Directory path for the original fastq files INPUT DIR= # Directory path for the outputs of the analysis OUTPUT DIR= OUTPUT NAME= # Name for analysis. Choose a single name without spaces LIST SAMPLES= # File directory for a list with samples and individual id, separated by space. Use LIST SAMPLES=none if no list is available. ANALYSIS= # ANALYSIS=1 (for WGS) or ANALYSIS=2 (for RNAseq) EXTENSION SE= # For single-end fastg files, write the extension of original fastg files. If paired- end files, le" blank (EXTENSION SE=). EXTENSION PE1= # For paired-end fastq files, write the extension of PE1 fastq files. If single-end files, le" blank (EXTENSION PE1=). EXTENSION PE2= # For paired-end fastq files, write the extension of PE2 fastq files. If single-end files, le" blank (EXTENSION PE2=). ASSEMBLY= # Give a name for the reference genome. Eg ASSEMBLY=hg38. # File directory for reference genome (*.fa). The reference genome must be uncompressed file and not (*.fa.gz). REFERENCE= # File directory for variants file (*.vcf.gz) for the reference genome. Use VARIANTS=none if no variants is available. **VARIANTS=** The variants must be (*.vcf.gz) and not (*.vcf). ANNOTATION= # File directory for annotation file (*.gtf) for the reference genome. Use ANNOTATION=none if no annotation is available. The annotation must be uncompressed file and not (*.gtf.gz)

```
# Activate quality control and adapter trimming step? STEP QC TRIM=yes or STEP QC TRIM=no (Default yes)
STEP QC TRIM=
STEP ALIGNMENT=
                       # Activate alignment step? STEP ALIGNMENT=yes or STEP ALIGNMENT=no (Default yes)
STEP GENECOUNT=
                       # Activate gene count step? STEP GENECOUNT=yes or STEP GENECOUNT=no (Default: yes for RNAseq
                        and no for WGS)
                       # Activate mark duplicated read step? STEP MDUP=yes or STEP MDUP=no (Default: yes). It includes
STEP MDUP=
                         splitncigar step for RNAseq
                       # Activate base quality score recalibration step? STEP BQSR=yes or STEP BQSR=no (Default: yes)
STEP BQSR=
STEP VCF CALL=
                       # Activate variant calling step? STEP VCF CALL=yes or STEP VCF CALL=no (Default: yes)
STEP VCF FILTER=
                       # Activate vcf filtering step? STEP VCF FILTER=yes or STEP VCF FILTER=no (Default: yes)
STEP VCF COMBINE=
                       # Activate combine filtered snps from different vcfs? STEP VCF COMBINE=yes or STEP VCF COMBINE=no
                         (Default: yes)
STEP VCF ANNOTATE= # Activate annotation of vcf files? STEP VCF ANNOTATE=yes or STEP VCF ANNOTATE=no (Default: yes)
```

ALIGNER PROGRAM= # Choose programs for alignment step. If more than one program are chosen, use ',' between them. (Options: bbmap, bowtie, bowtie2, bwa, bwa2, gsnap, hisat2, star) # Choose programs for variant calling step. If more than one program are chosen, use ',' between them. CALLER PROGRAM= (Options: bcftools, freebayes, freebayes-parallel, gatk4, gatk4GenomicsDBImport, gatk4CombineGVCFs, platypus, varscan) GENECOUNT PROGRAM= # Choose programs for gene count step. If more than one program are chosen, use ',' between them. (Options: htseq, featurecounts). Use GENECOUNT PROGRAM=none if you do not want gene counts. # Any feature of the third column of gtf file. (Default FEATURE TYPE=exon). Use FEATURE TYPE=none if FEATURE TYPE= you do not want gene counts. # Should variant calling be done by chromosome? CALL BY CHROM=no or CALL BY CHROM=yes (Default no). CALL BY CHROM= MIN READ LENGTH= # Minimum read length to be kept after adapter trimming (Default 36). MAX READ LENGTH= # Maximum read length to be kept after adapter trimming (Default 150). DOWN SAMPLING FASTQ= # Down sampling the fastq files. (Default 0). DOWN SAMPLING FASTQ=0 means no reads sampling from fastq files. Any number between 0.01 and 0.99 means to sampling the fraction of reads from fastq files. Any number equal or greater than 1 means to sampling the number of reads from fastg file. MIN DEPTH= # Minimum locus coverage for the filtering step (Default 3). # COMBINEVCF=none : do not combine different vcfs. COMBINE VCF= # COMBINEVCF=partial: If it was used more than one program for variant calling, combine common SNPs that appeared AT LEAST in two different vcf files. # COMBINEVCF=full: If it was used more than one program for variant calling, combine common SNPs that appeared IN ALL the vcf files. THREADS= # Number of threads for analysis. BATCH= # Number of samples to be processed at same time. # Keep the log files in the output folder? KEEP LOG=no or KEEP LOG=yes (Default no). KEEP LOG=

KEEP_INTERMEDIATE_DATA= # Keep intermediate fastq and bam files in the output folder? KEEP_INTERMEDIATE_DATA=no or KEEP_INTERMEDIATE_DATA=yes (Default no).