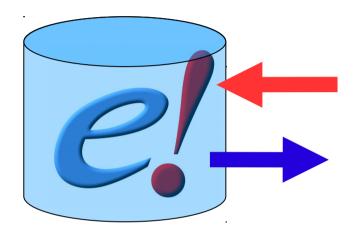


### **Ensembl Core API**





EMBL – European Bioinformatics Institute Wellcome Trust Genome Campus Hinxton, Cambridge, CB10 1SD, UK



## Slides and Examples

### Slides:

http://training.ensembl.org/events/2017/2017-02-09-APITaiwan

(https://goo.gl/ozhBs8)

### **Examples:**

https://github.com/Ensembl/ensembl-presentation/tree/master/API/Core





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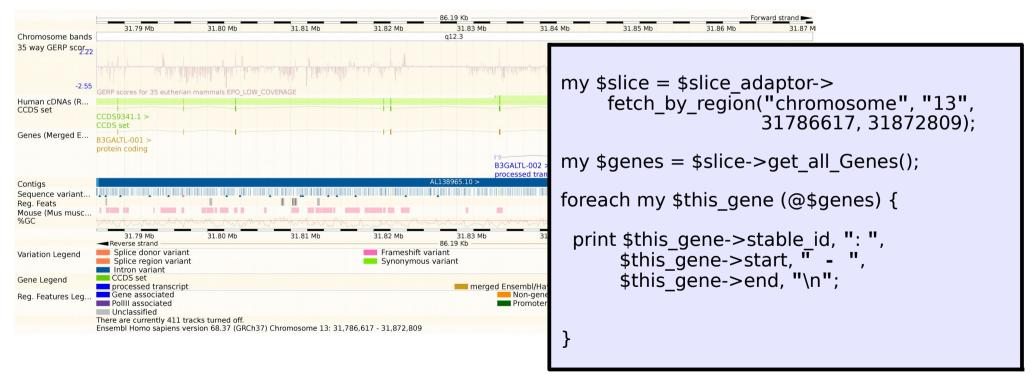
Slides: https://goo.gl/ozhBs8





### **Ensembl API**

- Written in Object-Oriented Perl.
- Used to retrieve data from and to store data in Ensemble databases.
- Foundation for the Ensembl Pipeline and Ensembl Web interface.









# Why use an API?

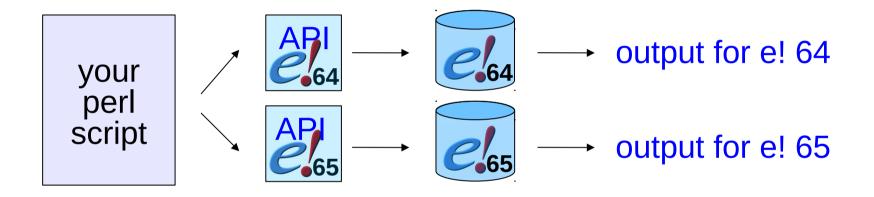
- Uniform method of access to the data.
- Avoid writing the same thing twice: reusable in different systems.
- Reliable: lots of hard work, testing and optimisation already done.
- Insulates developers from underlying changes at a lower level (i.e. the database).





### Using the correct API version

API version **must** match database version. Old scripts using the API *should* continue working with a newer API.



Run script ensembl/misc-scripts/ping\_ensembl.pl to test if you can contact the Ensembl database server. The script will help you resolve issues with your setup.





### **An Alternative - REST**



**Endpoints** 

User Guide

Change Log

About the Ensembl Project

Contact Ensembl

### **Sequences**

Resource	Description	
GET sequence/id/:id	Request multiple types of sequence by stable identifier.	
GET sequence/region/:species/:region	Returns the genomic sequence of the specified region of the given species.	

#### **Variation**

Resource	Description	
GET variation/:species/:id	Uses a variation identifier (e.g. rsID) to return the variation features	
GET vep/:species/id/:id	Fetch variant consequences based on a variation identifier	
POST vep/:species/id/	Fetch variant consequences for multiple ids	

## http://rest.ensembl.org







### Installing the Perl API from FTP

```
# cd to a location to install Ensembl to
mkdir src
cd src
# Get the latest API from FTP (always the live version) and BioPerl 1.6.1
wget ftp://ftp.ensembl.org/pub/ensembl-api.tar.gz
wget https://cpan.metacpan.org/authors/id/C/CJ/CJFIELDS/BioPerl-1.6.1.tar.gz
# untar both
tar zxvf ensembl-api.tar.qz
tar zxvf BioPerl-1.6.1.tar.gz
# open up .bashrc or .profile and add the following
PERL5LIB=${PERL5LIB}:${HOME}/src/bioperl-1.6.1
PERL5LIB=${PERL5LIB}:${HOME}/src/ensembl/modules
PERL5LIB=${PERL5LIB}:${HOME}/src/ensembl-compara/modules
PERL5LIB=${PERL5LIB}:${HOME}/src/ensembl-variation/modules
PERL5LIB=${PERL5LIB}:${HOME}/src/ensembl-functgenomics/modules
export PERL5LIB
# Checking your installation
perl $HOME/src/ensembl/misc-scripts/ping ensembl.pl
```







### **Alternative Methods of Installation**

- Git clone from https://github.com/Ensembl/
- Ensembl Virtual Machine
  - ftp://ftp.ensembl.org/pub/current\_virtual\_machine
  - 64bit only in OVA format
  - Available as a Vagrant box too
     vagrant init ensembl/ensembl; vagrant up --provider virtualbox







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## Ensembl API – Object Types

We have two main object types in Ensembl API:

### 1. Data Objects

Talk to a particular row in a data table, such as the BRCA2 gene to get (or set) information related to this gene only.

### 2. Object Adaptors

Talk to a particular data table, such as the gene table to retrieve or store genes in the gene table.

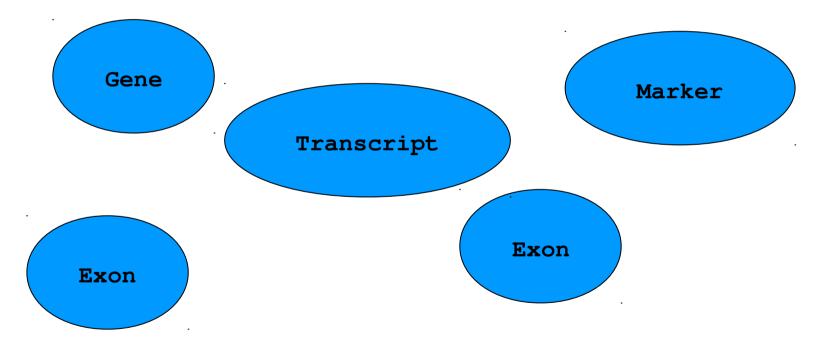




### **Data Objects**

Data objects model biological entities, e.g. genes, transcripts, exons...

A *Data Object* represents a piece of data that is (or can be) stored in the database.







## **Object Adaptors**

Data Objects are retrieved from and stored in the database using Object Adaptors.

Each *Object Adaptor* is responsible for creating objects of only one particular type. For instance:

- The GeneAdaptor is used to fetch Gene objects
- The ExonAdaptor is used to fetch Exon objects

Object Adaptor fetch, store, and remove methods are used to retrieve, save, and delete information in the database.

Two types of methods:

- fetch\_by\_.... returns 1 object (or undef)
- fetch\_all\_by\_... returns a ref. to an array of objects (or ref. to an empty array)





## Object methods and adaptors

- The API deals with objects representing database entities
- Adaptors are "factories" for generating objects
  - Adaptors are retrieved from the Registry

```
use strict;
                                     Use 'strict' and 'warnings'
Use warnings
use Bio::EnsEMBL::Registry;
my $reg = 'Bio::EnsEMBL::Registry';
$reg->load_registry_from_db(
                                             Change your host and port
   -host => 'ensembldb.ensembl.org',
   -user => 'anonymous'
);
my $ga = $reg->get_adaptor('human', 'core',
                                                     'gene');
                                                  object name
                             species
                                         group
my $gene= $ga->fetch_by_stable_id('ENSG00000139618');
```





## Object methods in Ensembl API

An Object has attributes and methods.

We avoid accessing object attributes directly, we use methods instead.

Methods are called using the "arrow" (->) operator:

```
my $exons = $gene->get_all_Exons();
```

Many methods can be used to either get or set an attribute value:

```
GET (no arg.):

my $gene_id = $gene->stable_id( );

SET (new value):

$gene->stable_id("ENSG000000123152");
```





# Object retrieval

- Using adaptors
  - Fetch object(s) according to some property e.g. name, location
    - "fetch all ..." -> returns a list reference of items
    - "fetch\_by\_..." usually returns only 1 item
  - Check documentation which methods the adaptor provides
- Using API objects: e.g. Slice, Gene, Transcript...
  - Get other object(s) from an API object
     e.g. \$gene->get\_all\_Transcripts()
     returns a list reference of Ensembl Transcript objects
  - Usually the object is written with a upper case in the method
     e.g. get\_all\_Transcripts()





## Data Objects & Object Adaptors

```
# fetch a gene by its stable identifier using a gene
adaptor

my $gene =
    $gene_adaptor->fetch_by_stable_id('ENSG00000139618');

# print out the name of a gene and its stable identifier

print $gene->external_name(), "\n";

print $gene->stable_id, "\n";
```





### **Ensembl Core modules**

### Name space for the modules:

- Object modules start with Bio::EnsEMBL
  - Bio::EnsEMBL::Gene for gene objects
  - Bio::EnsEMBL::Exon for exon objects
- ObjectAdaptors start with Bio::EnsEMBL::DBSQL
  - Bio::EnsEMBL::DBSQL::GeneAdaptor
  - Bio::EnsEMBL::DBSQL::ExonAdaptor





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# **Ensembl API Documentation (1)**

Go to http://www.ensembl.org/info/docs/Doxygen/index.html



#### Doxygen perl module documentation

These are the components of the Ensembl code base. Click on a module below for API documentation:

- Ensembl Ensembl core database API
- Core code base

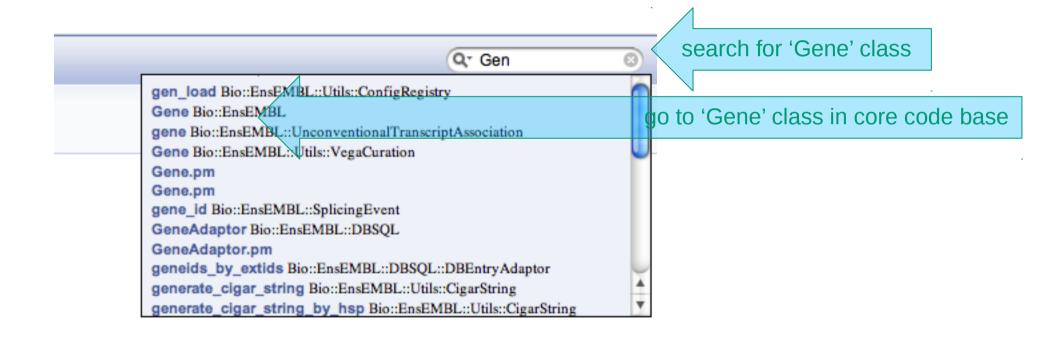
- Ensembl-analysis
- Ensembl-hive
- Ensembl-compara Ensembl comparative genomics API
- Ensembl-external
- Ensembl-functgenomics Ensembl functional genomics API
- Ensembl-pipeline Ensembl gene build pipeline\*
- Ensembl-variation Ensembl variation data API
- BioPerl





# **Ensembl API Documentation (2)**

#### Search for classes or methods

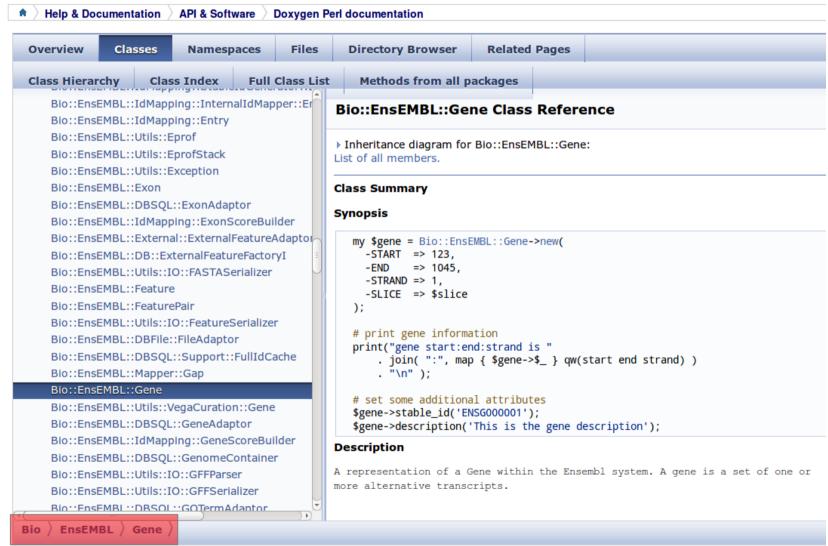






# **Ensembl API Documentation (3)**

Breadcrumbs help you keep track of which code base you're in.

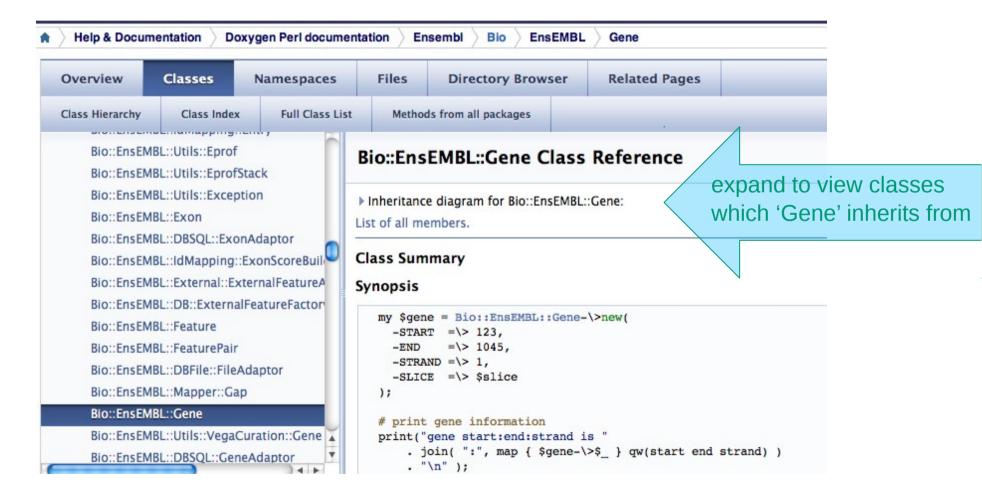






# **Ensembl API Documentation (4)**

Classes can *inherit* attributes and methods from other classes. For instance a Gene is also a Feature as it can be located on a region of DNA so it inherits from the **Bio::EnsEMBL::Feature** object.

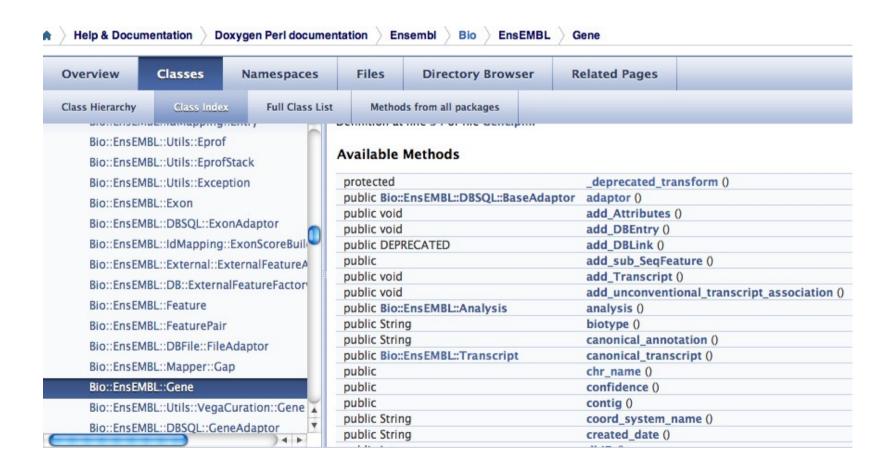






# **Ensembl API Documentation (5)**

Scroll down to see a list of available methods in the 'Gene' class. Can you find a method to retrieve all transcripts for a gene? Where possible, this list shows the returned data types for each method.







# **Ensembl API Documentation (6)**

Clicking on a method takes you to a description with associated arguments, return types and exceptions.

```
public Bio::EnsEMBL::Slice Bio::EnsEMBL::Feature::slice ( )
 Arg [1]
             : (optional) Bio::EnsEMBL::Slice $slice
  Example
  $seqname = $feature->slice()->name();
  Description: Getter/Setter for the Slice that is associated with this
               feature. The slice represents the underlying sequence that this
               feature is on. Note that this method call is analagous to the
               old SegFeature methods contig(), entire seg(), attach seg(),
               etc.
 Returntype : Bio::EnsEMBL::Slice
  Exceptions : thrown if an invalid argument is passed
 Caller : general
  Status : Stable
Code:
click to view
Reimplemented in Bio::EnsEMBL::Exon, and Bio::EnsEMBL::Map::DitagFeature.
```





# **Ensembl API Documentation (7)**

Clicking on **Code:** shows the method's implementation

#### Code:

```
sub slice {
 my ( $self, $slice ) = @ ;
 if ( defined($slice) ) {
   if ( !check ref( $slice, 'Bio::EnsEMBL::Slice' )
         && !check ref( $slice, 'Bio::EnsEMBL::LRGSlice' ) )
     throw('slice argument must be a Bio::EnsEMBL::Slice');
   $self->{'slice'} = $slice;
 } elsif ( @ > 1 ) {
   delete($self->{'slice'});
 return $self->{'slice'};
```

Reimplemented in Bio::EnsEMBL::Exon, and Bio::EnsEMBL::Map::DitagFeature.





## **Ensembl Core DB Documentation (1)**

### Go to http://www.ensembl.org/info/docs/api/core/core schema.html

#### Ensembl Core - Schema documentation

This document gives a high-level description of the tables that make up the EnsEMBL core schema. Tables are grouped into logical groups, and the purpose of each table is explained. It is intended to allow people to familiarise themselves with the schema when encountering it for the first time, or when they need to use some tables that they've not used before.

This document refers to version 74 of the EnsEMBL core schema.

The core database schema is available in several diagrams (PDF format) here:











#### List of the tables:

#### Assembly Tables

- assembly
- assembly exception
- coord system
- data file
- dna

#### **External References**

- associated group
- associated xref
- dependent xref
- external db
- external synonym

#### **Features**

- · density feature
- density type
- ditaq
- ditag feature

- prediction transcript
- repeat consensus
- repeat feature
- simple feature
- intron supporting evidence
   transcript intron supporting evidence







# **Ensembl Core DB Documentation (2)**

Scroll down to the list of Fundamental Tables and click on the gene table.

#### Fundamental Tables

- alt allele
- alt allele attrib
- alt\_allele\_group
- analysis
- analysis description
- attrib\_type
- dna align feature
- exon
- exon\_transcript
- gene
- gene\_attrib
- protein align feature
- protein\_feature
- splicing event
- splicing event feature

- splicing transcript pair
- supporting feature
- transcript
- transcript attrib
- transcript supporting feature
- translation
- translation\_attrib







## **Ensembl Core DB Documentation (3)**

Click on the 'show columns' link to the right to expand a list of gene table columns, their types, descriptions and indices over the columns.

gene 

→ Hide columns I [Back to top]

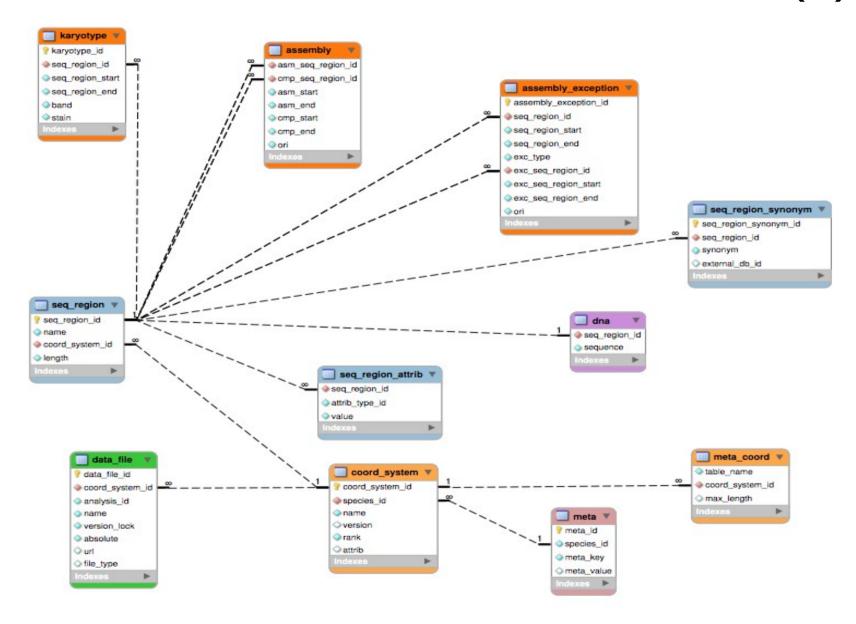
Allows transcripts to be related to genes.

Column	Туре	Default value	Description
gene_id	INT(10)		Primary key, internal identifier.
biotype	VARCHAR(40)		Biotype, e.g. protein_coding.
analysis_id	SMALLINT		Foreign key references to the analysis table.
seq_region_id	INT(10)		Foreign key references to the seq_region table.
seq_region_start	INT(10)		Sequence start position.
seq_region_end	INT(10)		Sequence end position.
seq_region_strand	TINYINT(2)		Sequence region strand: 1 - forward; -1 - reverse.
display_xref_id	INT(10)		External reference for EnsEMBL web site. Foreign key re
source	VARCHAR(20)		e.g ensembl, havana etc.
status	ENUM('KNOWN', 'NOVEL', 'PUTATIVE', 'PREDICTED', 'KNOWN_BY_PROJECTION', 'UNKNOWN', 'ANNOTATED')		Status, e.g. 'KNOWN', 'NOVEL', 'PUTATIVE', 'PREDICTEL' 'KNOWN_BY_PROJECTION', 'UNKNOWN'.
description	TEXT		Gene description





## **Ensembl Core DB Documentation (4)**









### **Exercise 1**

- a) Find documentation for the Exon class in the Ensembl core code base. Which method would you use to retrieve the DNA sequence for an exon? What is the return type for this method?
- b) Can you find a table which stores stable ids for transcripts? Which table stores DNA sequence? How many columns does this table have?





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## The Registry

We know how to use Data Objects and Object Adaptors. How do we make sure we get those from the right database? This is what we use the Registry for.

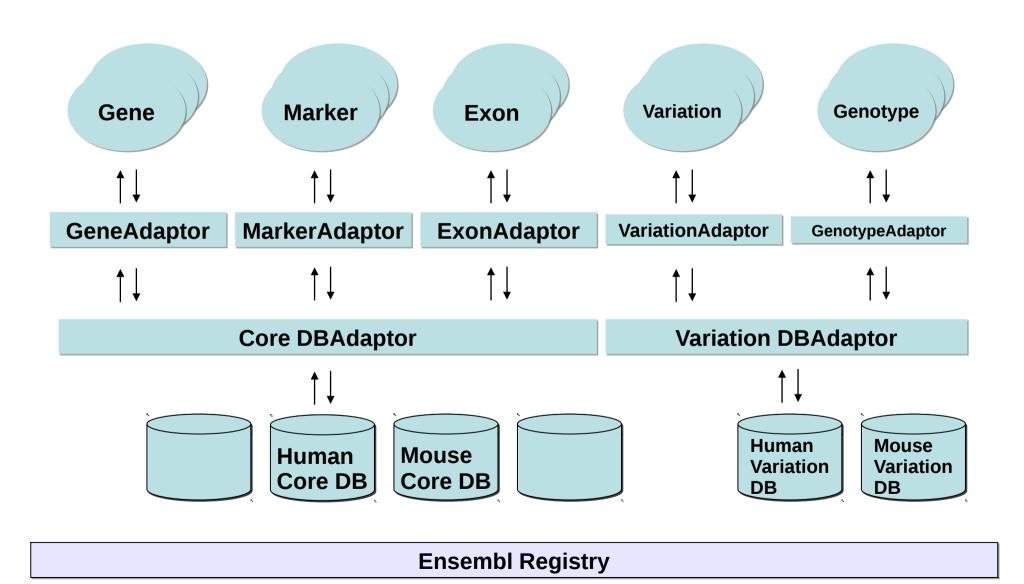
### The Registry:

- loads all databases of the same version as the API
- lazy loads so no connections are made until requested





### **Ensembl API Architecture**







### Review: The basic Ensembl script

```
use strict;
Use warnings
use Bio::EnsEMBL::Registry;
my $reg = 'Bio::EnsEMBL::Registry';
$reg->load_registry_from_db(
   -host => 'ensembldb.ensembl.org',
   -user => 'anonymous'
);
my $ga = $reg->get_adaptor('human','core','gene');
my $gene= $ga->fetch_by_stable_id('ENSG00000139618');
```





### **Exercise 2**

Create a script which uses the method load\_registry\_from\_db to load all databases into the Registry and prints the names of the databases loaded.

Hint: Have a look at the Doxygen documentation for the Registry object and method load\_registry\_from\_db (http://www.ensembl.org/info/docs/Doxygen/index.html).





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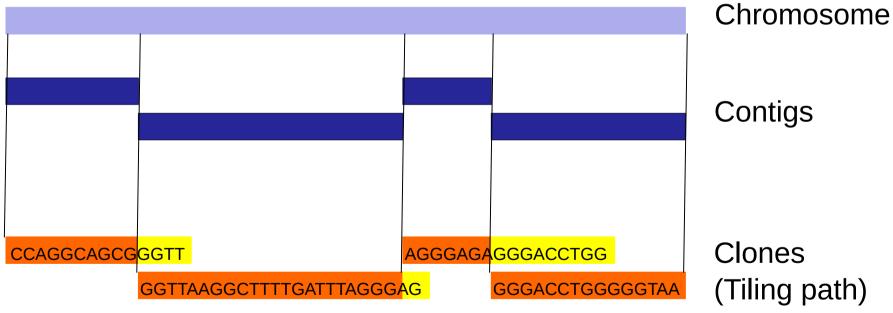




# **Coordinate Systems (1)**

Ensembl stores features and DNA sequence in a number of coordinate systems.

Top level coordinate system



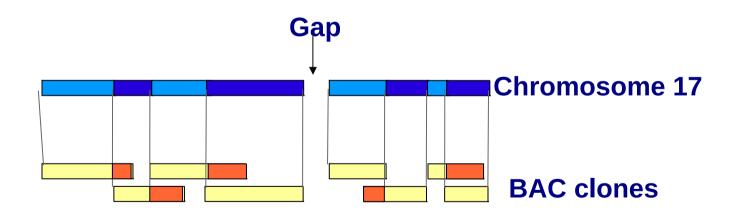
Sequence level coordinate system





# **Coordinate Systems (2)**

Regions in one coordinate system may be constructed from a tiling path of regions from another coordinate system.







# Coordinate Systems - Code Example

Obtain all coordinate systems for human #

```
use Bio::EnsEMBL::Registry;
my $registry = 'Bio::EnsEMBL::Registry';
$registry->load_registry_from_db(
    -host => 'ensembldb.ensembl.org',
    -user => 'anonymous'
);
my $coordsystem_adaptor = $registry->get_adaptor( 'Human', 'Core', 'CoordSystem' );
my $coordsystems = $coordsystem adaptor->fetch all;
while ( my $coordsystem = shift @{$coordsystems} ){
    print $coordsystem->name, "\t",
          $coordsystem->version, "\t",
          $coordsystem->rank ,"\n";
```

Note use of 'while' and 'shift' instead of 'foreach' – more memory efficient way for large datasets





# Coordinate Systems – Code Output

#### **OUTPUT:**

name	versi	on	rank
chromosome scaffold clone	GRCh38 GRCh38	1 2 3	Latest assembly, top level
contig		4	Latest assembly, sequence level
chromosome chromosome chromosome chromosome	GRCh37 NCBI36 NCBI35 NCBI34	5 6 7 7	Old assemblies, used for mapping features between assembly versions.
lrg		8	Locus-Reference Genes, used in clinical





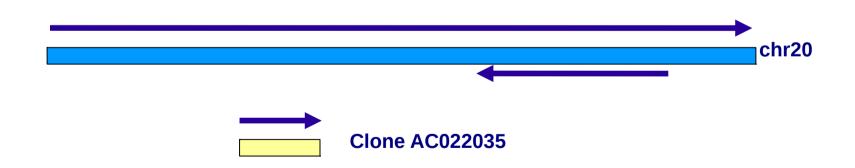
tests

#### Slices

A *Slice* Data Object represents an arbitrary region of a genome, a slice of a Sequence Region.

Slices are not directly stored in the database.

A *Slice* is used to request sequence or features from a specific region in a specific coordinate system.







# Slices - Code Example (1)

# get a slice covering the entire human Y chromosome

#### **OUTPUT:**

Coord system: chromosome

Seq region: Y

Start: 1

End: 57227415

Strand: 1

Slice: chromosome: GRCh38:Y:1:57227415:1





# Slices - Code Example (2)





### **Exercise 3**

- (a) Fetch all chromosomes for human. Determine their number and print the name and length for each of them. The number of chromosomes is probably not what you would expect! Why is this?
- (b) Use the gene stable id 'ENSG00000101266' to fetch a slice surrounding this gene with 2kb of flanking sequence.
  - (a)hint: use the Ensembl API documentation to find an appropriate method in SliceAdaptor class which retrieves a slice given a gene stable id (pay attention to the method's arguments)
- (c) Fetch the sequence of the first 10Mb of chromosome 20 and write it to a file in FASTA format. Print the number of genes in this region.

```
my $output = Bio::SeqIO->new( -file=>'>filename.fasta',
   -format=>'Fasta');
$output->write_seq($slice);
```





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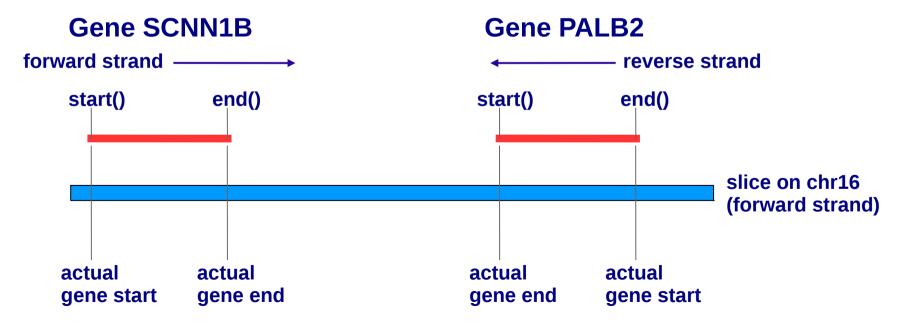


# Features (1)

Features have a defined location on the genome and are stored in a single coordinate system

All Features have a start, end, strand and slice

Start and end are plotted onto the forward strand: start < end





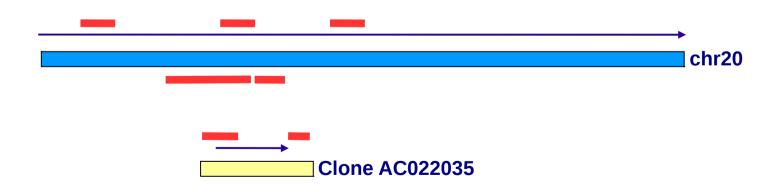


# Features (2)

slice() method returns the Slice object with which the Feature is associated

feature\_Slice() method returns the Slice object which covers only the Feature (will start at 1 and end with Feature length)

Features are retrieved from Object Adaptors using identifiers or regions (slices).







### Features Objects - Biological Correspondence

Object	Biological entity
Gene, Transcript, Exon	Ensembl gene models
PredictionTranscript, PredictionExon	Genscan gene models
DNAAlignFeature, ProteinAlignFeature	cDNAs, proteins
RepeatFeature	repeats
MarkerFeature	markers
KaryotypeBand	cytogenetic bands
SimpleFeature	results of cpg, Eponine, FirstEF and tRNAscan
MiscFeature	clones, ENCODE regions



**ProteinFeature** 



protein domains

# Align features

A sequence (mRNA or protein) is aligned against the genome

The result is stored in an align\_feature table (dna\_align\_feature or protein\_align\_feature)

A row represents the alignment between the genome sequence (a slice) and the target sequence (a hit)

seq_region_id	INT(10)		Foreign key references to the seq_region table.
seq_region_start	INT(10)		Sequence start position.
seq_region_end	INT(10)		Sequence end position.
seq_region_strand	TINYINT(1)	'1'	Sequence region strand: 1 - forward; -1 - reverse.
hit_start	INT(10)		Alignment hit start position.
hit_end	INT(10)		Alignment hit end position.
hit_name	VARCHAR(40)		Alignment hit name.





#### **Exercise 4**

- (a) Get all the repeat features from chromosome 20:1-500kb. Print out the name and position of each on the chromosome and the total number.
  - hint: create a slice, retrieve repeat features on this slice
- (b) Find which genomic region the RefSeq dna entry NM\_000059.3 was mapped to. Print the name of the region and coordinates of the alignment on the genome as well as the name of the region and coordinates of the alignment on the RefSeq dna entry. Print the score and percentage identity for the alignment.
  - hint: use DnaAlignFeatureAdaptor; use the core schema documentation as a guide to appropriate methods which correspond to columns in dna\_align\_feature table

A list of useful Feature methods is in the Appendix at the end of the presentation slides





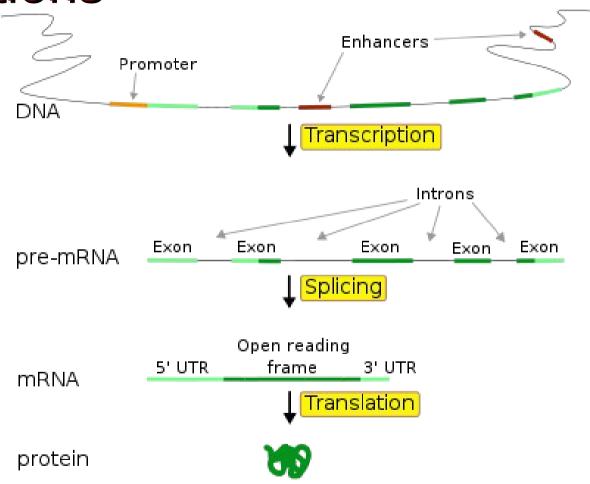
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# Genes, transcripts, exons & translations







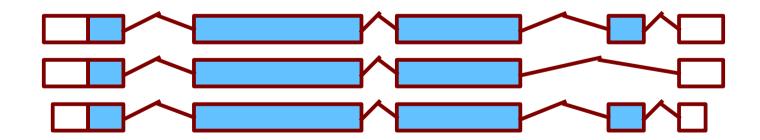
# Genes, Transcripts and Exons

Genes, Transcript and Exons are objects that can be used just like any other Feature object

A Gene is a set of alternatively spliced Transcripts

A Transcript is a set of Exons

Introns are not explicitly defined in the database







### Genes, Transcripts, Exons – Code Output

```
# helper function: returns location and stable id string for a feature
sub get string {
  my $feature = shift;
  my $stable_id = $feature->stable_id;
  my $seq region = $feature->slice->seq region name;
  mv $start = $feature->start;
  my $end = $feature->end;
  my $strand = $feature->strand;
  return "$stable id $seg region:$start-$end($strand)";
# fetch a gene by its stable identifier
my $gene = $gene adaptor->fetch by stable id('ENSG00000123427');
# print out the gene, its transcripts, and its exons
print "Gene: ", get_string($gene), "\n";
while ( my $transcript = shift @{$gene->get_all_Transcripts} ){
  print " Transcript: ", get_string($transcript), "\n";
  while ( my $exon = shift @{$transcript->get_all_Exons} ){
    print " Exon: ", get_string($exon), "\n";
}
```





### Genes, Transcripts, Exons – Code Output

#### OUTPUT:

```
Gene: ENSG00000123427 12:57771492-57782541(1)
 Transcript: ENST00000548256 12:57771492-57780430(1)
   Exon: ENSE00002360002 12:57771492-57771625(1)
   Fxon: ENSE00003631087
                           12:57773017-57773128(1)
   Exon: ENSE00003530124
                           12:57774629-57774767(1)
   Exon: ENSE00002406112
                           12:57780255-57780430(1)
 Transcript: ENST00000551420
                              12:57772200-57780780(1)
   Exon: ENSE00002355737
                           12:57772200-57772397(1)
   Exon: ENSE00003506271
                           12:57773017-57773128(1)
   Exon: ENSE00002376752
                           12:57780255-57780780(1)
 Transcript: ENST00000300209
                               12:57772600-57782403(1)
   Exon: ENSE00002301479
                           12:57772600-57772901(1)
   Exon: ENSE00003631087
                           12:57773017-57773128(1)
   Exon: ENSE00002393444
                           12:57780255-57782403(1)
                                                  etc.
```





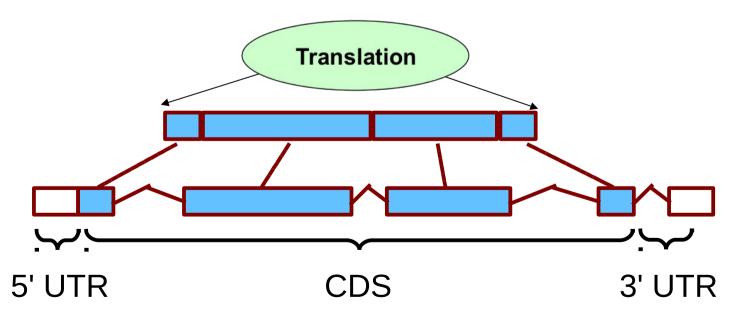
#### **Translations**

Translations are not Features.

A Translation object defines the UTR and CDS of a Transcript.

Peptides are not stored in the database, they are computed on the fly using Transcript objects.

Not all transcripts have a translation (e.g. ncRNAs)







### Translations – Code Example

```
my $transcript adaptor = $registry->get adaptor( 'Homo sapiens', 'Core',
   'Transcript');
# fetch a transcript from the database
my $transcript =
  $transcript_adaptor->fetch_by_stable_id('ENST00000333012');
# obtain the translation of the transcript
my $translation = $transcript->translation;
# print out the translation info
print "Translation: ", $translation->stable_id, "\n";
print "Start Exon: ",$translation->start_Exon->stable_id,"\n";
print "End Exon: ", $translation->end Exon->stable id, "\n";
# cDNA start and end (spliced sequence with UTR)
print "Start : ", $translation->cdna_start, "\n";
print "End : ", $translation->cdna end, "\n";
# print the peptide which is the product of the translation
print "Peptide : ", $transcript->translate->seq, "\n";
```





### Translations – Code Output

#### **OUTPUT:**

Translation: ENSP00000327425

Start Exon: ENSE00002340145

End Exon: ENSE00002428887

Start: 48

End : 497

Peptide:

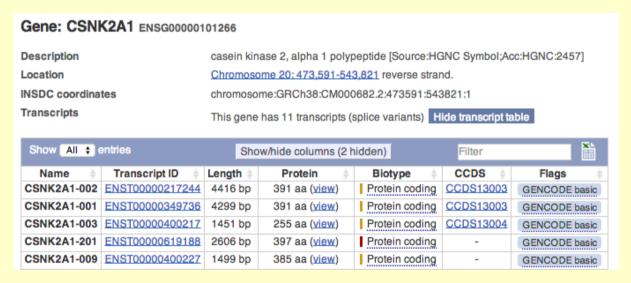
MADPGPDPESESESVFPREVGLFADSYSEKSQFCFCGHVLTITQNFGSRLGVAARVWDAALSLCNYFESQNVDFR GKKVIELGAGTGIVGILAALQGAYGLVRETEDDVIEQELWRGMRGACGHALSMSTMTPWESIKGSSVRGGCYHH





#### **Exercise 5**

- (a) Fetch gene 'CSNK2A1' and print the number of its transcripts and exons.
  - √ hint: use GeneAdaptor method fetch\_by\_display\_label; remember that not all transcripts have a translation



- (b) For the above gene, get all the transcripts and list the number of exons for each. Also show any translations associated with the transcripts.
- (c) Why do the exon numbers not match?





#### **Outline**

- a. Introduction
- b. Data objects & object adaptors
- c. Ensembl documentation
- d. The Registry & Ensembl API script design
- e. Coordinate systems & slices
- f. Features
- g. Genes, transcripts, exons & translations
- h. External references



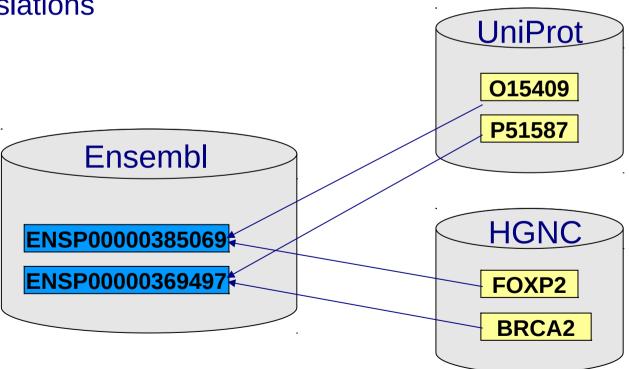


#### **External References**

Ensembl cross references its Gene models with identifiers from other databases, such as HGNC, WikiGenes, UniProtKB/Swiss-Prot, RefSeq, MIM etc.

External References (Xrefs) can be linked to genes, transcripts or

translations







# **Xrefs - Code Example (1)**

# Obtain external references for Ensembl gene ENSG00000139618

```
'ENSG0000139618');
my $gene = $gene_adaptor->fetch_by_stable_id(
                                                   this method will only return xrefs
my $gene_xrefs = $gene->get_all_DBEntries;
                                                   linked to the object it's called on
print "Xrefs on gene level: \n\n";
                                                   (e.g. gene)
while ( my $gene_xref = shift @{$gene_xrefs} ){
  print $gene_xref->dbname, ":", $gene_xref->display_id, "\n";
}
                                               this method will return xrefs on all
my $all_xrefs = $gene->get_all_DBLinks;
                                               levels (gene, transcript and
                                               translation)
print "\nXrefs on gene, transcript and protein level: \n\n";
while ( my $all_xref = shift @{$all_xrefs} ){
  print $all_xref->dbname, ":", $all_xref->display_id, "\n";
}
```





# Xrefs - Code Output (1)

#### **OUTPUT:**

Xrefs on gene level:

Vega\_gene:OTTHUMG00000017411

Vega\_gene:BRCA2 PUBMED:15057823 PUBMED:7581463

PUBMED: 8091231

RefSeq\_dna:NM\_000059

OTTG: OTTHUMG00000017411

ENS\_LRG\_gene: LRG\_293

ArrayExpress: ENSG00000139618

DBASS3:BRCA2
DBASS5:BRCA2

EntrezGene:BRCA2

**HGNC: BRCA2** 

MIM\_GENE: BRCA2 GENE [\*600185]

MIM\_MORBID: BREAST CANCER [#114480]

MIM\_MORBID: GLIOMA SUSCEPTIBILITY 3 [#613029]

MIM\_MORBID: PANCREATIC CANCER, SUSCEPTIBILITY [#613347]

UniGene:Hs.34012 UniGene:Hs.686439 Uniprot\_gn:BRCA2

WikiGene: BRCA2

Xrefs on gene, transcript and translation level:
(same as on gene level + transcript and translation
 level)

Vega\_gene: OTTHUMG00000017411

Vega\_gene:BRCA2
PUBMED:15057823
PUBMED:7581463
PUBMED:8091231

RefSeq\_dna:NM\_000059 OTTG:OTTHUMG00000017411 ENS\_LRG\_gene:LRG\_293

ArrayExpress: ENSG00000139618

DBASS3:BRCA2
DBASS5:BRCA2

EntrezGene: BRCA2

HGNC: BRCA2

Uniprot/SPTREMBL:E9PIQ1

Uniprot/SPTREMBL:K4JTT2

etc.





# **Xrefs - Code Example (2)**

# Retrieve Ensembl IDs for a list of UniProt protein IDs

Proteins map to Ensembl Translation objects so we will use a TranslationAdaptor

```
my @uniprot_ids = qw(P51587 P15056 B8A597 B8A595 B7ZW72);
while ( my $uniprot_id = shift @{$uniprot_ids} ){
   my @trans = @{
    $translation_adaptor->fetch_all_by_external_name($uniprot_id,'Uniprot%')
   };

while ( my $translation = shift @{$trans} ){
   print $translation->stable_id."\t".$uniprot_id."\n";
  }
}
```





# Xrefs - Code Output (2)

#### **OUTPUT:**

ENSP00000439902 P51587

ENSP00000369497 P51587

ENSP00000288602 P15056

ENSP00000387217 B8A597

ENSP00000386781 B8A595

ENSP00000307640 B7ZW72 Cross references can map to more than one Ensembl identifier





#### **Exercise 6**

Retrieve a list of GO term IDs and term names linked to the gene with stable id 'ENSG0000139618'

- ✓ Use *get\_all\_DBLinks* with an external database name argument to restrict the number of xrefs returned
- ✓ Ontology term data such as term name and definition are stored outside of the core database. Create an OntologyTerm Adaptor with the help of the Registry method *get\_adaptor* using arguments: 'Multi' (species), 'Ontology' (database type), 'OntologyTerm' (adaptor type)
- ✓ For all xrefs returned by *get\_all\_DBLinks* use the OntologyTerm Adaptor to fetch the relevant term and print its accession and name (xref display\_id is the same as term accession)





# Recap - Ensembl API script design

#### Always:

Load the registry

Which features (genes, repeats, SNPs, etc.) are in my particular region of interest?

- Get the SliceAdaptor
- Fetch the Slice for you region of interest
- Get the features from your Slice

What do we know about a particular gene (or any other feature)?

- Get the GeneAdaptor
- Fetch your Gene of interest
- Get more details about the gene:
  - Gene structure (transcripts, exons, translations)
  - Annotations: GO xrefs, HGNC symbols, etc.
  - Features in the same region -> get the Slice for the Gene!





# **Documentation & Help**

 Installation instructions, web-browsable version of the POD (Perldoc), database schema and tutorial:

http://www.ensembl.org/info/docs/api/index.html

- Inline Perl POD (Plain Old Documentation)
- dev@ensembl.org mailing list:

http://www.ensembl.org/info/about/contact/mailing.html searchable mailing list archive:

http://blog.gmane.org/gmane.science.biology.ensembl.devel

Ensembl helpdesk:

helpdesk@ensembl.org





### **Ensembl Acknowledgements**

#### The Entire Ensembl Team

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## Ensembl 2016







#### Feedback

Please give us your feedback on the course!

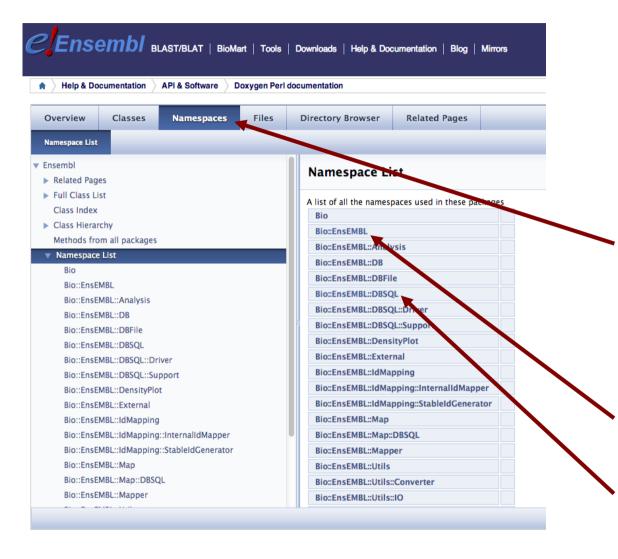
http://training.ensembl.org/events/2017/2017-02-09-APITaiwan

(https://goo.gl/ozhBs8)





# Appendix – Adaptors



"What kinds of adaptors are there?"

The online API documentation lists them all under their respective name spaces.

Bio::EnsEMBL for Data

Objects

Bio::EnsEMBL::DBSQL

for Adaptors





# Appendix – CoordSystem Methods

Attribute	Example value(s)	Method(s)
name	chromosome, scaffold, contig, clone	<pre>\$coordsystem-&gt;name</pre>
version	GRCh37, NCBI36, NCBIM37	<pre>\$coordsystem-&gt;version</pre>





# **Appendix – Feature Methods**

Attribute	Example value(s)	Method(s)	
			slice relative
name	AluSp, D1S2217	<pre>\$feature-&gt;display_id</pre>	_ chromosome relative
coordinates		Sfeature->seq_region_name Sfeature->start Steature->end Sfeature->seq_region_start Sfeature->sed_region_end Sfeature->strand	
sequence		\$feature->seq	
length	399	\$feature->length	



# Appendix – Gene Methods

Attribute	Example value(s)	Method(s)
stable ID	ENSG00000139618	\$gene->stable_id
name	BRCA2	\$gene->external_name
description	breast cancer 2, early onset	\$gene->description
biotype	protein_coding, miRNA	\$gene->biotype
analysis	ensembl, havana, ensembl_havana_gene	\$gene->analysis->logic_name

# **Appendix - Transcript Methods**

Attribute	Example value(s)	Method(s)
stable ID	ENST00000380152	<pre>\$transcript-&gt;stable_id</pre>
name	BRCA2-001	<pre>\$transcript-&gt;external_name</pre>
biotype	protein_coding honsense_mediated_decay	<pre>\$transcript-&gt;biotype</pre>
analysis	ensembl, havana ensembl_havana_transcript	<pre>\$transcript-&gt;analysis-&gt; logic_name</pre>
status	KNOWN, NOVEL	\$transcript->status
\		



# **Appendix - Translation Methods**

Attribute	Example value(s)	Method(s)
stable id	ENSP00000369497	<pre>\$translation-&gt;stable_id</pre>
length	3418	\$translation->length
sequence	MPIGSKERPTFFEIFKTRCNKADLG	\$translation->seq



