

CaRNeSS
4.8 (20131209.62)

Generated by Doxygen 1.8.5

Tue Dec 10 2013 10:10:07

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Chapter 1

Catalytic Rections Network Stochastic Simulator - CaRNeSS 4.8 (20131209.62)

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Version

4.8 (20131209.62)

Date

2013-12-09 sourceforge repository – <https://carness.svn.sourceforge.net/svnroot/carness/>
git repository – <https://github.com/paxelito/carness>

This manual is divided in the following sections:

- [Essential informations](#)
- [Initial Data Structures](#)
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Chapter 2

Essential informations

The **Catalytic Reactions Network Stochastic Simulator (CaRNeSS)** is a computational model devoted to the simulation of theoretical complex catalytic networks composed of different interacting molecular species. The model takes inspiration from the original model proposed by Stuart Kauffman in 1986, and describes systems composed of molecular species interacting by means of two possible reactions only, cleavage and condensation. One polymer is divided into two short polymers in the former case while two polymers are glued together forming a longer polymer in the latter case. Each reaction must be catalyzed by another species in the system to occur, and one of the assumptions is that any chemical has an independent probability to catalyze a randomly chosen reaction. Since the version 4.8 (20131026.60) spontaneous reactions are considered too. It is important to notice that there are not indications about the chemical nature of the molecules, species "A" may be both a polipeptide, an amminoacid, a particular protein domain or an RNA strenght.

2.1 Using the simulator

To run the simulator open a terminal shell and type:

```
$path/executiveFile <configuration_File_Folder> <output_folder> <reaction_structures_folder>
```

Examples:

- **Unix Based Systems:** `~/Documents/project/acsm2s ~/Documents/.../confFileFolder/ ~/Documents/.../resFolder/ ~/Documents/.../StructuresFolder/`
- **Win Systems:** `C:\Documents\project\acsm2s.exe C:\Documents ..\confFileFolder\ C-:\Documents ..\resFolder\ C:\Documents ..\StructuresFolder\`

2.2 System Requirement

In order to have the simulator run correctly the recommended staff is reported:

- MacOSX 10.4 or later, Linux (or in general a system UNIX based) or Windows OS (tests have been performed on Win7 and win Vista) as well
- GCC (G++) compiler, or similar, installed (if you need to compile the software on your machine)

On MacOS system compile using `g++ -Wall -ansi -lm -o CaRNeSS *.cpp`

2.3 Input Parameters :: acsm2s.conf

All the system parameters are stored in a file called **acsm2s.conf**. Anyone can create his own configuration file paying attention to put "=" char between the parameter name and the the parameter value (NO SPACE BETWEEN

THEM).

Notice that the simulator does not create the initial structures but it simply loads the structures created by an external software and process them. Nevertheless the configuration file is fundamental to supply all the parameters to the simulation (during the simulation new entities may be created). The simulator is provided with a structures initializer developed in MATLAB language by the group (a description of the initializer is provided in the main file "start.m") in which all the parameters we are going to describe are used to create the initial structures. All parameters are reported below divided in three categories:

- System
- Environment
- Dynamic

Categories are useful only to help users in the parameter recognition within the configuration file. The following parameters are used both by the initializer and the simulator. Nevertheless it is ALWAYS neces

2.3.1 System

Parameters

<i>nGen</i>	(> 0) Number of generations. This parameter indicate how many times the simulation is stopped, concentration are set to the initial ones and the simulazion restart for other n-Seconds seconds.
<i>nSIM</i>	(> 0) Number of simulations per generation starting with the same initial conditions (same data structures) but different random seed
<i>nSeconds</i>	(> 0) Number of seconds
<i>nReactions</i>	(> 0) Max number of reactions (the system will be stopped after nSeconds or after n-Reactions)
<i>randomSeed</i>	(>= 0) Random seed (if 0 the random seed is randomly created and the it is stored in the acsm2s.conf file saved in the results folder)
<i>nHours</i>	(>=0) Runtime limit (hours)
<i>nAttempts</i>	(>=0) Number of temptative in simulating the same network structure different random seed
<i>debugLevel</i>	(>= -1) Debug Level Runtime: different runTime message amounts (from -1 to 4, 0 is suggested)
<i>timeStructures-SavingInterval</i>	(> 0) All system structures (species, catalysis and reactions) are saved every <i>timeStructures-SavingInterval</i> seconds (simulation time)
<i>fileTimesSaveInterval</i>	(>= 0) Times data are stored in file times.csv every <i>fileTimesSaveInterval</i> seconds (If 0 reactions are stored continually)

2.3.2 Environment

Parameters

<i>newSpecies-ProbMin-Threshold</i>	(>=0) Minimal new species creation probability to allow system expansion
<i>lastFiringDisk-SpeciesID</i>	(> 0) The ID of the last firing disk species.
<i>overall-Concentration</i>	(> 0) The overall initial concentration that will be divided between all the initial species according to the selected initial distribution.
<i>EC-Concentration</i>	(> 0) Incoming concentration of charged molecules per second.
<i>alphabet</i>	(string) Alphabet used in the simulation (e.g. AGCT for DNA, ADEGFLYCWPHQIMTNKSRV for proteins)

<i>volume</i>	(> 0) Volume of the container or protocell
---------------	--

2.3.3 Dynamic

Parameters

<i>energy</i>	(0 or 1) 0 no energy in the system, 1 energy constraints are applied
<i>ratioSpecies-Energizable</i>	(%) The probability for a species to be potentially energized by the energy carriers
<i>nonCatalytic-MaxLength</i>	(>= 0) Max length of non catalytic species
<i>reaction-Probability</i>	(from 0 to 1) Probability for a species to catalyze a reaction
<i>cleavage-Probability</i>	(from 0 to 1) Cleavage probability (Condensation probability is 1 - cleavage probability)
<i>reverseReaction</i>	(0 or 1) Set to 1 to enable reverse reactions, 0 otherwise
<i>revRctRatio</i>	(>0) Ratio between forward and backward reactions, it is used in the creation of new reactions only (if reverseReactions = TRUE)
<i>spontRct</i>	(0 or 1) If 1 spontaneous reactions are considered, otherwise no
<i>K_ass</i>	(>= 0) Final Condensation kinetic constant ($C.A + B \rightarrow AB + C$) where A.C is the molecular complex composed of C (the catalyst) and A (the first substrate)
<i>K_diss</i>	(>= 0) Cleavage kinetic constant ($AB \rightarrow A + B$)
<i>K_cpx</i>	(>= 0) Complex formation kinetic constant ($A + C(\text{catalyst}) \rightarrow C.A$)
<i>K_cpxDiss</i>	(>= 0) Complex Dissociation kinetic constant ($C.A \rightarrow A + C$)
<i>K_nrg</i>	(>= 0) species phosphorylation kinetic constant
<i>K_nrg_decay</i>	(>= 0) de-energization kinetic constant
<i>moleculeDecay-KineticConstant</i>	(> 0) Molecule decay (efflux) kinetic Constant (Disregarded if the system is closed)
<i>K_spont_ass</i>	(>= 0) If spontaneous reactions are turned on this is the default kinetic rate for spontaneous condensations
<i>K_spont_diss</i>	(>= 0) If spontaneous reactions are turned on this is the default kinetic rate for spontaneous cleavages
<i>influx_rate</i>	(>= 0) Concentration per seconds (The species to insert in the system will be randomly chosen according to the <code>_acsinflux.csv</code> file). If equal to 0 the system is closed (<code>maxLOut=0</code>) or only the species that can cross the membrane come in and go out (<code>maxLOut>0</code>).
<i>maxLOut</i>	Maximum length of the species involved in the efflux process (<code>influx_rate > 0</code>), equal to 0 indicates that all the species can be involved in the efflux process (no filter). If <code>influx_rate = 0</code> the parameter indicates the species that can cross the semipermeable membrane of the protocell. THE COUPLING BETWEEN INFLUX_RATE AND MAXLOUT INDICATES IF WE ARE SIMULATING A PROTOCELL OR A FLOW REACTOR
<i>diffusion_contribute</i>	(KD) (0 or 0.5) if set to 0.5 the speed of molecules goes with the inverse of the square of the length, L^{-KD}
<i>solubility_threshold</i>	(> 0) Solubility Threshold, all the species longer than <code>solubility_threshold</code> precipitate

2.4 Acknowledgments

- University of Bologna, Interdepartment of industrial research (C.I.R.I.)
- European Centre for Living Technology <http://www.ecitech.org/>
- Fondazione Venezia <http://www.fondazionevenezia.it>
- Alex Graudenzi to take care of the initializer.
- Chiara Damiani to contribute to the development of the software.

- Roberto Serra, Marco Villani, Timoteo Carletti, Norman Packard, Ruedi Fuchslin and Stuart Kauffman for the essential hints.
- http://www.bedaux.net/mt_rand/ for the pseudo-random Marseinne-Twister library for C++.
- <http://perso.wanadoo.es/antlarr/otherapps.html> for the poisson distribution generator numbers (`acs_longInt random_poisson(acs_double tmpLambda, MTRand& tmpRandomGenerator)`).
- Dr. Luca Ansaloni (luca.ansaloni@unimore.it) for the support but especially for the file handling functions and new Python development.

Chapter 3

Initial Data Structures

In order to proceed with the simulation the system needs from **4 to 6 (open system and energy on)** initial data structures files (an example for each file is located into the source code folder):

- \c acsm2s.conf (described in the \ref parameters section)
- \c _acsspecies.csv - This file contains all the initial species with their proprieties
- \c _acsreactions.csv - This file contains all the initial reactions with their proprieties
- \c _acscatalysis.csv - This file contains all the correspondances between species and reactions with

and, if the system is open

- \c _acsinflux.csv - This file contains all the species belonging to the incoming flux

and. if the system is energy based

- \c _acsnrgbooleanfunctions.csv - This file contains all the possible boolean functions associated

3.1 _acsspecies.csv

Columns description (each field is delimited using "\t"):

Identification (ID)	Sequence	Concentration	Dif-fu-sion en-hance- ment	Pre-ci-pi-ta-tion flag	Com-plex Dis-so-cia-tion Ki-netic Con-stant	Com-plex Bind-ing Point	Al-ready eval-u-ated flag	Species Age	Num-ber of re-borns	Cat-a-lyst ID	Com-plex ID	Phos-pho-ri-la-tion Ki-netic con-stant	Charged Molecules Con-cen-tration	Con-cen-tration locked
------------------------	----------	---------------	----------------------------------	---------------------------	--	-------------------------------	---------------------------------	----------------	---------------------------	------------------	----------------	--	---	---------------------------

- *ID*: Species Index
- *Sequence*: Species sequence
- *Concentration*: Total concentration of the species
- *Diffusion enhancement*: Diffusion enhancement
- *Precipitation flag*: If 0 species is precipitated and each new molecules of this species will be precipitated
- *Dissociation Kinetic Constant*: Complex dissociation kinetic constant
- *Binding point*: If the species is a complex this field indicates the division point between catalyst and substrate
- *Evaluated*: If 1 the species is not virtual and all the reactions it catalyzes are created, if 0 the species is only potentially created
- *Species Age*: Age (in seconds) of the species since its last creation, each time that a species amount pass from 0 to > 1 the counter is resetted
- *Number of reborns*: Number of times that a species amount pass from 0 to > 0
- *Catalyst ID*: If the species is a complex (e.g. catalyst C forming a complex C.A with the substrate A) this is the CATALYST ID
- *Substrate ID*: If the species is a complex (e.g. catalyst C forming a complex C.A with the substrate A) this is the SUBSTATE ID
- *Phosphorilation Kinetic constant*: NOT USED NOW!!!
- *Charged Molecules Concentration*: Concentration of the charged molecules belonging to the species.
- *Concentration locked*: 0 -> Concentration of the species changes according to the reactions affecting it, 1 -> The concentration of the species is locked (permeable species, the concentration of the species is assumed to be constant within the cell and in the environment)

3.2 _acsreactions.csv

Columns description (each field is delimited using "\t"):

Identifica- tor	Reaction type	Species 1	Species 2	Species 3	Reaction counter	Energy type	k spont
--------------------	------------------	-----------	-----------	-----------	---------------------	----------------	---------

- *Identificator*: Reaction ID
- *Reaction type*: 0 Condensation, 1 Cleavage
- *Species 1*: Product ID if reaction type = 1, Substrate ID reaction type = 0
- *Species 2*: Product ID if reaction type = 0, Substrate ID reaction type = 1
- *Species 3*: Product ID if reaction type = 0, Substrate ID reaction type = 1
- *Reaction counter*: Reaction occurrence counter
- *Energy Type*: The reaction energetic configuration, 1 for endoergonic 0 for esoergonic
- *Spontaneous Constant*: Spontaneous reaction constant

3.3 _acscatalysis.csv

Columns description (each field is delimited using "\t"):

Identifica- tor	Catalyst ID	Reaction ID	Catalysis counter	K conden- sation	K cleavage	K Complex Associa- tion	Complex creation substrate target
--------------------	-------------	----------------	----------------------	---------------------	------------	-------------------------------	--

- *Identifier*: Catalysis ID
- *Catalyst ID*: species (as catalyst) ID
- *Reaction ID*: Reaction ID
- *Catalysis counter*: Catalysis counter
- *K final step end condensation kinetic constant*: Final step end condensation kinetic constant
- *K Cleavage*: Cleavage Kinetic constant
- *K complex association*: Complex association kinetic constant
- *Complex creation substrate target*: Which substrate will be involved in complex creation, 1 or 2

3.4 _acsinflux.csv

Columns description (each field is delimited using "\t"):

Identifier	Probability
------------	-------------

- *Identifier*: Species ID
- *Probability*: Probability to be selected when a species has to be inserted into the system

3.5 _acsnrgrbooleanfunctions.csv

Columns description (each field is delimited using "\t"):

Energetic Boolean Function (decimal form)	Probability
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- *Energetic Boolean Function (decimal form)*: Energetic Boolean Function decimal form, e.g. 1001011010100 = 4820
- *Probability*: Probability to be selected when a new reaction has to be created

Chapter 4

Outcomes

During the simulation **8** different files concerning species, reactions and catalysis tables, times, reactions parameters, living species, living species amounts and concentration are produced. In the following you find a brief description of all files and some indications about the different time storage.

4.1 Species

Every environment::timeStructuresSavingInterval seconds species structures will be saved in files called species_<generationNumber>_<simulationNumber>_<reactionNumber>.csv (e.g. **species_1_06_0016933.csv** means species structure, generation number 1, simulation number 6, reaction number 16933).

Columns description (each field is delimited using "\t") is the same described above, section [_acsspecies.csv](#).

4.2 Reactions

Every environment::timeStructuresSavingInterval seconds reactions structures will be saved in files called reactions-<generationNumber>_<simulationNumber>_<reactionNumber>.csv (e.g. **reactions_1_06_0016933.csv** means reactions structure, generation number 1, simulation number 6, reaction number 16933).

Columns description (each field is delimited using "\t") is the same described above, section [_acsreactions.csv](#).

4.3 Catalysis

Every environment::timeStructuresSavingInterval seconds catalysis structures will be saved in files called catalysis-<generationNumber>_<simulationNumber>_<reactionNumber>.csv (e.g. **catalysis_1_06_0016933.csv** means catalysis structure, generation number 1, simulation number 6, reaction number 16933).

Columns description (each field is delimited using "\t") is the same described above, section [_acscatalysis.csv](#) :

4.4 Times

Every simulation generates a times file called times_<generationNumber>_<simulationNumber>.csv (e.g. **times_1_03.csv** means times file, generation 1, simulation 3) containing values concerning **each events** occurred during the simulation. This file is **created at the beginning of the simulation** and it is **updated after each event**, hence this file will contain a number of rows equal to the number of reaction (of any type) occurred during the simulation.

Columns description (each field is delimited using "\t"):

Identifier (reaction)	Reaction Time	Gillespie ID selected	Reaction Type	Number of possible reactions	Computational Time (sec)	Number of Species	Number of Molecules	Number of complex species	Number of complexes	Number of bricks	Gillespie Computational Time	Reaction Process Computational Time	Various Processes Computational Time	New species creation probability
-----------------------	---------------	-----------------------	---------------	------------------------------	--------------------------	-------------------	---------------------	---------------------------	---------------------	------------------	------------------------------	-------------------------------------	--------------------------------------	----------------------------------

- *Identifier (reaction)*: Number of the reaction (STEP)
- *Reaction Time*: Time of the reaction
- *Gillespie ID selected*: Gillespie structure reaction ID
- *Reaction Type*: 0 condensation, 1 cleavage, 2 complex formation, 3 complex degradation, 4 species efflux, 6 endergonic cleavage, 7 endergonic condensation, 8 endergonic complex creation
- *Number of possible reactions*: Total number of possible reactions according to the Gillespie algorithm computation
- *Computational Time (ms)*: Computational time between two successive reactions
- *Number of Species*: Number of species with at least one molecule
- *Number of Molecules*: Number of molecules
- *Number of complex species*: Number of complex species with at least one molecule
- *Number of complexes*: Number of complex molecules
- *Number of bricks*: Number of single bricks (monomers) present in the system
- *Gillespie Computational Time*: Number of milliseconds necessary to complete the Gillespie task
- *Reaction Process Computational Time*: Number of milliseconds necessary to perform a reaction (and evaluate products) once that the Gillespie algorithm has selected the reaction
- *Various Processes Computational Time*: Number of milliseconds necessary to perform several tasks not correlated with the simulation of the phenomena
- *New species creation probability*: Given the state of the system, probability to create a new species
- *Reverse Reaction Probability*: Given the state of the system, probability for a reverse reaction to occur

4.5 Reactions_parameters

Every simulation generates a reactions parameters file called `reactions_parameters_<generationNumber>_<simulationNumber>.csv` (e.g. **reactions_parameters_1_07.csv** means reactions parameters file, generation 1 simulation number 7) containing informations on **cleavage and condensation** reactions. This file is **generated at the beginning of each simulation** and it is **updated each time that a cleavage or a condensation occur**.

Columns description (each field is delimited using "\t"):

Reaction ID	Reaction Time	Reaction Type	Catalyst ID	Species 1 ID	Species 2 ID	Species 3 ID	Charged Molecules	Charged Concentration	Gillespie Score Average	Gillespie Score Standard Deviation	Entropy	New species creation probability
-------------	---------------	---------------	-------------	--------------	--------------	--------------	-------------------	-----------------------	-------------------------	------------------------------------	---------	----------------------------------

- *Reaction ID*: This ID refers to the reaction number (step)
- *Reaction Time*: Time of the reaction
- *Reaction Type ID*: 0 condensation, 1 cleavage, 6 endoergonic cleavage, 7 endoergonic condensation
- *Catalyst ID*: Catalyst ID into the species table
- *Species 1 ID*: product ID in case of condensation, substrate in case of cleavage
- *Species 2 ID*: substrate ID in case of condensation, product in case of cleavage
- *Species 3 ID*: substrate ID in case of condensation, product in case of cleavage
- *Charged Molecules*: Number of charged molecules
- *Charged Concentration*: Concentration of the charged molecules
- *Gillespie Score Average*: Average of the all gillespie algorithm scores
- *Gillespie Score Standard Deviation*: Standard Deviation of the all gillespie algorithm scores
- *Entropy*: Entropy Misure (test) based on the probabilities that reactions occur
- *New species creation probability*: Given the state of the system, probability to create a new species

4.6 Living_species

Each simulation generates a living species file called `livingSpecies_<generationNumber>_<simulationNumber>.-csv` (e.g. **livingSpecies_1_07.csv** means living species file, generation 1 simulation number 7) containing all the living species IDs (living species are those species with concentration greater than 0) for each reaction in each row. This file is **generated at the beginning of each simulation** and it is **updated each time that a cleavage or a condensation occur**. This file has a variable number of columns equal to the number of living species plus 2 (Reaction ID and time of the reaction)

Columns description (each field is delimited using "\t"):

Reaction ID	Reaction Time	living species IDs..., one for column
-------------	---------------	---------------------------------------

- *Reaction ID*: This ID refers to the reaction number (step)
- *Reaction Time*: Time of the reaction
- *Living Species IDs..., one for column*: Each column from the thirth to the last one contains living species ID

4.7 Living_species_amounts

Each simulation generates a living species amount file called `livingAmount_<generationNumber>_<simulationNumber>.csv` (e.g. **livingAmount_1_07.csv** means living species amount file, generation 1, simulation number 7) containing all the living species amount (living species are those species with concentration greater than 0) for each reaction in each row. **This file is tightly correlated with the *Living_species* file**. Each row contains the total amount of molecules belonging to the species indicated by its ID in the `livingSpecies` file. This file is **created at the beginning of each simulation** and it is **updated each time that a cleavage or a condensation occur**. This file has a variable number of columns equals to the number of living species at time t

Columns description (each field is delimited using "\t"):

living species amounts..., one for column

- *Living Species amounts..., one for column*: Each values is related to the species indicated in the livingSpecies file. Reaction ID and time is that of the livingSpecies file

4.8 Living_species_concentration

Each simulation generates a living species amount file called livingConcentration_<generationNumber>_<simulationNumber>.csv (e.g. **livingConcentration_1_07.csv** means living species amount file, generation 1, simulation number 7) containing all the living species amount (living species are those species with concentration greater than 0) for each reaction in each raw. **This file is tightly correlated with the *Living_species* file.** Each row contains the total amount of molecules belonging to the species indicated by its ID in the livingSpecies file. This file is **created at the beginning of each simulation** and it is **updated each time that a cleavage or a condensation occur**. This file has a variable number of columns equals to the number of living species at time t

Columns description (each field is delimited using "\t"):

living species amounts..., one for column

- *Living Species amounts..., one for column*: Each values is related to the species indicated in the livingSpecies file. Reaction ID and time is that of the livingSpecies file

Chapter 5

Gillespie Class

The Gillespie class contains the information concerning the Gillespie algorithm propensity with other useful information. In particular it has 4 properties, namely mol_I, mol_II, mol_III, mol_IV containing the species ID's related to different species type according to the different admitted reactions. In the following a table summarizing the different meanings for the different reactions is reported.

Reaction Type	Reaction Type Code	MOL 1	MOL 2	MOL 3	MOL 4
Complex Formation	2	Catalyst	First Substrate	Catalysis ID	Second Substrate
Final Condensation	0	Catalyst	Substrate	Product	Complex
Cleavage	1	Substrate	Product 1	Product 2	Catalyst
Spontaneous Cleavage	11	Substrate	Product 1	Product 2	//
Spontaneous Condensation	10	Product	Substrate 1	Substrate 2	//

Chapter 6

The initializer (a very brief description)

The initializer provided with the simulator is located in the `initializer` folder (within the source code folder) and it is developed in Matlab code. All the parameters are set in the `start.m` file (from line 22 to line 63). In addition you find the parameters related to the name of the folder that will contain the simulation (`simFolder.name`), the path where that folder will be created (`simFolder.path`) and the number of different network ensambles to create (`simFolder.nets`). It is important to notice that the initializer has not been thought to be shared, so it is not too much user friendly to be manipulated. Nevertheless it could be very useful with a little bit of practice.

6.1 Screening Parameter

To initialize structures to perform a sensitivity analysis of a specific parameters follow the following instruction:

- Change the name of the array at row 13 with the name of the parameter you want analyze. This array contains the values of the paramter. Remember to insert the old parameter you are changing in the parameters list with its single value, otherwise it would be a missing parameter.
- In the array at row 14 (`nome_folder`) you have put the same number of elements of the array containing the screening values. This array contains a numeric tag of the values (used to create the simulations folder names) contained in the values array.
- In the code of the `start.m` file (rows 122 to 150) you must change:
 - row 121: `[rows,b]=size(reactionProbability); -> [rows,b]=size(name_of_the_parameter_you_have_trasformed_in_array);`
 - row 146: `inizializzatore_ACS(... reactionProbability(i), ...); -> inizializzatore_ACS(... name_of_the_parameter_you_have_trasformed_in_array(i), ...);` (remember to remove the index to the reaction-Probability parameter

Chapter 7

Namespace Index

7.1 Namespace List

Here is a list of all namespaces with brief descriptions:

acsAttractorAnalysis	??
acsAttractorAnalysisInTime	??
acsBufferedFluxes	??
acsDynStatInTime	??
acsFromWim2Carness	??
acsRAFanalysis	??
acsSCCanalysis	??
acsSpeciesActivities	??
acsStatesAnalysis	??
bufferedFluxAnalysis	??
fromWithin2Between	??
init	??
lib	??
lib.dyn	??
lib.dyn.dynamics	??
lib.graph	??
lib.graph.network	??
lib.graph.raf	??
lib.IO	??
lib.IO.readfiles	??
lib.IO.writefiles	??
main	??
prepareNewSim	??
resetForNewSimulations	??

Chapter 8

Hierarchical Index

8.1 Class Hierarchy

This inheritance list is sorted roughly, but not completely, alphabetically:

catalysis	??
commonFunctions	??
environment	??
gillespie	??
MTRand_int32	??
MTRand	??
MTRand53	??
MTRand_closed	??
MTRand_open	??
reactions	??
species	??

Chapter 9

Class Index

9.1 Class List

Here are the classes, structs, unions and interfaces with brief descriptions:

catalysis	CATALYSIS class	??
commonFunctions	This class contains all the common function of the system	??
environment	Environment class	??
gillespie	??
MTRand	??
MTRand53	??
MTRand_closed	??
MTRand_int32	??
MTRand_open	??
reactions	??
species	This class contains declarations of the species class	??

Chapter 10

File Index

10.1 File List

Here is a list of all files with brief descriptions:

/Users/alessandrofilisetti/Documents/GIT/carness/ acs_headers.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ catalysis.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ catalysis.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ commonFunctions.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ commonFunctions.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ environment.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ environment.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ gillespie.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ gillespie.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ main.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ mtrand.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ mtrand.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ reactions.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ reactions.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/ species.cpp	??
/Users/alessandrofilisetti/Documents/GIT/carness/ species.h	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ allTimesAnalysis.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ bufferedFluxAnalysis.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ concAnalysis.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ fromWithin2Between.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ garbageSearch.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ generalConcentrationOverThreshold.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ KillSpam.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ KSSearch.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ KSSearchLauncher.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ overallStats.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ readParameters.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ resetForNewSimulations.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ somma.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ stats.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ timesAnalysis.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/ timesAnalysis_PANINI.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/ acsAttractorAnalysis.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/ acsAttractorAnalysisInTime.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/ acsBufferedFluxes.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/ acsDynStatInTime.py	??

/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acsFromWim2Carness.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acsRAFanalysis.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acsSCCAnalysis.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acsSpeciesActivities.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acsStatesAnalysis.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/init.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/main.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/prepareNewSim.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/__init__.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/dyn/__init__.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/dyn/dynamics.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/graph/__init__.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/graph/network.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/graph/raf.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/IO/__init__.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/IO/readfiles.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/IO/writefiles.py	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_catalizzatori.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_concentrazioni_iniziali.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_e_controlla_i_catalizzatori.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_firing_disk.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_influx.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_influx_semplice.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_tutte_le_combinazioni_di_elementi.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/initial_distribution.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/inizializzatore_ACS.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/lancia_acs.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/lancia_inizializzatore_acs.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/start.m	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/catalysis.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/commonFunctions.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/environment.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/gillespie.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/main.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/mtrand.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/reactions.d	??
/Users/alessandrofilisetti/Documents/GIT/carness/Debug/species.d	??

Chapter 11

Namespace Documentation

11.1 `acsAttractorAnalysis` Namespace Reference

Functions

- def `zeroBeforeStrNum`

Variables

- tuple `StrPath` = `os.path.abspath(StrPath)`
- tuple `today` = `dt.date.today()`
- tuple `tmpDirs` = `sort(os.listdir(StrPath))`
- list `allSortedSpecies` = []
- list `allConcentrations` = []
- list `allSortedSpeciesNOINFLUX` = []
- list `allConcentrationsNOINFLUX` = []
- tuple `totDirName` = `os.path.join(StrPath,tmpDir)`
- tuple `resDirPath` = `os.path.abspath(os.path.join("./", "res"))`
- tuple `numberOfGen` = `len(glob.glob(os.path.join(resDirPath,'times_*')))`
- tuple `strZeros` = `zeroBeforeStrNum(nGen, numberOfGen)`
- string `strSpecies` = `'species_'`
- tuple `speciesFiles` = `sorted(glob.glob(os.path.join(resDirPath,strSpecies)))`
- list `speciesFile` = `speciesFiles[-1]`
- tuple `fidSpecies` = `open(speciesFile, 'r')`
- list `seq` = []
- list `overallConcList` = []
- list `overallConcListNOINFLUX` = []
- int `numberOfFolders` = 0
- tuple `pos` = `seq.index(key)`
- tuple `pos2` = `seq.index(key2)`
- tuple `ANG_overallResMatrix` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `ANG_overallResMatrixNOINFLUX` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `HAM_overallResMatrix` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `HAM_overallResMatrixNOINFLUX` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `EUC_overallResMatrix` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `EUC_overallResMatrixNOINFLUX` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `vecX` = `np.array(lx)`
- tuple `vecY` = `np.array(ly)`
- tuple `tmpCos` = `float(np.dot(vecX,vecY) / (np.linalg.norm(vecX) * np.linalg.norm(vecY)))`

- int `tmpHD` = 0
- int `tmpEU` = 0
- string `ndn` = '_0_new_allStatResults'
- tuple `newdirAllResults` = `os.path.join(os.getcwd(), ndn)`
- string `outFnameStat` = 'speciesVector.csv'
- tuple `saveFileStat` = `open(outFnameStat, 'w')`
- string `strTypes` = ''
- int `cnt` = 0

11.1.1 Function Documentation

11.1.1.1 `def acsAttractorAnalysis.zeroBeforeStrNum (tmpI, tmpL)`

Definition at line 25 of file `acsAttractorAnalysis.py`.

11.1.2 Variable Documentation

11.1.2.1 `list acsAttractorAnalysis.allConcentrations = []`

Definition at line 50 of file `acsAttractorAnalysis.py`.

11.1.2.2 `list acsAttractorAnalysis.allConcentrationsNOINFLUX = []`

Definition at line 52 of file `acsAttractorAnalysis.py`.

11.1.2.3 `tuple acsAttractorAnalysis.allSortedSpecies = []`

Definition at line 49 of file `acsAttractorAnalysis.py`.

11.1.2.4 `tuple acsAttractorAnalysis.allSortedSpeciesNOINFLUX = []`

Definition at line 51 of file `acsAttractorAnalysis.py`.

11.1.2.5 `tuple acsAttractorAnalysis.ANG_overallResMatrix = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 171 of file `acsAttractorAnalysis.py`.

11.1.2.6 `tuple acsAttractorAnalysis.ANG_overallResMatrixNOINFLUX = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 172 of file `acsAttractorAnalysis.py`.

11.1.2.7 `int acsAttractorAnalysis.cnt = 0`

Definition at line 262 of file `acsAttractorAnalysis.py`.

11.1.2.8 `tuple acsAttractorAnalysis.EUC_overallResMatrix = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 175 of file `acsAttractorAnalysis.py`.

11.1.2.9 `tuple acsAttractorAnalysis.EUC_overallResMatrixNOINFLUX = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 176 of file `acsAttractorAnalysis.py`.

11.1.2.10 `tuple acsAttractorAnalysis.fidSpecies = open(speciesFile, 'r')`

Definition at line 82 of file `acsAttractorAnalysis.py`.

11.1.2.11 `tuple acsAttractorAnalysis.HAM_overallResMatrix = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 173 of file `acsAttractorAnalysis.py`.

11.1.2.12 `tuple acsAttractorAnalysis.HAM_overallResMatrixNOINFLUX = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 174 of file `acsAttractorAnalysis.py`.

11.1.2.13 `string acsAttractorAnalysis.ndn = '_0_new_allStatResults'`

Definition at line 219 of file `acsAttractorAnalysis.py`.

11.1.2.14 `tuple acsAttractorAnalysis.newdirAllResults = os.path.join(os.curdir, ndn)`

Definition at line 220 of file `acsAttractorAnalysis.py`.

11.1.2.15 `int acsAttractorAnalysis.numberOfFolders = 0`

Definition at line 109 of file `acsAttractorAnalysis.py`.

11.1.2.16 `tuple acsAttractorAnalysis.numberOfGen = len(glob.glob(os.path.join(resDirPath,'times_*')))`

Definition at line 69 of file `acsAttractorAnalysis.py`.

11.1.2.17 `string acsAttractorAnalysis.outFnameStat = 'speciesVector.csv'`

Definition at line 230 of file `acsAttractorAnalysis.py`.

11.1.2.18 `list acsAttractorAnalysis.overallConcList = []`

Definition at line 107 of file `acsAttractorAnalysis.py`.

11.1.2.19 `list acsAttractorAnalysis.overallConcListNOINFLUX = []`

Definition at line 108 of file `acsAttractorAnalysis.py`.

11.1.2.20 `tuple acsAttractorAnalysis.pos = seq.index(key)`

Definition at line 153 of file `acsAttractorAnalysis.py`.

11.1.2.21 `tuple acsAttractorAnalysis.pos2 = seq.index(key2)`

Definition at line 159 of file `acsAttractorAnalysis.py`.

11.1.2.22 `tuple acsAttractorAnalysis.resDirPath = os.path.abspath(os.path.join(".", "res"))`

Definition at line 63 of file `acsAttractorAnalysis.py`.

11.1.2.23 `tuple acsAttractorAnalysis.saveFileStat = open(outFnameStat, 'w')`

Definition at line 231 of file `acsAttractorAnalysis.py`.

11.1.2.24 `list acsAttractorAnalysis.seq = []`

Definition at line 87 of file `acsAttractorAnalysis.py`.

11.1.2.25 `list acsAttractorAnalysis.speciesFile = speciesFiles[-1]`

Definition at line 78 of file `acsAttractorAnalysis.py`.

11.1.2.26 `tuple acsAttractorAnalysis.speciesFiles = sorted(glob.glob(os.path.join(resDirPath, strSpecies)))`

Definition at line 77 of file `acsAttractorAnalysis.py`.

11.1.2.27 `tuple acsAttractorAnalysis.StrPath = os.path.abspath(StrPath)`

Definition at line 42 of file `acsAttractorAnalysis.py`.

11.1.2.28 `string acsAttractorAnalysis.strSpecies = 'species_'`

Definition at line 74 of file `acsAttractorAnalysis.py`.

11.1.2.29 `string acsAttractorAnalysis.strTypes = "`

Definition at line 233 of file `acsAttractorAnalysis.py`.

11.1.2.30 `tuple acsAttractorAnalysis.strZeros = zeroBeforeStrNum(ngen, numberOfGen)`

Definition at line 73 of file `acsAttractorAnalysis.py`.

11.1.2.31 `tuple acsAttractorAnalysis.tmpCos = float(np.dot(vecX, vecY) / (np.linalg.norm(vecX) * np.linalg.norm(vecY)))`

Definition at line 184 of file `acsAttractorAnalysis.py`.

11.1.2.32 `tuple acsAttractorAnalysis.tmpDirs = sort(os.listdir(StrPath))`

Definition at line 48 of file `acsAttractorAnalysis.py`.

11.1.2.33 `int acsAttractorAnalysis.tmpEU = 0`

Definition at line 189 of file `acsAttractorAnalysis.py`.

11.1.2.34 `int acsAttractorAnalysis.tmpHD = 0`

Definition at line 188 of file `acsAttractorAnalysis.py`.

11.1.2.35 `tuple acsAttractorAnalysis.today = dt.date.today()`

Definition at line 44 of file `acsAttractorAnalysis.py`.

11.1.2.36 `tuple acsAttractorAnalysis.totDirName = os.path.join(StrPath,tmpDir)`

Definition at line 59 of file `acsAttractorAnalysis.py`.

11.1.2.37 `tuple acsAttractorAnalysis.vecX = np.array(lx)`

Definition at line 181 of file `acsAttractorAnalysis.py`.

11.1.2.38 `tuple acsAttractorAnalysis.vecY = np.array(ly)`

Definition at line 182 of file `acsAttractorAnalysis.py`.

11.2 acsAttractorAnalysisInTime Namespace Reference

Functions

- def [zeroBeforeStrNum](#)

Variables

- tuple [StrPath](#) = `os.path.abspath(StrPath)`
- tuple [today](#) = `dt.date.today()`
- tuple [tmpDirs](#) = `sort(os.listdir(StrPath))`
- list [allSortedSpecies](#) = []
- list [allConcentrations](#) = []
- list [allSortedSpeciesNOINFLUX](#) = []
- list [allConcentrationsNOINFLUX](#) = []
- tuple [totDirName](#) = `os.path.join(StrPath,tmpDir)`
- tuple [resDirPath](#) = `os.path.abspath(os.path.join("./", "res"))`
- tuple [numberOfGen](#) = `len(glob.glob(os.path.join(resDirPath,'times_*')))`
- tuple [strZeros](#) = `zeroBeforeStrNum(nGen, numberOfGen)`
- string [strSpeciesZero](#) = `'species_'`
- tuple [speciesFilesZero](#) = `sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))`
- string [strSpecies](#) = `'species_'`
- tuple [speciesFiles](#) = `sorted(glob.glob(os.path.join(resDirPath,strSpecies)))`
- list [speciesFile](#) = `speciesFiles[timeFileID]`
- tuple [fidSpecies](#) = `open(speciesFile, 'r')`
- list [seq](#) = []

- list `overallConcList` = []
- list `overallConcListNOINFLUX` = []
- int `numberOfFolders` = 0
- tuple `pos` = `seq.index(key)`
- tuple `pos2` = `seq.index(key2)`
- tuple `ANG_overallResMatrix` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `ANG_overallResMatrixNOINFLUX` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `HAM_overallResMatrix` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `HAM_overallResMatrixNOINFLUX` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `EUC_overallResMatrix` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `EUC_overallResMatrixNOINFLUX` = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple `vecX` = `np.array(lx)`
- tuple `vecY` = `np.array(ly)`
- tuple `tmpCos` = `float(np.dot(vecX,vecY) / (np.linalg.norm(vecX) * np.linalg.norm(vecY)))`
- int `tmpHD` = 0
- int `tmpEU` = 0
- string `ndn` = `'_0_new_allStatResults'`
- tuple `newdirAllResults` = `os.path.join(os.curdir, ndn)`
- tuple `tmpZeroSaving` = `zeroBeforeStrNum(timeFileID, tmpNOF)`
- string `outFnameStat` = `'speciesVector'`
- tuple `saveFileStat` = `open(outFnameStat, 'w')`
- string `strTypes` = ""
- int `cnt` = 0

11.2.1 Function Documentation

11.2.1.1 `def acsAttractorAnalysisInTime.zeroBeforeStrNum (tmpl, tmpl)`

Definition at line 23 of file `acsAttractorAnalysisInTime.py`.

11.2.2 Variable Documentation

11.2.2.1 `list acsAttractorAnalysisInTime.allConcentrations = []`

Definition at line 54 of file `acsAttractorAnalysisInTime.py`.

11.2.2.2 `list acsAttractorAnalysisInTime.allConcentrationsNOINFLUX = []`

Definition at line 56 of file `acsAttractorAnalysisInTime.py`.

11.2.2.3 `tuple acsAttractorAnalysisInTime.allSortedSpecies = []`

Definition at line 53 of file `acsAttractorAnalysisInTime.py`.

11.2.2.4 `tuple acsAttractorAnalysisInTime.allSortedSpeciesNOINFLUX = []`

Definition at line 55 of file `acsAttractorAnalysisInTime.py`.

11.2.2.5 `tuple acsAttractorAnalysisInTime.ANG_overallResMatrix = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 195 of file `acsAttractorAnalysisInTime.py`.

11.2.2.6 `tuple acsAttractorAnalysisInTime.ANG_overallResMatrixNOINFLUX = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 196 of file acsAttractorAnalysisInTime.py.

11.2.2.7 `int acsAttractorAnalysisInTime.cnt = 0`

Definition at line 298 of file acsAttractorAnalysisInTime.py.

11.2.2.8 `tuple acsAttractorAnalysisInTime.EUC_overallResMatrix = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 199 of file acsAttractorAnalysisInTime.py.

11.2.2.9 `tuple acsAttractorAnalysisInTime.EUC_overallResMatrixNOINFLUX = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 200 of file acsAttractorAnalysisInTime.py.

11.2.2.10 `tuple acsAttractorAnalysisInTime.fidSpecies = open(speciesFile, 'r')`

Definition at line 96 of file acsAttractorAnalysisInTime.py.

11.2.2.11 `tuple acsAttractorAnalysisInTime.HAM_overallResMatrix = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 197 of file acsAttractorAnalysisInTime.py.

11.2.2.12 `tuple acsAttractorAnalysisInTime.HAM_overallResMatrixNOINFLUX = np.zeros((numberOfFolders,numberOfFolders))`

Definition at line 198 of file acsAttractorAnalysisInTime.py.

11.2.2.13 `string acsAttractorAnalysisInTime.ndn = '_0_new_allStatResults'`

Definition at line 243 of file acsAttractorAnalysisInTime.py.

11.2.2.14 `tuple acsAttractorAnalysisInTime.newdirAllResults = os.path.join(os.curdir, ndn)`

Definition at line 244 of file acsAttractorAnalysisInTime.py.

11.2.2.15 `int acsAttractorAnalysisInTime.numberOfFolders = 0`

Definition at line 123 of file acsAttractorAnalysisInTime.py.

11.2.2.16 `tuple acsAttractorAnalysisInTime.numberOfGen = len(glob.glob(os.path.join(resDirPath,'times_*')))`

Definition at line 73 of file acsAttractorAnalysisInTime.py.

11.2.2.17 `string acsAttractorAnalysisInTime.outFnameStat = 'speciesVector'`

Definition at line 266 of file acsAttractorAnalysisInTime.py.

11.2.2.18 `list acsAttractorAnalysisInTime.overallConcList = []`

Definition at line 121 of file `acsAttractorAnalysisInTime.py`.

11.2.2.19 `list acsAttractorAnalysisInTime.overallConcListNOINFLUX = []`

Definition at line 122 of file `acsAttractorAnalysisInTime.py`.

11.2.2.20 `tuple acsAttractorAnalysisInTime.pos = seq.index(key)`

Definition at line 177 of file `acsAttractorAnalysisInTime.py`.

11.2.2.21 `tuple acsAttractorAnalysisInTime.pos2 = seq.index(key2)`

Definition at line 183 of file `acsAttractorAnalysisInTime.py`.

11.2.2.22 `tuple acsAttractorAnalysisInTime.resDirPath = os.path.abspath(os.path.join("./", "res"))`

Definition at line 67 of file `acsAttractorAnalysisInTime.py`.

11.2.2.23 `tuple acsAttractorAnalysisInTime.saveFileStat = open(outFnameStat, 'w')`

Definition at line 267 of file `acsAttractorAnalysisInTime.py`.

11.2.2.24 `list acsAttractorAnalysisInTime.seq = []`

Definition at line 101 of file `acsAttractorAnalysisInTime.py`.

11.2.2.25 `list acsAttractorAnalysisInTime.speciesFile = speciesFiles[timeFileID]`

Definition at line 91 of file `acsAttractorAnalysisInTime.py`.

11.2.2.26 `tuple acsAttractorAnalysisInTime.speciesFiles = sorted(glob.glob(os.path.join(resDirPath, strSpecies)))`

Definition at line 86 of file `acsAttractorAnalysisInTime.py`.

11.2.2.27 `tuple acsAttractorAnalysisInTime.speciesFilesZero = sorted(glob.glob(os.path.join(resDirPath, strSpeciesZero)))`

Definition at line 81 of file `acsAttractorAnalysisInTime.py`.

11.2.2.28 `tuple acsAttractorAnalysisInTime.StrPath = os.path.abspath(StrPath)`

Definition at line 41 of file `acsAttractorAnalysisInTime.py`.

11.2.2.29 `string acsAttractorAnalysisInTime.strSpecies = 'species_'`

Definition at line 83 of file `acsAttractorAnalysisInTime.py`.

11.2.2.30 `string acsAttractorAnalysisInTime.strSpeciesZero = 'species_'`

Definition at line 80 of file `acsAttractorAnalysisInTime.py`.

11.2.2.31 `string acsAttractorAnalysisInTime.strTypes = ''`

Definition at line 269 of file `acsAttractorAnalysisInTime.py`.

11.2.2.32 `tuple acsAttractorAnalysisInTime.strZeros = zeroBeforeStrNum(nngen, numberOfGen)`

Definition at line 77 of file `acsAttractorAnalysisInTime.py`.

11.2.2.33 `tuple acsAttractorAnalysisInTime.tmpCos = float(np.dot(vecX,vecY) / (np.linalg.norm(vecX) * np.linalg.norm(vecY)))`

Definition at line 208 of file `acsAttractorAnalysisInTime.py`.

11.2.2.34 `tuple acsAttractorAnalysisInTime.tmpDirs = sort(os.listdir(StrPath))`

Definition at line 52 of file `acsAttractorAnalysisInTime.py`.

11.2.2.35 `int acsAttractorAnalysisInTime.tmpEU = 0`

Definition at line 213 of file `acsAttractorAnalysisInTime.py`.

11.2.2.36 `int acsAttractorAnalysisInTime.tmpHD = 0`

Definition at line 212 of file `acsAttractorAnalysisInTime.py`.

11.2.2.37 `tuple acsAttractorAnalysisInTime.tmpZeroSaving = zeroBeforeStrNum(timeFileID, tmpNOF)`

Definition at line 263 of file `acsAttractorAnalysisInTime.py`.

11.2.2.38 `tuple acsAttractorAnalysisInTime.today = dt.date.today()`

Definition at line 43 of file `acsAttractorAnalysisInTime.py`.

11.2.2.39 `tuple acsAttractorAnalysisInTime.totDirName = os.path.join(StrPath,tmpDir)`

Definition at line 63 of file `acsAttractorAnalysisInTime.py`.

11.2.2.40 `tuple acsAttractorAnalysisInTime.vecX = np.array(lx)`

Definition at line 205 of file `acsAttractorAnalysisInTime.py`.

11.2.2.41 `tuple acsAttractorAnalysisInTime.vecY = np.array(ly)`

Definition at line 206 of file `acsAttractorAnalysisInTime.py`.

11.3 acsBufferedFluxes Namespace Reference

Functions

- def [zeroBeforeStrNum](#)

Variables

- tuple [parser](#)
- tuple [args](#) = parser.parse_args()
- tuple [strPath](#) = os.path.abspath(args.strPath)
- tuple [tmpDirs](#) = sort(os.listdir(StrPath))
- int [_CONDENSATION_](#) = 0
- int [_CLEAVAGE_](#) = 1
- int [_ENDOCONDENSATION_](#) = 7
- int [_ENDOCLEAVAGE_](#) = 6
- int [_SPONTCONDENSATION_](#) = 10
- int [_SPONTCLEAVAGE_](#) = 11
- int [chemistry](#) = 1
- string [ndn](#) = '_0_new_allStatResults'
- tuple [newdirAllResults](#) = os.path.join(StrPath, [ndn](#))
- tuple [totDirName](#) = os.path.join(StrPath, tmpDir)
- tuple [resDirPath](#) = os.path.abspath(os.path.join(".", "res"))
- tuple [numberOfGen](#) = len(glob.glob(os.path.join([resDirPath](#), 'times_*')))
- string [tmpFluxFile](#) = 'fluxDynamics_'
- tuple [tmpSpeciesStatsSummaryNameFID](#) = open(tmpFluxFile, 'w')
- tuple [strZeros](#) = [zeroBeforeStrNum](#)(ngen, [numberOfGen](#))
- string [strSpeciesZero](#) = 'species_'
- tuple [speciesFilesZero](#) = sorted(glob.glob(os.path.join([resDirPath](#), [strSpeciesZero](#))))
- string [strSpecies](#) = 'species_'
- tuple [speciesFiles](#) = sorted(glob.glob(os.path.join([resDirPath](#), [strSpecies](#))))
- list [lastfilespecies](#) = [speciesFiles](#)[-1]
- tuple [fidSpecies](#) = open([lastfilespecies](#), 'r')
- list [flux_seq](#) = []
- tuple [counters](#) = np.zeros((nSpecies, (len(seq)*2)+7))
- string [strRctPar](#) = 'reactions_parameters_'
- tuple [fidRctPar](#) = open([strRctPar](#), 'r')
- int [totIN](#) = 0
- int [totOUT](#) = 0
- int [deltaIO](#) = 0
- int [totBIN](#) = 0
- int [totBOUT](#) = 0
- tuple [rctTime](#) = int(tmpRctT)
- tuple [rctType](#) = int(tmpRctType)
- tuple [cat](#) = int(tmpCat)
- tuple [S1](#) = int(tmpS1)
- tuple [S2](#) = int(tmpS2)
- tuple [S3](#) = int(tmpS3)
- string [tmpFileName](#) = 'speciesStats_'
- tuple [tmpFileNameFID](#) = open([tmpFileName](#), 'w')
- int [ID](#) = 0
- string [tmpStr](#) = 'Total Number of Reactions\t\t\t\t\t'

11.3.1 Function Documentation

11.3.1.1 `def acsBufferedFluxes.zeroBeforeStrNum (tmpl, tmpL)`

Definition at line 20 of file `acsBufferedFluxes.py`.

11.3.2 Variable Documentation

11.3.2.1 `int acsBufferedFluxes._CLEAVAGE_ = 1`

Definition at line 42 of file `acsBufferedFluxes.py`.

11.3.2.2 `int acsBufferedFluxes._CONDENSATION_ = 0`

Definition at line 41 of file `acsBufferedFluxes.py`.

11.3.2.3 `int acsBufferedFluxes._ENDOCLEAVAGE_ = 6`

Definition at line 44 of file `acsBufferedFluxes.py`.

11.3.2.4 `int acsBufferedFluxes._ENDOCONDENSATION_ = 7`

Definition at line 43 of file `acsBufferedFluxes.py`.

11.3.2.5 `int acsBufferedFluxes._SPONTCLEAVAGE_ = 11`

Definition at line 46 of file `acsBufferedFluxes.py`.

11.3.2.6 `int acsBufferedFluxes._SPONTCONDENSATION_ = 10`

Definition at line 45 of file `acsBufferedFluxes.py`.

11.3.2.7 `tuple acsBufferedFluxes.args = parser.parse_args()`

Definition at line 34 of file `acsBufferedFluxes.py`.

11.3.2.8 `tuple acsBufferedFluxes.cat = int(tmpCat)`

Definition at line 133 of file `acsBufferedFluxes.py`.

11.3.2.9 `int acsBufferedFluxes.chemistry = 1`

Definition at line 50 of file `acsBufferedFluxes.py`.

11.3.2.10 `tuple acsBufferedFluxes.counters = np.zeros((nSpecies,(len(seq)*2)+7))`

Definition at line 111 of file `acsBufferedFluxes.py`.

11.3.2.11 `int acsBufferedFluxes.detalO = 0`

Definition at line 123 of file `acsBufferedFluxes.py`.

11.3.2.12 `tuple acsBufferedFluxes.fidRctPar = open(strRctPar, 'r')`

Definition at line 117 of file `acsBufferedFluxes.py`.

11.3.2.13 `tuple acsBufferedFluxes.fidSpecies = open(lastfilespecies, 'r')`

Definition at line 96 of file `acsBufferedFluxes.py`.

11.3.2.14 `list acsBufferedFluxes.flux_seq = []`

Definition at line 100 of file `acsBufferedFluxes.py`.

11.3.2.15 `int acsBufferedFluxes.ID = 0`

Definition at line 169 of file `acsBufferedFluxes.py`.

11.3.2.16 `list acsBufferedFluxes.lastfilespecies = speciesFiles[-1]`

Definition at line 92 of file `acsBufferedFluxes.py`.

11.3.2.17 `string acsBufferedFluxes.ndn = '_0_new_allStatResults'`

Definition at line 53 of file `acsBufferedFluxes.py`.

11.3.2.18 `tuple acsBufferedFluxes.newdirAllResults = os.path.join(StrPath, ndn)`

Definition at line 54 of file `acsBufferedFluxes.py`.

11.3.2.19 `tuple acsBufferedFluxes.numberOfGen = len(glob.glob(os.path.join(resDirPath, 'times_*')))`

Definition at line 73 of file `acsBufferedFluxes.py`.

11.3.2.20 `tuple acsBufferedFluxes.parser`

Initial value:

```
1 = ArgumentParser(
2     description='This script characterize the buffered flux dynamics.'
3     , epilog='''Buffered flux dynamics. '''
```

Definition at line 30 of file `acsBufferedFluxes.py`.

11.3.2.21 `tuple acsBufferedFluxes.rctTime = int(tmpRctT)`

Definition at line 131 of file `acsBufferedFluxes.py`.

11.3.2.22 `tuple acsBufferedFluxes.rctType = int(tmpRctType)`

Definition at line 132 of file `acsBufferedFluxes.py`.

11.3.2.23 `tuple acsBufferedFluxes.resDirPath = os.path.abspath(os.path.join("/", "res"))`

Definition at line 67 of file `acsBufferedFluxes.py`.

11.3.2.24 `tuple acsBufferedFluxes.S1 = int(tmpS1)`

Definition at line 134 of file `acsBufferedFluxes.py`.

11.3.2.25 `tuple acsBufferedFluxes.S2 = int(tmpS2)`

Definition at line 135 of file `acsBufferedFluxes.py`.

11.3.2.26 `tuple acsBufferedFluxes.S3 = int(tmpS3)`

Definition at line 136 of file `acsBufferedFluxes.py`.

11.3.2.27 `tuple acsBufferedFluxes.speciesFiles = sorted(glob.glob(os.path.join(resDirPath, strSpecies)))`

Definition at line 89 of file `acsBufferedFluxes.py`.

11.3.2.28 `tuple acsBufferedFluxes.speciesFilesZero = sorted(glob.glob(os.path.join(resDirPath, strSpeciesZero)))`

Definition at line 84 of file `acsBufferedFluxes.py`.

11.3.2.29 `tuple acsBufferedFluxes.strPath = os.path.abspath(args.strPath)`

Definition at line 37 of file `acsBufferedFluxes.py`.

11.3.2.30 `string acsBufferedFluxes.strRctPar = 'reactions_parameters_'`

Definition at line 113 of file `acsBufferedFluxes.py`.

11.3.2.31 `string acsBufferedFluxes.strSpecies = 'species_'`

Definition at line 86 of file `acsBufferedFluxes.py`.

11.3.2.32 `string acsBufferedFluxes.strSpeciesZero = 'species_'`

Definition at line 83 of file `acsBufferedFluxes.py`.

11.3.2.33 `tuple acsBufferedFluxes.strZeros = zeroBeforeStrNum(nngen, numberOfGen)`

Definition at line 81 of file `acsBufferedFluxes.py`.

11.3.2.34 `tuple acsBufferedFluxes.tmpDirs = sort(os.listdir(StrPath))`

Definition at line 38 of file `acsBufferedFluxes.py`.

11.3.2.35 `string acsBufferedFluxes.tmpFileName = 'speciesStats_'`

Definition at line 166 of file `acsBufferedFluxes.py`.

11.3.2.36 `tuple acsBufferedFluxes.tmpFileNameFID = open(tmpFileName, 'w')`

Definition at line 168 of file `acsBufferedFluxes.py`.

11.3.2.37 `string acsBufferedFluxes.tmpFluxFile = 'fluxDynamics_'`

Definition at line 78 of file `acsBufferedFluxes.py`.

11.3.2.38 `tuple acsBufferedFluxes.tmpSpeciesStatsSummaryNameFID = open(tmpFluxFile, 'w')`

Definition at line 79 of file `acsBufferedFluxes.py`.

11.3.2.39 `tuple acsBufferedFluxes.tmpStr = 'Total Number of Reactions\t\t\t\t'`

Definition at line 170 of file `acsBufferedFluxes.py`.

11.3.2.40 `int acsBufferedFluxes.totBIN = 0`

Definition at line 124 of file `acsBufferedFluxes.py`.

11.3.2.41 `int acsBufferedFluxes.totBOUT = 0`

Definition at line 125 of file `acsBufferedFluxes.py`.

11.3.2.42 `tuple acsBufferedFluxes.totDirName = os.path.join(StrPath,tmpDir)`

Definition at line 63 of file `acsBufferedFluxes.py`.

11.3.2.43 `int acsBufferedFluxes.totIN = 0`

Definition at line 121 of file `acsBufferedFluxes.py`.

11.3.2.44 `int acsBufferedFluxes.totOUT = 0`

Definition at line 122 of file `acsBufferedFluxes.py`.

11.4 `acsDynStatInTime` Namespace Reference

Variables

- tuple `parser`

- tuple `args` = `parser.parse_args()`
- tuple `strPath` = `os.path.abspath(args.strPath)`
- string `strSubStrKey` = `'*'`
- tuple `filesToAnal` = `sorted(glob.glob(os.path.join(strPath,strSubStrKey)))`
- tuple `lenFilesToAnal` = `len(filesToAnal)`
- int `numOfTraj` = 0
- tuple `x` = `np.array(map(list, np.loadtxt(fileToAnal, str)), float)`
- `xsize` = `x.shape`
- list `run4Chem` = `xsize[0]`
- tuple `y` = `np.zeros((lenFilesToAnal,numOfTraj))`
- int `pos` = 0
- int `chem` = 1
- string `tmpHead` = `"`
- string `filename` = `"_arranged_"`

11.4.1 Variable Documentation

11.4.1.1 tuple `acsDynStatInTime.args` = `parser.parse_args()`

Definition at line 24 of file `acsDynStatInTime.py`.

11.4.1.2 int `acsDynStatInTime.chem` = 1

Definition at line 47 of file `acsDynStatInTime.py`.

11.4.1.3 string `acsDynStatInTime.filename` = `"_arranged_"`

Definition at line 66 of file `acsDynStatInTime.py`.

11.4.1.4 tuple `acsDynStatInTime.filesToAnal` = `sorted(glob.glob(os.path.join(strPath,strSubStrKey)))`

Definition at line 34 of file `acsDynStatInTime.py`.

11.4.1.5 tuple `acsDynStatInTime.lenFilesToAnal` = `len(filesToAnal)`

Definition at line 35 of file `acsDynStatInTime.py`.

11.4.1.6 tuple `acsDynStatInTime.numOfTraj` = 0

Definition at line 36 of file `acsDynStatInTime.py`.

11.4.1.7 tuple `acsDynStatInTime.parser`

Initial value:

```
1 = ArgumentParser(
2     description='This script re-arrange results in a more friendly way from the
   angle analysis in time.'
3     , epilog='''File with angle trajectories are created. ''')
```

Definition at line 17 of file `acsDynStatInTime.py`.

11.4.1.8 `int acsDynStatInTime.pos = 0`

Definition at line 46 of file `acsDynStatInTime.py`.

11.4.1.9 `list acsDynStatInTime.run4Chem = xsize[0]`

Definition at line 42 of file `acsDynStatInTime.py`.

11.4.1.10 `tuple acsDynStatInTime.strPath = os.path.abspath(args.strPath)`

Definition at line 27 of file `acsDynStatInTime.py`.

11.4.1.11 `string acsDynStatInTime.strSubStrKey = '*'`

Definition at line 33 of file `acsDynStatInTime.py`.

11.4.1.12 `string acsDynStatInTime.tmpHead = "`

Definition at line 48 of file `acsDynStatInTime.py`.

11.4.1.13 `tuple acsDynStatInTime.x = np.array(map(list, np.loadtxt(fileToAnal, str)), float)`

Definition at line 39 of file `acsDynStatInTime.py`.

11.4.1.14 `acsDynStatInTime.xsize = x.shape`

Definition at line 40 of file `acsDynStatInTime.py`.

11.4.1.15 `tuple acsDynStatInTime.y = np.zeros((lenFilesToAnal,numOfTraj))`

Definition at line 44 of file `acsDynStatInTime.py`.

11.5 `acsFromWim2Carness` Namespace Reference

Variables

- tuple `parser`
- tuple `args` = `parser.parse_args()`
- string `ndn` = `'_0_new_allStatResults'`
- tuple `newdirAllResults` = `os.path.join(args.strOut, ndn)`
- tuple `fname_initRafRes` = `os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`
- tuple `fname_initRafResLIST` = `os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`
- tuple `fname_initRafResALL` = `os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`
- tuple `fid_initRafRes` = `open(fname_initRafRes, 'w')`
- tuple `fid_initRafResLIST` = `open(fname_initRafResLIST, 'w')`
- tuple `fid_initRafResALL` = `open(fname_initRafResALL, 'w')`
- string `strToWrite` = `"Folder\TP\TAC\TM\TRAfsize\Closure\Cats\tuRAF\n"`
- tuple `f` = `open(args.wimFile)`
- tuple `lines` = `f.readlines()`
- int `procedure` = 0

- list `speciesList` = []
- list `foodList` = []
- int `rct` = 0
- tuple `l` = `line.split()`
- tuple `catNums` = `len(l)`
- tuple `cats` = `np.vstack([cats,(int(rct), int(speciesList.index(l[6+catNums-1][0:len(l[6+catNums-1])-1])), int(rct), int(0), int(5), int(25), int(5), int(1))))]`
- tuple `rafsets` = `raf.rafComputation(fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, 'tmpDir', 0, rct/float(len(speciesList)), rcts, cats, foodList, 10)`

11.5.1 Variable Documentation

11.5.1.1 tuple `acsFromWim2Carness.args` = `parser.parse_args()`

Definition at line 33 of file `acsFromWim2Carness.py`.

11.5.1.2 tuple `acsFromWim2Carness.catNums` = `len(l)`

Definition at line 100 of file `acsFromWim2Carness.py`.

11.5.1.3 tuple `acsFromWim2Carness.cats` = `np.vstack([cats,(int(rct), int(speciesList.index(l[6+catNums-1][0:len(l[6+catNums-1])-1])), int(rct), int(0), int(5), int(25), int(5), int(1))))]`

Definition at line 110 of file `acsFromWim2Carness.py`.

11.5.1.4 tuple `acsFromWim2Carness.f` = `open(args.wimFile)`

Definition at line 54 of file `acsFromWim2Carness.py`.

11.5.1.5 tuple `acsFromWim2Carness.fid_initRafRes` = `open(fname_initRafRes, 'w')`

Definition at line 48 of file `acsFromWim2Carness.py`.

11.5.1.6 tuple `acsFromWim2Carness.fid_initRafResALL` = `open(fname_initRafResALL, 'w')`

Definition at line 50 of file `acsFromWim2Carness.py`.

11.5.1.7 tuple `acsFromWim2Carness.fid_initRafResLIST` = `open(fname_initRafResLIST, 'w')`

Definition at line 49 of file `acsFromWim2Carness.py`.

11.5.1.8 tuple `acsFromWim2Carness.fname_initRafRes` = `os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`

Definition at line 45 of file `acsFromWim2Carness.py`.

11.5.1.9 tuple `acsFromWim2Carness.fname_initRafResALL` = `os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`

Definition at line 47 of file `acsFromWim2Carness.py`.

11.5.1.10 tuple `acsFromWim2Carness.fname_initRafResLIST = os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`

Definition at line 46 of file `acsFromWim2Carness.py`.

11.5.1.11 list `acsFromWim2Carness.foodList = []`

Definition at line 58 of file `acsFromWim2Carness.py`.

11.5.1.12 tuple `acsFromWim2Carness.l = line.split()`

Definition at line 74 of file `acsFromWim2Carness.py`.

11.5.1.13 tuple `acsFromWim2Carness.lines = f.readlines()`

Definition at line 55 of file `acsFromWim2Carness.py`.

11.5.1.14 string `acsFromWim2Carness.ndn = '_0_new_allStatResults'`

Definition at line 36 of file `acsFromWim2Carness.py`.

11.5.1.15 tuple `acsFromWim2Carness.newdirAllResults = os.path.join(args.strOut, ndn)`

Definition at line 37 of file `acsFromWim2Carness.py`.

11.5.1.16 tuple `acsFromWim2Carness.parser`

Initial value:

```
1 = ArgumentParser(
2     description='From WIM format to my format'
3     , epilog='''From WIM format to my format. '''
```

Definition at line 26 of file `acsFromWim2Carness.py`.

11.5.1.17 int `acsFromWim2Carness.procedure = 0`

Definition at line 56 of file `acsFromWim2Carness.py`.

11.5.1.18 tuple `acsFromWim2Carness.rafsets = raf.rafComputation(fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, 'tmpDir', 0, rct/float(len(speciesList)), rcts, cats, foodList, 10)`

Definition at line 116 of file `acsFromWim2Carness.py`.

11.5.1.19 int `acsFromWim2Carness.rct = 0`

Definition at line 59 of file `acsFromWim2Carness.py`.

11.5.1.20 list `acsFromWim2Carness.speciesList = []`

Definition at line 57 of file `acsFromWim2Carness.py`.

11.5.1.21 `string acsFromWim2Carness.strToWrite = "Folder\\tP\\tAC\\tMtRAFsize\\tClosure\\tCats\\tuRAF\\n"`

Definition at line 51 of file `acsFromWim2Carness.py`.

11.6 acsRAFanalysis Namespace Reference

Variables

- tuple `parser`
- tuple `args` = `parser.parse_args()`
- tuple `strPath` = `os.path.abspath(args.strPath)`
- int `_CLOSE_` = 0
- int `_PROTO_` = 1
- int `_CSTR_` = 2
- tuple `conf` = `rf.readConfFile(strPath)`
- tuple `closure` = `dm.generateFluxList(strPath, sysType)`
- tuple `foodSet` = `deepcopy(closure)`
- tuple `rcts` = `rf.loadAllData(strPath, '_acsreactions.csv')`
- tuple `cats` = `rf.loadAllData(strPath, '_acscatalysis.csv')`
- tuple `RA` = `raf.RAcondition(strPath, closure, rcts, cats)`
- tuple `RAF` = `raf.Fcondition(strPath, closure, RA, rcts)`
- tuple `RAFlpre` = `len(RAF)`
- list `redRcts` = `rcts[RAF, :]`
- int `RAFlpost` = 0
- tuple `catalists` = `raf.findCatforRAF(cats, RAF, closure)`

11.6.1 Variable Documentation

11.6.1.1 `int acsRAFanalysis._CLOSE_ = 0`

Definition at line 31 of file `acsRAFanalysis.py`.

11.6.1.2 `int acsRAFanalysis._CSTR_ = 2`

Definition at line 33 of file `acsRAFanalysis.py`.

11.6.1.3 `int acsRAFanalysis._PROTO_ = 1`

Definition at line 32 of file `acsRAFanalysis.py`.

11.6.1.4 `tuple acsRAFanalysis.args = parser.parse_args()`

Definition at line 26 of file `acsRAFanalysis.py`.

11.6.1.5 `tuple acsRAFanalysis.catalists = raf.findCatforRAF(cats, RAF, closure)`

Definition at line 76 of file `acsRAFanalysis.py`.

11.6.1.6 `tuple acsRAFanalysis.cats = rf.loadAllData(strPath, '_acscatalysis.csv')`

Definition at line 45 of file `acsRAFanalysis.py`.

11.6.1.7 tuple `acsRAFanalysis.closure = dm.generateFluxList(strPath, sysType)`

Definition at line 41 of file `acsRAFanalysis.py`.

11.6.1.8 tuple `acsRAFanalysis.conf = rf.readConfFile(strPath)`

Definition at line 34 of file `acsRAFanalysis.py`.

11.6.1.9 tuple `acsRAFanalysis.foodSet = deepcopy(closure)`

Definition at line 42 of file `acsRAFanalysis.py`.

11.6.1.10 tuple `acsRAFanalysis.parser`

Initial value:

```
1 = ArgumentParser(
2     description='Script to perform a RAF analysis.'
3     , epilog='''File with angle trajectories are created.''' )
```

Definition at line 22 of file `acsRAFanalysis.py`.

11.6.1.11 tuple `acsRAFanalysis.RA = raf.RAcondition(strPath,closure,rcts,cats)`

Definition at line 50 of file `acsRAFanalysis.py`.

11.6.1.12 tuple `acsRAFanalysis.RAF = raf.Fcondition(strPath,closure,RA,rcts)`

Definition at line 52 of file `acsRAFanalysis.py`.

11.6.1.13 tuple `acsRAFanalysis.RAFIpost = 0`

Definition at line 60 of file `acsRAFanalysis.py`.

11.6.1.14 tuple `acsRAFanalysis.RAFIpre = len(RAF)`

Definition at line 53 of file `acsRAFanalysis.py`.

11.6.1.15 tuple `acsRAFanalysis.rcts = rf.loadAllData(strPath,'_acsreactions.csv')`

Definition at line 44 of file `acsRAFanalysis.py`.

11.6.1.16 list `acsRAFanalysis.redRcts = rcts[RAF,:]`

Definition at line 55 of file `acsRAFanalysis.py`.

11.6.1.17 tuple `acsRAFanalysis.strPath = os.path.abspath(args.strPath)`

Definition at line 29 of file `acsRAFanalysis.py`.

11.7 acsSCAnalysis Namespace Reference

Functions

- def [zeroBeforeStrNum](#)
- def [loadReactionGraph](#)
- def [loadSpecificReactionGraph](#)
- def [loadSpecificReactionSubGraph](#)
- def [saveGraphToFile](#)
- def [saveGraphSUBToFile](#)
- def [saveNrgToFile](#)
- def [saveGillToFile](#)

Variables

- tuple [StrPath](#) = os.path.abspath(StrPath)
- tuple [today](#) = dt.date.today()
- tuple [mswindows](#) = (sys.platform == "win32")
- string [cmdFileName](#) = StrPath+'/'
- tuple [cmdFileFid](#) = open(cmdFileName, 'a')
- string [strToWrite](#) = "\tReaction Probability"
- int [initilizeGraphStructure](#) = 0
- tuple [graph](#) = [loadSpecificReactionGraph](#)()
- tuple [graphSUB](#) = [loadSpecificReactionSubGraph](#)()
- tuple [tmpRctFileToLoadSplitString](#) = tmpRctFileToLoad.split("_")
- tuple [initRctId](#) = int(tmpRctFileToLoadSplitString[len(tmpRctFileToLoadSplitString)-2])
- list [initTempTime](#) = tmpRctFileToLoadSplitString[len(tmpRctFileToLoadSplitString)-1]
- tuple [initTime](#) = float(initTempTime[0:len(initTempTime)-4])
- tuple [tmpDirs](#) = sort(os.listdir(StrPath))
- tuple [totDirName](#) = os.path.join(StrPath,tmpDir)
- tuple [resDirPath](#) = os.path.abspath(os.path.join("./", "res"))
- tuple [numberOfGen](#) = len(glob.glob(os.path.join(resDirPath,'times_*')))
- tuple [strZeros](#) = [zeroBeforeStrNum](#)(nGen, [numberOfGen](#))
- string [paramFile](#) = "acsm2s.conf"
- int [simFolder](#) = 0
- tuple [fid](#) = open(paramFile, 'r')
- tuple [strLine](#) = line.split('=')
- tuple [rp](#) = float(strLine[1])
- tuple [decayTime](#) = int(strLine[1])
- tuple [totTimes](#) = int(strLine[1])
- tuple [nrgType](#) = int(strLine[1])
- tuple [totalRcts](#) = int(strLine[1])
- tuple [nrgConc](#) = float(strLine[1])
- tuple [influx_rate](#) = float(strLine[1])
- tuple [maxLOut](#) = float(strLine[1])
- int [_ANALTIMEINTERAVAL_](#) = totTimes/10
- int [_ANALTIMEINTERAVALNOSAVE_](#) = totTimes/100
- int [nrg](#) = 1
- tuple [speciesInFlux](#) = range(0,int(pow(2,(maxLOut+1)) - 2))
- tuple [fidflux](#) = open('_acsinflux.csv', 'r')
- tuple [overThreshold](#) = float(0)
- tuple [overThresholdTOT](#) = float(0)
- string [tmpFilesToSearch](#) = 'species_'
- tuple [speciesFiles](#) = sorted(glob.glob(os.path.join(resDirPath,tmpFilesToSearch)))

- list `speciesFile` = `speciesFiles[-1]`
- tuple `fidSpecies` = `open(speciesFile, 'r')`
- int `ok` = 0
- list `IDsOverThreshold` = []
- list `concVec` = []
- tuple `index` = `int(tmpID)`
- tuple `conc` = `float(tmpConc)`
- tuple `cpxCut` = `int(tmpCpxCut)`
- tuple `age` = `float(tmpAge)`
- `realT` = `threshold`
- string `folderCat` = `'__0_iGraph_CAT_'`
- string `folderSub` = `'__0_iGraph_SUB_'`
- tuple `newdir` = `os.path.join(os.curdir, folderCat)`
- tuple `newdirSUB` = `os.path.join(os.curdir, folderSub)`
- string `filextPre` = `'_'`
- string `rctParamFileQ` = `'reactions_parameters_'`
- tuple `rctParamFile` = `sorted(glob.glob(os.path.join(resDirPath,rctParamFileQ)))`
- float `rctIDshow` = 1.0
- float `rctIDshowNoSave` = 1.0
- `rctID` = `initRctId`
- int `previousTime` = 0
- int `endo_condensation_counter` = 0
- int `condensation_counter` = 0
- int `endo_cleavage_counter` = 0
- int `cleavage_counter` = 0
- tuple `nrgTimeSeries` = `np.array([[0,0,0]])`
- tuple `gillTimeSeries` = `np.array([[0,0,0,0]])`
- int `tmpFlagLastRctSaved` = 0
- tuple `reaction` = `int(tmpReaction)`
- tuple `rtime` = `float(tmpTime)`
- tuple `cc` = `int(tmpcc)`
- tuple `cat` = `int(tmpCat)`
- tuple `mol_I` = `int(tmpMol_I)`
- tuple `mol_II` = `int(tmpMol_II)`
- tuple `mol_III` = `int(tmpMol_III)`
- tuple `loadedMolsConc` = `float(tmpLoadedMolsConc)`
- tuple `loadedMols` = `int(tmpLoadedMols)`
- tuple `gillMean` = `float(tmpGillMean)`
- tuple `gillSD` = `float(tmpGillSD)`
- tuple `gillEntropy` = `float(tmpGillEntropy)`
- tuple `newSpeciesCreationProb` = `float(tmpNSCprob)`
- tuple `reverseProbability` = `float(tmpRevProb)`
- int `printTemporalMessage` = 1
- `timeInterval` = `rtime-previousTime`
- tuple `position` = `((graph[:,0] == cat) & (graph[:,1] == mol_I))`
- int `realSccs` = 0
- tuple `Gcatpro` = `nx.DiGraph()`
- tuple `scc` = `nx.strongly_connected_components(Gcatpro)`
- tuple `sccN` = `nx.number_strongly_connected_components(Gcatpro)`
- tuple `selfLoops` = `Gcatpro.number_of_selfloops()`
- tuple `selfLoopsEgdes` = `Gcatpro.selfloop_edges()`
- int `prod_inSCC` = 0
- int `prod_chain` = 0
- int `prod_bySCC` = 0
- int `prod_overlap` = 0

- int `sccID` = 0
- int `autocatalysis` = 0
- int `prod_inSCC_weight` = 0
- int `prod_chain_weight` = 0
- int `prod_bySCC_weight` = 0
- int `prod_overlap_weight` = 0
- int `self_loop_weight` = 0
- int `conc_inSCC` = 0
- int `conc_chain` = 0
- int `conc_bySCC` = 0
- int `conc_overLap` = 0
- int `conc_selfCat` = 0
- int `wasteSpecies` = 0
- int `alreadyAdded_ACS` = 0
- int `alreadyAdded_leaves` = 0
- int `alreadyAdded_chain` = 0
- int `tmpProd_chain` = 0
- tuple `incomingNodes` = `Gcatpro.predecessors(IdsOT)`
- int `tempProd_chain_weight` = 0
- int `nolnAcs` = 1
- int `inSCCFlag` = 0
- list `weightToDistribute` = `graph[((graph[:,0] == sngInNode) & (graph[:,1] == IdsOT)),5]`
- tuple `inDegreeMean` = `mean(Gcatpro.in_degree().values())`
- tuple `meanOverThreshold` = `float(overThreshold)`

11.7.1 Function Documentation

11.7.1.1 `def acsSCCanalysis.loadReactionGraph ()`

Definition at line 31 of file `acsSCCanalysis.py`.

11.7.1.2 `def acsSCCanalysis.loadSpecificReactionGraph ()`

Definition at line 64 of file `acsSCCanalysis.py`.

11.7.1.3 `def acsSCCanalysis.loadSpecificReactionSubGraph ()`

Definition at line 93 of file `acsSCCanalysis.py`.

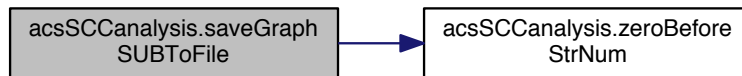
11.7.1.4 `def acsSCCanalysis.saveGillToFile ()`

Definition at line 154 of file `acsSCCanalysis.py`.

11.7.1.5 `def acsSCCanalysis.saveGraphSUBToFile ()`

Definition at line 133 of file `acsSCCanalysis.py`.

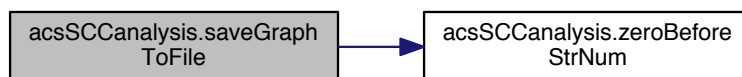
Here is the call graph for this function:



11.7.1.6 `def acsSCCanalysis.saveGraphToFile ()`

Definition at line 122 of file `acsSCCanalysis.py`.

Here is the call graph for this function:



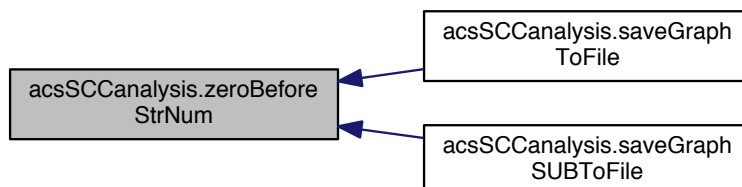
11.7.1.7 `def acsSCCanalysis.saveNrgToFile ()`

Definition at line 144 of file `acsSCCanalysis.py`.

11.7.1.8 `def acsSCCanalysis.zeroBeforeStrNum (tmpl, tmpL)`

Definition at line 22 of file `acsSCCanalysis.py`.

Here is the caller graph for this function:



11.7.2 Variable Documentation

11.7.2.1 `int acsSCCanalysis._ANALTIMEINTERAVAL_ = totTimes/10`

Definition at line 269 of file acsSCCanalysis.py.

11.7.2.2 `int acsSCCanalysis._ANALTIMEINTERAVALNOSAVE_ = totTimes/100`

Definition at line 270 of file acsSCCanalysis.py.

11.7.2.3 `tuple acsSCCanalysis.age = float(tmpAge)`

Definition at line 329 of file acsSCCanalysis.py.

11.7.2.4 `int acsSCCanalysis.alreadyAdded_ACS = 0`

Definition at line 600 of file acsSCCanalysis.py.

11.7.2.5 `int acsSCCanalysis.alreadyAdded_chain = 0`

Definition at line 602 of file acsSCCanalysis.py.

11.7.2.6 `int acsSCCanalysis.alreadyAdded_leaves = 0`

Definition at line 601 of file acsSCCanalysis.py.

11.7.2.7 `int acsSCCanalysis.autocatalysis = 0`

Definition at line 573 of file acsSCCanalysis.py.

11.7.2.8 `tuple acsSCCanalysis.cat = int(tmpCat)`

Definition at line 411 of file acsSCCanalysis.py.

11.7.2.9 `tuple acsSCCanalysis.cc = int(tmpcc)`

Definition at line 410 of file acsSCCanalysis.py.

11.7.2.10 `int acsSCCanalysis.cleavage_counter = 0`

Definition at line 392 of file acsSCCanalysis.py.

11.7.2.11 `tuple acsSCCanalysis.cmdFileFid = open(cmdFileName, 'a')`

Definition at line 185 of file acsSCCanalysis.py.

11.7.2.12 `string acsSCCanalysis.cmdFileName = StrPath+'/'`

Definition at line 182 of file acsSCCanalysis.py.

11.7.2.13 `tuple acsSCCanalysis.conc = float(tmpConc)`

Definition at line 327 of file acsSCCanalysis.py.

11.7.2.14 `int acsSCCanalysis.conc_bySCC = 0`

Definition at line 581 of file acsSCCanalysis.py.

11.7.2.15 `int acsSCCanalysis.conc_chain = 0`

Definition at line 580 of file acsSCCanalysis.py.

11.7.2.16 `int acsSCCanalysis.conc_inSCC = 0`

Definition at line 579 of file acsSCCanalysis.py.

11.7.2.17 `int acsSCCanalysis.conc_overLap = 0`

Definition at line 582 of file acsSCCanalysis.py.

11.7.2.18 `int acsSCCanalysis.conc_selfCat = 0`

Definition at line 583 of file acsSCCanalysis.py.

11.7.2.19 `list acsSCCanalysis.concVec = []`

Definition at line 318 of file acsSCCanalysis.py.

11.7.2.20 `int acsSCCanalysis.condensation_counter = 0`

Definition at line 390 of file acsSCCanalysis.py.

11.7.2.21 `tuple acsSCCanalysis.cpxCut = int(tmpCpxCut)`

Definition at line 328 of file acsSCCanalysis.py.

11.7.2.22 `tuple acsSCCanalysis.decayTime = int(strLine[1])`

Definition at line 255 of file acsSCCanalysis.py.

11.7.2.23 `int acsSCCanalysis.endo_cleavage_counter = 0`

Definition at line 391 of file acsSCCanalysis.py.

11.7.2.24 `int acsSCCanalysis.endo_condensation_counter = 0`

Definition at line 389 of file acsSCCanalysis.py.

11.7.2.25 `tuple acsSCCanalysis.fid = open(paramFile, 'r')`

Definition at line 244 of file acsSCCanalysis.py.

11.7.2.26 `tuple acsSCCanalysis.fidflux = open('_acsinflux.csv', 'r')`

Definition at line 285 of file acsSCCanalysis.py.

11.7.2.27 `tuple acsSCCanalysis.fidSpecies = open(speciesFile, 'r')`

Definition at line 311 of file acsSCCanalysis.py.

11.7.2.28 `string acsSCCanalysis.filextPre = '_'`

Definition at line 376 of file acsSCCanalysis.py.

11.7.2.29 `string acsSCCanalysis.folderCat = '__0_iGraph_CAT_'`

Definition at line 348 of file acsSCCanalysis.py.

11.7.2.30 `string acsSCCanalysis.folderSub = '__0_iGraph_SUB_'`

Definition at line 349 of file acsSCCanalysis.py.

11.7.2.31 `tuple acsSCCanalysis.Gcatpro = nx.DiGraph()`

Definition at line 555 of file acsSCCanalysis.py.

11.7.2.32 `tuple acsSCCanalysis.gillEntropy = float(tmpGillEntropy)`

Definition at line 419 of file acsSCCanalysis.py.

11.7.2.33 `tuple acsSCCanalysis.gillMean = float(tmpGillMean)`

Definition at line 417 of file acsSCCanalysis.py.

11.7.2.34 `tuple acsSCCanalysis.gillSD = float(tmpGillSD)`

Definition at line 418 of file acsSCCanalysis.py.

11.7.2.35 `tuple acsSCCanalysis.gillTimeSeries = np.array([[0,0,0,0]])`

Definition at line 399 of file acsSCCanalysis.py.

11.7.2.36 `tuple acsSCCanalysis.graph = loadSpecificReactionGraph()`

Definition at line 193 of file acsSCCanalysis.py.

11.7.2.37 tuple `acsSCCanalysis.graphSUB = loadSpecificReactionSubGraph()`

Definition at line 194 of file `acsSCCanalysis.py`.

11.7.2.38 list `acsSCCanalysis.IDsOverThreshold = []`

Definition at line 317 of file `acsSCCanalysis.py`.

11.7.2.39 tuple `acsSCCanalysis.incomingNodes = Gcatpro.predecessors(idsOT)`

Definition at line 605 of file `acsSCCanalysis.py`.

11.7.2.40 tuple `acsSCCanalysis.inDegreeMean = mean(Gcatpro.in_degree().values())`

Definition at line 682 of file `acsSCCanalysis.py`.

11.7.2.41 tuple `acsSCCanalysis.index = int(tmpID)`

Definition at line 326 of file `acsSCCanalysis.py`.

11.7.2.42 tuple `acsSCCanalysis.influx_rate = float(strLine[1])`

Definition at line 264 of file `acsSCCanalysis.py`.

11.7.2.43 int `acsSCCanalysis.initializeGraphStructure = 0`

Definition at line 189 of file `acsSCCanalysis.py`.

11.7.2.44 int `acsSCCanalysis.initRctId = int(tmpRctFileToLoadSplitString[len(tmpRctFileToLoadSplitString)-2])`

Definition at line 198 of file `acsSCCanalysis.py`.

11.7.2.45 list `acsSCCanalysis.initTempTime = tmpRctFileToLoadSplitString[len(tmpRctFileToLoadSplitString)-1]`

Definition at line 199 of file `acsSCCanalysis.py`.

11.7.2.46 int `acsSCCanalysis.initTime = float(initTempTime[0:len(initTempTime)-4])`

Definition at line 200 of file `acsSCCanalysis.py`.

11.7.2.47 int `acsSCCanalysis.inSCCFlag = 0`

Definition at line 616 of file `acsSCCanalysis.py`.

11.7.2.48 tuple `acsSCCanalysis.loadedMols = int(tmpLoadedMols)`

Definition at line 416 of file `acsSCCanalysis.py`.

11.7.2.49 `tuple acsSCCanalysis.loadedMolsConc = float(tmpLoadedMolsConc)`

Definition at line 415 of file acsSCCanalysis.py.

11.7.2.50 `tuple acsSCCanalysis.maxLOut = float(strLine[1])`

Definition at line 266 of file acsSCCanalysis.py.

11.7.2.51 `tuple acsSCCanalysis.meanOverThreshold = float(overThreshold)`

Definition at line 703 of file acsSCCanalysis.py.

11.7.2.52 `tuple acsSCCanalysis.mol_I = int(tmpMol_I)`

Definition at line 412 of file acsSCCanalysis.py.

11.7.2.53 `tuple acsSCCanalysis.mol_II = int(tmpMol_II)`

Definition at line 413 of file acsSCCanalysis.py.

11.7.2.54 `tuple acsSCCanalysis.mol_III = int(tmpMol_III)`

Definition at line 414 of file acsSCCanalysis.py.

11.7.2.55 `tuple acsSCCanalysis.mswindows = (sys.platform == "win32")`

Definition at line 179 of file acsSCCanalysis.py.

11.7.2.56 `tuple acsSCCanalysis.newdir = os.path.join(os.curdir, folderCat)`

Definition at line 356 of file acsSCCanalysis.py.

11.7.2.57 `tuple acsSCCanalysis.newdirSUB = os.path.join(os.curdir, folderSub)`

Definition at line 365 of file acsSCCanalysis.py.

11.7.2.58 `tuple acsSCCanalysis.newSpeciesCreationProb = float(tmpNSCprob)`

Definition at line 420 of file acsSCCanalysis.py.

11.7.2.59 `int acsSCCanalysis.nolnAcs = 1`

Definition at line 612 of file acsSCCanalysis.py.

11.7.2.60 `int acsSCCanalysis.nrg = 1`

Definition at line 273 of file acsSCCanalysis.py.

11.7.2.61 `tuple acsSCCanalysis.nrgConc = float(strLine[1])`

Definition at line 262 of file acsSCCanalysis.py.

11.7.2.62 `tuple acsSCCanalysis.nrgTimeSeries = np.array([[0,0,0]])`

Definition at line 397 of file acsSCCanalysis.py.

11.7.2.63 `tuple acsSCCanalysis.nrgType = int(strLine[1])`

Definition at line 258 of file acsSCCanalysis.py.

11.7.2.64 `tuple acsSCCanalysis.numberOfGen = len(glob.glob(os.path.join(resDirPath,'times_*')))`

Definition at line 230 of file acsSCCanalysis.py.

11.7.2.65 `int acsSCCanalysis.ok = 0`

Definition at line 316 of file acsSCCanalysis.py.

11.7.2.66 `tuple acsSCCanalysis.overThreshold = float(0)`

Definition at line 299 of file acsSCCanalysis.py.

11.7.2.67 `tuple acsSCCanalysis.overThresholdTOT = float(0)`

Definition at line 300 of file acsSCCanalysis.py.

11.7.2.68 `string acsSCCanalysis.paramFile = "acsm2s.conf"`

Definition at line 241 of file acsSCCanalysis.py.

11.7.2.69 `tuple acsSCCanalysis.position = ((graph[:,0] == cat) & (graph[:,1] == mol_l))`

Definition at line 482 of file acsSCCanalysis.py.

11.7.2.70 `acsSCCanalysis.previousTime = 0`

Definition at line 386 of file acsSCCanalysis.py.

11.7.2.71 `int acsSCCanalysis.printTemporalMessage = 1`

Definition at line 428 of file acsSCCanalysis.py.

11.7.2.72 `int acsSCCanalysis.prod_bySCC = 0`

Definition at line 570 of file acsSCCanalysis.py.

11.7.2.73 `int acsSCCanalysis.prod_bySCC_weight = 0`

Definition at line 576 of file acsSCCanalysis.py.

11.7.2.74 `int acsSCCanalysis.prod_chain = 0`

Definition at line 569 of file acsSCCanalysis.py.

11.7.2.75 `int acsSCCanalysis.prod_chain_weight = 0`

Definition at line 575 of file acsSCCanalysis.py.

11.7.2.76 `int acsSCCanalysis.prod_inSCC = 0`

Definition at line 568 of file acsSCCanalysis.py.

11.7.2.77 `int acsSCCanalysis.prod_inSCC_weight = 0`

Definition at line 574 of file acsSCCanalysis.py.

11.7.2.78 `int acsSCCanalysis.prod_overlap = 0`

Definition at line 571 of file acsSCCanalysis.py.

11.7.2.79 `int acsSCCanalysis.prod_overlap_weight = 0`

Definition at line 577 of file acsSCCanalysis.py.

11.7.2.80 `int acsSCCanalysis.rctID = initRctID`

Definition at line 385 of file acsSCCanalysis.py.

11.7.2.81 `int acsSCCanalysis.rctIDshow = 1.0`

Definition at line 383 of file acsSCCanalysis.py.

11.7.2.82 `int acsSCCanalysis.rctIDshowNoSave = 1.0`

Definition at line 384 of file acsSCCanalysis.py.

11.7.2.83 `tuple acsSCCanalysis.rctParamFile = sorted(glob.glob(os.path.join(resDirPath,rctParamFileQ)))`

Definition at line 380 of file acsSCCanalysis.py.

11.7.2.84 `string acsSCCanalysis.rctParamFileQ = 'reactions_parameters_'`

Definition at line 379 of file acsSCCanalysis.py.

11.7.2.85 `tuple acsSCCanalysis.reaction = int(tmpReaction)`

Definition at line 408 of file acsSCCanalysis.py.

11.7.2.86 `int acsSCCanalysis.realSccs = 0`

Definition at line 554 of file acsSCCanalysis.py.

11.7.2.87 `acsSCCanalysis.realT = threshold`

Definition at line 333 of file acsSCCanalysis.py.

11.7.2.88 `tuple acsSCCanalysis.resDirPath = os.path.abspath(os.path.join(".", "res"))`

Definition at line 223 of file acsSCCanalysis.py.

11.7.2.89 `tuple acsSCCanalysis.reverseProbability = float(tmpRevProb)`

Definition at line 421 of file acsSCCanalysis.py.

11.7.2.90 `tuple acsSCCanalysis.rp = float(strLine[1])`

Definition at line 252 of file acsSCCanalysis.py.

11.7.2.91 `tuple acsSCCanalysis.rtime = float(tmpTime)`

Definition at line 409 of file acsSCCanalysis.py.

11.7.2.92 `tuple acsSCCanalysis.scc = nx.strongly_connected_components(Gcatpro)`

Definition at line 561 of file acsSCCanalysis.py.

11.7.2.93 `int acsSCCanalysis.sccID = 0`

Definition at line 572 of file acsSCCanalysis.py.

11.7.2.94 `tuple acsSCCanalysis.sccN = nx.number_strongly_connected_components(Gcatpro)`

Definition at line 562 of file acsSCCanalysis.py.

11.7.2.95 `int acsSCCanalysis.self_loop_weight = 0`

Definition at line 578 of file acsSCCanalysis.py.

11.7.2.96 `tuple acsSCCanalysis.selfLoops = Gcatpro.number_of_selfloops()`

Definition at line 563 of file acsSCCanalysis.py.

11.7.2.97 `tuple acsSCCanalysis.selfLoopsEdges = Gcatpro.selfloop_edges()`

Definition at line 564 of file acsSCCanalysis.py.

11.7.2.98 `int acsSCCanalysis.simFolder = 0`

Definition at line 242 of file acsSCCanalysis.py.

11.7.2.99 `list acsSCCanalysis.speciesFile = speciesFiles[-1]`

Definition at line 305 of file acsSCCanalysis.py.

11.7.2.100 `tuple acsSCCanalysis.speciesFiles = sorted(glob.glob(os.path.join(resDirPath,tmpFilesToSearch)))`

Definition at line 304 of file acsSCCanalysis.py.

11.7.2.101 `list acsSCCanalysis.speciesInFlux = range(0,int(pow(2,(maxLOut+1)) - 2))`

Definition at line 280 of file acsSCCanalysis.py.

11.7.2.102 `tuple acsSCCanalysis.strLine = line.split('=')`

Definition at line 250 of file acsSCCanalysis.py.

11.7.2.103 `tuple acsSCCanalysis.StrPath = os.path.abspath(StrPath)`

Definition at line 177 of file acsSCCanalysis.py.

11.7.2.104 `string acsSCCanalysis.strToWrite = "\tReaction Probability"`

Definition at line 186 of file acsSCCanalysis.py.

11.7.2.105 `tuple acsSCCanalysis.strZeros = zeroBeforeStrNum(nGen, numberOfGen)`

Definition at line 238 of file acsSCCanalysis.py.

11.7.2.106 `int acsSCCanalysis.tempProd_chain_weight = 0`

Definition at line 607 of file acsSCCanalysis.py.

11.7.2.107 `acsSCCanalysis.timeInterval = rtime-previousTime`

Definition at line 451 of file acsSCCanalysis.py.

11.7.2.108 `tuple acsSCCanalysis.tmpDirs = sort(os.listdir(StrPath))`

Definition at line 209 of file acsSCCanalysis.py.

11.7.2.109 `string acsSCCanalysis.tmpFilesToSearch = 'species_'`

Definition at line 303 of file acsSCCanalysis.py.

11.7.2.110 `int acsSCCanalysis.tmpFlagLastRctSaved = 0`

Definition at line 401 of file acsSCCanalysis.py.

11.7.2.111 `int acsSCCanalysis.tmpProd_chain = 0`

Definition at line 603 of file acsSCCanalysis.py.

11.7.2.112 `tuple acsSCCanalysis.tmpRctFileToLoadSplitString = tmpRctFileToLoad.split("_")`

Definition at line 196 of file acsSCCanalysis.py.

11.7.2.113 `tuple acsSCCanalysis.today = dt.date.today()`

Definition at line 178 of file acsSCCanalysis.py.

11.7.2.114 `tuple acsSCCanalysis.totalRcts = int(strLine[1])`

Definition at line 260 of file acsSCCanalysis.py.

11.7.2.115 `tuple acsSCCanalysis.totDirName = os.path.join(StrPath,tmpDir)`

Definition at line 218 of file acsSCCanalysis.py.

11.7.2.116 `tuple acsSCCanalysis.totTimes = int(strLine[1])`

Definition at line 256 of file acsSCCanalysis.py.

11.7.2.117 `int acsSCCanalysis.wasteSpecies = 0`

Definition at line 584 of file acsSCCanalysis.py.

11.7.2.118 `list acsSCCanalysis.weightToDistribute = graph[((graph[:,0] == snglnNode) & (graph[:,1] == ldsOT)),5]`

Definition at line 622 of file acsSCCanalysis.py.

11.8 acsSpeciesActivities Namespace Reference

Functions

- def [zeroBeforeStrNum](#)

Variables

- tuple `StrPath` = `os.path.abspath(StrPath)`
- tuple `tmpDirs` = `sort(os.listdir(StrPath))`
- int `chemistry` = 1
- string `ndn` = `'_0_new_allStatResults'`
- tuple `newdirAllResults` = `os.path.join(StrPath, ndn)`
- tuple `newdirAllResultsInner` = `os.path.join(StrPath, '_0_new_allStatResults', ndn)`
- tuple `totDirName` = `os.path.join(StrPath, tmpDir)`
- tuple `resDirPath` = `os.path.abspath(os.path.join(".", "res"))`
- tuple `numberOfGen` = `len(glob.glob(os.path.join(resDirPath, 'times_*')))`
- string `tmpSpeciesStatsSummaryName` = `'speciesStatsSummary_'`
- tuple `tmpSpeciesStatsSummaryNameFID` = `open(tmpSpeciesStatsSummaryName, 'w')`
- string `tmpStr` = `'\n----- CHEMISTRY '`
- tuple `strZeros` = `zeroBeforeStrNum(nGen, numberOfGen)`
- string `strSpeciesZero` = `'species_'`
- tuple `speciesFilesZero` = `sorted(glob.glob(os.path.join(resDirPath, strSpeciesZero)))`
- string `strSpecies` = `'species_'`
- tuple `speciesFiles` = `sorted(glob.glob(os.path.join(resDirPath, strSpecies)))`
- list `lastfilespecies` = `speciesFiles[-1]`
- tuple `fidSpecies` = `open(lastfilespecies, 'r')`
- list `seq` = []
- tuple `counters` = `np.zeros((nSpecies, 3))`
- string `strRctPar` = `'reactions_parameters_'`
- tuple `fidRctPar` = `open(strRctPar, 'r')`
- tuple `rctType` = `int(tmpRctType)`
- tuple `cat` = `int(tmpCat)`
- tuple `S1` = `int(tmpS1)`
- tuple `S2` = `int(tmpS2)`
- tuple `S3` = `int(tmpS3)`
- string `tmpFileName` = `'speciesStats_'`
- tuple `tmpFileNameFID` = `open(tmpFileName, 'w')`
- int `ID` = 0

11.8.1 Function Documentation

11.8.1.1 `def acsSpeciesActivities.zeroBeforeStrNum (tmpl, tmpl)`

Definition at line 20 of file `acsSpeciesActivities.py`.

11.8.2 Variable Documentation

11.8.2.1 `tuple acsSpeciesActivities.cat = int(tmpCat)`

Definition at line 130 of file `acsSpeciesActivities.py`.

11.8.2.2 `int acsSpeciesActivities.chemistry = 1`

Definition at line 43 of file `acsSpeciesActivities.py`.

11.8.2.3 `tuple acsSpeciesActivities.counters = np.zeros((nSpecies, 3))`

Definition at line 116 of file `acsSpeciesActivities.py`.

11.8.2.4 `tuple acsSpeciesActivities.fidRctPar = open(strRctPar, 'r')`

Definition at line 122 of file acsSpeciesActivities.py.

11.8.2.5 `tuple acsSpeciesActivities.fidSpecies = open(lastfilespecies, 'r')`

Definition at line 105 of file acsSpeciesActivities.py.

11.8.2.6 `int acsSpeciesActivities.ID = 0`

Definition at line 162 of file acsSpeciesActivities.py.

11.8.2.7 `list acsSpeciesActivities.lastfilespecies = speciesFiles[-1]`

Definition at line 101 of file acsSpeciesActivities.py.

11.8.2.8 `string acsSpeciesActivities.ndn = '_0_new_allStatResults'`

Definition at line 46 of file acsSpeciesActivities.py.

11.8.2.9 `tuple acsSpeciesActivities.newdirAllResults = os.path.join(StrPath, ndn)`

Definition at line 47 of file acsSpeciesActivities.py.

11.8.2.10 `tuple acsSpeciesActivities.newdirAllResultsInner = os.path.join(StrPath, '_0_new_allStatResults', ndn)`

Definition at line 55 of file acsSpeciesActivities.py.

11.8.2.11 `tuple acsSpeciesActivities.numberOfGen = len(glob.glob(os.path.join(resDirPath, 'times_*')))`

Definition at line 74 of file acsSpeciesActivities.py.

11.8.2.12 `tuple acsSpeciesActivities.rctType = int(tmpRctType)`

Definition at line 129 of file acsSpeciesActivities.py.

11.8.2.13 `tuple acsSpeciesActivities.resDirPath = os.path.abspath(os.path.join("/", "res"))`

Definition at line 68 of file acsSpeciesActivities.py.

11.8.2.14 `tuple acsSpeciesActivities.S1 = int(tmpS1)`

Definition at line 131 of file acsSpeciesActivities.py.

11.8.2.15 `tuple acsSpeciesActivities.S2 = int(tmpS2)`

Definition at line 132 of file acsSpeciesActivities.py.

11.8.2.16 `tuple acsSpeciesActivities.S3 = int(tmpS3)`

Definition at line 133 of file `acsSpeciesActivities.py`.

11.8.2.17 `list acsSpeciesActivities.seq = []`

Definition at line 109 of file `acsSpeciesActivities.py`.

11.8.2.18 `tuple acsSpeciesActivities.speciesFiles = sorted(glob.glob(os.path.join(resDirPath,strSpecies)))`

Definition at line 98 of file `acsSpeciesActivities.py`.

11.8.2.19 `tuple acsSpeciesActivities.speciesFilesZero = sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))`

Definition at line 93 of file `acsSpeciesActivities.py`.

11.8.2.20 `tuple acsSpeciesActivities.StrPath = os.path.abspath(StrPath)`

Definition at line 36 of file `acsSpeciesActivities.py`.

11.8.2.21 `string acsSpeciesActivities.strRctPar = 'reactions_parameters_'`

Definition at line 118 of file `acsSpeciesActivities.py`.

11.8.2.22 `string acsSpeciesActivities.strSpecies = 'species_'`

Definition at line 95 of file `acsSpeciesActivities.py`.

11.8.2.23 `string acsSpeciesActivities.strSpeciesZero = 'species_'`

Definition at line 92 of file `acsSpeciesActivities.py`.

11.8.2.24 `tuple acsSpeciesActivities.strZeros = zeroBeforeStrNum(nngen, numberOfGen)`

Definition at line 90 of file `acsSpeciesActivities.py`.

11.8.2.25 `tuple acsSpeciesActivities.tmpDirs = sort(os.listdir(StrPath))`

Definition at line 38 of file `acsSpeciesActivities.py`.

11.8.2.26 `string acsSpeciesActivities.tmpFileName = 'speciesStats_'`

Definition at line 159 of file `acsSpeciesActivities.py`.

11.8.2.27 `tuple acsSpeciesActivities.tmpFileNameFID = open(tmpFileName, 'w')`

Definition at line 161 of file `acsSpeciesActivities.py`.

11.8.2.28 `string acsSpeciesActivities.tmpSpeciesStatsSummaryName = 'speciesStatsSummary_'`

Definition at line 79 of file `acsSpeciesActivities.py`.

11.8.2.29 `tuple acsSpeciesActivities.tmpSpeciesStatsSummaryNameFID = open(tmpSpeciesStatsSummaryName, 'w')`

Definition at line 80 of file `acsSpeciesActivities.py`.

11.8.2.30 `tuple acsSpeciesActivities.tmpStr = '\n----- CHEMISTRY '`

Definition at line 82 of file `acsSpeciesActivities.py`.

11.8.2.31 `tuple acsSpeciesActivities.totDirName = os.path.join(StrPath,tmpDir)`

Definition at line 64 of file `acsSpeciesActivities.py`.

11.9 acsStatesAnalysis Namespace Reference

Functions

- def [zeroBeforeStrNum](#)
- def [returnZeroSpeciesList](#)
- def [distanceMisures](#)

Variables

- tuple [today](#) = `dt.date.today()`
- tuple [StrPath](#) = `os.path.abspath(StrPath)`
- tuple [tmpDirs](#) = `sort(os.listdir(StrPath))`
- string [currentDir](#) = ""
- string [ndn](#) = `currentDir+'_0_new_allStatResults'`
- tuple [newdirAllResults](#) = `os.path.join(os.curdir, ndn)`
- tuple [previousFILE_FID](#) = `open('STAT_t_tminus_1.csv', 'w')`
- tuple [previousNOINFLUX_FILE_FID](#) = `open('STAT_t_tminus_1_NOINFLUX.csv', 'w')`
- tuple [startFILE_FID](#) = `open('STAT_t_start.csv', 'w')`
- tuple [startNOINFLUX_FILE_FID](#) = `open('STAT_t_start_NOINFLUX.csv', 'w')`
- tuple [HAM_previousFILE_FID](#) = `open('STAT_HAM_t_tminus_1.csv', 'w')`
- tuple [HAM_previousNOINFLUX_FILE_FID](#) = `open('STAT_HAM_t_tminus_1_NOINFLUX.csv', 'w')`
- tuple [HAM_startFILE_FID](#) = `open('STAT_HAM_t_start.csv', 'w')`
- tuple [HAM_startNOINFLUX_FILE_FID](#) = `open('STAT_HAM_t_start_NOINFLUX.csv', 'w')`
- tuple [EUC_previousFILE_FID](#) = `open('STAT_EUC_t_tminus_1.csv', 'w')`
- tuple [EUC_previousNOINFLUX_FILE_FID](#) = `open('STAT_EUC_t_tminus_1_NOINFLUX.csv', 'w')`
- tuple [EUC_startFILE_FID](#) = `open('STAT_EUC_t_start.csv', 'w')`
- tuple [EUC_startNOINFLUX_FILE_FID](#) = `open('STAT_EUC_t_start_NOINFLUX.csv', 'w')`
- tuple [ANG_middlePreviousFILE_FID](#) = `open('STAT_ANG_t_middle_NOINFLUX.csv', 'w')`
- tuple [HAM_middlePreviousFILE_FID](#) = `open('STAT_HAM_t_middle_NOINFLUX.csv', 'w')`
- tuple [EUC_middlePreviousFILE_FID](#) = `open('STAT_EUC_t_middle_NOINFLUX.csv', 'w')`
- tuple [previousFILE_FID_group](#) = `open('STAT_t_tminus_1_group.csv', 'w')`
- tuple [previousNOINFLUX_FILE_FID_group](#) = `open('STAT_t_tminus_1_NOINFLUX_group.csv', 'w')`
- tuple [startFILE_FID_group](#) = `open('STAT_t_start_group.csv', 'w')`
- tuple [startNOINFLUX_FILE_FID_group](#) = `open('STAT_t_start_NOINFLUX_group.csv', 'w')`

- tuple `HAM_previousFILE_FID_group` = open('STAT_HAM_t_tminus_1_group.csv', 'w')
- tuple `HAM_previousNOINFLUX_FILE_FID_group` = open('STAT_HAM_t_tminus_1_NOINFLUX_group.csv', 'w')
- tuple `HAM_startFILE_FID_group` = open('STAT_HAM_t_start_group.csv', 'w')
- tuple `HAM_startNOINFLUX_FILE_FID_group` = open('STAT_HAM_t_start_NOINFLUX_group.csv', 'w')
- tuple `EUC_previousFILE_FID_group` = open('STAT_EUC_t_tminus_1_group.csv', 'w')
- tuple `EUC_previousNOINFLUX_FILE_FID_group` = open('STAT_EUC_t_tminus_1_NOINFLUX_group.csv', 'w')
- tuple `EUC_startFILE_FID_group` = open('STAT_EUC_t_start_group.csv', 'w')
- tuple `EUC_startNOINFLUX_FILE_FID_group` = open('STAT_EUC_t_start_NOINFLUX_group.csv', 'w')
- tuple `newSpecies_FID` = open('STAT_GENERAL_newSpecies.csv', 'w')
- tuple `livingSpecies_FID` = open('STAT_GENERAL_livingSpecies.csv', 'w')
- tuple `mols_FID` = open('STAT_GENERAL_mols.csv', 'w')
- tuple `totMass_FID` = open('STAT_GENERAL_overallMass.csv', 'w')
- tuple `totOverallMass_FID` = open('STAT_GENERAL_overallTotMass.csv', 'w')
- tuple `complex_FID` = open('STAT_GENERAL_complex.csv', 'w')
- tuple `complexMols_FID` = open('STAT_GENERAL_complexMols.csv', 'w')
- tuple `evaluatedFID` = open('STAT_GENERAL_evaluated.csv', 'w')
- tuple `zeroOneSpeciesFID` = open('STAT_GENERAL_zeroOneSpecies.csv', 'w')
- tuple `biodeversityFID` = open('STAT_GENERAL_bioDiversity.csv', 'w')
- int `validDir` = 1
- tuple `totDirName` = os.path.join(StrPath,tmpDir)
- tuple `resDirPath` = os.path.abspath(os.path.join("./", "res"))
- tuple `numberOfGen` = len(glob.glob(os.path.join(resDirPath,'times_*')))
- list `group_A_prev` = []
- list `group_A_start` = []
- list `group_A_prev_NI` = []
- list `group_A_start_NI` = []
- tuple `strZeros` = zeroBeforeStrNum(ngen, numberOfGen)
- string `strSpeciesZero` = 'species_'
- tuple `speciesFilesZero` = sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))
- string `strSpecies` = 'species_'
- tuple `speciesFiles` = sorted(glob.glob(os.path.join(resDirPath,strSpecies)))
- tuple `zeroList` = returnZeroSpeciesList(speciesFiles[-1])
- tuple `speciesConcs` = np.zeros((len(speciesFiles)+1,len(zeroList)))
- list `seqOLD` = []
- list `seqSTART` = []
- list `totMass` = []
- list `seqMIDDLE_NOINFLUX` = []
- int `oldNumberOfSpecies` = 0
- tuple `fidSpecies` = open(sngSpeciesFile, 'r')
- list `seq` = []
- int `tmpMols` = 0
- int `bioDivInd` = 0
- `deltaNspecies` = numberOfSpecies-oldNumberOfSpecies
- tuple `strtoW` = str(deltaNspecies)
- tuple `tmpMisure` = distanceMisures(seq, conc, seqOLD, concOLD, idS)
- list `seqOLDNOINFLUX` = seqNOINFLUX[:]
- list `concOLD` = conc[:]
- string `filename` = "STAT_species_Concentrations_"

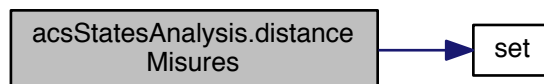
11.9.1 Function Documentation

11.9.1.1 `def acsStatesAnalysis.distanceMisures (tmpSeqX, tmpConcX, tmpSeqY, tmpConcY, tmpIDs)`

Function to compute the angle between two multidimensional vectors

Definition at line 45 of file `acsStatesAnalysis.py`.

Here is the call graph for this function:



11.9.1.2 `def acsStatesAnalysis.returnZeroSpeciesList (tmpLastSpeciesFile)`

Function to create a zero vector for each species (NO COMPLEXES)

Definition at line 31 of file `acsStatesAnalysis.py`.

11.9.1.3 `def acsStatesAnalysis.zeroBeforeStrNum (tmpI, tmpL)`

Function to create string zero string vector before graph filename.
According to the total number of reactions N zeros will be add before the instant reaction number
(e.g. reaction 130 of 10000 the string became '00130')

Definition at line 21 of file `acsStatesAnalysis.py`.

11.9.2 Variable Documentation

11.9.2.1 `tuple acsStatesAnalysis.ANG_middlePreviousFILE_FID = open('STAT_ANG_t_middle_NOINFLUX.csv', 'w')`

Definition at line 140 of file `acsStatesAnalysis.py`.

11.9.2.2 `tuple acsStatesAnalysis.biodeversityFID = open('STAT_GENERAL_bioDiversity.csv', 'w')`

Definition at line 168 of file `acsStatesAnalysis.py`.

11.9.2.3 `tuple acsStatesAnalysis.bioDivInd = 0`

Definition at line 273 of file `acsStatesAnalysis.py`.

11.9.2.4 `tuple acsStatesAnalysis.complex_FID = open('STAT_GENERAL_complex.csv', 'w')`

Definition at line 164 of file `acsStatesAnalysis.py`.

11.9.2.5 `tuple acsStatesAnalysis.complexMols_FID = open('STAT_GENERAL_complexMols.csv', 'w')`

Definition at line 165 of file `acsStatesAnalysis.py`.

11.9.2.6 `list acsStatesAnalysis.concOLD = conc[:]`

Definition at line 351 of file `acsStatesAnalysis.py`.

11.9.2.7 `string acsStatesAnalysis.currentDir = "`

Definition at line 113 of file `acsStatesAnalysis.py`.

11.9.2.8 `acsStatesAnalysis.deltaNspecies = numberOfSpecies-oldNumberOfSpecies`

Definition at line 279 of file `acsStatesAnalysis.py`.

11.9.2.9 `tuple acsStatesAnalysis.EUC_middlePreviousFILE_FID = open('STAT_EUC_t_middle_NOINFLUX.csv', 'w')`

Definition at line 142 of file `acsStatesAnalysis.py`.

11.9.2.10 `tuple acsStatesAnalysis.EUC_previousFILE_FID = open('STAT_EUC_t_tminus_1.csv', 'w')`

Definition at line 135 of file `acsStatesAnalysis.py`.

11.9.2.11 `tuple acsStatesAnalysis.EUC_previousFILE_FID_group = open('STAT_EUC_t_tminus_1_group.csv', 'w')`

Definition at line 154 of file `acsStatesAnalysis.py`.

11.9.2.12 `tuple acsStatesAnalysis.EUC_previousNOINFLUX_FILE_FID = open('STAT_EUC_t_tminus_1_NOINFLUX.csv', 'w')`

Definition at line 136 of file `acsStatesAnalysis.py`.

11.9.2.13 `tuple acsStatesAnalysis.EUC_previousNOINFLUX_FILE_FID_group = open('STAT_EUC_t_tminus_1_NOINFLUX_group.csv', 'w')`

Definition at line 155 of file `acsStatesAnalysis.py`.

11.9.2.14 `tuple acsStatesAnalysis.EUC_startFILE_FID = open('STAT_EUC_t_start.csv', 'w')`

Definition at line 137 of file `acsStatesAnalysis.py`.

11.9.2.15 `tuple acsStatesAnalysis.EUC_startFILE_FID_group = open('STAT_EUC_t_start_group.csv', 'w')`

Definition at line 156 of file `acsStatesAnalysis.py`.

11.9.2.16 `tuple acsStatesAnalysis.EUC_startNOINFLUX_FILE_FID = open('STAT_EUC_t_start_NOINFLUX.csv', 'w')`

Definition at line 138 of file `acsStatesAnalysis.py`.

11.9.2.17 `tuple acsStatesAnalysis.EUC_startNOINFLUX_FILE_FID_group = open('STAT_EUC_t_start_NOINFLUX_group.csv', 'w')`

Definition at line 157 of file `acsStatesAnalysis.py`.

11.9.2.18 `tuple acsStatesAnalysis.evaluatedFID = open('STAT_GENERAL_evaluated.csv', 'w')`

Definition at line 166 of file `acsStatesAnalysis.py`.

11.9.2.19 `tuple acsStatesAnalysis.fidSpecies = open(sngSpeciesFile, 'r')`

Definition at line 230 of file `acsStatesAnalysis.py`.

11.9.2.20 `string acsStatesAnalysis.filename = "STAT_species_Concentrations_"`

Definition at line 410 of file `acsStatesAnalysis.py`.

11.9.2.21 `list acsStatesAnalysis.group_A_prev = []`

Definition at line 186 of file `acsStatesAnalysis.py`.

11.9.2.22 `list acsStatesAnalysis.group_A_prev_NI = []`

Definition at line 188 of file `acsStatesAnalysis.py`.

11.9.2.23 `list acsStatesAnalysis.group_A_start = []`

Definition at line 187 of file `acsStatesAnalysis.py`.

11.9.2.24 `list acsStatesAnalysis.group_A_start_NI = []`

Definition at line 189 of file `acsStatesAnalysis.py`.

11.9.2.25 `tuple acsStatesAnalysis.HAM_middlePreviousFILE_FID = open('STAT_HAM_t_middle_NOINFLUX.csv', 'w')`

Definition at line 141 of file `acsStatesAnalysis.py`.

11.9.2.26 `tuple acsStatesAnalysis.HAM_previousFILE_FID = open('STAT_HAM_t_tminus_1.csv', 'w')`

Definition at line 130 of file `acsStatesAnalysis.py`.

11.9.2.27 `tuple acsStatesAnalysis.HAM_previousFILE_FID_group = open('STAT_HAM_t_tminus_1_group.csv', 'w')`

Definition at line 149 of file `acsStatesAnalysis.py`.

11.9.2.28 `tuple acsStatesAnalysis.HAM_previousNOINFLUX_FILE_FID = open('STAT_HAM_t_tminus_1_NOINFLUX.csv', 'w')`

Definition at line 131 of file `acsStatesAnalysis.py`.

11.9.2.29 `tuple acsStatesAnalysis.HAM_previousNOINFLUX_FILE_FID_group = open('STAT_HAM_t_tminus_1_NOINFLUX_group.csv', 'w')`

Definition at line 150 of file acsStatesAnalysis.py.

11.9.2.30 `tuple acsStatesAnalysis.HAM_startFILE_FID = open('STAT_HAM_t_start.csv', 'w')`

Definition at line 132 of file acsStatesAnalysis.py.

11.9.2.31 `tuple acsStatesAnalysis.HAM_startFILE_FID_group = open('STAT_HAM_t_start_group.csv', 'w')`

Definition at line 151 of file acsStatesAnalysis.py.

11.9.2.32 `tuple acsStatesAnalysis.HAM_startNOINFLUX_FILE_FID = open('STAT_HAM_t_start_NOINFLUX.csv', 'w')`

Definition at line 133 of file acsStatesAnalysis.py.

11.9.2.33 `tuple acsStatesAnalysis.HAM_startNOINFLUX_FILE_FID_group = open('STAT_HAM_t_start_NOINFLUX_group.csv', 'w')`

Definition at line 152 of file acsStatesAnalysis.py.

11.9.2.34 `tuple acsStatesAnalysis.livingSpecies_FID = open('STAT_GENERAL_livingSpecies.csv', 'w')`

Definition at line 160 of file acsStatesAnalysis.py.

11.9.2.35 `tuple acsStatesAnalysis.mols_FID = open('STAT_GENERAL_mols.csv', 'w')`

Definition at line 161 of file acsStatesAnalysis.py.

11.9.2.36 `string acsStatesAnalysis.ndn = currentDir+'_0_new_allStatResults'`

Definition at line 114 of file acsStatesAnalysis.py.

11.9.2.37 `tuple acsStatesAnalysis.newdirAllResults = os.path.join(os.curdir, ndn)`

Definition at line 115 of file acsStatesAnalysis.py.

11.9.2.38 `tuple acsStatesAnalysis.newSpecies_FID = open('STAT_GENERAL_newSpecies.csv', 'w')`

Definition at line 159 of file acsStatesAnalysis.py.

11.9.2.39 `tuple acsStatesAnalysis.numberOfGen = len(glob.glob(os.path.join(resDirPath, 'times_*')))`

Definition at line 184 of file acsStatesAnalysis.py.

11.9.2.40 `acsStatesAnalysis.oldNumberOfSpecies = 0`

Definition at line 222 of file acsStatesAnalysis.py.

11.9.2.41 `tuple acsStatesAnalysis.previousFILE_FID = open('STAT_t_tminus_1.csv', 'w')`

Definition at line 125 of file `acsStatesAnalysis.py`.

11.9.2.42 `tuple acsStatesAnalysis.previousFILE_FID_group = open('STAT_t_tminus_1_group.csv', 'w')`

Definition at line 144 of file `acsStatesAnalysis.py`.

11.9.2.43 `tuple acsStatesAnalysis.previousNOINFLUX_FILE_FID = open('STAT_t_tminus_1_NOINFLUX.csv', 'w')`

Definition at line 126 of file `acsStatesAnalysis.py`.

11.9.2.44 `tuple acsStatesAnalysis.previousNOINFLUX_FILE_FID_group = open('STAT_t_tminus_1_NOINFLUX_group.csv', 'w')`

Definition at line 145 of file `acsStatesAnalysis.py`.

11.9.2.45 `tuple acsStatesAnalysis.resDirPath = os.path.abspath(os.path.join("/", "res"))`

Definition at line 178 of file `acsStatesAnalysis.py`.

11.9.2.46 `list acsStatesAnalysis.seq = []`

Definition at line 234 of file `acsStatesAnalysis.py`.

11.9.2.47 `list acsStatesAnalysis.seqMIDDLE_NOINFLUX = []`

Definition at line 220 of file `acsStatesAnalysis.py`.

11.9.2.48 `list acsStatesAnalysis.seqOLD = []`

Definition at line 217 of file `acsStatesAnalysis.py`.

11.9.2.49 `list acsStatesAnalysis.seqOLDNOINFLUX = seqNOINFLUX[:]`

Definition at line 350 of file `acsStatesAnalysis.py`.

11.9.2.50 `list acsStatesAnalysis.seqSTART = []`

Definition at line 218 of file `acsStatesAnalysis.py`.

11.9.2.51 `list acsStatesAnalysis.speciesConcs = np.zeros((len(speciesFiles)+1,len(zeroList)))`

Definition at line 214 of file `acsStatesAnalysis.py`.

11.9.2.52 `acsStatesAnalysis.speciesFiles = sorted(glob.glob(os.path.join(resDirPath,strSpecies)))`

Definition at line 204 of file `acsStatesAnalysis.py`.

11.9.2.53 `tuple acsStatesAnalysis.speciesFilesZero = sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))`

Definition at line 199 of file acsStatesAnalysis.py.

11.9.2.54 `tuple acsStatesAnalysis.startFILE_FID = open('STAT_t_start.csv', 'w')`

Definition at line 127 of file acsStatesAnalysis.py.

11.9.2.55 `tuple acsStatesAnalysis.startFILE_FID_group = open('STAT_t_start_group.csv', 'w')`

Definition at line 146 of file acsStatesAnalysis.py.

11.9.2.56 `tuple acsStatesAnalysis.startNOINFLUX_FILE_FID = open('STAT_t_start_NOINFLUX.csv', 'w')`

Definition at line 128 of file acsStatesAnalysis.py.

11.9.2.57 `tuple acsStatesAnalysis.startNOINFLUX_FILE_FID_group = open('STAT_t_start_NOINFLUX_group.csv', 'w')`

Definition at line 147 of file acsStatesAnalysis.py.

11.9.2.58 `tuple acsStatesAnalysis.StrPath = os.path.abspath(StrPath)`

Definition at line 101 of file acsStatesAnalysis.py.

11.9.2.59 `string acsStatesAnalysis.strSpecies = 'species_'`

Definition at line 201 of file acsStatesAnalysis.py.

11.9.2.60 `string acsStatesAnalysis.strSpeciesZero = 'species_'`

Definition at line 198 of file acsStatesAnalysis.py.

11.9.2.61 `tuple acsStatesAnalysis.strtoW = str(deltaNspecies)`

Definition at line 281 of file acsStatesAnalysis.py.

11.9.2.62 `tuple acsStatesAnalysis.strZeros = zeroBeforeStrNum(nngen, numberOfGen)`

Definition at line 195 of file acsStatesAnalysis.py.

11.9.2.63 `tuple acsStatesAnalysis.tmpDirs = sort(os.listdir(StrPath))`

Definition at line 103 of file acsStatesAnalysis.py.

11.9.2.64 `tuple acsStatesAnalysis.tmpMisure = distanceMisures(seq, conc, seqOLD, concOLD, idS)`

Definition at line 302 of file acsStatesAnalysis.py.

11.9.2.65 `int acsStatesAnalysis.tmpMols = 0`

Definition at line 235 of file `acsStatesAnalysis.py`.

11.9.2.66 `tuple acsStatesAnalysis.today = dt.date.today()`

Definition at line 99 of file `acsStatesAnalysis.py`.

11.9.2.67 `tuple acsStatesAnalysis.totDirName = os.path.join(StrPath,tmpDir)`

Definition at line 174 of file `acsStatesAnalysis.py`.

11.9.2.68 `list acsStatesAnalysis.totMass = []`

Definition at line 219 of file `acsStatesAnalysis.py`.

11.9.2.69 `tuple acsStatesAnalysis.totMass_FID = open('STAT_GENERAL_overallMass.csv', 'w')`

Definition at line 162 of file `acsStatesAnalysis.py`.

11.9.2.70 `tuple acsStatesAnalysis.totOverallMass_FID = open('STAT_GENERAL_overallTotMass.csv', 'w')`

Definition at line 163 of file `acsStatesAnalysis.py`.

11.9.2.71 `int acsStatesAnalysis.validDir = 1`

Definition at line 171 of file `acsStatesAnalysis.py`.

11.9.2.72 `tuple acsStatesAnalysis.zeroList = returnZeroSpeciesList(speciesFiles[-1])`

Definition at line 207 of file `acsStatesAnalysis.py`.

11.9.2.73 `tuple acsStatesAnalysis.zeroOneSpeciesFID = open('STAT_GENERAL_zeroOneSpecies.csv', 'w')`

Definition at line 167 of file `acsStatesAnalysis.py`.

11.10 bufferedFluxAnalysis Namespace Reference

Variables

- list `StrPath` = `sys.argv[1]`
- list `tmpResFold` = `sys.argv[2]`
- tuple `today` = `dt.date.today()`
- string `simF` = `StrPath+'/'`
- tuple `tmpDirs` = `sort(glob.glob(simF))`
- tuple `newdir` = `os.path.join(os.getcwd(), '0_statistics')`
- tuple `matrixTimeLITE` = `np.zeros((101,len(tmpDirs)))`
- tuple `matrixFluxLITE` = `np.zeros((101,len(tmpDirs)))`
- tuple `matrixAbsLITE` = `np.zeros((101,len(tmpDirs)))`

- tuple `matrixExpLITE` = `np.zeros((101,len(tmpDirs)))`
- int `tmpDirsCnt` = 0
- tuple `speciesFiles` = `sort(glob.glob('species_*'))`
- tuple `fidSpecies` = `open(speciesFiles[0], 'r')`
- int `ok` = 0
- list `fluxIndexes` = []
- list `fluxLengths` = []
- tuple `index` = `int(tmpID)`
- tuple `speciesSeq` = `len(tmpSeq)`
- tuple `concFixed` = `int(tmpConcFixed)`
- tuple `fileslist` = `sort(glob.glob('reactions_parameters_*'))`
- int `rctParFileNum` = 1
- int `rctID` = 1
- list `totFluxDyn` = []
- list `absorbedBricks` = []
- int `tempAbsorbedBricks` = 0
- list `expelledBricks` = []
- int `tempExpelledBricks` = 0
- list `totTimes` = []
- list `totFluxDyn_LITE` = []
- list `absorbedBricks_LITE` = []
- list `expelledBricks_LITE` = []
- list `totTimes_LITE` = []
- tuple `fid` = `open(file, 'r')`
- int `okmonitor` = 1
- int `oksaveLite` = 0
- int `deltaRct` = 0
- tuple `reaction` = `int(tmpReaction)`
- tuple `rtime` = `float(tmpTime)`
- tuple `cc` = `int(tmpcc)`
- tuple `cat` = `int(tmpCat)`
- tuple `mol_I` = `int(tmpMol_I)`
- tuple `mol_II` = `int(tmpMol_II)`
- tuple `mol_III` = `int(tmpMol_III)`
- tuple `loadedMolsConc` = `float(tmpLoadedMolsConc)`
- tuple `loadedMols` = `int(tmpLoadedMols)`
- tuple `gillMean` = `float(tmpGillMean)`
- tuple `gillSD` = `float(tmpGillSD)`
- tuple `gillEntropy` = `float(tmpGillEntropy)`
- tuple `savingMatrix` = `np.zeros((len(totTimes),4))`
- tuple `tmpDirSplit` = `tmpDir.split("/")`
- string `cmdFileName` = `StrPath+'0_statistics/0_fluxDynamics_'`
- tuple `cmdFileFid` = `open(cmdFileName, 'a')`

11.10.1 Variable Documentation

11.10.1.1 list `bufferedFluxAnalysis.absorbedBricks` = []

Definition at line 99 of file `bufferedFluxAnalysis.py`.

11.10.1.2 list `bufferedFluxAnalysis.absorbedBricks_LITE` = []

Definition at line 107 of file `bufferedFluxAnalysis.py`.

11.10.1.3 `tuple bufferedFluxAnalysis.cat = int(tmpCat)`

Definition at line 131 of file bufferedFluxAnalysis.py.

11.10.1.4 `tuple bufferedFluxAnalysis.cc = int(tmpcc)`

Definition at line 130 of file bufferedFluxAnalysis.py.

11.10.1.5 `tuple bufferedFluxAnalysis.cmdFileFid = open(cmdFileName, 'a')`

Definition at line 206 of file bufferedFluxAnalysis.py.

11.10.1.6 `string bufferedFluxAnalysis.cmdFileName = StrPath+'0_statistics/0_fluxDynamics_'`

Definition at line 203 of file bufferedFluxAnalysis.py.

11.10.1.7 `tuple bufferedFluxAnalysis.concFixed = int(tmpConcFixed)`

Definition at line 85 of file bufferedFluxAnalysis.py.

11.10.1.8 `list bufferedFluxAnalysis.deltaRct = 0`

Definition at line 123 of file bufferedFluxAnalysis.py.

11.10.1.9 `list bufferedFluxAnalysis.expelledBricks = []`

Definition at line 101 of file bufferedFluxAnalysis.py.

11.10.1.10 `list bufferedFluxAnalysis.expelledBricks_LITE = []`

Definition at line 108 of file bufferedFluxAnalysis.py.

11.10.1.11 `tuple bufferedFluxAnalysis.fid = open(file, 'r')`

Definition at line 116 of file bufferedFluxAnalysis.py.

11.10.1.12 `tuple bufferedFluxAnalysis.fidSpecies = open(speciesFiles[0], 'r')`

Definition at line 70 of file bufferedFluxAnalysis.py.

11.10.1.13 `tuple bufferedFluxAnalysis.fileslist = sort(glob.glob('reactions_parameters_*'))`

Definition at line 94 of file bufferedFluxAnalysis.py.

11.10.1.14 `list bufferedFluxAnalysis.fluxIndexes = []`

Definition at line 76 of file bufferedFluxAnalysis.py.

11.10.1.15 `list bufferedFluxAnalysis.fluxLengths = []`

Definition at line 77 of file bufferedFluxAnalysis.py.

11.10.1.16 `tuple bufferedFluxAnalysis.gillEntropy = float(tmpGillEntropy)`

Definition at line 139 of file bufferedFluxAnalysis.py.

11.10.1.17 `tuple bufferedFluxAnalysis.gillMean = float(tmpGillMean)`

Definition at line 137 of file bufferedFluxAnalysis.py.

11.10.1.18 `tuple bufferedFluxAnalysis.gillSD = float(tmpGillSD)`

Definition at line 138 of file bufferedFluxAnalysis.py.

11.10.1.19 `tuple bufferedFluxAnalysis.index = int(tmpID)`

Definition at line 83 of file bufferedFluxAnalysis.py.

11.10.1.20 `tuple bufferedFluxAnalysis.loadedMols = int(tmpLoadedMols)`

Definition at line 136 of file bufferedFluxAnalysis.py.

11.10.1.21 `tuple bufferedFluxAnalysis.loadedMolsConc = float(tmpLoadedMolsConc)`

Definition at line 135 of file bufferedFluxAnalysis.py.

11.10.1.22 `tuple bufferedFluxAnalysis.matrixAbsLITE = np.zeros((101,len(tmpDirs)))`

Definition at line 48 of file bufferedFluxAnalysis.py.

11.10.1.23 `tuple bufferedFluxAnalysis.matrixExpLITE = np.zeros((101,len(tmpDirs)))`

Definition at line 49 of file bufferedFluxAnalysis.py.

11.10.1.24 `tuple bufferedFluxAnalysis.matrixFluxLITE = np.zeros((101,len(tmpDirs)))`

Definition at line 47 of file bufferedFluxAnalysis.py.

11.10.1.25 `tuple bufferedFluxAnalysis.matrixTimeLITE = np.zeros((101,len(tmpDirs)))`

Definition at line 46 of file bufferedFluxAnalysis.py.

11.10.1.26 `tuple bufferedFluxAnalysis.mol_I = int(tmpMol_I)`

Definition at line 132 of file bufferedFluxAnalysis.py.

11.10.1.27 `tuple bufferedFluxAnalysis.mol_II = int(tmpMol_II)`

Definition at line 133 of file bufferedFluxAnalysis.py.

11.10.1.28 `tuple bufferedFluxAnalysis.mol_III = int(tmpMol_III)`

Definition at line 134 of file bufferedFluxAnalysis.py.

11.10.1.29 `tuple bufferedFluxAnalysis.newdir = os.path.join(os.curdir, '0_statistics')`

Definition at line 39 of file bufferedFluxAnalysis.py.

11.10.1.30 `int bufferedFluxAnalysis.ok = 0`

Definition at line 75 of file bufferedFluxAnalysis.py.

11.10.1.31 `int bufferedFluxAnalysis.okmonitor = 1`

Definition at line 118 of file bufferedFluxAnalysis.py.

11.10.1.32 `int bufferedFluxAnalysis.oksaveLite = 0`

Definition at line 119 of file bufferedFluxAnalysis.py.

11.10.1.33 `int bufferedFluxAnalysis.rctID = 1`

Definition at line 96 of file bufferedFluxAnalysis.py.

11.10.1.34 `int bufferedFluxAnalysis.rctParFileNum = 1`

Definition at line 95 of file bufferedFluxAnalysis.py.

11.10.1.35 `tuple bufferedFluxAnalysis.reaction = int(tmpReaction)`

Definition at line 128 of file bufferedFluxAnalysis.py.

11.10.1.36 `tuple bufferedFluxAnalysis.rtime = float(tmpTime)`

Definition at line 129 of file bufferedFluxAnalysis.py.

11.10.1.37 `tuple bufferedFluxAnalysis.savingMatrix = np.zeros((len(totTimes),4))`

Definition at line 195 of file bufferedFluxAnalysis.py.

11.10.1.38 `string bufferedFluxAnalysis.simF = StrPath+'/'`

Definition at line 31 of file bufferedFluxAnalysis.py.

11.10.1.39 `tuple bufferedFluxAnalysis.speciesFiles = sort(glob.glob('species_*'))`

Definition at line 64 of file bufferedFluxAnalysis.py.

11.10.1.40 `tuple bufferedFluxAnalysis.speciesSeq = len(tmpSeq)`

Definition at line 84 of file bufferedFluxAnalysis.py.

11.10.1.41 `list bufferedFluxAnalysis.StrPath = sys.argv[1]`

Definition at line 18 of file bufferedFluxAnalysis.py.

11.10.1.42 `list bufferedFluxAnalysis.tempAbsorbedBricks = 0`

Definition at line 100 of file bufferedFluxAnalysis.py.

11.10.1.43 `list bufferedFluxAnalysis.tempExpelledBricks = 0`

Definition at line 102 of file bufferedFluxAnalysis.py.

11.10.1.44 `tuple bufferedFluxAnalysis.tmpDirs = sort(glob.glob(simF))`

Definition at line 32 of file bufferedFluxAnalysis.py.

11.10.1.45 `int bufferedFluxAnalysis.tmpDirsCnt = 0`

Definition at line 51 of file bufferedFluxAnalysis.py.

11.10.1.46 `tuple bufferedFluxAnalysis.tmpDirSplit = tmpDir.split("/")`

Definition at line 202 of file bufferedFluxAnalysis.py.

11.10.1.47 `list bufferedFluxAnalysis.tmpResFold = sys.argv[2]`

Definition at line 19 of file bufferedFluxAnalysis.py.

11.10.1.48 `tuple bufferedFluxAnalysis.today = dt.date.today()`

Definition at line 25 of file bufferedFluxAnalysis.py.

11.10.1.49 `list bufferedFluxAnalysis.totFluxDyn = []`

Definition at line 98 of file bufferedFluxAnalysis.py.

11.10.1.50 `list bufferedFluxAnalysis.totFluxDyn_LITE = []`

Definition at line 106 of file bufferedFluxAnalysis.py.

11.10.1.51 `list bufferedFluxAnalysis.totTimes = []`

Definition at line 103 of file bufferedFluxAnalysis.py.

11.10.1.52 `list bufferedFluxAnalysis.totTimes_LITE = []`

Definition at line 109 of file bufferedFluxAnalysis.py.

11.11 fromWithin2Between Namespace Reference

Functions

- def [zeroBeforeStrNum](#)

Variables

- tuple [zeroSIM](#) = [zeroBeforeStrNum](#)(i,numSim)
- tuple [zeroGEN](#) = [zeroBeforeStrNum](#)(j,numGen)
- string [folderNew](#) = "s_"
- tuple [resdir](#) = os.path.join(os.curdir, "res")

11.11.1 Function Documentation

11.11.1.1 `def fromWithin2Between.zeroBeforeStrNum (tpl, tplL)`

Definition at line 10 of file fromWithin2Between.py.

11.11.2 Variable Documentation

11.11.2.1 `string fromWithin2Between.folderNew = "s_"`

Definition at line 41 of file fromWithin2Between.py.

11.11.2.2 `tuple fromWithin2Between.resdir = os.path.join(os.curdir, "res")`

Definition at line 52 of file fromWithin2Between.py.

11.11.2.3 `tuple fromWithin2Between.zeroGEN = zeroBeforeStrNum(j,numGen)`

Definition at line 39 of file fromWithin2Between.py.

11.11.2.4 `tuple fromWithin2Between.zeroSIM = zeroBeforeStrNum(i,numSim)`

Definition at line 35 of file fromWithin2Between.py.

11.12 init Namespace Reference

Variables

- tuple `parser`
- tuple `args` = `parser.parse_args()`
- string `ndn` = `'_0_new_allStatResults'`
- tuple `newdirAllResults` = `os.path.join(args.strOut, ndn)`
- tuple `fname_initRafRes` = `os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`
- tuple `fname_initRafResSUM` = `os.path.join(newdirAllResults, '0_initRafAnalysisSUM.csv')`
- tuple `fname_initRafResLIST` = `os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`
- tuple `fname_initRafResALL` = `os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`
- tuple `fid_initRafRes` = `open(fname_initRafRes, 'w')`
- tuple `fid_initRafResSUM` = `open(fname_initRafResSUM, 'w')`
- tuple `fid_initRafResLIST` = `open(fname_initRafResLIST, 'w')`
- tuple `fid_initRafResALL` = `open(fname_initRafResALL, 'w')`
- string `strToWrite` = `"Folder\\tP\\tAC\\tM\\tRAFsize\\tClosure\\tCats\\tuRAF\\n"`
- tuple `foodList` = `range(args.lastFood+1)`
- tuple `avgCon` = `dn.rangeFloat(float(args.avgCon[0]), float(args.avgCon[1]), float(args.avgCon[2]))`
- int `raffound` = 0
- list `alphabet` = `['A', 'B']`
- list `species` = []
- int `totSpecies` = 2
- tuple `totCleavage` = `sum(map(lambda x: len(x)-1, species))`
- int `totCond` = `totSpecies**2`
- `totRcts` = `totCleavage+totCond`
- tuple `rctToCat` = `int(round(totRcts * totSpecies * prob))`
- int `nCleavage` = 0
- int `nCondensa` = 0
- tuple `initSpeciesListLength` = `len(species)`
- tuple `conf` = `(1,1,2000,0,200000,0,0,2,args.lastFood,prob)`
- int `rctType` = 1
- tuple `molToCleav` = `ran.choice(species[len(alphabet):initSpeciesListLength-1])`
- tuple `cutPt` = `ran.randint(1,len(molToCleav)-1)`
- list `tmp1` = `molToCleav[0:cutPt]`
- tuple `tmp1id` = `species.index(tmp1)`
- `find1` = True
- list `tmp2` = `molToCleav[cutPt:len(molToCleav)]`
- tuple `tmp2id` = `species.index(tmp2)`
- tuple `sub1` = `ran.choice(species[:initSpeciesListLength-1])`
- tuple `idsub1` = `species.index(sub1)`
- tuple `sub2` = `ran.choice(species[:initSpeciesListLength-1])`
- tuple `idsub2` = `species.index(sub2)`
- `prod` = `sub1+sub2`
- tuple `tmpprodid` = `species.index(prod)`
- int `catalyst` = -1
- `catFound` = False
- tuple `rafsets` = `raf.rafComputation(fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, 'tmpDir', prob, averageConn, rcts, cats, foodList, maxlength)`

11.12.1 Variable Documentation

11.12.1.1 list `init.alphabet` = `['A', 'B']`

Definition at line 79 of file `init.py`.

11.12.1.2 `tuple init.args = parser.parse_args()`

Definition at line 39 of file `init.py`.

11.12.1.3 `tuple init.avgCon = dn.rangeFloat(float(args.avgCon[0]), float(args.avgCon[1]), float(args.avgCon[2]))`

Definition at line 72 of file `init.py`.

11.12.1.4 `tuple init.catalyst = -1`

Definition at line 158 of file `init.py`.

11.12.1.5 `init.catFound = False`

Definition at line 159 of file `init.py`.

11.12.1.6 `tuple init.conf = (1,1,2000,0,200000,0,0,2,args.lastFood,prob)`

Definition at line 104 of file `init.py`.

11.12.1.7 `tuple init.cutPt = ran.randint(1,len(molToCleav)-1)`

Definition at line 121 of file `init.py`.

11.12.1.8 `tuple init.fid_initRafRes = open(fname_initRafRes, 'w')`

Definition at line 59 of file `init.py`.

11.12.1.9 `tuple init.fid_initRafResALL = open(fname_initRafResALL, 'w')`

Definition at line 62 of file `init.py`.

11.12.1.10 `tuple init.fid_initRafResLIST = open(fname_initRafResLIST, 'w')`

Definition at line 61 of file `init.py`.

11.12.1.11 `tuple init.fid_initRafResSUM = open(fname_initRafResSUM, 'w')`

Definition at line 60 of file `init.py`.

11.12.1.12 `init.find1 = True`

Definition at line 125 of file `init.py`.

11.12.1.13 `tuple init.fname_initRafRes = os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`

Definition at line 55 of file `init.py`.

11.12.1.14 `tuple init.fname_initRafResALL = os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`

Definition at line 58 of file `init.py`.

11.12.1.15 `tuple init.fname_initRafResLIST = os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`

Definition at line 57 of file `init.py`.

11.12.1.16 `tuple init.fname_initRafResSUM = os.path.join(newdirAllResults, '0_initRafAnalysisSUM.csv')`

Definition at line 56 of file `init.py`.

11.12.1.17 `tuple init.foodList = range(args.lastFood+1)`

Definition at line 69 of file `init.py`.

11.12.1.18 `tuple init.idsub1 = species.index(sub1)`

Definition at line 143 of file `init.py`.

11.12.1.19 `tuple init.idsub2 = species.index(sub2)`

Definition at line 146 of file `init.py`.

11.12.1.20 `tuple init.initSpeciesListLength = len(species)`

Definition at line 101 of file `init.py`.

11.12.1.21 `tuple init.molToCleav = ran.choice(species[len(alphabet):initSpeciesListLength-1])`

Definition at line 120 of file `init.py`.

11.12.1.22 `int init.nCleavage = 0`

Definition at line 99 of file `init.py`.

11.12.1.23 `int init.nCondensa = 0`

Definition at line 100 of file `init.py`.

11.12.1.24 `string init.ndn = '_0_new_allStatResults'`

Definition at line 45 of file `init.py`.

11.12.1.25 `tuple init.newdirAllResults = os.path.join(args.strOut, ndn)`

Definition at line 46 of file `init.py`.

11.12.1.26 tuple init.parser**Initial value:**

```

1 = ArgumentParser(
2     angle analysis in time.' description='This script re-arrange results in a more friendly way from the
3     , epilog='''File with angle trajectories are created. '''

```

Definition at line 23 of file init.py.

11.12.1.27 init.prod = sub1+sub2

Definition at line 147 of file init.py.

11.12.1.28 int init.raffound = 0

Definition at line 76 of file init.py.

11.12.1.29 tuple init.rafsets = raf.rafComputation(fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, 'tmpDir', prob, averageConn, rcts, cats, foodList, maxlength)

Definition at line 167 of file init.py.

11.12.1.30 tuple init.rctToCat = int(round(totRcts * totSpecies * prob))

Definition at line 96 of file init.py.

11.12.1.31 int init.rctType = 1

Definition at line 110 of file init.py.

11.12.1.32 list init.species = []

Definition at line 80 of file init.py.

11.12.1.33 tuple init.strToWrite = "Folder\tP\tAC\tM\tRAFsize\tClosure\tCats\ttuRAF\n"

Definition at line 63 of file init.py.

11.12.1.34 tuple init.sub1 = ran.choice(species[:initSpeciesListLength-1])

Definition at line 142 of file init.py.

11.12.1.35 tuple init.sub2 = ran.choice(species[:initSpeciesListLength-1])

Definition at line 145 of file init.py.

11.12.1.36 list init.tmp1 = molToCleav[0:cutPt]

Definition at line 122 of file init.py.

11.12.1.37 `tuple init.tmp1id = species.index(tmp1)`

Definition at line 124 of file `init.py`.

11.12.1.38 `list init.tmp2 = molToCleav[cutPt:len(molToCleav)]`

Definition at line 130 of file `init.py`.

11.12.1.39 `tuple init.tmp2id = species.index(tmp2)`

Definition at line 132 of file `init.py`.

11.12.1.40 `tuple init.tmpprodid = species.index(prod)`

Definition at line 149 of file `init.py`.

11.12.1.41 `tuple init.totCleavage = sum(map(lambda x: len(x)-1,species))`

Definition at line 86 of file `init.py`.

11.12.1.42 `int init.totCond = totSpecies**2`

Definition at line 87 of file `init.py`.

11.12.1.43 `init.totRcts = totCleavage+totCond`

Definition at line 88 of file `init.py`.

11.12.1.44 `int init.totSpecies = 2`

Definition at line 84 of file `init.py`.

11.13 lib Namespace Reference

Namespaces

- [dyn](#)
- [graph](#)
- [IO](#)

11.14 lib.dyn Namespace Reference

Namespaces

- [dynamics](#)

11.15 lib.dyn.dynamics Namespace Reference

Functions

- def [generateFluxList](#)
- def [rangeFloat](#)
- def [fluxAnalysis](#)

11.15.1 Function Documentation

11.15.1.1 `def lib.dyn.dynamics.fluxAnalysis (tmpDir, resDirPath, strZeros, ngen)`

Definition at line 37 of file dynamics.py.

11.15.1.2 `def lib.dyn.dynamics.generateFluxList (tmpPath, tmpSysType, tmpLastID=None)`

Definition at line 17 of file dynamics.py.

11.15.1.3 `def lib.dyn.dynamics.rangeFloat (start, step, stop)`

Definition at line 31 of file dynamics.py.

11.16 lib.graph Namespace Reference

Namespaces

- [network](#)
- [raf](#)

11.17 lib.graph.network Namespace Reference

Functions

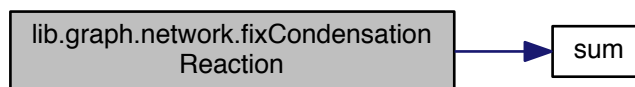
- def [removeRareRcts](#)
- def [fixCondensationReaction](#)

11.17.1 Function Documentation

11.17.1.1 `def lib.graph.network.fixCondensationReaction (m1, m2, m3, rcts)`

Definition at line 25 of file network.py.

Here is the call graph for this function:



11.17.1.2 `def lib.graph.network.removeRareRcts (graph, dt, life, nrg, deltat)`

Definition at line 16 of file network.py.

11.18 lib.graph.raf Namespace Reference

Functions

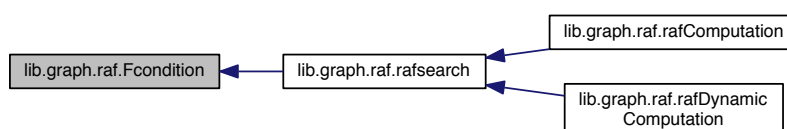
- `def generateClosure`
- `def RAcondition`
- `def Fcondition`
- `def findCatforRAF`
- `def rafsearch`
- `def rafComputation`
- `def rafDynamicComputation`
- `def findRAFrcts`

11.18.1 Function Documentation

11.18.1.1 `def lib.graph.raf.Fcondition (tmpCL, tmpRA, rcts, debug=False)`

Definition at line 60 of file raf.py.

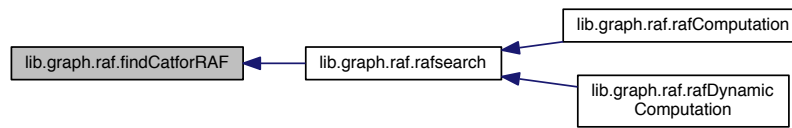
Here is the caller graph for this function:



11.18.1.2 `def lib.graph.raf.findCatforRAF (tmpCat, tmpRAF, tmpClosure, debug=False)`

Definition at line 82 of file raf.py.

Here is the caller graph for this function:



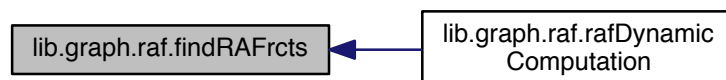
11.18.1.3 `def lib.graph.raf.findRAFrcts (RAF, rcts, actrcts)`

Definition at line 160 of file `raf.py`.

Here is the call graph for this function:



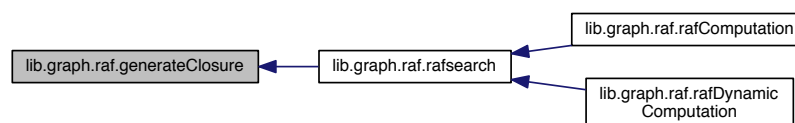
Here is the caller graph for this function:



11.18.1.4 `def lib.graph.raf.generateClosure (tmpF, rcts, debug=False)`

Definition at line 20 of file `raf.py`.

Here is the caller graph for this function:



11.18.1.5 `def lib.graph.raf.RAcondition (tmpCL, rcts, cats, debug=False)`

Definition at line 46 of file raf.py.

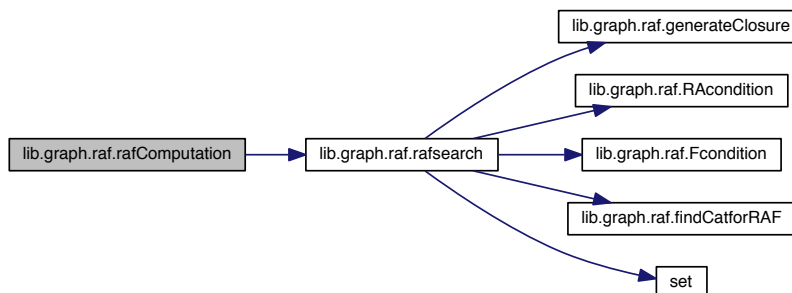
Here is the caller graph for this function:



11.18.1.6 `def lib.graph.raf.rafComputation (fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, tmpDir, rctProb, avgCon, rcts, cats, foodList, maxDim, debug=False)`

Definition at line 134 of file raf.py.

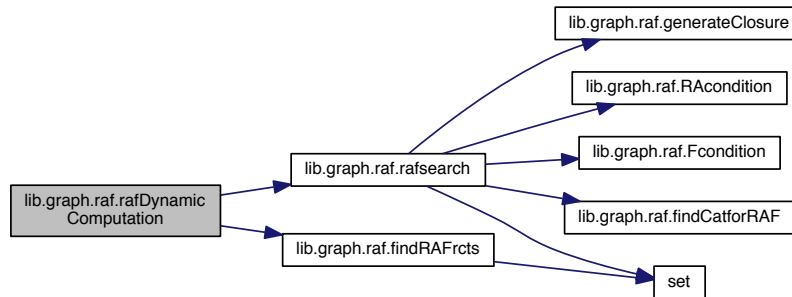
Here is the call graph for this function:



11.18.1.7 `def lib.graph.raf.rafDynamicComputation (fid_dynRafRes, tmpTime, rcts, cats, foodList, growth=False, rctsALL=None, catsALL=None, completeRCTS=None, debug=False)`

Definition at line 144 of file raf.py.

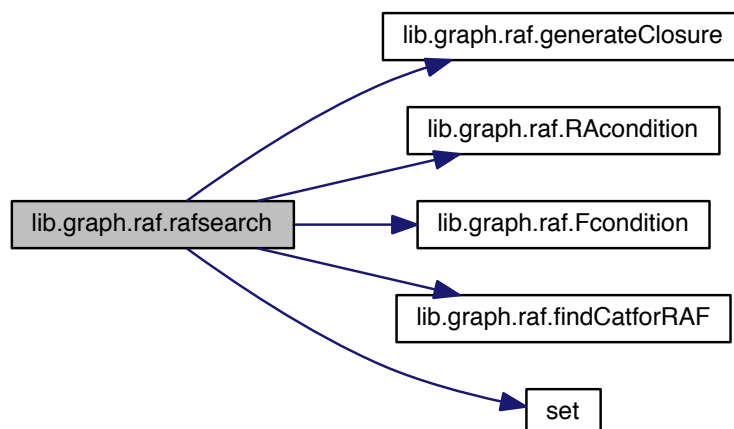
Here is the call graph for this function:



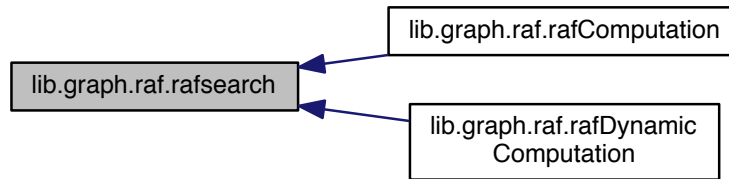
11.18.1.8 `def lib.graph.raf.rafsearch (rcts, cats, closure, debug = False)`

Definition at line 91 of file `raf.py`.

Here is the call graph for this function:



Here is the caller graph for this function:



11.19 lib.IO Namespace Reference

Namespaces

- [readfiles](#)
- [writefiles](#)

11.20 lib.IO.readfiles Namespace Reference

Functions

- [def readConfFile](#)
- [def readInitConfFile](#)
- [def readBufferedID](#)
- [def readCSTRflux](#)
- [def loadAllData](#)
- [def zeroBeforeStrNum](#)
- [def splitRctParsLine](#)

11.20.1 Function Documentation

11.20.1.1 `def lib.IO.readfiles.loadAllData (tmpPath, tmpFname)`

Definition at line 120 of file `readfiles.py`.

11.20.1.2 `def lib.IO.readfiles.readBufferedID (tmpPath)`

Definition at line 89 of file `readfiles.py`.

11.20.1.3 `def lib.IO.readfiles.readConfFile (tmpPath)`

Definition at line 14 of file `readfiles.py`.

11.20.1.4 `def lib.IO.readfiles.readCSTRflux (tmpPath)`

Definition at line 105 of file `readfiles.py`.

11.20.1.5 `def lib.IO.readfiles.readInitConfFile (tmpPath)`

Definition at line 52 of file readfiles.py.

11.20.1.6 `def lib.IO.readfiles.splitRctParsLine (tmpLine)`

Definition at line 135 of file readfiles.py.

11.20.1.7 `def lib.IO.readfiles.zeroBeforeStrNum (tmpl, tmpL)`

Definition at line 127 of file readfiles.py.

11.21 lib.IO.writefiles Namespace Reference

Functions

- `def write_init_raf_list`
- `def write_init_raf_all`

11.21.1 Function Documentation

11.21.1.1 `def lib.IO.writefiles.write_init_raf_all (fid, rafinfo, folder, rcts, cats)`

Definition at line 23 of file writefiles.py.

11.21.1.2 `def lib.IO.writefiles.write_init_raf_list (fid, rafinfo, folder)`

Definition at line 14 of file writefiles.py.

11.22 main Namespace Reference

Variables

- tuple `parser`
- tuple `args` = `parser.parse_args()`
- tuple `strPath` = `os.path.abspath(args.strPath)`
- tuple `tmpDirs` = `sort(os.listdir(strPath))`
- string `ndn` = `'_0_new_allStatResults'`
- tuple `newdirAllResults` = `os.path.join(strPath, ndn)`
- int `_CLOSE_` = 0
- int `_PROTO_` = 1
- int `_CSTR_` = 2
- tuple `fname_initRafRes` = `os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`
- tuple `fname_initRafResLIST` = `os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`
- tuple `fname_initRafResALL` = `os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`
- tuple `fid_initRafRes` = `open(fname_initRafRes, 'w')`
- tuple `fid_initRafResLIST` = `open(fname_initRafResLIST, 'w')`
- tuple `fid_initRafResALL` = `open(fname_initRafResALL, 'w')`
- string `strToWrite` = `"Folder\tP\tAC\tM\tM\tRAFSsize\tClosureSize\tCatsSize\tuRAF\n"`
- tuple `totDirName` = `os.path.join(strPath,tmpDir)`

- tuple `resDirPath` = `os.path.abspath(os.path.join("./", args.resFolder))`
- tuple `conf` = `readfiles.readConfFile(totDirName)`
- tuple `foodList` = `dm.generateFluxList(totDirName, sysType)`
- tuple `rcts` = `readfiles.loadAllData(totDirName, '_acsreactions.csv')`
- tuple `cats` = `readfiles.loadAllData(totDirName, '_acscatalysis.csv')`
- tuple `numberOfGen` = `len(glob.glob(os.path.join(resDirPath, 'times_*')))`
- tuple `strZeros` = `readfiles.zeroBeforeStrNum(nGen, numberOfGen)`
- string `fName` = `'RAF_structuresInTime_analysis_gen_'`
- tuple `fname_inTimeRafRes` = `os.path.join(newdirAllResults, fName)`
- tuple `fid_inTimeRafRes` = `open(fname_inTimeRafRes, 'w')`
- `potential` = `False`
- string `strRctZero` = `'reactions_'`
- string `strCatZero` = `'catalysis_'`
- tuple `rctFilesZero` = `sorted(glob.glob(os.path.join(resDirPath, strRctZero)))`
- tuple `catFilesZero` = `sorted(glob.glob(os.path.join(resDirPath, strCatZero)))`
- string `strRct` = `'reactions_'`
- string `strCat` = `'catalysis_'`
- tuple `rctFiles` = `sorted(glob.glob(os.path.join(resDirPath, strRct)))`
- tuple `catFiles` = `sorted(glob.glob(os.path.join(resDirPath, strCat)))`
- list `sngTime` = `conf[2]`
- int `actTime` = `0`
- list `procrcts` = `rcts[rcts[:,5] > 0,:]`
- list `proccats` = `cats[cats[:,3] > 0,:]`
- tuple `R` = `raf.rafDynamicComputation(fid_inTimeRafRes, actTime, procrcts[:,0:5], proccats[:,0:5], foodList, potential, rcts, cats, debug=args.debug)`
- tuple `lastRct` = `readfiles.loadAllData(totDirName, rctFiles[-1])`
- tuple `lastCat` = `readfiles.loadAllData(totDirName, catFiles[-1])`
- tuple `fname_dynRafRes` = `os.path.join(newdirAllResults, fName)`
- tuple `fid_dynRafRes` = `open(fname_dynRafRes, 'w')`
- string `strRctPar` = `'reactions_parameters_'`
- tuple `rctParamFile` = `sorted(glob.glob(os.path.join(resDirPath, strRctPar)))`
- tuple `fid` = `open(rctParamFile[0], 'r')`
- int `previousTime` = `0`
- `decayTime` = `args.decay`
- int `condensation_counter` = `0`
- int `endo_condensation_counter` = `0`
- int `cleavage_counter` = `0`
- int `endo_cleavage_counter` = `0`
- int `nAnal` = `1`
- int `rctCurrID` = `0`
- int `catCurrID` = `0`
- `timeInterval` = `rtime-previousTime`
- tuple `graph` = `network.removeRareRcts(graph, 2, 3, 4, timeInterval)`
- tuple `graphSUB` = `network.removeRareRcts(graphSUB, 2, 3, 4, timeInterval)`
- tuple `onrcts` = `network.removeRareRcts(onrcts, 5, 6, 7, timeInterval)`
- tuple `oncats` = `network.removeRareRcts(oncats, 3, 4, 5, timeInterval)`
- tuple `positionR` = `((onrcts[:,1] == cc) & (onrcts[:,2] == mol_I) & (onrcts[:,3] == mol_II))`
- tuple `position` = `((oncats[:,1] == cat) & (oncats[:,2] == onrcts[positionR,0]))`

11.22.1 Variable Documentation

11.22.1.1 int main._CLOSE_ = 0

Definition at line 57 of file main.py.

11.22.1.2 `int main._CSTR_ = 2`

Definition at line 59 of file main.py.

11.22.1.3 `int main._PROTO_ = 1`

Definition at line 58 of file main.py.

11.22.1.4 `int main.actTime = 0`

Definition at line 144 of file main.py.

11.22.1.5 `tuple main.args = parser.parse_args()`

Definition at line 40 of file main.py.

11.22.1.6 `int main.catCurID = 0`

Definition at line 191 of file main.py.

11.22.1.7 `tuple main.catFiles = sorted(glob.glob(os.path.join(resDirPath,strCat)))`

Definition at line 137 of file main.py.

11.22.1.8 `tuple main.catFilesZero = sorted(glob.glob(os.path.join(resDirPath,strCatZero)))`

Definition at line 132 of file main.py.

11.22.1.9 `tuple main.cats = readfiles.loadAllData(totDirName,'_acscatalysis.csv')`

Definition at line 95 of file main.py.

11.22.1.10 `int main.cleavage_counter = 0`

Definition at line 187 of file main.py.

11.22.1.11 `int main.condensation_counter = 0`

Definition at line 185 of file main.py.

11.22.1.12 `tuple main.conf = readfiles.readConfFile(totDirName)`

Definition at line 84 of file main.py.

11.22.1.13 `main.decayTime = args.decay`

Definition at line 184 of file main.py.

11.22.1.14 `int main.endo_cleavage_counter = 0`

Definition at line 188 of file main.py.

11.22.1.15 `int main.endo_condensation_counter = 0`

Definition at line 186 of file main.py.

11.22.1.16 `tuple main.fid = open(rctParamFile[0], 'r')`

Definition at line 182 of file main.py.

11.22.1.17 `tuple main.fid_dynRafRes = open(fname_dynRafRes, 'w')`

Definition at line 174 of file main.py.

11.22.1.18 `tuple main.fid_initRafRes = open(fname_initRafRes, 'w')`

Definition at line 65 of file main.py.

11.22.1.19 `tuple main.fid_initRafResALL = open(fname_initRafResALL, 'w')`

Definition at line 67 of file main.py.

11.22.1.20 `tuple main.fid_initRafResLIST = open(fname_initRafResLIST, 'w')`

Definition at line 66 of file main.py.

11.22.1.21 `tuple main.fid_inTimeRafRes = open(fname_inTimeRafRes, 'w')`

Definition at line 119 of file main.py.

11.22.1.22 `string main.fName = 'RAF_structuresInTime_analysis_gen_'`

Definition at line 117 of file main.py.

11.22.1.23 `tuple main.fname_dynRafRes = os.path.join(newdirAllResults, fName)`

Definition at line 173 of file main.py.

11.22.1.24 `tuple main.fname_initRafRes = os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`

Definition at line 62 of file main.py.

11.22.1.25 `tuple main.fname_initRafResALL = os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`

Definition at line 64 of file main.py.

11.22.1.26 `tuple main.fname_initRafResLIST = os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`

Definition at line 63 of file main.py.

11.22.1.27 `tuple main.fname_inTimeRafRes = os.path.join(newdirAllResults, fName)`

Definition at line 118 of file main.py.

11.22.1.28 `tuple main.foodList = dm.generateFluxList(totDirName, sysType)`

Definition at line 89 of file main.py.

11.22.1.29 `tuple main.graph = network.removeRareRcts(graph,2,3,4,timeInterval)`

Definition at line 205 of file main.py.

11.22.1.30 `tuple main.graphSUB = network.removeRareRcts(graphSUB,2,3,4,timeInterval)`

Definition at line 206 of file main.py.

11.22.1.31 `tuple main.lastCat = readfiles.loadAllData(totDirName,catFiles[-1])`

Definition at line 166 of file main.py.

11.22.1.32 `tuple main.lastRct = readfiles.loadAllData(totDirName,rctFiles[-1])`

Definition at line 165 of file main.py.

11.22.1.33 `int main.nAnal = 1`

Definition at line 189 of file main.py.

11.22.1.34 `string main.ndn = '_0_new_allStatResults'`

Definition at line 50 of file main.py.

11.22.1.35 `tuple main.newdirAllResults = os.path.join(strPath, ndn)`

Definition at line 51 of file main.py.

11.22.1.36 `tuple main.numberOfGen = len(glob.glob(os.path.join(resDirPath,'times_*')))`

Definition at line 102 of file main.py.

11.22.1.37 `tuple main.oncats = network.removeRareRcts(oncats,3,4,5,timeInterval)`

Definition at line 208 of file main.py.

11.22.1.38 tuple main.onrcts = network.removeRareRcts(onrcts,5,6,7,timeInterval)

Definition at line 207 of file main.py.

11.22.1.39 tuple main.parser

Initial value:

```
1 = ArgumentParser(
2                                     description='Main script of ACS analysis.'
3                                     , epilog='''ACS ANALYSIS Main File. ''')
```

Definition at line 28 of file main.py.

11.22.1.40 tuple main.position = ((oncats[:,1] == cat) & (oncats[:,2] == onrcts[positionR,0]))

Definition at line 233 of file main.py.

11.22.1.41 list main.positionR = ((onrcts[:,1] == cc) & (onrcts[:,2] == mol_I) & (onrcts[:,3] == mol_II))

Definition at line 221 of file main.py.

11.22.1.42 main.potential = False

Definition at line 122 of file main.py.

11.22.1.43 main.previousTime = 0

Definition at line 183 of file main.py.

11.22.1.44 list main.proccats = cats[cats[:,3] > 0,:]

Definition at line 153 of file main.py.

11.22.1.45 list main.procrcts = rcts[rcts[:,5] > 0,:]

Definition at line 152 of file main.py.

11.22.1.46 tuple main.R = raf.rafDynamicComputation(fid_inTimeRafRes, actTime, procrcts[:,0:5], proccats[:,0:5], foodList, potential, rcts, cats, debug=args.debug)

Definition at line 154 of file main.py.

11.22.1.47 int main.rctCurrID = 0

Definition at line 190 of file main.py.

11.22.1.48 tuple main.rctFiles = sorted(glob.glob(os.path.join(resDirPath,strRct)))

Definition at line 136 of file main.py.

11.22.1.49 `tuple main.rctFilesZero = sorted(glob.glob(os.path.join(resDirPath, strRctZero)))`

Definition at line 131 of file main.py.

11.22.1.50 `tuple main.rctParamFile = sorted(glob.glob(os.path.join(resDirPath, strRctPar)))`

Definition at line 181 of file main.py.

11.22.1.51 `tuple main.rcts = readfiles.loadAllData(totDirName, '_acsreactions.csv')`

Definition at line 94 of file main.py.

11.22.1.52 `tuple main.resDirPath = os.path.abspath(os.path.join(".", args.resFolder))`

Definition at line 79 of file main.py.

11.22.1.53 `list main.sngTime = conf[2]`

Definition at line 143 of file main.py.

11.22.1.54 `string main.strCat = 'catalysis_'`

Definition at line 134 of file main.py.

11.22.1.55 `string main.strCatZero = 'catalysis_'`

Definition at line 130 of file main.py.

11.22.1.56 `tuple main.strPath = os.path.abspath(args.strPath)`

Definition at line 43 of file main.py.

11.22.1.57 `string main.strRct = 'reactions_'`

Definition at line 133 of file main.py.

11.22.1.58 `string main.strRctPar = 'reactions_parameters_'`

Definition at line 180 of file main.py.

11.22.1.59 `string main.strRctZero = 'reactions_'`

Definition at line 129 of file main.py.

11.22.1.60 `string main.strToWrite = "Folder\tP\tAC\tM\tRAFsize\tClosureSize\tCatsSize\tuRAF\n"`

Definition at line 68 of file main.py.

11.22.1.61 `tuple main.strZeros = readfiles.zeroBeforeStrNum(ngen, numberOfGen)`

Definition at line 111 of file main.py.

11.22.1.62 `main.timeInterval = rtime-previousTime`

Definition at line 200 of file main.py.

11.22.1.63 `tuple main.tmpDirs = sort(os.listdir(strPath))`

Definition at line 44 of file main.py.

11.22.1.64 `tuple main.totDirName = os.path.join(strPath,tmpDir)`

Definition at line 73 of file main.py.

11.23 prepareNewSim Namespace Reference

Functions

- def [zeroBeforeStrNum](#)

Variables

- tuple [parser](#)
- tuple [args](#) = `parser.parse_args()`
- tuple [StrFrom](#) = `os.path.abspath(args.StrFrom)`
- tuple [StrTo](#) = `os.path.abspath(args.StrTo)`
- tuple [StrFileSpeciesToGetConc](#) = `os.path.abspath(args.FileSpeciesToGetConc)`
- tuple [origin](#) = `os.getcwd()`
- int [_LASTSPECIES_](#) = 29
- tuple [_REVRCTS_](#) = `int(args.revRct)`
- tuple [_RATIOREV_](#) = `int(args.k_revRct)`
- float [_CLEAVAGE_](#) = 25.0
- float [_CONDENSATION_](#) = 50.0
- float [_COMPLEXFORM_](#) = 50.0
- tuple [_COMPLEXDISS_](#) = `float(args.kDiss)`
- tuple [_INITSPECIESCONC_](#) = `float(args.singleInitConc)`
- tuple [fileDest](#) = `os.path.join(StrTo,"_acsinflux.csv")`
- tuple [sourceResFolder](#) = `os.path.join(StrFrom,"res")`
- tuple [lastSpeciesFile](#) = `sorted(glob.glob('species_*'))`
- tuple [lastReactionsFile](#) = `sorted(glob.glob('reactions_1*'))`
- tuple [lastCatalysisFile](#) = `sorted(glob.glob('catalysis_*'))`
- tuple [mod](#) = `open("acsm2s.conf")`
- int [id](#) = 0
- tuple [linesplitted](#) = `line.split("=")`
- tuple [file](#) = `open("acsm2s.conf", "w")`
- list [concs](#) = []
- tuple [specFileLines](#) = `open(StrFileSpeciesToGetConc)`
- tuple [mod_rct](#) = `open("_acsreactions.csv")`
- int [flag](#) = 0
- tuple [catRct](#) = `linecache.getline('_acsreactions.csv', int(linesplitted[2])+1)`
- tuple [carRctSplit](#) = `catRct.split("\t")`

11.23.1 Function Documentation

11.23.1.1 `def prepareNewSim.zeroBeforeStrNum (tmpl, tmpL)`

Definition at line 12 of file `prepareNewSim.py`.

11.23.2 Variable Documentation

11.23.2.1 `float prepareNewSim._CLEAVAGE_ = 25.0`

Definition at line 51 of file `prepareNewSim.py`.

11.23.2.2 `tuple prepareNewSim._COMPLEXDISS_ = float(args.kDiss)`

Definition at line 54 of file `prepareNewSim.py`.

11.23.2.3 `float prepareNewSim._COMPLEXFORM_ = 50.0`

Definition at line 53 of file `prepareNewSim.py`.

11.23.2.4 `float prepareNewSim._CONDENSATION_ = 50.0`

Definition at line 52 of file `prepareNewSim.py`.

11.23.2.5 `tuple prepareNewSim._INITSPECIESCONC_ = float(args.singleInitConc)`

Definition at line 55 of file `prepareNewSim.py`.

11.23.2.6 `int prepareNewSim._LASTSPECIES_ = 29`

Definition at line 48 of file `prepareNewSim.py`.

11.23.2.7 `tuple prepareNewSim._RATIOREV_ = int(args.k_revRct)`

Definition at line 50 of file `prepareNewSim.py`.

11.23.2.8 `tuple prepareNewSim._REVRCTS_ = int(args.revRct)`

Definition at line 49 of file `prepareNewSim.py`.

11.23.2.9 `tuple prepareNewSim.args = parser.parse_args()`

Definition at line 36 of file `prepareNewSim.py`.

11.23.2.10 `tuple prepareNewSim.carRctSplit = catRct.split("\t")`

Definition at line 217 of file `prepareNewSim.py`.

11.23.2.11 `tuple prepareNewSim.catRct = linecache.getline('_acsreactions.csv', int(linesplitted[2])+1)`

Definition at line 216 of file prepareNewSim.py.

11.23.2.12 `list prepareNewSim.concs = []`

Definition at line 142 of file prepareNewSim.py.

11.23.2.13 `tuple prepareNewSim.file = open("acsm2s.conf", "w")`

Definition at line 131 of file prepareNewSim.py.

11.23.2.14 `tuple prepareNewSim.fileDest = os.path.join(StrTo, "_acsinflux.csv")`

Definition at line 61 of file prepareNewSim.py.

11.23.2.15 `int prepareNewSim.flag = 0`

Definition at line 211 of file prepareNewSim.py.

11.23.2.16 `int prepareNewSim.id = 0`

Definition at line 100 of file prepareNewSim.py.

11.23.2.17 `tuple prepareNewSim.lastCatalysisFile = sorted(glob.glob('catalysis_*'))`

Definition at line 74 of file prepareNewSim.py.

11.23.2.18 `tuple prepareNewSim.lastReactionsFile = sorted(glob.glob('reactions_1*'))`

Definition at line 73 of file prepareNewSim.py.

11.23.2.19 `tuple prepareNewSim.lastSpeciesFile = sorted(glob.glob('species_*'))`

Definition at line 71 of file prepareNewSim.py.

11.23.2.20 `tuple prepareNewSim.linesplitted = line.split("=")`

Definition at line 103 of file prepareNewSim.py.

11.23.2.21 `tuple prepareNewSim.mod = open("acsm2s.conf")`

Definition at line 99 of file prepareNewSim.py.

11.23.2.22 `tuple prepareNewSim.mod_rct = open("_acsreactions.csv")`

Definition at line 208 of file prepareNewSim.py.

11.23.2.23 tuple prepareNewSim.origin = os.getcwd()

Definition at line 47 of file prepareNewSim.py.

11.23.2.24 tuple prepareNewSim.parser

Initial value:

```
1 = ArgumentParser(
2     description='Script to create new init simulation files starting from the
   end of a previous simulation.'
3     , epilog='''Species concentration are initilized according to the sequence
   uploaded, pay attention to the file selected. '''
)
```

Definition at line 22 of file prepareNewSim.py.

11.23.2.25 tuple prepareNewSim.sourceResFolder = os.path.join(StrFrom,"res")

Definition at line 67 of file prepareNewSim.py.

11.23.2.26 tuple prepareNewSim.specFileLines = open(StrFileSpeciesToGetConc)

Definition at line 143 of file prepareNewSim.py.

11.23.2.27 tuple prepareNewSim.StrFileSpeciesToGetConc = os.path.abspath(args.FileSpeciesToGetConc)

Definition at line 41 of file prepareNewSim.py.

11.23.2.28 tuple prepareNewSim.StrFrom = os.path.abspath(args.StrFrom)

Definition at line 39 of file prepareNewSim.py.

11.23.2.29 tuple prepareNewSim.StrTo = os.path.abspath(args.StrTo)

Definition at line 40 of file prepareNewSim.py.

11.24 resetForNewSimulations Namespace Reference

Functions

- def [zeroBeforeStrNum](#)

Variables

- int [foldersSIMS](#) = 10
- int [foldersREP](#) = 10
- tuple [zerosSIMS](#) = [zeroBeforeStrNum](#)(i,foldersSIMS)
- tuple [zerosREPS](#) = [zeroBeforeStrNum](#)(j,foldersREP)
- string [folderName](#) = "s_inv_1e-2_"
- string [folderNew](#) = "s_inv_1e-1_"
- tuple [resdir](#) = os.path.join(os.curdir, "res")

- tuple `crtSimFolder` = `os.path.join(StrTo, folderName)`
- tuple `fileDest` = `os.path.join(StrTo, folderNew, "_acsinflux.csv")`
- tuple `speciesFiles` = `sorted(glob.glob("species_1_*"))`
- tuple `mod` = `open("_acsspecies.csv")`
- int `id` = 0
- tuple `linesplitted` = `line.split("\t")`
- tuple `file` = `open("_acsspecies.csv", "w")`

11.24.1 Function Documentation

11.24.1.1 `def resetForNewSimulations.zeroBeforeStrNum (tmpl, tmpL)`

Definition at line 5 of file `resetForNewSimulations.py`.

11.24.2 Variable Documentation

11.24.2.1 `tuple resetForNewSimulations.crtSimFolder = os.path.join(StrTo, folderName)`

Definition at line 51 of file `resetForNewSimulations.py`.

11.24.2.2 `tuple resetForNewSimulations.file = open("_acsspecies.csv", "w")`

Definition at line 99 of file `resetForNewSimulations.py`.

11.24.2.3 `tuple resetForNewSimulations.fileDest = os.path.join(StrTo, folderNew, "_acsinflux.csv")`

Definition at line 57 of file `resetForNewSimulations.py`.

11.24.2.4 `string resetForNewSimulations.folderName = "s_inv_1e-2_"`

Definition at line 31 of file `resetForNewSimulations.py`.

11.24.2.5 `string resetForNewSimulations.folderNew = "s_inv_1e-1_"`

Definition at line 32 of file `resetForNewSimulations.py`.

11.24.2.6 `int resetForNewSimulations.foldersREP = 10`

Definition at line 22 of file `resetForNewSimulations.py`.

11.24.2.7 `int resetForNewSimulations.foldersSIMS = 10`

Definition at line 21 of file `resetForNewSimulations.py`.

11.24.2.8 `int resetForNewSimulations.id = 0`

Definition at line 83 of file `resetForNewSimulations.py`.

11.24.2.9 tuple `resetForNewSimulations.linesplitted = line.split("\t")`

Definition at line 85 of file `resetForNewSimulations.py`.

11.24.2.10 tuple `resetForNewSimulations.mod = open("_acsspecies.csv")`

Definition at line 82 of file `resetForNewSimulations.py`.

11.24.2.11 tuple `resetForNewSimulations.resdir = os.path.join(os.curdir, "res")`

Definition at line 42 of file `resetForNewSimulations.py`.

11.24.2.12 tuple `resetForNewSimulations.speciesFiles = sorted(glob.glob("species_1_*"))`

Definition at line 66 of file `resetForNewSimulations.py`.

11.24.2.13 tuple `resetForNewSimulations.zerosREPS = zeroBeforeStrNum(j,foldersREP)`

Definition at line 29 of file `resetForNewSimulations.py`.

11.24.2.14 tuple `resetForNewSimulations.zerosSIMS = zeroBeforeStrNum(i,foldersSIMS)`

Definition at line 28 of file `resetForNewSimulations.py`.

Chapter 12

Class Documentation

12.1 catalysis Class Reference

CATALYSIS class.

```
#include <catalysis.h>
```

Public Member Functions

- [catalysis](#) ()
- [catalysis](#) ([acs_longInt](#) tmpCatId, [acs_longInt](#) tmpCat, [acs_longInt](#) tmpRctId, [acs_longInt](#) tmpAmount, [acs_double](#) tmpKass, [acs_double](#) tmpKdiss, [acs_double](#) tmpK_cpx, [acs_int](#) tmpCpxTarget)
- [~catalysis](#) ()
- [acs_longInt](#) getCatId () const
- [acs_longInt](#) getCat () const
- [acs_longInt](#) getReactionID () const
- [acs_longInt](#) getTotAmount () const
- [acs_double](#) getKass () const
- [acs_double](#) getKdiss () const
- [acs_double](#) getK_cpx () const
- [acs_int](#) getCpxTarget () const
- void [updateTotAmount](#) ()
- void [resetEventsCounter](#) ()

12.1.1 Detailed Description

CATALYSIS class.

This class contains catalysis proprieties and methods

Author

Alessandro Filisetti

Version

0.1

Date

2009-04-16

Definition at line 16 of file catalysis.h.

12.1.2 Constructor & Destructor Documentation

12.1.2.1 catalysis::catalysis ()

12.1.2.2 catalysis::catalysis (*acs_longInt tmpCatId*, *acs_longInt tmpCat*, *acs_longInt tmpRctId*, *acs_longInt tmpAmount*, *acs_double tmpKass*, *acs_double tmpKdiss*, *acs_double tmpK_cpx*, *acs_int tmpCpxTarget*)

catalysis class constructor (FROM FILE)

Version

0.1

Date

2010-03-16

Definition at line 19 of file catalysis.cpp.

Here is the call graph for this function:



12.1.2.3 catalysis::~~catalysis () [inline]

Definition at line 35 of file catalysis.h.

12.1.3 Member Function Documentation

12.1.3.1 *acs_longInt* catalysis::getCat () const [inline]

Definition at line 39 of file catalysis.h.

12.1.3.2 *acs_longInt* catalysis::getCatId () const [inline]

Definition at line 38 of file catalysis.h.

12.1.3.3 *acs_int* catalysis::getCpxTarget () const [inline]

Definition at line 46 of file catalysis.h.

12.1.3.4 *acs_double* catalysis::getK_cpx () const [inline]

Definition at line 44 of file catalysis.h.

12.1.3.5 **acs_double** catalysis::getKass () const [inline]

Definition at line 42 of file catalysis.h.

12.1.3.6 **acs_double** catalysis::getKdiss () const [inline]

Definition at line 43 of file catalysis.h.

12.1.3.7 **acs_longInt** catalysis::getReactionID () const [inline]

Definition at line 40 of file catalysis.h.

12.1.3.8 **acs_longInt** catalysis::getTotAmount () const [inline]

Definition at line 41 of file catalysis.h.

12.1.3.9 **void** catalysis::resetEventsCounter () [inline]

Definition at line 50 of file catalysis.h.

12.1.3.10 **void** catalysis::updateTotAmount () [inline]

Definition at line 49 of file catalysis.h.

The documentation for this class was generated from the following files:

- /Users/alessandrofilisetti/Documents/GIT/carness/[catalysis.h](#)
- /Users/alessandrofilisetti/Documents/GIT/carness/[catalysis.cpp](#)

12.2 commonFunctions Class Reference

This class contains all the common function of the system.

```
#include <commonFunctions.h>
```

12.2.1 Detailed Description

This class contains all the common function of the system.

This class contains all the functions useful in general

Authors

alessandro filisetti

Date

2011/12/10

Version

1.0

The documentation for this class was generated from the following file:

- /Users/alessandrofilisetti/Documents/GIT/carness/[commonFunctions.h](#)

12.3 environment Class Reference

environment class

```
#include <environment.h>
```

Public Member Functions

- [environment](#) (string tmpInitialPath)
- [~environment](#) ()
- [acs_int getNgen](#) () const
- [acs_int getNsim](#) () const
- [acs_double getActualTime](#) () const
- [acs_double getNseconds](#) () const
- [acs_int getNreactions](#) () const
- [acs_double getMAXhours](#) () const
- [acs_int getMAXattempts](#) () const
- [acs_int getCurrentAttempts](#) () const
- [acs_double getTimeStructuresSavingInterval](#) () const
- [acs_double getFileTimesSavingInterval](#) () const
- [acs_int getLastFiringDiskSpeciesID](#) () const
- [acs_double getOverallConcentration](#) () const
- [acs_int getMaxNonCatalyticLength](#) () const
- [acs_double getRctProb](#) () const
- [acs_double getCleavProb](#) () const
- [bool getReverseReactions](#) () const
- [acs_int getEnergy](#) () const
- [acs_double getRatioSpeciesEnergizable](#) () const
- [acs_int getADP](#) () const
- [acs_int getATP](#) () const
- [acs_longInt getMols](#) () const
- [acs_longInt getNewMols](#) () const
- [acs_longInt getNspecies](#) () const
- [acs_longInt getNewSpecies](#) () const
- [acs_longInt getNcpx](#) () const
- [acs_longInt getNcpxMols](#) () const
- [acs_double getGillespieMean](#) () const
- [acs_double getgillespieSD](#) () const
- [acs_double getgillespieEntropy](#) () const
- [acs_double getRatioBetweenNewGillTotGill](#) () const
- [acs_double getRatioBetweenBackandForw](#) () const
- [bool getSystemExpFlag](#) () const
- [acs_double getKdiss](#) () const
- [acs_double getKass](#) () const
- [acs_double getKcpx](#) () const
- [acs_double getKcpxDiss](#) () const
- [acs_double getKnrg](#) () const
- [acs_double getKrrad](#) () const
- [acs_double getK_spont_diss](#) () const
- [acs_double getK_spont_ass](#) () const
- [acs_double getCleavageKC](#) () const
- [acs_double getComplexKC](#) () const
- [acs_double getCondensationKC](#) () const
- [acs_double getComplexDegKC](#) () const

- [acs_double getMoleculeDecayKC \(\) const](#)
- [acs_int getMaxLOut \(\) const](#)
- [acs_int getSolubilityThreshold \(\) const](#)
- [acs_double getDiffusionContribute \(\) const](#)
- [acs_double getInflux \(\) const](#)
- [acs_double getRefillInterval \(\) const](#)
- [string getAlphabet \(\) const](#)
- [acs_double getVolume \(\) const](#)
- [acs_double getRandomSeed \(\) const](#)
- [vector< species > getMoleculesPopulation \(\) const](#)
- [acs_longInt getTotalNumberOfSpecies \(\)](#)
- [acs_longInt getTotalNumberOfPossibleCatalysts \(\)](#)
- [acs_longInt getTotalNumberOfMolecules \(\)](#)
- [acs_longInt getTotalNumberOfComplexSpecies \(\)](#)
- [acs_longInt getTotalNumberOfComplexes \(\)](#)
- [acs_longInt getTotalNumberOfMonomers \(\)](#)
- [vector< reactions > getReactionsLayer \(\) const](#)
- [int getDebugLevel \(\) const](#)
- [acs_longInt getNumberOfTheoreticalSpecies \(\) const](#)
- [acs_longInt getNumberOfReactions \(\) const](#)
- [acs_longInt getNumberOfCatalysis \(\) const](#)
- [acs_longInt getNumberOfGillespieCOPYpossibleRcts \(\) const](#)
- [acs_longInt getNumberOfGillespiePossibleRcts \(\) const](#)
- [void setLivingSpeciesIDsAndAmounts \(\)](#)
- [void setNotChargedAndChargedSpeciesIDsAndAmounts \(\)](#)
- [acs_longInt getCleavageCounter \(\) const](#)
- [acs_longInt getEndoCleavageCounter \(\) const](#)
- [acs_longInt getCondensationCounter \(\) const](#)
- [acs_longInt getEndoCondensationCounter \(\) const](#)
- [acs_longInt getCpxFormCounter \(\) const](#)
- [acs_longInt getCpxDissCounter \(\) const](#)
- [acs_longInt getOverallLoadedMolsCounter \(\) const](#)
- [acs_longInt getSpontDissCounter \(\) const](#)
- [acs_longInt getSpontAssCounter \(\) const](#)
- [acs_int getTotNumberOfChargedMols \(\)](#)
- [void showGlobalParameter \(\)](#)
- [void printInitialCondition \(\)](#)
- [void printAllSpeciesIdAndSequence \(\)](#)
- [void printGillespieStructure \(\)](#)
- [void printNutrientsAndProbability \(\)](#)
- [void printAllChargeMols \(\)](#)
- [bool createInitialMoleculesPopulationFromFileSTD \(string tmpSpeciesFilePath\)](#)
- [bool createInitialReactionsLayerFromFileSTD \(string tmpSpeciesFilePath\)](#)
- [bool createInitialCatalysisLayerFromFileSTD \(string tmpCatalysisFilePath\)](#)
- [bool createInfluxLayersFromFileSTD \(string tmpInfluxFilePath\)](#)
- [bool createNrgBooleanFunctionsFromFileSTD \(string tmpBoolNrgFilePath\)](#)
- [bool createInitialMoleculesPopulationFromSpecificFileSTD \(string tmpSpeciesFilePath, \[acs_int\]\(#\) tmpActGEN, \[acs_int\]\(#\) tmpActSIM\)](#)
- [bool createInitialReactionsLayerFromSpecificFileSTD \(string tmpReactionsFilePath, \[acs_int\]\(#\) tmpActGEN, \[acs_int\]\(#\) tmpActSIM\)](#)
- [bool createInitialCatalysisLayerFromSpecificFileSTD \(string tmpCatalysisFilePath, \[acs_int\]\(#\) tmpActGEN, \[acs_int\]\(#\) tmpActSIM\)](#)
- [void nutrientsAmountsFixing \(\)](#)
- [acs_int computeSngSpeciesRctsNumber \(\[acs_longInt\]\(#\) tmpTotalNumberOfReactions, \[MTRand\]\(#\) &tmpRnd-DoubleGen\)](#)

- [acs_int selectWhetherCleavageOrCond](#) ([MTRand](#) &tmp__RndDoubleGen)
- [bool createReactionsForThisSpecies](#) ([acs_longInt](#) tmpsID, [acs_int](#) tmpReactionsForThisSpecies, [MTRand](#) &tmp_RndDoubleGen, [vector](#)< [acs_longInt](#) > &tmpIDOfCandidateSpecies, [acs_int](#) tmpRctCreationType)
- [bool updateReactions](#) ([acs_longInt](#) tmpIDtoUpdate, [acs_longInt](#) tmpNewSpecies, [acs_int](#) tmpRctType, [vector](#)< [acs_longInt](#) > &tmp_AlreadyEvaluatedSpeciesVector, [MTRand](#) &tmp_RndDoubleGen)
- [acs_double createDiffusionReinforcement](#) ([acs_double](#) tmpDiffEnh, [acs_int](#) tmpNewSpeciesLength)
- [bool setSolubility](#) ([acs_int](#) tmpNewSpeciesLength, [MTRand](#) &tmpRndDoubleGen)
- [acs_longInt returnPosSpeciesAlreadyPresent](#) ([string](#) tmpNewSequence)
- [acs_longInt returnPosReactionAlreadyPresent](#) ([acs_int](#) tmpReactionType, [acs_longInt](#) tmpIds_I, [acs_longInt](#) tmpIds_II, [acs_longInt](#) tmpIds_III)
- [bool checkIfTheReactionIsAlreadyCatalyzedByThisSpecies](#) ([acs_longInt](#) tmpSPeciesID, [acs_longInt](#) tmpID-Reaction)
- [bool performOPTGillespieComputation](#) ([MTRand](#) &tmpRndDoubleGen, [clock_t](#) &tmpTimeElapsed, [acs_int](#) tmpActGEN, [acs_int](#) tmpActSIM, [acs_int](#) tmpActSTEP, [string](#) tmpStoringPath)
- [bool performReaction](#) ([acs_longInt](#) reaction_u, [MTRand](#) &tmp_RndDoubleGen, [acs_int](#) tmp_ActGEN, [acs_int](#) tmp_ActSIM, [acs_int](#) tmp_ActSTEP, [string](#) tmp_StoringPath)
- [bool newSpeciesEvaluationIII](#) ([acs_longInt](#) tmpNewSpecies, [MTRand](#) &tmp__RndDoubleGen)
- [bool complexEvaluation](#) ([string](#) tmpComplex, [MTRand](#) &tmp__RndDoubleGen, [acs_int](#) tmpCuttingPnt, [acs_longInt](#) tmpCatalyst_ID, [acs_longInt](#) tmpCatID, [acs_longInt](#) tmpSubstrate_ID, [acs_longInt](#) tmpSecSub_ID, [bool](#) tmpCpxType)
- [acs_double computeSinglGilScore](#) ([acs_longInt](#) tmpAmountI, [acs_double](#) tmpDifI, [acs_int](#) tmpSolI, [acs_longInt](#) tmpAmountII, [acs_double](#) tmpDifII, [acs_int](#) tmpSolII, [acs_double](#) tmpK, [bool](#) tmpSameMol)
- [void performSingleGilleSpielIntroduction](#) ([acs_longInt](#) tmpAmountI, [acs_longInt](#) tmpAmountII, [acs_longInt](#) tmpIDI, [acs_longInt](#) tmpIDII, [acs_longInt](#) tmpIDCatalysis, [acs_int](#) tmp__rctType, [acs_longInt](#) tmpMol_I, [acs_longInt](#) tmpMol_II, [acs_longInt](#) tmpMol_III, [acs_longInt](#) tmpMol_IV, [acs_int](#) tmpNRGDirection, [acs_longInt](#) tmpRctID, [bool](#) tmpSameSpeciesControl)
- [void changeVolume](#) ([acs_int](#) tmpTimeSinceLastReaction)
- [void incNumberOfSpecies](#) ()
- [void decNumberOfSpecies](#) ([acs_int](#) tmpID)
- [void incNumberOfMols](#) ()
- [void decNumberOfMols](#) ()
- [void incNumberOfCpx](#) ()
- [void decNumberOfCpx](#) ([acs_int](#) tmpID)
- [void incNumberOfCpxMols](#) ()
- [void decNumberOfCpxMols](#) ()
- [void incNumberOfNewSpecies](#) ([acs_int](#) tmpID)
- [void decNumberOfNewSpecies](#) ([acs_int](#) tmpID)
- [void incNumberOfNewMols](#) ([acs_int](#) tmpID)
- [void decNumberOfNewMols](#) ([acs_int](#) tmpID)
- [void decMolSpeciesProcedure](#) ([acs_int](#) tmp_ID)
- [void decCpxProcedure](#) ([acs_int](#) tmp_ID)
- [void incMolProcedure](#) ([acs_int](#) tmp_ID)
- [void incSpeciesProcedure](#) ([acs_int](#) tmp_ID)
- [void unchargeMolProcess](#) ([acs_int](#) tmp_ID)
- [void incCleavageCounter](#) ()
- [void incEndoCleavageCounter](#) ()
- [void incCondensationCounter](#) ()
- [void incEndoCondensationCounter](#) ()
- [void incCpxFormCounter](#) ()
- [void incCpxDissCounter](#) ()
- [void incOverallLoadedMolsCounter](#) ()
- [void decOverallLoadedMolsCounter](#) ()
- [void incSpontDissCounter](#) ()
- [void incSpontAssCounter](#) ()
- [void resetCleavageCounter](#) ()

- void [resetEndoCleavageCounter](#) ()
- void [resetCondensationCounter](#) ()
- void [resetEndoCondensationCounter](#) ()
- void [resetOverallLoadedMolsCounter](#) ()
- void [resetCpxFormCounter](#) ()
- void [resetCpxDissCounter](#) ()
- void [resetSpontDissCounter](#) ()
- void [resetSpontAssCounter](#) ()
- void [resetReactionsCounter](#) ()
- bool [addChargeMolToList](#) ([acs_int](#) tmpSpeciesID)
- bool [removeChargeMolFromList](#) ([acs_int](#) tmpSpeciesID)
- void [clearAllStructures](#) ()
- void [resetConcentrationToInitialConditions](#) ()
- void [storeInitialStructures](#) ()
- bool [performRefill](#) ([acs_double](#) tmpTimeSinceTheLastInflux, [acs_double](#) tmpMinimalTimeForOneMols, [MTRand](#) &tmp__RndDoubleGen)
- bool [performMoleculesEfflux](#) ([acs_double](#) tmpTimeInterval, [MTRand](#) &tmp__RndDoubleGen)
- bool [performDETMoleculesCharging](#) ([acs_double](#) tmpTimeInterval, [MTRand](#) &tmp__RndDoubleGen)
- bool [performDETComplexDissociation](#) ([acs_double](#) tmpTimeInterval, [MTRand](#) &tmp__RndDoubleGen)
- void [setActualTime](#) ([acs_double](#) tmpActualTime)
- void [updateSpeciesAges](#) ()
- void [increaseAttempts](#) ()
- bool [performCondensation](#) ([acs_longInt](#) tmpCatalyst, [acs_longInt](#) tmpSubstrate, [acs_longInt](#) tmpProduct, [acs_longInt](#) tmpComplex, [acs_longInt](#) tmpIdReaction, [acs_longInt](#) tmpIdCatalysis, [MTRand](#) &tmp__RndDoubleGen)
- bool [perform_endo_Condensation](#) ([acs_longInt](#) tmpCatalyst, [acs_longInt](#) tmpSubstrate, [acs_longInt](#) tmpProduct, [acs_longInt](#) tmpComplex, [acs_int](#) tmpNRGside, [acs_longInt](#) tmpIdReaction, [acs_longInt](#) tmpIdCatalysis, [MTRand](#) &tmp__RndDoubleGen)
- bool [performCleavage](#) ([acs_longInt](#) tmpSubstrate, [acs_longInt](#) tmpProduct_I, [acs_longInt](#) tmpProduct_II, [acs_longInt](#) tmpIdReaction, [acs_longInt](#) tmpIdCatalysis, [MTRand](#) &tmp__RndDoubleGen)
- bool [perform_endo_Cleavage](#) ([acs_longInt](#) tmpSubstrate, [acs_longInt](#) tmpProduct_I, [acs_longInt](#) tmpProduct_II, [acs_int](#) tmpNRGside, [acs_longInt](#) tmpIdReaction, [acs_longInt](#) tmpIdCatalysis, [MTRand](#) &tmp__RndDoubleGen)
- bool [performComplexFormation](#) ([acs_longInt](#) tmpCatalyst, [acs_longInt](#) tmpSubstrate, [acs_longInt](#) tmpCatID, [acs_longInt](#) tmpSecSub, [MTRand](#) &tmp__RndDoubleGen)
- bool [perform_endo_ComplexFormation](#) ([acs_longInt](#) tmpCatalyst, [acs_longInt](#) tmpSubstrate, [acs_longInt](#) tmpCatID, [acs_longInt](#) tmpSecSub, [acs_int](#) tmpNRGside, [MTRand](#) &tmp__RndDoubleGen)
- bool [performComplexDissociation](#) ([acs_longInt](#) tmpComplex, [acs_longInt](#) tmpCatalyst, [acs_longInt](#) tmpSubstrate, [MTRand](#) &tmp__RndDoubleGen)
- bool [performSpontaneousCondensation](#) ([acs_longInt](#) tmpReaction, [MTRand](#) &tmp__RndDoubleGen)
- bool [performSpontaneousCleavage](#) ([acs_longInt](#) tmpReaction, [MTRand](#) &tmp__RndDoubleGen)
- bool [performMoleculeEfflux](#) ([acs_longInt](#) tmpSpecies, [MTRand](#) &tmp__RndDoubleGen)
- bool [performEnergyEfflux](#) ([MTRand](#) &tmp__RndDoubleGen)
- bool [structureCoherenceCheckUp](#) ()
- bool [notInverseReactionAlreadyCatalyzed](#) ([acs_int](#) tmpRct, [acs_longInt](#) tmpID_I, [acs_longInt](#) tmpID_II)
- bool [checkIfOnlyMutualCatalysis](#) ([acs_int](#) tmpCat, [acs_int](#) tmpCandidateProduct)
- bool [checkAvailability](#) ([acs_longInt](#) tmpMI, [acs_longInt](#) tmpMII, [acs_longInt](#) tmpQI, [acs_longInt](#) tmpQII)
- void [inserSubListInSpecies](#) ()
- void [showSubListInSpecies](#) ()
- void [showGillEngagementInSpecies](#) ()
- bool [saveConfigurationFileSTD](#) (string tmpStoringPath)
- bool [saveInfluxStructureSTD](#) (string tmpStoringPath)
- bool [saveNrgBoolFncStructureSTD](#) (string tmpStoringPath)
- string [zeroBeforeStringNumberSTD](#) ([acs_int](#) tmpTotN, [acs_int](#) tmpCurrentN)
- bool [saveSpeciesStructureSTD](#) ([acs_int](#) tmpCurrentGen, [acs_int](#) tmpCurrentSim, [acs_int](#) tmpCurrentStep, string tmpStoringPath)

- bool [saveReactionsStructureSTD](#) ([acs_int](#) tmpCurrentGen, [acs_int](#) tmpCurrentSim, [acs_int](#) tmpCurrentStep, string tmpStoringPath)
- bool [saveCatalysisStructureSTD](#) ([acs_int](#) tmpCurrentGen, [acs_int](#) tmpCurrentSim, [acs_int](#) tmpCurrentStep, string tmpStoringPath)
- bool [saveTimesSTD](#) ([acs_int](#) tmpCurrentGen, [acs_int](#) tmpCurrentSim, [acs_int](#) tmpCurrentStep, string tmpStoringPath)
- bool [saveReactionsParametersSTD](#) ([acs_int](#) tmp__CurrentGen, [acs_int](#) tmp__CurrentSim, [acs_int](#) tmp__CurrentStep, string tmp__StoringPath, [acs_int](#) tmpRctType, [acs_longInt](#) tmpCat, [acs_longInt](#) tmpMol_I, [acs_longInt](#) tmpMol_II, [acs_longInt](#) tmpMol_III)
- bool [saveLivingSpeciesIDSTD](#) ([acs_int](#) tmp__CurrentGen, [acs_int](#) tmp__CurrentSim, [acs_int](#) tmp__CurrentStep, string tmp__StoringPath)
- bool [saveLivingSpeciesAmountSTD](#) ([acs_int](#) tmp__CurrentGen, [acs_int](#) tmp__CurrentSim, string tmp__StoringPath)
- bool [saveLivingSpeciesConcentrationSTD](#) ([acs_int](#) tmp__CurrentGen, [acs_int](#) tmp__CurrentSim, string tmp__StoringPath)
- bool [devStd](#) ()
- bool [entropy](#) ()

12.3.1 Detailed Description

environment class

Author

Alessandro Filisetti

Version

2.4

Date

2010-06-10 *^*^*^*^*

This class contains environmental proprieties and methods; within this class all things occur

Author

Alessandro Filisetti

Version

0.2

Date

2011-12-15

Definition at line 20 of file environment.h.

12.3.2 Constructor & Destructor Documentation

12.3.2.1 `environment::environment (string tmpInitialPath)`

TEMPLATE DEACLARATION AND DEFINITION Test environment costructor

Version

2.4

Date

2010-06-27

Parameters

<i>tmpRndDouble-Gen</i>	randomGenerator reference Environment Constructor
-------------------------	---

Version

1.0

Parameters

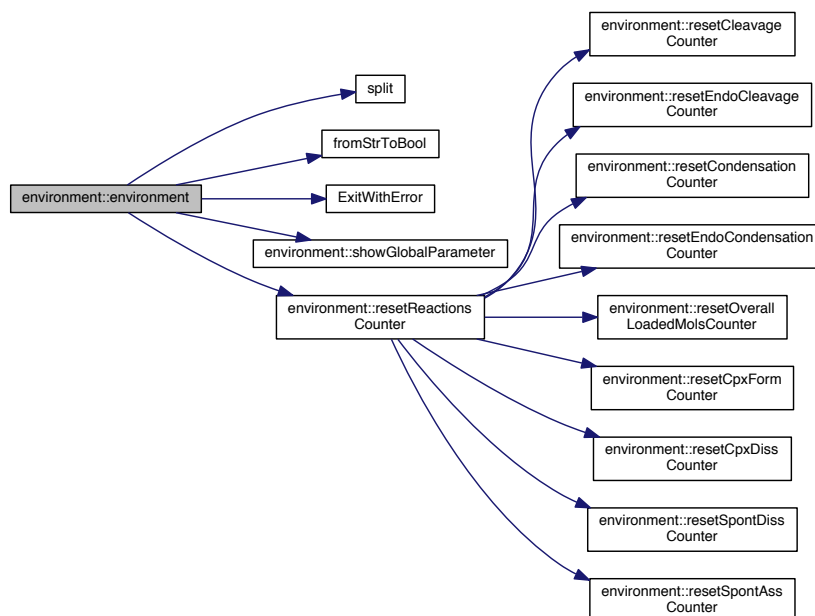
<i>tmpInitialPath</i>	
-----------------------	--

Date

2013/07/04

Definition at line 80 of file environment.cpp.

Here is the call graph for this function:



12.3.2.2 environment::~~environment () [inline]

Definition at line 148 of file environment.h.

12.3.3 Member Function Documentation

12.3.3.1 `bool environment::addChargeMolToList (acs_int tmpSpeciesID)`

Perform vector unchargedIDlist update adding a new charge molecule vector unchargedIDlist and cumUnchargedAmountList are involved

Version

1.0

Date

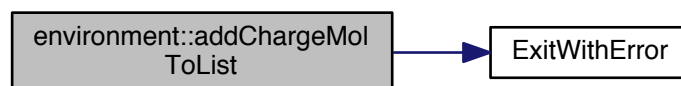
2010-10-10

Parameters

<i>acs_int</i>	<i>tmpSpeciesID</i> Specie to charge
----------------	--------------------------------------

Definition at line 4313 of file environment.cpp.

Here is the call graph for this function:



12.3.3.2 `void environment::changeVolume (acs_int tmpTimeSinceLastReaction)`

Change volume function 1.0

Date

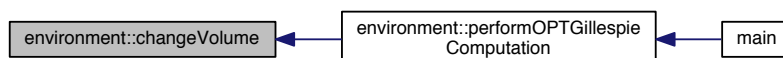
2013/07/17

Author

Alessandro Filisetti

Definition at line 6157 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.3 `bool environment::checkAvailability (acs_longInt tmpMI, acs_longInt tmpMII, acs_longInt tmpQI, acs_longInt tmpQII)`

Return TRUE if there are sufficient molecules for the reaction. It is used for the reaction in which catalyst and substrate are the same molecules

Version

1.0

Date

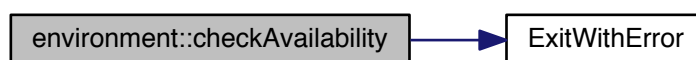
2011.07.25

Parameters

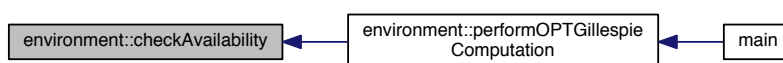
<i>tmpMI</i>	
<i>tmpMII</i>	
<i>tmpQI</i>	
<i>tmpQII</i>	

Definition at line 2635 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.4 `bool environment::checkIfOnlyMutualCatalysis (acs_int tmpCat, acs_int tmpCandidateProduct)`

This function return false if the tmpCandidateProduct is a catalyst of tmpCat

Version

1.0 - last update 2009/10/08 - build 009

Definition at line 1965 of file environment.cpp.

Here is the call graph for this function:



12.3.3.5 `bool environment::checkIfTheReactionIsAlreadyCatalyzedByThisSpecies (acs_longInt tmpSpeciesID, acs_longInt tmpIdReaction)`

If the reaction is not new this function checks if the reactions has been already catalysed by this species

Version

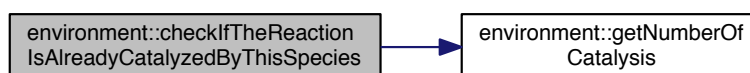
1.0

Parameters

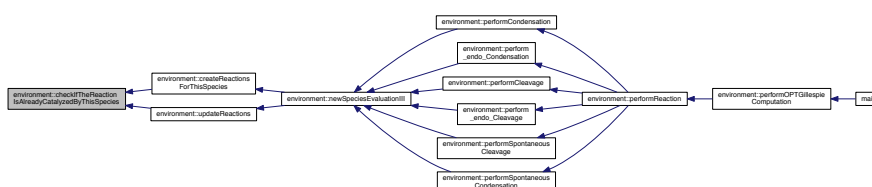
<i>acs_int</i>	tmpSpeciesID catalyst ID
<i>tmpIdReaction</i>	reaction ID

Definition at line 1937 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.6 `void environment::clearAllStructures ()`

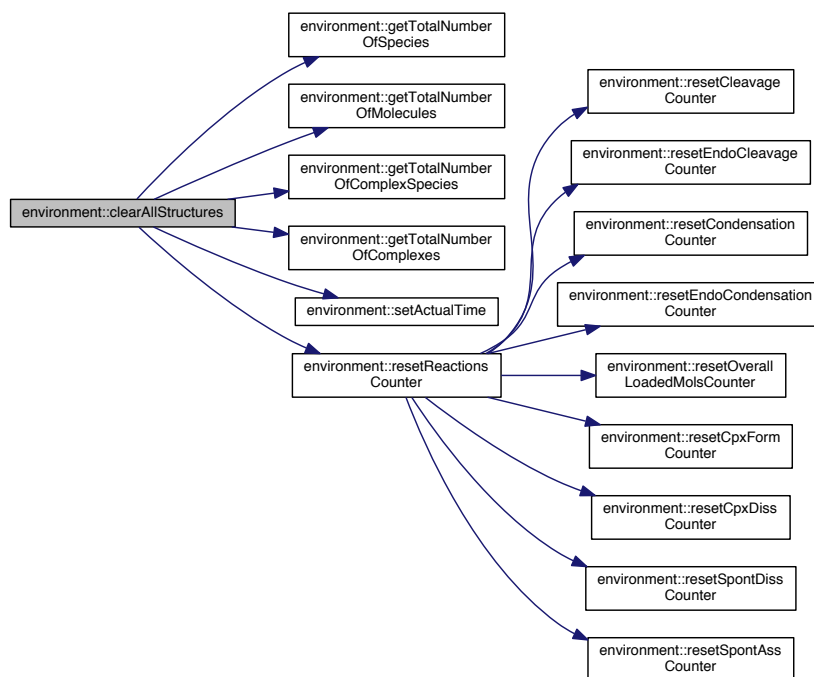
Clear all structures after each simulation

Version

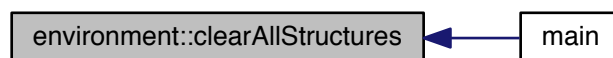
1.0

Definition at line 6405 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.7 `bool environment::complexEvaluation (string tmpComplex, MTRand & tmp__RndDoubleGen, acs_int tmpCuttingPnt, acs_longInt tmpCatalyst_ID, acs_longInt tmpSubstrate_ID, acs_longInt tmpCatID, acs_longInt tmpSecSub_ID, bool tmpCpxType)`

Complex evaluation

Version

1.1

Date

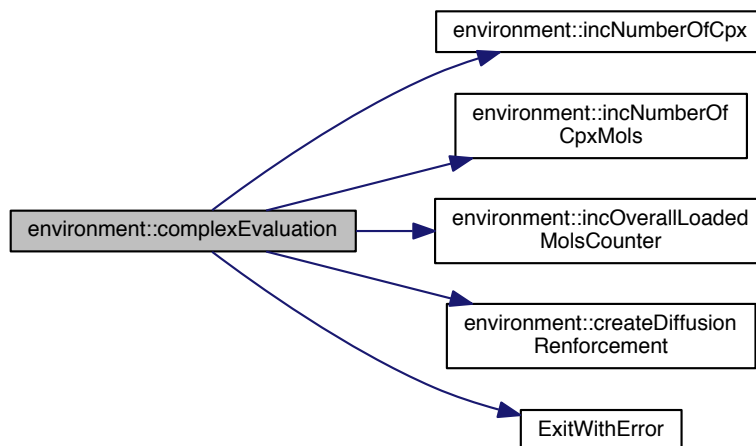
2010-06-04

Parameters

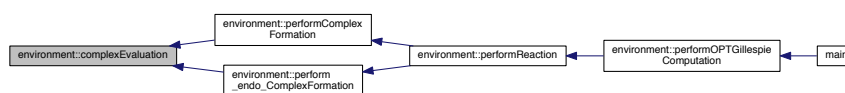
<i>string</i>	tmpNewSpecies New species sequence to evaluate
<i>MTRand&</i>	tmp___RndDoubleGen random number generator
<i>tmpCuttingPnt</i>	Complex cutting point
<i>tmpCatalyst_ID</i>	catalyst ID
<i>tmpSubstrate_ID</i>	substrate ID
<i>tmp_catalysisID</i>	catalysis ID
<i>tmpCpxType</i>	ENDOERGONIC or ESOERGONIC

Definition at line 6025 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.8 `acs_double environment::computeSinglGilScore (acs_longInt tmpAmountl, acs_double tmpDifl, acs_int tmpSoll, acs_longInt tmpAmountll, acs_double tmpDifll, acs_int tmpSolll, acs_double tmpK, bool tmpSameMol)`

Compute a single gillespie score according to the amount and peoprieties of the species involved

Version

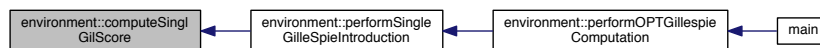
1.0

Date

20110214

Definition at line 3288 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.9 `acs_int environment::computeSngSpeciesRctsNumber (acs_longInt tmpTotalNumberOfReactions, MTRand & tmpRndDoubleGen)`

Initial molecule population creation. If the number of species stored in the configuration file is grater than the possible number of species according to the alphabet and maximum length all species up to the MAX length will be created

Version

1.0

Parameters

<i>MTRand&</i>	tmpRndDoubleGen initial layer initialization
--------------------	--

Version

1.0

Parameters

<i>MTRand&</i>	tmpRndDoubleGen Compute number of reaction catalysd by a catalyst according to the total number of reactions and reactions probabilities
--------------------	--

Version

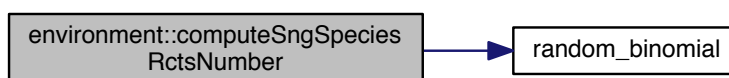
1.0

Parameters

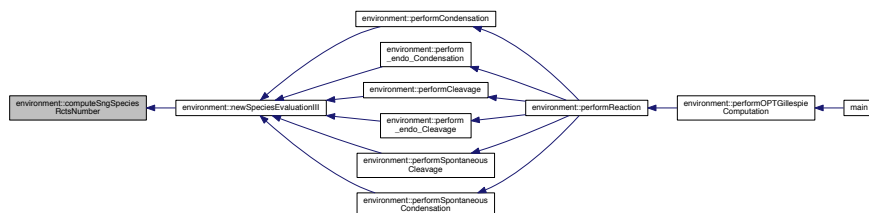
<i>acs_int</i>	tmpTotalNumberOfReactions Total number of conceivable reactions
<i>acs_double</i>	tmpRctsProb reaction probability

Definition at line 481 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.10 `acs_double environment::createDiffusionReinforcement (acs_double tmpDiffEnh, acs_int tmpNewSpeciesLength)`

Create the diffusion constant reinforcement according to the species length

Version

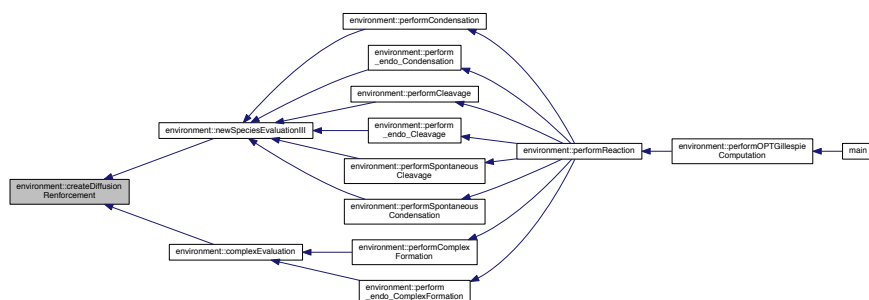
1.0

Parameters

<i>tmpDiffEnh</i>	diffusion enhancement from parameters
<i>MTRand&</i>	tmp_RndDoubleGen random number generator
<i>tmpNewSpeciesLength</i>	Length of the species

Definition at line 1819 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.11 `bool environment::createInfluxLayersFromFileSTD (string tmpInfluxFilePath)`

Create influx layer from file C++ libraries

Version

1.0

Parameters

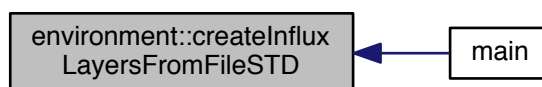
<i>string</i>	tmpInfluxFilePath file path
---------------	-----------------------------

Date

20130702

Definition at line 1572 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.12 bool environment::createInitialCatalysisLayerFromFileSTD (string tmpCatalysisFilePath)

Catalysis from file using standard C++ libraries

Version

1.0

Parameters

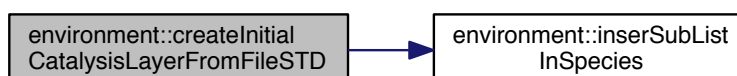
<i>string</i>	tmpSpeciesFilePath file path
---------------	------------------------------

Date

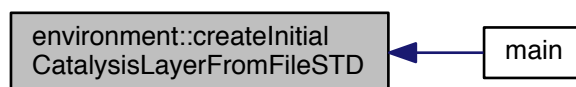
20130702

Definition at line 1724 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.13 `bool environment::createInitialCatalysisLayerFromSpecificFileSTD (string tmpCatalysisFilePath, acs_int tmpActGEN, acs_int tmpActSIM)`

catalysis from file using standard C++ libraries

Version

1.0

Parameters

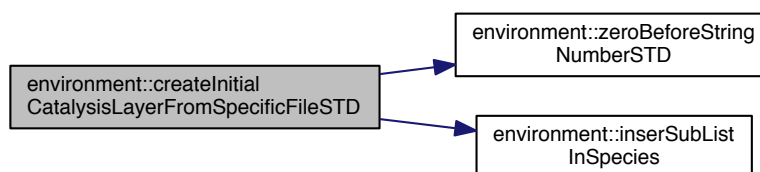
<i>string</i>	<i>tmpSpeciesFilePath</i> file path
---------------	-------------------------------------

Date

20130702

Definition at line 1766 of file environment.cpp.

Here is the call graph for this function:



12.3.3.14 `bool environment::createInitialMoleculesPopulationFromFileSTD (string tmpSpeciesFilePath)`

Initial molecule population creation from file using standard C++ libraries

Version

1.0

Parameters

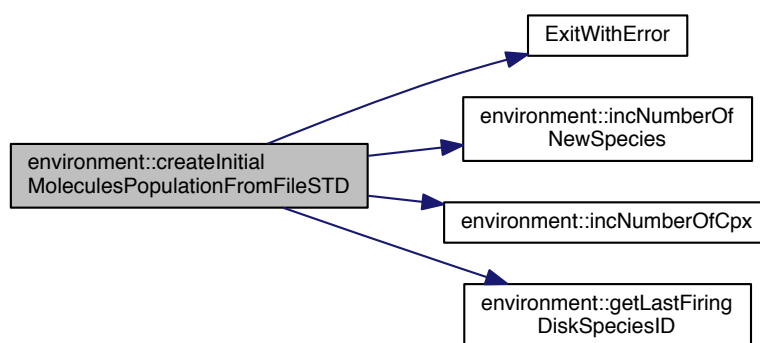
<i>string</i>	tmpSpeciesFilePath file path
---------------	------------------------------

Date

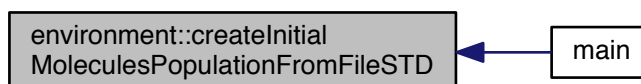
20130702

Definition at line 1249 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.15 `bool environment::createInitialMoleculesPopulationFromSpecificFileSTD (string tmpSpeciesFilePath, acs_int tmpActGEN, acs_int tmpActSIM)`

Initial molecule population creation from file

Version

1.0

Parameters

<i>QString</i>	tmpSpeciesFilePath file path Initial molecule population creation. Species are uploaded from a SPECIFIC file created using actual generation and simulation
----------------	---

Version

1.0

Parameters

<i>QString</i>	tmpSpeciesFilePath file path Initial molecule population creation. Species are uploaded from a SPECIFIC file created using actual generation and simulation (standard C++ libraries)
----------------	--

Version

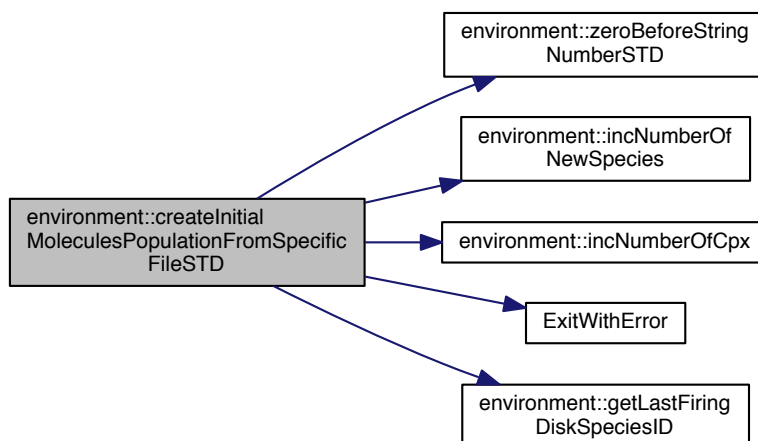
1.0

Parameters

<i>QString</i>	tmpSpeciesFilePath file path
----------------	------------------------------

Definition at line 1478 of file environment.cpp.

Here is the call graph for this function:



12.3.3.16 `bool environment::createInitialReactionsLayerFromFileSTD (string tmpSpeciesFilePath)`

Reactions from file using standard C++ libraries

Version

1.0

Parameters

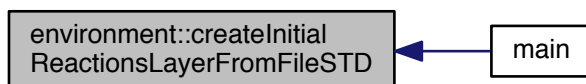
<i>string</i>	tmpSpeciesFilePath file path
---------------	------------------------------

Date

20130702

Definition at line 1635 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.17 `bool environment::createInitialReactionsLayerFromSpecificFileSTD (string tmpReactionsFilePath, acs_int tmpActGEN, acs_int tmpActSIM)`

Reactions from file using standard C++ libraries

Version

1.0

Parameters

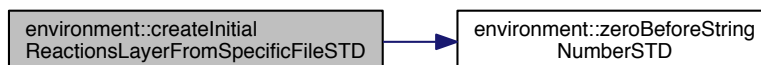
<i>string</i>	tmpSpeciesFilePath file path
---------------	------------------------------

Date

20130702

Definition at line 1674 of file environment.cpp.

Here is the call graph for this function:



12.3.3.18 `bool environment::createNrgBooleanFunctionsFromFileSTD (string tmpBoolNrgFilePath)`

load energy boolean function (in decimal format) - Standard C++

Version

1.0

Parameters

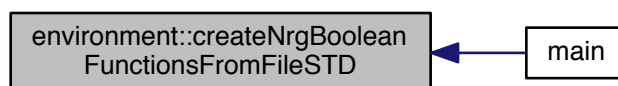
<i>string</i>	tmpBoolNrgFilePath file path
---------------	------------------------------

Date

20130702

Definition at line 1604 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.19 **bool** environment::createReactionsForThisSpecies (**acs_longInt** *tmpsID*, **acs_int** *tmpReactionsForThisSpecies*, **MTRand** & *tmp_RndDoubleGen*, **vector**< **acs_longInt** > & *tmpIDOfCandidateSpecies*, **acs_int** *tmpRctCreationType*)

Creation of all the reactions related to one specific species

Version

1.1

Date

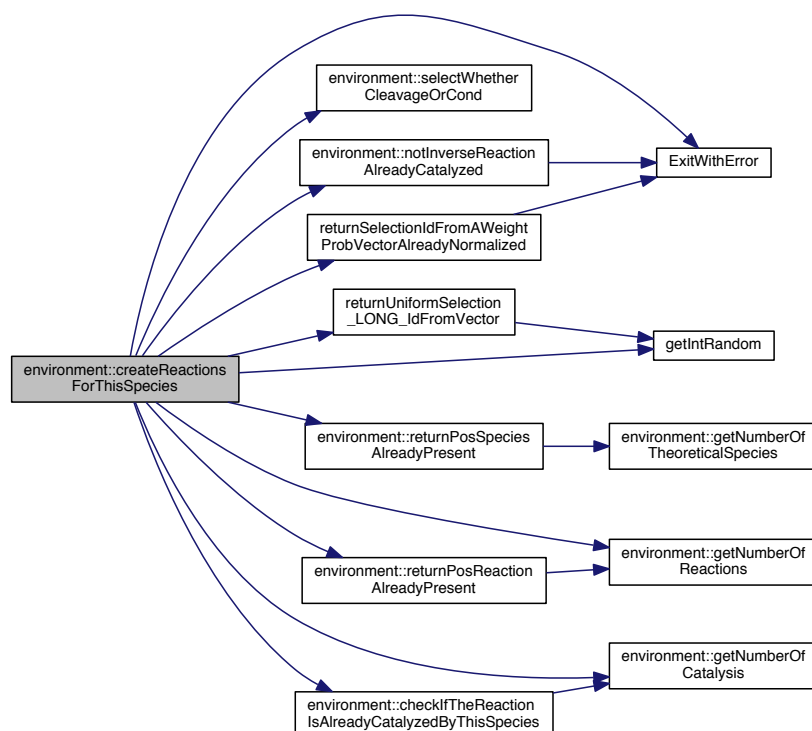
2011/07/07

Parameters

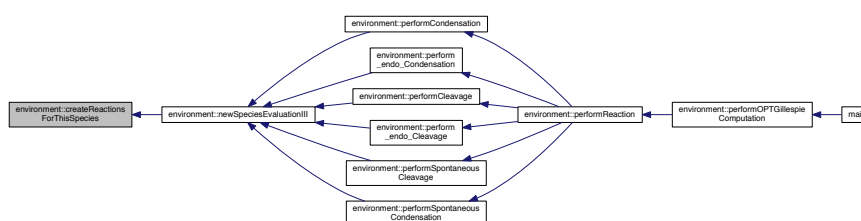
<i>acs_longInt</i>	tmpsID species vector ID
<i>acs_int</i>	tmpReactionsForThisSpecies number of reactions to create for this species
<i>MTRand</i> &	tmp_RndDoubleGen random number generator
<i>vector</i> < <i>acs_ - longInt</i> > &	tmpIDOfCandidateSpecies ID of the species available for the reaction
<i>acs_int</i>	tmpRctCreationType NEWREACTION or UPGRADEREACTIONS

Definition at line 525 of file environment.cpp.

Here is the call graph for this function:



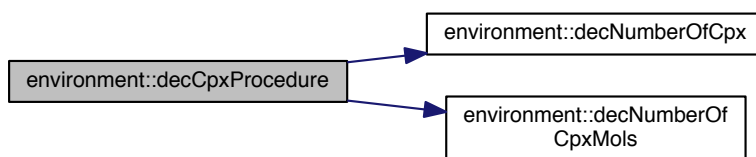
Here is the caller graph for this function:



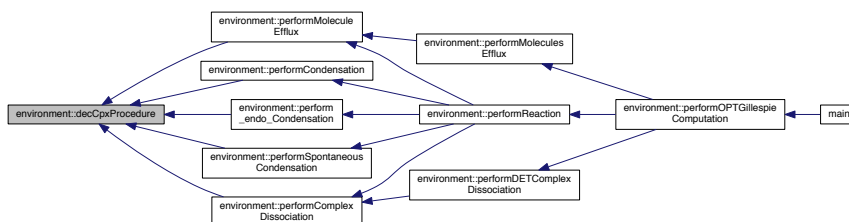
12.3.3.20 `void environment::decCpxProcedure (acs_int tmp_ID) [inline]`

Definition at line 298 of file `environment.h`.

Here is the call graph for this function:



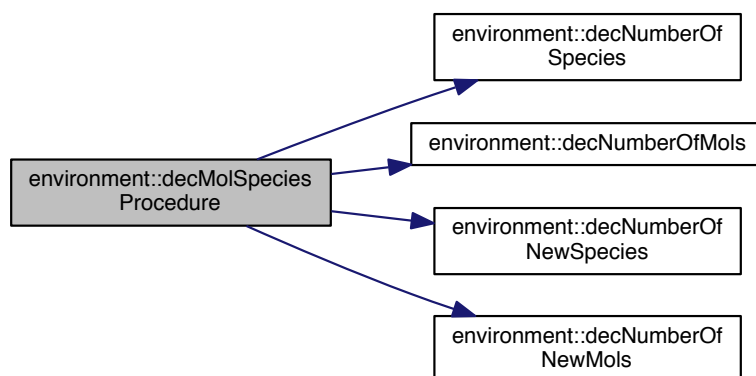
Here is the caller graph for this function:



12.3.3.21 `void environment::decMolSpeciesProcedure (acs_int tmp_ID) [inline]`

Definition at line 297 of file `environment.h`.

Here is the call graph for this function:



```

graph TD
    main --> env_opt_gillespie[environment:performOPTGillespieComputation]
    env_opt_gillespie --> env_molecules_efflux[environment:performMoleculesEfflux]
    env_opt_gillespie --> env_condensation[environment:performCondensation]
    env_opt_gillespie --> env_cleavage[environment:performCleavage]
    env_opt_gillespie --> env_reaction[environment:performReaction]
    env_opt_gillespie --> env_spontaneous_cleavage[environment:performSpontaneousCleavage]
    env_molecules_efflux --> env_molecule_efflux[environment:performMoleculeEfflux]
    env_condensation --> env_endo_condensation[environment:perform_endo_Condensation]
    env_cleavage --> env_endo_cleavage[environment:perform_endo_Cleavage]
    env_reaction --> env_endo_cleavage
    env_reaction --> env_complex_formation[environment:performComplexFormation]
    env_reaction --> env_endo_complex_formation[environment:perform_endo_ComplexFormation]
    env_reaction --> env_spontaneous_condensation[environment:performSpontaneousCondensation]
    env_spontaneous_cleavage --> env_spontaneous_condensation
    env_molecule_efflux --> env_dec_mol_spec[environment:decMolSpeciesProcedure]
    env_endo_condensation --> env_dec_mol_spec
    env_endo_cleavage --> env_dec_mol_spec
    env_complex_formation --> env_dec_mol_spec
    env_endo_complex_formation --> env_dec_mol_spec
    env_spontaneous_condensation --> env_dec_mol_spec
  
```

Definition at line 288 of file environment.h.

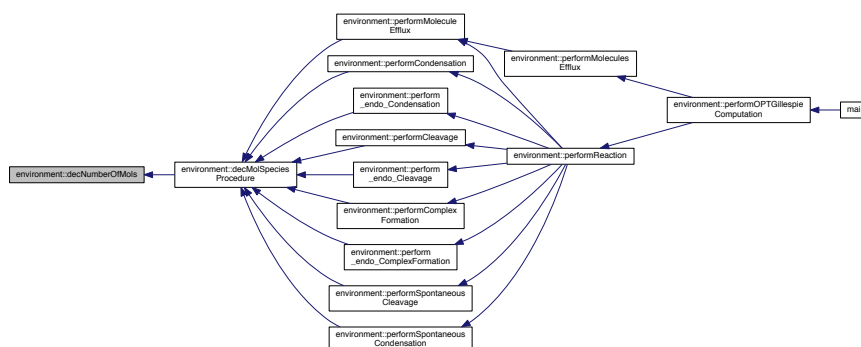
```

graph LR
    main((main)) --> envOPT[environment:performOPT Gillespie Computation]
    envOPT --> envMolEfflux[environment:performMoleculesEfflux]
    envMolEfflux --> envMolEfflux2[environment:performMoleculeEfflux]
    envMolEfflux2 --> envCondensation[environment:performCondensation]
    envCondensation --> envReaction[environment:performReaction]
    envReaction --> envDecCpx[environment:decCpxProcedure]
    envReaction --> envEndoCondensation[environment:perform_endo_Condensation]
    envReaction --> envSpontCondensation[environment:performSpontaneous Condensation]
    envReaction --> envComplexDissociation[environment:performComplex Dissociation]
    envReaction --> envDETComplexDissociation[environment:performDETComplex Dissociation]
    envDecCpx --> envDecNumberCpx[environment:decNumberOfCpx]
    envEndoCondensation --> envDecCpx
    envSpontCondensation --> envDecCpx
    envComplexDissociation --> envDecCpx
    envDETComplexDissociation --> envDecCpx
  
```

Definition at line 290 of file environment.h.

Definition at line 286 of file environment.h.

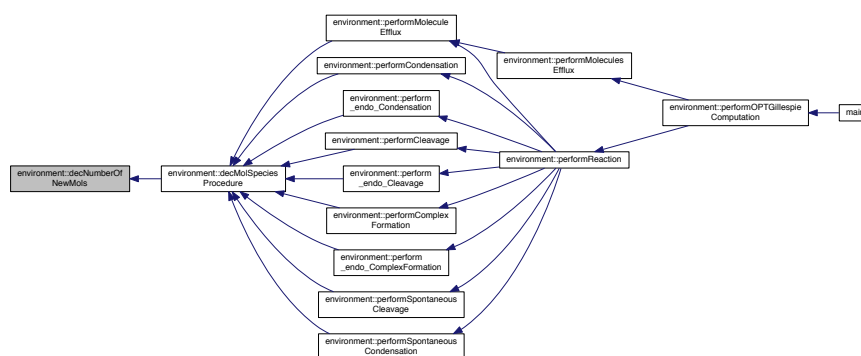
Here is the caller graph for this function:



12.3.3.25 void environment::decNumberOfNewMols (*acs_int tmpID*) [inline]

Definition at line 295 of file environment.h.

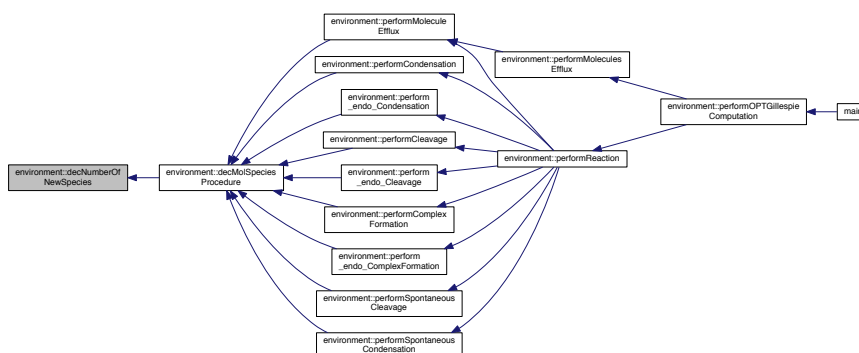
Here is the caller graph for this function:



12.3.3.26 void environment::decNumberOfNewSpecies (*acs_int tmpID*) [inline]

Definition at line 293 of file environment.h.

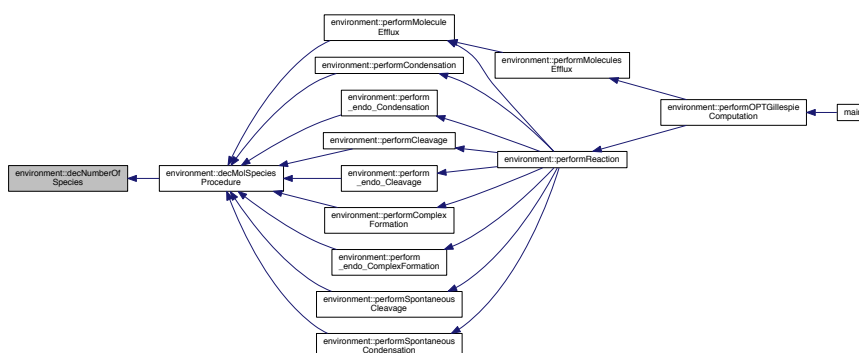
Here is the caller graph for this function:



```
12.3.3.27 void environment::decNumberOfSpecies ( acs_int tmpID ) [inline]
```

Definition at line 284 of file environment.h.

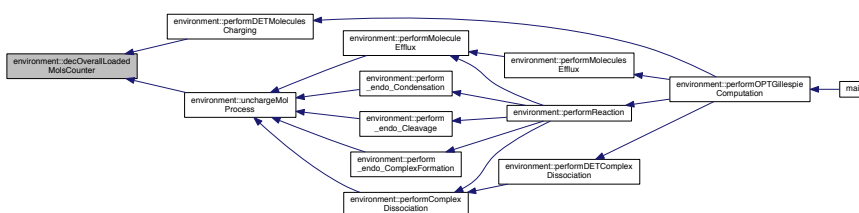
Here is the caller graph for this function:



12.3.3.28 void environment::decOverallLoadedMolsCounter () [inline]

Definition at line 311 of file environment.h.

Here is the caller graph for this function:



12.3.3.29 `bool environment::devStd ()`

Definition at line 7370 of file environment.cpp.

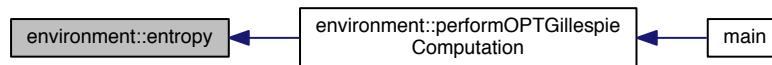
Here is the caller graph for this function:



12.3.3.30 `bool environment::entropy ()`

Definition at line 7388 of file environment.cpp.

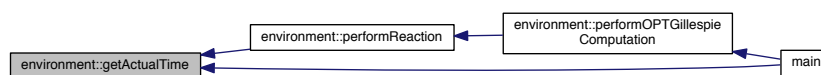
Here is the caller graph for this function:



12.3.3.31 `acs_double environment::getActualTime () const` `[inline]`

Definition at line 153 of file environment.h.

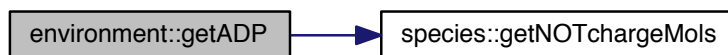
Here is the caller graph for this function:



12.3.3.32 `acs_int environment::getADP () const` `[inline]`

Definition at line 169 of file environment.h.

Here is the call graph for this function:



12.3.3.33 `string environment::getAlphabet () const [inline]`

Definition at line 207 of file `environment.h`.

12.3.3.34 `acs_int environment::getATP () const [inline]`

Definition at line 170 of file `environment.h`.

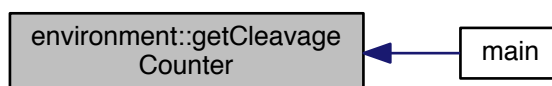
Here is the call graph for this function:



12.3.3.35 `acs_longInt environment::getCleavageCounter () const [inline]`

Definition at line 226 of file `environment.h`.

Here is the caller graph for this function:



12.3.3.36 `acs_double environment::getCleavageKC () const [inline]`

Definition at line 195 of file `environment.h`.

12.3.3.37 **acs_double** environment::getCleavProb () const [inline]

Definition at line 165 of file environment.h.

12.3.3.38 **acs_double** environment::getComplexDegKC () const [inline]

Definition at line 198 of file environment.h.

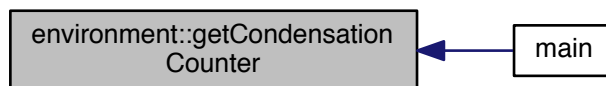
12.3.3.39 **acs_double** environment::getComplexKC () const [inline]

Definition at line 196 of file environment.h.

12.3.3.40 **acs_longInt** environment::getCondensationCounter () const [inline]

Definition at line 228 of file environment.h.

Here is the caller graph for this function:



12.3.3.41 **acs_double** environment::getCondensationKC () const [inline]

Definition at line 197 of file environment.h.

12.3.3.42 **acs_longInt** environment::getCpxDissCounter () const [inline]

Definition at line 231 of file environment.h.

Here is the caller graph for this function:



12.3.3.43 **acs_longInt** environment::getCpxFormCounter () const [inline]

Definition at line 230 of file environment.h.

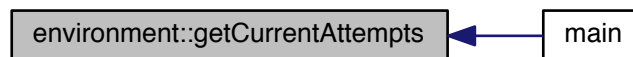
Here is the caller graph for this function:



12.3.3.44 `acs_int environment::getCurrentAttempts () const [inline]`

Definition at line 158 of file environment.h.

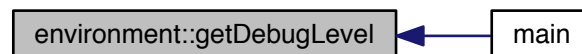
Here is the caller graph for this function:



12.3.3.45 `int environment::getDebugLevel () const [inline]`

Definition at line 218 of file environment.h.

Here is the caller graph for this function:



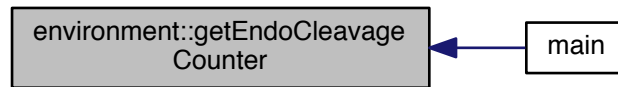
12.3.3.46 `acs_double environment::getDiffusionContribute () const [inline]`

Definition at line 204 of file environment.h.

12.3.3.47 `acs_longInt environment::getEndoCleavageCounter () const [inline]`

Definition at line 227 of file environment.h.

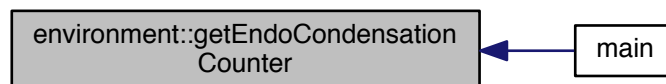
Here is the caller graph for this function:



12.3.3.48 **acs_longInt** environment::getEndoCondensationCounter () const [inline]

Definition at line 229 of file environment.h.

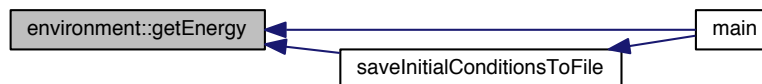
Here is the caller graph for this function:



12.3.3.49 **acs_int** environment::getEnergy () const [inline]

Definition at line 167 of file environment.h.

Here is the caller graph for this function:



12.3.3.50 **acs_double** environment::getFileTimesSavingInterval () const [inline]

Definition at line 160 of file environment.h.

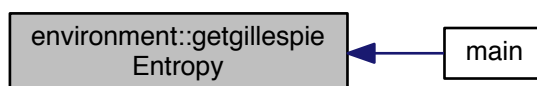
Here is the caller graph for this function:



12.3.3.51 `acs_double environment::getgillespieEntropy () const [inline]`

Definition at line 180 of file `environment.h`.

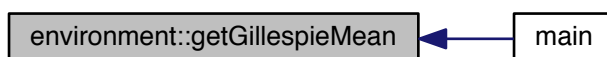
Here is the caller graph for this function:



12.3.3.52 `acs_double environment::getGillespieMean () const [inline]`

Definition at line 178 of file `environment.h`.

Here is the caller graph for this function:



12.3.3.53 `acs_double environment::getgillespieSD () const [inline]`

Definition at line 179 of file `environment.h`.

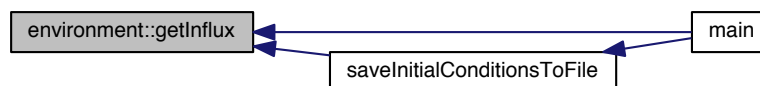
Here is the caller graph for this function:



12.3.3.54 **acs_double** `environment::getInflux () const` `[inline]`

Definition at line 205 of file `environment.h`.

Here is the caller graph for this function:



12.3.3.55 **acs_double** `environment::getK_spont_ass () const` `[inline]`

Definition at line 193 of file `environment.h`.

12.3.3.56 **acs_double** `environment::getK_spont_diss () const` `[inline]`

Definition at line 192 of file `environment.h`.

12.3.3.57 **acs_double** `environment::getKass () const` `[inline]`

Definition at line 187 of file `environment.h`.

12.3.3.58 **acs_double** `environment::getKcpx () const` `[inline]`

Definition at line 188 of file `environment.h`.

12.3.3.59 **acs_double** `environment::getKcpxDiss () const` `[inline]`

Definition at line 189 of file `environment.h`.

12.3.3.60 **acs_double** `environment::getKdiss () const` `[inline]`

Definition at line 186 of file `environment.h`.

12.3.3.61 **acs_double** environment::getKrrad () const [inline]

Definition at line 191 of file environment.h.

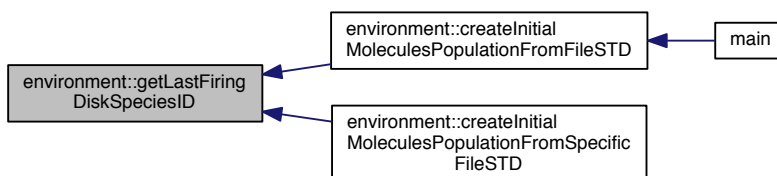
12.3.3.62 **acs_double** environment::getKnrg () const [inline]

Definition at line 190 of file environment.h.

12.3.3.63 **acs_int** environment::getLastFiringDiskSpeciesID () const [inline]

Definition at line 161 of file environment.h.

Here is the caller graph for this function:



12.3.3.64 **acs_int** environment::getMAXattempts () const [inline]

Definition at line 157 of file environment.h.

Here is the caller graph for this function:



12.3.3.65 **acs_double** environment::getMAXhours () const [inline]

Definition at line 156 of file environment.h.

Here is the caller graph for this function:



12.3.3.66 **acs_int** `environment::getMaxLOut () const` `[inline]`

Definition at line 200 of file `environment.h`.

12.3.3.67 **acs_int** `environment::getMaxNonCatalyticLength () const` `[inline]`

Definition at line 163 of file `environment.h`.

12.3.3.68 **acs_double** `environment::getMoleculeDecayKC () const` `[inline]`

Definition at line 199 of file `environment.h`.

12.3.3.69 **vector<species>** `environment::getMoleculesPopulation () const` `[inline]`

Definition at line 210 of file `environment.h`.

12.3.3.70 **acs_longInt** `environment::getMols () const` `[inline]`

Definition at line 171 of file `environment.h`.

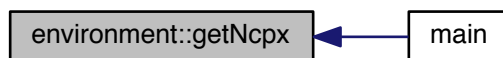
Here is the caller graph for this function:



12.3.3.71 **acs_longInt** `environment::getNcpx () const` `[inline]`

Definition at line 175 of file `environment.h`.

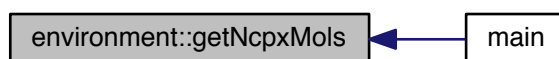
Here is the caller graph for this function:



12.3.3.72 `acs_longInt environment::getNcpxMols () const [inline]`

Definition at line 176 of file `environment.h`.

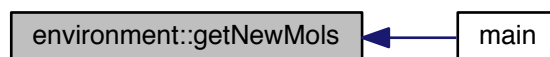
Here is the caller graph for this function:



12.3.3.73 `acs_longInt environment::getNewMols () const [inline]`

Definition at line 172 of file `environment.h`.

Here is the caller graph for this function:



12.3.3.74 `acs_int environment::getNgen () const [inline]`

Definition at line 151 of file `environment.h`.

Here is the caller graph for this function:



12.3.3.75 `acs_longInt environment::getNewSpecies () const [inline]`

Definition at line 174 of file environment.h.

Here is the caller graph for this function:



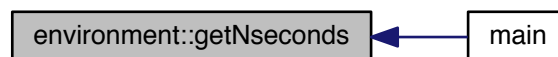
12.3.3.76 `acs_int environment::getNreactions () const [inline]`

Definition at line 155 of file environment.h.

12.3.3.77 `acs_double environment::getNseconds () const [inline]`

Definition at line 154 of file environment.h.

Here is the caller graph for this function:



12.3.3.78 `acs_int environment::getNsim () const [inline]`

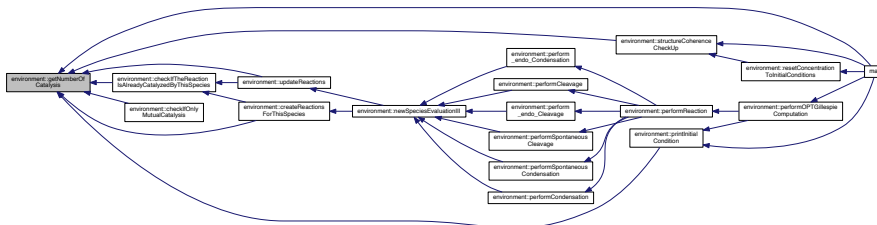
Definition at line 152 of file environment.h.


```
graph LR; main --> getNsim[environment::getNsim]
```

Here is the caller graph for this function:



Here is the caller graph for this function:

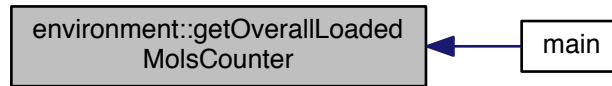


Generated on Tue Dec 10 2013 10:10:07 for CaRNeSS by Doxygen

12.3.3.86 **acs_longInt** environment::getOverallLoadedMolsCounter () const [inline]

Definition at line 232 of file environment.h.

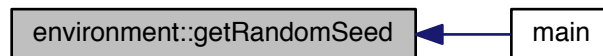
Here is the caller graph for this function:



12.3.3.87 **acs_double** environment::getRandomSeed () const [inline]

Definition at line 209 of file environment.h.

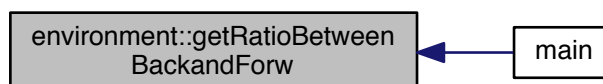
Here is the caller graph for this function:



12.3.3.88 **acs_double** environment::getRatioBetweenBackandForw () const [inline]

Definition at line 182 of file environment.h.

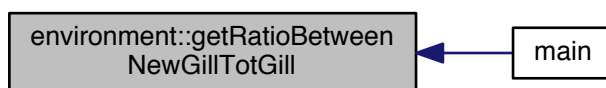
Here is the caller graph for this function:



12.3.3.89 **acs_double** environment::getRatioBetweenNewGillTotGill () const [inline]

Definition at line 181 of file environment.h.

Here is the caller graph for this function:



12.3.3.90 **acs_double** environment::getRatioSpeciesEnergizable () const [inline]

Definition at line 168 of file environment.h.

12.3.3.91 **acs_double** environment::getRctProb () const [inline]

Definition at line 164 of file environment.h.

12.3.3.92 **vector<reactions>** environment::getReactionsLayer () const [inline]

Definition at line 217 of file environment.h.

12.3.3.93 **acs_double** environment::getRefillInterval () const [inline]

Definition at line 206 of file environment.h.

12.3.3.94 **bool** environment::getReverseReactions () const [inline]

Definition at line 166 of file environment.h.

12.3.3.95 **acs_int** environment::getSolubilityThreshold () const [inline]

Definition at line 201 of file environment.h.

12.3.3.96 **acs_longInt** environment::getSpontAssCounter () const [inline]

Definition at line 234 of file environment.h.

Here is the caller graph for this function:



12.3.3.97 `acs_longInt environment::getSpontDissCounter () const [inline]`

Definition at line 233 of file environment.h.

Here is the caller graph for this function:

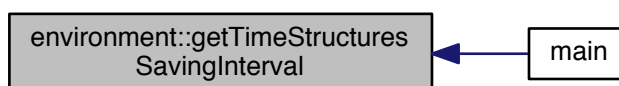
**12.3.3.98** `bool environment::getSystemExpFlag () const [inline]`

Definition at line 183 of file environment.h.

12.3.3.99 `acs_double environment::getTimeStructuresSavingInterval () const [inline]`

Definition at line 159 of file environment.h.

Here is the caller graph for this function:

**12.3.3.100** `acs_longInt environment::getTotalNumberOfComplexes ()`

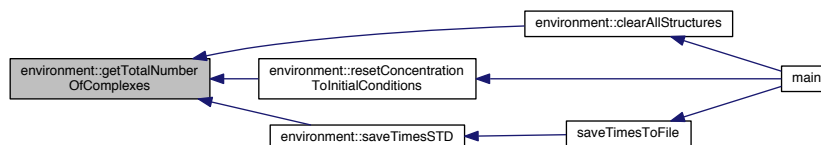
This Function returns the total amount of MOLECULES in the tmpSpeciesvector The file is saved in the directory indicated as a second parameter in the run command

Parameters

<code>vector<species>*</code>	tmpSpeciesvector pointer to tmpSpeciesvector
-------------------------------------	--

Definition at line 3578 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.101 `acs_longInt environment::getTotalNumberOfComplexSpecies ()`

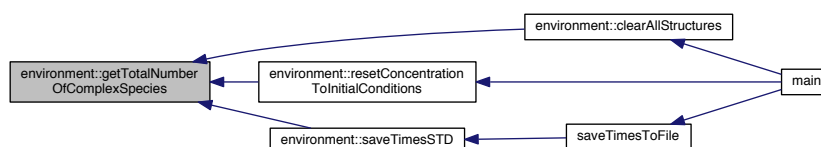
This Function returns the total amount of COMPLEX SPECIES in the tmpSpeciesvector The file is saved in the directory indicated as a second parameter in the run command

Parameters

<code>vector<species>*</code>	tmpSpeciesvector pointer to tmpSpeciesvector
-------------------------------------	--

Definition at line 3557 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.102 `acs_longInt environment::getTotalNumberOfMolecules ()`

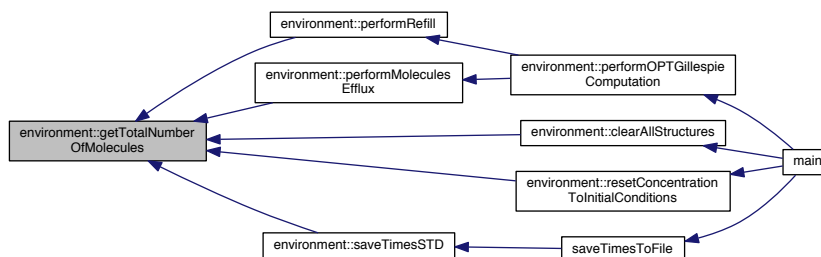
This Function returns the total amount of MOLECULES in the tmpSpeciesvector The file is saved in the directory indicated as a second parameter in the run command

Parameters

<code>vector<species>*</code>	tmpSpeciesvector pointer to tmpSpeciesvector
-------------------------------------	--

Definition at line 3533 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.103 `acs_longInt environment::getTotalNumberOfMonomers ()`

This Function returns the total amount of MONOMERS in the tmpSpeciesvector The file is saved in the directory indicated as a second parameter in the run command

Parameters

<code>vector<species>*</code>	tmpSpeciesvector pointer to tmpSpeciesvector
-------------------------------------	--

Definition at line 3599 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.104 `acs_longInt environment::getTotalNumberOfPossibleCatalysts ()`

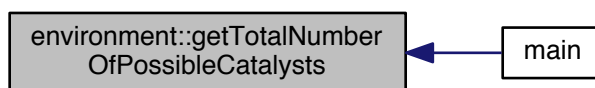
This Function returns the total amount of MOLECULES in the tmpSpeciesvector The file is saved in the directory indicated as a second parameter in the run command

Parameters

<code>vector<species>*</code>	tmpSpeciesvector pointer to tmpSpeciesvector
-------------------------------------	--

Definition at line 3510 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.105 `acs_longInt environment::getTotalNumberOfSpecies ()`

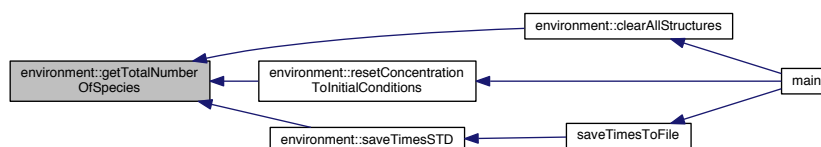
This Function returns the total amount of MOLECULES in the tmpSpeciesvector The file is saved in the directory indicated as a second parameter in the run command

Parameters

<code>vector<species>*</code>	tmpSpeciesvector pointer to tmpSpeciesvector
-------------------------------------	--

Definition at line 3486 of file environment.cpp.

Here is the caller graph for this function:

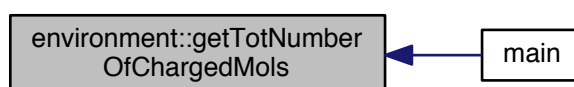


12.3.3.106 `acs_int environment::getTotNumberOfChargedMols ()`

Get the total number of charged molecules

Definition at line 3619 of file environment.cpp.

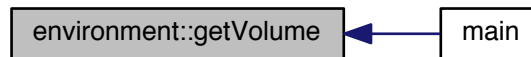
Here is the caller graph for this function:



12.3.3.107 `acs_double environment::getVolume () const [inline]`

Definition at line 208 of file environment.h.

Here is the caller graph for this function:



12.3.3.108 `void environment::incCleavageCounter () [inline]`

Definition at line 304 of file environment.h.

Here is the caller graph for this function:



12.3.3.109 `void environment::incCondensationCounter () [inline]`

Definition at line 306 of file environment.h.

Here is the caller graph for this function:



12.3.3.110 `void environment::incCpxDissCounter () [inline]`

Definition at line 309 of file environment.h.

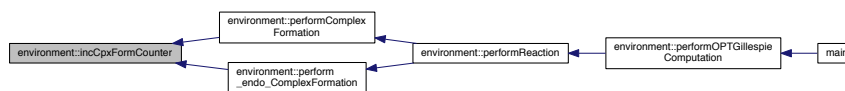
Here is the caller graph for this function:



12.3.3.111 void environment::incCpxFormCounter () [inline]

Definition at line 308 of file environment.h.

Here is the caller graph for this function:



12.3.3.112 void environment::incEndoCleavageCounter () [inline]

Definition at line 305 of file environment.h.

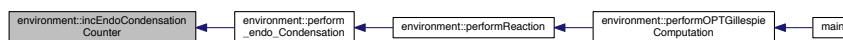
Here is the caller graph for this function:



12.3.3.113 void environment::incEndoCondensationCounter () [inline]

Definition at line 307 of file environment.h.

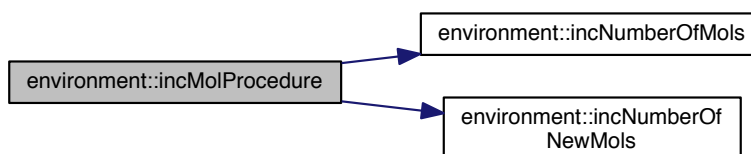
Here is the caller graph for this function:



12.3.3.114 void environment::incMolProcedure (*acs_int tmp_ID*) [inline]

Definition at line 299 of file environment.h.

Here is the call graph for this function:



```

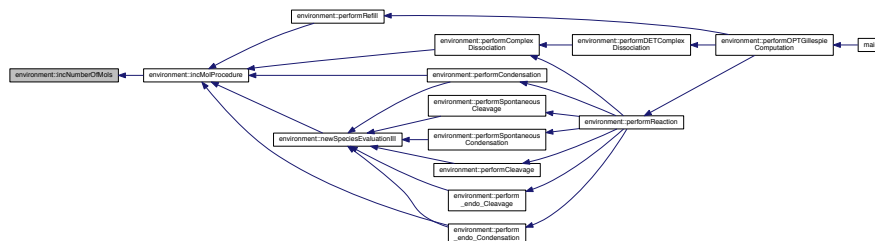
graph LR
    main((main)) --> env_create_initial_mol_pop[environment: createInitial  
MoleculesPopulationFromFileSTD]
    env_create_initial_mol_pop --> env_perform_reaction[environment: performReaction]
    env_perform_reaction --> env_perform_opt_gillespie[environment: performOPTGillespie  
Computation]
    env_perform_opt_gillespie --> env_perform_reaction
    env_perform_reaction --> env_perform_complex_formation[environment: perform  
_endo_ComplexFormation]
    env_perform_complex_formation --> env_perform_reaction
    env_perform_complex_formation --> env_complex_evaluation[environment: complexEvaluation]
    env_complex_evaluation --> env_perform_reaction
    env_perform_reaction --> env_create_initial_mol_pop
    env_perform_reaction --> env_include_nicops[environment: includeNICops]
    env_include_nicops --> env_create_initial_mol_pop

```

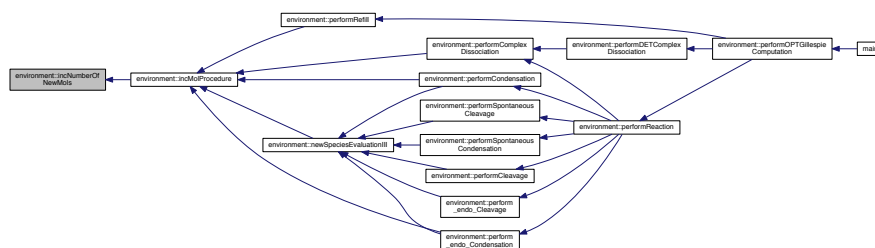
```

graph LR
    main --> OPTGillespieComputation[environment:performOPTGillespieComputation]
    OPTGillespieComputation --> performReaction[environment:performReaction]
    performReaction --> performComplexFormation[environment:performComplexFormation]
    performReaction --> perform_endo_ComplexFormation[environment:perform_endo_ComplexFormation]
    performComplexFormation --> complexEvaluation[environment:complexEvaluation]
    perform_endo_ComplexFormation --> complexEvaluation
    complexEvaluation --> incNumberOfCpdmols[environment:incNumberOfCpdmols]
  
```

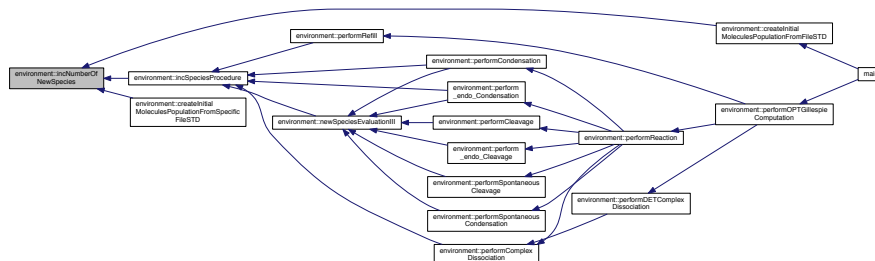
Generated on Tue Dec 10 2013 10:10:07 for CaRNeSS by Doxygen



Definition at line 294 of file environment.h.

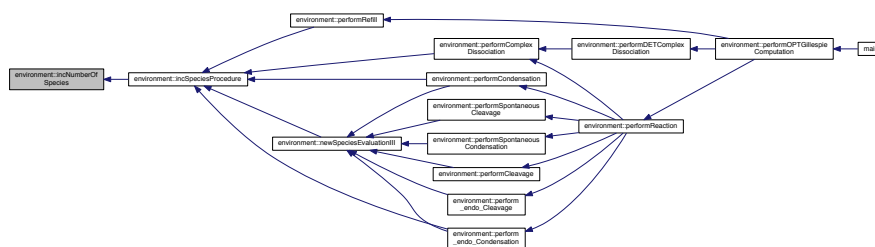


Definition at line 292 of file environment.h.



Definition at line 283 of file environment.h.

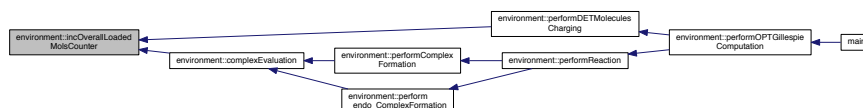
Here is the caller graph for this function:



12.3.3.121 void environment::incOverallLoadedMolsCounter () [inline]

Definition at line 310 of file environment.h.

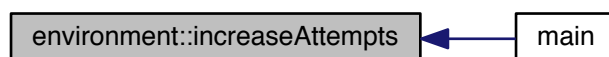
Here is the caller graph for this function:



12.3.3.122 void environment::increaseAttempts () [inline]

Definition at line 346 of file environment.h.

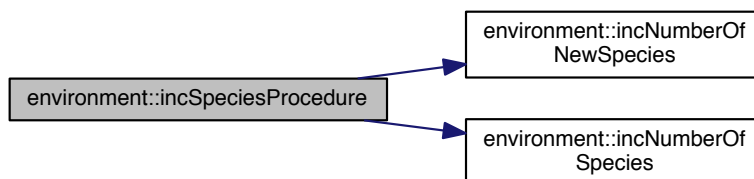
Here is the caller graph for this function:



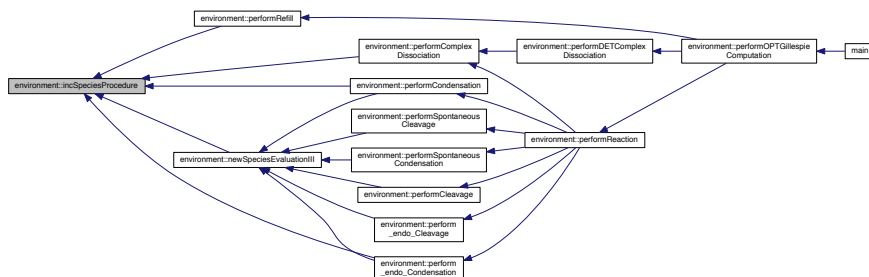
12.3.3.123 void environment::incSpeciesProcedure (acs_int tmp_ID) [inline]

Definition at line 300 of file environment.h.

Here is the call graph for this function:



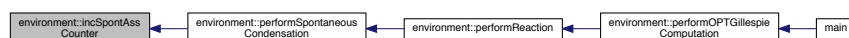
Here is the caller graph for this function:



12.3.3.124 void environment::incSpontAssCounter () [inline]

Definition at line 313 of file environment.h.

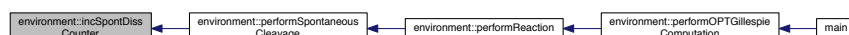
Here is the caller graph for this function:



12.3.3.125 void environment::incSpontDissCounter () [inline]

Definition at line 312 of file environment.h.

Here is the caller graph for this function:



12.3.3.126 void environment::insertSubListInSpecies ()

Insert substrate reactions information in species if complex

Version

1.0

Date

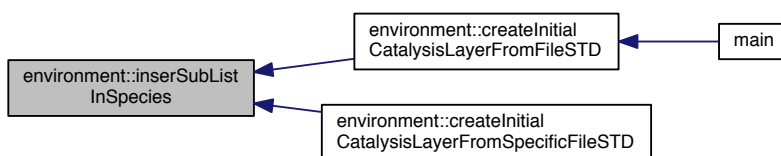
20130917

Author

Alessandro Filisetti

Definition at line 2462 of file environment.cpp.

Here is the caller graph for this function:

**12.3.3.127 bool environment::newSpeciesEvaluationIII (*acs_longInt tmpNewSpecies*, *MTRand* & *tmp__RndDoubleGen*)**

Evaluate new species (Optimized function with new species reactions update process)

Version

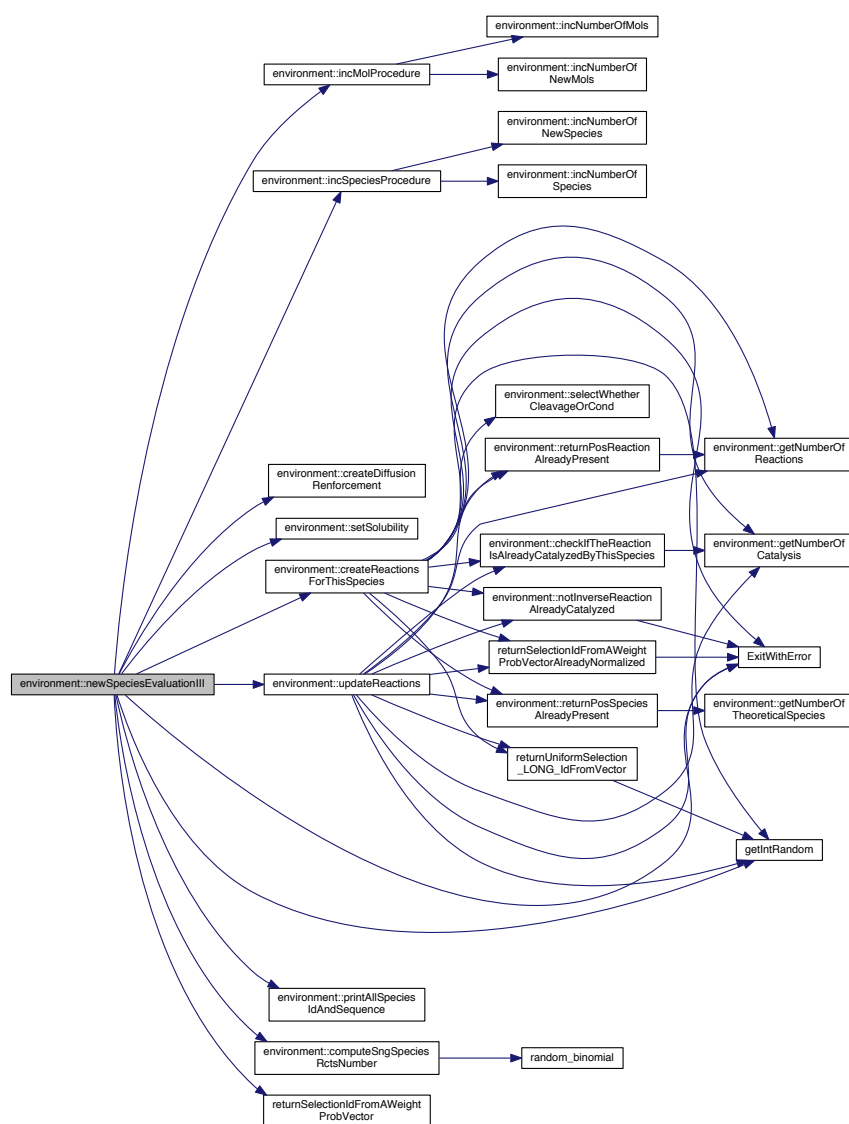
3.0

Parameters

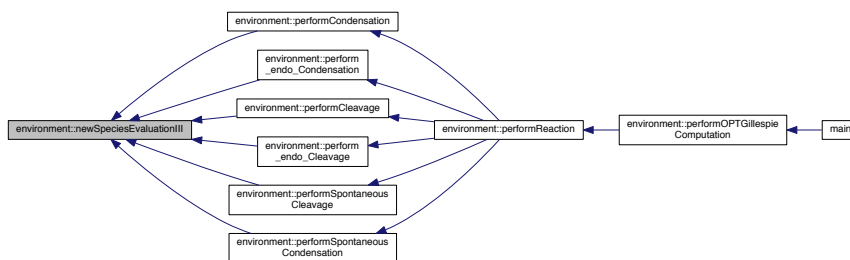
<i>acs_int</i>	tmpNewSpecies New species ID to evaluate
<i>MTRand&</i>	tmp__RndDoubleGen random number generator

Definition at line 5851 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.128 `bool environment::notInverseReactionAlreadyCatalyzed (acs_int tmpRct, acs_longInt tmpID_I, acs_longInt tmpID_II)`

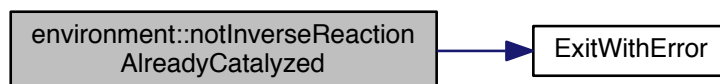
Check if the reaction catalyze both one reaction and the inverted one

Version

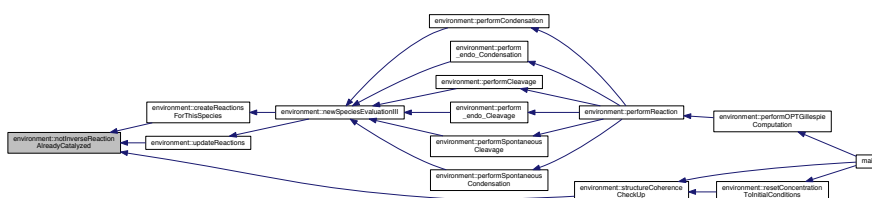
1.0

Definition at line 2578 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.129 `void environment::nutrientsAmountsFixing ()`

Nutrients amount fixing process. The amount of nutrients has to be fixed according to the initial theoretical distribution

Version

1.0

Definition at line 3685 of file environment.cpp.

```
12.3.3.130  bool environment::perform_endo_Cleavage ( acs_longInt tmpSubstrate, acs_longInt tmpProduct_I,  
                                                    acs_longInt tmpProduct_II, acs_int tmpNRGside, acs_longInt tmpIdReaction, acs_longInt tmpIdCatalysis,  
                                                    MTRand & tmp__RndDoubleGen )
```

Perform ENDO_CLEAVAGE reaction

Version

1.2

Date

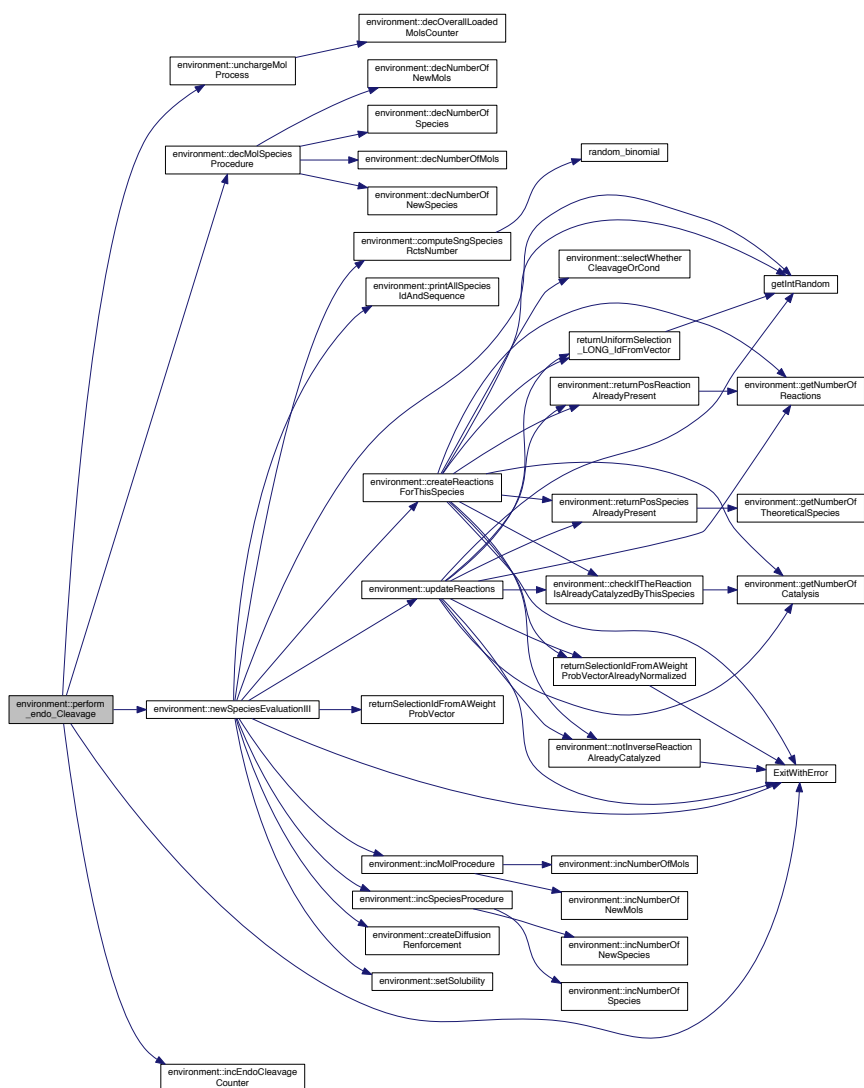
2010.11.08

Parameters

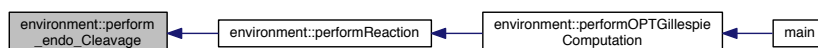
<i>acs_longInt</i>	tmpSubstrate Substrate ID
<i>acs_longInt</i>	tmpProduct_I Product 1 ID
<i>acs_longInt</i>	tmpProduct_II Product 2 ID
<i>acs_int</i>	tmpNrgTarget Energy Target -> catalyst loaded, substrate loaded or both
<i>tmpIdReaction</i>	Rections ID
<i>tmpIdCatalysis</i>	Catalysis ID
<i>MTRand&</i>	tmp__RndDoubleGen random number generator

Definition at line 5258 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.131 bool environment::perform_endo_ComplexFormation (acs_longInt tmpCatalyst, acs_longInt tmpSubstrate, acs_longInt tmpCatID, acs_longInt tmpSecSub, acs_int tmpNRGSide, MTRand & tmp_RndDoubleGen)

Perform ENDO COMPLEX FORMATION reaction

Version

1.2

Date

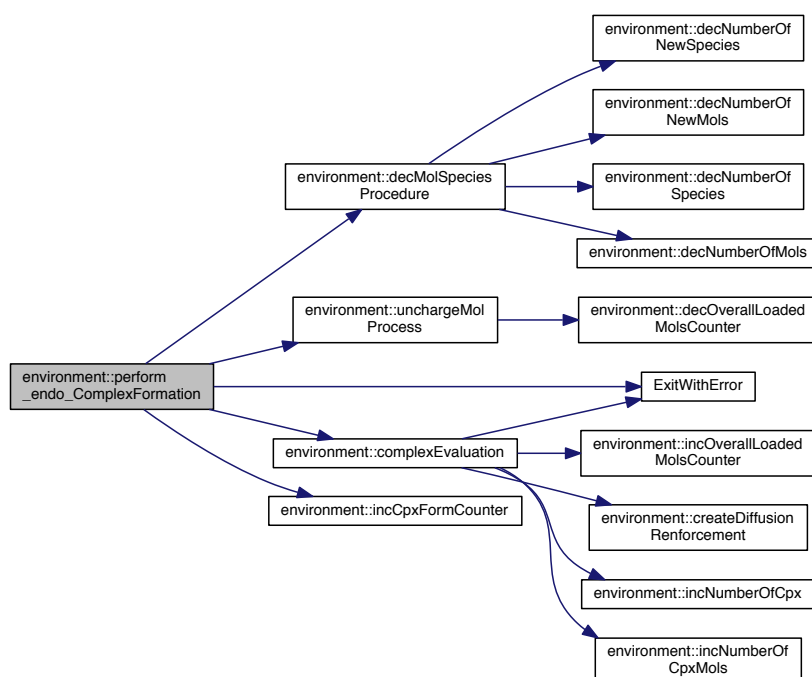
2011.04.13

Parameters

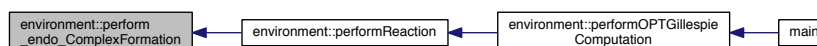
<i>acs_longInt</i>	tmpCatalyst Catalyst ID
<i>acs_longInt</i>	tmpSubstrate Substrate ID Catalysis ID
<i>MTRand</i>	tmp__RndDoubleGen random generator

Definition at line 5467 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



```

12.3.3.132 bool environment::perform_endo_Condensation ( acs_longInt tmpCatalyst, acs_longInt tmpSubstrate,
acs_longInt tmpProduct, acs_longInt tmpComplex, acs_int tmpNRGside, acs_longInt tmpIpdReaction,
acs_longInt tmpIpdCatalysis, MTRand & tmp__RndDoubleGen )

```

Perform ENDO CONDENSATION reaction

Version

1.2

Date

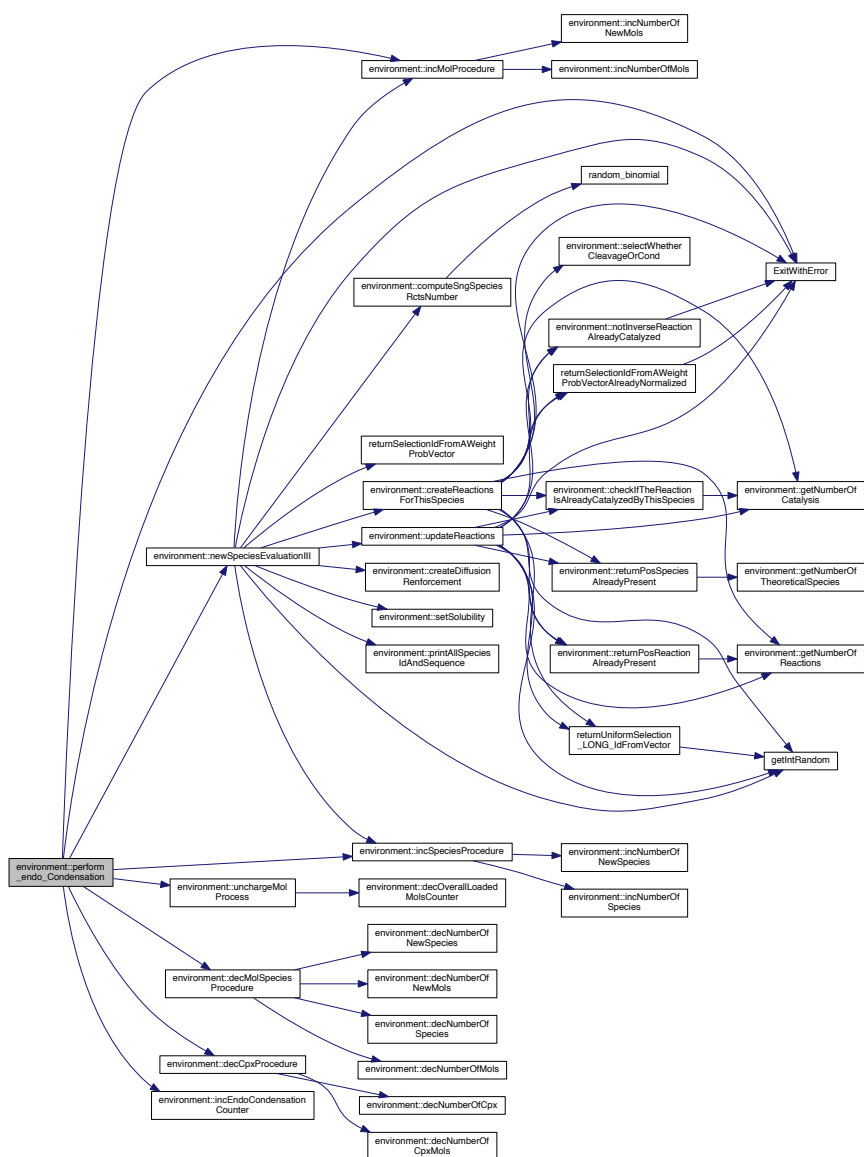
2011-02-12

Parameters

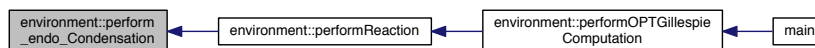
<i>tmpCatalyst</i>	Catalyst (bound in the complex) ID
<i>tmpSubstrate</i>	second (or first, depend on the condensation type) substrate ID
<i>tmpProduct</i>	product ID
<i>acs_longInt</i>	tmpComplex Complex ID
<i>acs_longInt</i>	tmpSubstrate Substrate ID
<i>acs_longInt</i>	tmpProduct Product ID
<i>MTRand</i>	tmp__RndDoubleGen random generator

Definition at line 5015 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.133 `bool environment::performCleavage (acs_longInt tmpSubstrate, acs_longInt tmpProduct_I, acs_longInt tmpProduct_II, acs_longInt tmpIdReaction, acs_longInt tmpIdCatalysis, MTRand & tmp__RndDoubleGen)`

Perform CLEAVAGE reaction

Version

1.2

Date

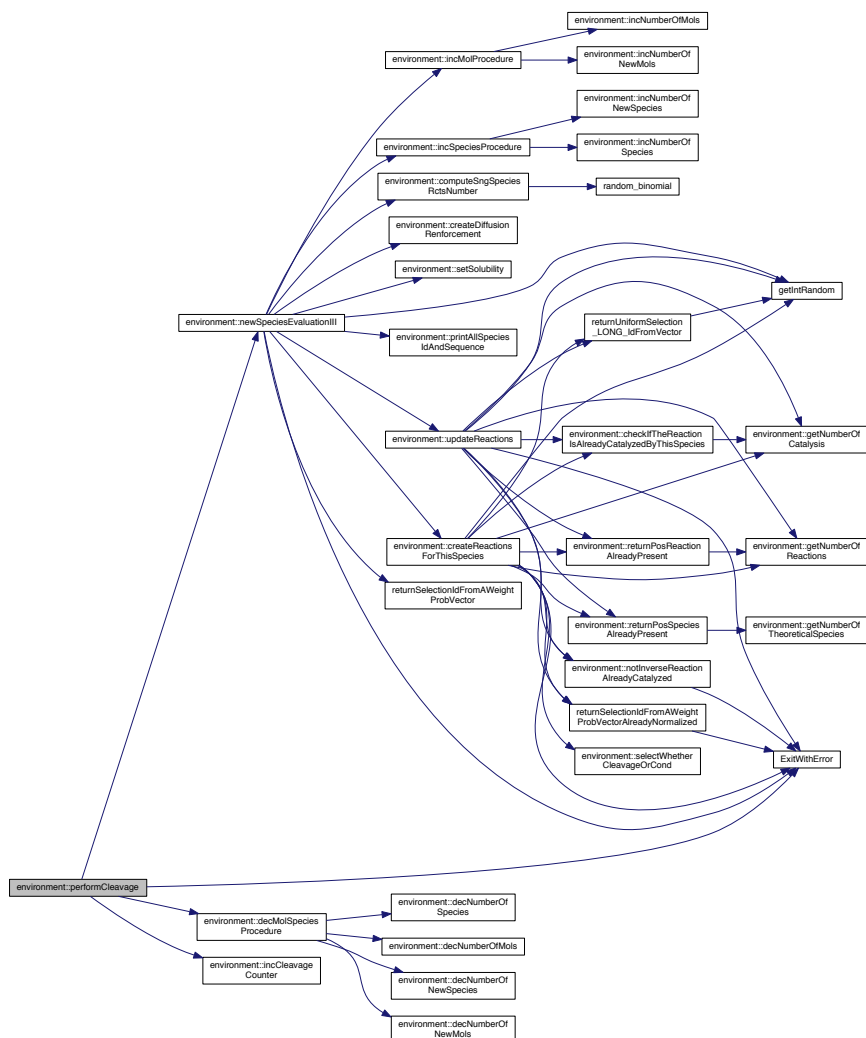
2011.02.12

Parameters

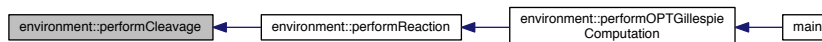
<i>acs_longInt</i>	tmpSubstrate Substrate ID
<i>acs_longInt</i>	tmpProduct_I Product 1 ID
<i>acs_longInt</i>	tmpProduct_II Product 2 ID
<i>tmpIdReaction</i>	Rections ID
<i>tmpIdCatalysis</i>	Catalysis ID
<i>MTRand&</i>	tmp__RndDoubleGen random number generator

Definition at line 5151 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.134 `bool environment::performComplexDissociation (acs_longInt tmpComplex, acs_longInt tmpCatalyst, acs_longInt tmpSubstrate, MTRand & tmp_RndDoubleGen)`

Perform COMPLEX DISASSOCIATION reaction

Version

1.1

Date

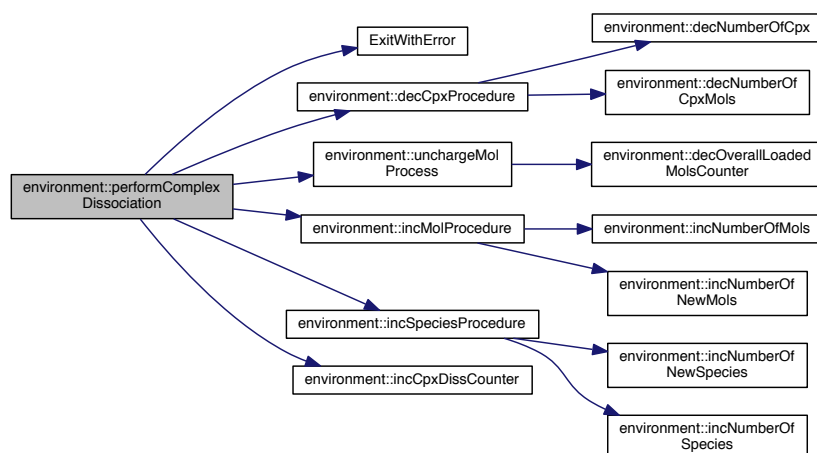
2010.06.08

Parameters

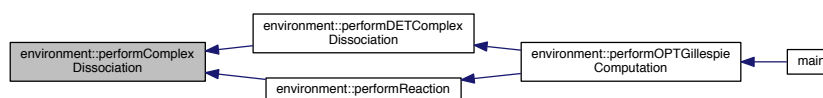
<i>acs_longInt</i>	tmpComplex Complex ID
<i>acs_longInt</i>	tmpCatalyst Catalyst ID
<i>acs_longInt</i>	tmpSubstrate Substrate ID
<i>MTRand</i> &	tmp__RndDoubleGen random generator

Definition at line 5565 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.135 `bool environment::performComplexFormation (acs_longInt tmpCatalyst, acs_longInt tmpSubstrate, acs_longInt tmpCatID, acs_longInt tmpSecSub, MTRand & tmp__RndDoubleGen)`

Perform COMPLEX FORMATION reaction

Version

1.2

Date

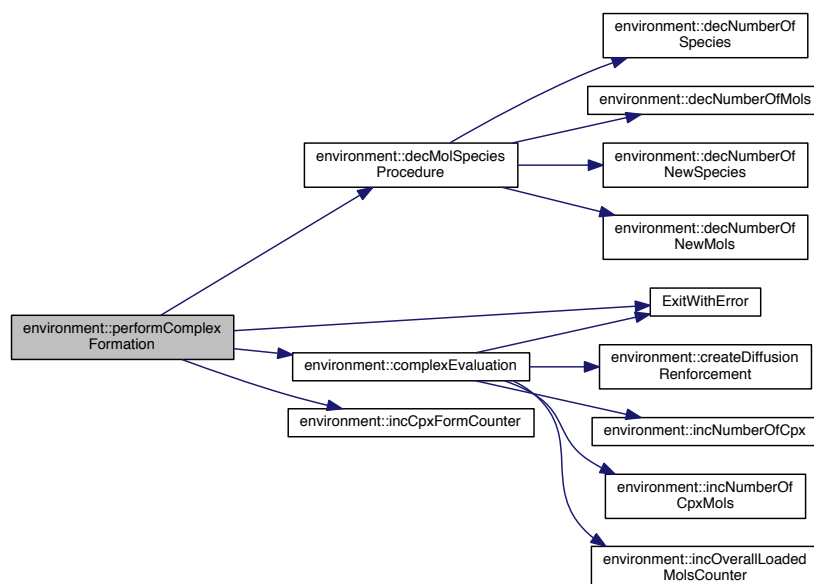
2011.02.13

Parameters

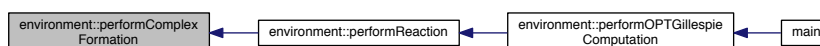
<i>acs_longInt</i>	tmpCatalyst Catalyst ID
<i>acs_longInt</i>	tmpSubstrate Substrate ID Catalysis ID
<i>MTRand&</i>	tmp__RndDoubleGen random generator

Definition at line 5379 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.136 `bool environment::performCondensation (acs_longInt tmpCatalyst, acs_longInt tmpSubstrate, acs_longInt tmpProduct, acs_longInt tmpComplex, acs_longInt tmpIpdReaction, acs_longInt tmpIpdCatalysis, MTRand & tmp__RndDoubleGen)`

Perform CONDENSATION reaction

Version

1.2

Date

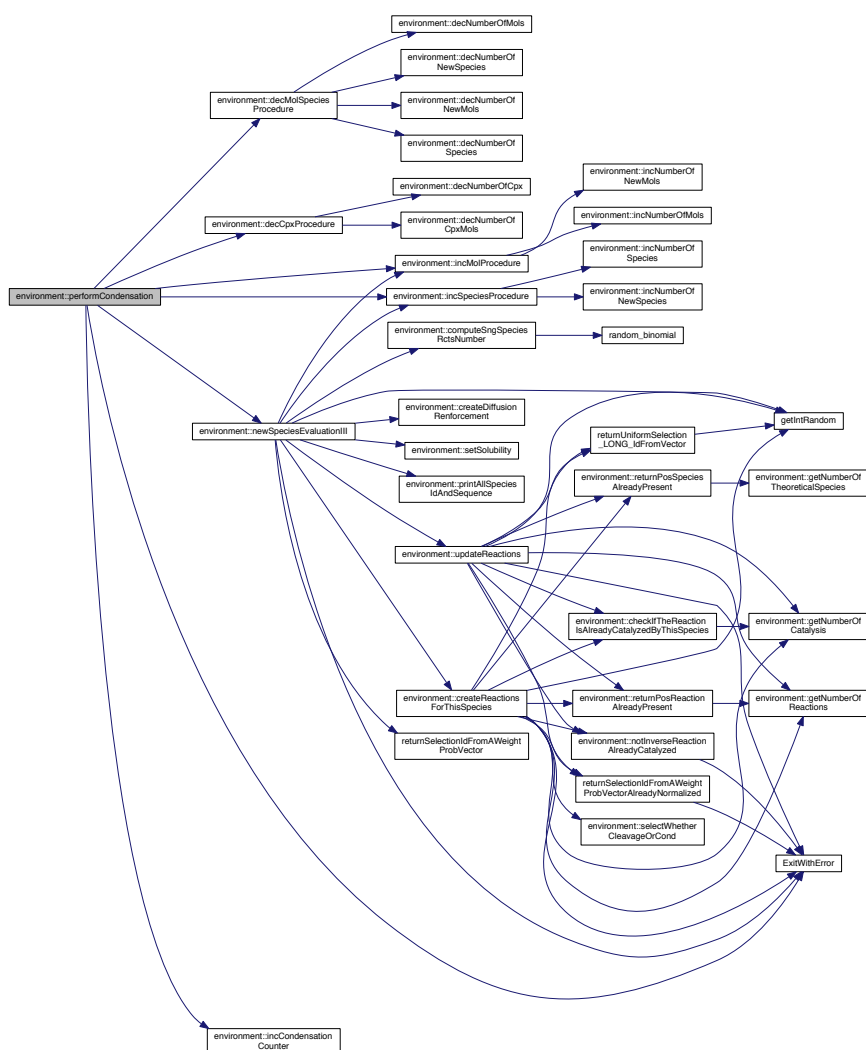
2011-02-12

Parameters

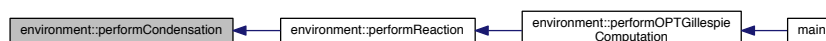
<i>acs_longInt</i>	tmpComplex Complex ID
<i>acs_longInt</i>	tmpSubstrate Substrate ID
<i>acs_longInt</i>	tmpProduct Product ID
<i>MTRand&</i>	tmp__RndDoubleGen random generator

Definition at line 4898 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.137 `bool environment::performDETComplexDissociation (acs_double tmpTimeInterval, MTRand & tmp_RndDoubleGen)`

This function perform the deterministic dissociation of the complex

Version

1.0

Date

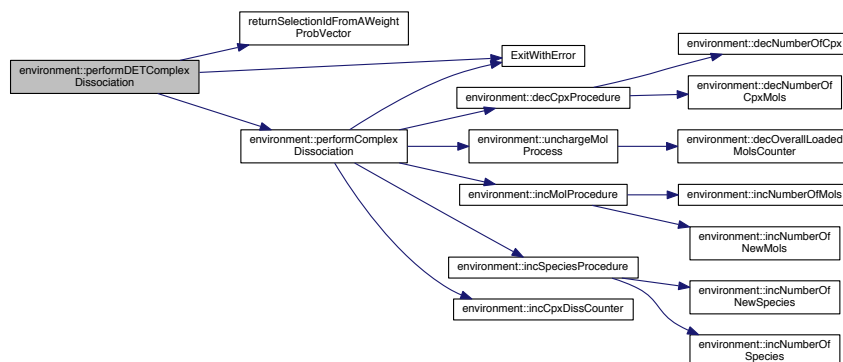
2013-09-18

Parameters

<i>tmpTimeInterval</i>	time elapsed since the last reaction
<i>MTRand</i>	&tmp_RndDoubleGen random number generator

Definition at line 3809 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.138 `bool environment::performDETMoleculesCharging (acs_double tmpTimeInterval, MTRand & tmp_RndDoubleGen)`

This function perform the pseudo-deterministic molecules charging process

Version

2.6

Date

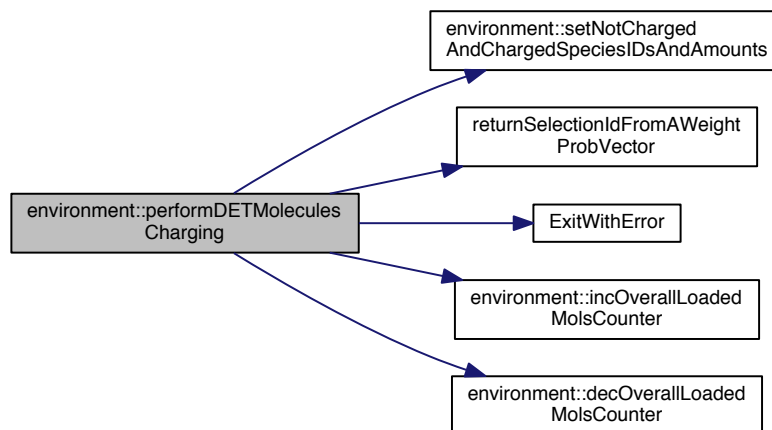
2011-02-24

Parameters

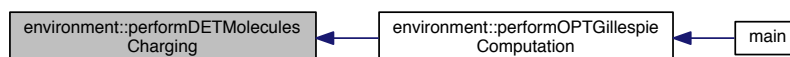
<i>tmpTimeInterval</i>	time elapsed since the last reaction
<i>MTrand</i>	&tmp_RndDoubleGen random number generator

Definition at line 3937 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.139 bool environment::performEnergyEfflux (MTRand & tmp_RndDoubleGen)

Perform ENERGY EFFLUX reaction

Version

2.4.1

Date

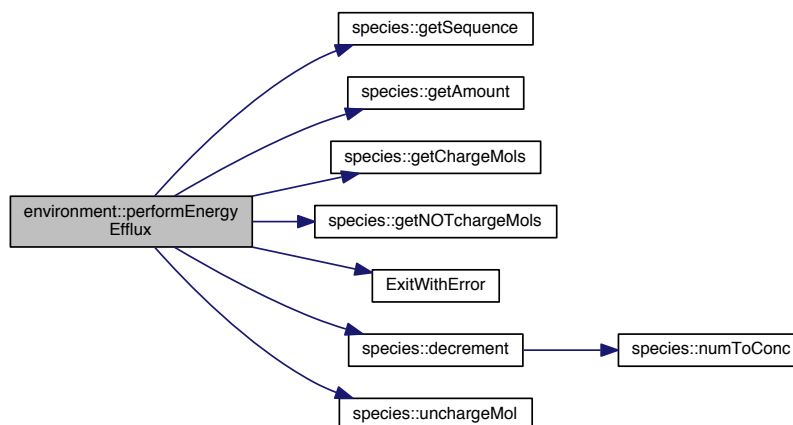
2010-06-27

Parameters

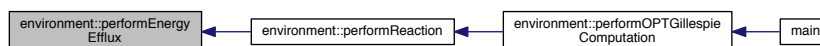
<i>MTRand</i> &	tmp__RndDoubleGen random number generator
-----------------	---

Definition at line 4611 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.140 `bool environment::performMoleculeEfflux (acs_longInt tmpSpecies, MTRand & tmp__RndDoubleGen)`

Perform MOLECULE EFFLUX reaction

Version

2.5.1

Date

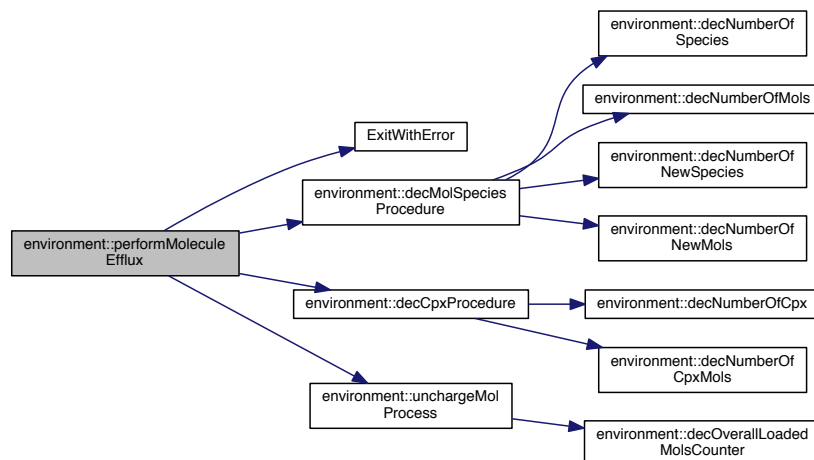
2010-06-27

Parameters

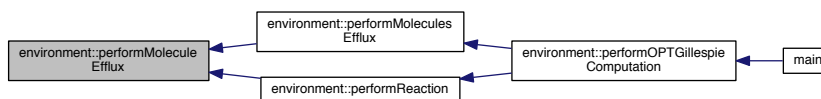
<i>acs_longInt</i>	tmpSpecies Species ID
<i>MTRand</i> &	tmp__RndDoubleGen random generator

Definition at line 4531 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.141 `bool environment::performMoleculesEfflux (acs_double tmpTimeInterval, MTRand & tmp_RndDoubleGen)`

This function perform the deterministic molecules efflux process

Version

2.5

Date

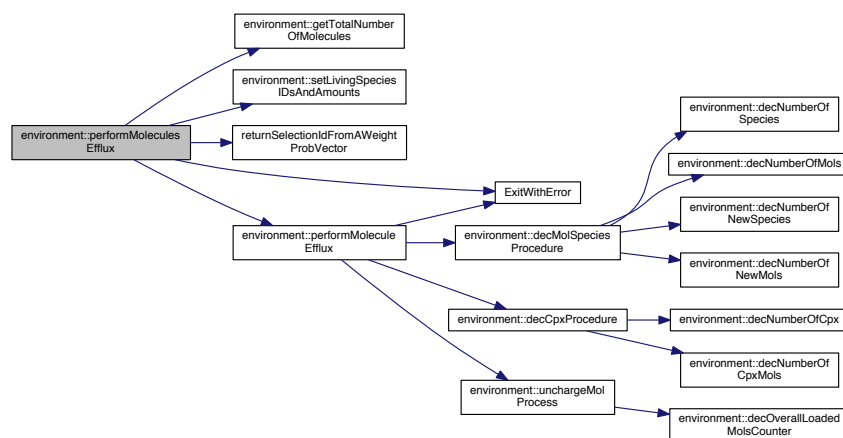
2010-06-25

Parameters

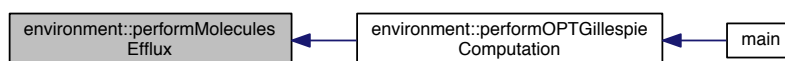
<i>tmpTimeInterval</i>	time elapsed since the last reaction
<i>MTrand</i>	&tmp_RndDoubleGen random number generator

Definition at line 4369 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.142 `bool environment::performOPTGillespieComputation (MTrand & tmpRndDoubleGen, clock_t & tmpTimeElapsed, acs_int tmpActGEN, acs_int tmpActSIM, acs_int tmpActSTEP, string tmpStoringPath)`

Perform all the gillespie algorithm procedure (OPTIMIZED VERSION)

Version

1.0

Date

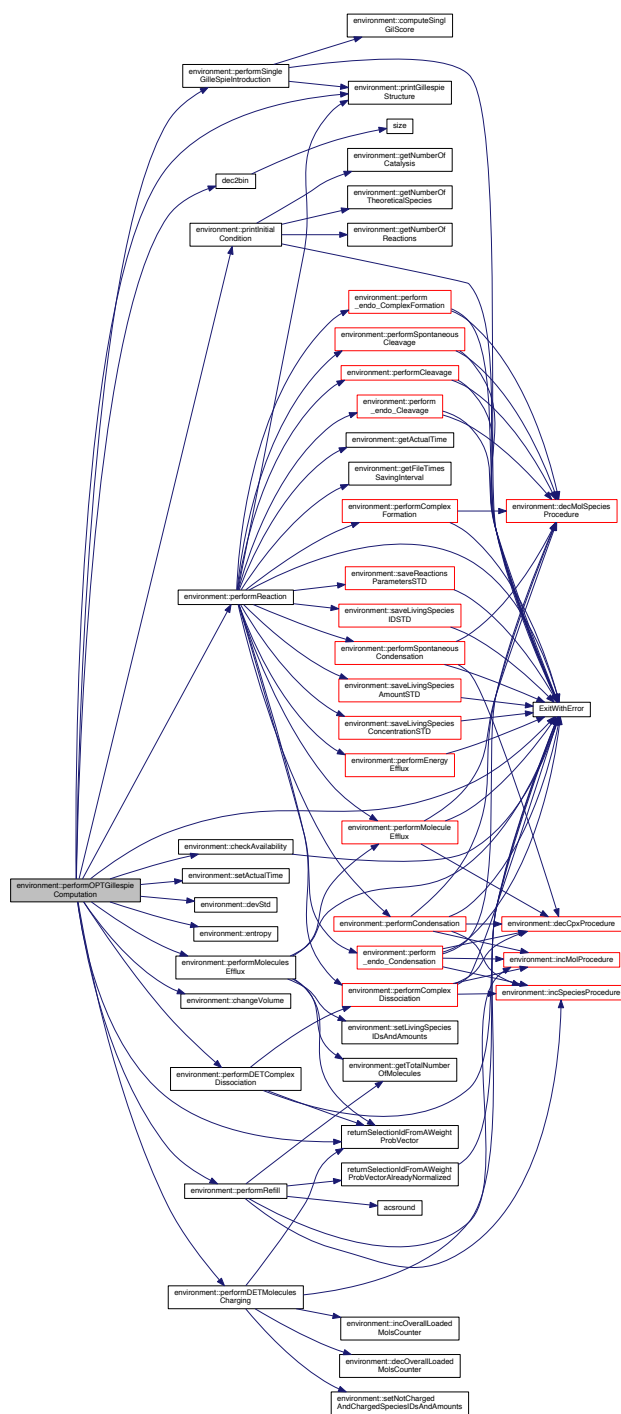
2013.09.17

Parameters

<i>tmpRndDouble-Gen</i>	random numbers generator
<i>tmpTimeElapsed</i>	Computation time elapsed from the T0
<i>tmpActGEN</i>	actual generation
<i>tmpActSIM</i>	actual simulation
<i>tmpActSTEP</i>	actual step (reaction)
<i>tmpStoringPath</i>	path where results are stored

Definition at line 2668 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.143 `bool environment::performReaction (acs_longInt reaction_u, MTRand & tmp_RndDoubleGen, acs_int tmp_ActGEN, acs_int tmp_ActSIM, acs_int tmp_ActSTEP, string tmp_StoringPath)`

Perform the reaction after the Gillespie computation

Version

1.1

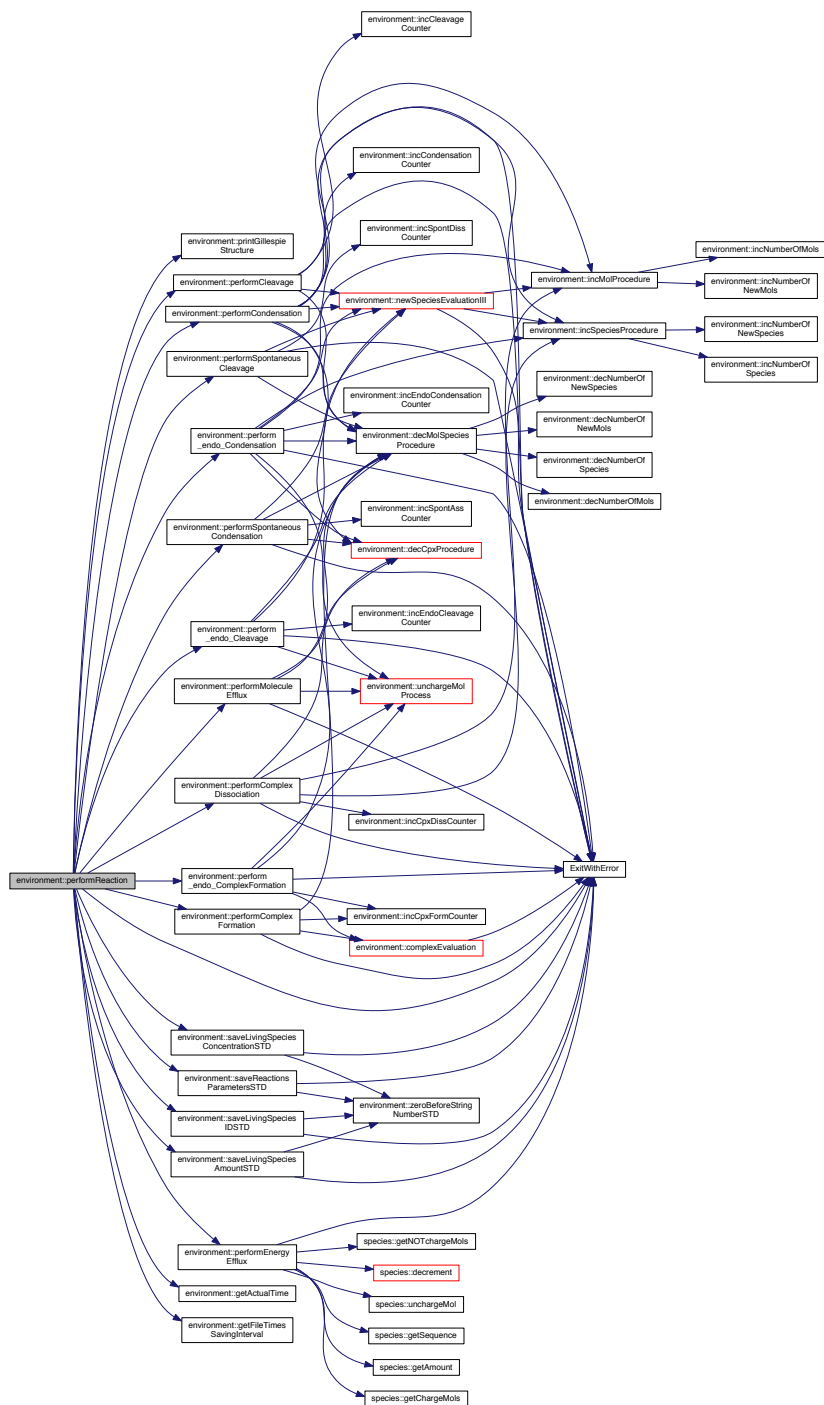
Date

2013-10-29

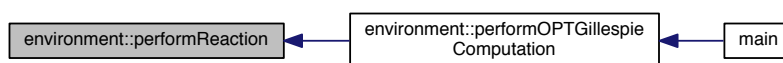
Parameters

<i>acs_longInt</i>	reaction_u reaction ID in Gillespie structure
<i>MTRand&</i>	tmp_RndDoubleGen Random number generator
<i>acs_int</i>	tmp_ActGEN Current generation
<i>acs_int</i>	tmp_ActSIM Current generation
<i>acs_int</i>	tmp_ActSTEP Current step (reaction)
<i>string</i>	Storing directory

Definition at line 4668 of file environment.cpp.



Here is the caller graph for this function:



12.3.3.144 **bool** environment::performRefill (**acs_double** *tmpTimeSinceTheLastInFlux*, **acs_double** *tmpMinimalTimeForOneMols*, **MTRand** & *tmp_RndDoubleGen*)

Perform Refill according to the time interval and the total amount of refill

Version

2.4

Date

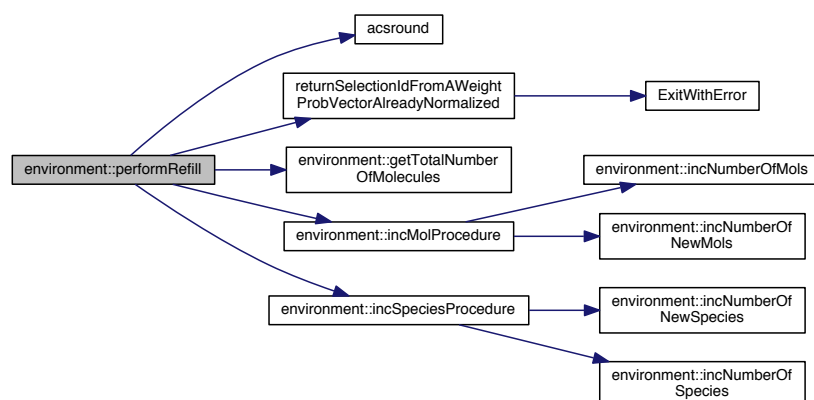
2010.06.10

Parameters

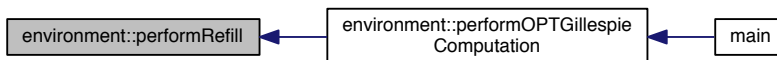
<i>tmpTimeSinceTheLastInFlux</i>	time elapsed since the last influx of at least one molcolule
<i>tmpMinimalTimeForOneMols</i>	time necessary to feed to the system one molecule
<i>tmp_RndDoubleGen</i>	random number generator

Definition at line 3718 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.145 `void environment::performSingleGilleSpielIntroduction (acs_longInt tmpAmountI, acs_longInt tmpAmountII, acs_longInt tmpIDI, acs_longInt tmpIDII, acs_longInt tmpIDCatalysis, acs_int tmp_rctType, acs_longInt tmpMol_I, acs_longInt tmpMol_II, acs_longInt tmpMol_III, acs_longInt tmpMol_IV, acs_int tmp_NRGDirection, acs_longInt tmpRctID, bool tmpSameSpeciesControl)`

Compute and introduce a single Gillespie entry within the Gillespie Structure

Version

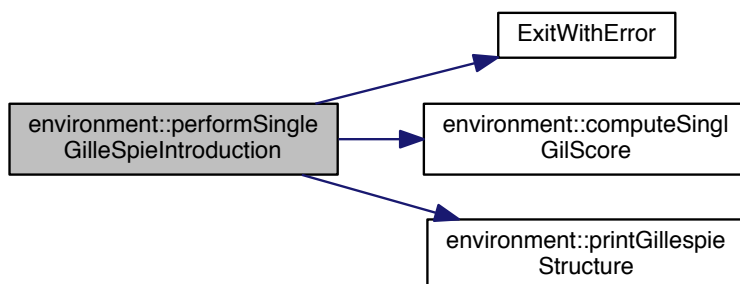
1.0

Date

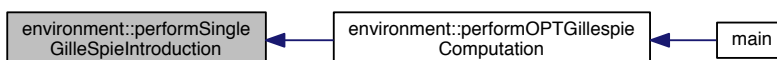
20110222

Definition at line 3314 of file `environment.cpp`.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.146 `bool environment::performSpontaneousCleavage (acs_longInt tmpReaction, MTRand & tmp__RndDoubleGen)`

Perform SPONTANEOUS CLEAVAGE reaction

Version

1.0

Date

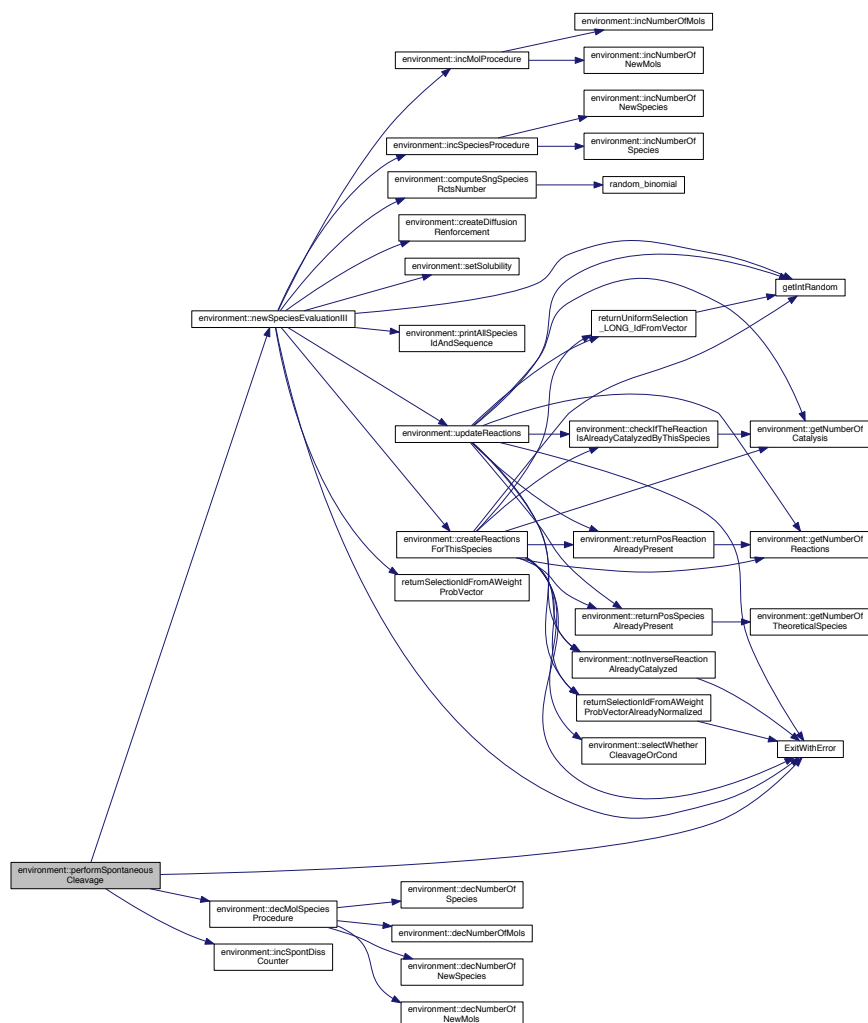
2013.10.28

Parameters

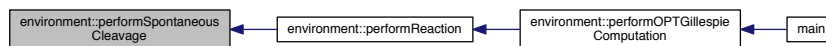
<i>acs_longInt</i>	<i>tmpReaction</i> Reaction ID
<i>MTRand</i> &	<i>tmp__RndDoubleGen</i> random generator

Definition at line 5657 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.147 `bool environment::performSpontaneousCondensation (acs_longInt tmpReaction, MTRand & tmp_RndDoubleGen)`

Perform SPONTANEOUS CONDENSATION reaction

Version

1.0

Date

2013-10-28

Parameters

<i>acs_longInt</i>	tmpReaction Reaction ID
<i>MTRand&</i>	tmp__RndDoubleGen random generator

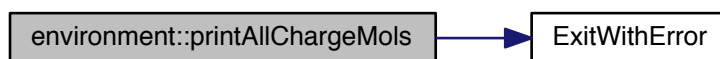
Definition at line 5727 of file environment.cpp.

Date

2010-11-14

Definition at line 6381 of file environment.cpp.

Here is the call graph for this function:



12.3.3.149 void environment::printAllSpeciesIdAndSequence ()

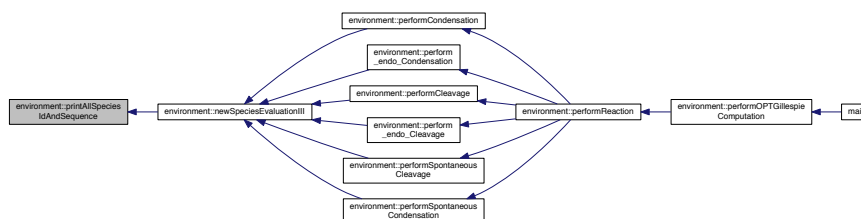
Show all the species with their ID

Version

1.0

Definition at line 6325 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.150 void environment::printGillespieStructure ()

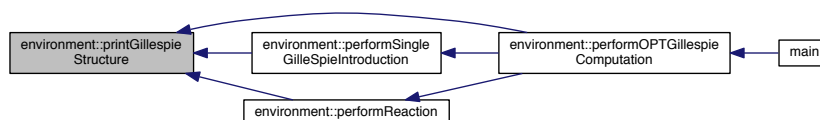
Show all the Gillespie Structure

Version

1.0

Definition at line 6353 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.151 void environment::printInitialCondition ()

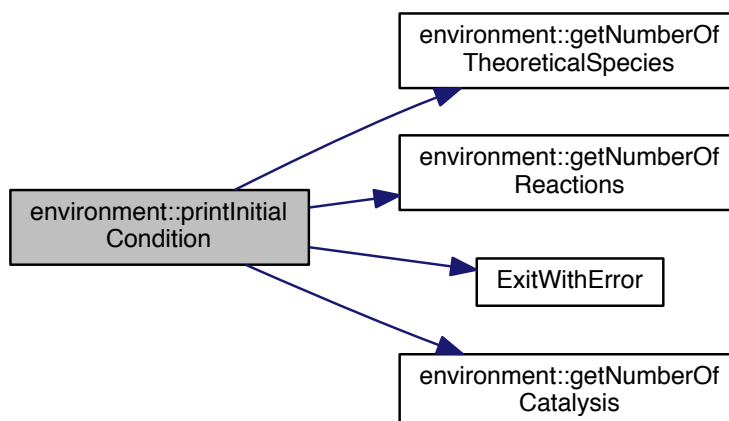
Show all initial species in table format

Version

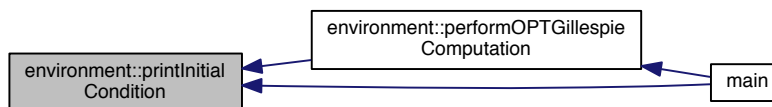
1.0

Definition at line 6267 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.152 void environment::printNutrientsAndProbability ()

This Function populates two vectors containing all the nutrients and all the probabilities for the influx selected species from the firing disk according to the max length of the influx This function print to monitor the content of the vectors `nutrientsForInflux` and `nutrientsProb2BeSelected`

Version

1.0

Date

2010-05-17

Definition at line 3660 of file `environment.cpp`.

12.3.3.153 bool environment::removeChargeMolFromList (*acs_int tmpSpeciesID*)

Perform vector `unchargedIDlist` update removing a new charge molecule vectors `unchargedIDlist` and `cum-UnchargedAmountList` are involved

Version

1.0

Date

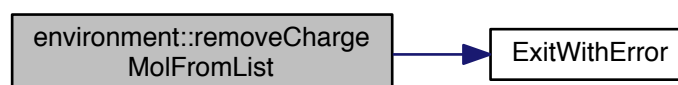
2010-10-10

Parameters

<i>acs_int</i>	<code>tmpSpeciesID</code> Specie to uncharge
----------------	--

Definition at line 4265 of file `environment.cpp`.

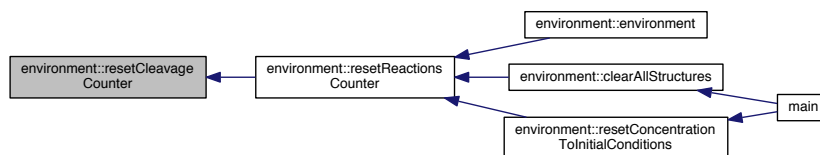
Here is the call graph for this function:



12.3.3.154 void environment::resetCleavageCounter () [inline]

Definition at line 315 of file environment.h.

Here is the caller graph for this function:



12.3.3.155 void environment::resetConcentrationToInitialConditions ()

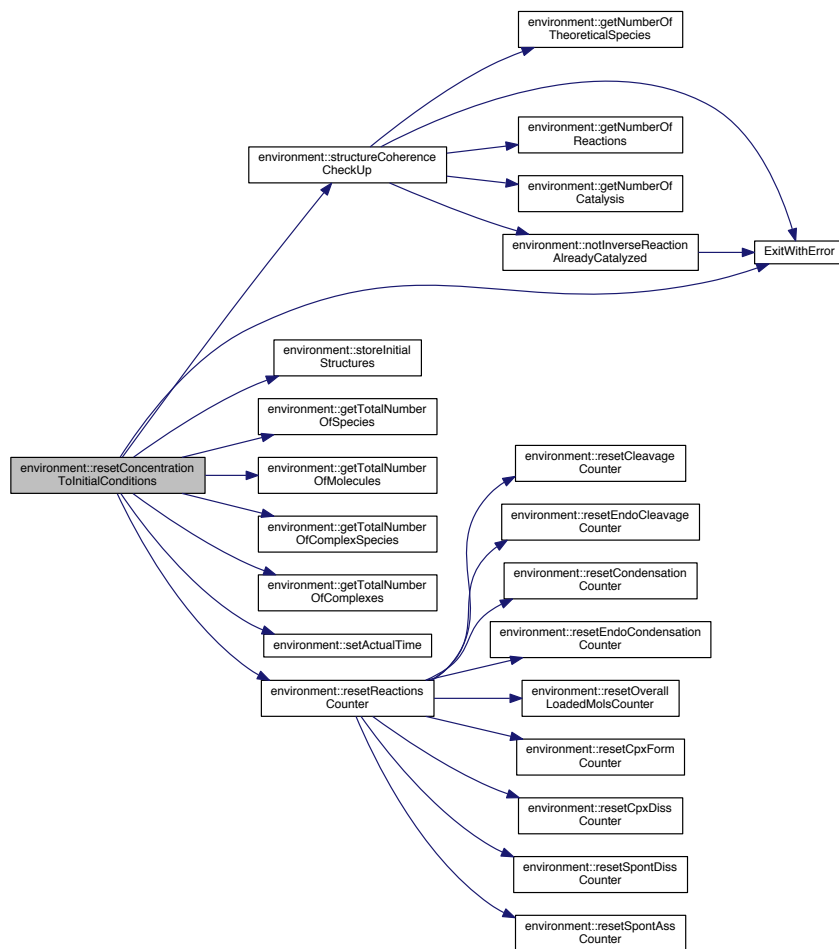
set the concentrations to the initial values and reset internal statistics and counter

Version

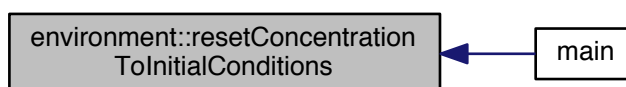
1.0

Definition at line 6472 of file environment.cpp.

Here is the call graph for this function:



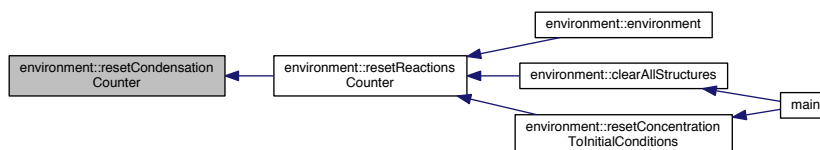
Here is the caller graph for this function:



12.3.3.156 `void environment::resetCondensationCounter () [inline]`

Definition at line 317 of file `environment.h`.

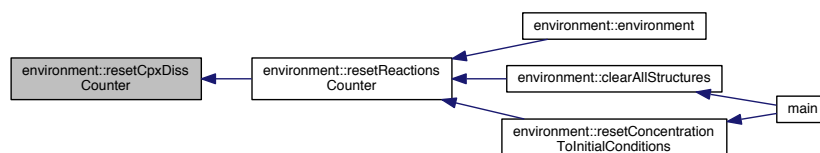
Here is the caller graph for this function:



12.3.3.157 void environment::resetCpxDissCounter () [inline]

Definition at line 321 of file `environment.h`.

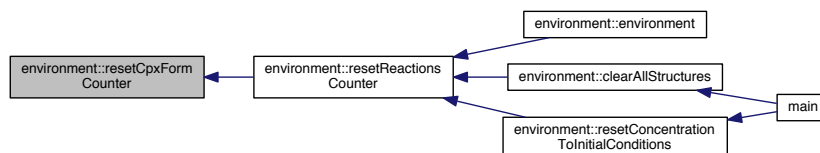
Here is the caller graph for this function:



12.3.3.158 void environment::resetCpxFormCounter () [inline]

Definition at line 320 of file `environment.h`.

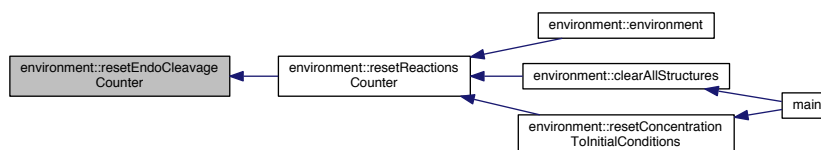
Here is the caller graph for this function:



12.3.3.159 void environment::resetEndoCleavageCounter () [inline]

Definition at line 316 of file `environment.h`.

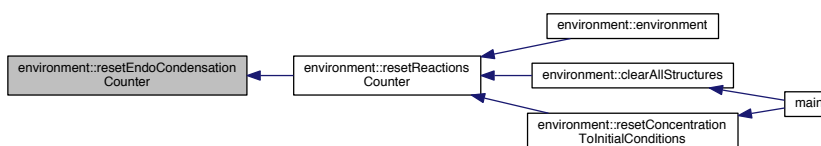
Here is the caller graph for this function:



12.3.3.160 void environment::resetEndoCondensationCounter () [inline]

Definition at line 318 of file environment.h.

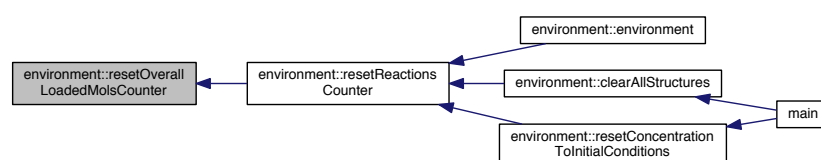
Here is the caller graph for this function:



12.3.3.161 void environment::resetOverallLoadedMolsCounter () [inline]

Definition at line 319 of file environment.h.

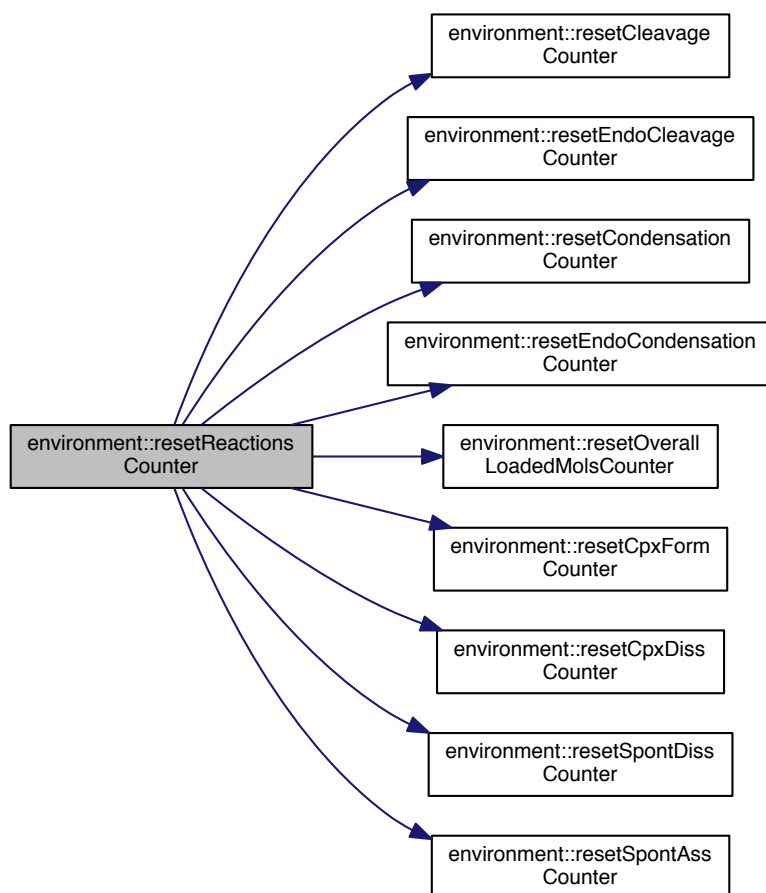
Here is the caller graph for this function:



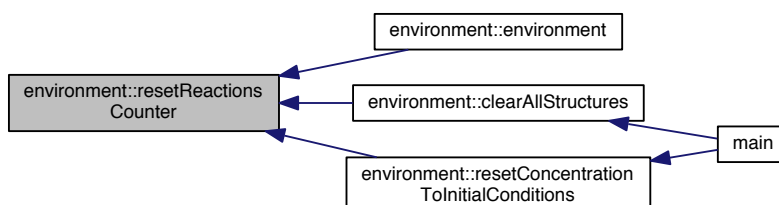
12.3.3.162 void environment::resetReactionsCounter () [inline]

Definition at line 325 of file environment.h.

Here is the call graph for this function:



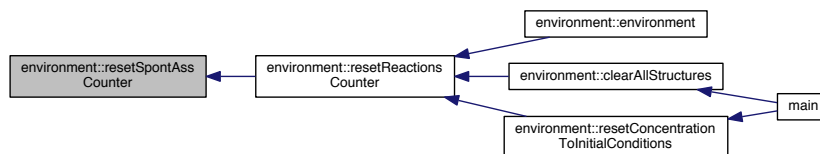
Here is the caller graph for this function:



12.3.3.163 `void environment::resetSpontAssCounter () [inline]`

Definition at line 323 of file `environment.h`.

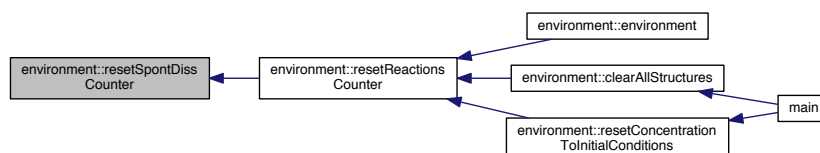
Here is the caller graph for this function:



12.3.3.164 void environment::resetSpontDissCounter () [inline]

Definition at line 322 of file environment.h.

Here is the caller graph for this function:



12.3.3.165 acs_longInt environment::returnPosReactionAlreadyPresent (acs_int tmpReactionType, acs_longInt tmpIds_I, acs_longInt tmpIds_II, acs_longInt tmpIds_III)

Return the reaction ID if the reaction is already present, otherwise it returns the new reaction ID

Version

1.0

Parameters

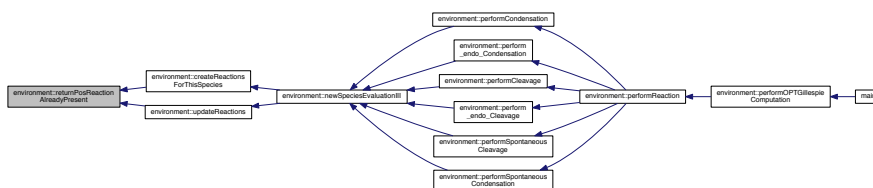
acs_int	tmpReactionType reaction type (cleavage or condensation)
acs_longInt	tmpIds_I species I ID
acs_longInt	tmpIds_II species II ID
acs_longInt	tmpIds_III species III ID

Definition at line 1904 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.166 `acs_longInt environment::returnPosSpeciesAlreadyPresent (string tmpNewSequence)`

This functions returns the ID of the species whether this one is already present, otherwise the new ID is created as the number of elements in the species vector

Version

1.0

Parameters

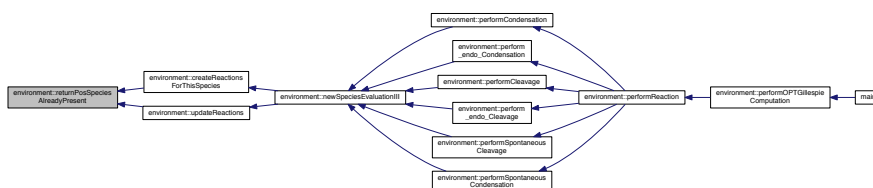
<i>tmpNew-Sequence</i>	New sequence to evaluate
<i>MTRand&</i>	tmp_RndDoubleGen random number generator
<i>tmpNewSpecies-Length</i>	Lenght of the species

Definition at line 1874 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.167 `bool environment::saveCatalysisStructureSTD (acs_int tmpCurrentGen, acs_int tmpCurrentSim, acs_int tmpCurrentStep, string tmpStoringPath)`

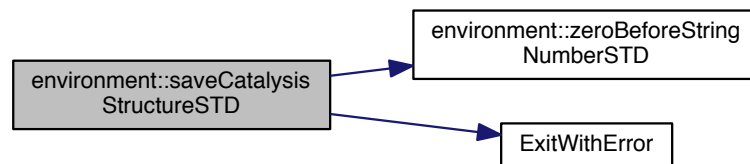
Save the catalysis structures in a file named catalysis_[currentSims]_[currentStep].csv. This file is equal to the acs_catalysis.csv input file. Standard C++ The file is saved in the directory indicated as a second parameter in the run command

Version

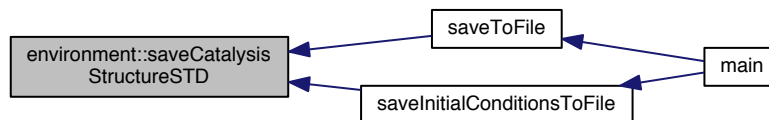
1.0

Definition at line 6978 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.168 `bool environment::saveConfigurationFileSTD (string tmpStoringPath)`

Save a file with the configuration parameters

Version

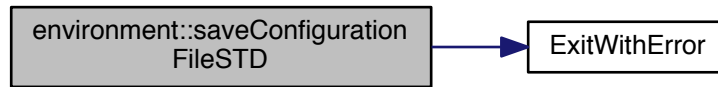
1.0

Date

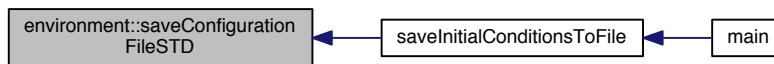
2013/07/03

Definition at line 6613 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.169 `bool environment::saveInfluxStructureSTD (string tmpStoringPath)`

Save influx structure on file, standard C++

Parameters

<i>bool</i>	<code>saveInfluxStructure(QString tmpStoringPath);</code>
-------------	---

Version

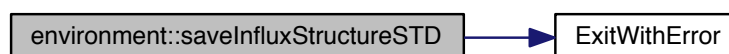
1.0

Date

2013-047-03

Definition at line 6783 of file `environment.cpp`.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.170 `bool environment::saveLivingSpeciesAmountSTD (acs_int tmp_CurrentGen, acs_int tmp_CurrentSim, string tmp_StoringPath)`

Save living species total AMOUNT in a file named `livingAmount_[CurrentGen]_[currentSim].csv`. The file is saved in the directory indicated as a second parameter in the run command - Standard C++

Version

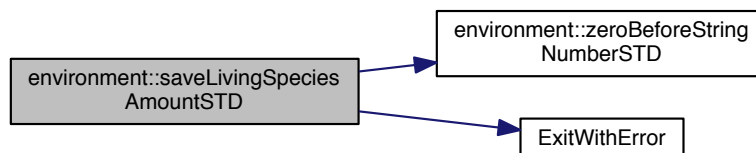
1.0

Date

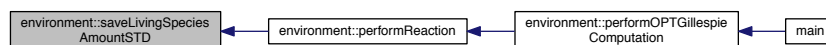
2013/07/03

Definition at line 7242 of file `environment.cpp`.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.171 `bool environment::saveLivingSpeciesConcentrationSTD (acs_int tmp_CurrentGen, acs_int tmp_CurrentSim, string tmp_StoringPath)`

Save living species total CONCENTRATION in a file named `livingAmount_[CurrentGen]_[currentSim].csv`. The file is saved in the directory indicated as a second parameter in the run command

Version

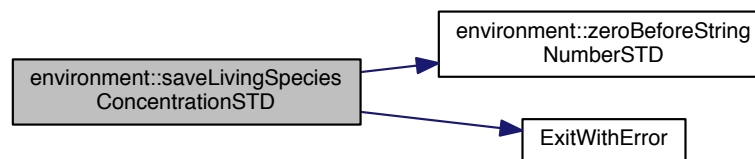
1.0

Date

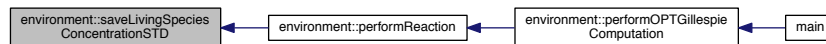
2013/07/03

Definition at line 7308 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.172 `bool environment::saveLivingSpeciesIDSTD (acs_int tmp_CurrentGen, acs_int tmp_CurrentSim, acs_int tmp_CurrentStep, string tmp_StoringPath)`

Save living species in a file named `living_species_[currentSim].csv`. Standard C++ The file is saved in the directory indicated as a second parameter in the run command

Version

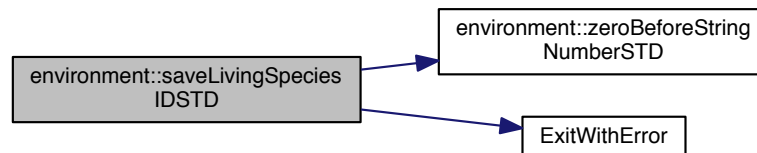
1.0

Date

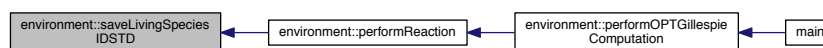
2013/07/03

Definition at line 7181 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.173 bool environment::saveNrgBoolFncStructureSTD (string tmpStoringPath)

Save Energetic Boolean Function on file, standard C++

Parameters

<i>bool</i>	saveInfluxStructure(QString tmpStoringPath);
-------------	--

Version

1.0

Date

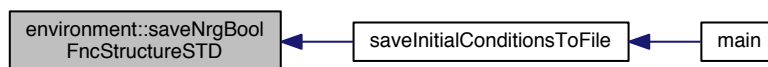
2013-047-03

Definition at line 6821 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.174 `bool environment::saveReactionsParametersSTD (acs_int tmp_CurrentGen, acs_int tmp_CurrentSim, acs_int tmp_CurrentStep, string tmp_StoringPath, acs_int tmpRctType, acs_longInt tmpCat, acs_longInt tmpMol_I, acs_longInt tmpMol_II, acs_longInt tmpMol_III)`

Save the reactions parameters in a file named reactions_parameters_[currentSim].csv. The file is saved in the directory indicated as a second parameter in the run command

Version

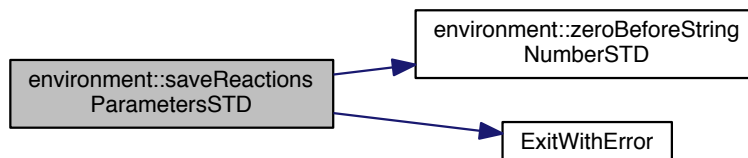
1.0

Date

2013/07/03

Definition at line 7122 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.175 `bool environment::saveReactionsStructureSTD (acs_int tmpCurrentGen, acs_int tmpCurrentSim, acs_int tmpCurrentStep, string tmpStoringPath)`

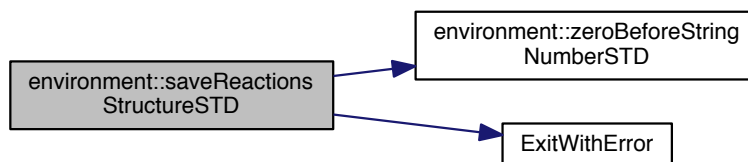
Save the reactions structures in a file named reactions_[currentSims]_[currentStep].csv. This file is equal to the acs_reactions.csv input file. The file is saved in the directory indicated as a second parameter in the run command

Version

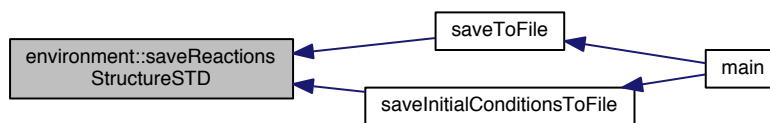
1.0

Definition at line 6924 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.176 `bool environment::saveSpeciesStructureSTD (acs_int tmpCurrentGen, acs_int tmpCurrentSim, acs_int tmpCurrentStep, string tmpStoringPath)`

Save the species structures in a file named `species_[currentSims]_[currentStep].csv`. This file is equal to the `acs_species.csv` input file. C++ standard The file is saved in the directory indicated as a second parameter in the run command

Version

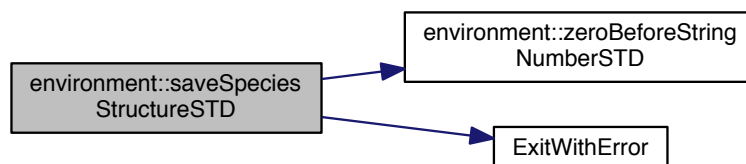
1.0

Date

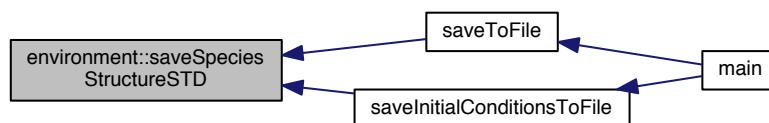
2013/07/03

Definition at line 6859 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.177 `bool environment::saveTimesSTD (acs_int tmpCurrentGen, acs_int tmpCurrentSim, acs_int tmpCurrentStep, string tmpStoringPath)`

Save the reactions times in a file named `times_[currentSim].csv`. Standard C++ The file is saved in the directory indicated as a second parameter in the run command

Version

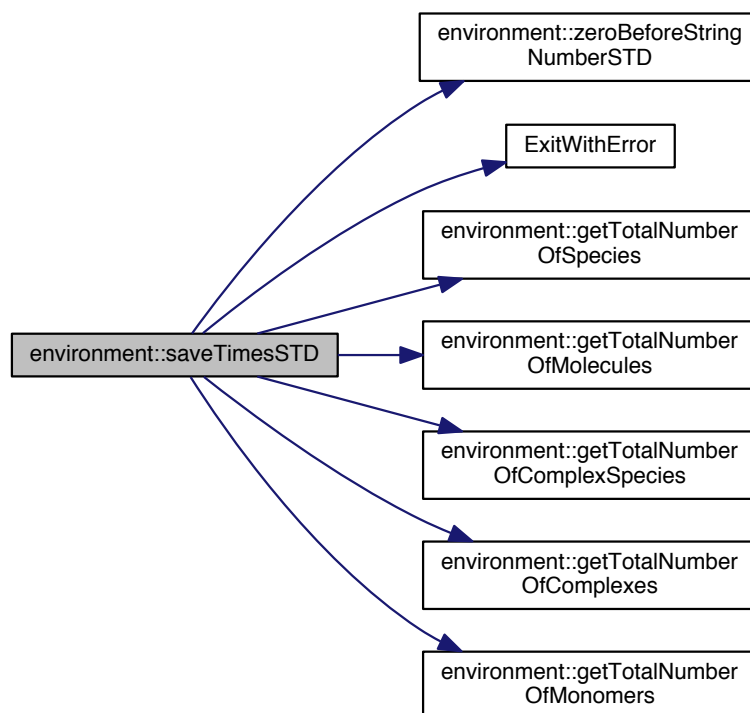
1.0

Date

2013/07/03

Definition at line 7035 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.178 `acs_int environment::selectWhetherCleavageOrCond (MTRand & tmp_RndDoubleGen)`

Select reaction type (cleavage or condensation) according to the cleavage probability

Version

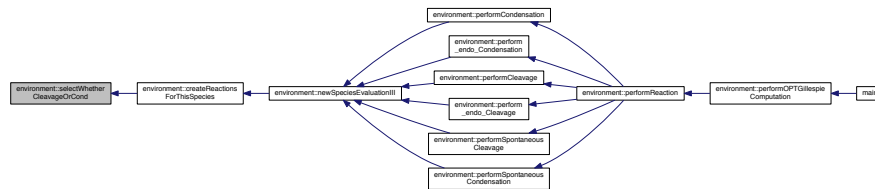
1.0

Parameters

<i>acs_int</i>	tmpTotalNumberOfReactions Total number of conceivable reactions
<i>acs_double</i>	tmpRctsProb reaction probability

Definition at line 1227 of file environment.cpp.

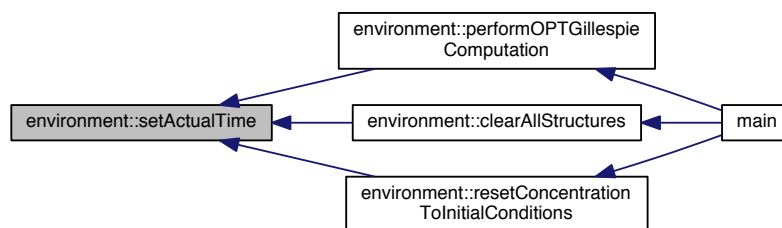
Here is the caller graph for this function:



12.3.3.179 void environment::setActualTime (*acs_double tmpActualTime*) [inline]

Definition at line 343 of file environment.h.

Here is the caller graph for this function:



12.3.3.180 void environment::setLivingSpeciesIDsAndAmounts ()

This function populates to lists, the first containing all the living species ID and the second one containing the cumulative number of living species

Version

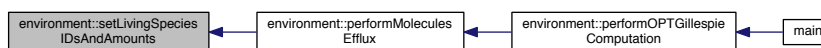
2.5.2

Date

2010-11-11

Definition at line 4176 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.181 void environment::setNotChargedAndChargedSpeciesIDsAndAmounts ()

This function populates to lists, the first one contains the IDs of the uncharged mols and the second one contains the cumulative amount of uncharged molecules

Version

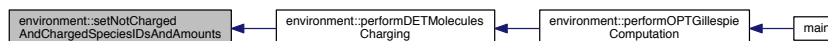
2.5.3

Date

2011-02-22

Definition at line 4217 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.182 bool environment::setSolubility (*acs_int tmpNewSpeciesLength*, *MTRand* & *tmpRndDoubleGen*)

Create the precipitation constant reinforcement according to the species length

Version

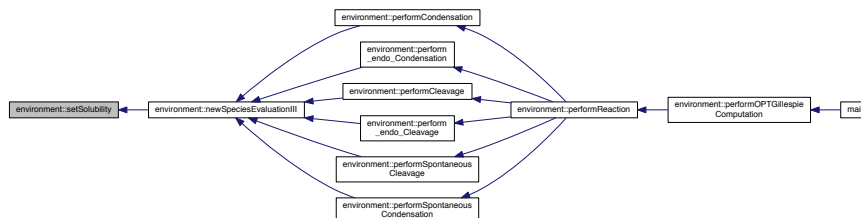
2.5.1

Parameters

<i>tmpPreEnh</i>	precipitation enhancement from parameters
<i>MTRand</i> &	tmp_RndDoubleGen random number generator
<i>tmpNewSpeciesLength</i>	Lenght of the species

Definition at line 1840 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.183 void environment::showGillEngagementInSpecies ()

Function to show the Gillespie engagement of each species.

Version

: 1.0

Date

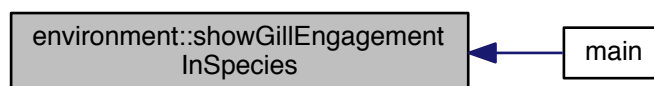
: 20131031

Author

: Alessandro Filisetti

Definition at line 2559 of file environment.cpp.

Here is the caller graph for this function:



12.3.3.184 void environment::showGlobalParameter ()

Shows all parameters uploaded from the configuration file

Version

1.0

Definition at line 6174 of file environment.cpp.

Here is the caller graph for this function:

**12.3.3.185 void environment::showSubListInSpecies ()**

Function to show the substrates list of the complex species

Definition at line 2523 of file environment.cpp.

12.3.3.186 void environment::storeInitialStructures ()

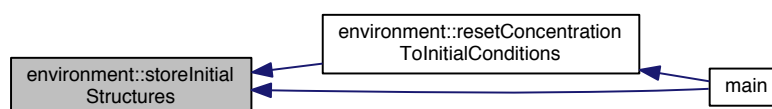
Store initial structures into storing variables

Version

1.0

Definition at line 6567 of file environment.cpp.

Here is the caller graph for this function:

**12.3.3.187 bool environment::structureCoherenceCheckUp ()**

This function check the coherence of the data structures. IF IT FAULTS THE SYSTEM RETURNS ERROR!!! IT IS VERY RECOMMENDED RUN THIS FUNCTION ALWAYS AFTER THE INITIALIZATION

Version

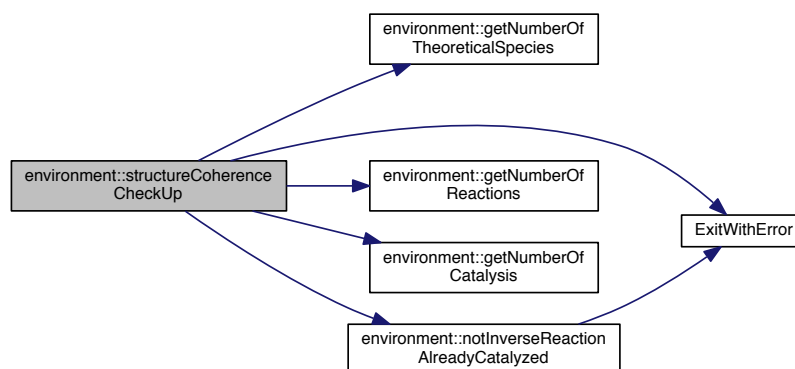
1.1

Date

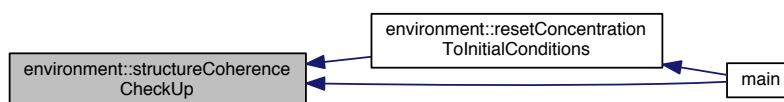
2011-04-13

Definition at line 2004 of file environment.cpp.

Here is the call graph for this function:



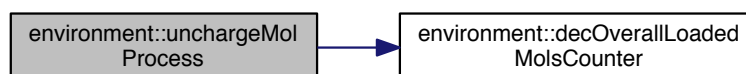
Here is the caller graph for this function:



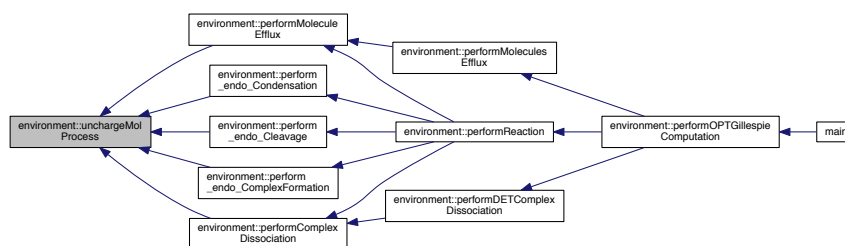
12.3.3.188 void environment::unchargeMolProcess (*acs_int tmp_ID*) [inline]

Definition at line 302 of file environment.h.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.189 `bool environment::updateReactions (acs_longInt tmpIDtoUpdate, acs_longInt tmpNewSpecies, acs_int tmpRctType, vector< acs_longInt > & tmp_AlreadyEvaluatedSpeciesVector, MTRand & tmp_RndDoubleGen)`

Old Species Reactions and Catalysis update process

Version

1.0

Date

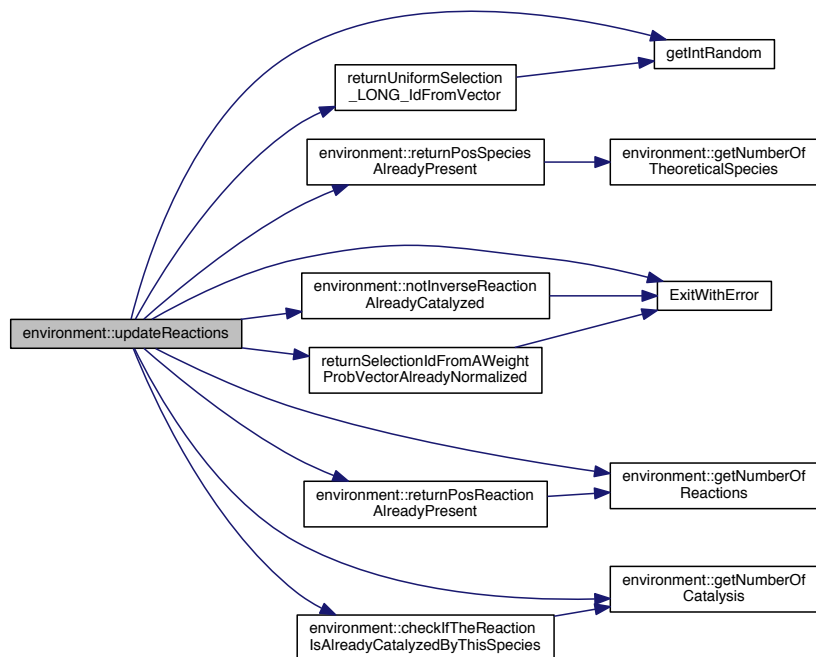
2013/07/30

Parameters

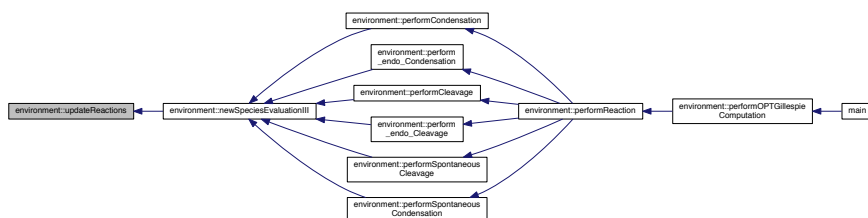
<i>acs_longInt</i>	tmpIDtoUpdate ID of the species to update
<i>acs_longInt</i>	ID of the new species involved in the reactions to update
<i>acs_int</i>	tmpRctType Reaction type, namely cleavage or condensation
<i>MTRand&</i>	tmp____RndDoubleGen Random number generator

Definition at line 941 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.190 void environment::updateSpeciesAges ()

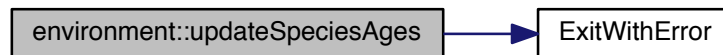
Update the species age

Version

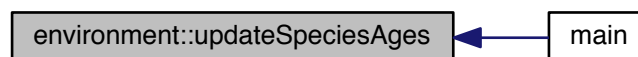
1.0

Definition at line 6125 of file environment.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.3.3.191 string environment::zeroBeforeStringNumberSTD (*acs_int tmpTotN*, *acs_int tmpCurrentN*)

This function creates a chain of zero as STRING according to `tmpTotN` and `tmpCurrent N` in order to make possible a sorting (e.g. `tmpTotN = 1000`, `tmpCurrentN = 3`, return `0003`)

Version

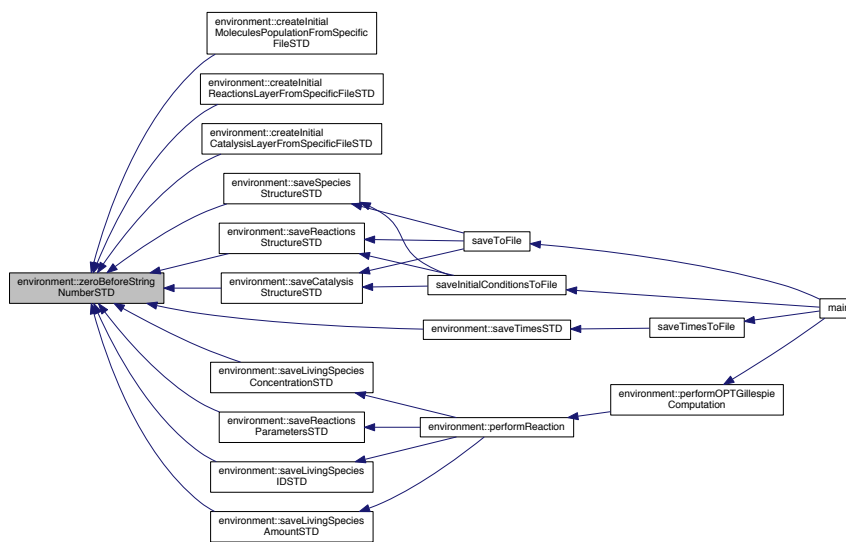
1.0

Parameters

<i>acs_int</i>	tmpTotN Total N
<i>acs_int</i>	tmpCurrentN current N

Definition at line 6590 of file environment.cpp.

Here is the caller graph for this function:



The documentation for this class was generated from the following files:

- /Users/alessandrofilisetti/Documents/GIT/carness/[environment.h](#)
- /Users/alessandrofilisetti/Documents/GIT/carness/[environment.cpp](#)

12.4 gillespie Class Reference

```
#include <gillespie.h>
```

Public Member Functions

- [gillespie](#) ()
- [gillespie](#) ([acs_longInt](#) tmpIDU, [acs_int](#) tmpIDReactionType, [acs_double](#) tmpScore, [acs_longInt](#) tmpMoll, [acs_longInt](#) tmpMolIII, [acs_longInt](#) tmpMolIII, [acs_longInt](#) tmpMolIV, [acs_longInt](#) tmpIDReaction, [acs_longInt](#) tmpIDCatalysis)
- [gillespie](#) ([acs_longInt](#) tmpIDU, [acs_int](#) tmpIDReactionType, [acs_double](#) tmpScore, [acs_longInt](#) tmpMoll, [acs_longInt](#) tmpMolIII, [acs_longInt](#) tmpMolIII, [acs_longInt](#) tmpMolIV, [acs_int](#) tmpNRGside, [acs_longInt](#) tmpIDReaction, [acs_longInt](#) tmpIDCatalysis)
- [~gillespie](#) ()
- [acs_longInt](#) [getID](#) () const
- [acs_int](#) [getIDReactionType](#) () const
- [acs_double](#) [getScore](#) () const

- [acs_longInt getMoll](#) () const
- [acs_longInt getMolII](#) () const
- [acs_longInt getMolIII](#) () const
- [acs_longInt getMolIV](#) () const
- [acs_longInt getReactionID](#) () const
- [acs_longInt getCatalysisID](#) () const
- [acs_int getNRGside](#) () const

12.4.1 Detailed Description

Author

Alessandro Filisetti

Version

0.1

Date

2009-04-21

Definition at line 12 of file gillespie.h.

12.4.2 Constructor & Destructor Documentation

12.4.2.1 gillespie::gillespie ()

12.4.2.2 gillespie::gillespie (*acs_longInt tmpIDU*, *acs_int tmpIDReactionType*, *acs_double tmpScore*, *acs_longInt tmpMoll*, *acs_longInt tmpMolII*, *acs_longInt tmpMolIII*, *acs_longInt tmpMolIV*, *acs_longInt tmpIDReaction*, *acs_longInt tmpIDCatalysis*)

Gillespie Obj constructor

Version

1.0

Parameters

<i>tmpRndDouble-Gen</i>	randomGenerator reference
-------------------------	---------------------------

Definition at line 21 of file gillespie.cpp.

12.4.2.3 gillespie::gillespie (*acs_longInt tmpIDU*, *acs_int tmpIDReactionType*, *acs_double tmpScore*, *acs_longInt tmpMoll*, *acs_longInt tmpMolII*, *acs_longInt tmpMolIII*, *acs_longInt tmpMolIV*, *acs_int tmpNRGside*, *acs_longInt tmpIDReaction*, *acs_longInt tmpIDCatalysis*)

Definition at line 38 of file gillespie.cpp.

12.4.2.4 gillespie::~~gillespie () [inline]

Definition at line 33 of file gillespie.h.

12.4.3 Member Function Documentation

12.4.3.1 `acs_longInt gillespie::getCatalysisID () const` `[inline]`

Definition at line 44 of file gillespie.h.

12.4.3.2 `acs_longInt gillespie::getID () const` `[inline]`

Definition at line 36 of file gillespie.h.

12.4.3.3 `acs_int gillespie::getIdReactionType () const` `[inline]`

Definition at line 37 of file gillespie.h.

12.4.3.4 `acs_longInt gillespie::getMolI () const` `[inline]`

Definition at line 39 of file gillespie.h.

12.4.3.5 `acs_longInt gillespie::getMolII () const` `[inline]`

Definition at line 40 of file gillespie.h.

12.4.3.6 `acs_longInt gillespie::getMolIII () const` `[inline]`

Definition at line 41 of file gillespie.h.

12.4.3.7 `acs_longInt gillespie::getMolIV () const` `[inline]`

Definition at line 42 of file gillespie.h.

12.4.3.8 `acs_int gillespie::getNRGside () const` `[inline]`

Definition at line 45 of file gillespie.h.

12.4.3.9 `acs_longInt gillespie::getReactionID () const` `[inline]`

Definition at line 43 of file gillespie.h.

12.4.3.10 `acs_double gillespie::getScore () const` `[inline]`

Definition at line 38 of file gillespie.h.

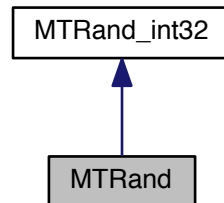
The documentation for this class was generated from the following files:

- [/Users/alessandrofilisetti/Documents/GIT/carness/gillespie.h](#)
- [/Users/alessandrofilisetti/Documents/GIT/carness/gillespie.cpp](#)

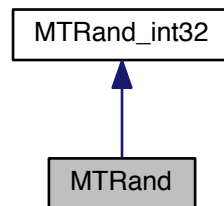
12.5 MTRand Class Reference

```
#include <mtrand.h>
```

Inheritance diagram for MTRand:



Collaboration diagram for MTRand:



Public Member Functions

- [MTRand](#) ()
- [MTRand](#) (unsigned long [seed](#))
- [MTRand](#) (const unsigned long *[seed](#), int [size](#))
- [~MTRand](#) ()
- double [operator\(\)](#) ()

Additional Inherited Members

12.5.1 Detailed Description

Definition at line 97 of file mtrand.h.

12.5.2 Constructor & Destructor Documentation

12.5.2.1 MTRand::MTRand () [inline]

Definition at line 99 of file mtrand.h.

12.5.2.2 MTRand::MTRand (unsigned long *seed*) [inline]

Definition at line 100 of file mtrand.h.

12.5.2.3 MTRand::MTRand (const unsigned long * *seed*, int *size*) [inline]

Definition at line 101 of file mtrand.h.

12.5.2.4 MTRand::~~MTRand () [inline]

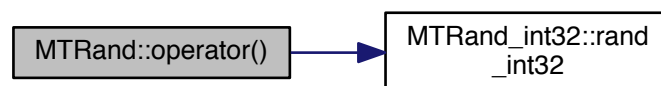
Definition at line 102 of file mtrand.h.

12.5.3 Member Function Documentation

12.5.3.1 double MTRand::operator() () [inline]

Definition at line 103 of file mtrand.h.

Here is the call graph for this function:



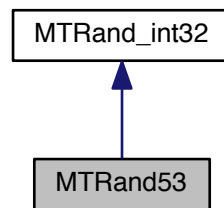
The documentation for this class was generated from the following file:

- [/Users/alessandrofilisetti/Documents/GIT/carness/mtrand.h](#)

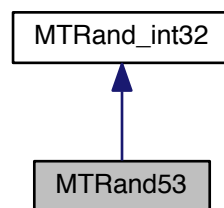
12.6 MTRand53 Class Reference

```
#include <mtrand.h>
```


Inheritance diagram for MTRand53:



Collaboration diagram for MTRand53:



Public Member Functions

- [MTRand53](#) ()
- [MTRand53](#) (unsigned long [seed](#))
- [MTRand53](#) (const unsigned long *[seed](#), int [size](#))
- [~MTRand53](#) ()
- double [operator\(\)](#) ()

Additional Inherited Members

12.6.1 Detailed Description

Definition at line 139 of file mtrand.h.

12.6.2 Constructor & Destructor Documentation

12.6.2.1 MTRand53::MTRand53 () [inline]

Definition at line 141 of file mtrand.h.

12.6.2.2 MTRand53::MTRand53 (unsigned long *seed*) [inline]

Definition at line 142 of file mtrand.h.

12.6.2.3 MTRand53::MTRand53 (const unsigned long * *seed*, int *size*) [inline]

Definition at line 143 of file mtrand.h.

12.6.2.4 MTRand53::~~MTRand53 () [inline]

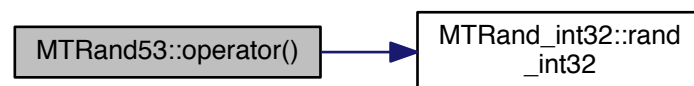
Definition at line 144 of file mtrand.h.

12.6.3 Member Function Documentation

12.6.3.1 double MTRand53::operator() () [inline]

Definition at line 145 of file mtrand.h.

Here is the call graph for this function:



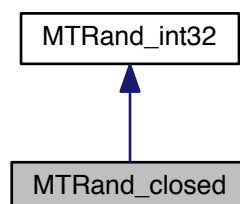
The documentation for this class was generated from the following file:

- </Users/alessandrofilisetti/Documents/GIT/carness/mtrand.h>

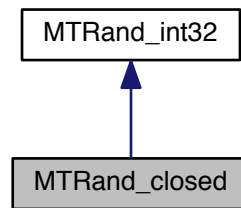
12.7 MTRand_closed Class Reference

```
#include <mtrand.h>
```

Inheritance diagram for MTRand_closed:



Collaboration diagram for MTRand_closed:



Public Member Functions

- [MTRand_closed](#) ()
- [MTRand_closed](#) (unsigned long *seed*)
- [MTRand_closed](#) (const unsigned long **seed*, int *size*)
- [~MTRand_closed](#) ()
- double [operator](#)() ()

Additional Inherited Members

12.7.1 Detailed Description

Definition at line 111 of file mtrand.h.

12.7.2 Constructor & Destructor Documentation

12.7.2.1 MTRand_closed::MTRand_closed () [inline]

Definition at line 113 of file mtrand.h.

12.7.2.2 MTRand_closed::MTRand_closed (unsigned long *seed*) [inline]

Definition at line 114 of file mtrand.h.

12.7.2.3 MTRand_closed::MTRand_closed (const unsigned long * *seed*, int *size*) [inline]

Definition at line 115 of file mtrand.h.

12.7.2.4 MTRand_closed::~~MTRand_closed () [inline]

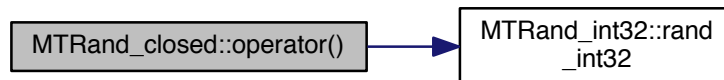
Definition at line 116 of file mtrand.h.

12.7.3 Member Function Documentation

12.7.3.1 `double MTRand_closed::operator() ()` `[inline]`

Definition at line 117 of file `mtrand.h`.

Here is the call graph for this function:



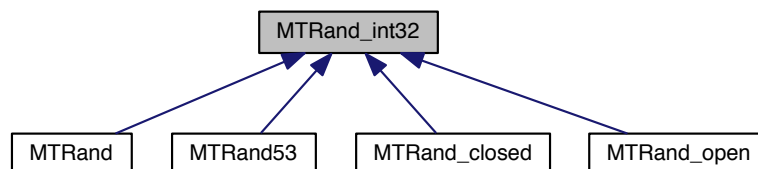
The documentation for this class was generated from the following file:

- </Users/alessandrofilisetti/Documents/GIT/carness/mtrand.h>

12.8 MTRand_int32 Class Reference

```
#include <mtrand.h>
```

Inheritance diagram for MTRand_int32:



Public Member Functions

- [MTRand_int32 \(\)](#)
- [MTRand_int32 \(unsigned long s\)](#)
- [MTRand_int32 \(const unsigned long *array, int size\)](#)
- void [seed](#) (unsigned long)
- void [seed](#) (const unsigned long *, int size)
- unsigned long [operator\(\) \(\)](#)
- virtual [~MTRand_int32 \(\)](#)

Protected Member Functions

- unsigned long [rand_int32 \(\)](#)

12.8.1 Detailed Description

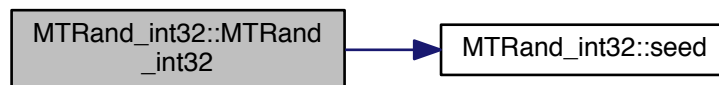
Definition at line 48 of file mtrand.h.

12.8.2 Constructor & Destructor Documentation

12.8.2.1 MTRand_int32::MTRand_int32 () [inline]

Definition at line 51 of file mtrand.h.

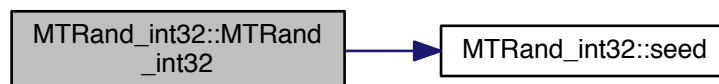
Here is the call graph for this function:



12.8.2.2 MTRand_int32::MTRand_int32 (unsigned long s) [inline]

Definition at line 53 of file mtrand.h.

Here is the call graph for this function:



12.8.2.3 MTRand_int32::MTRand_int32 (const unsigned long * array, int size) [inline]

Definition at line 55 of file mtrand.h.

Here is the call graph for this function:



12.8.2.4 `virtual MTRand_int32::~~MTRand_int32 () [inline],[virtual]`

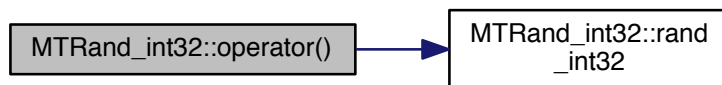
Definition at line 62 of file mtrand.h.

12.8.3 Member Function Documentation

12.8.3.1 `unsigned long MTRand_int32::operator() () [inline]`

Definition at line 60 of file mtrand.h.

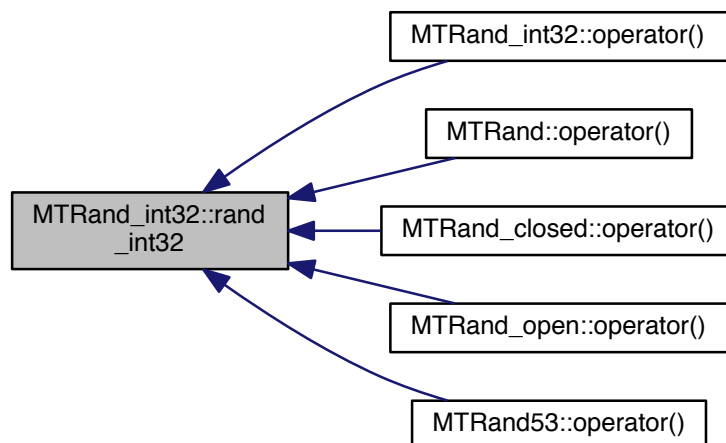
Here is the call graph for this function:



12.8.3.2 `unsigned long MTRand_int32::rand_int32 () [inline],[protected]`

Definition at line 85 of file mtrand.h.

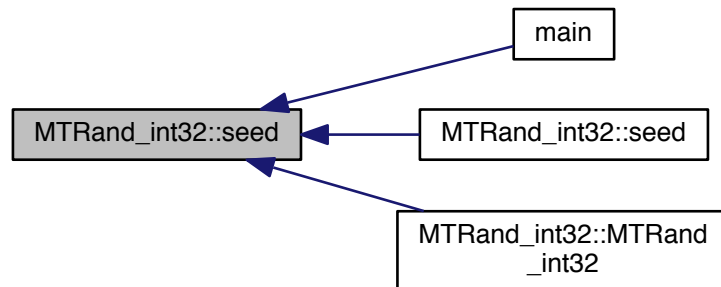
Here is the caller graph for this function:



12.8.3.3 `void MTRand_int32::seed (unsigned long s)`

Definition at line 23 of file mtrand.cpp.

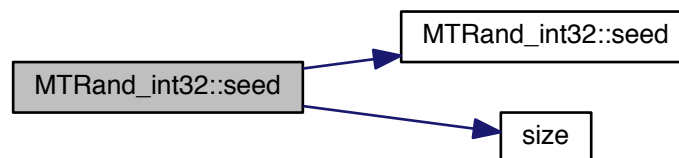
Here is the caller graph for this function:



12.8.3.4 void MTRand_int32::seed (const unsigned long * array, int size)

Definition at line 35 of file `mtrand.cpp`.

Here is the call graph for this function:



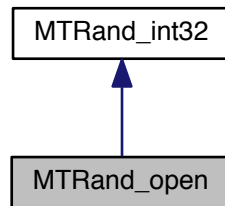
The documentation for this class was generated from the following files:

- [/Users/alessandrofilisetti/Documents/GIT/carness/mtrand.h](#)
- [/Users/alessandrofilisetti/Documents/GIT/carness/mtrand.cpp](#)

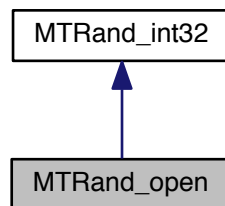
12.9 MTRand_open Class Reference

```
#include <mtrand.h>
```

Inheritance diagram for MTRand_open:



Collaboration diagram for MTRand_open:



Public Member Functions

- [MTRand_open](#) ()
- [MTRand_open](#) (unsigned long [seed](#))
- [MTRand_open](#) (const unsigned long *[seed](#), int [size](#))
- [~MTRand_open](#) ()
- double [operator\(\)](#) ()

Additional Inherited Members

12.9.1 Detailed Description

Definition at line 125 of file mtrand.h.

12.9.2 Constructor & Destructor Documentation

12.9.2.1 MTRand_open::MTRand_open () [inline]

Definition at line 127 of file mtrand.h.

12.9.2.2 MTRand_open::MTRand_open (unsigned long *seed*) [inline]

Definition at line 128 of file mtrand.h.

12.9.2.3 MTRand_open::MTRand_open (const unsigned long * *seed*, int *size*) [inline]

Definition at line 129 of file mtrand.h.

12.9.2.4 MTRand_open::~~MTRand_open () [inline]

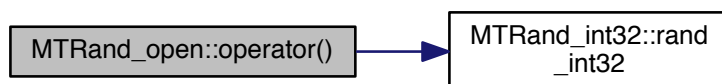
Definition at line 130 of file mtrand.h.

12.9.3 Member Function Documentation

12.9.3.1 double MTRand_open::operator() () [inline]

Definition at line 131 of file mtrand.h.

Here is the call graph for this function:



The documentation for this class was generated from the following file:

- </Users/alessandrofilisetti/Documents/GIT/carness/mtrand.h>

12.10 reactions Class Reference

```
#include <reactions.h>
```

Public Member Functions

- [reactions](#) ([acs_longInt](#) tmpID, [acs_int](#) tmpType, [acs_longInt](#) tmpM_I, [acs_longInt](#) tmpM_II, [acs_longInt](#) tmpM_III, [acs_int](#) tmpEvents, [acs_int](#) tmpEnergyType, [acs_double](#) tmpKspont)
Constructor.
- [~reactions](#) ()
- [acs_longInt](#) getID () const
- [acs_int](#) getType () const
- [acs_longInt](#) getSpecies_I () const
- [acs_longInt](#) getSpecies_II () const
- [acs_longInt](#) getSpecies_III () const
- [acs_int](#) getEvents () const
- [acs_int](#) getEnergyType () const

- [acs_double getKspont \(\)](#) const
- void [updateTotEvents \(\)](#)
- void [resetEventsCounter \(\)](#)

12.10.1 Detailed Description

Definition at line 17 of file reactions.h.

12.10.2 Constructor & Destructor Documentation

12.10.2.1 `reactions::reactions (acs_longInt tmpID, acs_int tmpType, acs_longInt tmpM_I, acs_longInt tmpM_II, acs_longInt tmpM_III, acs_int tmpEvents, acs_int tmpEnergyType, acs_double tmpKspont)`

Constructor.

Parameters

<i>tmpID</i>	reaction identifier
<i>tmpType</i>	condensation or cleavage
<i>tmpM_I</i>	product (if condensation) or substrates (if cleavage)
<i>tmpM_II</i>	product (if cleavage) or substrates (if condensation)
<i>tmpM_III</i>	product (if cleavage) or substrates (if condensation)
<i>tmpKeq</i>	equilibrium constant

Definition at line 20 of file reactions.cpp.

12.10.2.2 `reactions::~~reactions ()` `[inline]`

Definition at line 33 of file reactions.h.

12.10.3 Member Function Documentation

12.10.3.1 `acs_int reactions::getEnergyType ()` const `[inline]`

Definition at line 42 of file reactions.h.

12.10.3.2 `acs_int reactions::getEvents ()` const `[inline]`

Definition at line 41 of file reactions.h.

12.10.3.3 `acs_longInt reactions::getID ()` const `[inline]`

Definition at line 36 of file reactions.h.

12.10.3.4 `acs_double reactions::getKspont ()` const `[inline]`

Definition at line 43 of file reactions.h.

12.10.3.5 `acs_longInt reactions::getSpecies_I ()` const `[inline]`

Definition at line 38 of file reactions.h.

12.10.3.6 `acs_longInt reactions::getSpecies_II () const [inline]`

Definition at line 39 of file reactions.h.

12.10.3.7 `acs_longInt reactions::getSpecies_III () const [inline]`

Definition at line 40 of file reactions.h.

12.10.3.8 `acs_int reactions::getType () const [inline]`

Definition at line 37 of file reactions.h.

12.10.3.9 `void reactions::resetEventsCounter () [inline]`

Definition at line 47 of file reactions.h.

12.10.3.10 `void reactions::updateTotEvents () [inline]`

Definition at line 46 of file reactions.h.

The documentation for this class was generated from the following files:

- [/Users/alessandrofilisetti/Documents/GIT/carness/reactions.h](#)
- [/Users/alessandrofilisetti/Documents/GIT/carness/reactions.cpp](#)

12.11 species Class Reference

This class contains declarations of the species class.

```
#include <species.h>
```

Public Member Functions

- [species \(\)](#)
< New species constructor (IN AMOUNT)
- [species \(acs_longInt tmpID, string tmpSequence, acs_longInt tmpAmount, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpVolume, acs_double tmpK_phospho, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut\)](#)
New species constructor (IN CONCENTRATION)
- [species \(acs_longInt tmpID, string tmpSequence, acs_double tmpConcentration, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpVolume, acs_double tmpK_phospho, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut\)](#)
New species constructor in case of species structure file upload (IN AMOUNT)
- [species \(acs_longInt tmpID, string tmpSequence, acs_longInt tmpAmount, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpAge, acs_int tmpReborns, acs_double tmpVolume, acs_longInt tmpNotUsedCatID, acs_longInt tmpNotUsedSubID, acs_double tmpK_phospho, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut\)](#)
New species constructor in case of species structure file upload (IN CONCENTRATION)

- **species** ([acs_longInt](#) tmpID, string tmpSequence, [acs_double](#) tmpConcentration, [acs_double](#) tmpDiffusionEnh, [acs_int](#) tmpSoluble, [acs_double](#) tmpComplexDegEnh, [acs_int](#) tmpComplexCuttingPoint, [acs_int](#) tmpEvaluated, [acs_double](#) tmpAge, [acs_int](#) tmpReborns, [acs_double](#) tmpVolume, [acs_longInt](#) tmpNotUsedCatID, [acs_longInt](#) tmpNotUsedSubID, [acs_double](#) tmpK_phospho, [acs_double](#) tmpKLoadConc, [acs_int](#) tmpEnergizable, [acs_double](#) tmpInflux_rate, [acs_int](#) tmpMaxLOut)

New random species constructor.

- **species** ([acs_longInt](#) tmpID, string tmpSequence, [acs_longInt](#) tmpAmount, [acs_double](#) tmpDiffusionEnh, [acs_int](#) tmpSoluble, [acs_double](#) tmpComplexProb, [acs_double](#) tmpMaxComplexDegKinetic, [MTRand](#) &tmp_RandomGenerator, [acs_double](#) tmpVolume, [acs_double](#) tmpK_phospho, [acs_int](#) tmpEnergizable)

new Complex species constructor

- **species** ([acs_longInt](#) tmpID, string tmpSequence, [acs_double](#) tmpDiffusionEnh, [acs_int](#) tmpSoluble, [acs_double](#) tmpMaxComplexDegKinetic, [acs_int](#) tmpCuttingPoint, [MTRand](#) &tmp_RandomGenerator, [acs_longInt](#) tmpCatalyst_ID, [acs_longInt](#) tmpSubstrate_ID, [acs_double](#) tmpVolume, [acs_double](#) tmpK_phospho, [acs_int](#) tmpEnergizable)

This constructor is used to create a molecular complex.

- **~species** ()
- [acs_longInt](#) **getID** () const
- string **getSequence** () const
- [acs_int](#) **getSequenceLength** () const
- [acs_longInt](#) **getAmount** () const
- [acs_longInt](#) **getNOTchargeMols** () const
- [acs_longInt](#) **getChargeMols** () const
- [acs_double](#) **getConcentration** () const
- [acs_double](#) **getLoadedConcentration** ([acs_double](#) tmpVolume)
- [acs_double](#) **getAge** () const
- [acs_int](#) **getReborns** () const
- [acs_double](#) **getDiffusionEnh** () const
- [acs_int](#) **getSolubility** () const
- [acs_double](#) **getComplexDegEnh** () const
- [acs_int](#) **getComplexCutPnt** () const
- [acs_int](#) **getEvaluated** () const
- [acs_longInt](#) **getCatalyst_ID** () const
- [acs_longInt](#) **getSubstrate_ID** () const
- [acs_double](#) **getK_phospho** () const
- [acs_int](#) **getEnergizable** () const
- bool **getConcentrationFixed** () const
- [acs_double](#) **getFirstConcentration** () const
- [acs_int](#) **getLastSpeciesEvaluated** () const
- vector< [acs_longInt](#) > **getSecSubList** () const
- vector< [acs_double](#) > **getSec_k_SubList** () const
- vector< [acs_longInt](#) > **getCatalysisIfCpx** () const
- [acs_int](#) **getSecSubListSize** () const
- [acs_longInt](#) **getSecSubListID** ([acs_int](#) tmpID) const
- [acs_double](#) **getSec_k_SubListID** ([acs_int](#) tmpID) const
- [acs_longInt](#) **getCatalysisIfCpxID** ([acs_int](#) tmpID) const
- void **increment** ([acs_double](#) tmpVolume)
- void **specificIncrement** ([acs_int](#) tmpIncrement, [acs_double](#) tmpVolume)
- void **setAmount** ([acs_int](#) tmpAmount, [acs_double](#) tmpVolume)
- void **setConcentration** ([acs_double](#) tmpConc, [acs_double](#) tmpVolume)
- void **decrement** ([acs_double](#) tmpVolume)
- bool **setChargeMols** ([acs_int](#) tmpMolsToCharge)
- bool **setSpecificChargeMols** ([acs_int](#) tmpMolsToCharge)
- bool **chargeMol** ()
- bool **unchargeMol** ()
- void **setEvaluated** ()

- void [setDiffusion](#) ([acs_double](#) tmpDiff)
- void [setSolubility](#) ([acs_int](#) tmpSol)
- void [setKphospho](#) ([acs_double](#) tmpKphospho)
- void [setNewAge](#) ([acs_double](#) tmpLastTimeInterval)
- void [rebornsIncrement](#) ()
- void [concToNum](#) ([acs_double](#) tmpVolume)
- void [numToConc](#) ([acs_double](#) tmpVolume)
- void [resetAge](#) ()
- void [resetReborns](#) ()
- void [resetToInitConc](#) ([acs_double](#) tmpVolume)
- void [setLastSpeciesEvaluated](#) ([acs_int](#) tmpID)
- void [insertSecSub](#) ([acs_longInt](#) tmpID, [acs_double](#) tmpK, [acs_longInt](#) tmpCat)
- void [insertGillID](#) ([acs_longInt](#) tmpID)
- [acs_longInt](#) [getGillIDpos](#) ([acs_longInt](#) tmpID) const
- void [showGillEngagement](#) ()

12.11.1 Detailed Description

This class contains declarations of the species class.

class species

Authors

Alessandro Filisetti

Version

1.1 questa modifica è di prova per subversion

Created by Alessandro Filisetti on 19/02/09. Copyright 2009 European Centre for Living Technology. All rights reserved. Test paxelito SVN

Definition at line 18 of file species.h.

12.11.2 Constructor & Destructor Documentation

12.11.2.1 `species::species ()`

< New species constructor (IN AMOUNT)

This class containing the declaration of the species.

class species

Authors

Alessandro Filisetti

Version

0.1

Created by Alessandro Filisetti on 19/02/09. Copyright 2009 European Centre for Living Technology. All rights reserved. Default constructor

Definition at line 16 of file species.cpp.

12.11.2.2 `species::species (acs_longInt tmpID, string tmpSequence, acs_longInt tmpAmount, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpVolume, acs_double tmpK_phospho, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut)`

New species constructor (IN CONCENTRATION)

This constructor is used each time a new species is created (AMOUNT BASED)

Parameters

<i>tmpID</i>	species identifier
<i>tmpSequence</i>	species sequence (e.g. ABABAABABA)
<i>tmpAmount</i>	species initial amount
<i>tmpDiffusionEnh</i>	Diffusion enhancement degree
<i>tmpSoluble</i>	1 if the species is soluble, 0 otherwise
<i>tmpComplex-DegEnh</i>	complex dissociation kinetic constant
<i>tmpComplex-CuttingPoint</i>	complex cutting point (catalyst-substrate)
<i>tmpEvaluated</i>	This parameter indicates whether the species has been already evaluated (i.e. all the catalysis of the species are instantiated)
<i>tmpVolume</i>	the volume is necessary to convert numbers in concentrations
<i>tmpK_phospho</i>	phosphorilation kinetic constant (in case of energy based simulations)
<i>tmpEnergizable</i>	this is a flag indicating whether or not the species is energizable

Definition at line 53 of file species.cpp.

Here is the call graph for this function:



12.11.2.3 `species::species (acs_longInt tmpID, string tmpSequence, acs_double tmpConcentration, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpVolume, acs_double tmpK_phospho, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut)`

New species constructor in case of species structure file upload (IN AMOUNT)

This constructor is used each time a new species is created (CONCENTRATION BASED)

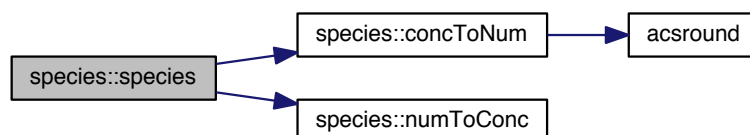
Parameters

<i>tmpID</i>	species identifier
<i>tmpSequence</i>	species sequence (e.g. ABABAABABA)
<i>tmp-Concentration</i>	species initial concentration

<i>tmpDiffusionEnh</i>	Diffusion enhancement degree
<i>tmpSoluble</i>	1 if the species is soluble, 0 otherwise
<i>tmpComplex-DegEnh</i>	complex dissociation kinetic constant
<i>tmpComplex-CuttingPoint</i>	complex cutting point (catalyst-substrate)
<i>tmpEvaluated</i>	This parameter indicates whether the species has been already evalutad (i.e. all the catalysis of the species are instantiated)
<i>tmpVolume</i>	the volume is necessary to convert concentrations in numbers
<i>tmpK_phospho</i>	phosphorilation kinetic constant (in case of energy based simulations)
<i>tmpEnergizable</i>	this is a flag indicating whether or not the species is energizable

Definition at line 99 of file species.cpp.

Here is the call graph for this function:



12.11.2.4 `species::species (acs_longInt tmpID, string tmpSequence, acs_longInt tmpAmount, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpAge, acs_int tmpReborns, acs_double tmpVolume, acs_longInt tmpNotUsedCatID, acs_longInt tmpNotUsedSubID, acs_double tmpK_phospho, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut)`

New species constructor in case of species structure file upload (IN CONCENTRATION)

This constructor is used when a new species is uploaded from file (TOTAL AMOUNT BASED)

Parameters

<i>tmpID</i>	species identificator
<i>tmpSequence</i>	species sequence (e.g. ABABAABABA)
<i>tmpAmount</i>	species initial amount of molecules
<i>tmpReactions_constant</i>	????

Definition at line 138 of file species.cpp.

Here is the call graph for this function:



12.11.2.5 `species::species (acs_longInt tmpID, string tmpSequence, acs_double tmpConcentration, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexDegEnh, acs_int tmpComplexCuttingPoint, acs_int tmpEvaluated, acs_double tmpAge, acs_int tmpReborns, acs_double tmpVolume, acs_longInt tmpNotUsedCatID, acs_longInt tmpNotUsedSubID, acs_double tmpK_phospho, acs_double tmpKLoadConc, acs_int tmpEnergizable, acs_double tmpInflux_rate, acs_int tmpMaxLOut)`

New random species constructor.

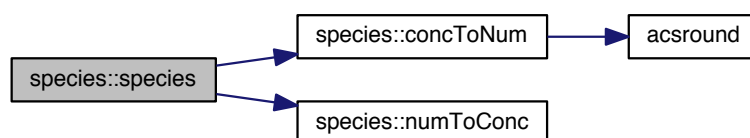
This constructor is used when a new species is uploaded from file (CONCENTRATION BASED)

Parameters

<i>tmpID</i>	species identifier
<i>tmpSequence</i>	species sequence (e.g. ABABAABABA)
<i>tmpAmount</i>	species initial amount of molecules
<i>tmpReactions_ - constant</i>	????

Definition at line 180 of file species.cpp.

Here is the call graph for this function:



12.11.2.6 `species::species (acs_longInt tmpID, string tmpSequence, acs_longInt tmpAmount, acs_double tmpDiffusionEnh, acs_int tmpSoluble, acs_double tmpComplexProb, acs_double tmpMaxComplexDegKinetic, MTRand & tmp_RandomGenerator, acs_double tmpVolume, acs_double tmpK_phospho, acs_int tmpEnergizable)`

new Complex species constructor

This constructor is used when a species is randomly created (!!! NOT USED NOW)

Version

0.1 (8 parameters)

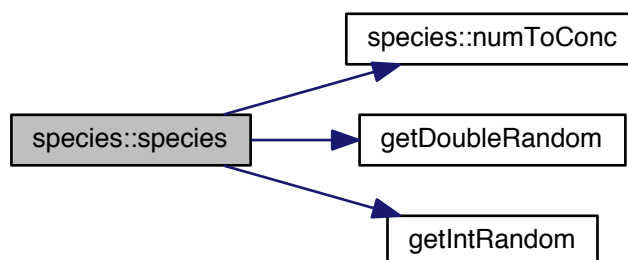
Parameters

<i>tmpID</i>	species identifier
<i>tmpSequence</i>	species sequence (e.g. ABABAABABA)
<i>tmpAmount</i>	species initial amount of molecules
<i>acs_double</i>	tmpDiffusionEnh Diffusion enhancement parameter
<i>acs_double</i>	tmpPrecipitationEnh Precipitation Enhancement parameters
<i>acs_double</i>	tmpComplexProb Probability to be a complex

<i>acs_double</i>	tmpMaxComplexDegKinetic max complex degradation constant
<i>MTRand</i> &	tmp_RandomGenerator random generator

Definition at line 228 of file species.cpp.

Here is the call graph for this function:



12.11.2.7 **species::species** (*acs_longInt tmpID*, *string tmpSequence*, *acs_double tmpDiffusionEnh*, *acs_int tmpSoluble*, *acs_double tmpMaxComplexDegKinetic*, *acs_int tmpCuttingPoint*, *MTRand* & *tmp_RandomGenerator*, *acs_longInt tmpCatalyst_ID*, *acs_longInt tmpSubstrate_ID*, *acs_double tmpVolume*, *acs_double tmpK_phospho*, *acs_int tmpEnergizable*)

This constructor is used to create a molecular complex.

Version

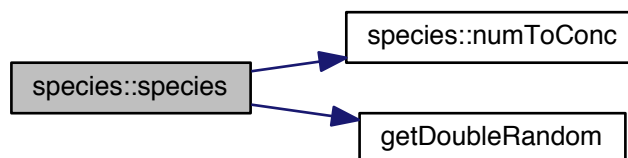
0.1 (10 paramters)

Parameters

<i>tmpID</i>	species identifier
<i>tmpSequence</i>	species sequence (e.g. ABABAABABA)
<i>acs_double</i>	tmpDiffusionEnh Diffusione enhancement parameter
<i>acs_double</i>	tmpPrecipitationEnh Precipitation Enhancement parameters
<i>acs_double</i>	tmpComplexProb Probability to be a complex
<i>acs_double</i>	tmpMaxComplexDegKinetic max complex degradation constant
<i>MTRand</i> &	tmp_RandomGenerator random generator
<i>acs_int</i>	tmpCatalyst_ID Catalyst ID
<i>acs_int</i>	tmpSubstrate_ID substrate ID

Definition at line 274 of file species.cpp.

Here is the call graph for this function:



12.11.2.8 `species::~~species ()` [inline]

Definition at line 83 of file `species.h`.

12.11.3 Member Function Documentation

12.11.3.1 `bool species::chargeMol ()`

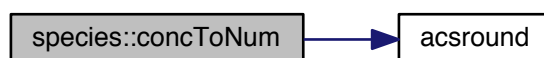
to charge molecules

Definition at line 350 of file `species.cpp`.

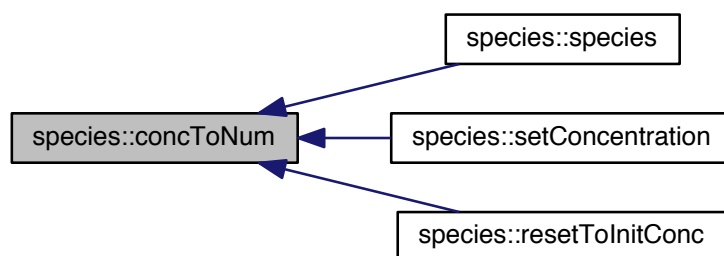
12.11.3.2 `void species::concToNum (acs_double tmpVolume)` [inline]

Definition at line 136 of file `species.h`.

Here is the call graph for this function:



Here is the caller graph for this function:



12.11.3.3 void species::decrement (acs_double tmpVolume)

Function to decrement the total number of molecules belonging to this species

Definition at line 313 of file species.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



12.11.3.4 acs_double species::getAge () const [inline]

Definition at line 94 of file species.h.

12.11.3.5 acs_longInt species::getAmount () const [inline]

Definition at line 89 of file species.h.

Here is the caller graph for this function:



12.11.3.6 `vector<acs_longInt> species::getCatalysisIfCpx () const [inline]`

Definition at line 110 of file species.h.

12.11.3.7 `acs_longInt species::getCatalysisIfCpxID (acs_int tmpID) const [inline]`

Definition at line 114 of file species.h.

12.11.3.8 `acs_longInt species::getCatalyst_ID () const [inline]`

Definition at line 101 of file species.h.

12.11.3.9 `acs_longInt species::getChargeMols () const [inline]`

Definition at line 91 of file species.h.

Here is the caller graph for this function:



12.11.3.10 `acs_int species::getComplexCutPnt () const [inline]`

Definition at line 99 of file species.h.

12.11.3.11 `acs_double species::getComplexDegEnh () const [inline]`

Definition at line 98 of file species.h.

12.11.3.12 `acs_double species::getConcentration () const [inline]`

Definition at line 92 of file species.h.

12.11.3.13 `bool species::getConcentrationFixed () const [inline]`

Definition at line 105 of file species.h.

12.11.3.14 **acs_double** species::getDiffusionEnh () const [inline]

Definition at line 96 of file species.h.

12.11.3.15 **acs_int** species::getEnergizable () const [inline]

Definition at line 104 of file species.h.

12.11.3.16 **acs_int** species::getEvaluated () const [inline]

Definition at line 100 of file species.h.

12.11.3.17 **acs_double** species::getFirstConcentration () const [inline]

Definition at line 106 of file species.h.

12.11.3.18 **acs_longInt** species::getGillIDpos (**acs_longInt** *tmpID*) const [inline]

Definition at line 145 of file species.h.

12.11.3.19 **acs_longInt** species::getID () const [inline]

Definition at line 86 of file species.h.

12.11.3.20 **acs_double** species::getK_phospho () const [inline]

Definition at line 103 of file species.h.

12.11.3.21 **acs_int** species::getLastSpeciesEvaluated () const [inline]

Definition at line 107 of file species.h.

12.11.3.22 **acs_double** species::getLoadedConcentration (**acs_double** *tmpVolume*)

return the concentration of the loaded molecules

Definition at line 377 of file species.cpp.

12.11.3.23 **acs_longInt** species::getNOTchargeMols () const [inline]

Definition at line 90 of file species.h.

Here is the caller graph for this function:



12.11.3.24 `acs_int species::getReborns () const` `[inline]`

Definition at line 95 of file species.h.

12.11.3.25 `vector<acs_double> species::getSec_k_SubList () const` `[inline]`

Definition at line 109 of file species.h.

12.11.3.26 `acs_double species::getSec_k_SubListID (acs_int tmpID) const` `[inline]`

Definition at line 113 of file species.h.

12.11.3.27 `vector<acs_longInt> species::getSecSubList () const` `[inline]`

Definition at line 108 of file species.h.

12.11.3.28 `acs_longInt species::getSecSubListID (acs_int tmpID) const` `[inline]`

Definition at line 112 of file species.h.

12.11.3.29 `acs_int species::getSecSubListSize () const` `[inline]`

Definition at line 111 of file species.h.

12.11.3.30 `string species::getSequence () const` `[inline]`

Definition at line 87 of file species.h.

Here is the caller graph for this function:



12.11.3.31 `acs_int species::getSequenceLength () const` `[inline]`

Definition at line 88 of file species.h.

12.11.3.32 `acs_int species::getSolubility () const` `[inline]`

Definition at line 97 of file species.h.

12.11.3.33 `acs_longInt species::getSubstrate_ID () const` `[inline]`

Definition at line 102 of file species.h.

12.11.3.34 void species::increment (*acs_double tmpVolume*)

Function to increment the total number of molecules belonging to this species

Definition at line 302 of file species.cpp.

Here is the call graph for this function:



12.11.3.35 void species::insertGillID (*acs_longInt tmpID*) [inline]

Definition at line 144 of file species.h.

12.11.3.36 void species::insertSecSub (*acs_longInt tmpID, acs_double tmpK, acs_longInt tmpCat*)

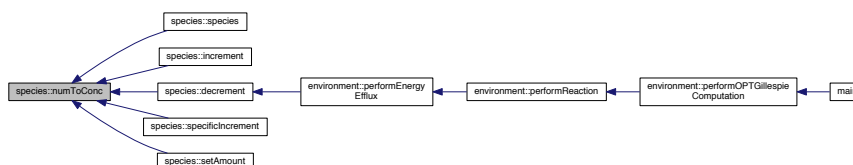
to insert the second substrate - and k reaction - to the species (complexes list)

Definition at line 386 of file species.cpp.

12.11.3.37 void species::numToConc (*acs_double tmpVolume*) [inline]

Definition at line 137 of file species.h.

Here is the caller graph for this function:



12.11.3.38 void species::rebornsIncrement () [inline]

Definition at line 134 of file species.h.

12.11.3.39 void species::resetAge () [inline]

Definition at line 139 of file species.h.

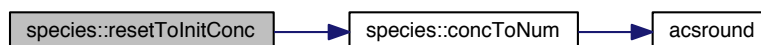
12.11.3.40 void species::resetReborns () [inline]

Definition at line 140 of file species.h.

12.11.3.41 `void species::resetToInitConc (acs_double tmpVolume) [inline]`

Definition at line 141 of file species.h.

Here is the call graph for this function:



12.11.3.42 `void species::setAmount (acs_int tmpAmount, acs_double tmpVolume) [inline]`

Definition at line 120 of file species.h.

Here is the call graph for this function:



12.11.3.43 `bool species::setChargeMols (acs_int tmpMolsToCharge)`

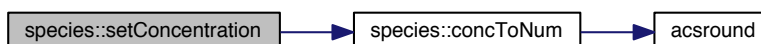
to charge a specific number of molecules

Definition at line 337 of file species.cpp.

12.11.3.44 `void species::setConcentration (acs_double tmpConc, acs_double tmpVolume) [inline]`

Definition at line 121 of file species.h.

Here is the call graph for this function:



12.11.3.45 `void species::setDiffusion (acs_double tmpDiff) [inline]`

Definition at line 130 of file species.h.

12.11.3.46 `void species::setEvaluated () [inline]`

Definition at line 129 of file species.h.

12.11.3.47 `void species::setKphospho (acs_double tmpKphospho) [inline]`

Definition at line 132 of file species.h.

12.11.3.48 `void species::setLastSpeciesEvaluated (acs_int tmpID) [inline]`

Definition at line 142 of file species.h.

12.11.3.49 `void species::setNewAge (acs_double tmpLastTimeInterval) [inline]`

Definition at line 133 of file species.h.

12.11.3.50 `void species::setSolubility (acs_int tmpSol) [inline]`

Definition at line 131 of file species.h.

12.11.3.51 `bool species::setSpecificChargeMols (acs_int tmpMolsToCharge)`

to charge a specific number of molecules

Definition at line 324 of file species.cpp.

12.11.3.52 `void species::showGillEngagement ()`

Function to show the species gillespie engagement

Definition at line 406 of file species.cpp.

12.11.3.53 `void species::specificIncrement (acs_int tmpIncrement, acs_double tmpVolume) [inline]`

Definition at line 119 of file species.h.

Here is the call graph for this function:



12.11.3.54 `bool species::unchargeMol ()`

to uncharge molecules

Definition at line 365 of file species.cpp.

Here is the caller graph for this function:



The documentation for this class was generated from the following files:

- [/Users/alessandrofilisetti/Documents/GIT/carness/species.h](#)
- [/Users/alessandrofilisetti/Documents/GIT/carness/species.cpp](#)

Chapter 13

File Documentation

13.1 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/allTimesAnalysis.m File Reference

Functions

- params do not prompt `figures` (0)
- OUT `disp ('|-----|')`
- `disp ('|-All times analysis...|')`

Variables

- `function [out]`
- params `figureVisible` = prompt `figures` (1)
- params `deltaT` = Delta T
- params `totT` = total time of the `simulation`
- `currentDir` = `cd()`

13.1.1 Function Documentation

13.1.1.1 `disp ('|-----|')`

13.1.1.2 `disp ('|-All times analysis...|')`

13.1.1.3 params do not prompt figures (0)

13.1.2 Variable Documentation

13.1.2.1 `currentDir = cd()`

Definition at line 17 of file allTimesAnalysis.m.

13.1.2.2 params `deltaT` = Delta T

Definition at line 9 of file allTimesAnalysis.m.

13.1.2.3 params figureVisible = prompt figures (1)

Definition at line 8 of file allTimesAnalysis.m.

13.1.2.4 function[out]

Initial value:

```
= allTimesAnalysis(params)
% function [out] = allTimesAnalysis(params)
%
%
% INPUT
%   params.tmpPath = simulations path
```

Definition at line 1 of file allTimesAnalysis.m.

13.1.2.5 params totT = total time of the simulation

Definition at line 10 of file allTimesAnalysis.m.

13.2 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/bufferedFluxAnalysis.py File Reference

Namespaces

- [bufferedFluxAnalysis](#)

Variables

- list [bufferedFluxAnalysis.StrPath](#) = sys.argv[1]
- list [bufferedFluxAnalysis.tmpResFold](#) = sys.argv[2]
- tuple [bufferedFluxAnalysis.today](#) = dt.date.today()
- string [bufferedFluxAnalysis.simF](#) = StrPath+''
- tuple [bufferedFluxAnalysis.tmpDirs](#) = sort(glob.glob(simF))
- tuple [bufferedFluxAnalysis.newdir](#) = os.path.join(os.getcwd(), '0_statistics')
- tuple [bufferedFluxAnalysis.matrixTimeLITE](#) = np.zeros((101,len(tmpDirs)))
- tuple [bufferedFluxAnalysis.matrixFluxLITE](#) = np.zeros((101,len(tmpDirs)))
- tuple [bufferedFluxAnalysis.matrixAbsLITE](#) = np.zeros((101,len(tmpDirs)))
- tuple [bufferedFluxAnalysis.matrixExpLITE](#) = np.zeros((101,len(tmpDirs)))
- int [bufferedFluxAnalysis.tmpDirsCnt](#) = 0
- tuple [bufferedFluxAnalysis.speciesFiles](#) = sort(glob.glob('species_*'))
- tuple [bufferedFluxAnalysis.fidSpecies](#) = open(speciesFiles[0], 'r')
- int [bufferedFluxAnalysis.ok](#) = 0
- list [bufferedFluxAnalysis.fluxIndexes](#) = []
- list [bufferedFluxAnalysis.fluxLengths](#) = []
- tuple [bufferedFluxAnalysis.index](#) = int(tmpID)
- tuple [bufferedFluxAnalysis.speciesSeq](#) = len(tmpSeq)
- tuple [bufferedFluxAnalysis.concFixed](#) = int(tmpConcFixed)
- tuple [bufferedFluxAnalysis.fileslist](#) = sort(glob.glob('reactions_parameters_*'))
- int [bufferedFluxAnalysis.rctParFileNum](#) = 1
- int [bufferedFluxAnalysis.rctID](#) = 1
- list [bufferedFluxAnalysis.totFluxDyn](#) = []

- list `bufferedFluxAnalysis.absorbedBricks` = []
- int `bufferedFluxAnalysis.tempAbsorbedBricks` = 0
- list `bufferedFluxAnalysis.expelledBricks` = []
- int `bufferedFluxAnalysis.tempExpelledBricks` = 0
- list `bufferedFluxAnalysis.totTimes` = []
- list `bufferedFluxAnalysis.totFluxDyn_LITE` = []
- list `bufferedFluxAnalysis.absorbedBricks_LITE` = []
- list `bufferedFluxAnalysis.expelledBricks_LITE` = []
- list `bufferedFluxAnalysis.totTimes_LITE` = []
- tuple `bufferedFluxAnalysis.fid` = open(file, 'r')
- int `bufferedFluxAnalysis.okmonitor` = 1
- int `bufferedFluxAnalysis.oksaveLite` = 0
- int `bufferedFluxAnalysis.deltaRct` = 0
- tuple `bufferedFluxAnalysis.reaction` = int(tmpReaction)
- tuple `bufferedFluxAnalysis.rtime` = float(tmpTime)
- tuple `bufferedFluxAnalysis.cc` = int(tmpcc)
- tuple `bufferedFluxAnalysis.cat` = int(tmpCat)
- tuple `bufferedFluxAnalysis.mol_I` = int(tmpMol_I)
- tuple `bufferedFluxAnalysis.mol_II` = int(tmpMol_II)
- tuple `bufferedFluxAnalysis.mol_III` = int(tmpMol_III)
- tuple `bufferedFluxAnalysis.loadedMolsConc` = float(tmpLoadedMolsConc)
- tuple `bufferedFluxAnalysis.loadedMols` = int(tmpLoadedMols)
- tuple `bufferedFluxAnalysis.gillMean` = float(tmpGillMean)
- tuple `bufferedFluxAnalysis.gillSD` = float(tmpGillSD)
- tuple `bufferedFluxAnalysis.gillEntropy` = float(tmpGillEntropy)
- tuple `bufferedFluxAnalysis.savingMatrix` = np.zeros((len(totTimes),4))
- tuple `bufferedFluxAnalysis.tmpDirSplit` = tmpDir.split("/")
- string `bufferedFluxAnalysis.cmdFileName` = StrPath+'0_statistics/0_fluxDynamics_'
- tuple `bufferedFluxAnalysis.cmdFileFid` = open(cmdFileName, 'a')

13.3 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/concAnalysis.m File Reference

Functions

- ID ho raggiunto la fine del file if isempty (itmp)
- fid,'%s', 1 fscanf ()
- Species sequence `concentrazione` (indice)
- Dissociation Kinetic Constant `bindpnt` (indice)
- Binding point `evaluated` (indice)
- Species `Age` (in seconds) `itmp = fscanf(fid,'%d',1)`
- fixed Concentration if (`evaluated(indice)==1`)&&(`bindpnt(indice)`)
- else speciesLENvec (indice)=0
- Species sequence `overallConcMatrix` (j, indice)
- Dissociation Kinetic Constant `tmpbindpnt` (indice)
- Binding point `tmpevaluated` (indice)
- fixed Concentration if (`tmpevaluated(indice)==1`)&&(`tmpbindpnt(indice)`)
- `coss` (j+1)
- `angles` (j+1)
- `overallConcMatrix` (end,:)
- `cd` (strcat(params.tmpPath,'0_statistics')) = size(`overallConcMatrix`)
- if exist (fileConcAllName,'file') delete(fileConcAllName)
- `fclose` (fidC)

- `cd (strcat('..', thisSimFolder))`
- INIT lengths `analysis file` (maxL, meanL, medianL) `lengthAnalysis`
- `lengthAnalysis (end,:)`
- Create vector containing all the `coseno` (nSpecies files%-1) from both the previous file and the first file `cos`
- `lengthAnalysis (j,:)`
- `fclose (fid)`
- `end disp ('Save concentrations...')`
- `:(and(evaluated >0, bindpnt==0 overallConcMatrix ()))`

Variables

- `function [out]`
- on each single species file `currentDir = cd()`
- if `nargin < 1` `params.path=currentDir();params.figureVisible=0;endcd(params.tmpPath);disp('-----
-----|');disp('|-Concentration Analysis...|');disp('|-----|');if ~isdir('0_statistics')
mkdir('0_statistics');end%READ ALL THE DIRECTORY CONTAINING SIMULATIONSsearch=strcat('*'
params.simFolder,'*');simDirs=dir(search);%For each folder the necessary computations are per-
formedfor i=1:length(simDirs)%Go into the results folderif isdir(strcat(simDirs(i).name)) cd(strcat(sim-
Dirs(i).name,'res'));thisSimFolder=strcat(simDirs(i).name,'res');%Read configuration fileconfParams=read-
Parameters();%File Containing all Timesteps=0:confParams.timeStructuresSavingInterval:confParams.-
nSeconds;speciesFiles=dir('species_ *');%speciesFiles=speciesFiles(1:end-1);to comment if species_-
2%does not exist%speciesFiles=speciesFiles(1:length(speciesFiles)-1);%For each species file, from the
last to the firstnFile=1:[nSpeciesFile, r]=size(speciesFiles);for j=nSpeciesFile:-1:1%FROM VILLANI MA-
RCOfid=fopen(speciesFiles(j).name,'r');%apro il primo filedisp(sprintf('Processing dir%s, file%s,%d/%d',
simDirs(i).name, speciesFiles(j).name, nSpeciesFile-j, nSpeciesFile));%leggo gli oggetti che ci sono in ogni
riga-alcuni li memorizzoindice=1;%definisco il parametro di controllo"continua"while indice >`
- `ID __pad0__`
- `end`
- if `nFile`
- Total concentration of the species `itmp = fscanf(fid,'%d',1)`
- Precipitation flag `ftmp = fscanf(fid,'%f',1)`
- Phosphorilation Kinetic `constant`
- Phosphorilation Kinetic `f`
- `end indice = indice+1`
- `else` per ora ho memorizzato solo nome specie e `concentrazione`
- if `nFile` Compute matrix indicators `tmpcos = (overallConcMatrix(j,:)*overallConcMatrix(j+1,:))/(norm(overall-
ConcMatrix(j,:))*norm(overallConcMatrix(j+1,:)))`
- `strZero = zeroBeforeStrNum(i,length(simDirs))`
- `fileConcAllName = strcat('0_concentrations_ALL_',strZero,int2str(i),'.csv')`
- `end fidC = fopen(fileConcAllName,'a')`
- for `a`
- `speciesLENvec = speciesLENvec(speciesLENvec > 0)`
- Create vector containing all the `angles = zeros(1,nSpeciesFile)`
- `cosFromInit = zeros(1,nSpeciesFile)`
- `anglesFromInit = zeros(1,nSpeciesFile)`
- `pause`
- Clear temp variable used to store species
- Clear temp variable used to store binding points and evaluation flags for each file
- clear `tmpspecie`
- clear `tmpbindpnt`
- clear `tmpevaluated`
- for `k`
- `overallConcMatrixEval = overallConcMatrix(:,evaluated>0)`

13.3.1 Function Documentation

13.3.1.1 Species Age (in *seconds*) =fscanf(fid,'%d',1)

13.3.1.2 angles (j+ 1)

13.3.1.3 Dissociation Kinetic Constant bindpnt (indice)

13.3.1.4 MOVING INTO THE STATISTIC FOLDER cd (strcat(params.tmpPath,'0_statistics')) = size(overallConcMatrix)

13.3.1.5 cd (strcat('..', thisSimFolder))

13.3.1.6 Species sequence concentrazione (indice)

13.3.1.7 Create vector containing all the coseno (nSpecies files%- 1)

13.3.1.8 coss (j+ 1)

13.3.1.9 end disp ('Save concentrations...')

13.3.1.10 Binding point evaluated (indice)

13.3.1.11 if exist (fileConcAllName , 'file')

13.3.1.12 fclose (fidC)

13.3.1.13 fclose (fid)

13.3.1.14 INIT lengths analysis file (maxL , meanL , medianL)

13.3.1.15 fid s fscanf () [virtual]

13.3.1.16 fixed Concentration if (evaluated(indice) ==1)

13.3.1.17 fixed Concentration if (tmpevaluated(indice) ==1)

13.3.1.18 ID ho raggiunto la fine del file if isempty (itmp)

13.3.1.19 lenghtAnalysis (end , :)

13.3.1.20 lenghtAnalysis (j , :)

13.3.1.21 Species sequence overallConcMatrix (j , indice)

13.3.1.22 overallConcMatrix (end , :)

13.3.1.23 :,(and(evaluated>0,bindpnt==0 overallConcMatrix () [virtual]

13.3.1.24 else speciesLENvec (indice) [pure virtual]

13.3.1.25 Dissociation Kinetic Constant tmpbindpnt (indice)

13.3.1.26 Binding point tmpevaluated (indice)

13.3.2 Variable Documentation

13.3.2.1 ID __pad0__

Definition at line 69 of file concAnalysis.m.

13.3.2.2 for a

Initial value:

```
=1:indice-1  
                fprintf(fidC,'%s \t',specie(a).nome)
```

Definition at line 149 of file concAnalysis.m.

13.3.2.3 angles = zeros(1,nSpeciesFile)

Definition at line 159 of file concAnalysis.m.

13.3.2.4 anglesFromInit = zeros(1,nSpeciesFile)

Definition at line 164 of file concAnalysis.m.

13.3.2.5 else per ora ho memorizzato solo nome specie e concentrazione

Definition at line 101 of file concAnalysis.m.

13.3.2.6 Phosphorilation Kinetic constant

Definition at line 90 of file concAnalysis.m.

13.3.2.7 cossFromInit = zeros(1,nSpeciesFile)

Definition at line 163 of file concAnalysis.m.

13.3.2.8 on each single species file currentDir = cd()

Definition at line 10 of file concAnalysis.m.

13.3.2.9 end

Definition at line 73 of file concAnalysis.m.

13.3.2.10 Phosphorilation Kinetic f

Definition at line 90 of file concAnalysis.m.

13.3.2.11 fidC = fopen(fileConcAllName,'a')

Definition at line 148 of file concAnalysis.m.

13.3.2.12 `fileConcAllName = strcat('0_concentrations_ALL_',strZero,int2str(i),'.csv')`

Definition at line 144 of file concAnalysis.m.

13.3.2.13 `Phosphorilation Kinetic Charged Molecules Concentration ftmp =fscanf(fid,'%f',1)`

Definition at line 82 of file concAnalysis.m.

13.3.2.14 `function[out]`

Initial value:

```
= concAnalysis(params)
% function [out] = concAnalysis(params)
%
% INPUT
% params.tmpPath = tmpPath of the simulations root
%
% This function performs statistics on each single simulation
```

Definition at line 1 of file concAnalysis.m.

13.3.2.15 `end indice =indice+1`

Definition at line 99 of file concAnalysis.m.

13.3.2.16 `Catalyst ID itmp =fscanf(fid,'%d',1)`

Definition at line 80 of file concAnalysis.m.

13.3.2.17 `end end case favorire quelle corte con una scale free di esponente gamma k`

Initial value:

```
= 1 : ss
    fprintf(fidC,' \n')
```

Definition at line 186 of file concAnalysis.m.

13.3.2.18 `if nargin< 1params.path=currentDir();params.figureVisible=0;endcd(params.tmpPath);disp('-----
-----');disp('Concentration Analysis...');disp('-----');if ~isdir('0_ -
statistics')mkdir('0_statistics');end%READ ALL THE DIRECTORY CONTAINING SIMULATIONSearch=strcat('*',
params.simFolder,'*');simDirs=dir(search);%For each folder the necessary computations are performedfor
i=1:length(simDirs)%Go into the results folderif isdir(strcat(simDirs(i).name))cd(strcat(simDirs(i).name,'res'));this-
SimFolder=strcat(simDirs(i).name,'res');%Read configuration fileconfParams=readParameters();%File Containing
all Times=0:confParams.timeStructuresSavingInterval:confParams.nSeconds;speciesFiles=dir('species_
*');%speciesFiles=speciesFiles(1:end-1);to comment if species_2%does not exist%speciesFiles=species-
Files(1:length(speciesFiles)-1);%For each species file, from the last to the firstnFile=1:[nSpeciesFile,
r]=size(speciesFiles);for j=nSpeciesFile:-1:1%FROM VILLANI MARCOfid=fopen(speciesFiles(j).name,'r');%apro il
primo filedisp(sprintf('Processing dir%s, file%s,%d/%d', simDirs(i).name, speciesFiles(j).name, nSpeciesFile-j,
nSpeciesFile));%leggo gli oggetti che ci sono in ogni riga-alcuni li memorizzoindice=1;%definisco il parametro di
controllo"continua"while indice >`

Initial value:

```
=1 % For each species
    itmp=fscanf(fid,'%d',1)
```

Definition at line 65 of file concAnalysis.m.

13.3.2.19 end if nFile

Initial value:

```
== 1
    % per ora ho memorizzato solo nome specie e concentrazione
```

Definition at line 75 of file concAnalysis.m.

13.3.2.20 overallConcMatrixEval = overallConcMatrix(:,evaluated>0)

Definition at line 195 of file concAnalysis.m.

13.3.2.21 pause

Definition at line 169 of file concAnalysis.m.

13.3.2.22 Clear temp variable used to store species

Definition at line 172 of file concAnalysis.m.

13.3.2.23 clear speciesLENvec = speciesLENvec(speciesLENvec > 0)

Definition at line 157 of file concAnalysis.m.

13.3.2.24 strZero = zeroBeforeStrNum(i,length(simDirs))

Definition at line 143 of file concAnalysis.m.

13.3.2.25 clear tmpbindpnt

Definition at line 174 of file concAnalysis.m.

13.3.2.26 if nFile Compute matrix indicators tmpcos = (overallConcMatrix(j,:)*overallConcMatrix(j+1,:)))/(norm(overallConcMatrix(j,:))*norm(overallConcMatrix(j+1,:)))

Definition at line 132 of file concAnalysis.m.

13.3.2.27 clear tmpevaluated

Definition at line 174 of file concAnalysis.m.

13.3.2.28 Clear temp variable used to store binding points and evaluation flags for each file clear tmpspecie

Definition at line 172 of file concAnalysis.m.

13.4 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/fromWithin2Between.py File Reference

Namespaces

- `fromWithin2Between`

Functions

- `def fromWithin2Between.zeroBeforeStrNum`

Variables

- tuple `fromWithin2Between.zeroSIM` = `zeroBeforeStrNum(i,numSim)`
- tuple `fromWithin2Between.zeroGEN` = `zeroBeforeStrNum(j,numGen)`
- string `fromWithin2Between.folderNew` = "s_"
- tuple `fromWithin2Between.resdir` = `os.path.join(os.curdir, "res")`

13.5 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/garbageSearch.m File Reference

Functions

- `function garbageSearch (param) currentDir`
- `cd (param.tmpPath)`
- `disp ('|-----|')`
- `disp ('|-Garbage analysis is started|')`
- READ ALL THE DIRECTORY CONTAINING SIMULATIONS `if ~isdir ('0_statistics') mkdir('0_statistics')`
- `cd ('0_statistics')`
- `fclose (fid)`
- `cd ('../')`
- `disp (strcat('|-Folder: ', simDirs(x).name,'/res...'))`
- `if conc && sValues (5)`
- `speciesMatrix (continua, 2)`
- `break end reactionsMatrix (continua, 1:6)`
- `cleavagesMatrix (cleavages, 1:6)`
- `condensationsMatrix (condensations, 1:6)`
- `end catalysisMatrix (continua, 1:4)`
- `garbageMatrix (gRows, 1:2)`
- `fprintf (fid,'NET%d\n\n', x)`
- `fprintf (fid,'%f\n\n', garbageMatrix(i, 2))`
- `end cd (currentDir)`

Variables

- `end tmpFolder` = `strcat('*',param.simFolder,'*')`
- `simDirs` = `dir(tmpFolder)`
- `fid` = `fopen('ALL_garbage.txt','w')`
- for `x`
- creating `species` matrix `sFiles` = `dir('species*')`

- `speciesFile = sFiles(length(sFiles)).name`
- `continua = 1`
- `speciesMatrix = 0`
- `while continua index = fscanf(fid,'%d',1)`
- `stop = isempty(index)`
- `break end name = fscanf(fid,'%s',1)`
- `conc = fscanf(fid,'%f',1)`
- `sValues = fscanf(fid,'%f',12)`
- creating reactions matrix `rFiles = dir('reactions*')`
- `reactionsFile = rFiles(length(rFiles)-1).name`
- `reactionsMatrix = 0`
- `while continua rValues = fscanf(fid,'%d',7)`
- dividing into condensations
and cleavages `condensations = 0`
- `cleavages = 0`
- `cleavagesMatrix = 0`
- `condensationsMatrix = 0`
- for `i`
- creating catalysis matrix `cFiles = dir('catalysis*')`
- `catalysisFile = cFiles(length(cFiles)).name`
- `catalysisMatrix = 0`
- `while continua cValues = fscanf(fid,'%f',7)`
- `break`
- identifying garbage `[rsm csm] = size(speciesMatrix)`
- `gRows = 0`
- `garbageMatrix = 0`
- `check2 = ismember(speciesMatrix(i,1),condensationsMatrix(:,4:5))`
- `check3 = ismember(speciesMatrix(i,1),catalysisMatrix(:,2))`
- `if check1`
- `else [r c] = size(garbageMatrix)`
- `clear r`
- `clear c`

13.5.1 Function Documentation

13.5.1.1 `end catalysisMatrix (continua , 1:4)`

13.5.1.2 `end cd (param. tmpPath)`

13.5.1.3 `cd ('0_statistics')`

13.5.1.4 `cd ('..')`

13.5.1.5 `end cd (currentDir)`

13.5.1.6 `cleavagesMatrix (cleavages , 1:6)`

13.5.1.7 `condensationsMatrix (condensations , 1:6)`

13.5.1.8 `disp ('|-----|')`

13.5.1.9 `disp ('|-Garbage analysis is started|')`

13.5.1.10 `disp (strcat('|-Folder: ', simDirs(x).name,'/res...'))`

13.5.1.11 `fclose (fid)`

13.5.1.12 `fprintf (fid , 'NET%d\n\n' , x)`

13.5.1.13 `fprintf (fid , '%f\n\n' , garbageMatrix(i,2))`

13.5.1.14 `garbageMatrix (gRows , 1:2)`

13.5.1.15 `function garbageSearch (param)`

13.5.1.16 `break end reactionsMatrix (continua , 1:6)`

13.5.1.17 `speciesMatrix (continua , 2)`

13.5.1.18 `if conc&& sValues (5)`

13.5.1.19 `READ ALL THE DIRECTORY CONTAINING SIMULATIONS if ~isdir ('0_statistics')`

13.5.2 Variable Documentation

13.5.2.1 `break`

Definition at line 98 of file garbageSearch.m.

13.5.2.2 `clear c`

Initial value:

```
= 1 : length(numScc) % for each ACS (if present)
                        if numScc(c) > 1 % IF the ACS contains more than one
species                                     inSCCFlag = 0
```

Definition at line 138 of file garbageSearch.m.

13.5.2.3 `catalysisFile = cFiles(length(cFiles)).name`

Definition at line 89 of file garbageSearch.m.

13.5.2.4 `catalysisMatrix = 0`

Definition at line 92 of file garbageSearch.m.

13.5.2.5 `creating catalysis matrix cFiles = dir('catalysis*')`

Definition at line 88 of file garbageSearch.m.

13.5.2.6 `if check1`

Initial value:

```
== 0 && check2 == 0 && check3 == 0
gRows = gRows+1
```

Definition at line 113 of file garbageSearch.m.

13.5.2.7 `check2 = ismember(speciesMatrix(i,1),condensationsMatrix(:,4:5))`

Definition at line 111 of file garbageSearch.m.

13.5.2.8 `check3 = ismember(speciesMatrix(i,1),catalysisMatrix(:,2))`

Definition at line 112 of file garbageSearch.m.

13.5.2.9 `cleavages = 0`

Definition at line 72 of file garbageSearch.m.

13.5.2.10 `cleavagesMatrix = 0`

Definition at line 74 of file garbageSearch.m.

13.5.2.11 `conc = fscanf(fid,'%f',1)`

Definition at line 42 of file garbageSearch.m.

13.5.2.12 `else condensations = 0`

Definition at line 71 of file garbageSearch.m.

13.5.2.13 `condensationsMatrix = 0`

Definition at line 75 of file garbageSearch.m.

13.5.2.14 `continua = 1`

Definition at line 32 of file garbageSearch.m.

13.5.2.15 `while continua cValues = fscanf(fid,'%f',7)`

Definition at line 94 of file garbageSearch.m.

13.5.2.16 `else[r c] = size(garbageMatrix)`

Definition at line 128 of file garbageSearch.m.

13.5.2.17 `fid = fopen('ALL_garbage.txt','w')`

Definition at line 20 of file garbageSearch.m.

13.5.2.18 `identifying garbage[rsm csm] = size(speciesMatrix)`

Definition at line 106 of file garbageSearch.m.

13.5.2.19 **if** garbageMatrix = 0

Definition at line 108 of file garbageSearch.m.

13.5.2.20 **gRows** = 0

Definition at line 107 of file garbageSearch.m.

13.5.2.21 **end** *****
T A R T *******for** i

Initial value:

```
= 1:rrm
    if reactionsMatrix(i,2) == 1
        cleavages = cleavages+1
```

Definition at line 76 of file garbageSearch.m.

13.5.2.22 **else** index = fscanf(fid,'%d',1)

Definition at line 35 of file garbageSearch.m.

13.5.2.23 **break end** name = fscanf(fid,'%s',1)

Definition at line 41 of file garbageSearch.m.

13.5.2.24 **clear** r

Definition at line 137 of file garbageSearch.m.

13.5.2.25 **reactionsFile** = rFiles(length(rFiles)-1).name

Definition at line 54 of file garbageSearch.m.

13.5.2.26 **reactionsMatrix** = 0

Definition at line 57 of file garbageSearch.m.

13.5.2.27 **creating reactions matrix** rFiles = dir('reactions*')

Definition at line 53 of file garbageSearch.m.

13.5.2.28 **while continua** rValues = fscanf(fid,'%d',7)

Definition at line 59 of file garbageSearch.m.

13.5.2.29 **creating species matrix** sFiles = dir('species*')

Definition at line 29 of file garbageSearch.m.

- `end` If there are `species` over threshold network `analysis` is performed `disp` ('|-Graph Creation')
- `if ~isdir` (strcat('.../', `folderCat`)) `mkdir`(strcat('.../'
- `end if ~isdir` (strcat('.../', `folderSub`)) `mkdir`(strcat('.../'
- `disp` (strcat('|-File ', `rcsFiles(rfileID).name`, 'processing...'))
- while `ischar` (`rlineb`) `if` `ischar`(`rlineb`)%Format lines `if` `isstrprop`(`rline`(`end`))
- while `rline` (`end`)
- `end gillTimeSeries` (`rlineID`,:)
- If the time is `righe` save `iGraph` structures to file `disp` (sprintf('|-%s|Reaction%d-time:%6.4f-Save structures', `simDirs(IDF).name`, `reaction`, `rtime`))
- `saveGraphToFile` (`folderCat`, `reaction`, `rtime`, `confparams.nReactions`, `graph`, `filextPre`)
- `saveGraphSUBToFile` (`folderSub`, `reaction`, `rtime`, `confparams.nReactions`, `graphSUB`, `filextPre`)
- `end` If the prompt time is `righe` a message on the screen indicating the `reaction` number and the time is shown `if and` ((`rtime` > `rctIDshowNoSave` *`analysisTimeIntervalNoSave`),(`printTemporal-Message`==1)) `msg`
- `disp` (`msg`)
- `;`, 4 `timeInterval` ()
- `graph` (`;`, 5)
- `graphSUB` (`;`, 5)
- `end end if` (`cc`==0)||(`cc`
- `end INFLUX ECONOMY ANALYSIS` `if` `sum` (`influx`==`mol_I`) >
- `params` `params` `Substrate` (If different from 1) `end else`%CAT-> PRO, Otherwise `if` the `reaction` is already present its parameters are updated `if` `sum`(`and`((`graph`(`;`, 1)==`cat`), `graph`(`;`, 2)==`mol_I`))==1 `position`=`and`((`graph`(`;`, 1)==`cat`), `graph`(`;`, 2)==`mol_I`)
- `graph` (`position`,:)
- `;`, 2 `mol_I` ()
- `graphSUB` (`position`,:)
- `;`, 2 `mol_II` ()
- `end fclose` (`fid`)
- `cd` ('.../0_statistics')
- e.g. sim 1 of 10 sims, `strZero` will be '0' in order to create `a%file` named XXX_01_XXX `strZero`=`zeroBeforeStrNum`(`IDF`, `length`(`simDirs` `names` ()
- grid on `set` (`gca`, 'fontsize', 15, 'fontname', 'times')
- `xlabel` ('Time', 'Interpreter', 'latex', 'fontsize', 15)
- `ylabel` ('Gillespie Mean', 'Interpreter', 'latex', 'fontsize', 15)
- `eval` ([`print-depsc` ', `fileName`])
- `saveas` (`figure1`, `fileName`)
- `ylabel` ('Gillespie SD', 'Interpreter', 'latex', 'fontsize', 15)
- `saveas` (`figure2`, `fileName`)
- `ylabel` ('Entropy', 'Interpreter', 'latex', 'fontsize', 15)
- `saveas` (`figure3`, `fileName`)
- `ylabel` ('New `species` Probability', 'Interpreter', 'latex', 'fontsize', 15)
- `saveas` (`figure4`, `fileName`)
- `ylabel` ('Flux Molecules Balance', 'Interpreter', 'latex', 'fontsize', 15)
- `saveas` (`figure5`, `fileName`)
- `ylabel` ('Ratio of backward `reactions`', 'Interpreter', 'latex', 'fontsize', 15)
- `saveas` (`figure6`, `fileName`)
- `ylabel` ('Flux Molecules', 'Interpreter', 'latex', 'fontsize', 15)

```

• legend ('Added','Removed')
• saveas (figure7, fileName)
• NET ANALYSIS disp ('|-*****NETWORK ANALYSIS *****')
• disp ('|-Strongly connected components analysis... ')
• disp (sprintf('<> SCC n.%d', c))
• end end disp (sprintf('|-Number of ACS:%d', realSccs))
• disp (sprintf('|-Number of ACS(length 1):%d', self))
• disp (sprintf('|-Species over threshold:%d', length(IDsOverThreshold)))
• if ~isempty (IDsOverThreshold) for idot
• if ~isempty (incomingNodes) wasteSpeciesFLAG
• if sum (find(scc==c)==IDsOverThreshold(idot))%If the species belong to the ACS inSCCFlag
• disp (fprintf('\t Within Acs%d->%d#%d-%d]%d%6.4f', incomingNodes(innode), IDsOverThreshold(idot),
weightToDistribute,...IDsOverThreshold(idot), IDsOverThreshold(idot), concVec(idot)))
• disp (fprintf('\t From Acs%d->%d#%d-%d]%d%6.4f', incomingNodes(innode), IDsOverThreshold(idot),
weightToDistribute,...IDsOverThreshold(idot), IDsOverThreshold(idot), concVec(idot)))
• end end end end if self if sum (selfID==idot) > 0 wasteSpeciesFLAG
• end end disp (fprintf('\t<> Number of Structural Autocatalytic set of molecules:%d', realSccs))
• disp (fprintf('\t<>-----'))
• disp (fprintf('\t<> Species over threshold produced by a CHAIN:%d', prod_chain))
• if realSccs disp (fprintf('\t<> Species over threshold produced INTO an ACS:%d', prod_inSCC))
• disp (fprintf('\t<> Species over threshold produced INTO an ACS(weigthed):%d', prod_inSCC_weight))
• disp (fprintf('\t<> Species over threshold produced BY an ACS:%d', prod_bySCC))
• disp (fprintf('\t<> Species over threshold produced BY an ACS(weigthed):%d', prod_bySCC_weight))
• disp (fprintf('\t<> Species over threshold produced by an overlap:%d', prod_overlap))
• disp (fprintf('\t<> Species over threshold produced by an overlap(weigthed):%d', prod_overlap_weight))
• disp (fprintf('\t<> Species over threshold produced by itself:%d', autocatalysis))
• disp (fprintf('\t<> Species over threshold produced by itself(weigthed):%d', self_loop_weight))
• disp (fprintf('\t<> Concentration in ACSs:%6.4f', conc_inSCC))
• disp (fprintf('\t<> Concentration in ACSs leaves:%6.4f', conc_bySCC))
• disp (fprintf('\t<> Concentration in chains:%6.4f', conc_chain))
• disp (fprintf('\t<> Concentration of autocatalyst:%6.4f', conc_selfCat))
• disp (fprintf('\t<> Number of endo condensations:%6.4f', endo_condensation_counter))
• disp (fprintf('\t<> Number of condensations:%6.4f', condensation_counter))
• disp (fprintf('\t<> Number of endo cleavages:%6.4f', endo_cleavage_counter))
• disp (fprintf('\t<> Number of cleavages:%6.4f', cleavage_counter))
• fprintf (fidFINAL,'\n%s\t%6.4f\t%6.4f\t%6.4f\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%6.-
4f\t%6.4f\t%6.4f\t%6.4f\t%d\t%d\t%d\t%d\t%d',...simDirs(IDF).name, rct, ecc, idOt, realSccs, prod_inS-
CC, prod_bySCC, prod_chain, prod_overlap, autocatalysis, prod_inSCC_weight,...prod_bySCC_weight,
prod_chain_weight, prod_overlap_weight, self_loop_weight, conc_inSCC, conc_bySCC, conc_chain, conc-
_selfCat, endo_condensation_counter,...condensation_counter, endo_cleavage_counter, cleavage_counter,
wasteSpecies)
• disp (fprintf('\n ANALYSIS of the SIMULATION%s IS FINISHED\n', simDirs(IDF).name))
• cd (currentDir)
• tmpL, tmpL_strZero ()
• cd ('./res')
• end function saveGraphToFile (tmpDir, tmpRct, tmpRTime, tmpRcts, tmpGraph, tmpFilextPre) currentDir
• cd (strcat('./../', tmpDir))
• if exist (outFname,'file') delete(outFname)
• end fclose (fid1)
• end function saveGraphSUBToFile (tmpDir, tmpRct, tmpRTime, tmpRcts, tmpGraph, tmpFilextPre) currentDir
• while ischar (tline) tline
• if isequal (tline(1:param-1),nSeconds') confparams.nSeconds
• elseif isequal (tline(1:param-1),nGEN') confparams.nGEN
• elseif isequal (tline(1:param-1),nSIM') confparams.nSIM
• elseif isequal (tline(1:param-1),nReactions') confparams.nReactions

```

- elseif `isequal (tline(1:param-1),'timeStructuresSavingInterval')` `confparams.timeStructuresSavingInterval`
- elseif `isequal (tline(1:param-1),'reactionProbability')` `confparams.reactionProbability`
- elseif `isequal (tline(1:param-1),'energy')` `confparams.energy`
- elseif `isequal (tline(1:param-1),'ECConcentration')` `confparams.ECConcentration` = `str2num(tline(param+1-length(tline)))`
- elseif `isequal (tline(1:param-1),'influx_rate')` `confparams.influx_rate`
- elseif `isequal (tline(1:param-1),'maxLOut')` `confparams.maxLOut`
- elseif `isequal (tline(1:param-1),'volume')` `confparams.volume`
- `end end fclose (fidConf)`

Variables

- `function out`
- `params threshold = 0`
- `params decayTime = 100`
- `params simFolder = 'K_cpx5_rete_n_'`
- `params tmpRctFileToLoad = ''`
- `params tmpRctSUBFileToLoad = ''`
- `params figureVisible = 0`
- `params interval = 10`
- `end` Set current date `and`
current directory `currentDate = date()`
- `currentDir = cd()`
- `end` CREATE HEADER ROW IN THE
FINAL OUTCOMES FILES `outFileName = strcat(currentDate, '__', int2str(params.threshold), '_convOver-`
`Threshold_', int2str(params.decayTime), '.csv')`
- `end fidFINAL = fopen(outFileName,'w')`
- Come back to the original
folder READ ALL THE DIRECTORY
CONTAINING SIMULATIONS `search = strcat('*',params.simFolder,'*')`
- `simDirs = dir(search)`
- File Containing all Times `times = 0:params.deltaT:params.totT`
- `IDsimFOLDER = 1`
- for `IDF`
- Read file conf `confparams = readParameters()`
- Select Times and reactions files `speciesFiles = dir('species_1*')`
- if `confparams energy < 2 nrg=1;else`
`nrg=0;end`%analysisTimeInterval
is 1/10 of the total decay
time `analysisTimeInterval=params.interval;analysisTimeIntervalNoSave=confparams.nSeconds./params.-`
`decayTime;%load`
incoming flux if
`confparams.influx_rate > influx = loadInflux()`
- `folderCat = strcat('__0_iGraph_CAT_', int2str(params.decayTime))`
- `folderSub = strcat('__0_iGraph_SUB_', int2str(params.decayTime))`
- `end` ID file species counter `fileSpeciesID = 1`
- Compute species over threshold `IDsOverThreshold = tmpID(and(tmpConc>params.threshold,tmpCpx-`
`Cut==0))`
- `concVec = tmpConc`
- reaction parameters files `rscFiles = dir('*reactions_parameters*')`
- `filextPre = strcat('_', zeroBeforeStrNum(IDsimFOLDER, length(simDirs)),int2str(IDsimFOLDER))`
- for `rfileID`
- `condensation_counter = 0`
- `endo_cleavage_counter = 0`
- `cleavage_counter = 0`

- `fid = fopen(rcsFiles(rfileID).name,'r')`
- apro il primo file initialize
useful variables `previousTime = 0`
- `rctIDshow = 1`
- `rctIDshowNoSave = 1`
- `rctID = 1`
- `rline = fgetl(fid)`
- `rlineb = rline`
- `rlineID = 1`
- FOR EACH REACTION `fluxEconomy = 0`
- `fluxPlus = 0`
- `fluxMinus = 0`
- `fluxEconomyArray = []`
- while `cntrl`
- Craete different variables `reaction = rline(1)`
- `rtime = rline(2)`
- `cc = rline(3)`
- `cat = rline(4)`
- `mol_I = rline(5)`
- `mol_II = rline(6)`
- `mol_III = rline(7)`
- `loadedMolsConc = rline(8)`
- `loadedMols = rline(9)`
- `gillMean = rline(10)`
- `gillSD = rline(11)`
- `gillEntropy = rline(12)`
- `newSpeciesProb = rline(13)`
- `ratioBackForward = rline(14)`
- end if `nrg`
- `printTemporalMessage = 1`
- end update time intervals `timeInterval = rtime - previousTime`
- `graph = graph(graph(:,5)>0,:)`
- `graphSUB = graphSUB(graphSUB(:,5)>0,:)`
- else `endo_condensation_counter = endo_condensation_counter + 1`
- `currentFolder = cd()`
- `fileName = strcat('25_gilleMean_',strZero,int2str(IDF),'.eps')`
- `figure1 = gcf`
- `figure2 = gcf`
- `figure3 = gcf`
- `figure4 = gcf`
- `figure5 = gcf`
- `figure6 = gcf`
- `figure7 = gcf`
- SAVE THE GILLESPIE MATRXI ON FILE `filename = strcat('0_gillespie_',strZero,int2str(IDF),'.txt')`
- `fluxEconomyMatrix = [gillTimeSeries(:,1), fluxEconomyArray]`
- Computing actual number od
strongly connected components `from = graph(:,1)+1`
- `to = graph(:,2)+1`
- `prod_inSCC = 0`
- `prod_chain = 0`
- `prod_bySCC = 0`
- `prod_overlap = 0`
- `sccID = 0`
- `autocatalysis = 0`
- `prod_inSCC_weight = 0`

```

• prod_chain_weight = 0
• prod_bySCC_weight = 0
• prod_overlap_weight = 0
• self_loop_weight = 0
• conc_inSCC = 0
• conc_chain = 0
• conc_bySCC = 0
• conc_selfCat = 0
• wasteSpecies = 0
• catSparse = sparse(from,to,true,max(max(from,to)),max(max(from,to)))
• numScc = histc(scc,1:max(scc))
• realSccs = 0
• for c
• alreadyAdded_leaves = 0
• alreadyAdded_chain = 0
• tmpProd_chain = 0
• incomingNodes = graph(graph(:,2)==IDsOverThreshold(idot),1)
• tempProd_chain_weight = 0
• wasteSpeciesFLAG = 0
• for innode
• nolnAcs = 1
• if alreadyAdded_ACS
• end end Reactions to
    distribuite in the different
    nature of the reactions weightToDistribute = graph(and((graph(:,1) == incomingNodes(innode)),(graph(:,2) ==
    IDsOverThreshold(idot))),6)
• if inSCCFlag == 1 % If the node is in an ACS
• else Otherwise it has been
    produced by an ACS
• end end If the species
    concentration but the species
    is not produced by other species
• rct = confparams.reactionProbability
• ecc = confparams.ECConcentration
• idOt = length(IDsOverThreshold)
• clear nrgTimeSeries
• clear gillTimeSeries = [rtime,gillMean,gillSD,gillEntropy,newSpeciesProb]
• nZeros = length(num2str(tmpL)) - length(num2str(tmpI))
• if nZeros for p
• tmpStrZeros = zeroBeforeStrNum(tmpRct, tmpRcts)
• outFile = strcat('_iGraph_CAT',tmpFilextPre,'_',tmpStrZeros,num2str(tmpRct),'_',num2str(tmpTime),'.csv')
• end fid1 = fopen(outFile,'a')
• for j
• end function [N, ids]
• ids = graph(graph(:,1)==graph(:,2),1)
• tline = fgets(fidConf)
• param = findstr(tline,'=')

```

13.6.1 Function Documentation

- 13.6.1.1 end If the prompt time is righe a message on the screen indicating the reaction number and the time is shown if and (`rtime > rctIDshowNoSave *analysisTimeIntervalNoSave`), (`printTemporalMessage==1`))
- 13.6.1.2 `cd (params. tmpPath)`
- 13.6.1.3 Go into the results folder `cd (strcat(simDirs(IDF).name,'/res'))`
- 13.6.1.4 `cd ('../0_statistics')`
- 13.6.1.5 `cd (currentDir)`
- 13.6.1.6 `cd ('/res')`
- 13.6.1.7 `cd (strcat('../', tmpDir))`
- 13.6.1.8 For each folder the necessary computations are performed `disp (`
`'*****'`)
- 13.6.1.9 `disp ('*NEW SIMULATION ANALYSIS PROCESS')`
- 13.6.1.10 end If there are species over theshold network analysis is performed `disp ('|-Graph Creation')`
- 13.6.1.11 `disp (strcat('|-File ', rcsFiles(rfileID).name, 'processing...'))`
- 13.6.1.12 If the time is righe save iGraph structures to file `disp (sprintf('|-%s|Reaction%d-time:%6.4f-Save structures',`
`simDirs(IDF).name, reaction, rtime))`
- 13.6.1.13 `disp (msg)`
- 13.6.1.14 NET ANALYSIS `disp ('|-*****NETWORK ANALYSIS *****')`
- 13.6.1.15 `disp ('|-Strongly connected components analysis...')`
- 13.6.1.16 `disp (sprintf('<> SCC n.%d', c))`
- 13.6.1.17 end end `disp (sprintf('|-Number of ACS:%d', realSccs))`
- 13.6.1.18 `disp (sprintf('|-Number of ACS(length 1):%d', self))`
- 13.6.1.19 `disp (sprintf('|-Species over threshold:%d', length(IDsOverThreshold)))`
- 13.6.1.20 `disp (fprintf('\t Within Acs%d->%d#%d-[%d]%d%6.4f', incomingNodes(innode), IDsOverThreshold(idot),`
`weightToDistribute,...IDsOverThreshold(idot), IDsOverThreshold(idot), concVec(idot)))`
- 13.6.1.21 `disp (fprintf('\t From Acs%d->%d#%d-[%d]%d%6.4f', incomingNodes(innode), IDsOverThreshold(idot),`
`weightToDistribute,...IDsOverThreshold(idot), IDsOverThreshold(idot), concVec(idot)))`
- 13.6.1.22 end end `disp (fprintf('\t<> Number of Structural Autocatalytic set of molecules:%d', realSccs))`
- 13.6.1.23 end `disp (fprintf('\t<>-----'))`
- 13.6.1.24 `disp (fprintf('\t<> Species over threshold produced by a CHAIN:%d', prod_chain))`
- 13.6.1.25 if `realSccs` `disp (fprintf('\t<> Species over threshold produced INTO an ACS:%d', prod_inSCC))`

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13.6.1.50 fprintf ( fidFINAL , '\n%s\t%6.4ft%6.4ft%6.4ft%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%d\t%6.4ft%6.4ft%6.4ft%6.4ft%d\t%d\t%d\t%d\t%d', ...simDirs(IDF). name, rct, ecc, idOt, realSccs,
prod_inSCC, prod_bySCC, prod_chain, prod_overlap, autocatalysis, prod_inSCC_weight,
prod_bySCC_weight, prod_chain_weight, prod_overlap_weight, self_loop_weight, conc_inSCC,
conc_bySCC, conc_chain, conc_selfCat, endo_condensation_counter, condensation_counter,
endo_cleavage_counter, cleavage_counter, wasteSpecies )

13.6.1.51 end gillTimeSeries ( rlineID , : )

13.6.1.52 graph ( : , 5 )

13.6.1.53 graph ( position , : )

13.6.1.54 graphSUB ( : , 5 )

13.6.1.55 graphSUB ( position , : )

13.6.1.56 end end if ( cc ==0 )

13.6.1.57 while ischar ( rlineb )

13.6.1.58 while ischar ( tline )

13.6.1.59 if isequal ( tline(1:param-1), 'nSeconds' )

13.6.1.60 elseif isequal ( tline(1:param-1), 'nGEN' )

13.6.1.61 elseif isequal ( tline(1:param-1), 'nSIM' )

13.6.1.62 elseif isequal ( tline(1:param-1), 'nReactions' )

13.6.1.63 elseif isequal ( tline(1:param-1), 'timeStructuresSavingInterval' )

13.6.1.64 elseif isequal ( tline(1:param-1), 'reactionProbability' )

13.6.1.65 elseif isequal ( tline(1:param-1), 'energy' )

13.6.1.66 elseif isequal ( tline(1:param-1), 'ECCConcentration' ) = str2num(tline(param+1:length(tline)))

13.6.1.67 elseif isequal ( tline(1:param-1), 'influx_rate' )

13.6.1.68 elseif isequal ( tline(1:param-1), 'maxLOut' )

13.6.1.69 elseif isequal ( tline(1:param-1), 'volume' )

13.6.1.70 legend ( 'Added', 'Removed' )

13.6.1.71 :,2 mol_I ( ) [virtual]

13.6.1.72 :,2 mol_II ( ) [virtual]

13.6.1.73 e.g. sim 1 of 10 sims, strZero will be '0' in order to create a % file named XXX_01_XXX strZero =
zeroBeforeStrNum(IDF,length(simDirs names ( ) [virtual]

13.6.1.74 while rline ( end )

```



```

13.6.1.75 saveas ( figure1 , fileName )

13.6.1.76 saveas ( figure2 , fileName )

13.6.1.77 saveas ( figure3 , fileName )

13.6.1.78 saveas ( figure4 , fileName )

13.6.1.79 saveas ( figure5 , fileName )

13.6.1.80 saveas ( figure6 , fileName )

13.6.1.81 saveas ( figure7 , fileName )

13.6.1.82 saveGraphSUBToFile ( folderSub , reaction , rtime , confparams. nReactions, graphSUB , filextPre )

13.6.1.83 end function saveGraphSUBToFile ( tmpDir , tmpRct , tmprTime , tmpRcts , tmpGraph , tmpFilextPre )

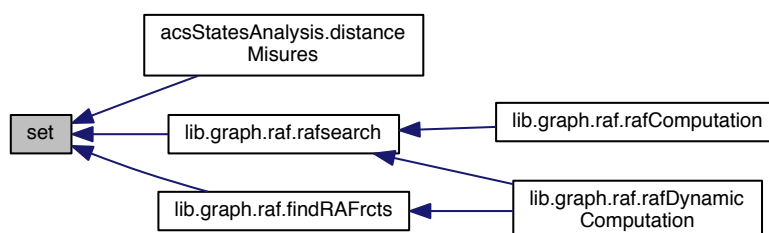
13.6.1.84 saveGraphToFile ( folderCat , reaction , rtime , confparams. nReactions, graph , filextPre )

13.6.1.85 end function saveGraphToFile ( tmpDir , tmpRct , tmprTime , tmpRcts , tmpGraph , tmpFilextPre )

13.6.1.86 grid on set ( gca , 'fontsize' , 15 , 'fontname' , 'times' )

```

Here is the caller graph for this function:



```

13.6.1.87 tmpL,tmpL strZero ( ) [virtual]

13.6.1.88 params params Substrate ( If different from 1 ) -> PRO, Otherwise if the reaction is already present its parameters
are updatedif sum(and((graph(:, 1)==cat), graph(:, 2)==mol_I))==1position=and((graph(:, 1)==cat), graph(:,
2)==mol_I)

13.6.1.89 end if sum ( influx ==mol_I )

```

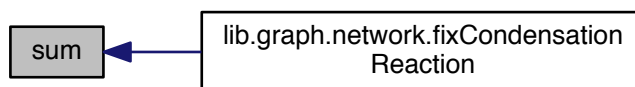
Initial value:

```

= 1
fluxEconomy = fluxEconomy + 1

```

Here is the caller graph for this function:



```

13.6.1.90 if sum ( find(scc==c) ==IDsOverThreshold (idot) )
13.6.1.91 end end end end if self if sum ( selfID ==idot )
13.6.1.92 :,4 timeInterval ( ) [virtual]
13.6.1.93 xlabel ( 'Time', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.94 ylabel ( 'Gillespie Mean', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.95 ylabel ( 'Gillespie SD', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.96 ylabel ( 'Entropy', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.97 ylabel ( 'New species Probability', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.98 ylabel ( 'Flux Molecules Balance', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.99 ylabel ( 'Ratio of backward reactions', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.100 ylabel ( 'Flux Molecules', 'Interpreter', 'latex', 'fontsize', 15 )
13.6.1.101 if ~isdir ( '0_statistics' )
13.6.1.102 if ~isdir ( strcat('..', folderCat) )
13.6.1.103 end if ~isdir ( strcat('..', folderSub) )
13.6.1.104 if ~isempty ( IDsOverThreshold )
13.6.1.105 if ~isempty ( incomingNodes )
  
```

13.6.2 Variable Documentation

13.6.2.1 else Otherwise it has been produced by an ACS

Definition at line 567 of file generalConcentrationOverThreshold.m.

13.6.2.2 alreadyAdded_ACS

Initial value:

```
== 0 % Add concentration to conc of ACSs
                                conc_inSCC =
    conc_inSCC + concVec(idot)
```

Definition at line 551 of file generalConcentrationOverThreshold.m.

13.6.2.3 if alreadyAdded_chain = 0

Definition at line 533 of file generalConcentrationOverThreshold.m.

13.6.2.4 else Otherwise it has been produced by an so it is a first layer leaf if alreadyAdded_leaves = 0

Definition at line 532 of file generalConcentrationOverThreshold.m.

13.6.2.5 autocatalysis = 0

Definition at line 501 of file generalConcentrationOverThreshold.m.

13.6.2.6 if realScCs If there are ACS for c

Initial value:

```
= 1 : length(numScC)
    if numScC(c) > 1
        sccID = sccID + 1
```

Definition at line 518 of file generalConcentrationOverThreshold.m.

13.6.2.7 cat = rline(4)

Definition at line 162 of file generalConcentrationOverThreshold.m.

13.6.2.8 catSparse = sparse(from,to,true,max(max(from,to)),max(max(from,to)))

Definition at line 513 of file generalConcentrationOverThreshold.m.

13.6.2.9 params params end end end else if cc = rline(3)

Definition at line 161 of file generalConcentrationOverThreshold.m.

13.6.2.10 else cleavage_counter = 0

Definition at line 131 of file generalConcentrationOverThreshold.m.

13.6.2.11 while cntrl

Definition at line 156 of file generalConcentrationOverThreshold.m.

13.6.2.12 conc_bySCC = 0

Definition at line 509 of file generalConcentrationOverThreshold.m.

13.6.2.13 `conc_chain = 0`

Definition at line 508 of file `generalConcentrationOverThreshold.m`.

13.6.2.14 `conc_inSCC = 0`

Definition at line 507 of file `generalConcentrationOverThreshold.m`.

13.6.2.15 `conc_selfCat = 0`

Definition at line 510 of file `generalConcentrationOverThreshold.m`.

13.6.2.16 `concVec = tmpConc`

Definition at line 117 of file `generalConcentrationOverThreshold.m`.

13.6.2.17 `condensation_counter = 0`

Definition at line 129 of file `generalConcentrationOverThreshold.m`.

13.6.2.18 `end function confparams = readParameters()`

Definition at line 71 of file `generalConcentrationOverThreshold.m`.

13.6.2.19 `end Set current date and current directory currentDate = date()`

Definition at line 21 of file `generalConcentrationOverThreshold.m`.

13.6.2.20 `currentDir = cd()`

Definition at line 22 of file `generalConcentrationOverThreshold.m`.

13.6.2.21 `currentFolder = cd()`

Definition at line 352 of file `generalConcentrationOverThreshold.m`.

13.6.2.22 `params params decayTime = 100`

Definition at line 12 of file `generalConcentrationOverThreshold.m`.

13.6.2.23 `ecc = confparams.ECConcentration`

Definition at line 640 of file `generalConcentrationOverThreshold.m`.

13.6.2.24 `endo_cleavage_counter = 0`

Definition at line 130 of file `generalConcentrationOverThreshold.m`.

13.6

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File Reference

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13.6.2.25 `else endo_condensation_counter = endo_condensation_counter + 1`

Definition at line 227 of file generalConcentrationOverThreshold.m.

13.6.2.26 `fid = fopen(rcsFiles(rfileID).name,'r')`

Definition at line 135 of file generalConcentrationOverThreshold.m.

13.6.2.27 `end fid1 = fopen(outFname,'a')`

Definition at line 696 of file generalConcentrationOverThreshold.m.

13.6.2.28 `end fidFINAL = fopen(outFileName,'w')`

Definition at line 38 of file generalConcentrationOverThreshold.m.

13.6.2.29 `figure1 = gcf`

Definition at line 374 of file generalConcentrationOverThreshold.m.

13.6.2.30 `figure2 = gcf`

Definition at line 390 of file generalConcentrationOverThreshold.m.

13.6.2.31 `figure3 = gcf`

Definition at line 406 of file generalConcentrationOverThreshold.m.

13.6.2.32 `figure4 = gcf`

Definition at line 422 of file generalConcentrationOverThreshold.m.

13.6.2.33 `figure5 = gcf`

Definition at line 438 of file generalConcentrationOverThreshold.m.

13.6.2.34 `figure6 = gcf`

Definition at line 454 of file generalConcentrationOverThreshold.m.

13.6.2.35 `figure7 = gcf`

Definition at line 471 of file generalConcentrationOverThreshold.m.

13.6.2.36 `if params figureVisible = 0`

Definition at line 16 of file generalConcentrationOverThreshold.m.

13.6.2.37 `fileName = strcat('25_gilleMean_',strZero,int2str(IDF),'.eps')`

Definition at line 372 of file `generalConcentrationOverThreshold.m`.

13.6.2.38 `SAVE THE GILLESPIE MATRXI ON FILE filename = strcat('0_gillespie_',strZero,int2str(IDF),'.txt')`

Definition at line 476 of file `generalConcentrationOverThreshold.m`.

13.6.2.39 `fileSpeciesID = 1`

Definition at line 107 of file `generalConcentrationOverThreshold.m`.

13.6.2.40 `filextPre = strcat('_',zeroBeforeStrNum(IDsimFOLDER, length(simDirs)),int2str(IDsimFOLDER))`

Definition at line 122 of file `generalConcentrationOverThreshold.m`.

13.6.2.41 `fluxEconomy = 0`

Definition at line 148 of file `generalConcentrationOverThreshold.m`.

13.6.2.42 `params params end end end end fluxEconomyArray = []`

Definition at line 151 of file `generalConcentrationOverThreshold.m`.

13.6.2.43 `fluxEconomyMatrix = [gillTimeSeries(:,1), fluxEconomyArray]`

Definition at line 479 of file `generalConcentrationOverThreshold.m`.

13.6.2.44 `fluxMinus = 0`

Definition at line 150 of file `generalConcentrationOverThreshold.m`.

13.6.2.45 `fluxPlus = 0`

Definition at line 149 of file `generalConcentrationOverThreshold.m`.

13.6.2.46 `if folderCat = strcat('__0_iGraph_CAT_', int2str(params.decayTime))`

Definition at line 95 of file `generalConcentrationOverThreshold.m`.

13.6.2.47 `end if folderSub = strcat('__0_iGraph_SUB_', int2str(params.decayTime))`

Definition at line 96 of file `generalConcentrationOverThreshold.m`.

13.6.2.48 `Computing actual number od strongly connected components from = graph(:,1)+1`

Definition at line 493 of file `generalConcentrationOverThreshold.m`.

13.6.2.49 **end function**[N, ids]
Initial value:

```
= self_loops(graph)
%Compute self-loop
N = sum(graph(:,1)==graph(:,2))
```

Definition at line 722 of file generalConcentrationOverThreshold.m.

13.6.2.50 **gillEntropy = rline(12)**

Definition at line 170 of file generalConcentrationOverThreshold.m.

13.6.2.51 **gillMean = rline(10)**

Definition at line 168 of file generalConcentrationOverThreshold.m.

13.6.2.52 **gillSD = rline(11)**

Definition at line 169 of file generalConcentrationOverThreshold.m.

13.6.2.53 **gillTimeSeries = [rtime,gillMean,gillSD,gillEntropy,newSpeciesProb]**

Definition at line 649 of file generalConcentrationOverThreshold.m.

13.6.2.54 **else graph = graph(graph(:,5)>0,:)**

Definition at line 214 of file generalConcentrationOverThreshold.m.

13.6.2.55 **else graphSUB = graphSUB(graphSUB(:,5)>0,:)**

Definition at line 220 of file generalConcentrationOverThreshold.m.

13.6.2.56 **for IDF**
Initial value:

```
=1:length(simDirs)
if isdir(strcat(simDirs(IDF).name))
    disp(sprintf('|- Processing dir %s',simDirs(IDF).name))
```

Definition at line 61 of file generalConcentrationOverThreshold.m.

13.6.2.57 **idOt = length(IDsOverThreshold)**

Definition at line 641 of file generalConcentrationOverThreshold.m.

13.6.2.58 **ids = graph(graph(:,1)==graph(:,2),1)**

Definition at line 725 of file generalConcentrationOverThreshold.m.

13.6.2.69 `mol_I = rline(5)`

Definition at line 163 of file generalConcentrationOverThreshold.m.

13.6.2.70 `params params end if mol_II = rline(6)`

Definition at line 164 of file generalConcentrationOverThreshold.m.

13.6.2.71 `mol_III = rline(7)`

Definition at line 165 of file generalConcentrationOverThreshold.m.

13.6.2.72 `newSpeciesProb = rline(13)`

Definition at line 171 of file generalConcentrationOverThreshold.m.

13.6.2.73 `end end end end end If both the species over threshold and the incoming node are not belonging to an ACS
if nInAcs = 1`

Definition at line 544 of file generalConcentrationOverThreshold.m.

13.6.2.74 `end if nrg`

Initial value:

```
== 1
nrgTimeSeries(rlineID,:) = [rtime,loadedMolsConc,loadedMols]
```

Definition at line 175 of file generalConcentrationOverThreshold.m.

13.6.2.75 `clear nrgTimeSeries`

Definition at line 648 of file generalConcentrationOverThreshold.m.

13.6.2.76 `numScc = histc(scc,1:max(scc))`

Definition at line 515 of file generalConcentrationOverThreshold.m.

13.6.2.77 `nZeros = length(num2str(tmpL)) - length(num2str(tmpI))`

Definition at line 673 of file generalConcentrationOverThreshold.m.

13.6.2.78 `Come back to the original folder end end out`

Initial value:

```
= generalConcentrationOverThreshold(params)

% param.threshold          # Threshold above which the concentration has
%   to be
% param.decayTime          # Reaction decay time
% param.simFolder          # Subtring identifiycating the folders containing sims
% param.tmpRctFileToLoad   # Reaction file to load
% param.tmpRctSUBFileToLoad # Reaction SUB file to load
```

```
if nargin < 1
    params.tmpPath = '~/Documents/simChiara/variaK_cpx/K_cpx_05'
```

Definition at line 1 of file generalConcentrationOverThreshold.m.

13.6.2.79 `end CREATE HEADER ROW IN THE FINAL OUTCOMES FILES` `outFileName = strcat(currentDate, '_',`
`int2str(params.threshold), '_convOverThreshold_', int2str(params.decayTime), '.csv')`

Definition at line 34 of file generalConcentrationOverThreshold.m.

13.6.2.80 `outFname = strcat('_iGraph_CAT', tmpFileExtPre, '_', tmpStrZeros, num2str(tmpRct), '_', num2str(tmpTime), '.csv')`

Definition at line 692 of file generalConcentrationOverThreshold.m.

13.6.2.81 `if nZeros for p`

Initial value:

```
=1:nZeros
    strZero = strcat(strZero, '0')
```

Definition at line 675 of file generalConcentrationOverThreshold.m.

13.6.2.82 `param = findstr(tline, '=')`

Definition at line 736 of file generalConcentrationOverThreshold.m.

13.6.2.83 `previousTime = 0`

Definition at line 138 of file generalConcentrationOverThreshold.m.

13.6.2.84 `printTemporalMessage = 1`

Definition at line 182 of file generalConcentrationOverThreshold.m.

13.6.2.85 `end prod_bySCC = 0`

Definition at line 498 of file generalConcentrationOverThreshold.m.

13.6.2.86 `prod_bySCC_weight = 0`

Definition at line 504 of file generalConcentrationOverThreshold.m.

13.6.2.87 `prod_chain = 0`

Definition at line 497 of file generalConcentrationOverThreshold.m.

13.6.2.88 `prod_chain_weight = 0`

Definition at line 503 of file generalConcentrationOverThreshold.m.

13.6.2.89 if then it has been produced within an ACS prod_inSCC = 0

Definition at line 496 of file generalConcentrationOverThreshold.m.

13.6.2.90 prod_inSCC_weight = 0

Definition at line 502 of file generalConcentrationOverThreshold.m.

13.6.2.91 end Compute the overlap between the different counter prod_overlap = 0

Definition at line 499 of file generalConcentrationOverThreshold.m.

13.6.2.92 prod_overlap_weight = 0

Definition at line 505 of file generalConcentrationOverThreshold.m.

13.6.2.93 ratioBackForward = rline(14)

Definition at line 172 of file generalConcentrationOverThreshold.m.

13.6.2.94 reaction parameters files rcsFiles = dir('*reactions_parameters*')

Definition at line 120 of file generalConcentrationOverThreshold.m.

13.6.2.95 rct = confparams.reactionProbability

Definition at line 639 of file generalConcentrationOverThreshold.m.

13.6.2.96 rctID = 1

Definition at line 141 of file generalConcentrationOverThreshold.m.

13.6.2.97 rctIDshow = 1

Definition at line 139 of file generalConcentrationOverThreshold.m.

13.6.2.98 rctIDshowNoSave = 1

Definition at line 140 of file generalConcentrationOverThreshold.m.

13.6.2.99 Craete different variables reaction = rline(1)

Definition at line 159 of file generalConcentrationOverThreshold.m.

13.6.2.100 realScCs = 0

Definition at line 517 of file generalConcentrationOverThreshold.m.

13.6.2.101 for rfileID**Initial value:**

```
=1:length(rcsFiles)
    % read species file

    %Initialize reaction type counter
    endo_condensation_counter = 0
```

Definition at line 124 of file generalConcentrationOverThreshold.m.

13.6.2.102 end rline = fgetl(fid)

Definition at line 143 of file generalConcentrationOverThreshold.m.

13.6.2.103 rlineb = rline

Definition at line 144 of file generalConcentrationOverThreshold.m.

13.6.2.104 Update rlineID rlineID = 1

Definition at line 145 of file generalConcentrationOverThreshold.m.

13.6.2.105 rtime = rline(2)

Definition at line 160 of file generalConcentrationOverThreshold.m.

13.6.2.106 sccID = 0

Definition at line 500 of file generalConcentrationOverThreshold.m.

**13.6.2.107 Come back to the original folder READ ALL THE DIRECTORY CONTAINING SIMULATIONS search =
strcat(' ',params.simFolder,'*')**

Definition at line 48 of file generalConcentrationOverThreshold.m.

13.6.2.108 self_loop_weight = 0

Definition at line 506 of file generalConcentrationOverThreshold.m.

13.6.2.109 simDirs = dir(search)

Definition at line 49 of file generalConcentrationOverThreshold.m.

13.6.2.110 params.simFolder = 'K_cpx5_rete_n_'

Definition at line 13 of file generalConcentrationOverThreshold.m.

13.6.2.111 end end If the species concentration but the species is not produced by other species

Definition at line 602 of file generalConcentrationOverThreshold.m.

13.6.2.112 LOAD FIRST SPECIES FILE `speciesFiles = dir('species_1*')`

Definition at line 74 of file `generalConcentrationOverThreshold.m`.

13.6.2.113 `tempProd_chain_weight = 0`

Definition at line 537 of file `generalConcentrationOverThreshold.m`.

13.6.2.114 `params.threshold = 0`

Definition at line 11 of file `generalConcentrationOverThreshold.m`.

13.6.2.115 : `timeInterval = rtime - previousTime`

Definition at line 201 of file `generalConcentrationOverThreshold.m`.

13.6.2.116 File Containing all Times `times = 0:params.deltaT:params.totT`

Definition at line 52 of file `generalConcentrationOverThreshold.m`.

13.6.2.117 `tline = fgets(fidConf)`

Definition at line 732 of file `generalConcentrationOverThreshold.m`.

13.6.2.118 `tmpProd_chain = 0`

Definition at line 534 of file `generalConcentrationOverThreshold.m`.

13.6.2.119 `params.tmpRctFileToLoad = ''`

Definition at line 14 of file `generalConcentrationOverThreshold.m`.

13.6.2.120 `params.tmpRctSUBFileToLoad = ''`

Definition at line 15 of file `generalConcentrationOverThreshold.m`.

13.6.2.121 `tmpStrZeros = zeroBeforeStrNum(tmpRct, tmpRcts)`

Definition at line 691 of file `generalConcentrationOverThreshold.m`.

13.6.2.122 `to = graph(:,2)+1`

Definition at line 494 of file `generalConcentrationOverThreshold.m`.

13.6.2.123 `wasteSpecies = 0`

Definition at line 511 of file `generalConcentrationOverThreshold.m`.

13.6.2.124 **end end** If the species concentration but the species is not produced by other so it is waste if
 wasteSpeciesFLAG = 0

Definition at line 538 of file generalConcentrationOverThreshold.m.

13.6.2.125 **end end** Reactions to distribute in the different nature of the reactions weightToDistribute =
 graph(and((graph(:,1) == incomingNodes(innode)),(graph(:,2) == IDsOverThreshold(idot))),6)

Definition at line 558 of file generalConcentrationOverThreshold.m.

13.7 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/KillSpam.m File Reference

Functions

- `function KillSpam (param) currentDir`
- `cd (param.tmpPath)`
- `disp ('|-----|')`
- `disp ('|-Killer and Spammer analysis is started|')`
- `READ ALL THE DIRECTORY`
- `CONTAINING SIMULATIONS if ~isdir ('0_statistics') mkdir('0_statistics')`
- `cd ('0_statistics')`
- `fclose (fid)`
- `cd ('../')`
- `disp (strcat('|-Folder: ', simDirs(x).name,'/res...'))`
- `break end reactionsMatrix (continua, 1:6)`
- `cleavagesMatrix (cleavages, 1:6)`
- `condensationsMatrix (condensations, 1:6)`
- `end catalysisMatrix (continua, 1:4)`
- `outliersMatrix (outRows, 1:6)`
- `if ((outliersMatrix(i, 2)==1)&&(check2==1 &&check3==1))|(outliersMatrix(i`
- `killersMatrix (killRows, 1:6)`
- `killerCatalysts (KCRows, 1)`
- `killerCatalysts (KCRows, 2)`
- `killerCatalysts (KCRows, 3)`
- `id KSRows ()`
- `if outliersMatrix (i, 2)`
- `KSMatrix (KSRows, 1:6)`
- `KSMatrix (i,:)`
- `KSCatalysts (KSCRows, 1)`
- `KSCatalysts (KSCRows, 2)`
- `KSCatalysts (KSCRows, 3)`
- `if ((outliersMatrix(i, 2)==1)&&(check2==0 &&check3==0))|(outliersMatrix(i`
- `SpammersMatrix (SpamRows, 1:6)`
- `if SpamRows SpammersMatrix (:, 7)`
- `SpammersMatrix (i,:)`
- `else SpammersMatrix (i, 7)`
- `spammerCatalysts (KSCRows, 1)`
- `spammerCatalysts (KSCRows, 2)`
- `spammerCatalysts (KSCRows, 3)`
- `fprintf (fid,'NET%d\n\n', x)`
- `fprintf (fid,'KILLERS\n\n')`
- `end fprintf (fid,'\n\n')`

- `fprintf(fid,'Catalysts\n\n')`
- `fprintf(fid,'%d\t', killerCatalysts(i, 2))`
- `fprintf(fid,'%d\n\n', killerCatalysts(i, 3))`
- `end fprintf(fid,'KILLERS-SPAMMERS\n\n')`
- `fprintf(fid,'%d\t', KSCatalysts(KSCRows, 2))`
- `fprintf(fid,'%d\n\n', KSCatalysts(KSCRows, 3))`
- `end fprintf(fid,'SPAMMERS\n\n')`
- `fprintf(fid,'%d\t', spammerCatalysts(SCRows, 2))`
- `fprintf(fid,'%d\n\n', spammerCatalysts(SCRows, 3))`
- `end cd(currentDir)`

Variables

- `end tmpFolder = strcat('*',param.simFolder,'*')`
- `simDirs = dir(tmpFolder)`
- `fid = fopen('ALL_results_TS1.txt','w')`
- `for x`
- `blocked = 0:param.lastSpecies`
- creating `reactions` matrix `rFiles = dir('reactions*')`
- `reactionsFile = rFiles(length(rFiles)-1).name`
- `continua = 1`
- `reactionsMatrix = 0`
- `while continua rValues = fscanf(fid,'%d',7)`
- `stop = isempty(rValues)`
- dividing into condensations
and cleavages condensations = 0
- `cleavages = 0`
- `cleavagesMatrix = 0`
- `condensationsMatrix = 0`
- `for i`
- creating `catalysis` matrix `cFiles = dir('catalysis*')`
- `catalysisFile = cFiles(length(cFiles)).name`
- `catalysisMatrix = 0`
- `while continua cValues = fscanf(fid,'%f',7)`
- `break`
- identifying outliers `reactionsCounters = reactionsMatrix(:,6)`
- `outRows = 0`
- `outliersMatrix = 0`
- `if possible`
- identifying killers `killRows = 0`
- `killersMatrix = 0`
- `check2 = ismember(outliersMatrix(i,4),blocked)`
- `check3 = ismember(outliersMatrix(i,5),blocked)`
- `killerCatalysts = 0`
- `KCRows = 0`
- `for j`
- `KSMatrix = 0`
- `check4 = xor(check2,check3)`
- `check6 = ismember(KSMatrix(i,5),cleavagesMatrix(:,3))`
- `check7 = ismember(KSMatrix(i,5),catalysisMatrix(:,2))`
- `if check5`
- `checkKSM = isempty(KSMatrix)`
- `KSCatalysts = 0`
- `KSCRows = 0`

- `end` identifying spammers `SpamRows = 0`
- `SpammersMatrix = 0`
- `end checkSM = isempty(SpammersMatrix)`
- `spammersCatalysts = 0`
- `SCRows = 0`
- `checkKM = isempty(killersMatrix)`
- `else [r c] = size(killersMatrix)`
- `clear r`
- `clear c`

13.7.1 Function Documentation

13.7.1.1 `end catalysisMatrix (continua , 1:4)`

13.7.1.2 `cd (param. tmpPath)`

13.7.1.3 `cd ('0_statistics')`

13.7.1.4 `cd ('..')`

13.7.1.5 `end cd (currentDir)`

13.7.1.6 `cleavagesMatrix (cleavages , 1:6)`

13.7.1.7 `condensationsMatrix (condensations , 1:6)`

13.7.1.8 `disp ('|-----|')`

13.7.1.9 `disp ('|-Killer and Spammer analysis is started|')`

13.7.1.10 `disp (strcat('|-Folder: ', simDirs(x).name,'res...'))`

13.7.1.11 `fclose (fid)`

13.7.1.12 `fprintf (fid , 'NET%d\n\n' , x)`

13.7.1.13 `fprintf (fid , 'KILLERS\n\n')`

13.7.1.14 `end fprintf (fid , '\n\n')`

13.7.1.15 `fprintf (fid , 'Catalysts\n\n')`

13.7.1.16 `fprintf (fid , '%d\t' , killerCatalysts(i,2))`

13.7.1.17 `fprintf (fid , '%d\n\n' , killerCatalysts(i,3))`

13.7.1.18 `end fprintf (fid , 'KILLERS-SPAMMERS\n\n')`

13.7.1.19 `fprintf (fid , '%d\t' , KSCatalysts(KSCRows,2))`

13.7.1.20 `fprintf (fid , '%d\n\n' , KSCatalysts(KSCRows,3))`

13.7.1.21 `end fprintf (fid , 'SPAMMERS\n\n')`

13.7.1.22 `fprintf (fid , '%d\t' , spammerCatalysts(SCRows,2))`

13.7.1.23 `fprintf (fid , '%d\n\n' , spammerCatalysts(SCRows,3))`

13.7.1.24 `if ((outliersMatrix(i,2)==1)&&(check2==1 &&check3==1))`

13.7.1.25 `if ((outliersMatrix(i,2)==1)&&(check2==0 &&check3==0))`

13.7.1.26 `killerCatalysts (KCRows , 1)`

13.7.1.27 `killerCatalysts (KCRows , 2)`

13.7.1.28 `killerCatalysts (KCRows , 3)`

13.7.1.29 `killersMatrix (killRows , 1:6)`

13.7.1.30 `function KillSpam (param)`

13.7.1.31 `KSCatalysts (KSCRows , 1)`

13.7.1.32 `KSCatalysts (KSCRows , 2)`

13.7.1.33 `KSCatalysts (KSCRows , 3)`

13.7.1.34 `KSMatrix (KSRows , 1:6)`

13.7.1.35 `KSMatrix (i , :)`

13.7.1.36 `id KSRows () [virtual]`

13.7.1.37 `outliersMatrix (outRows , 1:6)`

13.7.1.38 `if outliersMatrix (i , 2)`

13.7.1.39 `break end reactionsMatrix (continua , 1:6)`

13.7.1.40 `spammerCatalysts (KSCRows , 1)`

13.7.1.41 `spammerCatalysts (KSCRows , 2)`

13.7.1.42 `spammerCatalysts (KSCRows , 3)`

13.7.1.43 `SpammersMatrix (SpamRows , 1:6)`

13.7.1.44 `if SpamRows SpammersMatrix (: , 7)`

13.7.1.45 `SpammersMatrix (i , :)`

13.7.1.46 `else SpammersMatrix (i , 7)`

13.7.1.47 `READ ALL THE DIRECTORY CONTAINING SIMULATIONS if ~isdir ('0_statistics')`

13.7.2 Variable Documentation

13.7.2.1 `blocked = 0:param.lastSpecies`

Definition at line 27 of file KillSpam.m.

13.7.2.2 break

Definition at line 74 of file KillSpam.m.

13.7.2.3 clear c

Definition at line 273 of file KillSpam.m.

13.7.2.4 catalysisFile = cFiles(length(cFiles)).name

Definition at line 65 of file KillSpam.m.

13.7.2.5 catalysisMatrix = 0

Definition at line 68 of file KillSpam.m.

13.7.2.6 creating catalysis matrix cFiles = dir('catalysis*')

Definition at line 64 of file KillSpam.m.

13.7.2.7 check2 = ismember(outliersMatrix(i,4),blocked)

Definition at line 106 of file KillSpam.m.

13.7.2.8 check3 = ismember(outliersMatrix(i,5),blocked)

Definition at line 107 of file KillSpam.m.

13.7.2.9 check4 = xor(check2,check3)

Definition at line 139 of file KillSpam.m.

13.7.2.10 if check5

Initial value:

```
== 1 || check6 == 1 || check7 == 1
      KSRows = KSRows-1
```

Definition at line 152 of file KillSpam.m.

13.7.2.11 check6 = ismember(KSMatrix(i,5),cleavagesMatrix(:,3))

Definition at line 150 of file KillSpam.m.

13.7.2.12 check7 = ismember(KSMatrix(i,5),catalysisMatrix(:,2))

Definition at line 151 of file KillSpam.m.

13.7.2.13 if checkKM = isempty(killersMatrix)

Definition at line 259 of file KillSpam.m.

13.7.2.14 if checkKSM = isempty(KSMatrix)

Definition at line 168 of file KillSpam.m.

13.7.2.15 if checkSM = isempty(SpammersMatrix)

Definition at line 234 of file KillSpam.m.

13.7.2.16 cleavages = 0

Definition at line 48 of file KillSpam.m.

13.7.2.17 cleavagesMatrix = 0

Definition at line 50 of file KillSpam.m.

13.7.2.18 else condensations = 0

Definition at line 47 of file KillSpam.m.

13.7.2.19 condensationsMatrix = 0

Definition at line 51 of file KillSpam.m.

13.7.2.20 continua = 1

Definition at line 32 of file KillSpam.m.

13.7.2.21 while continua cValues = fscanf(fid,'%f',7)

Definition at line 70 of file KillSpam.m.

13.7.2.22 else[r c] = size(killersMatrix)

Definition at line 263 of file KillSpam.m.

13.7.2.23 fid = fopen('ALL_results_TS1.txt','w')

Definition at line 20 of file KillSpam.m.

13.7.2.24 end clear i**Initial value:**

```
= 1:rrm
    if reactionsMatrix(i,2) == 1
        cleavages = cleavages+1
```

Definition at line 52 of file KillSpam.m.

13.7.2.25 clear j

Initial value:

```
= 1:rcm
    if killerReaction == catalysisMatrix(j,3) &&
        catalysisMatrix(j,4) > 0
        KCRows = KCRows+1
```

Definition at line 120 of file KillSpam.m.

13.7.2.26 KCRows = 0

Definition at line 117 of file KillSpam.m.

13.7.2.27 killerCatalysts = 0

Definition at line 116 of file KillSpam.m.

13.7.2.28 killersMatrix = 0

Definition at line 103 of file KillSpam.m.

13.7.2.29 if killRows = 0

Definition at line 102 of file KillSpam.m.

13.7.2.30 KSCatalysts = 0

Definition at line 171 of file KillSpam.m.

13.7.2.31 KSCRows = 0

Definition at line 172 of file KillSpam.m.

13.7.2.32 KSMatrix = 0

Definition at line 135 of file KillSpam.m.

13.7.2.33 outliersMatrix = 0

Definition at line 91 of file KillSpam.m.

13.7.2.34 outRows = 0

Definition at line 90 of file KillSpam.m.

13.7.2.35 if possible

Initial value:

```
== 1
    outRows = outRows+1
```

Definition at line 94 of file KillSpam.m.

13.7.2.36 clear r

Definition at line 272 of file KillSpam.m.

13.7.2.37 identifying outliers reactionsCounters = reactionsMatrix(:,6)

Definition at line 83 of file KillSpam.m.

13.7.2.38 reactionsFile = rFiles(length(rFiles)-1).name

Definition at line 30 of file KillSpam.m.

13.7.2.39 reactionsMatrix = 0

Definition at line 33 of file KillSpam.m.

13.7.2.40 creating reactions matrix rFiles = dir('reactions*')

Definition at line 29 of file KillSpam.m.

13.7.2.41 while continua rValues = fscanf(fid,'%d',7)

Definition at line 35 of file KillSpam.m.

13.7.2.42 SCRows = 0

Definition at line 238 of file KillSpam.m.

13.7.2.43 simDirs = dir(tmpFolder)

Definition at line 17 of file KillSpam.m.

13.7.2.44 spammersCatalysts = 0

Definition at line 237 of file KillSpam.m.

13.7.2.45 SpammersMatrix = 0

Definition at line 190 of file KillSpam.m.

13.7.2.46 end identifying spammers SpamRows = 0

Definition at line 189 of file KillSpam.m.

13.7.2.47 if stop = isempty(rValues)

Definition at line 36 of file KillSpam.m.

13.7.2.48 end tmpFolder = strcat('*',param.simFolder,'*')

Definition at line 16 of file KillSpam.m.

13.7.2.49 for x

Initial value:

```
= 1:length(simDirs)
    cd(strcat(simDirs(x).name,'/res'))
```

Definition at line 24 of file KillSpam.m.

13.8 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/KSSearch.m File Reference

Functions

- break end reactionsMatrix (continua, 1:6)
- end fclose (fid)
- cleavagesMatrix (i, 1:6)
- condensationsMatrix (i, 1:6)
- break end catalysisMatrix (continua, 1:7)
- outliersMatrix (outRows, 1:6)
- if ((outliersMatrix(i, 2)==0)&&(check2==1 &&check3==1))||(outliersMatrix(i
- killersMatrix (killRows, 1:6)
- id KSRows ()
- if outliersMatrix (i, 2)
- KSMatrix (KSRows, 1:6)
- KSMatrix (i,:)
- if ((outliersMatrix(i, 2)==0)&&(check2==0 &&check3==0))||(outliersMatrix(i
- SpammersMatrix (SpamRows, 1:6)
- if SpamRows SpammersMatrix (:, 7)
- SpammersMatrix (i,:)
- else SpammersMatrix (i, 7)

Variables

- blocked = 0:5
- creating reactions matrix rFiles = dir('reactions*')
- reactionsFile = rFiles(length(rFiles)-1).name
- fid = fopen(reactionsFile)
- continua = 1

- `reactionsMatrix = 0`
- while `continua rValues = fscanf(fid,'%d',7)`
- `stop = isempty(rValues)`
- dividing into condensations
and `cleavages condensations = 0`
- `cleavages = 0`
- `cleavagesMatrix = 0`
- `condensationsMatrix = 0`
- for `i`
- creating `catalysis` matrix `cFiles = dir('catalysis*')`
- `catalysisFile = cFiles(length(cFiles)).name`
- `catalysisMatrix = 0`
- while `continua cValues = fscanf(fid,'%f',7)`
- identifying outliers `reactionsCounters = sort(reactionsMatrix(:,6))'`
- `fpc = ceil(rrm/20*19)`
- `outliersCounter = reactionsCounters(fpc:rrm)`
- `outRows = 0`
- `outliersMatrix = 0`
- if possible
- identifying killers `killRows = 0`
- `killersMatrix = 0`
- `check2 = ismember(outliersMatrix(i,4),blocked)`
- `check3 = ismember(outliersMatrix(i,5),blocked)`
- `KSMatrix = 0`
- `check4 = xor(check2,check3)`
- `check6 = ismember(KSMatrix(i,5),cleavagesMatrix(:,3))`
- `check7 = ismember(KSMatrix(i,5),catalysisMatrix(:,2))`
- if `check5`
- identifying spammers `SpamRows = 0`
- `SpammersMatrix = 0`

13.8.1 Function Documentation

13.8.1.1 `break end catalysisMatrix (continua , 1:7)`

13.8.1.2 `cleavagesMatrix (i , 1:6)`

13.8.1.3 `condensationsMatrix (i , 1:6)`

13.8.1.4 `end fclose (fid)`

13.8.1.5 `if ((outliersMatrix(i,2)==0)&&(check2==1 &&check3==1))`

13.8.1.6 `if ((outliersMatrix(i,2)==0)&&(check2==0 &&check3==0))`

13.8.1.7 `killersMatrix (killRows , 1:6)`

13.8.1.8 `KSMatrix (KSRows , 1:6)`

13.8.1.9 `KSMatrix (i , :)`

13.8.1.10 `id KSRows () [virtual]`

13.8.1.11 `outliersMatrix (outRows , 1:6)`

13.8.1.12 **if outliersMatrix (i , 2)**

13.8.1.13 **break end reactionsMatrix (continua , 1:6)**

13.8.1.14 **SpammersMatrix (SpamRows , 1:6)**

13.8.1.15 **if SpamRows SpammersMatrix (: , 7)**

13.8.1.16 **SpammersMatrix (i , :)**

13.8.1.17 **else SpammersMatrix (i , 7)**

13.8.2 Variable Documentation

13.8.2.1 **blocked = 0:5**

Definition at line 1 of file KSSearch.m.

13.8.2.2 **catalysisFile = cFiles(length(cFiles)).name**

Definition at line 40 of file KSSearch.m.

13.8.2.3 **catalysisMatrix = 0**

Definition at line 43 of file KSSearch.m.

13.8.2.4 **creating catalysis matrix cFiles = dir('catalysis*')**

Definition at line 39 of file KSSearch.m.

13.8.2.5 **check2 = ismember(outliersMatrix(i,4),blocked)**

Definition at line 82 of file KSSearch.m.

13.8.2.6 **check3 = ismember(outliersMatrix(i,5),blocked)**

Definition at line 83 of file KSSearch.m.

13.8.2.7 **check4 = xor(check2,check3)**

Definition at line 97 of file KSSearch.m.

13.8.2.8 **if check5**

Initial value:

```
== 1 || check6 == 1 || check7 == 1
      KSRows = KSRows-1
```

Definition at line 110 of file KSSearch.m.

13.8.2.9 `check6 = ismember(KSMatrix(i,5),cleavagesMatrix(:,3))`

Definition at line 108 of file KSSearch.m.

13.8.2.10 `check7 = ismember(KSMatrix(i,5),catalysisMatrix(:,2))`

Definition at line 109 of file KSSearch.m.

13.8.2.11 `cleavages = 0`

Definition at line 23 of file KSSearch.m.

13.8.2.12 `cleavagesMatrix = 0`

Definition at line 25 of file KSSearch.m.

13.8.2.13 `else condensations = 0`

Definition at line 22 of file KSSearch.m.

13.8.2.14 `condensationsMatrix = 0`

Definition at line 26 of file KSSearch.m.

13.8.2.15 `continua = 1`

Definition at line 7 of file KSSearch.m.

13.8.2.16 `while continua cValues = fscanf(fid,'%f',7)`

Definition at line 45 of file KSSearch.m.

13.8.2.17 `fid = fopen(reactionsFile)`

Definition at line 6 of file KSSearch.m.

13.8.2.18 `fpc = ceil(rrm/20*19)`

Definition at line 58 of file KSSearch.m.

13.8.2.19 `end end end clear i`

Initial value:

```
= 1:rrm
    if reactionsMatrix(i,2) == 0
        cleavages = cleavages+1
```

Definition at line 27 of file KSSearch.m.

13.8.2.20 killersMatrix = 0

Definition at line 79 of file KSSearch.m.

13.8.2.21 identifying killers killRows = 0

Definition at line 78 of file KSSearch.m.

13.8.2.22 KSMatrix = 0

Definition at line 93 of file KSSearch.m.

13.8.2.23 outliersCounter = reactionsCounters(fpc:rrm)

Definition at line 59 of file KSSearch.m.

13.8.2.24 outliersMatrix = 0

Definition at line 67 of file KSSearch.m.

13.8.2.25 outRows = 0

Definition at line 66 of file KSSearch.m.

13.8.2.26 if possible**Initial value:**

```
== 1      outRows = outRows+1
```

Definition at line 70 of file KSSearch.m.

13.8.2.27 identifying outliers reactionsCounters = sort(reactionsMatrix(:,6))'

Definition at line 57 of file KSSearch.m.

13.8.2.28 reactionsFile = rFiles(length(rFiles)-1).name

Definition at line 5 of file KSSearch.m.

13.8.2.29 reactionsMatrix = 0

Definition at line 8 of file KSSearch.m.

13.8.2.30 creating reactions matrix rFiles = dir('reactions*')

Definition at line 4 of file KSSearch.m.

13.8.2.31 while continua rValues = fscanf(fid,'%d',7)

Definition at line 10 of file KSSearch.m.

13.8.2.32 SpammersMatrix = 0

Definition at line 129 of file KSSearch.m.

13.8.2.33 identifying spammers SpamRows = 0

Definition at line 128 of file KSSearch.m.

13.8.2.34 if stop = isempty(rValues)

Definition at line 11 of file KSSearch.m.

13.9 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/KSSearchLauncher.m File Reference

Functions

- [fclose](#) (fid)
- [cd](#) (currentDir)
- [fprintf](#) (fid,'NET%d\n\n', x)
- [fprintf](#) (fid,'KILLERS\n\n')
- [end fprintf](#) (fid,'\n\n')
- [fprintf](#) (fid,'KILLERS-SPAMMERS\n\n')
- [fprintf](#) (fid,'SPAMMERS\n\n')

Variables

- [currentDir](#) = [cd](#)()
- [simDirs](#) = [dir](#)('sim_*')
- [fid](#) = [fopen](#)('results.txt','w')
- [for](#) x
- [KSSearch](#)
- [checkKM](#) = [isempty](#)(killersMatrix)
- [else](#) [r c] = [size](#)(killersMatrix)
- [for](#) i
- [clear](#) j
- [clear](#) r
- [clear](#) c
- [checkKSM](#) = [isempty](#)(KSMatrix)
- [checkSM](#) = [isempty](#)(SpammersMatrix)

13.9.1 Function Documentation

13.9.1.1 `cd(currentDir)`

13.9.1.2 `fclose(fid)`

13.9.1.3 `fprintf(fid, 'NET%d\n\n', x)`

13.9.1.4 `fprintf(fid, 'KILLERS\n\n')`

13.9.1.5 `end fprintf(fid, '\n\n')`

13.9.1.6 `fprintf(fid, 'KILLERS-SPAMMERS\n\n')`

13.9.1.7 `fprintf(fid, 'SPAMMERS\n\n')`

13.9.2 Variable Documentation

13.9.2.1 `clear c`

Definition at line 27 of file `KSSearchLauncher.m`.

13.9.2.2 `if checkKM = isempty(killersMatrix)`

Definition at line 12 of file `KSSearchLauncher.m`.

13.9.2.3 `if checkKSM = isempty(KSMatrix)`

Definition at line 29 of file `KSSearchLauncher.m`.

13.9.2.4 `if checkSM = isempty(SpammersMatrix)`

Definition at line 46 of file `KSSearchLauncher.m`.

13.9.2.5 `currentDir = cd()`

Definition at line 1 of file `KSSearchLauncher.m`.

13.9.2.6 `else[r c] = size(killersMatrix)`

Definition at line 16 of file `KSSearchLauncher.m`.

13.9.2.7 `fid = fopen('results.txt','w')`

Definition at line 3 of file `KSSearchLauncher.m`.

13.9.2.8 `end end clear i`

Initial value:

```
= 1:r
    for j = 1:c
        fprintf(fid,'%d\t',killersMatrix(i,j))
```

Definition at line 17 of file KSSearchLauncher.m.

13.9.2.9 clear j

Definition at line 25 of file KSSearchLauncher.m.

13.9.2.10 KSSearch

Definition at line 7 of file KSSearchLauncher.m.

13.9.2.11 clear r

Definition at line 26 of file KSSearchLauncher.m.

13.9.2.12 simDirs = dir('sim_*')

Definition at line 2 of file KSSearchLauncher.m.

13.9.2.13 for x

Initial value:

```
= 1:length(simDirs)
    cd(strcat(simDirs(x).name,'/res'))
```

Definition at line 5 of file KSSearchLauncher.m.

13.10 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/overallStats.m File Reference

Functions

- [function overallStats \(\)](#) close all clear all [p.tmpAnalysis](#)
- Used in KillSpan it represents the last [influx species](#) LATEST FILES ANALYSIS if [p tmpAnalysis](#) (1)
- [end if p tmpAnalysis](#) (2)
- [end if p tmpAnalysis](#) (3)
- [end if p tmpAnalysis](#) (4)
- [if p tmpAnalysis](#) (6)
- [if exist](#) ('MATLAB2R.Rout','file') delete('MATLAB2R.Rout')
- [eval](#) ([!R CMD BATCH--no-save--no-restore--slave"--args sf=",[scriptAnalDir](#)," ls=',num2str([p.lastSpecies](#)),...' sss=",[p.simFolder](#)," gfs=",[gfs](#)," sd=",[p.tmpPath](#),""MATLAB2R.R'])
- [cd](#) (crtDir)

Variables

- [p tmpPath](#) = '/Users/alessandrofilisetti/Documents/results/VILLANI/'
- [p figureVisible](#) = 0
- [p simFolder](#) = 'ACS2'
- [p lastSpecies](#) = 2

- Used in KillSpan [analysis](#)
- `p.threshold = 0`
- `p.tmpRctFileToLoad = ''`
- `p.tmpRctSUBFileToLoad = ''`
- `end scriptAnalDir = cd()`
- `gfs = strcat('_',num2str(p.decayTime))`

13.10.1 Function Documentation

13.10.1.1 `cd (crtDir)`

13.10.1.2 `eval ()`

13.10.1.3 `if exist ('MATLAB2R.Rout' , 'file')`

13.10.1.4 `function overallStats ()`

13.10.1.5 Used in KillSpan it represents the last influx species LATEST FILES ANALYSIS if `p.tmpAnalysis (1)`

13.10.1.6 `end if p.tmpAnalysis (2)`

13.10.1.7 `end if p.tmpAnalysis (3)`

13.10.1.8 `end if p.tmpAnalysis (4)`

13.10.1.9 `end Need p.tmpAnalysis (6)`

Initial value:

```
== 1
% Using an R-Cran script this part is a little bit different
crtDir = cd()
```

13.10.2 Variable Documentation

13.10.2.1 Used in KillSpan analysis

Definition at line 10 of file overallStats.m.

13.10.2.2 `p.figureVisible = 0`

Definition at line 8 of file overallStats.m.

13.10.2.3 `gfs = strcat('_',num2str(p.decayTime))`

Definition at line 50 of file overallStats.m.

13.10.2.4 `p.lastSpecies = 2`

Definition at line 10 of file overallStats.m.

13.10.2.5 `end scriptAnalDir = cd()`

Definition at line 49 of file overallStats.m.

13.10.2.6 `p simFolder = 'ACS2'`

Definition at line 9 of file overallStats.m.

13.10.2.7 `p threshold = 0`

Definition at line 36 of file overallStats.m.

13.10.2.8 `concAnalysis(params) clear all close all tmpPath = '/Users/alessandrofilisetti/Documents/results/VILLANI/'`

Definition at line 7 of file overallStats.m.

13.10.2.9 `p tmpRctFileToLoad = ''`

Definition at line 37 of file overallStats.m.

13.10.2.10 `p tmpRctSUBFileToLoad = ''`

Definition at line 38 of file overallStats.m.

13.11 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/readParameters.m File Reference

Functions

- `while ischar (tline) tline`
- `if isequal (tline(1:param-1),'nSeconds') confparams.nSeconds`
- `elseif isequal (tline(1:param-1),'nGEN') confparams.nGEN`
- `elseif isequal (tline(1:param-1),'nSIM') confparams.nSIM`
- `elseif isequal (tline(1:param-1),'nReactions') confparams.nReactions`
- `elseif isequal (tline(1:param-1),'timeStructuresSavingInterval') confparams.timeStructuresSavingInterval`
- `elseif isequal (tline(1:param-1),'fileTimesSaveInterval') confparams.fileTimesSaveInterval`
- `elseif isequal (tline(1:param-1),'reactionProbability') confparams.reactionProbability`
- `elseif isequal (tline(1:param-1),'energy') confparams.energy`
- `elseif isequal (tline(1:param-1),'ECConcentration') confparams.ECConcentration`
- `elseif isequal (tline(1:param-1),'influx_rate') confparams.influx_rate`
- `elseif isequal (tline(1:param-1),'maxLOut') confparams.maxLOut`
- `elseif isequal (tline(1:param-1),'volume') confparams.volume`
- `end end fclose (fidConf)`

Variables

- `function confparams`
- `tline = fgets(fidConf)`
- `param = findstr(tline,'=')`

13.11.1 Function Documentation

13.11.1.1 `end end fclose (fidConf)`

13.11.1.2 `while ischar (tline)`

13.11.1.3 `if isequal (tline(1:param-1), 'nSeconds')`

13.11.1.4 `elseif isequal (tline(1:param-1), 'nGEN')`

13.11.1.5 `elseif isequal (tline(1:param-1), 'nSIM')`

13.11.1.6 `elseif isequal (tline(1:param-1), 'nReactions')`

13.11.1.7 `elseif isequal (tline(1:param-1), 'timeStructuresSavingInterval')`

13.11.1.8 `elseif isequal (tline(1:param-1), 'fileTimesSaveInterval')`

13.11.1.9 `elseif isequal (tline(1:param-1), 'reactionProbability')`

13.11.1.10 `elseif isequal (tline(1:param-1), 'energy')`

13.11.1.11 `elseif isequal (tline(1:param-1), 'ECConcentration')`

13.11.1.12 `elseif isequal (tline(1:param-1), 'influx_rate')`

13.11.1.13 `elseif isequal (tline(1:param-1), 'maxLOut')`

13.11.1.14 `elseif isequal (tline(1:param-1), 'volume')`

13.11.2 Variable Documentation

13.11.2.1 `function confparams`

Initial value:

```
= readParameters()
    fidConf=fopen('acsm2s.conf','r')
```

Definition at line 1 of file readParameters.m.

13.11.2.2 `param = findstr(tline,'=')`

Definition at line 9 of file readParameters.m.

13.11.2.3 `tline = fgets(fidConf)`

Definition at line 5 of file readParameters.m.

13.12 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/resetForNew-Simulations.py File Reference

Namespaces

- [resetForNewSimulations](#)

Functions

- def [resetForNewSimulations.zeroBeforeStrNum](#)

Variables

- int [resetForNewSimulations.foldersSIMS](#) = 10
- int [resetForNewSimulations.foldersREP](#) = 10
- tuple [resetForNewSimulations.zerosSIMS](#) = zeroBeforeStrNum(i,foldersSIMS)
- tuple [resetForNewSimulations.zerosREPS](#) = zeroBeforeStrNum(j,foldersREP)
- string [resetForNewSimulations.folderName](#) = "s_inv_1e-2_"
- string [resetForNewSimulations.folderNew](#) = "s_inv_1e-1_"
- tuple [resetForNewSimulations.resdir](#) = os.path.join(os.curdir, "res")
- tuple [resetForNewSimulations.crtSimFolder](#) = os.path.join(StrTo,folderName)
- tuple [resetForNewSimulations.fileDest](#) = os.path.join(StrTo,folderNew,"_acsinflux.csv")
- tuple [resetForNewSimulations.speciesFiles](#) = sorted(glob.glob("species_1_*"))
- tuple [resetForNewSimulations.mod](#) = open("_acsspecies.csv")
- int [resetForNewSimulations.id](#) = 0
- tuple [resetForNewSimulations.linesplitted](#) = line.split("\t")
- tuple [resetForNewSimulations.file](#) = open("_acsspecies.csv", "w")

13.13 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/somma.m File Reference

Functions

- function [somma](#) (N) close all [a](#)
- [a](#) (i)
- if [r b](#) (i)
- end [c](#) (i)

Variables

- [b](#) = zeros(1,N)
- [c](#) = zeros(1,N)
- for [i](#)

13.13.1 Function Documentation

13.13.1.1 [a](#) ([i](#))

13.13.1.2 if [r b](#) ([i](#))

13.13.1.3 end [c](#) ([i](#))

13.13.1.4 function [somma](#) ([N](#))

13.13.2 Variable Documentation

13.13.2.1 `b = zeros(1,N)`

Definition at line 7 of file `somma.m`.

13.13.2.2 `c = zeros(1,N)`

Definition at line 8 of file `somma.m`.

13.13.2.3 `for i`

Initial value:

```
= 1 : N
    r = rand()
```

Definition at line 10 of file `somma.m`.

13.14 `/Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/stats.m` File Reference

Functions

- `function stats()` %clear all%close all%%`params.path`
- `concAnalysis(params)` %clear all%close all%%`params.path`

Variables

- `params deltaT = 10`
- `params totT = 1000`
- `params showFig = 0`
- `params figureVisible = 0`
- `params threshold = 0`
- `params decayTime = 10`
- `params tmpResFold = 'res'`
- `params distinctiveSubStr = 'sim'`
- `params tmpIDsim = '5'`
- `params tmpRctFileToLoad = ''`
- `params tmpRctSUBFileToLoad = ''`

13.14.1 Function Documentation

13.14.1.1 `concAnalysis(params)`

13.14.1.2 `function stats()`

13.14.2 Variable Documentation

13.14.2.1 `params decayTime = 10`

Definition at line 84 of file `stats.m`.

13.14.2.2 `params deltaT = 10`

Definition at line 7 of file stats.m.

13.14.2.3 `params distinctiveSubStr = 'sim'`

Definition at line 86 of file stats.m.

13.14.2.4 `params figureVisible = 0`

Definition at line 57 of file stats.m.

13.14.2.5 `params showFig = 0`

Definition at line 9 of file stats.m.

13.14.2.6 `params threshold = 0`

Definition at line 83 of file stats.m.

13.14.2.7 `params tmpIDsim = '5'`

Definition at line 87 of file stats.m.

13.14.2.8 `params tmpRctFileToLoad = ''`

Definition at line 88 of file stats.m.

13.14.2.9 `params tmpRctSUBFileToLoad = ''`

Definition at line 89 of file stats.m.

13.14.2.10 `params tmpResFold = 'res'`

Definition at line 85 of file stats.m.

13.14.2.11 `params totT = 1000`

Definition at line 8 of file stats.m.

13.15 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/timesAnalysis.m File Reference

Variables

- [function](#) [out]

- hence different scatterplot between different dimensions are performed Output `out` is a structure containing two variables

13.15.1 Variable Documentation

13.15.1.1 function[out]

Initial value:

```
= timesAnalysis(param)
% function [out] = timesAnalysis(tmpPath)
%
% This function collects all the last reactions contained in both the
%   times and
% reactions_parameter files
```

Definition at line 1 of file timesAnalysis.m.

13.15.1.2 hence different scatterplot between different dimensions are performed Output out is a structure containing two variables

Definition at line 1 of file timesAnalysis.m.

13.16 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/old/timesAnalysis_P-ANINI.m File Reference

Variables

- function [out]

13.16.1 Variable Documentation

13.16.1.1 function[out]

Initial value:

```
= timesAnalysis(tmpPath)
% function [out] = timesAnalysis(tmpPath)
%
% This function collects all the last reactions contained in the times
%   and
% reactions_parameter files and performs different scatterplot between different dimensions.
% Output out is a structure containing two variables:
% out.timesMatrix = all the last row of the times files
% out.cct = the correlation coefficients of the out.timesMatrix matrix
%
% INPUT
% tmpPath = path of the simulations root
currentDir = cd()
```

Definition at line 1 of file timesAnalysis_PANINI.m.

13.17 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-AttractorAnalysis.py File Reference

Namespaces

- [acsAttractorAnalysis](#)

Functions

- def [acsAttractorAnalysis.zeroBeforeStrNum](#)

Variables

- tuple [acsAttractorAnalysis.StrPath](#) = os.path.abspath(StrPath)
- tuple [acsAttractorAnalysis.today](#) = dt.date.today()
- tuple [acsAttractorAnalysis.tmpDirs](#) = sort(os.listdir(StrPath))
- list [acsAttractorAnalysis.allSortedSpecies](#) = []
- list [acsAttractorAnalysis.allConcentrations](#) = []
- list [acsAttractorAnalysis.allSortedSpeciesNOINFLUX](#) = []
- list [acsAttractorAnalysis.allConcentrationsNOINFLUX](#) = []
- tuple [acsAttractorAnalysis.totDirName](#) = os.path.join(StrPath,tmpDir)
- tuple [acsAttractorAnalysis.resDirPath](#) = os.path.abspath(os.path.join(".", "res"))
- tuple [acsAttractorAnalysis.numberOfGen](#) = len(glob.glob(os.path.join(resDirPath,'times_*')))
- tuple [acsAttractorAnalysis.strZeros](#) = zeroBeforeStrNum(nngen, numberOfGen)
- string [acsAttractorAnalysis.strSpecies](#) = 'species_'
- tuple [acsAttractorAnalysis.speciesFiles](#) = sorted(glob.glob(os.path.join(resDirPath,strSpecies)))
- list [acsAttractorAnalysis.speciesFile](#) = [speciesFiles](#)[-1]
- tuple [acsAttractorAnalysis.fidSpecies](#) = open([speciesFile](#), 'r')
- list [acsAttractorAnalysis.seq](#) = []
- list [acsAttractorAnalysis.overallConcList](#) = []
- list [acsAttractorAnalysis.overallConcListNOINFLUX](#) = []
- int [acsAttractorAnalysis.numberOfFolders](#) = 0
- tuple [acsAttractorAnalysis.pos](#) = [seq.index](#)(key)
- tuple [acsAttractorAnalysis.pos2](#) = [seq.index](#)(key2)
- tuple [acsAttractorAnalysis.ANG_overallResMatrix](#) = np.zeros((numberOfFolders,numberOfFolders))
- tuple [acsAttractorAnalysis.ANG_overallResMatrixNOINFLUX](#) = np.zeros((numberOfFolders,numberOfFolders))
- tuple [acsAttractorAnalysis.HAM_overallResMatrix](#) = np.zeros((numberOfFolders,numberOfFolders))
- tuple [acsAttractorAnalysis.HAM_overallResMatrixNOINFLUX](#) = np.zeros((numberOfFolders,numberOfFolders))
- tuple [acsAttractorAnalysis.EUC_overallResMatrix](#) = np.zeros((numberOfFolders,numberOfFolders))
- tuple [acsAttractorAnalysis.EUC_overallResMatrixNOINFLUX](#) = np.zeros((numberOfFolders,numberOfFolders))
- tuple [acsAttractorAnalysis.vecX](#) = np.array(lx)
- tuple [acsAttractorAnalysis.vecY](#) = np.array(ly)
- tuple [acsAttractorAnalysis.tmpCos](#) = float(np.dot(vecX,vecY) / (np.linalg.norm(vecX) * np.linalg.norm(vecY)))
- int [acsAttractorAnalysis.tmpPHD](#) = 0
- int [acsAttractorAnalysis.tmpEU](#) = 0
- string [acsAttractorAnalysis.ndn](#) = '_0_new_allStatResults'
- tuple [acsAttractorAnalysis.newdirAllResults](#) = os.path.join(os.getcwd(), ndn)
- string [acsAttractorAnalysis.outFnameStat](#) = 'speciesVector.csv'
- tuple [acsAttractorAnalysis.saveFileStat](#) = open(outFnameStat, 'w')
- string [acsAttractorAnalysis.strTypes](#) = "
- int [acsAttractorAnalysis.cnt](#) = 0

13.18 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-AttractorAnalysisInTime.py File Reference

Namespaces

- [acsAttractorAnalysisInTime](#)

Functions

- def [acsAttractorAnalysisInTime.zeroBeforeStrNum](#)

Variables

- tuple [acsAttractorAnalysisInTime.StrPath](#) = `os.path.abspath(StrPath)`
- tuple [acsAttractorAnalysisInTime.today](#) = `dt.date.today()`
- tuple [acsAttractorAnalysisInTime.tmpDirs](#) = `sort(os.listdir(StrPath))`
- list [acsAttractorAnalysisInTime.allSortedSpecies](#) = []
- list [acsAttractorAnalysisInTime.allConcentrations](#) = []
- list [acsAttractorAnalysisInTime.allSortedSpeciesNOINFLUX](#) = []
- list [acsAttractorAnalysisInTime.allConcentrationsNOINFLUX](#) = []
- tuple [acsAttractorAnalysisInTime.totDirName](#) = `os.path.join(StrPath,tmpDir)`
- tuple [acsAttractorAnalysisInTime.resDirPath](#) = `os.path.abspath(os.path.join(".", "res"))`
- tuple [acsAttractorAnalysisInTime.numberOfGen](#) = `len(glob.glob(os.path.join(resDirPath,'times_*')))`
- tuple [acsAttractorAnalysisInTime.strZeros](#) = `zeroBeforeStrNum(nngen, numberOfGen)`
- string [acsAttractorAnalysisInTime.strSpeciesZero](#) = `'species_'`
- tuple [acsAttractorAnalysisInTime.speciesFilesZero](#) = `sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))`
- string [acsAttractorAnalysisInTime.strSpecies](#) = `'species_'`
- tuple [acsAttractorAnalysisInTime.speciesFiles](#) = `sorted(glob.glob(os.path.join(resDirPath,strSpecies)))`
- list [acsAttractorAnalysisInTime.speciesFile](#) = `speciesFiles[timeFileID]`
- tuple [acsAttractorAnalysisInTime.fidSpecies](#) = `open(speciesFile, 'r')`
- list [acsAttractorAnalysisInTime.seq](#) = []
- list [acsAttractorAnalysisInTime.overallConcList](#) = []
- list [acsAttractorAnalysisInTime.overallConcListNOINFLUX](#) = []
- int [acsAttractorAnalysisInTime.numberOfFolders](#) = 0
- tuple [acsAttractorAnalysisInTime.pos](#) = `seq.index(key)`
- tuple [acsAttractorAnalysisInTime.pos2](#) = `seq.index(key2)`
- tuple [acsAttractorAnalysisInTime.ANG_overallResMatrix](#) = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple [acsAttractorAnalysisInTime.ANG_overallResMatrixNOINFLUX](#) = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple [acsAttractorAnalysisInTime.HAM_overallResMatrix](#) = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple [acsAttractorAnalysisInTime.HAM_overallResMatrixNOINFLUX](#) = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple [acsAttractorAnalysisInTime.EUC_overallResMatrix](#) = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple [acsAttractorAnalysisInTime.EUC_overallResMatrixNOINFLUX](#) = `np.zeros((numberOfFolders,numberOfFolders))`
- tuple [acsAttractorAnalysisInTime.vecX](#) = `np.array(lx)`
- tuple [acsAttractorAnalysisInTime.vecY](#) = `np.array(ly)`
- tuple [acsAttractorAnalysisInTime.tmpCos](#) = `float(np.dot(vecX,vecY) / (np.linalg.norm(vecX) * np.linalg.norm(vecY)))`
- int [acsAttractorAnalysisInTime.tmpHD](#) = 0
- int [acsAttractorAnalysisInTime.tmpEU](#) = 0
- string [acsAttractorAnalysisInTime.ndn](#) = `'_0_new_allStatResults'`

- tuple `acsAttractorAnalysisInTime.newdirAllResults` = `os.path.join(os.getcwd(), ndn)`
- tuple `acsAttractorAnalysisInTime.tmpZeroSaving` = `zeroBeforeStrNum(timeFileID, tmpNOF)`
- string `acsAttractorAnalysisInTime.outFnameStat` = 'speciesVector'
- tuple `acsAttractorAnalysisInTime.saveFileStat` = `open(outFnameStat, 'w')`
- string `acsAttractorAnalysisInTime.strTypes` = "
- int `acsAttractorAnalysisInTime.cnt` = 0

13.19 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-BufferedFluxes.py File Reference

Namespaces

- `acsBufferedFluxes`

Functions

- def `acsBufferedFluxes.zeroBeforeStrNum`

Variables

- tuple `acsBufferedFluxes.parser`
- tuple `acsBufferedFluxes.args` = `parser.parse_args()`
- tuple `acsBufferedFluxes.strPath` = `os.path.abspath(args.strPath)`
- tuple `acsBufferedFluxes.tmpDirs` = `sort(os.listdir(StrPath))`
- int `acsBufferedFluxes._CONDENSATION_` = 0
- int `acsBufferedFluxes._CLEAVAGE_` = 1
- int `acsBufferedFluxes._ENDOCONDENSATION_` = 7
- int `acsBufferedFluxes._ENDOCLEAVAGE_` = 6
- int `acsBufferedFluxes._SPONTCONDENSATION_` = 10
- int `acsBufferedFluxes._SPONTCLEAVAGE_` = 11
- int `acsBufferedFluxes.chemistry` = 1
- string `acsBufferedFluxes.ndn` = '_0_new_allStatResults'
- tuple `acsBufferedFluxes.newdirAllResults` = `os.path.join(StrPath, ndn)`
- tuple `acsBufferedFluxes.totDirName` = `os.path.join(StrPath, tmpDir)`
- tuple `acsBufferedFluxes.resDirPath` = `os.path.abspath(os.path.join(".", "res"))`
- tuple `acsBufferedFluxes.numberOfGen` = `len(glob.glob(os.path.join(resDirPath, 'times_*')))`
- string `acsBufferedFluxes.tmpFluxFile` = 'fluxDynamics_'
- tuple `acsBufferedFluxes.tmpSpeciesStatsSummaryNameFID` = `open(tmpFluxFile, 'w')`
- tuple `acsBufferedFluxes.strZeros` = `zeroBeforeStrNum(nGen, numberOfGen)`
- string `acsBufferedFluxes.strSpeciesZero` = 'species_'
- tuple `acsBufferedFluxes.speciesFilesZero` = `sorted(glob.glob(os.path.join(resDirPath, strSpeciesZero)))`
- string `acsBufferedFluxes.strSpecies` = 'species_'
- tuple `acsBufferedFluxes.speciesFiles` = `sorted(glob.glob(os.path.join(resDirPath, strSpecies)))`
- list `acsBufferedFluxes.lastfilespecies` = `speciesFiles[-1]`
- tuple `acsBufferedFluxes.fidSpecies` = `open(lastfilespecies, 'r')`
- list `acsBufferedFluxes.flux_seq` = []
- tuple `acsBufferedFluxes.counters` = `np.zeros((nSpecies, (len(seq)*2)+7))`
- string `acsBufferedFluxes.strRctPar` = 'reactions_parameters_'
- tuple `acsBufferedFluxes.fidRctPar` = `open(strRctPar, 'r')`
- int `acsBufferedFluxes.totIN` = 0
- int `acsBufferedFluxes.totOUT` = 0
- int `acsBufferedFluxes.deltaIO` = 0

- int `acsBufferedFluxes.totBIN` = 0
- int `acsBufferedFluxes.totBOUT` = 0
- tuple `acsBufferedFluxes.rctTime` = int(tmpRctT)
- tuple `acsBufferedFluxes.rctType` = int(tmpRctType)
- tuple `acsBufferedFluxes.cat` = int(tmpCat)
- tuple `acsBufferedFluxes.S1` = int(tmpS1)
- tuple `acsBufferedFluxes.S2` = int(tmpS2)
- tuple `acsBufferedFluxes.S3` = int(tmpS3)
- string `acsBufferedFluxes.tmpFileName` = 'speciesStats_'
- tuple `acsBufferedFluxes.tmpFileNameFID` = open(tmpFileName, 'w')
- int `acsBufferedFluxes.ID` = 0
- string `acsBufferedFluxes.tmpStr` = 'Total Number of Reactions\t\t\t\t'

13.20 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-DynStatInTime.py File Reference

Namespaces

- `acsDynStatInTime`

Variables

- tuple `acsDynStatInTime.parser`
- tuple `acsDynStatInTime.args` = parser.parse_args()
- tuple `acsDynStatInTime.strPath` = os.path.abspath(args.strPath)
- string `acsDynStatInTime.strSubStrKey` = '*'
- tuple `acsDynStatInTime.filesToAnal` = sorted(glob.glob(os.path.join(strPath,strSubStrKey)))
- tuple `acsDynStatInTime.lenFilesToAnal` = len(filesToAnal)
- int `acsDynStatInTime.numOfTraj` = 0
- tuple `acsDynStatInTime.x` = np.array(map(list, np.loadtxt(fileToAnal, str)), float)
- `acsDynStatInTime.xsize` = x.shape
- list `acsDynStatInTime.run4Chem` = xsize[0]
- tuple `acsDynStatInTime.y` = np.zeros((lenFilesToAnal,numOfTraj))
- int `acsDynStatInTime.pos` = 0
- int `acsDynStatInTime.chem` = 1
- string `acsDynStatInTime.tmpHead` = "
- string `acsDynStatInTime.filename` = "_arranged_"

13.21 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-FromWim2Carness.py File Reference

Namespaces

- `acsFromWim2Carness`

Variables

- tuple `acsFromWim2Carness.parser`
- tuple `acsFromWim2Carness.args` = `parser.parse_args()`
- string `acsFromWim2Carness.ndn` = `'_0_new_allStatResults'`
- tuple `acsFromWim2Carness.newdirAllResults` = `os.path.join(args.strOut, ndn)`
- tuple `acsFromWim2Carness.fname_initRafRes` = `os.path.join(newdirAllResults, '0_initRafAnalysis.csv')`
- tuple `acsFromWim2Carness.fname_initRafResLIST` = `os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')`
- tuple `acsFromWim2Carness.fname_initRafResALL` = `os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')`
- tuple `acsFromWim2Carness.fid_initRafRes` = `open(fname_initRafRes, 'w')`
- tuple `acsFromWim2Carness.fid_initRafResLIST` = `open(fname_initRafResLIST, 'w')`
- tuple `acsFromWim2Carness.fid_initRafResALL` = `open(fname_initRafResALL, 'w')`
- string `acsFromWim2Carness.strToWrite` = `"Folder\\tP\\tAC\\tM\\tRAFsize\\tClosure\\tCats\\tuRAF\\n"`
- tuple `acsFromWim2Carness.f` = `open(args.wimFile)`
- tuple `acsFromWim2Carness.lines` = `f.readlines()`
- int `acsFromWim2Carness.procedure` = 0
- list `acsFromWim2Carness.speciesList` = []
- list `acsFromWim2Carness.foodList` = []
- int `acsFromWim2Carness.rct` = 0
- tuple `acsFromWim2Carness.l` = `line.split()`
- tuple `acsFromWim2Carness.catNums` = `len(l)`
- tuple `acsFromWim2Carness.cats` = `np.vstack([cats,(int(rct), int(speciesList.index(l[6+catNums-1])[0:len(l[6+catNums-1])-1])), int(rct), int(0), int(5), int(25), int(5), int(1))])`
- tuple `acsFromWim2Carness.rafses` = `raf.rafComputation(fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, 'tmpDir', 0, rct/float(len(speciesList)), rcts, cats, foodList, 10)`

13.22 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-RAFanalysis.py File Reference

Namespaces

- `acsRAFanalysis`

Variables

- tuple `acsRAFanalysis.parser`
- tuple `acsRAFanalysis.args` = `parser.parse_args()`
- tuple `acsRAFanalysis.strPath` = `os.path.abspath(args.strPath)`
- int `acsRAFanalysis._CLOSE_` = 0
- int `acsRAFanalysis._PROTO_` = 1
- int `acsRAFanalysis._CSTR_` = 2
- tuple `acsRAFanalysis.conf` = `rf.readConfFile(strPath)`
- tuple `acsRAFanalysis.closure` = `dm.generateFluxList(strPath, sysType)`
- tuple `acsRAFanalysis.foodSet` = `deepcopy(closure)`
- tuple `acsRAFanalysis.rcts` = `rf.loadAllData(strPath, '_acsreactions.csv')`
- tuple `acsRAFanalysis.cats` = `rf.loadAllData(strPath, '_acscatalysis.csv')`
- tuple `acsRAFanalysis.RA` = `raf.RAcondition(strPath,closure,rcts,cats)`
- tuple `acsRAFanalysis.RAF` = `raf.Fcondition(strPath,closure,RA,rcts)`
- tuple `acsRAFanalysis.RAFIpre` = `len(RAF)`
- list `acsRAFanalysis.redRcts` = `rcts[RAF,:]`
- int `acsRAFanalysis.RAFIpost` = 0
- tuple `acsRAFanalysis.catalists` = `raf.findCatforRAF(cats, RAF, closure)`

13.23 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-SCCanalysis.py File Reference

Namespaces

- [acsSCCanalysis](#)

Functions

- def [acsSCCanalysis.zeroBeforeStrNum](#)
- def [acsSCCanalysis.loadReactionGraph](#)
- def [acsSCCanalysis.loadSpecificReactionGraph](#)
- def [acsSCCanalysis.loadSpecificReactionSubGraph](#)
- def [acsSCCanalysis.saveGraphToFile](#)
- def [acsSCCanalysis.saveGraphSUBToFile](#)
- def [acsSCCanalysis.saveNrgToFile](#)
- def [acsSCCanalysis.saveGillToFile](#)

Variables

- tuple [acsSCCanalysis.StrPath](#) = os.path.abspath(StrPath)
- tuple [acsSCCanalysis.today](#) = dt.date.today()
- tuple [acsSCCanalysis.mswindows](#) = (sys.platform == "win32")
- string [acsSCCanalysis.cmdFileName](#) = StrPath+'/'
- tuple [acsSCCanalysis.cmdFileFid](#) = open(cmdFileName, 'a')
- string [acsSCCanalysis.strToWrite](#) = "\tReaction Probability"
- int [acsSCCanalysis.initializeGraphStructure](#) = 0
- tuple [acsSCCanalysis.graph](#) = loadSpecificReactionGraph()
- tuple [acsSCCanalysis.graphSUB](#) = loadSpecificReactionSubGraph()
- tuple [acsSCCanalysis.tmpRctFileToLoadSplitString](#) = [tmpRctFileToLoad.split](#)("_")
- tuple [acsSCCanalysis.initRctId](#) = int(tmpRctFileToLoadSplitString[len(tmpRctFileToLoadSplitString)-2])
- list [acsSCCanalysis.initTempTime](#) = tmpRctFileToLoadSplitString[len(tmpRctFileToLoadSplitString)-1]
- tuple [acsSCCanalysis.initTime](#) = float(initTempTime[0:len(initTempTime)-4])
- tuple [acsSCCanalysis.tmpDirs](#) = sort(os.listdir(StrPath))
- tuple [acsSCCanalysis.totDirName](#) = os.path.join(StrPath,tmpDir)
- tuple [acsSCCanalysis.resDirPath](#) = os.path.abspath(os.path.join("./", "res"))
- tuple [acsSCCanalysis.numberOfGen](#) = len(glob.glob(os.path.join(resDirPath,'times_*')))
- tuple [acsSCCanalysis.strZeros](#) = zeroBeforeStrNum(nGen, numberOfGen)
- string [acsSCCanalysis.paramFile](#) = "acsm2s.conf"
- int [acsSCCanalysis.simFolder](#) = 0
- tuple [acsSCCanalysis.fid](#) = open(paramFile, 'r')
- tuple [acsSCCanalysis.strLine](#) = [line.split](#)('=')
- tuple [acsSCCanalysis.rp](#) = float(strLine[1])
- tuple [acsSCCanalysis.decayTime](#) = int(strLine[1])
- tuple [acsSCCanalysis.totTimes](#) = int(strLine[1])
- tuple [acsSCCanalysis.nrgType](#) = int(strLine[1])
- tuple [acsSCCanalysis.totalRcts](#) = int(strLine[1])
- tuple [acsSCCanalysis.nrgConc](#) = float(strLine[1])
- tuple [acsSCCanalysis.influx_rate](#) = float(strLine[1])
- tuple [acsSCCanalysis.maxLOut](#) = float(strLine[1])
- int [acsSCCanalysis._ANALTIMEINTERAVAL](#) = totTimes/10
- int [acsSCCanalysis._ANALTIMEINTERAVALNOSAVE](#) = totTimes/100
- int [acsSCCanalysis.nrg](#) = 1

- tuple `acsSCCanalysis.speciesInFlux` = range(0,int(pow(2,(maxLOut+1)) - 2))
- tuple `acsSCCanalysis.fidflux` = open('_acsinflux.csv', 'r')
- tuple `acsSCCanalysis.overThreshold` = float(0)
- tuple `acsSCCanalysis.overThresholdTOT` = float(0)
- string `acsSCCanalysis.tmpFilesToSearch` = 'species_'
- tuple `acsSCCanalysis.speciesFiles` = sorted(glob.glob(os.path.join(resDirPath,tmpFilesToSearch)))
- list `acsSCCanalysis.speciesFile` = `speciesFiles`[-1]
- tuple `acsSCCanalysis.fidSpecies` = open(`speciesFile`, 'r')
- int `acsSCCanalysis.ok` = 0
- list `acsSCCanalysis.IDsOverThreshold` = []
- list `acsSCCanalysis.concVec` = []
- tuple `acsSCCanalysis.index` = int(tmpID)
- tuple `acsSCCanalysis.conc` = float(tmpConc)
- tuple `acsSCCanalysis.cpxCut` = int(tmpCpxCut)
- tuple `acsSCCanalysis.age` = float(tmpAge)
- `acsSCCanalysis.realT` = `threshold`
- string `acsSCCanalysis.folderCat` = '__0_iGraph_CAT_'
- string `acsSCCanalysis.folderSub` = '__0_iGraph_SUB_'
- tuple `acsSCCanalysis.newdir` = os.path.join(os.curdir, `folderCat`)
- tuple `acsSCCanalysis.newdirSUB` = os.path.join(os.curdir, `folderSub`)
- string `acsSCCanalysis.filextPre` = '_'
- string `acsSCCanalysis.rctParamFileQ` = 'reactions_parameters_'
- tuple `acsSCCanalysis.rctParamFile` = sorted(glob.glob(os.path.join(resDirPath,rctParamFileQ)))
- float `acsSCCanalysis.rctIDshow` = 1.0
- float `acsSCCanalysis.rctIDshowNoSave` = 1.0
- `acsSCCanalysis.rctID` = `initRctId`
- int `acsSCCanalysis.previousTime` = 0
- int `acsSCCanalysis.endo_condensation_counter` = 0
- int `acsSCCanalysis.condensation_counter` = 0
- int `acsSCCanalysis.endo_cleavage_counter` = 0
- int `acsSCCanalysis.cleavage_counter` = 0
- tuple `acsSCCanalysis.nrgTimeSeries` = np.array([[0,0,0]])
- tuple `acsSCCanalysis.gillTimeSeries` = np.array([[0,0,0,0]])
- int `acsSCCanalysis.tmpFlagLastRctSaved` = 0
- tuple `acsSCCanalysis.reaction` = int(tmpReaction)
- tuple `acsSCCanalysis.rtime` = float(tmpTime)
- tuple `acsSCCanalysis.cc` = int(tmpcc)
- tuple `acsSCCanalysis.cat` = int(tmpCat)
- tuple `acsSCCanalysis.mol_I` = int(tmpMol_I)
- tuple `acsSCCanalysis.mol_II` = int(tmpMol_II)
- tuple `acsSCCanalysis.mol_III` = int(tmpMol_III)
- tuple `acsSCCanalysis.loadedMolsConc` = float(tmpLoadedMolsConc)
- tuple `acsSCCanalysis.loadedMols` = int(tmpLoadedMols)
- tuple `acsSCCanalysis.gillMean` = float(tmpGillMean)
- tuple `acsSCCanalysis.gillSD` = float(tmpGillSD)
- tuple `acsSCCanalysis.gillEntropy` = float(tmpGillEntropy)
- tuple `acsSCCanalysis.newSpeciesCreationProb` = float(tmpNSCprob)
- tuple `acsSCCanalysis.reverseProbability` = float(tmpRevProb)
- int `acsSCCanalysis.printTemporalMessage` = 1
- `acsSCCanalysis.timeInterval` = `rtime-previousTime`
- tuple `acsSCCanalysis.position` = ((`graph[:,0]` == `cat`) & (`graph[:,1]` == `mol_I`))
- int `acsSCCanalysis.realSccs` = 0
- tuple `acsSCCanalysis.Gcatpro` = nx.DiGraph()
- tuple `acsSCCanalysis.scc` = nx.strongly_connected_components(`Gcatpro`)
- tuple `acsSCCanalysis.sccN` = nx.number_strongly_connected_components(`Gcatpro`)

- tuple `acsSCCanalysis.selfLoops` = `Gcatpro.number_of_selfloops()`
- tuple `acsSCCanalysis.selfLoopsEdges` = `Gcatpro.selfloop_edges()`
- int `acsSCCanalysis.prod_inSCC` = 0
- int `acsSCCanalysis.prod_chain` = 0
- int `acsSCCanalysis.prod_bySCC` = 0
- int `acsSCCanalysis.prod_overlap` = 0
- int `acsSCCanalysis.sccID` = 0
- int `acsSCCanalysis.autocatalysis` = 0
- int `acsSCCanalysis.prod_inSCC_weight` = 0
- int `acsSCCanalysis.prod_chain_weight` = 0
- int `acsSCCanalysis.prod_bySCC_weight` = 0
- int `acsSCCanalysis.prod_overlap_weight` = 0
- int `acsSCCanalysis.self_loop_weight` = 0
- int `acsSCCanalysis.conc_inSCC` = 0
- int `acsSCCanalysis.conc_chain` = 0
- int `acsSCCanalysis.conc_bySCC` = 0
- int `acsSCCanalysis.conc_overLap` = 0
- int `acsSCCanalysis.conc_selfCat` = 0
- int `acsSCCanalysis.wasteSpecies` = 0
- int `acsSCCanalysis.alreadyAdded_ACS` = 0
- int `acsSCCanalysis.alreadyAdded_leaves` = 0
- int `acsSCCanalysis.alreadyAdded_chain` = 0
- int `acsSCCanalysis.tmpProd_chain` = 0
- tuple `acsSCCanalysis.incomingNodes` = `Gcatpro.predecessors(IdsOT)`
- int `acsSCCanalysis.tempProd_chain_weight` = 0
- int `acsSCCanalysis.noInAcs` = 1
- int `acsSCCanalysis.inSCCFlag` = 0
- list `acsSCCanalysis.weightToDistribute` = `graph[(((graph[:,0] == sngInNode) & (graph[:,1] == IdsOT)),5]`
- tuple `acsSCCanalysis.inDegreeMean` = `mean(Gcatpro.in_degree().values())`
- tuple `acsSCCanalysis.meanOverThreshold` = `float(overThreshold)`

13.24 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acs-SpeciesActivities.py File Reference

Namespaces

- `acsSpeciesActivities`

Functions

- def `acsSpeciesActivities.zeroBeforeStrNum`

Variables

- tuple `acsSpeciesActivities.StrPath` = `os.path.abspath(StrPath)`
- tuple `acsSpeciesActivities.tmpDirs` = `sort(os.listdir(StrPath))`
- int `acsSpeciesActivities.chemistry` = 1
- string `acsSpeciesActivities.ndn` = `'_0_new_allStatResults'`
- tuple `acsSpeciesActivities.newdirAllResults` = `os.path.join(StrPath, ndn)`
- tuple `acsSpeciesActivities.newdirAllResultsInner` = `os.path.join(StrPath, '_0_new_allStatResults', ndn)`
- tuple `acsSpeciesActivities.totDirName` = `os.path.join(StrPath, tmpDir)`
- tuple `acsSpeciesActivities.resDirPath` = `os.path.abspath(os.path.join(".", "res"))`

- tuple `acsSpeciesActivities.numberOfGen` = `len(glob.glob(os.path.join(resDirPath,'times_*')))`
- string `acsSpeciesActivities.tmpSpeciesStatsSummaryName` = `'speciesStatsSummary_'`
- tuple `acsSpeciesActivities.tmpSpeciesStatsSummaryNameFID` = `open(tmpSpeciesStatsSummaryName, 'w')`
- string `acsSpeciesActivities.tmpStr` = `'\n----- CHEMISTRY '`
- tuple `acsSpeciesActivities.strZeros` = `zeroBeforeStrNum(nGen, numberOfGen)`
- string `acsSpeciesActivities.strSpeciesZero` = `'species_'`
- tuple `acsSpeciesActivities.speciesFilesZero` = `sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))`
- string `acsSpeciesActivities.strSpecies` = `'species_'`
- tuple `acsSpeciesActivities.speciesFiles` = `sorted(glob.glob(os.path.join(resDirPath,strSpecies)))`
- list `acsSpeciesActivities.lastfilespecies` = `speciesFiles[-1]`
- tuple `acsSpeciesActivities.fidSpecies` = `open(lastfilespecies, 'r')`
- list `acsSpeciesActivities.seq` = `[]`
- tuple `acsSpeciesActivities.counters` = `np.zeros((nSpecies,3))`
- string `acsSpeciesActivities.strRctPar` = `'reactions_parameters_'`
- tuple `acsSpeciesActivities.fidRctPar` = `open(strRctPar, 'r')`
- tuple `acsSpeciesActivities.rctType` = `int(tmpRctType)`
- tuple `acsSpeciesActivities.cat` = `int(tmpCat)`
- tuple `acsSpeciesActivities.S1` = `int(tmpS1)`
- tuple `acsSpeciesActivities.S2` = `int(tmpS2)`
- tuple `acsSpeciesActivities.S3` = `int(tmpS3)`
- string `acsSpeciesActivities.tmpFileName` = `'speciesStats_'`
- tuple `acsSpeciesActivities.tmpFileNameFID` = `open(tmpFileName, 'w')`
- int `acsSpeciesActivities.ID` = `0`

13.25 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/acsStatesAnalysis.py File Reference

Namespaces

- `acsStatesAnalysis`

Functions

- def `acsStatesAnalysis.zeroBeforeStrNum`
- def `acsStatesAnalysis.returnZeroSpeciesList`
- def `acsStatesAnalysis.distanceMisures`

Variables

- tuple `acsStatesAnalysis.today` = `dt.date.today()`
- tuple `acsStatesAnalysis.StrPath` = `os.path.abspath(StrPath)`
- tuple `acsStatesAnalysis.tmpDirs` = `sort(os.listdir(StrPath))`
- string `acsStatesAnalysis.currentDir` = `"`
- string `acsStatesAnalysis.ndn` = `currentDir+'_0_new_allStatResults'`
- tuple `acsStatesAnalysis.newdirAllResults` = `os.path.join(os.curdir, ndn)`
- tuple `acsStatesAnalysis.previousFILE_FID` = `open('STAT_t_tminus_1.csv', 'w')`
- tuple `acsStatesAnalysis.previousNOINFLUX_FILE_FID` = `open('STAT_t_tminus_1_NOINFLUX.csv', 'w')`
- tuple `acsStatesAnalysis.startFILE_FID` = `open('STAT_t_start.csv', 'w')`
- tuple `acsStatesAnalysis.startNOINFLUX_FILE_FID` = `open('STAT_t_start_NOINFLUX.csv', 'w')`
- tuple `acsStatesAnalysis.HAM_previousFILE_FID` = `open('STAT_HAM_t_tminus_1.csv', 'w')`

```

• tuple acsStatesAnalysis.HAM_previousNOINFLUX_FILE_FID = open('STAT_HAM_t_tminus_1_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.HAM_startFILE_FID = open('STAT_HAM_t_start.csv', 'w')
• tuple acsStatesAnalysis.HAM_startNOINFLUX_FILE_FID = open('STAT_HAM_t_start_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.EUC_previousFILE_FID = open('STAT_EUC_t_tminus_1.csv', 'w')
• tuple acsStatesAnalysis.EUC_previousNOINFLUX_FILE_FID = open('STAT_EUC_t_tminus_1_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.EUC_startFILE_FID = open('STAT_EUC_t_start.csv', 'w')
• tuple acsStatesAnalysis.EUC_startNOINFLUX_FILE_FID = open('STAT_EUC_t_start_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.ANG_middlePreviousFILE_FID = open('STAT_ANG_t_middle_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.HAM_middlePreviousFILE_FID = open('STAT_HAM_t_middle_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.EUC_middlePreviousFILE_FID = open('STAT_EUC_t_middle_NOINFLUX.csv', 'w')
• tuple acsStatesAnalysis.previousFILE_FID_group = open('STAT_t_tminus_1_group.csv', 'w')
• tuple acsStatesAnalysis.previousNOINFLUX_FILE_FID_group = open('STAT_t_tminus_1_NOINFLUX_group.csv', 'w')
• tuple acsStatesAnalysis.startFILE_FID_group = open('STAT_t_start_group.csv', 'w')
• tuple acsStatesAnalysis.startNOINFLUX_FILE_FID_group = open('STAT_t_start_NOINFLUX_group.csv', 'w')
• tuple acsStatesAnalysis.HAM_previousFILE_FID_group = open('STAT_HAM_t_tminus_1_group.csv', 'w')
• tuple acsStatesAnalysis.HAM_previousNOINFLUX_FILE_FID_group = open('STAT_HAM_t_tminus_1_NOINFLUX_group.csv', 'w')
• tuple acsStatesAnalysis.HAM_startFILE_FID_group = open('STAT_HAM_t_start_group.csv', 'w')
• tuple acsStatesAnalysis.HAM_startNOINFLUX_FILE_FID_group = open('STAT_HAM_t_start_NOINFLUX_group.csv', 'w')
• tuple acsStatesAnalysis.EUC_previousFILE_FID_group = open('STAT_EUC_t_tminus_1_group.csv', 'w')
• tuple acsStatesAnalysis.EUC_previousNOINFLUX_FILE_FID_group = open('STAT_EUC_t_tminus_1_NOINFLUX_group.csv', 'w')
• tuple acsStatesAnalysis.EUC_startFILE_FID_group = open('STAT_EUC_t_start_group.csv', 'w')
• tuple acsStatesAnalysis.EUC_startNOINFLUX_FILE_FID_group = open('STAT_EUC_t_start_NOINFLUX_group.csv', 'w')
• tuple acsStatesAnalysis.newSpecies_FID = open('STAT_GENERAL_newSpecies.csv', 'w')
• tuple acsStatesAnalysis.livingSpecies_FID = open('STAT_GENERAL_livingSpecies.csv', 'w')
• tuple acsStatesAnalysis.mols_FID = open('STAT_GENERAL_mols.csv', 'w')
• tuple acsStatesAnalysis.totMass_FID = open('STAT_GENERAL_overallMass.csv', 'w')
• tuple acsStatesAnalysis.totOverallMass_FID = open('STAT_GENERAL_overallTotMass.csv', 'w')
• tuple acsStatesAnalysis.complex_FID = open('STAT_GENERAL_complex.csv', 'w')
• tuple acsStatesAnalysis.complexMols_FID = open('STAT_GENERAL_complexMols.csv', 'w')
• tuple acsStatesAnalysis.evaluatedFID = open('STAT_GENERAL_evaluated.csv', 'w')
• tuple acsStatesAnalysis.zeroOneSpeciesFID = open('STAT_GENERAL_zeroOneSpecies.csv', 'w')
• tuple acsStatesAnalysis.biodeversityFID = open('STAT_GENERAL_bioDiversity.csv', 'w')
• int acsStatesAnalysis.validDir = 1
• tuple acsStatesAnalysis.totDirName = os.path.join(StrPath,tmpDir)
• tuple acsStatesAnalysis.resDirPath = os.path.abspath(os.path.join(".", "res"))
• tuple acsStatesAnalysis.numberOfGen = len(glob.glob(os.path.join(resDirPath,'times_*')))
• list acsStatesAnalysis.group_A_prev = []
• list acsStatesAnalysis.group_A_start = []
• list acsStatesAnalysis.group_A_prev_NI = []
• list acsStatesAnalysis.group_A_start_NI = []
• tuple acsStatesAnalysis.strZeros = zeroBeforeStrNum(nngen, numberOfGen)
• string acsStatesAnalysis.strSpeciesZero = 'species_'
• tuple acsStatesAnalysis.speciesFilesZero = sorted(glob.glob(os.path.join(resDirPath,strSpeciesZero)))
• string acsStatesAnalysis.strSpecies = 'species_'
• tuple acsStatesAnalysis.speciesFiles = sorted(glob.glob(os.path.join(resDirPath,strSpecies)))
• tuple acsStatesAnalysis.zeroList = returnZeroSpeciesList(speciesFiles[-1])
• tuple acsStatesAnalysis.speciesConcs = np.zeros((len(speciesFiles)+1,len(zeroList)))

```

- list `acsStatesAnalysis.seqOLD` = []
- list `acsStatesAnalysis.seqSTART` = []
- list `acsStatesAnalysis.totMass` = []
- list `acsStatesAnalysis.seqMIDDLE_NOINFLUX` = []
- int `acsStatesAnalysis.oldNumberOfSpecies` = 0
- tuple `acsStatesAnalysis.fidSpecies` = open(sngSpeciesFile, 'r')
- list `acsStatesAnalysis.seq` = []
- int `acsStatesAnalysis.tmpMols` = 0
- int `acsStatesAnalysis.bioDivInd` = 0
- `acsStatesAnalysis.deltaNspecies` = numberOfSpecies-oldNumberOfSpecies
- tuple `acsStatesAnalysis.strtoW` = str(deltaNspecies)
- tuple `acsStatesAnalysis.tmpMisure` = distanceMisures(seq, `conc`, seqOLD, concOLD, idS)
- list `acsStatesAnalysis.seqOLDNOINFLUX` = seqNOINFLUX[:]
- list `acsStatesAnalysis.concOLD` = `conc`[:]
- string `acsStatesAnalysis.filename` = "STAT_species_Concentrations_"

13.26 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/init.py File Reference

Namespaces

- `init`

Variables

- tuple `init.parser`
- tuple `init.args` = parser.parse_args()
- string `init.ndn` = '_0_new_allStatResults'
- tuple `init.newdirAllResults` = os.path.join(args.strOut, ndn)
- tuple `init.fname_initRafRes` = os.path.join(newdirAllResults, '0_initRafAnalysis.csv')
- tuple `init.fname_initRafResSUM` = os.path.join(newdirAllResults, '0_initRafAnalysisSUM.csv')
- tuple `init.fname_initRafResLIST` = os.path.join(newdirAllResults, '0_initRafAnalysisLIST.csv')
- tuple `init.fname_initRafResALL` = os.path.join(newdirAllResults, '0_initRafAnalysisALL.csv')
- tuple `init.fid_initRafRes` = open(fname_initRafRes, 'w')
- tuple `init.fid_initRafResSUM` = open(fname_initRafResSUM, 'w')
- tuple `init.fid_initRafResLIST` = open(fname_initRafResLIST, 'w')
- tuple `init.fid_initRafResALL` = open(fname_initRafResALL, 'w')
- string `init.strToWrite` = "Folder\\P\\tAC\\tM\\tRAFsize\\tClosure\\tCats\\tuRAF\\n"
- tuple `init.foodList` = range(args.lastFood+1)
- tuple `init.avgCon` = dn.rangeFloat(float(args.avgCon[0]), float(args.avgCon[1]), float(args.avgCon[2]))
- int `init.raffound` = 0
- list `init.alphabet` = ['A', 'B']
- list `init.species` = []
- int `init.totSpecies` = 2
- tuple `init.totCleavage` = sum(map(lambda x: len(x)-1, species))
- int `init.totCond` = totSpecies**2
- `init.totRcts` = totCleavage+totCond
- tuple `init.rctToCat` = int(round(totRcts * totSpecies * prob))
- int `init.nCleavage` = 0
- int `init.nCondensa` = 0
- tuple `init.initSpeciesListLength` = len(species)
- tuple `init.conf` = (1,1,2000,0,200000,0,0,2,args.lastFood,prob)

- int `init.rctType` = 1
- tuple `init.molToCleav` = `ran.choice(species[len(alphabet):initSpeciesListLength-1])`
- tuple `init.cutPt` = `ran.randint(1,len(molToCleav)-1)`
- list `init.tmp1` = `molToCleav[0:cutPt]`
- tuple `init.tmp1id` = `species.index(tmp1)`
- `init.find1` = True
- list `init.tmp2` = `molToCleav[cutPt:len(molToCleav)]`
- tuple `init.tmp2id` = `species.index(tmp2)`
- tuple `init.sub1` = `ran.choice(species[:initSpeciesListLength-1])`
- tuple `init.idsub1` = `species.index(sub1)`
- tuple `init.sub2` = `ran.choice(species[:initSpeciesListLength-1])`
- tuple `init.idsub2` = `species.index(sub2)`
- `init.prod` = `sub1+sub2`
- tuple `init.tmpprodid` = `species.index(prod)`
- int `init.catalyst` = -1
- `init.catFound` = False
- tuple `init.rafsets` = `raf.rafComputation(fid_initRafRes, fid_initRafResALL, fid_initRafResLIST, 'tmpDir', prob, averageConn, rcts, cats, foodList, maxlength)`

13.27 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/- __init__.py File Reference

Namespaces

- `lib`

13.28 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/dyn/- __init__.py File Reference

Namespaces

- `lib.dyn`

13.29 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/graph/- __init__.py File Reference

Namespaces

- `lib.graph`

13.30 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/- IO/__init__.py File Reference

Namespaces

- `lib.IO`

13.31 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/dyn/dynamics.py File Reference

Namespaces

- [lib.dyn.dynamics](#)

Functions

- def [lib.dyn.dynamics.generateFluxList](#)
- def [lib.dyn.dynamics.rangeFloat](#)
- def [lib.dyn.dynamics.fluxAnalysis](#)

13.32 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/graph/network.py File Reference

Namespaces

- [lib.graph.network](#)

Functions

- def [lib.graph.network.removeRareRcts](#)
- def [lib.graph.network.fixCondensationReaction](#)

13.33 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/graph/raf.py File Reference

Namespaces

- [lib.graph.raf](#)

Functions

- def [lib.graph.raf.generateClosure](#)
- def [lib.graph.raf.RAcondition](#)
- def [lib.graph.raf.Fcondition](#)
- def [lib.graph.raf.findCatforRAF](#)
- def [lib.graph.raf.rafsearch](#)
- def [lib.graph.raf.rafComputation](#)
- def [lib.graph.raf.rafDynamicComputation](#)
- def [lib.graph.raf.findRAFrcts](#)

13.34 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/- IO/readfiles.py File Reference

Namespaces

- [lib.IO.readfiles](#)

Functions

- def [lib.IO.readfiles.readConfFile](#)
- def [lib.IO.readfiles.readInitConfFile](#)
- def [lib.IO.readfiles.readBufferedID](#)
- def [lib.IO.readfiles.readCSTRflux](#)
- def [lib.IO.readfiles.loadAllData](#)
- def [lib.IO.readfiles.zeroBeforeStrNum](#)
- def [lib.IO.readfiles.splitRctParsLine](#)

13.35 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/lib/-IO/writefiles.py File Reference

Namespaces

- [lib.IO.writefiles](#)

Functions

- def [lib.IO.writefiles.write_init_raf_list](#)
- def [lib.IO.writefiles.write_init_raf_all](#)

13.36 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/main.py File Reference

Namespaces

- [main](#)

Variables

- tuple [main.parser](#)
- tuple [main.args](#) = [parser.parse_args\(\)](#)
- tuple [main.strPath](#) = [os.path.abspath\(args.strPath\)](#)
- tuple [main.tmpDirs](#) = [sort\(os.listdir\(strPath\)\)](#)
- string [main.ndn](#) = ['_0_new_allStatResults'](#)
- tuple [main.newdirAllResults](#) = [os.path.join\(strPath, ndn\)](#)
- int [main._CLOSE_](#) = 0
- int [main._PROTO_](#) = 1
- int [main._CSTR_](#) = 2
- tuple [main.fname_initRafRes](#) = [os.path.join\(newdirAllResults, '0_initRafAnalysis.csv'\)](#)
- tuple [main.fname_initRafResLIST](#) = [os.path.join\(newdirAllResults, '0_initRafAnalysisLIST.csv'\)](#)
- tuple [main.fname_initRafResALL](#) = [os.path.join\(newdirAllResults, '0_initRafAnalysisALL.csv'\)](#)
- tuple [main.fid_initRafRes](#) = [open\(fname_initRafRes, 'w'\)](#)
- tuple [main.fid_initRafResLIST](#) = [open\(fname_initRafResLIST, 'w'\)](#)
- tuple [main.fid_initRafResALL](#) = [open\(fname_initRafResALL, 'w'\)](#)
- string [main.strToWrite](#) = ["Folder\\tP\\tAC\\tM\\tRAFsize\\tClosureSize\\tCatsSize\\tuRAF\\n"](#)
- tuple [main.totDirName](#) = [os.path.join\(strPath, tmpDir\)](#)
- tuple [main.resDirPath](#) = [os.path.abspath\(os.path.join\(".", args.resFolder\)\)](#)
- tuple [main.conf](#) = [readfiles.readConfFile\(totDirName\)](#)

- tuple `main.foodList` = `dm.generateFluxList(totDirName, sysType)`
- tuple `main.rcts` = `readfiles.loadAllData(totDirName, '_acsreactions.csv')`
- tuple `main.cats` = `readfiles.loadAllData(totDirName, '_acscatalysis.csv')`
- tuple `main.numberOfGen` = `len(glob.glob(os.path.join(resDirPath, 'times_*')))`
- tuple `main.strZeros` = `readfiles.zeroBeforeStrNum(nGen, numberOfGen)`
- string `main.fName` = `'RAF_structuresInTime_analysis_gen_'`
- tuple `main.fname_inTimeRafRes` = `os.path.join(newdirAllResults, fName)`
- tuple `main.fid_inTimeRafRes` = `open(fname_inTimeRafRes, 'w')`
- `main.potential` = `False`
- string `main.strRctZero` = `'reactions_'`
- string `main.strCatZero` = `'catalysis_'`
- tuple `main.rctFilesZero` = `sorted(glob.glob(os.path.join(resDirPath, strRctZero)))`
- tuple `main.catFilesZero` = `sorted(glob.glob(os.path.join(resDirPath, strCatZero)))`
- string `main.strRct` = `'reactions_'`
- string `main.strCat` = `'catalysis_'`
- tuple `main.rctFiles` = `sorted(glob.glob(os.path.join(resDirPath, strRct)))`
- tuple `main.catFiles` = `sorted(glob.glob(os.path.join(resDirPath, strCat)))`
- list `main.sngTime` = `conf[2]`
- int `main.actTime` = `0`
- list `main.procrcts` = `rcts[rcts[:,5] > 0,:]`
- list `main.proccats` = `cats[cats[:,3] > 0,:]`
- tuple `main.R` = `raf.rafDynamicComputation(fid_inTimeRafRes, actTime, procrcts[:,0:5], proccats[:,0:5], foodList, potential, rcts, cats, debug=args.debug)`
- tuple `main.lastRct` = `readfiles.loadAllData(totDirName, rctFiles[-1])`
- tuple `main.lastCat` = `readfiles.loadAllData(totDirName, catFiles[-1])`
- tuple `main.fname_dynRafRes` = `os.path.join(newdirAllResults, fName)`
- tuple `main.fid_dynRafRes` = `open(fname_dynRafRes, 'w')`
- string `main.strRctPar` = `'reactions_parameters_'`
- tuple `main.rctParamFile` = `sorted(glob.glob(os.path.join(resDirPath, strRctPar)))`
- tuple `main.fid` = `open(rctParamFile[0], 'r')`
- int `main.previousTime` = `0`
- `main.decayTime` = `args.decay`
- int `main.condensation_counter` = `0`
- int `main.endo_condensation_counter` = `0`
- int `main.cleavage_counter` = `0`
- int `main.endo_cleavage_counter` = `0`
- int `main.nAnal` = `1`
- int `main.rctCurrID` = `0`
- int `main.catCurrID` = `0`
- `main.timeInterval` = `rtime-previousTime`
- tuple `main.graph` = `network.removeRareRcts(graph, 2, 3, 4, timeInterval)`
- tuple `main.graphSUB` = `network.removeRareRcts(graphSUB, 2, 3, 4, timeInterval)`
- tuple `main.onrcts` = `network.removeRareRcts(onrcts, 5, 6, 7, timeInterval)`
- tuple `main.oncats` = `network.removeRareRcts(oncats, 3, 4, 5, timeInterval)`
- tuple `main.positionR` = `((onrcts[:,1] == cc) & (onrcts[:,2] == mol_I) & (onrcts[:,3] == mol_II))`
- tuple `main.position` = `((oncats[:,1] == cat) & (oncats[:,2] == onrcts[positionR,0]))`

13.37 /Users/alessandrofilisetti/Documents/GIT/carness/_analysis/underDevelopment/prepare-NewSim.py File Reference

Namespaces

- `prepareNewSim`

Functions

- def `prepareNewSim.zeroBeforeStrNum`

Variables

- tuple `prepareNewSim.parser`
- tuple `prepareNewSim.args` = `parser.parse_args()`
- tuple `prepareNewSim.StrFrom` = `os.path.abspath(args.StrFrom)`
- tuple `prepareNewSim.StrTo` = `os.path.abspath(args.StrTo)`
- tuple `prepareNewSim.StrFileSpeciesToGetConc` = `os.path.abspath(args.FileSpeciesToGetConc)`
- tuple `prepareNewSim.origin` = `os.getcwd()`
- int `prepareNewSim._LASTSPECIES_` = 29
- tuple `prepareNewSim._REVRCTS_` = `int(args.revRct)`
- tuple `prepareNewSim._RATIOREV_` = `int(args.k_revRct)`
- float `prepareNewSim._CLEAVAGE_` = 25.0
- float `prepareNewSim._CONDENSATION_` = 50.0
- float `prepareNewSim._COMPLEXFORM_` = 50.0
- tuple `prepareNewSim._COMPLEXDISS_` = `float(args.kDiss)`
- tuple `prepareNewSim._INITSPECIESCONC_` = `float(args.singleInitConc)`
- tuple `prepareNewSim.fileDest` = `os.path.join(StrTo, "_acsinflux.csv")`
- tuple `prepareNewSim.sourceResFolder` = `os.path.join(StrFrom, "res")`
- tuple `prepareNewSim.lastSpeciesFile` = `sorted(glob.glob('species_*'))`
- tuple `prepareNewSim.lastReactionsFile` = `sorted(glob.glob('reactions_1*'))`
- tuple `prepareNewSim.lastCatalysisFile` = `sorted(glob.glob('catalysis_*'))`
- tuple `prepareNewSim.mod` = `open("acsm2s.conf")`
- int `prepareNewSim.id` = 0
- tuple `prepareNewSim.linesplitted` = `line.split("=")`
- tuple `prepareNewSim.file` = `open("acsm2s.conf", "w")`
- list `prepareNewSim.concs` = []
- tuple `prepareNewSim.specFileLines` = `open(StrFileSpeciesToGetConc)`
- tuple `prepareNewSim.mod_rct` = `open("_acsreactions.csv")`
- int `prepareNewSim.flag` = 0
- tuple `prepareNewSim.catRct` = `linecache.getline('_acsreactions.csv', int(linesplitted[2])+1)`
- tuple `prepareNewSim.carRctSplit` = `catRct.split("\t")`

13.38 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializator/crea_catalizzatori.m File Reference

Functions

- distribuendo le catalisi a
caso viene una distribuzione
uniforme `switch (decisione_catalizzatori)%distribuzione dei catalizzatori` 1 random 2 con distribuzione case 1
`k=0`
- `end catalizzatore (kk, 1)`
- id catalizzatore `catalizzatore (kk, 2)`
- id specie `catalizzatore (kk, 3)`

Variables

- `function [catalizzatore reazione specie_non_esistenti]`
- `numero_specie = length(firing_disk(:,1))`
- `for i`
- `end tot_cond = numero_specie^2`
- `tot_reaz = tot_cond + tot_cleav`
- `controllo while tot_reaz`
- `*reactionProbability`
- `*cleavageProbability tot_cleav reactionProbability = input('introduci la nuova reactionProbability \n')`
- `cleavageProbability = input('introduci la nuova cleavageProbability \n')`
- `end catalisi_reali = round(tot_reaz*reactionProbability*numero_specie)`
- `reazione = -9999`
- `specie_non_esistenti {1,1} = -9999`
- `else h = 0`
- `indice reazione kk = 0`
- `indice catalizzatore while`
- `catalisi_reali k = k+1`
- `numero_specie_da_togliere = 0`
- `end numero_specie_da_togliere index = ceil(rand*(numero_specie-numero_specie_da_togliere))+numero_specie_da_togliere`
- `if fino_a_che_lunghezza_i_polimeri_non_catalizzano == 0 %i monomeri non catalizzano 0`

13.38.1 Function Documentation

13.38.1.1 `end catalizzatore (kk , 1)`

13.38.1.2 `id catalizzatore catalizzatore (kk , 2)`

13.38.1.3 `id specie catalizzatore (kk , 3)`

13.38.1.4 `distribuendo le catalisi a caso viene una distribuzione uniforme switch (decisione_catalizzatori) [pure virtual]`

13.38.2 Variable Documentation

13.38.2.1 `if catalisi_reali = round(tot_reaz*reactionProbability*numero_specie)`

Definition at line 21 of file `crea_catalizzatori.m`.

13.38.2.2 `cleavageProbability = input('introduci la nuova cleavageProbability \n')`

Definition at line 18 of file `crea_catalizzatori.m`.

13.38.2.3 `if fino_a_che_lunghezza_i_polimeri_non_catalizzano == 0 %i monomeri non catalizzano 0`

Definition at line 54 of file `crea_catalizzatori.m`.

13.38.2.4 `function[catalizzatore reazione specie_non_esistenti]`

Initial value:

```
=crea_catalizzatori (firing_disk, reactionProbability,
    decisione_catalizzatori,
    fino_a_che_lunghezza_i_polimeri_non_catalizzano,
    alphabet, cleavageProbability, specie_def)
%function [catalizzatore reazione specie_non_esistenti]=crea_catalizzatori (
    firing_disk, reactionProbability,
    decisione_catalizzatori,
    fino_a_che_lunghezza_i_polimeri_non_catalizzano,
    alphabet, cleavageProbability, specie_def)

tot_cleav = 0
```

Definition at line 1 of file crea_catalizzatori.m.

13.38.2.5 else h = 0

Definition at line 29 of file crea_catalizzatori.m.

13.38.2.6 for i

Initial value:

```
=1:numero_specie
    tot_cleav = tot_cleav + firing_disk(i,2)-1
```

Definition at line 7 of file crea_catalizzatori.m.

13.38.2.7 else index = ceil(rand*(numero_specie-numero_specie_da_togliere))+numero_specie_da_togliere

Definition at line 49 of file crea_catalizzatori.m.

13.38.2.8 indice catalizzatore while catalisi_reali k = k+1

Definition at line 40 of file crea_catalizzatori.m.

13.38.2.9 kk = 0

Definition at line 36 of file crea_catalizzatori.m.

13.38.2.10 numero_specie = length(firing_disk(:,1))

Definition at line 5 of file crea_catalizzatori.m.

13.38.2.11 numero_specie_da_togliere = 0

Definition at line 43 of file crea_catalizzatori.m.

13.38.2.12 controllo while tot_reaz* reactionProbability* cleavageProbability tot_cleav reactionProbability = input('introduci la nuova reactionProbability \n')

Definition at line 17 of file crea_catalizzatori.m.

13.38.2.13 reazione = -9999

Definition at line 25 of file crea_catalizzatori.m.

```
13.38.2.14 id reazione if rand< cleavageProbability%cleavage o condensazione??catalizzatore(kk,
4)=1%caso cleavage trovato=0 while trovato==0 index_reaz=ceil(rand *(numero_specie-
length(alphabet)))+length(alphabet) punto_di_taglio=ceil(rand *(firing_disk(index_reaz, 2)-1)) reazione(k,
1)=k reazione(k, 2)=1%id del cleavage:1 cleavage 0 condensazione reazione(k, 3)=firing_disk(index_reaz,
1) sp=char(specie_def(:, :)) sp1=char(sp(index_reaz, 1:punto_di_taglio)) sp2=char(sp(index_reaz,
punto_di_taglio+1:firing_disk(index_reaz, 2))) for i=1:length(specie_def(:, 1)) if strcmp(deblank(sp(i,:)), sp1)==1
reazione(k, 4)=i end if strcmp(deblank(sp(i,:)), sp2)==1 reazione(k, 5)=i end end for i=length(reazione(:,
1)):-1:1 if((sum(reazione(k, 2:5))==reazione(i, 2:5)))==4 &&i~=k reazione(k,:)=[] k=k-1 catalizzatore(kk,
3)=i trovato=1 break else%CONTROLLARE IL CONTROLLO!!!!DOPO IL||%TOGLIERE IL CONTROLLO
DA QUI E DA SOTTO!!!!if(sum(reazione(k, 3:5))==reazione(i, 3:5))==3 &&reazione(k, 2)~=reazione(i,
2))||((reazione(k, 3)==reazione(i, 3)&&reazione(k, 4)==reazione(i, 5)&&reazione(k, 5)==reazione(i, 4)))
trovato=0 break else trovato=1 end end end end else%caso condensazione catalizzatore(kk,
4)=0 trovato=0 while trovato==0 reazione(k, 1)=k reazione(k, 2)=0%id della condensazione:1 cleavage 0
condensazione index_specie_1=ceil(rand *(numero_specie)) index_specie_2=ceil(rand *(numero_specie))
nuova_specie=[deblank(specie_def(index_specie_1,:)), deblank(specie_def(index_specie_2,:))]
nuova_specie=char(nuova_specie) sp=char(specie_def(:, :)) found=0%controllo per vedere se trovo una
specie che esiste gir i=1:length(specie_def(:, 1)) if strcmp(deblank(sp(i,:)), deblank(nuova_specie))==1
reazione(k, 3)=i found=1 end if strcmp(deblank(sp(i,:)), deblank(sp(index_specie_1,:)))==1 reazione(k,
4)=i end if strcmp(deblank(sp(i,:)), deblank(sp(index_specie_2,:)))==1 reazione(k, 5)=i end end if
found==0%se ho trovato una specie che NON esiste h=h+1 reazione(k, 3)=length(specie_def(:,
1))+h specie_non_esistenti(h)=[nuova_specie]for iii=h:-1:1 if strcmp(specie_non_esistenti(h),
specie_non_esistenti(iii))==1 &&h~=iii specie_non_esistenti(h)=[] h=h-1 reazione(k, 3)=length(specie_def(:,
1))+iii end end end%controllo che la reazione non esista gir i=length(reazione(:, 1)):-1:1 if(sum(reazione(k,
2:5))==reazione(i, 2:5)))==4 &&i~=k reazione(k,:)=[] k=k-1 catalizzatore(kk, 3)=i trovato=1 break
else if(sum(reazione(k, 3:5))==reazione(i, 3:5))==3 &&reazione(k, 2)~=reazione(i, 2))||((reazione(k,
3)==reazione(i, 3)&&reazione(k, 4)==reazione(i, 5)&&reazione(k, 5)==reazione(i, 4))) trovato=0 break
else trovato=1 end end end end end%controllo per evitare che lo stesso catalizzatore catalizzi
la%stessa reazione trovato_uguale=1 for iii=length(catalizzatore(:, 1)):-1:1 if sum(catalizzatore(kk,
2:3))==catalizzatore(iii, 2:3))==2 &&kk~=iii catalizzatore(kk,:)=[] kk=kk-1 trovato_uguale=0 break end
end if trovato_uguale==1 catalisi_reali=catalisi_reali-1 end end if exist('specie_non_esistenti')>
specie_non_esistenti {1,1} =9999
```

Definition at line 26 of file crea_catalizzatori.m.

```
13.38.2.15 end tot_cond = numero_specie^2
```

Definition at line 10 of file crea_catalizzatori.m.

```
13.38.2.16 tot_reaz=tot_cond+tot_cleav
```

Definition at line 12 of file crea_catalizzatori.m.

13.39 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializator/crea_concentrazioni_iniziali.m File Reference

Functions

- [firing_disk](#) (:, 3)
- [if firing_disk](#) (index, 3)
- [else concentrazioni_iniziali](#) (i)=0
- [firing_disk_reale](#) (k,:)=firing_disk(i,:)
- [id](#) Z ()
- [valore](#) (1:max(firing_disk_bck(:, 2)))=0
- [valore](#) (i)

- `end end end` STAMPA
DISTRIBUZIONI DELLE
CONCENTRAZIONI `figure` (123)%plot(valore

Variables

- `function` [concentrazioni_iniziali] = crea_concentrazioni_iniziali(alphabet,firing_disk,initialMaxLength,lunghezza_max_fd,ratio_firing_disk, scelta_concentrazioni, overallConcentration, gamma)
- le specie fittizie create fino alla lunghezza_massima_per_calcolare_le_reazioni vengono poste uguali a in `concentrazione tot_species = 0`
- `for i`
- `end` `lunghezza_totale` = length(firing_disk(:,1))
- `firing_disk_bck` =firing_disk
- `firing_disk_2` =firing_disk(1:tot_species,:)
- `firing_disk` =[]
- check di esistenza nel firing `disk` `numero_molecole`
- `a` seconda dei casi seleziono le specie esistenti del firing `disk`
- `a` seconda dei casi seleziono le specie esistenti del firing quelle non esistenti andranno semplicemente `a concentrazione remaining_species = tot_species`
- `l_max` =0
- `species_to_delete` = round(ratio_firing_disk*remaining_species)
- la percentuale `reazioni` fra quelle che restano `species_to_keep` = remaining_species - species_to_delete + l_max
- `while trovato`
- `end end end switch`
`scelta_concentrazioni` case
distribuzione uniforme su tutte le specie esistenti del firing `disk` `probabilita_uniforme` = 1/species_to_keep
- `end end` case uniforme sulle `lunghezza k` =0
- `end end` `vettore_ordinato_lunghezze` = unique(firing_disk_reale(:,2))
- `probabilita_per_lunghezza` = 1/length(vettore_ordinato_lunghezze)
- `end end` normalizzazione per avere la `concentrazione concentrazioni_iniziali` =concentrazioni_iniziali/sum(concentrazioni_iniziali)*overall-Concentration

13.39.1 Function Documentation

13.39.1.1 `else` `concentrazioni_iniziali` (`i`) [pure virtual]

13.39.1.2 `end end end` STAMPA DISTRIBUZIONI DELLE CONCENTRAZIONI `figure` (123)

13.39.1.3 `firing_disk` (: , 3)

13.39.1.4 `if` `firing_disk` (`index` , 3)

13.39.1.5 `firing_disk_reale` (`k` , :) =firing_disk(i,:)

13.39.1.6 `valore (1: maxfiring_disk_bck(:, 2)) [pure virtual]`

13.39.1.7 `valore (i)`

13.39.1.8 `id Z () [virtual]`

13.39.2 Variable Documentation

13.39.2.1 `sum(concentrazioni_iniziali) end concentrazioni_iniziali =concentrazioni_iniziali/sum(concentrazioni_iniziali)*overallConcentration`

Definition at line 129 of file `crea_concentrazioni_iniziali.m`.

13.39.2.2 `a seconda dei casi seleziono le specie esistenti del firing disk`

Definition at line 25 of file `crea_concentrazioni_iniziali.m`.

13.39.2.3 `firing_disk =[]`

Definition at line 17 of file `crea_concentrazioni_iniziali.m`.

13.39.2.4 `firing_disk_2 =firing_disk(1:tot_species,:)`

Definition at line 16 of file `crea_concentrazioni_iniziali.m`.

13.39.2.5 `firing_disk_bck =firing_disk`

Definition at line 14 of file `crea_concentrazioni_iniziali.m`.

13.39.2.6 `function[concentrazioni_iniziali] = crea_concentrazioni_iniziali(alphabet,firing_disk,initialMaxLength,lunghezza_max_fd,radio_firing_disk, scelta_concentrazioni, overallConcentration, gamma)`

Definition at line 1 of file `crea_concentrazioni_iniziali.m`.

13.39.2.7 `for i`

Initial value:

```
= 1:initialMaxLength
    tot_species = tot_species+length(alphabet)^i
```

Definition at line 9 of file `crea_concentrazioni_iniziali.m`.

13.39.2.8 `end end case favorire quelle corte con una scale free di esponente gamma k =0`

Definition at line 73 of file `crea_concentrazioni_iniziali.m`.

13.39.2.9 `I_max =0`

Definition at line 31 of file `crea_concentrazioni_iniziali.m`.

13.39.2.10 **end** lunghezza_totale = length(firing_disk(:,1))

Definition at line 13 of file crea_concentrazioni_iniziali.m.

13.39.2.11 **check** di esistenza nel firing disk numero_molecole

Initial value:

```
= overallConcentration*volume
numero_specie = length(firing_disk(:,1))
```

Definition at line 22 of file crea_concentrazioni_iniziali.m.

13.39.2.12 **probabilita_per_lunghezza** = 1/length(vettore_ordinato_lunghezze)

Definition at line 82 of file crea_concentrazioni_iniziali.m.

13.39.2.13 **end end end switch scelta_concentrazioni** case distribuzione uniforme su tutte le specie esistenti del firing disk **probabilita_uniforme** = 1/species_to_keep

Definition at line 60 of file crea_concentrazioni_iniziali.m.

13.39.2.14 **end** remaining_species = tot_species

Definition at line 30 of file crea_concentrazioni_iniziali.m.

13.39.2.15 **end end** species_to_delete = round(ratio_firing_disk*remaining_species)

Definition at line 38 of file crea_concentrazioni_iniziali.m.

13.39.2.16 **la percentuale reazioni** fra quelle che restano **species_to_keep** = remaining_species - species_to_delete + l_max

Definition at line 39 of file crea_concentrazioni_iniziali.m.

13.39.2.17 **le specie fittizie create fino alla lunghezza_massima_per_calcolare_le_reazioni** vengono poste uguali a **a concentrazione tot_species** = 0

Definition at line 8 of file crea_concentrazioni_iniziali.m.

13.39.2.18 **while** trovato

Initial value:

```
==0
index = l_max+ceil(rand*remaining_species)
```

Definition at line 44 of file crea_concentrazioni_iniziali.m.

13.39.2.19 **end end** vettore_ordinato_lunghezze = unique(firing_disk_reale(:,2))

Definition at line 81 of file crea_concentrazioni_iniziali.m.

13.40 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializator/crea_e_controlla_i_catalizzatori.m File Reference

Functions

- `1:length(influx(:, 1 matrice_adiacenza_sub_prod: (influx(:, 1 length)`
- `matrice_adiacenza_sub_prod (1:max(influx(:, 1)), 1:max(influx(:, 1)))=0`
- `matrice_adiacenza_sub_prod (reazione(i, 5), reazione(i, 3))`
- `else matrice_adiacenza_sub_prod (reazione(i, 3), reazione(i, 4))`
- `matrice_adiacenza_sub_prod (reazione(i, 3), reazione(i, 5))`
- `end end end if max (real(eig(matrice_adiacenza_sub_prod))) > 0 clear catalizzatore clear reazione clear specie_non_esistenti clear matrice_adiacenza_sub_prod check_ACS=0`
- `id check_ACS ()`
- `matrice_adiacenza_cat_prod (1:max(influx(:, 1)), 1:max(influx(:, 1)))=0`
- `else matrice_adiacenza_cat_prod (catalizzatore(i, 2), reazione(catalizzatore(i, 3), 4))`
- `matrice_adiacenza_cat_prod (catalizzatore(i, 2), reazione(catalizzatore(i, 3), 5))`
- `end end end if max (real(eig(matrice_adiacenza_cat_prod))) > 0 clear catalizzatore clear reazione clear specie_non_esistenti clear matrice_adiacenza_cat_prod check_ACS=0 = 1`
- `end end end size (matrice_adiacenza_sub_prod)%size(matrice_adiacenza_cat_prod) if max(real(eig(matrice_adiacenza_sub_prod))) > 0||max(real(eig(matrice_adiacenza_cat_prod))) > 0 clear catalizzatore clear reazione clear specie_non_esistenti clear matrice_adiacenza_cat_prod clear matrice_adiacenza_sub_prod check_ACS=0`

Variables

- `function [catalizzatore reazione specie_non_esistenti matrice_adiacenza_sub_prod matrice_adiacenza_cat_prod] = crea_e_controlla_i_catalizzatori (controllo_ACS_nel_ciclo, firing_disk, reactionProbability, decisione_catalizzatori, fino_a_che_lunghezza_i_polimeri_non_catalizzano, alphabet, cleavageProbability, specie_def, influx)`
- `controllo no ACS nell influx`
- `switch controllo_ACS_nel_ciclo`
- `case nessun controllo matrice_adiacenza_sub_prod =0`
- `matrice_adiacenza_cat_prod = 0`
- `check_ACS = 0`
- `for i`
- `inpudda = input("")`
- `counter_cicli = 0`

13.40.1 Function Documentation

13.40.1.1 `id check_ACS () [virtual]`

13.40.1.2 `matrice_adiacenza_cat_prod (1: maxinflux(:, 1), 1: maxinflux(:, 1)) [pure virtual]`

13.40.1.3 `else matrice_adiacenza_cat_prod (catalizzatore(i, 2) , reazione(catalizzatore(i, 3), 4))`

13.40.1.4 `matrice_adiacenza_cat_prod (catalizzatore(i, 2) , reazione(catalizzatore(i, 3), 5))`

13.40.1.5 `matrice_adiacenza_sub_prod (1: maxinflux(:, 1), 1: maxinflux(:, 1)) [pure virtual]`

13.40.1.6 `matrice_adiacenza_sub_prod (reazione(i, 5) , reazione(i, 3))`

13.40.1.7 `else matrice_adiacenza_sub_prod (reazione(i, 3) , reazione(i, 4))`

13.40.1.8 `matrice_adiacenza_sub_prod (reazione(i, 3) , reazione(i, 5))`

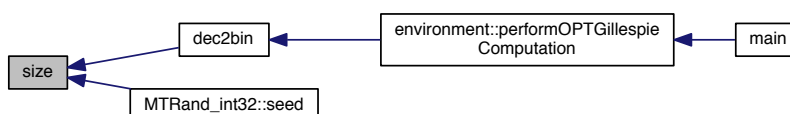
13.40.1.9 `1:length(influx(:,1 matrice_adiacenza_sub_prod: () [virtual]`

13.40.1.10 `end end end if max (real(eig(matrice_adiacenza_sub_prod))) [pure virtual]`

13.40.1.11 `end end end if max (real(eig(matrice_adiacenza_cat_prod))) = 1 [pure virtual]`

13.40.1.12 `end end end size (matrice_adiacenza_sub_prod) [pure virtual]`

Here is the caller graph for this function:



13.40.2 Variable Documentation

13.40.2.1 `while check_ACS = 0`

Definition at line 18 of file `crea_e_controlla_i_catalizzatori.m`.

13.40.2.2 `counter_cicli = 0`

Definition at line 82 of file `crea_e_controlla_i_catalizzatori.m`.

13.40.2.3 `function[catalizzatore reazione specie_non_esistenti] = crea_e_controlla_i_catalizzatori
(controllo_ACS_nel_ciclo, firing_disk, reactionProbability, decisione_catalizzatori,
fino_a_che_lunghezza_i_polimeri_non_catalizzano, alphabet, cleavageProbability, specie_def,
influx)`

Definition at line 1 of file `crea_e_controlla_i_catalizzatori.m`.

13.40.2.4 `for i`

Initial value:

```

= 1:length(reazione(:,1))
    if sum(reazione(i,3)==influx(:,1))==1 && sum(
        reazione(i,4)==influx(:,1))==1 && sum(reazione(i,5)==influx(:,1))==1
        if reazione(i,2)==0
            matrice_adiacenza_sub_prod(
                reazione(i,4),reazione(i,3))=1
  
```

Definition at line 26 of file `crea_e_controlla_i_catalizzatori.m`.

13.40.2.5 `inpudda = input('')`

Definition at line 44 of file `crea_e_controlla_i_catalizzatori.m`.

13.40.2.6 `matrice_adiacenza_cat_prod = 0`

Definition at line 11 of file `crea_e_controlla_i_catalizzatori.m`.

13.40.2.7 `matrice_adiacenza_sub_prod = 0`

Definition at line 10 of file `crea_e_controlla_i_catalizzatori.m`.

13.41 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_firing_disk.m File Reference

Functions

- [step](#) (i)

Variables

- [function](#) [`firing_disk`]
- `for i`
- `end k = 1`
- `end firing_disk = id_species`

13.41.1 Function Documentation

13.41.1.1 `step (i)`

13.41.2 Variable Documentation

13.41.2.1 `end firing_disk = id_species`

Definition at line 24 of file `crea_firing_disk.m`.

13.41.2.2 `function[firing_disk]`

Initial value:

```
= crea_firing_disk(alphabet,  
    massima_lunghezza_su_cui_calcolare_le_reazioni)  
%function [firing_disk] = crea_firing_disk(alphabet,  
    massima_lunghezza_su_cui_calcolare_le_reazioni)  
  
tot_species = 0
```

Definition at line 3 of file `crea_firing_disk.m`.

13.41.2.3 `for i`

Initial value:

```
= 1:massima_lunghezza_su_cui_calcolare_le_reazioni  
    tot_species = tot_species+length(alphabet)^i
```

Definition at line 8 of file `crea_firing_disk.m`.

13.41.2.4 $k = 1$

Definition at line 18 of file crea_firing_disk.m.

13.42 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_influx.m File Reference

Functions

- `influx(i, 2)`
- `influx(i,:)`
- `if sum(influx(i, 1)==influx(:, 1))`
- `influx(1, 1:2)=0`
- `if influx(1, 1)>0%%%for i`

Variables

- `function [influx]`
- `end for j`
- `end end tot_species = length(firing_disk(:,1))`
- `switch scelta_influx casuale case species_to_delete = round(ratio_influx*tot_species)`
- `species_to_keep = tot_species - species_to_delete`
- `for i`
- `while trovato == 0`
- `end end end influx =sort(influx)`
- `casuale tenendo i polimeri`
- `fino a lunghezza x e poi ne`
- `toglie l x case remaining_species = tot_species`
- `end proporzionale alla lunghezza`

13.42.1 Function Documentation

13.42.1.1 `influx(i , 2)`

13.42.1.2 `influx(i , :)`

13.42.1.3 `influx(1 , 1:2)` [pure virtual]

13.42.1.4 `if influx(1 , 1)`

13.42.1.5 `if sum(influx(i,1) ==influx(: , 1))`

Initial value:

```
==1
%          trovato = 1
```

13.42.2 Variable Documentation

13.42.2.1 `function[influx]`

Initial value:

```

= crea_influx(concentrazioni_iniziali)
%function [influx] = crea_influx(concentrazioni_iniziali)

for i =1:length(concentrazioni_iniziali)
    influx(i,1)=i

```

Definition at line 3 of file crea_influx.m.

13.42.2.2 for i

Initial value:

```

= 1:species_to_keep
%          trovato = 0

```

Definition at line 56 of file crea_influx.m.

13.42.2.3 else clear influx influx =sort(influx)

Definition at line 65 of file crea_influx.m.

13.42.2.4 end for j

Initial value:

```

= length(concentrazioni_iniziali):-1:1
    if influx (j,2)==0
        influx(j,:)=[]
    end

```

Definition at line 13 of file crea_influx.m.

13.42.2.5 end proporzionale alla lunghezza

Definition at line 108 of file crea_influx.m.

13.42.2.6 remaining_species = tot_species

Definition at line 71 of file crea_influx.m.

13.42.2.7 end end species_to_delete = round(ratio_influx*tot_species)

Definition at line 53 of file crea_influx.m.

13.42.2.8 species_to_keep = tot_species - species_to_delete

Definition at line 54 of file crea_influx.m.

13.42.2.9 end end tot_species = length(firing_disk(:,1))

Definition at line 44 of file crea_influx.m.

13.42.2.10 while trovato == 0

Definition at line 58 of file crea_influx.m.

13.43 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/crea_influx- _semplice.m File Reference

Functions

- [influx](#) (j, 2)

Variables

- [function](#) [[influx](#)]
- for i
- [end influx](#) = zeros(sn,2)
- for j

13.43.1 Function Documentation

13.43.1.1 [influx](#) (j , 2)

13.43.2 Variable Documentation

13.43.2.1 [function](#)[[influx](#)]

Initial value:

```
= crea_influx_semplice(lMaxInflux,alphabet)
% total Number of species
sn = 0
```

Definition at line 3 of file crea_influx_semplice.m.

13.43.2.2 for i

Initial value:

```
= 1:lMaxInflux
sn = sn + length(alphabet)^i
```

Definition at line 7 of file crea_influx_semplice.m.

13.43.2.3 [end influx](#) = zeros(sn,2)

Definition at line 11 of file crea_influx_semplice.m.

13.43.2.4 for j

Initial value:

```
= 1:sn
influx(j,1)=j
```

Definition at line 12 of file crea_influx_semplice.m.

13.44 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializator/crea_tutte_le_combinazioni_di_elementi.m File Reference

Variables

- prova m `function` [specie]
- `numero_elementi` = length(vettore_elementi)
- `righe` = `numero_elementi`^`lunghezza_stringa`
- for i

13.44.1 Variable Documentation

13.44.1.1 prova m function[specie]

Initial value:

```
=crea_tutte_le_combinazioni_di_elementi(vettore_elementi,K)
%function [specie]=crea_tutte_le_combinazioni_di_elementi(vettore_elementi)

%dato un vettore di elementi e una lunghezza massima K restituisce tutte le
%possibili combinazioni di elementi ordinati

lunghezza_stringa = K
```

Definition at line 4 of file crea_tutte_le_combinazioni_di_elementi.m.

13.44.1.2 for i

Initial value:

```
=1:lunghezza_stringa %colonne
for j = 1:numero_elementi
    for z = (numero_elementi^(i-1))*(j-1)+1:numero_elementi^(
        i):righe
        specie(z+z*numero_elementi^(i-1)-1,i)=(vettore_elementi(
            j))
```

Definition at line 14 of file crea_tutte_le_combinazioni_di_elementi.m.

13.44.1.3 numero_elementi = length(vettore_elementi)

Definition at line 11 of file crea_tutte_le_combinazioni_di_elementi.m.

13.44.1.4 righe = numero_elementi^lunghezza_stringa

Definition at line 12 of file crea_tutte_le_combinazioni_di_elementi.m.

13.45 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/initial_distribution.m File Reference

13.46 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/inizializzatore_ACS.m File Reference

Functions

- `mkdir` (nome_cartella)
- `mkdir` ('res')
- `cd` (thisFolder)
- `fid1`, `nGEN` = `fprintf` ()
- perch sono input da file (i prossimi 4) `count`
- id `i`: (`fid10`, '%d\t', `funzioni_booleane_in_dec`(`i`, 1) `fprintf`)
- `alphabet`,
 `massima_lunghezza_su_cui_calcolare_le_reazioni` `crea_firing_disk` ()
- `alphabet`, `firing_disk`,
 `initialMaxLength`,
 `lunghezza_max_fd`,
 `ratio_firing_disk`,
 `scelta_concentrazioni`,
 `overallConcentration`,
 `gamma_powerlaw_concentrazioni` `crea_concentrazioni_iniziali` ()
- `concentrazioni_iniziali` `crea_influx` ()
- id `k` ()
- `specie_def_2` (`k`,:)
- `controllo_ACS_nel_ciclo`,
 `firing_disk`,
 `reactionProbability`,
 `decisione_catalizzatori`,
 `fino_a_che_lunghezza_i_polimeri_non_catalizzano`,
 `alphabet`, `cleavageProbability`,
 `specie_def`, `influx` `crea_e_controlla_i_catalizzatori` ()
- id `kk` ()
- Punto di tagli del `complesso` (1--L-1) `if` `concentrazioni_iniziali`(`i`) > 0||`sum`(`i`)
- `end else if` `reazione` (`i`, 2)
- id `specie_non_esistenti` ()
- Coefficiente di `degradazione` (per complessi) `count`

Variables

- `function` [`firing_disk` `concentrazioni_iniziali` `specie_def` `influx` `catalizzatore` `reazione` `specie_non_esistenti` `matrice_adiacenza_sub_prod` `matrice_adiacenza_cat_prod`]
- `fid1` = `fopen`('acsm2s.conf','w')
- `fid2` = `fopen`('_acsspecies.csv','a')
- `fid3` = `fopen`('_acsreactions.csv','a')
- `fid4` = `fopen`('_acscatalysis.csv','a')
- `fid5` = `fopen`('_acsinflux.csv','a')
- `fid10` = `fopen`('_acsnrbooleanfunctions.csv','a')
- `count` = `fprintf`(`fid1`, '%d\n', `nGEN`)
- `st` = `fclose`(`fid1`)
- `vettore_rand` = [0.5 1 2]
- counter id specie for `i`

```

• for j
• for z
• end end clear specie_temp end k = 0
• end specie_def =specie_def_2
• lunghezza_stringa = length(specie_def(1,:))
• for jjjj
• for xxxx
• end end if controllo
• end end if gg
• end end tmpStr = specie_def(i,:)
• Coefficiente di degradazione del complesso = fprintf(fid2,'%d\t',0)
• concentrazione molecole
  cariche if rand
    < ratioSpeciesEnergizable%count=fprintf(fid2,'%d\n',
    1);%specie energizzabile%else%count=fprintf(fid2,'%d\n',
    0);%specie NON energizzabile%end
  if influx_rate==0 if maxLOut >
    if i<=(2^(maxLOut+1)-2) count=fprintf(fid2,'%d\n',
    1);else count=fprintf(fid2,'%d\n',
    0);end else count=fprintf(fid2,'%d\n',
    0);end else count=fprintf(fid2,'%d\n',
    0);endend%-----%inizializzazione
  del file dell'influx" _influx.csv"%-----for
  i=1:length(influx(:, 1)) count=fprintf(fid5,'%d\t',
  influx(i, 1)-1);count=fprintf(fid5,'%d\n',
  influx(i, 2));endst=fclose(fid5);%-----%inizializzazione
  del file delle reazioni e dei
  catalizzatori%-----if
  catalizzatore(1,
  1)==-9999;else for i=1:length(catalizzatore(:,
  1)) count=fprintf(fid4,'%d\t',
  catalizzatore(i, 1)-1);count=fprintf(fid4,'%d\t',
  catalizzatore(i, 2)-1);count=fprintf(fid4,'%d\t',
  catalizzatore(i, 3)-1);count=fprintf(fid4,'%d\t',
  0);%quante volte if
  catalizzatore(i,
  4)==0 tmpKdiss=Kdiss/revRctRatio;count=fprintf(fid4,'%g\t',
  Kass);%kass count=fprintf(fid4,'%g\t',
  tmpKdiss);%kdiss count=fprintf(fid4,'%g\t',
  Kcpx);%k complex else tmpKass=Kass/revRctRatio;tmpKcpx=Kcpx/revRctRatio;count=fprintf(fid4,'%g\t',
  tmpKass);%kass count=fprintf(fid4,'%g\t',
  Kdiss);%kdiss count=fprintf(fid4,'%g\t',
  tmpKcpx);%k complex end
  index_rand=ceil(rand *2);count=fprintf(fid4,'%g\n',
  index_rand);%nuovo parametro
  end[righe_xx colonne_xx]=size(funzioni_booleane_in_dec);for
  i=1:length(reazione(:,
  1)) indexx=ceil(rand *righe_xx);funzione_giusta=funzioni_booleane_in_dec(indexx);count=fprintf(fid3,'%d\t',
  reazione(i, 1)-1);count=fprintf(fid3,'%d\t',
  reazione(i, 2));count=fprintf(fid3,'%d\t',
  reazione(i, 3)-1);count=fprintf(fid3,'%d\t',
  reazione(i, 4)-1);count=fprintf(fid3,'%d\t',
  reazione(i, 5)-1);count=fprintf(fid3,'%d\t',
  0);if energy==2 eso_endo=1;%perch?se
  non c'?l'energia sono tutte
  eso else if rand > energy if reazione (i, 2)
• else eso_endo =0
• temporal = specie_non_esistenti{i}

```

13.46.1 Function Documentation

13.46.1.1 `cd (thisFolder)`

13.46.1.2 `Punto di tagli del complesso (1--L- 1)`

13.46.1.3 `alphabet,firing_disk,initialMaxLength,lunghezza_max_fd,ratio_firing_disk,scelta_concentrazioni,overallConcentration,gamma_powerlaw_concentrazioni crea_concentrazioni_iniziali ()` [virtual]

13.46.1.4 `controllo_ACS_nel_ciclo, firing_disk, reactionProbability, decisione_catalizzatori, fino_a_che_lunghezza_i_polimeri_non_catalizzano, alphabet, cleavageProbability, specie_def, influx crea_e_controlla_i_catalizzatori ()` [virtual]

13.46.1.5 `alphabet,massima_lunghezza_su_cui_calcolare_le_reazioni crea_firing_disk ()` [virtual]

13.46.1.6 `concentrazioni_iniziali crea_influx ()` [virtual]

13.46.1.7 `Coefficiente di degradazione (per complessi)`

13.46.1.8 `perch sono input da file (i prossimi 4)`

13.46.1.9 `fid2 d t fprintf ()` [virtual]

13.46.1.10 `id i: (fid10, '%d\t')` [virtual]

13.46.1.11 `id k ()` [virtual]

13.46.1.12 `id kk ()` [virtual]

13.46.1.13 `mkdir (nome_cartella)`

13.46.1.14 `mkdir ('res')`

13.46.1.15 `end else if reazione (i, 2)`

13.46.1.16 `specie_def_2 (k, :)`

13.46.1.17 `id specie_non_esistenti ()` [virtual]

Definition at line 366 of file `inizializzatore_ACS.m`.

13.46.2 Variable Documentation

13.46.2.1 `Punto di tagli del complesso = fprintf(fid2,'%d\t',0)`

Definition at line 252 of file `inizializzatore_ACS.m`.

13.46.2.2 `end end if controllo`

Initial value:

```
== 1
count = fprintf(fid2,'%c',specie_def(i,
jjjj))
```

Definition at line 223 of file `inizializzatore_ACS.m`.

13.46.2.3 `k_fosforilazione velocit con cui l atp count = fprintf(fid1,'%d\n',nGEN)`

Definition at line 35 of file `inizializzatore_ACS.m`.

13.46.2.4 `else eso_endo =0`

Definition at line 348 of file `inizializzatore_ACS.m`.

13.46.2.5 `fid1 = fopen('acsm2s.conf','w')`

Definition at line 19 of file `inizializzatore_ACS.m`.

13.46.2.6 `fid10 = fopen('_acsnrgbooleanfunctions.csv','a')`

Definition at line 24 of file `inizializzatore_ACS.m`.

13.46.2.7 `fid2 = fopen('_acsspecies.csv','a')`

Definition at line 20 of file `inizializzatore_ACS.m`.

13.46.2.8 `fid3 = fopen('_acsreactions.csv','a')`

Definition at line 21 of file `inizializzatore_ACS.m`.

13.46.2.9 `fid4 = fopen('_acscatalysis.csv','a')`

Definition at line 22 of file `inizializzatore_ACS.m`.

13.46.2.10 `fid5 = fopen('_acsinflux.csv','a')`

Definition at line 23 of file `inizializzatore_ACS.m`.

13.46.2.11 `function[firing_disk concentrazioni_iniziali specie_def influx catalizzatore reazione
specie_non_esistenti matrice_adiacenza_sub_prod matrice_adiacenza_cat_prod]`

Initial value:

```
= inizializzatore_ACS(nGEN, nSIM, nSeconds,
    nReactions, initialMaxLength,
    massima_lunghezza_su_cui_calcolare_le_reazioni,
    overallConcentration, alphabet,
    complexFormationSymmetry,
    fino_a_che_lunghezza_i_polimeri_non_catalizzano,
    reactionProbability, cleavageProbability,
    diffusion_contribute, solubility_threshold,
    influx_rate, reverseReactions,K_nrg,
    moleculeDecay_KineticConstant, ratio_firing_disk,
    lunghezza_max_fd, scelta_concentrazioni,
    gamma_powerlaw_concentrazioni,
    decisione_catalizzatori, lastFiringDiskSpeciesID,
    ECConcentration, volume, energy, controllo_ACS_nel_ciclo, K_nrg_decay, nome_cartella,
    funzioni_booleane_in_dec, ratioSpeciesEnergizable,
    Kass,Kdiss,Kcpx,K_cpx, randomSeed, debugLevel,
    timeStructuresSavingInterval,maxLOut,
    simFolder,lMaxInflux,fileTimesSaveInterval,nHours,nAttempts,
    revRctRatio,newSpeciesProbMinThreshold,volumeGrowth,
    stochDivision)
%function [firing_disk concentrazioni_iniziali specie_def influx catalizzatore reazione
    specie_non_esistenti matrice_adiacenza_sub_prod matrice_adiacenza_cat_prod] = inizializzatore_ACS(
```

```

nGEN, nSIM, nSeconds, nReactions, initialMaxLength,
massima_lunghezza_su_cui_calcolare_le_reazioni,
overallConcentration, alphabet,
complexFormationSymmetry,
fino_a_che_lunghezza_i_polimeri_non_catalizzano,
reactionProbability, cleavageProbability,
diffusion_contribute, solubility_threshold,
influx_rate, reverseReactions,K_nrg,
moleculeDecay_KineticConstant, ratio_firing_disk,
lunghezza_max_fd, scelta_concentrazioni,
gamma_powerlaw_concentrazioni,
decisione_catalizzatori, lastFiringDiskSpeciesID,
ECConcentration, volume, energy, controllo_ACS_nel_ciclo, K_nrg_decay, nome_cartella,
funzioni_booleane_in_dec, ratioSpeciesEnergizable,
Kass,Kdiss,Kcpx,K_cpx, randomSeed, debugLevel,
timeStructuresSavingInterval,maxLOut,
simFolder,lMaxInflux,fileTimesSaveInterval,nHours,nAttempts,
revRctRatio,newSpeciesProbMinThreshold,volumeGrowth,
stochDivision)

rand

%-----
%apertura file
%-----
thisFolder = pwd

```

Definition at line 1 of file iniziatore_ACS.m.

13.46.2.12 end end if gg

Initial value:

```

= input('')
% %      specie_def(i,jjjj)
% %      isspace(specie_def(i,jjjj))
% %      specie_def(i,jjjj)==' '
%      if sum(alphabetspecie_def(i,jjjj)) || specie_def(
i,jjjj)==' '
%      else
%      count = fprintf(fid2,'%c',specie_def(
i,jjjj))

```

Definition at line 232 of file iniziatore_ACS.m.

13.46.2.13 for i

Initial value:

```

= massima_lunghezza_su_cui_calcolare_le_reazioni:-1:1
specie_temp=crea_tutte_le_combinazioni_di_elementi(alphabet,i)

```

Definition at line 174 of file iniziatore_ACS.m.

13.46.2.14 for j

Initial value:

```

= length(specie_temp(:,1)):-1:1
k = k+1

```

Definition at line 177 of file iniziatore_ACS.m.

13.46.2.15 for jjjj

Initial value:

```

=1:length(specie_def(1,:))
    controllo = 0

```

Definition at line 216 of file inizializzatore_ACS.m.

13.46.2.16 end end clear specie_temp end k = 0

Definition at line 188 of file inizializzatore_ACS.m.

13.46.2.17 lunghezza_stringa = length(specie_def(1,:))

Definition at line 215 of file inizializzatore_ACS.m.

```

13.46.2.18 concentrazione molecole cariche if rand < ratioSpeciesEnergizable% count = fprintf(fid2,'%d\n',1);
% specie energizzabile% else% count = fprintf(fid2,'%d\n',0); % specie NON energizzabile% end
if influx_rate == 0 if maxLOut > if i <= (2^(maxLOut+1)-2) count = fprintf(fid2,'%d\n',1);
else count = fprintf(fid2,'%d\n',0); end else count = fprintf(fid2,'%d\n',0); end else count =
fprintf(fid2,'%d\n',0); endend%-----%inizializzazione del file
dell'influx " _influx.csv"%-----for i = 1:length(influx(:,1))
count = fprintf(fid5,'%d \t', influx(i,1)-1); count = fprintf(fid5,'%d \n', influx(i,2));endst =
fclose(fid5);%-----%inizializzazione del file delle reazioni
e dei catalizzatori%-----if catalizzatore(1,1) ==
-9999;else for i=1:length(catalizzatore(:,1)) count = fprintf(fid4,'%d\t',catalizzatore(i,1)-1); count
= fprintf(fid4,'%d\t',catalizzatore(i,2)-1); count = fprintf(fid4,'%d\t',catalizzatore(i,3)-1); count
= fprintf(fid4,'%d\t',0); %quante volte if catalizzatore(i,4)==0 tmpKdiss = Kdiss / revRctRatio;
count = fprintf(fid4,'%g\t',Kass); %kass count = fprintf(fid4,'%g\t',tmpKdiss); %kdiss count =
fprintf(fid4,'%g\t',Kcpx); %k complex else tmpKass = Kass / revRctRatio; tmpKcpx = Kcpx / revRctRatio;
count = fprintf(fid4,'%g\t',tmpKass); %kass count = fprintf(fid4,'%g\t',Kdiss); %kdiss count =
fprintf(fid4,'%g\t',tmpKcpx); %k complex end index_rand = ceil(rand*2); count = fprintf(fid4,'%g\n',index_rand);
%nuovo parametro end [righe_xx colonne_xx]=size(funzioni_booleane_in_dec); for
i=1:length(reazione(:,1)) indexx = ceil(rand*righe_xx); funzione_giusta = funzioni_booleane_in_dec(indexx);
count = fprintf(fid3,'%d\t',reazione(i,1)-1); count = fprintf(fid3,'%d\t',reazione(i,2)); count
= fprintf(fid3,'%d\t',reazione(i,3)-1); count = fprintf(fid3,'%d\t',reazione(i,4)-1); count =
fprintf(fid3,'%d\t',reazione(i,5)-1); count = fprintf(fid3,'%d\t',0); if energy == 2 eso_endo = 1; %perch? se non
c'? l'energia sono tutte eso else if rand > energy if reazione(i, 2)

```

Initial value:

```

==1
    eso_endo =1

```

Definition at line 345 of file inizializzatore_ACS.m.

13.46.2.19 end specie_def =specie_def_2

Definition at line 193 of file inizializzatore_ACS.m.

13.46.2.20 inserisco il numero decimale relativo alla funzione booleana della reazione end end st = fclose(fid1)

Definition at line 126 of file inizializzatore_ACS.m.

13.46.2.21 `temporal = specie_non_esistenti{i}`

Definition at line 376 of file `inizializzatore_ACS.m`.

13.46.2.22 `tmpStr = specie_def(i,:)`

Definition at line 243 of file `inizializzatore_ACS.m`.

13.46.2.23 `vettore_rand = [0.5 1 2]`

Definition at line 127 of file `inizializzatore_ACS.m`.

13.46.2.24 `for xxxx`

Initial value:

```
= 1:length(alphabet)
    if sum(alphabet (xxxx) == specie_def (i,
        jjjj)) > 0
        controllo = 1
```

Definition at line 218 of file `inizializzatore_ACS.m`.

13.46.2.25 `for z`

Initial value:

```
= 1:length(specie_temp(1,:))
    specie_def(k,z) = specie_temp(j,z)
```

Definition at line 179 of file `inizializzatore_ACS.m`.

13.47 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/lancia_acs.m File Reference

Functions

- `Keq(k,j)`

Variables

- `lancia_acs.m` clear all close
all parametri `kdiss = 0`
- `kcomplex = 1e6`
- `kcond = 6e8`
- `k_complex = 1e-4`
- `AB = 0`
- `C = 2e-7`
- `CA = 0`
- `k = 0`
- for `A`
- for `B`

13.47.1 Function Documentation

13.47.1.1 $K_{eq}(k, j)$

13.47.2 Variable Documentation

13.47.2.1 for A

Initial value:

```
= 10e-7:10e-7:10e-6
```

```
    k=k+1
    j=0
```

Definition at line 17 of file lancia_acs.m.

13.47.2.2 $AB = 0$

Definition at line 12 of file lancia_acs.m.

13.47.2.3 for B

Initial value:

```
= 10e-7:10e-7:10e-6
```

```
    j=j+1
```

```
    [t y] = ACS_reverse_reaction_con_input(A,B,AB,C,CA,kdiss,
    kcomplex,kcond,kcomplex)
```

Definition at line 21 of file lancia_acs.m.

13.47.2.4 $C = 2e-7$

Definition at line 13 of file lancia_acs.m.

13.47.2.5 $CA = 0$

Definition at line 14 of file lancia_acs.m.

13.47.2.6 $k = 0$

Definition at line 16 of file lancia_acs.m.

13.47.2.7 $k_{complex} = 1e-4$

Definition at line 10 of file lancia_acs.m.

13.47.2.8 $k_{complex} = 1e6$

Definition at line 8 of file lancia_acs.m.

13.47.2.9 kcond = 6e8

Definition at line 9 of file lancia_acs.m.

13.47.2.10 lancia_acs m clear all close all parametri kdiss = 0

Definition at line 7 of file lancia_acs.m.

13.48 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/lancia_inizializzatore_acs.m File Reference

Functions

- id [nGEN](#) ()
- experiment all condensation are [eso](#) (and cleavage endo)) [energy](#)
- id [ratio_firing_disk](#) ()

Variables

- [nSIM](#) =1
- [nSeconds](#) =400
- [nReactions](#) =200000000
- [initialPopulationNumber](#) =0
- [initialMaxLength](#) =3
- [massima_lunghezza_su_cui_calcolare_le_reazioni](#) = 3
- experiment [__pad1__](#)
- [alphabet](#) = ['AB']
- [lastFiringDiskSpeciesID](#) = 0
- for i
- [overallConcentration](#) =1e-4
- [volume](#) =1e-15
- experiment [__pad2__](#)
- [energyTarget](#) =0
- [complexFormationSymmetry](#) =0
- [**NEW ** fino_a_che_lunghezza_i_polimeri_non_catalizzano](#) = 2
- [reactionProbability](#) =0.004
- experiment [__pad3__](#)
- [reverseReactions](#) =0
- costanti CINETICHE [K_eq](#) =1000
- parte da rivedere e correggere [Kass_o_Kdiss](#) = 0
- [se](#) cleavage [kdiss](#) = 100
- [se](#) cleavage [se](#) a
[condensazione kass](#) =100
- [rapporto_Kfor_Kback](#) = 0
- e g [Kass](#)
- [K_cpxDiss](#) =0
- coefficiente di [fosforilazione](#)
- [K_nrg](#) = 0
- [K_irrad](#) = 0
- altri parametri [moleculeDecay_KineticConstant](#) =0.02
- [diffusion_contribute](#) =0
- [solubility_threshold](#) =0

- experimentveri `influx` =2
- `influx_rate` =1e-6
- percentuale di specie da cancellare rispetto a tutte quelle che restano dopo aver conservato i polimeri fino a `lunghezzamaxfd` `lunghezza_max_fd` = 1
- `lunghezza` dei monomeri polimeri da conservare `scelta_concentrazioni` =1
- parametro `switch`
- parametro uniforme sulle `lunghezze`
- parametro uniforme sulle powerlaw con esponente gamma `gamma_powerlaw_concentrazioni` = 2.1
- esponente della powerlaw in caso parametri per la distribuzione dei `catalizzatori`
- richiamo la funzione `inizializzatore_ACS` [fd concentrazioni `specie_def` `influx` `catalizzatore` `reazione` `specie-non_esistenti` `matrice_adiacenza_sub_prod` `matrice_adiacenza_cat_prod`] = `inizializzatore_ACS`(nGEN, nSIM, nSeconds, nReactions, initialPopulationNumber, initialMaxLength, `massima_lunghezza_su_cui_calcolare_le_reazioni`, overallConcentration, alphabet, complexFormationSymmetry, `fino_a_che_lunghezza_i_polimeri_non_catalizzano`, reactionProbability, cleavageProbability, `diffusion_contribute`, `solubility_threshold`, `influx`, `influx_rate`, `reverseReactions`, `Kass_o_Kdiss`,rapporto_Kfor_Kback,rapporto_Kcpx_K-ass,K_cpxDiss, `K_nrg`, `moleculeDecay_KineticConstant`, `ratio_firing_disk`, `lunghezza_max_fd`, `scelta_concentrazioni`, `gamma_powerlaw_concentrazioni`,`decisione_catalizzatori`, `lastFiringDiskSpeciesID`, EC-Concentration, `volume`, `energy`, `energyTarget`, `controllo_ACS_nel_ciclo`, `K_irrad`)

13.48.1 Function Documentation

13.48.1.1 experiment all condensation are eso (and cleavage endo)

13.48.1.2 id nGEN () [virtual]

13.48.1.3 id ratio_firing_disk () [virtual]

13.48.2 Variable Documentation

13.48.2.1 experiment __pad1__

Definition at line 27 of file `lancia_inizializzatore_acs.m`.

13.48.2.2 experiment __pad2__

Definition at line 45 of file `lancia_inizializzatore_acs.m`.

13.48.2.3 experiment __pad3__

Definition at line 57 of file `lancia_inizializzatore_acs.m`.

13.48.2.4 alphabet = ['AB']

Definition at line 29 of file `lancia_inizializzatore_acs.m`.

13.48.2.5 esponente della powerlaw in caso parametri per la distribuzione dei catalizzatori

Definition at line 89 of file lancia_inizializzatore_acs.m.

13.48.2.6 complexFormationSymmetry =0

Definition at line 51 of file lancia_inizializzatore_acs.m.

13.48.2.7 diffusion_contribute =0

Definition at line 75 of file lancia_inizializzatore_acs.m.

13.48.2.8 energyTarget =0

Definition at line 49 of file lancia_inizializzatore_acs.m.

13.48.2.9 ** NEW fino_a_che_lunghezza_i_polimeri_non_catalizzano = 2**

Definition at line 54 of file lancia_inizializzatore_acs.m.

13.48.2.10 coefficiente di fosforilazione

Definition at line 68 of file lancia_inizializzatore_acs.m.

13.48.2.11 parametro uniforme sulle powerlaw con esponente gamma gamma_powerlaw_concentrazioni = 2.1

Definition at line 85 of file lancia_inizializzatore_acs.m.

13.48.2.12 for i**Initial value:**

```
= 1:massima_lunghezza_su_cui_calcolare_le_reazioni  
   lastFiringDiskSpeciesID = lastFiringDiskSpeciesID +  
   length(alphabet)^i
```

Definition at line 33 of file lancia_inizializzatore_acs.m.

13.48.2.13 controllo che non ci siano cicli nell influx =2

Definition at line 77 of file lancia_inizializzatore_acs.m.

13.48.2.14 influx_rate =1e-6

Definition at line 78 of file lancia_inizializzatore_acs.m.

13.48.2.15 initialMaxLength =3

Definition at line 23 of file lancia_inizializzatore_acs.m.

13.48.2.16 initialPopulationNumber = 0

Definition at line 21 of file lancia_inizializzatore_acs.m.

13.48.2.17 richiamo la funzione inizializzatore_ACS[fd concentrazioni specie_def influx catalizzatore reazione specie_non_esistenti matrice_adiacenza_sub_prod matrice_adiacenza_cat_prod] = inizializzatore_ACS(nGEN, nSIM, nSeconds, nReactions, initialPopulationNumber, initialMaxLength, massima_lunghezza_su_cui_calcolare_le_reazioni, overallConcentration, alphabet, complexFormationSymmetry, fino_a_che_lunghezza_i_polimeri_non_catalizzano, reaction-Probability, cleavageProbability, diffusion_contribute, solubility_threshold, influx, influx_rate, reverseReactions, Kass_o_Kdiss, rapporto_Kfor_Kback, rapporto_Kcpx_K_ass, K_cpxDiss, K_nrg, moleculeDecay_KineticConstant, ratio_firing_disk, lunghezza_max_fd, scelta_concentrazioni, gamma_powerlaw_concentrazioni, decisione_catalizzatori, lastFiringDiskSpeciesID, ECConcentration, volume, energy, energyTarget, controllo_ACS_nel_ciclo, K_irrad)

Definition at line 100 of file lancia_inizializzatore_acs.m.

13.48.2.18 K_cpxDiss = 0

Definition at line 66 of file lancia_inizializzatore_acs.m.

13.48.2.19 costanti CINETICHE K_eq = 1000

Definition at line 61 of file lancia_inizializzatore_acs.m.

13.48.2.20 K_irrad = 0

Definition at line 71 of file lancia_inizializzatore_acs.m.

13.48.2.21 K_nrg = 0

Definition at line 69 of file lancia_inizializzatore_acs.m.

13.48.2.22 se cleavage se a condensazione kass = 100

Definition at line 63 of file lancia_inizializzatore_acs.m.

13.48.2.23 e g Kass

Initial value:

```
= 100 --> Kdiss = 100/100 = 1  
rapporto_Kcpx_K_ass = 0
```

Definition at line 64 of file lancia_inizializzatore_acs.m.

13.48.2.24 parte da rivedere e correggere Kass_o_Kdiss = 0

Definition at line 63 of file lancia_inizializzatore_acs.m.

13.48.2.25 **se cleavage kdiss = 100**

Definition at line 63 of file lancia_inizializzatore_acs.m.

13.48.2.26 **end lastFiringDiskSpeciesID = 0**

Definition at line 32 of file lancia_inizializzatore_acs.m.

13.48.2.27 **percentuale di specie da cancellare rispetto a tutte quelle che restano dopo aver conservato i polimeri fino a
lunghezzamaxfd lunghezza_max_fd = 1**

Definition at line 83 of file lancia_inizializzatore_acs.m.

13.48.2.28 **parametro uniforme sulle lunghezze**

Definition at line 84 of file lancia_inizializzatore_acs.m.

13.48.2.29 **massima_lunghezza_su_cui_calcolare_le_reazioni = 3**

Definition at line 24 of file lancia_inizializzatore_acs.m.

13.48.2.30 **altri parametri moleculeDecay_KineticConstant =0.02**

Definition at line 74 of file lancia_inizializzatore_acs.m.

13.48.2.31 **nReactions =200000000**

Definition at line 20 of file lancia_inizializzatore_acs.m.

13.48.2.32 **nSeconds =400**

Definition at line 19 of file lancia_inizializzatore_acs.m.

13.48.2.33 **nSIM =1**

Definition at line 18 of file lancia_inizializzatore_acs.m.

13.48.2.34 **overallConcentration =1e-4**

Definition at line 38 of file lancia_inizializzatore_acs.m.

13.48.2.35 **rapporto_Kfor_Kback = 0**

Definition at line 64 of file lancia_inizializzatore_acs.m.

13.48.2.36 **reactionProbability =0.004**

Definition at line 56 of file lancia_inizializzatore_acs.m.

13.48.2.37 reverseReactions =0

Definition at line 58 of file lancia_inizializzatore_acs.m.

13.48.2.38 lunghezza dei monomeri polimeri da conservare scelta_concentrazioni =1

Definition at line 84 of file lancia_inizializzatore_acs.m.

13.48.2.39 solubility_threshold =0

Definition at line 76 of file lancia_inizializzatore_acs.m.

13.48.2.40 parametro switch

Definition at line 84 of file lancia_inizializzatore_acs.m.

13.48.2.41 volume =1e-15

Definition at line 40 of file lancia_inizializzatore_acs.m.

13.49 /Users/alessandrofilisetti/Documents/GIT/carness/_matlabinitializer/start.m File Reference

Functions

- Numero di [simulazioni](#) (diversi semi random) [nSeconds](#)
- Numero massimo di [reazioni](#) (secondo parametro di [stop](#) oltre al numero di secondi) [nHours=0](#)
- Numero massimo di ore per la [simulazione](#) (=0 no vincolo) [nAttempts=0](#)
- Numero di volte in cui sistema ritenta la stessa [rete](#) (=0 no vincolo) [initialMaxLength](#)
- Number of different networks for every network will be performed [nSim](#) different [simulation](#) (differnt random seeds) [simFolder.name](#)
- [if exist](#) ([simFolder.name](#), 'dir')
- introduzione delle FUNZIONI BOOLEANE nell che verranno successivamente assegnate con probabilit uniforme alle [reazioni](#) la funzione divisa a la prima [met](#) corrisponde alla [condensazione](#) (8 bit)
- converto in decimale for per default uniforme [funzioni_booleane_in_dec](#) (i, 2)

Variables

- lancia_serie_di_inizializzatore
m clear all close all
*****Inserimento
dei PARAMETRI VARIABILI sui
quali fare lo [SCREENING](#)

- lancia_serie_di_inizializzatore
m clear all close all
*****Inserimento
dei PARAMETRI VARIABILI sui
quali fare lo sottoforma di
matrici o vettori `reactionProbability` = [0.000516529
- `nome_prob` = [0.125
- `IMaxInflux` = [2]
- `parametro_screening` = `IMaxInflux`
- `nome_folder` = [2]
- *****
dei PARAMETRI FISSI
- *****
dei PARAMETRI quelli che
restano cio costanti in tutti
gli esperimenti della serie `nSIM` =1
- Numero di secondi `randomSeed` =0
- lasciare `a debugLevel` =0
- livello di dettaglio messaggi
durante `simulazione`
- livello di dettaglio messaggi
durante lasciare `a`
- livello di dettaglio messaggi
durante lasciare per debug
software `timeStructuresSavingInterval` =`nSeconds`/100
- definisce il tempo in cui
vengono salvati i file durante
la `simulazione` `fileTimesSaveInterval` =0
- Definisce il tempo in cui
vengono salvati i dati sui
`file times`
- Definisce il tempo in cui
vengono salvati i dati sui
`file` `reaction_parameter` e i
vari living `nReactions` =200000000
- Lunghezza massima delle specie
da creare `massima_lunghezza_su_cui_calcolare_le_reazioni` = 6
- Lunghezza massima fino alla
quale creare le `reazioni` `maxLOut` = 2
- `se` =0 non viene considerato
- altrimenti Quando `influx_rate`
indica la `lunghezza` massima
delle molecole che possono
uscire dal `contenitore`
- altrimenti Quando `influx_rate`
indica la `lunghezza` massima
delle molecole che possono
uscire dal quando `influx_rate`
- `alphabet` = ['AB']
- `overallConcentration` =0.0333
- `volume` =1e-18
- `energy` =0
- `energia considerata`
- `energia` non `considerata` `complexFormationSymmetry` =0
- `fino_a_che_lunghezza_i_polimeri_non_catalizzano` = 2
- `cleavageProbability` =0.5

- `reverseReactions` =0
- `Kass` = 50
- `Kdiss` = 25
- `Kcpx` = 50
- `K_cpx` = 1
- `K_nrg` = 0
- `K_nrg_decay` =0
- coefficiente di decadimento delle molecole o dei carrier dalla propria componente energetica `revRctRatio` = 10
- `ratioSpeciesEnergizable` = 0
- percentuale di specie presenti nel sistema che possono essere energizzate per ogni specie create `c` una certa probabilit di essere energizzabile o meno `moleculeDecay_KineticConstant` =0.02
- `diffusion_contribute` =0
- `solubility_threshold` =0
- `lunghezza` massima delle molecole presenti nell `influx ratio_firing_disk` =0
- percentuale di specie da cancellare rispetto `a` tutte quelle che restano dopo aver conservato `i` polimeri fino `a` `lunghezzamaxfd` `lunghezza_max_fd` = 7
- `lunghezza` dei monomeri polimeri da conservare `scelta_concentrazioni` =1
- parametro `switch`
- parametro uniforme sulle `lunghezze`
- parametro uniforme sulle powerlaw con esponente gamma `gamma_powerlaw_concentrazioni` = 2.1
- esponente della powerlaw in caso `decisione_catalizzatori` =1
- parametri per la distribuzione dei `catalizzatori`
- `controllo` che non ci siano cicli nell `influx`
- NUOVO PARAMETRO `volumeGrowth` = 0
- NUOVO PARAMETRO `stochDivision` = 0
- NUOVO PARAMETRO `simFolder nets` = 10
- Number of different networks `ensambles`
- Nome della cartella dove verr salvata la `simulazione` `simFolder path` = 'SIMS'
- Percorso dove verr creata la cartella `simFolder` dove verranno salvati tutti i file *****
`nGEN` =10
- Numero di `generazioni`
- Numero di al momento significa che alla fine di ogni generazione da ogni `file` di fine sim partono altre `Nsim`
- Numero di al momento significa che alla fine di ogni generazione da ogni `file` di fine sim partono altre lasciare ad `lLastFiringDiskSpeciesID` = 0

- calcolata in automatico for `i`
- `end lastFiringDiskSpeciesID` = `lastFiringDiskSpeciesID -1`
- `thisFolder` = `pwd`
- `end fid20` = `fopen('lanciatore.sh','w')`
- introduzione delle FUNZIONI BOOLEANE nell `energia`
- introduzione delle FUNZIONI BOOLEANE nell che verranno successivamente assegnate con probabilit uniforme alle `reazioni` la funzione divisa `a met`
- introduzione delle FUNZIONI BOOLEANE nell che verranno successivamente assegnate con probabilit uniforme alle `reazioni` la funzione divisa `a` la prima `met` corrisponde alla la seconda parte al cleavage(4 bit).%esempio `funzioni_booleane_in_dec` = `bi2de(funzioni_booleane,'left-msb')`
- `parte2_nome_cartella` = `num2str(nome_folder(i))`
- `parte3_nome_cartella` = `('rete_n_')`

13.49.1 Function Documentation

- 13.49.1.1 introduzione delle FUNZIONI BOOLEANE nell che verranno successivamente assegnate con probabilit uniforme alle `reazioni` la funzione divisa `a` la prima `met` corrisponde alla condensazione (8 bit)
- 13.49.1.2 `if exist (simFolder. name, 'dir')`
- 13.49.1.3 converto in decimale for per default uniforme `funzioni_booleane_in_dec (i , 2)`
- 13.49.1.4 Numero massimo di reazioni (secondo parametro di `stop` oltre al numero di `secondi`) [pure virtual]
- 13.49.1.5 Numero di volte in cui sistema ritenta la stessa rete ()
- 13.49.1.6 Number of different networks for every network will be performed nSim different simulation (differnt random *seeds*)
- 13.49.1.7 Numero massimo di ore per la simulazione () [pure virtual]
- 13.49.1.8 Numero di simulazioni (diversi semi *random*)

13.49.2 Variable Documentation

- 13.49.2.1 Numero di al momento significa che alla fine di ogni generazione da ogni file di fine sim partono altre lasciare ad `!lastFiringDiskSpeciesID = 0`

Definition at line 79 of file `start.m`.

- 13.49.2.2 livello di dettaglio messaggi durante lasciare a

Definition at line 26 of file `start.m`.

- 13.49.2.3 `alphabet = ['AB']`

Definition at line 36 of file `start.m`.

13.49.2.4 parametri per la distribuzione dei catalizzatori

Definition at line 63 of file start.m.

13.49.2.5 cleavageProbability =0.5

Definition at line 42 of file start.m.

13.49.2.6 energia non considerata complexFormationSymmetry =0

Definition at line 40 of file start.m.

13.49.2.7 energia considerata

Definition at line 39 of file start.m.

13.49.2.8 altrimenti Quando influx_rate indica la lunghezza massima delle molecole che possono uscire dal contenitore

Definition at line 34 of file start.m.

13.49.2.9 lasciare a debugLevel =0

Definition at line 26 of file start.m.

13.49.2.10 esponente della powerlaw in caso decisione_catalizzatori =1

Definition at line 62 of file start.m.

13.49.2.11 diffusion_contribute =0

Definition at line 54 of file start.m.

13.49.2.12 introduzione delle FUNZIONI BOOLEANE nell energia

Definition at line 111 of file start.m.

13.49.2.13 energy =0

Definition at line 39 of file start.m.

13.49.2.14 Number of different networks ensambles

Definition at line 72 of file start.m.

13.49.2.15 end fid20 = fopen('lanciatore.sh','w')

Definition at line 92 of file start.m.

13.49.2.16 definisce il tempo in cui vengono salvati i file durante la simulazione fileTimesSaveInterval =0

Definition at line 28 of file start.m.

13.49.2.17 fino_a_che_lunghezza_i_polimeri_non_catalizzano = 2

Definition at line 41 of file start.m.

13.49.2.18 *****

 Inserimento dei PARAMETRI FISSI

Definition at line 20 of file start.m.

13.49.2.19 converto in decimale funzioni_booleane_in_dec = bi2de(funzioni_booleane,'left-msb')

Definition at line 119 of file start.m.

13.49.2.20 parametro uniforme sulle powerlaw con esponente gamma gamma_powerlaw_concentrazioni = 2.1

Definition at line 61 of file start.m.

13.49.2.21 Numero di generazioni

Definition at line 78 of file start.m.

13.49.2.22 end*****
 ***** START***** for i

Initial value:

```
= 1:massima_lunghezza_su_cui_calcolare_le_reazioni
    lastFiringDiskSpeciesID = lastFiringDiskSpeciesID +
    length(alphabet)^i
```

Definition at line 80 of file start.m.

13.49.2.23 controllo che non ci siano cicli nell'influx

Definition at line 68 of file start.m.

13.49.2.24 influx_rate

Initial value:

```
= 0 indica fino a quale lunghezza le molecole non variano in quantita' (simulazione membrana
    permeabile)
ECCConcentration=0
```

Definition at line 34 of file start.m.

13.49.2.25 `K_cpx = 1`

Definition at line 48 of file start.m.

13.49.2.26 `K_nrg = 0`

Definition at line 49 of file start.m.

13.49.2.27 `K_nrg_decay = 0`

Definition at line 50 of file start.m.

13.49.2.28 `Kass = 50`

Definition at line 45 of file start.m.

13.49.2.29 `Kcpx = 50`

Definition at line 47 of file start.m.

13.49.2.30 `Kdiss = 25`

Definition at line 46 of file start.m.

13.49.2.31 `end lastFiringDiskSpeciesID = lastFiringDiskSpeciesID -1`

Definition at line 83 of file start.m.

13.49.2.32 `se il sistema chiuso lMaxInflux = [2]`

Definition at line 13 of file start.m.

13.49.2.33 `percentuale di specie da cancellare rispetto a tutte quelle che restano dopo aver conservato i polimeri fino a
lunghezzamaxfd lunghezza_max_fd = 7`

Definition at line 59 of file start.m.

13.49.2.34 `parametro uniforme sulle lunghezze`

Definition at line 60 of file start.m.

13.49.2.35 `Lunghezza massima delle specie da creare massima_lunghezza_su_cui_calcolare_le_reazioni = 6`

Definition at line 33 of file start.m.

13.49.2.36 `Lunghezza massima fino alla quale creare le reazioni maxLOut = 2`

Definition at line 34 of file start.m.

13.49.2.37 introduzione delle FUNZIONI BOOLEANE nell che verranno successivamente assegnate con probabilit uniforme alle reazioni la funzione divisa a met

Definition at line 111 of file start.m.

13.49.2.38 percentuale di specie presenti nel sistema che possono essere energizzate per ogni specie create c una certa probabilit di essere energizzabile o meno moleculeDecay_KineticConstant =0.02

Definition at line 53 of file start.m.

13.49.2.39 NUOVO PARAMETRO simFolder nets = 10

Definition at line 72 of file start.m.

13.49.2.40 Percorso dove verr creata la cartella simFolder dove verranno salvati tutti i file***** nGEN =10

Definition at line 78 of file start.m.

13.49.2.41 nome_folder = [2]

Definition at line 15 of file start.m.

13.49.2.42 nome_prob = [0.125]

Definition at line 12 of file start.m.

13.49.2.43 Definisce il tempo in cui vengono salvati i dati sui file reaction_parameter e i vari living nReactions =200000000

Definition at line 29 of file start.m.

13.49.2.44 *****

Inserimento dei PARAMETRI quelli che restano cio costanti in tutti gli esperimenti della serie nSIM =1

Definition at line 23 of file start.m.

13.49.2.45 Numero di al momento significa che alla fine di ogni generazione da ogni file di fine sim partono altre Nsim

Definition at line 78 of file start.m.

13.49.2.46 overallConcentration =0.0333

Definition at line 37 of file start.m.

13.49.2.47 parametro_screening = IMaxInflux

Definition at line 14 of file start.m.

13.49.2.48 `parte2_nome_cartella = num2str(nome_folder(i))`

Definition at line 142 of file start.m.

13.49.2.49 `parte3_nome_cartella = ('_rete_n_')`

Definition at line 143 of file start.m.

13.49.2.50 `concAnalysis(params) clear all close all params path = 'SIMS'`

Definition at line 74 of file start.m.

13.49.2.51 `Numero di secondi randomSeed =0`

Definition at line 25 of file start.m.

13.49.2.52 `lunghezza massima delle molecole presenti nell influx ratio_firing_disk =0`

Definition at line 58 of file start.m.

13.49.2.53 `ratioSpeciesEnergizable = 0`

Definition at line 52 of file start.m.

13.49.2.54 `reactionProbability = [0.000516529`

Definition at line 11 of file start.m.

13.49.2.55 `reverseReactions =0`

Definition at line 43 of file start.m.

13.49.2.56 `coefficiente di decadimento delle molecole o dei carrier dalla propria componente energetica revRctRatio = 10`

Definition at line 51 of file start.m.

13.49.2.57 `lunghezza dei monomeri polimeri da conservare scelta_concentrazioni =1`

Definition at line 60 of file start.m.

13.49.2.58 `lancia_serie_di_inizializzatore m clear all close`

`all*****
Inserimento dei PARAMETRI VARIABILI sui quali fare lo SCREENING`

Definition at line 8 of file start.m.

13.49.2.59 `se =0 non viene considerato`

Definition at line 34 of file start.m.

13.49.2.60 livello di dettaglio messaggi durante simulazione

Definition at line 26 of file start.m.

13.49.2.61 solubility_threshold =0

Definition at line 55 of file start.m.

13.49.2.62 NUOVO PARAMETRO stochDivision = 0

Definition at line 70 of file start.m.

13.49.2.63 parametro switch

Definition at line 60 of file start.m.

13.49.2.64 thisFolder = pwd

Definition at line 86 of file start.m.

13.49.2.65 Definisce il tempo in cui vengono salvati i dati sui file times

Definition at line 28 of file start.m.

13.49.2.66 livello di dettaglio messaggi durante lasciare per debug software timeStructuresSavingInterval =nSeconds/100

Definition at line 27 of file start.m.

13.49.2.67 volume =1e-18

Definition at line 38 of file start.m.

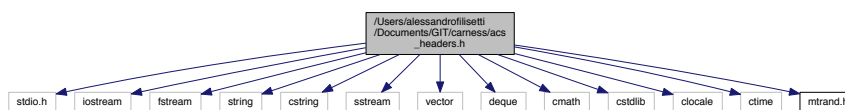
13.49.2.68 NUOVO PARAMETRO volumeGrowth = 0

Definition at line 69 of file start.m.

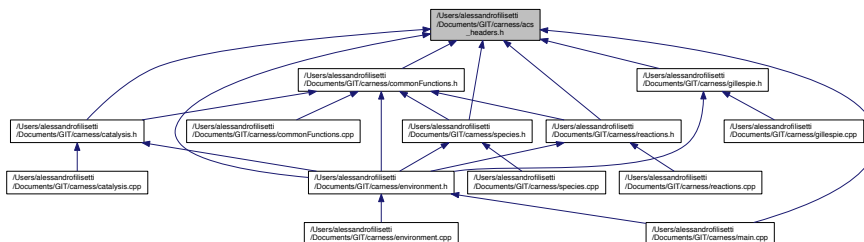
13.50 /Users/alessandrofilisetti/Documents/GIT/carness/acs_headers.h File Reference

```
#include <stdio.h>
#include <iostream>
#include <fstream>
#include <string>
#include <cstring>
#include <sstream>
#include <vector>
#include <deque>
#include <cmath>
#include <cstdlib>
#include <locale>
#include <ctime>
#include "mtrand.h"
```


Include dependency graph for acs_headers.h:



This graph shows which files directly or indirectly include this file:



Macros

- #define `__SOFTWARE_VERSION__` "4.8b20131209.62"
- #define `MINIMAL_PROMPT` -1
- #define `RUNNING_VERSION` 0
- #define `SMALL_DEBUG` 1
- #define `MEDIUM_DEBUG` 2
- #define `HIGH_DEBUG` 3
- #define `FINDERERRORDURINGRUNTIME` -10
- #define `COMPLEXSTUFF` -20
- #define `GILLESPIESTUFF` -30
- #define `RANDOMRANGE` random()
- #define `PROPORTIONALMOLECULEAMOUNT` 1
- #define `UNIFORMMOLECULEAMOUNT` 2
- #define `INVPROPORTIONALMOLECULEAMOUNT` 3
- #define `CONDENSATION` 0
- #define `CLEAVAGE` 1
- #define `COMPLEXFORMATION` 2
- #define `COMPLEXDEGRADATION` 3
- #define `SPECIESDECAY` 4
- #define `PHOSPHORILATION` 5
- #define `ENDO_CLEAVAGE` 6
- #define `ENDO_CONDENSATION` 7
- #define `ENDO_COMPLEXFORMATION` 8
- #define `ENERGYEFFLUX` 9
- #define `SPONTANEOUS_CONDENSATION` 10
- #define `SPONTANEOUS_CLEAVAGE` 11
- #define `SOLUBLE` 1
- #define `PRECIPITATED` 0
- #define `ESOERGONIC` 1
- #define `ENDOERGONIC` 0
- #define `CLEAVAGEBASED` 1

- `#define CONDENSATIONBASED 0`
- `#define ENERGYBASED 1`
- `#define ENERGYFREE 0`
- `#define TRUENRG '1'`
- `#define FALSENRG '0'`
- `#define ENERGIZABLE 1`
- `#define NOTENERGIZABLE 0`
- `#define SUBSTRATELOAD 0`
- `#define CATALYSTLOAD 1`
- `#define BOTHLOAD 2`
- `#define COMPLEXLOAD 3`
- `#define NOTHINGLOAD 4`
- `#define NEWREACTIONS 1`
- `#define UPGRADEREACTIONS 0`
- `#define NOSPONTANEOUS 0`
- `#define NEP 2.7182818284590452353602874`
- `#define AVO 6.02214179e+23`
- `#define MINIMALRCTTIMEMULTI 100`

Typedefs

- typedef long double [acs_double](#)
- typedef unsigned long int [acs_longInt](#)
- typedef unsigned int [acs_int](#)

13.50.1 Macro Definition Documentation

13.50.1.1 `#define __SOFTWARE__ "4.8b20131209.62"`

Definition at line 42 of file `acs_headers.h`.

13.50.1.2 `#define AVO 6.02214179e+23`

Definition at line 108 of file `acs_headers.h`.

13.50.1.3 `#define BOTHLOAD 2`

Definition at line 95 of file `acs_headers.h`.

13.50.1.4 `#define CATALYSTLOAD 1`

Definition at line 94 of file `acs_headers.h`.

13.50.1.5 `#define CLEAVAGE 1`

Definition at line 65 of file `acs_headers.h`.

13.50.1.6 `#define CLEAVAGEBASED 1`

Definition at line 84 of file `acs_headers.h`.

13.50.1.7 #define COMPLEXDEGRADATION 3

Definition at line 67 of file acs_headers.h.

13.50.1.8 #define COMPLEXFORMATION 2

Definition at line 66 of file acs_headers.h.

13.50.1.9 #define COMPLEXLOAD 3

Definition at line 96 of file acs_headers.h.

13.50.1.10 #define COMPLEXSTUFF -20

Definition at line 52 of file acs_headers.h.

13.50.1.11 #define CONDENSATION 0

Definition at line 64 of file acs_headers.h.

13.50.1.12 #define CONDENSATIONBASED 0

Definition at line 85 of file acs_headers.h.

13.50.1.13 #define ENDO_CLEAVAGE 6

Definition at line 70 of file acs_headers.h.

13.50.1.14 #define ENDO_COMPLEXFORMATION 8

Definition at line 72 of file acs_headers.h.

13.50.1.15 #define ENDO_CONDENSATION 7

Definition at line 71 of file acs_headers.h.

13.50.1.16 #define ENDOERGONIC 0

Definition at line 83 of file acs_headers.h.

13.50.1.17 #define ENERGIZABLE 1

Definition at line 90 of file acs_headers.h.

13.50.1.18 #define ENERGYBASED 1

Definition at line 86 of file acs_headers.h.

13.50.1.19 `#define ENERGYEFLUX 9`

Definition at line 73 of file `acs_headers.h`.

13.50.1.20 `#define ENERGYFREE 0`

Definition at line 87 of file `acs_headers.h`.

13.50.1.21 `#define ESOERGONIC 1`

Definition at line 82 of file `acs_headers.h`.

13.50.1.22 `#define FALSENRG '0'`

Definition at line 89 of file `acs_headers.h`.

13.50.1.23 `#define FINDERERRORDURINGRUNTIME -10`

Definition at line 51 of file `acs_headers.h`.

13.50.1.24 `#define GILLESPIESTUFF -30`

Definition at line 53 of file `acs_headers.h`.

13.50.1.25 `#define HIGH_DEBUG 3`

Definition at line 50 of file `acs_headers.h`.

13.50.1.26 `#define INVPROPORTIONALMOLECULEAMOUNT 3`

Definition at line 61 of file `acs_headers.h`.

13.50.1.27 `#define MEDIUM_DEBUG 2`

Definition at line 49 of file `acs_headers.h`.

13.50.1.28 `#define MINIMAL_PROMPT -1`

Definition at line 46 of file `acs_headers.h`.

13.50.1.29 `#define MINIMALRCTTIMEMULTI 100`

Definition at line 111 of file `acs_headers.h`.

13.50.1.30 `#define NEP 2.7182818284590452353602874`

Definition at line 107 of file `acs_headers.h`.

13.50.1.31 #define NEWREACTIONS 1

Definition at line 100 of file acs_headers.h.

13.50.1.32 #define NOSPONTANEOUS 0

Definition at line 104 of file acs_headers.h.

13.50.1.33 #define NOTENERGIZABLE 0

Definition at line 91 of file acs_headers.h.

13.50.1.34 #define NOTHINGLOAD 4

Definition at line 97 of file acs_headers.h.

13.50.1.35 #define PHOSPHORILATION 5

Definition at line 69 of file acs_headers.h.

13.50.1.36 #define PRECIPITATED 0

Definition at line 79 of file acs_headers.h.

13.50.1.37 #define PROPORTIONALMOLECULEAMOUNT 1

Definition at line 59 of file acs_headers.h.

13.50.1.38 #define RANDOMRANGE random()

Definition at line 56 of file acs_headers.h.

13.50.1.39 #define RUNNING_VERSION 0

Definition at line 47 of file acs_headers.h.

13.50.1.40 #define SMALL_DEBUG 1

Definition at line 48 of file acs_headers.h.

13.50.1.41 #define SOLUBLE 1

Definition at line 78 of file acs_headers.h.

13.50.1.42 #define SPECIESDECAY 4

Definition at line 68 of file acs_headers.h.

13.50.1.43 **#define SPONTANEOUS_CLEAVAGE 11**

Definition at line 75 of file acs_headers.h.

13.50.1.44 **#define SPONTANEOUS_CONDENSATION 10**

Definition at line 74 of file acs_headers.h.

13.50.1.45 **#define SUBSTRATELOAD 0**

Definition at line 93 of file acs_headers.h.

13.50.1.46 **#define TRUENRG '1'**

Definition at line 88 of file acs_headers.h.

13.50.1.47 **#define UNIFORMMOLECULEAMOUNT 2**

Definition at line 60 of file acs_headers.h.

13.50.1.48 **#define UPGRADEREACTIONS 0**

Definition at line 101 of file acs_headers.h.

13.50.2 Typedef Documentation

13.50.2.1 **typedef long double acs_double**

Definition at line 34 of file acs_headers.h.

13.50.2.2 **typedef unsigned int acs_int**

Definition at line 36 of file acs_headers.h.

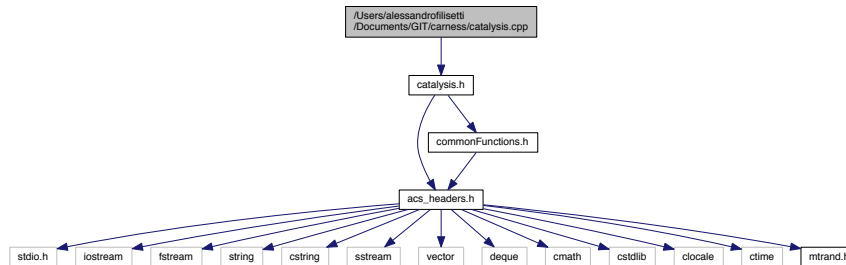
13.50.2.3 **typedef unsigned long int acs_longInt**

Definition at line 35 of file acs_headers.h.

13.51 /Users/alessandrofilisetti/Documents/GIT/carness/catalysis.cpp File Reference

```
#include "catalysis.h"
```

Include dependency graph for catalysis.cpp:

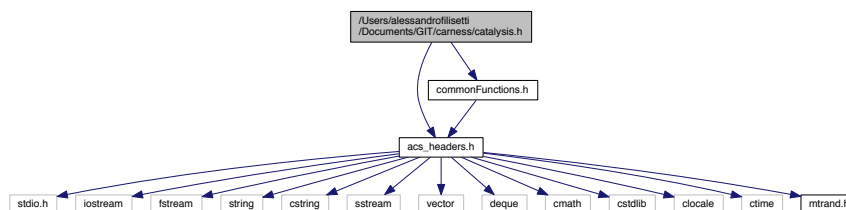


13.52 /Users/alessandrofilisetti/Documents/GIT/carness/catalysis.h File Reference

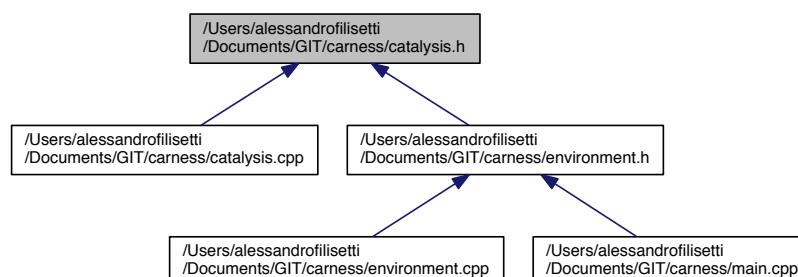
```
#include "acs_headers.h"
```

```
#include "commonFunctions.h"
```

Include dependency graph for catalysis.h:



This graph shows which files directly or indirectly include this file:



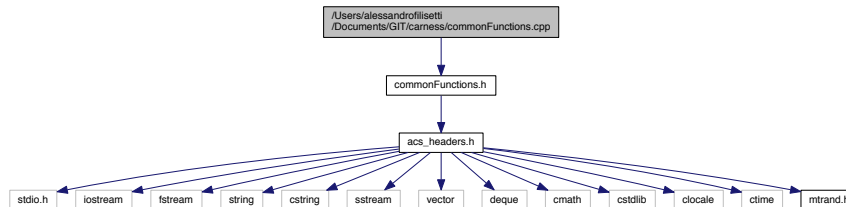
Classes

- class [catalysis](#)
CATALYSIS class.

13.53 /Users/alessandrofilisetti/Documents/GIT/carness/commonFunctions.cpp File Reference

```
#include "commonFunctions.h"
```

Include dependency graph for commonFunctions.cpp:



Functions

- `acs_longInt returnSelectionIdFromAWeightProbVector (acs_double *tmpArray, MTRand &tmpRandomGenerator, acs_int tmpRow)`
This function returns a random position in a probability weight array of N elements.
- `acs_longInt returnSelectionIdFromAWeightProbVector (vector< acs_double > &tmpVector, acs_double tmpMaxValue, MTRand &tmpRandomGenerator, acs_int tmpRow)`
- `acs_longInt returnSelectionIdFromAWeightProbVectorAlreadyNormalized (vector< acs_double > &tmpVector, MTRand &tmpRandomGenerator)`
- `acs_longInt returnUniformSelection_LONG_IdFromVector (vector< acs_longInt > &tmpVector, MTRand &tmpRandomGenerator)`
- `acs_longInt getIntRandom (acs_longInt tmpFromNum, acs_longInt tmpToNum, MTRand &tmpRandomGenerator)`
- `acs_double getDoubleRandom (acs_double tmpFromNum, acs_double tmpToNum, MTRand &tmpRandomGenerator)`
- `acs_longInt random_poisson (acs_double tmpLambda, MTRand &tmpRandomGenerator)`
- `acs_longInt random_binomial (acs_longInt n, acs_double tmpP, MTRand &tmpRandomGenerator)`
- `acs_double acsround (acs_double tmpX)`
- `string dec2bin (acs_int tmpInt)`
- `bool ExitWithError (string strFunctionName, string strError)`
- `vector< string > split (string str, const char *delim)`
- `bool fromStrToBool (string const &string)`

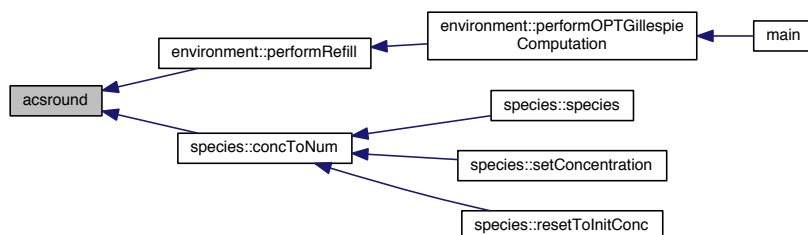
13.53.1 Function Documentation

13.53.1.1 `acs_double acsround (acs_double tmpX)`

Function to round double numbers in integers

Definition at line 229 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.53.1.2 string dec2bin (*acs_int tmpInt*)

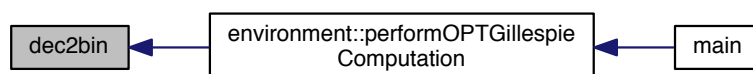
Function to convert a decimal number in a binary string composed of 12 bit such Example -> input: 10, binary 1010 -> 000000001010

Definition at line 241 of file commonFunctions.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



13.53.1.3 bool ExitWithError (string *strFunctionName*, string *strError*)

Function to close the program after having en error

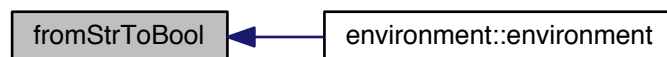
Definition at line 269 of file commonFunctions.cpp.

13.53.1.4 bool fromStrToBool (string const & *string*)

Function to convert from string to boolean

Definition at line 302 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.53.1.5 `acs_double getDoubleRandom (acs_double tmpFromNum, acs_double tmpToNum, MTRand & tmpRandomGenerator)`

Definition at line 151 of file commonFunctions.cpp.

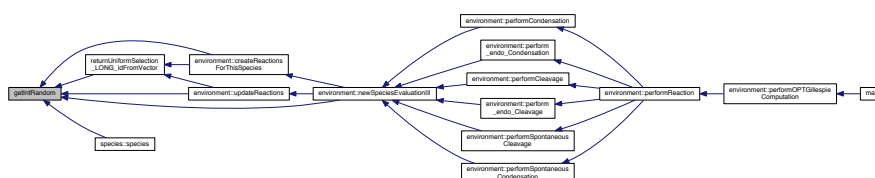
Here is the caller graph for this function:



13.53.1.6 `acs_longInt getIntRandom (acs_longInt tmpFromNum, acs_longInt tmpToNum, MTRand & tmpRandomGenerator)`

Definition at line 131 of file commonFunctions.cpp.

Here is the caller graph for this function:

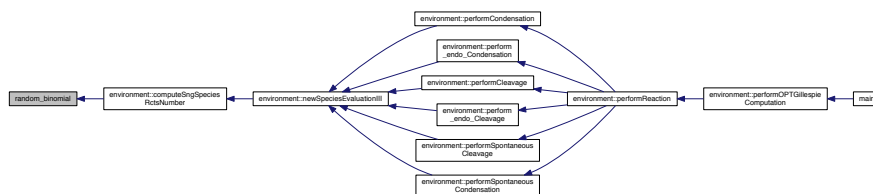


13.53.1.7 `acs_longInt random_binomial (acs_longInt n, acs_double tmpP, MTRand & tmpRandomGenerator)`

Function to return a number from a binomial distribution

Definition at line 203 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.53.1.8 `acs_longInt random_poisson (acs_double tmpLambda, MTRand & tmpRandomGenerator)`

Function to return a number from a poisson random distribution

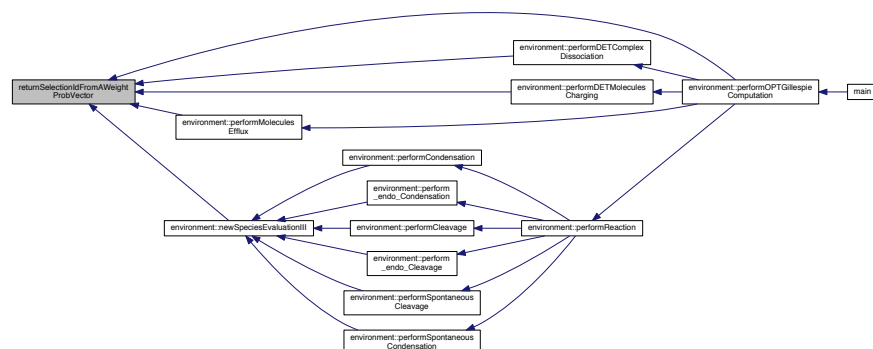
Definition at line 183 of file commonFunctions.cpp.

13.53.1.9 `acs_longInt returnSelectionIdFromAWeightProbVector (acs_double * tmpArray, MTRand & tmpRandomGenerator, acs_int tmpRow)`

This funtion returns a random position in a probability weight array of N elements.

Definition at line 15 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.53.1.10 `acs_longInt returnSelectionIdFromAWeightProbVector (vector< acs_double > & tmpVector, acs_double tmpMaxValue, MTRand & tmpRandomGenerator, acs_int tmpRow)`

Return position of a randomly selected element from a vector containing cumulative values for each element

Version

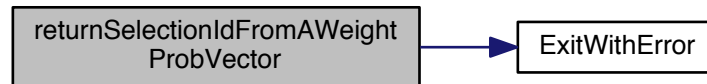
1.0

Parameters

<i>vector<acs_ - double> &</i>	tmpVector
<i>acs_double</i>	MAX VALUE contained within the QList (being a cumulative list this is the last value)
<i>MTRand&</i>	tmpRandomGenerator

Definition at line 40 of file commonFunctions.cpp.

Here is the call graph for this function:



13.53.1.11 **acs_longInt** returnSelectionIdFromAWeightProbVectorAlreadyNormalized (**vector< acs_double > & tmpVector**, **MTRand** & **tmpRandomGenerator**)

Return position of a randomly selected element from a normalized vector containing cumulative values for each element

Version

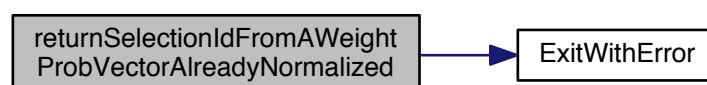
1.0

Parameters

<i>vector<acs_ - double> &</i>	tmpQList
<i>MTRand&</i>	tmpRandomGenerator

Definition at line 72 of file commonFunctions.cpp.

Here is the call graph for this function:

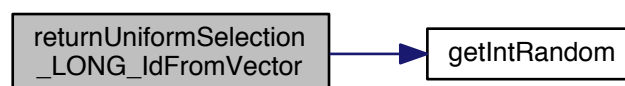


Return position of a LONG INT randomly selected element from a normalized vector containing cumulative values for each element

1.0

<i>vector<acs_-double>&</i>	tmpVector
<i>MTRand&</i>	tmpRandomGenerator

Here is the call graph for this function:

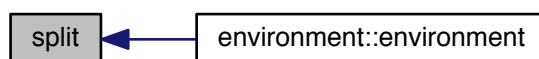


13.53.1.13 `vector<string> split (string str, const char * delim)`

Function to split a string and save tokens in a vector

Definition at line 279 of file `commonFunctions.cpp`.

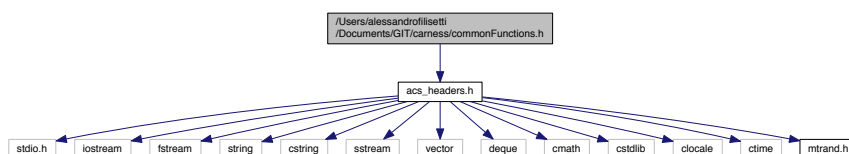
Here is the caller graph for this function:



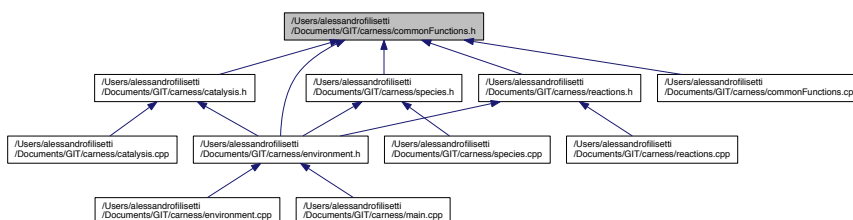
13.54 `/Users/alessandrofilisetti/Documents/GIT/carness/commonFunctions.h` File Reference

```
#include "acs_headers.h"
```

Include dependency graph for `commonFunctions.h`:



This graph shows which files directly or indirectly include this file:



Functions

- `acs_longInt returnSelectionIdFromAWeightProbVector (acs_double *tmpArray, MTRand &tmpRandomGenerator, acs_int tmpRow=0)`
This funtion returns a random position in a probability weight array of N elements.
- `acs_longInt returnSelectionIdFromAWeightProbVector (vector< acs_double > &tmpVector, acs_double tmpMaxValue, MTRand &tmpRandomGenerator, acs_int tmpRow=0)`
- `acs_longInt returnSelectionIdFromAWeightProbVectorAlreadyNormalized (vector< acs_double > &tmpVector, MTRand &tmpRandomGenerator)`

- [acs_longInt returnUniformSelection_LONG_IdFromVector](#) (vector< [acs_longInt](#) > &tmpVector, [MTRand](#) &tmpRandomGenerator)
- [acs_longInt getIntRandom](#) ([acs_longInt](#) tmpFromNum, [acs_longInt](#) tmpToNum, [MTRand](#) &tmpRandomGenerator)
- [acs_double getDoubleRandom](#) ([acs_double](#) tmpFromNum, [acs_double](#) tmpToNum, [MTRand](#) &tmpRandomGenerator)
- [acs_longInt random_poisson](#) ([acs_double](#) tmpLambda, [MTRand](#) &tmpRandomGenerator)
- [acs_longInt random_binomial](#) ([acs_longInt](#) n, [acs_double](#) tmpP, [MTRand](#) &tmpRandomGenerator)
- [acs_double acsround](#) ([acs_double](#) tmpX)
- string [dec2bin](#) ([acs_int](#) tmpInt)
- vector< string > [split](#) (string str, const char *delim)
- bool [ExitWithError](#) (string strFunctionName, string strError)
- bool [fromStrToBool](#) (string const &string)

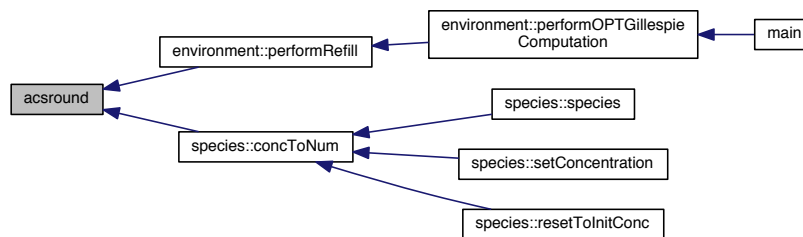
13.54.1 Function Documentation

13.54.1.1 [acs_double acsround \(\[acs_double\]\(#\) tmpX \)](#)

Function to round double numbers in integers

Definition at line 229 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.54.1.2 [string dec2bin \(\[acs_int\]\(#\) tmpInt \)](#)

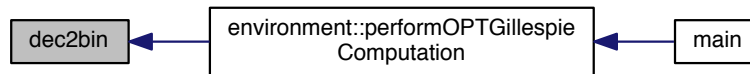
Function to convert a decimal number in a binary string composed of 12 bit such Example -> input: 10, binary 1010 -> 000000001010

Definition at line 241 of file commonFunctions.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



13.54.1.3 `bool ExitWithError (string strFunctionName, string strError)`

Function to close the program after having an error

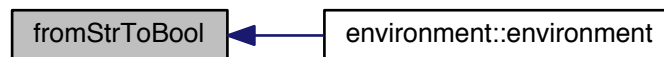
Definition at line 269 of file `commonFunctions.cpp`.

13.54.1.4 `bool fromStrToBool (string const & string)`

Function to convert from string to boolean

Definition at line 302 of file `commonFunctions.cpp`.

Here is the caller graph for this function:



13.54.1.5 `acs_double getDoubleRandom (acs_double tmpFromNum, acs_double tmpToNum, MTRand & tmpRandomGenerator)`

Definition at line 151 of file `commonFunctions.cpp`.

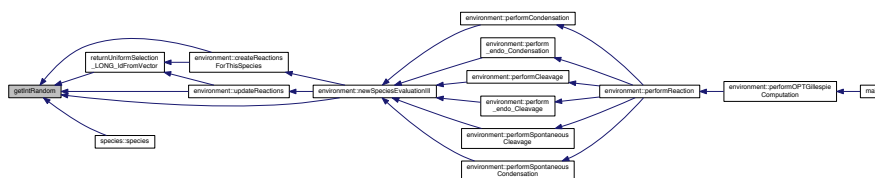
Here is the caller graph for this function:



13.54.1.6 `acs_longInt getIntRandom (acs_longInt tmpFromNum, acs_longInt tmpToNum, MTRand & tmpRandomGenerator)`

Definition at line 131 of file commonFunctions.cpp.

Here is the caller graph for this function:

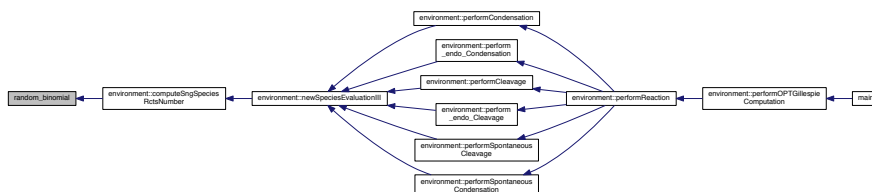


13.54.1.7 `acs_longInt random_binomial (acs_longInt n, acs_double tmpP, MTRand & tmpRandomGenerator)`

Function to return a number from a binomial distribution

Definition at line 203 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.54.1.8 `acs_longInt random_poisson (acs_double tmpLambda, MTRand & tmpRandomGenerator)`

Function to return a number from a poisson random distribution

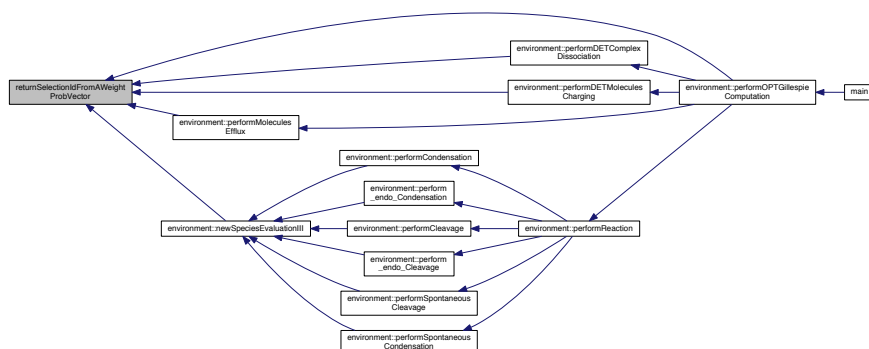
Definition at line 183 of file commonFunctions.cpp.

13.54.1.9 `acs_longInt returnSelectionIdFromAWeightProbVector (acs_double * tmpArray, MTRand & tmpRandomGenerator, acs_int tmpRow = 0)`

This funtion returns a random position in a probability weight array of N elements.

Definition at line 15 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.54.1.10 `acs_longInt returnSelectionIdFromAWeightProbVector (vector< acs_double > & tmpVector, acs_double tmpMaxValue, MTRand & tmpRandomGenerator, acs_int tmpRow)`

Return position of a randomly selected element from a vector containing cumulative values for each element

Version

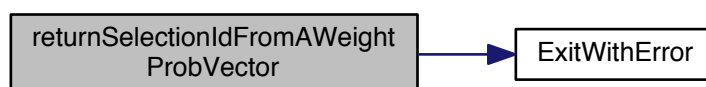
1.0

Parameters

<code>vector< acs_double > &</code>	tmpVector
<code>acs_double</code>	MAX VALUE contained within the QList (being a cumulative list this is the last value)
<code>MTRand &</code>	tmpRandomGenerator

Definition at line 40 of file commonFunctions.cpp.

Here is the call graph for this function:



13.54.1.11 `acs_longInt returnSelectionIdFromAWeightProbVectorAlreadyNormalized (vector< acs_double > & tmpVector, MTRand & tmpRandomGenerator)`

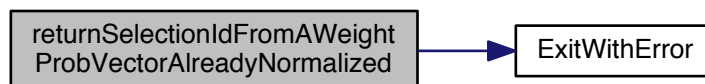
Return position of a randomly selected element from a normalized vector containing cumulative values for each element

Version

1.0

<i>vector<acs_ - double>&</i>	tmpQList
<i>MTRand&</i>	tmpRandomGenerator

Here is the call graph for this function:

[illegible]

Return position of a LONG INT randomly selected element from a normalized vector containing cumulative values for each element

Version

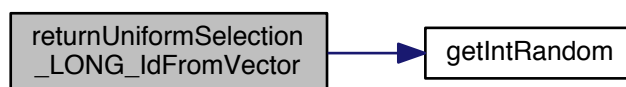
1.0

Parameters

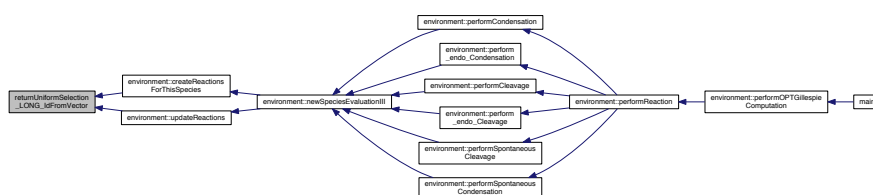
<code>vector<acs_ - double>&</code>	tmpVector
<code>MTRand&</code>	tmpRandomGenerator

Definition at line 124 of file commonFunctions.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:

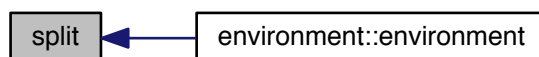


13.54.1.13 `vector<string> split (string str, const char * delim)`

Function to split a string and save tokens in a vector

Definition at line 279 of file commonFunctions.cpp.

Here is the caller graph for this function:



13.56 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/commonFunctions.d File Reference

13.57 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/environment.d File Reference

13.58 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/gillespie.d File Reference

13.59 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/main.d File Reference

13.60 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/mtrand.d File Reference

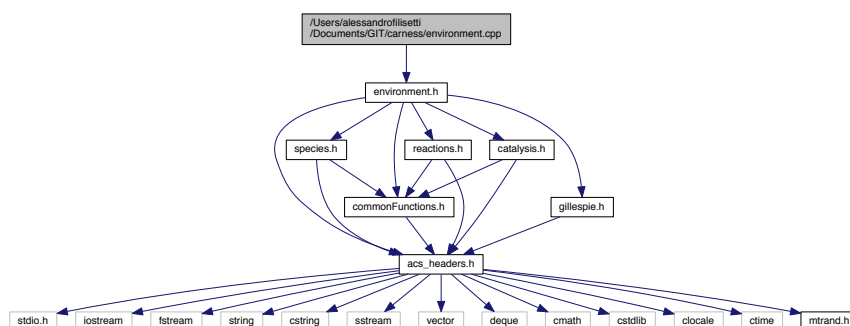
13.61 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/reactions.d File Reference

13.62 /Users/alessandrofilisetti/Documents/GIT/carness/Debug/species.d File Reference

13.63 /Users/alessandrofilisetti/Documents/GIT/carness/environment.cpp File Reference

```
#include "environment.h"
```

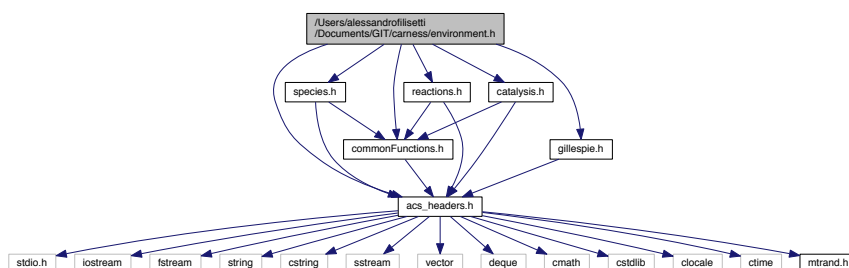
Include dependency graph for environment.cpp:



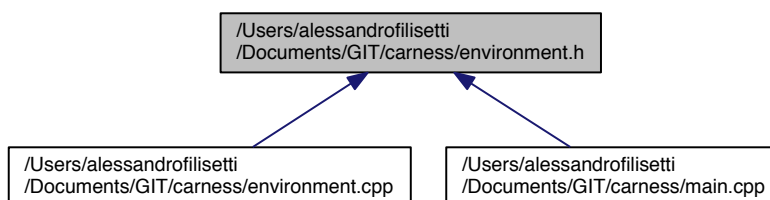
13.64 /Users/alessandrofilisetti/Documents/GIT/carness/environment.h File Reference

```
#include "acs_headers.h"
#include "species.h"
#include "reactions.h"
#include "commonFunctions.h"
#include "catalysis.h"
#include "gillespie.h"
```

Include dependency graph for environment.h:



This graph shows which files directly or indirectly include this file:



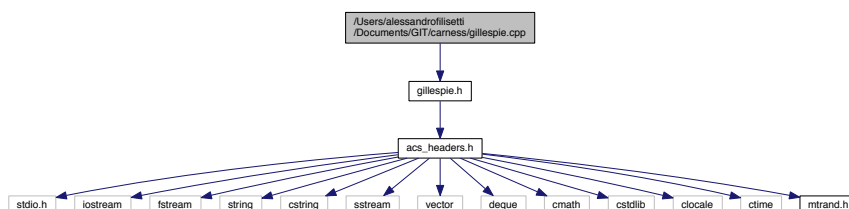
Classes

- class [environment](#)
environment class

13.65 /Users/alessandrofilisetti/Documents/GIT/carness/gillespie.cpp File Reference

```
#include "gillespie.h"
```

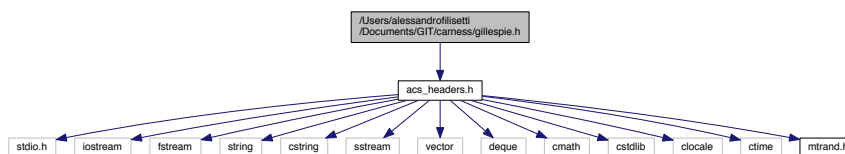
Include dependency graph for gillespie.cpp:



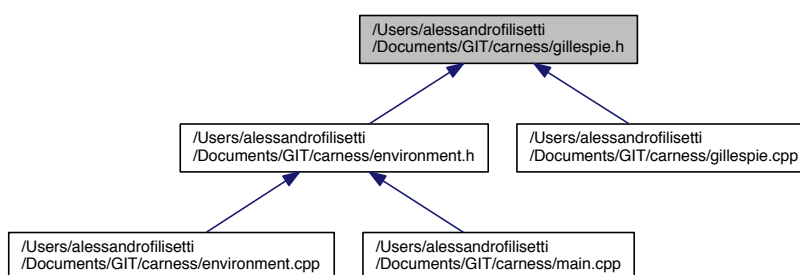
13.66 /Users/alessandrofilisetti/Documents/GIT/carness/gillespie.h File Reference

```
#include "acs_headers.h"
```

Include dependency graph for gillespie.h:



This graph shows which files directly or indirectly include this file:



Classes

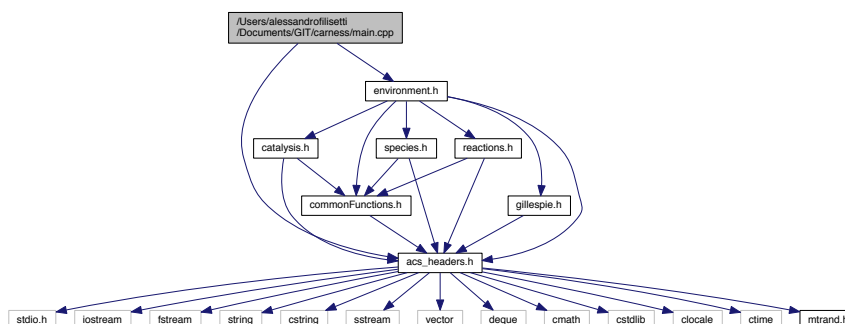
- class [gillespie](#)

13.67 /Users/alessandrofilisetti/Documents/GIT/carness/main.cpp File Reference

```
#include "acs_headers.h"
```

```
#include "environment.h"
```

Include dependency graph for main.cpp:



Functions

- void `saveToFile` (string tmpSavingPath, `environment` *tmpEnvironment, `acs_int` tmpGen, `acs_int` tmpSim, `acs_int` tmpStep)
- void `saveTimesToFile` (string tmpSavingPath, `environment` *tmpEnvironment, `acs_int` tmpGen, `acs_int` tmpSim, `acs_int` tmpStep)
- void `saveInitialConditionsToFile` (string tmpSavingPath, `environment` *tmpEnvironment, `acs_int` tmpGen, `acs_int` tmpSim, `acs_int` tmpStep)
- int `main` (int argc, char *argv[])

13.67.1 Function Documentation

13.67.1.1 int main (int *argc*, char * *argv*[])

double random number generator

Definition at line 354 of file main.cpp.

13.67.1.2 void saveInitialConditionsToFile (string *tmpSavingPath*, `environment` * *tmpEnvironment*, `acs_int` *tmpGen*, `acs_int` *tmpSim*, `acs_int` *tmpStep*)

Save to file all the INITIAL structures

Version

2.0

Parameters

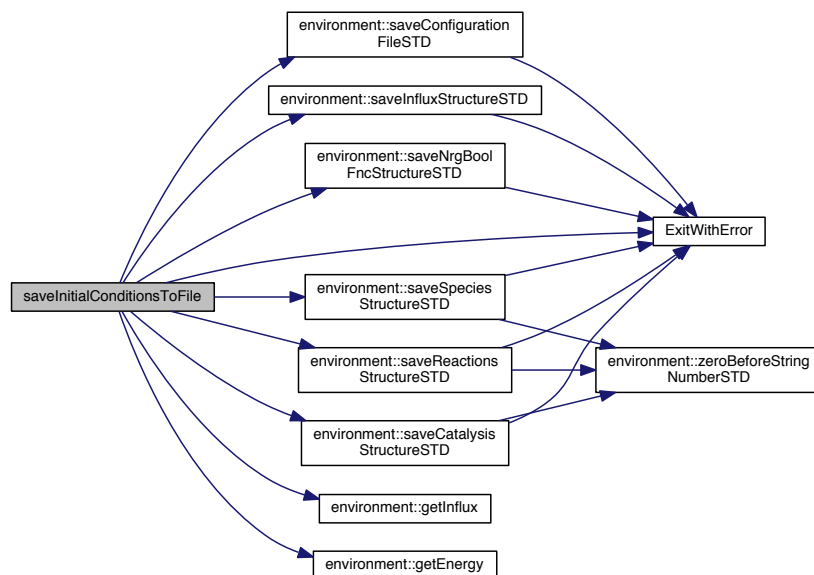
<i>string</i>	tmpSavingPath Saving files path
<i>environment</i>	*tmpEnvironment environment instance reference
<i>tmpSim</i>	Current simulation
<i>acs_int</i>	Current step

Date

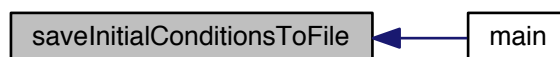
2013/07/03

Definition at line 669 of file main.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



13.67.1.3 `void saveTimesToFile (string tmpSavingPath, environment * tmpEnvironment, acs_int tmpGen, acs_int tmpSim, acs_int tmpStep)`

Save TIMES to file

Version

2.0

Parameters

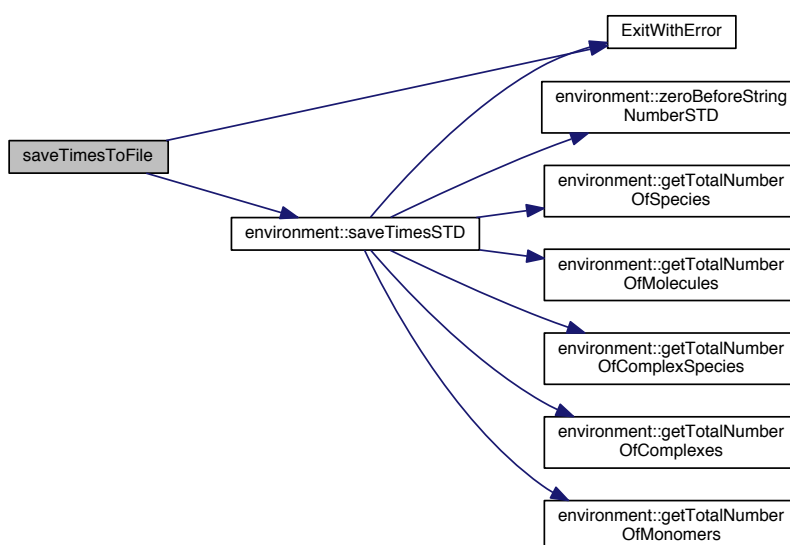
<i>string</i>	tmpSavingPath Saving files path
<i>environment</i>	*tmpEnvironment environment instance reference
<i>tmpSim</i>	Current simulation
<i>acs_int</i>	Current step

Date

2013/07/03

Definition at line 653 of file main.cpp.

Here is the call graph for this function:



Here is the caller graph for this function:



13.67.1.4 void saveToFile (string tmpSavingPath, environment * tmpEnvironment, acs_int tmpGen, acs_int tmpSim, acs_int tmpStep)

Save to file structures at step tmpStep

Version

2.0

Parameters

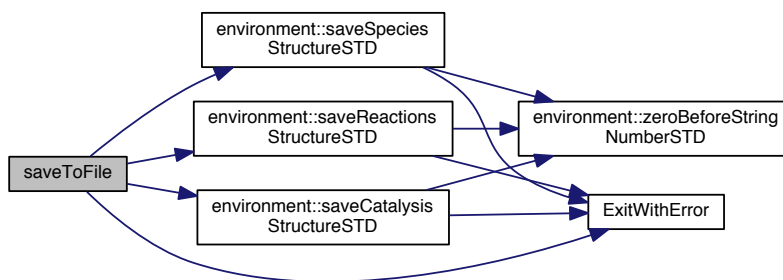
<i>string</i>	tmpSavingPath Saving files path
<i>environment</i>	*tmpEnvironment environment instance reference
<i>tmpSim</i>	Current simulation
<i>acs_int</i>	Current step

Date

2013/07/13

Definition at line 633 of file main.cpp.

Here is the call graph for this function:



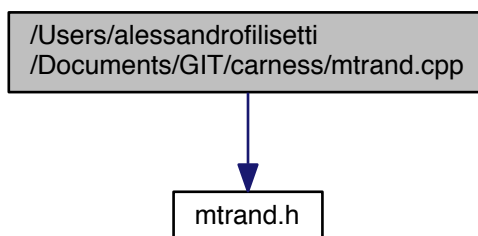
Here is the caller graph for this function:



13.68 /Users/alessandrofilisetti/Documents/GIT/carness/mtrand.cpp File Reference

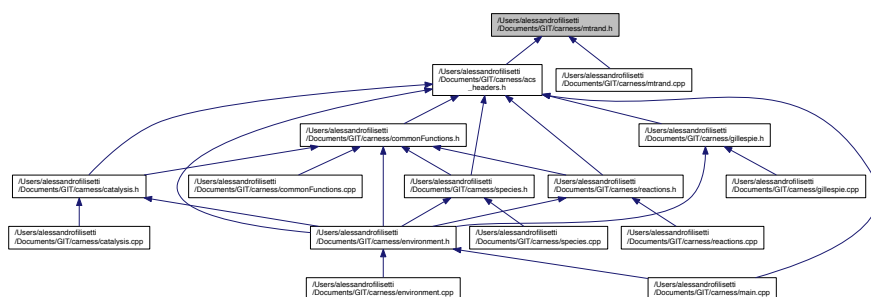
```
#include "mtrand.h"
```

Include dependency graph for mtrand.cpp:



13.69 /Users/alessandrofilisetti/Documents/GIT/carness/mtrand.h File Reference

This graph shows which files directly or indirectly include this file:



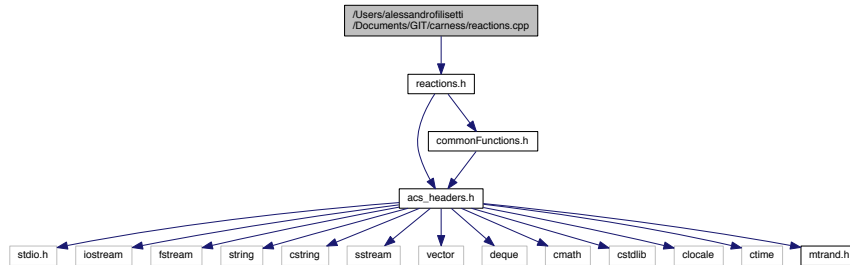
Classes

- class [MTRand_int32](#)
- class [MTRand](#)
- class [MTRand_closed](#)
- class [MTRand_open](#)
- class [MTRand53](#)

13.70 /Users/alessandrofilisetti/Documents/GIT/carness/reactions.cpp File Reference

```
#include "reactions.h"
```

Include dependency graph for reactions.cpp:

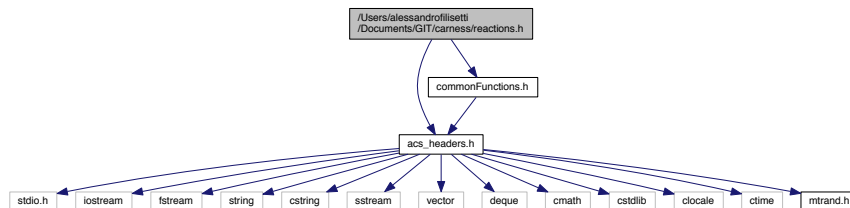


13.71 /Users/alessandrofilisetti/Documents/GIT/carness/reactions.h File Reference

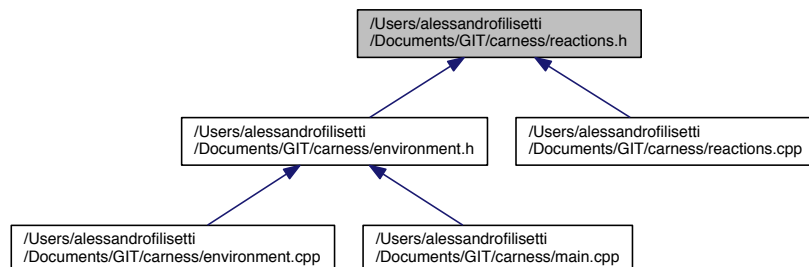
```
#include "acs_headers.h"
```

```
#include "commonFunctions.h"
```

Include dependency graph for reactions.h:



This graph shows which files directly or indirectly include this file:



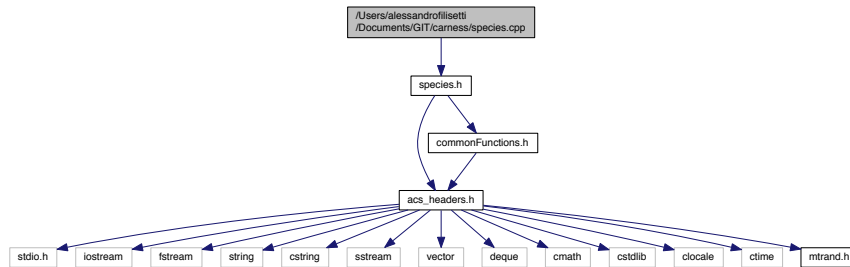
Classes

- class [reactions](#)

13.72 /Users/alessandrofilisetti/Documents/GIT/carness/species.cpp File Reference

```
#include "species.h"
```

Include dependency graph for species.cpp:

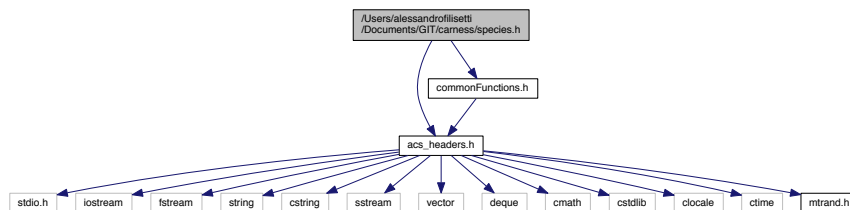


13.73 /Users/alessandrofilisetti/Documents/GIT/carness/species.h File Reference

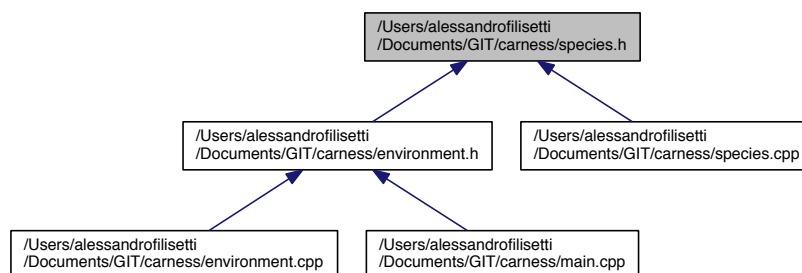
```
#include "acs_headers.h"
```

```
#include "commonFunctions.h"
```

Include dependency graph for species.h:



This graph shows which files directly or indirectly include this file:



Classes

- class [species](#)

This class contains declarations of the species class.