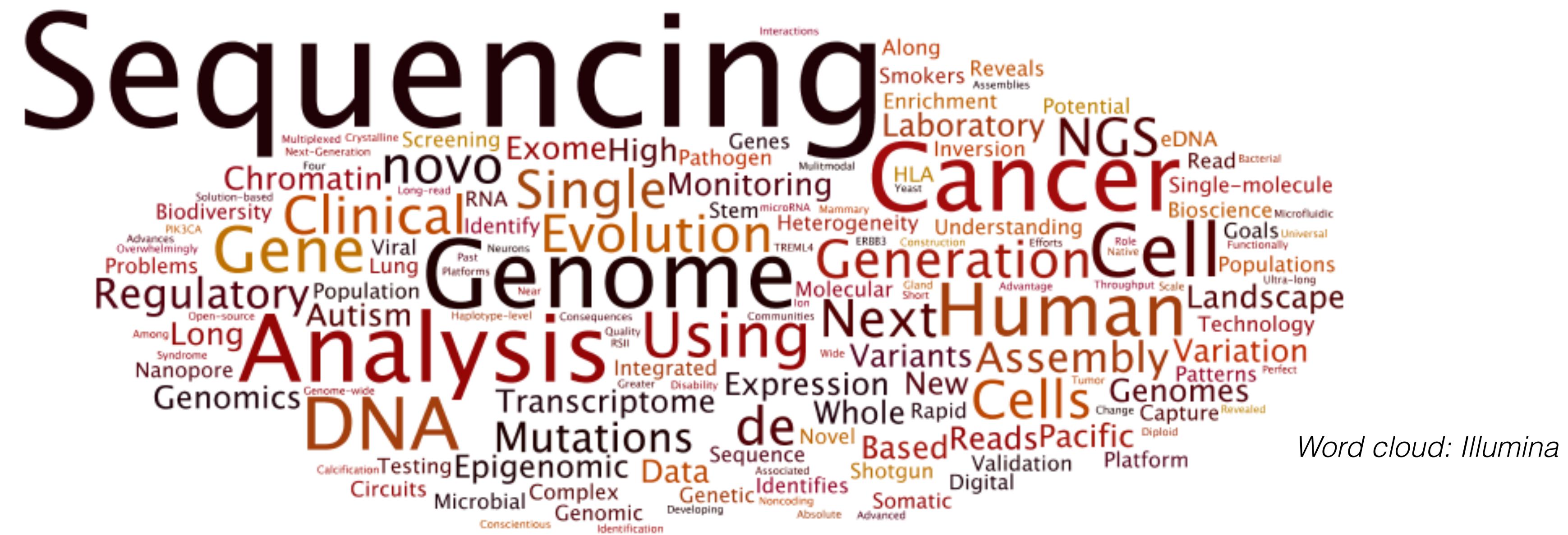


# An overview of genomics and sequencing terminology and practices

Mark Stenglein, GDW 2017



The jargon and terminology associated with genomics and ‘next gen’ sequencing can be confusing and intimidating



The goal of this lecture is to explain and demystify some common jargon  
and explain how sequencing works

There is a glossary available online that explains many of these terms

- [Transcriptome](#)
- [Variant](#)
- [WGS](#)

## 16S

The **16S** ribosomal RNA gene is present in all bacterial and archaeal genomes. This gene is sufficiently conserved that primers that anneal to conserved regions of the gene will amplify essentially any prokaryotic 16S rRNA gene. These PCR products (amplicons) can be sequenced to provide a survey of microbial diversity in a sample.

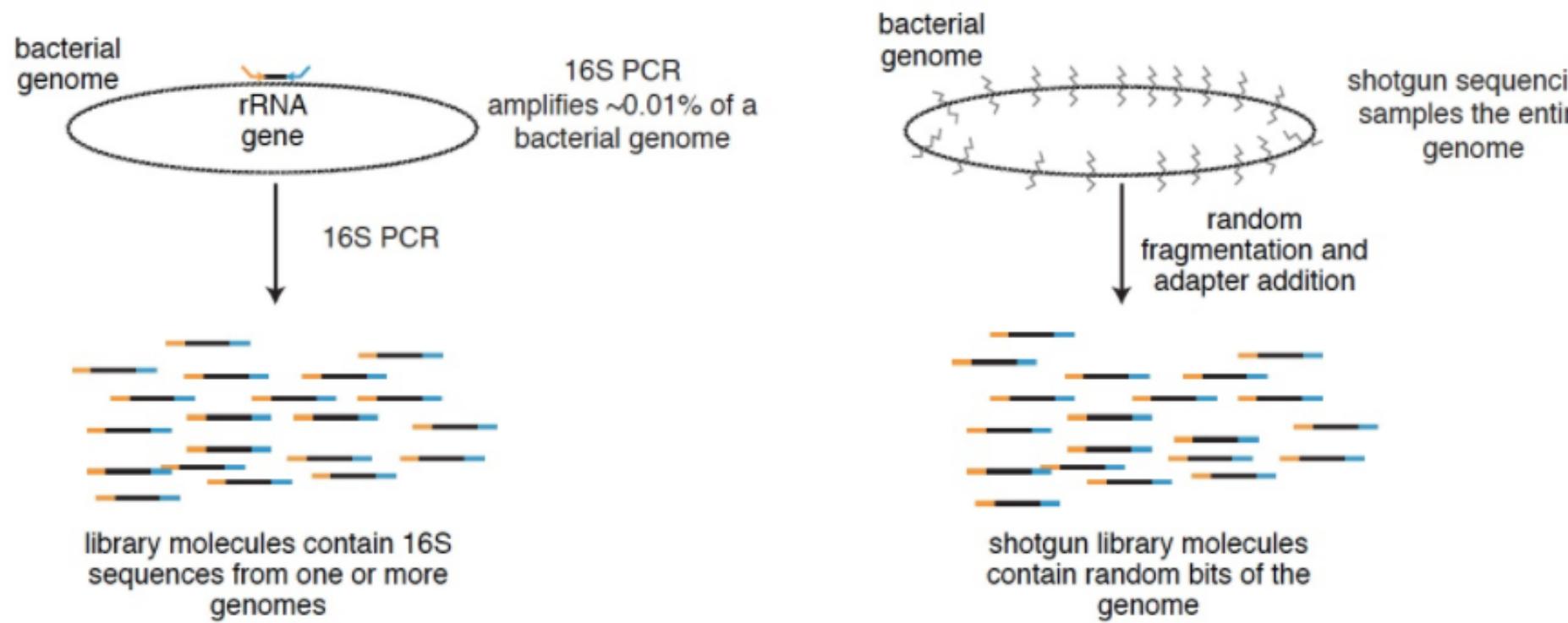


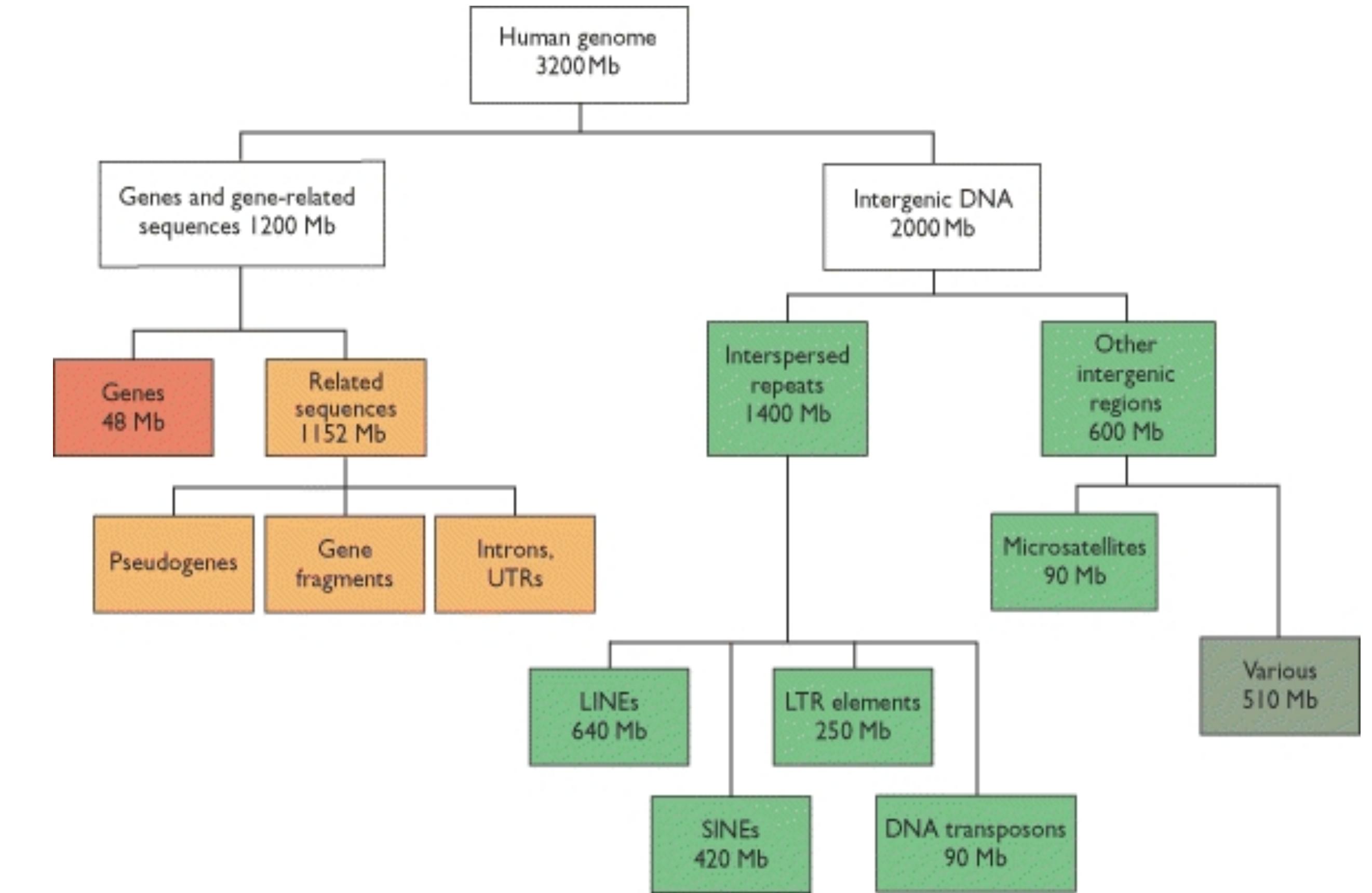
Figure: 16S vs. shotgun sequencing.

## Adapter

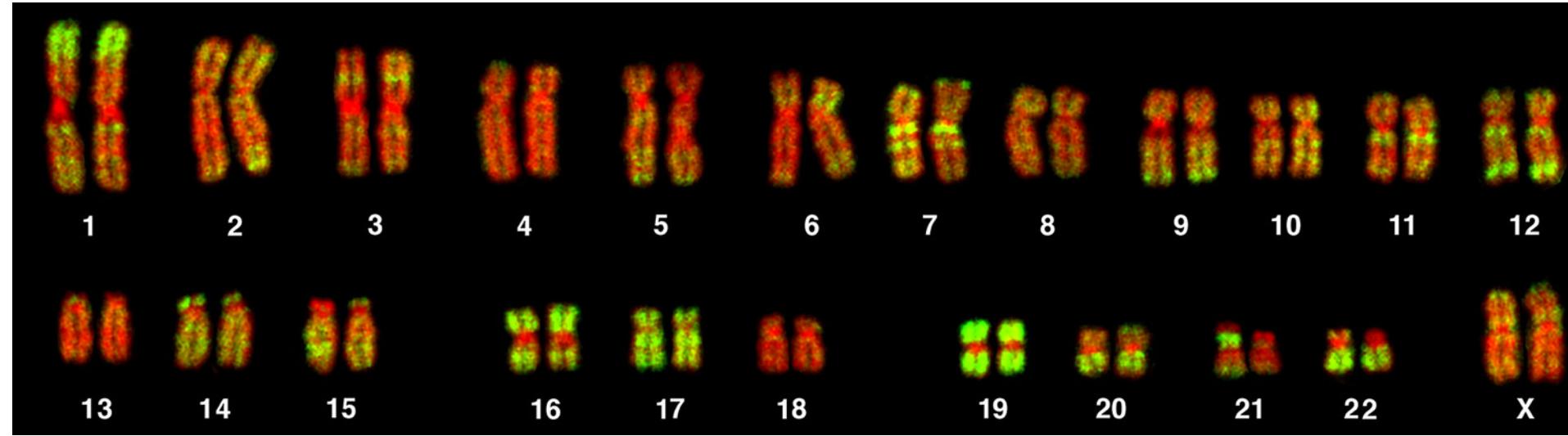
Most NGS instruments require that dsDNA of known sequence be added to the 2 ends of [library](#) molecules that will be sequenced on the instrument. Adapters can be added in a variety of ways to starting nucleic acid molecules during [library](#)

# Genomics is the study of any of a number of attributes of genome or genomes

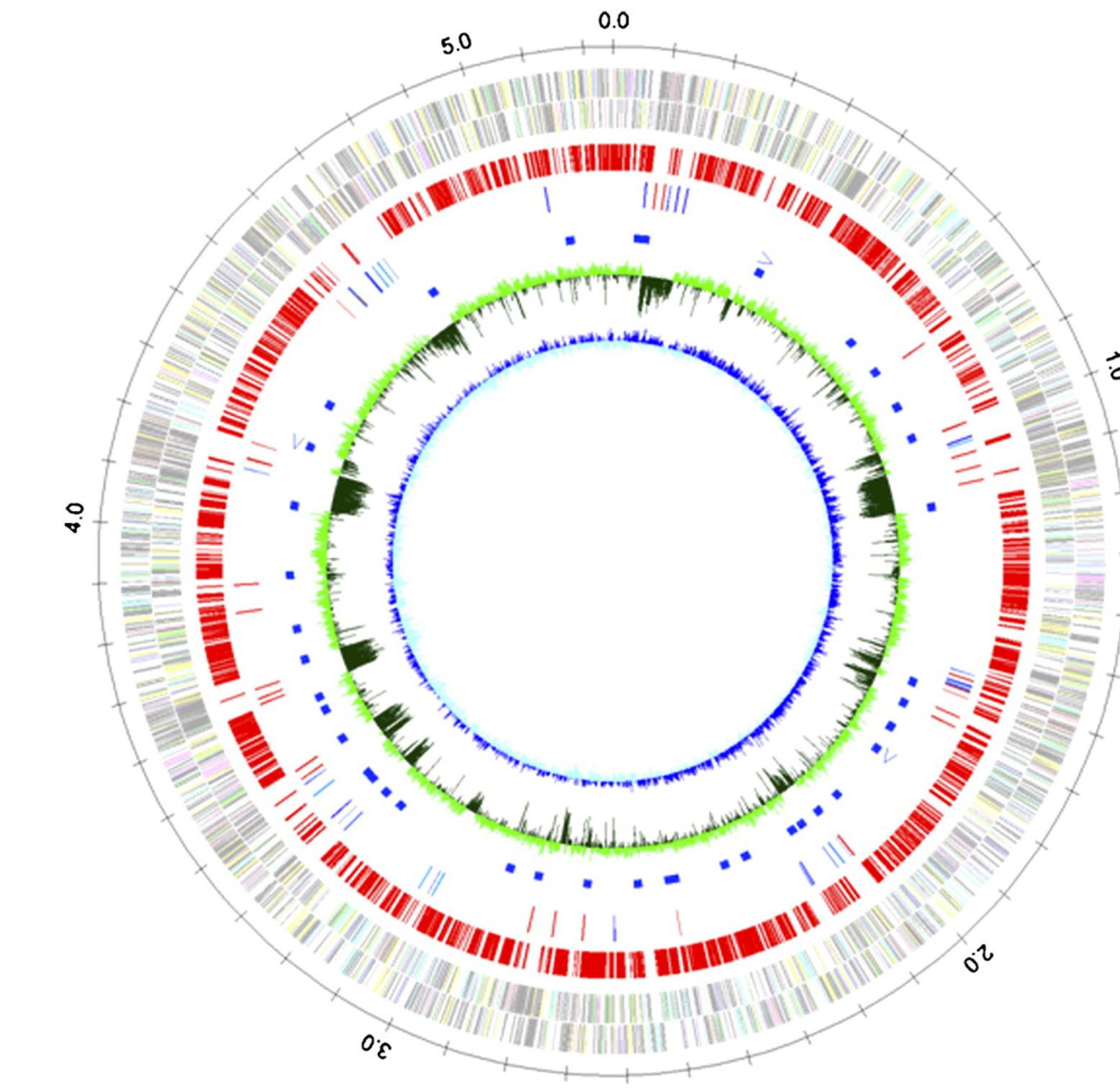
- Genome:
  - size
  - sequence
  - structure
  - evolution
- Gene:
  - structure
  - expression
- Comparative genomics
- Epigenomics
- Metagenomics
- Transcriptomics
- Other -omics: Proteomics/Metabolomics



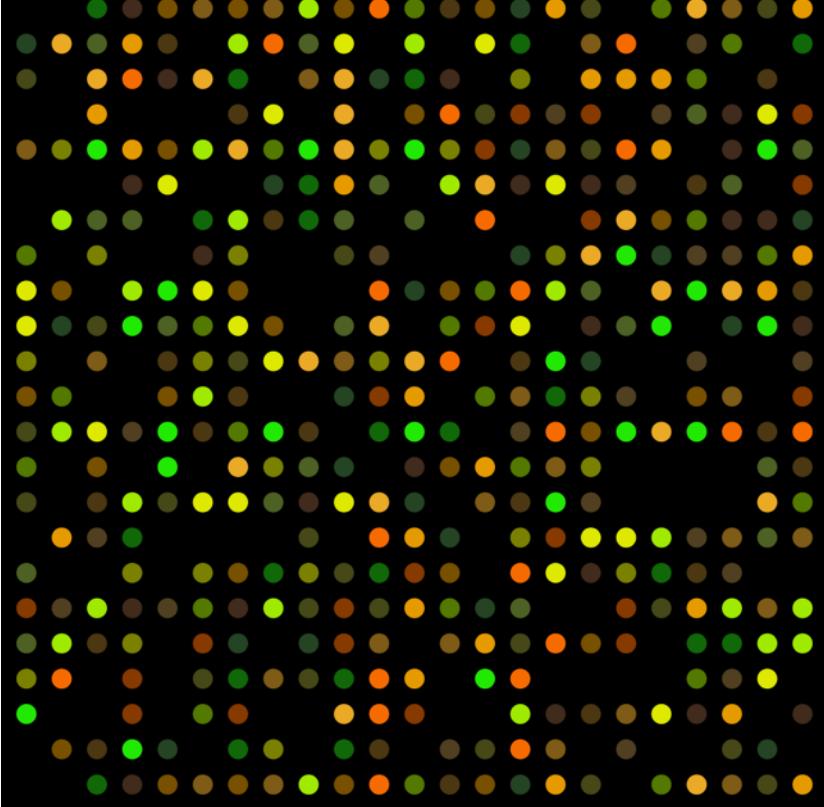
# Genomics isn't the same thing as sequencing, but they're increasingly related



Bolzer et al (2005) PLoS Biol

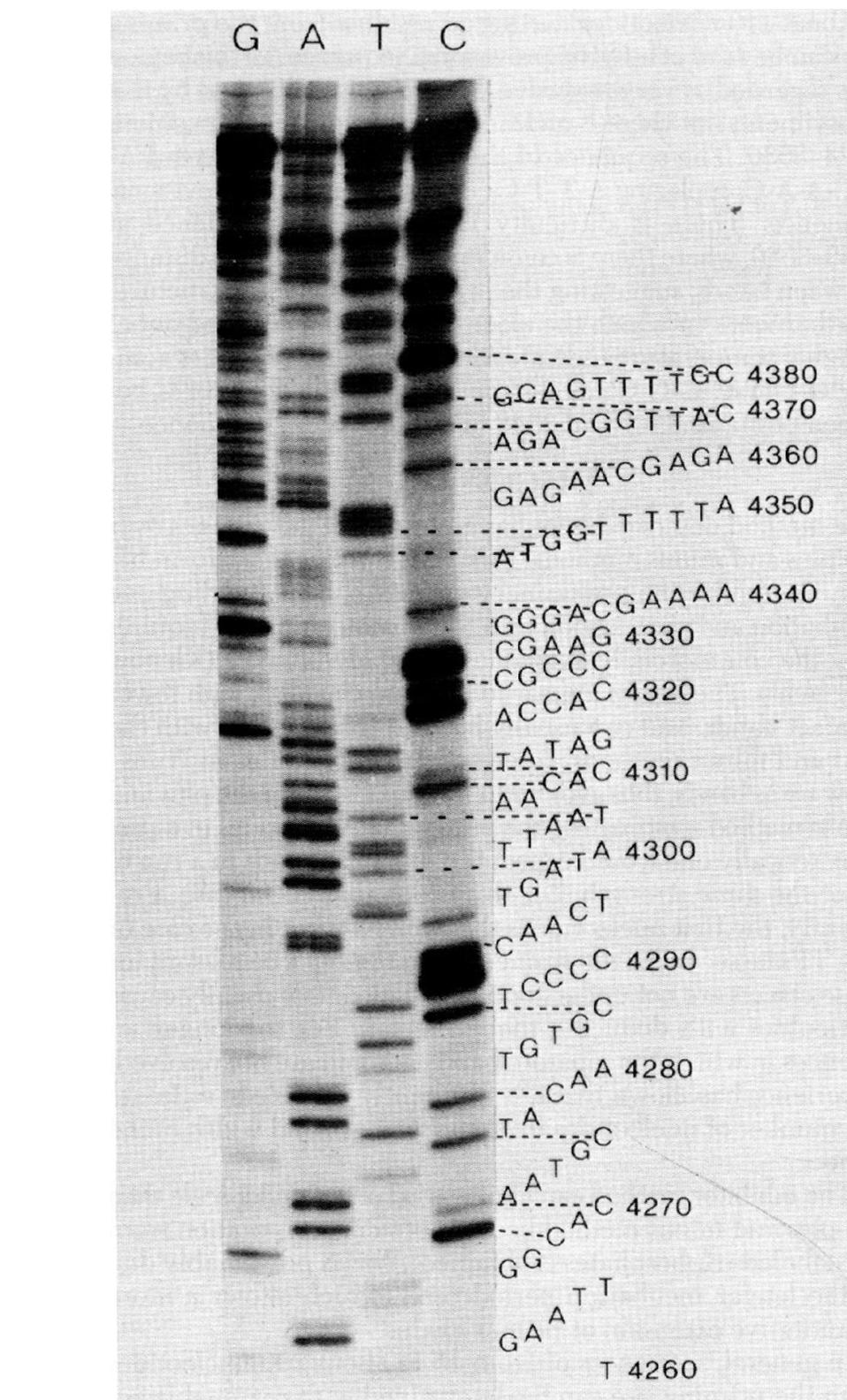
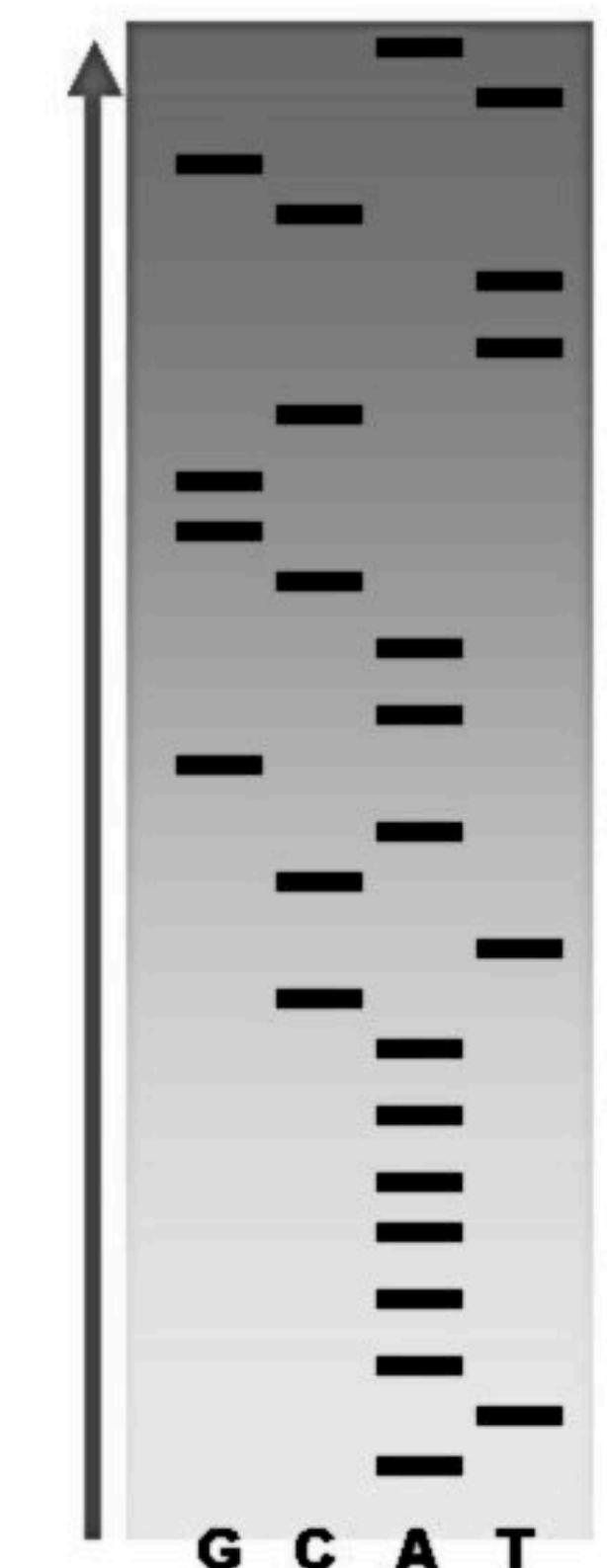
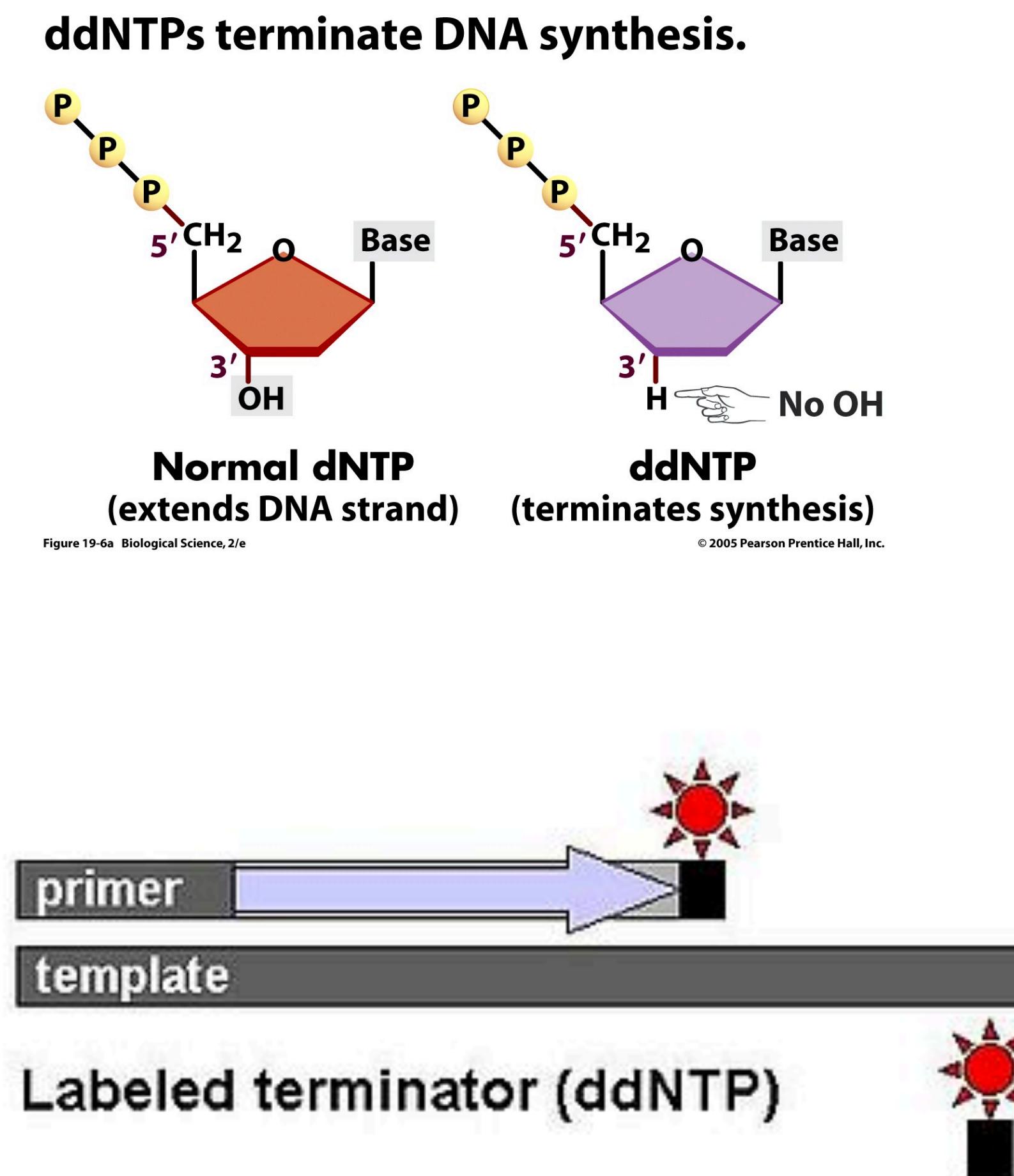


Nakazawa et al (2009) Genome Research



Wikimedia commons

# Sanger Sequencing (1977): sequencing 1 target at a time



*Slide courtesy Dan Sloan. Image credits: Sanger et al (1977) and Wikipedia*

# Improvements to Sanger sequencing and molecular methods allowed the sequencing of large genomes

1965 – First nucleic acid sequenced: Yeast trnA

1976 – First complete genome sequenced (RNA virus: bacteriophage MS2)

1977 – Maxam-Gilbert and Sanger DNA sequencing methods introduced and first complete DNA genome (Phage  $\Phi$ -X174)

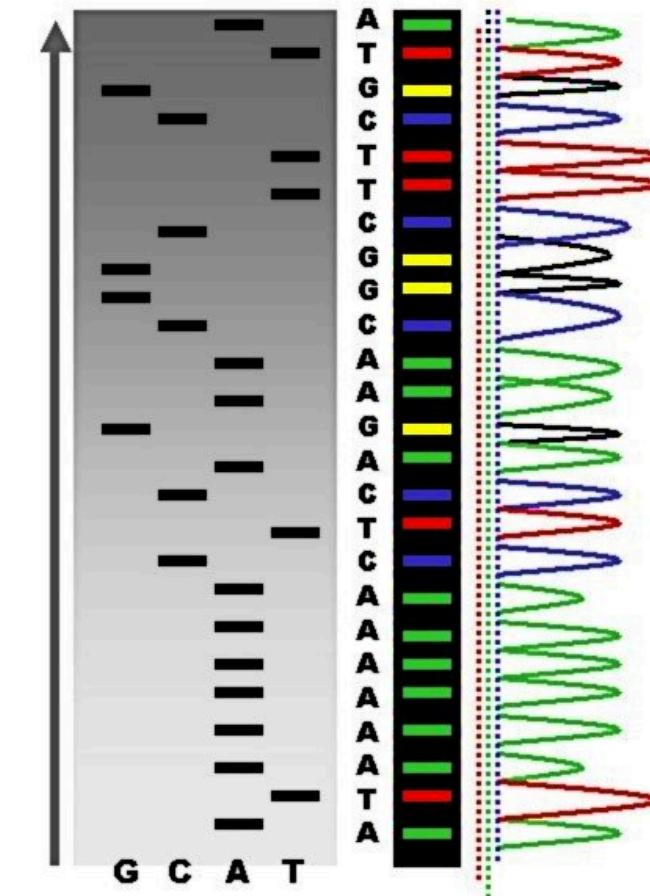
1983 – PCR introduced

1995 – First complete cellular genome (*Haemophilus influenzae*) and eukaryotic genome (yeast) sequenced

2001 – Publication of the first sequenced human genomes

2005 – Introduction of 454 Sequencing and the NGS Revolution

2017 – Genomics of Disease in Wildlife Workshop first offered



Slide courtesy Dan Sloan.

Next generation sequencing (NGS) ~ deep sequencing ~ high throughput sequencing (HTS)

All simultaneously sequence many molecules in parallel

## Short read sequencing

- Millions of reads
- Relatively short: ~50-300 nt (Illumina)
- Relative low error rates
- Illumina has virtually all of the market share



MiSeq

## Long read sequencing

- Fewer, longer reads
- >1 kb (PacBio), up to 100s of kb (Oxford Nanopore)
- Relative high error rates

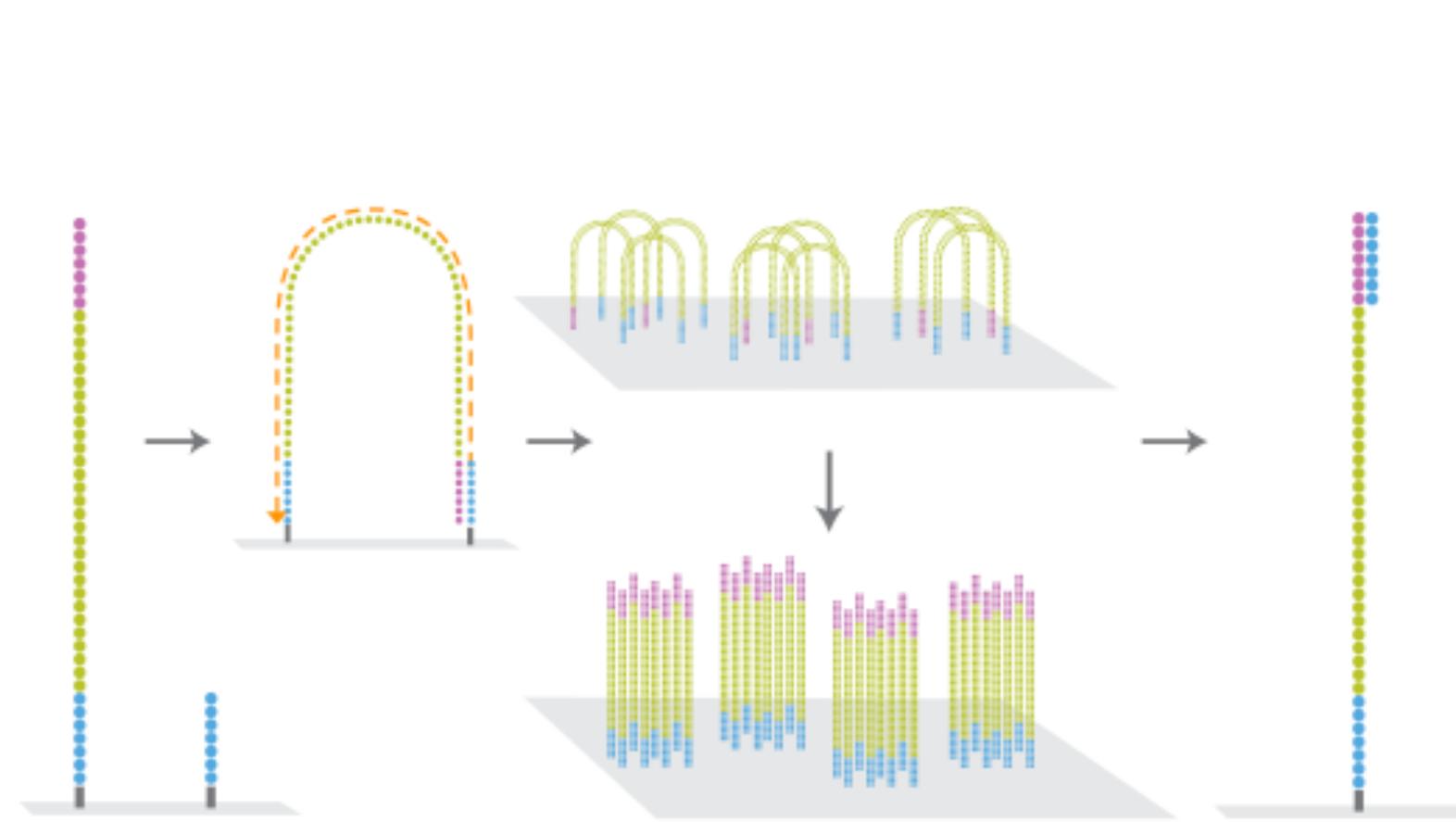
ONP MinION



PacBio RS-II

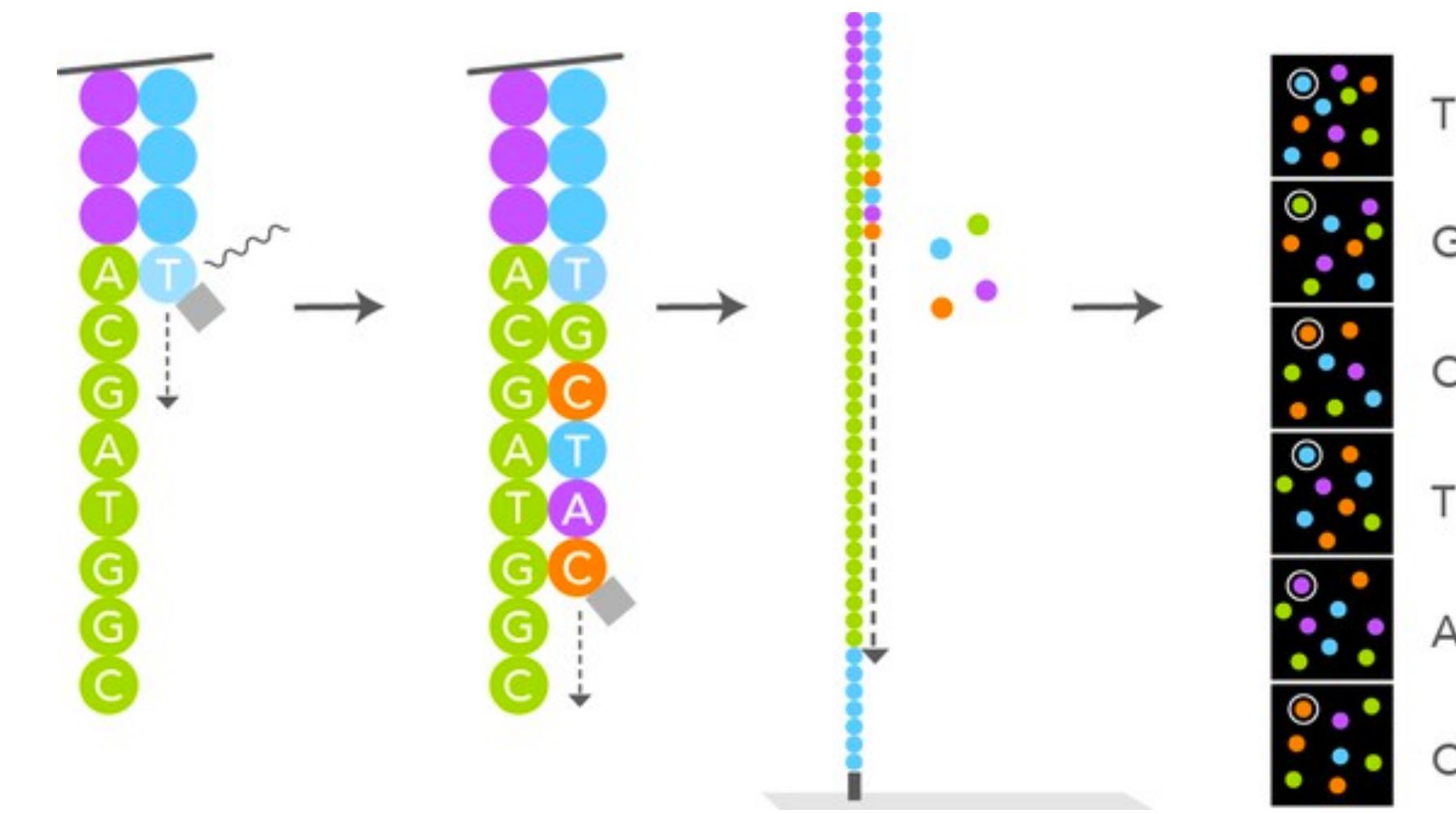


# Illumina instruments use sequencing by synthesis (SBS)



Millions of clusters per flow cell

Each cluster contains 1000s of clonal copies of a library molecule



Library molecules are sequenced by primer extension reactions that incorporate chain-terminated, fluorescent nucleotides

real raw Illumina sequencing data

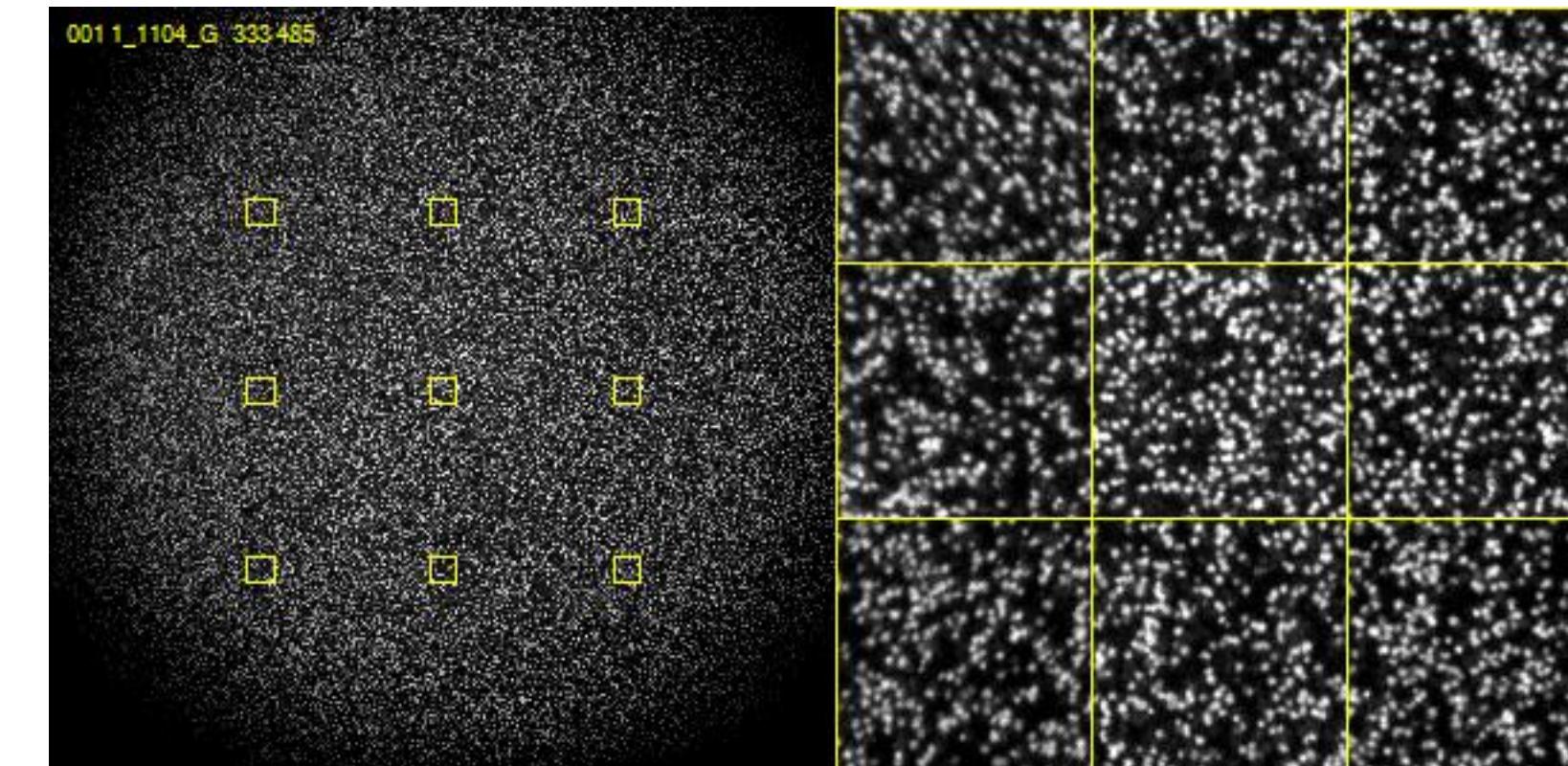
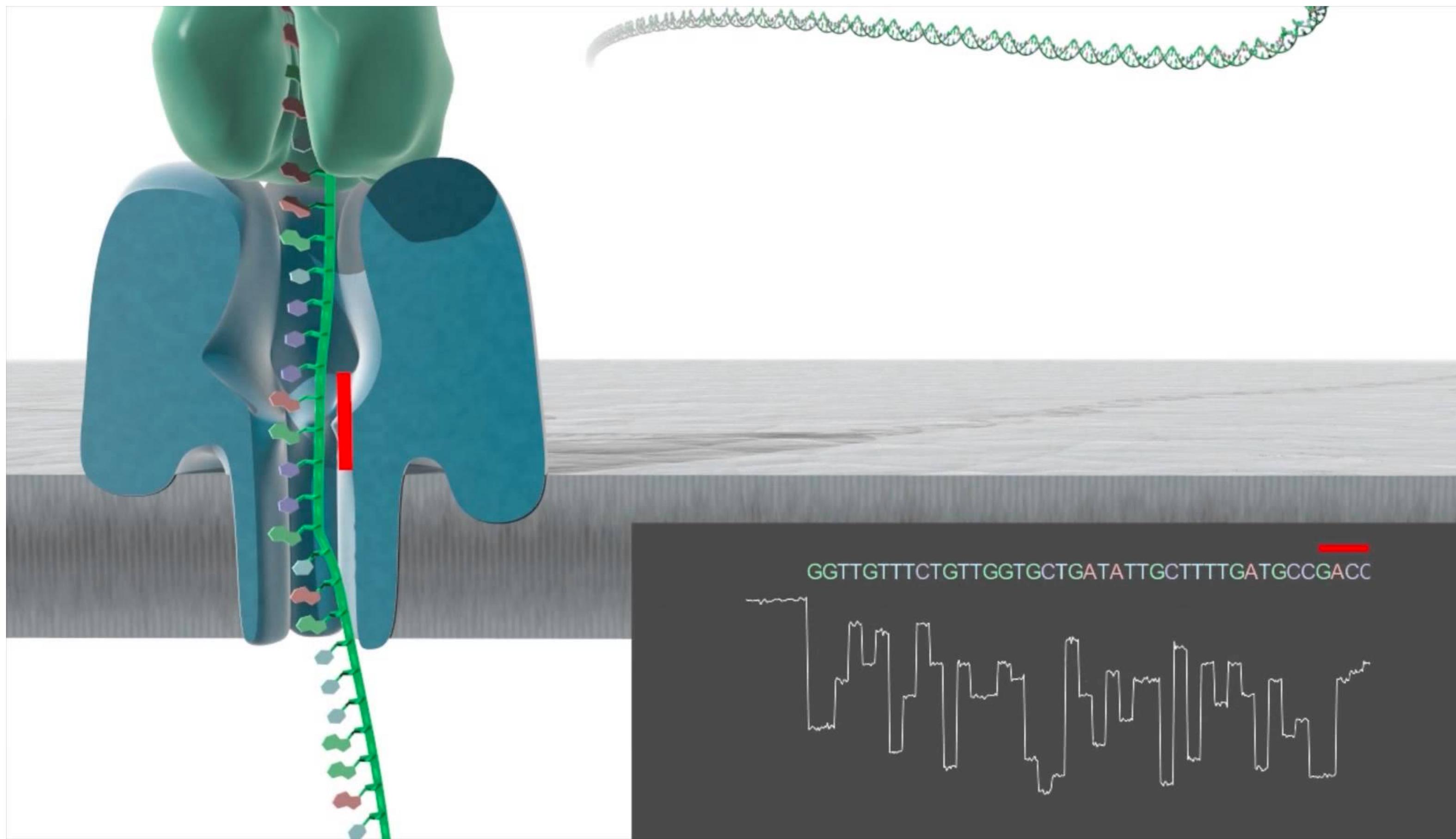


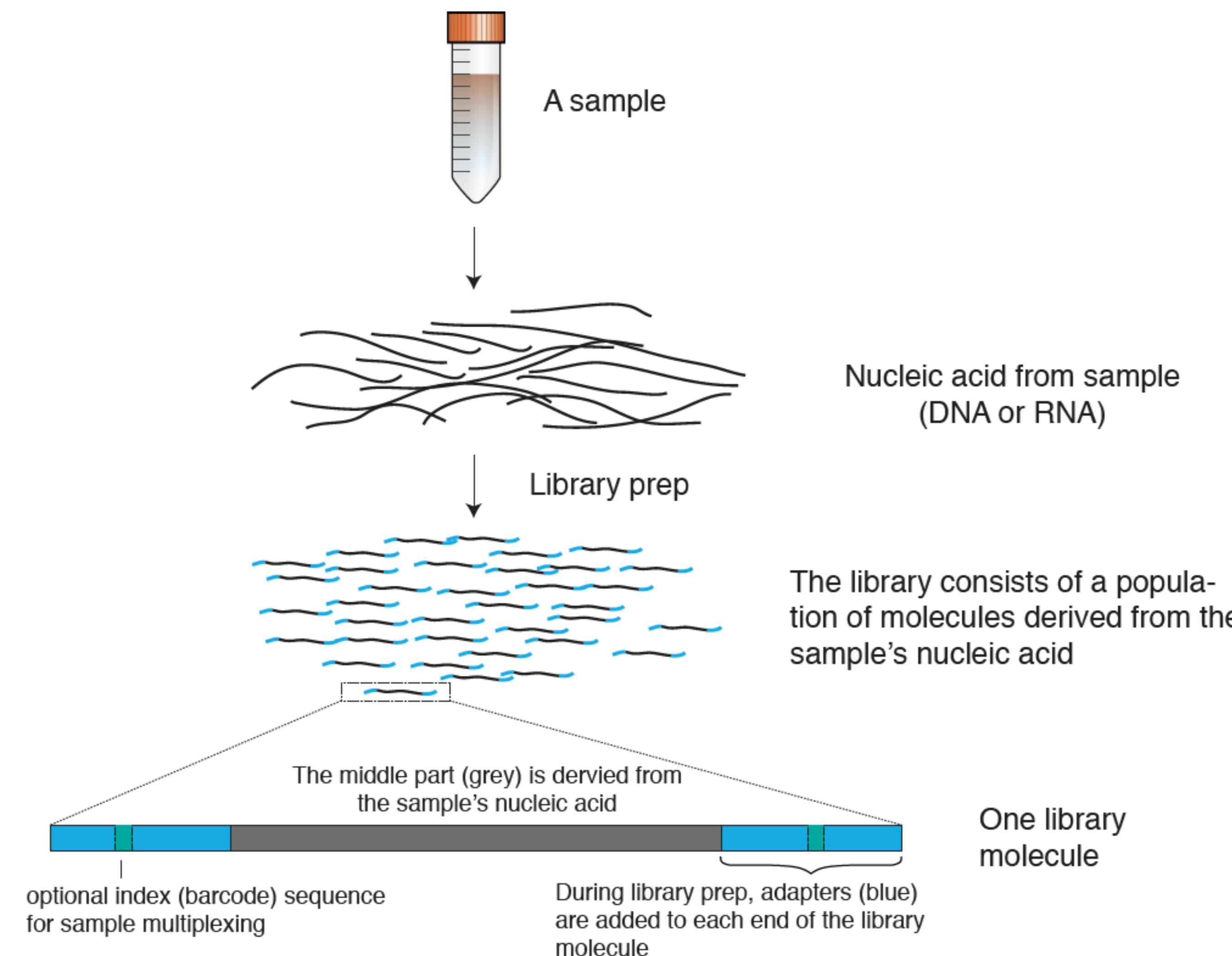
Image credit: Illumina

# Long read sequencers sequence single molecules

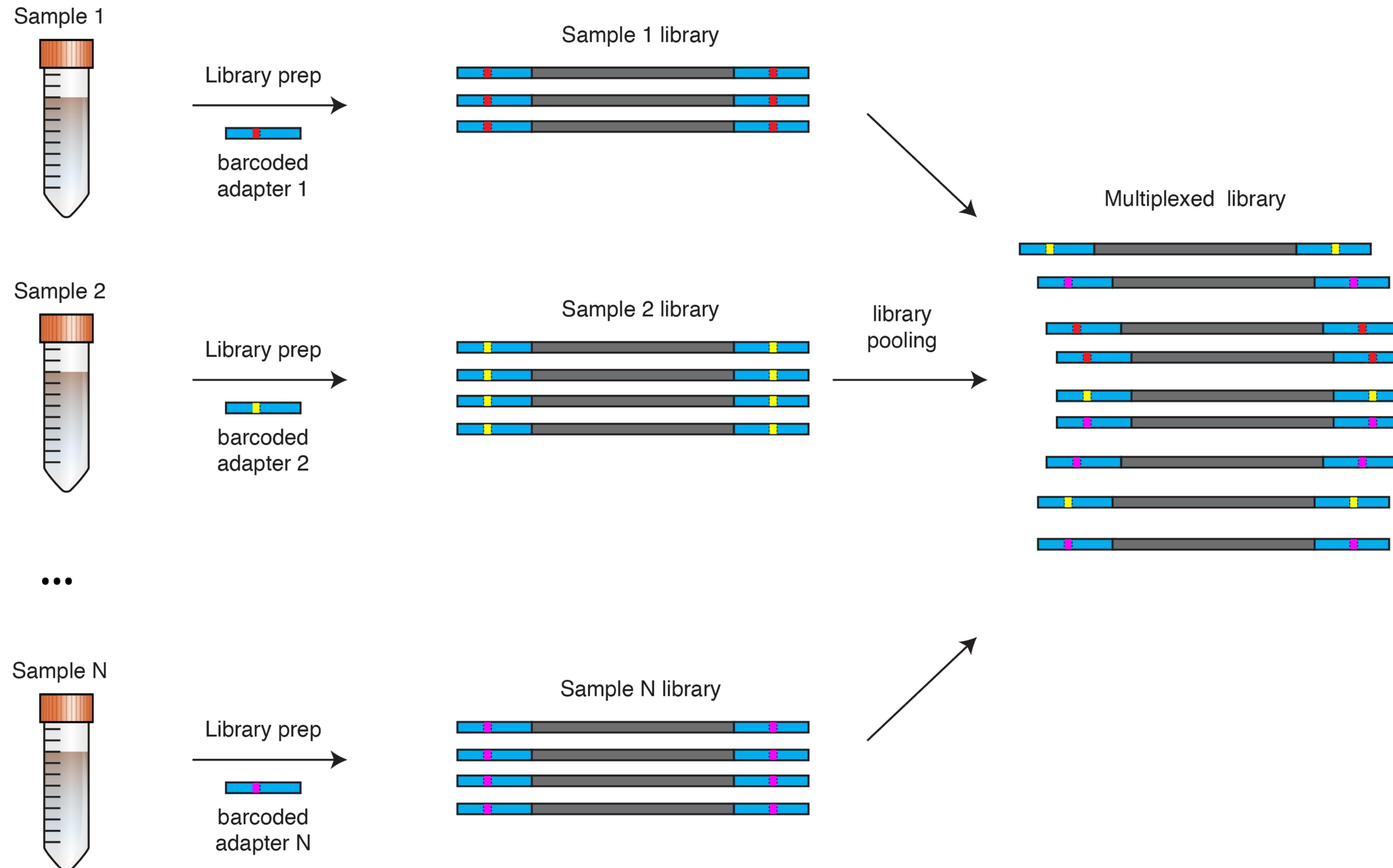


Much longer reads, but with much higher error rates

# Library preparation converts nucleic acids into a form suitable to be sequenced

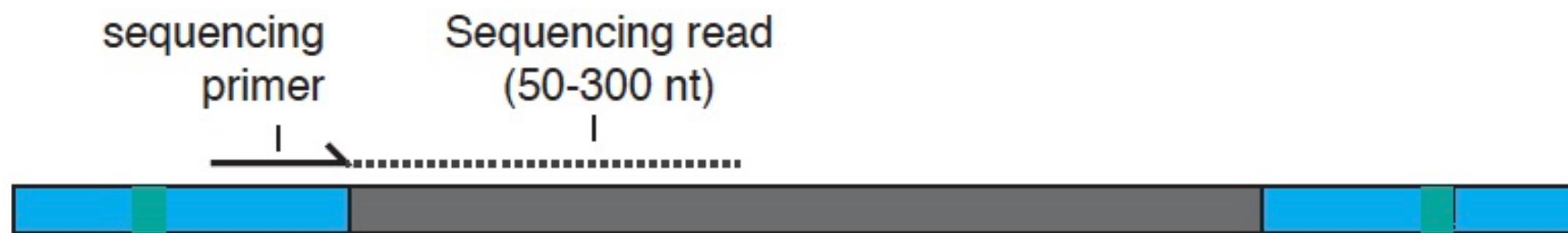


# Barcodes (or indexes) allow sample multiplexing



# Illumina sequencing produces 1-4 reads per library molecule

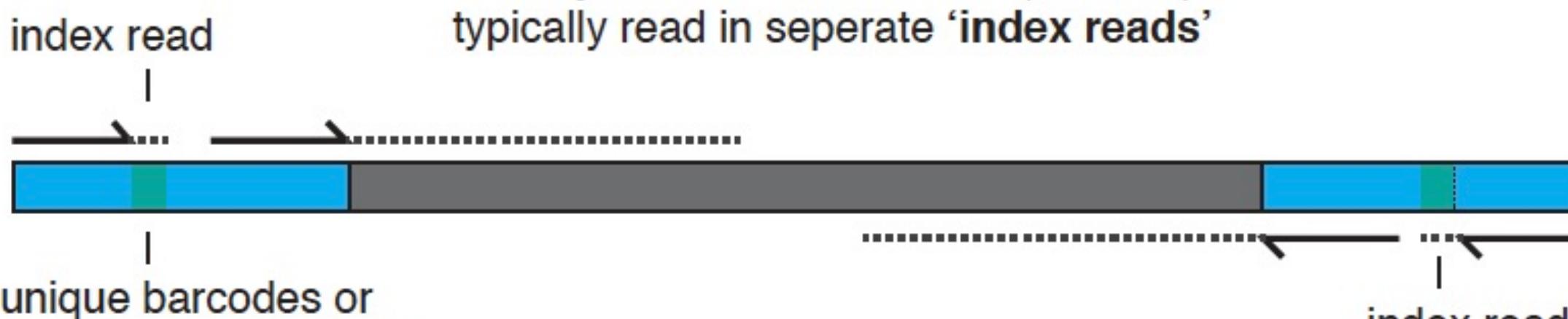
In **single end sequencing**, a library molecule is sequenced from one end



In **paired end sequencing**, a library molecule is sequenced from both ends



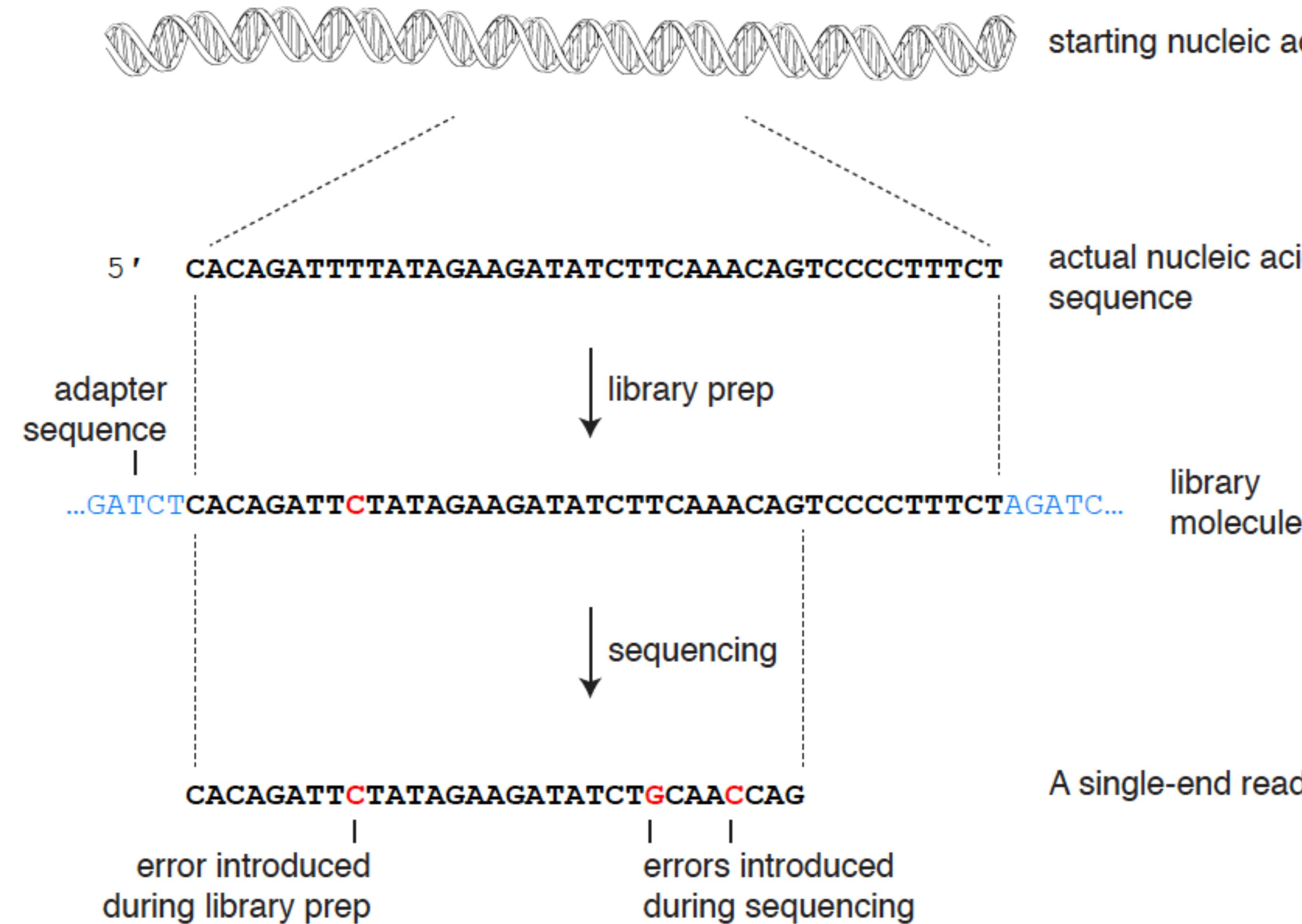
The library molecule's barcodes (indexes) are typically read in separate 'index reads'



unique barcodes or barcode pairs can be used to differentiate multiplexed samples

index read

Reads are sub-sequences of the starting nucleic acid that often contain errors

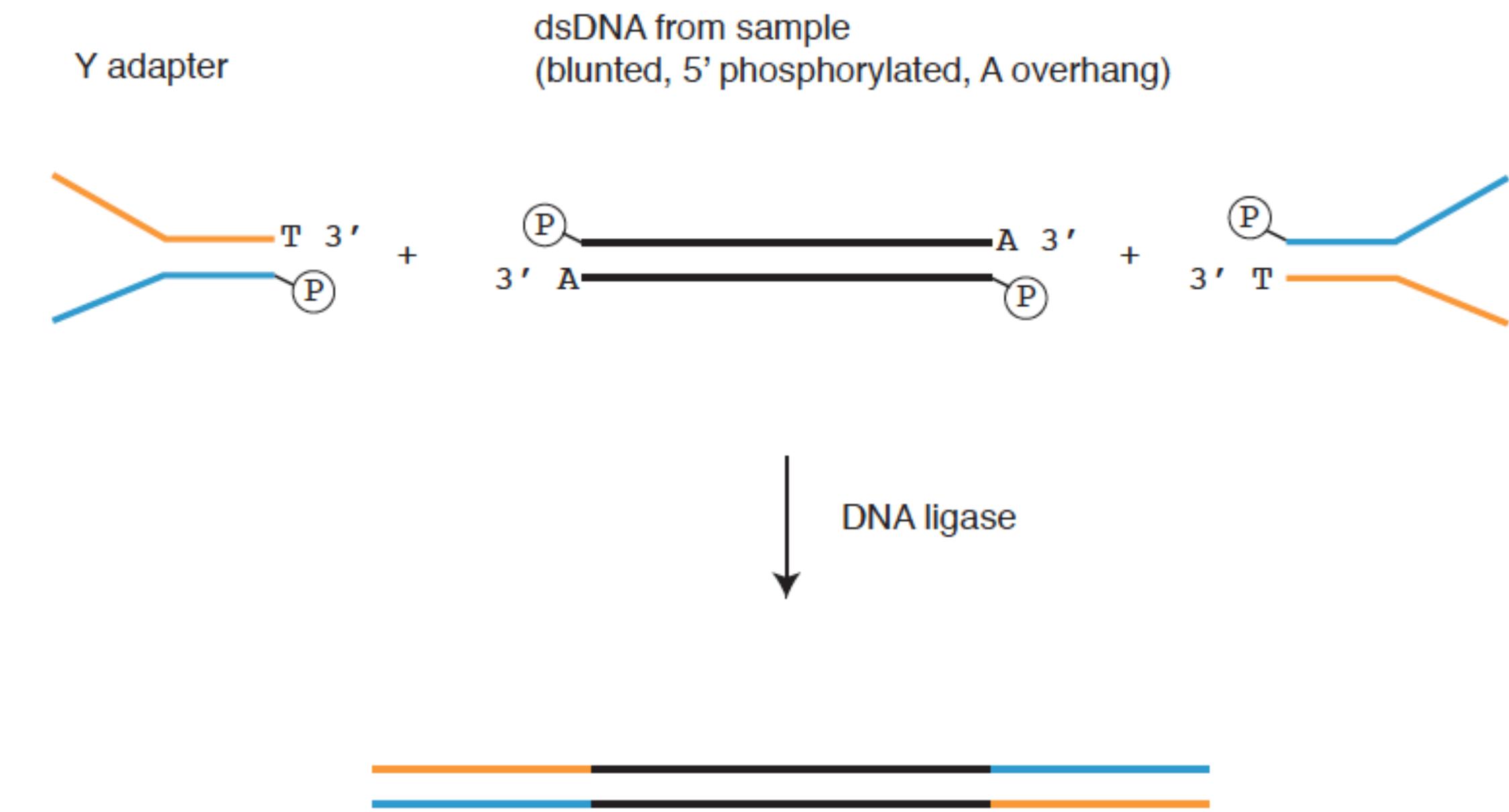


# There are many good ways to make sequencing libraries

## Common library prep steps (not always included and not always in this order)

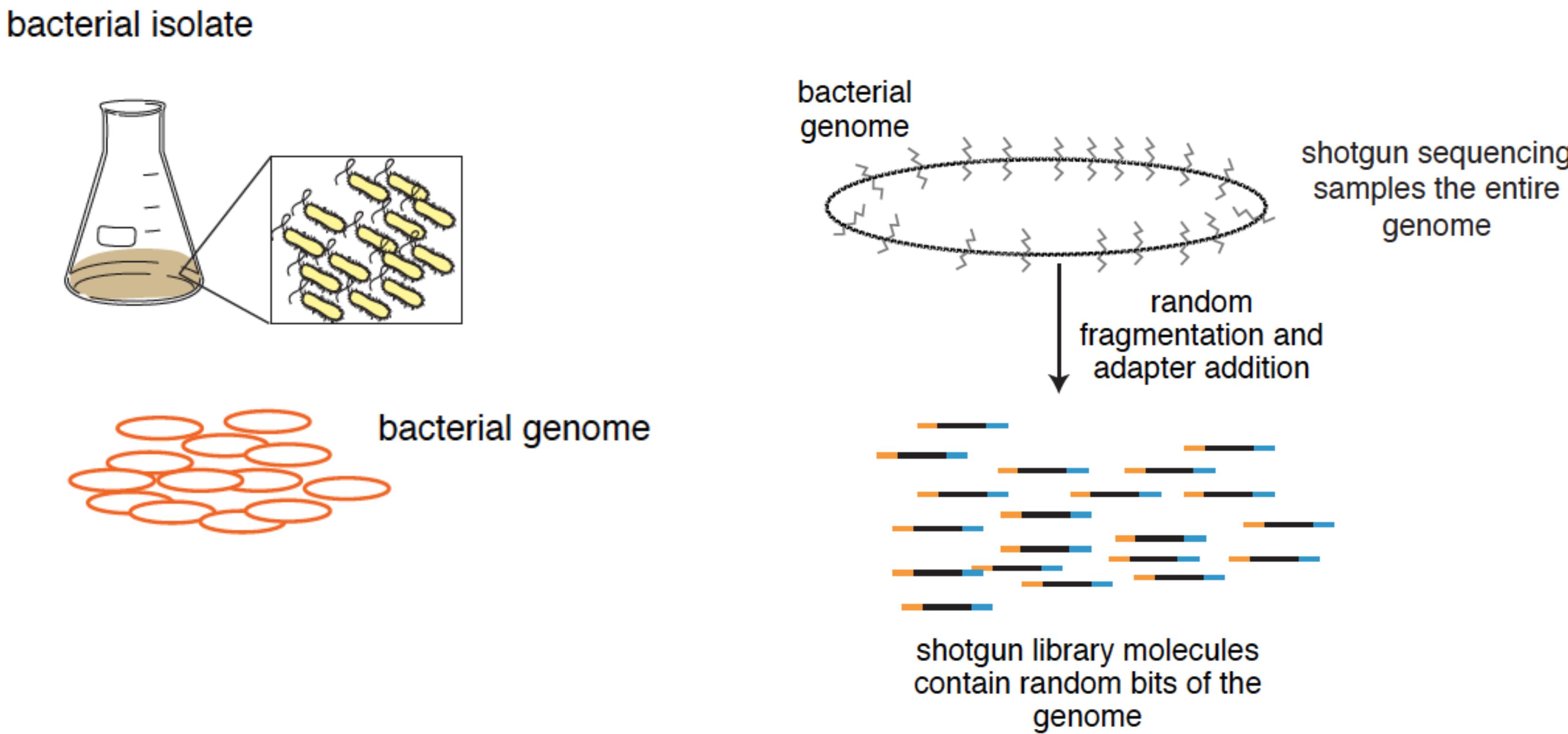
- Nucleic acid isolation
- Enrichment (of nucleic acid subtypes you want) or subtraction (of those you don't want)
- Nucleic acid fragmentation
- Conversion of RNA into dsDNA (for RNA sequencing)
- Addition of adapters to ends of library molecules, possibly with barcodes for multiplexing
- Library amplification
- Pooling of multiplexed samples
- Library QC / quantification

Adapters can be added to sample-derived dsDNA by ligation



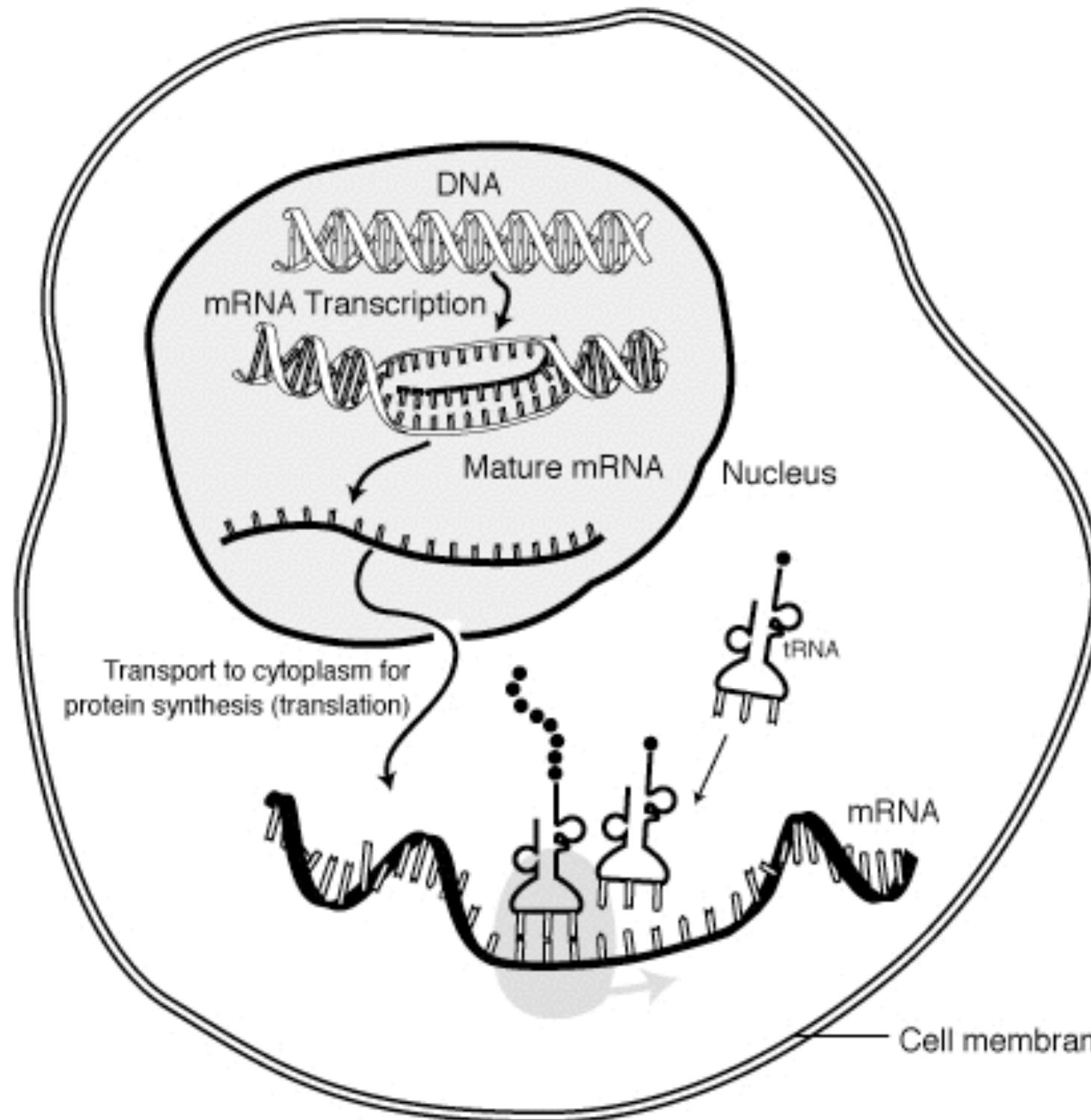
# How you make a library determines what type of sequencing you're doing

For instance, if you make a ‘shotgun library’ from a single organism, you’re doing whole genome sequencing (WGS)



# How you make a library determines what type of sequencing you're doing

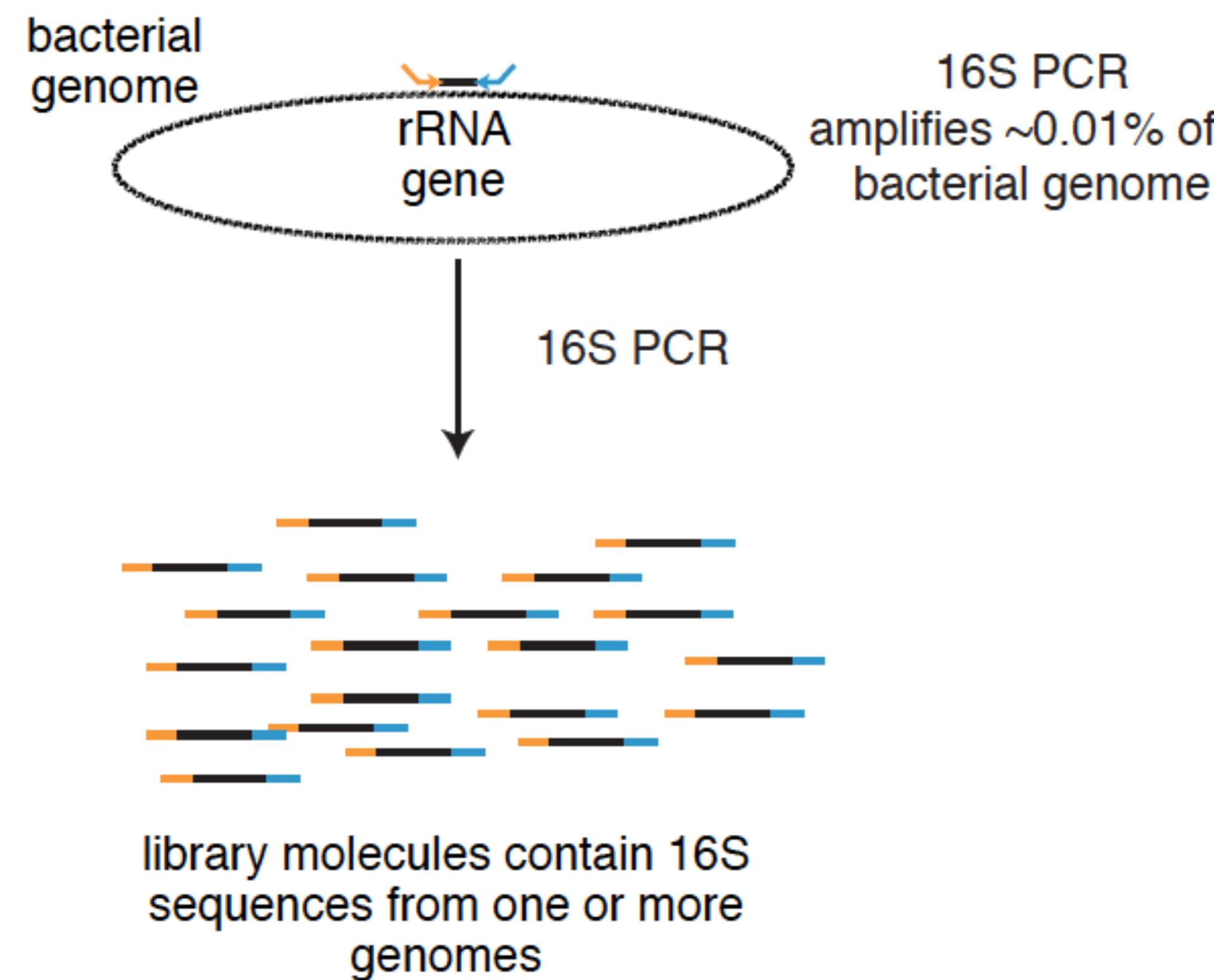
If you make a library from mRNA, that is RNA-Seq (transcriptome sequencing)



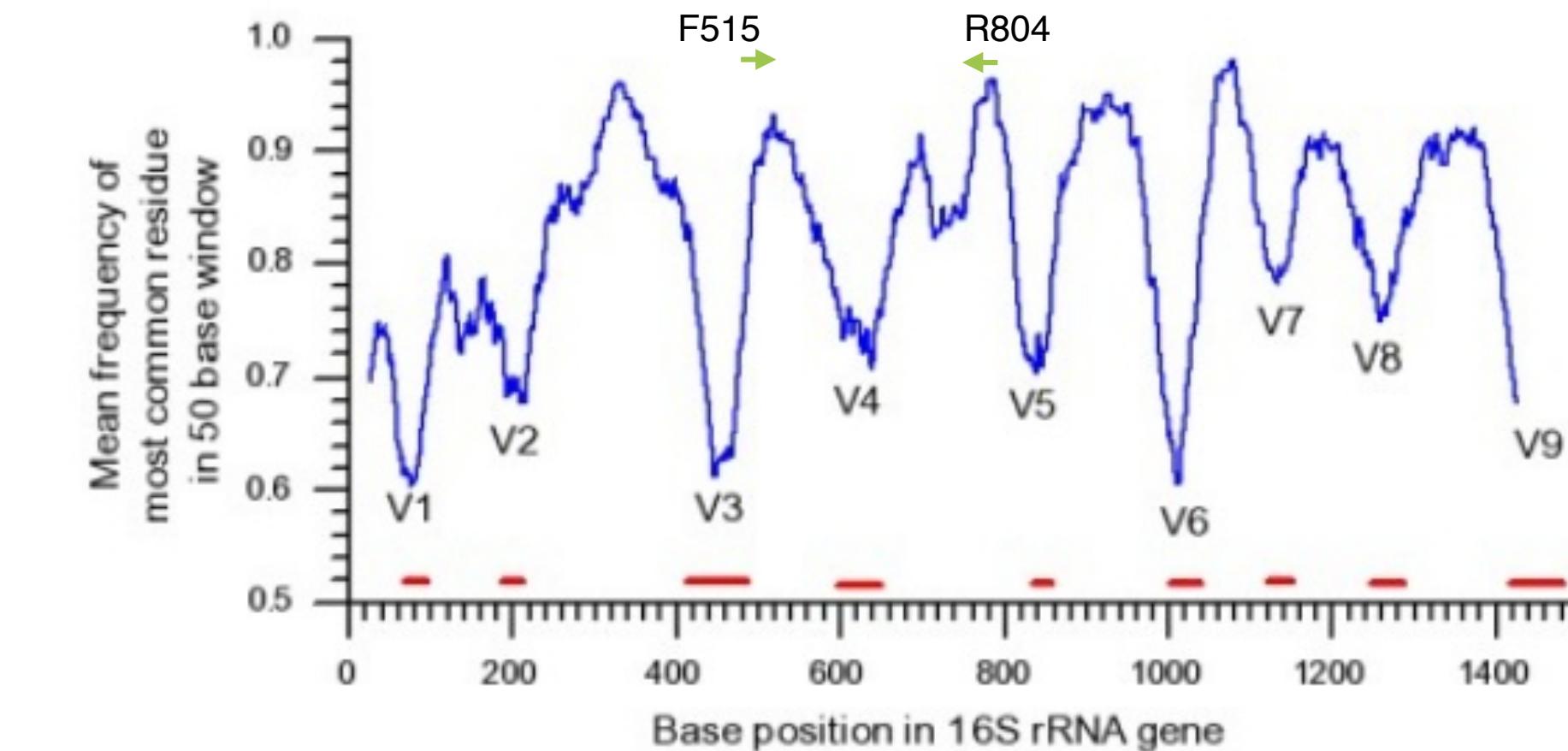
The abundance of reads from a particular mRNA is proportional to that mRNA's abundance in the cell

# Microbiome sequencing often means **16S rRNA** sequencing

16S sequencing is one type of  
'amplicon sequencing'

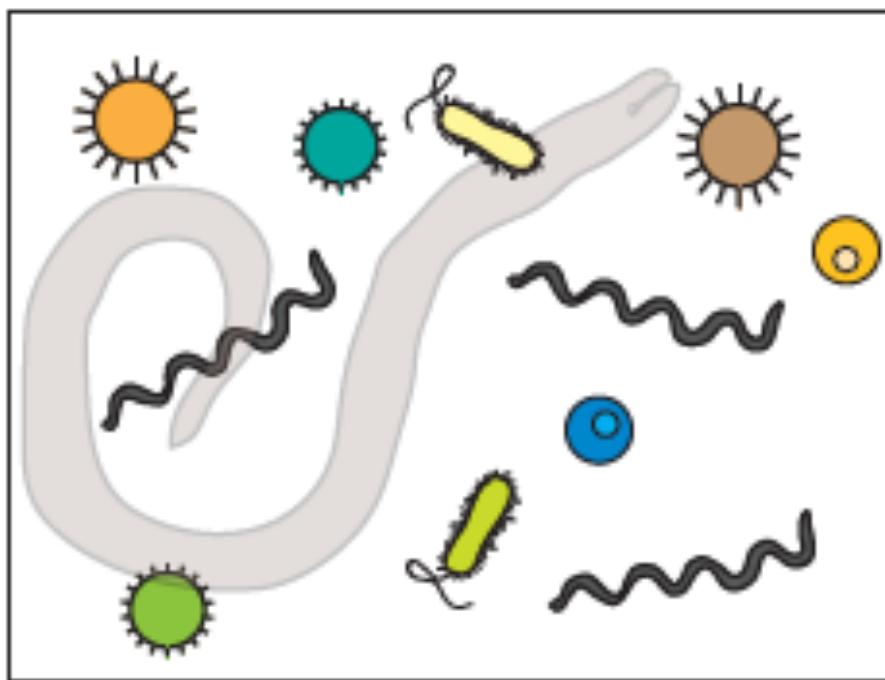


16S rRNA genes have highly conserved regions flanking variable regions



# Metagenomic sequencing involves sequencing of genomes from more than one organism

soil community



Could make a 16S or a shotgun library from these genomes

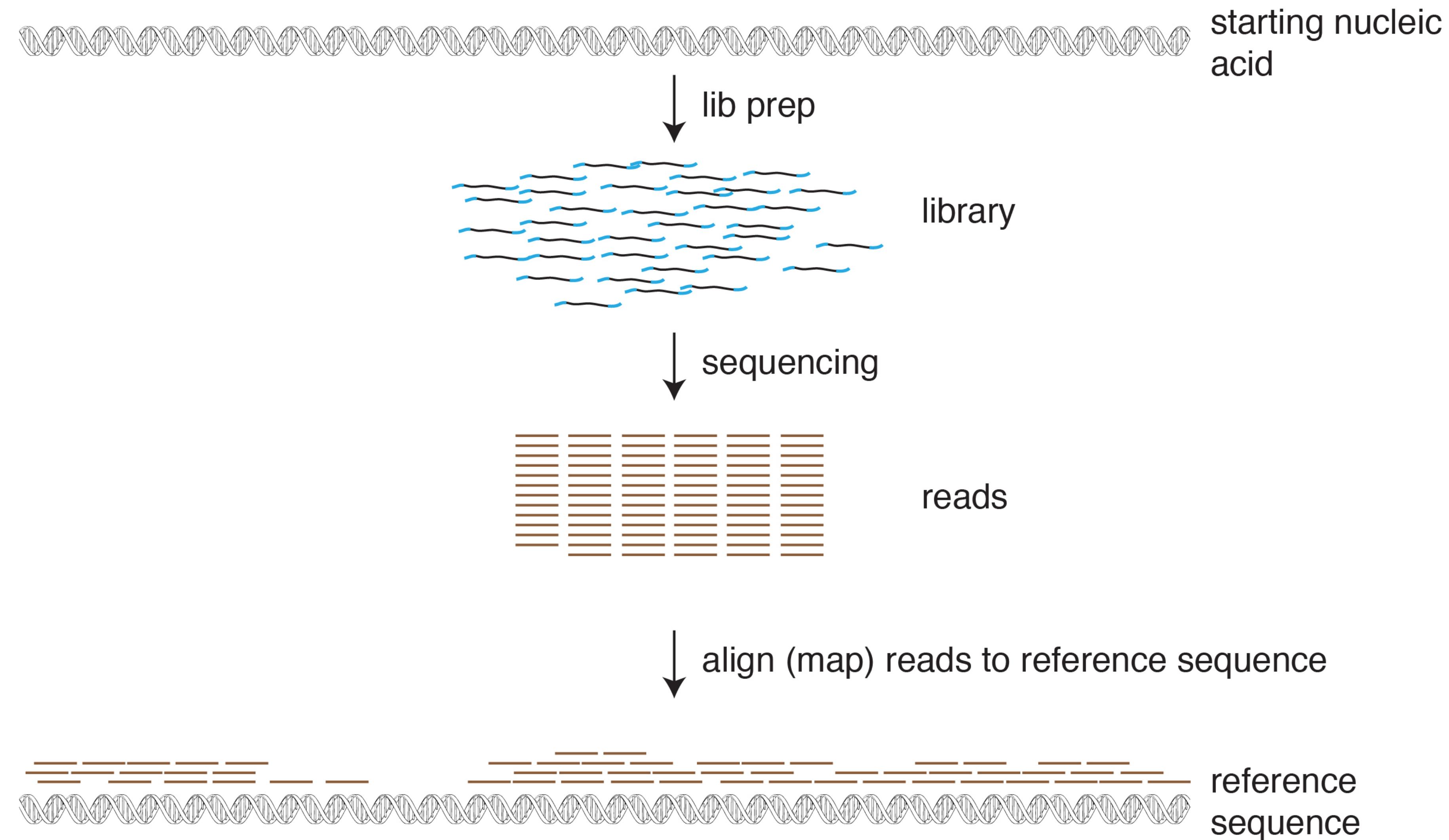
soil 'metagenome'



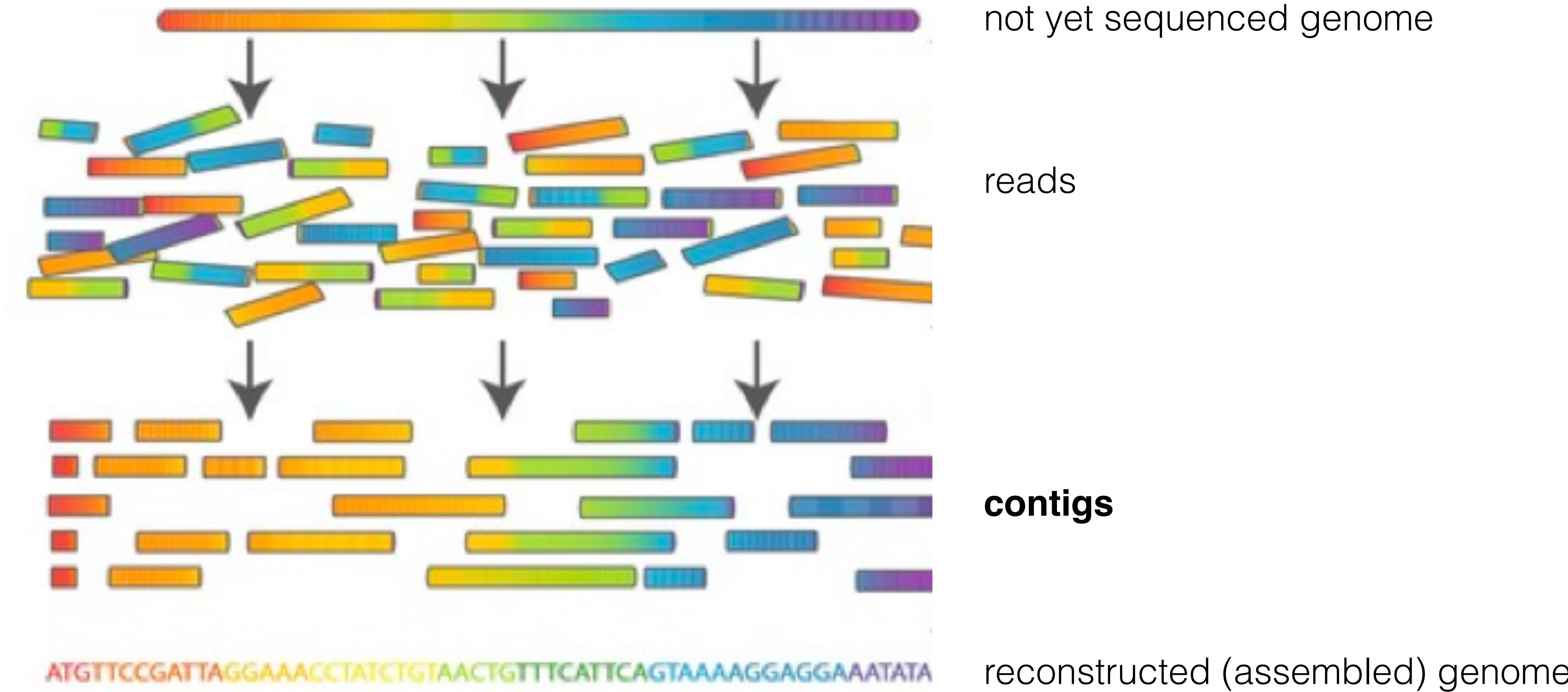
Sequencing of RNAs from a complex sample like this is metatranscriptomics

You can see that different sequencing methods are all variations on a few themes

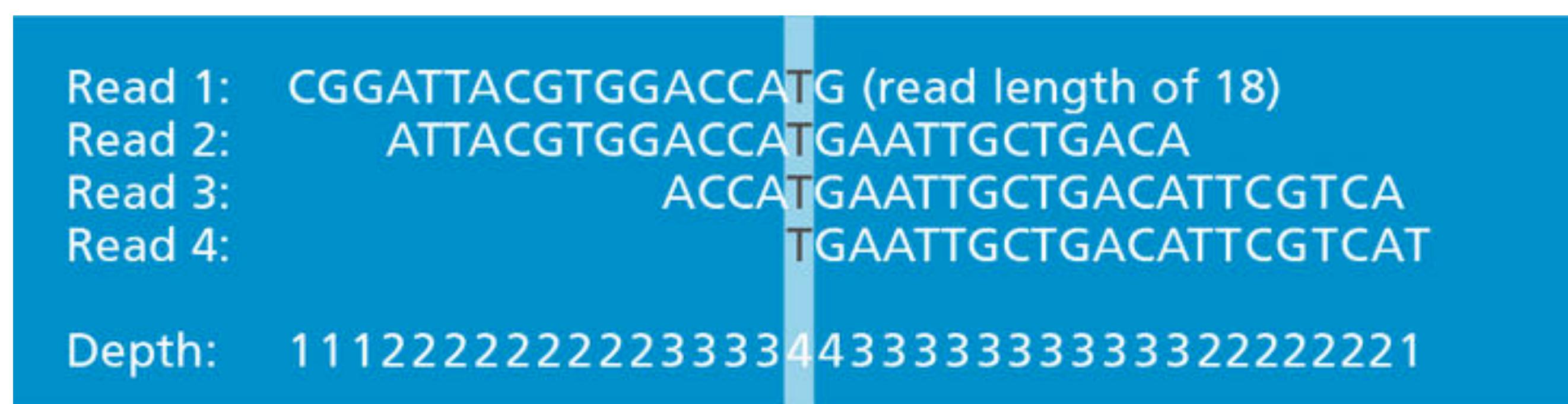
**Mapping** is the process by which sequencing reads are aligned to the region of a genome from which they derive.



(De novo) **assembly** is the process of trying to reconstruct a genome sequence from reads



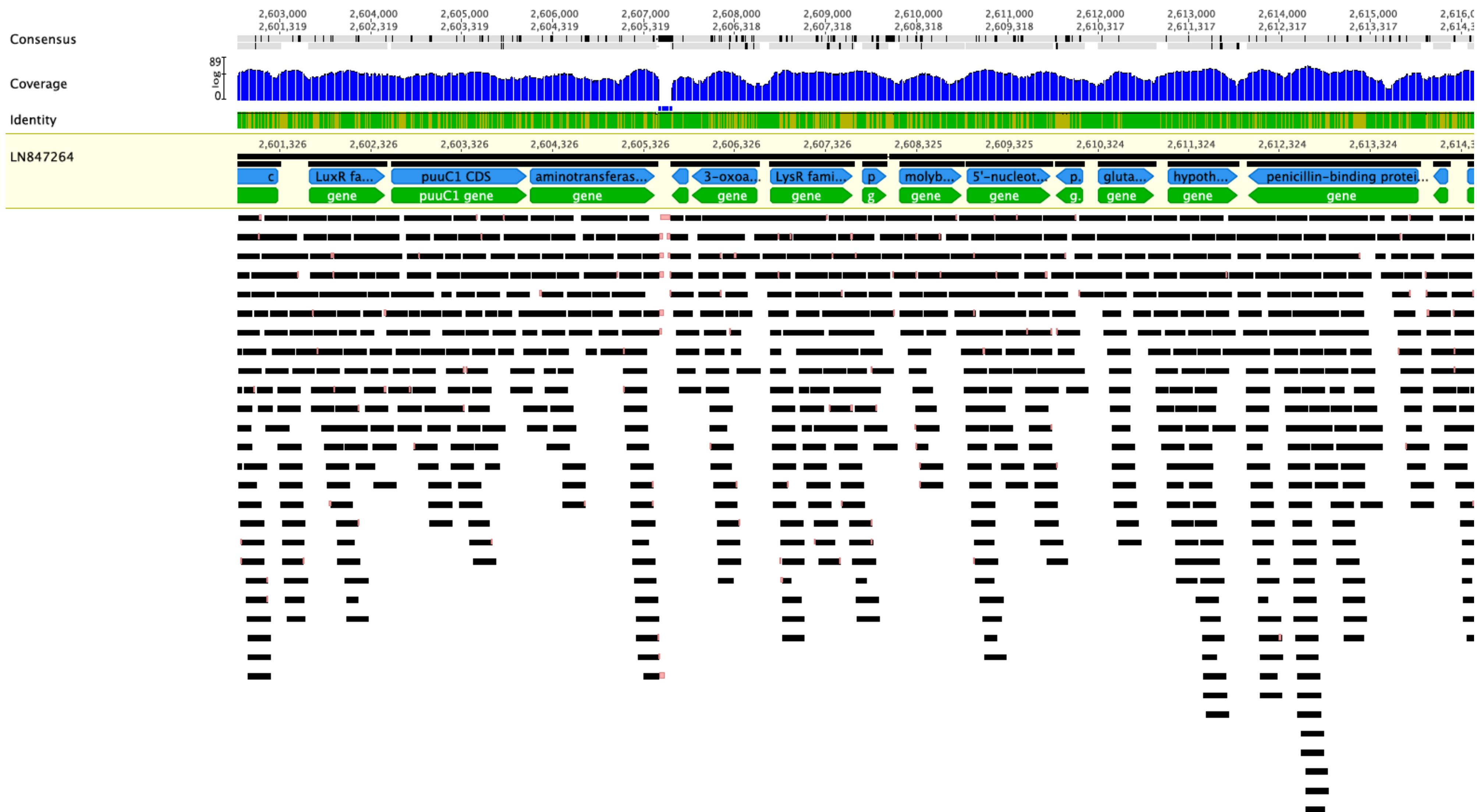
Coverage is the number of individual reads that support a particular nucleotide in an assembled (reconstructed) sequence or that align to a particular nucleotide in a reference sequence



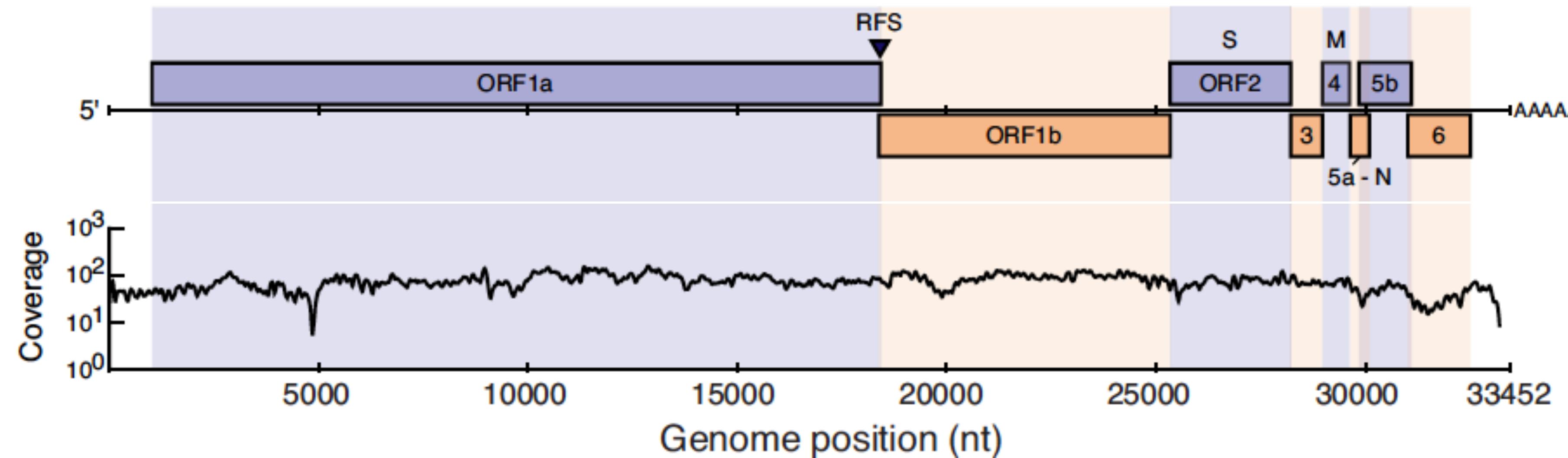
coverage is often referred to as  
'depth' or 'depth of coverage'

*image credit: [wikimedia.org](#)*

# Coverage from WGS of a bacterial isolate



Coverage is sometimes used to represent support for a genome assembly



## Questions?

Is there genomics or sequencing jargon that about which you're not certain?