How to predict new compounds

(1) About the models of the prediction system:

The seven models of the multi-layer sweetness prediction system are shown in **Table 1**.

Table 1. Seven models of the multi-layer sweetness prediction system

Category	Natural	Artificial	Carbohydrate	Non-	Nutritive	Non-	LogSw
				carbohydrate		nutritive	
Model	MOE2d-	MACCS-	Atompairs-	MOE2d-	MOE2d-	MOE2d-	Atompairs-
	XGBoost	RF	XGBoost	XGBoost	XGBoost	XGBoost	SVR

(2) Descriptor and fingerprints:

The models were built on KNIME (version 4.1.0). Atompairs (1024 bits), ECFP4 (1024 bits) and MACCS (167 bits) fingerprints were calculated by the RDKit node of KNIME. The MOE2d descriptors were calculated by MOE (version 2018). The details of the selected MOE2d descriptors related to the above seven models are as follows:

• Natural (105):

apol, ast_fraglike, ast_fraglike_ext, ast_violation, a_acid, a_aro, a_base, a_donacc, a_ICM, a_nCl, a_nF, a_nN, a_nP, a_nS, balabanJ, BCUT_SLOGP_0, BCUT_SLOGP_2, BCUT_SLOGP_3, BCUT_SMR_0, BCUT_SMR_1, b_1rotN, b_1rotR, b_double, b_max1len, b_triple, chiral, chiral_u, density, diameter, FCharge, GCUT_PEOE_0, GCUT_PEOE_1, GCUT_PEOE_2, GCUT_PEOE_3, GCUT_SLOGP_0, GCUT_SLOGP_1, GCUT_SLOGP_2, GCUT_SMR_0, GCUT_SMR_1, GCUT_SMR_2, h_emd_C, h_logP, h_logS, h_log_dbo, h_log_pbo, h_pavgQ, h_pKa, h_pKb, h_pstates, h_pstrain, Kier3, KierA2, lip_violation, logP(o/w), mutagenic, opr_brigid, opr_leadlike, PEOE_RPC+, PEOE_RPC-, PEOE_VSA+0, PEOE_VSA+1, PEOE_VSA+2, PEOE_VSA+3, PEOE_VSA+5, PEOE_VSA+6, PEOE_VSA-0, PEOE_VSA-1, PEOE_VSA-2, PEOE_VSA-3, PEOE_VSA-4, PEOE_VSA-5, PEOE_VSA_FHYD, PEOE_VSA_FNEG, PEOE_VSA_FPNEG, PEOE_VSA_NEG, PEOE_VSA_PNEG, PEOE_VSA-1, SlogP_VSA3, SlogP_VSA4, SlogP_VSA5, SlogP_VSA6, SlogP_VSA6, SlogP_VSA8, SlogP_VSA9, SMR_VSA0, SMR_VSA1, SMR_VSA2, SMR_VSA3, SMR_VSA4, SMR_VSA5, SMR_VSA6, SMR_VSA6, SMR_VSA7, VAdjEq, VAdjMa, VDistEq, vsa_acc, vsa_base, vsa_don, vsa_hyd, vsa_other, weinerPath

• Non-carbohydrate (124):

apol, ast_fraglike, ast_fraglike_ext, ast_violation, a_acc, a_acid, a_aro, a_base, a_don, a_ICM, a_nB, a_nBr, a_nCl, a_nF, a_nI, a_nN, a_nP, a_nS, balabanJ, BCUT_SLOGP_0, BCUT_SLOGP_1, BCUT_SLOGP_2, BCUT_SLOGP_3, BCUT_SMR_0, BCUT_SMR_1, BCUT_SMR_2, BCUT_SMR_3, b_1rotN, b_1rotR, b_double, b_max1len, b_triple, chi1_C, chiral, chiral_u, density, diameter, FCharge, GCUT_SLOGP_0, GCUT_SLOGP_1, GCUT_SLOGP_2, GCUT_SLOGP_3, GCUT_SMR_0, GCUT_SMR_1, GCUT_SMR_2, h_ema, h_emd, h_emd_C, h_logD, h_logP, h_logS, h_log_dbo, h_log_pbo, h_pavgQ, h_pKa, h_pKb, h_pstates, h_pstrain, Kier2, Kier3, KierFlex, lip_druglike, lip_violation, logP(o/w), logS, mutagenic, opr_brigid, opr_leadlike, opr_nrot, opr_violation, PEOE_RPC-, PEOE_VSA+0, PEOE_VSA+1, PEOE_VSA+2, PEOE_VSA+3, PEOE_VSA+4, PEOE_VSA+5, PEOE_VSA+6, PEOE_VSA-0, PEOE_VSA-1, PEOE_VSA-2, PEOE_VSA-3, PEOE_VSA-4, PEOE_VSA-5, PEOE_VSA-6, PEOE_VSA_FHYD, PEOE_VSA_FNEG, PEOE_VSA_FPNEG, PEOE_VSA_FPPOS, PEOE_VSA_NEG, PEOE_VSA_PNEG, PEOE_VSA_PNEG, PEOE_VSA_SlogP_VSA0,

SlogP_VSA1, SlogP_VSA2, SlogP_VSA3, SlogP_VSA4, SlogP_VSA5, SlogP_VSA6, SlogP_VSA7, SlogP_VSA8, SlogP_VSA9, SMR_VSA0, SMR_VSA1, SMR_VSA2, SMR_VSA3, SMR_VSA4, SMR_VSA5, SMR_VSA6, SMR_VSA7, TPSA, VAdjEq, VAdjMa, VDistEq, vsa_acc, vsa_base, vsa_don, vsa_hyd, vsa_other, weinerPath

• **Nutritive (102):**

ast_fraglike, ast_fraglike_ext, ast_violation, ast_violation_ext, a_acid, a_aro, a_count, a_don, a_ICM, a_nF, a_nI, a_nN, a_nP, a_nS, balabanJ, BCUT_PEOE_0, BCUT_PEOE_1, BCUT_PEOE_3, BCUT_SLOGP_0, BCUT_SLOGP_2, BCUT_SLOGP_3, BCUT_SMR_2, b_1rotN, b_1rotR, b_double, b_max1len, b_triple, chi0v_C, chi1_C, chiral, chiral_u, density, diameter, GCUT_PEOE_0, GCUT_PEOE_1, GCUT_PEOE_2, GCUT_SLOGP_0, GCUT_SLOGP_1, GCUT_SLOGP_2, GCUT_SMR_0, GCUT_SMR_1, GCUT_SMR_3, h_emd_C, h_logP, h_logS, h_log_dbo, h_log_pbo, h_pavgQ, h_pKa, h_pKb, h_pstates, h_pstrain, Kier3, KierA2, lip_druglike, lip_violation, mutagenic, opr_brigid, opr_leadlike, PEOE_RPC+, PEOE_RPC-, PEOE_VSA+0, PEOE_VSA+1, PEOE_VSA+2, PEOE_VSA+3, PEOE_VSA+5, PEOE_VSA+6, PEOE_VSA-0, PEOE_VSA-1, PEOE_VSA-2, PEOE_VSA-4, PEOE_VSA-5, PEOE_VSA_FHYD, PEOE_VSA_FNEG, PEOE_VSA_FPNEG, PEOE_VSA_SNEG, PEOE_VSA_RPOS, petitjean, reactive, rsynth, SlogP_VSA1, SlogP_VSA2, SlogP_VSA3, SlogP_VSA4, SlogP_VSA5, SlogP_VSA5, SlogP_VSA8, SlogP_VSA9, SMR_VSA0, SMR_VSA2, SMR_VSA4, SMR_VSA5, SMR_VSA5, SMR_VSA7, TPSA, vsa_acc, vsa_don, vsa_hyd, vsa_other, weinerPath, weinerPol

• Non-nutritive (122):

apol, ast_fraglike, ast_fraglike_ext, ast_violation, a_acid, a_aro, a_base, a_don, a_ICM, a_nB, a_nBr, a_nCl, a_nF, a nI, a nN, a nP, a nS, balabanJ, BCUT SLOGP 0, BCUT SLOGP 1, BCUT SLOGP 2, BCUT SLOGP 3, BCUT_SMR_0, BCUT_SMR_1, BCUT_SMR_2, BCUT_SMR_3, b_1rotN, b_1rotR, b_double, b_max1len, b triple, chi1 C, chiral, chiral u, density, diameter, FCharge, GCUT SLOGP 0, GCUT SLOGP 1, GCUT SLOGP 2, GCUT SLOGP 3, GCUT SMR 0, GCUT SMR 1, GCUT SMR 2, h emd, h emd C, h logD, h logP, h logS, h log dbo, h log pbo, h pavgQ, h pKa, h pKb, h pstates, h pstrain, Kier2, Kier3, KierFlex, lip druglike, lip violation, logP(o/w), logS, mutagenic, opr brigid, opr leadlike, opr nrot, opr violation, PEOE_PC-, PEOE_RPC-, PEOE_VSA+0, PEOE_VSA+1, PEOE_VSA+2, PEOE_VSA+3, PEOE_VSA+4, PEOE VSA+5, PEOE VSA+6, PEOE VSA-0, PEOE VSA-1, PEOE VSA-2, PEOE VSA-3, PEOE VSA-4, PEOE VSA-5, PEOE VSA-6, PEOE VSA FHYD, PEOE VSA FNEG, PEOE VSA FPNEG, PEOE VSA FPPOS, PEOE VSA NEG, PEOE VSA POL, petitjean, reactive, rsynth, SlogP, SlogP VSA0, SlogP_VSA1, SlogP_VSA2, SlogP_VSA3, SlogP_VSA4, SlogP_VSA5, SlogP_VSA6, SlogP_VSA7, SlogP_VSA8, SlogP_VSA9, SMR_VSA0, SMR_VSA1, SMR_VSA2, SMR_VSA3, SMR_VSA4, SMR_VSA5, SMR VSA6, SMR VSA7, VAdjEq, VAdjMa, VDistEq, vsa acc, vsa base, vsa don, vsa hyd, vsa other, vsa pol, weinerPath

(3) How to form your own prediction pipeline:

The details of constructing your own workflow are shown in **Figure 1** and **Figure 2**. Explanation of workflow:

- 1) The local model file is read by the **Model Reader** node above, while the below reads the model-Normalizer.zip file for normalizer;
 - 2) The node of File Reader is used to read the data that needs to be predicted;
 - 3) Convert numbers of label to strings using Number to String node is required in

classification models;

- 4) Select the corresponding prediction node according to the model read by the model reader;
- 5) Output for the prediction result. In addition, evaluation nodes can be chosen according to your task.

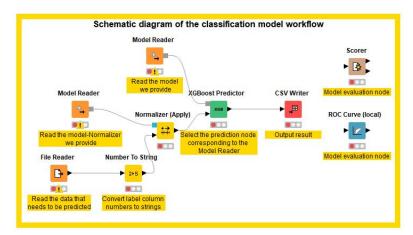


Figure 1. KNIME usage example of classification model.

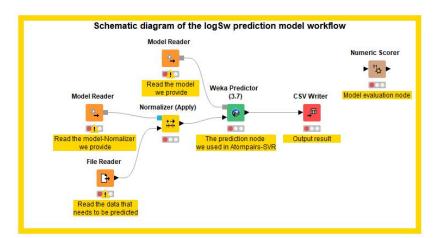


Figure 2. KNIME usage example of logSw prediction model.