresponding alignment appear as diagonal stretches, while insertions and deletions are horizontal or vertical stretches. See Figure ??.

Figure 1.3: Example of dotplot of the strings $x = AAAAA$ and $y = CCCCC$. The highlighted trace corresponds to the alignment of Example 1.2.



1.2.3 Distance functions

We can assign a cost to any alignment and to its associated edit transformation by defining a weight function $\omega : (\Sigma \cup \{-\}) \times (\Sigma \cup \{-\}) \rightarrow \mathbb{R}_0^+$, where:

- $\omega(\alpha, \beta)$ for all $(\alpha, \beta) \in \Sigma \times \Sigma$ defines the cost of substituting α with β ,
- $\omega(\alpha, -)$ for all $\alpha \in \Sigma$ defines the cost of deleting the symbol α ,
- $\omega(-, \beta)$ for all $\beta \in \Sigma$ defines the cost of inserting the symbol β ,

and by defining the total cost $C(z)$ of an alignment z between two strings as the sum of the weights of all its alignment symbols:

$$C(z) = \sum_{i=0}^{|z|} \omega(z_i) \quad (1.1)$$

Consequently, we can define the distance function $d : \Sigma^* \times \Sigma^* \rightarrow \mathbb{R}_0^+$ by taking the minimum cost over all possible alignments of x, y :

$$d(x, y) = \sum_{z \in A(x, y)} C(z) \quad (1.2)$$

In particular, the edit or *Levenshtein distance* between two strings $x, y \in \Sigma^*$ is defined as the function $d_E : \Sigma^* \times \Sigma^* \rightarrow \mathbb{N}_0$ counting the *minimum* number of edit operation necessary to transform x into y . It is obtained by defining for all $(\alpha, \beta) \in \Sigma \times \Sigma$, $\omega(\alpha, \beta) = 1$ iff $\alpha \neq \beta$ and 0 otherwise, and $\omega(\alpha, -)$ and $\omega(-, \beta)$ as 1. The *Hamming distance* between two strings $x, y \in \Sigma^n$ is defined as the function $d_H : \Sigma^n \times \Sigma^n \rightarrow \mathbb{N}_0$ counting the number of substitutions necessary to transform x into y . We obtain it by defining $\omega(\alpha, \beta)$ as in the edit distance, and by setting all $\omega(\alpha, -)$ and $\omega(-, \beta)$ to be ∞ in order to disallow indels.

Example 1.3. TODO: example of edit and hamming distance.

1.2.4 Optimal alignments

The problem of finding an optimal alignment between two strings is equivalent to the problem of finding their minimum distance [Gusfield, 1997]. A solution to this optimization problem can be efficiently computed via dynamic programming (DP). Below we describe the three essential components of the DP approach: the recurrence relation, the DP table, and the traceback.

Given two strings x, y of length m, n , for all $1 \leq i \leq m$ and $1 \leq j \leq n$ we define with $d(x_{1..i}, y_{1..j})$ the distance between their prefixes $x_{1..i}$ and $y_{1..j}$. The base conditions of the recurrence relation are:

$$d(\epsilon, \epsilon) = 0 \quad (1.3)$$

$$d(x_{1..i}, \epsilon) = \sum_{l=1}^i \omega(x_l, -) \text{ for all } 1 \leq i \leq m \quad (1.4)$$

$$d(\epsilon, y_{1..j}) = \sum_{l=1}^j \omega(-, y_l) \text{ for all } 1 \leq j \leq n \quad (1.5)$$

and the recursive case is defined as follows:

$$d(x_{1..i}, y_{1..j}) = \min \begin{cases} d(x_{1..i-1}, y_{1..j}) & + \omega(x_i, -) \\ d(x_{1..i}, y_{1..j-1}) & + \omega(-, y_j) \\ d(x_{1..i-1}, y_{1..j-1}) & + \omega(x_i, y_j) \end{cases} \quad (1.6)$$

We can compute the above recurrence relation in time $\mathcal{O}(nm)$ using a dynamic programming table D of $(m+1) \times (n+1)$ cells, where cell $D[i, j]$ stores the value of $d(x_{1..i}, y_{1..j})$. The sole distance without any alignment can be computed in space $\mathcal{O}(\min\{n, m\})$, as we only need column $D[: j - 1]$ to compute column $D[: j]$ (or row $D[i - 1 :]$ to compute

$D[i : j]$) and we can fill the table D either column-wise or row-wise¹. An optimal alignment can be computed in time $\mathcal{O}(m + n)$ via *traceback* on the table D : We start in the cell $D[m, n]$ and go backwards (either left, up-left, or up) to the previous cell by deciding which condition of Equation 1.6 yielded the value of $D[m, n]$.

Figure 1.4: DP table representing the computation of the edit distance $d_E(x_{1..5}, y_{1..4})$.

| ϵ | C | G | C | A | N | A | T | A | T | C | A | G | | |
|------------|---|---|---|---|---|---|---|---|---|---|----|----|----|----|
| ϵ | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 |
| C | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| G | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| G | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| C | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| A | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| A | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| T | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | • | | | | |
| T | | | | | | | | | | | | | | |
| A | | | | | | | | | | | | | | |
| T | | | | | | | | | | | | | | |
| C | | | | | | | | | | | | | | |
| A | | | | | | | | | | | | | | |
| G | | | | | | | | | | | | | | |

1.3 Overview of string matching

1.3.1 Problem definition

We can now define *exact string matching*, perhaps the most fundamental problem in stringology. Given a string p (with $|p| = m$) called the *pattern* and a longer string t (with $|t| = n$) called the *text*, the exact string matching problem is to find all occurrences, if any, of pattern p into text t [Gusfield, 1997]. This problem has been extensively studied from the theoretical standpoint and is well solved in practice.

The definition of distance functions between strings let us generalize exact string matching into a more challenging problem: *approximate string matching*. Given a text t , a pattern p , and a *distance threshold* $k \in \mathbb{N}$, the approximate string matching (a.s.m.) problem is to find all occurrences of p into t within distance k . The a.s.m. problem under the Hamming distance is commonly referred as the *k-mismatches* problem and under the edit distance as the *k-differences* problem. For *k-mismatches* and *k-differences*, it must hold $k > 0$ as the case $k = 0$ corresponds to exact string matching, and $k < m$ as a pattern occurs at any position in the text if we substitute all its m characters. Under these distances, we define the *error rate* as $\epsilon = \frac{k}{m}$, with $0 < \epsilon < 1$ given the above conditions, and we alternatively refer to these a.s.m. problems as ϵ -mismatches and ϵ -differences.

We can classify string matching problems in two categories, *online* and *offline*, depending on which string, the pattern or the text, is given first. Algorithms for online string matching work by preprocessing the pattern and scanning the text from left to right (or right to left); their worst-case runtime complexity ranges from $\mathcal{O}(nm)$ to $\mathcal{O}(n)$ while their worst-case memory complexity ranges from $\mathcal{O}(\sigma^k m^k)$ to $\mathcal{O}(m)$. Algorithms

¹ Note that D can be filled also diagonal-wise or antidiagonal-wise.

for offline string matching are instead allowed to preprocess the text; their worst-case runtime complexity ranges from $\mathcal{O}(m)$ to $\mathcal{O}(\sigma^k m^k)$ while their worst-case memory complexity is usually $\mathcal{O}(n)$. In practice, if the text is long, static and searched frequently, offline methods largely outperform online methods in terms of runtime, provided the necessary amount of memory. Therefore, we concentrate on offline algorithms.

We can subdivide algorithms for offline string matching in two categories: *fully-indexed* and *filtering*. Fully-indexed algorithms work solely on the index of the text, while filtering methods first use the index to discard uninteresting portions of the text and subsequently use an online method to verify narrow areas of the text. Filtering methods outperform fully-indexed methods for a vast range of inputs² and are thus very interesting from a practical standpoint. Nonetheless, filtering methods are just opportunistic combinations of online and fully-indexed methods.

In the following of this section we thus give a brief and non-exhaustive overview of the fundamental techniques for online and (both fully-indexed and filtering) offline string matching. This overview serves as an introduction to the more involved algorithms presented in chapter ?? and directly used in applications of part ?. For an extensive treatment of the subject, we refer the reader to complete surveys on exact [?] and approximate [?] online string matching methods, as well as to a succinct survey on indexed methods [Navarro *et al.*, 2001].

1.3.2 Online methods

We consider two classes of algorithms for online string matching, those based on automata and those based on dynamic programming.

Automata

Exact search of one pattern. Knuth-Morris-Pratt automaton.

Exact search of multiple patterns. Aho-corasick automaton.

Approximate search of one pattern. Ukkonen automaton.

Dynamic programming

The dynamic programming algorithm ?? to compute the distance of two strings can be easily turned into a string matching algorithm. Since an occurrence of the pattern can start and end anywhere in the text, a.s.m. consists of computing the edit distance between the pattern and all substrings of the text. The problem can be thus solved by computing the edit distance between the text and the pattern without penalizing leading and trailing deletions in the text.

We pose $x = p$ and $y = t$ and consider Equation 1.6. Since an occurrence of the pattern can start anywhere in the text, we change the initialization of the top row $D[0 :]$ of the DP matrix according to the condition:

$$d(\epsilon, y_{1..j}) = 0 \text{ for all } 1 \leq j \leq n \quad (1.7)$$

² When the error rate is low.

and since an occurrence of the pattern can end anywhere in the text, we check all cells $D[m, j]$ for all $1 \leq j \leq n$ in the bottom row of D for the condition $D[m, j] \leq k$.

Figure 1.5: DP table representing the match of $p = \dots$ in $t = \dots$

| | ϵ | C | G | C | A | N | A | T | A | A | T | C | A | G | A | A | A |
|------------|------------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| ϵ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| C | 1 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | | | | |
| G | 2 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | | | | |
| G | 3 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | | | | |
| C | 4 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | | | | |
| A | 5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | | | | |
| A | 6 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | • | | | | | | | |

1.3.3 Indexed methods

No matter how efficient online methods can be, these approaches quickly become impractical when the text is long and searched frequently. If the text is static and given in advance, we are allowed to preprocess it. Therefore we build an index of the text beforehand and use it to speed up subsequent searches. To this intent we introduce the *suffix tree*, an optimal data structure to index all substrings of a text. We take the suffix tree as an idealized data structure to elegantly expose our indexed algorithms solving string matching problems. In chapter ?? we will consider other substring indices to replace the suffix tree in practice.

Suffix tree and suffix trie

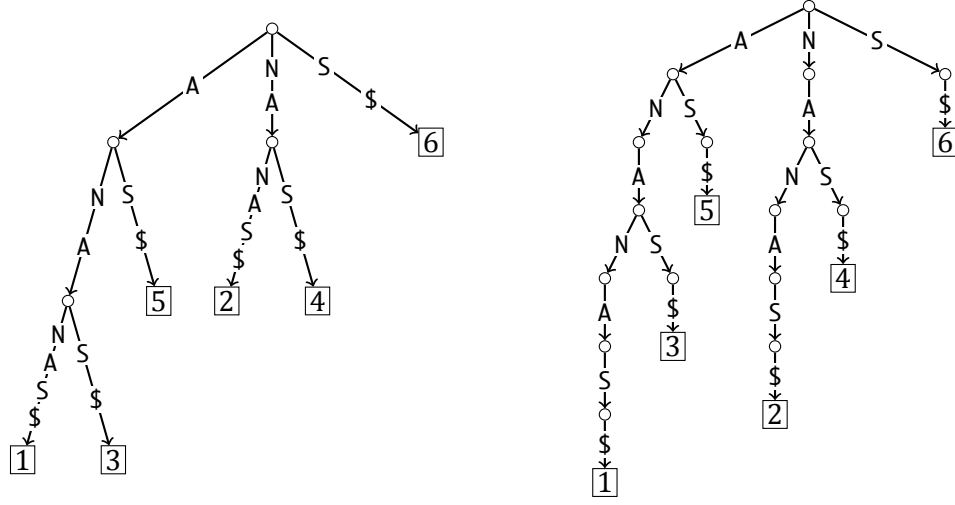
The suffix tree [Morrison, 1968] is a lexicographically ordered tree data structure representing all suffixes of a string. Assume w.l.o.g. a string s of length n , padded with a *terminator symbol* $\$$ not being part of the string alphabet Σ^3 . The suffix tree \mathcal{S} of the string s has one node designated as the root and n leaves, each one pointing to a distinct suffix of s , denoted as $l_1 \dots l_n$. Each internal node has more than one child, and each edge is labeled with a non-empty substring of s . Each path from the root to a leaf l_i spells the suffix $s_{i..n}$. Figure 1.6 illustrates.

In the following of this manuscript we consider w.l.o.g. *suffix tries* instead of suffix trees. On suffix tries, internal nodes can have only one child and each edge is labeled by one single character (see Figure ??). This fact simplifies the exposition of all given algorithms without affecting their runtime complexity nor their result. However, we remark that all given algorithms can be generalized to work on trees.

Therefore, from now on we assume the text t to be indexed using a suffix trie \mathcal{T} . Given a node x of \mathcal{T} , we denote with $label(x)$ the label of the edge entering into x , with $\mathbb{C}(x)$ the set of children of x being internal nodes, with $\mathbb{E}(x)$ the set of children of x being leaves, and with $\mathbb{L}(x)$ the set of all leaves of the subtree rooted in x . We remark that entering edges of internal nodes in $\mathbb{C}(x)$ are always labeled with symbols in Σ , while entering edges of leaves in $\mathbb{L}(x)$ and $\mathbb{E}(x)$ are always labeled with terminator symbols.

³ The terminator symbol is necessary to ensure that no suffix $s_{i..n}$ is a prefix of another suffix $s_{j..n}$.

Figure 1.6: Suffix tree and suffix trie for the string ANANAS.



Exact string matching

Using the suffix trie \mathcal{T} of the text t , we can find all occurrences of a pattern p into t in optimal time $\mathcal{O}(m)$, thus independently of n . Algorithm 1.1 searches a pattern p by starting in the root node of \mathcal{T} and following the path spelling the pattern. If the search ends up in a node x , each leaf $l_i \in \mathbb{L}(x)$ points to a distinct suffix $t_{i..n}$ such that $t_{i..i+m} = p$. Algorithm 1.1 is correct since each path from the root to any internal node of the suffix trie \mathcal{T} spells a different unique substring of t ; consequently all equal substrings of t are represented by a single common path.

Algorithm 1.1 Exact string matching on a suffix trie.

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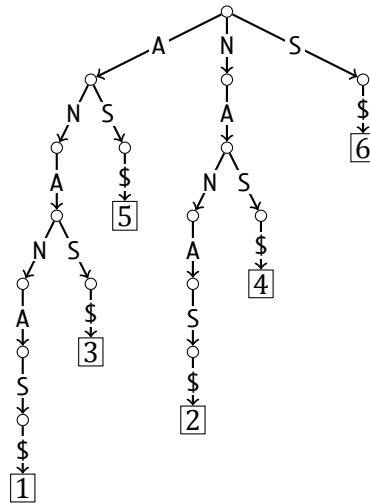
1: procedure EXACTSEARCH( $x, p$ )
2:   if  $p = \epsilon$  then
3:     report  $\mathbb{L}(x)$ 
4:   else if  $\exists c_x \in \mathbb{C}(x) : \text{label}(c_x) = p_1$  then
5:     EXACTSEARCH( $c_x, p_{2..|p|}$ )
6:   end if
7: end procedure

```

Backtracking k -mismatches

We can solve k -mismatches by backtracking [Ukkonen, 1993; Baeza-Yates and Gonnet, 1999] on the suffix trie \mathcal{T} , in average time sublinear in n [Navarro and Baeza-Yates, 2000]. A top-down traversal on \mathcal{T} spells incrementally all distinct substrings of t . While traversing each branch of the trie, we incrementally compute the distance between the query and the spelled string. If the computed distance exceeds k , we stop the traversal and proceed on the next branch. Conversely, if we completely spelled the pattern p and we ended

Figure 1.7: Exact string matching on a suffix tree. The pattern NA is searched exactly in the text ANANAS.



up in a node x , each leaf $l_i \in \mathbb{L}(x)$ points to a distinct suffix $t_{i..n}$ such that $d_H(t_{i..i+m}, p) \leq k$. See algorithm 1.2.

Algorithm 1.2 k -mismatches on a suffix trie.

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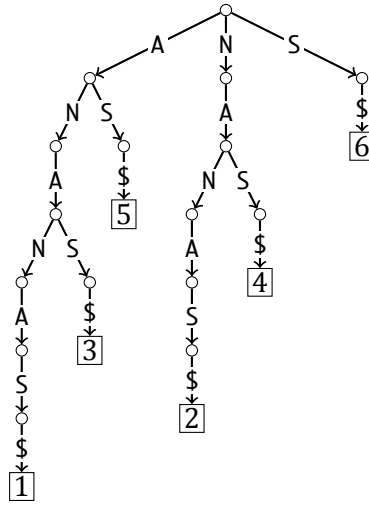
1: procedure KMISMATCHES( $x, p, e$ )
2:   if  $e = 0$  then
3:     EXACTSEARCH( $x, p$ )
4:   else
5:     for all  $c_x \in \mathbb{C}(x)$  do
6:       if  $\text{label}(c_x) = p_1$  then
7:         KMISMATCHES( $c_x, p_{2..|p|}, e$ )
8:       else
9:         KMISMATCHES( $c_x, p_{2..|p|}, e - 1$ )
10:      end if
11:   end if
12: end procedure

```

Backtracking k -differences

We can compute k -differences on a suffix tree in two different ways. Algorithm 1.3 explicitly enumerates errors by recursing on the suffix trie. Algorithm 1.4 computes the edit distance on the suffix trie.

In algorithm 1.4, any online method can be used to compute the edit distance. However, for theoretical considerations, it is important to consider an algorithm which computes in $\mathcal{O}(1)$ per node. Note that it is sufficient to have an algorithm capable of checking whether the current edit distance is within the imposed threshold k .



```

1: procedure KDIFFERENCES( $x, p, e$ )
2:   if  $e = 0$  then
3:     EXACTSEARCH( $x, p$ )
4:   else
5:     KDIFFERENCES( $x, p_{2..|p|}, e - 1$ )
6:     for all  $c_x \in \mathbb{C}(x)$  do
7:       KDIFFERENCES( $c_x, p, e - 1$ )
8:       if  $\text{label}(c_x) = p_1$  then
9:         KDIFFERENCES( $c_x, p_{2..|p|}, e$ )
10:      else
11:        KDIFFERENCES( $c_x, p_{2..|p|}, e - 1$ )
12:      end if
13:   end if
14: end procedure

```

Algorithm 1.4 k -difference on a suffix trie.

```

1: procedure KDIFFERENCES( $x, p, e$ )
2:   for all  $c_x \in \mathbb{C}(x)$  do
3:     KDIFFERENCES( $c_x, p_{2..|p|}, e - d_E(\text{repr}(x), p)$ )
4: end procedure

```

Filtering methods work under the assumption that given patterns occur in the text with a *low average probability*. Such probability is a function of the error rate ϵ , in addition to the alphabet size σ , and can be computed or estimated under the assumption of the text being generated by a specific random source. Under the uniform Bernoulli model, where each symbol of Σ occurs with probability $\frac{1}{\sigma}$, Navarro and Baeza-Yates [2000] estimates that $\epsilon < 1 - \frac{1}{\sigma}$ is a conservative bound on the error rate which ensures few matches, and for which filtering algorithms are effective. For higher error rates, non-filtering on-line and indexed methods work better.

We call a filter *lossless* or *full-sensitive* if it guarantees not to discard any occurrence of the pattern, otherwise we call it *lossy*. Lossy filters can be designed to solve approximately ϵ -differences. We focus our attention on lossless filters.

We now consider two classes of filtering methods: those based on *seeds* and those based on q -grams. Filters based on seeds partition the pattern into *non-overlapping* factors called seeds. We can derive full-sensitive partitioning strategies by application of the pigeonhole principle. Instead, filters based on q -grams consider all *overlapping* substrings of the pattern having length q , the so-called q -grams. Eventually, simple lemmas gives us lower bounds on the number of q -grams that must be present in a narrow window of the text as necessary condition for an occurrence of the pattern.

Seed filters

We start by considering the case of two arbitrary strings x, y s.t. $d_E(x, y) \leq k$. If we partition w.l.o.g. y into $k + 1$ non-overlapping seeds, then at least one seed will occur as a factor of x .

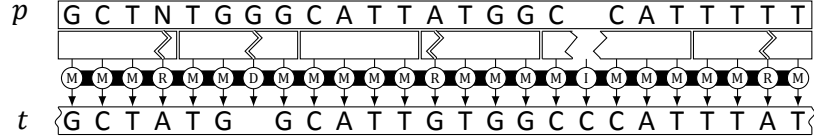
Lemma 1.1. [Baeza-Yates and Navarro, 1999] *Let x, y be two strings s.t. $d_E(x, y) = k$ for some $k \in \mathbb{N}_0$. If $y = y^1 y^2 \dots y^{k+1}$ then $x = ay^i b$ for some a, b .*

Proof. We proceed by induction on k . For $k = 0$, the string y is partitioned into only one factor, y itself. The condition $d_E(x, y) = 0$ implies $x = y$, which is true for $a = \epsilon$ and $b = \epsilon$. We suppose the case $k = j - 1$ to be true, thus since $d_E(x, y) = j - 1$ and $y = y^1 y^2 \dots y^j$ then $x = ay^i b$ for some a, b . We consider the case $k = j$. The j -th error can be in (i) $y^1 \dots y^{i-1}$, (ii) y^i , or (iii) $y^{i+1} \dots y^j$. In case i or iii, $x = ay^i b$ clearly holds. In case ii, if we partition y^i in two factors $y^{i'}$ and $y^{i''}$, then either $x = ay^{i'} b'$ or $x = a'y^{i''} b$. \square

Thus, we solve k -differences by partitioning the pattern into $k + 1$ seeds and searching all seeds into the text, e.g. with the help of a substring index. Figure 1.9 shows an example. Note that we are reducing one approximate search into many smaller exact

searches. As Lemma 1.1 gives us a necessary but not sufficient condition, we must verify whether any candidate location induced by an occurrence of some seed corresponds to an approximate occurrence of the pattern in the text. Thus, we verify any substring s of the text of length $m - k \leq |s| \leq m + k$ containing one seed of p .

Figure 1.9: Filtration with exact seeds.



How many verifications we expect to have?

$$p_\alpha = \frac{1}{\sigma} \text{ for all } \alpha \in \Sigma \quad (1.8)$$

$$\Pr(H > 0) = \frac{1}{\sigma^q} \quad (1.9)$$

$$E(H) = \sum_{i=1}^{n-q+1} \Pr(H > 0) = \frac{n-q+1}{\sigma^q} \leq \frac{n}{\sigma^q} \quad (1.10)$$

$$E(V) = (k+1) \cdot E(H) < \frac{n(k+1)}{\sigma^q} \quad (1.11)$$

Which is the runtime of the algorithm?

How to choose the partitioning? Which length of q makes filtration lossless? $q = \lfloor \frac{m}{k+1} \rfloor$.

q -Gram filters

q -Gram filters are based on the so-called q -gram similarity measure $\tau_q : \Sigma^* \times \Sigma^* \rightarrow \mathbb{N}_0$, defined as the number of substrings of length q common to two given strings. The following lemma relates q -gram similarity to edit distance⁴. It gives a lower bound on the q -gram similarity $\tau_q(x, y)$ for any two strings x, y for which $d_E(x, y) = k$. This means that $\tau_q(x, y) \geq k$ is a necessary but not sufficient condition for $d_E(x, y) \leq k$.

Lemma 1.2. [?] *Let x, y be two strings with edit distance k and $\min\{|x|, |y|\} = m$, then x and y have q -gram similarity $\tau_q(m, k) \geq m - q + 1 - kq$.*

Proof. By induction on k . □

How can we use this result to solve approximate string matching? The lemma itself does not give us the direct solution, indeed it considers the edit distance between two arbitrary strings, while in a.s.m. the pattern can match any substring of the text. In the case of Hamming distance, if the pattern matches any substring s of t , then s must have

⁴ Thus it relates q -gram similarity also to Hamming distance.

Online methods

The problem can be solved by checking whether $d_E(d, q) \leq k$ for all $d \in \mathbb{D}$. Answering the question whether the distance $d_E(d, q) \leq k$ is an easier problem than computing the edit distance $d_E(d, q)$: a band of size $k + 1$ is sufficient.

Lemma 1.3. *The k -differences global alignment problem can be solved by computing only a diagonal band of the DP matrix of width $k + 1$, where the leftmost band diagonal is $\lfloor \frac{m-n+k}{2} \rfloor$ cells left of the main diagonal (see Figure ??).*

Proof. Indirect. Assume that a cell outside the band is part of a global alignment with at most k errors. If the cell is left of the band, the traceback that starts in the top left corner would go down at least $c = \lfloor \frac{m-n+k}{2} \rfloor + 1$ cells. Then it needs to go right at least $n - m + c$ cells to end in the bottom right corner. Hence it contains at least $n - m + 2c > n - m + 2 \frac{m-n+k}{2} = k$ errors. The assumption that the cell is right of the band can be falsified analogously. \square

Figure 1.12: DP table representing the match of $p = \dots$ in $t = \dots$

| ϵ | C | G | C | A | N | A | T | A | T | C | A | G | | |
|------------|---|---|---|---|---|---|---|---|---|---|----|----|----|----|
| ϵ | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | | | | | | |
| C | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | | | | | |
| G | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | | | |
| G | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | | | |
| C | | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | | |
| A | | | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | |
| A | | | | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 |
| T | | | | | | | | | | | | | | |
| T | | | | | | | | | | | | | | |
| T | | | | | | | | | | | | | | |
| C | | | | | | | | | | | | | | |
| A | | | | | | | | | | | | | | |
| G | | | | | | | | | | | | | | |

Indexed methods

Using a radix tree \mathcal{D} we can find all strings in \mathbb{D} equal to a query string q , in optimal time $\mathcal{O}(|q|)$ and independently of $|\mathbb{D}|$.

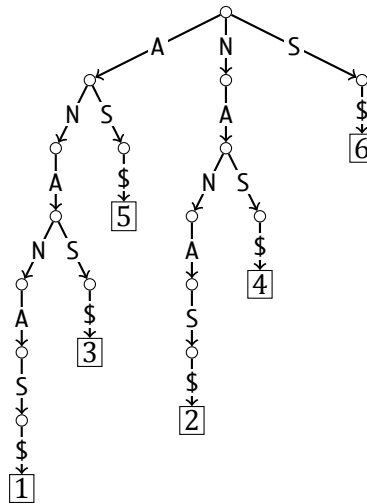
Algorithm 1.5 Exact dictionary search on a radix trie.

```

1: procedure EXACTSEARCH( $x, p$ )
2:   if  $p = \epsilon$  then
3:     report  $\mathbb{E}(x)$ 
4:   else if  $\exists c_x \in \mathbb{C}(x) : \text{label}(c_x) = p_1$  then
5:     EXACTSEARCH( $c_x, p_{2..|p|}$ )
6:   end if
7: end procedure

```

Figure 1.13: Exact dictionary search on a suffix trie.



Filtering methods

Filtering methods of section 1.3.4 can be directly applied to solve the dictionary search problem. Database strings satisfying the filtering condition can be verified with algorithm ??.

1.4.2 Local similarity search

Define score and scoring scheme.

Define local similarity.

Online methods

Give dynamic programming solution.

Indexed methods

Backtracking over substring index. BWT-SW.

Filtering methods

SWIFT/Stellar is based on the q -gram lemma.

1.4.3 Overlaps computation

Define problem.

Online methods

DP solution.

Indexed methods

Indexed solution, exact and approximate.

Next generation sequencing is a terrific technology. A wealth of applications have been developed on top of it. Data analysis pipelines for variant calling and structural variation discovery from DNA-seq, mRNA transcripts abundance estimation and novel non-coding RNA discovery from RNA-seq, transcription factor binding-sites prediction from ChIP-seq. All these applications rely on a common prerequisite step: mapping NGS reads to a known reference genome.

Read mapping is a critical step in all NGS data analysis pipelines. NGS reads produced by all current technologies contain sequencing errors, in form of single miscalled bases or stretches of oligonucleotides. Moreover, the donor genome from which reads have been sequenced contains small genomic variations (SNVs, Indels) in addition to CNV, inversions and translocations. After all, spotting genomic variation is one reason for which we resequence genomes. Thus, when mapping a read to a reference genome, it is not sufficient to consider the loci where the reads map exactly; it is necessary to consider any loci of relevant sequence similarity, being possible origins of the sequenced reads.

2.1 Sequencing technologies

2.1.1 Illumina / Solexa

2.1.2 Life Technologies / Ion Torrent

2.1.3 Roche / 454 Life Sciences

2.1.4 ABI / SOLiD

2.2 Sequencing quality

2.2.1 Phred base quality values

Phred base quality values have been introduced in [Ewing *et al.*, 1998; Ewing and Green, 1998] to assess the quality of sequencing single bases in capillary reads. Instead of directly discarding low-quality regions present in capillary reads, Phred calls each base and annotates it with a quality score encoding the probability that it has been wrongly

called. As this method has been widely accepted, base callers annotate reads issued by all sequencing technologies with Phred base quality scores.

To formally define Phred base quality values, let us fix the alphabet $\Sigma = \{A, C, G, T\}$, and consider a known donor genome g over Σ and a read r sequenced at location l from the template $g_{l \dots l+|r|-1}$. We define the base calling error ϵ_i at position i in the read r , as the probability ϵ_i of miscalling a base r_i instead of calling its corresponding base g_{l+i-1} in the donor genome. Therefore, we define the Phred base quality Q_i at position i as:

$$Q_i = -10 \log_{10} \epsilon_i. \quad (2.1)$$

Given the above, the probability $p(r_i | g_{l+i-1})$ of calling the base r_i in the read r , given the donor genome base g_{l+i-1} , is:

$$p(r_i | g_{l+i-1}) = \begin{cases} 1 - \epsilon_i & \text{if } g_{l+i-1} = r_i \\ \frac{\epsilon_i}{|\Sigma|-1} & \text{if } g_{l+i-1} \in \Sigma \setminus \{r_i\} \end{cases} \quad (2.2)$$

and assuming i.i.d. base calling errors, it follows that the probability $p(r | g, l)$ of observing the read r , given the donor genome template $g_{l \dots l+|r|-1}$, is:

$$p(r | g, l) = \prod_{i=1}^{|r|} p(r_i | g_{l+i-1}) \quad (2.3)$$

2.2.2 Mapping quality score

Mapping quality has been introduced in [Li *et al.*, 2008]. The study considers short reads of length ranging from 30 bp to 40 bp, produced by early Illumina/Solexa and ABI/SOLiD sequencing technologies, whose sequencing error rates were quite high. Given the short lengths and high error rates, a significant fraction of such reads can be aligned to multiple mapping locations, even considering only co-optimal Hamming distance locations.

The key point is that the Hamming distance is not an adequate scoring scheme to guess the correct mapping location of many reads. The authors claim¹ that:

It is possible to act conservatively by discarding reads that map ambiguously at some level, but this leaves no information in the repetitive regions and it also discards data, reducing coverage in an uneven fashion, which may complicate the calculation of coverage.

Since base callers output base call probabilities in Phred-scale along with the reads, Li *et al.* propose a novel probabilistic scoring scheme called mapping quality, giving the probability that a given read has been aligned correctly at a given mapping location in the reference genome.

By applying Bayes' theorem, we can derive the posterior probability $p(l | g, r)$, that location l in the reference genome g is the correct mapping location of read r . Assuming

¹ Li *et al.* do not show in their study what is the effect of relying on mapping quality rather than on mapping uniqueness.

uniform coverage, each location $l \in [1, |g| - |r| + 1]$ has equal probability of being the origin of a read in the donor genome, thus the prior probability $p(l)$ is simply:

$$p(l) = \frac{1}{|g| - |r| + 1} \quad (2.4)$$

Therefore, recalling $p(r|g, l)$ from equation 2.3, the posterior probability $p(l|g, r)$ equals the probability of the read r originating at location l normalized over all possible locations in the reference genome:

$$p(l|g, r) = \frac{p(r|g, l)}{\sum_{i=1}^{|g|-|r|+1} p(r|g, i)} \quad (2.5)$$

which in Phred-scale becomes:

$$Q(l|g, r) = -10 \log_{10}[1 - p(l|g, r)] \quad (2.6)$$

Computing the exact mapping quality as in equation requires aligning each read to all positions in the reference genome. On one hand, this computation would not be practical, indeed the vast majority of a reference genome is discarded when mapping reads by means of filtering and fully-indexed methods. On the other hand, the contribution of discarded locations to the sum in equation 2.5 can be neglected. Therefore, equation 2.5 is approximated using only relevant mapping locations found by the read mapper.

Mapping quality has been initially used in [Li *et al.*, 2008] and [Li *et al.*, 2009a] to maximize variant calling confidence by discarding reads whose best mapping location is below a given mapping quality threshold. This measure has been widely accepted: nowadays it is computed by most popular read mappers and used by almost all variant calling pipelines e.g. the Genome Analysis ToolKit (GATK) [DePristo *et al.*, 2011].

Nonetheless, some important objections can be moved against mapping quality. First, the mapping quality score is derived under the unlikely assumption of the reference genome being equal to the donor genome. In other words, mapping quality considers only errors due to base miscalls and disregards genetic variation; thus the risk is to prefer mapping locations supported by known low base qualities rather than by true but unknown SNVs. Second, mapping quality is nonetheless strongly correlated to mapping uniqueness, as discussed in section 2.3; it is easy to see that the mapping probability in equation 2.5 is diluted in presence of a large number of co-optimal mapping locations. Third, mapping quality tends to become less relevant as base calls improve, due to advances of sequencing technologies, and thus degenerates in a shallow measure of uniqueness.

2.3 Mappability

The difficulty of unambiguously finding the correct mapping location of next-generation sequencing reads comes from the non-random nature of genomes. Genomes evolved through multiple types of duplication events, including (i) whole-genome duplications

[?] or large-scale segmental duplications in chromosomes [?], (ii) transposition of repetitive elements as short tandem repeats (microsatellites) and interspersed nuclear elements (LINE, SINE) [?], (iii) proliferation of repetitive structural elements as telomeres and centromeres [?]. As a result of these events, about 50 % of the human genome is composed of repeats.

An analysis of the k -mer spectra of the genomes of some model organisms shows how genomes are statistically different from texts randomly generated according to uniform bernoulli models.

Repeats present in general technical challenges for all *de novo* assembly and sequence alignment programs [Lee and Schatz, 2012]. In the case of short reads mapping, the first evident effect of the heavy tail in the k -mer distribution of reference genomes is the dramatic loss of specificity in certain regions, which increases the computational cost of programs based on filtration methods. But the most subtle challenge lies in the interpretation of these results: it is not evident how to consider reads mapping to multiple locations.

Common strategies to deal with multi-reads are (i) to discard them all, (ii) to randomly pick one best mapping location, (iii) to consider all or up to k best mapping locations within a given distance threshold.

The next practical challenge is represented by reporting and handling the resulting datasets of mapping locations, which can have a size up to two orders of magnitude bigger compared to the corresponding input read sets.

Genome mappability can bias NGS analysis more than we might think at a first glance. Two recent studies [Derrien *et al.*, 2012; Lee and Schatz, 2012] show which is the bias of mappability.

2.3.1 Genome mappability

We now give a definition of genome mappability analogous to [Derrien *et al.*, 2012]. Let fix a q -gram length, a distance measure as the Hamming or edit distance, and a distance threshold k . Given a genomic sequence g , we defined the (q, k) -frequency $F_k^q(l)$ of the q -gram $g_{l...l+q-1}$ at location l in g as the number of occurrences of the q -gram in g and its reverse complement \bar{g} . We define the (q, k) -mappability $M_k^q(l)$ as the inverse (q, k) -frequency, i.e. $M_k^q(l) = F_k^q(l)^{-1}$ with $M_k^q : \mathbb{N} \rightarrow]0, 1]$. Note that $M_k^q(l)$ can be seen as the prior probability that any read of length q originating at location l will be mapped correctly. The values of (q, k) -frequency and mappability obviously vary with the distance threshold k . Nonetheless, under any distance measure, it hold that the q -gram at location l is unique up to the distance k iff $M_k^q(l) = 1$ and repeated otherwise.

Which is the minimum q -gram length from which we expect (q, k) -mappability to be 1 for the genomes of model organisms? Let consider the simple case of exact $(q, 0)$ -mappability. By assuming a genomic sequence of length n as being randomly generated under the uniform bernoulli model, the emission probability of any nucleotide is $p = \frac{1}{4}$ and, under i.i.d. assumptions, the emission probability of any q -gram is $p_q = \frac{1}{4^q}$. It follows that the expected value of $(q, 0)$ -frequency is $E[F_0^q] \simeq \frac{2n}{4^q}$. Thus, for $E[F_0^q] \leq 1$ it must

hold:

$$2n \leq 4^q \quad (2.7)$$

$$\log 2n \leq \log 4^q \quad (2.8)$$

$$q \geq \log_4 2n \quad (2.9)$$

Thus, we would expect any q -gram of length $\log_4 2n$ to occur about once in a genomic sequence of length n . Sticking to these assumption, in the human genome ($n \approx 3 \cdot 10^9$) almost all 17-mers would be unique, on fly ($n \approx 1.2 \cdot 10^8$) all 15-mers, on worm ($n \approx 4.2 \cdot 10^7$) all 13-mers. However, the q -gram distribution of model genomes does not fit the uniform bernoulli distribution. In [?] the k -mers distribution can be approximated by a double Pareto log-normal distribution, i.e. a distribution with a heavy tail. This is a result of the evolution of genomes being driven by gene duplications, retrotransposons [?].

Consequently, we would expect 36 bp reads produced by early Illumina sequencers to induce an almost perfect mappability. However, reads have to be mapped approximately to the reference genome. The expected number of approximate occurrences of a k -mer is higher than the exact one. Thus the above estimate is a lower bound.

Uniqueome

Derrien *et al.* quantified the whole genome unique mappability for human, mouse, fly, and worm. At a (36, 2) mapping, about 30 % of the human genome is not uniquely mappable. Unique mappability rises to 83 % by increasing the read length to 75 bp; however to map a significant fraction of the reads, we should consider 3–4 edit distance errors.

| | H.sapiens (hg19) | M.musculus (mm9) | D.mel (dm3) |
|------------------------------|---------------------|---------------------|----------------|
| Repeats content [%] | 45.25 | 42.33 | 26.50 |
| Uniqueome (36 bp, 2 msm) [%] | 69.99 | 72.07 | 68.09 |
| Uniqueome (50 bp, 2 msm) [%] | 76.59 | 77.06 | 69.44 |
| Uniqueome (75 bp, 2 msm) [%] | 83.09 | 81.65 | 71.00 |

The uniqueome plays an important role in ChIP-seq experiments. It is common practice [?] to rely on short (36 bp) reads and discard the non-unique ones. Not only a significant fraction of the sequencing data is thrown out. Worse than that, we end up with holes in 30 % of the genome. A ChIP-seq peak caller considering multi-reads calls up to 30 % more peaks.

Cite regions of clinical relevance, e.g. HLA-A. Cite regions of biological relevance, e.g. 5S rRNA.

Paired-end mappability

Pileup mappability

If we focus our attention to the resequencing accuracy at a single locus, we have to consider the mappability of all the possible reads spanning that given locus. Pileup mappability [Derrien *et al.*, 2012] at position i is the average mappability of all reads spanning position i .

$$M_p(i) = 1/q \sum_{j=i}^{i+1} M(j)$$

2.3.2 Genome mappability score

Genome mappability score (GMS) [Lee and Schatz, 2012] is analogous to pileup mappability. Instead of considering the inverse mapping frequency (q, k) -mappability, we can interpret mapping quality (see subsection 2.2.2) as the probability that a read originating at a given position can be mapped correctly. Therefore, we consider the average mapping probability of any read spanning a location l of a reference genome g^2 :

$$p(l|g) = \sum_{r \in \mathcal{R}(l)} \frac{p(l|g, r)}{|\mathcal{R}(l)|} \quad (2.10)$$

which in Phread-scale becomes:

$$Q(l|g) = \sum_{r \in \mathcal{R}(l)} \frac{1 - 10^{-\frac{Q(l|g, r)}{10}}}{|\mathcal{R}(l)|} \quad (2.11)$$

and thus, fixed a genomic sequence g , we define the genome mappability score $GMS(l)$ as its percentual value:

$$GMS(l) = 100Q(l|g) \quad (2.12)$$

Lee and Schatz simulate reads having length and error profiles similar to those issued by actual sequencing technologies, define low GMS regions as those locations for which $GMS(l) \leq 10$, and measure the percentage of such locations in the human genome.

2.4 Categorization of existing read mappers

Critic the surveys classifying hundreds of mappers [Li and Homer, 2010], [Fonseca *et al.*, 2012].

Critic the benchmarks [Hatem *et al.*, 2013] [Holtgrewe *et al.*, 2011].

² Equation 3 in [Lee and Schatz, 2012] is not precise, please refer to our equation 2.10.

| Sequencing technology | Read length [bp] | Error rate [%] (msm, ins, del) | Low GMS [%] | High GMS [%] |
|-----------------------|------------------|--------------------------------|-------------|--------------|
| SOLiD-like | 75 | (0.10, 0.00, 0.00) | 11.14 | 88.86 |
| Illumina-like | 100 | (0.10, 0.00, 0.00) | 10.51 | 89.49 |
| Ion Torrent-like | 200 | (0.04, 0.01, 0.95) | 9.35 | 90.65 |
| Roche/454-like | 800 | (0.18, 0.54, 0.36) | 8.91 | 91.09 |
| PacBio-like | 2000 | (1.40, 11.47, 3.43) | 100.0 | 0.00 |
| PacBio EC-like | 2000 | (0.33, 0.33, 0.33) | 8.61 | 91.39 |

2.4.1 Best mappers versus all mappers

The task of a read mapper is to guess where a read originates. Fixed a similarity scoring scheme that confidently models this problem, the optimal alignment under this scoring scheme correspond to the most likely explanation and induces a locus being the origin of the read. The simplest scoring scheme is the edit distance; more involved scoring schemes take into account base quality values, score gaps using affine cost functions, or allow to trim for free a prefix or a suffix of the read.

The above definition does not consider two problems: what if there are many co-optimal candidates, and what if the correct solution corresponds to a sub-optimal candidate. The former problem is exacerbated by genome mappability. One would expect such situations to arise very rarely, but instead it is a relevant problem. The latter problem arises whenever our model is not adequate to explain the difference between a read and its genomic origin. For instance, an evolutionary event producing an indel of length l might be considered as a unit, whether edit distance would consider it as l independent events. Under the edit distance, an alignment with less than l independent point mutations would be considered more likely than an alignment containing only one indel of length l .

From the former problem, we conclude that considering only one optimal mapping location is not sufficient, no matter how good our scoring scheme can be. The latter problem tells us to be careful about relying on strict optimality. Therefore, in general a read mapper should return a comprehensive set of relevant mapping locations along with the likelihood that they correspond to the original location.

2.5 Read mappers

2.5.1 Bowtie

Bowtie [Langmead *et al.*, 2009] is a mapper designed to have a small memory footprint and quickly report a few good mapping locations for early generation Illumina/Solexa and ABI/SOLiD short reads of length up to 50 bp. It achieves the former goal by indexing the reference genome with an FM-index and the latter goal by performing a greedy depth-first traversal on it.

The greedy depth-first traversal visits first the subtree yielding the least number of

mismatches and stops after having found a candidate (not guaranteed to be optimal when $k > 1$). In addition, Bowtie speeds up backtracking by applying case pruning, a simple application of the pigeonhole principle. However this technique is mostly suited for $k = 1$ and requires the index of the forward and reverse text.

Bowtie can be configured to search by strata, however the search time increases significantly while the traversal still misses a large fraction of the search space due to seeding heuristics. Main practical drawbacks of the tool are too many cryptic options.

Bowtie 2 [Langmead and Salzberg, 2012] has been designed to quickly report a couple of mapping locations for recent Illumina/Solexa, Ion Torrent and Roche/454 reads, usually having lengths in the range from 100 bp to 400 bp.

This tool uses an heuristic seed-and-extend approach, collecting seeds of fixed length, partially overlapping, and searching them exactly in the reference genome using an FM-index. Candidate locations to verify are chosen randomly, to avoid uncompressing large CSA intervals and executing many DP instances. Each mapping location is verified using a striped vectorial dynamic programming algorithm, implemented using SIMD instructions, previously introduced by [Farrar, 2007] and extended to compute end-to-end alignments.

Bowtie 2 can be configured to report end-to-end or local alignments, scored using a tunable affine scoring scheme. For this reason, it is believed to be good at reporting alignments containing indels. However, its completely heuristic filtration strategy, independent of the scoring scheme, makes it hard to believe what it promises.

2.5.2 BWA

BWA-backtrack [Li and Durbin, 2009] is designed to map Illumina/Solexa reads up to 100 bp and report a few best end-to-end alignments. The program performs a greedy breadth-first search on an FM-index of the reference genome. Nodes to be visited are ranked by edit distance score: the best node is popped from a priority queue and visited, its children are then inserted again in the queue. The traversal considers indels using a more involved 9-fold recursion. Backtracking is sped up by adopting a more stringent pruning strategy that nonetheless takes some preprocessing time and requires the index of the reverse reference genome.

BWA performs paired-end alignments by trying to anchor both paired-end reads and verifying the corresponding mate, within an estimated insert size, using the classic DP-based Smith-Waterman algorithm. Consequently, the program in paired-end mode aligns reads at a slower rate than in single-end mode. The program is not fully multi-threaded, therefore BWA scales poorly on modern multi-core machines.

BWA-SW [Li and Durbin, 2010] is designed to map Roche/454 reads, which have an average length of 400 bp. It is an heuristic version of BWT-SW, designed to report a few good local alignments.

This version of BWA adopts a double indexing strategy: it indexes all substrings of one read in a DAWG. It performs Smith-Waterman of all read substrings directly on the FM-index, by backtracking as soon as no viable alignment can be obtained. As in BWA-backtrack, the traversal proceeds in a greedy fashion. In addition, BWA-SW implements

some seeding heuristics to limit backtracking and jump in the reference genome to verify candidate locations whenever this becomes favorable.

This version of BWA does not support paired-end reads, presumably because it was meant for Roche/454 reads.

2.5.3 Soap

Soap 2 [Li *et al.*, 2009b] is very similar to Bowtie: it has been designed to produce a very quick but shallow mapping of Illumina/Solexa reads up to 75 bp with no more than 2 mismatches and no indels. However, its underlying algorithm is based on the so-called bi-directional (or 2-way) BWT. The tool support paired-end mapping but at a slower alignment rate. Practical drawbacks are the lack of native output in the de-facto standard SAM format and is closed source. Soap 3 [Liu *et al.*, 2012] is algorithmically similar to Soap 2 but targets only NVIDIA CUDA accelerators.

2.5.4 SHRiMP

SHRiMP 2 TODO.

2.5.5 RazerS

RazerS [Weese *et al.*, 2009] has been designed to report all mapping locations within a fixed hamming or edit distance error rate. It is based on a full-sensitive q -gram filtration method (SWIFT semi-global) combined with the Myers edit distance verification algorithm. On demand, the SWIFT filter can be configured to become lossy within a fixed loss rate. The lossy filter becomes more stringent and produces a lower number of candidates to verify, thus improving the overall speed of the program. All in all, the SWIFT filter is very slow while not highly specific.

RazerS 3 [Weese *et al.*, 2012] is a faster version featuring shared-memory parallelism, a faster banded-Myers verification algorithm, and a faster filtration scheme based on exact seeds that however turns out to be very weak on mammal genomes. Because of this, RazerS 3 is one-two orders of magnitude slower than Bowtie 2 and BWA-backtrack on mammal genomes.

All RazerS versions index the reads and scan the reference genome. One positive aspect of this strategy is that no preprocessing of the reference genome is required. However, other mapping strategies beyond all-mapping, e.g. mapping by strata, cannot be efficiently implemented. Moreover, the program exhibit an high memory footprint as it must remember the mapping locations of all input reads until the whole reference genome has been scanned.

2.5.6 mr(s)Fast

The tools mrFast [Ahmadi *et al.*, 2012] and mrsFast [Hach *et al.*, 2010] are designed to report all mapping locations within a fixed absolute number errors, respectively under

the hamming and edit distance, given Illumina/Solexa reads of length ranging from 50 bp to 125 bp. Similarly to RazerS 3, they are based on a full-sensitive filtration strategy using exact seeds, which turns out to be very weak on mammal genomes.

The peculiarity of their underlying method is a cache-oblivious strategy to mitigate the high cost of verifying clusters of candidate locations. In addition, mrsFast computes the edit distance between one read and one mapping location in the reference genome with an antidiagonal-wise vectorial dynamic programming algorithm, implemented using SIMD instructions.

These tools are as slow as RazerS 3 and appealing for nothing more than all-mapping. They lack multi-threading support and exhibit various bugs. Furthermore, they only accept reads of fixed length and produce files of impractical size.

2.5.7 GEM

The GEM mapper [Marco-Sola *et al.*, 2012] is a flexible read aligner for Illumina/Solexa, ABI/SOLiD, and Ion Torrent reads. It is full-sensitive and can be configured either as an all-mapper, as a best/unique-mapper, or to search by strata.

GEM uses a combination of state of the art approximate string matching methods, e.g. approximate seeds and suffix filters. The program indexes the reference genome with an FM-index, tries to find an optimal filtration strategy per read, and verifies candidate locations using Myers algorithm. Paired-reads are either mapped independently and then combined, or left/right are mapped and their mates verified using an online strategy.

Unfortunately the tool lacks direct SAM output, it is not open source, and provides many obscure parameters.

| | platform | | | | strategy | | | method | | | index | | |
|----------|------------|-----|-----|-------|----------|--------|-----|------------|---------|--------------|-----------------|-----------|-------|
| | Illumina | Ion | 454 | SOLID | best | strata | all | alignment | optimal | algorithm | type | reference | reads |
| Bowtie | ≤ 50 | X | X | ✓ | ✓ | ✓ | • | mismatches | X | backtracking | FM-index | ✓ | X |
| Bowtie 2 | ≥ 75 | ✓ | ✓ | X | ✓ | • | • | local | X | exact seeds | FM-index | ✓ | X |
| BWA | ≤ 100 | X | X | X | ✓ | X | • | indels | X | backtracking | FM-index | ✓ | X |
| BWA-SW | X | ✓ | ✓ | X | ✓ | X | X | local | X | backtracking | FM-index | ✓ | ✓ |
| Soap 2 | ≤ 75 | X | X | X | ✓ | ✓ | • | mismatches | X | backtracking | FM-index | ✓ | X |
| RazerS | ≥ 50 | X | X | X | • | • | ✓ | indels | ✓ | q -grams | q -gram index | X | ✓ |
| RazerS 3 | ≥ 50 | ✓ | X | X | • | • | ✓ | indels | ✓ | exact seeds | q -gram index | X | ✓ |
| SHRiMP 2 | ✓ | ✓ | ✓ | ✓ | ✓ | • | • | local | X | q -grams | q -gram index | ✓ | X |
| mrsFast | ≤ 75 | X | X | X | • | X | ✓ | mismatches | ✓ | exact seeds | q -gram index | ✓ | ✓ |
| mrFast | ≤ 125 | X | X | X | • | X | ✓ | indels | ✓ | exact seeds | q -gram index | ✓ | ✓ |
| GEM | ✓ | ✓ | X | ✓ | ✓ | ✓ | ✓ | indels | ✓ | apx seeds | FM-index | ✓ | X |
| Masai | ✓ | X | X | X | ✓ | • | ✓ | indels | ✓ | apx seeds | generic | ✓ | ✓ |
| Yara | ✓ | ✓ | X | X | ✓ | ✓ | ✓ | indels | ✓ | apx seeds | generic | ✓ | X |

2.6 Masai

2.6.1 Single-end mapping

All-mapping

Mapping by strata

2.6.2 Paired-end mapping

2.7 Yara

2.7.1 Single-end mapping

All-mapping

Mapping by strata

2.7.2 Paired-end mapping

All-mapping

Mapping by strata

2.7.3 Parallelization

2.7.4 Hardware acceleration

2.8 Assessment of read mappers performance

2.8.1 Comparison of filtration strategies

2.8.2 Rabema benchmark results

2.8.3 Variant detection results

2.8.4 Runtime results

2.9 Discussion

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