

Introduction to Exome Sequencing and Variant Analysis

**Bioinformatics and Computational
Bioscience Branch (BCBB)**

2019-6-13



National Institute of
Allergy and
Infectious Diseases

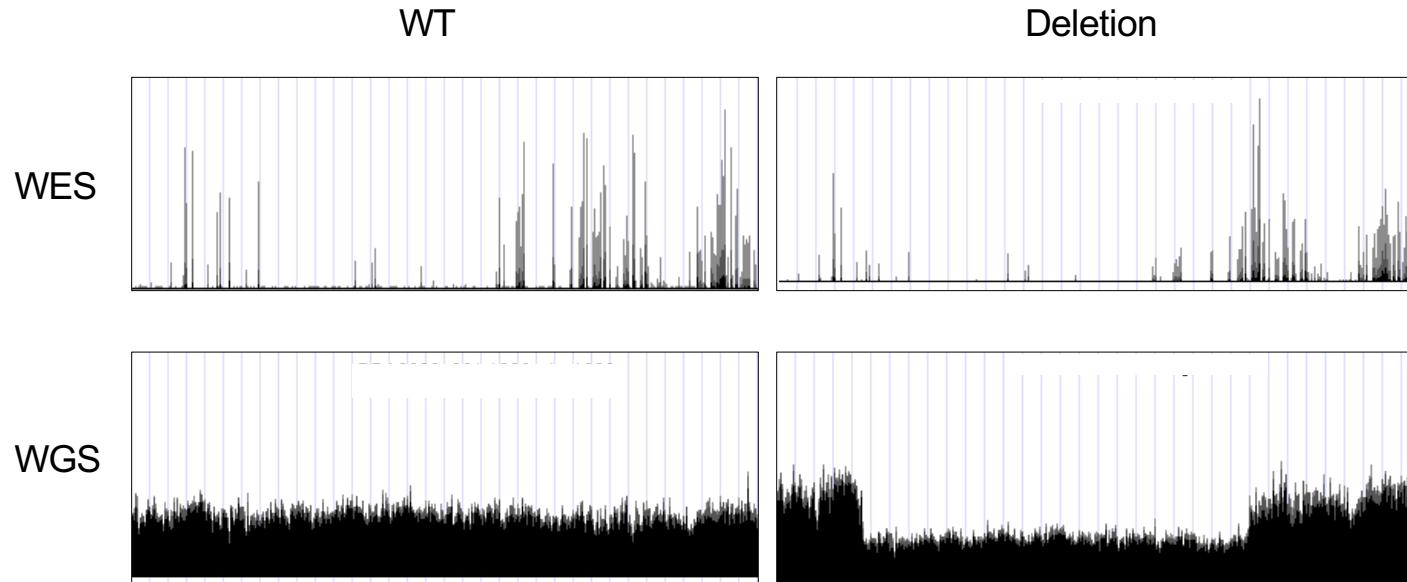
NIAID

Overview

- Part 1: *Seminar - Introduction to Whole Exome Sequencing*
- Part 2: *Hands on with Command Line*
 - Data Preprocessing and QC
 - Variant Discovery
 - Annotation and Downstream Analysis
- Part 3: *Seminar – Introduction to GWAS and CNV analysis*

Difference between WES and WGS

- Whole genome sequencing (WGS) has even coverage
- Even a deletion is observed by eye



National Institute of
Allergy and
Infectious Diseases

NIAID

All about exomes

- What is exome?
 - Sequencing targeted to exonic regions
 - ~2% of genome
- Important to know
 - You will NOT get a whole exome!
 - Not all exons in all genes are captured!
 - Important to know- Negative results vs no data
- Coverage will vary with targets

What to know about your data

- What is sequence depth?
 - The depth of your sequence
 - 10x, 30x, 50x, 100x
- Read length
 - How long is your read length?
- What software was used to align?
- What variant caller was used to call variants?
- Which reference sequence was used?
- Which capture kit was used? What is covered?



National Institute of
Allergy and
Infectious Diseases

NIH

Sequencers

Vendors

Illumina NovaSeq 6000



PacBio



Ion Torrent GeneStudio S5



Applications

- Large Whole-Genome Sequencing (human, plant, animal)
- Small Whole-Genome Sequencing
- Exome Sequencing
- Targeted Gene Sequencing
- Whole-Transcriptome Sequencing
- mRNA-Seq for gene expression

- Targeted Gene Expression Profiling
- Long-Range Amplicon Sequencing
- miRNA & Small RNA Analysis
- DNA-Protein Interaction Analysis
- Methylation
- 16S Metagenome

Applications determine platform choice



National Institute of
Allergy and
Infectious Diseases

NIH

Sequencers

Machine	Cost of Machine	Methods/Sample Throughput	Reads per run	Read length (bp)	Error rate
Ion Torrent GeneStudio S5	\$80K	Small genomes, targeted gene	2-130 Million	200-600	1.71%
Illumina NextSeq 550	\$250K	3 Exomes 13 Transcriptomes 30 Small Genomes	130-400 Million	2x150	0.8%
Illumina HiSeq 4000	\$654K	12 Genomes 96 Exomes 100 Transcriptomes	2.5-5 Billion	2X150	0.76%
Illumina NovaSeq 6000	\$900 K	48 Genomes 500 Exomes 400 Transcriptomes	16-20 Billion	2X250	0.8 %
PacBio Sequel II	\$695K	Small Genomes, targeted gene, methylation	100 MB	15 kb	13-15%



National Institute of
Allergy and
Infectious Diseases

Error rates vary between different platforms as well as the types of error (indel versus mismatches etc) and in biases depending on the sequence content (for example, GC skewing) requiring different error handling algorithms

NIH

Comparison between WES and WGS

	Exomes (ES/WES)	Genomes (GS/WGS)
DNA	Targeted and captured	Sheared DNA
What you can get	Most coding regions (+UTR)	coding and non-coding
Variants that can be examined	SNVs, indels (CNVs)	SNVs, indels, CNVs, structural variations
Size of bam files	~10 Gb	~200 Gb
Cost (research vs clinical, platform, coverage etc.)	\$300 - \$1000	\$500 - \$6500



National Institute of
Allergy and
Infectious Diseases

NIAID

Partial list of capture enrichment kits

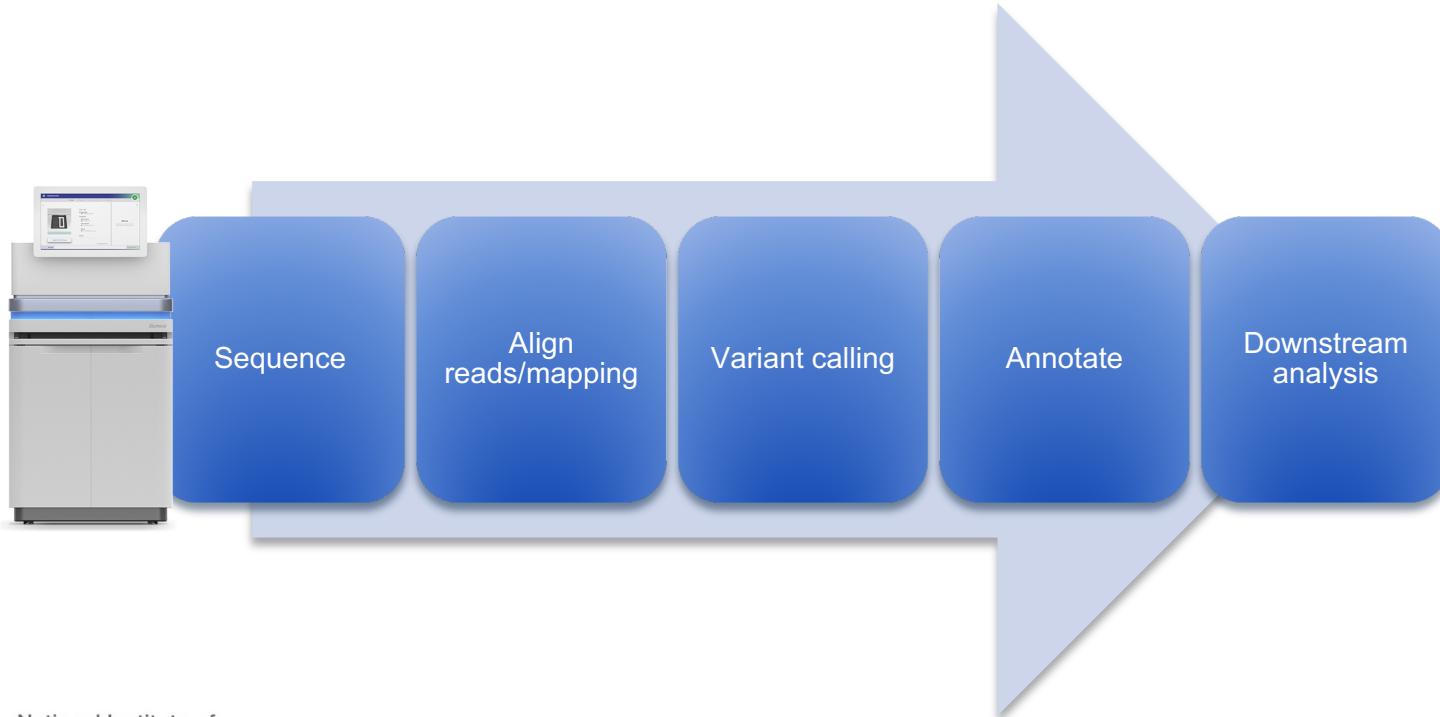
Manufacturer	Kits	Regions targeted	Bases covered
IDT	xGen Exome Research Panel	Exons	39Mb
Illumina	Nextera Rapid capture	Exons + UTRs + miRNA	62 Mb
Nimblegen	SeqCap EZ Exome	Exons + UTR	96 Mb
	SeqCap EZ MedExome	Disease-associated regions	47 Mb
Agilent	SureSelect Human All Exon V6	Exons + UTRs	60Mb
	Clinical Research Exome	Disease-relevant targets	51Mb



National Institute of
Allergy and
Infectious Diseases

NIAID

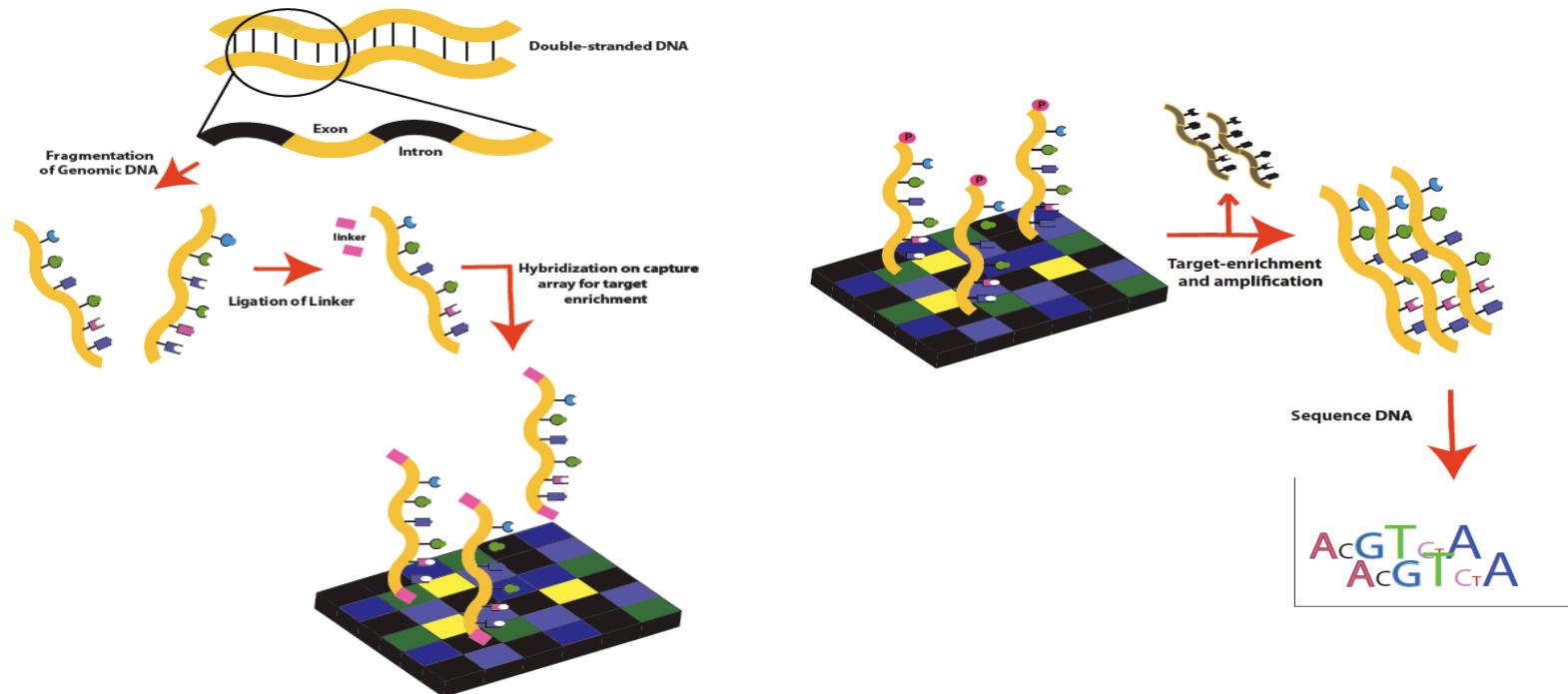
Overview of next-generation sequence processing



National Institute of
Allergy and
Infectious Diseases

NIAID

Exome sequencing overview



National Institute of
Allergy and
Infectious Diseases

Source: https://en.wikipedia.org/wiki/Exome_sequencing#/media/File:Exome_Sequencing_workflow_1b.png. 2012 Jan; 71(1):5-14

NIH

Popular tools

Task	Popular tools
Align reads/mapping	BWA-mem Bowtie NovoAlign BLASR (for PacBio reads)
Variant calling	GATK4 (Broad) DeepVariant (Google) SpeedSeq (Wash. Univ. St. Louis) Platypus (Wellcome Trust) Starling (Illumina)
Annotating variants	VEP (EMBL-EBI) AnnoVar snpEff Funcotator (Broad)



National Institute of
Allergy and
Infectious Diseases

NIAID

Popular resources

Task	Popular sources
Control population frequency	gnomAD TGAC ExAC 1000GP ESP
Annotation	RefSeq Ensembl UCSC genes GENCODE
Visualization	UCSC genome browser IGV
Clinical relevance	HGMD OMIM CGD ClinVar



National Institute of
Allergy and
Infectious Diseases

NIAID

GATK Best Practices Website

gatk [Home](#) [About](#) [Guide](#) [Blog](#) [Forum](#) [Download](#)

Guide

GATK Best Practices

[Presentations](#)
[Methods and Algorithms](#)
[Tool Documentation](#)

[FAQs](#)
[Tutorials](#)
[Dictionary](#)
[Common Problems](#)
[Bug Tracker](#)

[Version History](#)

[Guide to the Guide](#)
[Search Tags](#)

[Pipelining w/ Queue](#)
[Developer Zone](#)

[Third-Party Tools](#)

Current version is 3.5

The [Best Practices](#) have been updated for GATK version 3. If you are running an older version, you should seriously consider upgrading. For more details about what has changed in each version, please see the [Version History](#) section. If you cannot upgrade your version of GATK for any reason, please look up the corresponding version of the [GuideBook PDF](#) (also in the [Version History](#) section) to ensure that you are using the appropriate

GATK Best Practices

Recommended workflows for variant discovery analysis with GATK

What can you do with this?

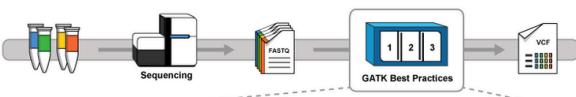
The GATK Best Practices workflows provide step-by-step recommendations for performing variant discovery analysis in high-throughput sequencing (HTS) data. They enable discovery of SNPs and small indels (size limit in theory but adjustments may be required to call indels > 50 bp) in DNA and RNAseq. They do not yet enable discovery of structural variants (SVs) or copy number variants (CNVs). Although they were originally designed for human genome research, the GATK Best Practices are widely used (with adaptations as described in the documentation) for analysis of non-human organisms of all kinds, including non-diploids.

What's in the box?

Our recommendations describe in detail the core processing steps required to go from raw reads coming off the sequencing machine, all the way to a variant callset that can be used in downstream analyses. Wherever we can, we try to provide guidance regarding experimental design, quality control (QC) and pipeline implementation options (how to set it up in practice), but please understand that those are dependent on many factors including sequencing technology and the hardware infrastructure that are at your disposal, so you may need to adapt our recommendations to your specific situation.

Break it down

We currently have two separate workflows for Germline DNA and for RNAseq, and we are developing a workflow for Somatic DNA. All of them are divided into three sequential phases:



1. PRE-PROCESSING
Pre-processing starts from raw sequence data, either in FASTQ or uBAM format, and produces analysis-ready BAM files. Processing steps include alignment to a reference genome as well as some data cleanup operations to correct for technical biases and make the data suitable for analysis.

2. VARIANT DISCOVERY
Variant Discovery starts from analysis-ready BAM files and produces a callset in VCF format. Processing involves identifying sites where one or more individuals display possible genomic variation, and applying filtering methods appropriate to the experimental design.

3. CALLSET REFINEMENT
Callset Refinement starts and ends with a VCF callset. Processing involves using meta-data to assess and improve genotyping accuracy, attach additional information and evaluate the overall quality of the callset.

Use the buttons in the figure above to start exploring the Best Practices workflows in detail.

<https://www.broadinstitute.org/gatk/guide/best-practices>

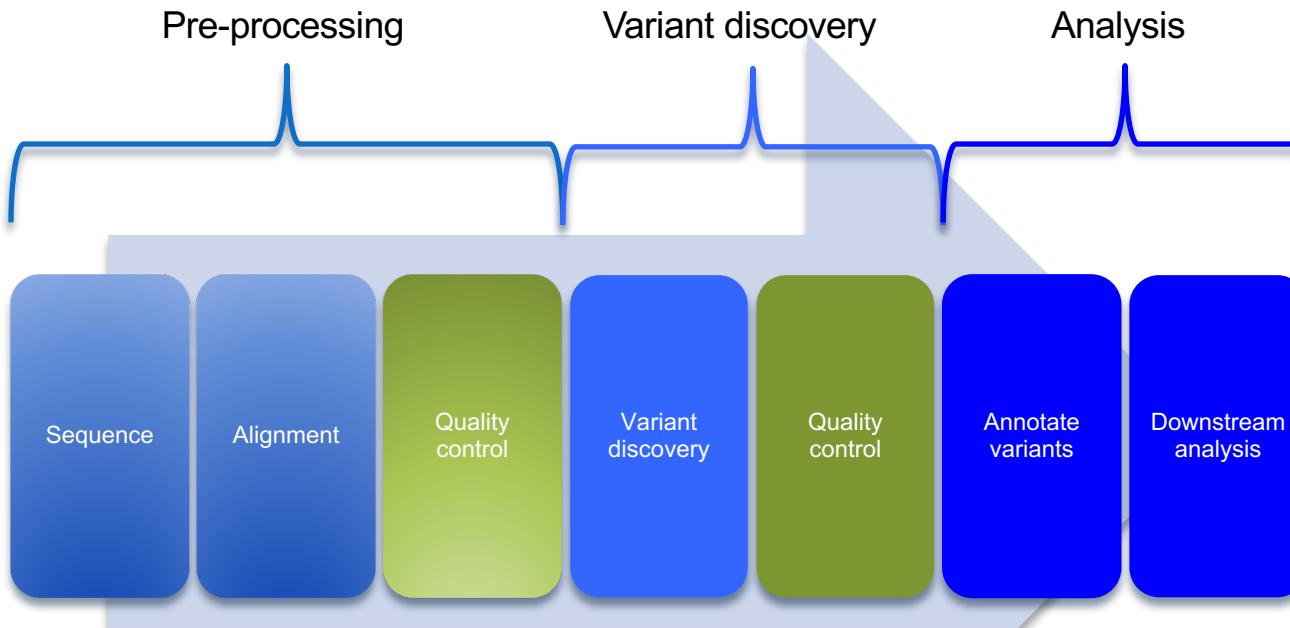
Introduction to WES Analysis Workflow



National Institute of
Allergy and
Infectious Diseases

NIAID

WES pipeline overview



National Institute of
Allergy and
Infectious Diseases

NIAID

Files formats

File type	Origin
FASTA	Text-based format for reference
FASTQ	Raw reads from sequencer
SAM BAM CRAM	Sequence Alignment/Map Binary version of SAM Compressed BAM
gVCF/VCF	Variant call format

Index files

- Index files are needed for files in next-generation analysis, as file sizes are big!
- Enables program to efficiently access the data, rather than having to read the whole file

File type	Index
FASTA	*fai
BAM	*bai
CRAM	*crai
VCF	*vcf.idx

Tools

Tool	Purpose
BWA mem	Read alignment to reference
Picard	Mark duplicates
FastQC	Short reads QC
samtools	Sort sam/bam, convert sam<->bam
bcftools	VCF query, normalization
GATK4 (Haplotype caller)	QC/variant calling
verifyBamId	Contamination detection
PEDDY	Family relationship & gender check
VEP	Variant annotation
Gemini	Mendelian inheritance analysis

Reference genome

- There are different versions of human reference genome

Reference Name	Chromosome Notation	Mitochondrial Sequence	Additional Sequences Included
GRCh37 (Genome Reference Consortium)	1, 2, ..., X, Y, MT	Yes	<ul style="list-style-type: none">UnlocalizedUnplacedAlternate loci
hg19 (UCSC Genome Browser)	chr1, chr2, ..., chrX, chrY, chrM	Copied from previous release	<ul style="list-style-type: none">(same as above)
b37/b37+decoy/hs37d5 (1000 Genomes Project)	1, 2, ..., X, Y, MT	Yes	<ul style="list-style-type: none">UnlocalizedUnplaced“decoy” sequenceHuman herpesvirus 4 type 1
GRCh38 (Genome Reference Consortium)	chr1, chr2, ..., chrX, chrY, chrM	Yes	<ul style="list-style-type: none">UnlocalizedUnplacedAlternate lociModel centromeresGRCh37 Assembly corrections
hg38 (UCSC Genome Browser)	chr1, chr2, ..., chrX, chrY, chrM	Yes	<ul style="list-style-type: none">(same as above)



National Institute of
Allergy and
Infectious Diseases

* Unlocalized: chromosome known, exact location unknown

* Unplaced: known to originate from human genome, chromosome unknown

* Alternate loci: alternate representation of specific human regions

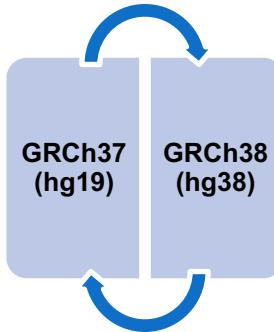
Reference FASTA index file

Contig name	1	249250621	52	60	61
	2	243199373	253404903	60	61
	3	198022430	500657651	60	61
	4	191154276	701980507	60	61
	5	180915260	896320740	60	61
	6	171115067	1080251307	60	61
The number of bases in the contig	7	159138663	1254218344	60	61
	8	146364022	1416009371	60	61
	9	141213431	1564812846	60	61
	10	135534747	1708379889	60	61
	11	135006516	1846173603	60	61
	12	13	1983430282	60	61
	13	11	2119513096	60	61
	14	10	2236602526	60	61
	15	102531392	2345741279	60	61
	16	90354753	2440081581	60	61
	17	81195210	25	60	61
	18	78077248	26	60	61
	19	59128983	27	60	61
	20	63025520	2763883926	60	61
	21	48129895	2827959925	60	61
	22	51304566	2876892038	60	61
X	155270560	2929051733			61
Y	59373566	3086910193			61
MT	16569	3147273397	70		
	GL000207.1	4262	3147290265	60	61
	GL000226.1	15008	3147294661	60	61
	GL000229.1	19913	3147309982	60	61
	GL000231.1	27386	3147330289	60	61
	GL000210.1	27682	3147358194	60	61
	GL000239.1	33824	3147386400	60	61
	GL000235.1	34474	3147420850	60	61
	GL000221.1	34474	3147420850	60	61



Converting Coordinates from One Genome Version to Another

- UCSC provides tools ([liftOver](#)) to convert BED file from one genome assembly to another



Lift over can fail for different reasons:

- 1) **Genome position cannot be lifted** -- a SNV resides in a contig that only exists in older/newer reference build, liftOver cannot give it new genome.
- 2) **Reference allele and alternative allele flipped in different reference build** -- G(ref)/C(alt) in GRCh37 and C(ref)/G(alt) in GRCh38

Why BWA+ GATK Haplotype caller?

- Widely accepted as the “conventional” way of processing NGS data
- Well assessed (many citations)
- Well documented
- Software is supported and still being updated
- Community support for troubleshooting or information

QC Checks for Data Quality

- Sequencer generated short reads – FastQC
- Coverage at different levels of partitioning and aggregation – GATK3 DepthOfCoverage
- Sample contamination – verifyBamId
- Familial-relationships and gender check – peddy

QC on FASTQ/BAMs by fastQC

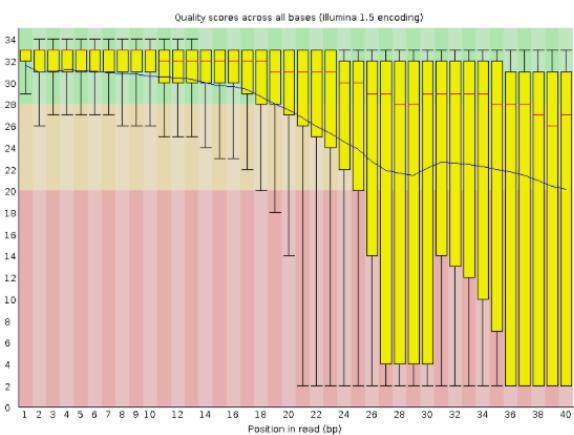
Summary

- ✓ Basic Statistics
- ✗ Per base sequence quality
- ✗ Per tile sequence quality
- ✓ Per sequence quality scores
- ! Per base sequence content
- ! Per sequence GC content
- ✓ Per base N content
- ✓ Sequence Length Distribution
- ! Sequence Duplication Levels
- ! Overrepresented sequences
- ✓ Adapter Content
- ! Kmer Content

Basic Statistics

Measure	Value
Filename	bad_sequence.txt
File type	Conventional base calls
Encoding	Illumina 1.5
Total Sequences	395288
Sequences flagged as poor quality	0
Sequence length	40
%GC	47

Per base sequence quality



Bad quality

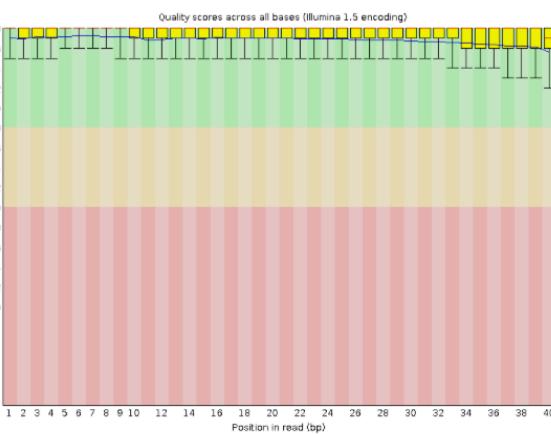
Summary

- ✓ Basic Statistics
- ✓ Per base sequence quality
- ✓ Per tile sequence quality
- ✓ Per sequence quality scores
- ✓ Per base sequence content
- ✓ Per sequence GC content
- ✓ Per base N content
- ✓ Sequence Length Distribution
- ✓ Sequence Duplication Levels
- ✓ Overrepresented sequences
- ✓ Adapter Content
- ! Kmer Content

Basic Statistics

Measure	Value
Filename	good_sequence_short.txt
File type	Conventional base calls
Encoding	Illumina 1.5
Total Sequences	250000
Sequences flagged as poor quality	0
Sequence length	40
%GC	45

Per base sequence quality



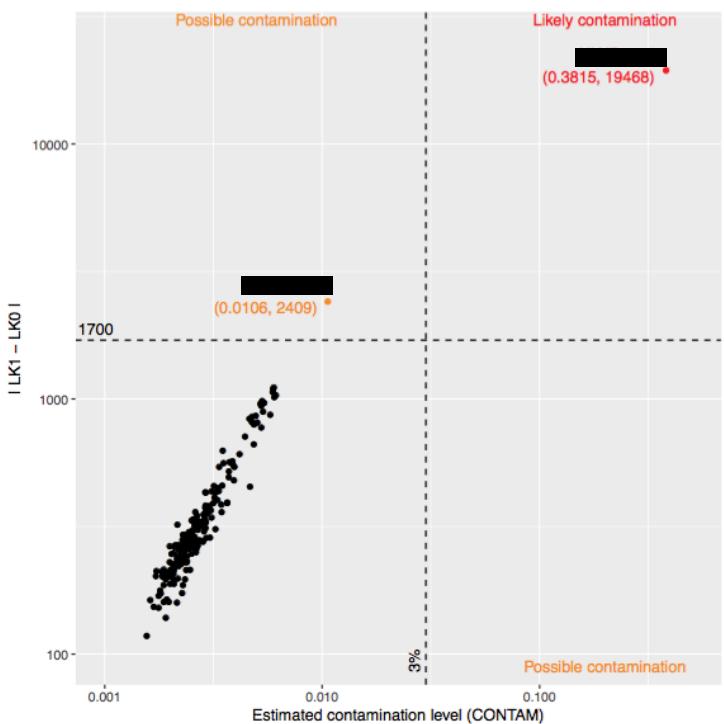
Good quality



National Institute of
Allergy and
Infectious Diseases

NIAID

QC on sample contamination by verifyBamId



Likely contaminated

ID	CONTAM	LK1-LK0	NumSNPs	NumReads	AvgDP
	0.38148	19468.34	15627	441693	28.26

CONTAM > 0.03 AND |LK1-LK0| > 1700.

CONTAM is the estimated fraction of contamination based on verifyBamID.

Possibly contaminated

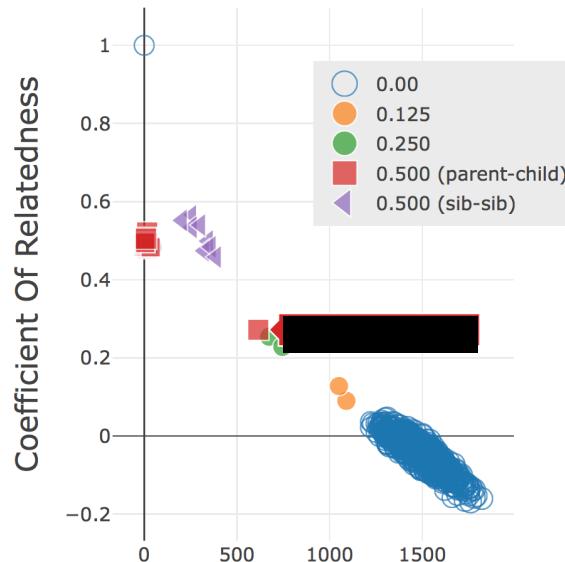
ID	CONTAM	LK1-LK0	NumSNPs	NumReads	AvgDP
	0.01062	2409.17	15627	634552	40.61

$\text{CONTAM} \geq 0.03$ OR $|LK1-LK0| \geq 1700$ (not both)

CONTAM is the estimated fraction of contamination based on verifyBamID.

QC on family-relationship and gender by peddy

Example of pedigree conflict



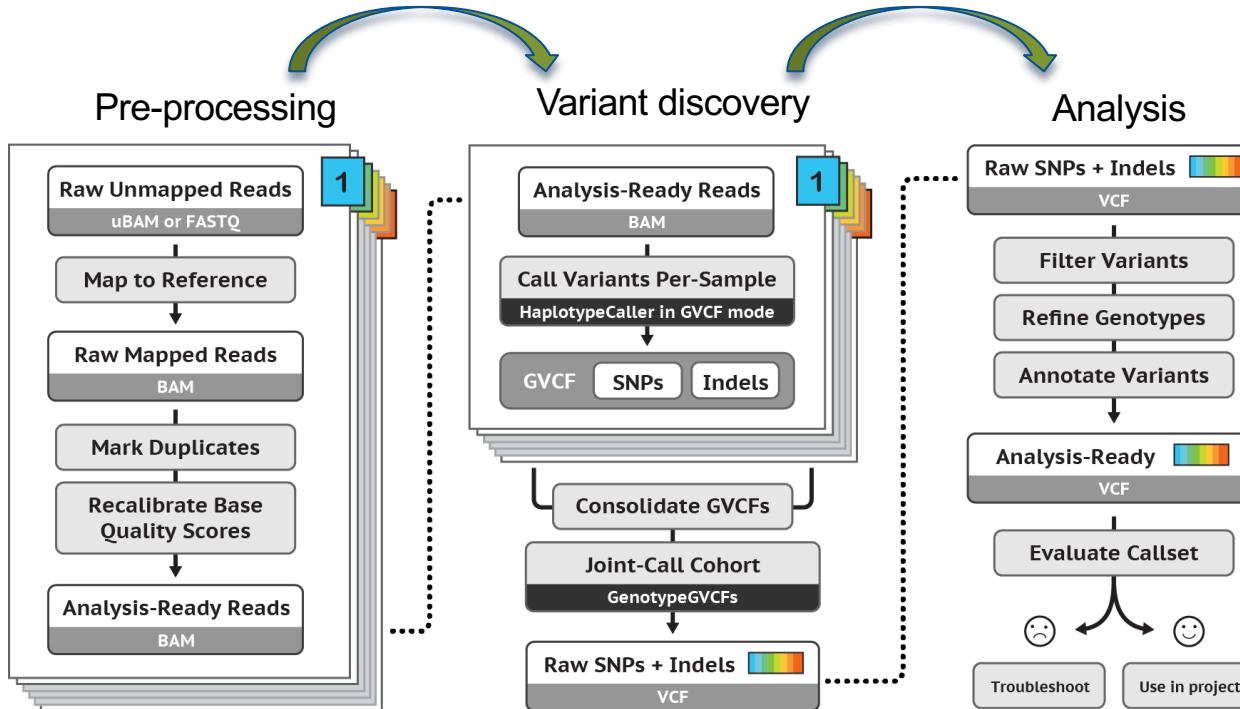
Example of gender conflict



National Institute of
Allergy and
Infectious Diseases

NIH

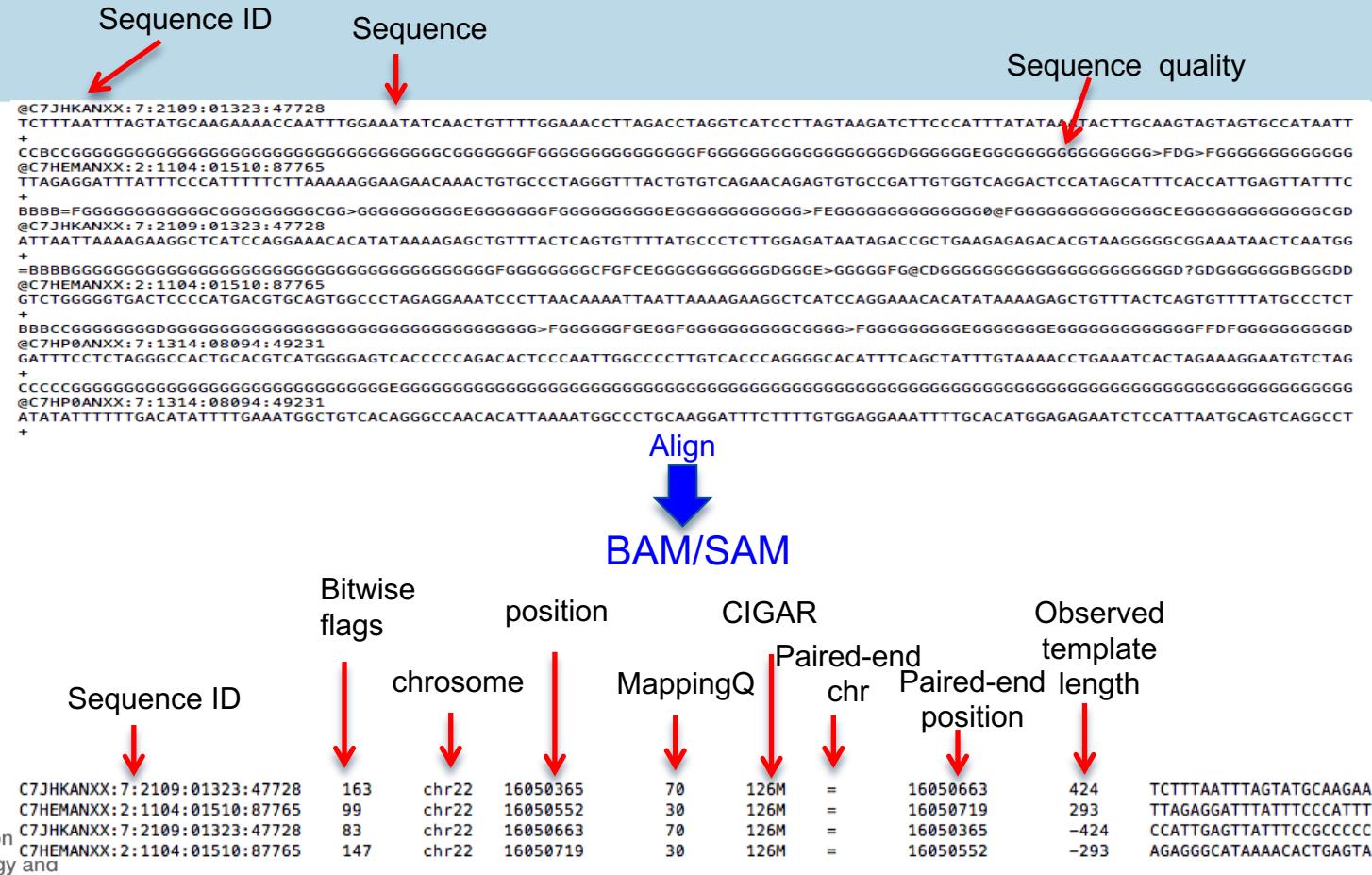
GATK Best Practices Workflow



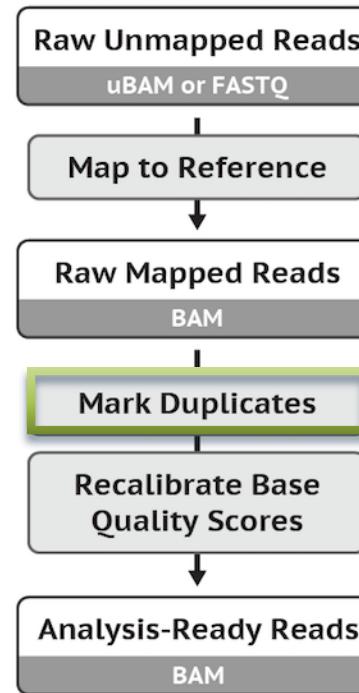
National Institute of
Allergy and
Infectious Diseases

NIAID

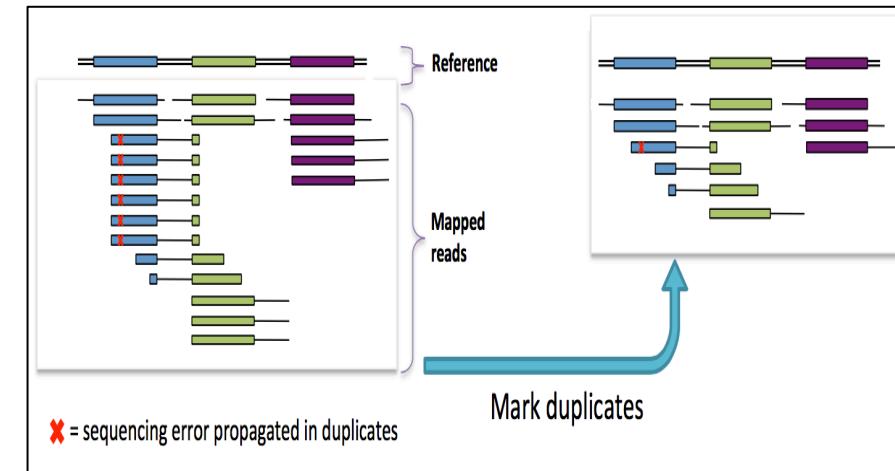
FASTQ



Marking duplicates and why is it necessary?



- It is assumed that each read corresponds to an independent DNA fragment from *randomly* sheared DNA
- However, PCR amplification and optic sensor artifact can cause duplicates
 - Identify based on start + stop of reads
 - Choose the best and ignore the rest

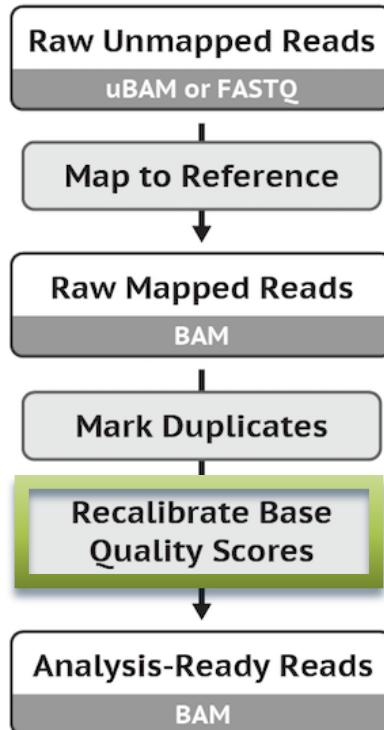


Recalibrate Base Quality Scores

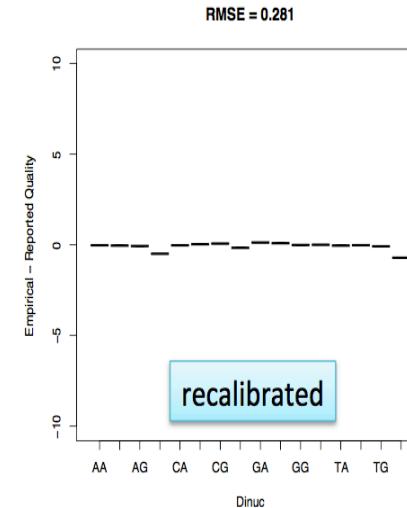
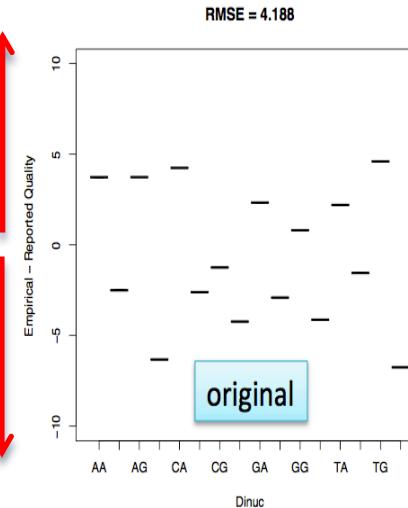


- Base scores are produced by sequencers
- Quality scores are **inaccurate and biased**
 - Prone to various technical errors
 - QS are often over- or under estimated
- To identify and correct non-random technical error
 - Physics or the chemistry of sequencing reactions
 - Manufacturing flaws in the equipment
- Error covariates e.g.
 - Reported quality score
 - Position within the read (machine cycle)
 - Preceding and current nucleotide (sequencing chemistry)

Recalibrate Base Quality Scores



Over-estimation ↑
↓ Under-estimation



- GATK BSQR builds model based on the known variants set
- Adjusts the base quality scores in the data based on the model

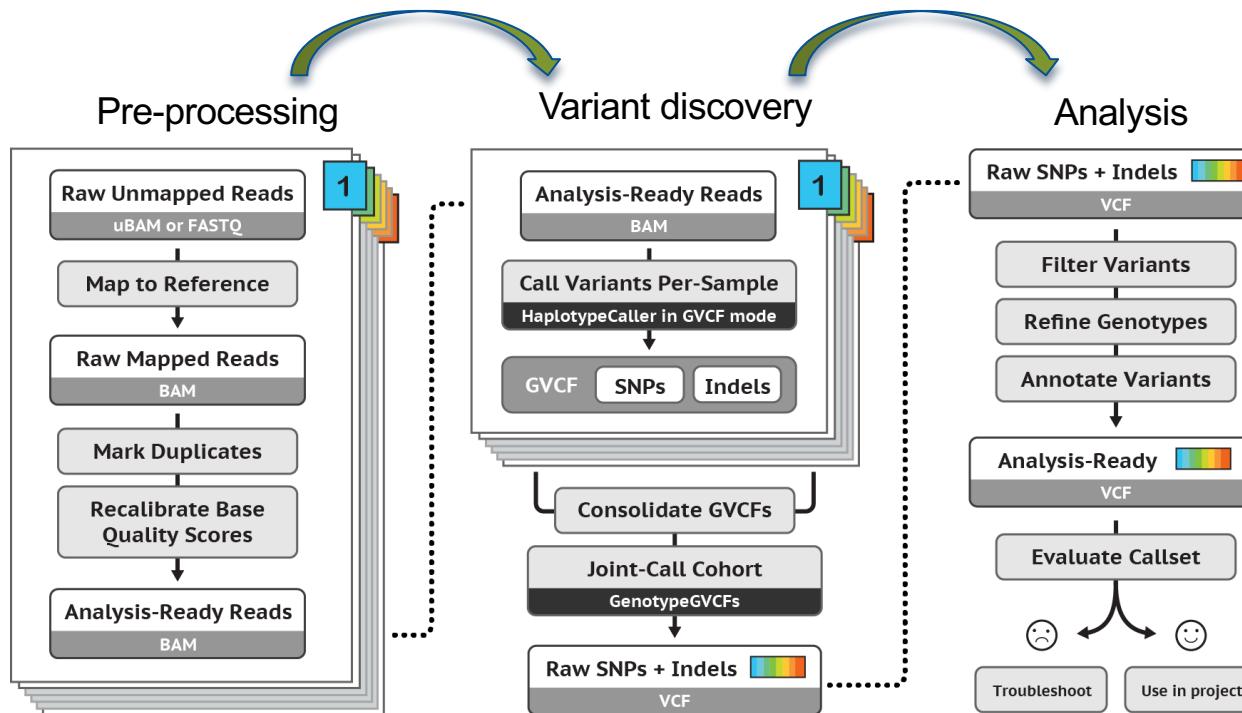


National Institute of
Allergy and
Infectious Diseases

Broad Institute

NIAID

GATK Best Practices Workflow

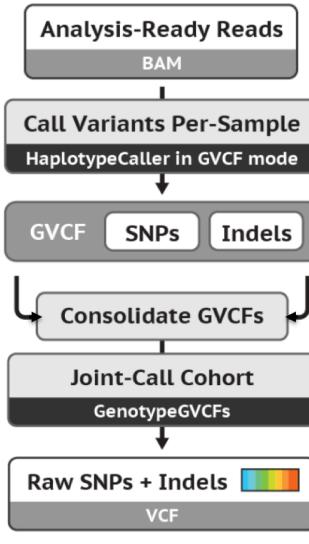


National Institute of
Allergy and
Infectious Diseases

Broad Institute

NIAID

Generate genotype likelihoods in each sample (gVCF)



- For a single sample, calculates normalized Phred-scaled likelihoods (PL) for genotypes:
 - “likelihood of the genotype” = “the probability that the genotype is *not* correct”
 - Normalized so that the most likely genotype’s PL is 0



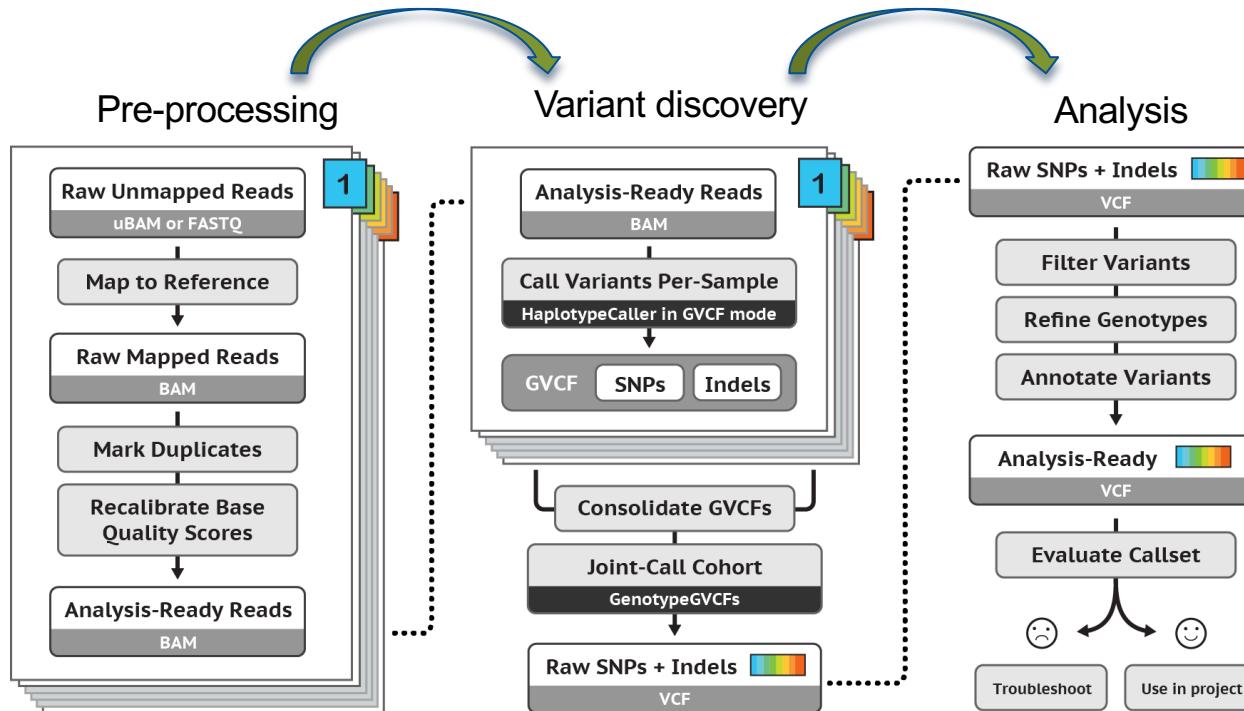
16	105459	.	G	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:60,0:60:99:0,120,1800
16	105460	.	T	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:60,0:60:99:0,120,1800
16	105461	.	T	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:61,0:61:99:0,120,1800
16	105462	.	A	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:61,1:62:99:0,120,1800
16	105463	.	C	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:64,0:64:99:0,120,1800
16	105464	.	T	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:64,0:64:99:0,120,1800
16	105465	.	C	<NON_REF>	.	.	.	GT:AD:DP:GQ:PL	0/0:66,0:66:99:0,120,1800

What is joint genotyping?



- If we analyze Sample 1 or Sample N alone, we are not confident that the variant is real
- If we see the variant in both samples, we are more confident that there is real variation at this site in this cohort

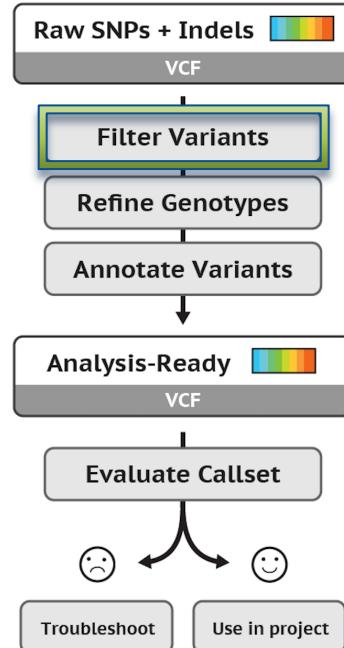
GATK Best Practices Workflow



National Institute of
Allergy and
Infectious Diseases

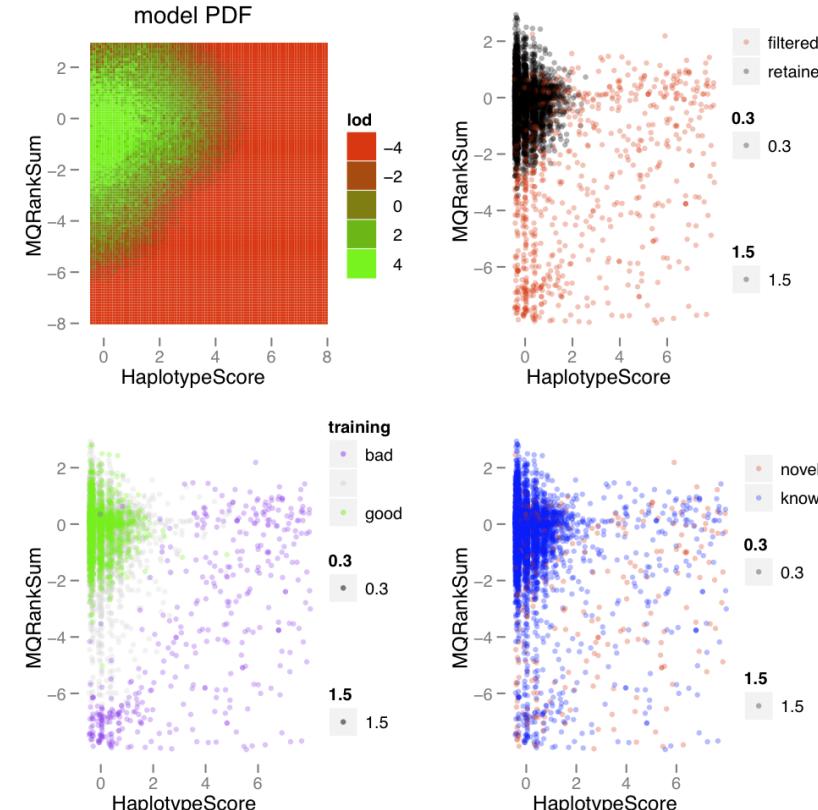
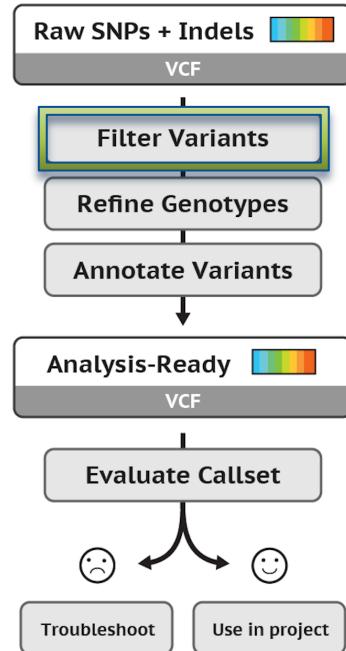
NIAID

Generate new variant quality score using VQSR



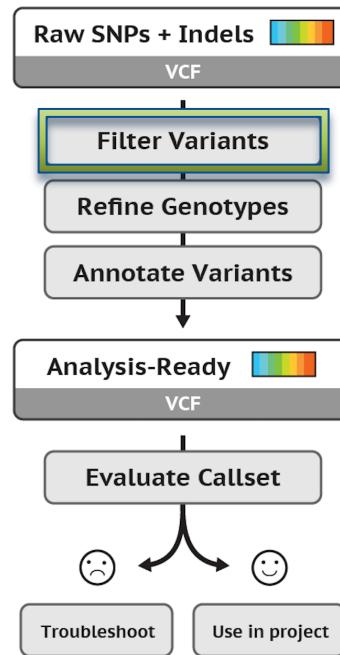
- What is Variant Quality Score Recalibration
 - NOT adjusting scores!!
 - Generate new score VQSLOD (variant quality score log-odds)
- Approach
 - Machine learning to profile good variants vs bad variants
 - Using multiple dimensions (5-8, typically)
 - Uses INFO annotations for each variant (eg. Allele count, allele frequency, etc)

Generate new variant quality score using VQSR



National Institute of
Allergy and
Infectious Diseases

Alternatively! If you have few samples (< 30 exomes)



- Apply hard filter!
- Define how to filter your variant or use default filter parameters
 - QualByDepth (QD) 2.0
 - FisherStrand (FS) 60.0
 - RMSMappingQuality (MQ) 40.0
 - MappingQualityRankSumTest (MQRankSum) -12.5 (only het calls)
 - ReadPosRankSumTest (ReadPosRankSum) -8.0 (only het calls)

Final VCF

Header

```
##fileformat=VCFv4.1
##FILTER=<ID=LowQual,Description="Low quality">
##FORMAT=<ID=AD,Number=.,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth (reads with MQ=255 or with bad mates are filtered)">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=PL,Number=6,Type=Integer,Description="Normalized, Phred-scaled likelihoods for genotypes as defined in the VCF specification">
##GATKCommandLine.HaplotypeCaller=<ID=HaplotypeCaller,Version=3.4-3-gd1ac142,Date="Mon May 18 17:36:4
.
.
.

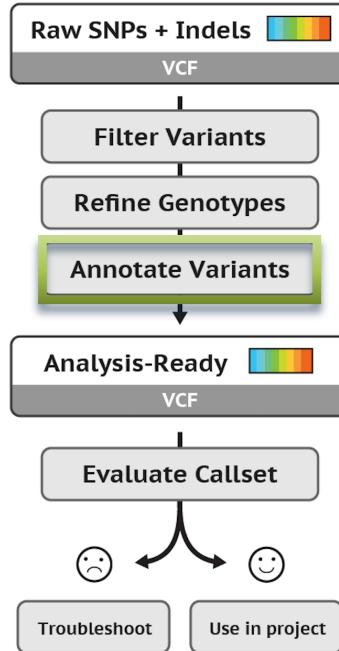
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes, for each ALT allele, in the same order as listed">
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##contig=<ID=chr1,length=249250621,assembly=b37>
##reference=file:human_genome_b37.fasta
```

Body

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	NA12878
1	873762	.	T	G	5231.78	PASS	[ANNOTATIONS]	GT:AD:DP:GQ:PL	0/1:173,141:282:99:255,0,255
1	877664	rs3828047	A	G	3931.66	PASS	[ANNOTATIONS]	GT:AD:DP:GQ:PL	1/1:0,105:94:99:255,255,0
1	899282	rs28548431	C	T	71.77	PASS	[ANNOTATIONS]	GT:AD:DP:GQ:PL	0/1:1,3:4:26:103,0,26
1	974165	rs9442391	T	C	29.84	LowQual	[ANNOTATIONS]	GT:AD:DP:GQ:PL	0/1:14,4:14:61:61,0,255



Annotating Variants with VEP



[VEP](#) determines the effect of your variants (SNPs, insertions, deletions, CNVs or structural variants) on genes, transcripts, and protein sequence, as well as regulatory regions.

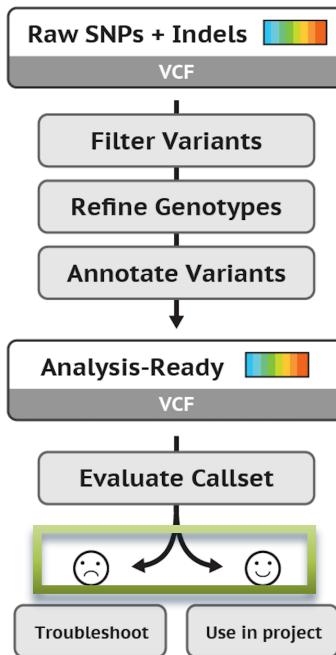
Paper: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4893825/>

The specific annotations retrieved are:

- Genes and transcripts affected by the variants
- Location of the variants:
 - upstream of a transcript
 - in coding sequences
 - in non-coding RNAs
 - in regulatory regions
- Impact of variants on the protein sequence (e.g. stop gained, missense, stop lost, frameshift)
- Known variants that match the variant, and associated minor allele frequencies from the 1000 Genomes Project
- [SIFT](#) and [PolyPhen](#) scores for changes to protein sequence

Family Based Study

Prioritize candidate variants with GEMINI



GEMINI (GEnome MINIng), a flexible software package for exploring all forms of human genetic variation.

It integrates genetic variation using an adaptable set of genome annotation sources (e.g., dbSNP, ENCODE, UCSC, ClinVar, KEGG)

Researchers can compose complex queries based on sample genotypes, inheritance patterns, and both pre-installed and custom genome annotations.

Both command line and graphical tools for common analyses are supported

Paper:

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3715403/>

Similar tool to consider: pVAAST <http://www.hufflab.org/software/pvaast/>



National Institute of
Allergy and
Infectious Diseases

NIH



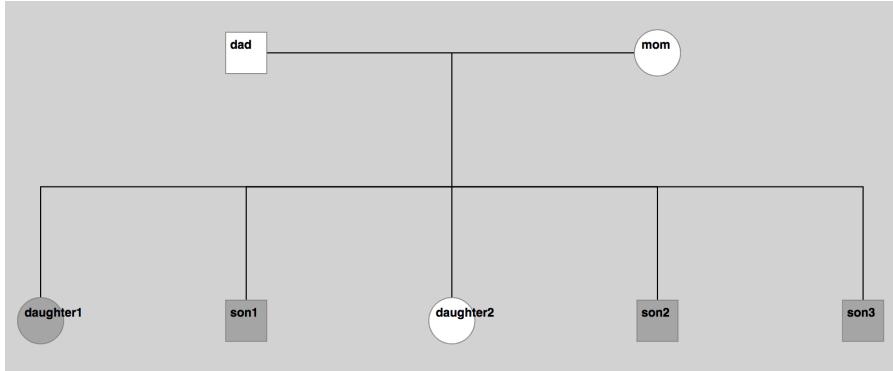
Hands-on exercises

Please visit this URL for walkthrough:

https://github.com/niaid/ACE/blob/master/WES_Data_Analysis/wes_data_analysis.md

Hands on settings

- We have FASTQ files from a family with 7 members



- We'll only be looking at subset regions on chr6, 11, X, and Y
- Are there any deleterious variants in this family?***



Steps to get started on LOCUS...

1. Log into the Locus Cluster: `ssh username@ai-submit1.niaid.nih.gov`

-Note: if you want to use an X-windows application see the walkthrough about adding the `-X` flag to your ssh command and using Xquartz or Xceed!

2. Start an Interactive Session: `qrsh -pe threaded 8`

3. Make a Tutorial Folder: `mkdir WES_Tutorial`

4. Move into Folder: `cd WES_Tutorial`

5. Copy Example Files:

```
cp -r /hpcdata/bcbb/wes_training/data/* .
```

6. Verify Files Copied: `ls -l`

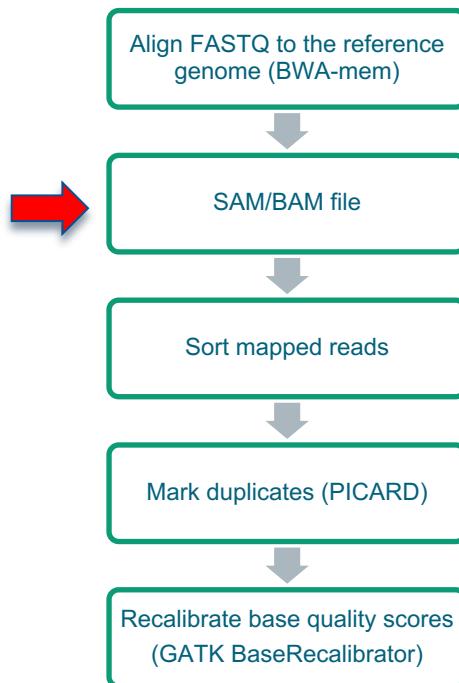
-You should see files like: `dad, daughter1, daughter2, etc.`

Modules we will use...

(*load only as needed in walkthrough!*)

- gatk/3.8.1-Java-1.8.0_92
- gatk/4.0-8-Java-1.8.0_92
- bwa
- samtools
- picard
- FastQC
- IGV
- verifyBamID
- peddy
- VEP/89-goolf-1.7.20
- GEMINI

Step 1: align fastq files



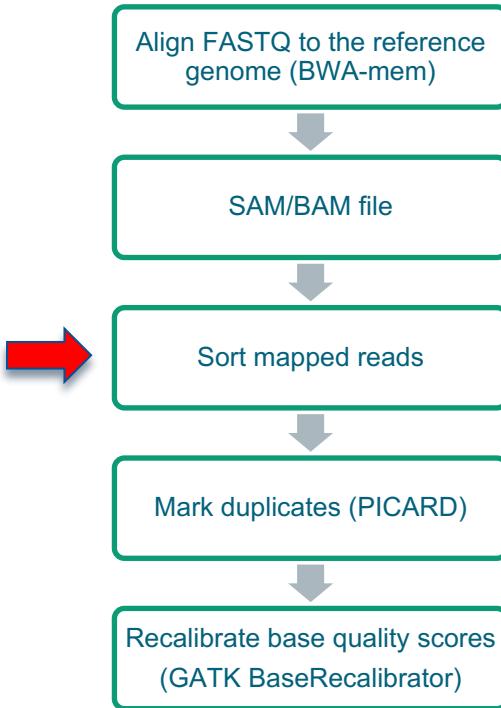
- Align with BWA MEM using **-M** to mark secondary alignments and **-R** to annotate Read Groups (e.g., different samples)
 - **ID, LB, SM, and PL tags are required**

```
module load bwa  
sample='son1'
```

```
bwa mem -t 8 -M -R  
"@RG\tID:${sample}\tLB:${sample}\tSM:${sample}\tPL:ILLUMINA" \  
/hpcdata/bcbb/wes_training/reference/human_g1k_v37_decoy.fasta \  
\ ${sample}/${sample}.R1.fq ./${sample}/${sample}.R2.fq \  
-o ${sample}/${sample}.sam
```

```
ls son1
```

Step 2: sort sam, convert to bam



- Aligned reads need to be sorted

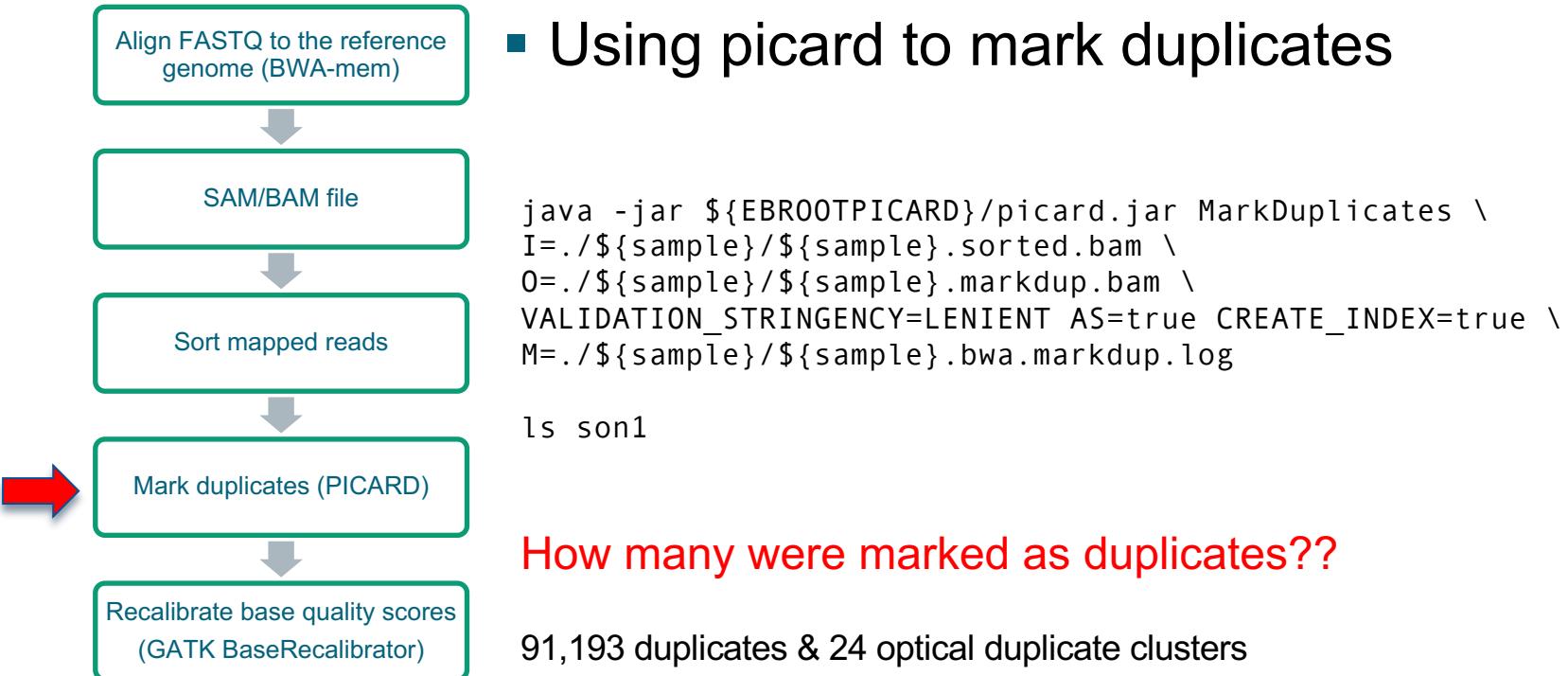
module load picard

```
java -jar ${EBROOTPICARD}/picard.jar SortSam \
I=./${sample}/${sample}.sam \
O=./${sample}/${sample}.sorted.bam \
MAX_RECORDS_IN_RAM=1000000 CREATE_INDEX=true \
VALIDATION_STRINGENCY=LENIENT \
SORT_ORDER=coordinate
```

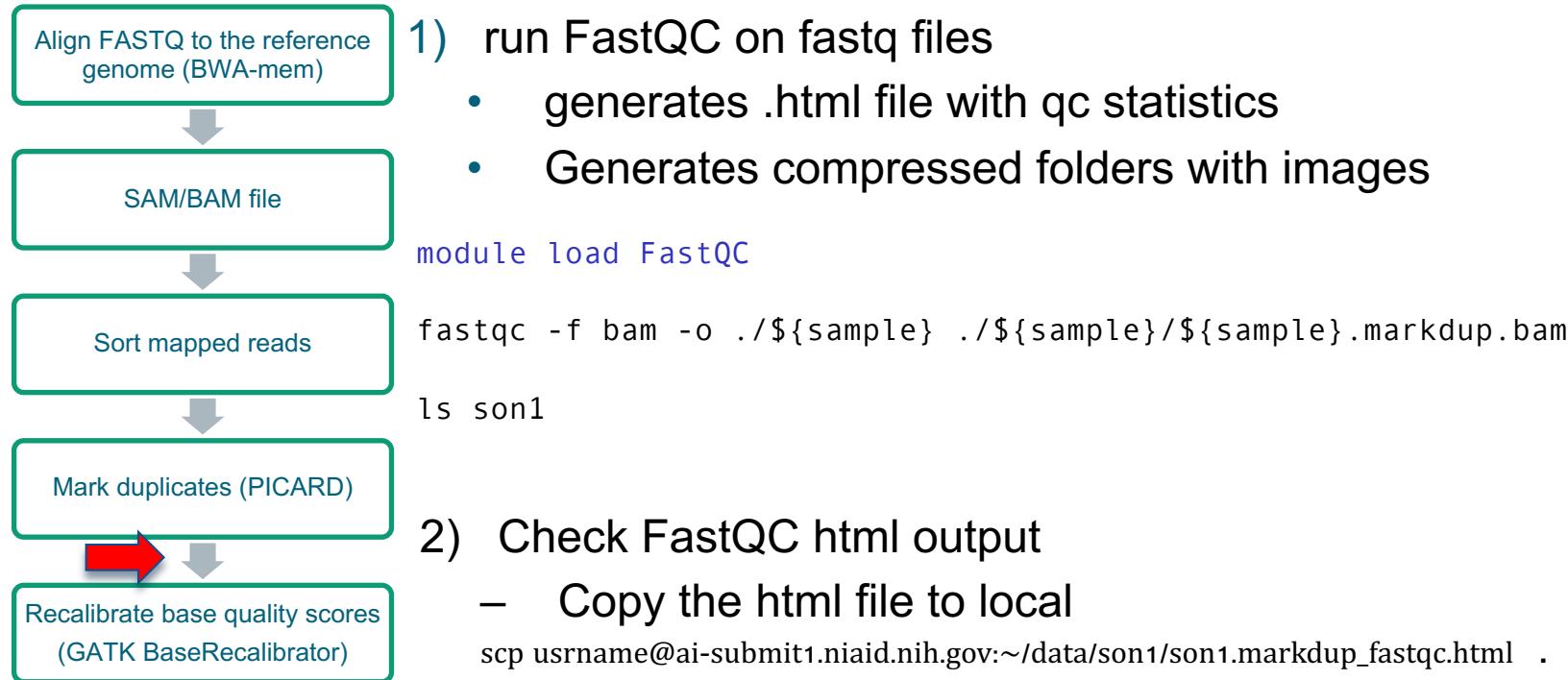
```
ls son1
```

Now we have ***mapped and sorted*** reads

Step 3: Mark duplicates



Step 4: QC - FastQC



Step 5: QC - DepthOfCoverage

NOTE: GenomeAnalysisTK is currently only in GATK3 – Broad suggests it will be released for GATK4 later this year!

Align FASTQ to the reference genome (BWA-mem)



SAM/BAM file



Sort mapped reads



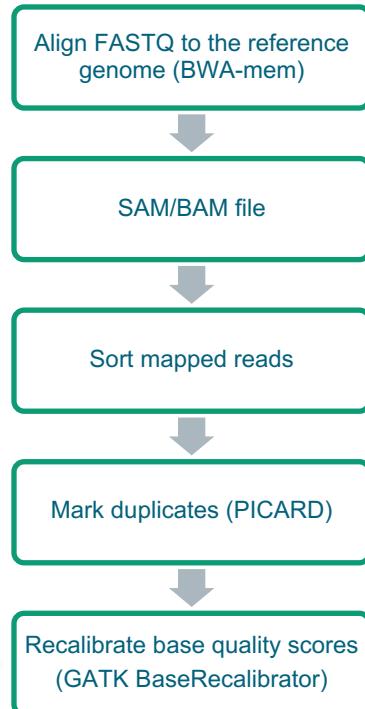
Mark duplicates (PICARD)



Recalibrate base quality scores (GATK BaseRecalibrator)

```
module load gatk/3.8.1-Java-1.8.0_92  
find . -name "*markdup.bam" > fam001_bams.list  
cat fam001_bams.list  
  
java -jar $EBROOTGATK/GenomeAnalysisTK.jar -T DepthOfCoverage \  
-R /hpcdata/bcbb/wes_training/reference/human_g1k_v37_decoy.fasta \  
-o ./fam001_dov \  
-I fam001_bams.list \  
-geneList:REFSEQ /hpcdata/bcbb/wes_training/reference/refSeq.sorted.txt \  
-ct 4 -ct 10 -ct 15 -ct 20 \  
-L /hpcdata/bcbb/wes_training/reference/exome.interval_list  
  
module unload gatk/3.8.1-Java-1.8.0_92
```

Step 6: Recalibrate base QS



Will need references data sets in [GATK resource bundles](#)

- Build a model

```
module load gatk/4.0-8-Java-1.8.0_92
```

```
#Part1
```

```
 ${EBROOTGATK}/gatk BaseRecalibrator \
-R /hpcdata/bcbb/wes_training/reference/human_g1k_v37_decoy.fasta \
-I ./${sample}/${sample}.markdup.bam \
-O ./${sample}/${sample}.recal_data.table \
--known-sites /hpcdata/bcbb/wes_training/reference/dbsnp_138.b37.vcf.gz \
--known-sites/hpcdata/bcbb/wes_training/reference/Mills_and_1000G_gold_standard.indels.b37.vcf.gz
```

- Recalibrate scores

```
#Part2
```

```
 ${EBROOTGATK}/gatk ApplyBQSR \
-R /hpcdata/bcbb/wes_training/reference/human_g1k_v37_decoy.fasta \
-I ./${sample}/${sample}.markdup.bam \
-bqsr ./${sample}/${sample}.recal_data.table \
-O ./${sample}/${sample}.bqsr.bam
```



QC Bonus: Sample contamination estimates!

```
module load verifyBamID

verifyBamID --ignoreRG --noPhoneHome \
--vcf /hpcdata/bcbb/wes_training/reference/1kg.sites.coding.ExAC.vcf.gz \
--bam ${sample}/${sample}.bqsr.bam \
--out ${sample}/${sample}.selfSM \
--maxDepth 1000 -precise

cat ./son1/son1.selfSM.selfSM

module unload verifyBamID
```

When genotype data is not available but allele-frequency-based estimates of [FREEMIX] ≥ 0.03 and [FREELK1]-[FREELK0] is large, then it is possible that the sample is contaminated with other samples!

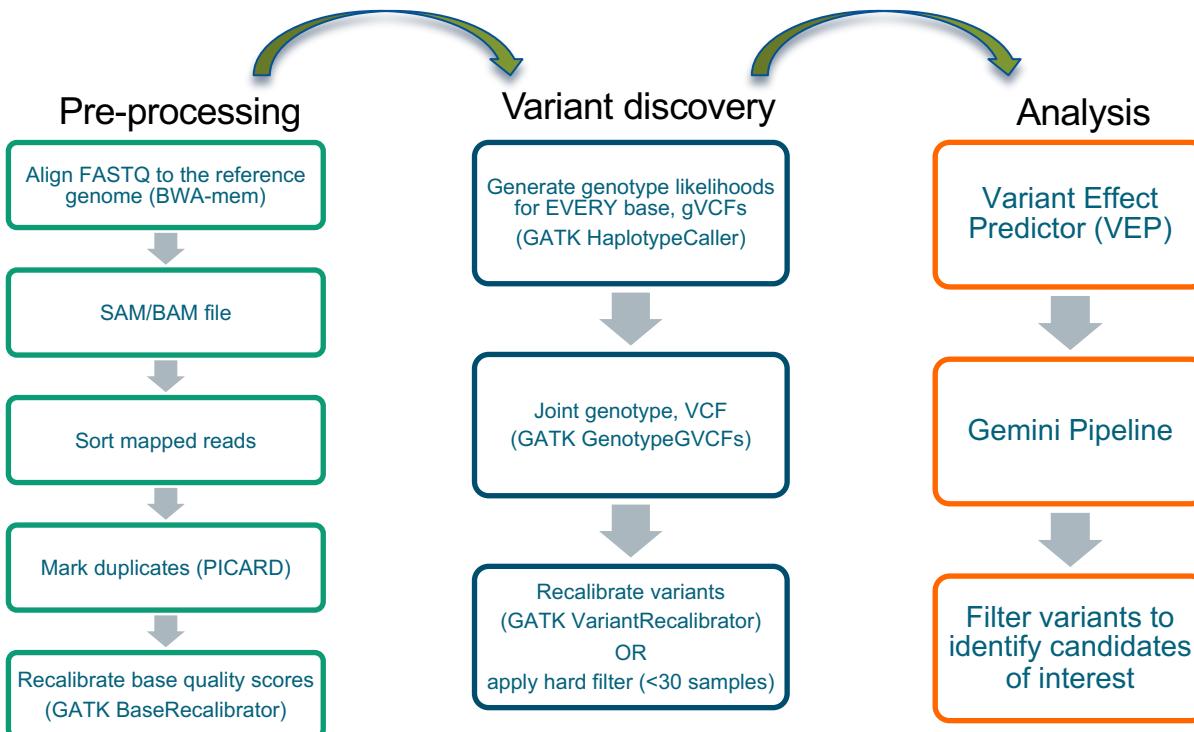
Lunch Break!



National Institute of
Allergy and
Infectious Diseases

NIAID

Data processing



National Institute of
Allergy and
Infectious Diseases

NIAID

Step 1: Generate gVCF



Generate *genotype likelihoods*
for EVERY base, gVCFs
(GATK HaplotypeCaller)



Joint genotype, raw SNP &
indel VCF
(GATK GenotypeGVCFs)



Recalibrate variants
(GATK VariantRecalibrator)
OR
apply hard filter (<30 samples)

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	son1
6	82204749	.	A	<NON_REF>	.	.	END=82204869	GT:DP:GQ:MIN_DP:PL	0/0:116:99:79:0,120,1800
6	82459379	.	A	<NON_REF>	.	.	END=82460199	GT:DP:GQ:MIN_DP:PL	0/0:75:99:42:0,108,1620
6	82460200	.	A	<NON_REF>	.	.	END=82460205	GT:DP:GQ:MIN_DP:PL	0/0:41:90:39:0,90,1350
6	82460206	.	A	<NON_REF>	.	.	END=82460219	GT:DP:GQ:MIN_DP:PL	0/0:34:84:31:0,84,1260
6	82460790	.	C	<NON_REF>	.	.	END=82460910	GT:DP:GQ:MIN_DP:PL	0/0:62:99:40:0,102,1482
6	82461270	.	C	<NON_REF>	.	.	END=82461272	GT:DP:GQ:MIN_DP:PL	0/0:33:96:33:0,96,1440
6	82461273	.	G	<NON_REF>	.	.	END=82461458	GT:DP:GQ:MIN_DP:PL	0/0:52:99:33:0,99,1211
6	82461459	.	C	<NON_REF>	.	.	END=82461463	GT:DP:GQ:MIN_DP:PL	0/0:37:93:36:0,93,1395
6	82461464	.	T	<NON_REF>	.	.	END=82461464	GT:DP:GQ:MIN_DP:PL	0/0:37:99:37:0,99,1485

```
$ ${EBROOTGATK}/gatk HaplotypeCaller \
-R /hpcdata/bcbb/wes_training/reference/human_g1k_v37_decoy.fasta \
-I ./${sample}/${sample}.bqsr.bam \
-O ./${sample}/${sample}.g.vcf.gz \
-stand-call-conf 20.0 \
--dbsnp /hpcdata/bcbb/wes_training/reference/dbsnp_138.b37.vcf.gz \
-L /hpcdata/bcbb/wes_training/reference/exome.interval_list \
-ERC GVCF
```



National Institute of
Allergy and
Infectious Diseases

Broad Institute

NIAID

Step 2: Make Joint Calling VCF

Generate *genotype likelihoods*
for EVERY base, gVCFs
(GATK HaplotypeCaller)

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	dad	daughter1	daughter2	mom
			son1	son2	son3							
6	82930437	.	G	GA	4774.86	.	AC=7;AF=0.500;AN=14;BaseQRankSum=-2.850e-01;DP=582;ExcessHet=17.2937;FS=0.534;MLEAC=7;MLEAF=0.500;MQ=59.97;MQRankSum=0.00;QD=13.08;ReadPosRankSum=-8.880e-01;SOR=0.651	GT				
			:AD:DP:GQ:PL	0/1:12,25:40:99:730,0,172		0/1:28,14:42:99:279,0,583		0/1:8,39:51:20:1145,0,20		0/1:44,20:68:99:468,0,895		0/1:24,34:66:99:824,0,457
6	82933309	.	C	T	7796.17	.	AC=9;AF=0.643;AN=14;BaseQRankSum=-5.360e-01;DP=386;ExcessHet=7.7512;FS=28.470;MLEAC=9;MLEAF=0.643;MQ=60.00;MQRankSum=0.00;QD=20.20;ReadPosRankSum=0.836;SOR=0.177	GT				
			:AD:DP:GQ:PL	1/1:0,39:39:99:1316,117,0		0/1:21,15:36:99:426,0,601		1/1:0,46:46:99:1621,138,0		0/1:39,43:82:99:1325,0,1159		0/1:24,32:56:99:950,0,649
6	83075914	.	G	A	12212.17	.	AC=9;AF=0.643;AN=14;BaseQRankSum=1.38;DP=630;	GT				

Joint genotype, raw SNP &
indel VCF
(GATK GenotypeGVCFs)

```
#Set up our samples for processing
sample_list=("dad" "mom" "daughter1" "son1" "daughter2" "son2" "son3")
gvcf_list=""
for i in ${sample_list[@]}; do gvcf_list=${gvcf_list}" ${gvcf_list}" -V ./${i}/.${i}.g.vcf.gz"; done
echo "$gvcf_list"
```

Recalibrate variants
(GATK VariantRecalibrator)
OR
apply hard filter (<30 samples)

```
#${EBROOTGATK}/gatk GenomicsDBImport ${gvcf_list} \
--genomicsdb-workspace-path demo_database \
-L /hpcdata/bcbb/wes_training/reference/exome.interval_list

#Perform joint genotyping on one or more samples pre-called with HaplotypeCaller
#${EBROOTGATK}/gatk GenotypeGVCFs -R /hpcdata/bcbb/wes_training/reference/human_g1k_v37_decoy.fasta \
-V gendb://demo_database -O fam001.joint.vcf.gz
```

```
#Sort the VCF
#${EBROOTGATK}/gatk SortVcf -I fam001.joint.vcf.gz \
-O fam001.joint.sorted.vcf.gz
```



National Institute of
Allergy and
Infectious Diseases

QC Bonus: Family and Gender Check!

- Check that the PED file looks normal...

```
cat fam001.ped
# fam001 dad 0 0 1 1
# fam001 mom 0 0 2 1
# fam001 daughter1 dad mom 2 2
# fam001 daughter2 dad mom 2 1
# fam001 son1 dad mom 1 2
# fam001 son2 dad mom 1 2
# fam001 son3 dad mom 1 2
```

- Run PEDDY to check status (<https://github.com/brentp/peddy>)

```
module load peddy
python -m peddy -p 8 --prefix fam001 --plot fam001.joint.vcf.gz fam001.ped
```

PEDDY will print any errors to STDOUT (e.g. 'Individual 123 is marked as dad but has female sex' etc.)

Step 3: Recalibrate variants with VQSR or Deep Learning

Generate genotype likelihoods for EVERY base, gVCFs
(GATK HaplotypeCaller)

Joint genotype, raw SNP & indel VCF
(GATK GenotypeGVCFs)

Recalibrate variants
(GATK VariantRecalibrator)
OR
apply hard filter (<30 samples)

At this stage we need a way to evaluate the quality of our variant calls. However, how we do this depends heavily on how many samples we have to consider. The GATK team offers a number of options...

- Working on individual (non-joint called) samples: You can use VQSR or get better results using the new “GATK4 CNN” (convolutional neural network) tool ([more details on the tool here](#))
- Working on **>= 30** joint-called samples: You can use VQSR ([tool details here](#)) to train a model on known SNPs and INDELs and apply this model to your samples to score and rank variant calls
- For **< 30** samples (*our case today!*) use HARD FILTERS – set thresholds and throw away any variants that fall above/below your cutoffs

Variant Call Performance Comparison [GATK4 CNN team]

Area Under Receiver Operating Characteristic Curve Table (**high number = better performance**)

-	INDEL AUC	INDEL AUC	INDEL AUC	SNP AUC	SNP AUC	SNP AUC	Metric
Architecture	NA12878	NA24385	CHM WGS1	NA12878	NA24385	CHM WGS1	Sample
VQSR Single Sample	.808	.557	.531	.954	.917	.808	
VQSR gnomAD	.869	.766	.571	.979	.955	.940	
Deep Variant	.874	.739	.779	.987	.985	.967	
GATK4 CNN	.949	.836	.817	.987	.979	.973	



National Institute of
Allergy and
Infectious Diseases

NIH

Step 3 Continued: Apply hard filters (<30 samples)

Generate genotype likelihoods for EVERY base, gVCFs
(GATK HaplotypeCaller)

- Extract SNPs

```
 ${EBROOTGATK}/gatk SelectVariants -V fam001.joint.sorted.vcf.gz \  
 -select-type SNP -O fam001.snps.vcf.gz
```

Joint genotype, raw SNP &
indel VCF
(GATK GenotypeGVCFs)

- Extract non-SNPs

```
 ${EBROOTGATK}/gatk SelectVariants -V fam001.joint.sorted.vcf.gz \  
 -xl-select-type SNP -O fam001.nonsnps.vcf.gz
```

Recalibrate variants
(GATK VariantRecalibrator)
OR
apply hard filter (<30 samples)



Step 3 Continued: Apply hard filters (<30 samples)

Generate genotype likelihoods for EVERY base, gVCFs
(GATK HaplotypeCaller)

Joint genotype, raw SNP & indel VCF
(GATK GenotypeGVCFs)

Recalibrate variants
(GATK VariantRecalibrator)
OR
apply hard filter (<30 samples)

- Filter SNPs

```
`${EBROOTGATK}/gatk VariantFiltration \  
-V fam001.snps.vcf.gz \  
-filter "QD < 2.0" --filter-name "QD2" \  
-filter "QUAL < 30.0" --filter-name "QUAL30" \  
-filter "SOR > 3.0" --filter-name "SOR3" \  
-filter "FS > 60.0" --filter-name "FS60" \  
-filter "MQ < 40.0" --filter-name "MQ40" \  
-filter "MQRankSum < -12.5" --filter-name "MQRankSum12.5" \  
-filter "ReadPosRankSum < -8.0" --filter-name "ReadPosRankSum8" \  
-O fam001.snps_filtered.vcf.gz
```

- Filter non-SNPs

```
`${EBROOTGATK}/gatk VariantFiltration \  
-V fam001.nonsnps.vcf.gz \  
-filter "QD < 2.0" --filter-name "QD2" \  
-filter "QUAL < 30.0" --filter-name "QUAL30" \  
-filter "FS > 200.0" --filter-name "FS200" \  
-filter "ReadPosRankSum < -20.0" --filter-name "ReadPosRankSum20" \  
-O fam001.nonsnps_filtered.vcf.gz
```



Step 3 Continued: Apply hard filters (<30 samples)

Generate genotype likelihoods for EVERY base, gVCFs
(GATK HaplotypeCaller)

- Now combine filtered variants into a single VCF ...

```
$ {EBR00TGATK}/gatk MergeVcfs \
-I fam001.snps_filtered.vcf.gz \
-I fam001.nonsnps_filtered.vcf.gz \
-O fam001.combined_filtered.vcf.gz
```

Joint genotype, raw SNP & indel VCF
(GATK GenotypeGVCFs)

Example of filtered output

Y 15027529 . T G 9330.14 SOR3 AC=8;AF=1.00;AN=8;DP=268;ExcessHet=3.0103;FS=0.000;MLEAC=8;MLEAF=1.00;MQ=59.98;QD=34.81;SOR=3.158 ./.:0,0:0:.:0,0,0 ./:0,0:0:.:0,0,0 1/1:0,65:65:99:2184,195,0 1/1:0,74:74:99:2758,223,0 1/1:0,67:67:99:2314,202,0
Y 15467824 . A G 2203.14 PASS AC=8;AF=1.00;AN=8;DP=64;ExcessHet=3.0103;FS=0.000;MLEAC=8;MLEAF=1.00;MQ=60.00;QD=34.42;SOR=1.721 ./:0,0:0:.:0,0,0 ./:0,0:0:.:0,0,0 1/1:0,10:10:30:319,30,0 ./:0,0:0:.:0,0,0 1/1:0,18:18:54:670,54,0 1/1:0,21:21:63:707,63,0 1/1:0,15:15:45:532,45,0
Y 15591537 . G C 1237.16 SOR3 AC=8;AF=1.00;AN=8;DP=41;ExcessHet=3.0103;FS=0.000;MLEAC=8;MLEAF=1.00;MQ=60.00;QD=30.17;SOR=3.312 ./:0,0:0:.:0,0,0 ./:0,0:0:.:0,0,0 1/1:0,17:17:51:536,51,0 1/1:0,6:6:18:179,18,0 1/1:0,8:8:24:239,24,0

Recalibrate variants
(GATK VariantRecalibrator)
OR
apply hard filter (<30 samples)

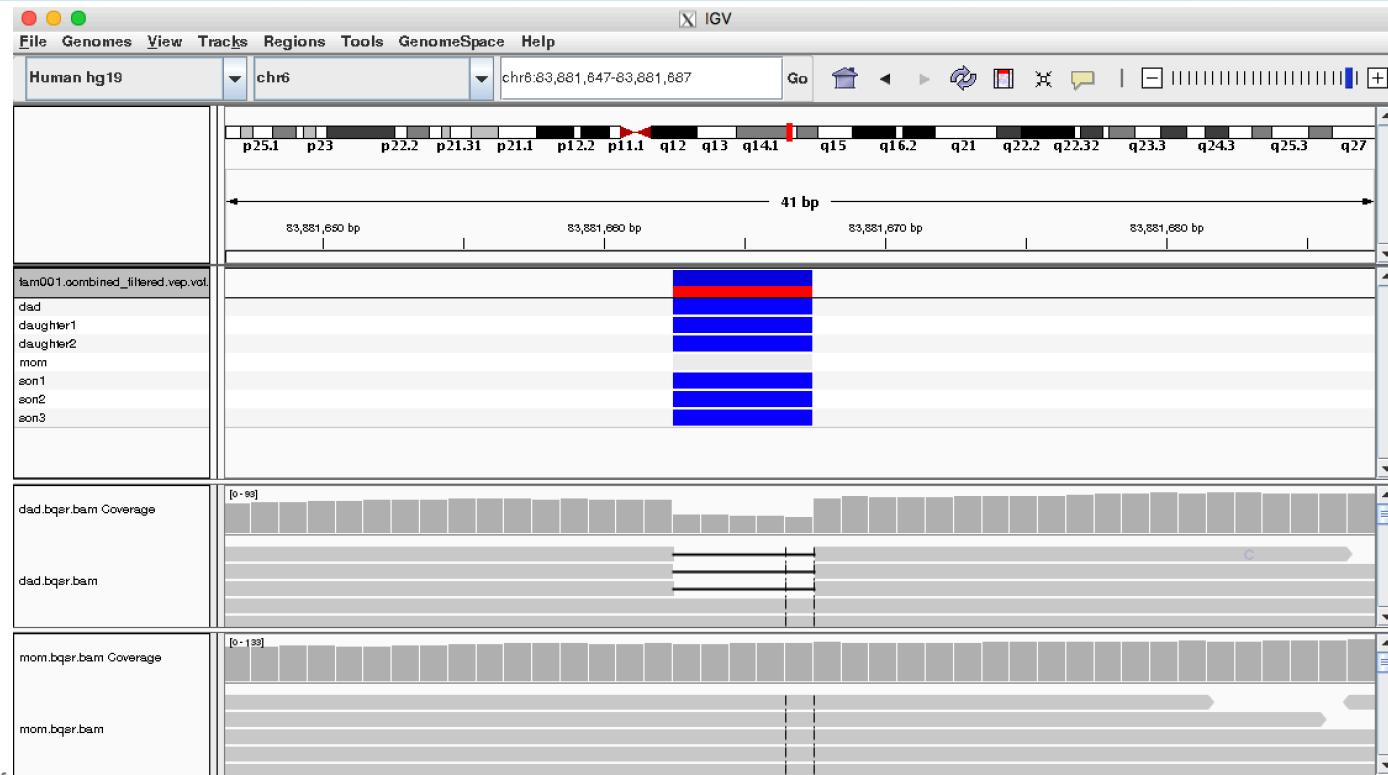


National Institute of
Allergy and
Infectious Diseases

Broad Institute

NIAID

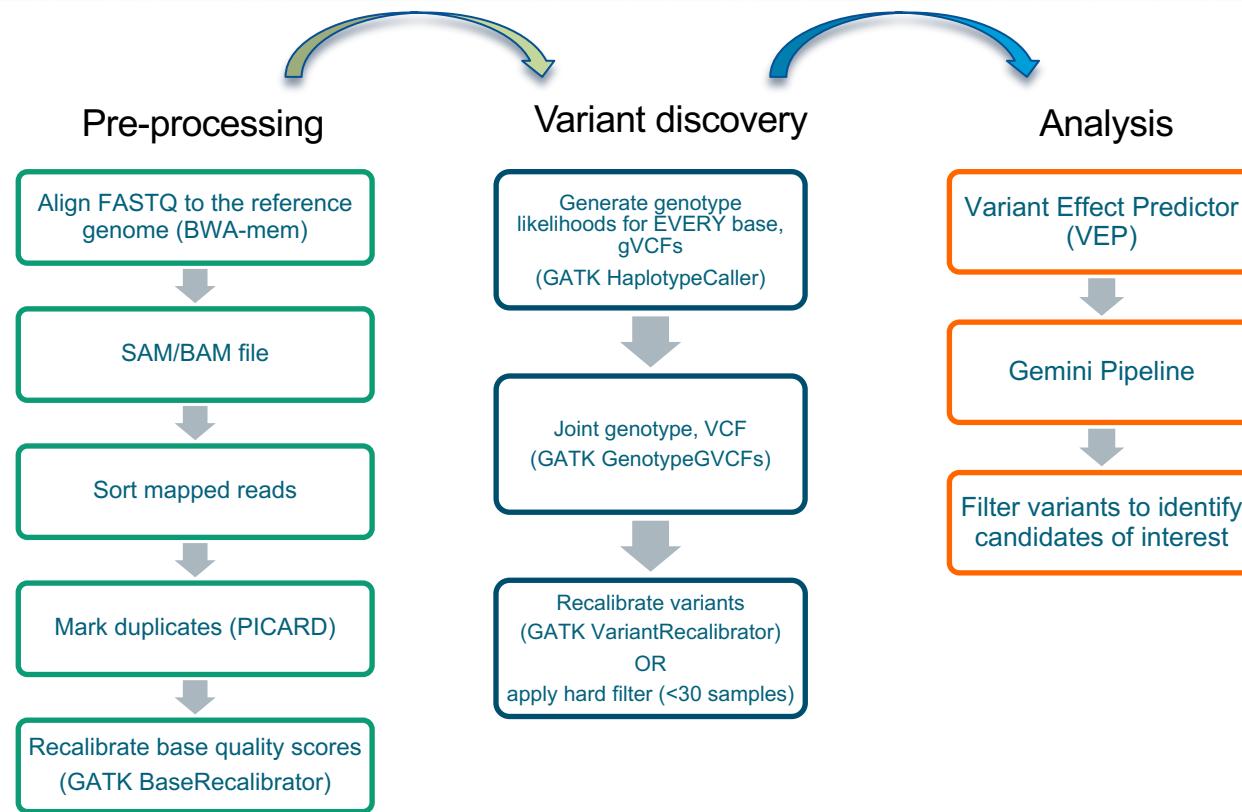
Check Output in IGV



National Institute of
Allergy and
Infectious Diseases

NIH

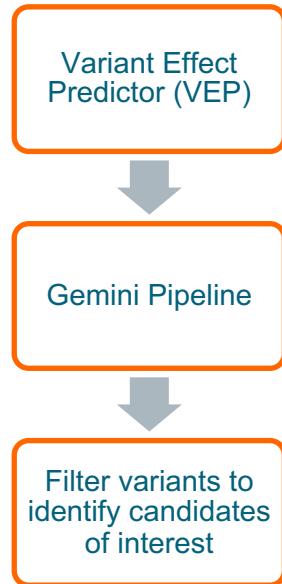
Data processing



National Institute of
Allergy and
Infectious Diseases

NIH

VEP



```
module load bcftools  
  
bcftools norm -m -any --threads 8 -Oz \  
-o fam001.combined_filtered.norm.vcf.gz fam001.combined_filtered.vcf.gz  
  
tabix -p vcf fam001.combined_filtered.norm.vcf.gz  
  
module load VEP/89-goolf-1.7.20  
  
dbNSFP_dir="/sysapps/cluster/software/VEP/84-goolf-1.7.20/.vep/dbNSFP/2.9.3"  
  
${EBROOTVEP}/vep --fork 8 --dir ${EBROOTVEP}/.vep --assembly GRCh37 --merged \  
--everything --vcf --allele_number --no_stats --cache --offline --force_overwrite \  
--compress_output bgzip --plugin \  
dbNSFP,${dbNSFP_dir}/dbNSFP.gz,Polyphen2_HVAR_pred,CADD_phred,SIFT_pred,FATHMM_pred,MutationTas  
ter_pred,MetaSVM_score,MetaSVM_pred,MetaLR_score,MetaLR_pred,Reliability_index --plugin\  
LoF,human_ancestor_fa:${EBROOTVEP}/.vep/Plugins/loftee/human_ancestor.fa.gz,filter_position:0.05,  
min_intron_size:15 \  
-i fam001.combined_filtered.norm.vcf.gz -o fam001.combined_filtered.vep.vcf.gz  
  
tabix -p vcf fam001.combined_filtered.vep.vcf.gz # create index for the normalized VCF
```

Now we have annotated VCF file!!

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	dad	daughter1	daughter2	mom
		son1	son2	son3								
6	82930437		.	G	GA	4774.86	PASS		AC=7;AF=0.5;AN=14;BaseQRankSum=-0.285;DP=582;ExcessHet=17.2937;FS=0.534;MLEAC=7;MLEAF=0.5;MQ=59.97;MQRankSum=0;QD=13.08;ReadPosRankSum=-0.888;SOR=0.651;CSQ=A splice_region_variant&intron_variant LOW IBTK ENSG00000005700 Transcript ENST00000306270 protein_coding 8/28 ENST00000306270.7:c.1125-8dupT rs34032511 1 -1 insertion HGNC 17853 YES CCDS34490.1 ENSP00000305721 Q9P2D0 UPI000041929F Ensembl ,A splice_region_variant&intron_variant&non_coding_transcript_variant LOW IBTK ENSG00000005700 Transcript ENST00000369751 retained_intron 8/23 ENST00000369751.2:n.1675-8dupT rs34032511 1 -1 insertion HGNC 17853 Ensembl ,A splice_region_variant&intron_variant&NMD_transcript_variant LOW IBTK ENSG00000005700 Transcript ENST00000503400 nonsense-mediated_decay 8/28 ENST00000503400.1:c.*436-8dupT rs34032511 1 -1 insertion HGNC 17853 ENSP00000422136 D6R9F4 UPI0001D3BC1A Ensembl ,A splice_region_variant&intron_variant LOW IBTK ENSG00000005700 Transcript ENST00000503631 protein_coding 7/26 ENST00000503631.1:c.1125-8dupT rs34032511 1 -1 insertion HGNC 17853 ENSP00000422762 E9PDR5 UPI0001D3BC1B Ensembl ,A splice_region_variant&intron_variant&non_coding_transcript_variant LOW IBTK ENSG00000005700 Transcript ENST0000050222 retained_intron 7/25 ENST0000050222.1:n.1204-8dupT rs34032511 1 -1 insertion HGNC 17853 Ensembl ,A splice_region_variant&intron_variant LOW IBTK ENSG00000005700 Transcript ENST00000510291 protein_coding 7/27 ENST00000510291.1:c.1125-8dupT rs34032511 1 -1 insertion HGNC 17853 ENSP00000426405 E7EPI0 UPI0001D3BBE0 Ensembl ,A splice_region_variant&intron_variant LOW IBTK 25998 Transcript NM_015525.2 protein_coding 8/28 NM_015525.2:c.1125-8dupT rs34032511 1 -1 insertion YES NP_056340.2 RefSeq GT:AD:DP:GQ:PL 0/1:12,25:40:99:730,0,172 0/1:28,14:42:99:279,0,583 0/1:8,39:51:20:1145,0,20 0/1:44,20:68:99:468 ,0,895 0/1:41,38:88:99:869,0,846 0/1:13,25:41:99:508,0,246 0/1:24,34:66:99:824,0,457			



Now what?

What do you do with all these variants?

Variant Analysis... like finding a needle in a 'deep' haystack



National Institute of
Allergy and
Infectious Diseases

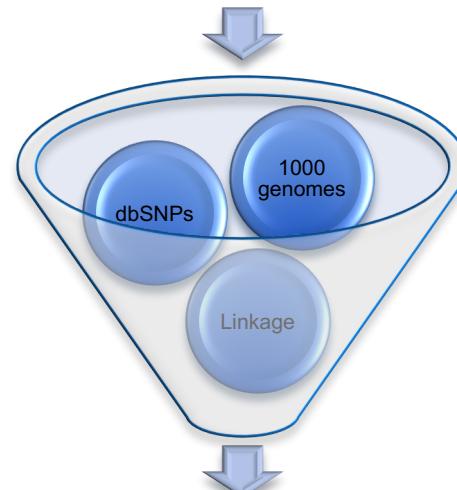
Look for evidence of variants of interest



Further filters needed

- High number of variants
- The goal is to narrow down your list of variants
- Eliminate variants that are not interesting

17,687 coding SNPs



2 novel in chr10

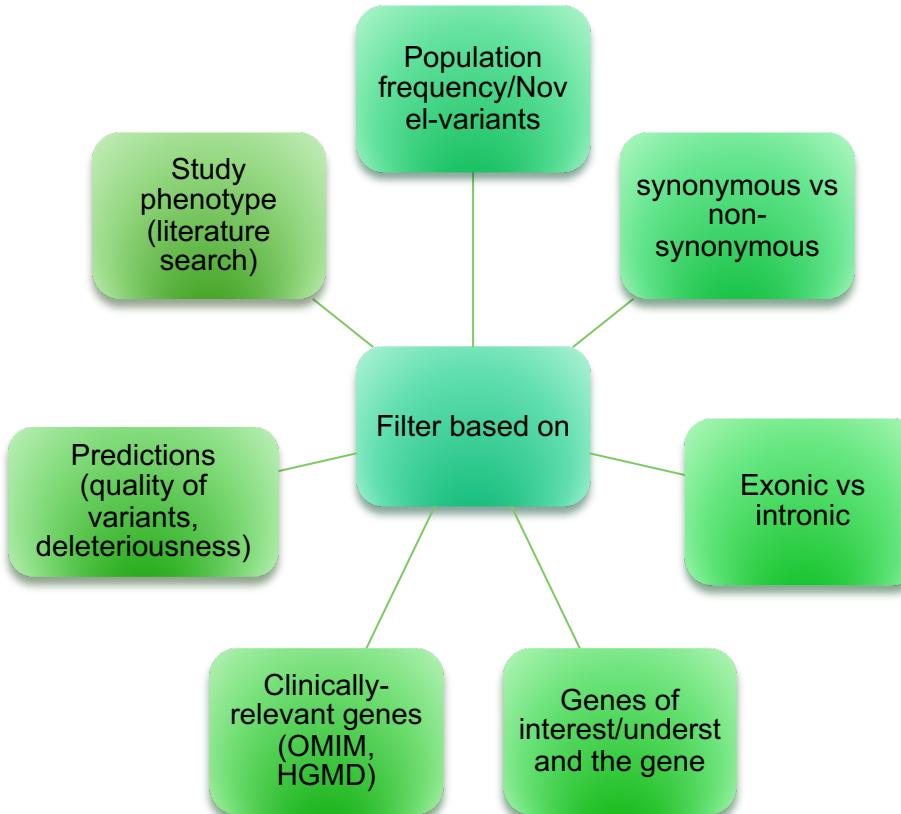
PLoS One. 2012;7(1):e29708



National Institute of
Allergy and
Infectious Diseases

NIAID

Things to consider



GEMINI

- GEMINI can be used to consider inheritance models to find candidate variants to explore further!
- Step1 - Create a reference DB:

```
module load GEMINI
gemini load -v fam001.combined_filtered.vep.vcf.gz \
-t VEP -p fam001.ped --cores 8 --tempdir /hpcdata/scratch/ fam001.db
```

- Step2 - Try different inheritance models:

```
gemini <INSERT_MODEL> --columns \
"gene, chrom, start, end, ref, alt, impact_severity, vep_cadd_phred, impact, in_1kg" fam001.db
```

Where <INSERT_MODEL> above can be “**autosomal_dominant**”, “**autosomal_recessive**”, “**comp_hets**”, “**de_novo**”, “**x_linked_de_novo**”, etc. ([type “`gemini -h`” on the command line for the full list of options](#))

Did any of your results show any promising candidates???

GEMINI Output

```
gemini autosomal_recessive --columns "gene, chrom, start, end, ref, alt, impact_severity,vep_cadd_phred, impact, in_1kg" fam001.db
```

OUTPUT:

```
gene chrom start end ref alt impact_severity vep_cadd_phred impact in_1kg variant_id family_id family_members
family_genotypes samples family_count
OR5AN1 chr11 59132797 59132798 G C MED 24.2 missense_variant 1 27 fam001
dad(dad;unaffected;male),daughter1(daughter1;affected;female),daughter2(daughter2;unaffected;female),mom(mom;unaffected;fe
male),son1(son1;affected;male),son2(son2;affected;male),son3(son3;affected;male) G/C,C/C,G/C,G/C,C/C,C/C,C/C
daughter1,son1,son2,son3 1
OR5A1 chr11 59211187 59211188 G A MED 27.2 missense_variant 1 33 fam001
dad(dad;unaffected;male),daughter1(daughter1;affected;female),daughter2(daughter2;unaffected;female),mom(mom;unaffected;fe
male),son1(son1;affected;male),son2(son2;affected;male),son3(son3;affected;male) G/A,A/A,G/A,G/A,A/A,A/A,A/A
daughter1,son1,son2,son3 1
OR5A1 chr11 59211264 59211265 T C LOW synonymous_variant 1 34 fam001
dad(dad;unaffected;male),daughter1(daughter1;affected;female),daughter2(daughter2;unaffected;female),mom(mom;unaffected;fe
male),son1(son1;affected;male),son2(son2;affected;male),son3(son3;affected;male) T/C,C/C,T/C,T/C,C/C,C/C,C/C
daughter1,son1,son2,son3 1
```



National Institute of
Allergy and
Infectious Diseases

NIH

GEMINI Output

```
gemini comp_hets --columns "gene, chrom, start, end, ref, alt, impact_severity,vep_cadd_phred, impact, in_1kg" fam001.db
```

OUTPUT:

```
gene chrom start end ref alt impact_severity vep_cadd_phred impact in_1kg variant_id family_id family_members
family_genotypes samples family_count comp_het_id priority
PGM3 chr6 83880061 83880062 C G MED 28.4 missense_variant 0 13 fam001
dad(dad;unaffected;male),daughter1(daughter1;affected;female),daughter2(daughter2;unaffected;female),mom(mom;unaffected;
female),son1(son1;affected;male),son2(son2;affected;male),son3(son3;affected;male) C/C,G|C,C/C,C/G,G|C,G|C,G|C
daughter1,son1,son2,son3 1 1_13_15 1
PGM3 chr6 83881661 83881667 TTTAAG T HIGH frameshift_variant 0 15 fam001
dad(dad;unaffected;male),daughter1(daughter1;affected;female),daughter2(daughter2;unaffected;female),mom(mom;unaffected;
female),son1(son1;affected;male),son2(son2;affected;male),son3(son3;affected;male)
TTTAAG/T,TTTAAG|T,TTTAAG|T,TTTAAG/TTTAAG,TTTAAG|T,TTTAAG|T,TTTAAG|T daughter1,son1,son2,son3 1 1_13_15 1
```

GEMINI

- Step3 - Get basic stats for your DB:

```
gemini stats --tstv fam001.db
```

```
gemini stats --tstv-coding fam001.db
```

```
gemini stats --tstv-noncoding fam001.db
```

- Step4 – Get number of variants for each sample:

```
gemini stat -vars-by-sample fam001.db
```



National Institute of
Allergy and
Infectious Diseases

NIAID

Follow-up

- A literature search *IS A MUST!* It is time consuming, but required to interpret for prioritization and further follow up of variants.
 - Valuable resources for exploring variants further:
 - gnomAD Browser: <https://gnomad.broadinstitute.org>
 - Human Gene Mutation Database (*NIH employees get a free subscription through the library!*):
<http://www.hgmd.cf.ac.uk/ac/index.php>
 - dbSNP: <https://www.ncbi.nlm.nih.gov/snp/>



National Institute of
Allergy and
Infectious Diseases

NIAID

Additional Analysis

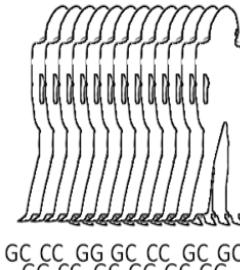
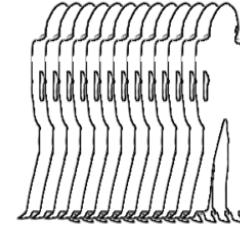
- Association analysis
 - Single variant based case-control study (common variants)
 - Group-wise burden test (rare variants)
- Copy number variation analysis
- Somatic Variation
 - MuTect2
 - VarDict

Population-based Cohort Study

GWAS Association studies

■ A typical analysis:

- Identify SNPs where one allele is significantly more common in cases than controls
- Hardy-Weinberg equilibrium (HWE) testing
- Population stratification
 - Genomic Control λ (λ_{GC})
 - Principal components analysis (PCA)
 - Mixed-Model
- Multiple-testing correction
 - Bonferroni
 - Benjamini & Hochberg FDR

SNP1	SNP2	SNP ...
Cases Count of G: 2104 of 4000	Cases Count of G: 1648 of 4000	<i>Repeat for all SNPs</i>
Frequency of G: 52.6%	Frequency of G: 41.2%	
 GC CC GG GC CC GC GC GG CC GC GG GC GG	 GC CC GC GC GG CC CC CC GC GC GG GC GG	
Controls Count of G: 2676 of 6000	Controls Count of G: 2532 of 6000	
Frequency of G: 44.6%	Frequency of G: 42.2%	
P-value: $5.0 \cdot 10^{-15}$	P-value: 0.33	

http://en.wikipedia.org/wiki/Genome-wide_association_study

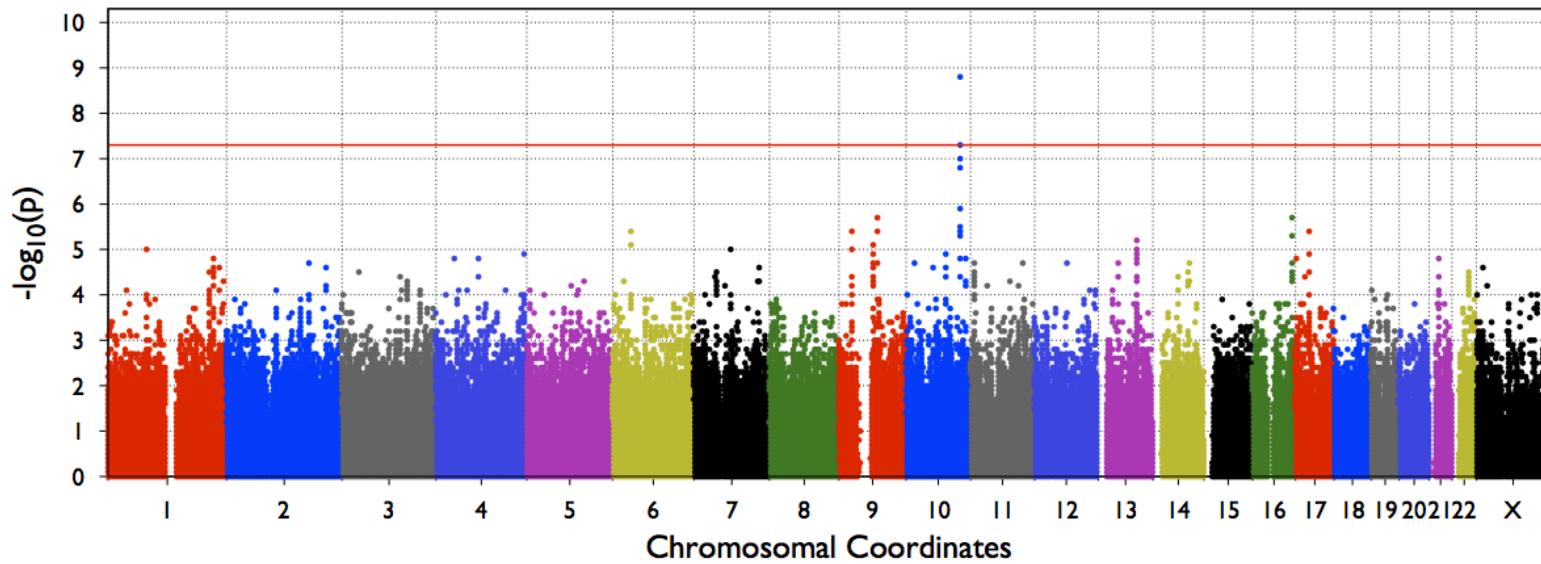
<http://www.ebi.ac.uk/gwas/>



National Institute of
Allergy and
Infectious Diseases

GWAS

GWAS Manhattan plot



- Only applicable for large cohort

Analysis Tools for genotype-phenotype association from sequencing data

- PLINK/SEQ - <https://atgu.mgh.harvard.edu/plinkseq/>
- EPACTS - <https://genome.sph.umich.edu/wiki/EPACTS>
- VAAST 2 - <http://www.yandell-lab.org/software/vaast.html>
- SNPTEST – https://mathgen.stats.ox.ac.uk/genetics_software/snptest/snptest.html

CNVs from exome

- High variability of read-depth in exomes
- CNV prediction is very challenging problem
 - High false positive rate
 - Ambiguous break-points due to targets
 - Long range CNVs have higher positive predictive value
- Useful when other alternatives (SNPs or aCGH) are not available
- Lots of prediction tools!! (some examples below)

Population Caller	Somatic Caller
CNVnator	ADTEx
CONTRA	CONTRA
CoNIFER	Control-FREEC
ExomeDepth	ExomeCNV
GATK	EXCAVATOR
XHMM	VarScan2



National Institute of
Allergy and
Infectious Diseases

NIAID

Evaluation of structural variation detection algorithms for WGS

NIH NIAID

Table 1 List of tools providing good SV calling results for both the simulated and NA12878 real datasets

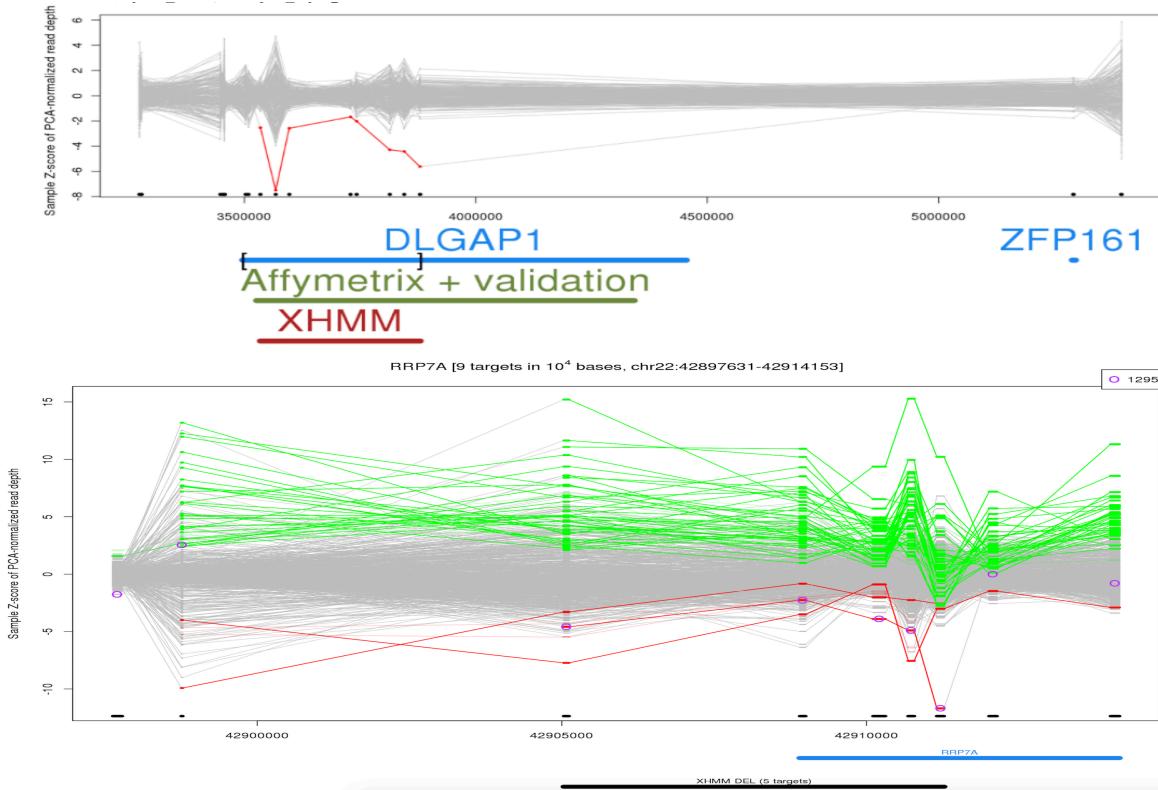
SV type	Tools	Simulated data		Real data		nF ^{*1}
		Precision	Recall	Precision	Recall	
DEL	GRIDSS	98.9 (5)	86.6 (2)	87.6 (7)	28.9 (2)	3.57 (1)
	Lumpy	99.1 (4)	81.4 (6)	87.1 (8)	26.1 (4)	3.41 (2)
	SVseq2	96.2 (11)	86.1 (3)	75.7 (17)	24.9 (5)	3.28 (3)
	SoftSV	96.8 (10)	83.6 (4)	80.2 (13)	23.2 (8)	3.25 (7)
	Manta	95.9 (12)	83.1 (5)	74.2 (20)	24.3 (6)	3.21 (5)
	MATCHCLIP	99.4 (2)	71.7 (10)	91.6 (4)	20.9 (11)	3.12 (6)
	inGAP-sv	91.1 (18)	78.6 (7)	78.3 (14)	22.5 (8)	3.10 (7)
DUP	Wham	96.9 (4)	81.7 (4)	57.1 (4)	10.2 (5)	3.92 (1)
	SoftSV	84.2 (14)	67.8 (13)	47.3 (6)	14.3 (3)	3.91 (2)
	MATCHCLIP	87.6 (11)	77.5 (8)	58.0 (3)	9.9 (6)	3.79 (3)
	GRIDSS	91.1 (9)	77.9 (7)	58.4 (2)	9.6 (7)	3.78 (4)
	Manta	99.0 (1)	83.2 (1)	40.4 (9)	6.5 (11)	3.35 (5)
INS [Unspecified]	SvABA	82.6 (15)	69.6 (11)	42.7 (8)	7.2 (9)	3.02 (6)
	pbsv	89.7 (3)	38.2 (5)	72.7 (8)	27.5 (2)	6.68 (1)
	inGAP-sv	99.7 (1)	58.5 (2)	85.5 (2)	11.8 (3)	6.27 (2)
	Sniffles	74.8 (5)	52.5 (3)	65.9 (10)	9.0 (5)	5.08 (3)
INS [MEI]	SVseq2	70.4 (8)	64.2 (1)	38.5 (19)	7.1 (9)	4.87 (4)
	MELT	99.7 (3)	68.9 (3)	88.9 (1)	85.6 ^{*2} (1)	3.21 (1)
	Mobster	100 (1)	67.1 (4)	88.3 (2)	71.9 ^{*2} (2)	3.04 (2)
INV	DELLY	94.7 (8)	81.8 (4)	38.9 (4)	15.6 (2)	3.07 (1)
	TIDDIT	89.2 (14)	77.9 (8)	49.1 (1)	11.7 (5)	2.89 (2)
	1–2–3-SV	70.7 (19)	81.2 (5)	31.8 (9)	14.8 (3)	2.67 (3)
	GRIDSS	96.6 (6)	84.7 (3)	34.2 (8)	10.4 (7)	2.67 (4)

^{*1}Sum of normalized F-measures of the simulated and the real data. Normalized F-measure = F-measure/the mean F-measure for the corresponding category

^{*2}Provisional recall value: the number of true positives was calculated by dividing by the provisional number of reference MEIs (1350), which was estimated using the data from the 1000 Genome project

Ranks of tools for each result (precision, recall, or F-measure) are indicated within parentheses

Popular utility of exome sequencing eXome Hidden Markov Model



National Institute of
Allergy and
Infectious Diseases

Suggestions

- Request for a LOCUS/Biowulf account (if you don't have one already, you will automatically receive notification when logging into the Locus Support Portal (<https://locus.niaid.nih.gov>) for the first time and will be given instructions to request an account.)
 - Programs are already installed
 - Easy to keep track of software version
 - You can request programs to be installed

NIAIDHPCSUPPORT@niaid.nih.gov

Find more information on BCBB

- bioinformatics @NIAID portal
 - <https://bioinformatics.niaid.nih.gov/>
- Inside NIAID
 - <http://inside.niaid.nih.gov/topic/IT/bioinformatics>

Questions ?



National Institute of
Allergy and
Infectious Diseases

NIAID

A large, colorful word cloud centered around the words "thank you". The word "thank" is at the top center in blue, and "you" is below it in yellow. Numerous other words in different languages are scattered around, such as "danke" (German), "merci" (French), "gracias" (Spanish), "teşekkür ederim" (Turkish), "mochchakkeram" (Korean), and many more. The words are in various colors and sizes, creating a dense and diverse visual representation of the phrase.



National Institute of
Allergy and
Infectious Diseases

Credit: <http://omogemura.com/thank-you/>

NIAID