

# QTL mapping in MAGIC populations with R/qtl2

Karl Broman

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[kbroman.org](http://kbroman.org)

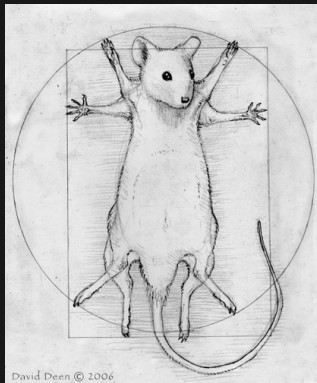
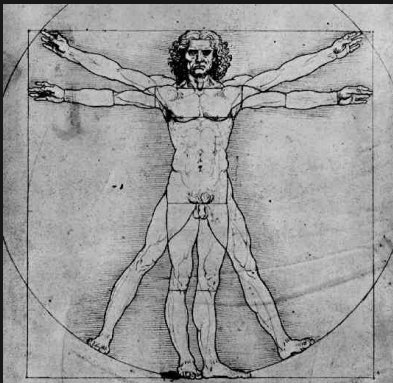
[github.com/kbroman](https://github.com/kbroman)

@kwbroman

Slides: [bit.ly/msu2019-12](https://bit.ly/msu2019-12)

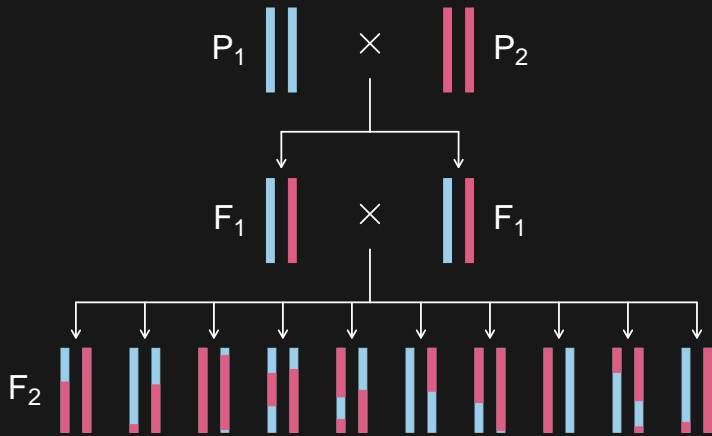




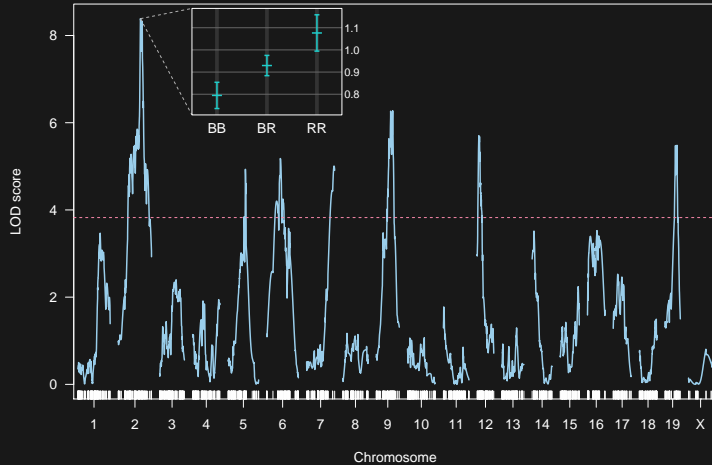


daviddeen.com

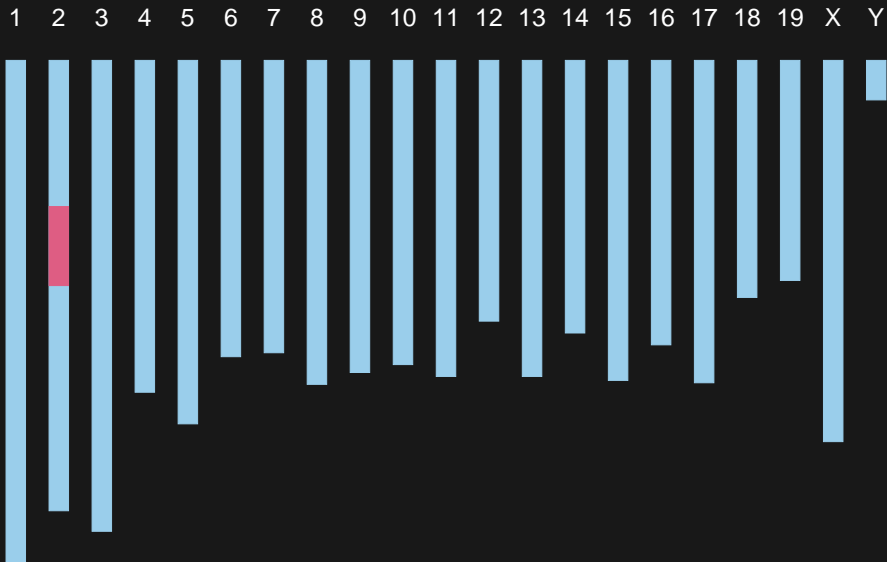
# Intercross



# QTL mapping



# Congenic line/NIL



# Improving precision

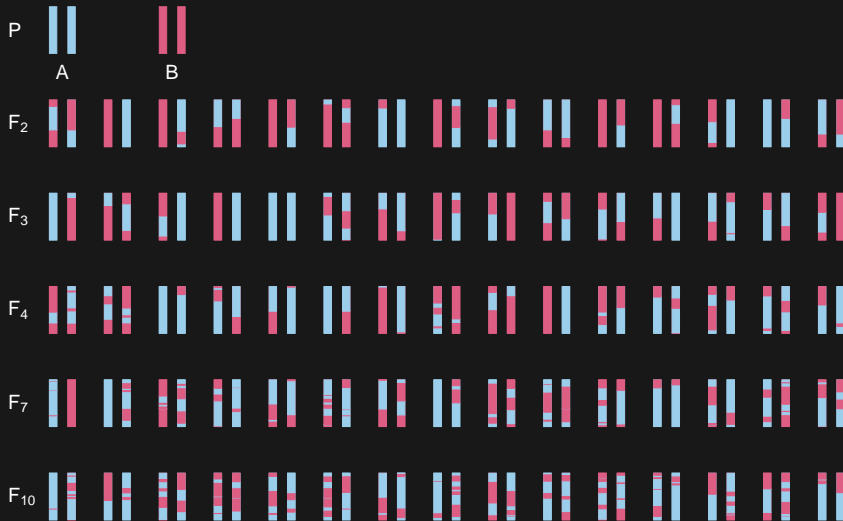
- ▶ more recombinations
- ▶ more individuals
- ▶ more precise phenotype
- ▶ lower-level phenotypes
  - transcripts, proteins, metabolites

# Genome-scale phenotypes

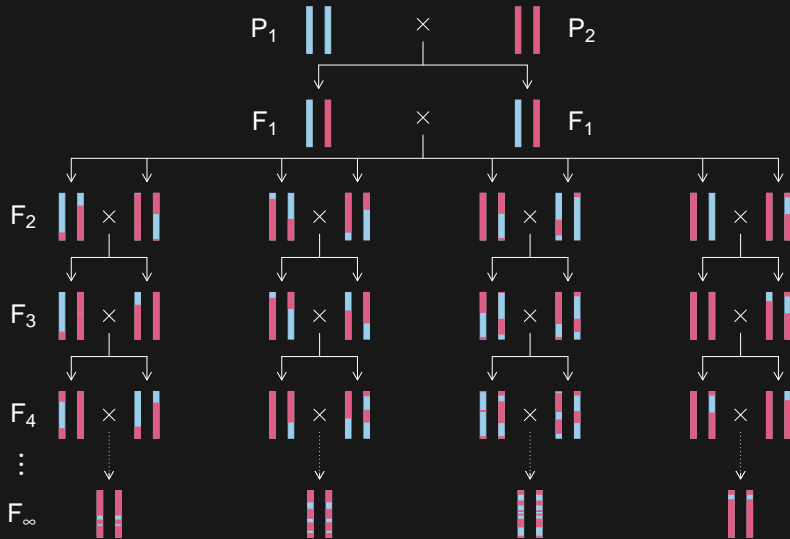




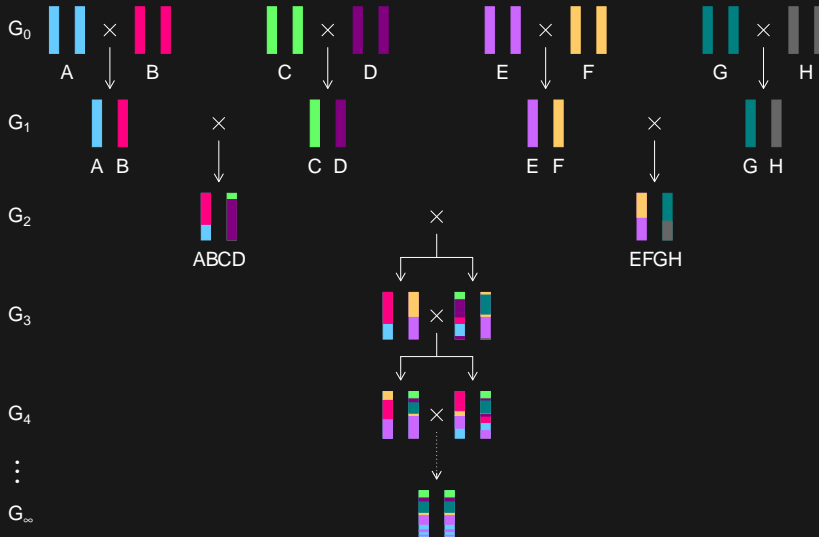
# Advanced intercross lines



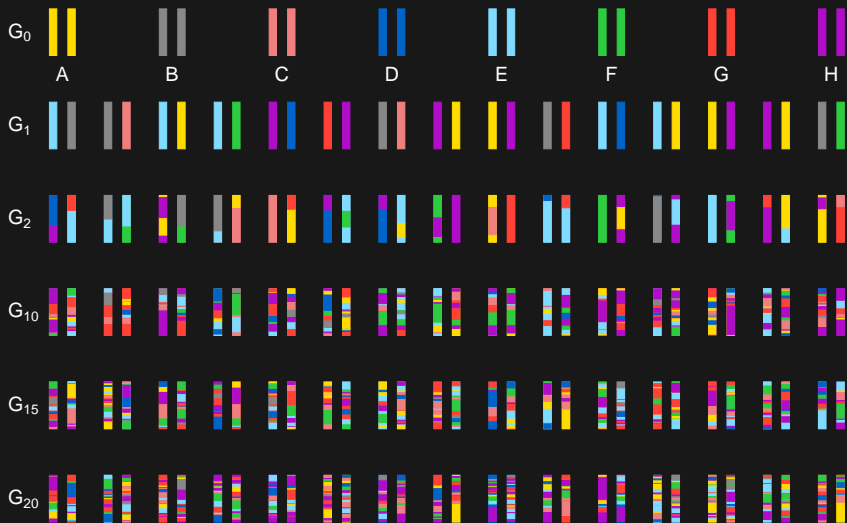
# Recombinant inbred lines



# Collaborative Cross



# Heterogeneous stock



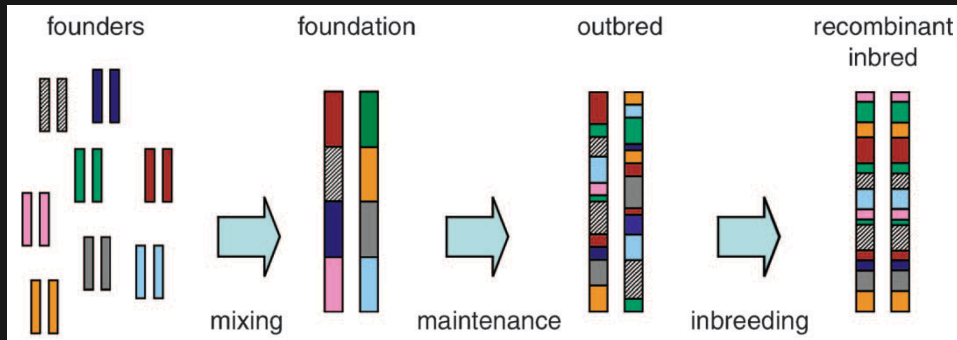
# MAGIC is magic

- ▶ Genetic diversity
- ▶ High-precision mapping
- ▶ Predictable linkage disequilibrium
- ▶ Phenotype replicates to reduce individual variation
- ▶ Pool phenotypes from multiple labs, environments, treatments
- ▶ Genotype once

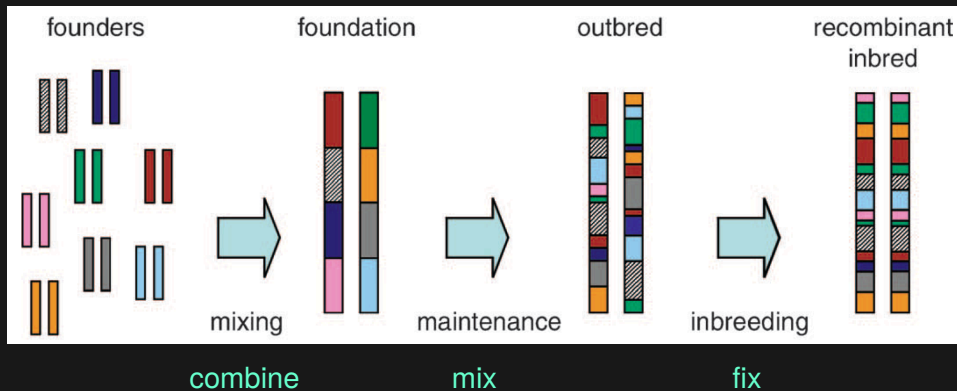
# MAGIC is magic

- ▶ Genetic diversity
- ▶ High-precision mapping
- ▶ Predictable linkage disequilibrium
- ▶ Phenotype replicates to reduce individual variation
- ▶ Pool phenotypes from multiple labs, environments, treatments
- ▶ Genotype once
- ▶ Cool name

# MAGIC lines

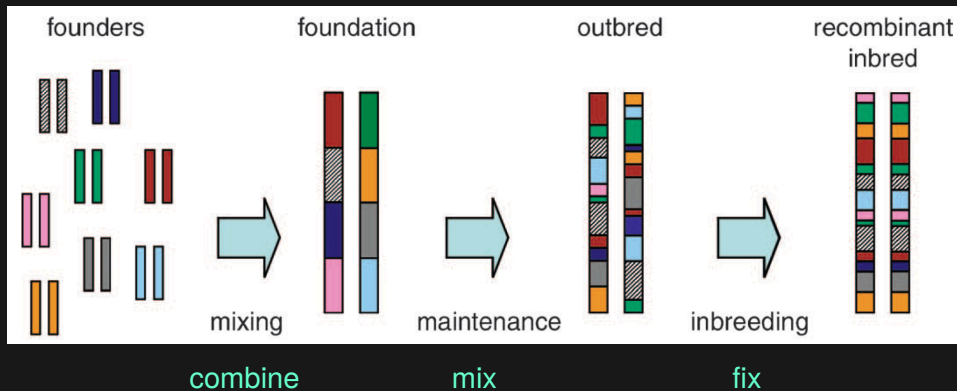


# MAGIC lines



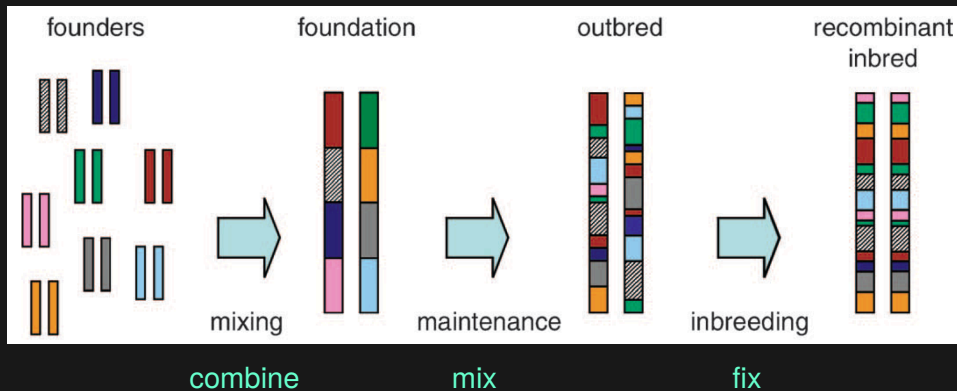


# MAGIC lines



How many?

# MAGIC lines

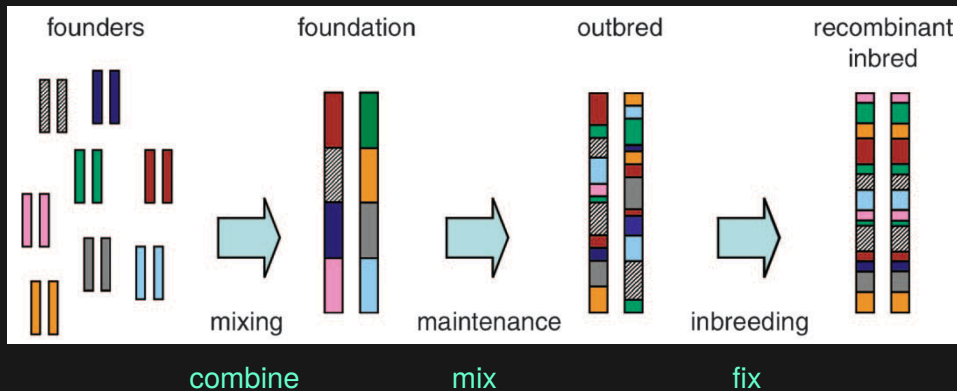


How many?

Which?

Valdar et al., Genetics 172:1783, 2006

# MAGIC lines



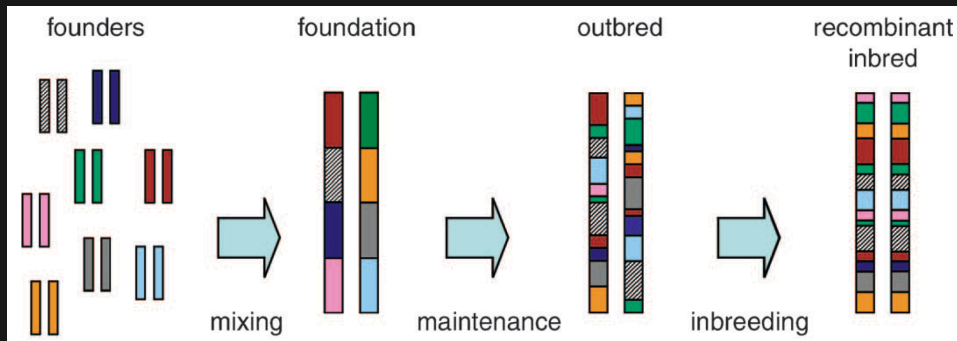
How many?

How long?

Which?

Valdar et al., Genetics 172:1783, 2006

# MAGIC lines



combine

mix

fix

How many?

How long?

How?

Which?

Valdar et al., Genetics 172:1783, 2006

# The goal

Identify QTL

- ▶ Power
- ▶ Mapping precision

# The goal

Identify QTG

- ▶ Power
- ▶ Mapping precision

# The goal

Identify QT**G**

- ▶ Power
- ▶ Mapping precision
- ▶ Estimate QTL allele frequencies

# Principles

- ▶ Avoid population structure
- ▶ Tradeoff between *power for de novo discovery* and *mapping precision*
- ▶ More QTL to find  $\Rightarrow$  more QTL getting in the way?
- ▶ More QTL alleles  $\Rightarrow$  less information about each
- ▶ Are QTL alleles common or rare?



# How many founders?

## More

- ▶ More general use
- ▶ More QTL
- ▶ Greater precision
- ▶ Estimate allele frequencies
- ▶ Haplotype analysis in founders

## Fewer

- ▶ Lower residual variance
- ▶ Greater power for a particular QTL?
- ▶ Better power for epistasis
- ▶ Rare alleles are less rare

# Which founders?

- ▶ Diverse
- ▶ Interesting
- ▶ No breeding problems
- ▶ Balanced: star phylogeny

# How much mixing?

- ▶ More mixing  $\Rightarrow$  Greater mapping precision
- ▶ ...but lower power for *de novo* mapping
- ▶ Potential for population structure, missing alleles
- ▶ Random mating or curated mating?
- ▶ Start with many random cross directions?

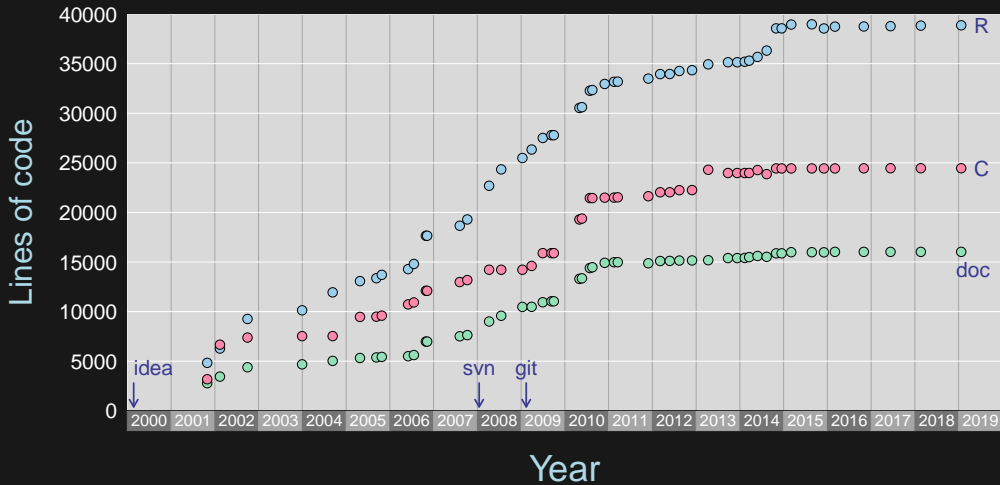
# Selfing or DH?

- ▶ Inbreeding gives added recombination
- ▶ But not so much as at the mixing stage
- ▶ If doubled haploids are feasible, use them

# Sharing is also key

- ▶ The greatest power of MAGIC comes from sharing  
Pooling data, exploring multiple environments/treatments
- ▶ Common software needs  
Analysis software, database infrastructure
- ▶ Many students need to learn the same stuff  
Joint training opportunities

# 19 years of R/qtl



# R/qtl cross types

- ▶ backcross, doubled haploids, haploid
- ▶ intercross
- ▶ 2-way RIL by selfing or sibling mating
- ▶ phase-known 4-way cross





## R/qtl2 cross types

- ▶ backcross, doubled haploids, haploid
- ▶ intercross
- ▶ 2-, 4-, 8-, 16-way RIL by selfing
- ▶ 2-, 4-, 8-way RIL by sibling mating
- ▶ 2-, 3-, 8-way advanced intercross
- ▶ 6- and 19-way MAGIC
- ▶ Diversity Outbred (DO) mice
- ▶  $F_1$  of DO  $\times$  inbred
- ▶ general RIL or AIL

# Data files

	A	B	C	D	E	F	G
1	id	bolting_days	seed_weight	seed_area	ttl_seedspruit	branches	height
2	MAGIC.1	15.33	17.15	0.64	45.11	10.5	NA
3	MAGIC.2	22	22.71	0.75	49.11	4.33	42.33
4	MAGIC.3	23	21.03	0.68	57	4.67	50
5	MAGIC.4	18.67	22.45	0.74	54.33	6.33	NA
6	MAGIC.5	18.67	25.36	0.82	38.33	5.67	42.25
7	MAGIC.6	25	21.53	0.71	52	4.33	NA
8	MAGIC.7	15.33	20.92	0.71	39	4	37.35
9	MAGIC.8	14.33	24.2	0.79	50.56	7.33	43.23
10	MAGIC.9	16.33	18.86	0.63	75.78	5.33	42.4
11	MAGIC.10	30	30.46	0.91	56.33	3	48.5
12	MAGIC.11	14	20.58	0.66	41.56	5.67	36.25
13	MAGIC.12	21.33	19.05	0.67	52.33	4.67	49.35
14	MAGIC.14	18.67	21.14	0.72	49.78	6	48.5
15	MAGIC.15	15	21.71	0.73	49	6.33	41.9
16	MAGIC.16	18.33	21.37	0.71	66.67	3.33	18.17

# Data files

	A	B	C	D	E	F	G	
1	id	bolting_days	seed_weight	seed_area	ttl_seedspruit	branches	height	
2		A	B	C	D	E	F	G
3	1	marker	MAGIC.1	MAGIC.10	MAGIC.100	MAGIC.101	MAGIC.102	MAGIC.103
4	2	MN1_29291	A	A	A	B	B	A
5	3	MN1_29716	A	A	A	B	B	A
6	4	MN1_112907	A	A	A	B	B	A
7	5	MASC03771	A	B	A	A	B	A
8	6	MN1_197787	A	B	A	B	B	A
9	7	MN1_340810	A	B	A	A	A	A
10	8	MN1_395107	A	A	A	A	B	A
11	9	MN1_444820	A	A	–	A	A	A
12	10	MN1_494205	A	A	A	A	A	A
13	11	MN1_592863	A	B	A	A	A	A
14	12	MN1_592760	A	A	A	A	A	A
15	13	BKN118	A	A	A	A	A	A
16	14	MN1_1042427	A	B	B	A	B	B
	15	CRY2_1021	A	A	A	A	A	A
	16	CRY2_429	A	A	A	A	B	A

# Data files

	A	B	C	D	E	F	G															
1	id	bolting_days	seed_weight	seed_area	ttl_seedspruit	branches	height															
2		A	B	C	D	E	F	G														
3	1	marker	MAGIC.1	MAGIC.10	MAGIC.100	MAGIC.101	MAGIC.102	MAGIC.103														
4	2		A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T
5	3	1	marker	Bur	Can	Col	Ct	Edi	Hi	Kn	Ler	Mt	No	Oy	Po	Rsch	Sf	Tsu	Wil	Ws	Wu	Zu
6	4	2	MN1_29291	B	A	A	B	A	A	B	A	A	A	B	B	A	B	A	A	A	A	A
7	5	3	MN1_29716	B	A	A	B	A	A	B	A	A	A	B	B	A	B	A	A	A	A	A
8	6	4	MN1_112907	B	A	A	A	B	B	B	B	A	A	B	B	A	B	B	A	A	A	A
9	7	5	MASC03771	A	B	A	A	B	A	B	A	A	A	A	A	A	A	A	A	A	A	A
10	8	6	MN1_197787	B	B	A	A	A	B	B	A	A	A	A	A	A	B	A	A	A	B	B
11	9	7	MN1_340810	B	B	A	B	A	B	A	A	A	A	A	A	A	A	A	B	B	A	A
12	10	8	MN1_395107	A	A	A	A	A	A	B	A	A	A	A	A	A	A	A	A	A	A	A
13	11	9	MN1_444820	B	A	A	B	B	A	A	A	A	A	A	A	A	A	B	A	B	B	A
14	12	10	MN1_494205	B	A	A	A	B	A	A	A	A	A	A	A	A	A	A	A	B	A	A
15	13	11	MN1_592863	A	B	A	B	A	A	A	B	A	A	A	A	A	A	B	A	B	A	A
16	14	12	MN1_592760	B	A	A	A	B	A	A	A	A	A	A	A	A	A	A	A	A	A	A
	15	13	BKN118	B	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
	16	14	MN1_1042427	A	B	A	B	A	B	B	A	B	A	A	A	B	A	A	B	A	B	A
		15	CRY2_1021	A	A	A	A	B	A	A	A	A	A	A	A	A	A	A	A	A	A	A
		16	CRY2_429	A	A	A	A	A	B	B	B	A	B	A	A	A	A	A	A	B	A	A
		17	MASC07014	A	A	B	A	A	A	A	A	A	A	A	A	B	A	A	B	A	A	A
		18	MASC03609	A	A	A	A	A	A	B	B	A	B	B	B	A	A	A	B	A	A	A
		19	MN1_1296068	A	A	A	A	A	A	B	A	A	A	A	A	A	A	A	B	B	A	A
		20	MN1_1399466	A	A	A	A	A	A	B	A	A	A	A	A	A	A	-	B	A	A	A
		21	AXR1_381	A	B	A	A	A	B	A	A	A	B	A	A	A	A	A	B	A	B	A
		22	MASC07424	A	B	B	A	A	A	A	A	A	A	A	A	A	B	-	A	A	A	A

# Data files

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3	1	marker	MAGIC.1	MAGIC.10	MAGIC.100	MAGIC.101	MAGIC.102	MAGIC.103															
4	2		A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	
5	3	1	marker	Bur	Can	Col	Ct	Edi	Hi	Kn	Ler	Mt	No	Ov	Po	Rsch	Sf	Tsu	Wil	Ws	Wu	Zu	
6	4	2	MN1_292				A			B			C			B	A	B	A	A	A	A	A
7	5	3	MN1_297	1				marker			chr			pos			B	A	B	A	A	A	A
8	6	4	MN1_112	2				MN1_29291			1			0.029291			B	A	B	B	A	A	A
9	7	5	MASC037	3				MN1_29716			1			0.029757			A	A	A	A	A	A	A
10	8	6	MN1_197	4				MN1_112907			1			0.112907			A	A	B	A	A	A	B
11	9	7	MN1_340	5				MASC03771			1			0.174605			A	A	A	A	A	A	A
12	10	8	MN1_444	6				MN1_197787			1			0.197787			A	A	A	B	A	B	A
13	11	9	MN1_494	7				MN1_340810			1			0.34081			A	A	A	A	A	A	B
14	12	10	MN1_592	8				MN1_395107			1			0.395107			A	A	A	A	A	A	A
15	13	11	MN1_592	9				MN1_444820			1			0.444764			A	A	A	A	A	A	A
16	14	12	BKN118	10				MN1_494205			1			0.494205			A	A	A	A	A	A	A
	15	13	MN1_1042	11				MN1_592863			1			0.592867			A	A	A	A	A	B	A
	16	14	CRY2_1021	12				MN1_592760			1			0.592984			A	A	A	A	A	B	A
		15	CRY2_429	13				BKN118			1			0.761584			B	A	A	A	A	B	A
		16	MN1_1296	14				MN1_1042427			1			1.042428			A	A	A	A	A	B	B
			MN1_1399	15				CRY2_1021			1			1.187841			A	A	A	A	A	B	A
			AXR1_3	16				CRY2_429			1			1.188433			A	A	B	-	A	A	A
			MASC07014	17				MASC07014			1			1.189374									
			MASC03609	18				MASC03609			1			1.22655									

# Control file (json or yaml)

```
{
  "description": "Arabidopsis MAGIC data, Gnan et al (2014)",
  "crosstype": "magic19",
  "sep": ",",
  "na.strings": ["-", "NA"],
  "comment.char": "#",
  "geno": "arabmagic_geno.csv",
  "founder_geno": "arabmagic_foundergeno.csv",
  "gmap": "arabmagic_pmap_tair9.csv",
  "pmap": "arabmagic_pmap_tair9.csv",
  "pheno": "arabmagic_pheno.csv",
  "genotypes":
    "A": 1
    "H": 2
    "B": 3
},
  "geno_transposed": true,
  "founder_geno_transposed": true
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```

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  "gmap": "arabmagic_pmap_tair9.csv",  
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    "B": 3  
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  "pmap": "arabmagic_pmap_tair9.csv",
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  "genotypes":
    "A": 1
    "H": 2
    "B": 3
},
  "geno_transposed": true,
  "founder_geno_transposed": true
}
```

## Reading data into R

```
library(qt12)  
arab <- read_cross2("arab_magic.json")
```

# Reading data into R

```
library(qtl2)  
arab <- read_cross2("arab_magic.json")
```

19-way Arabidopsis MAGIC

Kover et al. (2009) PLoS Genet

Gnan et al. (2014) Genetics

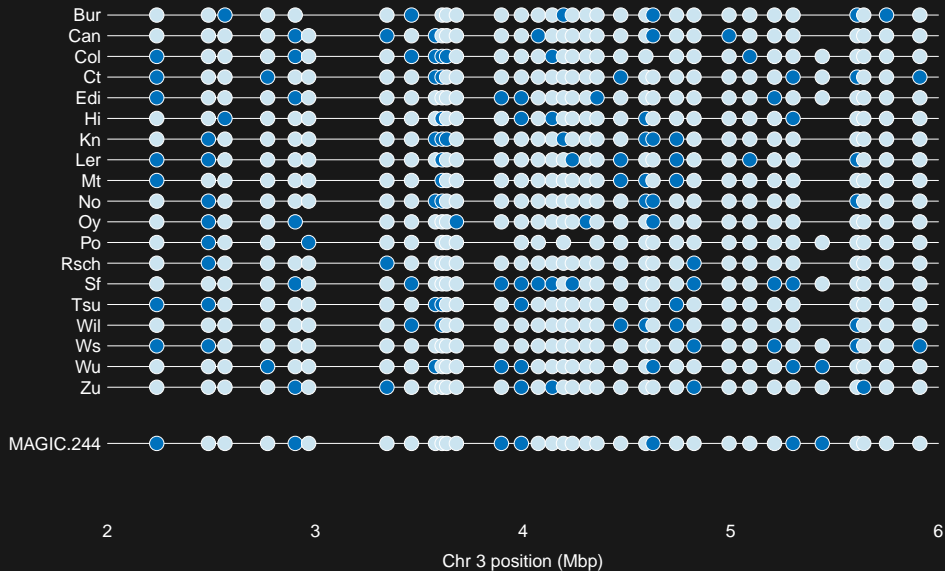
[github.com/rqtl/qtl2data](https://github.com/rqtl/qtl2data)

# Data diagnostics

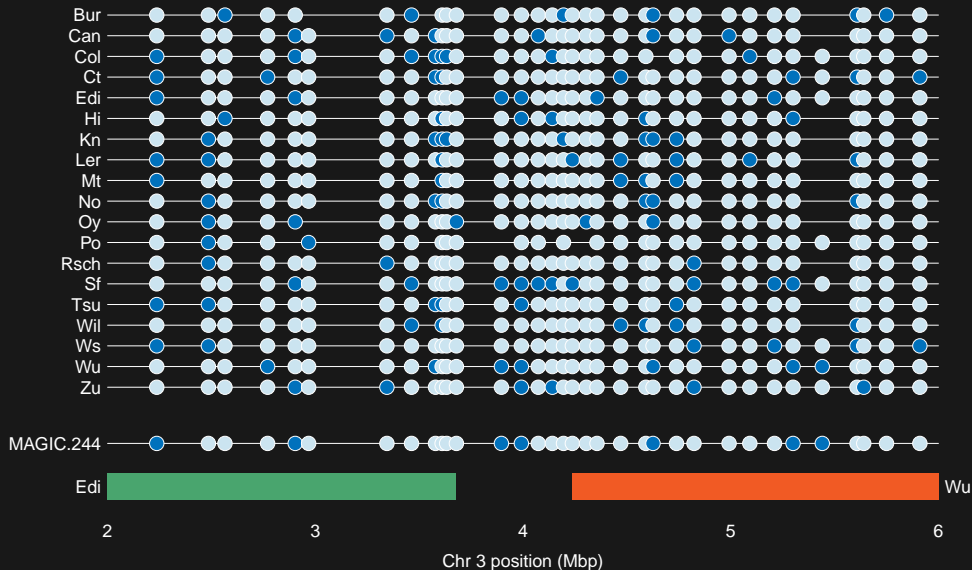
See Broman et al. (2019) Cleaning genotype data from  
Diversity Outbred mice. G3 9:1571–1579

doi: [10.1534/g3.119.400165](https://doi.org/10.1534/g3.119.400165)

# Genotype reconstruction



# Genotype reconstruction

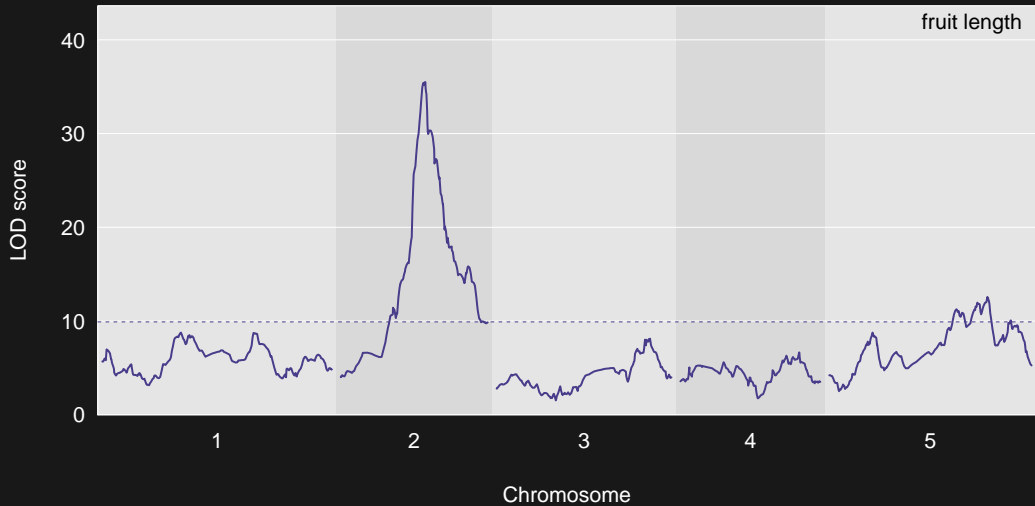


# Genotype reconstruction

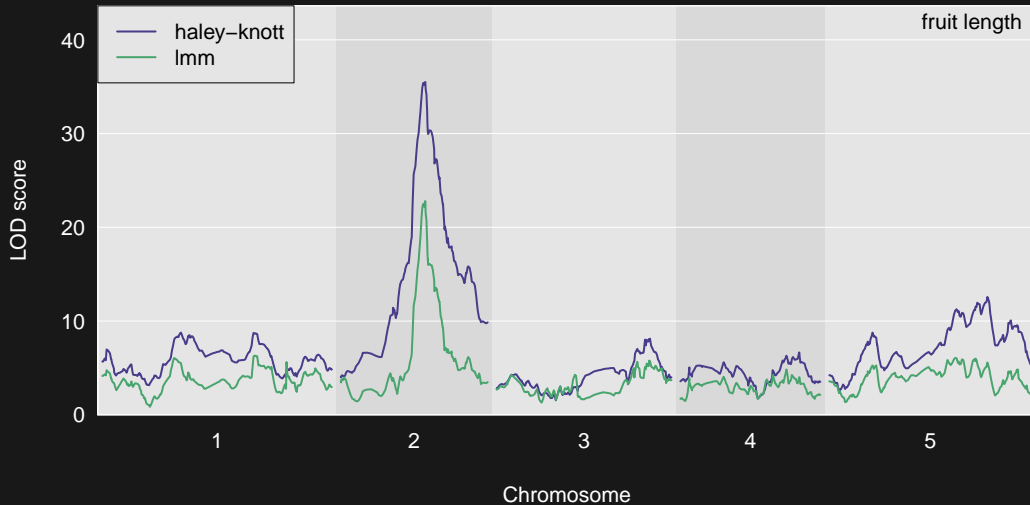
```
gmap <- insert_pseudomarkers(arab$gmap, step=0.2, stepwidth="max")  
pmap <- interp_map(gmap, arab$gmap, arab$pmap)  
  
pr <- calc_genoprob(arab, gmap, error_prob=0.002, cores=24)
```



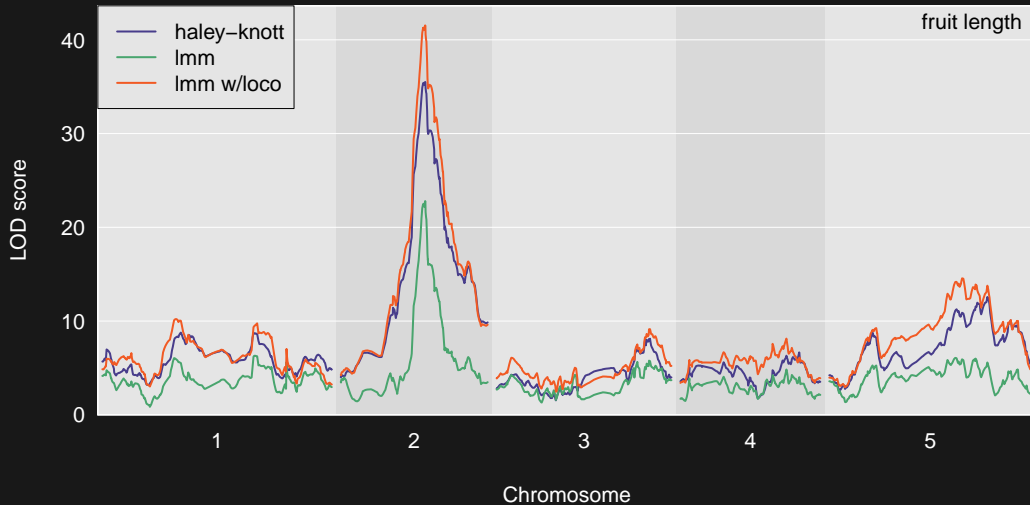
# Genome scan



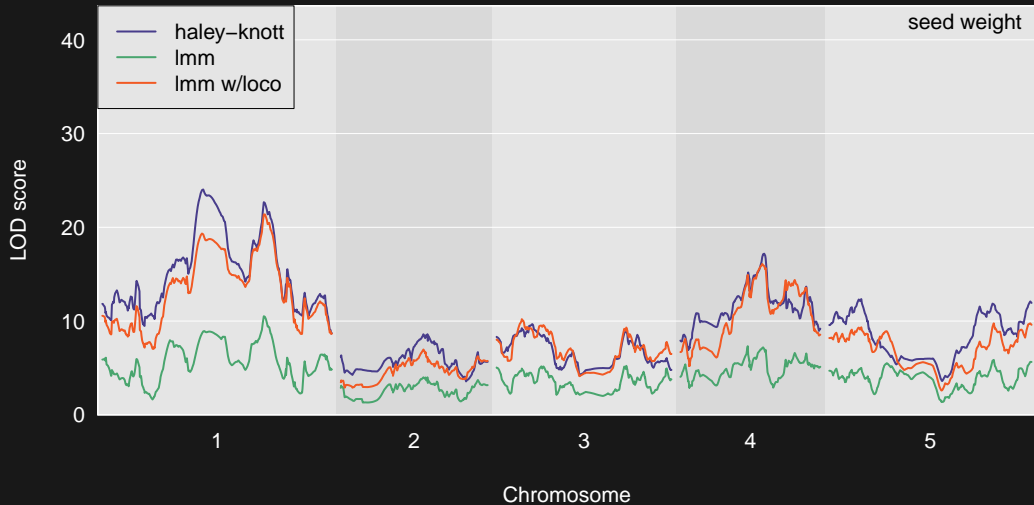
# Genome scan



# Genome scan



# Genome scan



# Genome scan

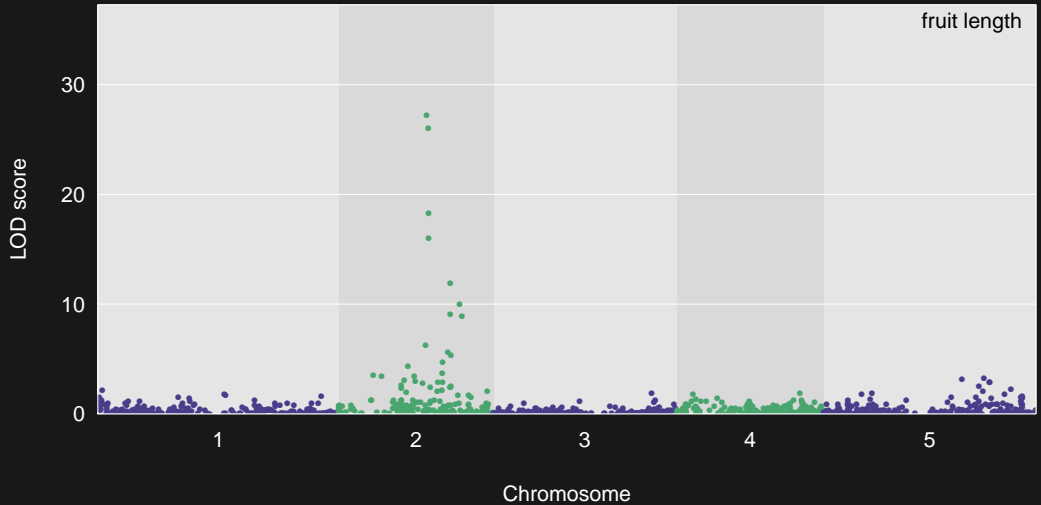
```
out_hk <- scan1(pr, arab$pheno, cores=24)

operm_hk <- scan1perm(pr, arab$pheno, n_perm=1000, cores=24)

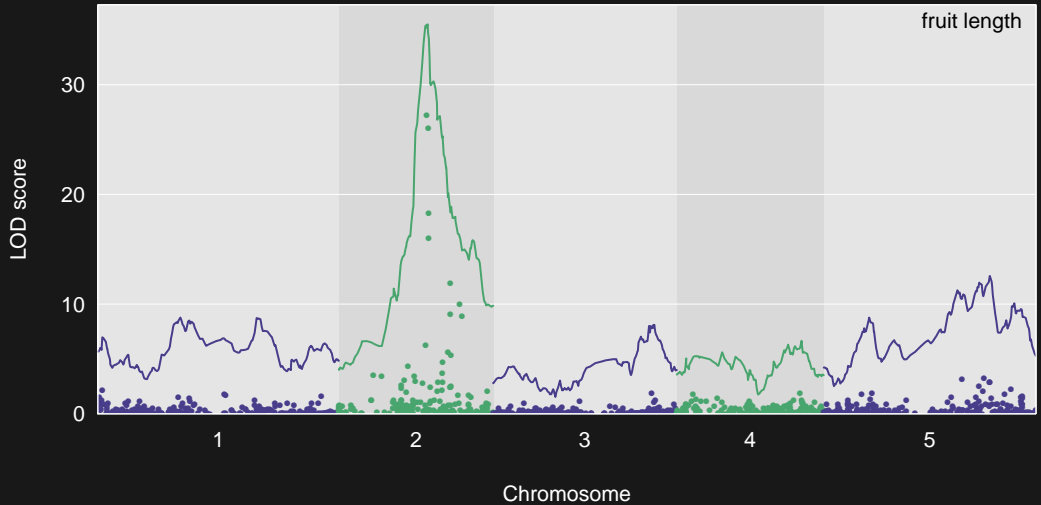
k <- calc_kinship(pr, cores=24)
out_lmm <- scan1(pr, arab$pheno, k, cores=24)

k_loco <- calc_kinship(pr, "loco", cores=24)
out_loco <- scan1(pr, arab$pheno, k_loco, cores=24)
```

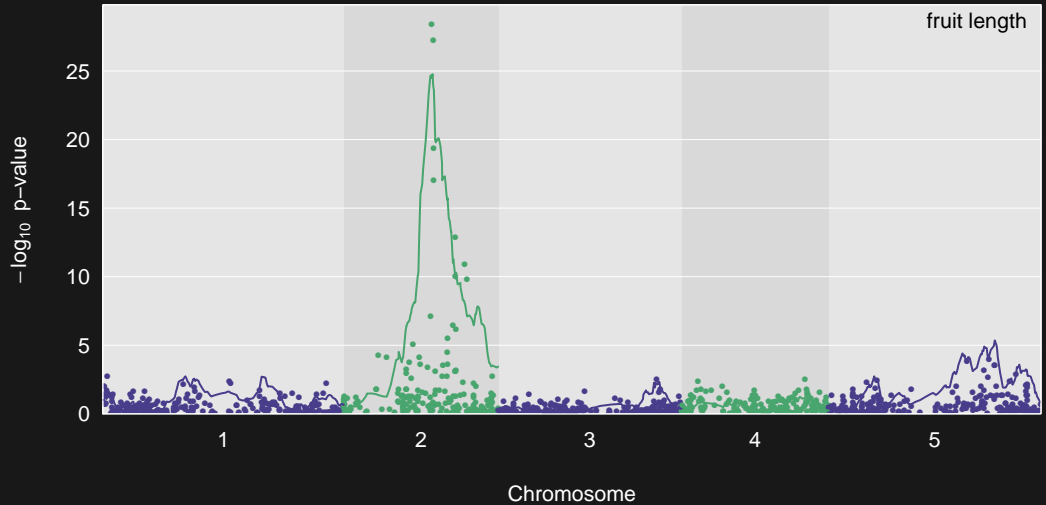
# SNP association scan



# SNP association scan

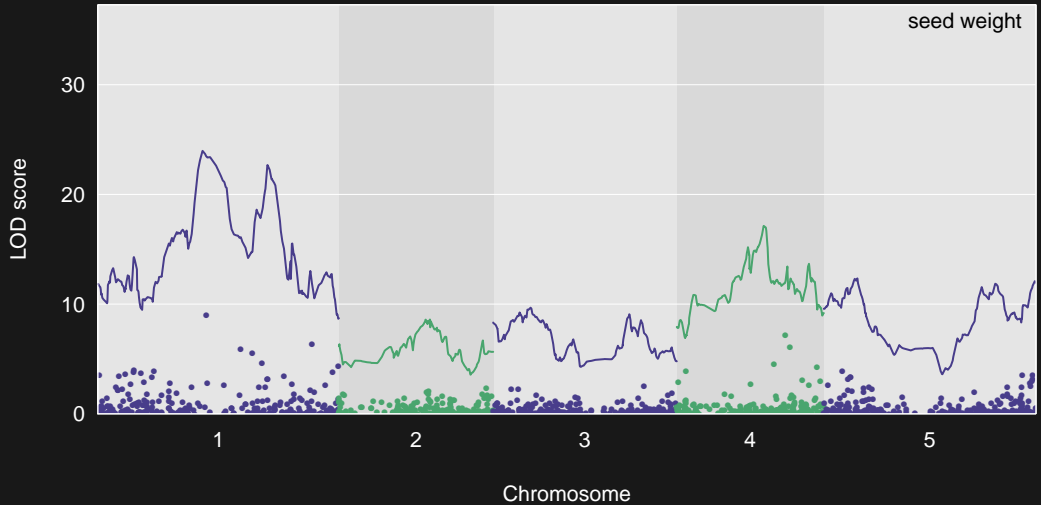


# SNP association scan

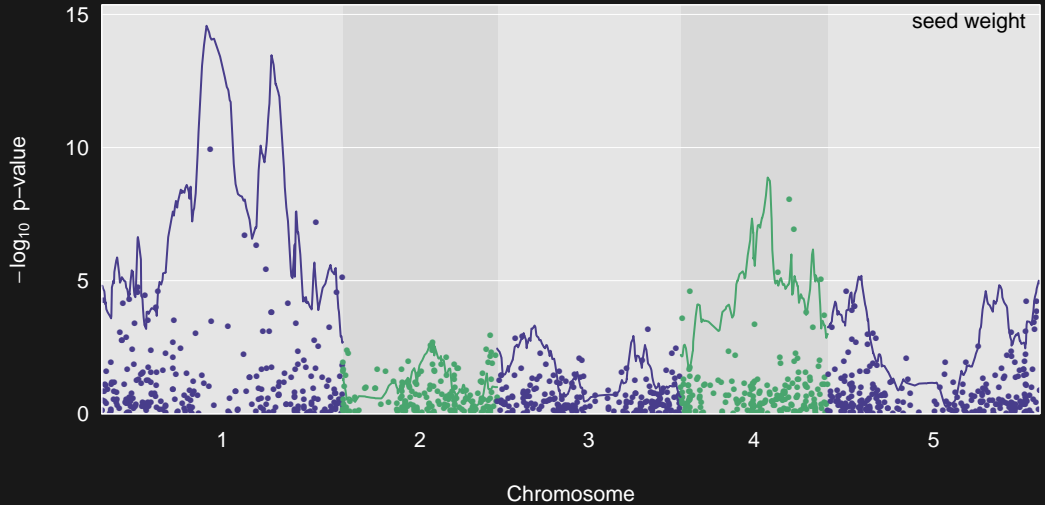




# SNP association scan



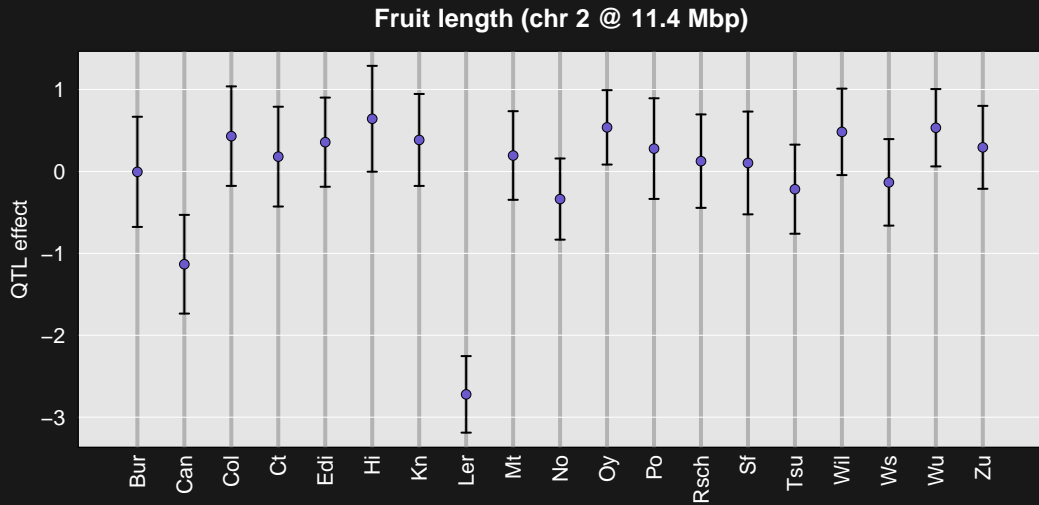
# SNP association scan



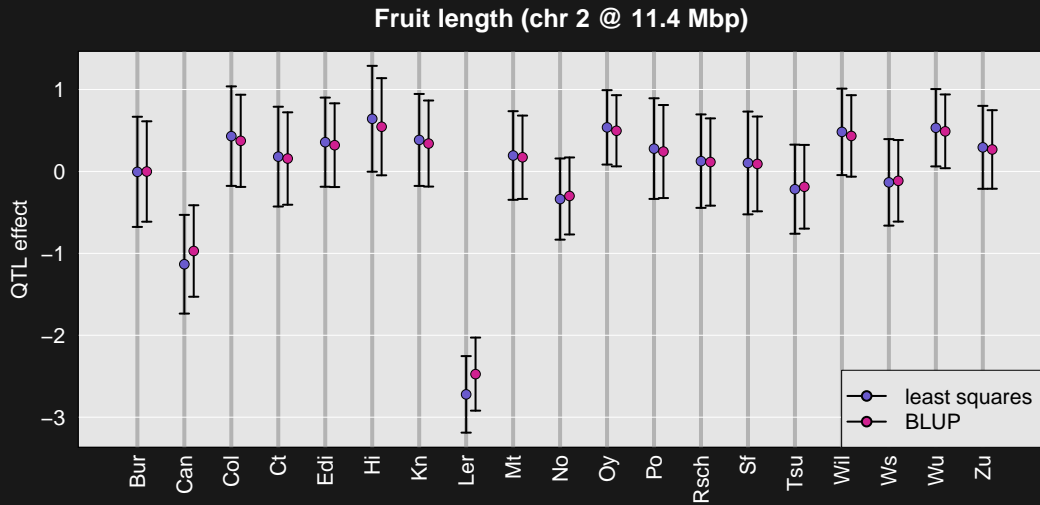
# SNP association scan

```
snp_pr <- genoprob_to_snpprob(pr, arab)  
out_snps <- scan1(snp_pr, arab$fruit, cores=24)
```

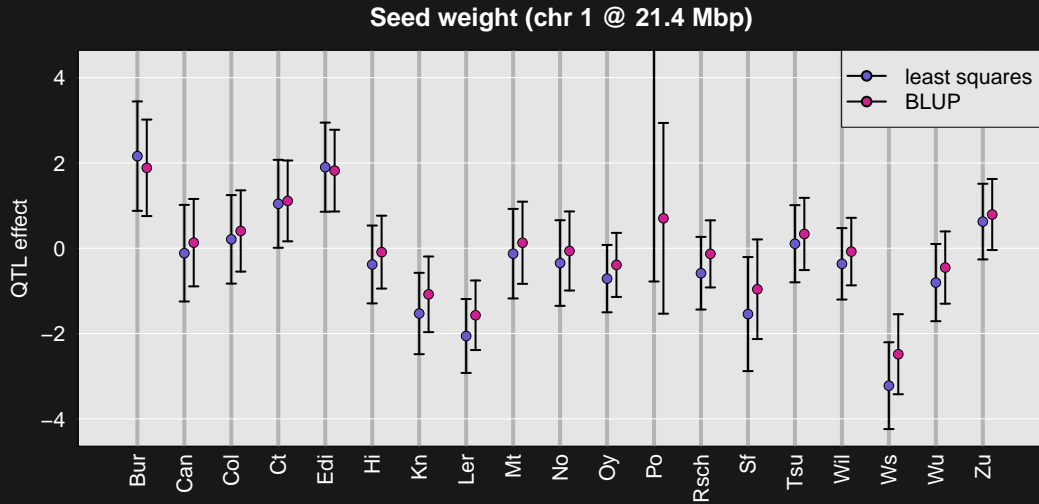
# QTL effects



# QTL effects



# QTL effects



# QTL effects

```
fl_peak <- max(out_hk, pmap, lodcolumn="fruit_length")
fl_pr <- pull_genoprobpos(pr, pmap, fl_peak$chr, fl_peak$pos)

fl_fit1 <- fit1(fl_pr, arab$pheno[, "fruit_length"])
fl_blup <- fit1(fl_pr, arab$pheno[, "fruit_length"], blup=TRUE)
```

# Goals

- ▶ Genotype reconstructions from external software
- ▶ General models for RIL and AIL
- ▶ Sequencing-based genotype data
- ▶ Multiple-QTL models
- ▶ QTL  $\times$  environment interactions
- ▶ Interactive data visualization



Slides: [bit.ly/msu2019-12](https://bit.ly/msu2019-12)



`kbroman.org`

[kbroman.org/qt12](https://kbroman.org/qt12)

`github.com/kbroman`

`@kwbroman`