

## Recap From Last Time:

- Bioinformatics is computer aided biology.
  - Deals with the collection, archiving, organization, and interpretation of a wide range of biological data.
- The NCBI and EBI are major online bioinformatics service providers.
- Introduced via **hands-on session** the BLAST, Entrez, GENE, OMIM, UniProt, Muscle and PDB bioinformatics tools and databases.
  - Muddy point assessment (see [results](#))
- There are a large number of bioinformatics databases (see [handout!](#)!).
- Also covered: Course structure; Supporting course website, Ethics code, and Introductions...

## Today's Menu

<b>Classifying Databases</b>	Primary, secondary and composite Bioinformatics databases
<b>Using Databases</b>	<b>Vignette</b> demonstrating how major Bioinformatics databases intersect
<b>Major Biomolecular Formats</b>	How nucleotide and protein sequence and structure data are represented
<b>Alignment Foundations</b>	<b>Introducing the why and how of comparing sequences</b>
<b>Alignment Algorithms</b>	<b>Hands-on</b> exploration of alignment algorithms and applications

## Primary, secondary & composite databases

Bioinformatics databases can be usefully classified into *primary*, *secondary* and *composite* according to their data source.

- **Primary databases** (or *archival databases*) consist of data derived experimentally.
  - **GenBank**: NCBI's primary nucleotide sequence database.
  - **PDB**: Protein X-ray crystal and NMR structures.
- **Secondary databases** (or *derived databases*) contain information derived from a primary database.
  - **RefSeq**: non redundant set of curated reference sequences primarily from GenBank
  - **PFAM**: protein sequence families primarily from UniProt and PDB
- **Composite databases** (or *metadatabases*) join a variety of different primary and secondary database sources.
  - **OMIM**: catalog of human genes, genetic disorders and related literature
  - **GENE**: molecular data and literature related to genes with extensive links to other databases.

# DATABASE VIGNETTE

You have just come out a seminar about gastric cancer and one of your co-workers asks:

**"What do you know about that 'Kras' gene the speaker kept taking about?"**

You have some recollection about hearing of 'Ras' before. How would you find out more?

- Google?
- Library?
- **Bioinformatics databases at NCBI and EBI!**

<http://www.ncbi.nlm.nih.gov/>

<http://www.ncbi.nlm.nih.gov/>

The screenshot shows the NCBI homepage with a search bar containing 'ras'. The search results are displayed on the right, including sections for Genotypes and Phenotypes and NCBI Announcements.

## Example Vignette Questions:

- What chromosome location and what genes are in the vicinity of a given query gene? **NCBI GENE**
- What can you find out about molecular functions, biological processes, and prominent cellular locations? **EBI GO**
- What amino acid positions in the protein are responsible for ligand binding? **EBI UniProt**
- What variants of this gene are associated with gastric cancer and other human diseases? **NCBI OMIN**
- What is known about the protein family, its species distribution, number in humans and residue-wise conservation? **EBI PFAM**
- Are high resolution protein structures available to examine the details of these mutations? How might we explain their potential molecular effects? **RCSB PDB**

The screenshot shows the search results for 'ras' on the NCBI Global Cross Query page. The results are categorized by database type, such as Literature, Genes, Health, and Proteins. The 'Genes' section is highlighted with a red box around the 'Gene' entry, which has a count of 87,165.

[www.ncbi.nlm.nih.gov/gene/?term=ras](http://www.ncbi.nlm.nih.gov/gene/?term=ras)

Gene Search: ras

Display Settings: Tabular, 20 per page, Sorted by Relevance

Filters: Manage Filters

Top Organisms: Homo sapiens (1126)

Results: 1 to 20 of 85633

Name/Gene ID	Description	Location	Aliases
ras	resistance to audiogenic seizures [Mus musculus (house mouse)]		asr
ras	rasberry [Drosophila melanogaster (fruit fly)]	Chromosome X, NC_004354.4 (10744502..10749097)	Dmel_CG1799, CG11485, CG1799, DmelCG1799, EP(X)1093,

Find related data

Search details: ras[All Fields] AND alive[property]

[www.ncbi.nlm.nih.gov/gene/?term=\(ras\)&term=\(Homo+sapiens\)\[porgn:\\_txid9606\]](http://www.ncbi.nlm.nih.gov/gene/?term=(ras)&term=(Homo+sapiens)[porgn:_txid9606])

Gene Search: (ras) AND "Homo sapiens"[porgn:\_txid9606]

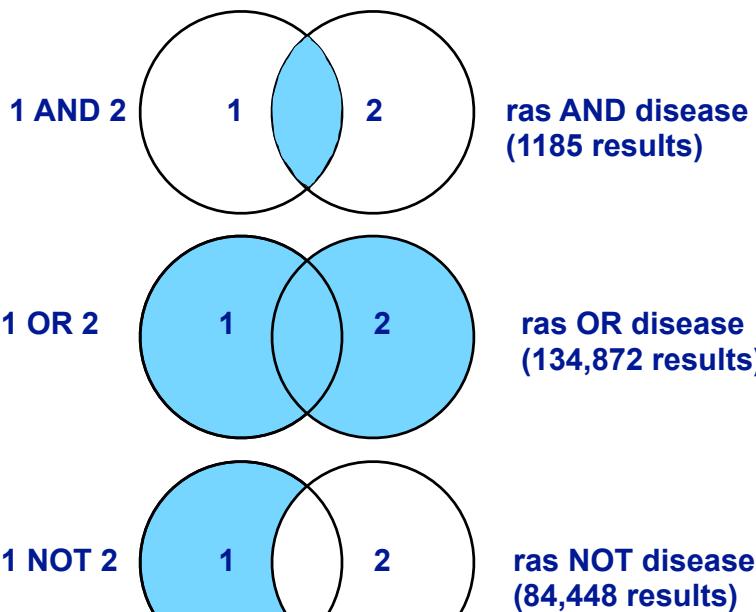
Display Settings: Tabular, 20 per page, Sorted by Relevance

Results: 1 to 20 of 1126

Name/Gene ID	Description	Location	Aliases
NRAS	neuroblastoma RAS viral (v-ras) oncogene homolog [Homo sapiens (human)]	Chromosome 1, NC_000001.11 (11470446..114716894, complement)	RP5-1000E10.2, ALPS4, CMNS, N-ras, NCMS1, NS6, NRAS
KRAS	Kirsten rat sarcoma viral oncogene homolog [Homo sapiens (human)]	Chromosome 12, NC_000012.12 (25205246..25250923, complement)	C-K-RAS, CFC2, K-RAS2A, K-RAS2B, K-RAS4A, K-RAS4B, K-RAS1, KRAS2, NS, NS2, RAS2

Find related data

Search details: ras[All Fields] AND "Homo sapiens"[porgn] AND alive[property]



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[www.ncbi.nlm.nih.gov/gene/?term=\(ras\)&term=\(Homo+sapiens\)\[porgn:\\_txid9606\]](http://www.ncbi.nlm.nih.gov/gene/?term=(ras)&term=(Homo+sapiens)[porgn:_txid9606])

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Find related data

Search details: ras[All Fields] AND "Homo sapiens"[porgn] AND alive[property]

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KRAS Kirsten rat sarcoma viral oncogene homolog [Homo sapiens (human)]

Gene ID: 3845, updated on 4-Jan-2015

**Summary**

Official Symbol KRAS provided by HGNC  
 Official Full Name Kirsten rat sarcoma viral oncogene homolog provided by HGNC  
 Primary source HGNC:HGNC:6407  
 See related Ensembl:ENSG0000133703; HPRD:01817; MIM:190070; Vega:OTTHUMG00000171193

Gene type protein coding  
 RefSeq status REVIEWED  
 Organism Homo sapiens  
 Lineage Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo  
 Also known as NS; NS3; CFC2; KRAS1; KRAS2; RASK2; KI-RAS; C-K-RAS; K-RAS2A; K-

**Table of contents**

- Summary
- Genomic context
- Genomic regions, transcripts, and products
- Bibliography
- Phenotypes
- Variation
- HIV-1 interactions
- Pathways from BioSystems
- Interactions
- General gene information
- Markers, Related pseudogene(s), Homology, Gene Ontology
- General protein information
- NCBI Reference Sequences (RefSeq)

**Example Questions:**  
 What chromosome location and what genes are in the vicinity?

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**Genomic context**

Location: 12p12.1  
 Exon count: 6

Annotation release	Status	Assembly	Chr	Location
106	current	GRCh38 (GCF_000001405.26)	12	NC_000012.12 (2505246..25250923, complement)
105	previous assembly	GRCh37.p13 (GCF_000001405.25)	12	NC_000012.11 (2535180..25403870, complement)

**Chromosome 12 - NC\_000012.12**

**Genomic regions, transcripts, and products**

Genomic Sequence: NC\_000012.12 chromosome 12 reference GRCh38 Primary Assembly  
 Go to nucleotide: Graphics Fasta GenBank

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**Example Questions:**  
 What 'molecular functions', 'biological processes', and 'cellular component' information is available?

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Function	Evidence Code	Pubs
GDP binding	IEA	
GMP binding	IEA	
GTP binding	IEA	
LRR domain binding	IEA	
protein binding	IPI	PubMed
protein complex binding	IDA	PubMed

Process	Evidence Code	Pubs
Fc-epsilon receptor signaling pathway	TAS	
GTP catabolic process	IEA	
MAPK cascade	TAS	
Ras protein signal transduction	TAS	
actin cytoskeleton organization	IEA	
activation of MAPKK activity	TAS	
axon guidance	TAS	
blood coagulation	TAS	

## GO: Gene Ontology

GO provides a controlled vocabulary of terms for describing gene product characteristics and gene product annotation data

The UniProt GO annotation program aims to provide high-quality Gene Ontology (GO) annotations to proteins in the UniProt Knowledgebase (UniProtKB). The assignment of GO terms to UniProt records is an integral part of UniProt biocuration. UniProt manual and electronic GO annotations are supplemented with manual annotations supplied by external collaborating GO Consortium groups, to ensure a comprehensive GO annotation dataset is supplied to users.

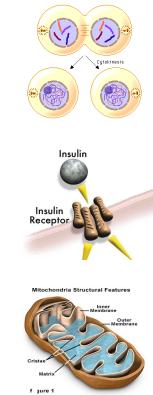
UniProt is a member of the GO Consortium.

## Why do we need Ontologies?

- Annotation is essential for capturing the understanding and knowledge associated with a sequence or other molecular entity
- Annotation is traditionally recorded as “free text”, which is easy to read by humans, but has a number of disadvantages, including:
  - Difficult for computers to parse
  - Quality varies from database to database
  - Terminology used varies from annotator to annotator
- Ontologies are annotations using standard vocabularies that try to address these issues
- GO is integrated with UniProt and many other databases including a number at NCBI

## GO Ontologies

- There are three ontologies in GO:
  - Biological Process**  
A commonly recognized series of events e.g. cell division, mitosis,
  - Molecular Function**  
An elemental activity, task or job e.g. kinase activity, insulin binding
  - Cellular Component**  
Where a gene product is located e.g. mitochondrion, mitochondrial membrane



The 'Gene Ontology' or GO is actually maintained by the EBI so lets switch or link over to UniProt also from the EBI.

UniProt will detail much more information for protein coding genes such as this one

UniProtKB/Swiss-Prot:P0116

Scroll down to Very bottom for UniProt link

UniProt will detail much more information for protein coding genes

P0116 - RASK\_HUMAN

Protein | GTPase KRas  
Gene | KRAS  
Organism | Homo sapiens (Human)  
Status | Reviewed - ●●●●● - Experimental evidence at protein level<sup>1</sup>

Display | None

FUNCTION NAMES & TAXONOMY SUBCELL LOCATION PATHOL/BIOTECH PTM / PROCESSING EXPRESSION INTERACTION STRUCTURE FAMILY & DOMAINS SEQUENCES (2) CROSS-REFERENCES

Function

Ras proteins bind GDP/GTP and possess intrinsic GTPase activity. Plays an important role in the regulation of cell proliferation (PubMed:23698361, PubMed:22711838). 2 Publications Curated

Enzyme regulation Alternates between an inactive form bound to GDP and an active form bound to GTP. Activated by a guanine nucleotide-exchange factor (GEF) and inactivated by a GTPase-activating protein (GAP). Interaction with SOS1 promotes exchange of bound GDP by GTP. 3 Publications

Regions

Feature key	Position(s)	Length	Description	Graphical view	Feature identifier	Actions
Nucleotide binding <sup>i</sup>	10 – 18	9 GTP	2 Publications			
Nucleotide binding <sup>i</sup>	29 – 35	7 GTP	2 Publications			
Nucleotide binding <sup>i</sup>	59 – 60	2 GTP	2 Publications			

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>sp|P0116|RASK\_HUMAN GTPase KRas OS=Homo sapiens GN=KRAS PE=1 SV=1 MTEYKLVVVGGAGGVGKSAITIQLIQNHFDEYDPTIEDSYRSRQVWVLDGETCLLDILDTAG QQEYSAMRDQYMTRTEGFLCVFAINTNKSFEDIHRYRQEIKRKVKSDEDVPVMVLGNKCDL PSEPVVDTRQADLARSYGIPPIETTSAKTRQRQVEDAFYTTLVREIRQYRLKKISKEEKTFCG VKIKRCIM

View FASTA file format

UniProt will detail much more information for protein coding genes

**Example Questions:**  
What variants of this enzyme are involved in gastric cancer and other human diseases?

**Example Questions:**  
Are high resolution protein structures available to examine the details of these mutations?

Open link in a new tab!

**Lets view the 3D structure:**  
Can we find where in the structure our mutations are located and infer their potential molecular effects?

**4EPV**  
Discovery of Small Molecules that Bind to K-Ras and Inhibit Sos-mediated Activation  
DOI: 10.2210/pdb4epv/pdb  
Classification: HYDROLASE  
Deposited: 2012-04-17 Released: 2012-05-23  
Deposition author(s): Sun, Q., Burke, J.P., Phan, J., Burns, M.C., Olejniczak, E.T., Waterson, A.G., Lee, T., Rossanese, O.W., Fesik, S.W.  
Organism: Homo sapiens  
Expression System: Escherichia coli  
Mutation(s): 1  
Experimental Data Snapshot    wwPDB Validation    3D Report    Full Report  
Method: X-RAY DIFFRACTION    Metric    Percentile Ranks    Value

**Lets view the 3D structure:**  
Can we find where in the structure our mutations are located and infer their potential molecular effects?

**4EPV**  
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Note: Use your mouse to drag, rotate, and zoom in and out of the structure. Click to identify atoms and bonds.  
Bond: [GLY]12:A-O - [GLY]12:A-C

**Display Options**  
Assembly: Biossembly 1  
Model: Model 1  
Symmetry: None  
Interaction: IGDPJ201-A  
Style: Cartoon  
Color: Rainbow  
Ligand: None  
Quality: Automatic  
Water:  Ions:   
Hydrogens:  Clashes:

**Back to UniProt:**  
What is known about the protein family, its species distribution, number in humans and residue-wise conservation, etc... ?

**FAMILY & DOMAINS**  
PFAM: PF00071: Ras. 1 hit. [Graphical view]  
PRINTS: PR00449. RASTRNSFRMNG.  
SMART: SM00173. RAS. 1 hit. [Graphical view]  
SUPFAM: SSF52540. SSF52540. 1 hit.  
TIGRFAMS: TIGR00231. small\_GTP. 1 hit.  
PROSITE: PS51421. RAS. 1 hit. [Graphical view]

**PFAM is one of the best protein family databases**

**Example Questions:**  
What is known about the protein family, its **species distribution**, number in humans and residue-wise conservation, etc... ?

**Family: Ras (PF00071)**  
Summary Domain organisation Clan Alignments HMM logo Trees Curation & model Species Interactions Structures  
Jump to... enter ID/acc Go  
1 History 2 Structure 3 Function 3.1 Activation and deactivation 4 Membrane attachment 4 Members 5 Ras in cancer 5.1 Inappropriate activation 5.2 Constitutively active Ras  
Contents [Read]  
Symbol Ras  
Pfam PF00071.0  
InterPro IPR013753.0  
PROSITE POC00017.0  
SCOP Sp21.0  
SUPERFAMILY Sp21.0

## Example Questions:

What is known about the protein family, its **species distribution, number in humans** and residue-wise conservation, etc... ?

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UniProt entry	UniProt residues	PDB ID	PDB chain ID	PDB residues	View
ABBKD1_GIALA	11 - 335	2vvg	A	11 - 335	<a href="#">Jmol AstexViewer SPICE</a>
			B	11 - 335	<a href="#">Jmol AstexViewer SPICE</a>
CENPE_HUMAN	12 - 329	1t5c	A	12 - 329	<a href="#">Jmol AstexViewer SPICE</a>
			B	12 - 329	<a href="#">Jmol AstexViewer SPICE</a>
KAR3_YEAST	392 - 723	1f9t	A	392 - 723	<a href="#">Jmol AstexViewer SPICE</a>
			A	392 - 723	<a href="#">Jmol AstexViewer SPICE</a>
			19u	392 - 723	<a href="#">Jmol AstexViewer SPICE</a>
			19v	392 - 723	<a href="#">Jmol AstexViewer SPICE</a>
			19w	392 - 723	<a href="#">Jmol AstexViewer SPICE</a>
			3kar	392 - 723	<a href="#">Jmol AstexViewer SPICE</a>
KI13B_HUMAN	11 - 352	3gbi	A	11 - 352	<a href="#">Jmol AstexViewer SPICE</a>
			B	11 - 352	<a href="#">Jmol AstexViewer SPICE</a>
			C	11 - 352	<a href="#">Jmol AstexViewer SPICE</a>
			1i16	24 - 359	<a href="#">Jmol AstexViewer SPICE</a>
			A	24 - 359	<a href="#">Jmol AstexViewer SPICE</a>
			B	24 - 359	<a href="#">Jmol AstexViewer SPICE</a>
			A	24 - 359	<a href="#">Jmol AstexViewer SPICE</a>
			B	24 - 359	<a href="#">Jmol AstexViewer SPICE</a>
			A	24 - 359	<a href="#">Jmol AstexViewer SPICE</a>
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<b>Alignment Algorithms</b>	<b>Hands-on</b> exploration of alignment algorithms and applications

## ALIGNMENT FOUNDATIONS

- **Why...**
  - Why compare biological sequences?
- **What...**
  - Alignment view of sequence changes during evolution (matches, mismatches and gaps)
- **How...**
  - Dot matrices
  - Dynamic programming
    - Global alignment
    - Local alignment
  - BLAST heuristic approach

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Basic Idea: Display one sequence above another with spaces (termed gaps) inserted in both to reveal similarity of nucleotides or amino acids.

**Seq1:** C A T T C A C

**Seq2:** C T C G C A G C

[Screencast Material]

Basic Idea: Display one sequence above another with spaces (termed gaps) inserted in both to reveal similarity of nucleotides or amino acids.

**Seq1:** C A T T C A C

**Seq2:** C T C G C A G C

mismatch  
match

Two types of character correspondence

Basic Idea: Display one sequence above another with spaces (termed gaps) inserted in both to reveal similarity of nucleotides or amino acids.

**Seq1:** C A T - T C A - C

**Seq2:** C - T C G C A G C

match  
mismatch  
gaps

Add gaps to increase number of matches

Basic Idea: Display one sequence above another with spaces (termed gaps) inserted in both to reveal similarity of nucleotides or amino acids.

**Seq1:** C A T - T C A - C

**Seq2:** C - T C G C A G C

match  
mismatch } mutation  
insertion  
deletion } indels

Gaps represent 'indels'  
mismatch represent mutations

## Why compare biological sequences?

- To obtain functional or mechanistic insight about a sequence by inference from another potentially better characterized sequence
- To find whether two (or more) genes or proteins are evolutionarily related
- To find structurally or functionally similar regions within sequences (e.g. catalytic sites, binding sites for other molecules, etc.)
- Many practical bioinformatics applications...

## Practical applications include...

- Similarity searching of databases
  - Protein structure prediction, annotation, etc...
- Assembly of sequence reads into a longer construct such as a genomic sequence
- Mapping sequencing reads to a known genome
  - "Resequencing", looking for differences from reference genome - SNPs, indels (insertions or deletions)
  - Mapping transcription factor binding sites via ChIP-Seq (chromatin immuno-precipitation sequencing)
  - Pretty much all next-gen sequencing data analysis

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N.B. Pairwise sequence alignment is arguably the most fundamental operation of bioinformatics!

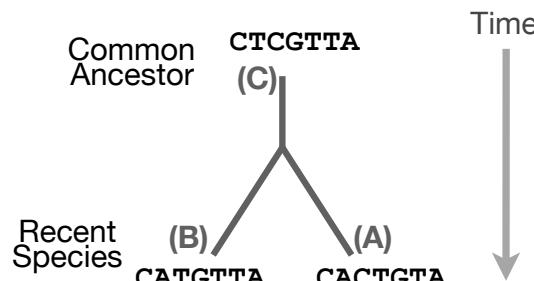
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## Sequence changes during evolution

There are three major types of sequence change that can occur during evolution.

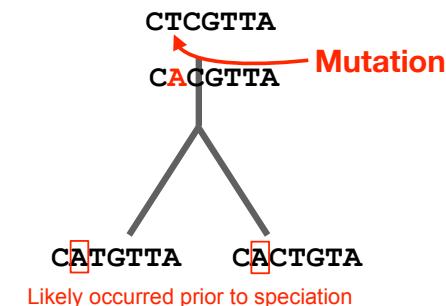
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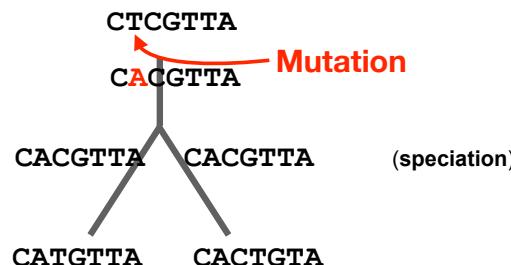
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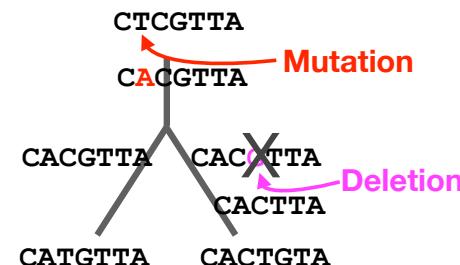
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- Insertions

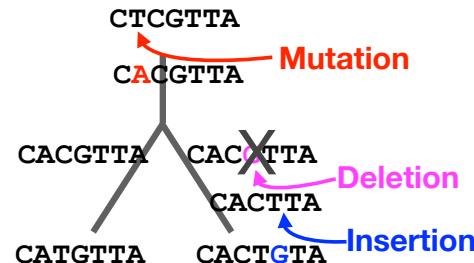


## Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- Mutations/Substitutions
- Deletions
- Insertions

CTCGTTA → CACGTTA  
CACGTTA → CACTTA  
CACTTA → CACTGTA

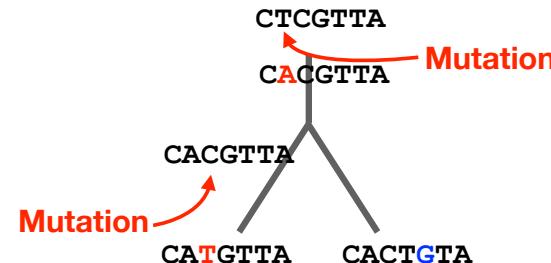


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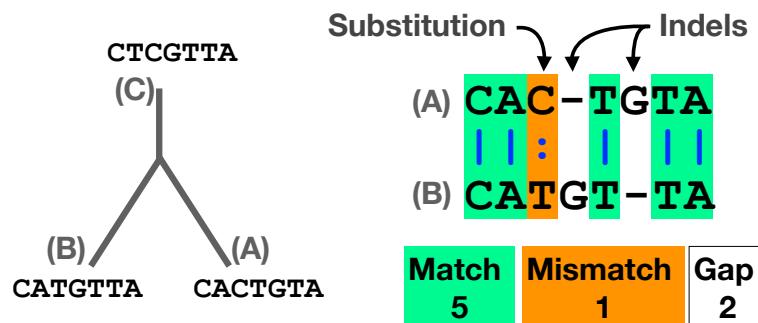
CTCGTTA → CACGTTA  
CACGTTA → CATGTTA



## Alignment view

Alignments are great tools to visualize sequence similarity and evolutionary changes in homologous sequences.

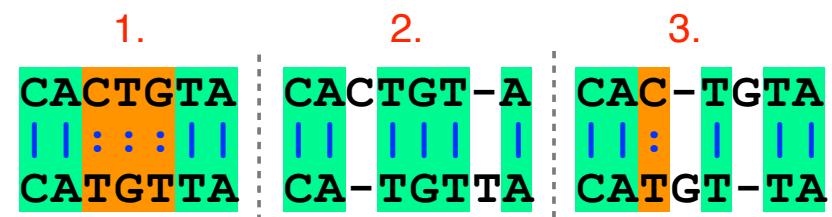
- Mismatches represent mutations/substitutions
- Gaps represent insertions and deletions (indels)



## Alternative alignments

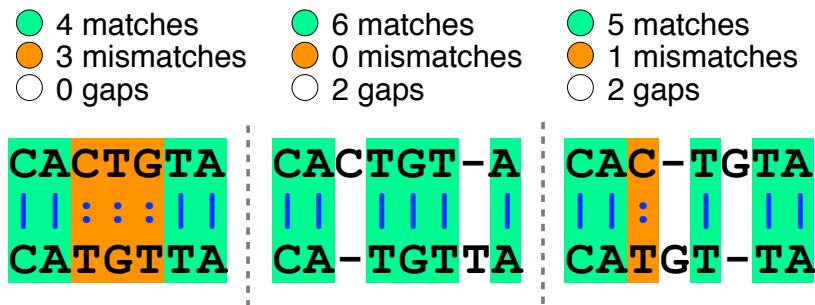
- Unfortunately, finding the correct alignment is difficult if we do not know the evolutionary history of the two sequences

Q. Which of these 3 possible alignments is best?



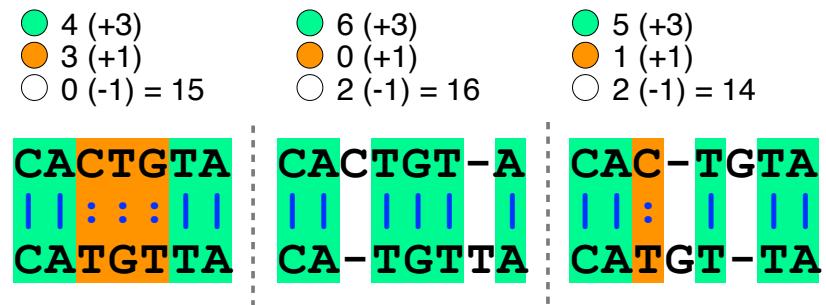
## Alternative alignments

- One way to judge alignments is to compare their number of matches, insertions, deletions and mutations



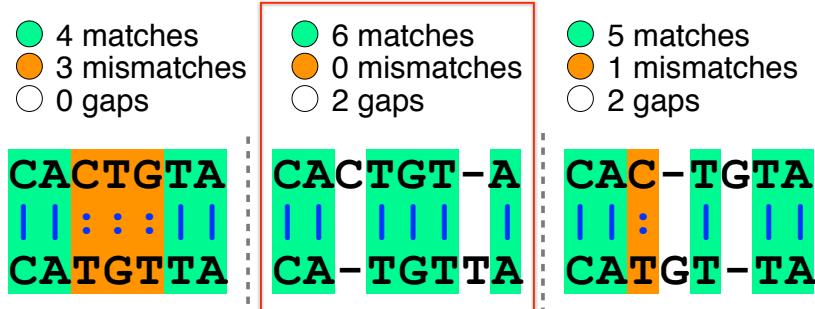
## Scoring alignments

- We can assign a score for each match (+3), mismatch (+1) and indel (-1) to identify the optimal alignment for this scoring scheme



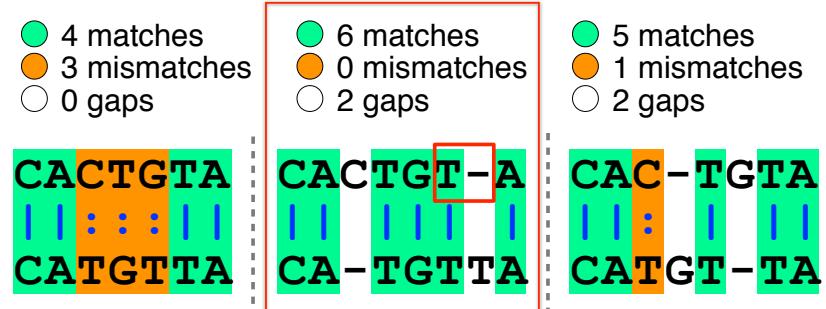
## Optimal alignments

- Biologists often prefer parsimonious alignments, where the number of postulated sequence changes is minimized.



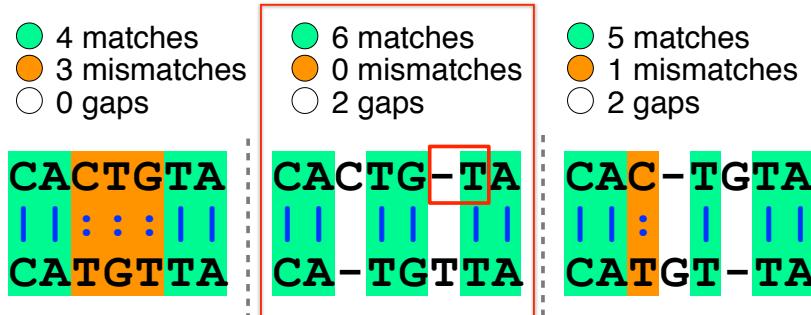
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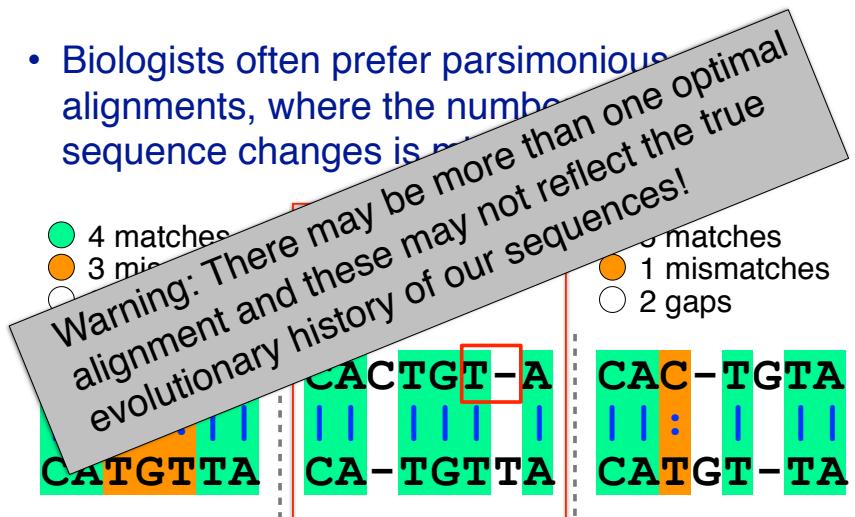
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## ALIGNMENT FOUNDATIONS

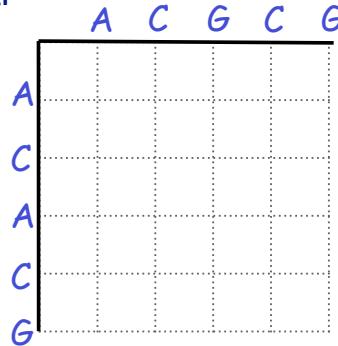
- Why...
  - Why compare biological sequences?
- What...
  - Alignment view of sequence changes during evolution (matches, mismatches and gaps)
- How...
  - Dot matrices
  - Dynamic programming
    - Global alignment
    - Local alignment
  - BLAST heuristic approach

## ALIGNMENT FOUNDATIONS

- Why...
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  - Dynamic programming
    - How do we compute the optimal alignment between two sequences?
  - BLAST heuristic approach

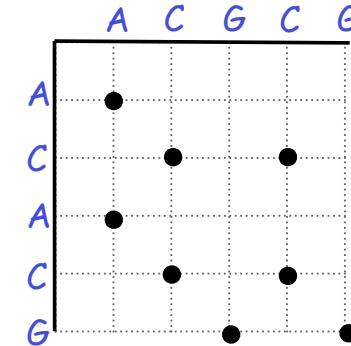
## Dot plots: simple graphical approach

- Place one sequence on the vertical axis of a 2D grid (or matrix) and the other on the horizontal



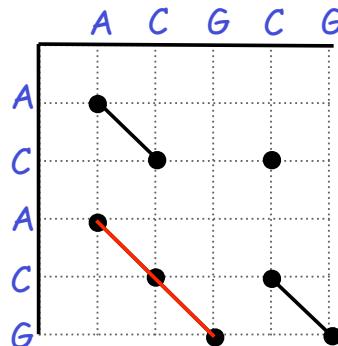
## Dot plots: simple graphical approach

- Now simply put dots where the horizontal and vertical sequence values match



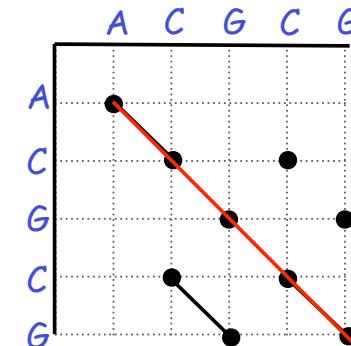
## Dot plots: simple graphical approach

- Diagonal runs of dots indicate matched segments of sequence



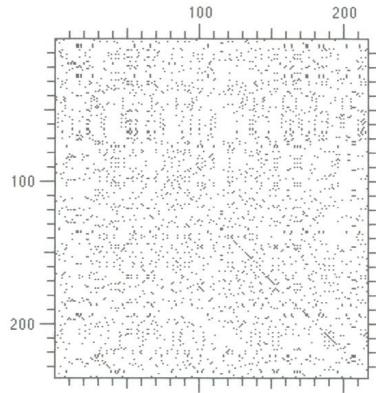
## Dot plots: simple graphical approach

- Q. What would the dot matrix of a two identical sequences look like?



## Dot plots: simple graphical approach

- Dot matrices for long sequences can be noisy

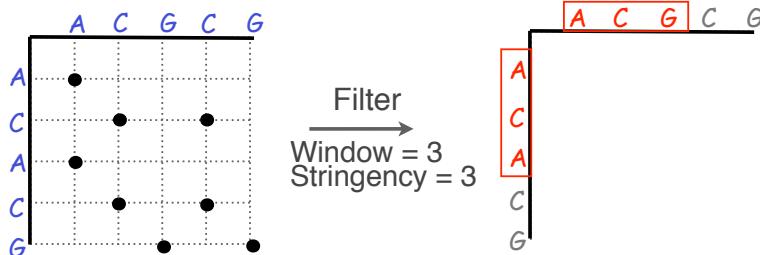


## Dot plots: window size and match stringency

Solution: use a window and a threshold

- compare character by character within a window
- require certain fraction of matches within window in order to display it with a dot.

- You have to choose window size and stringency

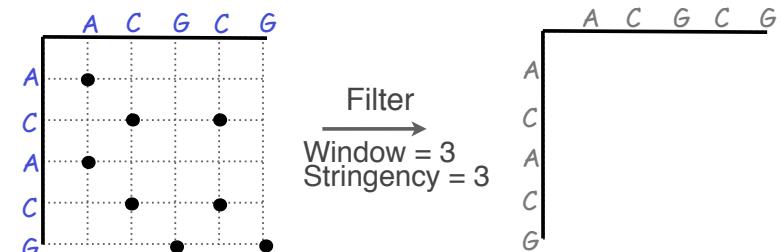


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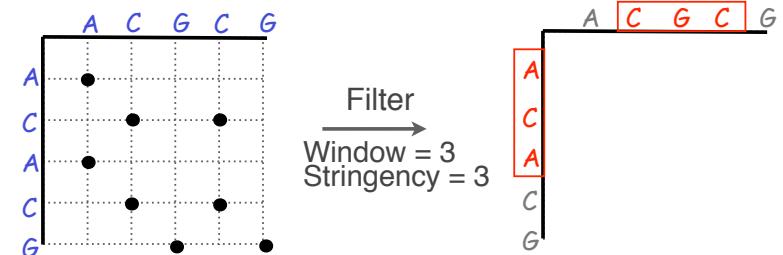


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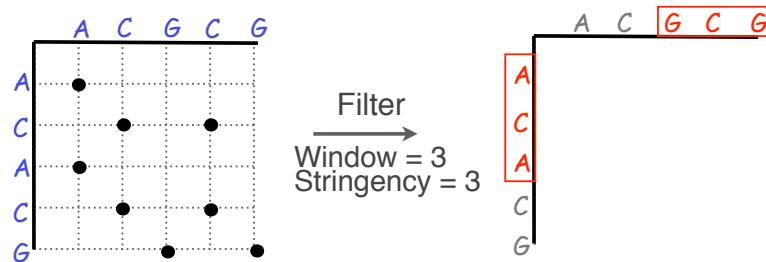
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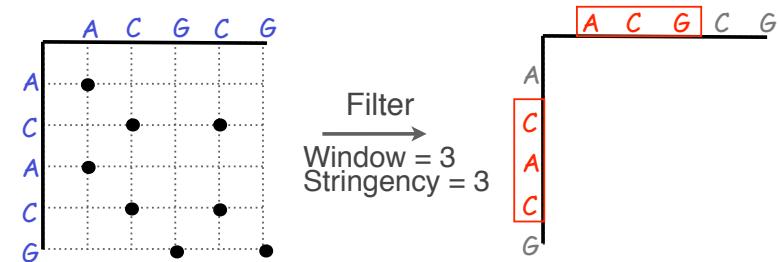
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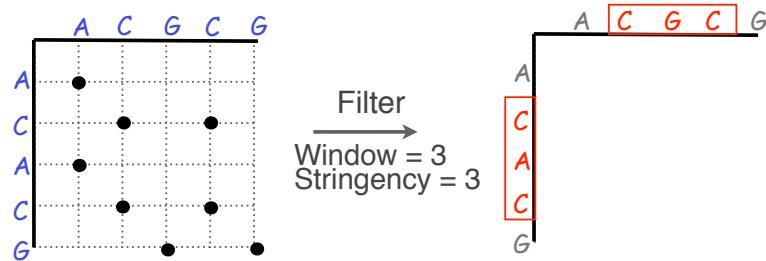
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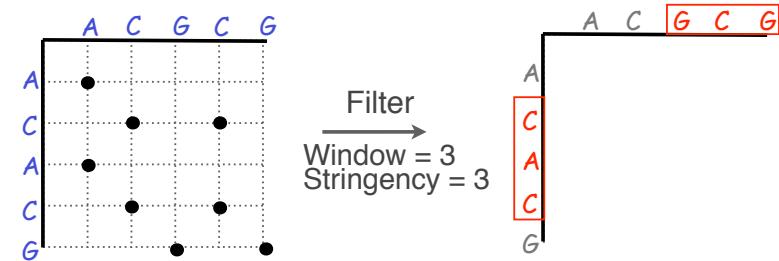
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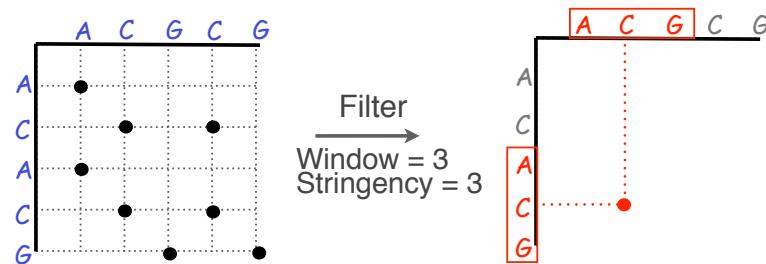
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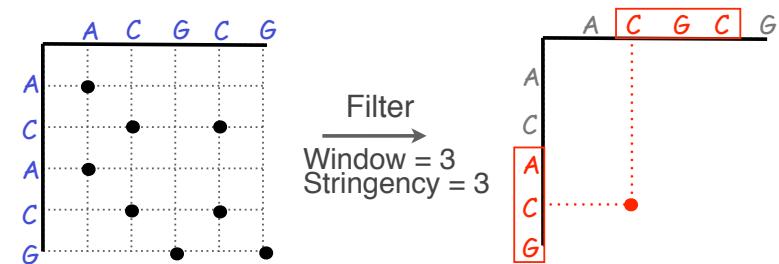
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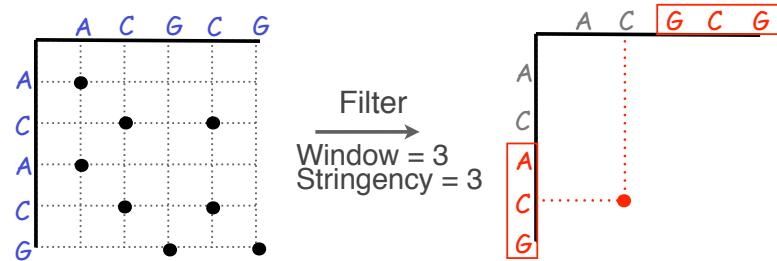
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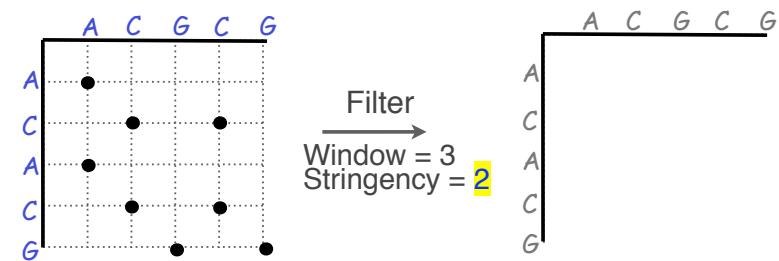
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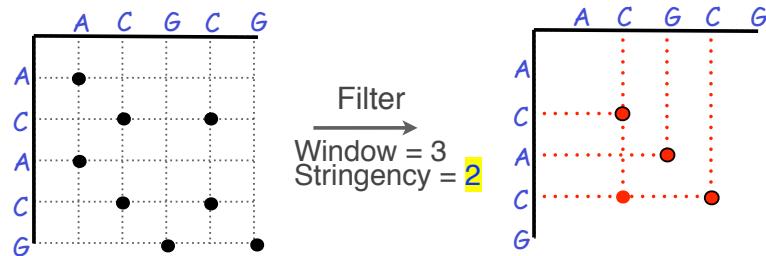
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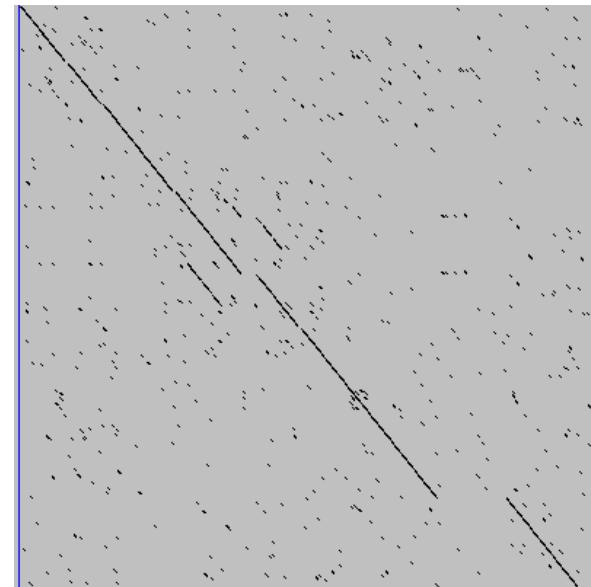
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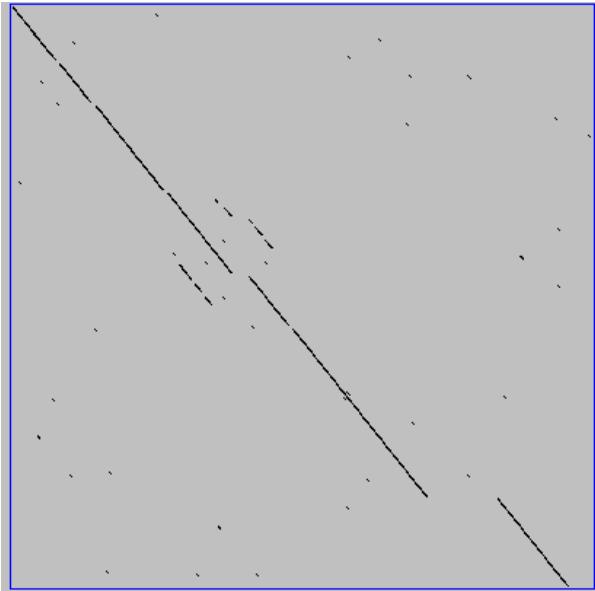
## Window size = 5 bases



A dot plot simply puts a dot where two sequences match. In this example, dots are placed in the plot if 5 bases in a row match perfectly. Requiring a 5 base perfect match is a **heuristic** – only look at regions that have a certain degree of identity.

Do you expect evolutionarily related sequences to have more word matches (matches in a row over a certain length) than random or unrelated sequences?

## Window size = 7 bases



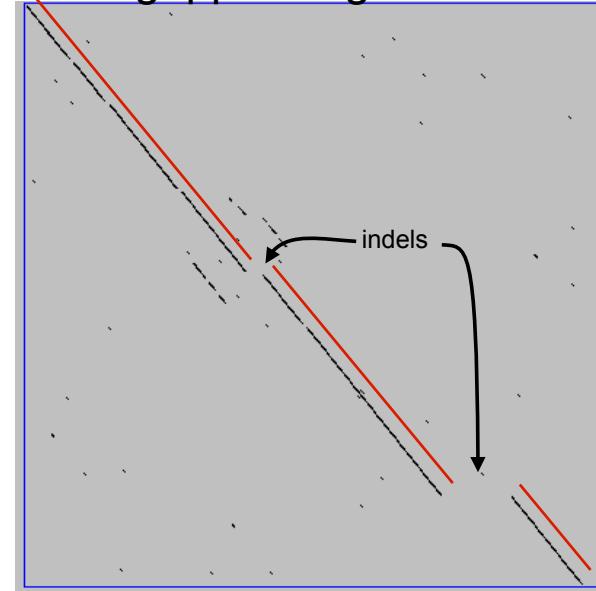
This is a dot plot of the same sequence pair. Now 7 bases in a row must match for a dot to be placed. Noise is reduced.

Using windows of a certain length is very similar to using words (kmers) of N characters in the heuristic alignment search tools

Bigger window (kmer)  
fewer matches to consider

Web site used: <http://www.vivo.colostate.edu/molkit/dnadot/>

## Ungapped alignments



Only **diagonals** can be followed.

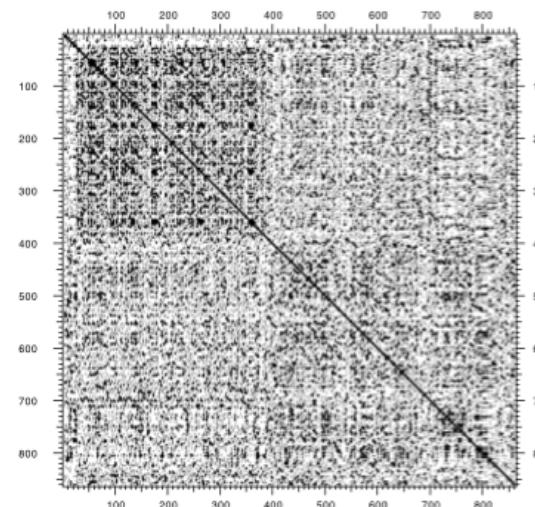
Downward or rightward paths represent **insertion** or **deletions** (gaps in one sequence or the other).

Web site used: <http://www.vivo.colostate.edu/molkit/dnadot/>

## Uses for dot matrices

- Visually assessing the similarity of two protein or two nucleic acid sequences
- Finding local repeat sequences within a larger sequence by comparing a sequence to itself
  - Repeats appear as a set of diagonal runs stacked vertically and/or horizontally

## Repeats

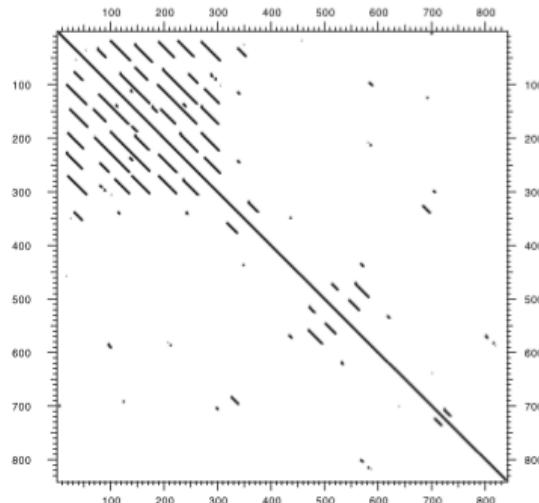


Human LDL receptor  
protein sequence  
(Genbank P01130)

$W = 1$   
 $S = 1$

(Figure from Mount, "Bioinformatics sequence and genome analysis")

## Repeats



Human LDL receptor  
protein sequence  
(Genbank P01130)

$W = 23$   
 $S = 7$

(Figure from Mount, "Bioinformatics sequence and genome analysis")

## Your Turn!

Exploration of dot plot parameters (hands-on worksheet **Section 1**)

<http://bio3d.ucsd.edu/dotplot/>    <https://bioboot.shinyapps.io/dotplot/>

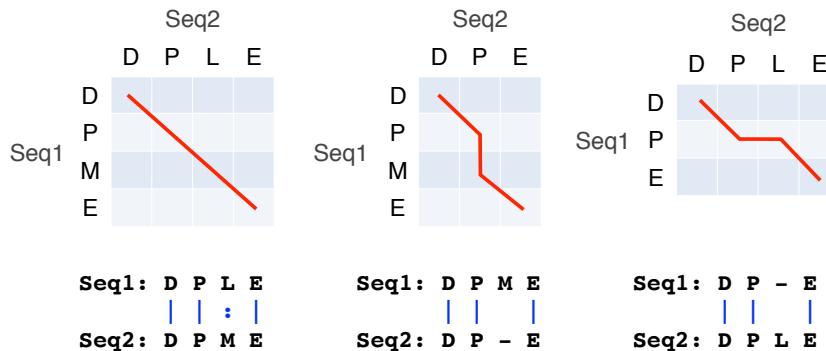
A screenshot of a web application titled 'BGGN-213: Dot Plot Comparison of Two Sequences'. The interface includes a 'Dot Plot Parameters' section with sliders for 'Window Size' (set to 3), 'Moving window step size' (set to 3), and 'Match stringency' (set to 2). To the right, there are two dot plots: a 'Protein Dot Plot' comparing Sequence 1 and Sequence 2, and a 'DNA Dot Plot' comparing Sequence 1 and Sequence 2. Both plots show a diagonal line of dots representing matches. At the bottom, there is a link to 'https://bioboot.shinyapps.io/dotplot2/'. A 'Questions for discussion:' section follows, containing three bullet points.

# ALIGNMENT FOUNDATIONS

- Why...
  - Why compare biological sequences?
- What...
  - Alignment view of sequence changes during evolution (matches, mismatches and gaps)

## • How...

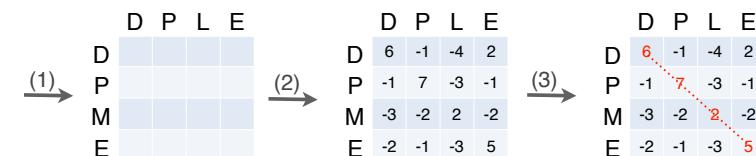
- Dot matrices
- Dynamic programming
  - Global alignment
  - Local alignment
- BLAST heuristic approach



Matches are represented by diagonal paths & indels with horizontal or vertical path segments

# The Dynamic Programming Algorithm

- The dynamic programming algorithm can be thought of an extension to the dot plot approach
  - One sequence is placed down the side of a grid and another across the top
  - Instead of placing a dot in the grid, we compute a score for each position
  - Finding the optimal alignment corresponds to finding the path through the grid with the best possible score

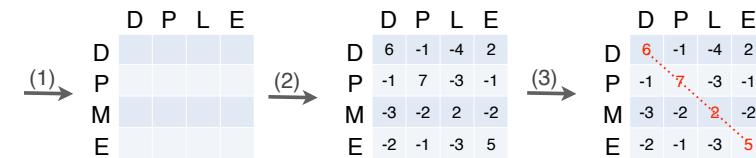


Needleman, S.B. & Wunsch, C.D. (1970) "A general method applicable to the search for similarities in the amino acid sequences of two proteins." J. Mol. Biol. 48:443-453.

90

# Algorithm of Needleman and Wunsch

- The Needleman–Wunsch approach to global sequence alignment has three basic steps:
  - (1) setting up a 2D-grid (or alignment matrix),
  - (2) scoring the matrix, and
  - (3) identifying the optimal path through the matrix



Needleman, S.B. & Wunsch, C.D. (1970) "A general method applicable to the search for similarities in the amino acid sequences of two proteins." J. Mol. Biol. 48:443-453.

## Scoring the alignment matrix

- Start by filling in the first row and column – these are all indels (gaps).
  - Each step you take you will add the gap penalty to the score ( $S_{i,j}$ ) accumulated in the previous cell

		Sequence 2				Scores: match = +1, mismatch = -1, gap = -2	
		-	D	P	L	E	
Sequence 1	-	0	-2	-4	-6	-8	
	D	-2					
	P	-4					
	M	-6					
	E	-8					

## Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
  - Now can ask which of the three directions gives the highest score?
  - keep track of this score and direction

		Sequence 2				Scores: match = +1, mismatch = -1, gap = -2	
		-	D	P	L	E	
Sequence 1	-	0	-2	-4	-6	-8	
	D	-2	?				
	P	-4					
	M	-6					
	E	-8					

The diagram illustrates the three possible paths to the cell  $S(i, j)$  from the cell  $S(i-1, j-1)$ . Path 1 is vertical (up), Path 2 is horizontal (left), and Path 3 is diagonal (up-left).

## Scoring the alignment matrix

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		Sequence 2				Scores: match = +1, mismatch = -1, gap = -2	
		-	D	P	L	E	
Sequence 1	-	0	-2	-4	-6	-8	
	D	-2					
	P	-4					
	M	-6					
	E	-8					

$$S_{i+4} = (-2) + (-2) + (-2) + (-2)$$

Seq1: DPME  
Seq2: -----

## Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
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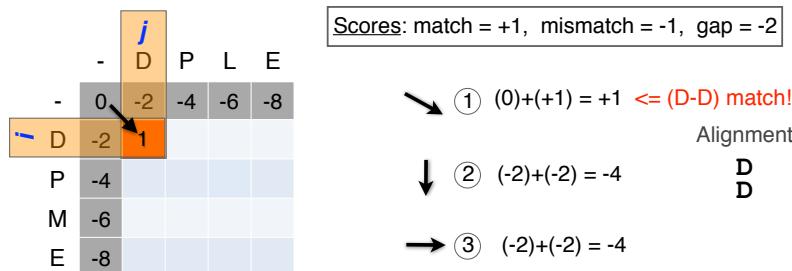
$$S(i, j) = \text{Max} \begin{cases} S(i-1, j-1) + (\text{mis})\text{match} \\ S(i-1, j) + \text{gap penalty} \\ S(i, j-1) + \text{gap penalty} \end{cases}$$

Diagram illustrating the scoring logic for cell  $S(i, j)$ :

- Path 1:  $S(i-1, j-1) + (\text{mis})\text{match}$  (labeled ①)
- Path 2:  $S(i-1, j) + \text{gap penalty}$  (labeled ②)
- Path 3:  $S(i, j-1) + \text{gap penalty}$  (labeled ③)

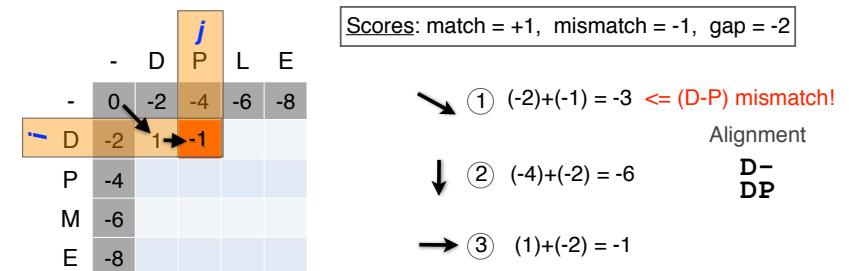
## Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
  - Now can ask which direction gives the highest score
  - keep track of direction and score



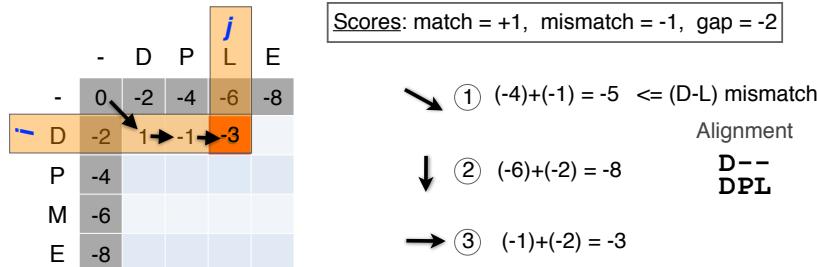
## Scoring the alignment matrix

- At each step, the score in the current cell is determined by the scores in the neighboring cells
  - The maximal score and the direction that gave that score is stored (we will use these later to determine the optimal alignment)



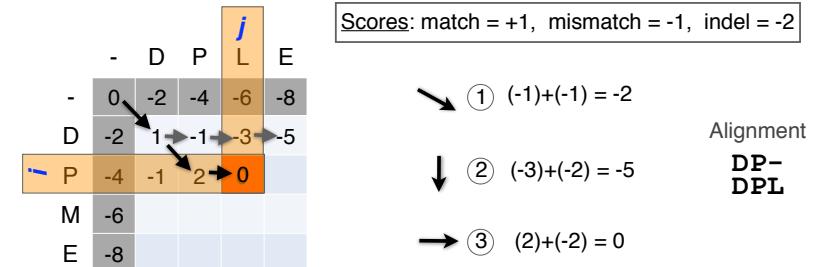
## Scoring the alignment matrix

- We will continue to store the alignment score ( $S_{i,j}$ ) for all possible alignments in the alignment matrix.



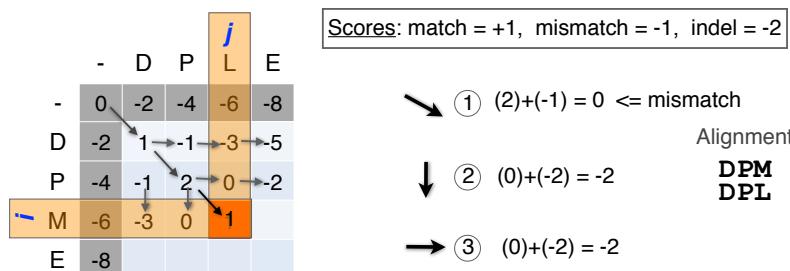
## Scoring the alignment matrix

- For the highlighted cell, the corresponding score ( $S_{i,j}$ ) refers to the score of the optimal alignment of the first  $i$  characters from sequence1, and the first  $j$  characters from sequence2.



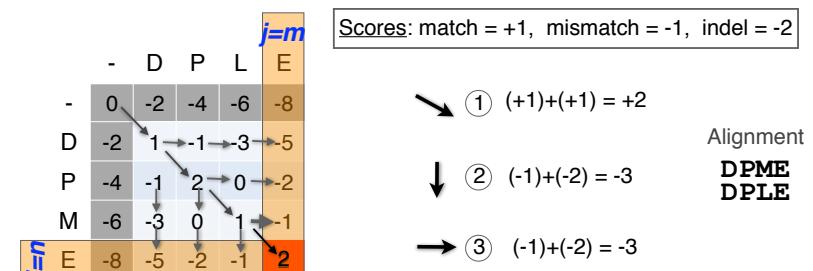
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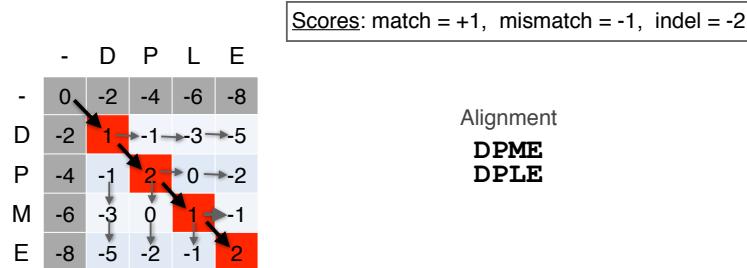
## Scoring the alignment matrix

- The score of the best alignment of the entire sequences corresponds to  $S_{n,m}$ 
  - (where n and m are the length of the sequences)



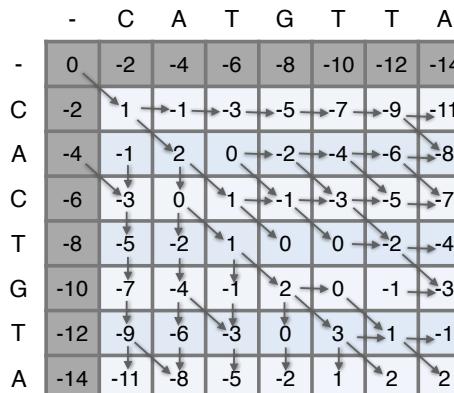
## Scoring the alignment matrix

- To find the best alignment, we retrace the arrows starting from the bottom right cell
  - N.B. The optimal alignment score and alignment are dependent on the chosen scoring system



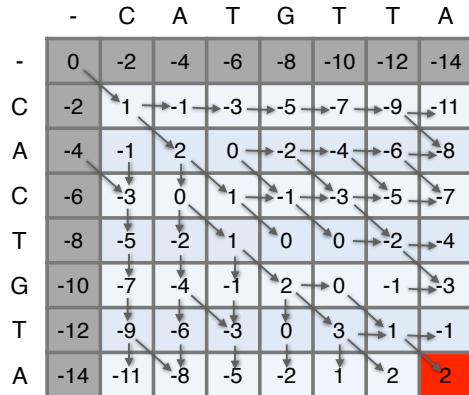
## Questions:

- What is the optimal score for the alignment of these sequences and how do we find the optimal alignment?



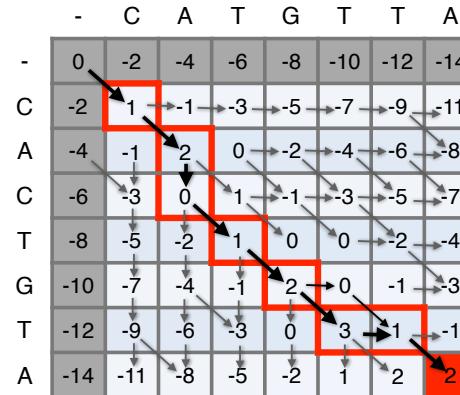
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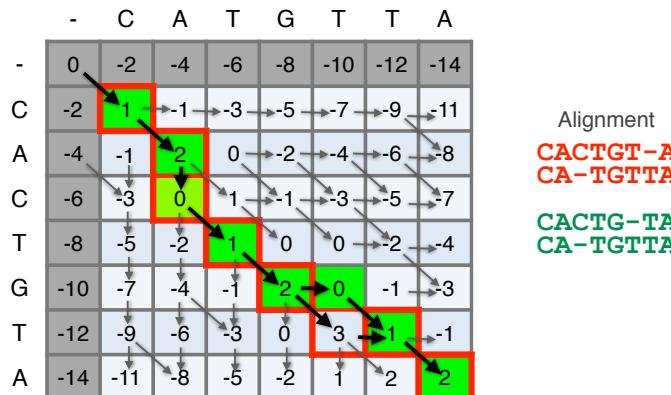
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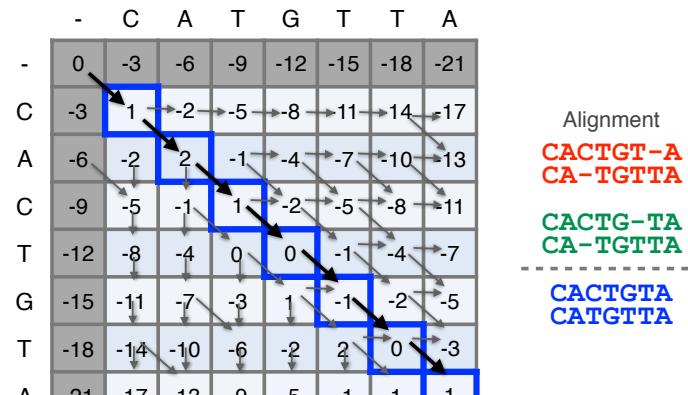
## More than one alignment possible

- Sometimes more than one alignment can result in the same optimal score



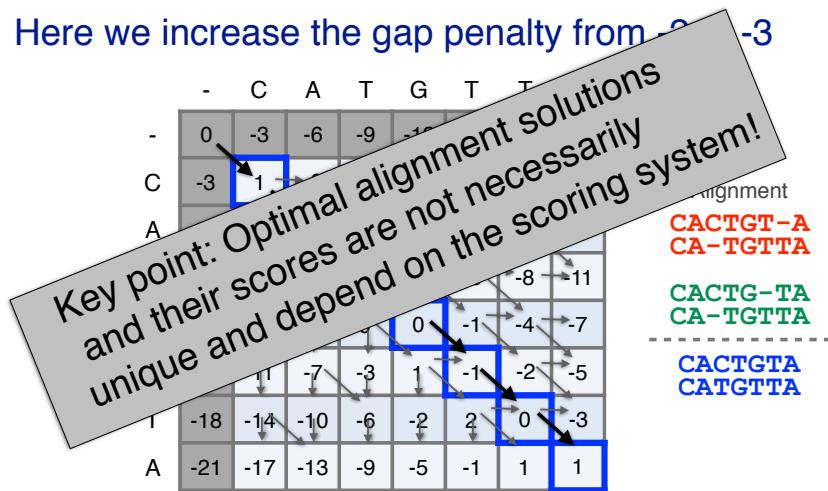
The alignment and score are dependent on the scoring system

- Here we increase the gap penalty from -2 to -3



## The alignment and score are dependent on the scoring system

- Here we increase the gap penalty from -2 to -3



## NW DYNAMIC PROGRAMMING

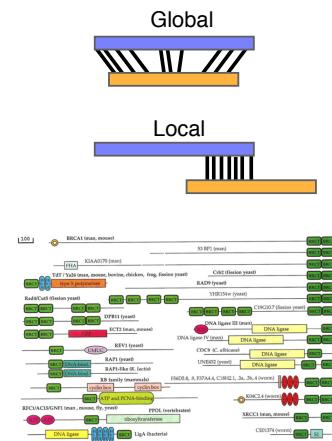
	A	G	T	T	C
A	0				
T					
T					
G					
C					

## ALIGNMENT FOUNDATIONS

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  - Dynamic programming
    - Global alignment
    - Local alignment
  - BLAST heuristic approach

## Global vs local alignments

- Needleman-Wunsch is a global alignment algorithm
  - Resulting alignment spans the complete sequences end to end
  - This is appropriate for closely related sequences that are similar in length
- For many practical applications we require local alignments
  - Local alignments highlight sub-regions (e.g. protein domains) in the two sequences that align well

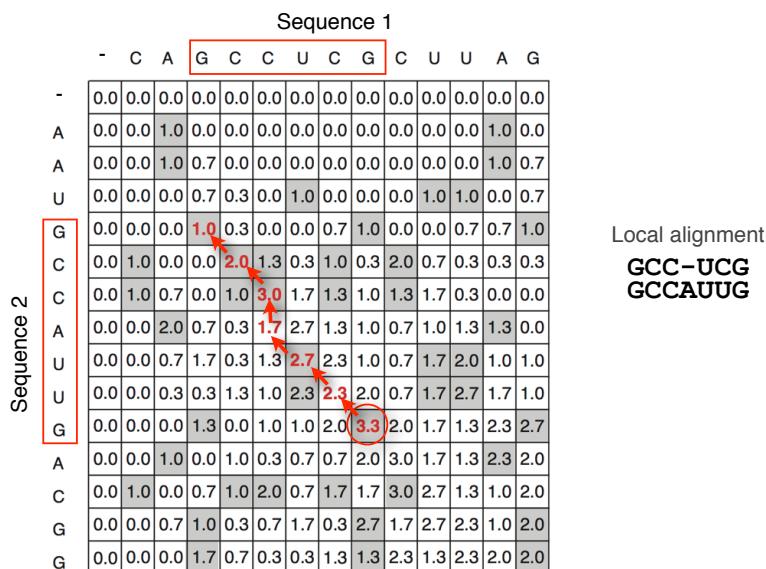


## Local alignment: Definition

- Smith & Waterman proposed simply that a local alignment of two sequences allow arbitrary-length segments of each sequence to be aligned, with no penalty for the unaligned portions of the sequences. Otherwise, the score for a local alignment is calculated the same way as that for a global alignment

Smith, T.F. & Waterman, M.S. (1981) "Identification of common molecular subsequences." J. Mol. Biol. 147:195-197.

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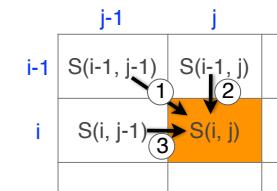


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## The Smith-Waterman algorithm

- Three main modifications to Needleman-Wunsch:
  - Allow a node to start at 0
  - The score for a particular cell cannot be negative
    - if all other score options produce a negative value, then a zero must be inserted in the cell
  - Record the highest-scoring node, and trace back from there

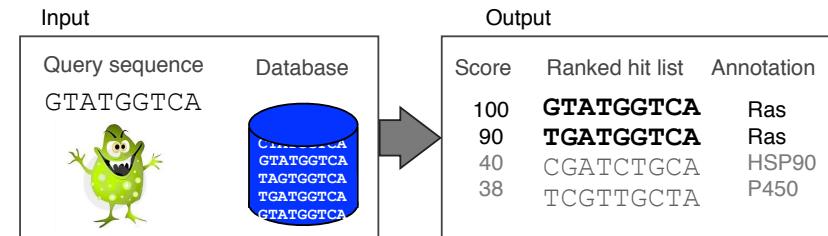
$$S(i, j) = \text{Max} \begin{cases} S(i-1, j-1) + (\text{mis})\text{match} \\ S(i-1, j) - \text{gap penalty} \\ S(i, j-1) - \text{gap penalty} \\ 0 \end{cases}$$



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## Local alignments can be used for database searching

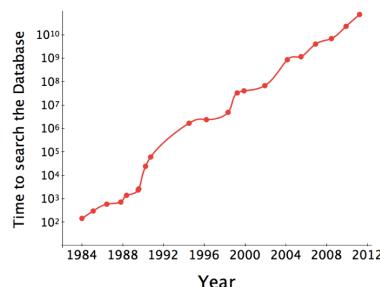
- Goal: Given a query sequence (Q) and a sequence database (D), find a list of sequences from D that are most similar to Q
  - Input: Q, D and scoring scheme
  - Output: Ranked list of hits



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## The database search problem

- Due to the rapid growth of sequence databases, search algorithms have to be both efficient and sensitive
  - Time to search with SW is proportional to  $m \times n$  ( $m$  is length of query,  $n$  is length of database), too slow for large databases!

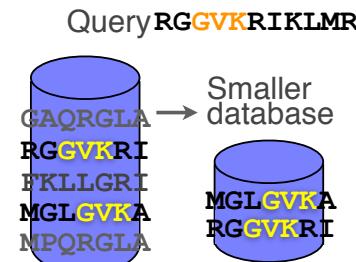


To reduce search time heuristic algorithms, such as BLAST, first remove database sequences without a strong local similarity to the query sequence in a quick initial scan.

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## The database search problem

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To reduce search time heuristic algorithms, such as BLAST, first remove database sequences without a strong local similarity to the query sequence in a quick initial scan.

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## ALIGNMENT FOUNDATIONS

- Why...
  - Why compare biological sequences?
- What...
  - Alignment view of sequence changes during evolution (matches, mismatches and gaps)
- How...
  - Dot matrices
  - Dynamic programming
    - Global alignment
    - Local alignment
  - BLAST heuristic approach

## Rapid, heuristic versions of Smith–Waterman: BLAST

- BLAST (Basic Local Alignment Search Tool) is a simplified form of Smith-Waterman (SW) alignment that is popular because it is fast and easily accessible
  - BLAST is a heuristic approximation to SW - It examines only part of the search space
  - BLAST saves time by restricting the search by scanning database sequences for likely matches before performing more rigorous alignments
  - Sacrifices some sensitivity in exchange for speed
  - In contrast to SW, BLAST is not guaranteed to find optimal alignments

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## Rapid, heuristic versions of Smith–Waterman: BLAST

- BLAST (Basic Local Alignment Search Technique) is a simplified form of Smith-Waterman local alignment. It is a fast algorithm that is popular because it is faster than SW.
    - BLAST finds regions of similarity between two sequences
    - BLAST uses a database search by scanning for local alignments to sequence pairs that contain an initial word pair match”  
“The central idea of the BLAST algorithm is to confine attention to sequence pairs that contain an initial word pair match”  
Altschul et al. (1990)  
“The search by scanning for local alignments is much faster than the search by scanning for all matches before performing local alignments. The sensitivity in exchange for speed is contrast to SW, BLAST is not guaranteed to find optimal alignments

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## How BLAST works

- Four basic phases
    - Phase 1: compile a list of query word pairs ( $w=3$ )

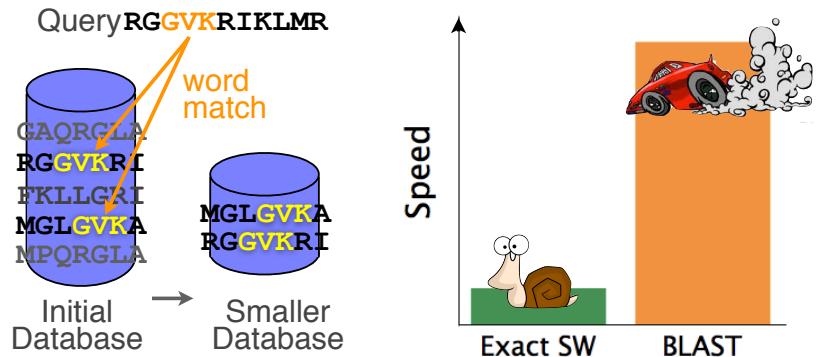
**RGGVKRI**    Query sequence  
**RGG**  
**GGV**  
**GVK**  
**VKR**  
**KRI**

generate list  
of w=3 words  
for query

generate list  
of w=3 words  
for query

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- BLAST uses this pre-screening heuristic approximation resulting in an approach that is about 50 times faster than the Smith-Waterman



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## Blast

- Phase 2: expand word pairs to include those similar to query (defined as those above a similarity threshold to original word, i.e. match scores in substitution matrix)

```

RGGVKRI Query sequence
RGG RAG RIG RLG ...
GGV GAV GTV GCV ...
GVK GAK GIK GGK ...
VKR VRR VHR VER ...
KRI KKI KHI KDI ...

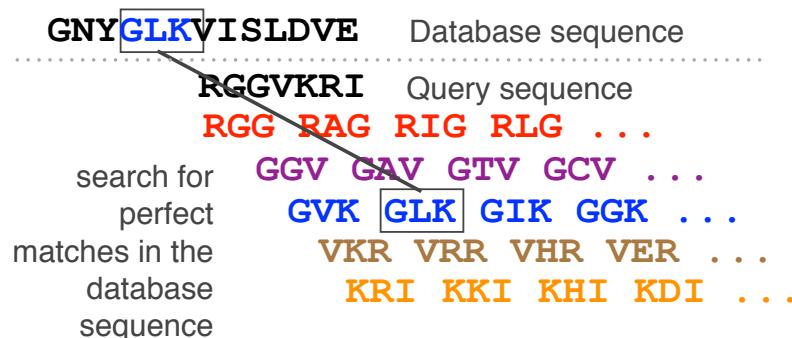
```

extend list of words similar to query

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## Blast

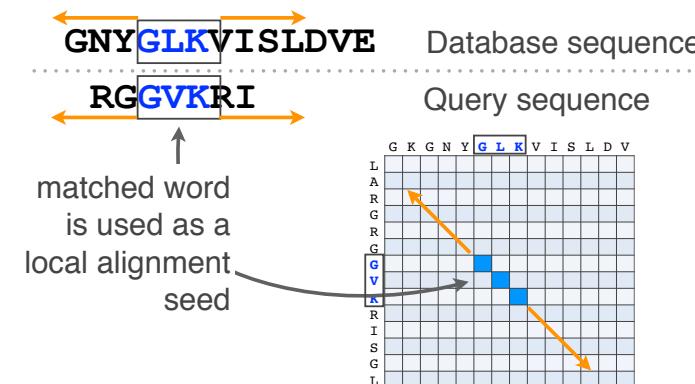
- Phase 3: a database is scanned to find sequence entries that match the compiled word list



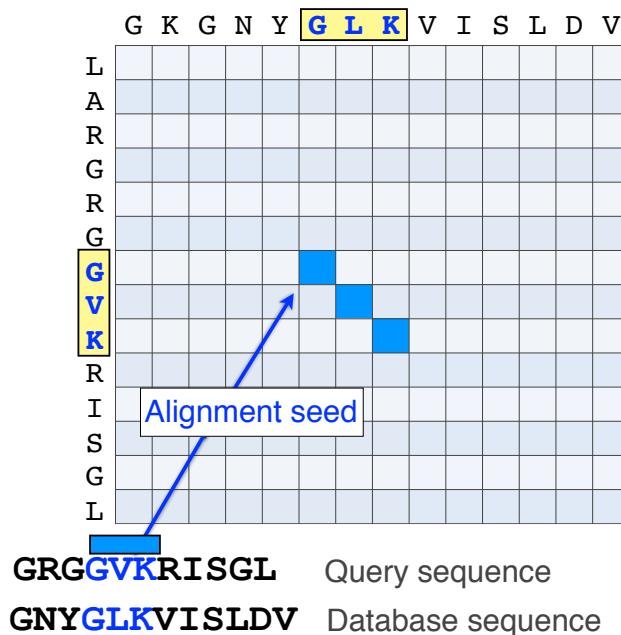
125

## Blast

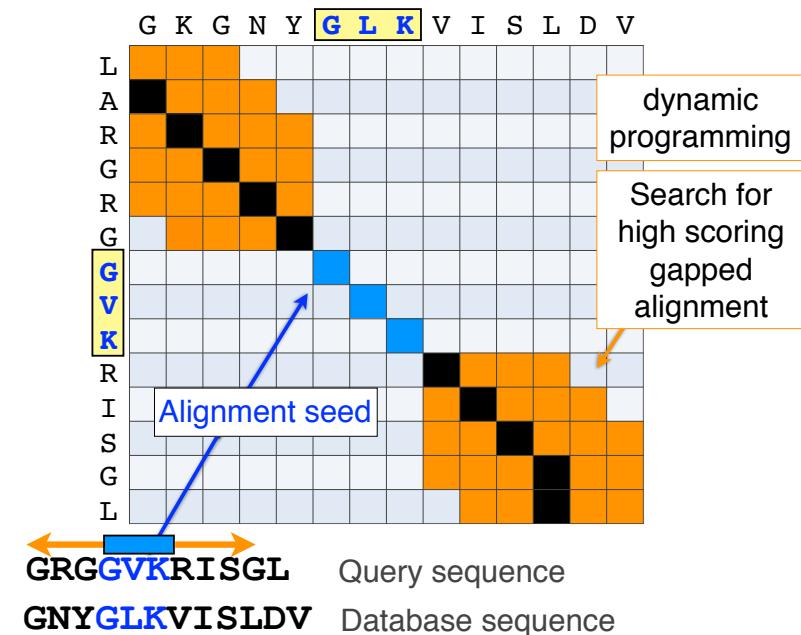
- Phase 4: the initial database hits are extended in both directions using dynamic programming



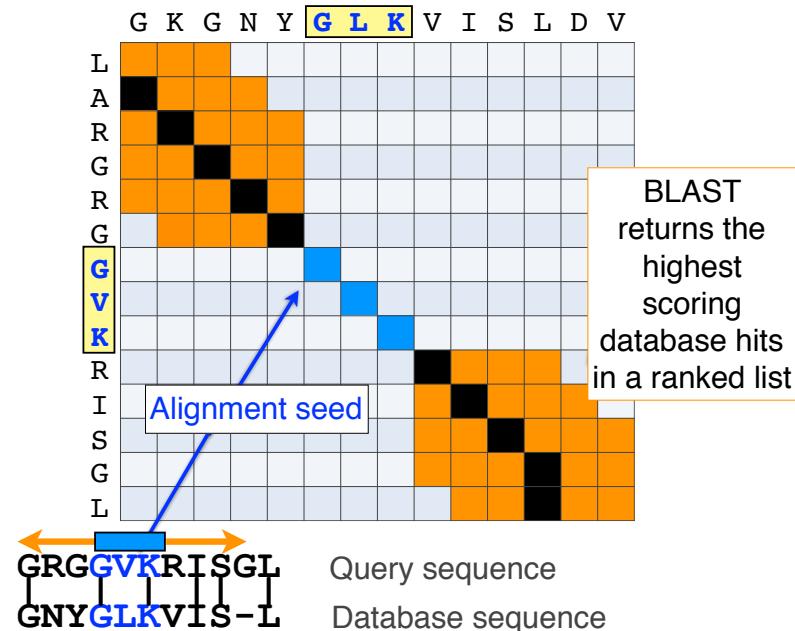
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## BLAST output

- BLAST returns the highest scoring database hits in a ranked list along with details about the target sequence and alignment statistics

Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
Kinesin-14 heavy chain [Danio rerio]	595	88%	0	78%	XP_00320703
hypothetical protein EGK_18589	48.2	40%	0.03	32%	ELK35081.1
mKIAA4102 protein [Mus musculus]	42.7	38%	3.02	24%	EHH28205.1

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## Statistical significance of results

- An important feature of BLAST is the computation of statistical significance for each hit. This is described by the E value (expect value)

Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
Kinesin-14 heavy chain [Danio rerio]	595	88%	0	78%	XP_00320703
hypothetical protein EGK_18589	48.2	40%	0.03	32%	ELK35081.1
mKIAA4102 protein [Mus musculus]	42.7	38%	3.02	24%	EHH28205.1

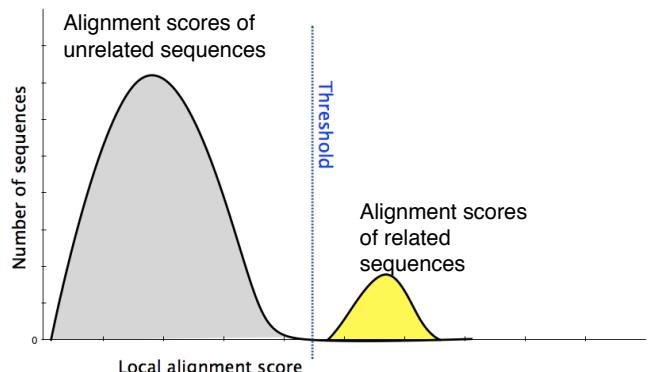
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## BLAST scores and E-values

- The E value is the expected number of hits that are as good or better than the observed local alignment score (with this score or better) if the query and database are random with respect to each other
  - i.e. the number of alignments expected to occur by chance with equivalent or better scores
- Typically, only hits with E value below a significance threshold are reported
  - This is equivalent to selecting alignments with score above a certain score threshold

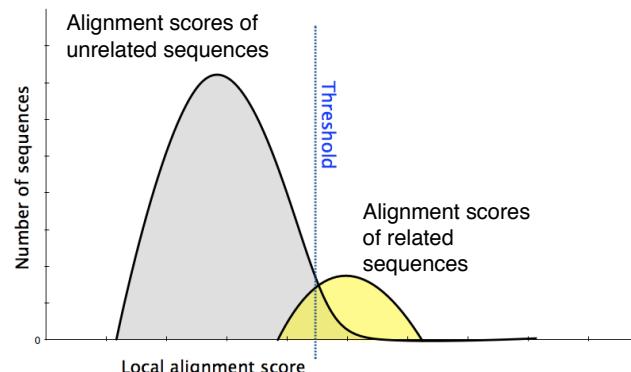
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- Ideally, a threshold separates all query related sequences (yellow) from all unrelated sequences (gray)



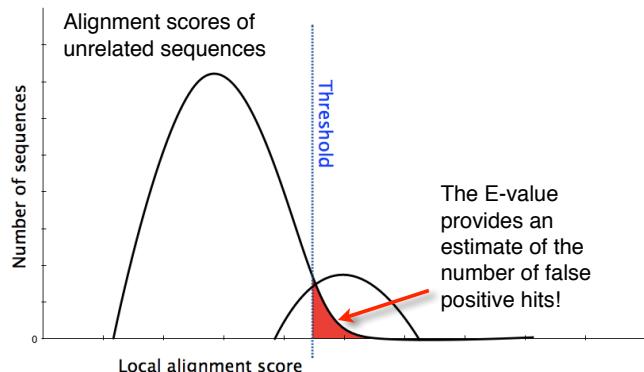
133

- Unfortunately, often both score distributions overlap
  - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



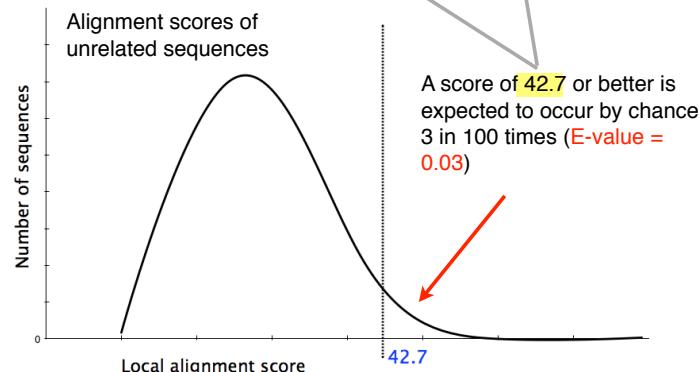
134

- Unfortunately, often both score distributions overlap
  - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



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Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
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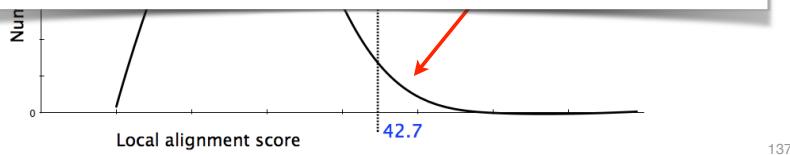
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Description	Max score	Total score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	676	100%	0	98%	AAA20133.1

In general *E* values < 0.005 are usually significant.

To find out more about *E* values see: “*The Statistics of Sequence Similarity Scores*” available in the help section of the NCBI BLAST site:

<http://www.ncbi.nlm.nih.gov/blast/tutorial/Altschul-1.html>



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## Summary

- Sequence alignment is a fundamental operation underlying much of bioinformatics.
- Even when optimal solutions can be obtained they are not necessarily unique or reflective of the biologically correct alignment.
- Dynamic programming is a classic approach for solving the pairwise alignment problem.
- Reviewed global and local alignment approaches and their major application areas.
- Heuristic approaches are often necessary for large database searches and many genomic applications.

## Your Turn!

Hands-on worksheet **Sections 6**

- Please do answer the last lab review question (**Q19**).
- We encourage discussion and exploration!

## FOR NEXT CLASS...

Check out the online:

- ✓ **Reading:** Sean Eddy’s “What is dynamic programming?”
- ✓ **Homework:** (1) [Quiz](#), (2) [Alignment Exercise](#).

## Homework Grading

Both (1) quiz questions and (2) alignment exercise carry equal weights (i.e. 50% each).

(Homework 2) Assessment Criteria	Points
Setup labeled alignment matrix	1
Include initial column and row for <b>GAPs</b>	1
All alignment matrix elements <b>scored</b> (i.e. filled in)	1
Evidence for correct use of <b>scoring scheme</b>	1
<b>Direction arrows</b> drawn between all cells	1
Evidence of multiple arrows to a given cell if appropriate	1
Correct <b>optimal score</b> position in matrix used	1
Correct optimal score obtained for given scoring scheme	1
<b>Traceback path(s)</b> clearly highlighted	1
Correct <b>alignment(s)</b> yielding optimal score listed	1
	A+

## REFERENCE SLIDES...

Additional reference slides for the motivated student

## Practical database searching with BLAST

- There are four basic components to a traditional BLAST search
  - (1) Choose the sequence (query)
  - (2) Select the BLAST program
  - (3) Choose the database to search
  - (4) Choose optional parameters
- Then click “BLAST”

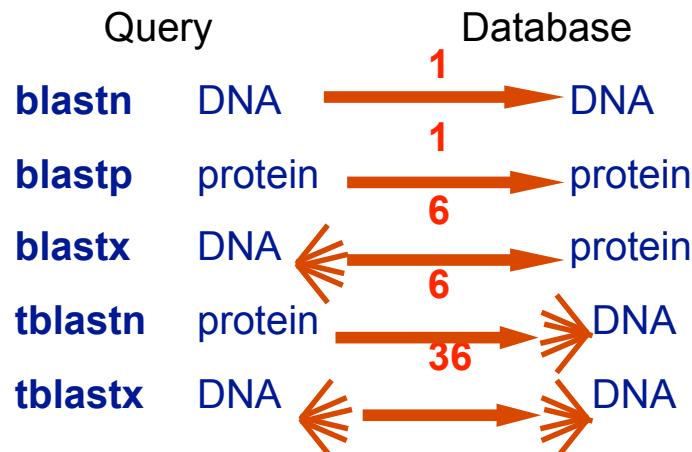
## Step 1: Choose your sequence

- Sequence can be input in FASTA format or as accession number

The screenshot shows the NCBI Protein search interface. The search bar is set to "Protein". The "Display Settings" dropdown is set to "FASTA". The search results for "hemoglobin subunit beta [Homo sapiens]" are shown, with the NCBI Reference Sequence (NP\_000509.1) highlighted. The sequence itself is displayed below.

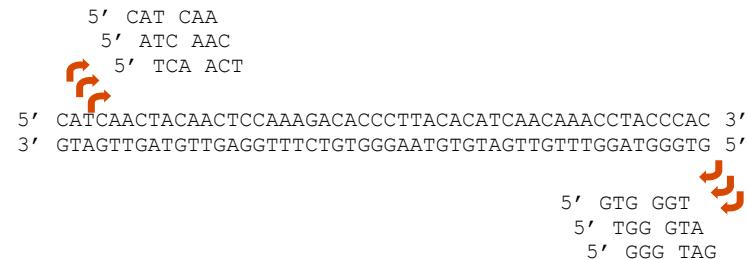
```
>gi|4504349|ref|NP_000509.1| hemoglobin subunit beta [Homo sapiens]
MVHLTPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFESFGDLSTPDAMGNPKVKAHGKVKVLG
AFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGNGVLVCVLAAHFGKEFTPPVQAYQKVVAGVAN
ALAHKYH
```

## Step 2: Choose the BLAST program



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## DNA potentially encodes six proteins



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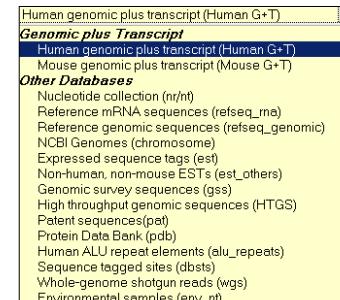
The screenshot shows the Protein BLAST search interface with the following highlights:

- Enter Query Sequence:** A grey arrow points to the input field containing the protein sequence: >gi|4504349|ref|NP\_000509.1| hemoglobin subunit beta [Homo sapiens] MVHLPEEKSAVTALWGKVNVDEVGGEALGRLLVYPWPTORFESFGDLSPTDAVMGNPKVKAHGCK KVLAGFSQGLAHLDNLKGTFATLSELHCDKLHVDPNFRLLGVLCVLAHHFGEKTPVQAAYQK VVAGVANALAHKYH.
- Choose Search Set:** An orange arrow points to the "Database" dropdown menu set to "Non-redundant protein sequences (nr)".
- Program Selection:** A red arrow points to the "Algorithm" section where "blastp (protein-protein BLAST)" is selected.
- BLAST Button:** A large red arrow points to the "BLAST" button at the bottom left.

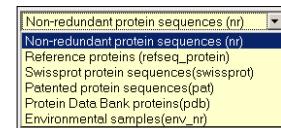
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## Step 3: Choose the database

**nr** = non-redundant (most general database)  
**dbest** = database of expressed sequence tags  
**dbsts** = database of sequence tag sites  
**gss** = genomic survey sequences



nucleotide databases



protein databases

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Protein BLAST: search protein databases using a protein query

Enter Query Sequence

Enter accession number(s), g(s), or FASTA sequence(s)  Clear

>gi|4504349|ref|NP\_000509.1| hemoglobin subunit beta [Homo sapiens]  
MVLHTREKSAVTALWCKVNDEVCGEALGRLLVVPPWTQRFESFGDLSTPDAVMCNPKVKAHK  
KVLCAFSFDLALHDNLKGCTATSELICDKLHVDPENRLLGNVLVCLAHHFKEETPPVQAAYQX  
VVAGVANALAHKYH

Or, upload file  no file selected

Job Title

Enter a descriptive title for your BLAST search

Align two or more sequences

Choose Search Set

Database Non-redundant protein sequences (nr)  Exclude

Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown.

Organism Optional

Exclude Optional

Entrez Query Optional

Enter an Entrez query to limit search

Program Selection

Algorithm

blastp (protein-protein BLAST)  
 PSI-BLAST (Position-Specific Iterated BLAST)  
 PHI-BLAST (Pattern Hit Initiated BLAST)  
 DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST)

Choose a BLAST algorithm

**BLAST** Search database Non-redundant protein sequences (nr) using Blastp (protein-protein BLAST)  
 Show results in a new window

**Algorithm parameters**

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## Step 4a: Select optional search parameters

Algorithm parameters

General Parameters

Max target sequences  Select the maximum number of aligned sequences to display

Short queries  Automatically adjust parameters for short input sequences

Expect threshold  **Expect**

Word size  **Word size**

Max matches in a query range

Scoring Parameters

Matrix BLOSUM62 **Scoring matrix**

Gap Costs Existence: 11 Extension: 1

Compositional adjustments Conditional compositional score matrix adjustment

Filters and Masking

Filter Low complexity regions

Mask Mask for lookup table only  
Mask lower case letters

**BLAST** Search database Non-redundant protein sequences (nr) using Blastp  
 Show results in a new window

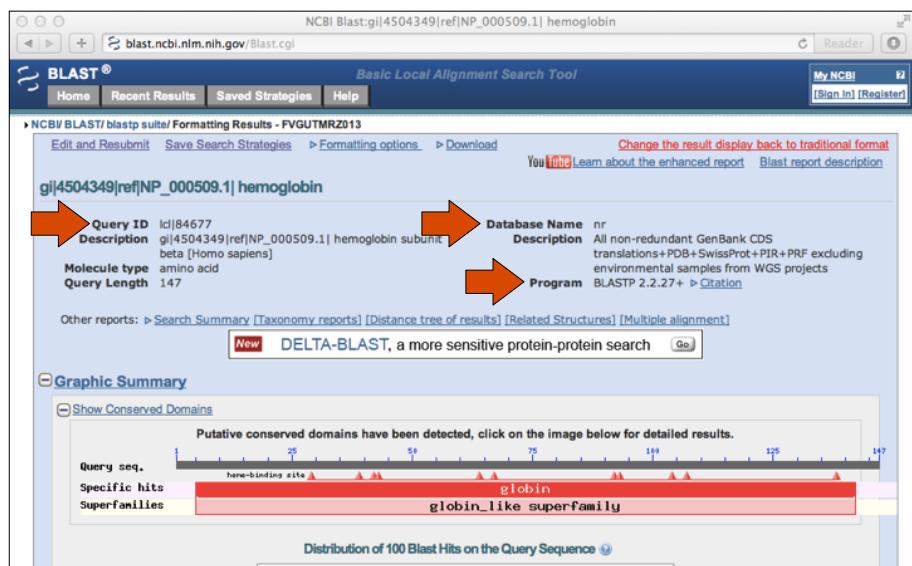
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## Step 4: Optional parameters

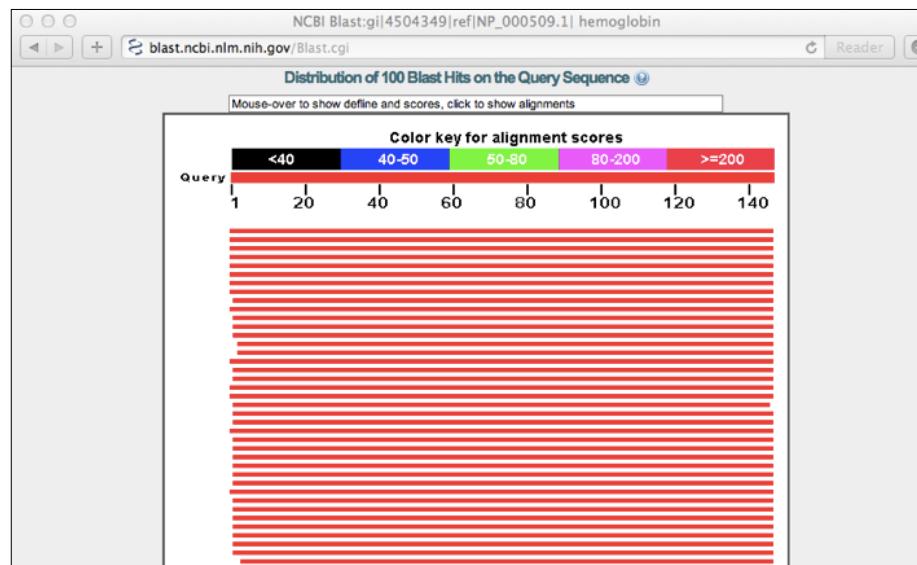
- You can...
  - choose the organism to search
  - change the substitution matrix
  - change the expect (E) value
  - change the word size
  - change the output format

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## Results page



## Further down the results page...



## Further down the results page...

NCBI Blast:gi|4504349|ref|NP\_000509.1| hemoglobin

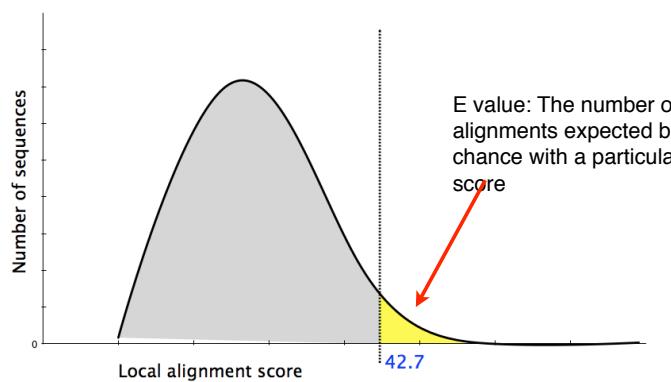
blast.ncbi.nlm.nih.gov/Blast.cgi Reader

Sequences producing significant alignments:

Select: All None Selected:0

Description	Max score	Total score	Query cover	E value	Max ident	Accession
hemoglobin beta [synthetic construct]	301	301	100%	9e-103	100%	AAX37051.1
hemoglobin beta [synthetic construct]	301	301	100%	1e-102	100%	AAX29557.1
hemoglobin subunit beta [Homo sapiens] >ref XP_508242.1  PREDICTED: hemoglobin_s	301	301	100%	1e-102	100%	NP_000509.1
RecName: Full=Hemoglobin subunit beta; AltName: Full=Beta-globin; AltName: Full=Hem	300	300	100%	4e-102	99%	P02024.2
beta globin chain variant [Homo sapiens]	299	299	100%	5e-102	99%	AAN84548.1
beta globin [Homo sapiens] >gb AAZ39781.1  beta globin [Homo sapiens] >gb AAZ39782	299	299	100%	5e-102	99%	AAZ39780.1
beta-globin [Homo sapiens]	299	299	100%	5e-102	99%	ACU56984.1
hemoglobin beta chain [Homo sapiens]	299	299	100%	6e-102	99%	AAD19696.1
Chain B. Structure Of Haemoglobin In The Deoxy Quaternary State With Ligand Bound A	298	298	99%	9e-102	100%	1COH_B
hemoglobin beta subunit variant [Homo sapiens] >gb AAA88054.1  beta-globin [Homo sa	298	298	100%	1e-101	99%	AAF00489.1
Chain B. Human Hemoglobin D Los Angeles: Crystal Structure >pdb 2YRS D.Chain D_H	298	298	99%	2e-101	99%	2YRS_B
Chain B. High-Resolution X-Ray Study Of Deoxy Recombinant Human Hemoglobins Syn	297	297	99%	3e-101	99%	1DXU_B
Chain B. Analysis Of The Crystal Structure, Molecular Modeling And Infrared Spectroscop	297	297	99%	3e-101	99%	1HDB_B

Description	Max score	Total score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo	677	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	676	100%	0	98%	AAA20133.1
Kinesin-14 heavy chain [Danio rerio]	595	595	88%	0	78%	XP_00320703
hypothetical protein EGK_18589	48.2	52	40%	0.03	32%	ELK35081.1
mKIAA4102 protein [Mus musculus]	42.7	42.7	38%	3.02	24%	EHH28205.1



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## E values in BLAST

- Each alignment gets a score determined from the alignment and doesn't take into account the full length of the query, target or database
- The E value is what you want to look at
- E value = Expect
  - How often do I expect an alignment with this score given the length of my query and the size of the database
  - $E = Kmne^{-\lambda s}$ 
    - K and  $\lambda$  are scaling factors
    - S is the score
    - m – length of query, n – length of database
  - E corrects for multiple comparisons, i.e., query compared to many sequences – proportional to length of database and query for a given S (score)

Further down the results page...

The screenshot shows two separate search results from the NCBI Blast search interface. Both results are for the same query sequence, "hemoglobin subunit beta [Homo sapiens]".

**Result 1:** Sequence ID: ref|NP\_000509.1, Length: 147, Number of Matches: 1. The alignment shows a single hit with an expect value of 1e-102. The alignment details are as follows:

Range	Score	Expect	Method	Identities	Positives	Gaps
1 to 147	301 bits(770)	1e-102	Compositional matrix adjust.	147/147(100%)	147/147(100%)	0/147(0%)

**Result 2:** RecName: Full=Hemoglobin subunit beta; AltName: Full=Beta-globin; AltName: Full=Hemoglobin beta chain. Sequence ID: sp|P02024.2|HBB\_GORG, Length: 147, Number of Matches: 1. The alignment shows a single hit with an expect value of 4e-102. The alignment details are as follows:

Range	Score	Expect	Method	Identities	Positives	Gaps
1 to 147	300 bits(767)	4e-102	Compositional matrix adjust.	146/147(99%)	147/147(100%)	0/147(0%)

Different output formats are available

The screenshot shows the "Formatting options" section of the NCBI BLAST search results page. This section allows users to customize how search results are displayed. A red circle highlights the "Formatting options" link in the top navigation bar.

**Formatting options:**

- Show: Alignment as (HTML or Old View) - HTML is selected.
- Alignment View: Query-anchored with letters for identities.
- Display: Graphical Overview (selected), Sequence Retrieval, NCBI-gi.
- Masking: Character (Lower Case or Upper Case), Color (Grey or Black).
- Limit results: Descriptions (50), Graphical overview (50), Alignments (50).
- Organism: Type common name, binomial, taxid, or group name. Only 20 top taxa will be shown.
- Entrez query: Text input field.
- Expect Min: Text input field.
- Expect Max: Text input field.
- Percent Identity Min: Text input field.
- Percent Identity Max: Text input field.
- Format for: PSI-BLAST (selected) with inclusion threshold.

E.g. Query anchored alignments

The screenshot shows a list of query sequences (Query) and their corresponding alignment scores. The alignment score for each query is 59, indicating a high similarity to the reference sequence.

Query	Score
AAX37051	59
AAX29557	59
NP_000509	59
P02024	59
AAN84548	59
AAZ39780	59
ACU56984	59
AAD19696	59
Icoh_B	59
AAF00489	59
ZYRS_B	59
IDXU_B	59
1HDB_B	59
IDKV_B	59
JKMF_C	59
AAL68978	59
INOP_B	59
I1K1K_B	59
AAAN11320	59
XP_002822173	59
IYB5_B	59
IYE0_B	59
I1O10_B	59
CAA23759	59
IYE2_B	59
IY5F_B	59
I1A00_B	59
IHB5_B	59
IABY_B	59
I1CMY_B	59

... and alignments with dots for identities

The screenshot shows the same list of query sequences as the previous screenshot, but the alignment is displayed using dots to represent identities. Dots are placed at positions where the query sequence matches the reference sequence. The alignment score for each query is 60, indicating a perfect match.

Query	Score
AAX37051	60
AAX29557	60
NP_000509	60
P02024	60
AAN84548	60
AAZ39780	60
ACU56984	60
AAD19696	60
Icoh_B	60
AAF00489	60
ZYRS_B	60
IDXU_B	60
1HDB_B	60
IDKV_B	60
JKMF_C	60
AAL68978	60
INOP_B	60
I1K1K_B	60
AAAN11320	60
XP_002822173	60
IYB5_B	60
IYE0_B	60
I1O10_B	60
CAA23759	60
IYE2_B	60
IY5F_B	60
I1A00_B	60

## Common problems

- Selecting the wrong version of BLAST
- Selecting the wrong database
- Too many hits returned
- Too few hits returned
- Unclear about the significance of a particular result - are these sequences homologous?

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## How to handle too many results

- Focus on the question you are trying to answer
  - select “refseq” database to eliminate redundant matches from “nr”
  - Limit hits by organism
  - Use just a portion of the query sequence, when appropriate
  - Adjust the expect value; lowering E will reduce the number of matches returned

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## How to handle too few results

- Many genes and proteins have no significant database matches
  - remove Entrez limits
  - raise E-value threshold
  - search different databases
  - try scoring matrices with lower BLOSUM values (or higher PAM values)
  - use a search algorithm that is more sensitive than BLAST (e.g. PSI-BLAST or HMMer)

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